

# **OUTBREAK REPORTING**

**Molly Mitchell, PhD, Bioinformatics Supervisor**

# FUNDING STATEMENT

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# INTROS – BRR TEAM



Molly Mitchell, PhD  
Bioinformatics Supervisor  
Molly.Mitchell@flhealth.gov



Nikhil Yengala Reddy, MS  
Bioinformatician  
Nikhil.Yengala@flhealth.gov



Sam Bernhoft, MPH  
Domain Lead Bioinformatician  
Samantha.Bernhoft@flhealth.gov

For any BRR requests, please send an email to [bphl-sebioinformatics@flhealth.gov](mailto:bphl-sebioinformatics@flhealth.gov)  
Based on the requests one of us from the team will respond as soon as possible.

# OVERVIEW

When creating an outbreak report, consider the following:

- The best reports include a cumulation of genotyping, AMR profiling, phylogenetic analysis, and corresponding metadata
- Make conclusions based on considerations from all analyses results
- Use audience appropriate language when making the summary report
- Make sure to omit PPI or identifiable information
  - Only include what may be helpful for drawing conclusions (i.e., collection dates and source)

# FLAQ-AMR

- Using raw [.fastq.gz](#) (zipped *.fastq*), run FL's FLAQ-AMR pipeline
  - [BPHL-Molecular FLAQ-AMR](#)
  - Follow the directions on this GitHub page for git cloning the repository and executing the script.
  - This pipeline also needs python to run, make sure to load using **module load python**
- The FLAQ-AMR output will produce a directory called 'amrfinder\_results'
  - Use this directory to analyze AMR genes found within your sample set
- The output from FLAQ-AMR will also produce a directory called 'annotations' with a [.gff](#) file for each isolate.
  - Copy and paste these files to a new directory using the code below

```
$ cp *.gff /path/to/new/directory/ .
```

# AMRFINDERPLUS

- Used to identify AMR genes and resistance-associated point mutations
- Has an organism flag which will provide extra info related to stress, heat, biocide, and virulence

For the following organisms, run amrfinder –organism

- <https://github.com/evolarjun/amr/wiki/Curated-organisms>

Either use singularity or conda to install and run AMRFinderPlus

```
$ module load conda
$ conda create -yp /blue/bphl-<state>/<user>/conda_envs/amrfinder
$ conda activate /blue/bphl-<state>/<user>/conda_envs/amrfinder
$ conda install -c bioconda ncbi-amrfinderplus
```

# AMRFINDERPLUS

We also covered this on the Office Hours on June 6, 2023

- This is on our [GitHub - StaPH-B Southeast-region](#) page if you need further information

I wrote a loop to run AMRFinderPlus in a directory of **.fasta** files

- This is in /blue/bphl-<state>/share/amrfinderplus/directory
- Edit the **-organism**, your email, and path to amrfinder conda env then **sbatch amrfinder\_plus\_loop.sh**

# FL\_CG\_SNP

In this newly created directory, run FL's FL-cgSNP pipeline

- [BPHL-Molecular FL-cgSNP](#)
- Follow the directions on this GitHub page for git cloning the repository and executing the script.
- This pipeline requires python to run; load using **module load python** prior to running the pipeline
- **Note: this script needs to be run on the group of isolates you're comparing. This pipeline will produce comparative output that can't be merged.**

The output of this pipeline includes a **SNPs\_boot.treefile**

- This file will produce a ML tree using the cgSNP's from a multiple sequence alignment of the shared core genes across your isolates.
- Upload this file to R/RStudio for visualization



