

IntelliFold-2 Release Notes

Surpassing AlphaFold 3 via Architectural Refinement and Structural Consistency

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IntelliGen-AI — February 7, 2026

Introduction

Following our initial release [1], we are excited to present **IntelliFold-2**. This major architectural update establishes a new state-of-the-art as one of the first open-source models to outperform AlphaFold 3 [2] on Foldbench [3].

Three model variants are provided:

- **IntelliFold-2-Flash**: A fast, efficient model intended primarily for academic use and easy fine-tuning, featuring our updated data curation and multiscale structural representations with 12 standard Pairformer blocks.
- **IntelliFold-2**: Our most accurate open-source model, featuring 48 widened Pairformer blocks with latent space scaling.
- **IntelliFold-2-Pro**: Our server-side flagship model. In addition to all architectural improvements, it incorporates exclusive PPO-enhanced sampling and Difficulty-Aware Loss optimization for maximum precision.

Benchmark Performance

This release emphasizes performance improvements in structurally challenging yet therapeutically relevant categories, with a focus on **antibody-antigen interactions** and **protein-ligand co-folding**.

Building on the capabilities of the initial release, IntelliFold-2 now establishes a new state-of-the-art by outperforming AlphaFold 3 in these critical categories.

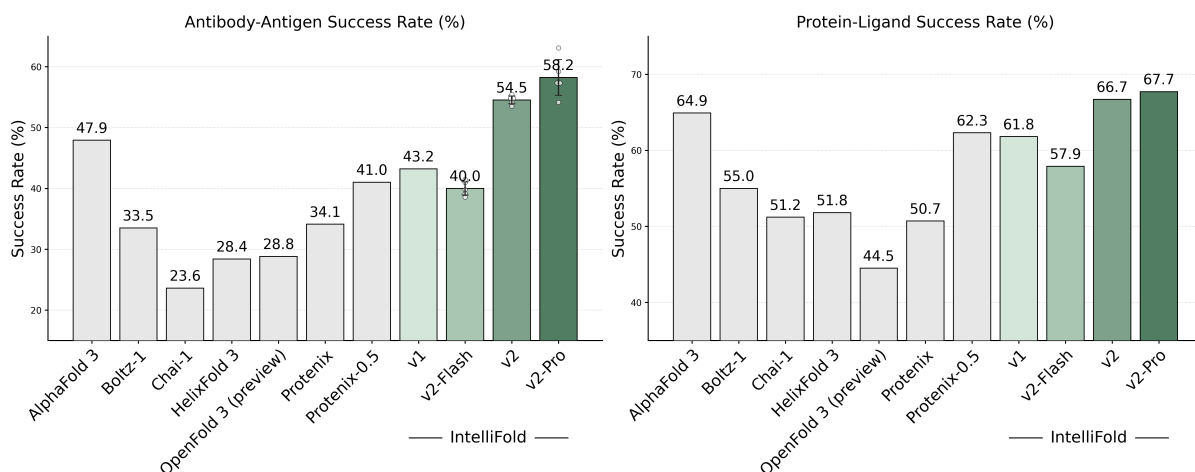


Figure 1: Performance on Foldbench [1, 2, 4–8]. IntelliFold-2 (v2 and v2-Pro) demonstrates a significant lead over AlphaFold 3 in Antibody-Antigen interactions and Protein-Ligand co-folding. Success is defined as DockQ > 0.23 for Antibody-Antigen and IRMSD < 2Å with LDDT-PLI > 0.8 for Protein-Ligand interactions. For ABAG, the reported v2-model results are aggregated over 5 runs, with raw datapoints provided.

Please send correspondence regarding this report to contact@intfold.com.

