



Differential splicing analysis

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Workshop outline

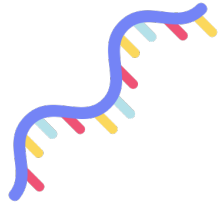


Introduction into identifying and quantifying splicing events



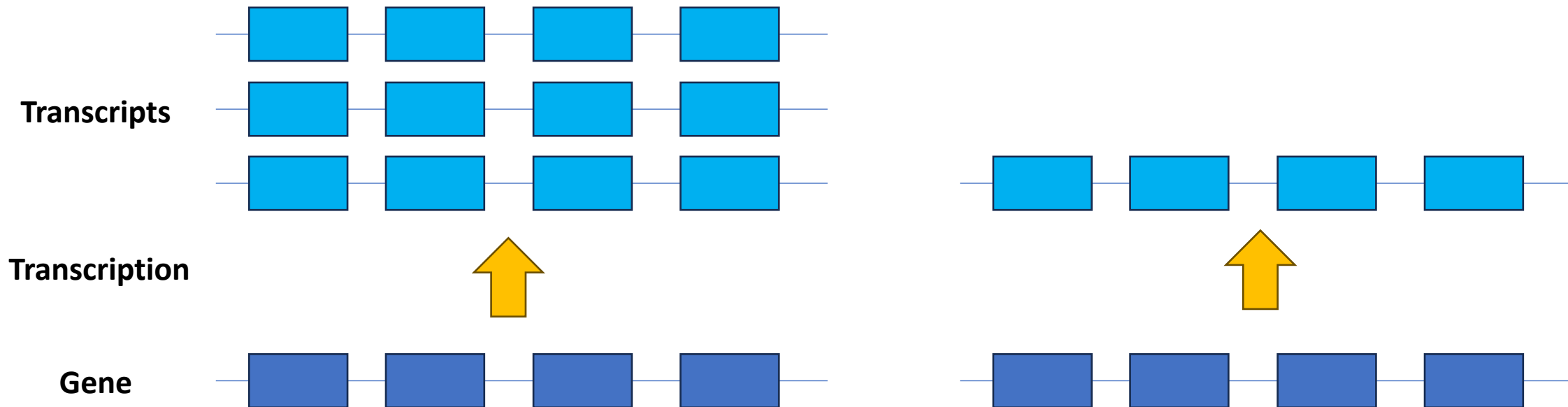
Processing and visualisation of splicing events

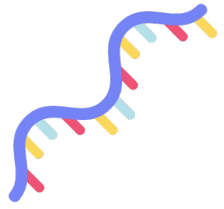




Differential gene expression

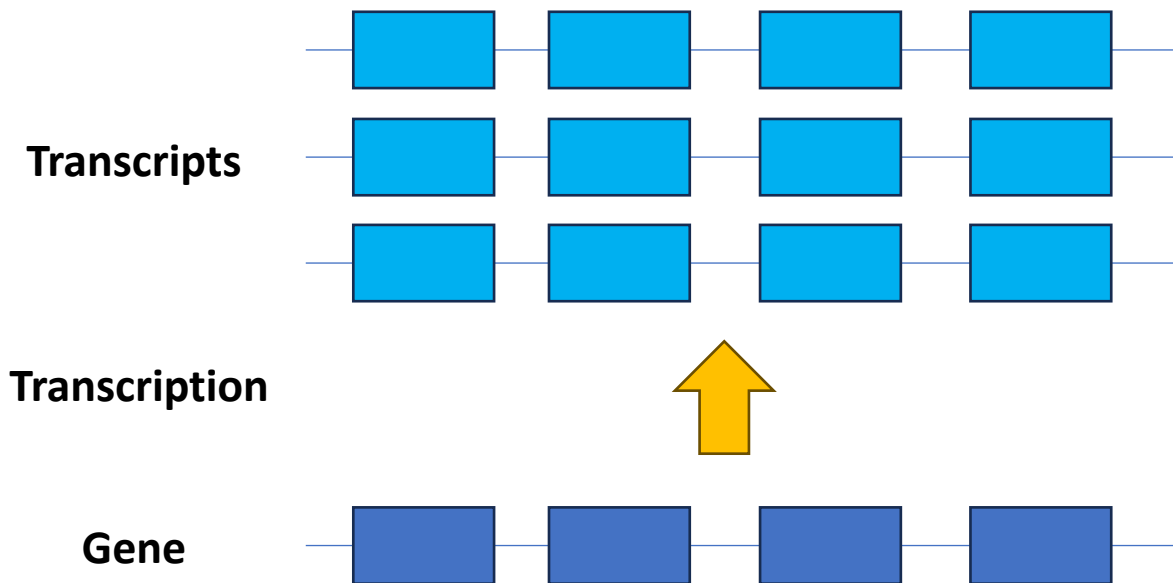
Differences in **amplitude**
(i.e., the amount a gene is transcribed)



