

Package ‘heteroplasmy’

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Title Calculation of the standard error of the variance for heteroplasmy data

Version 0.0.1.0

Description The package offers different methods to quantify uncertainty by calculating the standard errors of the variance of given data. The included functions are primarily aimed to heteroplasmy data, which are not assumed to follow a predefined distribution and the sample size is usually low. Included, there is a set of synthetic datasets to use, as well as real heteroplasmy data from mouse specimen, found in (xxx Iain's paper xxx) The code and the methods described in the package are used in the (xxx our report xxx), so please cite that work in case you use the heteroplasmy package.

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Depends R (>= 3.1.0)

URL <https://github.com/kostasgian21/heteroplasmy>

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R topics documented:

analyticVar	2
bootstrapVar	3
bootstrapVarCor	4
heteroplasmyShift	5
jackVar	6
joint_neg_log_lik	7
kimura_neg_loglik	7
maxlik	8
maxlikboot	8
mousedataHB	9
mousedataLE	10
plotStdErrVar	10
readHeteroplasmyData	11
transfun	11

analyticVar

Analytic calculation of the standard error of the variance

Description

This function calculates analytically the standard error of the variance. It's based on the use of the appropriate h-statistic as an estimator

Usage

```
analyticVar(data, normal = FALSE)
```

Arguments

data	The input data in the form of a dataframe or matrix (which will be transformed into a dataframe). NA values are omitted.
normal	Parameter that indicates if the normal approximation should be used instead of the general formula from (Wilks, S. S. (1962).Mathematical Statistics). Default is FALSE.

Value

The analytically derived standard error of the variance of data.

Examples

```
# size of the sample
n=50
#generate a random sample of size n from a normal distribution
data_ex=rnorm(n,0.5,0.1)
analyticVar(data)

mouseData=readHeteroplasmyData("HB")
mouseData1 = mouseData[which(!is.na(mouseData[,1])),1]
analyticVar(mouseData1)

# use the package data and load it to variable mouseData
mouseData=mousedataLE
# calculate the standard error of the variance for the LE oocyte sample #3
bootstrapVar(mouseData[,3])
```

bootstrapVar	<i>A bootstrap method to calculate the standard error of the variance</i>
--------------	---

Description

This function uses the bootstrap method to calculate the uncertainty of the variance of a given sample based on random resampling. The number of the resamples is a parameter (default is 1000). Given that the resampling methods underestimate the uncertainty and thus provide a biased estimation, we offer the unbiased method as a default, although the user may change this option through the biased parameter for experimental purposes (they are strongly advised not to do for real problems with small samples).

Usage

```
bootstrapVar(data, nrep = 1000, biased = FALSE)
```

Arguments

data	The input data in the form of a dataframe or matrix (which will be transformed into a dataframe). NA values are omitted.
nrep	The number of bootstrap resamples. Default is 1000. The higher the number of the samples, the better the bootstrap outcome.
biased	A logical parameter to indicate if the user wants the biased version. Resampling techniques always underestimate statistics like the variance or the standard error of it for small samples.

Value

The standard error of the variance of data.

Examples

```
# size of the sample
n=50
#generate a random sample of size n from a normal distribution
data_ex=rnorm(n,0.5,0.1)
bootstrapVar(data)

mouseData=readHeteroplasmyData("HB")
mouseData1 = mouseData[which(!is.na(mouseData[,1])),1]
bootstrapVar(mouseData1)

# use the package data and load it to variable mouseData
mouseData=mousedataLE
# calculate the standard error of the variance for the LE oocyte sample #3
bootstrapVar(mouseData[,3])
```

bootstrapVarCor

*The bootstrapVar function with default correction***Description**

This function is simply the bootstrapVar with the correction argument being TRUE. it is provided as a separate function for usability. The function can be used beyond heteroplasmy data, therefore one can use it to calculate the standard error of the variance for samples where other approaches may not fit, eg when the sample size is too small and/or the population distribution is not Gaussian (or not known at all).

