

# MAGERI benchmark using reference standard DNA library

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Load metadata

```
df.vmeta <- read.table("hd734_variant_metadata.txt", sep="\t", header=T) # variants observed in HD734 a
df.smeta <- read.table("sample_metadata.txt", sep="\t", header=T) # metadata for amplicon sequencing sa
```

VCF parsing function

```
library(stringr)
library(pROC)
```

```
## Type 'citation("pROC")' for a citation.
```

```
##
```

```
## Attaching package: 'pROC'
```

```
## The following objects are masked from 'package:stats':
```

```
##
```

```
##      cov, smooth, var
```

```
library(ggplot2)
```

```
library(ggbeeswarm)
```

```
library(dplyr)
```

```
##
```

```
## Attaching package: 'dplyr'
```

```
## The following objects are masked from 'package:stats':
```

```
##
```

```
##      filter, lag
```

```
## The following objects are masked from 'package:base':
```

```
##
```

```
##      intersect, setdiff, setequal, union
```

```
read_vcf <- function(file_name) {
```

```
  .vcf <- read.table(file_name, header = F, sep = "\t", stringsAsFactors = F)
```

```
  colnames(.vcf) <- c("chromosome", "position", "skip1", "from", "to", "qual", "skip2", "info", "skip3"
```

```
  .vcf$skip1 <- NULL
```

```
  .vcf$skip2 <- NULL
```

```
  .vcf$skip3 <- NULL
```

```
  .vcf$skip4 <- NULL
```

```

.vcf$qual <- as.integer(.vcf$qual)
.vcf <- subset(.vcf, nchar(from) == 1 &
               nchar(to) == 1 & !is.na(qual)) # no indels

.infosplit <- str_split_fixed(.vcf$info, regex("[=;]"), 15)[,c(2, 4, 12, 14)]

.vcf$coverage <- as.numeric(.infosplit[,1])
.vcf$frequency <- as.numeric(.infosplit[,2])
.vcf$bb.a <- as.numeric(.infosplit[,3])
.vcf$bb.b <- as.numeric(.infosplit[,4])
.vcf$info <- NULL

.vcf$count <- as.integer(round(.vcf$coverage * .vcf$frequency))

.vcf
}

head(read_vcf("p126.h4_2_ballast_m1.vcf"))

```

```
## Warning in read_vcf("p126.h4_2_ballast_m1.vcf"): NAs introduced by coercion
```

```

##   chromosome position from to qual coverage   frequency      bb.a
## 1      chr2 212295704    C  A     8      2265 0.0004415011 0.9957058
## 2      chr2 212295705    C  A     8      2265 0.0004415011 0.9957058
## 3      chr2 212295713    A  G     7      2265 0.0004415011 1.0295689
## 4      chr2 212295718    G  A     6      2265 0.0004415011 2.1314828
## 5      chr2 212295725    C  A     8      2264 0.0004416961 0.9957058
## 6      chr2 212295732    C  T    24      2264 0.0013250883 2.1314828
##           bb.b count
## 1 21862.23      1
## 2 21862.23      1
## 3 16394.12      1
## 4 24344.82      1
## 5 21862.23      1
## 6 24344.82      3

```

Read samples with HD734 standard DNA and control human DNA, append metadata

```
library(TailRank) # For betabinom.
```

```
## Loading required package: oompaBase
```

```

# installing:
# source("https://bioconductor.org/biocLite.R")
# biocLite("Biobase")
# install.packages("TailRank", repos="http://R-Forge.R-project.org")

df <- data.frame()

read_vcf_with_metadata <- function(file_name, primer_set, replica, ratio, type) {

```

```

.vcf <- read_vcf(file_name)
.vcf <- merge(.vcf, df.vmeta, all.x = type != "standard", all.y = F)
.vcf$known.frequency <- .vcf$known.frequency * ratio
.vcf$known.frequency[is.na(.vcf$known.frequency)] <- 0
.vcf$primer_set <- primer_set
.vcf$replica <- primer_set
.vcf$type <- type

.vcf <- subset(.vcf, frequency < 0.4 & count > 0) # remove alleles in control

.vcf
}

for (i in 1:nrow(df.smeta)) {
  df <- with(df.smeta, rbind(df,
                             read_vcf_with_metadata(paste(prefix[i], "vcf", sep="."),
                                                         primer_set[i],
                                                         replica[i],
                                                         ratio[i],
                                                         type[i])))
}

