

Authorship Information for GEP Scientific Publications

By checking this box, I/we grant permission for the Genomics Education
Partnership (GEP) to use the annotation data produced in this report in future scientific publications.

Note: Please skip the rest of this section if **more than three students** contribute to this annotation report. When more than three students contribute to an annotation project, the class as a whole will be acknowledged in future GEP scientific publications.

Co-authors Responsibilities

In order to be a co-author on a GEP publication, you must review, critique, and approve the final gene models and manuscript, responding promptly to requests to read and approve. As part of the preparations for the microPublication article, co-authors are required to validate specific data within the manuscript, supplemental materials, and GenBank submission (the specific details will depend on each annotation project). In most cases, the manuscript preparation process will take approximately 3–5 hours of your time.

The above requirements mean that we must be able to contact you when the GEP microPublication, and later, the scientific paper with meta-analysis, is ready for your review and approval. **If we cannot reach you at that time, you will not be a co-author on our GEP scientific publications,** as scientific journals require all co-authors to have read and approved the manuscript.

Please provide your contact information below. Note that your name and contact information will be publicly available through the scientific publication and the GenBank record (this is standard for all scientific publications.). Please list the authors in ascending alphabetical order by last name. (The actual order of the student co-authors in the scientific publication will be determined by a random number generator.)

Contact information for Author #1 (The student who completes this report)

First name	<u>Steven</u>
Middle initials	C
Last name	Smith
Author name (name that will appear on the publication):	Steven Smith
Permanent Email address (one you will use five years from now):	Panzerfaust412@hotmail.com
Alternative Email address (optional):	
Check this box to indicate that you have read and accept the co-authors responsibilities	
Project Details	
Project name: <u>contig48</u>	
Project species: D. ananassae	
Date of submission:	
Size of project in base pairs: 1-60,000 Number of genes in project: 8	
Does this report cover all of the genes or is it a partia	al report? All
If this is a partial report, please indicate the region o From base to base	i the project covered by this report:

Note: For each gene described in this annotation report, you should also prepare the corresponding **GFF**, **transcript and peptide sequence files** as part of your submission.

Complete the following Gene Report Form for each gene in your project. Copy and paste the sections below to create as many copies as needed within this report. Be sure to create enough Isoform Report Forms within your Gene Report Form for all isoforms. If you cannot find evidence for any protein-coding genes in the project, jump to the "Check for additional features in your project" section.

Gene Report Form		
Gene name (e.g., D. ananassae eyeless)	: D. ananassae CG14452-PA	
Gene symbol (<i>e.g.</i> , <i>dana_ey</i>):	name of gene if not given in record finder	
	netical gene model, reported below, I have doubts as pecies. Here are my reasons:	
No RNA-seq to support gene expression. This may be a pseudogene or is expressed in different stages of the life cycle.		
Number of isoforms in <i>D. melanogaste</i> Number of isoforms in this project:	ding all of the isoforms in this project:	
Name(s) of unique isoform(s) based on coding sequence	List of isoforms with identical coding sequences	
	CG12546-PA	
Names of the isoforms with unique so	ding sequences in <i>D. melanogaster</i> that are absent in	

this species:

Provide the evidence (text and figures) which support the hypothesis that these isoforms are absent in this species (*e.g.*, changes in canonical splice sites, gene structure, etc.):

Note: For isoforms with identical coding sequence, you only need to complete the Isoform Report Form for one of these isoforms (i.e. using the name of the isoform listed in the left column of the table above). However, you should **generate GFF, transcript, and peptide sequence files for <u>ALL</u> isoforms, irrespective of whether their coding sequence is identical to that of another isoform.**

Isoform Report Form

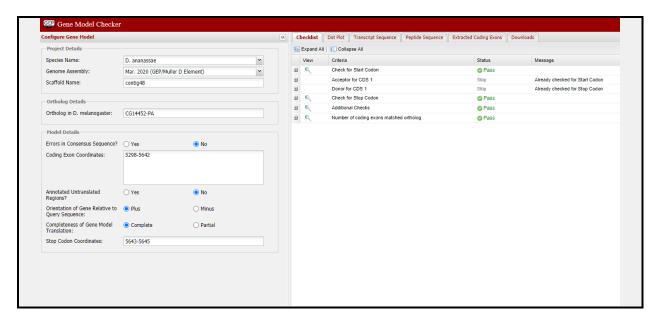
Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.

Gene-isoform symbol (<i>e.g.</i> , dana_ey-PA):	
Names of any additional isoforms with identical coding sequences:	
Is the 5' end of this isoform missing from the end of the project?	
(Define "putative exons" based on the exons present in the <i>D. melanogaster</i> ortholog)	

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the Gene Model Checker and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the <u>original project sequence</u>. Include the VCF file you have generated above when you submit the gene model to the Gene Model Checker. The Gene Model Checker will use this VCF file to automatically revise the submitted exon coordinates.

