

# Project Development Phase

Utilization Of Algorithms,Dynamic  
Programming,Optimal Memory Utilization

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Project Name	Block Chain Technology For Electronic Health Records

## Background:

Covariance models (CMs) are probabilistic models of RNA secondary structure, analogous to profile hidden Markov models of linear sequence. The dynamic programming algorithm for aligning a CM to an RNA sequence of length  $N$  is  $O(N^3)$  in memory. This is only practical for small RNAs.

## Results:

**I describe a divide and conquer variant of the alignment algorithm that is analogous to memory-efficient Myers/Miller dynamic programming algorithms for linear sequence alignment. The new algorithm has an  $O(N^2 \log N)$  memory complexity, at the expense of a small constant factor in time.**

## Conclusions:

**Optimal ribosomal RNA structural alignments that previously required up to 150 GB of memory now require less than 270 MB.**

*From guide tree to covariance model:*

The final CM is an array of  $M$  states, connected as a directed graph by transitions  $t_v(y)$  (or probability 1 transitions  $v \rightarrow (y,z)$  for bifurcations) with the states numbered such that  $(y,z) \geq v$ . There are no cycles in the directed graph other than cycles of length one (e.g. the self-transitions of the insert states). We can think of the CM as an array of states in which all transition dependencies run in one direction; we can do an iterative dynamic programming calculation through the model states starting with the last numbered end state  $M$  and ending in the root state 1. An example CM, corresponding to the input alignment of Figure [Figure1,1](#), is shown in Figure [Figure33](#).

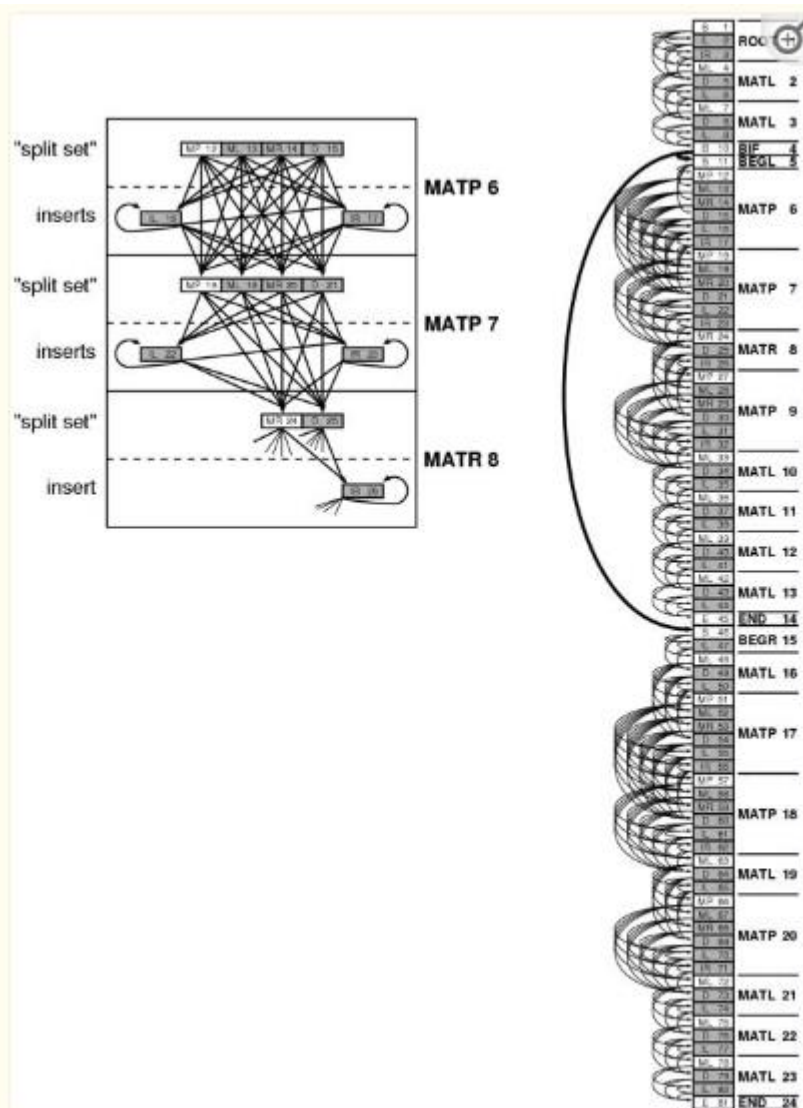
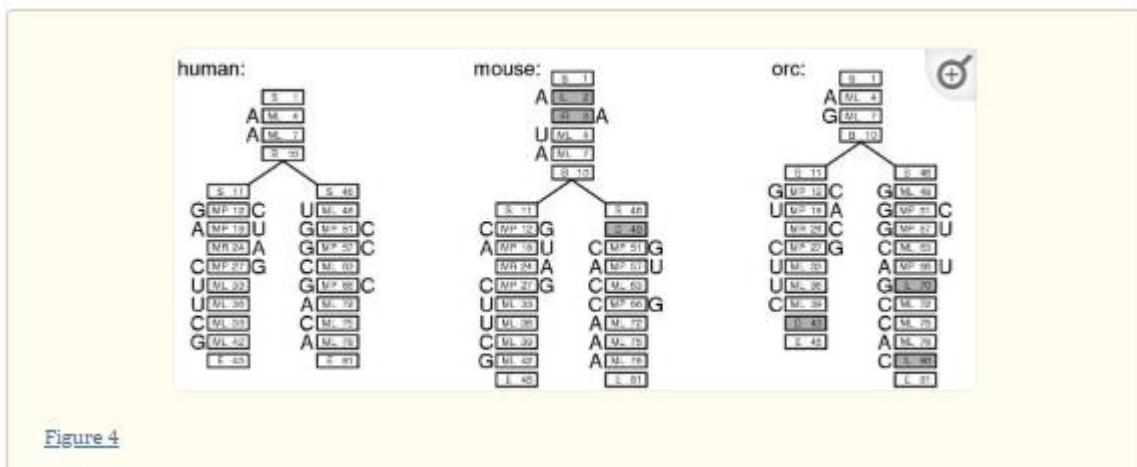


