

RMANOVA

=====

This R markdown document provides an example of performing a repeated measures ANOVA using the `jmv::anovaRM()` function in the `jmv` (Jamovi) package.

Package management in R

-----

```
``` r
# keep a list of the packages used in this script
packages <- c("tidyverse","rio","jmv")
```
```

This next code block has `eval=FALSE` because you don't want to run it when knitting the file. Installing packages when knitting an R notebook can be problematic.

```
``` r
# check each of the packages in the list and install them if they're not
installed already
for (i in packages){
  if(! i %in% installed.packages()){
    install.packages(i,dependencies = TRUE)
  }
  # show each package that is checked
  print(i)
}
```
```

```
``` r
# load each package into memory so it can be used in the script
for (i in packages){
  library(i,character.only=TRUE)
  # show each package that is loaded
  print(i)
}
```
```

```
## -- Attaching packages ----- tidyverse
1.3.0 --
```

```
## v ggplot2 3.3.3      v purrr  0.3.4
## v tibble  3.0.6      v dplyr  1.0.4
## v tidyr   1.1.2      v stringr 1.4.0
## v readr   1.4.0      v forcats 0.5.1
```

```
## -- Conflicts -----
tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()

## [1] "tidyverse"
## [1] "rio"
## [1] "jmv"
```

RMANOVA is a linear model

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The RMANOVA is a type of linear model. However the RMANOVA is a simplified for with additional constraints which allows us to analyze data which violate the independence of observations assumption which is required for the linear models we have used to this point. A nice explanation is provided in Field chapter 15.4.

Open data file

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The rio package works for importing several different types of data files. We're going to use it in this class. There are other packages which can be used to open datasets in R. You can see several options by clicking on the Import Dataset menu under the Environment tab in RStudio. (For a csv file like we have this week we'd use either From Text(base) or From Text (readr). Try it out to see the menu dialog.)

```
``` r
# Using the file.choose() command allows you to select a file to import from
another folder.
dataset <- rio::import(file.choose())
# This command will allow us to import a file included in our project folder.
# dataset <- rio::import("Album Sales.sav")
```
```

Get R code from Jamovi output

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You can get the R code for most of the analyses you do in Jamovi.

1. Click on the three vertical dots at the top right of the Jamovi window.
2. Click on the Syndax mode check box at the bottom of the Results section.
3. Close the Settings window by clicking on the Hide Settings arrow at the top right of the settings menu.
4. you should now see the R code for each of the analyses you just ran.

#### Visualization with ggplot

```
``` r
# plots for outcome split by groups
ggplot(dataset, aes(x = stick))+
  geom_histogram(binwidth = 1, color = "black", fill = "white")
```
```



```
``` r
ggplot(dataset, aes(x = testicle))+
  geom_histogram(binwidth = 1, color = "black", fill = "white")
```
```

```
![] (Week-12-RMANOVA-Assignment_files/figure-markdown_github/unnamed-
chunk-5-2.png)
```

```
``` r
ggplot(dataset, aes(x = eye))+
  geom_histogram(binwidth = 1, color = "black", fill = "white")
```
```

```
![] (Week-12-RMANOVA-Assignment_files/figure-markdown_github/unnamed-
chunk-5-3.png)
```

```
``` r
ggplot(dataset, aes(x = witchetty))+
  geom_histogram(binwidth = 1, color = "black", fill = "white")
```
```

```
![] (Week-12-RMANOVA-Assignment_files/figure-markdown_github/unnamed-
chunk-5-4.png)
```

RMANOVA in jmv package

```
-----

``` r
output = jmv::anovaRM(
  data = dataset,
  rm = list(
    list(
      label="Food",
      levels=c(
        "stick",
        "testicle",
        "eye",
        "witchetty"))),
  rmCells = list(
    list(
      measure="stick",
      cell="stick"),
    list(
      measure="testicle",
      cell="testicle"),
    list(
      measure="eye",
      cell="eye"),
    list(
      measure="witchetty",
      cell="witchetty")),
  effectSize = c("ges", "eta", "partEta"),
  depLabel = "Seconds",
  rmTerms = ~ Food,
  spherTests = TRUE,
  spherCorr = c("none", "GG", "HF"),
  postHoc = list(
    "Food"),
  postHocCorr = c(
    "bonf",
```