```

```

## Warning in read_vcf(file_name): NAs introduced by coercion
## Warning in read_vcf(file_name): NAs introduced by coercion
## Warning in read_vcf(file_name): NAs introduced by coercion
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## Warning in read_vcf(file_name): NAs introduced by coercion

```

```
## Warning in read_vcf(file_name): NAs introduced by coercion
## Warning in read_vcf(file_name): NAs introduced by coercion
## Warning in read_vcf(file_name): NAs introduced by coercion
```

```
# Compute scores

df$qual <- with(df,
# mapply(function(x,y,a,b) 1 - pbb(x,y,a,b) + 0.5 * dbb(x,y,a,b),
#         count, coverage, bb.a, bb.b)
# )

df$qual <- ifelse(df$qual <= 1e-100, 999, -10*log10(df$qual))
```

Group observed variants into tiers

```
df$tier <- cut(df$known.frequency, c(-1, 0, 0.009, 0.02, 1))
levels(df$tier) <- c("error", "0.1%", "1%", "5%")
summary(df$tier)
```

```
## error 0.1% 1% 5%
## 2468 43 46 12
```

Number of detected variants and sampling

```
df.v.summary <- df.vmeta
df.v.summary$known.frequency <- df.v.summary$known.frequency * 0.1 # 1:9 diluted samples
df.v.summary$ratio <- "1:10"
df.v.summary.2 <- df.vmeta
df.v.summary.2$ratio <- "1:1"
df.v.summary <- rbind(df.v.summary, df.v.summary.2)
df.v.summary$tier <- cut(df.v.summary$known.frequency, c(-1, 0, 0.009, 0.02, 1))
levels(df.v.summary$tier) <- c("error", "0.1%", "1%", "5%")

print(df.v.summary)
```

```
## chromosome position from to
## 1 chr7 140453136 A T
## 2 chr7 140453136 A C
## 3 chr7 140453137 C T
## 4 chr7 55241707 G A
## 5 chr7 55259515 T G
## 6 chr7 55259524 T A
## 7 chr7 55249071 C T
## 8 chr19 3118942 A T
## 9 chr2 209113112 C T
## 10 chr2 209113113 G A
## 11 chr12 25378562 C T
## 12 chr12 25398284 C G
## 13 chr12 25398285 C A
```

## 14	chr12	25398284	C	T
## 15	chr12	25398285	C	G
## 16	chr12	25398285	C	T
## 17	chr12	25398284	C	A
## 18	chr12	25398281	C	T
## 19	chr12	25380275	T	G
## 20	chr12	25380276	T	A
## 21	chr1	115256528	T	A
## 22	chr1	115256530	G	T
## 23	chr1	115256529	T	A
## 24	chr1	115256529	T	C
## 25	chr3	178936082	G	A
## 26	chr3	178936091	G	A
## 27	chr3	178952085	A	G
## 28	chr7	55242465	GGGAATTAAGAGAAGC	G
## 29	chr7	140453136	A	T
## 30	chr7	140453136	A	C
## 31	chr7	140453137	C	T
## 32	chr7	55241707	G	A
## 33	chr7	55259515	T	G
## 34	chr7	55259524	T	A
## 35	chr7	55249071	C	T
## 36	chr19	3118942	A	T
## 37	chr2	209113112	C	T
## 38	chr2	209113113	G	A
## 39	chr12	25378562	C	T
## 40	chr12	25398284	C	G
## 41	chr12	25398285	C	A
## 42	chr12	25398284	C	T
## 43	chr12	25398285	C	G
## 44	chr12	25398285	C	T
## 45	chr12	25398284	C	A
## 46	chr12	25398281	C	T
## 47	chr12	25380275	T	G
## 48	chr12	25380276	T	A
## 49	chr1	115256528	T	A
## 50	chr1	115256530	G	T
## 51	chr1	115256529	T	A
## 52	chr1	115256529	T	C
## 53	chr3	178936082	G	A
## 54	chr3	178936091	G	A
## 55	chr3	178952085	A	G
## 56	chr7	55242465	GGGAATTAAGAGAAGC	G
##			id known.frequency	reference ratio tier
## 1		BRAF_V600E,BRAF_V600K	0.0080	BRAF_E15 1:10 0.1%
## 2		BRAF_V600G,BRAF_V600R	0.0010	BRAF_E15 1:10 0.1%
## 3		BRAF_V600M,BRAF_V600K,BRAF_V600R	0.0010	BRAF_E15 1:10 0.1%
## 4		EGFR_G719S	0.0167	EGFR_E18 1:10 1%
## 5		EGFR_L858R	0.0010	EGFR_E21 1:10 0.1%
## 6		EGFR_L861Q	0.0010	EGFR_E21 1:10 0.1%
## 7		EGFR_T790M	0.0010	EGFR_E20 1:10 0.1%
## 8		GNA11_Q209L	0.0013	GNA11_E5 1:10 0.1%
## 9		IDH1_R132C	0.0013	IDH1_E4 1:10 0.1%
## 10		IDH1_R132H	0.0013	IDH1_E4 1:10 0.1%