Figure 3

## Parameterization:

Using the guide tree and the final CM, each individual sequence in the input multiple alignment can be converted unambiguously to a CM parse tree, as shown in Figure [Figure4.4](#). Counts for observed state transitions and singlet/pair emissions are then collected from these parse trees. The observed counts are converted to transition and

emission probabilities by standard procedures. I calculate maximum a posteriori parameters, using Dirichlet priors.



*Using CYK/inside and CYK/outside to divide and conquer:*

- The bifurcation-dependent strategy is a special case of this more general splitting strategy, where the B state is the only member of its split set, and where we also take advantage of the fact that  $\alpha_v(i,j) = \max_k \alpha_w(i, k) + \alpha_y(k + 1, j)$ . By carrying out the  $\max_k$  operation during the split, rather than before, we can split the current problem into three optimal pieces instead of just two.
- If we look at the consequences of these splitting strategies, we see we will have to deal with three types of problems (Figure [Figure55](#)):

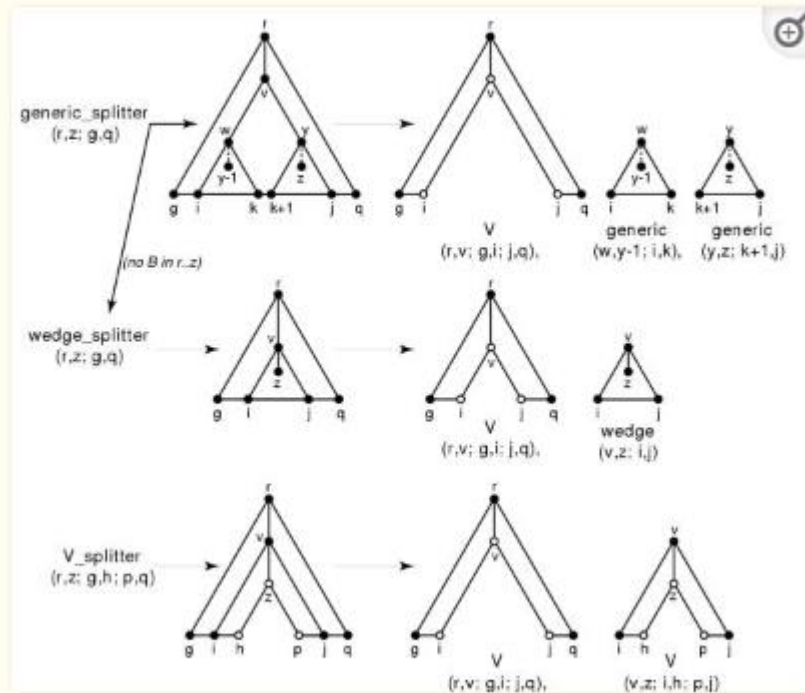


Figure 5

## Empirical results:

The same results are plotted in Figure [Figure 6.6](#). Memory requirements scale as expected:  $N^3$  for standard CYK alignment, and better than  $N^2 \log N$  for the divide and conquer algorithm. Empirical CPU time requirements scale similarly for the two algorithms ( $N^{3.24} - N^{3.29}$ ). The observed performance is better than the theoretical worst case of  $O(N^4)$ . The proportion of extra time required by divide and conquer is roughly constant over a wide range of RNAs. The difference shown in

Figure [Figure66](#) is exaggerated because times are plotted for score-only CYK, not complete CYK alignment, in order to include CPU times for SSU and LSU rRNA. Because score-only CYK does not keep a shadow traceback matrix nor perform the traceback, it is about 20% faster than CYK alignment, as seen in the data in Table [Table44](#).

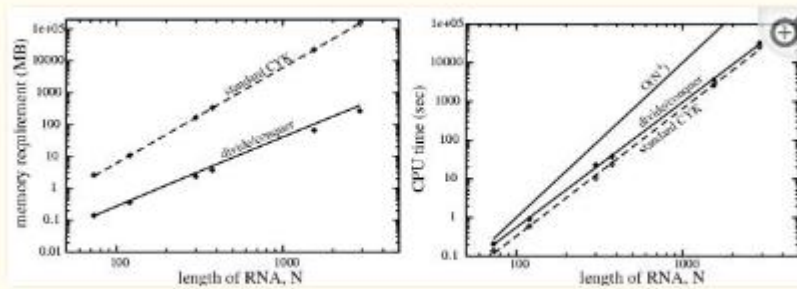


Figure 6