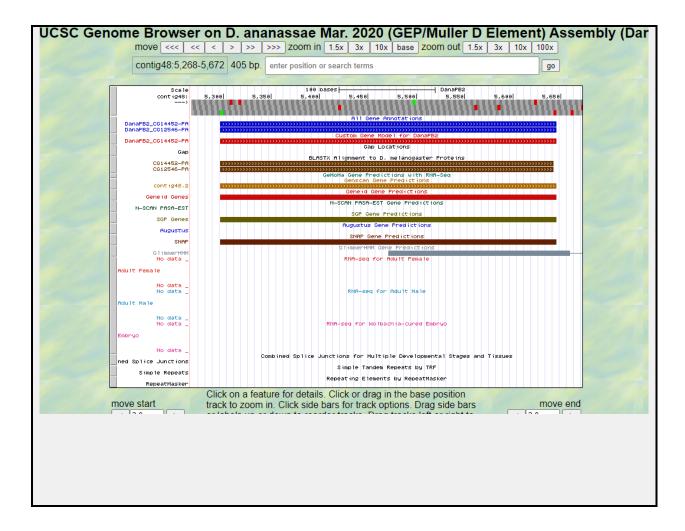


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this</u> <u>isoform is in the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

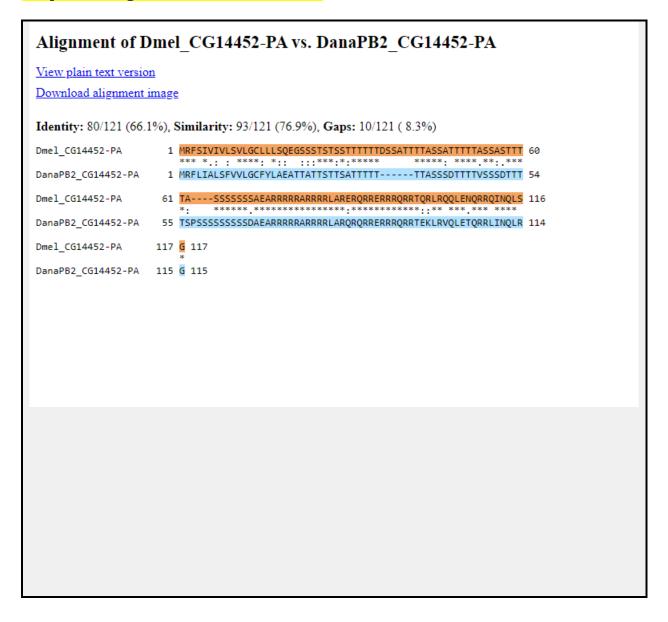
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

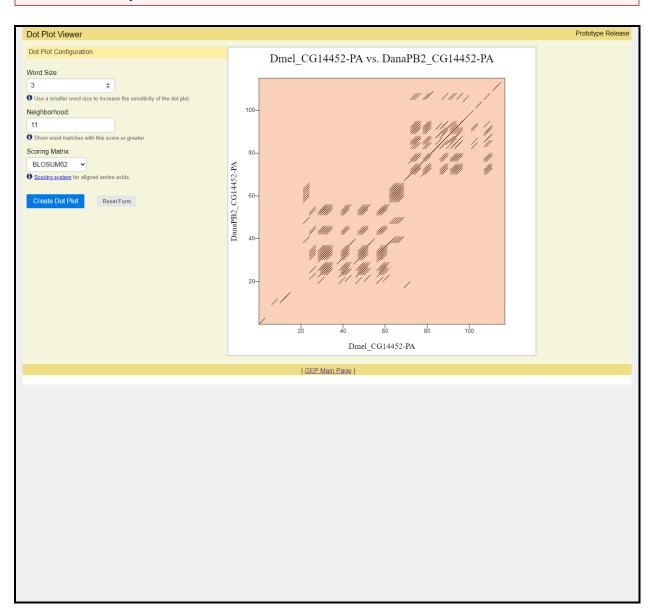
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the Gene Model Checker (available through the "**View protein alignment**" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST web site. **Paste a screenshot of the protein alignment into the box below:**



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the Gene Model Checker) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, regions with no sequence similarity, indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Gene	Re	port	: Fo	rm
------	----	------	------	----

Gene name (e.g., D. ananassae eyeless):		
Gene symbol (e.g., dana_ey):Dan	naPB2_CG32453-PB	name of gene if not given in
record finder		
Although I was able to create a hypoth	otical gono model, rono	rtod bolow. I boyo doubte as
to whether this gene is active in this sp	_	
to whether this gene is delive in this sp	recies. Here are my rea	301131
No RNA-seq in adult females to support stages of the life cycle.	rt gene expression. Like	ly expressed in different
Approximate location in project (from Number of isoforms in <i>D. melanogaste</i> . Number of isoforms in this project:		
Number of isoforms in this project:	3	
Complete the following table, include	ling all of the isoforms	s in this project:
Name(s) of unique isoform(s) based on coding sequence	List of isoforms with	identical coding sequences
	CG32453-PA	
	CG14454-PA	
	CG14454-PB	
Names of the isoforms with unique coothis species:		clanogaster that are absent in
Provide the evidence (text and figures are absent in this species (e.g., changes lsoform Report Form		
·		
Complete this report form for each uppaste this form to create as many cop	•	~ ~
Gene-isoform symbol (<i>e.g.</i> , dana_ey-PA	n):	
Names of any additional isoforms with	identical coding seque	nces:

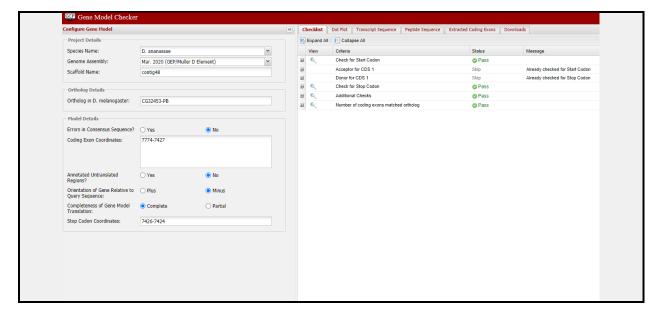
Is the 5' end of this isoform missing from the end of the project?	
If so, how many putative exons are missing from the 5' end:	
Is the 3' end of this isoform missing from the end of the project?	
If so, how many putative exons are missing from the 3' end:	

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the Gene Model Checker and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the Gene Model Checker. The Gene Model Checker will use this VCF file to automatically revise the submitted exon coordinates.



