

Biocomputing

Elements of Biotechnology

Content

- Introduce the methods used in the lab and in the cell itself to manipulate DNA
 - The availability of such tools made the rapid development of the field possible
 - It is not a lab manual, only present a number of available tools and the ideas supporting them
 - Essential to know about elementary lab tools also for mathematicians and computer scientists
- Topics: measuring DNA, testing for/isolating DNA molecules, separating and fusing DNA strands, lengthening, shortening, cutting, linking DNA, modifying nucleotides, multiplying, sequencing

Measuring the length of DNA molecules

- **Problem:** measure the length of a molecule without sequencing it
- Length – definition:
 - Single stranded DNA: the number of nucleotides (ex: 12 mer)
 - Double stranded DNA: the number of base pairs (ex: 12 bp)

Measuring the length of DNA

- **Central fact:** DNA molecules are negatively charged: in electric fields, they migrate towards + electrodes
- The force needed to move DNA molecules – proportional to its length (due to friction)
- Idea: gel electrophoresis technique

The gel electrophoresis technique

- Pour some gel in a rectangular container with electrodes on the sides
- In the cooling process: a comb is inserted on the negative side
- Gel cooled down, comb removed: row of small wells

Gel electrophoresis

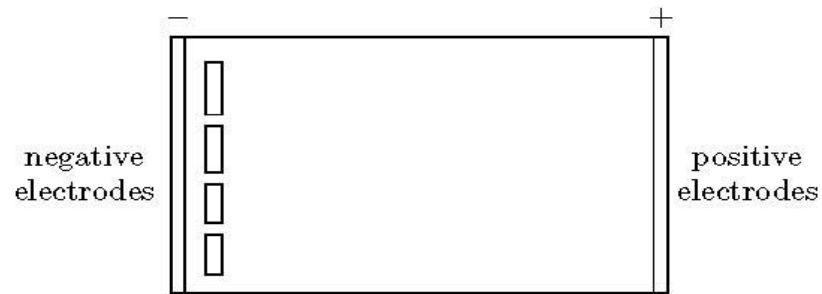


Figure 1.8: Gel prepared for electrophoresis

The gel electrophoresis technique

- The solution with the DNA to be measured brought in the wells
- Activate the electric field: DNA molecules travel towards +
- Gel acts as a molecular sieve – *big friction*
 - *small molecules move faster than big molecules*

The gel electrophoresis technique

- Deactivate the field when the first molecules have reached +
- Small molecules – longer distance than bigger molecules
- Molecules of same length – same distance

Gel electrophoresis

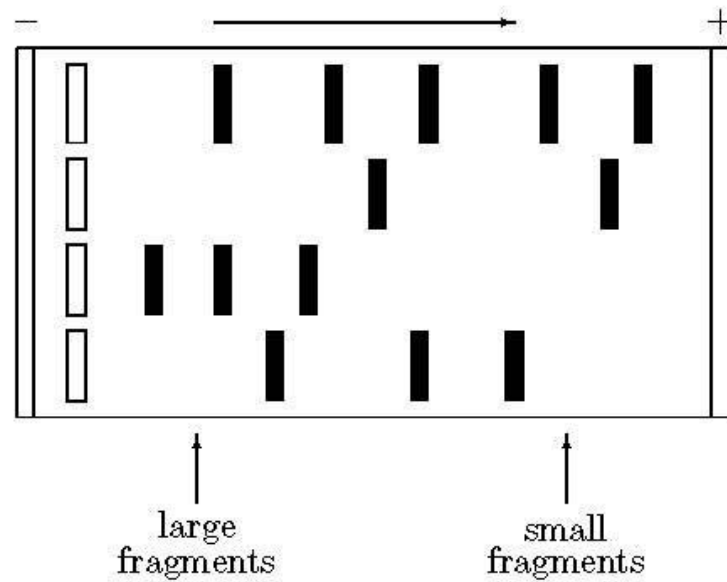


Figure 1.9: Gel electrophoresis

The gel electrophoresis technique

- Reading the resulting gel
- DNA molecules are colorless: mark them
- Two techniques for marking:
 - Staining with ethridium bromide → fluorescent mark under ultraviolet light
 - Attaching radioactive markers to the ends of DNA molecules → expose a film

Computing the length

- Based on the distance traveled
- Using a calibrating solution in one of the wells → compare the location of bands on the other paths with the calibrating path

Testing for known molecules

- **Problem:** test if some known single stranded molecules *s* exist in a given DNA solution;
variant: isolate some of them if so
- **Solution:** Attach the complementary strands (*probes*) to a filter and pour the solution through the filter
- *s* molecules bind to their complements to form double strands (*annealing*) and stay on the filter

Testing for/isolating known molecules

- Other methods:
 - Attach probes to tiny glass beads, place them in a glass column and pour the solution over them
 - Attach probes to tiny magnetic beads, throw them in the solution and stir; attract them to one side using a magnet

Operations on DNA molecules

- Separating and fusing DNA strands
- Lengthening DNA
- Shortening DNA
- Cutting DNA
- Linking DNA
- Modifying nucleotides
- Multiplying DNA (cloning, PCR)
- Sequencing

Separating and fusing DNA strands

- Hydrogen bond between complementary bases is weaker than the covalent bond between consecutive nucleotides within one strand
- One can separate the two strands of a DNA molecule without breaking the single strands
- Separating the two strands: *denaturation*
- Fusing two complementary strands: *annealing/renaturation/hybridization*

Denaturation and annealing

- *Denaturation*: heat the DNA solution until DNA melts
 - Melting temperature: 85 – 95 C
 - Use of chemicals (e.g., *formamide*) lowers the melting temperature
 - Melting temperature of DNA = the temperature at which half of the molecules separate
- *Annealing*: solution cooled down (slowly), the separate strands fuse again by hydrogen bonds (also called *renaturation* or *hybridization*)

