

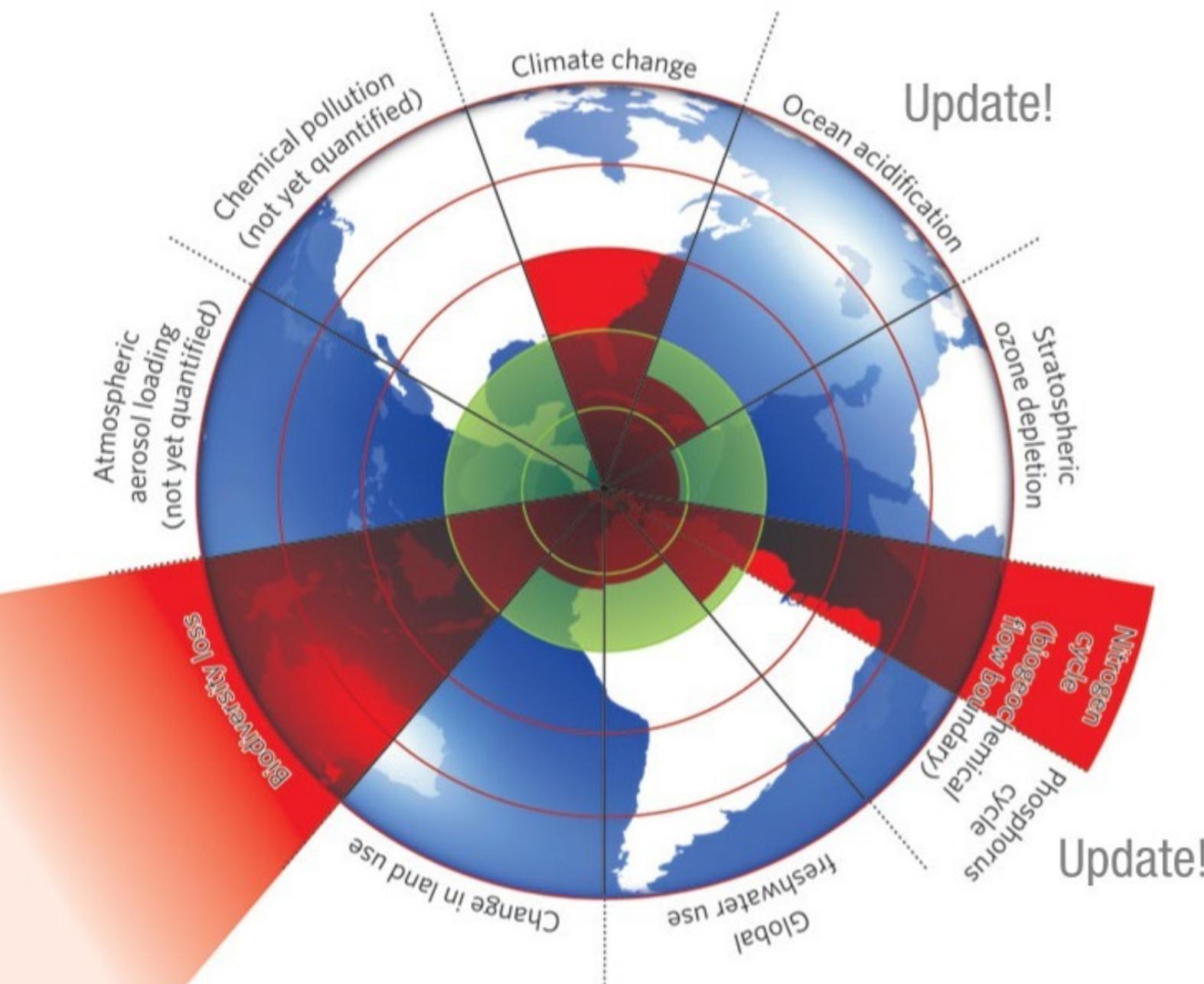
Sequencing technologies & metagenomics

| bioinformatics progresses & challenges

Frédéric Mahé
Nov. 17, 2016
Ouagadougou

Planetary boundaries and human development

Rockström et al. (2009) Nature 461 and Steffen et al., (2015) Science 347:6223



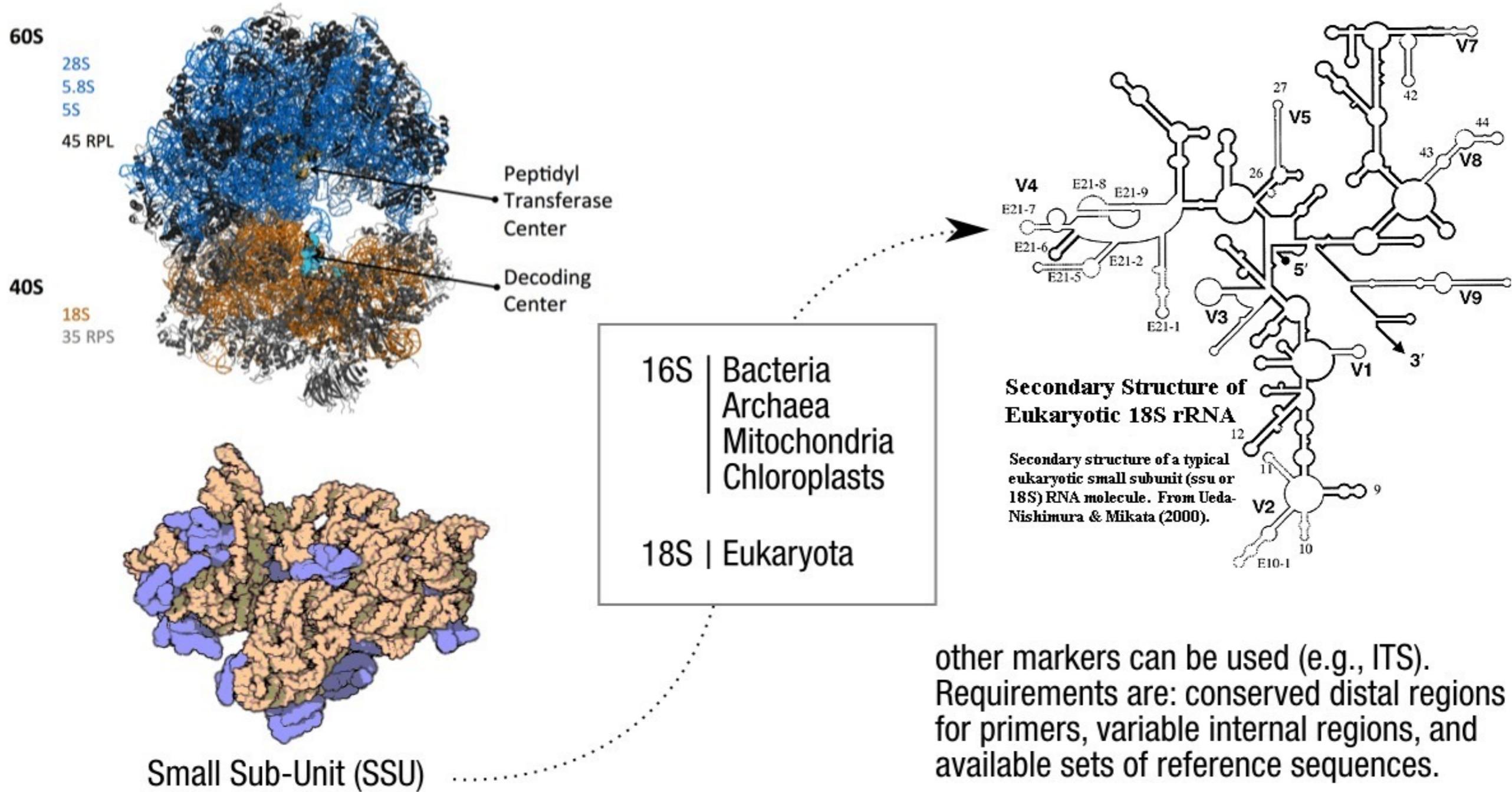
Biodiversity loss (genetic and functional) is a major short-term threat

How can we measure biodiversity?



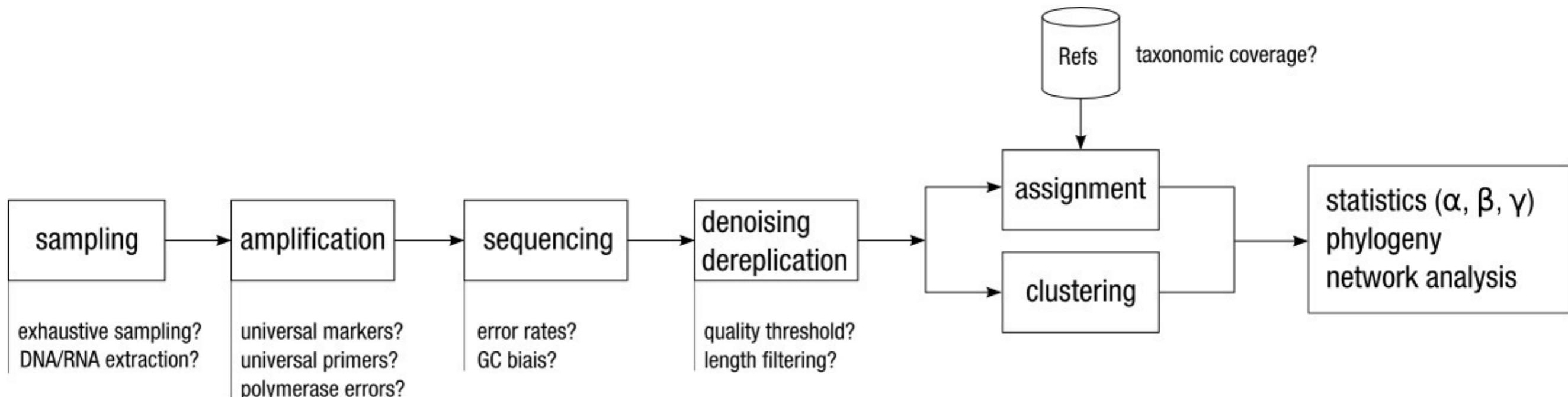
Small organisms play a major role but are hard to inventory

A universal gene: ribosomal RNA



Environmental Metagenomics

targeted amplification



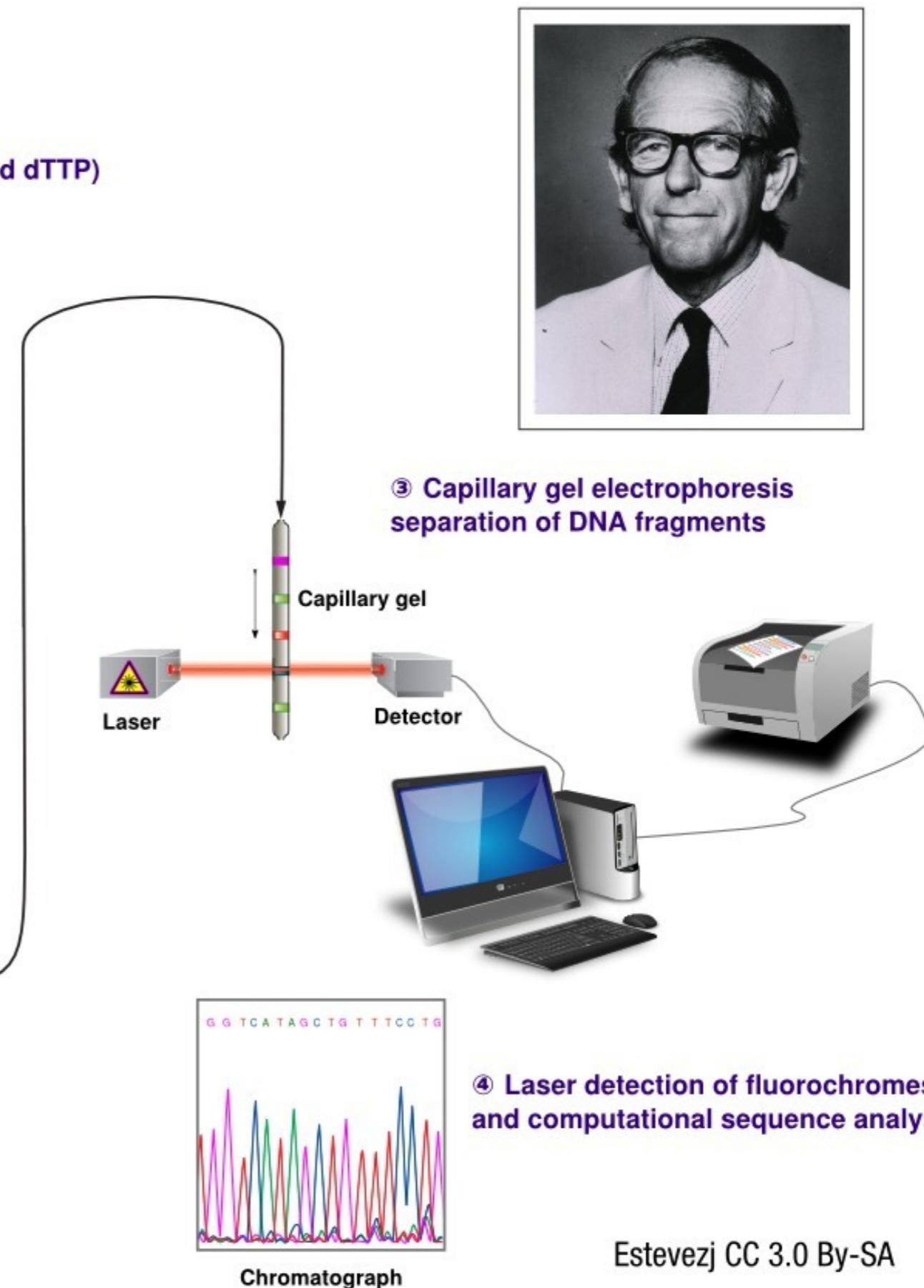
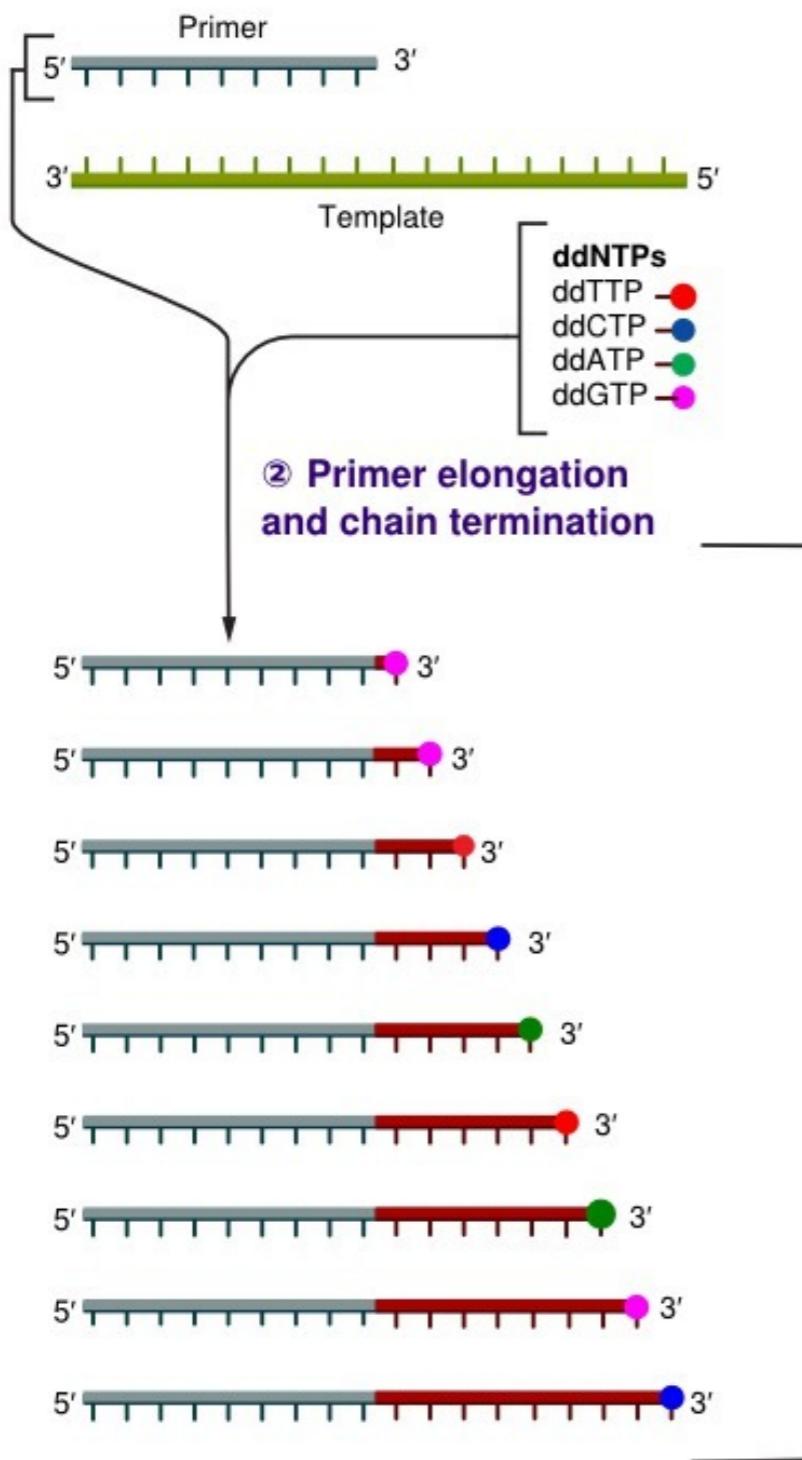
chemistry, physics and randomness

bioinformatics
(preparation)

biostatistics
(actual analyses)

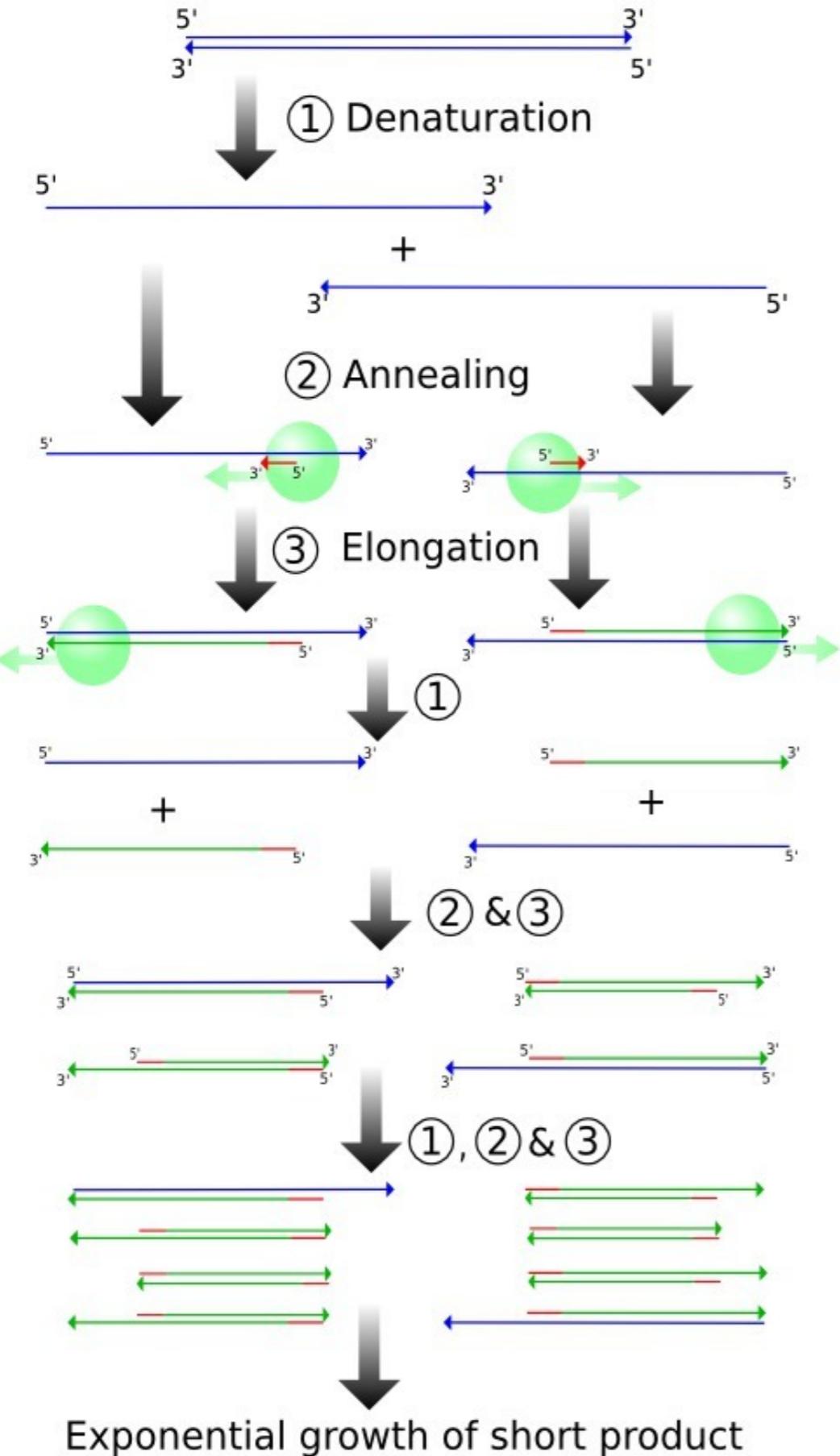
Sanger sequencing technique (1977)