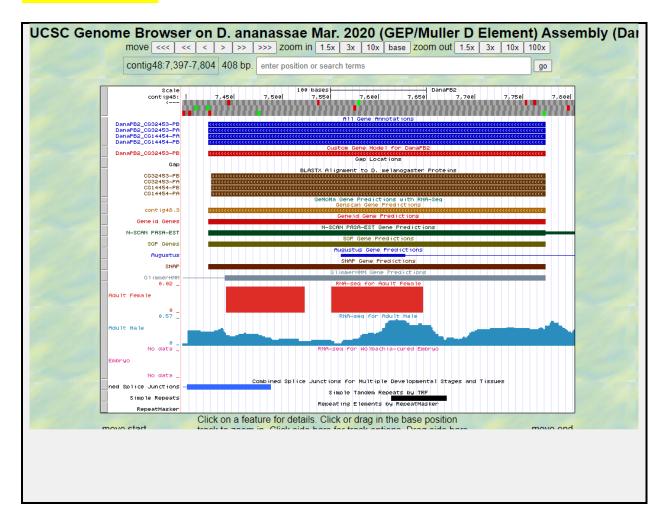
2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this</u> <u>isoform is in the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan)
- 3. At least one RNA-Seg track (e.g., RNA-Seg Coverage)

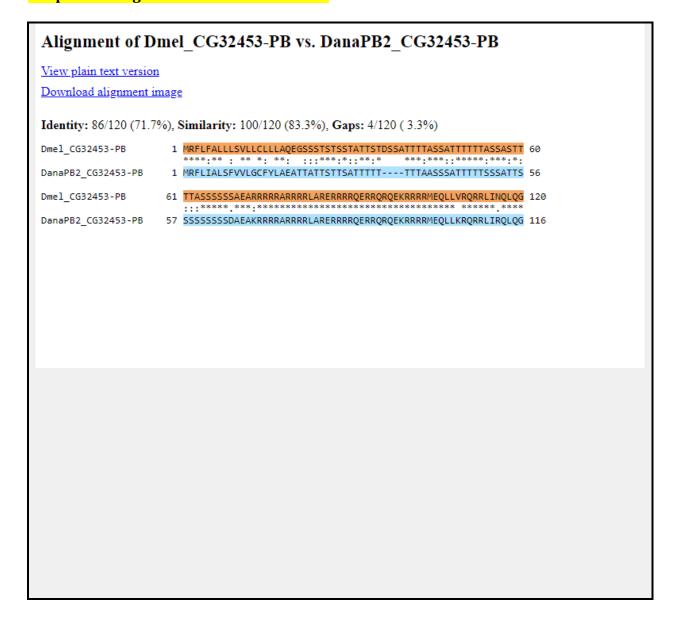
4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

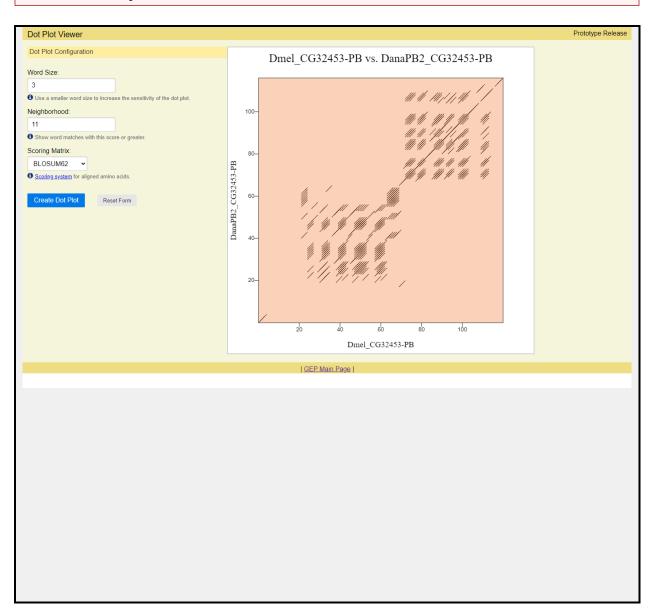
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the Gene Model Checker (available through the "**View protein alignment**" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST web site. **Paste a screenshot of the protein alignment into the box below:**



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the Gene Model Checker) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, regions with no sequence similarity, indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.

