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(54)	NOVEL ANTIBIOTIC COMPOSITION
	COMPRISING FLUFENAMIC ACID AS AN
	ACTIVE INGREDIENT

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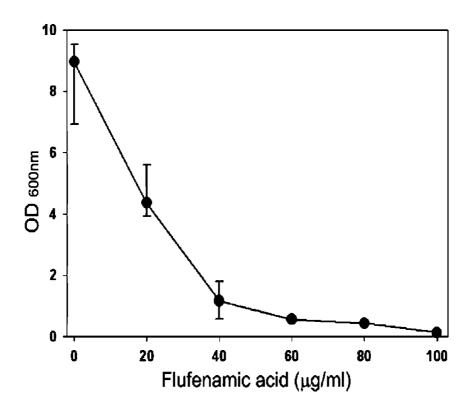
(57) **ABSTRACT**

The present invention relates to a novel antibiotic, and more specifically relates to an antibiotic composition comprising flufenamic acid as an active ingredient, which can reduce drug toxicity and side effects and the problem of antibiotic resistance caused by excessive use of antibiotics since the composition exhibits a good therapeutic effect, even in a small dose, when it is administered either alone or together with another antibiotic of the prior art.

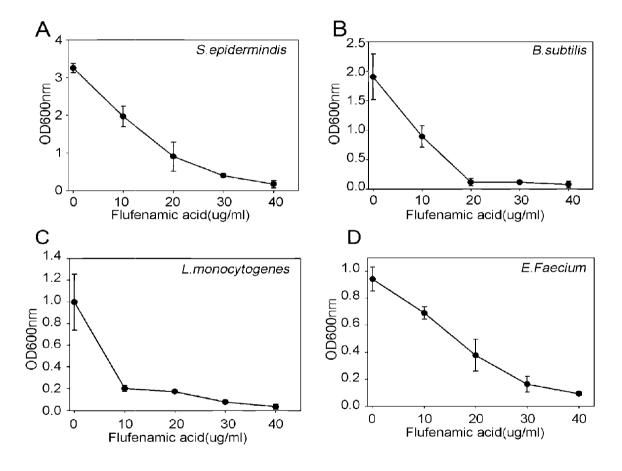
[Fig. 1]

Flufenamic acid

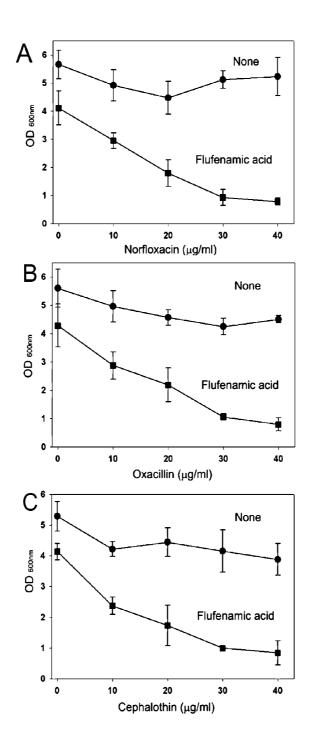
[Fig. 2]



[Fig. 3]



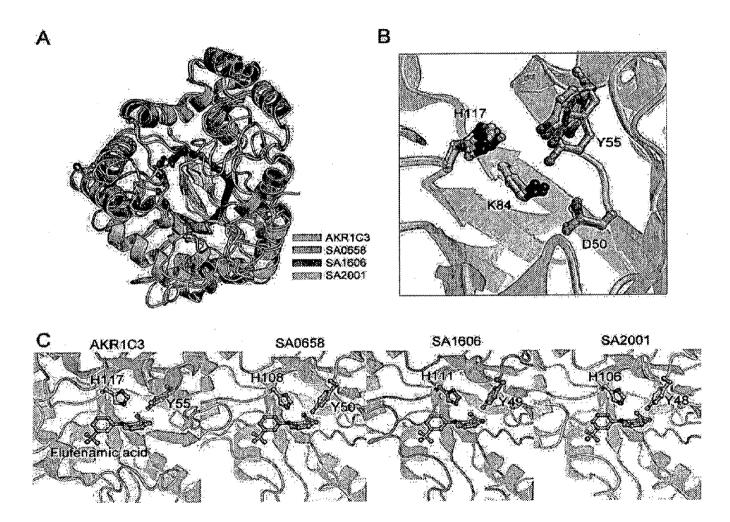
[Fig. 4]