# R/RSTUDIO FOR TREE GENERATION

Using the following as a guide,  
use **ggtree** to visualize your  
**SNPs\_boot.treefile**

```
Untitled1* x
Source on Save
Run
Source

1 library(readr)
2 SNPs_boot <- read_csv("path/to/SNPs_boot.treefile")
3 View(SNPs_boot)
4
5 library(ggtree)
6 library(ggplot2)
7 library(tidyverse)
8
9 #Read in tree file with ggtree"
10 tree <- read.tree("path/to/SNPs_boot.treefile")
11
12 ggtree(tree, right=TRUE) + geom_treescale() + geom_tiplab(size=6)
13
14 #Save plot as image
15 ggsave("SNPs_boot_tree.tiff", width = 85, height = 25, units = "cm")

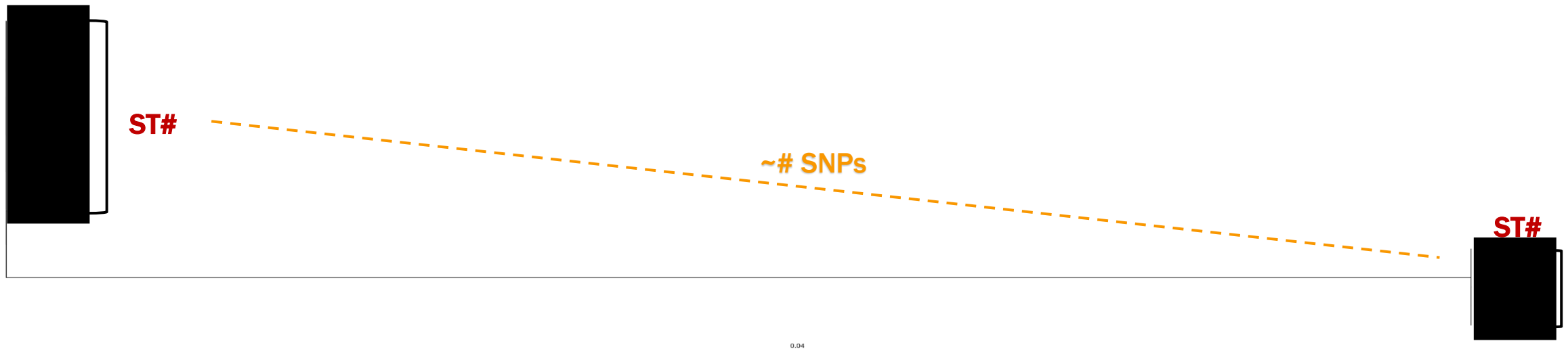
15:23 (Top Level) R Script
```

# ADVANCED R/RSTUDIO FOR TREE GENERATION

If you're proficient in R, use this script written by Dr. Schmedes for labeling clades, etc.

[Data visualization - R scripts \(ggtree\) · GitHub](#)

# ML CGSNP PHYLOGENETIC TREE



- An example of the ggtree output
- Adding ST values and numbers of SNPs between clades can help with concluding results

**\*\*Strain numbers blacked out for privacy**

# SNP MATRIX

- Use the **pairwise\_matrix.tsv** file from the FL-cgSNP output to determine SNP differences between your isolates
- Copy this matrix to an excel file for easy visualization
- Then, use conditional formatting to add a heatmap to the values

snp-dists 0.6.2	Strain 1	Strain 2	Strain 3
Strain 1	0	22	13
Strain 2	22	0	17
Strain 3	13	17	0

# FASTANI

## Start an interactive session

```
$ srun -qos=bphl-umbrella -account=bphl-umbrella -cpus-per-task=10 -mem=50gb -  
time=10:00:00 -pty bash -i
```

## Use conda to install fastANI

```
$ module load conda  
$ conda create -yp /your/path/to/conda/envs/fastani  
$ conda activate /your/path/to/conda/envs/fastani  
$ conda install -c bioconda fastani
```

# FASTANI

Change to the directory of *.fasta* files and run fastANI

```
$ ls *.fasta > strains.txt
```

```
$ fastANI -ql strains.txt -rl strains.txt -output ANIresults.txt -matrix
```

# FASTANI

Copy the results of the **ANIresults.txt.matrix** to excel and apply conditional formatting for easy visualization of the results

	Strain 1	Strain 2	Strain 3
Strain 1	100		
Strain 2	99.99758	100	
Strain 3	99.9997	99.99881	100

# SANIBEL AND TALBOT

- Nextflow versions of both FLAQ-AMR and FL\_cg\_SNP are Sanibel and Talbot – same output, slightly different format
- Both available on our Github: [BPHL-Molecular · GitHub](#)



# OTHER TOOLS – SPECIES DEPENDENT

- mycoSNP - CDC pipeline for *C. auris*
- Ksnp3 or Ksnp4 – highly divergent/variable (*Legionella*)
- Important to research pipelines before using them!
  - Pick what's best for your species or the question you're asking!