- ① Reaction mixture
- Primer and DNA template ► DNA polymerase
 - ddNTPs with fluorochromes ► dNTPs (dATP, dCTP, dGTP, and dTTP)



Estevezj CC 3.0 By-SA

Polymerase chain reaction (1983)

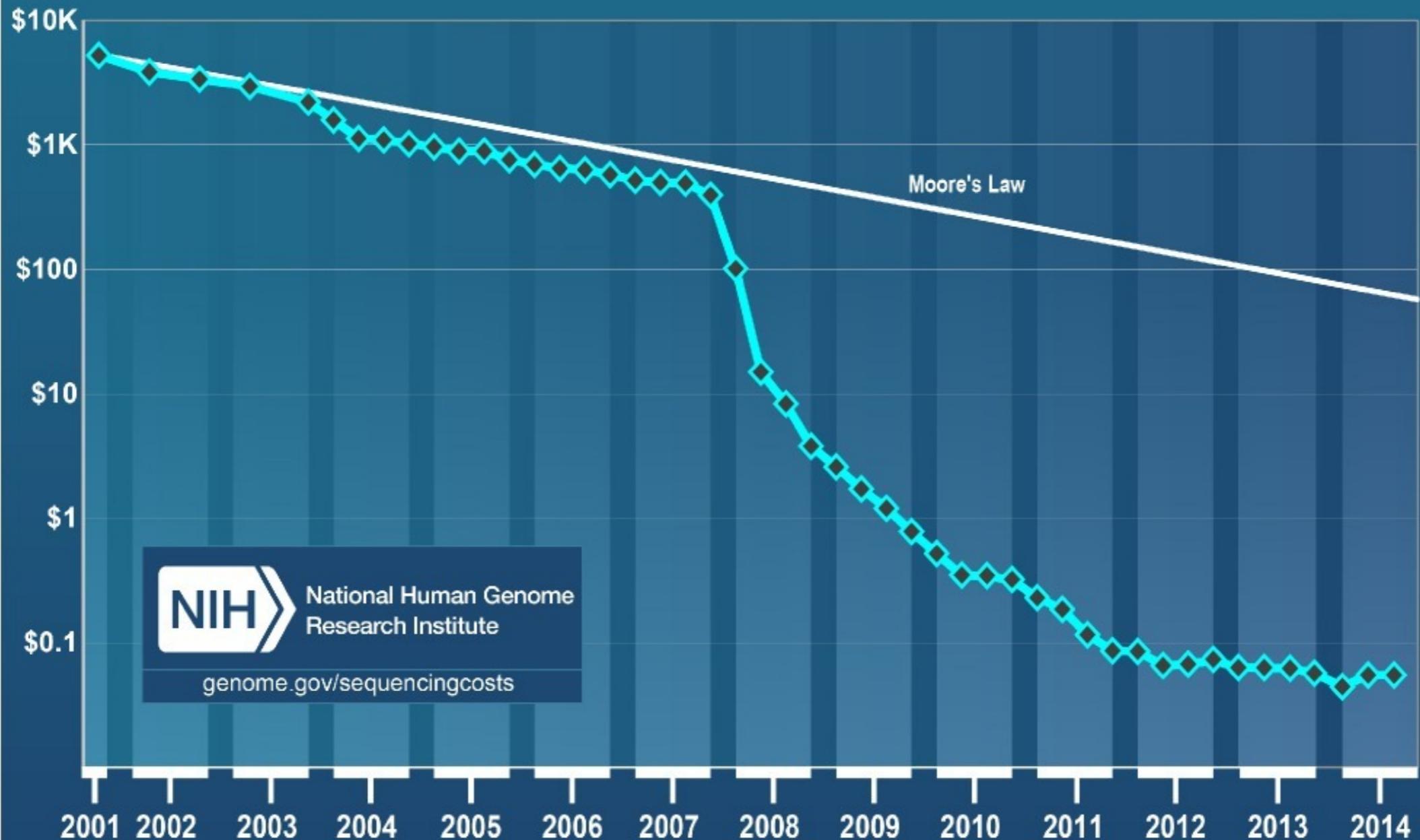


Kary Mullis (Nobel Prize 1993)



Yikrazuul CC 3.0 By-SA

Cost per Raw Megabase of DNA Sequence

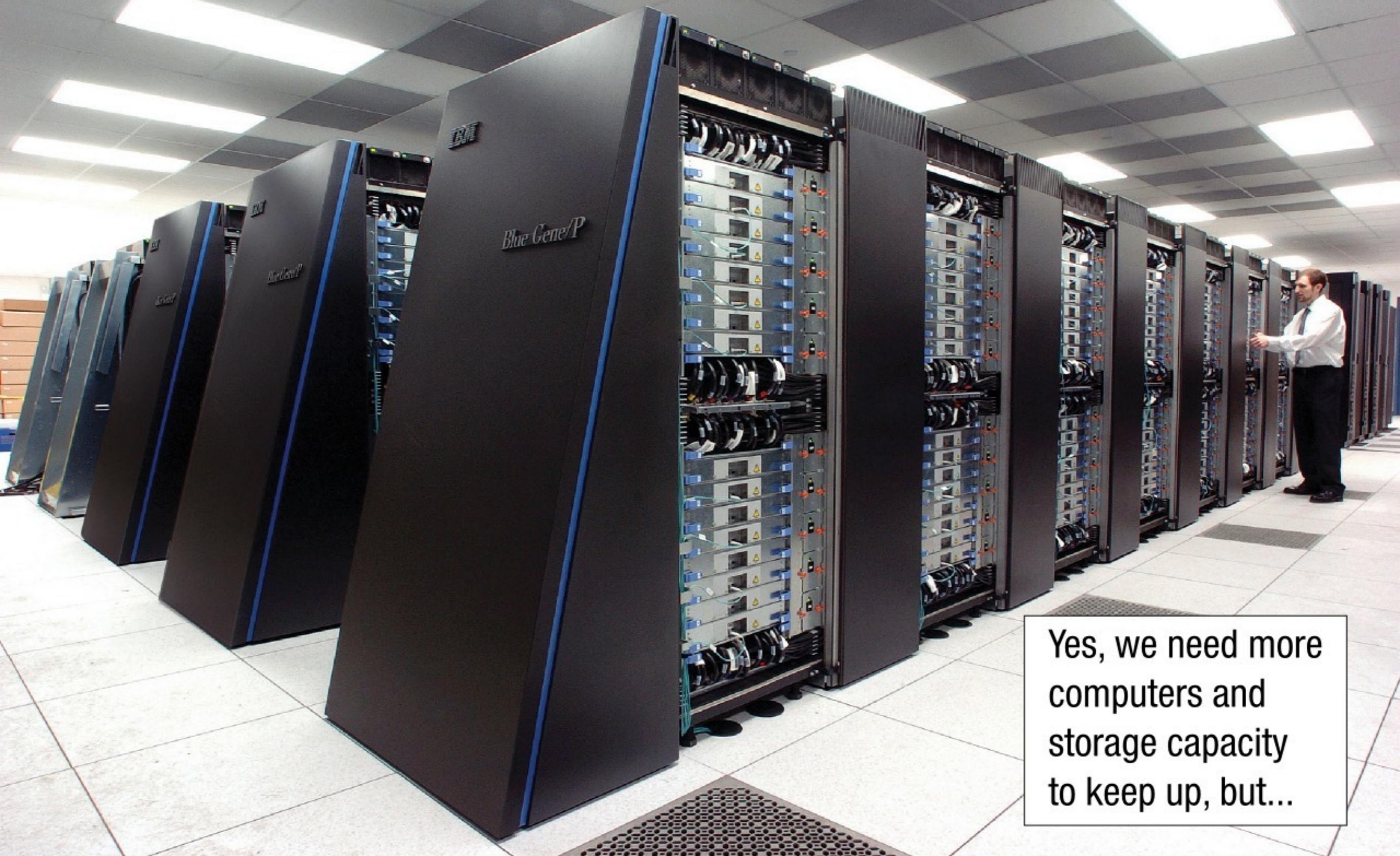


IonTorrent
Ion PGM, Ion Proton

Illumina
HiSeq 3000/4000, MiSeq 3000
allows lots of samples and deep sequencing

Pacific Biosciences

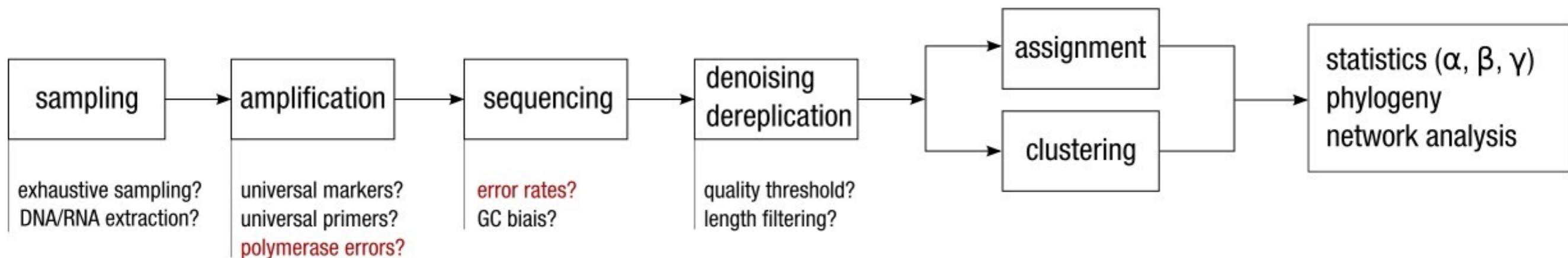
Oxford Nanopore Technologies



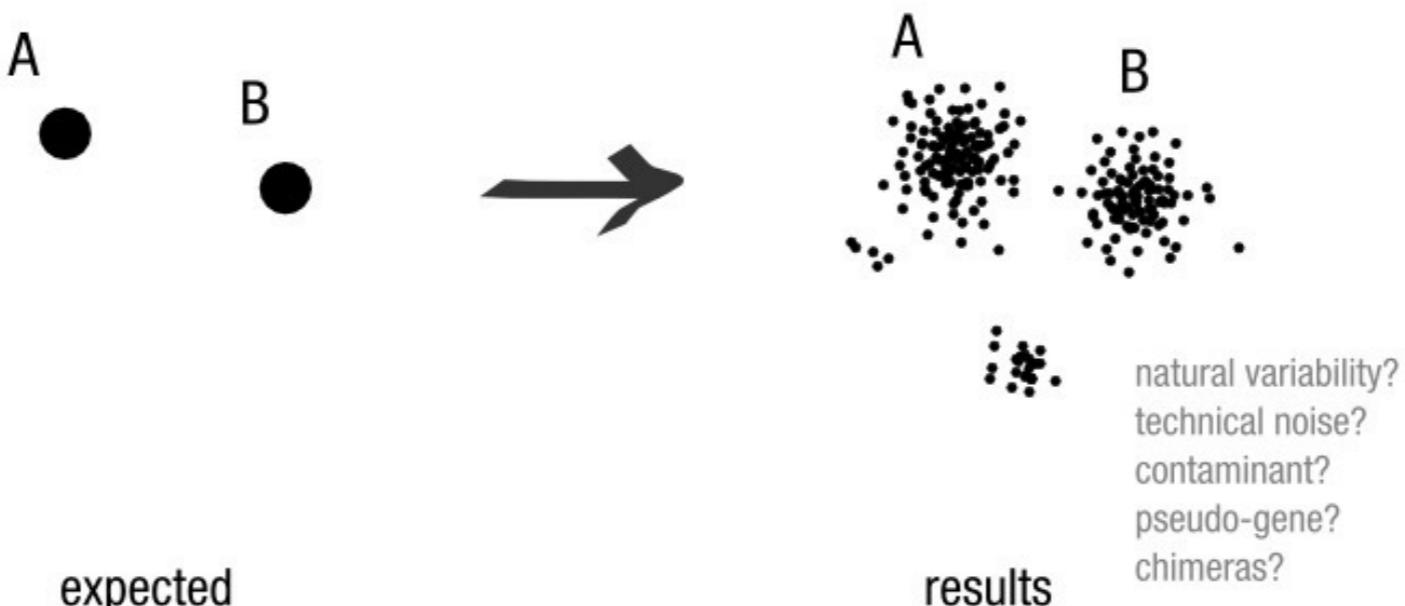
Yes, we need more
computers and
storage capacity
to keep up, but...

Noise is the real challenge

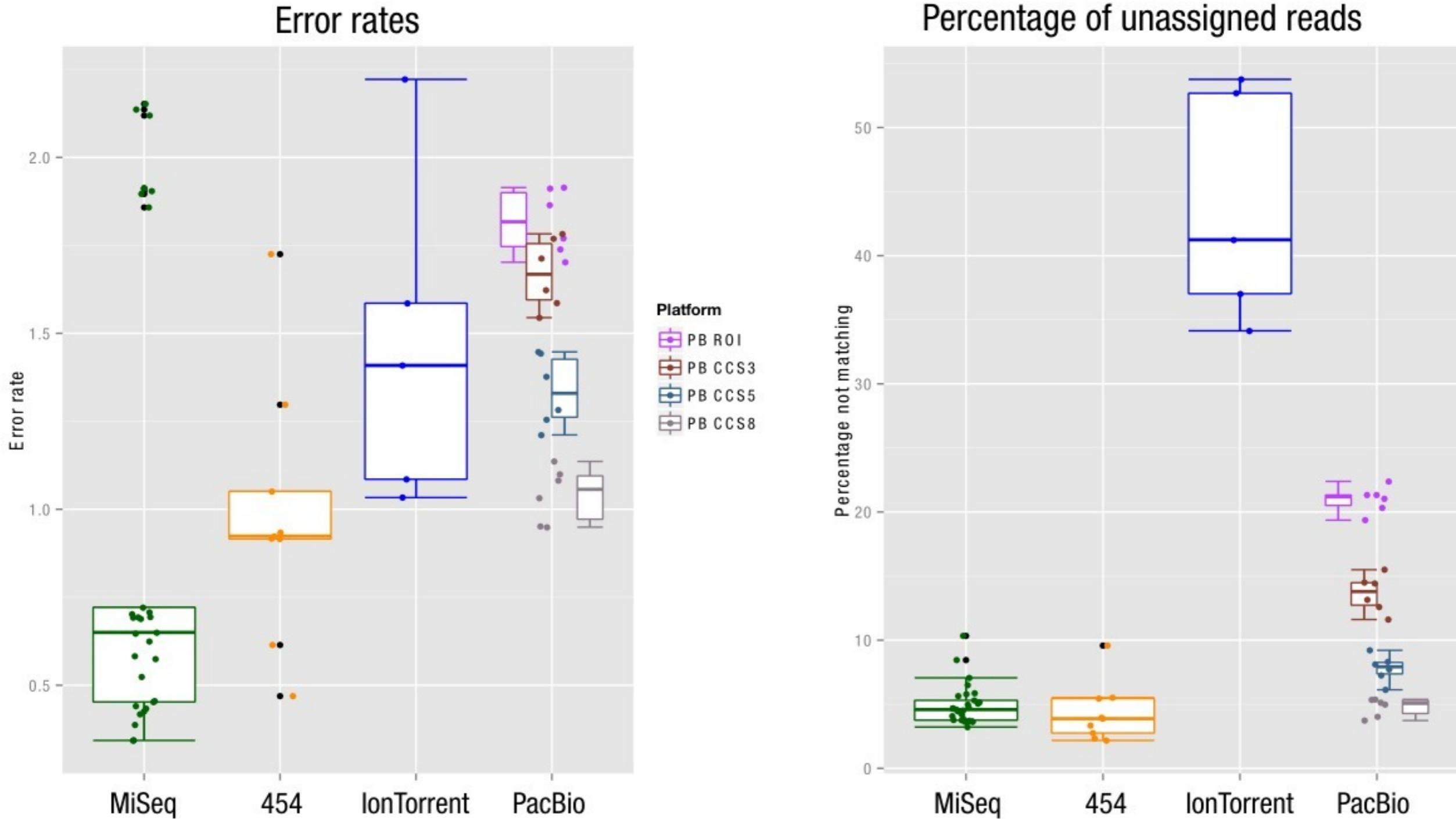
amplification and sequencing are imperfect processes



chemistry, physics and randomness



Error rate per sequencing platform



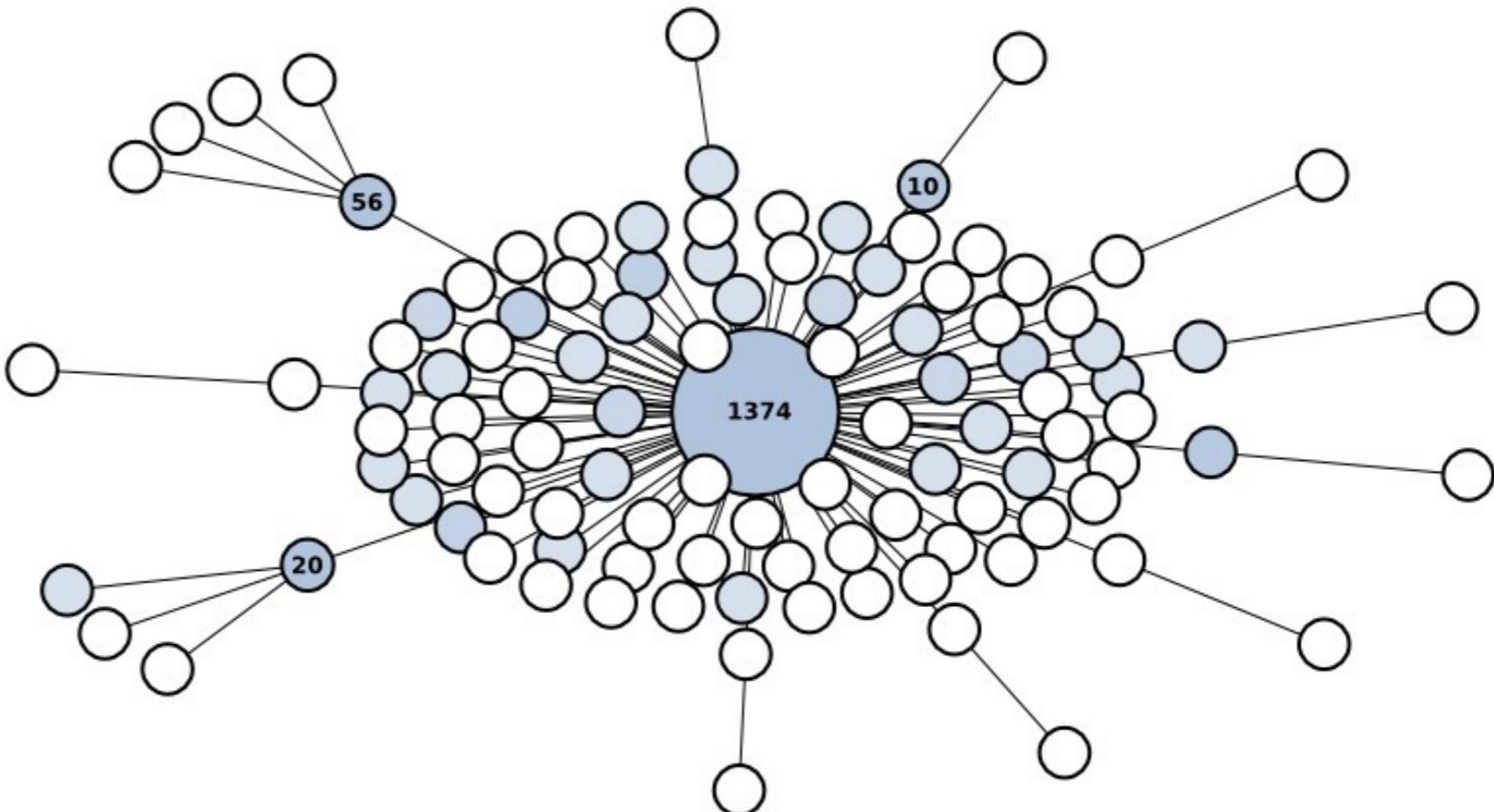
What is the nature of these errors?

