

# Report: Hybrid vigor in response to Eimeria in the HMHZ

Alice

19 June 2018

## Contents

<b>General informations on HMHZ</b>	<b>1</b>
Improving Eimeria oocysts detection . . . . .	2
Missing data (to complete with Victor) . . . . .	3
PCR . . . . .	3
qPCR . . . . .	3
Comparison oocysts flotation, PCR, qPCR . . . . .	4
<b>Testing hybrid vigor along HMHZ</b>	<b>4</b>
Oocyst shedding proxy . . . . .	4
qPCR proxy . . . . .	4
BCI proxy . . . . .	4

## General informations on HMHZ

- 485 mice were captured over three years and had fecal samples processed, from 146 farms.
- From these mice, (*tbc* : *N* had colon content and intestinal tissues collected for PCR and qPCR detection)
- 3.79 mice were caught on average by farm (95% ci : 0.36)
- **Hybrid indexes** were calculated as ratio of M.m.d/M.m.m alleles (between 4 and 14, on average 13 loci)

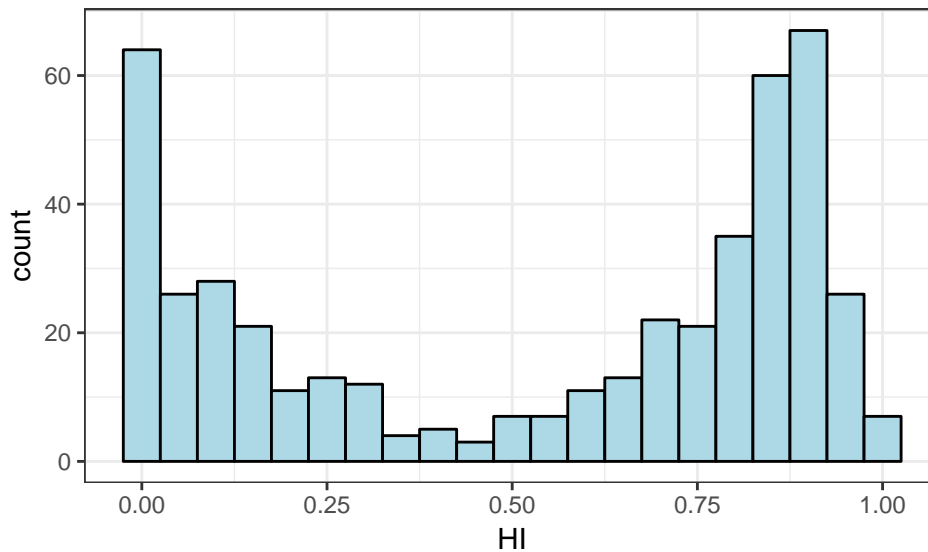


Figure 1: Number of animals caught along the hybrid index

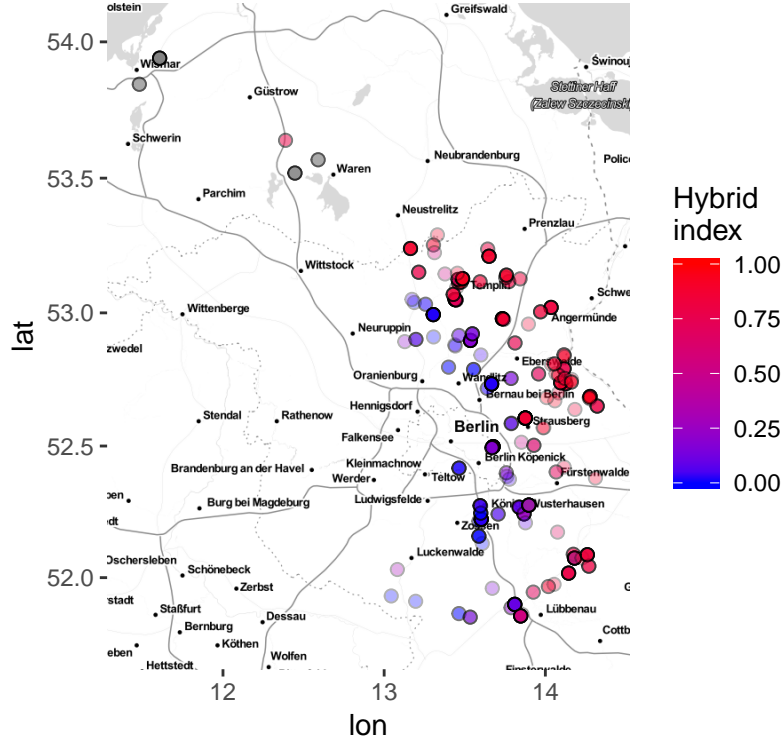


Figure 2: Samples map. Each point correspond to one location, a less pronounced transparency indicating more animals sampled at this location. Hybrid index is represented by a gradient from blue (M.m.d) to red (M.m.m)

The average *Eimeria* prevalence per farm based on oocysts flotation is 15.22. We observed, based on this technique, a variation between years (Table 1).