Usage

```
bootstrapVarCor(data, nrep = 1000)
```

Arguments

data	The input data in the form of a dataframe or matrix (which will be transformed into a dataframe). NA values are omitted.
nrep	The number of bootstrap resamples. Default is 1000. The higher the number of the samples, the better the bootstrap outcome (see par).

Value

The analytically derived standard error of the variance of data.

Author(s)

Kostas and Iain, <us@example.com>

References

<https://en.wikipedia.org/>

See Also

[bootstrapVar](#)

Examples

```
# size of the sample
n=50
#generate a random sample of size n from a normal distribution
data_ex=rnorm(n,0.5,0.1)
bootstrapVarCor(data)

mouseData=readHeteroplasmyData("HB")
mouseData1 = mouseData[which(!is.na(mouseData[,1])),1]
bootstrapVarCor(mouseData1,nrep=10000)

# use the package data and load it to variable mouseData
mouseData=mousedataLE
# calculate the standard error of the variance for the LE oocyte sample #3
bootstrapVar(mouseData[,3])
```

heteroplasmyShift	<i>Transformed heteroplasmy shift</i>
-------------------	---------------------------------------

Description

A numerical transformation of the heteroplasmy samples in order to work with the heteroplasmy shifts across diverse samples (e.g., due to time or different tissue samples). This transformation is used for comparing a heteroplasmy observation h to a reference value h_0 . It corresponds to the formula:

$$\Delta h = \ln \left(\frac{h(h_0 - 1)}{h_0(h - 1)} \right)$$

Usage

```
heteroplasmyShift(h, h0)
```

Arguments

<code>h</code>	The heteroplasmy observation. Can be either a single value or a vector of observations. Every observation should be in $[0, 1]$.
<code>h0</code>	The reference heteroplasmy value. Should be in $[0, 1]$.

Value

The Transformed heteroplasmy shift.

Author(s)

Kostas and Iain, <us@example.com>

References

[Site or paper](#)

See Also

[readHeteroplasmyData](#)

Examples

```
# size of the sample
n=50
#generate a random sample of size n from a normal distribution
data_ex=rnorm(n,0.5,0.1)
heteroplasmyShift(data)

mouseData=readHeteroplasmyData("HB")
mouseData1 = mouseData[which(!is.na(mouseData[,1])),1]
heteroplasmyShift(mouseData1,nrep=10000)
```

 jackVar

A jackknife method to compute the uncertainty of heteroplasmy data

Description

Similarly to the main bootstrapVar function that implements the bootstrap method to measure the standard error of the variance, the jackknife technique is another resampling method that can be used for the same purpose. Unlike bootstrapVar, jackVar (and very jackknife method) is deterministic and doesn't rely on randomness, but instead it uses removals of the sample points, one each time to calculate different sub-samples of size $(n-1)$. Note that the size of the input data should be strictly greater than 1.

Usage

```
jackVar(data)
```

Arguments

data	The input data in the form of a dataframe or matrix (which will be transformed into a dataframe). Its size should be ≥ 2 . NA values are omitted.
------	--

Value

The analytically derived standard error of the variance of data.

Examples

```
# size of the sample
n=50
#generate a random sample of size n from a normal distribution
data_ex=rnorm(n,0.5,0.1)
jackVar(data)

mouseData=readHeteroplasmyData("HB")
mouseData1 = mouseData[which(!is.na(mouseData[,1])),1]
jackVar(mouseData1)

# use the package data and load it to variable mouseData
mouseData=mousedataLE
# calculate the standard error of the variance for the LE oocyte sample #3
bootstrapVar(mouseData[,3])
## Not run:
#input data of size 1 will fail
data_ex=rnorm(1,0.5,0.1)
jackVar(data)

## End(Not run)
```

joint_neg_log_lik	<i>Joint negative log likelihood function for several heteroplasmy measurements</i>
-------------------	---

Description

joint negative log likelihood function for several families' heteroplasmy measurements $\theta = [b, h_{0.1}, h_{0.2}, \dots]$ (use h values if `use.h0s=F`, otherwise initial heteroplasms are enforced via `h0s`).