2
ACGT
AAGT
AGGT
ATGT
A-GT
AACGT
...

At each position in a sequence:

- 3 substitutions (C → A, G or T)
- 1 deletion,
- 4 insertions

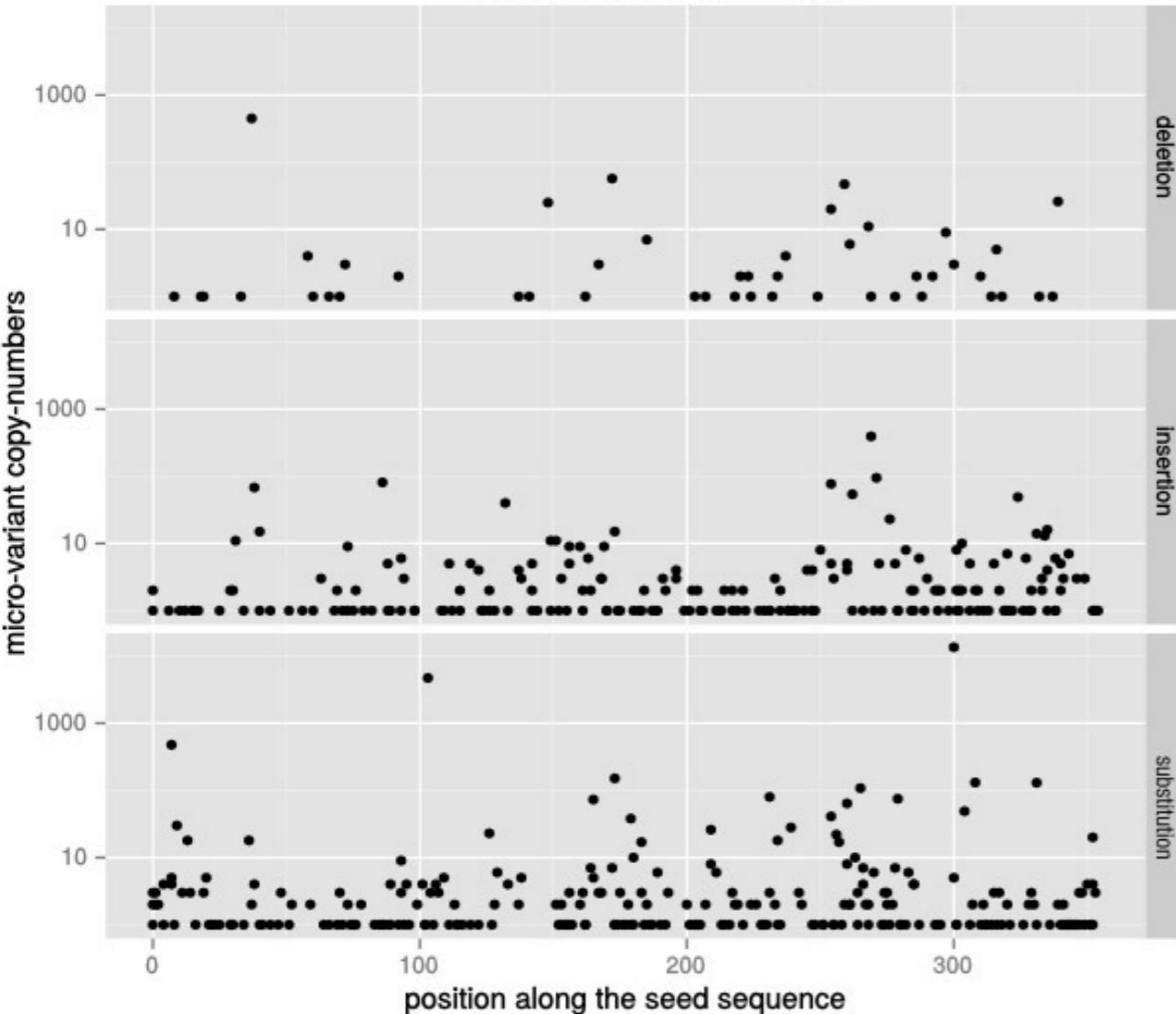
A 380 nucleotide seed (V4), can have up to 2,664 first degree unique micro-variants.



A 130 nucleotide seed (V9), can have up to 914 first degree unique micro-variants.

How are errors distributed?

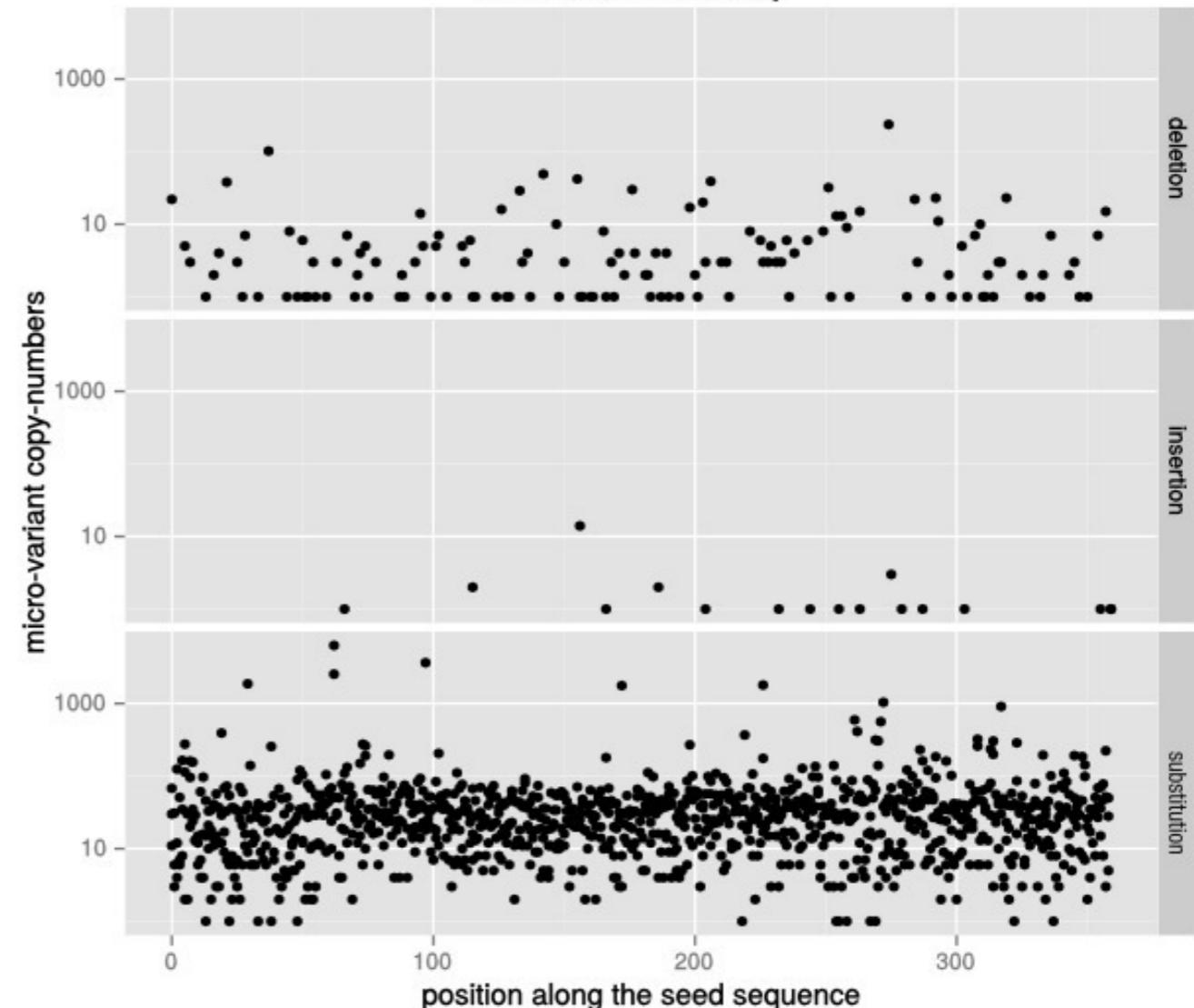
Roche 454 Titanium



Roche 454 Titanium (for 100 bp):

- 0.4011 insertions/deletions,
- 0.0543 substitutions.

Illumina MiSeq

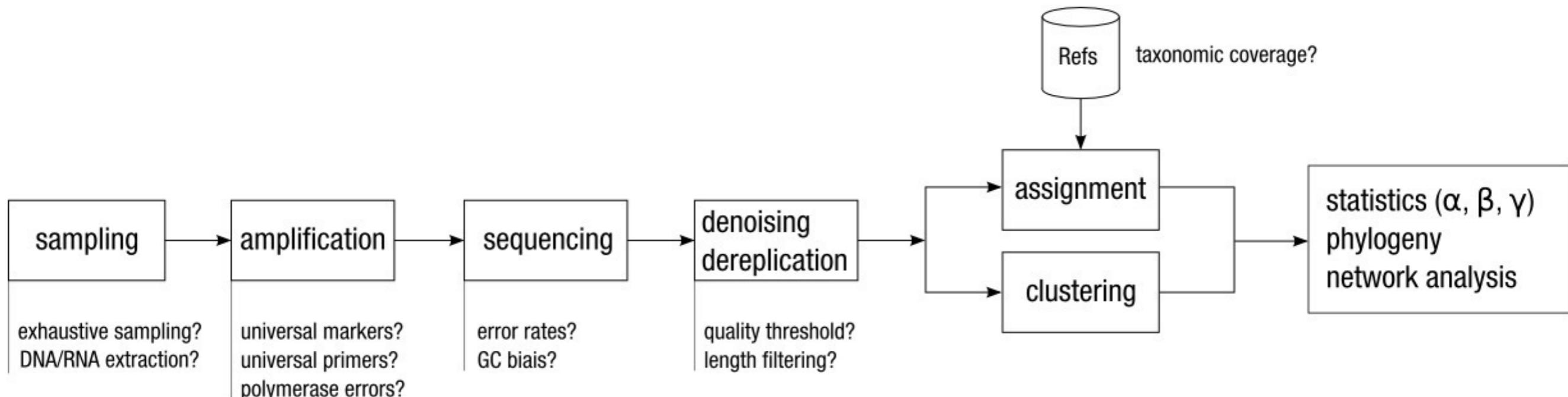


Illumina MiSeq v2 (for 100 bp):

- 0.0009 insertions/deletions,
- 0.0940 substitutions.

Environmental Metagenomics

targeted amplification



chemistry, physics and randomness

bioinformatics
(preparation)

biostatistics
(actual analyses)

Major metagenomics pipelines

targeted amplification



Mothur (2009)
Patrick Schloss
open-source
single piece
most cited
stats

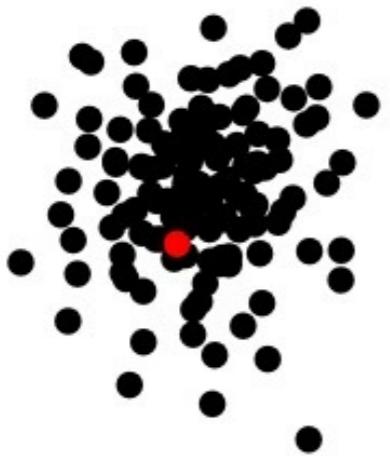


Qiime (2010)
Gregory Caporaso
open-source
python wrapper
most used(?)
stats

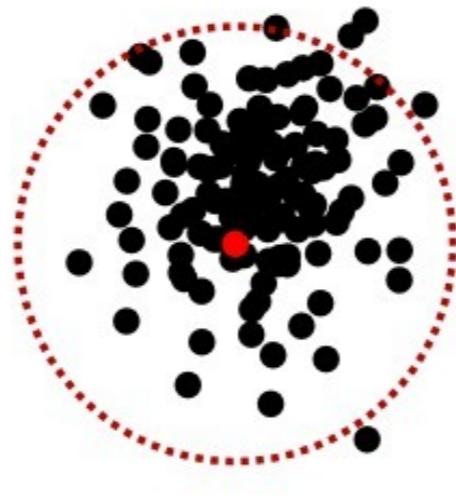
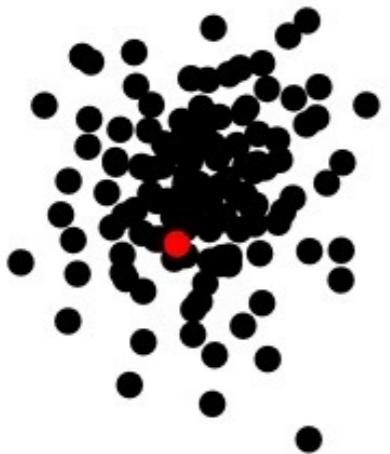
Uparse

Uparse (2013)
Robert Edgar
closed-source
usearch commands
popular
no stats

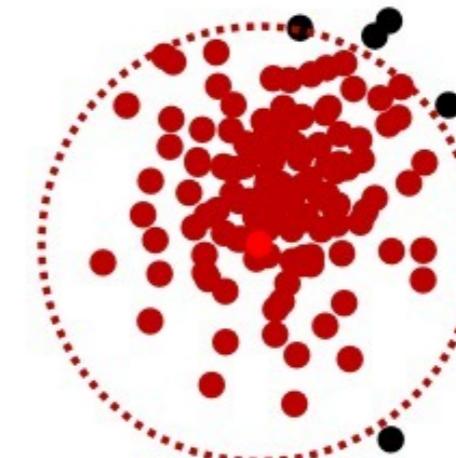
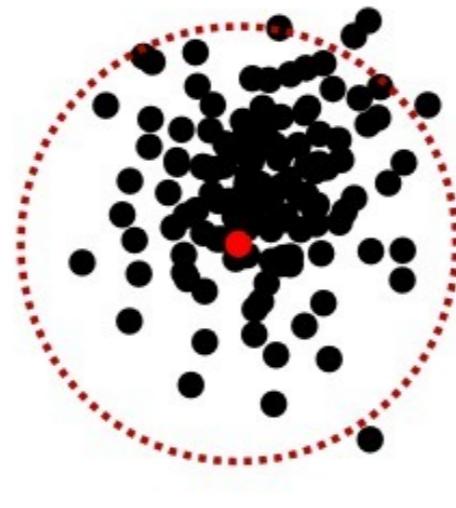
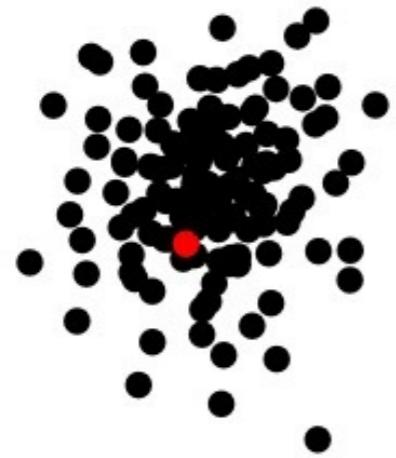
How traditional clustering works?



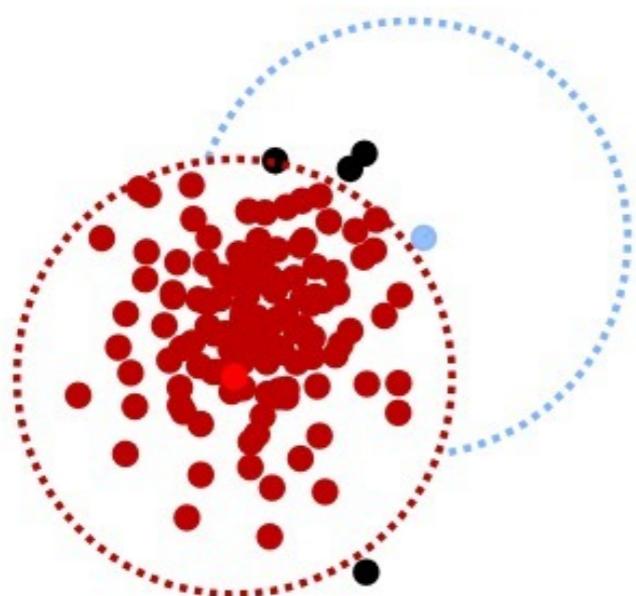
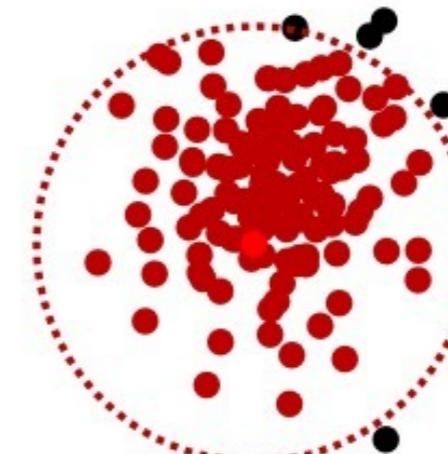
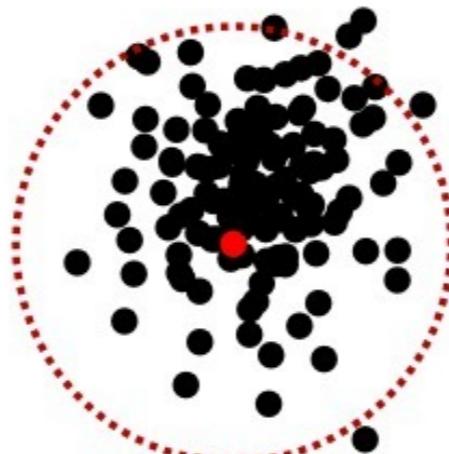
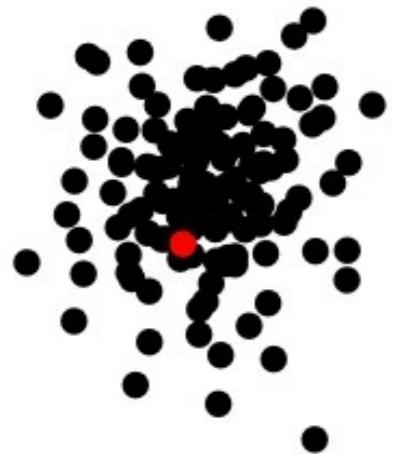
How traditional clustering works?



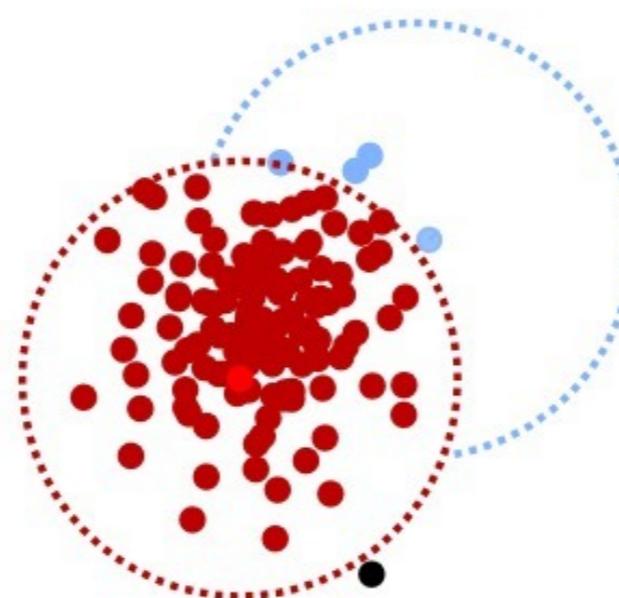
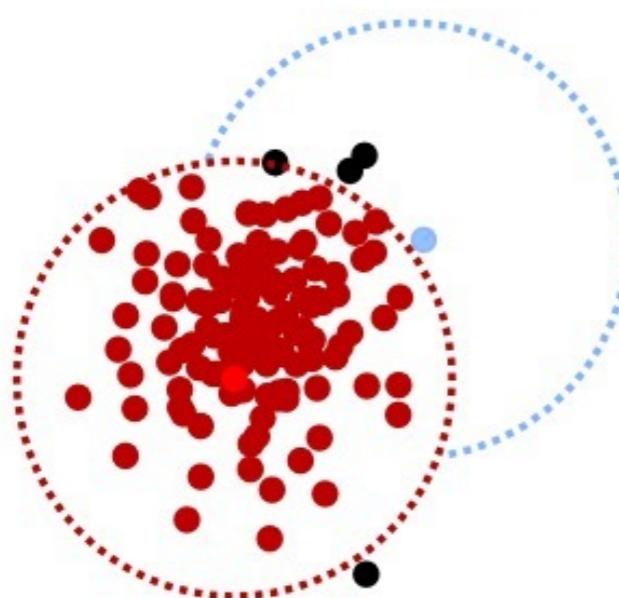
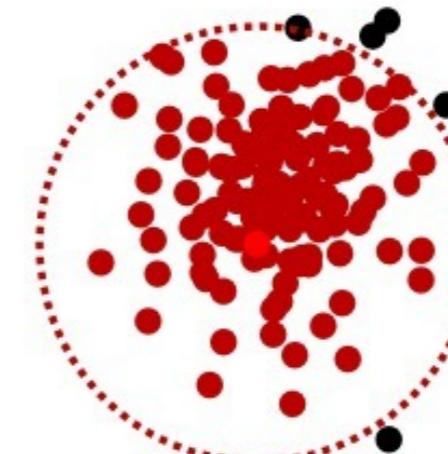
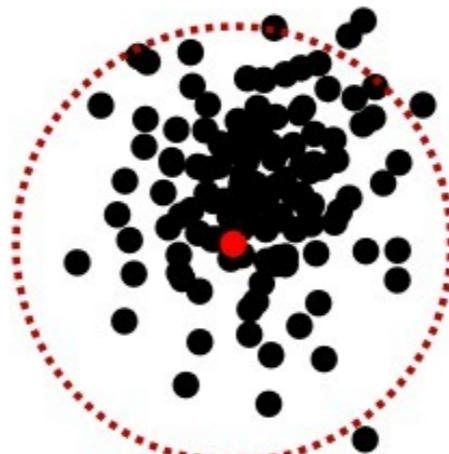
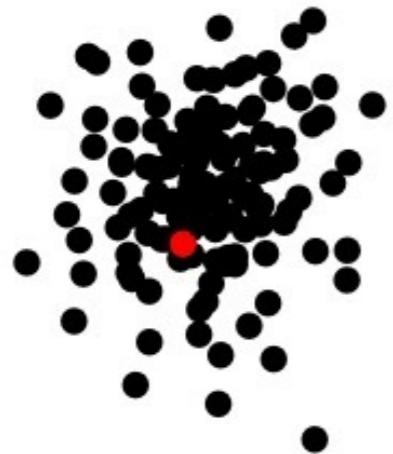
How traditional clustering works?



