

# 432 Class 03 Slides

[thomaseLove.github.io/432](https://thomaseLove.github.io/432)

2022-01-18

# Today's Agenda

- Create a data set for week 2 analyses from `smart_ohio`
- Making cleaning / tidying decisions, then saving our work
- Simple imputation
- Splitting the sample with `rsample` tools
- Fitting a model (and then several more models) with `lm`
  - Incorporating an interaction between factors
  - Incorporating polynomial terms
- Regression Diagnostics via Residual Plots
- Evaluating results in holdout sample with `yardstick`

# Creating and Managing the Data for Week 2

# Setup

```
knitr::opts_chunk$set(comment = NA)
options(width = 60)

library(here); library(knitr)
library(janitor); library(patchwork)
library(naniar); library(simputation)
library(skimr)           ## for a specific summary
library(equatiomatic)    ## print equations
library(broom)
library(rsample)          ## new today: data splitting
library(yardstick)        ## new today: evaluating fits
library(tidyverse)

theme_set(theme_bw())
options(dplyr.summarise.inform = FALSE) ## avoid message
```

## Similar approach as last time...

```
smart_ohio <- read_csv(here("data/smart_ohio.csv"))

week2 <- smart_ohio %>%
  filter(hx_diabetes == 0,
         mmsa == "Cleveland-Elyria",
         complete.cases(bmi)) %>%
  select(bmi, inc_imp, fruit_day, drinks_wk,
         female, exerany, genhealth, race_eth,
         hx_diabetes, mmsa, SEQNO) %>%
  type.convert(as.is = FALSE) %>%
  mutate(ID = as.character(SEQNO - 2017000000)) %>%
  relocate(ID)
```

```
# A tibble: 894 x 12
```

	ID	bmi	inc_imp	fruit_day	drinks_wk	female	exerany
	<chr>	<dbl>	<int>	<dbl>	<dbl>	<int>	<int>
1	2	23.0	86865	4	0	1	0
2	3	26.9	NA	3	0	1	1
3	4	26.5	NA	2	4.67	1	1
4	5	24.2	58311	0.57	0.93	0	1
5	7	23.0	2318	2	2	0	1
6	8	28.4	79667	1	0	0	1
7	9	30.1	47880	0.23	0	0	1
8	10	19.8	100136	0.77	0.47	1	1
9	11	27.2	73145	0.71	0	0	1
10	12	24.6	76917	1.07	0	1	1

```
# ... with 884 more rows, and 5 more variables:
```

```
#   genhealth <fct>, race_eth <fct>, hx_diabetes <int>,  
#   mmsa <fct>, SEQNO <int>
```

# Codebook for useful week2 variables

- 894 subjects in Cleveland-Elyria with `bmi` and no history of diabetes

Variable	Description
<code>bmi</code>	(outcome) Body-Mass index in $\text{kg}/\text{m}^2$ .
<code>inc_imp</code>	income (imputed from grouped values) in \$
<code>fruit_day</code>	average fruit servings consumed per day
<code>drinks_wk</code>	average alcoholic drinks consumed per week
<code>female</code>	sex: 1 = female, 0 = male
<code>exerany</code>	any exercise in the past month: 1 = yes, 0 = no
<code>genhealth</code>	self-reported overall health (5 levels)
<code>race_eth</code>	race and Hispanic/Latinx ethnicity (5 levels)

- plus `ID`, `SEQNO`, `hx_diabetes` (all 0), `MMSA` (all Cleveland-Elyria)
- See Chapter 2 of the Course Notes for details on the variables

Available approaches include:

- `summary`
- mosaic package's `inspect()`
- skimr package's `skim_without_charts()`
- Hmisc package's `describe`

all of which can work nicely in an HTML presentation, but none of them fit well on one of these slides.



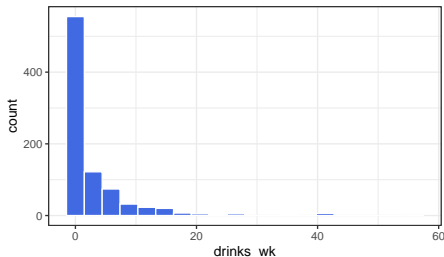
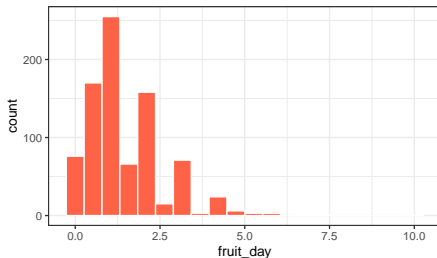
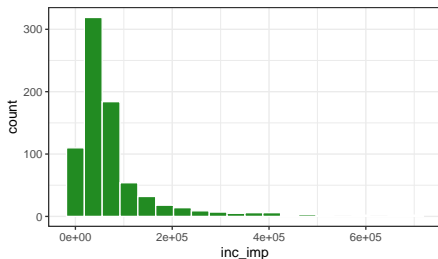
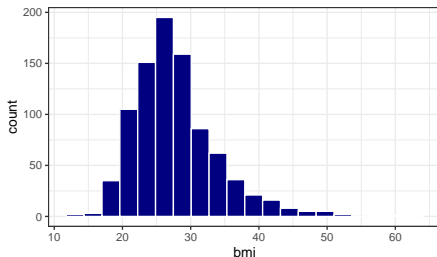
# Summarizing the Quantities (Raw week2)

```
week2 %>% select(bmi, inc_imp, fruit_day, drinks_wk) %>%  
  skim_without_charts() %>%  
  yank(., "numeric") %>%  
  select(var = skim_variable, n_missing, min = p0,  
         median = p50, max = p100, mean, sd) %>%  
  kable(digits = 1)
```

var	n_missing	min	median	max	mean	sd
bmi	0	13.3	26.8	63	27.9	6.3
inc_imp	120	216.0	48224.5	700676	75673.5	90695.8
fruit_day	41	0.0	1.1	10	1.4	1.1
drinks_wk	39	0.0	0.5	56	3.0	6.1

- Any signs of trouble? (What are we looking for?)