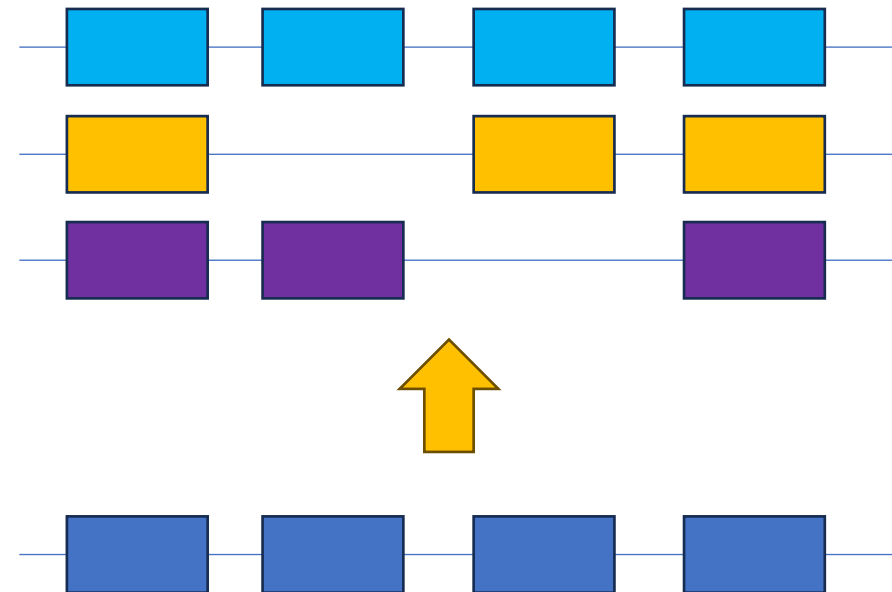


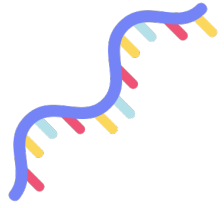
Differential gene expression

Changes in *amplitude*
(i.e., the amount a gene is transcribed)



Changes in *isoforms*
(i.e., what a gene is transcribing)





Common splicing events

Alternative Splicing Events

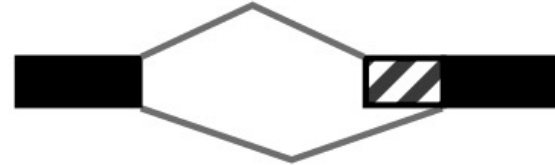
Skipped exon (SE)



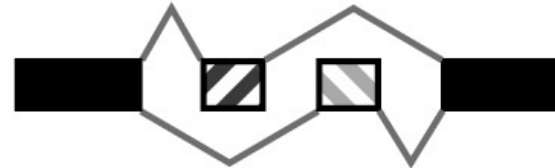
Alternative 5' splice site (A5SS)



Alternative 3' splice site (A3SS)



Mutually exclusive exons (MXE)