# REPORT GENERATION - BEGINNER

1. Use PowerPoint presentation or Microsoft publisher
2. Have an introductory slide with the main points
3. Include a methodology slide with pipelines and versions used in the analysis
4. List table with relevant metadata, without PPI or identifiable information
5. Also include AMR results from the FLAQ-AMR report if applicable
6. Include a ML phylogeny, SNP, and ANI matrix within report
  - Use a slide for each
7. Snipping Tool/screenshots help for imaging

# REPORT GENERATION - INTERMEDIATE

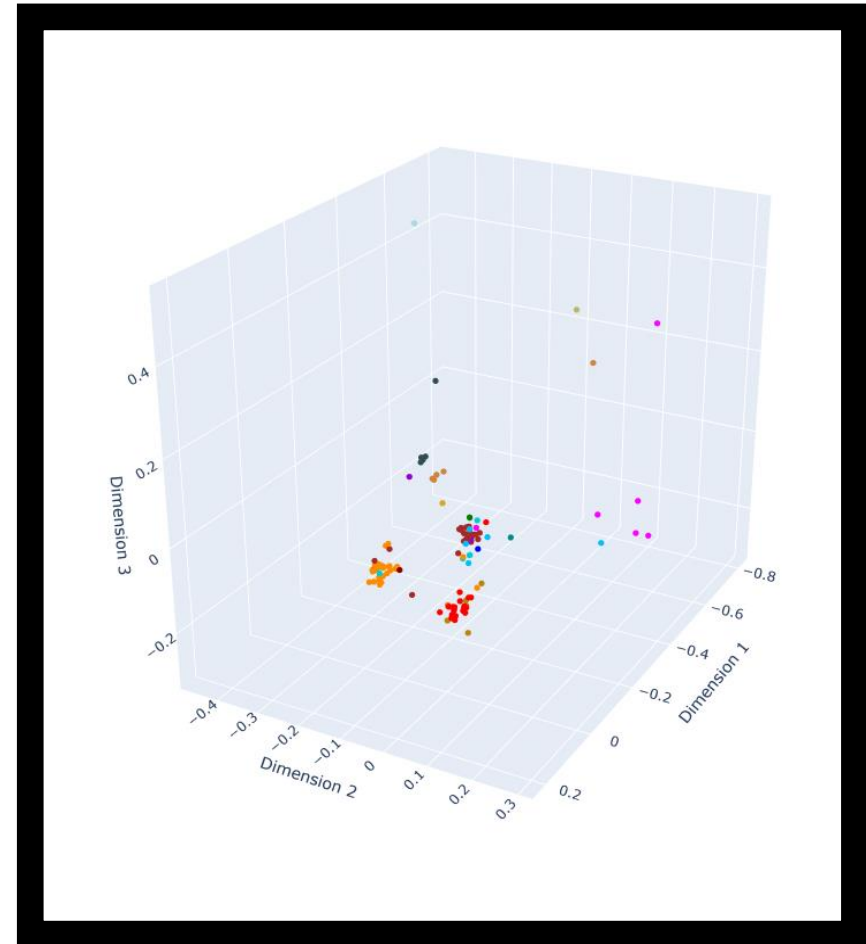
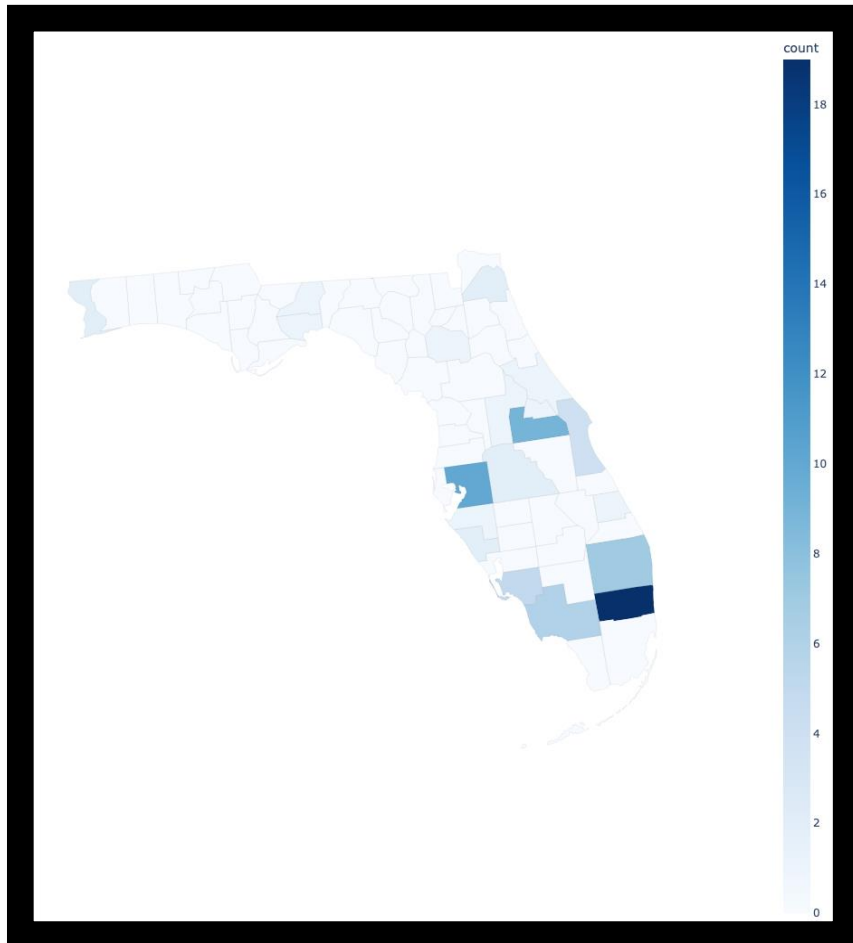
For those of you proficient in R, there is a report generation package called R Markdown