# Quick Histogram of each quantitative variable



# Code for previous slide

```
p1 <- ggplot(week2, aes(x = bmi)) +  
  geom_histogram(fill = "navy", col = "white", bins = 20)  
p2 <- ggplot(week2, aes(x = inc_imp)) +  
  geom_histogram(fill = "forestgreen", col = "white",  
                bins = 20)  
p3 <- ggplot(week2, aes(x = fruit_day)) +  
  geom_histogram(fill = "tomato", col = "white", bins = 20)  
p4 <- ggplot(week2, aes(x = drinks_wk)) +  
  geom_histogram(fill = "royalblue", col = "white",  
                bins = 20)  
(p1 + p2) / (p3 + p4)
```

I also used `warning = FALSE` in the plot's code chunk label to avoid warnings about missing values, like this one for `inc_imp`:

Warning: Removed 120 rows containing non-finite values

# Binary variables in raw week2

```
week2 %>% tabyl(female, exerany) %>% adorn_title()
```

	exerany		
female	0	1	NA_
0	95	268	20
1	128	361	22

- female is based on biological sex (1 = female, 0 = male)
- exerany comes from a response to “During the past month, other than your regular job, did you participate in any physical activities or exercises such as running, calisthenics, golf, gardening, or walking for exercise?” (1 = yes, 0 = no, don’t know and refused = missing)
- Any signs of trouble here?

# Binary variables in raw week2

```
week2 %>% tabyl(female, exerany) %>% adorn_title()
```

	exerany		
female	0	1	NA_
0	95	268	20
1	128	361	22

- female is based on biological sex (1 = female, 0 = male)
- exerany comes from a response to “During the past month, other than your regular job, did you participate in any physical activities or exercises such as running, calisthenics, golf, gardening, or walking for exercise?” (1 = yes, 0 = no, don’t know and refused = missing)
- Any signs of trouble here?
- I think the 1/0 values and names are OK choices.

# Multicategorical genhealth in raw week2

```
week2 %>% tabyl(genhealth)
```

genhealth	n	percent	valid_percent
1_Excellent	148	0.165548098	0.16573348
2_VeryGood	324	0.362416107	0.36282195
3_Good	274	0.306487696	0.30683091
4_Fair	112	0.125279642	0.12541993
5_Poor	35	0.039149888	0.03919373
<NA>	1	0.001118568	NA

- The variable is based on “Would you say that in general your health is ...” using the five specified categories (Excellent -> Poor), numbered for convenience after data collection.
- Don't know / not sure / refused were each treated as missing.
- How might we manage this variable?

# Changing the levels for genhealth

```
week2 <- week2 %>%  
  mutate(health =  
    fct_recode(genhealth,  
      E = "1_Excellent",  
      VG = "2_VeryGood",  
      G = "3_Good",  
      F = "4_Fair",  
      P = "5_Poor"))
```

Might want to run a sanity check here, just to be sure...

# Checking health vs. genhealth in week2

```
week2 %>% tabyl(genhealth, health) %>% adorn_title()
```

	health					
genhealth	E	VG	G	F	P	NA_
1_Excellent	148	0	0	0	0	0
2_VeryGood	0	324	0	0	0	0
3_Good	0	0	274	0	0	0
4_Fair	0	0	0	112	0	0
5_Poor	0	0	0	0	35	0
<NA>	0	0	0	0	0	1

- OK. We've preserved the order and we have much shorter labels. Sometimes, that's helpful.



# Multicategorical race\_eth in raw week2

```
week2 %>% count(race_eth)
```

```
# A tibble: 6 x 2
```

	race_eth	n
	<fct>	<int>
1	Black non-Hispanic	167
2	Hispanic	27
3	Multiracial non-Hispanic	19
4	Other race non-Hispanic	22
5	White non-Hispanic	646
6	<NA>	13

“Don’t know”, “Not sure”, and “Refused” were treated as missing.

- What is this variable actually about?

# Multicategorical race\_eth in raw week2

```
week2 %>% count(race_eth)
```

```
# A tibble: 6 x 2
```

	race_eth	n
	<fct>	<int>
1	Black non-Hispanic	167
2	Hispanic	27
3	Multiracial non-Hispanic	19
4	Other race non-Hispanic	22
5	White non-Hispanic	646
6	<NA>	13

“Don’t know”, “Not sure”, and “Refused” were treated as missing.

- What is this variable actually about?
- What is the most common thing people do here?

# What is the question you are asking?

Collapsing `race_eth` levels *might* be rational for *some* questions.

- We have lots of data from two categories, but only two.
- Systemic racism affects people of color in different ways across these categories, but also *within* them.
- Is combining race and Hispanic/Latinx ethnicity helpful?