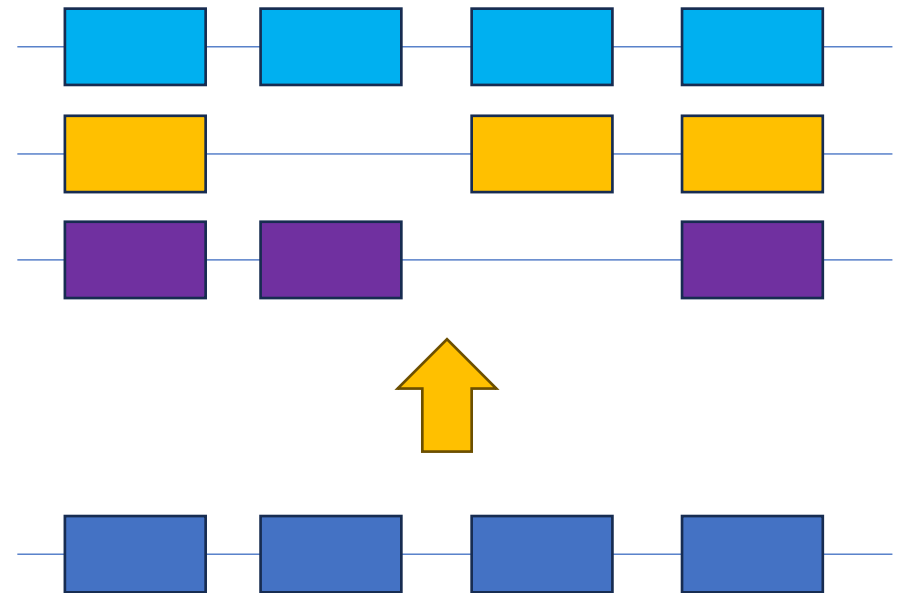
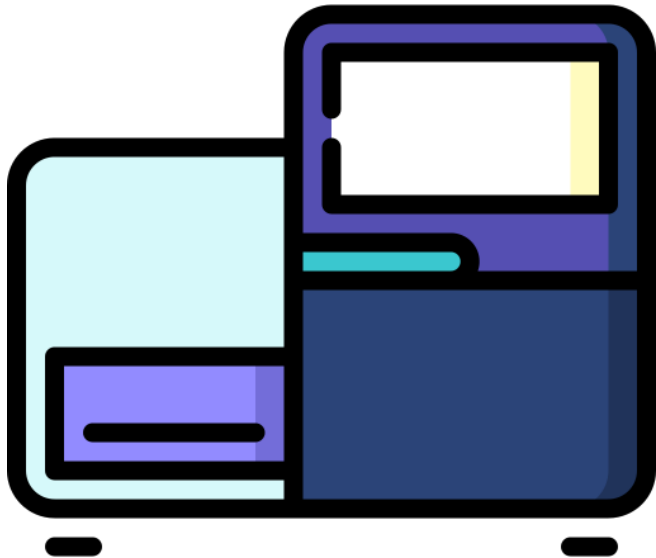
Retained intron (RI)





How can we measure alternative splicing?

The basic principle in RNA-Seq analysis of alternative splicing is to use RNA-Seq reads mapped to different isoforms to estimate the isoform proportion