- [R Markdown \(rstudio.com\)](https://rmarkdown.rstudio.com/)
- Fully reproducible
- HTML and PDF options

# REPORT GENERATION - ADVANCED

- Jupyter Notebook & plotly to create an interactive html report
- Geojson interactive maps where we add # of cases
- Plot phylogenies with
- Add 3D MDS modeling plots – another cool way of displaying distance between samples!

# SOME EXAMPLES



# SOME THINGS TO REMEMBER

## Use audience appropriate language

- Use "similar" or "related" instead of "identical" or "from the same source"
- Be ready to describe in detail your analyses and findings to epis or providers

## Add a "surveillance only" disclaimer

- Our NGS analyses and bioinformatics pipelines are not clinically validated and are for surveillance only
- Here's ours: "The results in this report were obtained by WGS, using research use only procedures that were not CLIA validated. They may be used for surveillance and research purposes only. They are not to be used for diagnosis, treatment, or management of patient care."

# NEXT STEPS

**An internal SOP that will be adjusted to external use**

**Files will be hosted on our GitHub - [BPHL-Molecular · GitHub](#)**

- **Geojson, conda environment yml, data processing and report generation ipynb**

**We will provide a training via our Office Hours hosted by Nikhil**



# **Advanced Molecular Detection**

## **Southeast Region Bioinformatics**

# **QUESTIONS?**

[bphl-sebioinformatics@flhealth.gov](mailto:bphl-sebioinformatics@flhealth.gov)

**Molly Mitchell, PhD**

Bioinformatics Supervisor

[Molly.Mitchell@flhealth.gov](mailto:Molly.Mitchell@flhealth.gov)

**Nikhil Yengala Reddy, MS**

Bioinformatician

[Nikhil.Yengala@flhealth.gov](mailto:Nikhil.Yengala@flhealth.gov)