



Advanced Molecular Detection

Southeast Region Bioinformatics

Outbreak/Cluster Report Training

06/12/2023

Outline



Updates - APHL



Agenda



Overview



FLAQ-AMR



AMRFinderPlus



FL-cgSNP



Tree Generation



FastANI



Report Generation



Questions

Updates - APHL

- Easy Genomics and Nextflow Training Application
 - [APHL Easy Genomics and Nextflow Training Application Survey \(surveymonkeys.com\)](https://surveymonkeys.com/s/1234567890)
 - Potentially one person per jurisdiction
 - In-person training in Arlington, VA - **August 14-16**
 - Applications are due by **June 15**
 - Must complete 8-10 hours of pre-work
 - Requires command line knowledge – proficient
- DataCamp subscription renewal/application
 - [DataCamp Membership Application 2023 Survey \(surveymonkeys.com\)](https://surveymonkeys.com/s/1234567890)

Agenda

June 26 – Git & GitHub

July 10 – Bactopia tools

Future Trainings

- ONT & FL's Flisochar pipeline
- StaPH-B Toolkit Programs/Pipelines
- GISAID flagged SARS-CoV-2
- R Training Series
- Dryad pipeline
- Generating R figures
- ...and more

Overview

- When asked to create an outbreak report take the following into consideration
 - The best reports include a cumulation of genotyping, AMR profiling, phylogenetic analysis, and corresponding metadata
 - Make conclusions based on considerations from all results from the analyses
 - Use appropriate language when making the summary report
 - Make sure to omit PPI or identifiable information
 - Only include what may be helpful in drawing conclusions (i.e., collection dates and source)

FLAQ-AMR

- Using raw [.fastq.gz](#), run FL's FLAQ-AMR pipeline
 - [BPHL-Molecular FLAQ-AMR](#)
 - Follow the directions on this GitHub page for git cloning the repository and steps to executing the script.
 - This pipeline also needs python to run, make sure to load using **module load python**
- The FLAQ-AMR output will produce a directory called 'amrfinder_results'
 - Use this to analyze AMR genes found within your sample set
- The output from FLAQ-AMR will produce a directory called 'annotations' with a [.gff](#) file for each isolate.
 - Copy and paste these files to a new directory

```
$ cp *.gff /path/to/new/directory/ .
```

AMRFinderPlus

- AMRFinderPlus has an organism flag which will provide extra info related to stress, heat, biocide, and virulence
- For the following organisms, run amrfinder --organism
 - <https://github.com/evolarjun/amr/wiki/Curated-organisms>
- Either use singularity or conda to install and run AMRFinderPlus

```
$ module load conda  
$ conda create -yp /blue/bphl-<state>/<user>/conda_envs/amrfinder  
$ conda activate /blue/bphl-<state>/<user>/conda_envs/amrfinder  
$ conda install -c bioconda ncbi-amrfinderplus
```

AMRFinderPlus

- We also covered this on the Office Hours on June 6, 2023
 - This is on our [GitHub - StaPH-B Southeast-region](#) page if you need further information
- I wrote a loop to run AMRFinderPlus in a directory of *.fasta* files
 - This is in /blue/bphl-<state>/share/amrfinderplus/ directory
 - Edit the **--organism**, your email, and path to amrfinder conda env then **sbatch amrfinder_plus_loop.sh**

FL-cgSNP

- In this new directory, run FL's FL-cgSNP pipeline
 - [BPHL-Molecular FL-cgSNP](#)
 - Follow the directions on this GitHub page for git cloning the repository and steps to executing the script.
 - This pipeline also needs python to run, make sure to load using **module load python**
 - **Note: this script needs to be run on the group of isolates you're comparing. This pipeline will produce comparative output that can't be merged.**
- The output of this pipeline includes a [SNPs_boot.treefile](#), This file will produce a ML tree using the cgSNP's from a multiple sequence alignment of the shared core genes across your isolates. Upload this file to R/RStudio