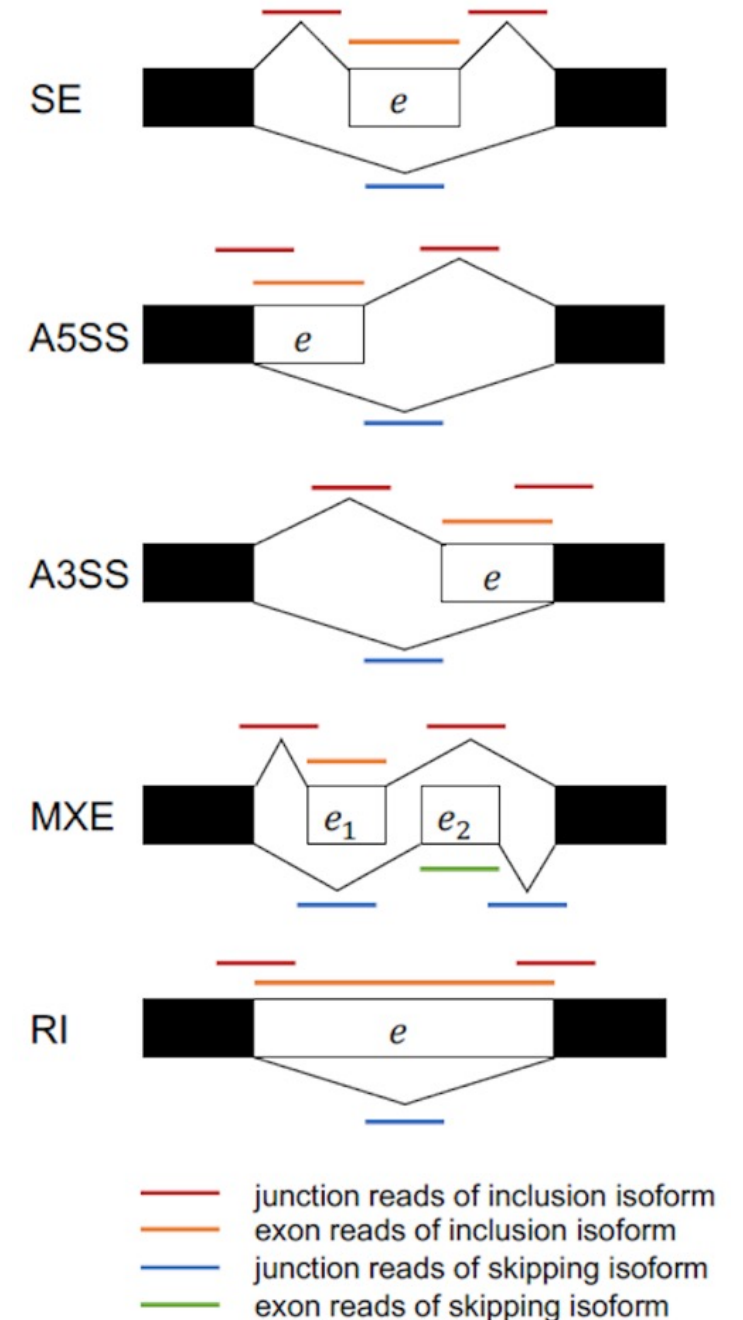
rMATs

Based on MATs (Multivariate analysis of transcript splicing)

Detect differential alternative splicing events from RNA-Seq data.

The statistical model of MATS calculates the P-value and false discovery rate that the difference in the isoform ratio of a gene between two conditions exceeds a given user-defined threshold.

From the RNA-Seq data, MATS can automatically detect and analyze alternative splicing events corresponding to all major types of alternative splicing patterns

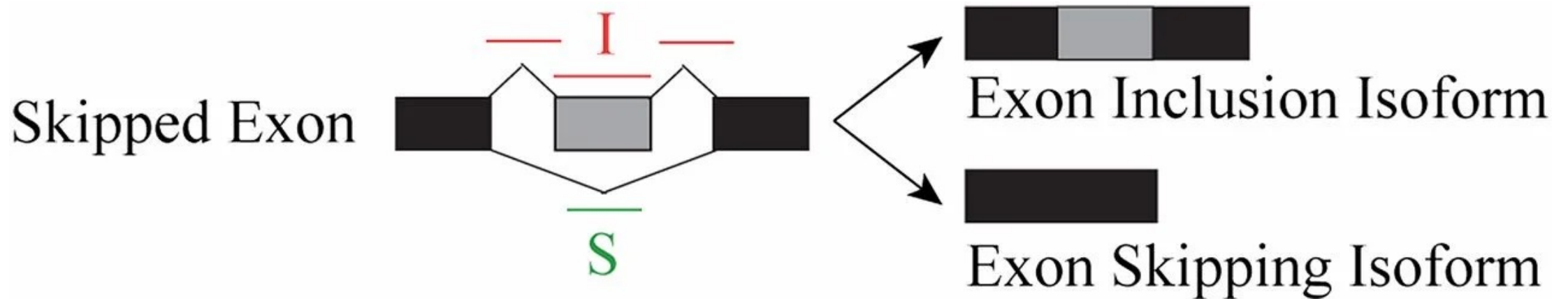




How can we measure alternative splicing?

Percent Spliced in (PSI): Ratio between reads including (I) or excluding exons (S)

