



Bioinformatics

doi.10.1093/bioinformatics/xxxxxx

Advance Access Publication Date: Day Month Year

Application Note

OXFORD

node.dating: timing ancestral dates in phylogenetic trees in R

Bradley R. Jones 1, 2,* and Art F. Y. Poon 2,3

Associate Editor: XXXXXXX

Received on XXXXX: revised on XXXXX: accepted on XXXXX

Abstract

Summary: Phylogenetic trees encode the evolutionary distances between species or populations. With sufficient information, these evolutionary distances can be rescaled over time to provide estimates of the dates of the most recent ancestors of the species. Here we present the R software node.dating, which uses a maximum likelihood method to estimate the dates of the internal nodes of a phylogenetic tree.

Availability and Implementation: node.dating is written in R and requires the R package, APE.

node.dating is available ()
Contact: brj1@sfu.ca

Supplementary information: Supplementary data are available at *Bioinformatics* online.

1 Introduction

Phylogenetic trees represent the evolutionary relationships among populations or species through their common ancestors. The length of a branch in the phylogeny corresponds to the expected amount of evolution between the ancestor and its descendant, where the passage of time and the rate of evolution are confounded. However, when there is external information available on the location of nodes in the tree in time, then the branch lengths can be rescaled with respect to time given sufficient variation in node timings for measurable evolution to occur. Thus, the internal nodes of a time-scaled tree estimate the dates that the respective lineages diverged from their common ancestor (Kumar and Hedges, 2016). These date estimates are an important resource for reconstructing the evolutionary history of species (Shapiro et al., 2004). In molecular epidemiology, these dates can also provide a rough approximation of transmission times during an outbreak of infectious disease (Ypma et al., 2013).

A multitude of software has been developed to date MRCA's and create time trees using various techniques such as: linear regression (Rambaut et al., 2016), maximum likelihood (Rambaut, 2000; Sanderson, 2003; Yang, 2007), Bayesian analysis (Drummond and Rambaut, 2007), heuristics (Drummond and Rodrigo, 2000; Yang et al., 2007), and least squares (To et al., 2016). However, most of this software is difficult to

access or use. Our software, node.dating, uses a maximum likelihood approach to date the internal nodes of a phylogenetic tree. node.dating is written in R and is compatible with the R package APE (Paradis et al., 2004); it is available (). APE has the capability to estimate the dates of internal nodes via the chronos function; however chronos does not incorporate the dates of the tips into its solution and thus does not offer the same precision as node.dating.

2 Algorithm

We start with a rooted phylogenetic tree with edge length equal to genetic distance and the dates when each tip of the tree was sampled. A simple linear regression is used to estimate the mutation rate assuming a strict molecular clock. To estimate the dates of the internal nodes, we follow an approach given by Felsenstein (1981) and inspired by *TipDates* (Rambaut, 2000). A maximum likelihood method is applied locally to date each internal node. Then using these estimates, the algorithm iterates until a sufficiently approximate maximum likelihood solution is obtained.

3 Runtime and accuracy

In order to test the viability of node.dating, we ran node.dating on 50 simulated trees. We evaluated the results of node.dating using a weighted root mean squared error (RMSE). The simulation method and weighted RMSE are detailed in the Supplementary Information.

We compared the results of node.dating of the 50 simulated trees against a modified version of *TempEst*'s (formerly *Path-O-Gen*) internal

© The Author 2016. Published by Oxford University Press. All rights reserved. For permissions, please e-mail: journals.permissions@oup.com





¹Faculty of Health Sciences, Simon Fraser University, Burnaby, V5A 1S6, Canada,

²BC Centre for Excellence in HIV/AIDS, Vancouver, V6Z 1Y6, Canada and

²Department of Medicine, University of British Columbia, V5Z 1M9, Canada

^{*}To whom correspondence should be addressed.





2 Jones et al.

Table 1. The runtime and accuracy of node . dating and comparable software on $50\ \mathrm{simulated}$ trees

Software	Runtime (s)	RMSE (days)
node.dating(initial)	1.64	29.2
node.dating(1 step)	2.26	26.7
node.dating(10 steps)	7.62	23.7
node.dating (100 steps)	62.1	22.8
node.dating(1000 steps)	596.	22.6
node.dating $(10^4~{ m steps})$	5950	22.1
TempEst	11.2	132.
LSD	0.707	24.9
$BEAST (10^4 \text{ steps})$	181	419.
$BEAST (10^6 \text{ steps})$	6840	20.1

node dating, *LSD* using a weighted root mean squared error (RMSE). We also compared our results using *BEAST*, but on the simulated sequences instead of their reconstructed phylogenies. *TempEst* uses the prediction given by a linear regression to estimate the dates of the internal nodes and *LSD* uses a least squares method.

The average RMSE's of TempEst and LSD are higher than the average weighted RMSE of the MRCA using node.dating with 1000 steps, though the weighted RMSE of LSD is comparable to the RMSE of node.dating. However, the main purpose of TempEst is not to date internal nodes, but to detect/verify the presence of a strict molecular clock. We did not use TempEst or LSD's root-to-tip regression in our experiments because we merely wanted to compare the internal node dating.