Enzymes

- Enzymes: proteins that catalyze biochemical reactions
 - Very specific: most catalyze just a single chemical reaction
 - Highly efficient: speed up the chemical reaction by as much as a *trillion* times
- If no enzymes existed, bio-reactions would most likely be too slow to support life

Lengthening DNA

- The enzymes: *polymerases* - add nucleotides to an existing DNA molecule
- Requirements:
 - Existing single stranded template describing the chain of nucleotides to be added
 - Existing sequence (primer) bonded to a part of the template

Lengthening DNA

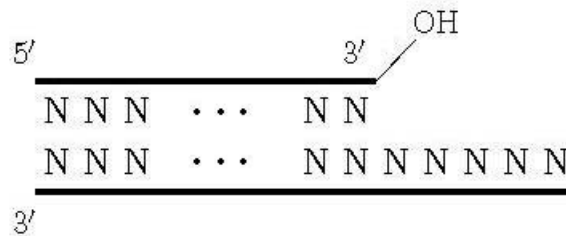


Figure 1.10: A DNA molecule with an incomplete upper strand

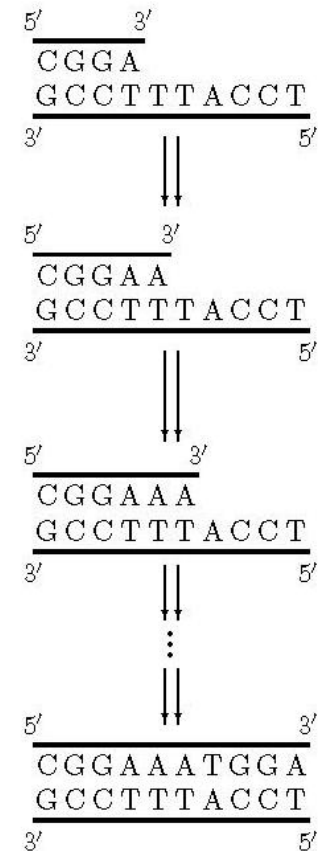


Figure 1.11: A polymerase in action

Lengthening DNA - polymerases

- Polymerases only extend the DNA molecules in the 5'-3' direction
- They extend repeatedly the 3' end, provided that the required nucleotides are available in the vicinity
- Given one strand, to produce the corresponding double stranded molecule, one needs the primer and the polymerases

Lengthening DNA

- Exceptions: *terminal transferase* does not need a template: extends double strands at both ends by some single stranded tails

Lengthening DNA

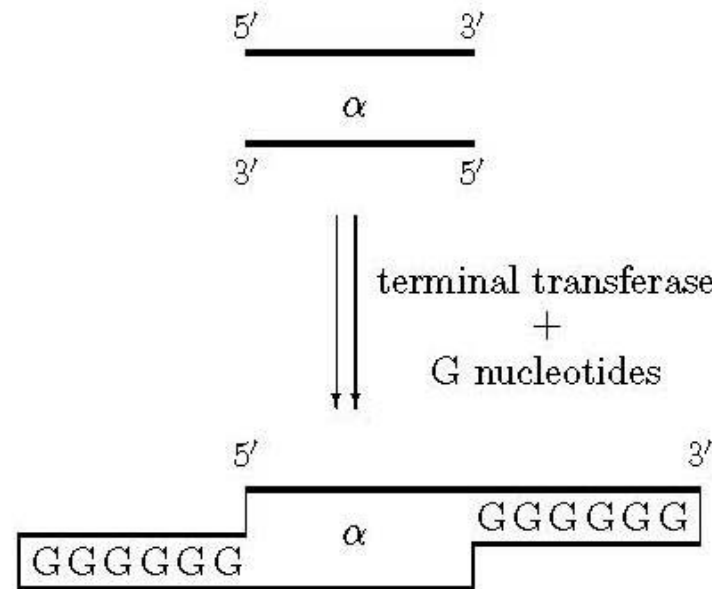


Figure 1.12: Transferase activity

Lengthening DNA

- Exceptions: *terminal transferase* does not need a template: extends double strands at both ends by some single stranded tails
- Engineering single strands is possible: a procedure leading to automation exists → “synthesizing robots”
 - Short such single strands are called *oligonucleotides* (*oligos* in short); they are essential in PCR

Shortening DNA

- The enzymes: *DNA nucleases* – degrade DNA
- Two types of enzymes:
 - *DNA exonucleases*: cleave (remove) one nucleotide at a time from the ends of the DNA
 - *DNA endonucleases*: destroy internal bonds in the DNA molecule (cutting DNA)

Shortening DNA

- *Exonucleases*:
 - some remove nucleotides from 5', others from 3'
 - some degrade single strands, others on double strands, other operate on both
- Examples:
 - Polymerases can act as exonucleases: error correction in DNA replication
 - Exonuclease III: 3'-nuclease
 - Bal 31