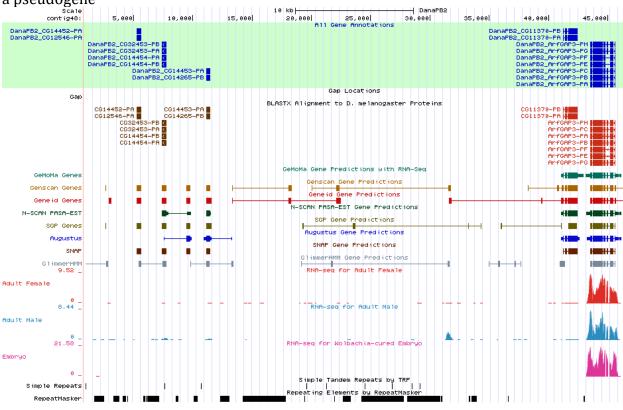


Gene Report Form

Gene name (<i>e.g., D. ananassae eyeless</i>): _	D. ananassae CG14453-PA	
Gene symbol (<i>e.g., dana_ey</i>):	name of gene if not given in record finder	
	9	

Although I was able to create a hypothetical gene model, reported below, I have doubts as to whether this gene is active in this species. Here are my reasons:

Little or no RNA-seq throughout entirety of exons to support gene expression. This may be a pseudogene



Approximate location in project (from 5' end to 3' end):	11,135-11,503	
Number of isoforms in <i>D. melanogaster:</i>		
Number of isoforms in this project:		

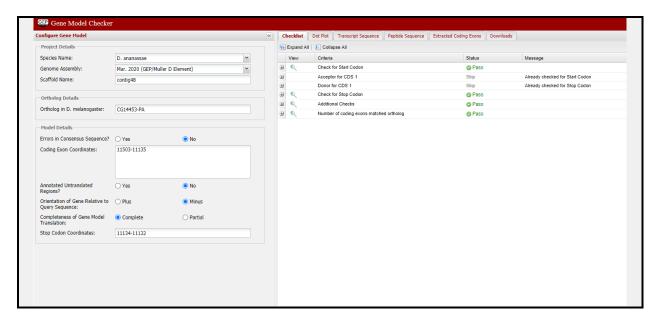
Complete the following table, including all of the isoforms in this project:

Name(s) of unique isoform(s) based on coding sequence	List of isoforms with identical coding sequences
	CG14265-PB
Names of the isoforms with unique co this species:	ding sequences in <i>D. melanogaster</i> that are absent in
, , ,	s) which support the hypothesis that these isoforms s in canonical splice sites, gene structure, etc.):
Isoform Report Form	
•	nique isoform listed in the table above. Copy and pies of this Isoform Report Form as needed.
Gene-isoform symbol (<i>e.g.</i> , dana_ey-P <i>r</i>	A):
Names of any additional isoforms with	n identical coding sequences:
Is the 5' end of this isoform missing from the end of the project? If so, how many putative exons are missing from the 5' end: Is the 3' end of this isoform missing from the end of the project? If so, how many putative exons are missing from the 3' end:	
(Define "putative exons" based on the exons present in the <i>D. melanogaster</i> ortholog)	

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the Gene Model Checker and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the <u>original project sequence</u>. Include the VCF file you have generated above when you submit the gene model to the Gene Model Checker. The Gene Model Checker will use this VCF file to automatically revise the submitted exon coordinates.

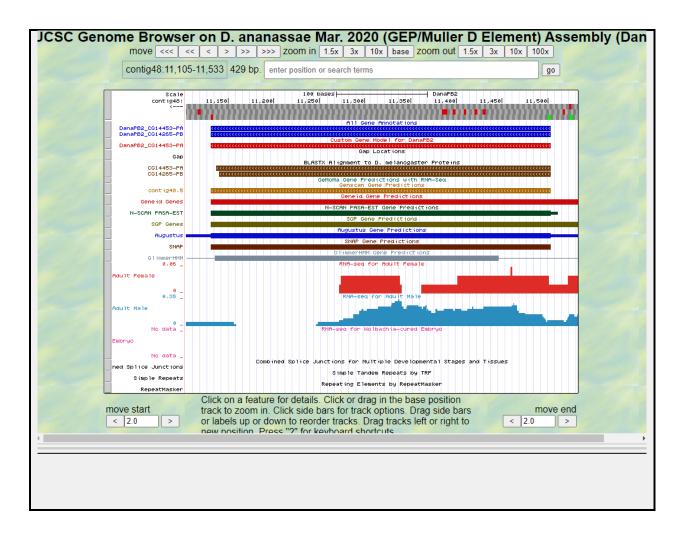


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this</u> <u>isoform is in the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

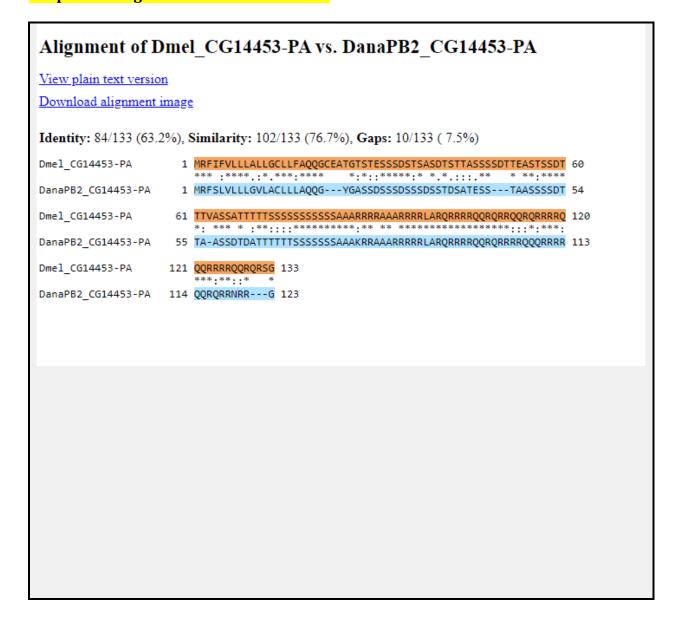
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