BEAST performed the best of all methods when running 10^6 steps, but this also took the most time. With 10^4 steps BEAST performed terribly and in around the same amount of time as node.dating at 100 steps. However, the results using BEAST are not completely comparable since we used the sequence data for BEAST instead of the reconstructed phylogenies.

4 Visualization

In this section, we exhibit a visualization of sequence data using the estimated dates of internal nodes. We retrieved intra-host patient-derived sequences from Patient 16617 on the LANL HIV database (LANL, accessed June 24, 2015). This patient's sequence data was collected in Llewellyn et al. (2006) as Patient 1180. We reconstructed the sequences' phylogenetic tree using the method described in the Supplementary Information. We then estimated the dates of the internal nodes using node.dating with 1000 iterations. Finally we plotted the genetic distance from the root versus time of the internal nodes and the sampled sequences. This plot displayed shown in Figure 1.

5 Future work

One drawback of our methodology is that it assumes that the phylogeny follows a strict molecular clock. However, the local likelihood model can be extended to incorporate a variable molecular clock; future work would like to include this extension. The molecular clock assumption also implies that mutations are strictly additive over time, which is not true. It may also be possible to incorporate this "negative" evolution into the model.

Acknowledgements

We would like to thank Richard Liang for his aid in automating *TempEst*.

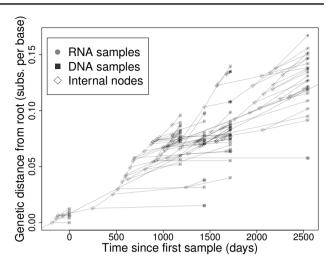


Fig. 1. Genetic distance from the root versus time of the sequences from Patient 16617 from the LANL HIV database. The dates of the internal nodes were estimated using node, dating and the internal nodes are included in the plot. The edges between adjacent nodes of the tree are drawn as solid lines and the molecular clock is drawn by a dashed line.

Funding

This work was supported by the Canadian Institutes of Health Research (CIHR Team Grant: HIV Cure Research — The Canadian HIV Cure Research Enterprise; CanCure), by the Bill and Melinda Gates Foundation Award Number OPP1110049, and by a CIHR operating grant to Art Poon (HOP-111406).

References

Drummond, A. and Rodrigo, R., A G (2000) Reconstructing genealogies of serial samples under the assumption of a molecular clock using serial-sample UPGMA. *Molecular Biology and Evolution*, 17(12), 1807–1815.

Drummond, A. J. and Rambaut, A. (2007) BEAST: Bayesian evolutionary analysis by sampling trees. BMC Evolutionary Biology, 7, 214.

Felsenstein, J. (1981) Evolutionary trees from DNA sequences: a maximum likelihood approach. *Journal of Molecular Evolution*, 17, 368–376.

Kumar, S. and Hedges, S. B. (2016) Advances in time estimation methods for molecular data. *Molecular Biology and Evolution*, 33(4), 863–869.

molecular data. *Molecular Biology and Evolution*, 33(4), 863–869.

LANL (accessed June 24, 2015) Los Alamos HIV Database. URL http://www.hiv.lanl.gov/.

Llewellyn, N. et al. (2006) Continued evolution of HIV-1 circulating in blood monocytes with antiretroviral therapy: genetic analysis of HIV-1 in monocytes and CD4+ T cells of patients with discontinued therapy. *J Leukoc Biol*, 80(5), 1118–1126.

Paradis, E. et al. (2004) APE: analyses of phylogenetics and evolution in R language. Bioinformatics, 20(2), 289–290.

Rambaut, A. (2000) Estimating the rate of molecular evolution: incorporating non-contemporaneous sequences into maximum likelihood phylogenies. *Bioinformatics*, 16(4), 395–399.

Rambaut, A. et al. (2016) Exploring the temporal structure of heterochronous sequences using TempEst (formerly Path-O-Gen). Virus Evolution, 2, vew007.

Sanderson, M. J. (2003) r8s: inferring absolute rates of molecular evolution and divergence times in the absence of a molecular clock. *Bioinformatics*, 19(2), 301– 302.

Shapiro, B. et al. (2004) Rise and fall of the Beringian Steppe bison. *Science*, 306, 1561–1565.

To, T.-H. et al. (2016) Fast dating using least-squares criteria and algorithms. Systematic Biology, 65(1), 82–97.

Yang, Y. (2007) PAML 4: phylogenetic analysis by maximum likelihood. *Molecular Biology and Evolution*, 24(8), 1585–1591.
 Yang, Z. et al. (2007) Tree and rate estimation by local evaluation of heterochronous

rang, z. et al. (2007) Free and rate estimation by local evaluation of neterochronous nucleotide data. *Bioinformatics*, 23(2), 169–176.

Ypma, R. J. F. et al. (2013) Relating phylogenetic trees to transmission trees of

Ypma, R. J. F. et al. (2013) Relating phylogenetic trees to transmission trees of infectious disease outbreaks. *Genetics*, 195(3), 1055–1062.