Exonucleases

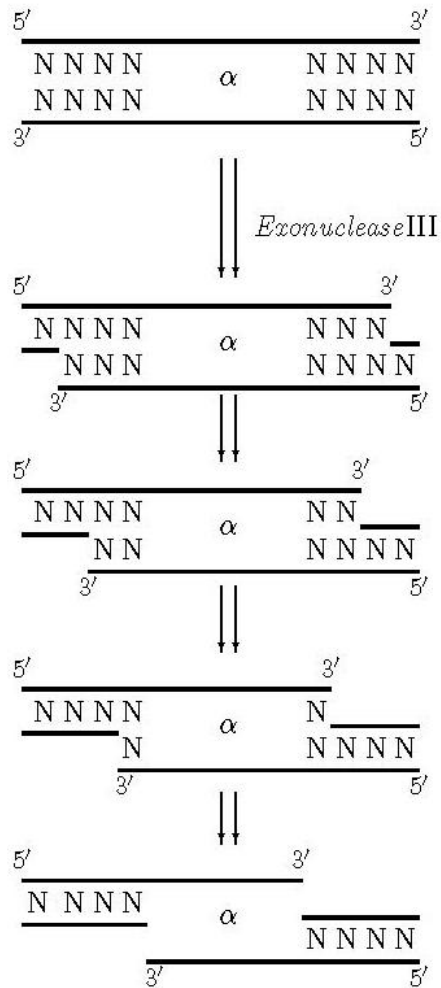


Figure 1.13: Exonuclease III in action

Exonucleases

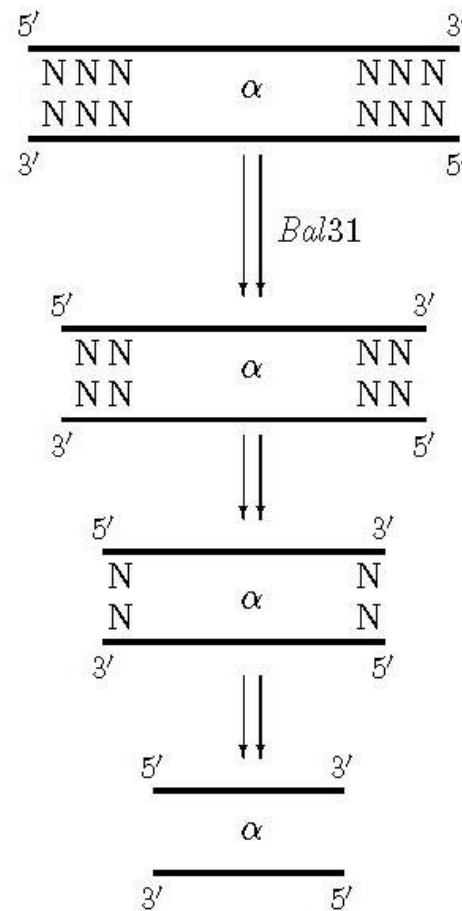


Figure 1.14: Exonuclease *Bal31* in action

Cutting DNA

- *Endonucleases*: destroy internal covalent bonds in DNA molecules
- Can be quite specialized: what/where/how they cut
- Examples:
 - S1 – not site specific
 - DnaseI – not site specific

Endonucleases

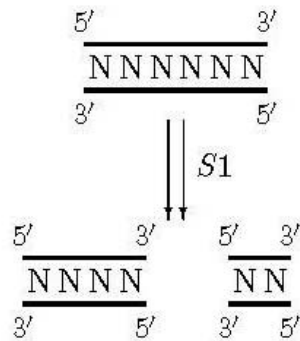


Figure 1.15: *S1* endonuclease in action (i)

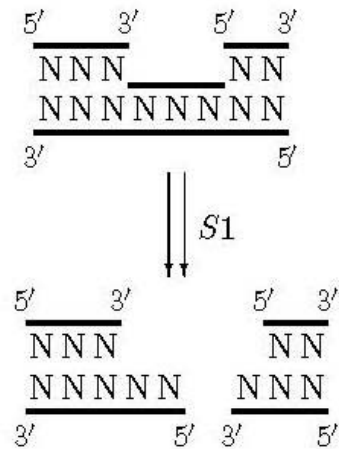


Figure 1.16: *S1* endonuclease in action (ii)

Restriction enzymes

- Restriction endonucleases: much more specific
- Cut only double stranded molecules, only at very specific set of sites
- Bind to the *recognition site* and then cut (sometimes even outside the binding site)
- The cut: blunt or staggered
- Examples:
 - EcoRI: 5'-GAATTC
 - XmaI: 5'-CCCGGG
 - SmaI: 5'-CCCGGG