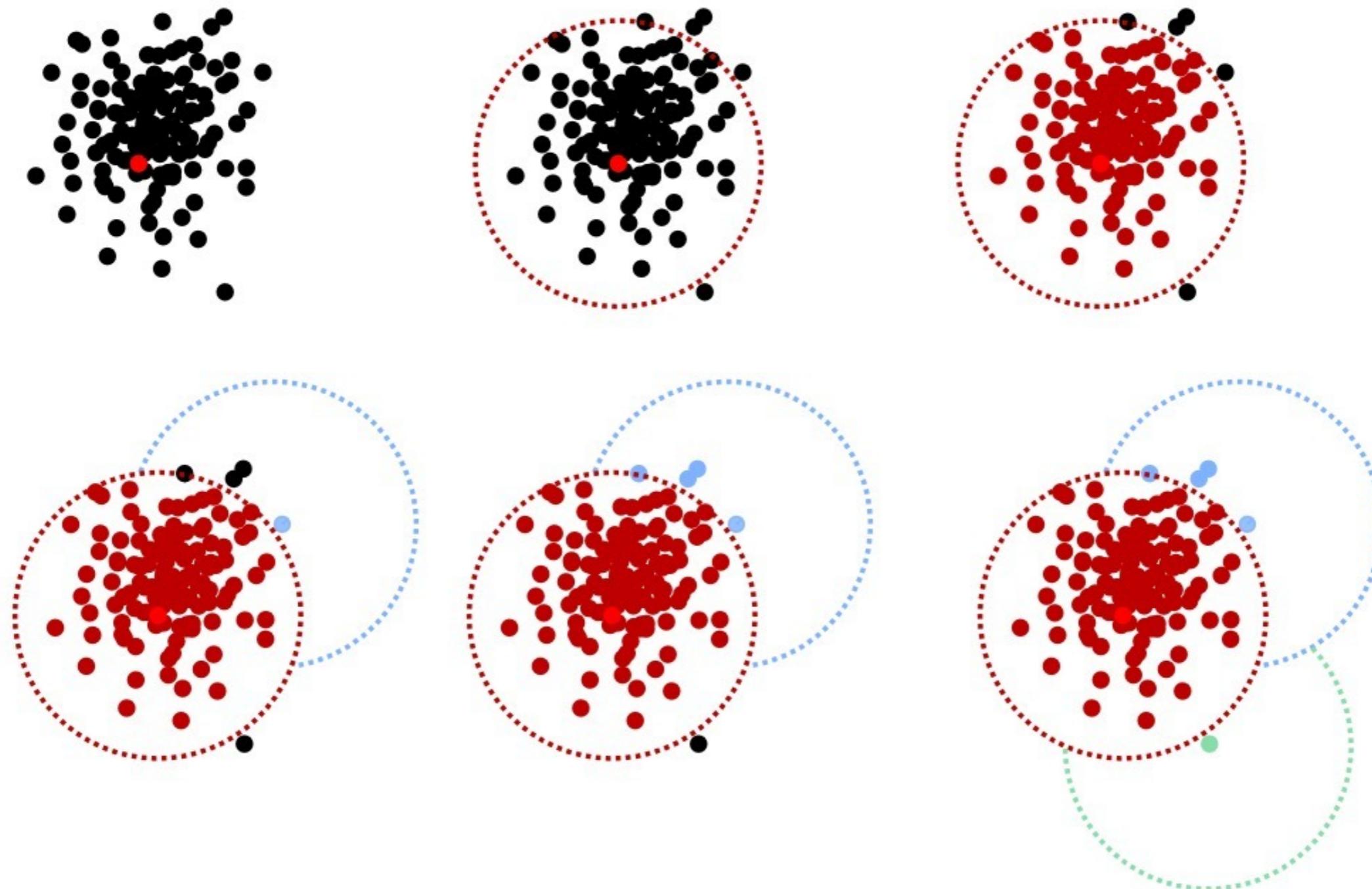
How traditional clustering works?



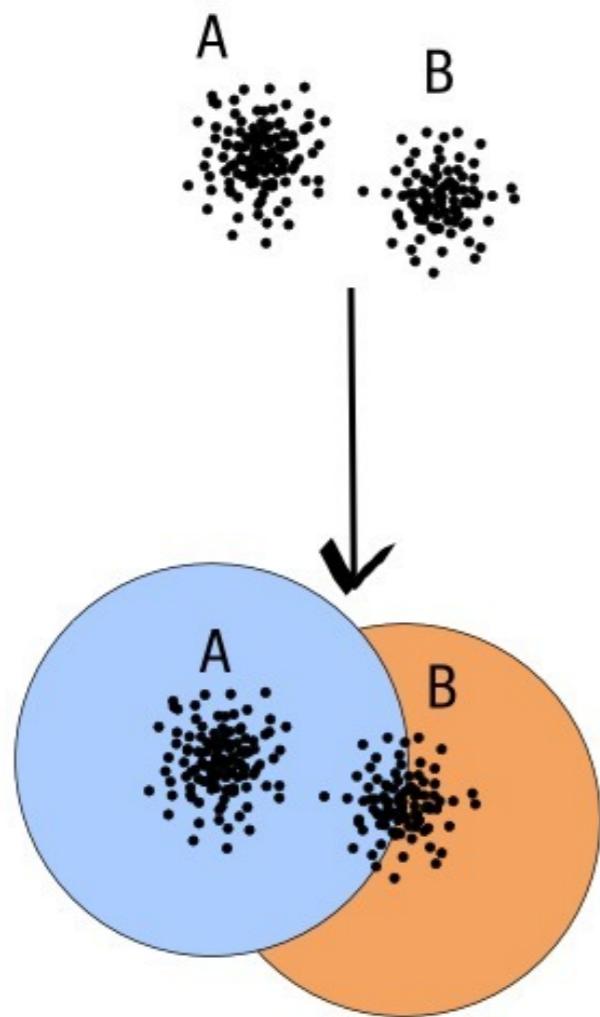
How traditional clustering works?



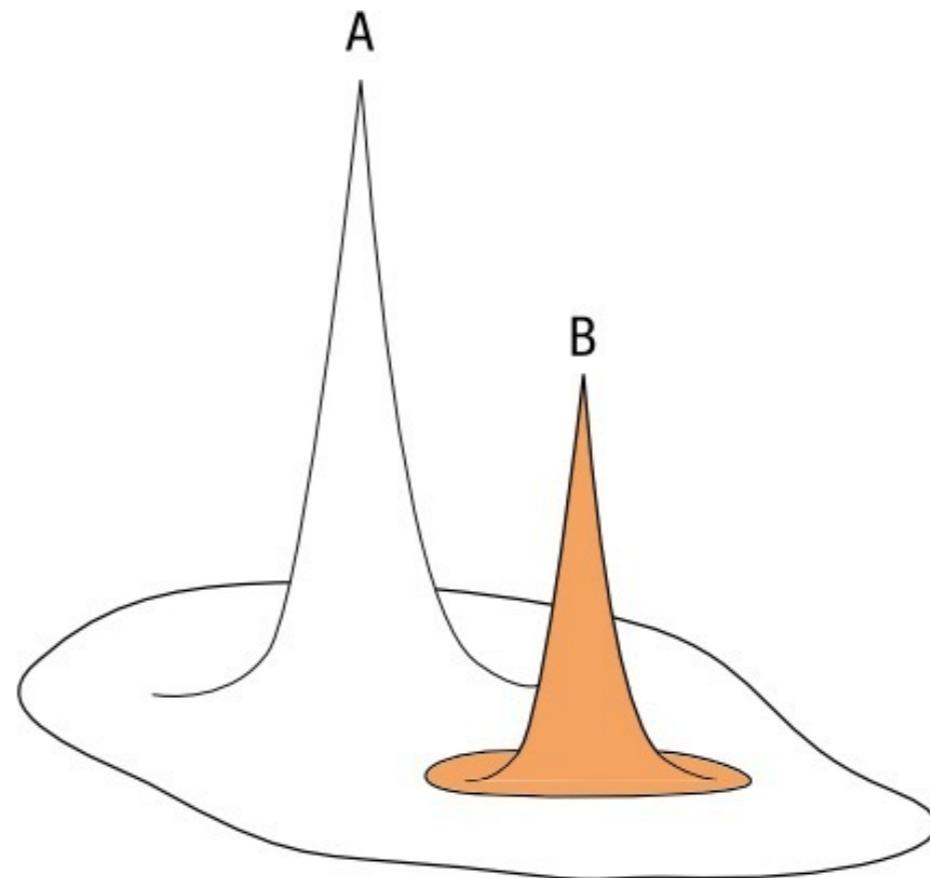
How traditional clustering works?



Swarm: fast, exact and high-resolution clustering



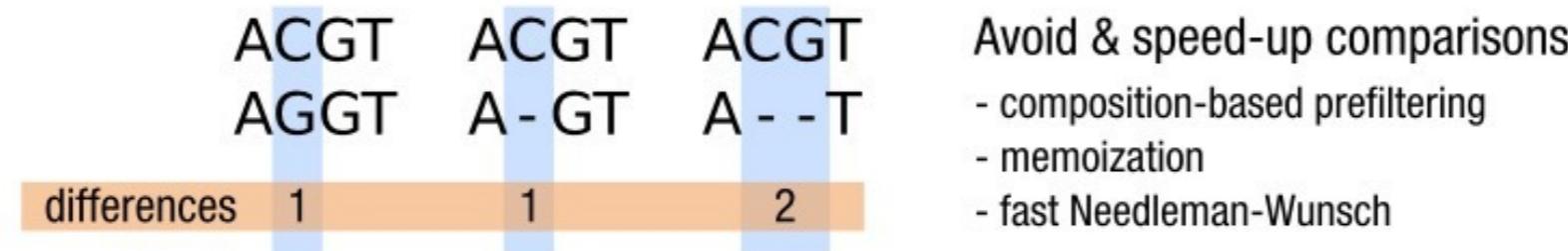
clustering threshold (often 97%)
is most of the time unadapted and
can mask diversity.



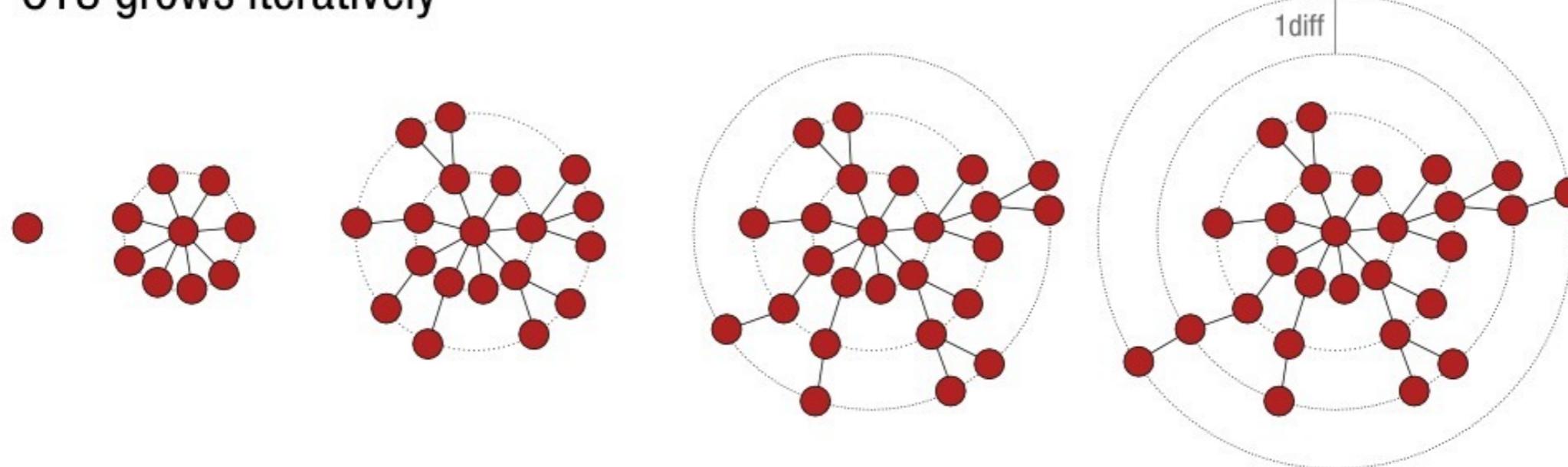
swarm uses abundance values and a new
clustering strategy to delineate natural
high-quality OTUs.

Swarm clustering method

growth phase



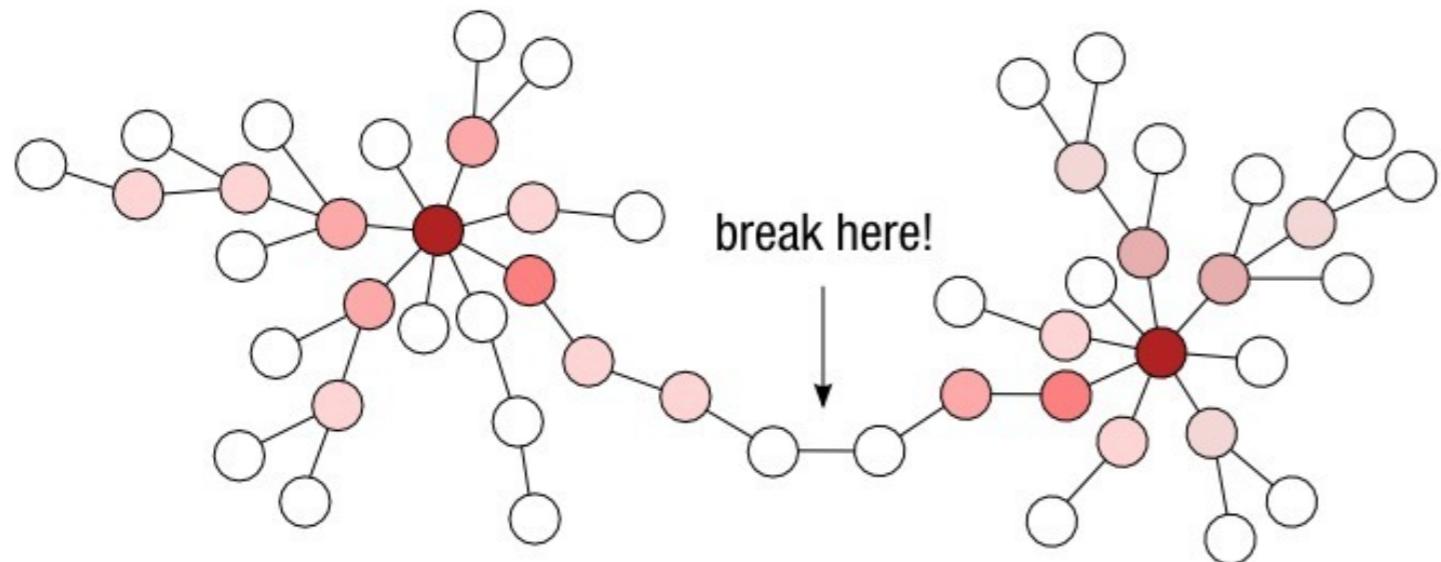
OTU grows iteratively



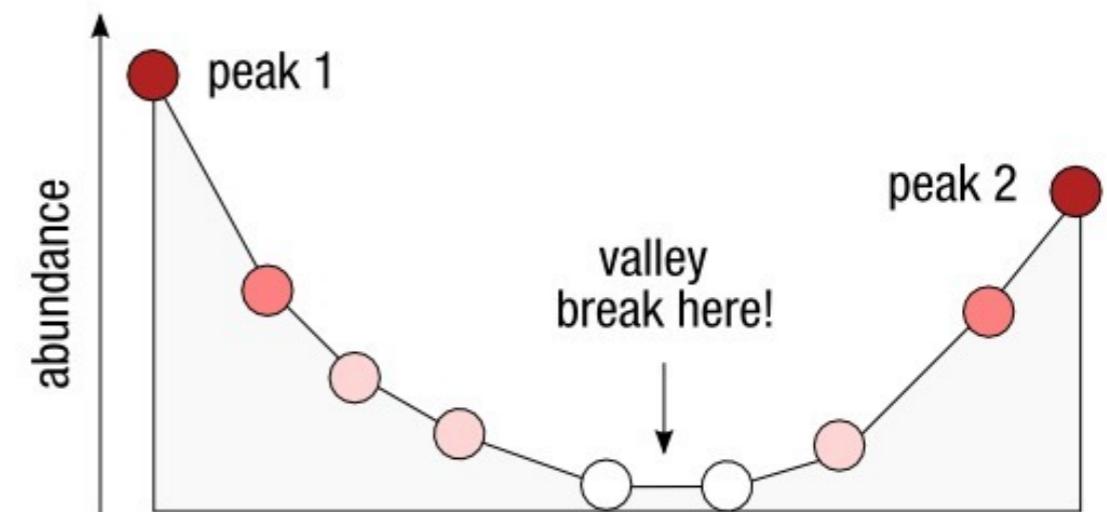
initial seed (randomly picked
from amplicon dataset)

no more closely related amplicons,
the process stops

Swarm clustering method breaking phase



Take into account the abundance of amplicons
to produce higher-resolution clusters.



Assuming that original sequences are more
abundant than erroneous copies.