Compare between mean PSI between two groups using likelihood ratio test

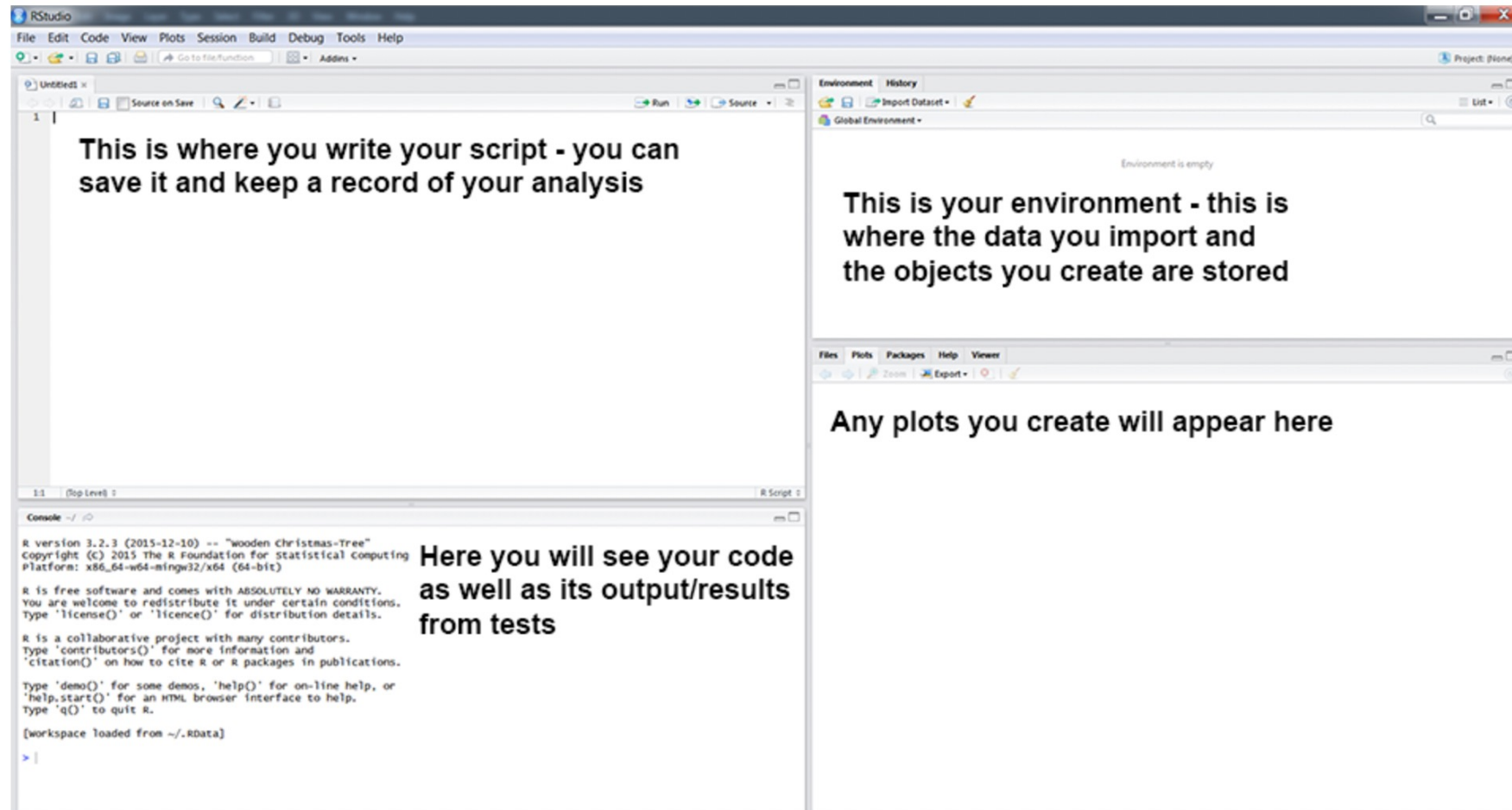


Introduction to R



Today, we will do our analysis in RStudio:

- A standard RStudio consists of four panels



Introduction to R



R is an **object-based language**

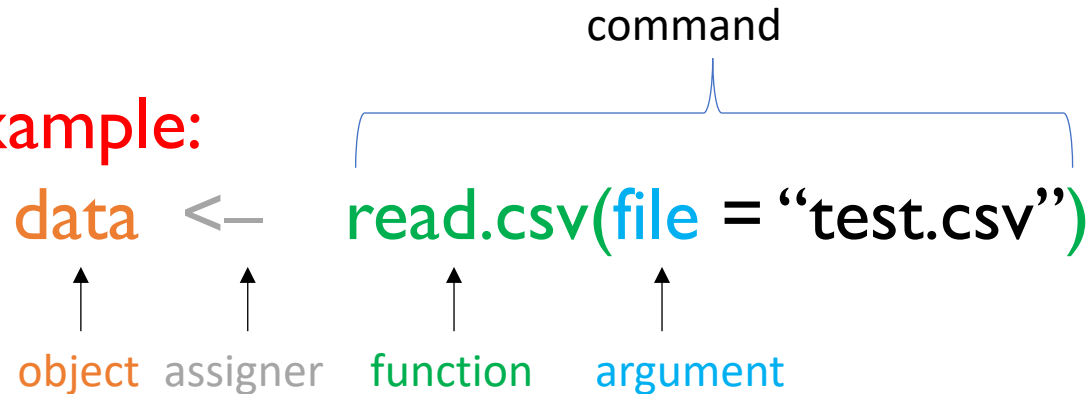
- This means the data you import or values you create during your R session are stored in **objects**