R/RStudio for Tree Generation

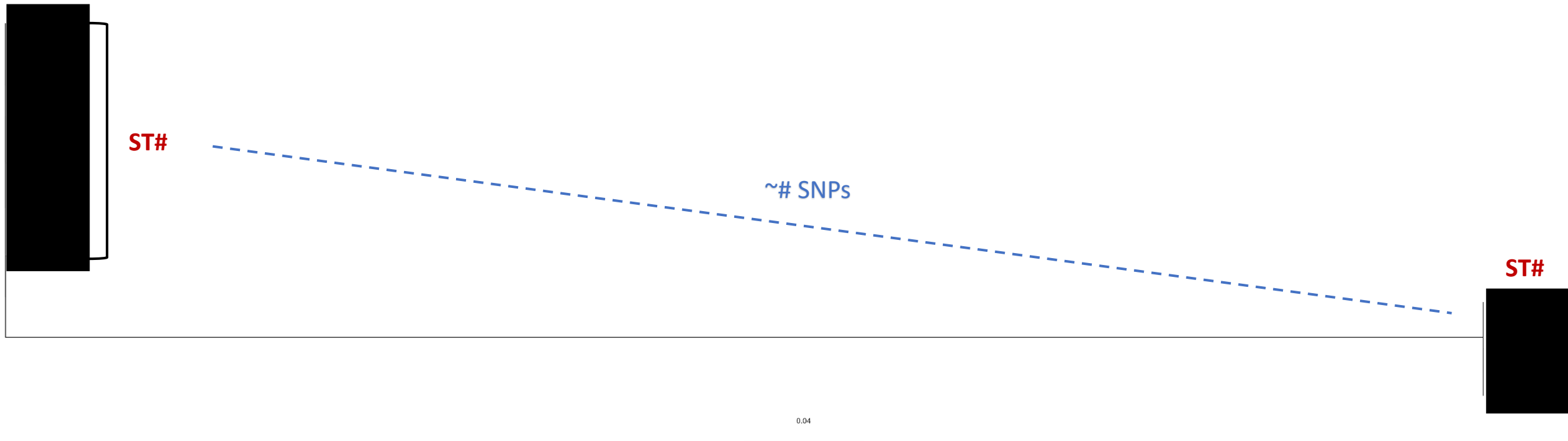
Using the following as a guide, use ggtree to visualize your **SNPs_boot.treefile**

```
1 library(readr)
2 SNPs_boot <- read_csv("path/to/SNPs_boot.treefile")
3 View(SNPs_boot)
4
5 library(ggtree)
6 library(ggplot2)
7 library(tidyverse)
8
9 #Read in tree file with ggtree"
10 tree <- read.tree("path/to/SNPs_boot.treefile")
11
12 ggtree(tree, right=TRUE) + geom_treescale() + geom_tiplab(size=6)
13
14 #Save plot as image
15 ggsave("SNPs_boot_tree.tiff", width = 85, height = 25, units = "cm")
```

R/RStudio for Tree Generation

- If you're proficient in R, use this script written by Dr. Schmedes for labeling clades, etc.
- [Data visualization - R scripts \(ggtree\) · GitHub](#)

ML cgSNP Phylogenetic Tree



- An example of the ggtree output
- Adding ST values and numbers of SNPs between clades can help with concluding results
- Strain numbers blacked out for privacy

SNP Matrix

- Use the [pairwise_matrix.tsv](#) file from the FL-cgSNP output to determine SNP differences between your isolates
- Copy this matrix to an excel file for easy visualization
- Then, use conditional formatting to add a heatmap to the values

snp-dists 0.6.2	Strain 1	Strain 2	Strain 3
Strain 1	0	22	13
Strain 2	22	0	17
Strain 3	13	17	0

FastANI

- Start an interactive session

```
$ srun --qos=bphl-umbrella --account=bphl-umbrella --cpus-per-task=10 --  
mem=50gb --time=10:00:00 --pty bash -i
```

- Use conda to install fastANI

```
$ module load conda  
$ conda create -yp /your/path/to/conda/envs/fastani  
$ conda activate /your/path/to/conda/envs/fastani  
$ conda install -c bioconda fastani
```

FastANI

- Change to the directory of *.fasta* files and run fastANI

```
$ ls *.fasta > strains.txt
```

```
$ fastANI --ql strains.txt --rl strains.txt --output ANIresults.txt --matrix
```

FastANI

- Copy the results of the [ANIresults.txt.matrix](#) to excel and apply conditional formatting for easy visualization of the results

	Strain 1	Strain 2	Strain 3
Strain 1	100		
Strain 2	99.99758	100	
Strain 3	99.9997	99.99881	100

Report Generation

- Use PowerPoint presentation
- Have an introductory slide with the main points
- Include a methodology slide with pipelines and versions used in the analysis
- List table with relevant metadata, without PPI or identifiable information
 - Also include AMR results from the FLAQ-AMR report if applicable
- Include a ML phylogeny, SNP, and ANI matrix within report
 - Use a slide for each
 - Snip Tool helps for imaging
- Use appropriate language
 - Use "similar" or "related" instead of "identical" or "from the same source"

R Markdown

- For those of you proficient in R, there is a report generation package called R Markdown
 - [R Markdown \(rstudio.com\)](https://rmarkdown.rstudio.com/)
 - Fully reproducible
 - HTML and PDF options
- Here in Florida, we're in the process of transitioning from PowerPoint to R Markdown
- Look for an Office Hours on R Markdown

Time for Questions & Feedback

- Questions?
 - Do you need help with anything?
 - Requests for separate trainings?
- Feedback
 - What would you like to see?



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Questions?

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