## 11	KRAS_A146T	0.0013	KRAS_E4	1:10	0.1%
## 12	KRAS_G12A	0.0013	KRAS_E2	1:10	0.1%
## 13	KRAS_G12C	0.0013	KRAS_E2	1:10	0.1%
## 14	KRAS_G12D	0.0013	KRAS_E2	1:10	0.1%
## 15	KRAS_G12R	0.0013	KRAS_E2	1:10	0.1%
## 16	KRAS_G12S	0.0013	KRAS_E2	1:10	0.1%
## 17	KRAS_G12V	0.0013	KRAS_E2	1:10	0.1%
## 18	KRAS_G13D	0.0250	KRAS_E2	1:10	5%
## 19	KRAS_Q61H	0.0013	KRAS_E3	1:10	0.1%
## 20	KRAS_Q61L	0.0013	KRAS_E3	1:10	0.1%
## 21	NRAS_Q61H	0.0013	NRAS_E3	1:10	0.1%
## 22	NRAS_Q61K	0.0013	NRAS_E3	1:10	0.1%
## 23	NRAS_Q61L	0.0013	NRAS_E3	1:10	0.1%
## 24	NRAS_Q61R	0.0013	NRAS_E3	1:10	0.1%
## 25	PIK3CA_E542K	0.0013	PIK3CA_E9	1:10	0.1%
## 26	PIK3CA_E545K	0.0013	PIK3CA_E9	1:10	0.1%
## 27	PIK3CA_H1047R	0.0300	PIK3CA_E20	1:10	5%
## 28	EGFR_dE746-A750	0.0010	EGFR_E19	1:10	0.1%
## 29	BRAF_V600E,BRAF_V600K	0.0800	BRAF_E15	1:1	5%
## 30	BRAF_V600G,BRAF_V600R	0.0100	BRAF_E15	1:1	1%
## 31	BRAF_V600M,BRAF_V600K,BRAF_V600R	0.0100	BRAF_E15	1:1	1%
## 32	EGFR_G719S	0.1670	EGFR_E18	1:1	5%
## 33	EGFR_L858R	0.0100	EGFR_E21	1:1	1%
## 34	EGFR_L861Q	0.0100	EGFR_E21	1:1	1%
## 35	EGFR_T790M	0.0100	EGFR_E20	1:1	1%
## 36	GNA11_Q209L	0.0130	GNA11_E5	1:1	1%
## 37	IDH1_R132C	0.0130	IDH1_E4	1:1	1%
## 38	IDH1_R132H	0.0130	IDH1_E4	1:1	1%
## 39	KRAS_A146T	0.0130	KRAS_E4	1:1	1%
## 40	KRAS_G12A	0.0130	KRAS_E2	1:1	1%
## 41	KRAS_G12C	0.0130	KRAS_E2	1:1	1%
## 42	KRAS_G12D	0.0130	KRAS_E2	1:1	1%
## 43	KRAS_G12R	0.0130	KRAS_E2	1:1	1%
## 44	KRAS_G12S	0.0130	KRAS_E2	1:1	1%
## 45	KRAS_G12V	0.0130	KRAS_E2	1:1	1%
## 46	KRAS_G13D	0.2500	KRAS_E2	1:1	5%
## 47	KRAS_Q61H	0.0130	KRAS_E3	1:1	1%
## 48	KRAS_Q61L	0.0130	KRAS_E3	1:1	1%
## 49	NRAS_Q61H	0.0130	NRAS_E3	1:1	1%
## 50	NRAS_Q61K	0.0130	NRAS_E3	1:1	1%
## 51	NRAS_Q61L	0.0130	NRAS_E3	1:1	1%
## 52	NRAS_Q61R	0.0130	NRAS_E3	1:1	1%
## 53	PIK3CA_E542K	0.0130	PIK3CA_E9	1:1	1%
## 54	PIK3CA_E545K	0.0130	PIK3CA_E9	1:1	1%
## 55	PIK3CA_H1047R	0.3000	PIK3CA_E20	1:1	5%
## 56	EGFR_dE746-A750	0.0100	EGFR_E19	1:1	1%

```