Restriction enzymes

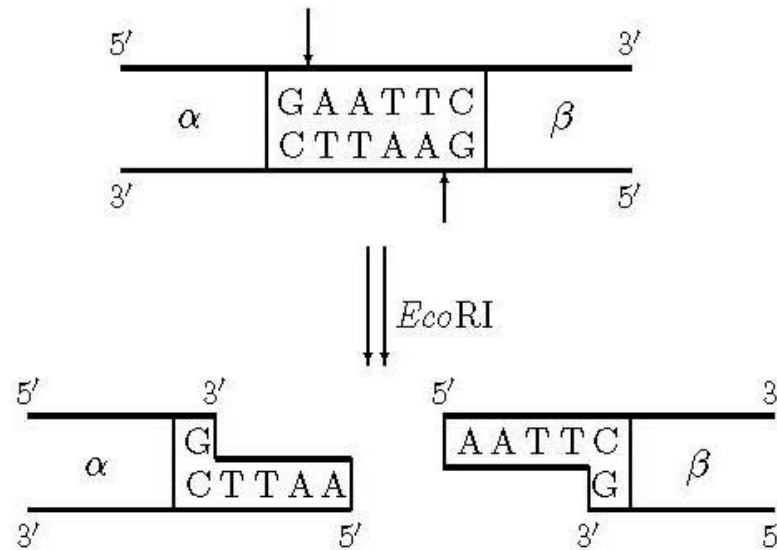


Figure 1.17: *Eco*RI in action

Restriction enzymes

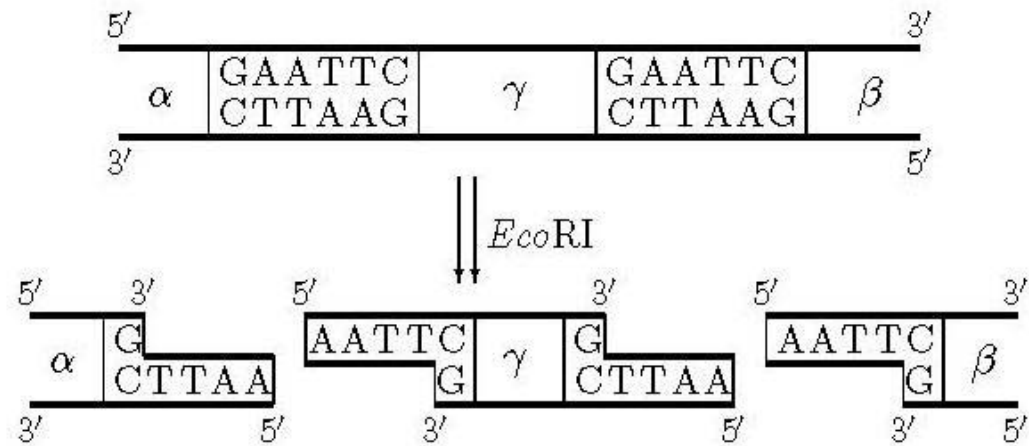


Figure 1.18: Multiple cut by *Eco*RI

Restriction enzymes

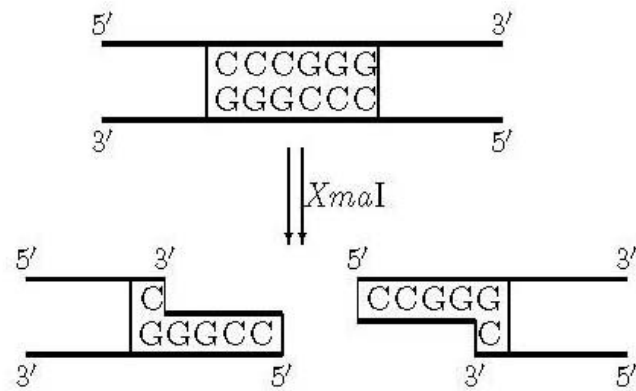


Figure 1.19: *Xma*I in action

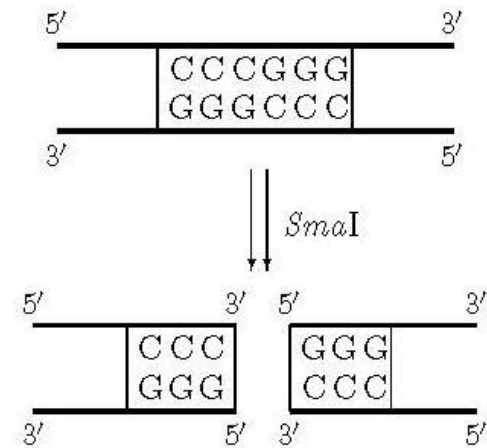


Figure 1.20: *Sma*I in action

Restriction enzymes

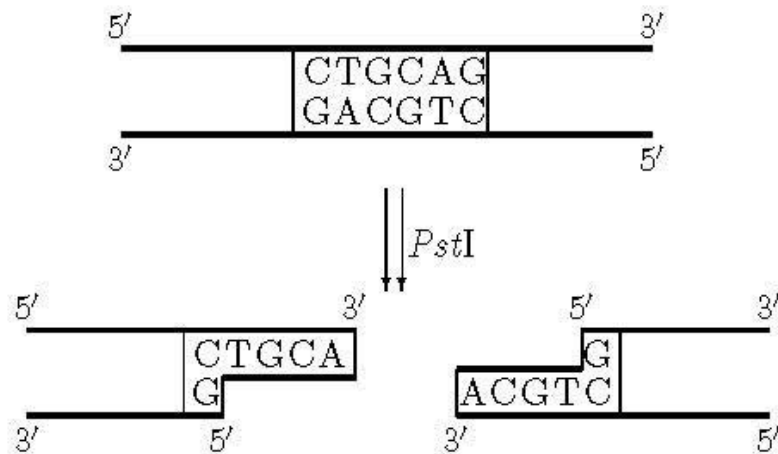


Figure 1.21: *Pst*I in action

Restriction enzymes

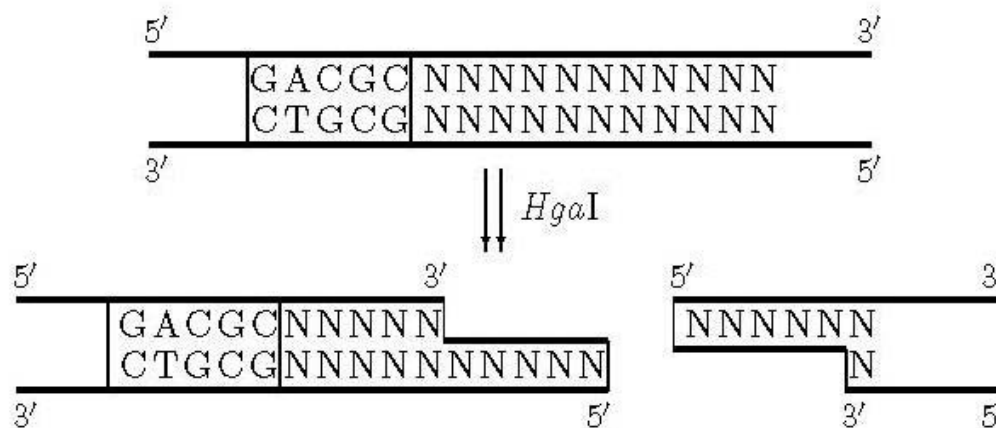


Figure 1.22: *HgaI* in action

Linking DNA

- Fuse together two pieces of DNA
- Add the necessary chemical links
- *Examples*
 - Hydrogen bonding
 - Ligation (enzyme: *ligase*)
 - Hybridization: link two complementary single strands
 - Blunt end ligation