Table 1: Prevalence of *Eimeria* based on OPG per year

	2015	2016	2017
FALSE	92.0	126	167.00
TRUE	10.0	24	66.00
prevalence(%)	9.8	16	28.33

## Improving *Eimeria* oocysts detection

22 new samples were detected while diluting by 0.1mL PBS instead of 1mL before counting in Neubauer chamber.

Adjusted R-squared = 0.81 represents the amount of variation in y explained by x.

<https://www.r-bloggers.com/correlation-and-linear-regression/> (for Lorenzo)

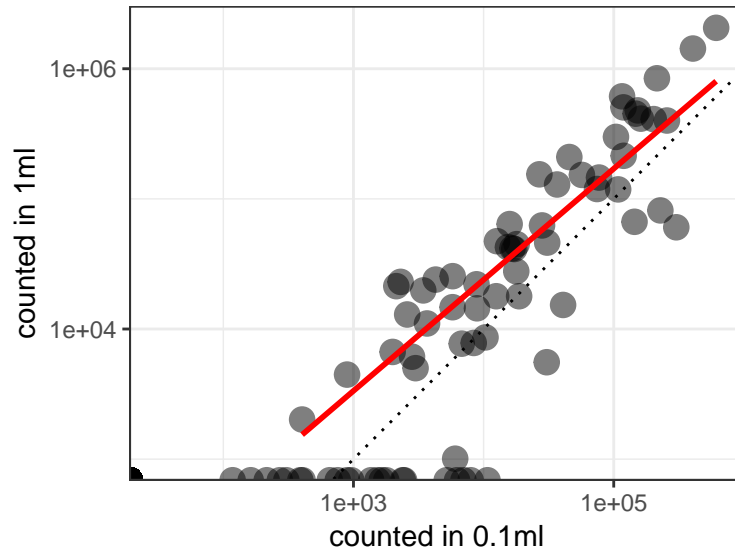


Figure 3: Comparison of OPG depending on dilution level. Red line represents linear relationship between both axis, dotted line represents

## Missing data (to complete with Victor)

Some mice do not have an hybrid index yet: SK\_3174, AA\_0411, AA\_0412, AA\_0489, AA\_0490, AA\_0491, AA\_0495, AA\_0496, AA\_0497, AA\_0498, AA\_0499, AA\_0500, AA\_0501, AA\_0502, AA\_0503, AA\_0504, AA\_0505, AA\_0506, AA\_0511, AA\_0512, AA\_0513, AA\_0514, AA\_0515

## PCR

```
getPrevalenceTable(table(myData$Ap5_PCR, myData$year))
```

```
##           2015 2016 2017
## negative      0    0 33.00
## positive      0    0  8.00
## prevalence(%) NaN  NaN 19.51
```

```
getPrevalenceTable(table(myData$PCR.positive, myData$year))
```

```
##           2015 2016 2017
## FALSE      0    0 36.0
## TRUE       0    0  5.0
## prevalence(%) NaN  NaN 12.2
```

## qPCR

```
getPrevalenceTable(table(myData$qPCRstatus, myData$year))
```

```
##           2015 2016 2017
## negative      0 125.00    0
## positive      0  19.00    0
## prevalence(%) NaN 13.19  NaN
```

## Comparison oocysts flotation, PCR, qPCR

```
#getPrevalenceTable(table(myData$qPCRstatus, myData$year))
```

## Testing hybrid vigor along HMHz

### Oocyst shedding proxy

First approximation:

```
## `geom_smooth()` using method = 'loess' and formula 'y ~ x'
```