It's hard to see the justice in collecting this information and not using it in as granular a form as possible, though this leaves some small sample sizes. There is no magic number for “too small a sample size.”

- Most people identified themselves in one of the categories.
- These data are not ordered, and (I'd argue) ordering them isn't helpful.
- Regression models are easier to interpret, though, if the “baseline” category is a common one.

# Resorting the factor for race\_eth

Let's sort all five levels, from most observations to least...

```
week2 <- week2 %>%  
  mutate(race_eth = fct_infreq(race_eth))
```

```
week2 %>% tabyl(race_eth)
```

	race_eth	n	percent	valid_percent
	White non-Hispanic	646	0.72259508	0.73325766
	Black non-Hispanic	167	0.18680089	0.18955732
	Hispanic	27	0.03020134	0.03064699
	Other race non-Hispanic	22	0.02460850	0.02497162
	Multiracial non-Hispanic	19	0.02125280	0.02156640
	<NA>	13	0.01454139	NA

- Not a perfect solution, certainly, but we'll try it out.

# “Cleaned” Data and Missing Values

```
week2 <- week2 %>%  
  select(ID, bmi, inc_imp, fruit_day, drinks_wk,  
         female, exerany, health, race_eth, everything())  
  
miss_var_summary(week2)
```

# A tibble: 13 x 3

	variable <chr>	n_miss <int>	pct_miss <dbl>
1	inc_imp	120	13.4
2	exerany	42	4.70
3	fruit_day	41	4.59
4	drinks_wk	39	4.36
5	race_eth	13	1.45
6	health	1	0.112
7	genhealth	1	0.112
8	ID	0	0

# Single Imputation Approach?

```
set.seed(43203)
week2im <- week2 %>%
  select(ID, bmi, inc_imp, fruit_day, drinks_wk,
         female, exerany, health, race_eth) %>%
  data.frame() %>%
  impute_cart(health ~ bmi + female) %>%
  impute_pmm(exerany ~ female + health + bmi) %>%
  impute_rlm(inc_imp + drinks_wk + fruit_day ~
            bmi + female + health + exerany) %>%
  impute_cart(race_eth ~ health + inc_imp + bmi) %>%
  tibble()

prop_miss_case(week2im)

[1] 0
```

# Saving the tidied data

Let's save both the unimputed and the imputed tidy data as R data sets.

```
saveRDS(week2, here("data", "week2.Rds"))
```

```
saveRDS(week2im, here("data", "week2im.Rds"))
```

To reload these files, we'd use `readRDS`.

- The main advantage here is that we've saved the whole R object, including all characteristics that we've added since the original download.

# Splitting the Sample

Use `initial_split` from `rsample` to partition the data into:

- Model development (training) sample where we'll build models
- Model evaluation (testing) sample which we'll hold out for a while

```
set.seed(432)      ## to make the work replicable in the future
week2im_split <- initial_split(week2im, prop = 3/4)
```

```
train_w2im <- training(week2im_split)
test_w2im <- testing(week2im_split)
```

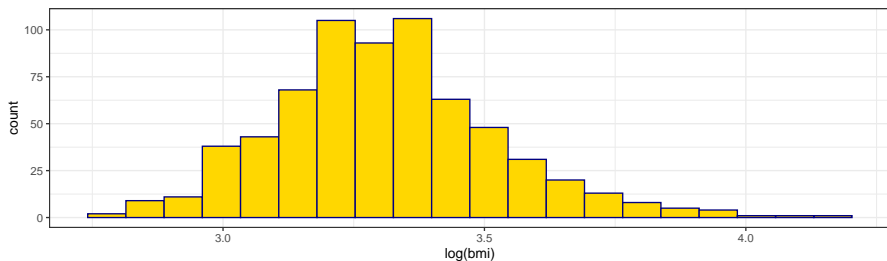
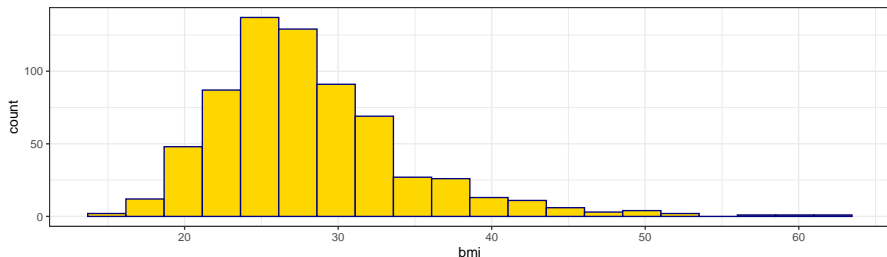
```
dim(train_w2im); dim(test_w2im)
```

```
[1] 670    9
```

```
[1] 224    9
```



# Should we transform our outcome?



**Outcome: bmi, with key predictors exerany and health both categorical (two-way ANOVA!)**

## bmi means by exerany and health

```
summaries_1 <- train_w2im %>%  
  group_by(exerany, health) %>%  
  summarise(n = n(), mean = mean(bmi), stdev = sd(bmi))  
summaries_1 %>% kable(digits = 2)
```

exerany	health	n	mean	stdev
0	E	18	27.49	3.56
0	VG	54	26.87	5.27
0	G	58	30.33	7.45
0	F	31	35.12	9.95
0	P	8	36.21	12.11
1	E	92	25.80	4.49
1	VG	191	26.80	4.89
1	G	152	29.12	6.26
1	F	49	27.21	5.55
1	P	17	28.50	8.61