Swarm clustering method

grafting phase

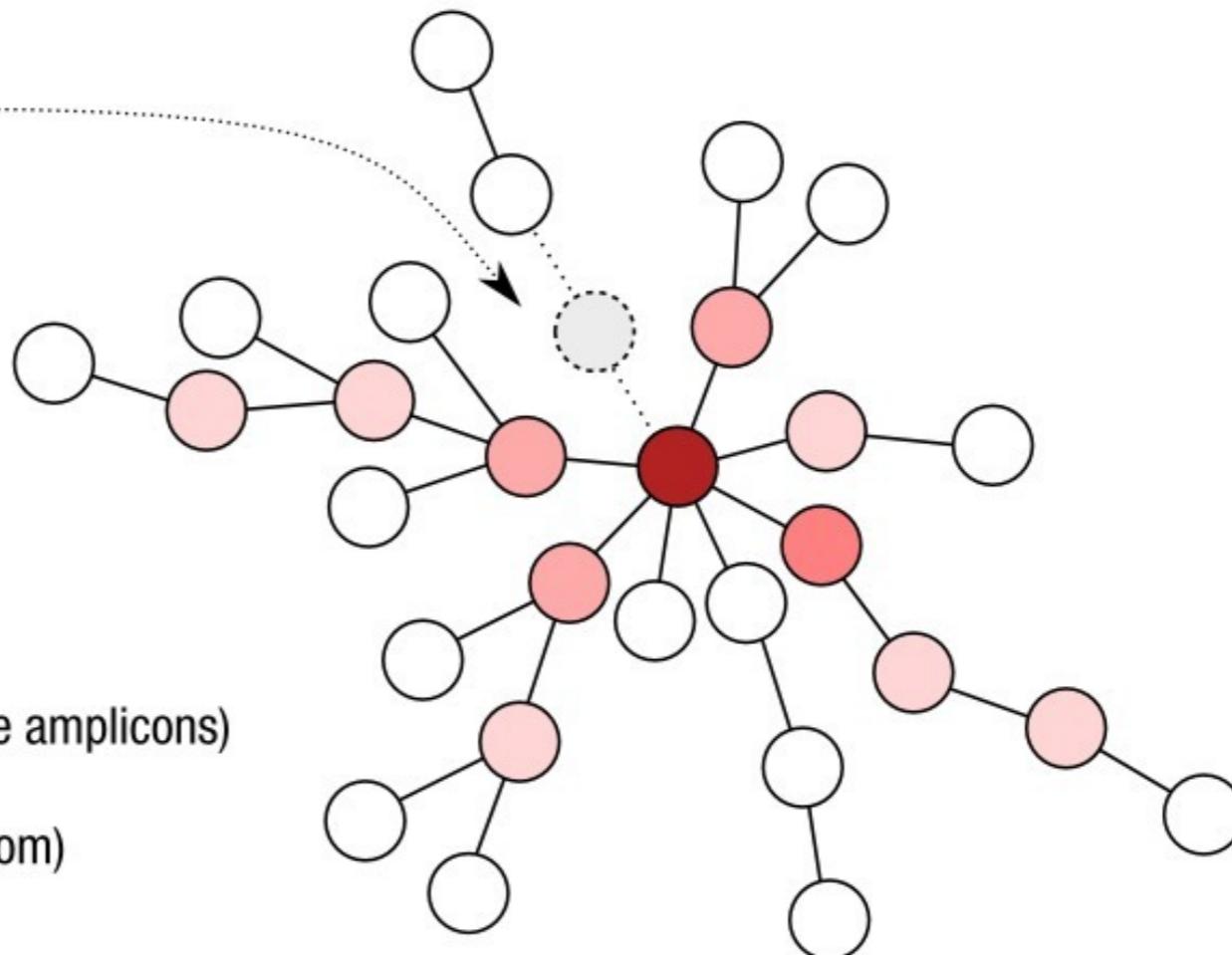
Postulate the existence of an intermediate amplicon to be able to graft a small OTU onto a bigger one.



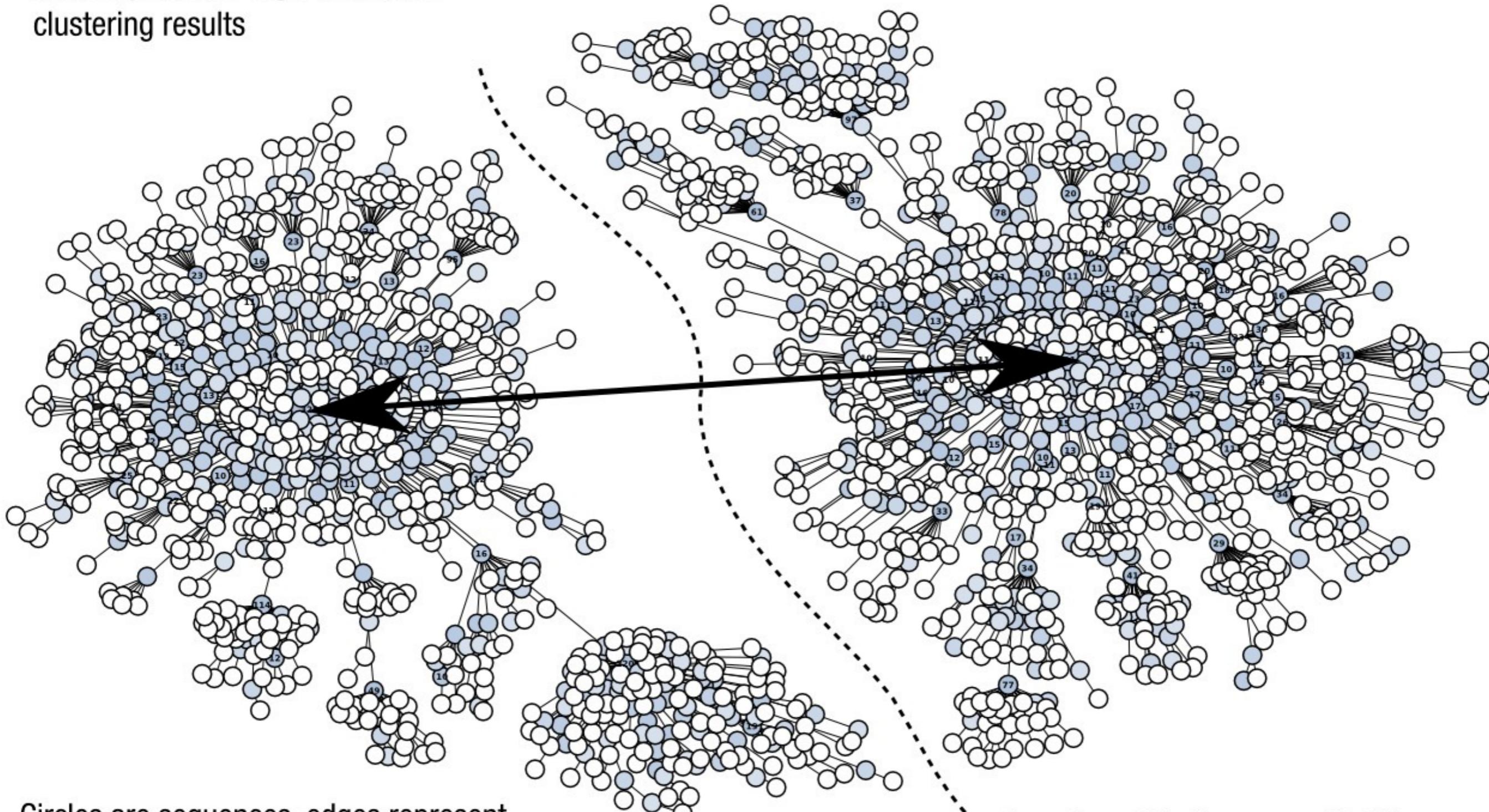
small OTU (made of 2 rare amplicons)



virtual amplicon (or phantom)



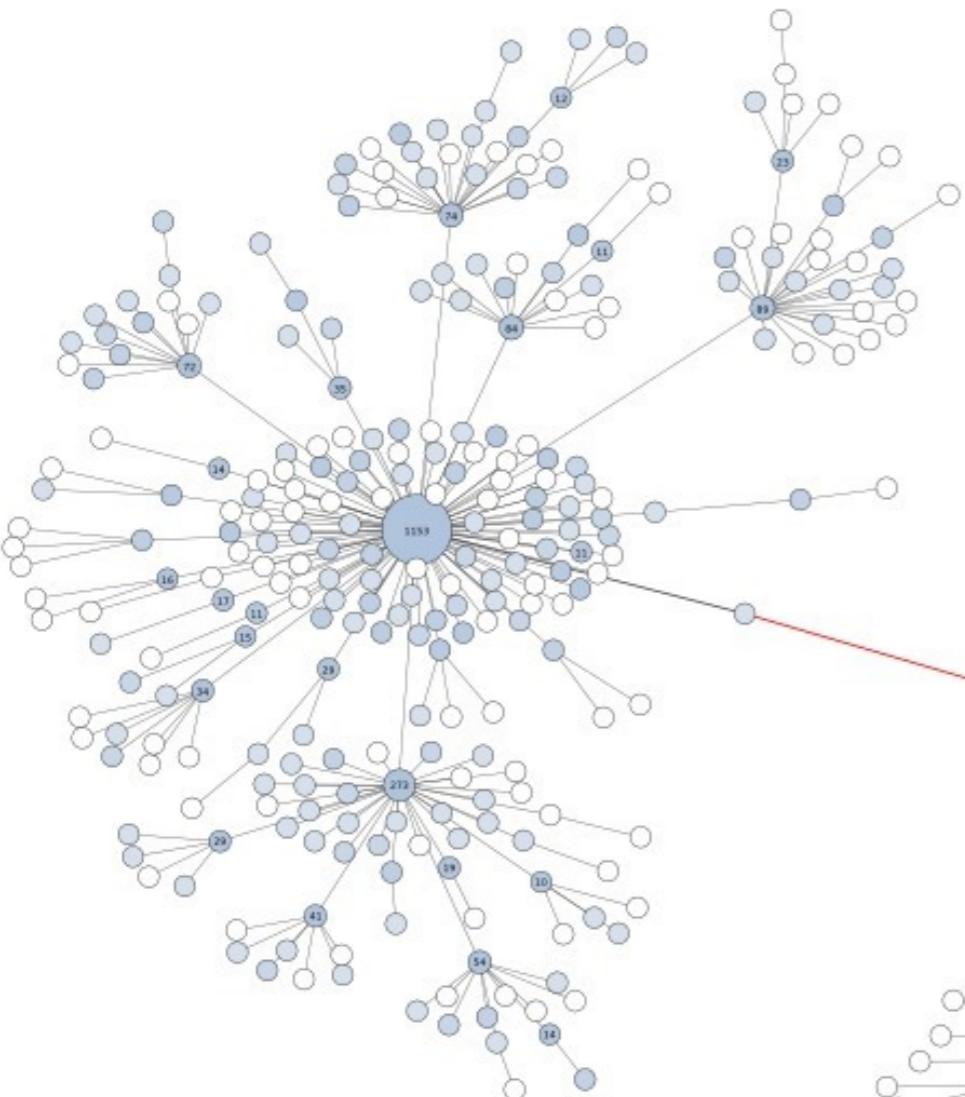
Swarm produces high-resolution clustering results



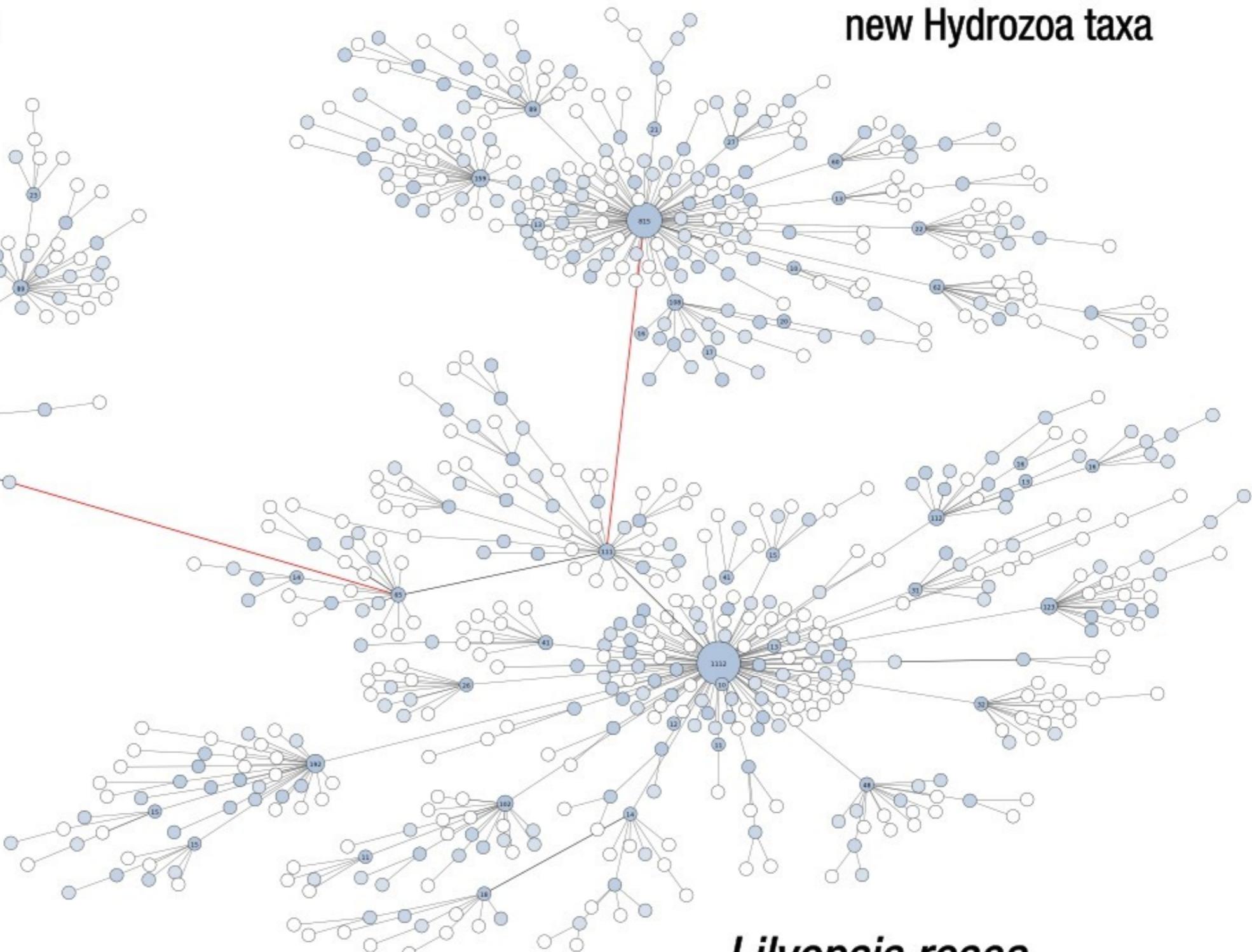
Circles are sequences, edges represent
one difference (substitution or indel)

Less than 1% divergence (3 differences)
between the two peaks of abundance

other Hydrozoa genus



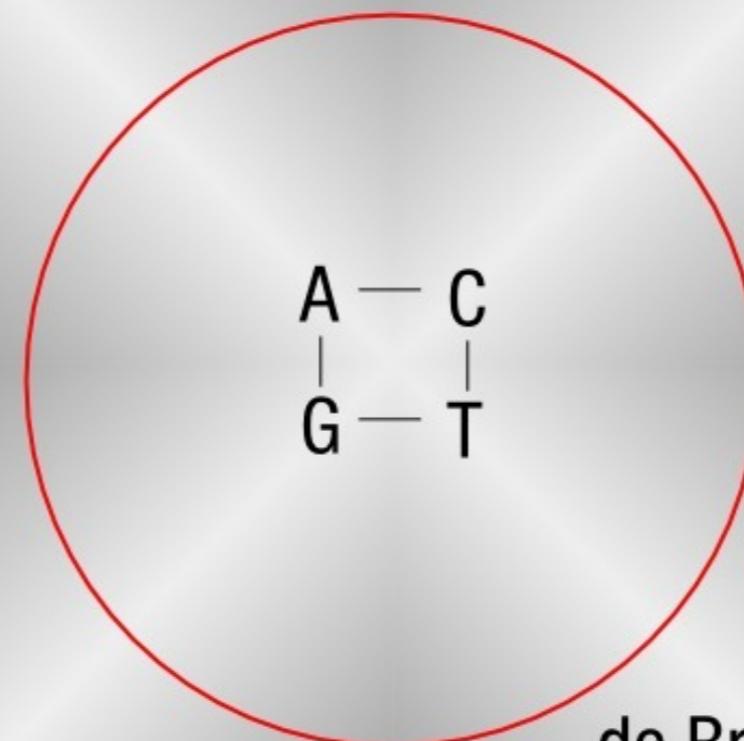
new Hydrozoa taxa



Cnidaria (Metazoa)

sequence-space

swarm derives from basic
observations of the
sequence-space



de Bruijn graph in
genome assembly

Swarm 2.0 is a highly scalable denoising-clustering method

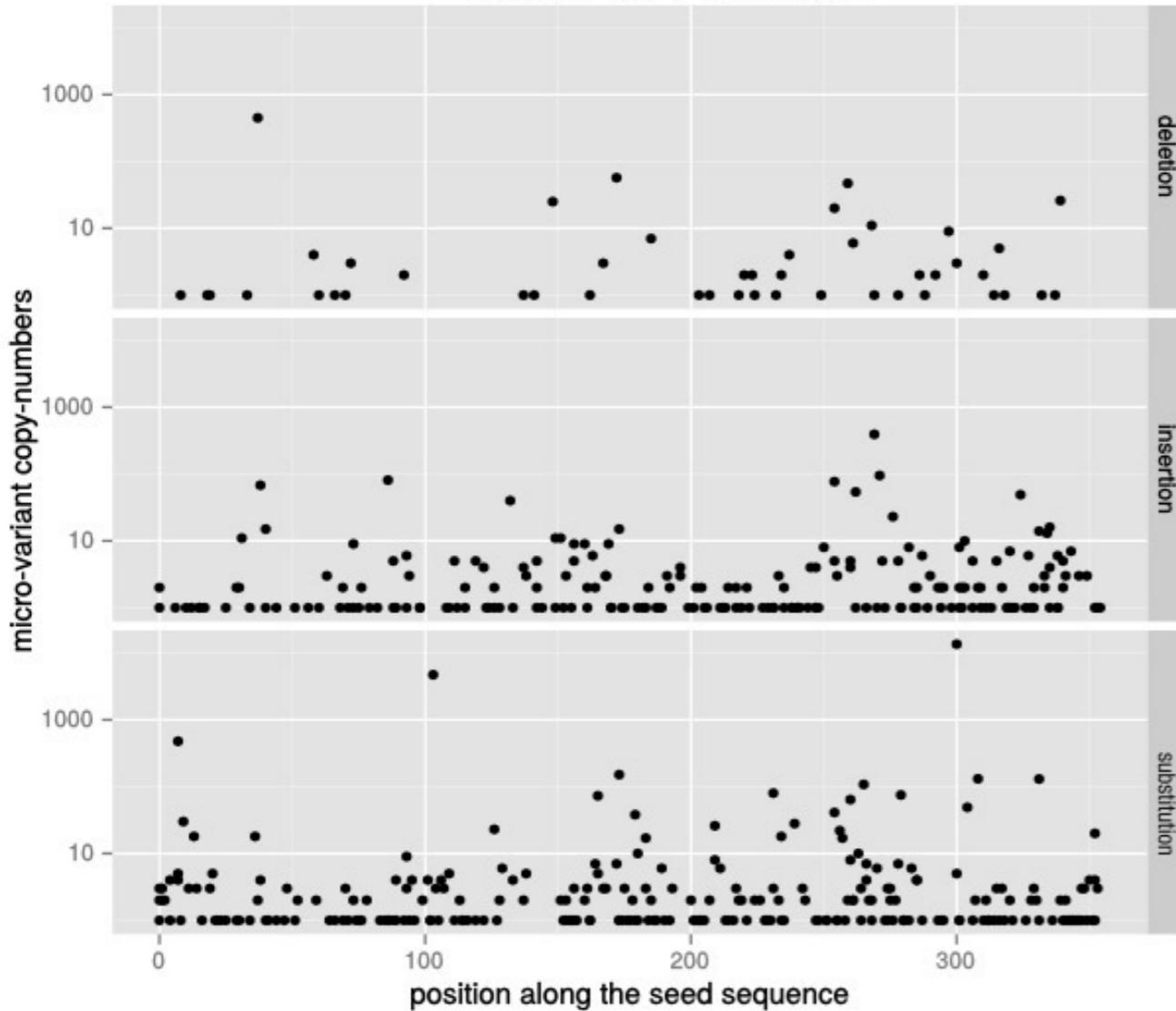


 earth
microbiome project

swarm: 5 hours
usearch: >150 days

How are errors distributed?