For example:

command

```
data <- read.csv(file = "test.csv")
```

object assigner function argument

A diagram illustrating the components of the R command 'data <- read.csv(file = "test.csv")'. The word 'data' is labeled as an 'object' with an upward arrow. The '<-' symbol is labeled as an 'assigner' with an upward arrow. The 'read.csv' part is labeled as a 'function' with an upward arrow. The '(file = "test.csv")' part is labeled as an 'argument' with an upward arrow. A bracket above the entire command is labeled 'command'.

In this command:

- the '**read.csv()**' takes the file 'test.csv' as input and assigns the information to the **object** '**data**' where the information is stored.
- objects can contain different data such as vectors, data frames, matrices, and lists.

Introduction to R



How data are stored:

-There are four main classes of data:

- **Integer:** only whole numbers (1, 2, 3, 4, 5, 6, 7, 8, 9, 100, 1000, 260000)
- **Numerical:** whole and floating values (0.1, 2.3, 3.5556, 5, 6, 7.8, 8.9, 0.0007)
- **Character:** ('a', 'b', 'dog', 'bee', 'I', '7.8', '4000')
- **Factor:**
 - Objects where you can add additional information (categories) to your data structure

For example: Categorise cities by country

Cities	Country
Berlin	Germany
Paris	France
Frankfurt	Germany
Mainz	Germany
London	UK

These categories are then known as **levels**

You can find out the class of an object by using the 'str()' command, which stands for structure

Introduction to R



Functions:

There are three main classes of functions:

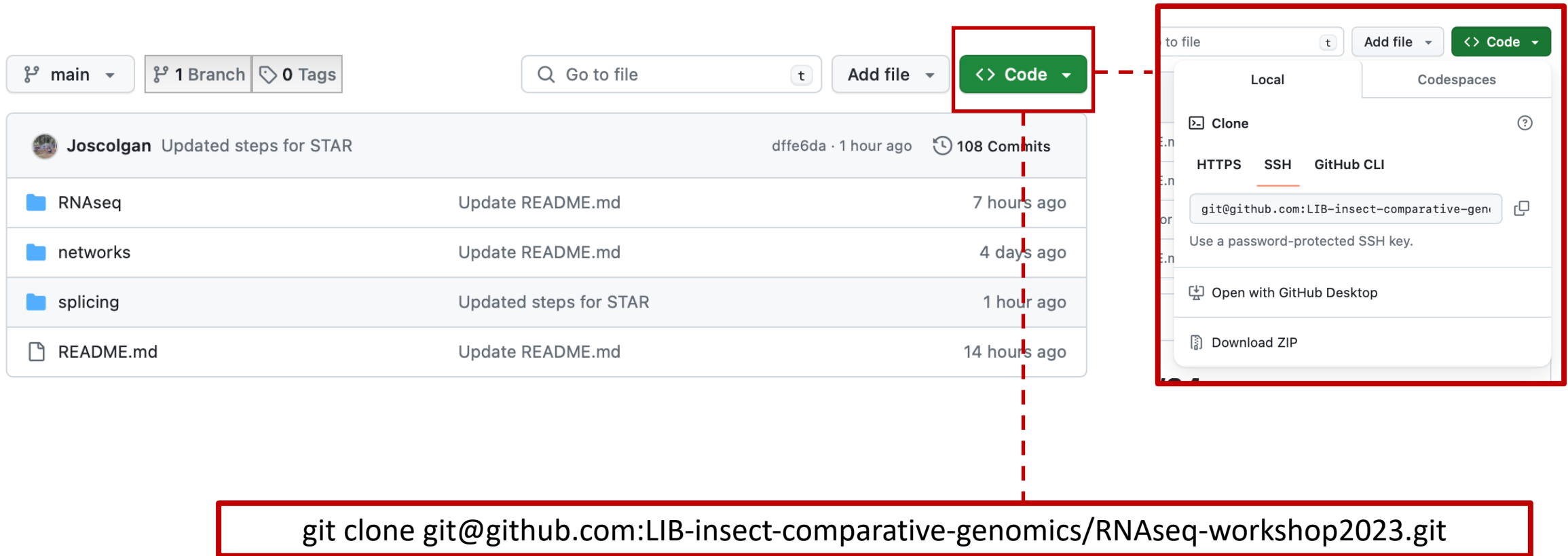
- **Base R functions:** these are functions that come already come with Rstudio (e.g., `nrow()`, `dim()`, `str()`)
- **Package functions:** these are functions that come with an R package and the package must be loaded before the function can be used (e.g., `ggplot()`, `dplyr()`, `euler()`).
 - NB: Sometimes functions from different packages may be called the same name (e.g., `plot()`) – to ensure you are using the right function, we can specify the function from a particular function by using the syntax: `package::function()` (e.g., `dplyr::select()`)
- **Custom functions:** functions that you can write yourself