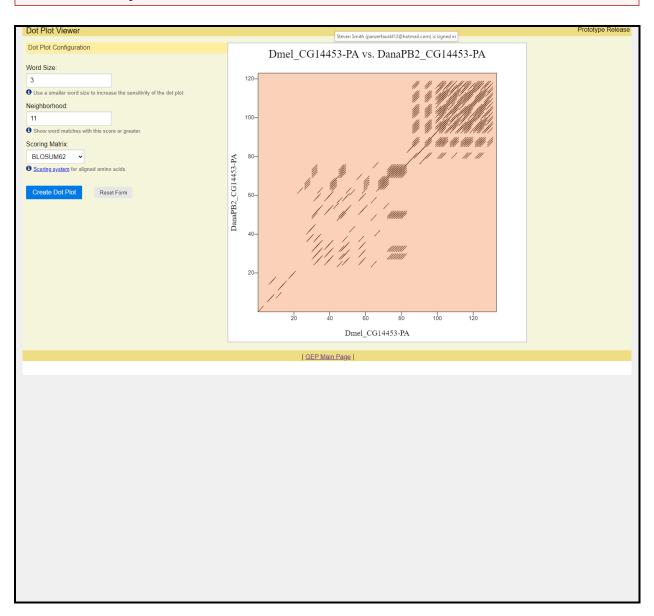
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the Gene Model Checker (available through the "**View protein alignment**" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST web site. **Paste a screenshot of the protein alignment into the box below:**



4. Dot plot between the submitted model and the *D. melanogaster* ortholog

Paste a screenshot of the dot plot (generated by the Gene Model Checker) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, regions with no sequence similarity, indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Gene Report Form		
Gene name (e.g., D. ananassae eyeless	s): D. ananassa	e CG11370-PB 1
Gene symbol (<i>e.g., dana_ey</i>):Drecord finder		name of gene if not given ir
Although I was able to create a hypoto whether this gene is active in this		
Approximate location in project (from Number of isoforms in <i>D. melanogas</i> ; Number of isoforms in this project:	ter:1	
Name(s) of unique isoform(s) based on coding sequence	_	identical coding sequences
CG11370-PA		
Names of the isoforms with unique c this species:		
Provide the evidence (text and figure are absent in this species (e.g., chang		_
Isoform Report Form		
Complete this report form for each paste this form to create as many co	•	2.5

Gene-isoform symbol (e.g., dana_ey-PA): CG11370-PA

Names of any additional isoforms with identical coding sequences:

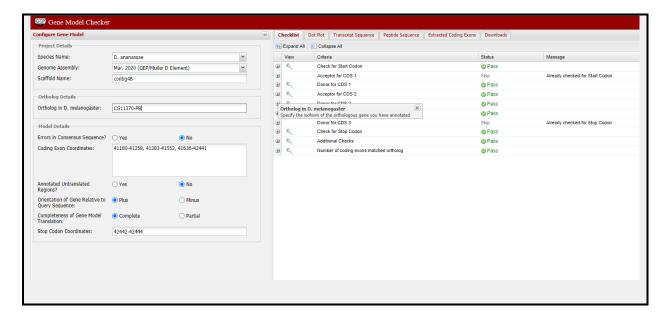
Is the 5' end of this isoform missing from the end of the project?	No		
If so, how many putative exons are missing from the 5' end:			
Is the 3' end of this isoform missing from the end of the project?		No	
If so, how many putative exons are missing from the 3' end: _			

(Define "putative exons" based on the exons present in the *D. melanogaster* ortholog)

1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the Gene Model Checker and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the <u>original project sequence</u>. Include the VCF file you have generated above when you submit the gene model to the Gene Model Checker. The Gene Model Checker will use this VCF file to automatically revise the submitted exon coordinates.



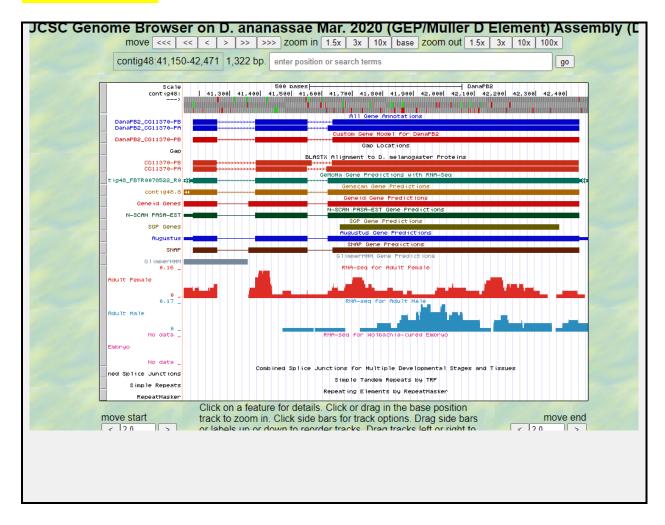
2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this</u> <u>isoform is in the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

- 5. A sequence alignment track (e.g., D. mel Proteins)
- 6. At least one gene prediction track (e.g., Genscan)

- 7. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 8. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