df.v.summary <- df.v.summary %>%
  group_by(tier) %>%
  summarise(total = n())
df.v.summary$total <- 2 * df.v.summary$total # we have 2 independent experiments

df.v.summary <- merge(df.v.summary,
  df %>% group_by(tier) %>%
```

```

        summarise(found = n(),
                  detect.prob.m = mean(1-exp(-frequency * coverage)),
                  detect.prob.sd = sd(1-exp(-frequency * coverage)))

df.v.summary$detection.prob.lb <- with(df.v.summary,
                                     detect.prob.m - 1.96*detect.prob.sd - 5e-3)
df.v.summary$detection.prob.ub <- with(df.v.summary,
                                     detect.prob.m + 1.96*detect.prob.sd)

df.v.summary$detection.prob.ub <- ifelse(df.v.summary$detection.prob.ub > 1,
                                     1,
                                     df.v.summary$detection.prob.ub )

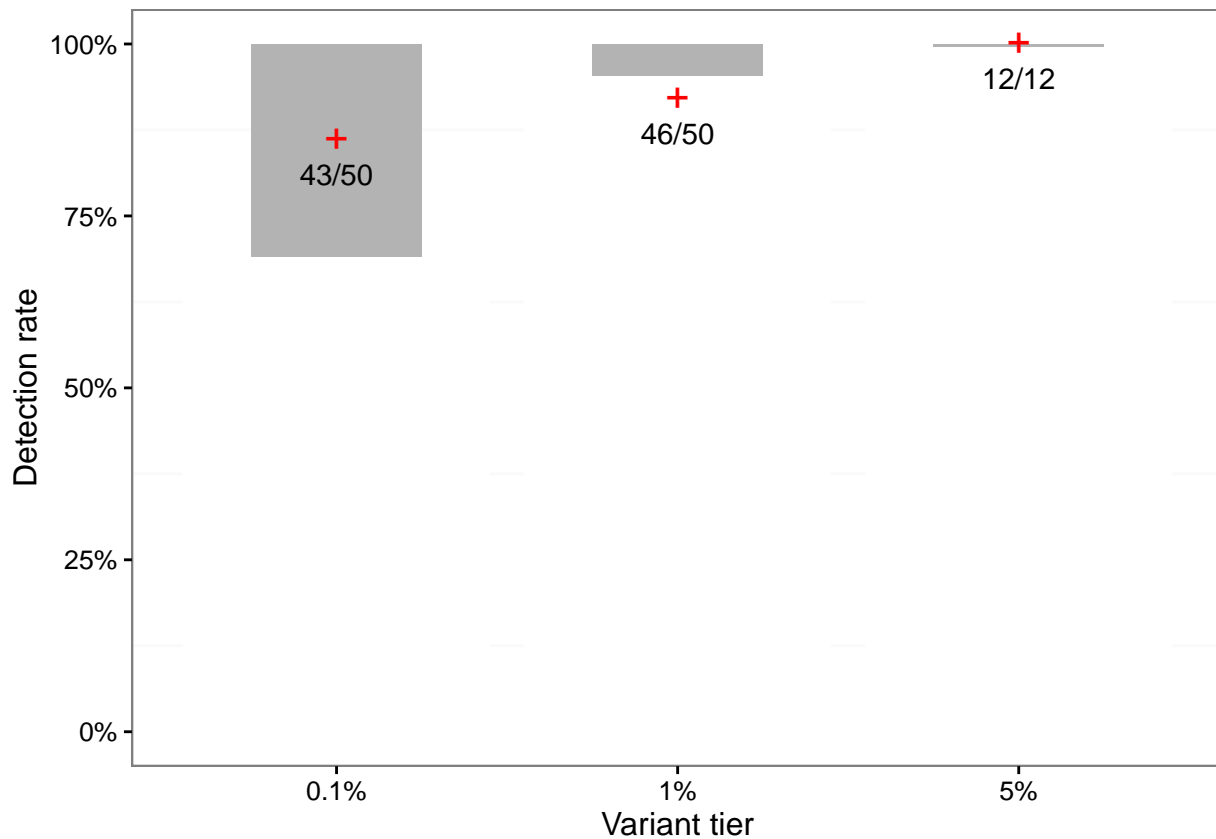
library(scales)

fig3 <- ggplot(df.v.summary, aes(x=tier)) +
  geom_bar(stat="identity", aes(y=detection.prob.ub), fill = "grey70", width=0.5)+
  geom_bar(stat="identity", aes(y=detection.prob.lb), fill = "white")+
  geom_point(aes(y = found/total), size = 5, color="red", shape="+") +
  geom_text(aes(y = found/total - 0.05, label = paste(found, total, sep="/"))) +
  scale_y_continuous("Detection rate", labels=percent) +
  xlab("Variant tier") +
  theme_bw() +
  theme(panel.grid.major.x = element_blank(),
        panel.grid.major.y = element_blank())

save(file = "../figures/fig3.Rda", fig3)

fig3