Hydrogen bonding

- Complementary base pairing

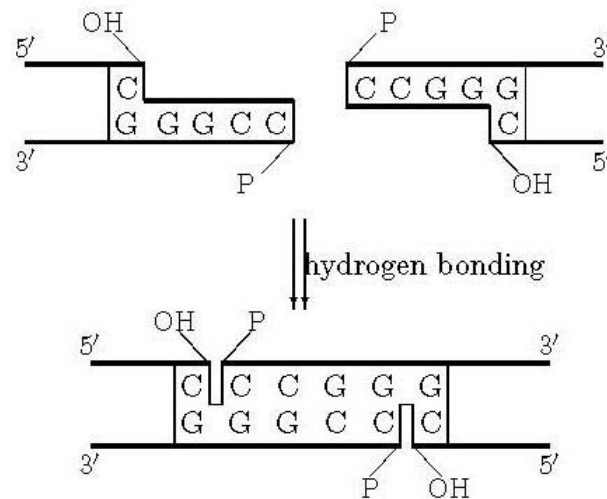


Figure 1.23: Complementary base pairing

Ligation

- Fix the phosphodiester bonds in single strands (*nick*)

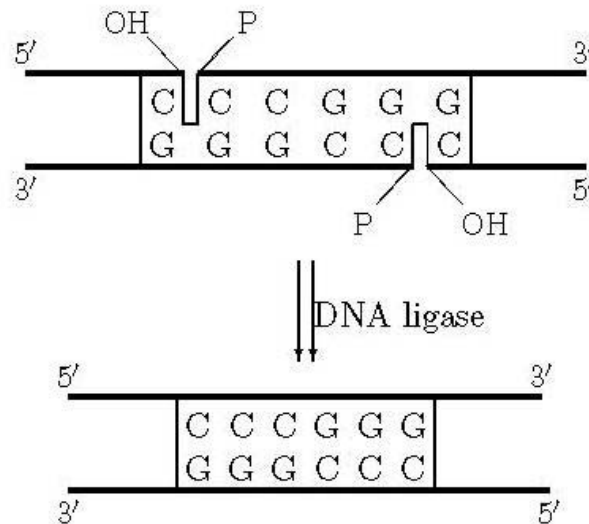


Figure 1.24: Ligation

Hybridization

- Double strands produced out of two single strands (*sticky ends*)