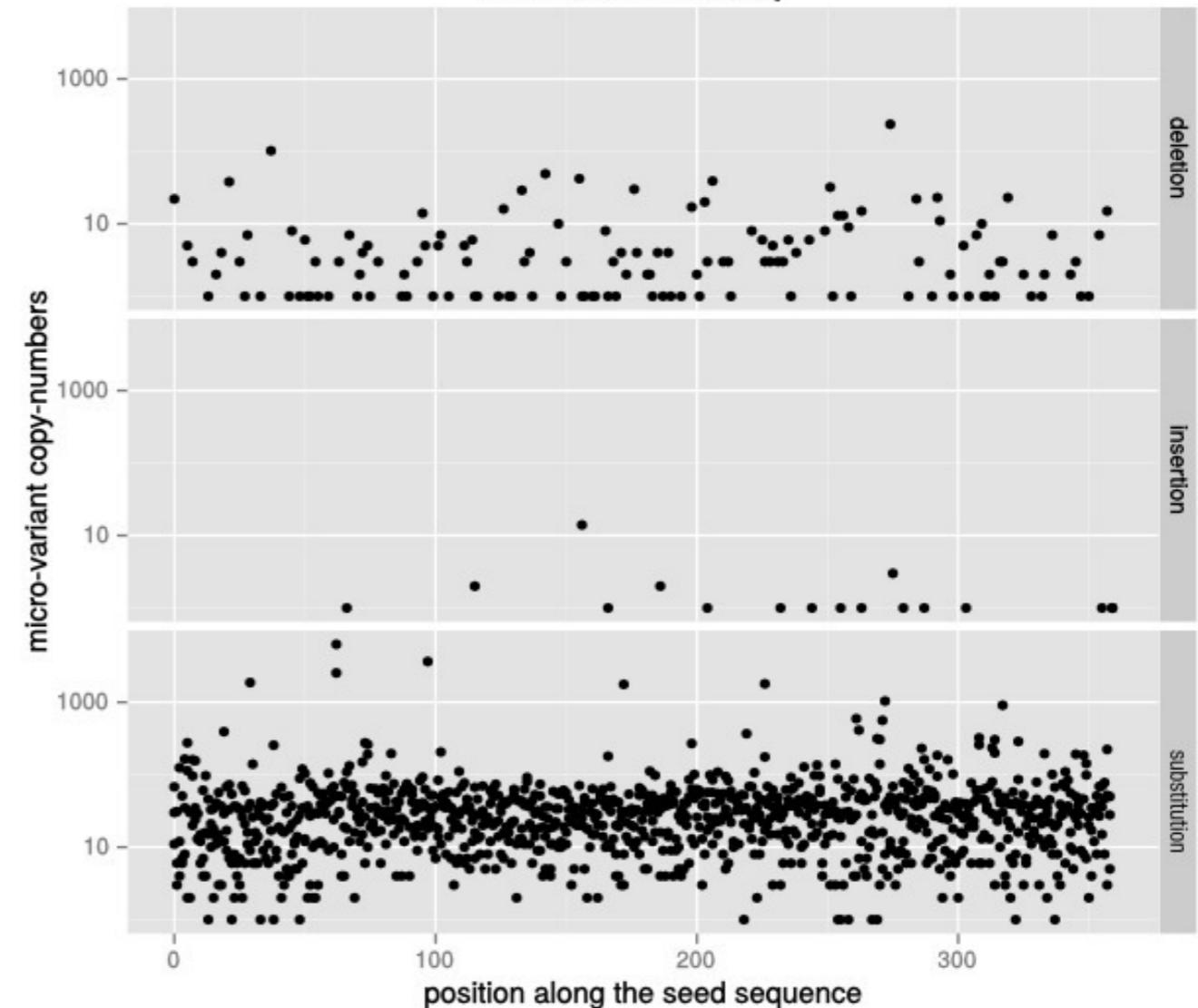
Roche 454 Titanium



Roche 454 Titanium (for 100 bp):

- 0.4011 insertions/deletions,
- 0.0543 substitutions.

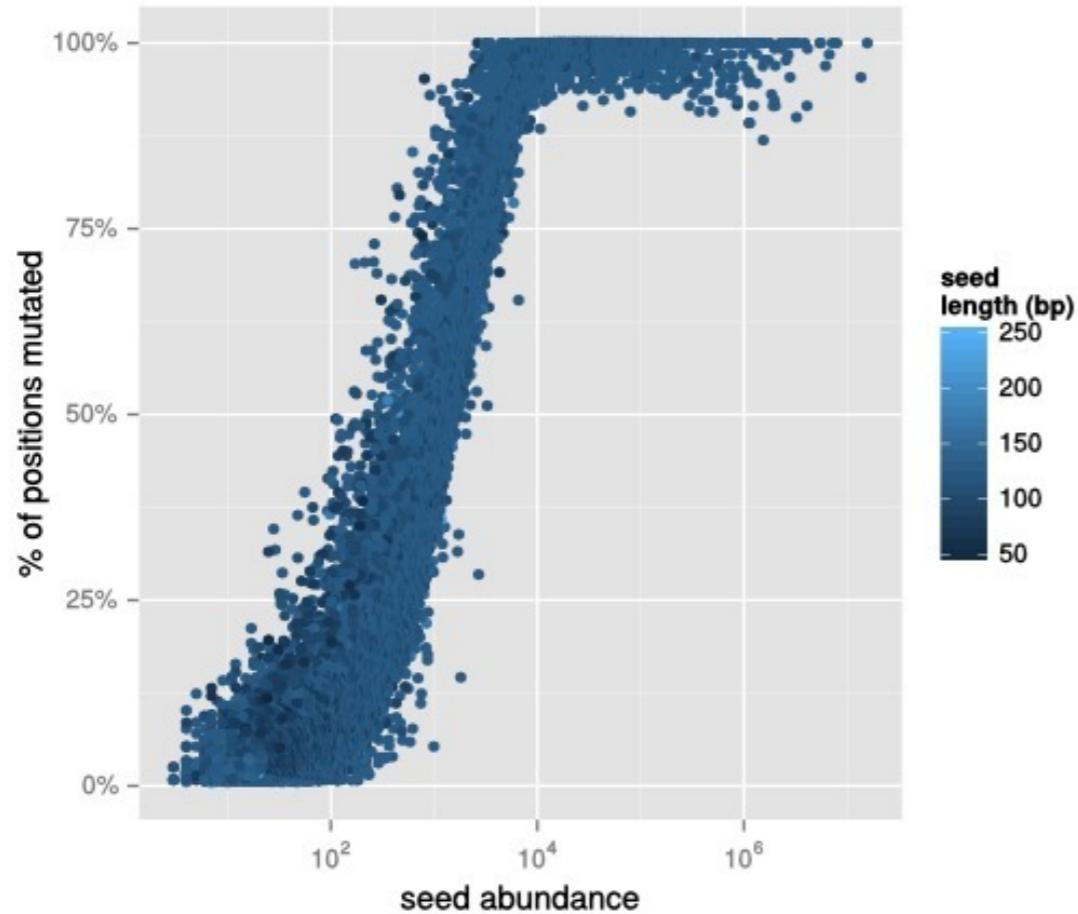
Illumina MiSeq



Illumina MiSeq v2 (for 100 bp):

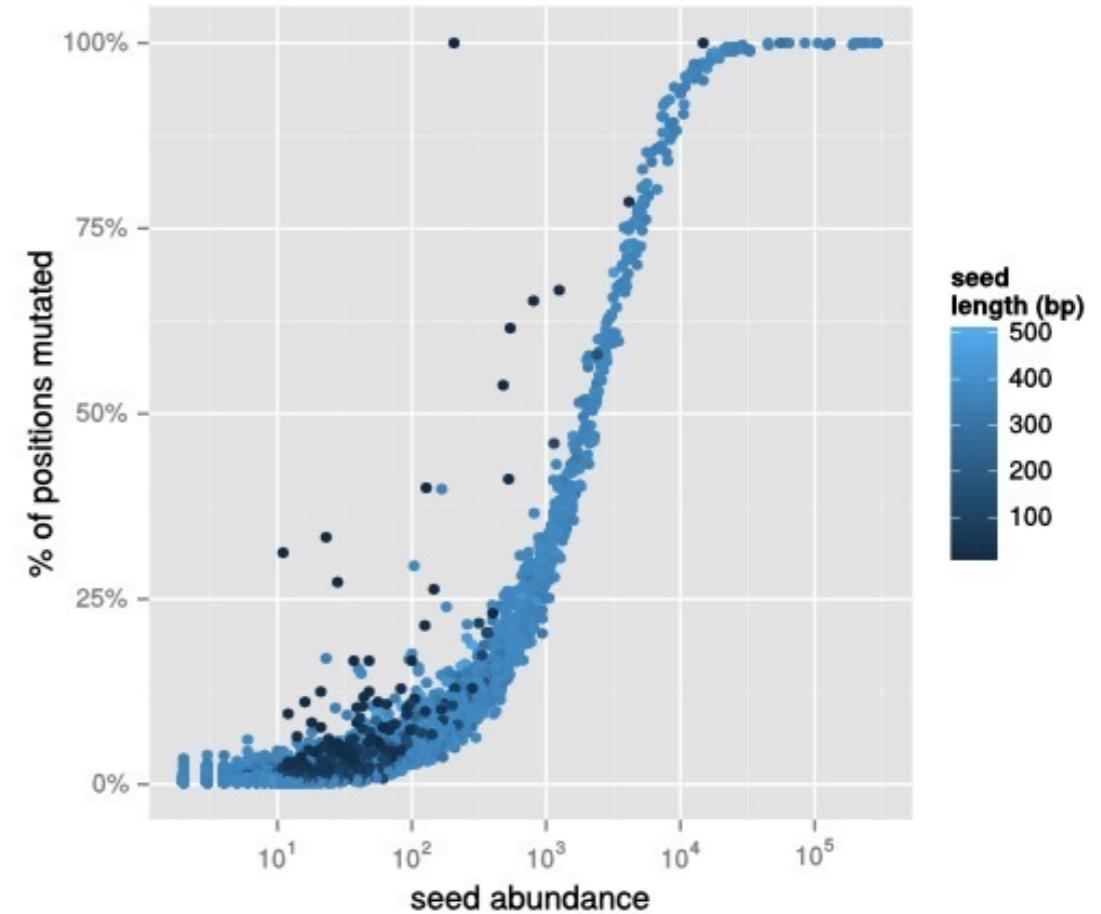
- 0.0009 insertions/deletions,
- 0.0940 substitutions.

How fast errors accumulate with sequencing depth?



TARA OCEANS
V9

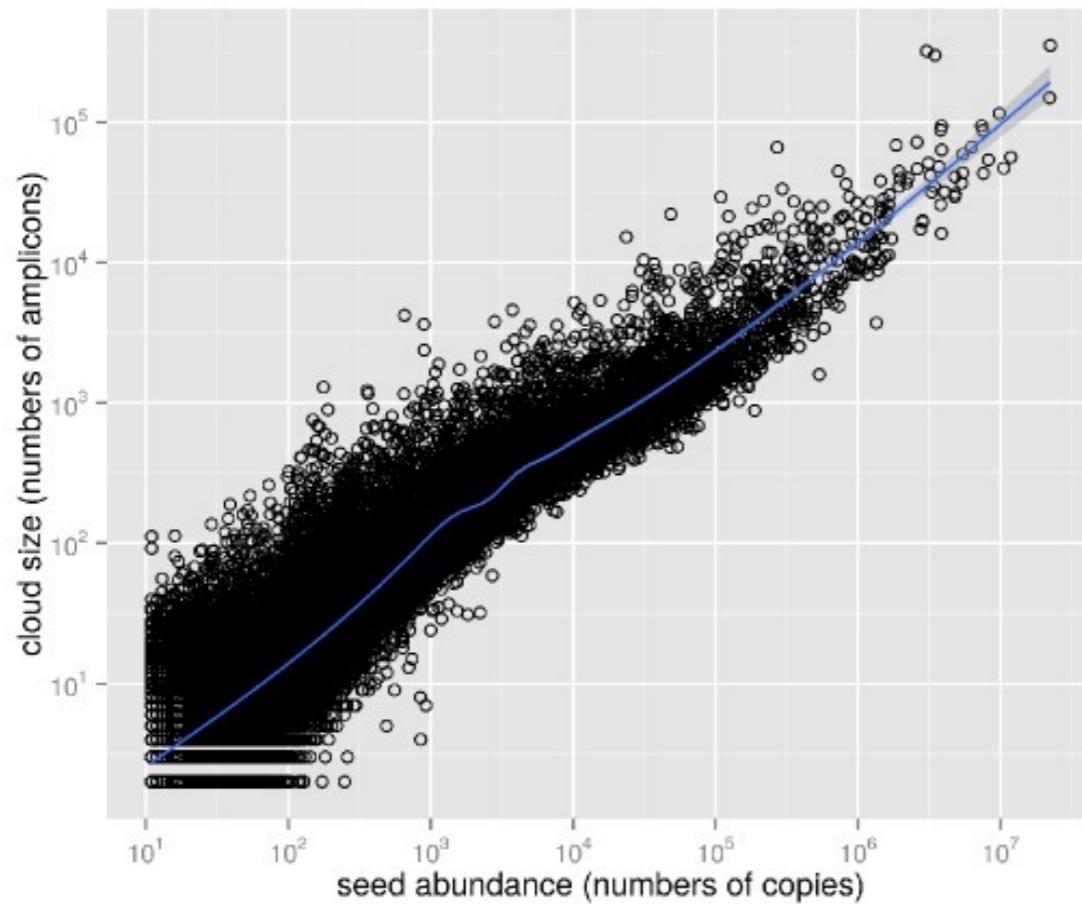
Saturation curve depends on the average seed length and sequencing platform, not on sequencing depth.



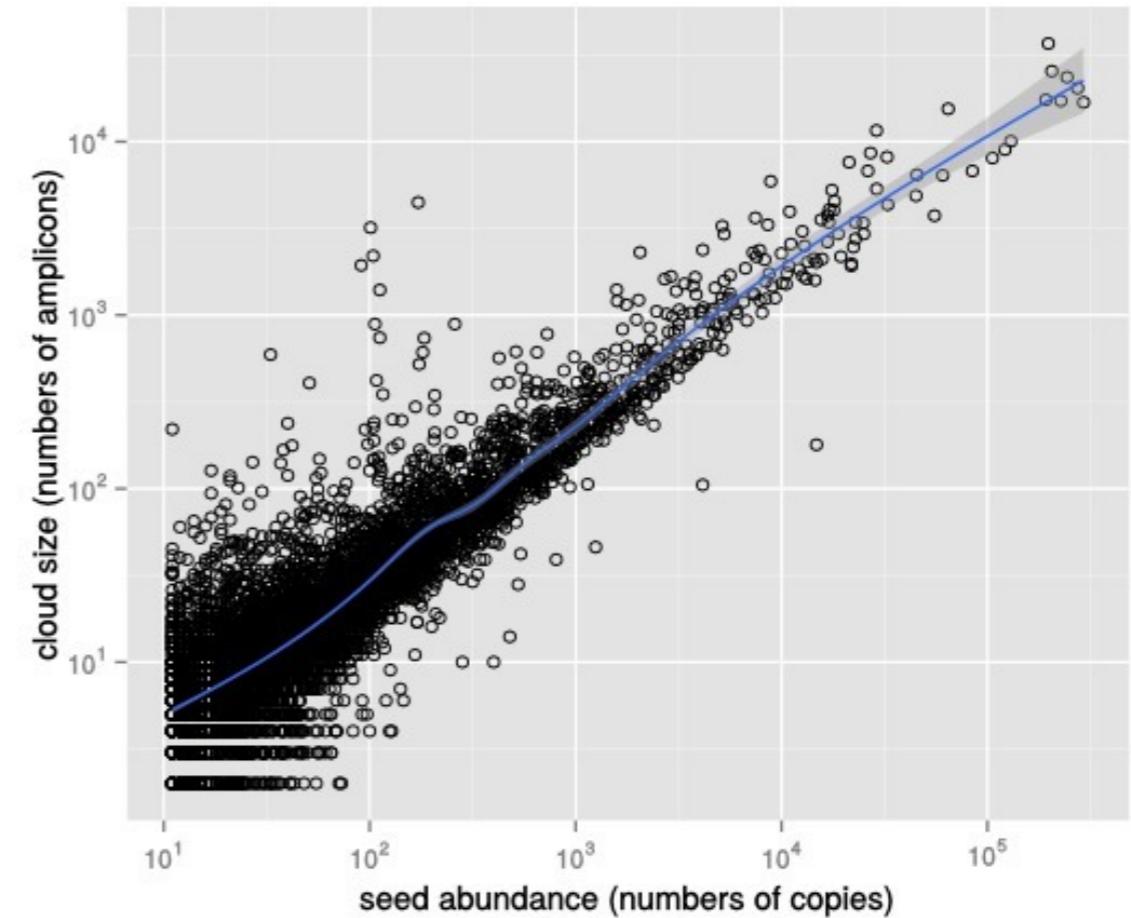
Neotropical Forest Soils
V4

Cloud size (and diameter) will vary with the taxa abundance. There is no optimal per-taxa clustering threshold.

Seed versus Cloud



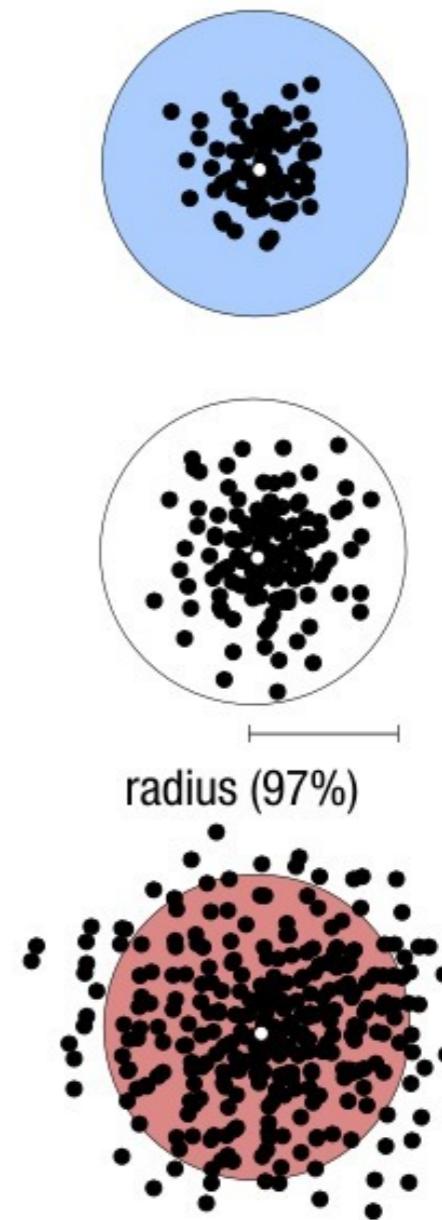
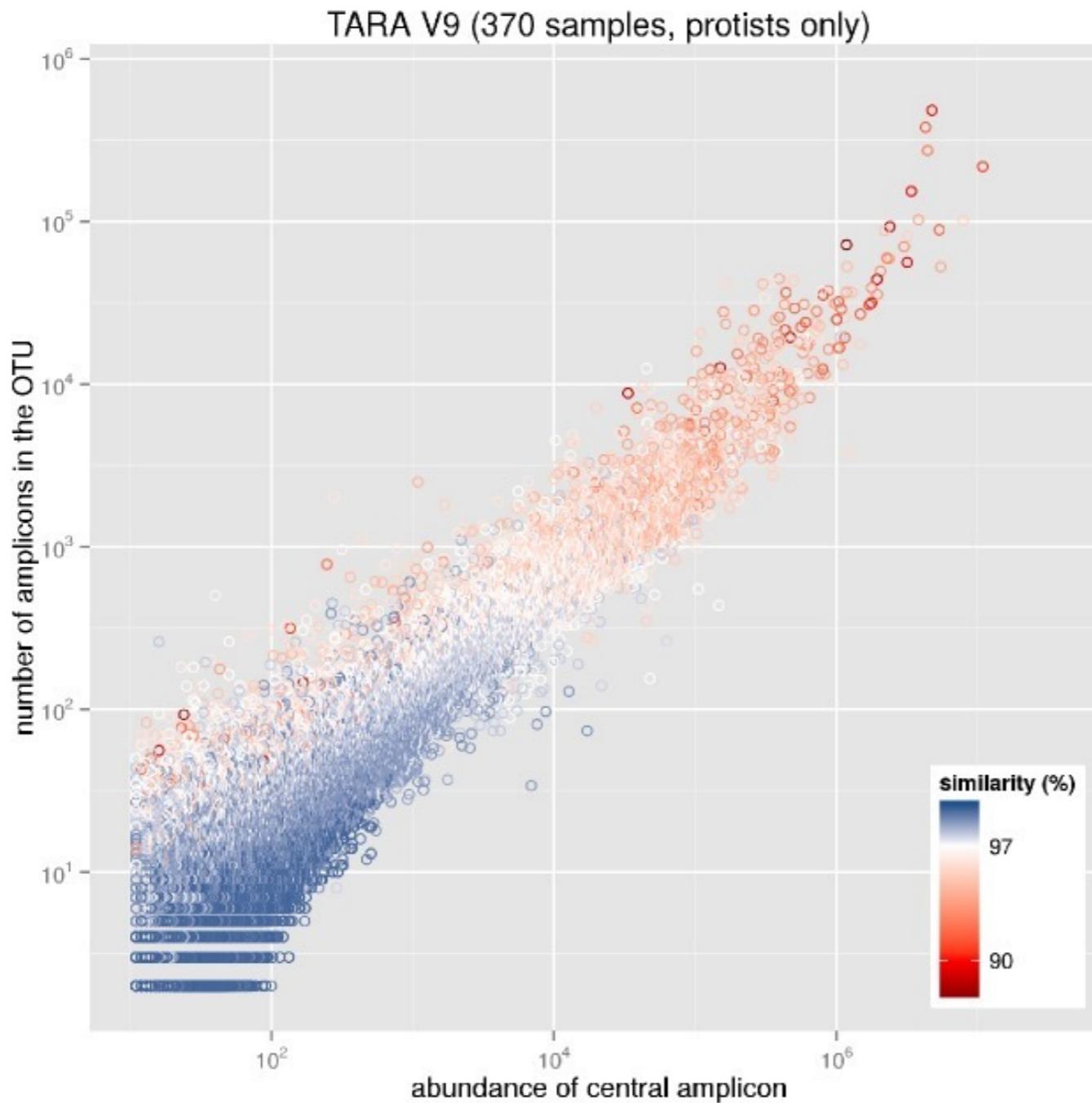
TARA OCEANS
V9