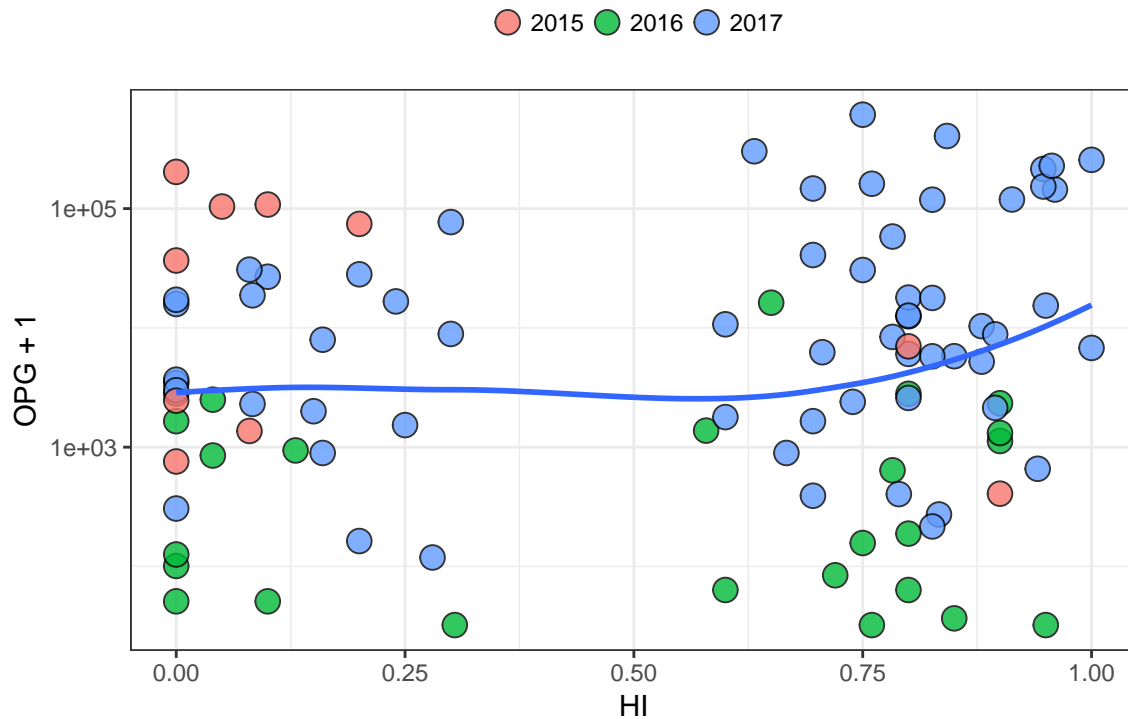


Figure 4: OPG along HI, colored per year. Blue line represent a smooth function (method = loess)

Statistical model (dvp...)

### qPCR proxy

tbc

### BCI proxy

First approximation:

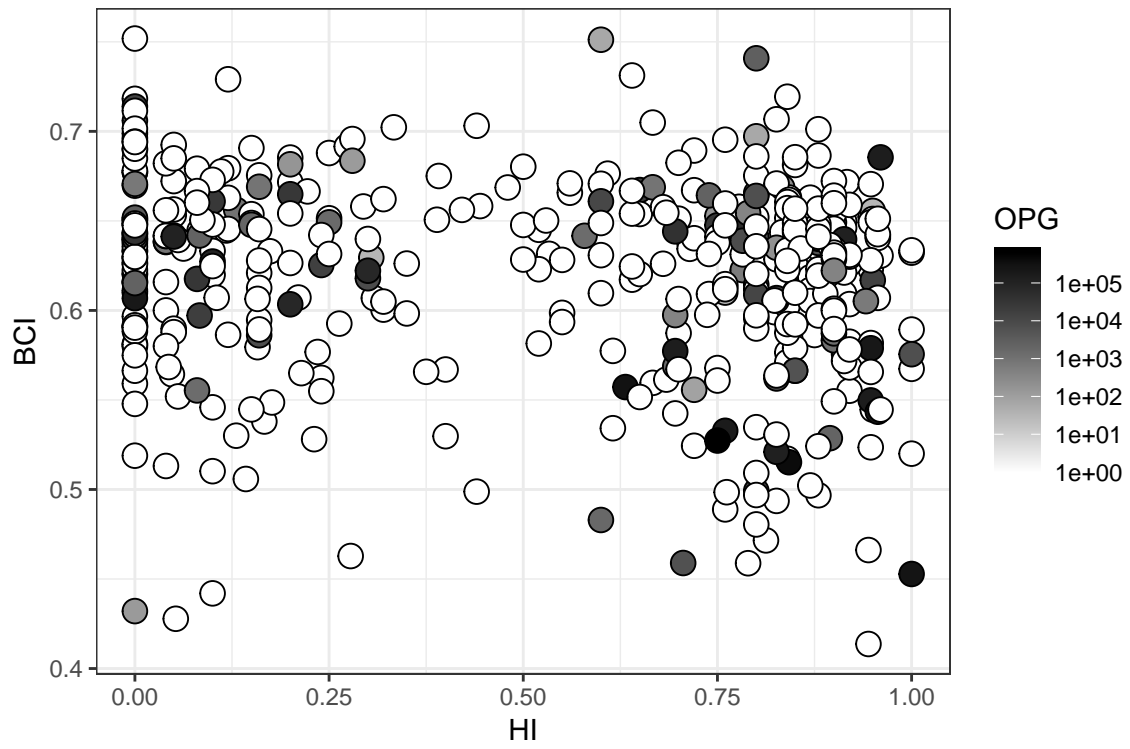


Figure 5: BCI along HI, colored per level of OPG