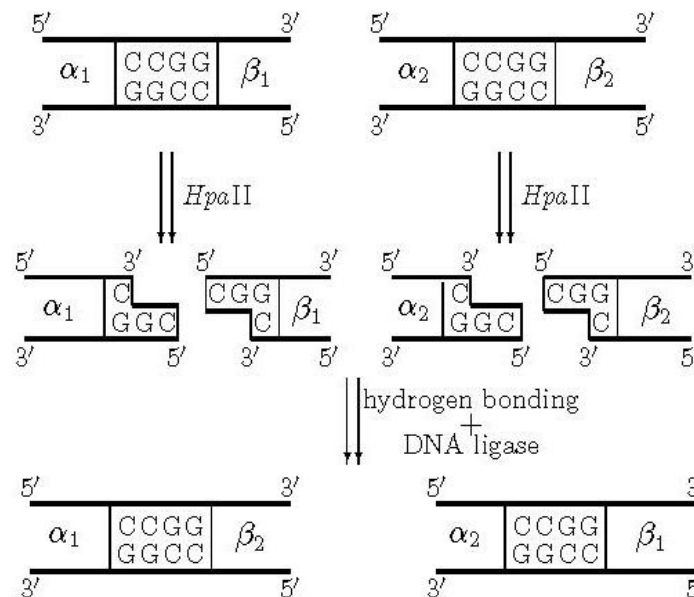


Figure 1.25: Hybridization

Blunt ligation

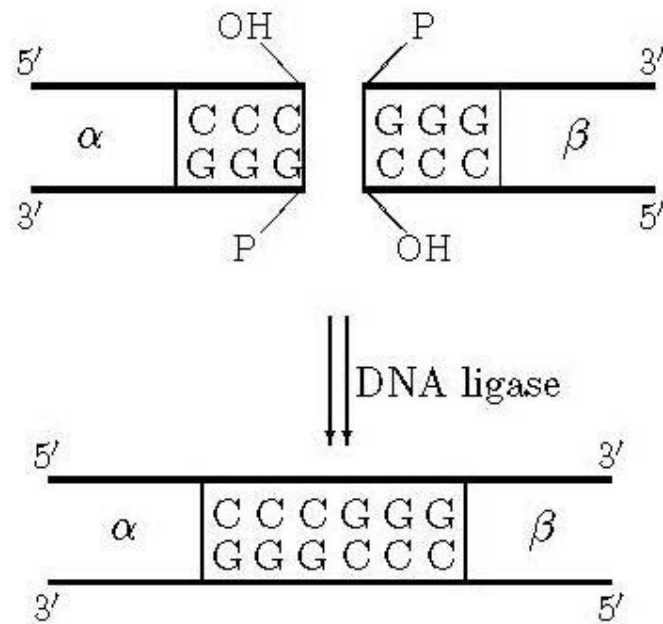


Figure 1.26: Blunt ligation

Blunt ligation

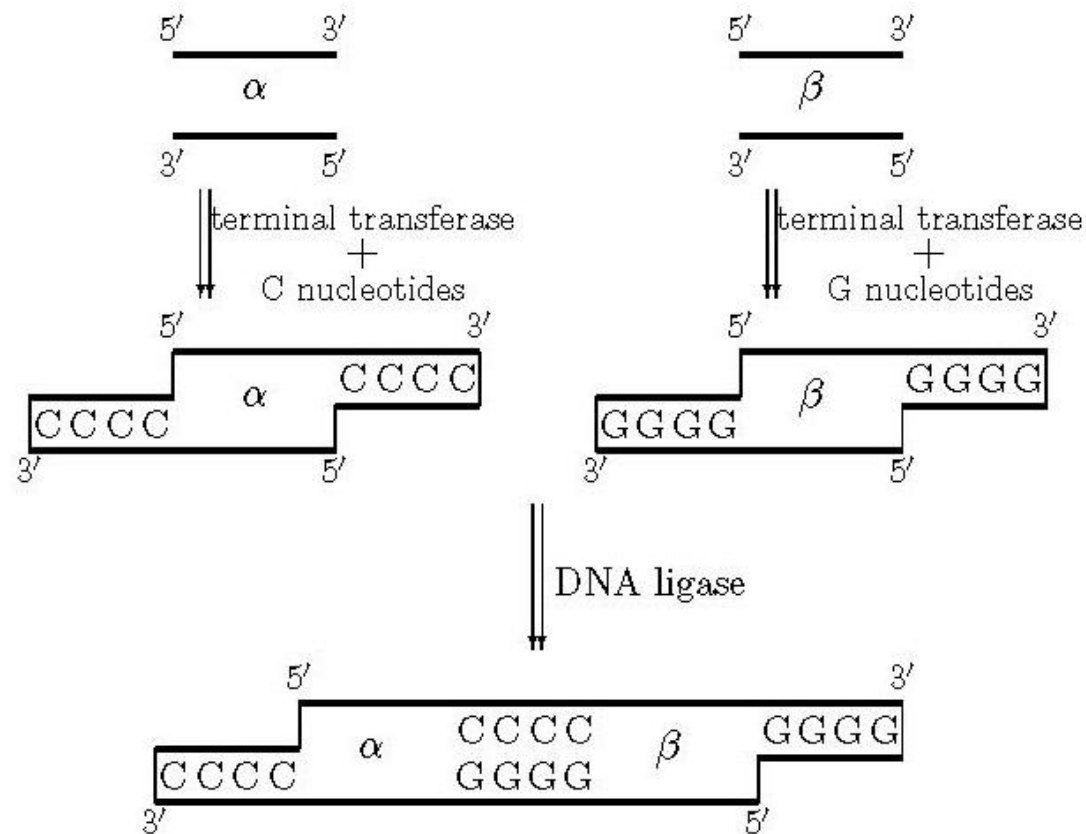


Figure 1.27: Joining blunt ended molecules using homopolymer tailing

Modifying nucleotides of DNA

- The enzymes: *modifying enzymes*
- Examples:
 - *Methylases*: used *in vivo* as partners of restriction enzymes – the same recognition site
 - It adds a methyl group to one of the nucleotides
 - *Alkaline phosphatase*: modifies the 5' end so that the molecule cannot ligate with itself
 - *Polynucleotide kinase*: reverses the effect of alkaline phosphate

Example

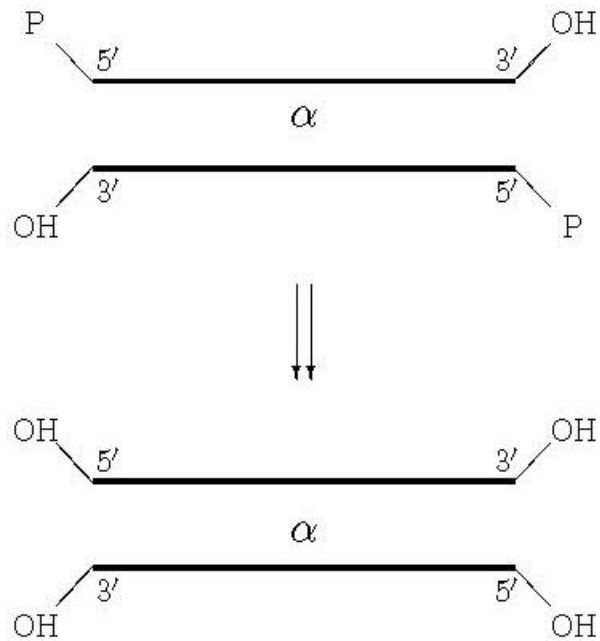


Figure 1.28: Alkaline phosphatase in action

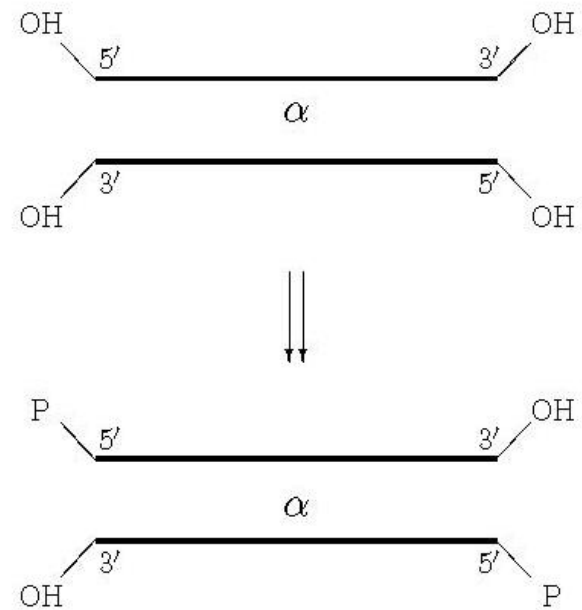


Figure 1.29: Polynucleotide kinase in action

Multiplying DNA

- Major problem in biochemical research:
obtain sufficient quantities of some substance
- Solution: cloning techniques, PCR

Cloning

- Clone: a collection of identical organisms, all replicas of a single ancestor
- Cloning: insert the DNA fragment in the DNA of some fast reproducing cell and the fragment will be replicated with the cell
 - Use viruses or hybridization