```



Quality score and error P-values

```
df.e <- df %>%
  filter(type == "blank") %>%
  mutate(true.p.value = -10 * log10(1 - (rank(frequency) - 0.5) / n()))

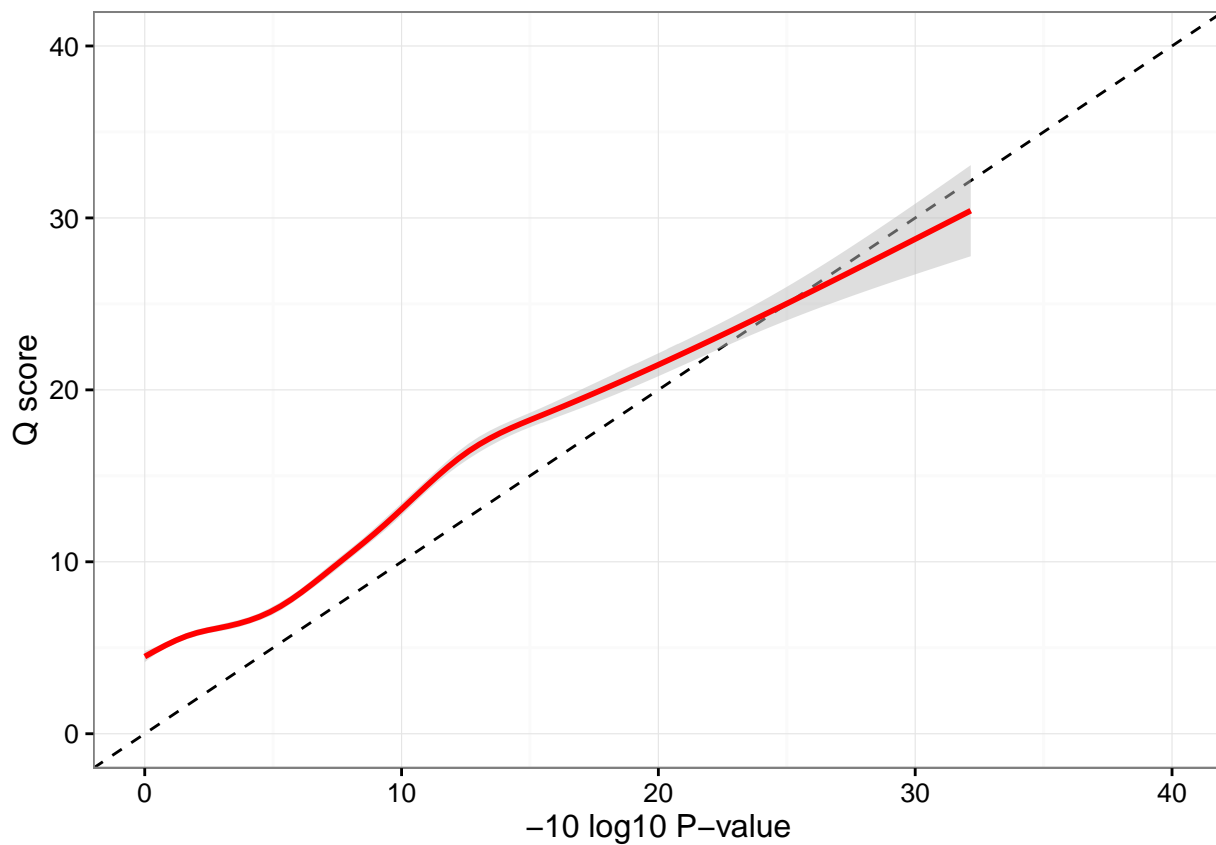
fig4 <- ggplot(df.e, aes(true.p.value, qual)) +
  geom_abline(slope = 1, intercept = 0, linetype = "dashed") +
  geom_smooth(color="red", fill="grey", alpha=0.5) +
  scale_x_continuous("-10 log10 P-value", limits = c(0, 40)) +
  scale_y_continuous("Q score", limits = c(0, 40)) +
  theme_bw()

save(file = "../figures/fig4.Rda", fig4)

fig4
```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```



