Steven Maharaj 695281 Assignment 2, Question 2

Due: Friday 20 September 2019

There are places in this assignment where R code will be required. Therefore set the random seed so assignment is reproducible.

```
set.seed(695281) #Please change random seed to your student id number.
library(dplyr)
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
       intersect, setdiff, setequal, union
##
library(mvtnorm)
library(coda)
library(ggplot2)
library(tidyr)
PART A
# Read the Data
Hiron <- read.csv("Hiron.csv")</pre>
HironHomo <- Hiron %>%filter(homc282y==1) %>% select(-idnum)
HironWild <- Hiron %>%filter(homc282y==0) %>% select(-idnum)
#fit linear regression using lm
model <- lm(logsf ~ time,data = HironHomo)</pre>
model$coefficients
## (Intercept)
## 4.23987253 0.07126085
intercept <-matrix(1,length(HironHomo$time),1)</pre>
X <- cbind(intercept, HironHomo$time)</pre>
XTX <- crossprod(X)</pre>
XTXinv <-solve(XTX)
# (XTX) ^-1
XTXinv
                 [,1]
                                [,2]
## [1,] 0.033734956 -0.0015415009
## [2,] -0.001541501 0.0001062175
```

```
#Estimated error variance
Sigma2<-sigma(model)^2
Sigma2
```

[1] 1.715042

PART B

For This Question we define proper priors for β , τ using the results from part a. That is using the following formula.

$$p(\theta|\mathbf{y}) = \frac{p(\mathbf{y_2}|\theta)p(\theta|\mathbf{y_1})}{p(\mathbf{y_2}|\mathbf{y_1})}$$

Using the posterior from the previous part we let the prior for this part be

$$p(\boldsymbol{\beta}|\tau) = \mathcal{N}(\hat{\boldsymbol{\beta}}_1, (\boldsymbol{X}_1 \boldsymbol{X}_1)^{-1}/\tau)$$
$$p(\boldsymbol{\beta}|\tau) = Ga(\frac{n_1 - p}{2}, \frac{(n_1 - p)s^2}{2})$$

where the subscript 1 indicates the results are from group 1 (analysed in 2a) alone. Using the results from lecture 13, we drop the prior for τ_{β} , and in all other places in the joint distribution, replace τ_{β} and τ_{e} with τ .

- $K = (X_1 X_1)^{-1}$
- $\beta_0 = \hat{\beta}_1$ $\alpha = \frac{n_1 p}{2}$
- $\gamma = \frac{(n_1-p)s^2}{2}$

Thus we get the following conditional posteriors

$$p\left(\tau|\mathbf{y},\boldsymbol{\beta},\boldsymbol{\beta}_{0},\mathbf{K}\right) = \operatorname{Ga}\left(\alpha + \frac{n+p}{2}, \gamma + \frac{(\mathbf{y} - \mathbf{X}\boldsymbol{\beta})'(\mathbf{y} - \mathbf{X}\boldsymbol{\beta}) + (\boldsymbol{\beta} - \boldsymbol{\beta}_{0})'\mathbf{K}^{-1}(\boldsymbol{\beta} - \boldsymbol{\beta}_{0})}{2}\right)$$
$$p\left(\boldsymbol{\beta}|\mathbf{y},\boldsymbol{\beta}_{0},\mathbf{K},\tau\right) = \mathcal{N}\left(\left(\mathbf{X}'\mathbf{X} + \mathbf{K}^{-1}\right)^{-1}\left(\mathbf{X}'\mathbf{y} + \mathbf{K}^{-1}\boldsymbol{\beta}_{0}\right), \left(\mathbf{X}'\mathbf{X} + \mathbf{K}^{-1}\right)^{-1}/\tau\right)$$