Cloning

- Separate the cell with the DNA of interest from the others using an antibiotic
- This technique provides high quantities of DNA fragments
- It also gives a mean to preserve the fragments for long time (keeping alive the “host”)

Polymerase chain reaction - PCR

- 1985, Kary Mullis: Nobel prize
- *In vitro* multiplication method
- Made possible by the technique of designing oligos
- Very efficient: produce in a short time billions of copies of even a single strand

The PCR technique

- We want to amplify a DNA molecule alpha with known borders beta and gamma

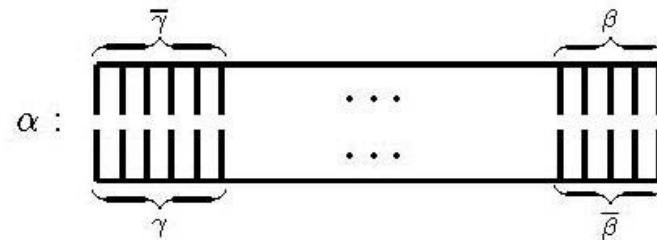


Figure 1.30: DNA with borders

The PCR technique

- We want to amplify a DNA molecule alpha with known borders beta and gamma
- **Process:** repeat a cycle of *denaturation*, *priming*, *extension*
- **Start:** prepare a solution containing alpha (the target), oligonucleotides beta' and gamma' (complements of beta and gamma), polymerase enzymes, and many nucleotides (A, C, G, T)

The PCR technique

- **Denaturation:** heat the solution to 85-95 C: alpha denatures into two single strands α_1 and α_2

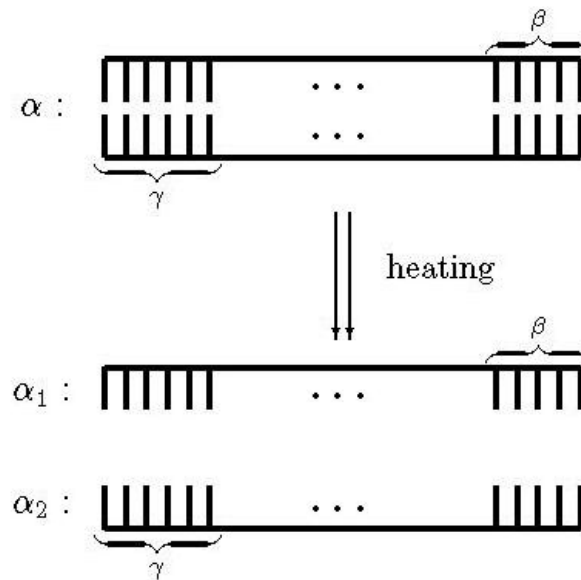


Figure 1.31: Denaturation

The PCR technique

- **Priming:** Cool down the solution to 55C: the primers beta' and gamma' anneal to their complementary borders

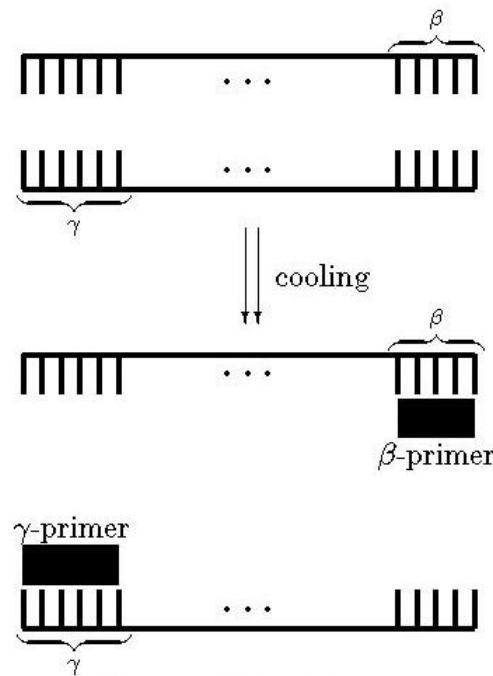


Figure 1.32: Priming

The PCR technique

- **Extension:** Heat the solution to 72C: polymerase extend the primers to produce two double stranded DNA molecules alpha

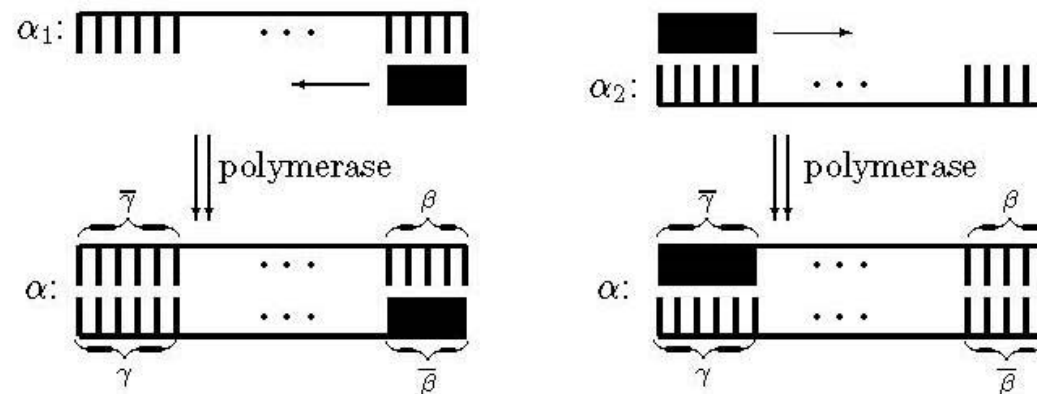


Figure 1.33: Extension

The PCR technique

- Repeat the cycle n times: 2^n copies (in principle): highly efficient bio-copy machine!
- A single cycle takes about 5 minutes: obtain billions of copies in several hours (days for cloning)
- The polymerase must be heat resistant – nature's solution: thermophilic bacteria
- Important observation:
 - one needs to know the borders of the DNA segment to be copied