Usage

```
joint_neg_log_lik(theta, hlist, use.h0s = F, h0s = -1)
```

Arguments

<code>theta</code>	Kimura parameters p (or h_0 here) and b .
<code>hlist</code>	TBDD list of different sets of heteroplasmy measurements
<code>use.h0s</code>	TBDD
<code>h0s</code>	TBDD

Value

The negative log likelihood for the list of inputs.

Examples

```
X.1 = rnorm(50,0.5,0.1)
joint_neg_log_lik(c(0.5,0.91),X.1)
```

kimura_neg_loglik	<i>Calculate negative log likelihood for heteroplasmy measurements</i>
-------------------	--

Description

calculate negative log likelihood for a given set of heteroplasmy measurements h and parameters $\theta = \text{logit}(p), \text{logit}(b)$ (we write h_0 for p) we can do this enforcing a particular h_0 value (passed as an argument) or treating h_0 as a fit parameter (default) the logit transform is used to ensure h_0 and b remain in the [0,1](#) interval regardless of what real-valued argument the numerical optimiser attempts

Usage

```
kimura_neg_loglik(theta, h, h0 = F)
```

Arguments

<code>theta</code>	Kimura parameters p (or h_0 here) and b .
<code>h</code>	The heteroplasmy measurements.
<code>h0</code>	A particular h_0 value Default is to treat h_0 as a fit parameter

Value

The negative log likelihood for the input.

Examples

```
X.1 = rnorm(50,0.5,0.1)
kimura_neg_loglik(c(0.5,0.91),X.1)
```

maxlik	<i>compute maximum likelihood parameters and confidence intervals for heteroplasmy data</i>
--------	---

Description

compute maximum likelihood parameters and confidence intervals for a given heteroplasmy set. We can do this while imposing a specific h0 as an argument or allowing a search over h0 values.

Usage

```
maxlik(h, conf.level = 0.95, h0 = F)
```

Arguments

h	The heteroplasmy measurements.
conf.level	The preferred confidence interval calculation, Default value is 0,95 (95%).
h0	A particular h0 value Default is to treat h0 as a fit parameter

Value

The maximum likelihood for the input data according to the Kimura distribution

Examples

```
X.1 = rnorm(50,0.5,0.1)
joint_neg_log_lik(c(0.5,0.91),X.1)
```

maxlikboot	<i>bBootstrap estimates for parameters and confidence intervals for heteroplasmy data</i>
------------	---

Description

compute bootstrap estimates for parameters and confidence intervals for a given heteroplasmy set. We can do this while imposing a specific h0 as an argument or allowing a search over h0 values

Usage

```
maxlikboot(h, nboot = 200, conf.level = 0.95, h0 = F)
```


Arguments

<code>h</code>	The heteroplasmy measurements.
<code>nboot</code>	The number of bootstrap samples.
<code>conf.level</code>	The preferred confidence interval calculation, Default value is 0.95 (95%).
<code>h0</code>	A particular h0 value Default is to treat h0 as a fit parameter

Value

The maximum likelihood for the input data according to the Kimura distribution (using bootstrapping)

Examples

```
X.1 = rnorm(50,0.5,0.1)
joint_neg_log_lik(c(0.5,0.91),X.1)
```

mouseDataHB

HB oocyte heteroplasmy data

Description

A dataset containing heteroplasmy values for the HB oocyte mouse lines. Each column corresponds to a different specimen. NA values have been added to make the number of rows equal. Please remove them after loading. Heteroplasmy data of the package are in the range $[0, 1]$.

Usage

```
mouseDataHB
```

Format

A data frame with 25 rows and 56 columns (TO FIX):

price price, in US dollars

carat weight of the diamond, in carats

Source

<http://www.example.info/>

mousedataLE	<i>LE oocyte heteroplasmy data</i>
-------------	------------------------------------

Description

A dataset containing hetoplasmy values for the HB oocyte mouse lines. Each column corresponds to a different specimen. NA values have been added to make the number of rows equal. Please remove them after loading. Heteroplasmy data of the package are in the range $[0, 1]$.