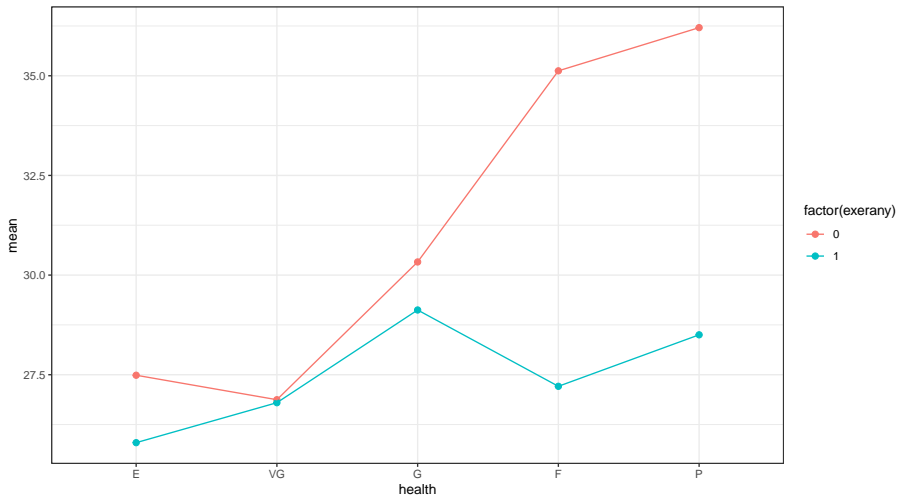
# Code for Interaction Plot

```
ggplot(summaries_1, aes(x = health, y = mean,  
                        col = factor(exerany))) +  
  geom_point(size = 2) +  
  geom_line(aes(group = factor(exerany))) +  
  labs(title = "Observed Means of BMI",  
       subtitle = "by Exercise and Overall Health")
```

- Note the use of factor here since the exerany variable is in fact numeric, although it only takes the values 1 and 0.
  - Sometimes it's helpful to treat 1/0 as a factor, and sometimes not.
- Where is the evidence of serious non-parallelism (if any) in the plot on the next slide that results from this code?

# Resulting Interaction Plot

Observed Means of BMI  
by Exercise and Overall Health



# Models we'll build today

- `m_1` a linear model without interaction using `exerany` and `health` to predict `bmi`
- `m_1int` add the interaction term for `exerany` and `health` to `m_1`

We'll assess these models carefully (today) in the training sample and (next time) in the test sample.

- We'll also explore adding a covariate `fruit_day` to the models in several different ways.

## Fitting ANOVA model $m_1$ without interaction

# Building a Model (m\_1) without interaction

```
m_1 <- lm(bmi ~ exerany + health,  
          data = train_w2im)
```

- How well does this model fit the training data?

```
glance(m_1) %>%  
  select(r.squared, adj.r.squared, sigma, nobs,  
         df, df.residual, AIC, BIC) %>%  
  kable(digits = c(3, 3, 2, 0, 0, 0, 1, 1))
```

r.squared	adj.r.squared	sigma	nobs	df	df.residual	AIC	BIC
0.089	0.082	6.12	670	5	664	4335.9	4367.5



# ANOVA for the `m_1` model

```
anova(m_1)
```

## Analysis of Variance Table

Response: bmi

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
exerany	1	895.7	895.71	23.948	1.243e-06	***
health	4	1528.5	382.12	10.217	4.952e-08	***
Residuals	664	24834.7	37.40			

---

Signif. codes:

0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

# Tidied ANOVA for the `m_1` model

```
tidy(anova(m_1)) %>%  
  kable(dig = c(0, 0, 2, 2, 2, 3))
```

term	df	sumsq	meansq	statistic	p.value
exerany	1	895.71	895.71	23.95	0
health	4	1528.47	382.12	10.22	0
Residuals	664	24834.72	37.40	NA	NA

# A summary of $m_1$ coefficients

```
summary(m_1)$coeff
```

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	27.9094987	0.7428015	37.5732944	1.704557e-166
exerany	-2.1966833	0.5501802	-3.9926613	7.262660e-05
healthVG	0.6176707	0.7026030	0.8791177	3.796555e-01
healthG	3.1372434	0.7224634	4.3424255	1.629287e-05
healthF	3.7122198	0.9070315	4.0927131	4.788419e-05
healthP	4.5514459	1.3577495	3.3521985	8.472164e-04

# Tidied summary of m\_1 coefficients

```
tidy(m_1, conf.int = TRUE, conf.level = 0.90) %>%  
  kable(digits = c(0,2,2,2,3,2,2))
```

term	estimate	std.error	statistic	p.value	conf.low	conf.high
(Intercept)	27.91	0.74	37.57	0.000	26.69	29.13
exerany	-2.20	0.55	-3.99	0.000	-3.10	-1.29
healthVG	0.62	0.70	0.88	0.380	-0.54	1.77
healthG	3.14	0.72	4.34	0.000	1.95	4.33
healthF	3.71	0.91	4.09	0.000	2.22	5.21
healthP	4.55	1.36	3.35	0.001	2.32	6.79

# Equation for Model without Interaction

From `m1` our equation is ...

```
extract_eq(m_1, use_coefs = TRUE, wrap = TRUE)
```

$$\widehat{\text{bmi}} = 27.91 - 2.2(\text{exerany}) + 0.62(\text{health}_{\text{VG}}) + 3.14(\text{health}_{\text{G}}) + 3.71(\text{health}_{\text{F}}) + 4.55(\text{health}_{\text{P}}) \quad (1)$$

- You need to use `results = "asis"` in the code chunk label to get this to work.
- This function `extract_eq` comes from the `equatiomatic` package.