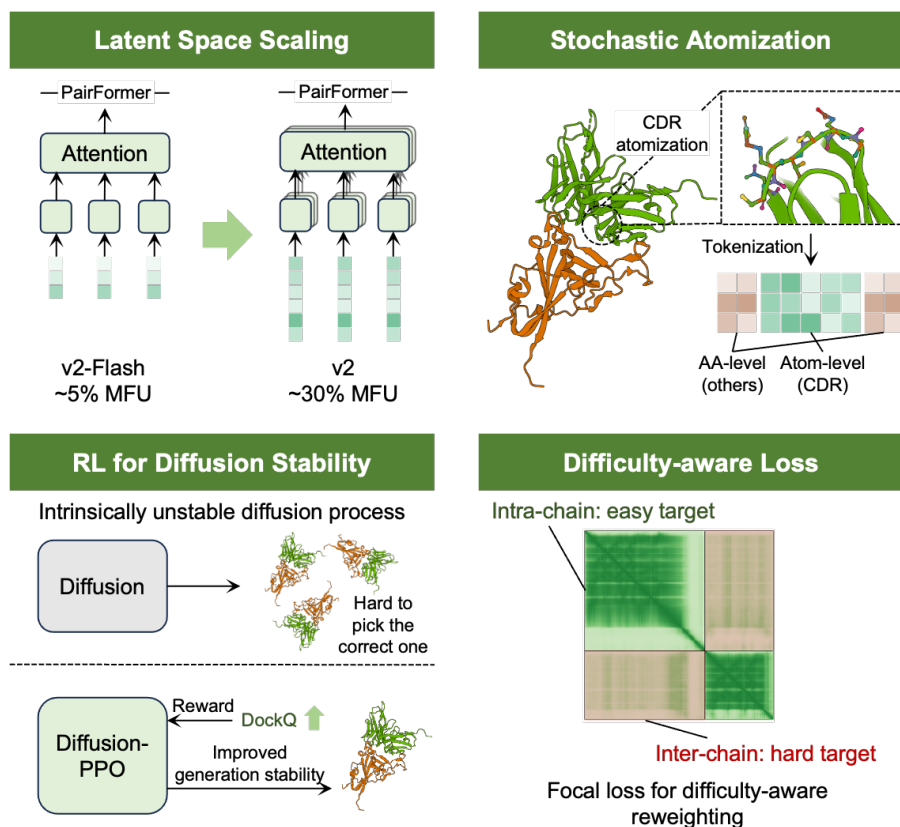


Figure 2: Key architectural and algorithmic innovations in IntelliFold-2

Key Technical Changes

Scaling Model Capacity and Computational Efficiency

Empirical analysis revealed that the previous hidden dimension bottlenecked both representational capacity and computational efficiency.

- **Latent Space Scaling:** We increase the dimensionality of latent representations within the PairFormer blocks, significantly enhancing the model’s capacity to capture complex biological interactions.
- **Improved Hardware Utilization:** Larger hidden dimensions increase arithmetic intensity, leading to substantially higher Model FLOPs Utilisation (MFU) and more effective use of modern GPU architectures.

Principled Multiscale Structural Representations

We refine the training procedure to better align atomic-level accuracy with global structural consistency.

- **Consistent Representations Across Scales:** We revise the atom attention mechanism to a more principled formulation, ensuring robust and self-consistent model behaviour during training and inference.
- **Stochastic Atomization:** Atom-level tokenization is applied stochastically to improve robustness and strengthen the model’s ability to capture fine-grained atomic interactions.

Policy-Guided Optimization for Diffusion Sampling

We apply reinforcement learning, instantiated via Proximal Policy Optimization (PPO), to fine-tune the diffusion module and improve sampling reliability at inference time.

- **Policy-Guided Sampling:** We frame the diffusion sampler as a stochastic policy and apply PPO updates to encourage trajectories that yield structurally coherent and physically plausible conformations.
- **Reduction of Random Sampling Failures:** By constraining policy updates through the clipped PPO objective, the fine-tuning process suppresses unstable or low-quality trajectories, leading to more robust sampling and fewer random failures during inference.

Difficulty-Aware Loss Reweighting

To address imbalance in training signals across structural difficulty levels, we adopt a focal-loss-style reweighting formulation.