Usage

```
mousedataLE
```

Format

A data frame with 20 rows and 43 columns (TO FIX):

price price, in US dollars

carat weight of the diamond, in carats

Source

<http://www.example.info/>

plotStdErrVar	<i>An example plotting function</i>
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Description

This function is used as a toy example on how to represent the data statistics regarding the variance of the sample. The mean variance and its standard error are depicted (XX maybe plot $2 \times \text{SEM}$? XX). Note that this is just an illustration to show that the analytic and the resampling approaches almost match each other.

Usage

```
plotStdErrVar(
  data,
  functions = c("normalApr", "analytic", "bootstrap", "jackknife"),
  ...
)
```

Arguments

data	The input data in the form of a dataframe or matrix (which will be transformed into a dataframe).
functions	Choose the subset of the functions you wish use for the calculation and subsequent plot of the standard error of the variance. You can use one or a combination of "normalApr", "analytic", "bootstrap", "correctedBoot", and "jackknife". For now, it outputs all of the aforementioned methods!

Warning

This is a plotting function just for demonstration purposes.

Examples

```
# size of the sample
n=50
#generate a random sample of size n from a normal distribution
data_ex=rnorm(n,0.5,0.1)
plotStdErrVar(data_ex)
```

`readHeteroplasmyData` *A Function to read mouse heteroplasmy data (not finished!)*

Description

This function allows you to read mouse heteroplasmy data from external files. Use with caution (for now).

Usage

```
readHeteroplasmyData(nameD = "HB")
```

Arguments

named	Either "HB" or "LE".
-------	----------------------

Value

A dataframe containing mouse heteroplasmy data.

Examples

```
readHeteroplasmyData(nameD="LE")
```

transfun	<i>A Function to cast real numbers to $[0, 1]$</i>
----------	---

Description

A transformation function to cast any real number onto the interval $[0, 1]$. Equivalent to the inverse logit transform.

Usage

transfun(x)

Arguments

x an integer number to be transformed.

Value

The transformed cast of the input value to the interval $[0, 1]$.

Examples

```
transfun(5.1)
```

Index

- * **bootstrapVarCor**,
bootstrapVarCor, 4
- * **bootstrap**
bootstrapVar, 3
- * **datasets**
mousedataHB, 9
mousedataLE, 10
- * **data**
readHeteroplasmyData, 11
- * **error**
analyticVar, 2
- * **fitted**
bootstrapVarCor, 4
- * **h-statistic**
analyticVar, 2
- * **heteroplasmy,transformation,shift**
heteroplasmyShift, 5
- * **heteroplasmy**
analyticVar, 2
bootstrapVar, 3
jackVar, 6
readHeteroplasmyData, 11
- * **jackknife**
jackVar, 6
- * **joint**
joint_neg_log_lik, 7
maxlik, 8
maxlikboot, 8
- * **kimura**
joint_neg_log_lik, 7
kimura_neg_loglik, 7
maxlik, 8
maxlikboot, 8
- * **likelihood**
joint_neg_log_lik, 7
kimura_neg_loglik, 7
maxlik, 8
maxlikboot, 8
- * **logit**
transfun, 11
- * **log**
joint_neg_log_lik, 7
kimura_neg_loglik, 7
maxlik, 8
maxlikboot, 8
- * **negative**
joint_neg_log_lik, 7
kimura_neg_loglik, 7
maxlik, 8
maxlikboot, 8
- * **plot,standard,error**
plotStdErrVar, 10
- * **resampling**
bootstrapVar, 3
jackVar, 6
- * **reverse**
transfun, 11
- * **standard**
analyticVar, 2
- * **uncertainty**
bootstrapVar, 3
jackVar, 6
- * **variance**
analyticVar, 2
0,1, 7
analyticVar, 2
bootstrapVar, 3, 4
bootstrapVarCor, 4
heteroplasmyShift, 5
jackVar, 6
joint_neg_log_lik, 7
kimura_neg_loglik, 7
maxlik, 8
maxlikboot, 8
mousedataHB, 9
mousedataLE, 10
par, 4
plotStdErrVar, 10
readHeteroplasmyData, 5, 11
transfun, 11