# Interpreting the `m_1` model

$$\widehat{\text{bmi}} = 27.91 - 2.2(\text{exerany}) + 0.62(\text{health}_{\text{VG}}) + 3.14(\text{health}_{\text{G}}) + 3.71(\text{health}_{\text{F}}) + 4.55(\text{health}_{\text{P}}) \quad (2)$$

Name	exerany	health	predicted bmi
Harry	0	Excellent	26.66
Sally	1	Excellent	$26.66 - 1.37 = 25.29$
Billy	0	Fair	$26.66 + 3.82 = 30.48$
Meg	1	Fair	$26.66 - 1.37 + 3.82 = 29.11$

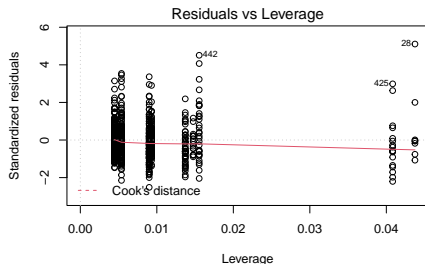
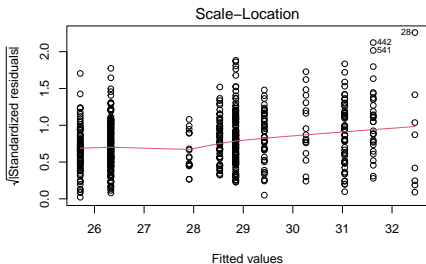
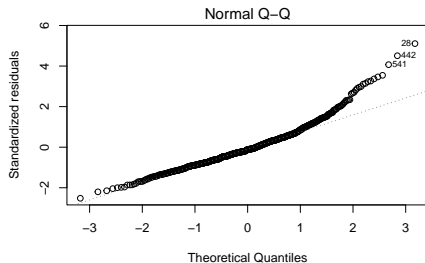
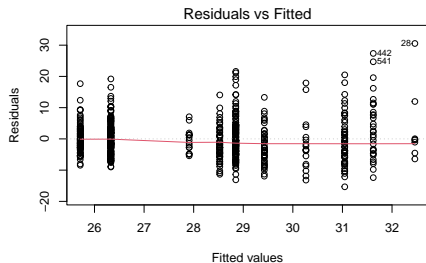
- Effect of `exerany`?
- Effect of `health` = Fair instead of Excellent?

# Plot the Residuals from model `m_1`?

```
par(mfrow = c(2,2))  
plot(m_1)  
par(mfrow = c(1,1))
```

That's the simplest code to get the four key plots to show up in the most familiar pattern, as shown on the next slide...

# m\_1 Residual Plots (conclusions?)





## Fitting ANOVA model `m_1int` including interaction

# Adding the interaction term to m\_1

```
m_1int <- lm(bmi ~ exerany * health,  
             data = train_w2im)
```

- How does this model compare in terms of fit to the training data?

```
bind_rows(glance(m_1), glance(m_1int)) %>%  
  mutate(mod = c("m_1", "m_1int")) %>%  
  select(mod, r.sq = r.squared, adj.r.sq = adj.r.squared,  
         sigma, nobs, df, df.res = df.residual, AIC, BIC) %>%  
  kable(digits = c(0, 3, 3, 2, 0, 0, 0, 1, 1))
```

mod	r.sq	adj.r.sq	sigma	nobs	df	df.res	AIC	BIC
m_1	0.089	0.082	6.12	670	5	664	4335.9	4367.5
m_1int	0.126	0.114	6.01	670	9	660	4315.8	4365.4

# ANOVA for the `m_1int` model

```
tidy(anova(m_1int)) %>%  
  kable(dig = c(0, 0, 2, 2, 2, 3))
```

term	df	sumsq	meansq	statistic	p.value
exerany	1	895.71	895.71	24.82	0
health	4	1528.47	382.12	10.59	0
exerany:health	4	1020.50	255.13	7.07	0
Residuals	660	23814.22	36.08	NA	NA

# ANOVA test comparing m\_1 to m\_lint

```
anova(m_1, m_lint)
```

Analysis of Variance Table

Model 1: bmi ~ exerany + health

Model 2: bmi ~ exerany \* health

	Res.Df	RSS	Df	Sum of Sq	F	Pr(>F)
1	664	24835				
2	660	23814	4	1020.5	7.0707	1.411e-05 ***

---

Signif. codes:

0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

# A summary of `m_lint` coefficients

```
summary(m_lint)$coeff
```

	Estimate	Std. Error	t value
(Intercept)	27.4872222	1.415826	19.4142627
exerany	-1.6917874	1.548148	-1.0927817
healthVG	-0.6140741	1.634855	-0.3756137
healthG	2.8419157	1.620700	1.7535108
healthF	7.6366487	1.780029	4.2901815
healthP	8.7202778	2.552417	3.4164785
exerany:healthVG	1.6167545	1.803846	0.8962818
exerany:healthG	0.4865311	1.804508	0.2696198
exerany:healthF	-6.2226958	2.072938	-3.0018726
exerany:healthP	-6.0145361	3.004914	-2.0015667
	Pr(> t )		
(Intercept)	9.227071e-67		
exerany	2.748884e-01		
healthVG	7.073248e-01		