What you have for today



We have a directory on Github that you can download/clone:

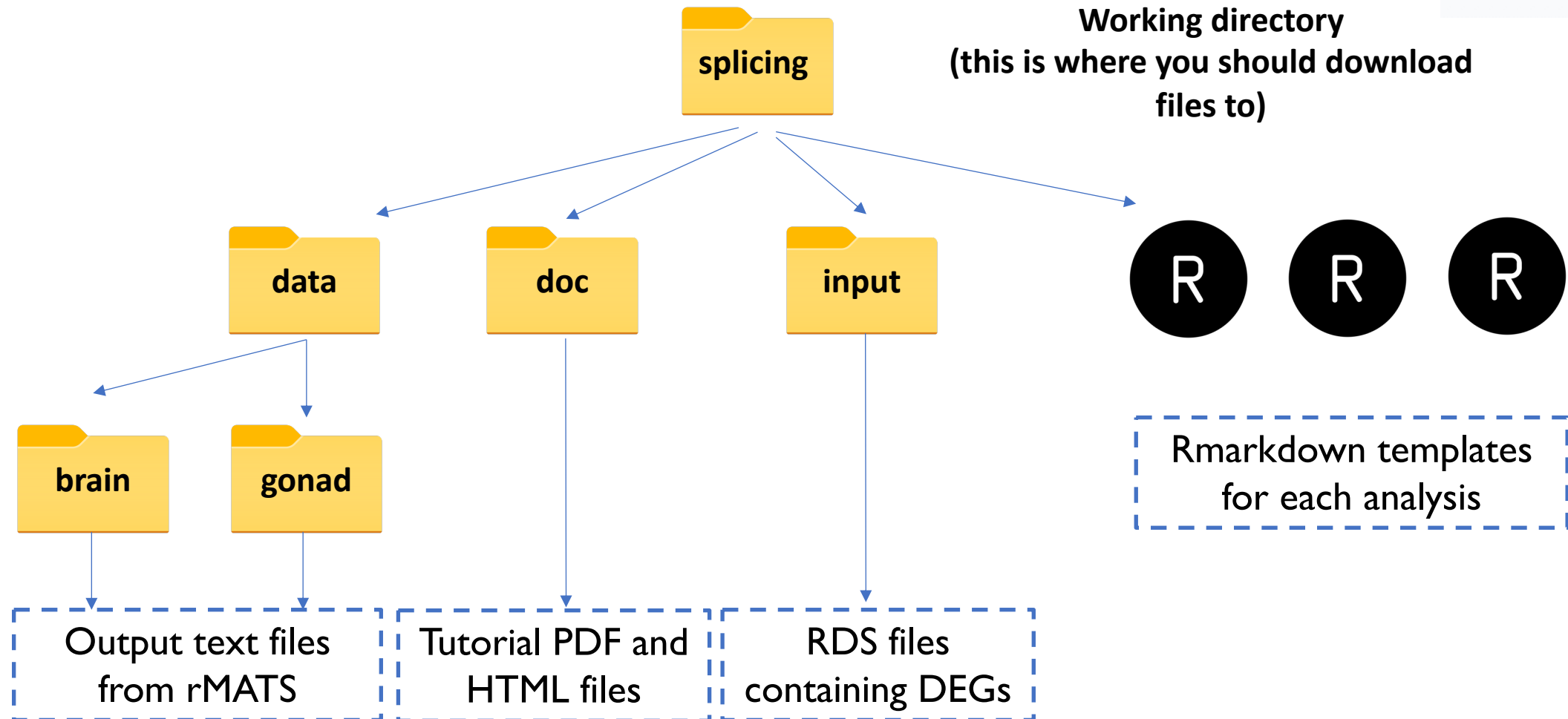
<https://github.com/LIB-insect-comparative-genomics/RNAseq-workshop2023>