```

    "none",
    "tukey",
    "scheffe",
    "holm"),
  emMeans = ~ Food,
  emmTables = TRUE)
```

## Warning: attributes are not identical across measure variables; they
will be
## dropped

``` r
output
```

##
## REPEATED MEASURES ANOVA
##
## Within Subjects Effects
##
-----
##          Sphericity Correction      Sum of Squares      df
Mean Square      F          p      <U+03B7>^2-G      <U+03B7>^2
<U+03B7>^2-p
##
-----
##      Food      None      83.12500      3
27.708333      3.793806      0.0255703      0.3274249      0.3274249      0.3514799
##      Greenhouse-Geisser      83.12500      1.598537
52.00068      3.793806      0.0625841      0.3274249      0.3274249      0.3514799
##      Huynh-Feldt      83.12500      1.997291
41.61888      3.793806      0.0483306      0.3274249      0.3274249      0.3514799
##
##      Residual      None      153.37500      21
7.303571
##      Greenhouse-Geisser      153.37500      11.189757
13.70673
##      Huynh-Feldt      153.37500      13.981036
10.97022
##
-----
##      Note. Type 3 Sums of Squares
##
##
## Between Subjects Effects
##
-----
##          Sum of Squares      df      Mean Square      F      p
<U+03B7>^2-G      <U+03B7>^2      <U+03B7>^2-p
##
-----
##      Residual      17.37500      7      2.482143
##
-----
##      Note. Type 3 Sums of Squares

```

```

##
##
## ASSUMPTIONS
##
## Tests of Sphericity
##
-----
##          Mauchly's W      p          Greenhouse-Geisser e      Huynh-
Feldt e
##
-----
##      Food      0.1362480      0.0468458          0.5328456
0.6657636
##
-----

##
##
## POST HOC TESTS
##
## Post Hoc Comparisons - Food
##
-----
t      ##      Food      Food      Mean Difference      SE      df
      ##      p      p-tukey      p-scheffe      p-bonferroni      p-holm
-----
##      stick      -      testicle      3.8750000      1.351256
21.00000      2.86770228      0.0092136      0.0422912      0.0688661      0.0552818
0.0460681
##      -      eye      4.0000000      1.351256
21.00000      2.96020881      0.0074701      0.0347801      0.0578305      0.0448205
0.0448205
##      -      witchetty      2.3750000      1.351256
21.00000      1.75762398      0.0933811      0.3206962      0.3996166      0.5602868
0.3735245
##      testicle      -      eye      0.1250000      1.351256
21.00000      0.09250653      0.9271724      0.9997067      0.9997827      1.0000000
0.9271724
##      -      witchetty      -1.5000000      1.351256
21.00000      -1.11007830      0.2795153      0.6874563      0.7469435      1.0000000
0.7275647
##      eye      -      witchetty      -1.6250000      1.351256
21.00000      -1.20258483      0.2425216      0.6320079      0.6982714      1.0000000
0.7275647
##
-----

##
##
## ESTIMATED MARGINAL MEANS
##
## FOOD
##
## Estimated Marginal Means - Food
##
-----
##      Food      Mean      SE      Lower      Upper
##
-----

```

```
##      stick      8.125000    0.8730846    6.327076    9.922924
##      testicle   4.250000    0.8730846    2.452076    6.047924
##      eye        4.125000    0.8730846    2.327076    5.922924
##      witchetty  5.750000    0.8730846    3.952076    7.547924
##      -----
```

![] (Week-12-RMANOVA-Assignment\_files/figure-markdown\_github/unnamed-chunk-6-1.png)

Calculate omega squared  
-----

Field prefers to use omega squared for the effect size. He shows how to calculate it using the information from the RMANOVA tables in section 15.11

```
``` r
omegaSquare = (((4-1)/(8*4))*(27.71 - 7.3))/(7.3 + ((2.48 - 7.3)/4) + (((4-1)/(8*4))*(27.71 - 7.3)))
omegaSquare
```

## [1] 0.2389277
```