Neotropical Forests Soils
V4

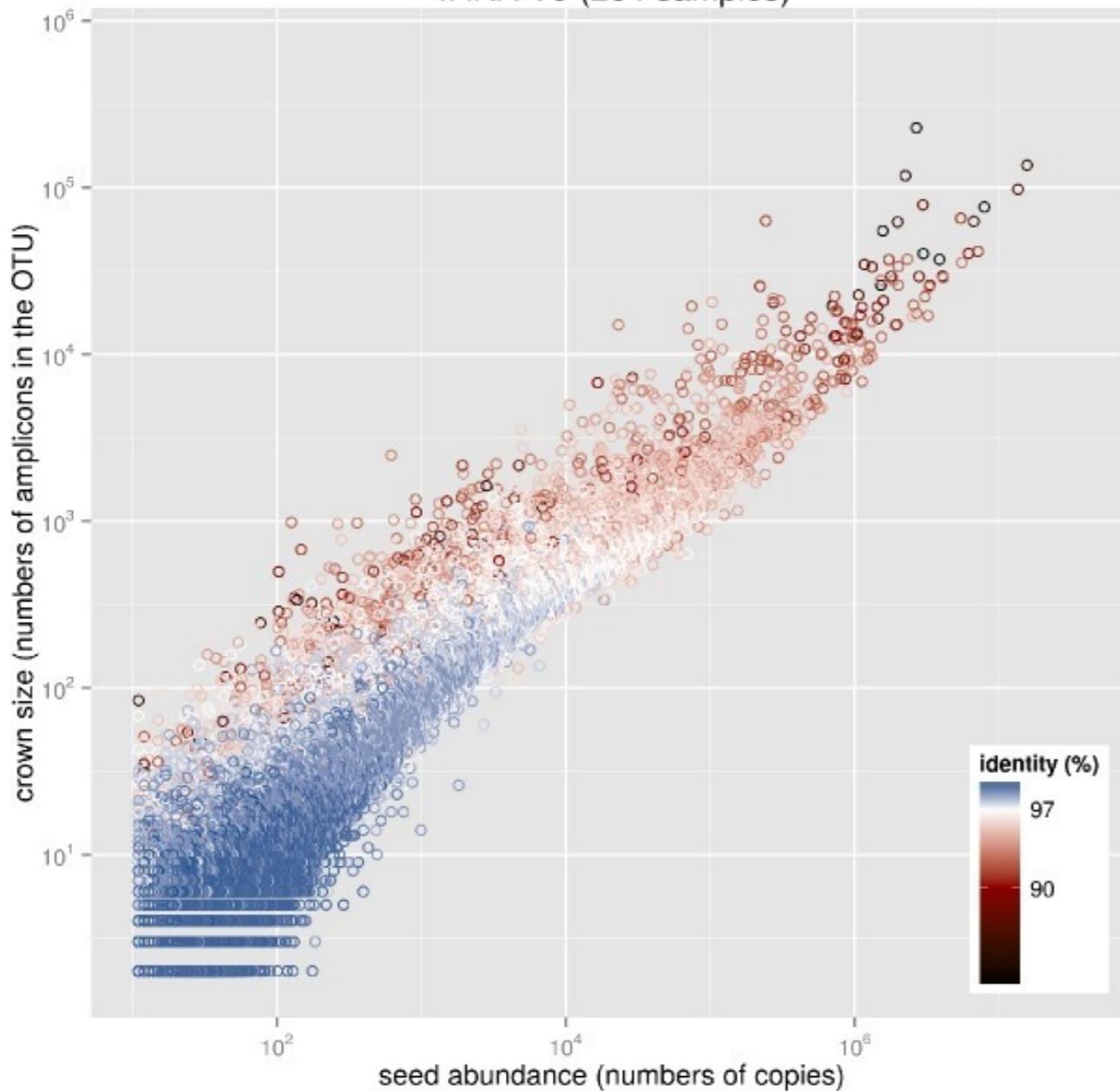
Strong correlation between abundance and the number of micro-variants. Abundant taxa are not known to have more diversified V9 sequences than less abundant taxa. What we see are technical errors.

What if we'd used a fix 97%-clustering threshold?

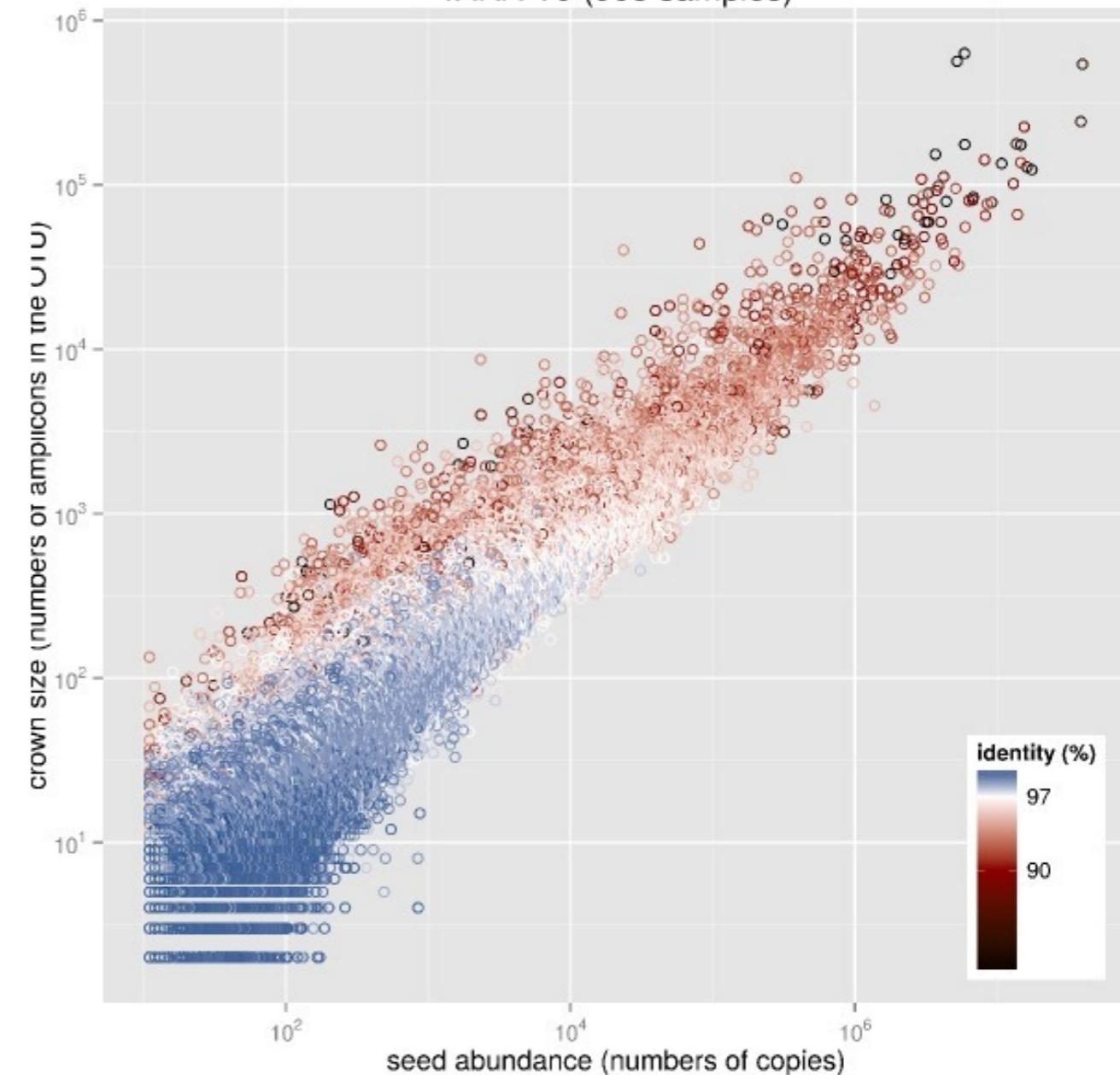


Seed abundance vs cloud vs cluster radius shows 97%-threshold inadequacy

TARA V9 (264 samples)

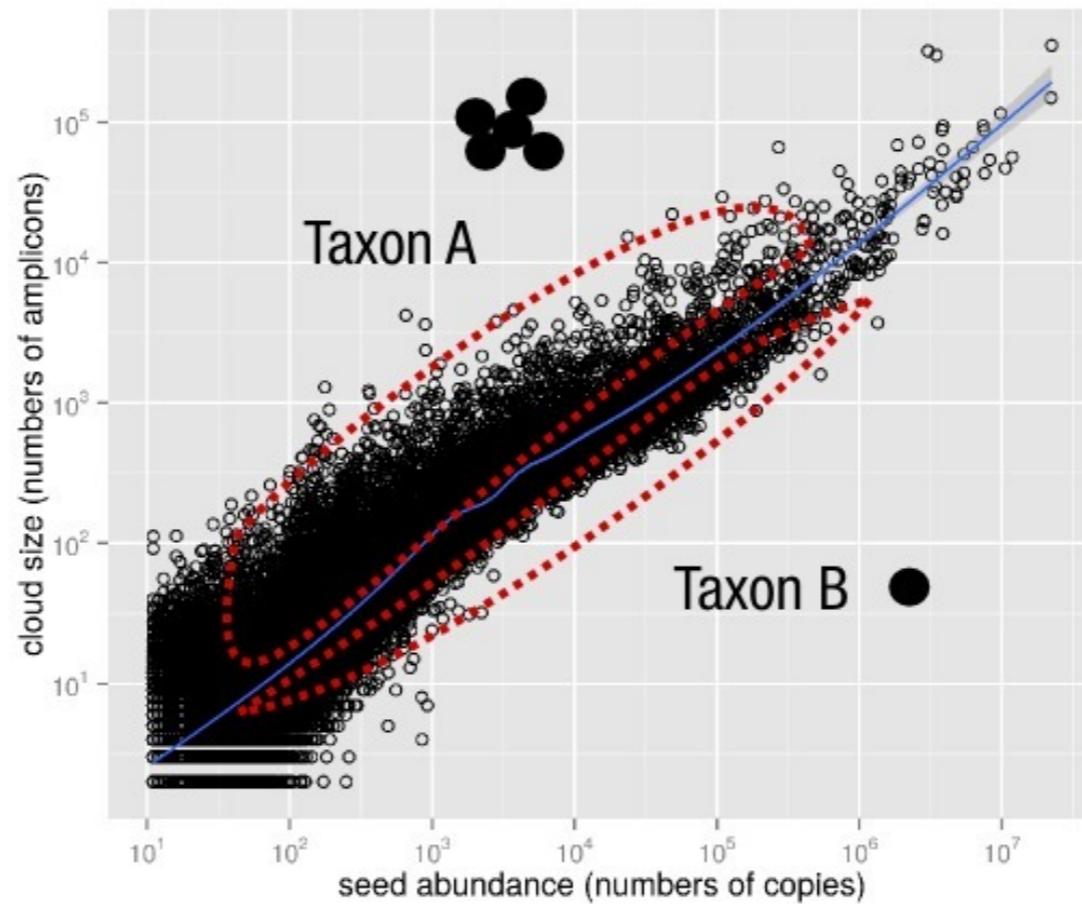


TARA V9 (908 samples)



clusters produced with swarm using $d = 1$

Natural variability?



TARA OCEANS
V9

New algorithms can deal with big data.
Now, how can we reduce noise further?

chimera detection

A: GTCGCTACTACCGATTGAACGTTTAGTGAGGTCTCGGACTGTGAGCCTGGCGGGTTG
|||||||||||

B: TACTACCAAACTGAGTTAGCGTTTAGTGAGGT AAGACGACCAAATGTAGCGTTAG

C: GTCGCTACTACCGATTGAACGTTTAGTGAGGT AAGACGACCAAATGTAGCGTTAG

uchime

UCHIME improves sensitivity and speed of chimera detection

Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R.

Bioinformatics. 2011 Aug 15;27(16):2194-200. doi: 10.1093/bioinformatics/btr381

R.C.Edgar et al.

A	81	CCTTGGTAGGCCGtTGCCCTGCCAAGTAGCTAATCAGACGCgggtCCATCtcaCACCaccggAgtTTTtcTCaCTgTacc	160	
Q	81	CCTTGGTAGGCCGCTGCCCTGCCAAGTAGCTAATCAGACGCATCCCCATCCATCACCGATAAAATCTTAATCTCTTCAG	160	
B	81	TCTTGGTgGGCCGtTaCCCcGCCAACaAGCTAATCAGACGCATCCCCATCCATCACCGATAAAATCTTAAaCTCTTCAG	160	
Diffs	A	A	p A A A	BBBB BBB BBBB BB BBa B B BBB
Votes	+	+	0 + + +	++++ +++ +++++ ++ ++ ! + + +++
Model	AAAAAAAAAAAAAAAAAAAAAxxxxxxBB			

Fig. 3. Chimeric alignment showing diffs and votes. This figure shows a region from an alignment generated by UCHIME. Diffs and votes are annotated. The ‘Model’ row indicates the three segments of the alignment which are closer to A, the crossover (X) and closer to B, respectively. Diffs are ‘A’ = diff with Q closer to A in the A segment, ‘a’ = diff with Q closer to A in the B segment, and similarly for ‘B’ and ‘b’. A ‘p’ diff indicates that the parents agree but are different from Q. Votes are ‘+’ (yes), ‘!’ (no) and ‘0’ (abstain), indicating whether the corresponding diff supports or contradicts the model.

vsearch: open-source alternative for usearch

clustering, chimera detection, dereplication, searching, sorting, masking and shuffling

usearch (Rob Edgar):

- very important for metagenomics,
- 1,000 citations,
- fundation for QIIME,
- closed-source & costly



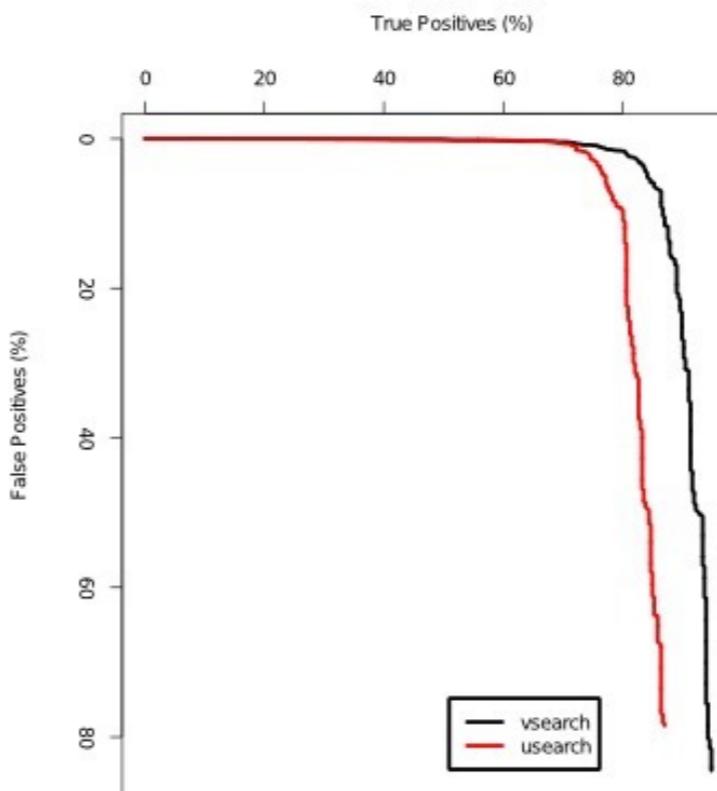
growing success:

- many happy users,
- faster and improved,
- fundation for QIIME 2.0

vsearch:

- free and open-source,
- fast,
- documented,
- revive the research field

Torbjørn Rognes
Oslo University



vsearch: open-source alternative for usearch

clustering, chimera detection, dereplication, searching, sorting, masking and shuffling

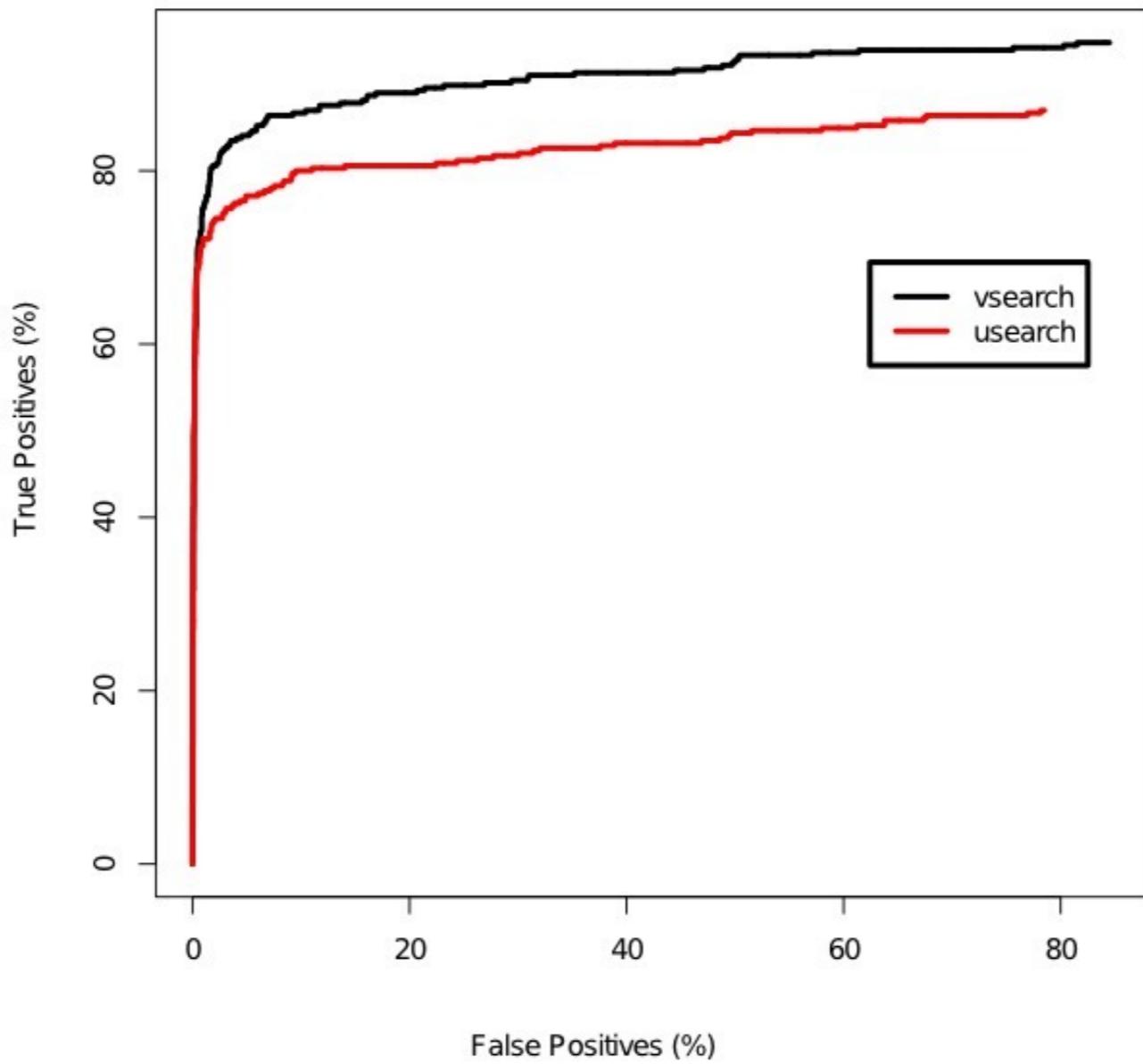
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- very important
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growing success