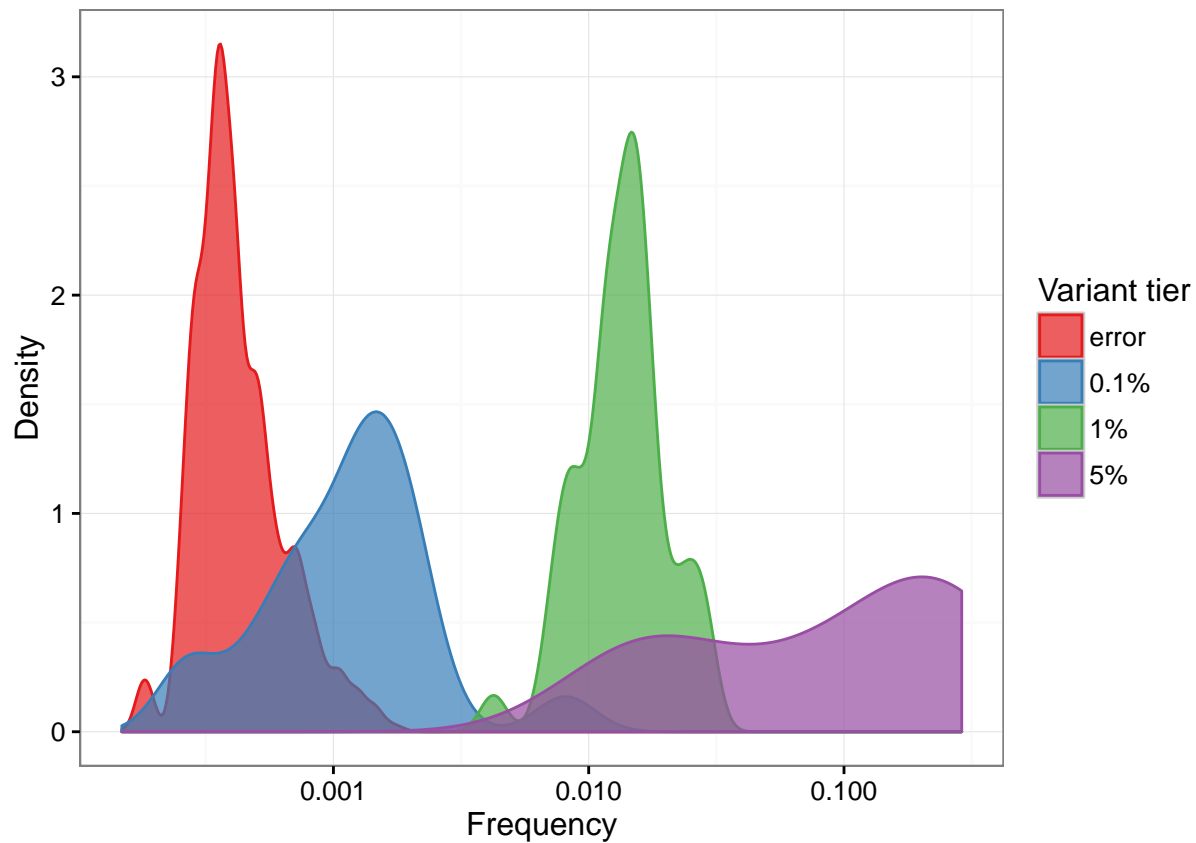


Frequency of variants and errors

```
fig5 <- ggplot(df, aes(x=frequency, color = tier)) +
  geom_density(aes(fill=tier), alpha=0.7) +
  ylab("Density") +
  scale_x_log10("Frequency") +
  scale_color_brewer("Variant tier", palette = "Set1") +
  scale_fill_brewer("Variant tier", palette = "Set1") +
  theme_bw()

save(file = "../figures/fig5.Rda", fig5)

fig5
```