Sequencing

- Learning the exact sequence of nucleotides of a DNA molecule
- The most popular method of sequencing: the *Sanger method*
 - 1940s: Sanger, insulin sequencing (Nobel prize)
 - 1977: Sanger, Gilbert, DNA sequencing (Nobel prize)
 - 1985: Mullis, PCR (Nobel prize)
- Main tool: nucleotide analogues – nucleotides chemically altered so that no other nucleotide can attach to their 3' end: ddA, ddC, ddG, ddT (dideoxynucleotides)

Sequencing – Sanger method

- Problem: sequence a single stranded molecule *alpha*
- Extend it to 3' by *gamma* and add the primer *gamma'* (labeled):
let *beta* be the new molecule

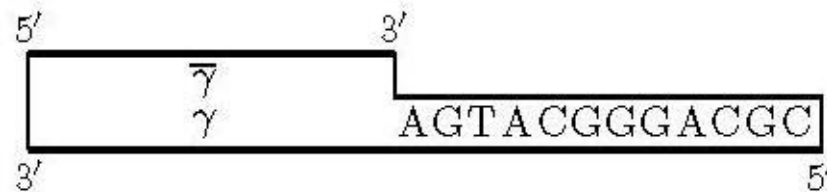


Figure 1.34: β' molecule

Sequencing – Sanger method

- Problem: sequence a single stranded molecule *alpha*
- Extend it to 3' by *gamma* and add the primer *gamma'* (labeled): let *beta* be the new molecule
- Prepare 4 tubes (A,C,G,T) containing:
 - *beta* molecules
 - Primers *gamma'*
 - Nucleotides

Sequencing

- If only nucleotides are used: produce full duplex



Figure 1.35: Full duplex

Sequencing

- If only nucleotides are used: produce full duplex
- Solution: tube X contains a *limited* amount of nucleotide analogues ddX, for all X in the set {A,C,T,G}

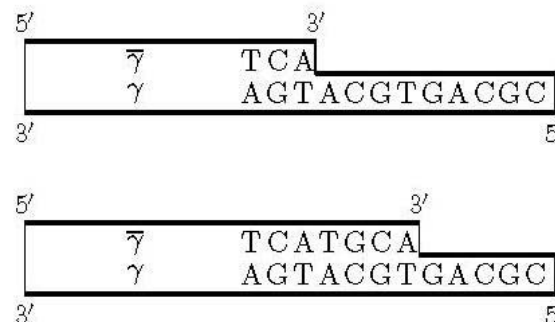


Figure 1.36: Incomplete molecules in Tube A

Example: alpha=3'-AGTACGTGACGC

- Tube A:
 - 5'-*gamma*'TCATGCACTGCG
 - 5'-*gamma*'TCA
 - 5'-*gamma*'TCATGCA
- Tube T:
 - 5'-*gamma*'TCATGCACTGCG
 - 5'-*gamma*'T
 - 5'-*gamma*'TCAT
 - 5'-*gamma*' TCATGCACT
- Tube C:
 - 5'-*gamma*'TCATGCACTGCG
 - 5'-*gamma*'TC
 - 5'-*gamma*'TCATGC
 - 5'-*gamma*'TCATGCAC
 - 5'-*gamma*'TCATGCACTGC
- Tube G:
 - 5'-*gamma*'TCATGCACTGCG
 - 5'-*gamma*'TCATG
 - 5'-*gamma*'TCATGCACTG

Sequencing – Sanger method

- Gel electrophoresis using 4 wells - one for each tube
- Read the bands (the primers were marked)
- Obtain the sequence

Result: 5'-TCATGCACTGCG

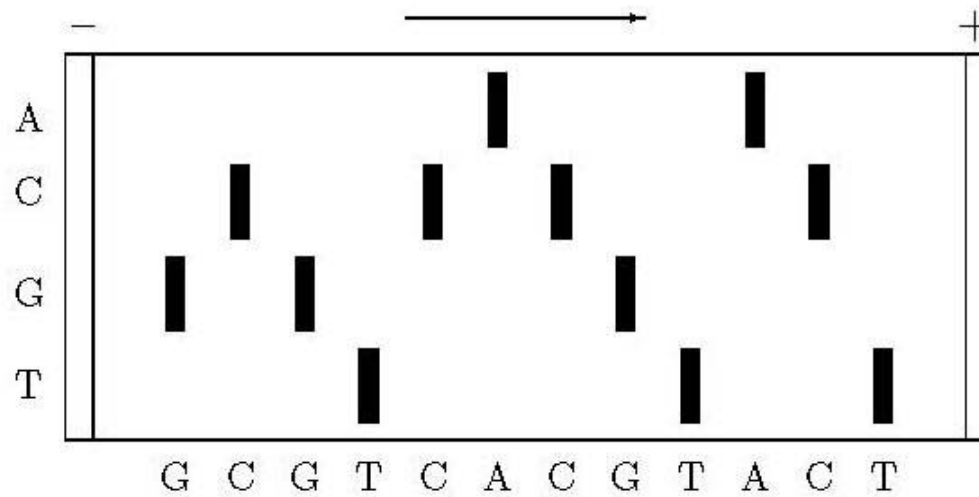


Figure 1.37: Sequencing ladder

Observations

- This was only a very simplified presentation
 - Use polymerase not having the associated exonuclease activity: do not cut the ddX
 - Limited amount of nucleotide analogues: too many make the polymerase before then end of the strand
 - Impossible to sequence long molecules with this method
- Sequencing a longer molecule: divide it in overlapping short segments, sequence them and then reassemble the original sequence: computational (math) problem !