- many happy users
- faster and improved
- fundation for QIIME



Torbjørn Rognes
Oslo University



quality filtering

```
@M00185:171:000000000-AJ75U:1  
TACGGGAGGCAGCAGTGGGAATCTTGCAGCGAAAGCGTGACGCAGAACGCCG  
+  
BCCCC@BBCCCCGGGGGGGGHHHHHGGGGHFEGGGGGHGGGEGGGGHHFGGG
```

```
@M00185:171:000000000-AJ75U:2  
CGGCGTTGCTGCGTCACGCTTCGCGCATTGCGCAAGATTCCCCACTGCTGCCTCCGTA  
+  
HEGCG-BCFGGGGGGGGGFFFFFAGAAGAAAAADFFFFFFFEDAF;
```

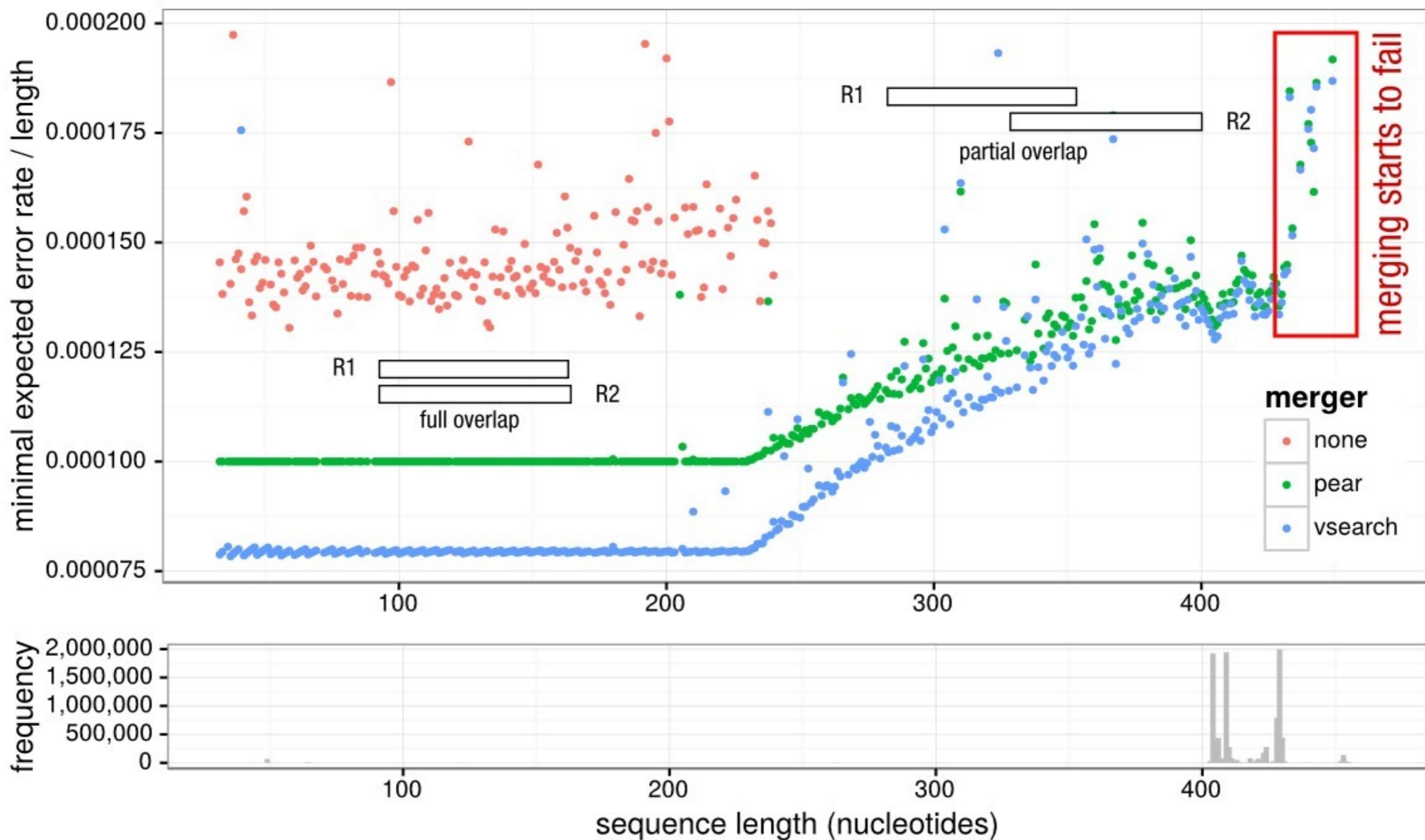
$$\text{expected error rate} = \sum Qv$$

ee = 1.0 (50% chance to have zero error)

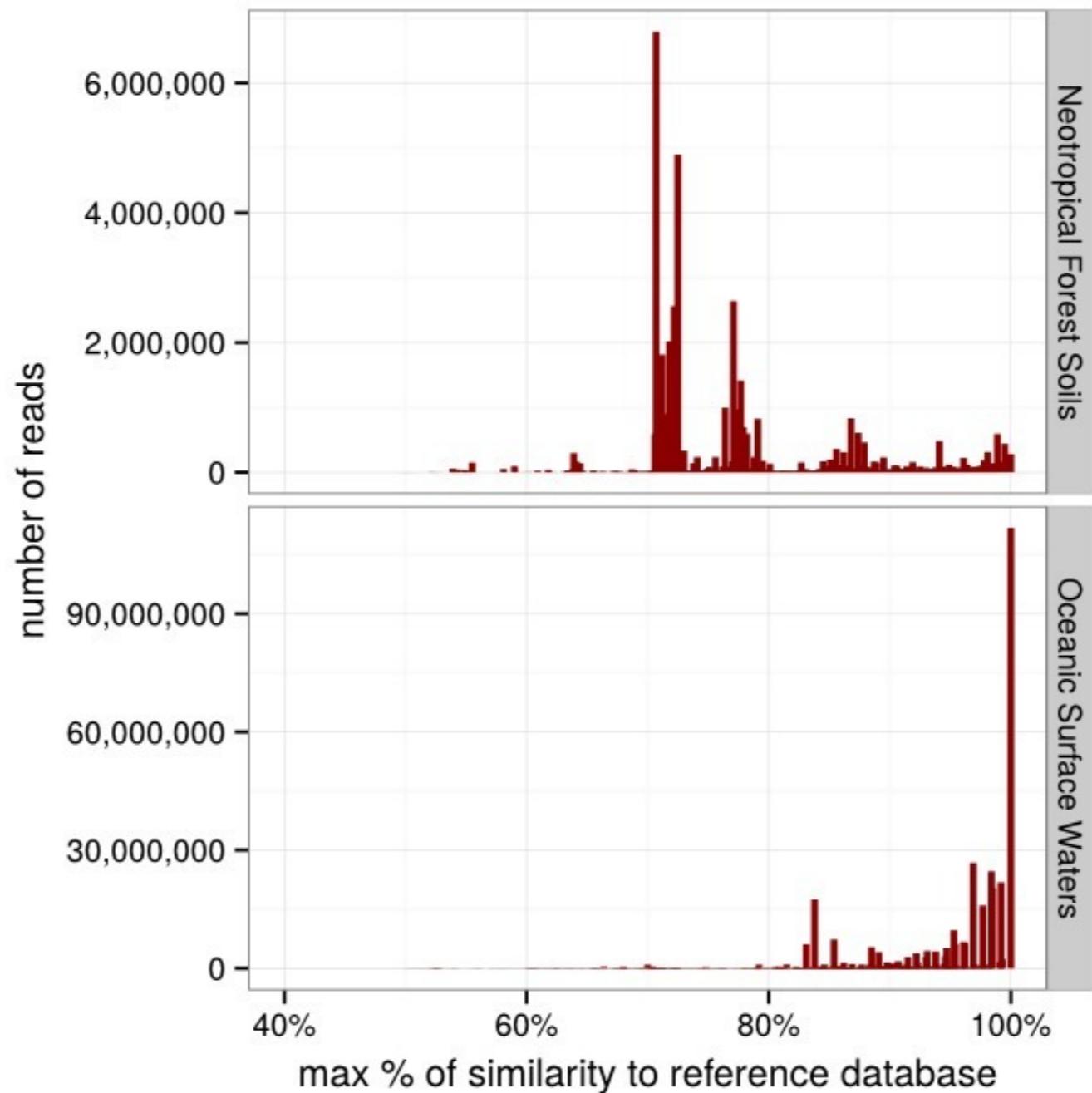
Unique sequences	Σ	Q	length
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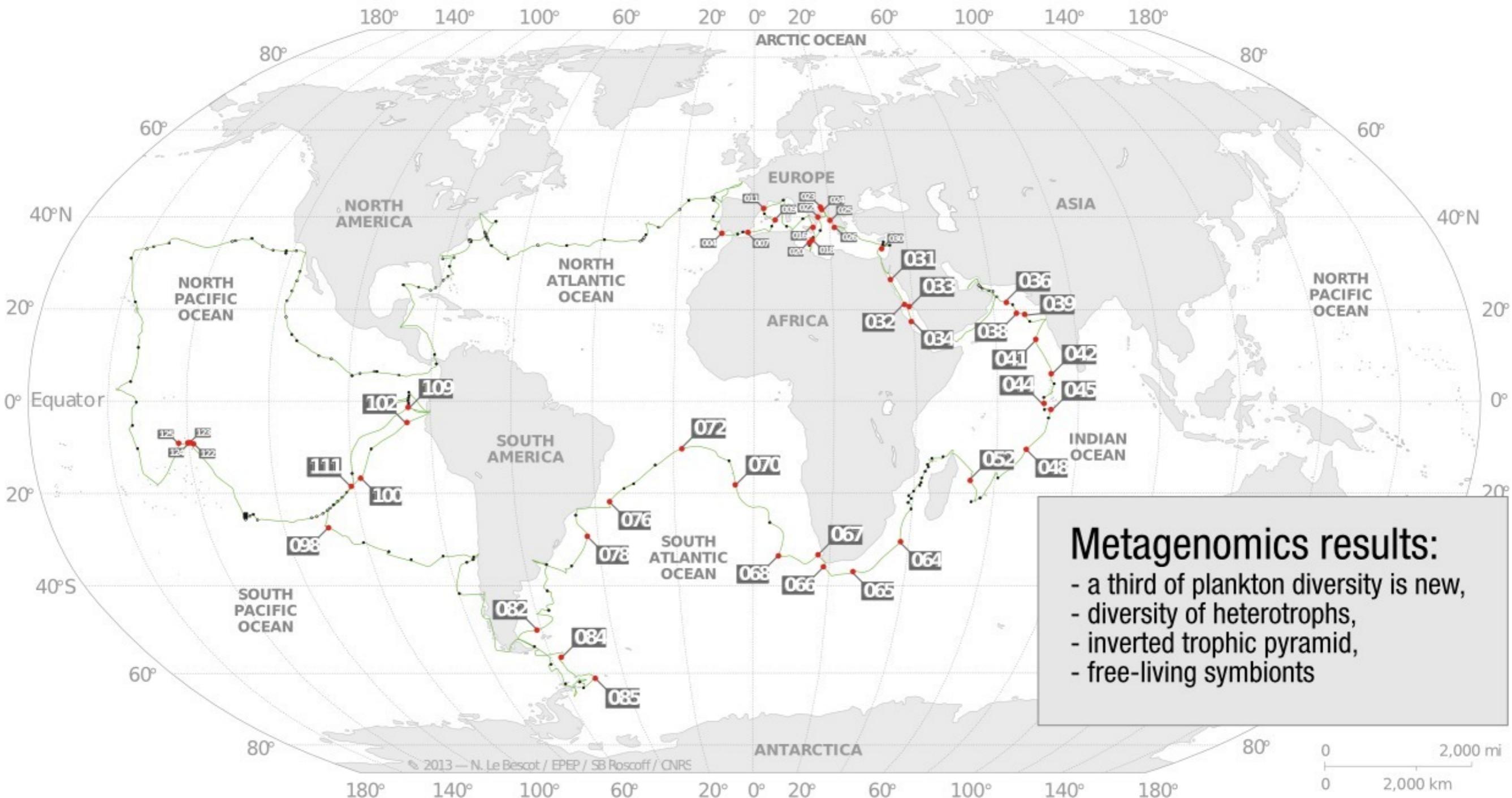
ea51412efcd0b3cc8510436791667c67f186d983	0.0037	48
57e9fa79a7d578ee3fa5dbd4960f8d283b4020b1	0.0042	54
bc42f3dc4a98988d39c23fa5bbf2f3f20c372b66	0.0044	57
8e6a5352d2ba02eb5291ce5a2c4fccc95e5279c6	0.0048	61
282382d34ca8346e1b7968b2084f75249840f819	0.0048	62
092bf0fdd2cf634aba6e3cc5b6ae323db37bbf62	0.0057	73
41873e83fb3666536a7bf0f702cd24805cd11e17	0.0057	73
448b14d7a8657f19f7340da3a31d25652f40b67e	0.0057	73
61c4bb94bf70f1f86e62cad5d7951941a67b8479	0.0057	73
6fc98ad4798e85e9b8b6643786d786701e1eb0d7	0.0057	73
ad967f92c57493caf41f8b86f5bf3397e6bd4a80	0.0057	73
b6b4e3fd88e23a68773ea00a530dcf320826322b	0.0687	73
2a46cc936ef8a263360257420a72921a769440c0	0.0063	80
03dcb427634a4fe6c204a276a9237a3a20c3fe5e	0.0072	92
10dbb1a0a9555c64ad2c0b4f91aec6055175a02b	0.0072	92
121c9a9ac6f8a6fdf1dcf3ca5ecb7c484d8e7fb0	0.0072	92
1222c57cba5c99b5b6013716ec42954f872c27e4	0.0072	92
2600601b600500415f515e11000561051051051051	0.0072	92

Lowest expected error observed per sequence length in a 16S V3-V4 MiSeq 2x250 bp dataset



Filtering by taxonomic assignment



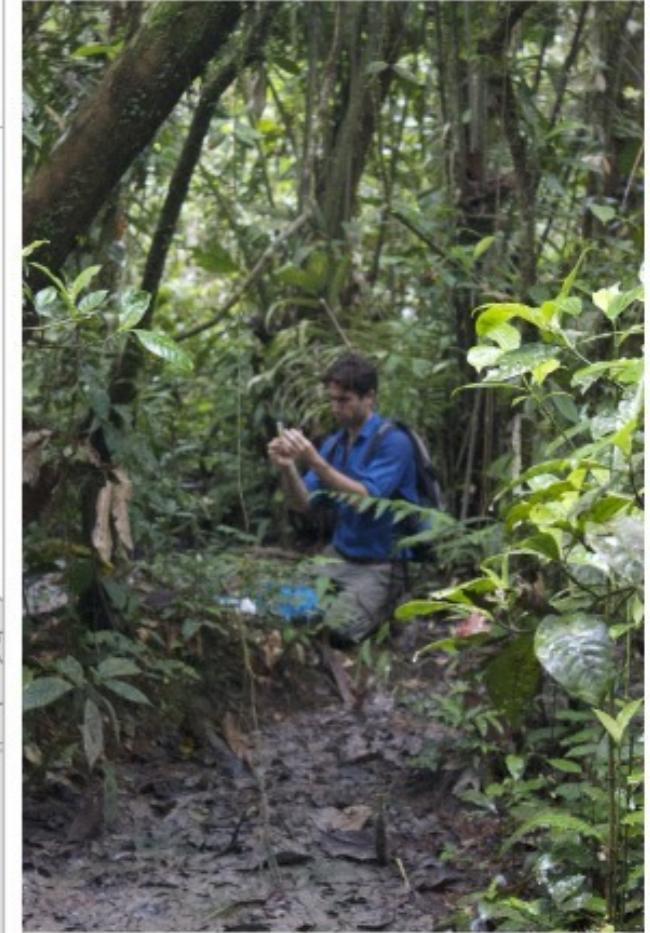


First trip of the TARA OCEANS project
(1.3 billion reads, the 47 sampling stations published so far are in red)

de Vargas et al., 2015 Science

Neotropical Forests Soil Sampling Project

Costa Rica
Panama
Ecuador



Early results

- half of unknowns,
- not-so-many fungi,
- dominance of parasites,
- notable endemism,
- hyperdominant taxa

Microbial diversity at the tree line level

David Wardle & Jordan Mayor, Swedish University of Agricultural Sciences



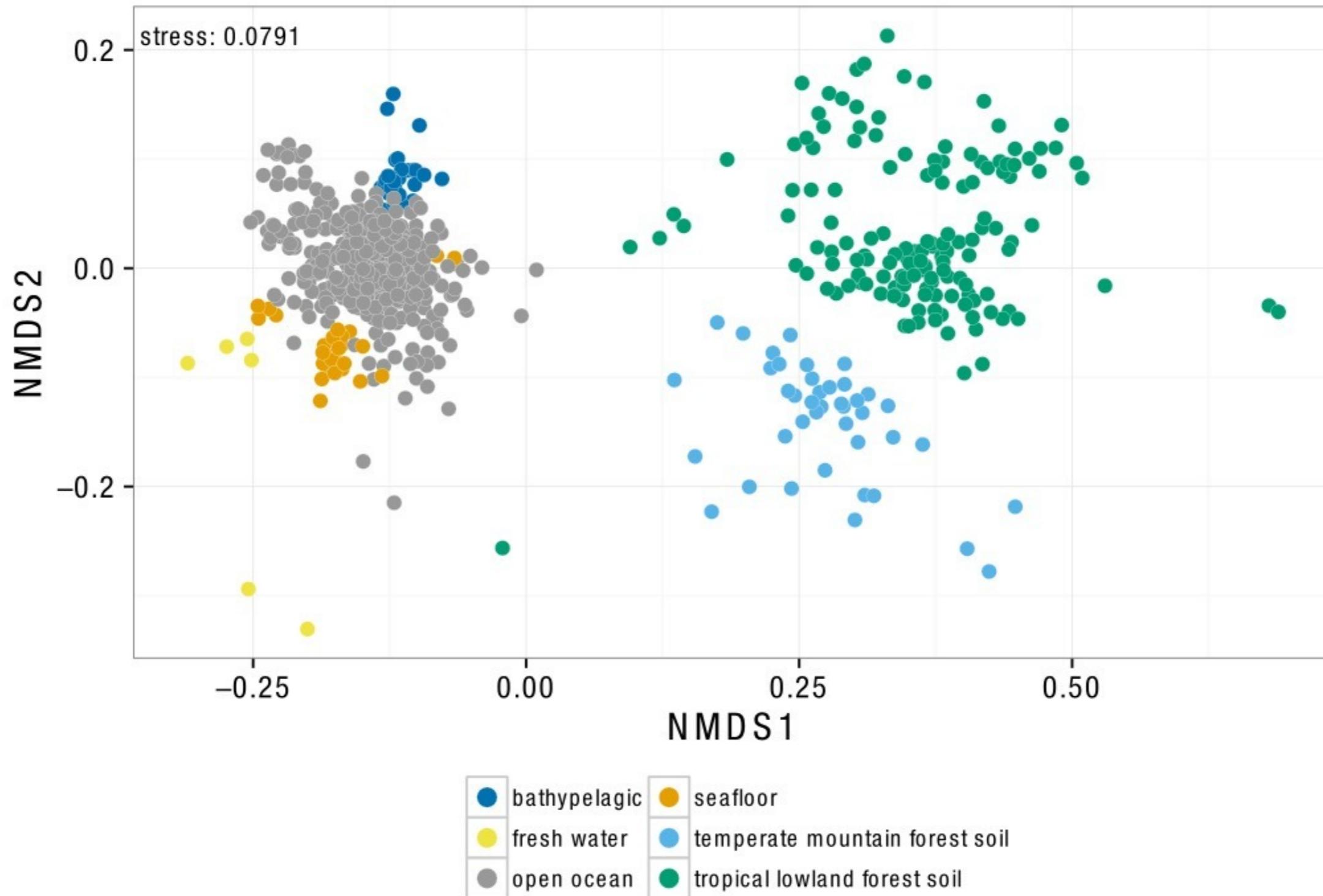
Sampling

- 7 countries (Australia, Austria, Canada, Chile, Japan, New Zealand, USA),
- 5 transects per country,
- 8 replicates per transect,
- 5 soil samples per replicate,
- soil chemistry

Early results

- few unknowns,
- dominance of fungi,
- weak endemism,
- microbial communities?
- geographical analysis?

Meta-analysis of 18S rRNA V4 (672 samples)



What do we need to deal with noise?

- faster and more efficient filters (denoising and clustering)
- better understanding of PCR/sequencer noise
- stronger mathematical background (sequence-space)
- robust α , β statistics able to deal with noise
- repeated experiments (technical/biological replicates)

Technical noise and robust stats
are our main challenges

Thanks!

<https://github.com/torognes/swarm>

<https://github.com/torognes/vsearch>



1 μm