The screenshot shows the GitHub repository page for `LIB-insect-comparative-genomics/RNAseq-workshop2023`. The repository is owned by `Joscolgan` and has 108 commits. The file list includes `RNAseq`, `networks`, `splicing`, and `README.md`. The `Code` button is highlighted with a red box, and its dropdown menu is open, showing options to clone the repository using HTTPS, SSH, or GitHub CLI, or to download the ZIP file. A red dashed line connects the `Code` button to the terminal command below.

```
git clone git@github.com:LIB-insect-comparative-genomics/RNAseq-workshop2023.git
```

What you have for today

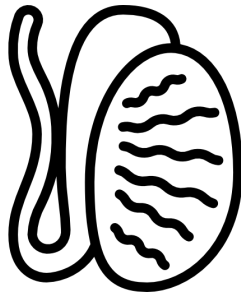




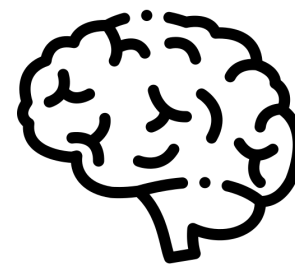
Dataset for today: Honeybees



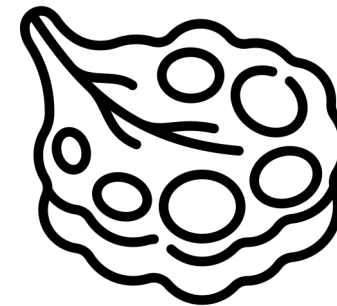
N = 5



N = 5



N = 4



N = 3



JOURNAL ARTICLE

Haploid and Sexual Selection Shape the Rate of Evolution of Genes across the Honey Bee (*Apis mellifera* L.) Genome

Garett P. Slater ✉, Amy L. Dapper, Brock A. Harpur ✉

Genome Biology and Evolution, Volume 14, Issue 6, June 2022, evac063,

<https://doi.org/10.1093/gbe/evac063>

Published: 12 May 2022 Article history ▼

- Reads downloaded from NCBI SRA database [sratool]
- Aligned against latest reference genome [STAR]
- Identification of splicing events [rMATs]

<https://academic.oup.com/gbe/article/14/6/evac063/6584681>

What is an R Markdown script?



It looks like a fancier version of a regular R script but has some great features:

- You can clearly separate sections with and without code (easier to read and add notes/annotations)
- Using the knitr function, you can convert your script into a html, PDF, or word document

```
1 ---
2 author: 'Put your name here'
3 title: '**Alternative-based splicing analysis of _Apis mellifera_ queens and drones - Part 3: Overlap**'
4 output:
5   html_document: default
6   pdf_document:
7     toc: true
8 fig_width: 5
9 fig_height: 10
10 fontsize: 20pt
11 ---
12
13 ## Introduction
14 The scripts today are written in RMarkdown which allows you to write text sections, which
15 are not run as code while you can specify text that will be run by code by writing in
16 specific sections like this one:
17
18 ```{r, message = FALSE}
19 print("You can run code here like you would in a normal R script")
20 ## If you want to write things that are not run as code, you have to start the
21 ## sentence with a hash/pound ('#') symbol
22
23 ```
24
25 We can then write sections again that we don't want to run as code.
```


What you have for today



We have three analyses today – three scripts that we want you to work through that will cover:

- **Processing** and **visualising** results of splicing events between queens and males for two tissues (brains and gonads)
- **Comparison** between alternatively spliced genes and those that differ in amplitude

Helpful resources for self-learning



- [Structuring folders for computational analyses](#)
- Improving in R and statistics (<https://ourcodingclub.github.io>)
- Introduction to Unix shell (<https://swcarpentry.github.io/shell-novice/>)
- Statistics & Machine learning (<https://www.youtube.com/@statquest>)
- Scripts for non-model organisms (mainly bees and fish): <https://github.com/Joscolgan>

Questions or feedback: tcolgan@uni-mainz.de