# Tidied summary of m\_1int coefficients

```
tidy(m_1int, conf.int = TRUE, conf.level = 0.90) %>%  
  rename(se = std.error, t = statistic, p = p.value) %>%  
  kable(digits = c(0,2,2,2,3,2,2))
```

term	estimate	se	t	p	conf.low	conf.high
(Intercept)	27.49	1.42	19.41	0.000	25.16	29.82
exerany	-1.69	1.55	-1.09	0.275	-4.24	0.86
healthVG	-0.61	1.63	-0.38	0.707	-3.31	2.08
healthG	2.84	1.62	1.75	0.080	0.17	5.51
healthF	7.64	1.78	4.29	0.000	4.70	10.57
healthP	8.72	2.55	3.42	0.001	4.52	12.92
exerany:healthVG	1.62	1.80	0.90	0.370	-1.35	4.59
exerany:healthG	0.49	1.80	0.27	0.788	-2.49	3.46
exerany:healthF	-6.22	2.07	-3.00	0.003	-9.64	-2.81
exerany:healthP	-6.01	3.00	-2.00	0.046	-10.96	-1.06

# Equation for Interaction Model

From `m1_int` our equation is ...

```
extract_eq(m1_int, use_coefs = TRUE,  
           wrap = TRUE, terms_per_line = 2)
```

$$\begin{aligned}\widehat{\text{bmi}} = & 27.49 - 1.69(\text{exerany}) - \\ & 0.61(\text{health}_{\text{VG}}) + 2.84(\text{health}_{\text{G}}) + \\ & 7.64(\text{health}_{\text{F}}) + 8.72(\text{health}_{\text{P}}) + \\ & 1.62(\text{exerany} \times \text{health}_{\text{VG}}) + 0.49(\text{exerany} \times \text{health}_{\text{G}}) - \\ & 6.22(\text{exerany} \times \text{health}_{\text{F}}) - 6.01(\text{exerany} \times \text{health}_{\text{P}})\end{aligned}\tag{3}$$

Don't forget to use `results = "asis"` in the code chunk label.

# Interpreting the `m_1int` model

$$\begin{aligned}\widehat{\text{bmi}} = & 27.49 - 1.69(\text{exerany}) - \\ & 0.61(\text{health}_{\text{VG}}) + 2.84(\text{health}_{\text{G}}) + \\ & 7.64(\text{health}_{\text{F}}) + 8.72(\text{health}_{\text{P}}) + \\ & 1.62(\text{exerany} \times \text{health}_{\text{VG}}) + 0.49(\text{exerany} \times \text{health}_{\text{G}}) - \\ & 6.22(\text{exerany} \times \text{health}_{\text{F}}) - 6.01(\text{exerany} \times \text{health}_{\text{P}})\end{aligned}\tag{4}$$

Name	exerany	health	predicted bmi
Harry	0	Excellent	25.82
Sally	1	Excellent	$25.82 - 0.41 = 25.41$
Billy	0	Fair	$25.82 + 6.88 = 32.70$
Meg	1	Fair	$25.82 - 0.41 + 6.88 - 4.83 = 27.46$

- How do we interpret effect sizes here?

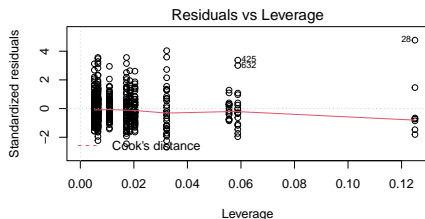
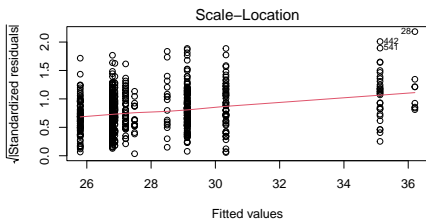
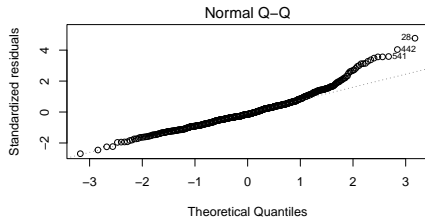
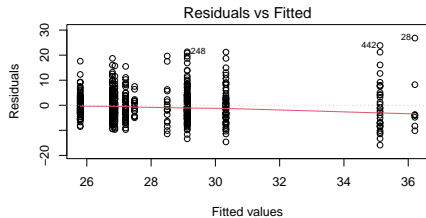


# Interpreting the `m_1int` model

Name	exerany	health	predicted bmi
Harry	0	Excellent	25.82
Sally	1	Excellent	$25.82 - 0.41 = 25.41$
Billy	0	Fair	$25.82 + 6.88 = 32.70$
Meg	1	Fair	$25.82 - 0.41 + 6.88 - 4.83 = 27.46$

- How do we interpret effect sizes here? **It depends.**
- Effect of `exerany`?
  - If `health` = Excellent, effect is -0.41
  - If `health` = Fair, effect is  $(-0.41 - 4.83) = -5.24$
- Effect of `health` = Fair instead of Excellent?
  - If `exerany` = 0 (no), effect is 6.88
  - If `exerany` = 1 (yes), effect is  $(6.88 - 4.83) = 2.05$

# Plot the Residuals from model `m_1int`?



# Incorporating a Covariate into our two-way ANOVA models

# Taking Stock

So far, we've fit two models to predict bmi, using exerany and health, one with an interaction term and one without.

```
m_1 <- lm(bmi ~ exerany + health, data = train_w2im)
m_1int <- lm(bmi ~ exerany * health, data = train_w2im)
```

Next, we'll fit models incorporating a covariate, specifically, fruit\_day, a quantity (servings/day).