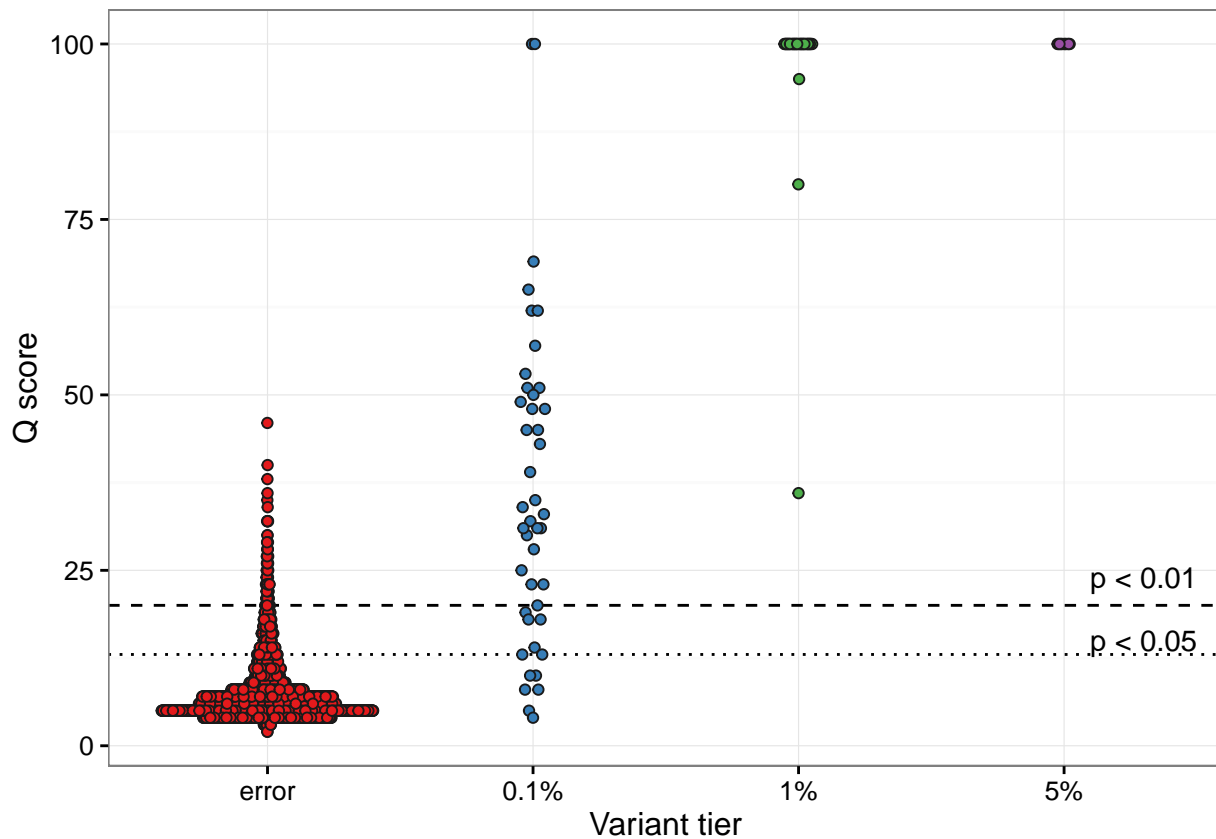


Variant quality scores

```
fig6 <- ggplot(df,aes(tier, qual, fill=tier)) +
  geom_hline(yintercept = 13.0103, linetype = "dotted") +
  geom_hline(yintercept = 20.0103, linetype = "dashed") +
  geom_quasirandom(varwidth = T, shape=21, color="grey10") +
  annotate("text", label = "p < 0.05", x=4.3,y=13.0103+2) +
  annotate("text", label = "p < 0.01", x=4.3,y=20.0103+4) +
  scale_fill_brewer("", palette = "Set1", guide=F) +
  xlab("Variant tier") + ylab("Q score") +
  theme_bw()
```

```
save(file = "../figures/fig6.Rda", fig6)
```

```
fig6
```



ROC curve for rare variants

```
df.1 <- subset(df, tier %in% c("error", "0.1%"))
df.1$type <- ifelse(df.1$tier == "error", 0, 1)

make_roc <- function(rocobj, type) {
  .df.roc <- data.frame(spec = seq(0, 1, 0.01))
  sens.ci <- ci.se(rocobj, specificities = .df.roc$spec)

  .df.roc$sens.lo <- sens.ci[,1]
  .df.roc$sens.me <- sens.ci[,2]
  .df.roc$sens.hi <- sens.ci[,3]
  .df.roc$type <- type

  .df.roc
}

rocobj <- roc(type ~ qual, df.1, ci=T)