[Fig. 5]

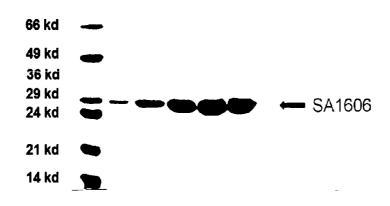
AKR1C3	MDSKQQCVKLNDGHFMPVLGFGTYAPPEVPRSKALEVTKLAIEAGFRHIDSAHLYNNEEQ
SA0658	MINEIQILNNGYPMPSVGLCVYKISDEDMTKVVNAAIDAGYRAFDTAYFYDN S
SA1606	MEVKTFYNCNTMPQIGLGTFRVENDENCMESVKYAIEQGYRSIDTAKVYGNEEQ
SA2001	MNHIEISKOVKIPVLGFGVFQIPQEQTAEAVXEAIXAGYRHIDTAQSYLNETE
	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
AKR1C3	VGLAIRSKIADGSVKREDIFYTSKLWSTFHRPELVRPALENSLKKAQLDYVDLYLIHSPM
SA0658	LGRALKDNGVDREDLFITTKLWNDYQGYEKTFEYFNKSIENLQTDYLDLFLIHWPC
SA1606	VGAGIRAGLESTGIAREDLFITSKLYFEDFGRENVAAAYEASLSRIGLKYLDLYLVHWPG
SA2001	VGQGIEASGIDRSELFITTKVWIENVNYEDTIKSIERSLQRLNLDYLDLVLIHQPY
	1*
AKR1C3	${\tt SLKPGEELSPTDENGKVIFDIVDLCTTWEAMEKCKDAGLAKSIGVSNFNRRQLEMILNKP}$
SA0658	EADGLFLETYKAMEELYEQGKVKAIGVCNFNVHHLEKLMAQS
SA1606	TNEAVMVD TWKGMEDLYKNNKAKNIGVSNFEPEHLEALLAQV
SA2001	NDVYGSWRALEELKENGKIKAIGVSNFGVDRIVDLGIHN
	:::*, * ***,** :: : :
AKR1C3	${\tt GLKYKPVCNQVECHPYFNRSKLLDFCKSKDIVLVAYSALGSQRDKRWVDPNSFVLLEDPV}$
SA0658	SIKP VNQIEVEPYFNQQELQEFCDRHDIKVTAWMPLMRNRGLLDNPV
SA1606	SIKPVINQVEYHPYLTQHKLKLYLAAQHIVMESWSPLNAQILNDET
SA2001	QIQPQVNQIEINPFHQQEEQVAALQQENVVVEAWAPFAEGKNQLFQNQL
AKR1C3	LCALAKKAKRTPALIALRYQLQRGVVVLAKSYNEQRIRQNVQVFEFQLTAEOMKAIDGLD
SA0658	${\tt IVKIAEKYBKTPAQVVLRWBLAHNRIIIPKSQTPKRIQENIDILDFNLELTEVAEIDALD}$
SA1606	IKD IAQ ELGKSP AQ VV LRWN VQ HGVVI IPKSVTPNRISENFQ IFDFELSDEQMTRIDGLN
SA2001	LQAIADKYNKSIAQVILRWLVERDIVVLAKSVNPERMAQNLDIFDFELTEEDKQQIATLE
	1 14.1 11 * 1 **1 1 1. 111.** . 1*1 1*.1111*1* . 1 . * . *
AKR1C3	RNLHYFNSDSFASHPNYPYSDEYLEHHHHHH
SA0658	RNARQGKNPDDVKIGDLK
SA1606	QDKRIGPDPKTFEG
SA2001	ESNSQFFSHADPEMIKALTSRELDV-
	• • • • • • • • • • • • • • • • • • • •

[Fig. 6]

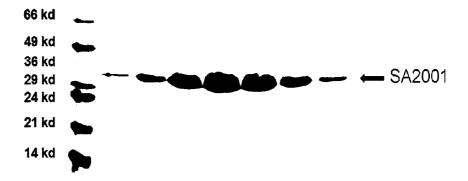


[Fig. 7]