Below we fit a Bayesian regression using a Gibbs sampler to only the wildtype (homc282y=0) data.

```
# define our Gibbs sampler function
#Arguments are
#X: matrix of predictors dimension n times p with flat prior. Includes the intercept.
#Z: matrix of predictors dimension n times q with normal prior for u.
#y: response vector, length p.
#taue_0, tauu_0: initial value for the residual and random effect precision.
#a.u,b.u. Hyper-parameter for gamma prior for tau_u
#a.e,b.e. Hyper-parameter for gamma prior for tau_e
#iter: number of iterations
#burnin: number of initial iterations to remove
normalmm.Gibbs<-function(iter,Z,X,y,burnin,taue_0,tauu_0,a.u,b.u,a.e,b.e){
      <-length(y) #no. observations</pre>
      <-dim(X)[2] #no of fixed effect predictors.
  р
      <-dim(Z)[2] #no of random effect levels
```

```
tauu<-tauu_0
  taue<-taue_0
  #starting value for u.
  u0 <-rnorm(q,0,sd=1/sqrt(tauu))</pre>
  #Building combined predictor matrix.
  W<-cbind(X,Z)
                            #for the joint conditional posterior for b,u
  WTW <-crossprod(W)
  library(mvtnorm)
  #storing results.
  par <-matrix(0,iter,p+q+2) #p beta coefficient, q u coefficients and 2 precision coefficient.
  #Create modified identity matrix for joint posterior.
  IO <-diag(p+q)
  diag(I0)[1:p]<-0
  for(i in 1:iter){
    #Conditional posteriors.
    tauu \-rgamma(1,a.u+0.5*q,b.u+0.5*sum(u0^2)) #sample tau_u
    #Updating component of normal posterior for beta, u
    Prec <-WTW + tauu*I0/taue</pre>
    P.mean <- solve(Prec) ** crossprod(W,y)
    P.var <-solve(Prec)/taue
    betau <-rmvnorm(1,mean=P.mean,sigma=P.var) #sample beta, u
    betau <-as.numeric(betau)</pre>
    err <- y-W%*%betau
    taue <-rgamma(1,a.e+0.5*n,b.e+0.5*sum(err^2)) #sample tau_e
    #storing iterations for beta, u, and standard deviation of e, u.
    par[i,]<-c(betau,1/sqrt(tauu),1/sqrt(taue))</pre>
           <-betau[p+1:q] #extracting u so we can update tau_u.</pre>
    u0
  }
par <-par[-c(1:burnin),] #removing initial iterations</pre>
colnames(par)<-c(paste('beta',1:p,sep=''),paste('u',1:q,sep=''),'sigma_b','sigma_e')</pre>
return(par)
# Define inputs for Gibbs sampler
Kinv <- XTX
X2 <- cbind(matrix(1,length(HironWild$time),1),HironWild$time)</pre>
y <- HironWild$logsf
b0 <- model$coefficients
n1 < -dim(X)[1]
p < -dim(X2)[2]
a \leftarrow (n1-p)*0.5
s2 \leftarrow sum((HironHomo\$logsf - X\%*\%b0)^2)/(n1-p)
g <- (n1-p)*s2*0.5
Gibbsq2 <- function(iter, X, y, burnin, tau_0, a, g, b0, Kinv){</pre>
  n <-length(y) #no. observations
  p <-dim(X)[2] #no of fixed effect predictors.
  XXkii <- solve(crossprod(X) + Kinv)</pre>
```

```
P.mean <- XXkii%*%(t(X)%*%y + Kinv%*%b0)
  b <- rnorm(p,0,sd=1/sqrt(tau_0))</pre>
  tau<- tau_0
  #storing results.
  par <-matrix(0,iter,p+3)</pre>
  for (i in 1:iter) {
    err <- y-X%*%b
    T_y <- sum(err^2)</pre>
    tau -\text{rgamma}(1,a+(n+p)*0.5, g + 0.5*T_y + 0.5*t(b-b0)\%*\%Kinv\%*\%(b-b0))
    b <- rmvnorm(1,mean=P.mean,sigma=XXkii/tau)
    b <-as.numeric(b)</pre>
    # posterior checking
  Xb <- X%*%as.vector(b)</pre>
  y_rep <- Xb+rnorm(length(y))*sqrt(1/tau)</pre>
  T_{rep} \leftarrow sum((y_{rep}-Xb)^2)
  #storing iterations for beta, tau,.
  par[i,] <- c(b[1],b[2],1/tau,T_y,T_rep)
  }
  par <-par[-c(1:burnin),] #removing initial iterations</pre>
  colnames(par)<-c("beta0","beta1","sigma2","T_y","T_rep")</pre>
  return(par)
chain1 <- Gibbsq2(iter=10000, X = X2, y=y, burnin=1000, tau 0=1, a=a, g=g, b0=b0, Kinv=Kinv)
chain2 <- Gibbsq2(iter=10000,X = X2,y=y,burnin=1000,tau_0=1,a=a,g=g,b0=b0,Kinv=Kinv)
# Remove every second iteration to reduce auto - corrlation
chain1t <- chain1[seq(1,dim(chain1)[1],by=2),]</pre>
chain2t <- chain2[seq(1,dim(chain2)[1],by=2),]</pre>
```

Reporting posterior means, standard deviations and 95 % central credible intervals for $\beta_0, \beta_1, \sigma^2$ by combining results for the two chains.

```
chain12t <- rbind(chain1t,chain2t)
chain_stats <- data.frame(matrix(nrow = 3,ncol =4 ))
para_names <- c('beta0','beta1','sigma2','lower_CI95','upper_CI95')
for (i in 1:3) {
   chain <- chain12t[,i]
   quat <- quantile(sort(chain), c(0.05, 0.975))
   chain_stats[i,] <- c(mean(chain),sd(chain),quat)
}
names(chain_stats)<- c("posterior mean","std",'lower_CI95','upper_CI95')</pre>
```

```
row.names(chain_stats) <- c('beta0','beta1','sigma2')
# chain results
chain_stats</pre>
```

```
## beta0 posterior mean std lower_CI95 upper_CI95
## beta0 4.12918718 0.111765425 3.94303272 4.35034827
## beta1 0.02963795 0.005814982 0.02024566 0.04106363
## sigma2 1.33588228 0.103746049 1.17458517 1.55165944
```