print(rocobj)

##
## Call:
## roc.formula(formula = type ~ qual, data = df.1, ci = T)
##
## Data: qual in 2468 controls (type 0) < 43 cases (type 1).
## Area under the curve: 0.9277
## 95% CI: 0.8727-0.9827 (DeLong)
```

```
df.roc <- make_roc(rocobj, "Q score")
rocobj <- roc(type ~ frequency, df.1, ci=T)
print(rocobj)
```

```
##
## Call:
## roc.formula(formula = type ~ frequency, data = df.1, ci = T)
##
## Data: frequency in 2468 controls (type 0) < 43 cases (type 1).
## Area under the curve: 0.8637
## 95% CI: 0.7839-0.9435 (DeLong)
```

```
df.roc <- rbind(df.roc, make_roc(rocobj, "Frequency"))

fig7 <- ggplot(df.roc, aes(x=spec)) +
  geom_ribbon(aes(ymin=sens.lo, ymax=sens.hi, group=type), fill="grey", alpha=0.5) +
  geom_abline(slope = 1, intercept = 1, linetype = "dashed") +
  geom_line(aes(y=sens.me, color = type)) +
  scale_x_reverse("Specificity") +
  scale_y_continuous("Sensitivity", limits=c(0,1)) +
  scale_color_brewer("Threshold", palette = "Set1") +
  theme_bw()

save(file = "../figures/fig7.Rda", fig7)
```

fig7