- m\_2 and m\_2int will add a linear term for fruit\_day
- Later models (we'll fit next time) will add various non-linear terms in fruit\_day
- We'll assess these models in our testing sample (next time) as well as our training sample.

**Giving away the ending:** We'll see that none of these augmented models will clearly improve the fit in our test sample over the performance of m\_1 and m\_1int.

# Adding in the covariate fruit\_day to m\_1

```
m_2 <- lm(bmi ~ fruit_day + exerany + health,  
          data = train_w2im)
```

- How well does this model fit the training data?

```
bind_rows(glance(m_1), glance(m_2)) %>%  
  mutate(mod = c("m_1", "m_2")) %>%  
  select(mod, r.sq = r.squared, adj.r.sq = adj.r.squared,  
         sigma, df, df.res = df.residual, AIC, BIC) %>%  
  kable(digits = c(0, 3, 3, 2, 0, 0, 1, 1))
```

mod	r.sq	adj.r.sq	sigma	df	df.res	AIC	BIC
m_1	0.089	0.082	6.12	5	664	4335.9	4367.5
m_2	0.098	0.090	6.09	6	663	4331.2	4367.3

- Also available in glance for a model fit with lm are statistic, p.value, logLik, and deviance.

# ANOVA for the m\_2 model

```
tidy(anova(m_2)) %>%  
  kable(dig = c(0, 0, 2, 2, 2, 3))
```

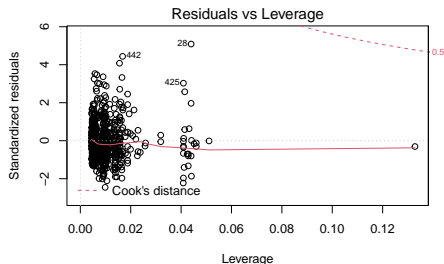
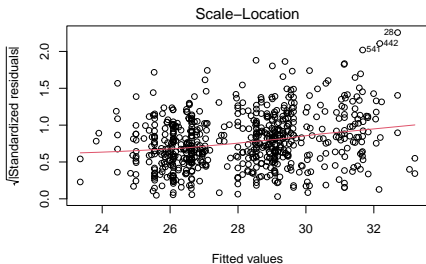
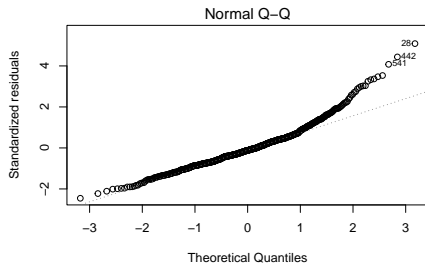
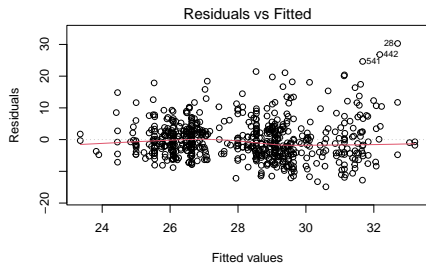
term	df	sumsq	meansq	statistic	p.value
fruit_day	1	468.10	468.10	12.62	0
exerany	1	760.50	760.50	20.51	0
health	4	1441.63	360.41	9.72	0
Residuals	663	24588.68	37.09	NA	NA

# Tidied summary of m\_2 coefficients

```
tidy(m_2, conf.int = TRUE, conf.level = 0.90) %>%  
  kable(digits = c(0,2,2,2,3,2,2))
```

term	estimate	std.error	statistic	p.value	conf.low	conf.high
(Intercept)	28.68	0.80	35.94	0.000	27.37	30.00
fruit_day	-0.55	0.21	-2.58	0.010	-0.90	-0.20
exerany	-2.05	0.55	-3.71	0.000	-2.95	-1.14
healthVG	0.55	0.70	0.79	0.430	-0.60	1.71
healthG	3.00	0.72	4.16	0.000	1.81	4.19
healthF	3.55	0.91	3.92	0.000	2.06	5.04
healthP	4.57	1.35	3.38	0.001	2.34	6.79

# m\_2 Residual Plots (non-constant variance?)





# Who is that poorest fit case?

Plot suggests we look at row 28

```
train_w2im %>% slice(28) %>%  
  select(ID, bmi, fruit_day, exerany, health) %>% kable()
```

ID	bmi	fruit_day	exerany	health
320	63	1	0	P

What is unusual about this subject?

```
train_w2im %$% sort(bmi) %>% tail()
```

```
[1] 50.46 51.22 51.54 56.31 58.98 63.00
```

# What if we included the interaction term?

```
m_2int <- lm(bmi ~ fruit_day + exerany * health,  
             data = train_w2im)
```

Compare m\_2int fit to previous models...