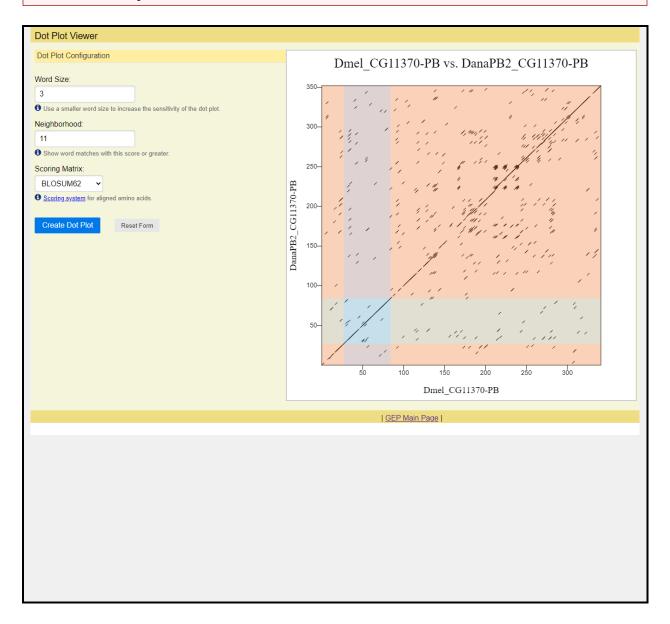
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the Gene Model Checker (available through the "**View protein alignment**" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST web site. **Paste a screenshot of the protein alignment into the box below:**

Alignment of Dmel CG11370-PB vs. DanaPB2 CG11370-PB View plain text version Download alignment image Identity: 284/354 (80.2%), Similarity: 304/354 (85.9%), Gaps: 15/354 (4.2%) Dmel_CG11370-PB 1 MKFSTALALLVIAGIAATGDALPARKRMVYYQQPAAAEWYGSAPQQRIMYMQYVQPGRTH 60 DanaPB2_CG11370-PB Dmel CG11370-PB 61 ARSTOAASALVAGETVATGTYLKESDTSAEGVPADDVLAAAGAHGATSVAEAYPDOAPVV 120 ****************************** 61 ARSTQAASALVAGETVATGTYLRESEVSAEAVQADDTLTAAGAHSATSVAEAYPDSEPVV 120 DanaPB2_CG11370-PB 121 QVATNSDVAPQAESEAEPEPEAA----DDAAKVPRDFNFAAEEASV-----GSAAEEESV 171 **: *:***** **: * **:: * **:: * *.:: * .:*:***: Dmel_CG11370-PB 121 OVSINADVAPOVETEAEPAPESNPSENDDASKVPRDLVFNDEEVAAAVPAPGPVADEESI 180 DanaPB2 CG11370-PB 172 PLPVA-EAELPAPAPIAPVAAVVPANRYLPAKKKVIVELDQ---EEEEPQAAAIEDEEEV 227 Dmel_CG11370-PB DanaPB2_CG11370-PB 181 VAPVAVESELPAP--IAPVASVVPANRYLPAKKKVIVELDQSPEEDEEPQAAAFEDEQEV 238 Dmel CG11370-PB 228 ENAVADDVEEDEELSVPVKPINPVRVPNARRPADKKPVKAASPAGKPSKKPAAPLPAGT 287 239 ENAVSDDVEEDEELSVPVKPVNPVRVPNARRPAVKKPVKAAPAGGKPAKKPAAPLPAGT 298 DanaPB2 CG11370-PB Dmel_CG11370-PB 288 FFPIDFGGTNGGAIAIANSFSTGEGGSATSHAIAYGSPESAVRRARPNPSKFRH 341 ********************************* DanaPB2 CG11370-PB 299 FFPIDFGGTNGGAIAIANSFSTGEGGSATSHAIAYGSPESASRRVRPNPSKFRH 352

4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the Gene Model Checker) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, regions with no sequence similarity, indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of donor and acceptor sites.



Gene Report Form	
Gene name (e.a., D. ananassae eveless):	D. ananassae ArfGAP3-PH
Gene symbol (e.g., dana_ey):	name of gene if not given in record finder
	etical gene model, reported below, I have doubts as pecies. Here are my reasons:
Number of isoforms in <i>D. melanogaste</i>	5' end to 3' end): <u>45,635-43,495</u> r:
Number of isoforms in this project:	7
Name(s) of unique isoform(s) based on coding sequence	ling all of the isoforms in this project: List of isoforms with identical coding sequences
ArfGAP3-PF	ArfGAP3-PC
ArfGAP3-PG	ArfGAP3-PA
midni 5 i d	ArfGAP3-PB
	ArfGAP3-PG
	ArfGAP3-PE (identical to ArfGAP3-PF)
	Thront 5 1 D (tachetear to final in 5 11)
Provide the evidence (text and figures are absent in this species (e.g., change	ding sequences in <i>D. melanogaster</i> that are absent in) which support the hypothesis that these isoforms in canonical splice sites, gene structure, etc.):
Isoform Report Form	
-	nique isoform listed in the table above. Copy and pies of this Isoform Report Form as needed.
Gene-isoform symbol (e.g., dana_ey-PA	A):ArfGAP3-PF

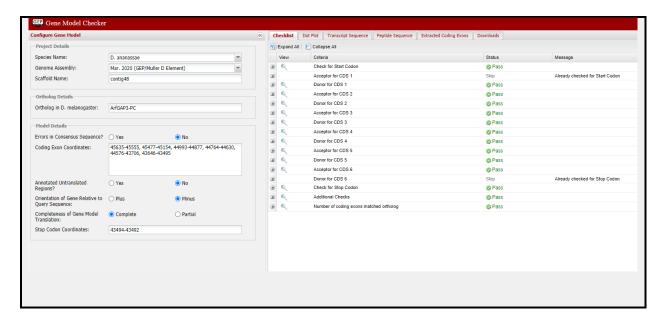
Names of any additional isoforms with identical coding sequences:

ArfGAP3-PE

Is the 5' end of this isoform missing from the end of the project? No
If so, how many putative exons are missing from the 5' end:
Is the 3' end of this isoform missing from the end of the project? No
If so, how many putative exons are missing from the 3' end:
(Define "putative exons" based on the exons present in the <i>D. melanogaster</i> ortholog)
Isoform Report Form
Complete this report form for each unique isoform listed in the table above. Copy and paste this form to create as many copies of this Isoform Report Form as needed.
Gene-isoform symbol (<i>e.g.</i> , dana_ey-PA):ArfGAP3-PG
Names of any additional isoforms with identical coding sequences:
Is the 5' end of this isoform missing from the end of the project?No
If so, how many putative exons are missing from the 5' end:
Is the 3' end of this isoform missing from the end of the project?No
If so, how many putative exons are missing from the 3' end:
if so, now many putative exons are missing from the 5' end:
(Define "putative exons" based on the exons present in the <i>D. melanogaster</i> ortholog)
1. Gene Model Checker checklist

Enter the coordinates of your final gene model for this isoform into the Gene Model Checker and paste a screenshot of the checklist results into the box below:

Note: For projects with consensus sequence errors, report the exon coordinates relative to the **original project sequence**. Include the VCF file you have generated above when you submit the gene model to the Gene Model Checker. The Gene Model Checker will use this VCF file to automatically revise the submitted exon coordinates.

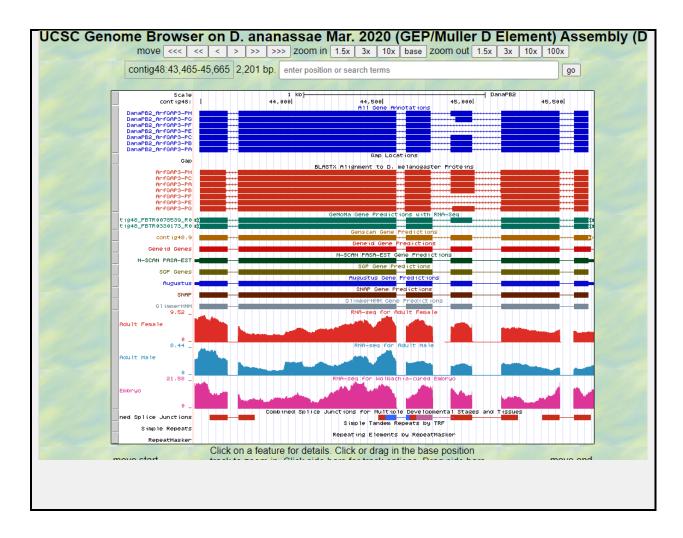


2. View the gene model on the Genome Browser

Click on the magnifying glass icon under the "Checklist" tab of the <u>Gene Model Checker</u> to view your gene model on the GEP UCSC Genome Browser. Zoom in so that <u>only this</u> <u>isoform is in the genome browser window, and capture a screenshot that includes the following evidence tracks if they are available:</u>

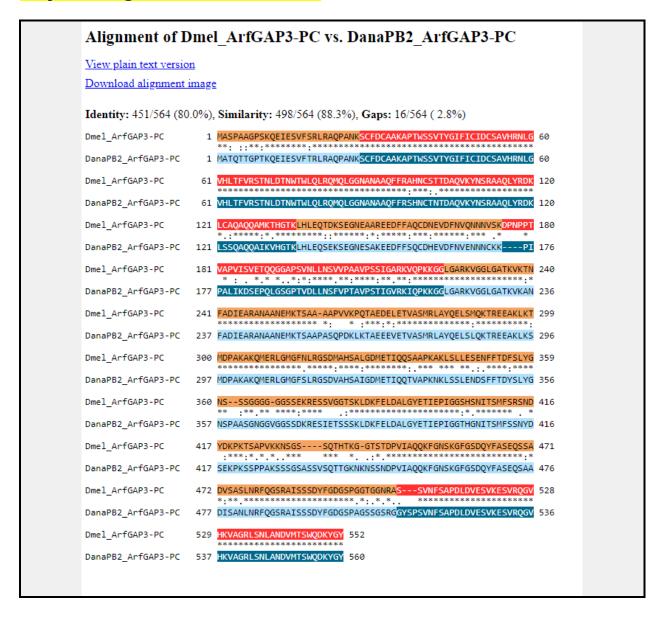
- 1. A sequence alignment track (e.g., D. mel Proteins)
- 2. At least one gene prediction track (e.g., Genscan)
- 3. At least one RNA-Seq track (e.g., RNA-Seq Coverage)
- 4. A comparative genomics track (e.g., D. mel. Net Alignment, Conservation)

Paste a screenshot of your gene model as shown on the GEP UCSC Genome Browser into the box below:



3. Alignment between the submitted model and the D. melanogaster ortholog

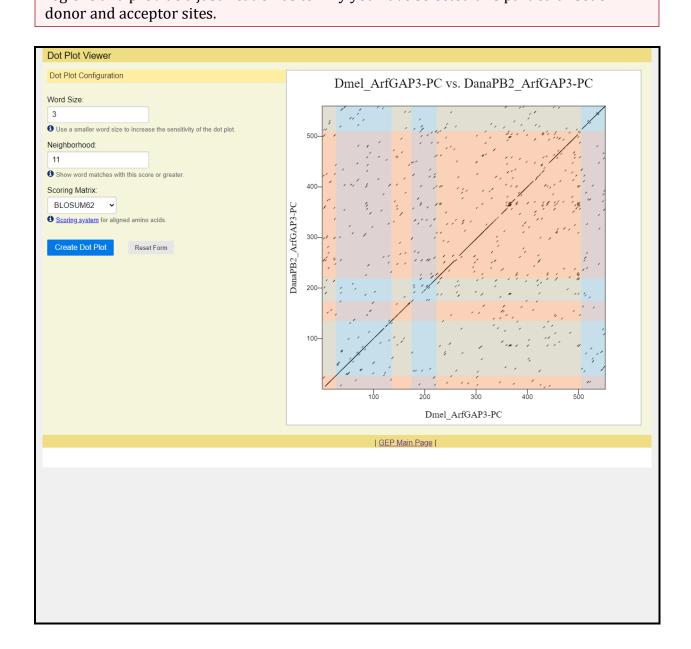
Show an alignment between the protein sequence for your gene model and the protein sequence from the putative *D. melanogaster* ortholog. You can either use the protein alignment generated by the Gene Model Checker (available through the "**View protein alignment**" link under the "Dot Plot" tab) or you can generate a new alignment using the "Align two or more sequences" feature at the NCBI BLAST web site. **Paste a screenshot of the protein alignment into the box below:**



4. Dot plot between the submitted model and the D. melanogaster ortholog

Paste a screenshot of the dot plot (generated by the Gene Model Checker) of your submitted model against the putative *D. melanogaster* ortholog into the box below. Provide an explanation for any anomalies on the dot plot (*e.g.*, large gaps, regions with no sequence similarity, indications of significant insertions or deletions).

Note: Large <u>vertical and horizontal gaps</u> near exon boundaries in the dot plot often indicate that an incorrect splice site might have been picked. Please re-examine these regions and provide a justification as to why you have selected this particular set of



Check for additional features in your project

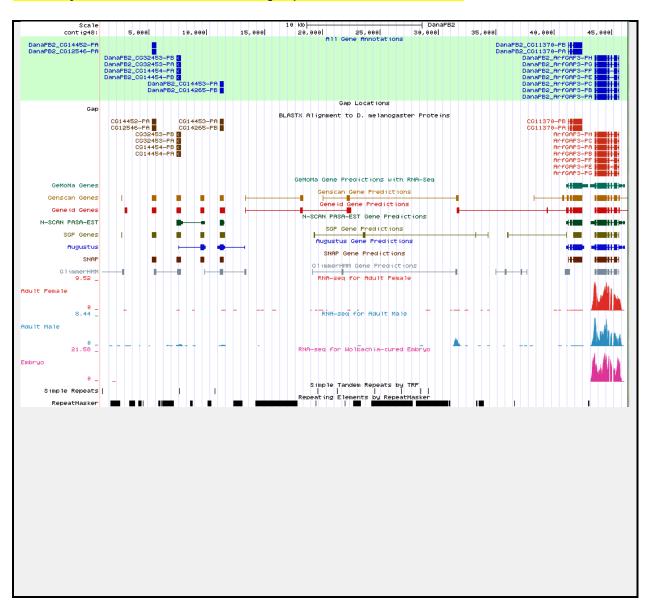
For each Genscan gene prediction that does not overlap with the genes you have already annotated, perform the following analyses to determine if the feature corresponds to a protein-coding gene, pseudogene, or partial gene duplication.

- 1. Perform a FlyBase BLASTP search of the predicted protein sequence from Genscan against the *D. melanogaster* "**Annotated proteins**" database. Report the significant matches (E-value < 1e-5) to protein sequences in *D. melanogaster*:
- 2. If there are significant matches to *D. melanogaster* proteins, analyze the genomic region immediately surrounding the Genscan prediction using the exon-by-exon strategy. Report your findings:
 - If the feature is a functional protein-coding gene, construct the gene model in the target species and provide the supporting evidence for the gene model in a new Gene Report Form
 - If the feature is a pseudogene or a partial gene duplication, provide the evidence (text and figures) which support these hypotheses:
 - Evidence for a pseudogene includes in-frame stop codons, and frame shifts within coding exons
 - Changes in gene structure from a multi-exon gene in *D. melanogaster* to a single exon gene in the target species could indicate a retrotransposed pseudogene
- 3. Perform a NCBI BLASTP search of the predicted protein sequence from Genscan against the "**Reference proteins (refseq_protein)**" database. Report the significant matches (E-value < 1e-5) to <u>curated RefSeq gene models</u>:
 - Protein records curated by the NCBI RefSeq database have the prefix "NP_"
- 4. Examine the gene expression tracks (*e.g.*, RNA-Seq data) for evidence of transcribed regions that do not correspond to the features you have already annotated or transposon remnants identified by RepeatMasker. Perform an NCBI BLASTX search of these genomic regions against the **refseq_protein** database to determine if they show significant similarity (E-value < 1e-5) to curated RefSeq gene models (i.e. protein records with the prefix "**NP_**"). Report as above:

Preparing the Project for Submission

For each project, you should prepare the project GFF, transcript, and peptide sequence files for <u>ALL</u> isoforms along with this report. You can combine the individual files generated by the Gene Model Checker into a single file using the <u>Annotation Files Merger</u>. Once you have combined the GFF files into a single file, click on the "**Show Track**" button to view all the gene models in the combined GFF file within the Genome Browser.

Paste a screenshot (generated by the Annotation Files Merger) with all the gene models you have annotated in this project into the box below.



Thank you for your submission, and congratulations on completing your analysis of this region of this genome. Our planned GEP meta-analysis of the genes and genomes in this study depends on the high quality annotations accomplished by GEP students.