- **Emphasis on Hard Examples:** Focal loss down-weights well-predicted samples and allocates more gradient signal to challenging regions of the structural space, such as flexible loops and ambiguous side-chain configurations.
- **Improved Optimization Dynamics:** This re-weighting leads to more stable convergence and improves accuracy on structurally complex targets without disproportionately affecting easy cases.

High-Fidelity and Scalable Data Curation

We re-processed the Protein Data Bank (PDB) dataset using an updated pipeline to ensure high-fidelity supervisory signals. Furthermore, we scaled up the self-distillation dataset and optimised its distribution. This expansion enriches the diversity of training examples and enhances the model’s generalisation capabilities across complex targets.

Usage and Availability

IntelliFold-2-Flash and IntelliFold-2 are available at <https://github.com/IntelliGen-AI/IntelliFold>. IntelliFold-2-Pro is available to Pro users via our online server at <https://server.intfold.com>.

Table 1: Comparative performance metrics across diverse biomolecular systems. We report LDDT for monomers and % DockQ > 0.23 for interaction systems.

Model	Protein Monomer	RNA Monomer	Protein-Protein	Protein-RNA
AlphaFold 3	0.88	0.61	72.9	62.3
IntelliFold-1	0.88	0.63	72.9	58.9
IntelliFold-2-Flash	0.88	0.55	73.6	56.5
IntelliFold-2	0.89	0.58	71.9	68.3

Author Contributions

Lifeng Qiao developed the majority of the technical changes. **He Yan** refined the atom local attention and validated the impact of self-distillation. **Gary Liu** and **Gaoxing Guo** contributed to building the distillation datasets. **Siqi Sun** led the project.

References

- [1] The IntFold Team, Leon Qiao, Wayne Bai, He Yan, Gary Liu, Nova Xi, Xiang Zhang, and Siqi Sun. Intfold: A controllable foundation model for general and specialized biomolecular structure prediction. *arXiv preprint arXiv:2507.02025*, 2025.
- [2] Josh Abramson, Jonas Adler, Jack Dunger, Richard Evans, Tim Green, Alexander Pritzel, Olaf Ronneberger, Lindsay Willmore, Andrew J Ballard, Joshua Bambrick, et al. Accurate structure prediction of biomolecular interactions with alphafold 3. *Nature*, 630(8016):493–500, 2024.
- [3] Sheng Xu, Qiantai Feng, Lifeng Qiao, Hao Wu, Tao Shen, Yu Cheng, Shuangjia Zheng, and Siqi Sun. Benchmarking all-atom biomolecular structure prediction with foldbench. *Nature Communications*, 2025.
- [4] Jeremy Wohlwend, Gabriele Corso, Saro Passaro, Noah Getz, Mateo Reveiz, Ken Leidal, Wojtek Swiderski, Liam Atkinson, Tally Portnoi, Itamar Chinn, et al. Boltz-1 democratizing biomolecular interaction modeling. *BioRxiv*, pages 2024–11, 2025.
- [5] Chai Discovery team, Jacques Boitreaud, Jack Dent, Matthew McPartlon, Joshua Meier, Vinicius Reis, Alex Rogozhonikov, and Kevin Wu. Chai-1: Decoding the molecular interactions of life. *BioRxiv*, pages 2024–10, 2024.
- [6] Lihang Liu, Shanzhuo Zhang, Yang Xue, Xianbin Ye, Kunrui Zhu, Yuxin Li, Yang Liu, Jie Gao, Wenlai Zhao, Hongkun Yu, et al. Technical report of helixfold3 for biomolecular structure prediction. *arXiv preprint arXiv:2408.16975*, 2024.
- [7] The OpenFold3 Team. Openfold3-preview, 2025. URL <https://github.com/aqlaboratory/openfold-3>.
- [8] ByteDance AML AI4Science Team, Xinshi Chen, Yuxuan Zhang, Chan Lu, Wenzhi Ma, Jiaqi Guan, Chengyue Gong, Jincai Yang, Hanyu Zhang, Ke Zhang, et al. Protenix-advancing structure prediction through a comprehensive alphafold3 reproduction. *BioRxiv*, pages 2025–01, 2025.