mod	r.sq	adj.r.sq	sigma	df	df.res	AIC	BIC
m_1	0.089	0.082	6.12	5	664	4335.9	4367.5
m_2	0.098	0.090	6.09	6	663	4331.2	4367.3
m_1int	0.126	0.114	6.01	9	660	4315.8	4365.4
m_2int	0.138	0.125	5.97	10	659	4309.1	4363.2

- m\_1 = no fruit\_day, no exerany\*health interaction
- m\_2 = fruit\_day, but no interaction
- m\_1int = no fruit\_day, with interaction
- m\_2int = both fruit\_day and interaction

# ANOVA for the `m_2int` model

```
tidy(anova(m_2int)) %>%  
  kable(dig = c(0, 0, 2, 2, 2, 3))
```

term	df	sumsq	meansq	statistic	p.value
fruit_day	1	468.10	468.10	13.12	0
exerany	1	760.50	760.50	21.32	0
health	4	1441.63	360.41	10.10	0
exerany:health	4	1080.39	270.10	7.57	0
Residuals	659	23508.29	35.67	NA	NA

# Tidied summary of m\_2int coefficients

```
tidy(m_2int, conf.int = TRUE, conf.level = 0.90) %>%  
  rename(se = std.error, t = statistic, p = p.value) %>%  
  kable(digits = c(0,2,2,2,3,2,2))
```

term	estimate	se	t	p	conf.low	conf.high
(Intercept)	28.28	1.43	19.73	0.000	25.91	30.64
fruit_day	-0.61	0.21	-2.93	0.004	-0.96	-0.27
exerany	-1.43	1.54	-0.93	0.353	-3.97	1.11
healthVG	-0.66	1.63	-0.40	0.686	-3.34	2.02
healthG	2.75	1.61	1.71	0.088	0.10	5.41
healthF	7.59	1.77	4.29	0.000	4.67	10.50
healthP	9.12	2.54	3.59	0.000	4.93	13.30
exerany:healthVG	1.59	1.79	0.88	0.377	-1.37	4.54
exerany:healthG	0.41	1.79	0.23	0.819	-2.54	3.37
exerany:healthF	-6.41	2.06	-3.11	0.002	-9.81	-3.02
exerany:healthP	-6.55	2.99	-2.19	0.029	-11.48	-1.62

# ANOVA comparison of m\_2 and m\_2int

```
anova(m_2, m_2int)
```

Analysis of Variance Table

Model 1: bmi ~ fruit\_day + exerany + health

Model 2: bmi ~ fruit\_day + exerany \* health

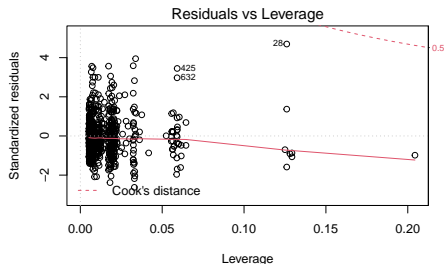
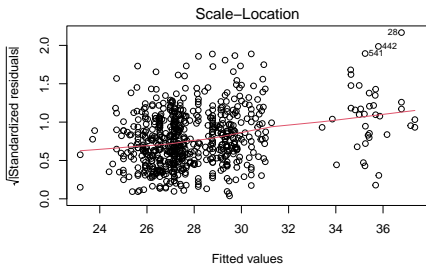
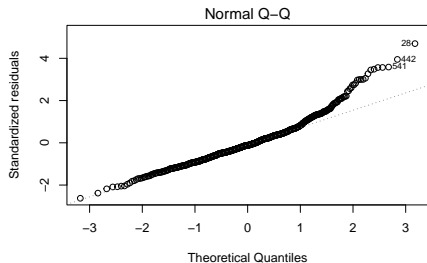
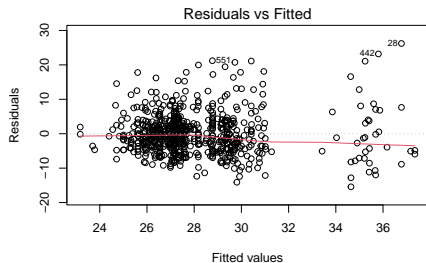
	Res.Df	RSS	Df	Sum of Sq	F	Pr(>F)
1	663	24589				
2	659	23508	4	1080.4	7.5716	5.751e-06 ***

---

Signif. codes:

0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

# Residual plots for model m\_2int?



# Which of the four models fits best?

In the **training** sample, we have...

mod	r.sq	adj.r.sq	sigma	df	df.res	AIC	BIC
m_1	0.089	0.082	6.12	5	664	4335.9	4367.5
m_2	0.098	0.090	6.09	6	663	4331.2	4367.3
m_1int	0.126	0.114	6.01	9	660	4315.8	4365.4
m_2int	0.138	0.125	5.97	10	659	4309.1	4363.2

- The interaction models look better by Adjusted  $R^2$  and  $\sigma$ ; AIC likes m\_2int while BIC likes m1. What to do?
- More importantly, the testing sample cannot judge between models accurately. Our models have already *seen* that data.
- For fairer comparisons, we'll need to consider the (held out) testing sample.

# Next Time

- Feedback from the Minute Paper after Class 03, due tomorrow at Noon, please.
- Assessing the models we've fit so far in the testing sample
- Incorporating polynomial terms and splines into linear regression (ANCOVA) models