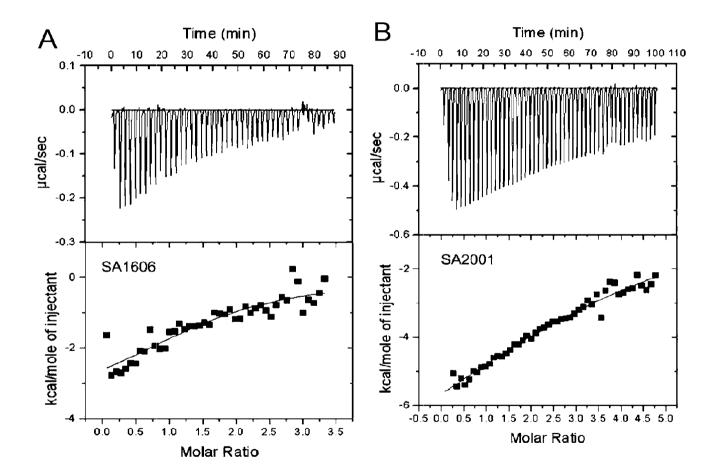




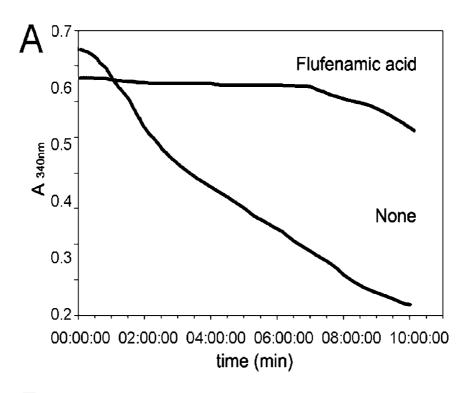
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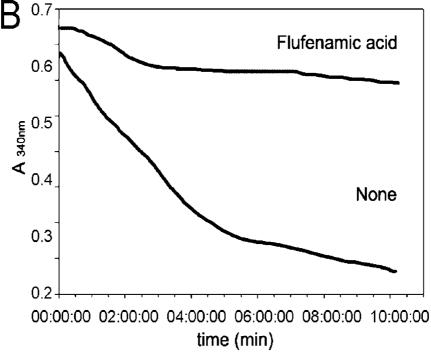


[Fig. 8]



[Fig. 9]





NOVEL ANTIBIOTIC COMPOSITION COMPRISING FLUFENAMIC ACID AS AN ACTIVE INGREDIENT

TECHNICAL FIELD

[0001] The present invention is to provide a novel antibiotic material effective for killing or inhibiting the growth or metabolism of Gram-positive bacteria.

BACKGROUND ART

[0002] Gram staining is a method of differentiating bacterial species into two large groups which are the Gram-positive and Gram-negative bacteria, which was devised by Danish physician H. C. J. Gram (1853-1938) in 1884 not for the purpose of distinguishing one type of bacterium from another but to enable bacteria to be more visible in stained sections. These bacterial groups are generally different in various aspects including sensitivity to chemical agents, nutrients necessary for their growth, response to physical and chemical stimuli, toxins produced thereby, and lesions caused thereby. Representative among Gram-positive bacteria, characterized by a purple color stain, are Staphylococcus spp., Pneumococcus spp, Mycobacterium leprae, Corynebacterium diphtheria, Clostridium tetani, Bacillus anthracis, and Actinomyces. Most of these bacteria are leading pathogens responsible for serious disease such as diphtheria, tuberculosis, pneumonia,

[0003] For example, Staphylococcus aureus, a Gram-positive bacterium, is a pathogen which causes various diseases including food poisoning, impetigo, cellulitis, scalded skin syndrome, mastitis, bacteremia, sepsis, staphylococcal pneumonia, endocarditis, heart failure, osteomylitis, Staphylococci sepsis, circulatory collapse, and toxic shock syndrome. In the United States, staphylococcal infection is diagnosed in more than two million people every year and accounts for the death of more than ninety thousand people.

[0004] Most bacterial infections are readily cured by antibiotics such as penicillin, etc., but some have recently been reported to be difficult to treat. Bacteria become resistant to antibiotic drugs through a spontaneous DNA mutation or the uptake of foreign DNA so that they are able to survive exposure to an antibiotic. This situation is called "antibiotic resistance" and the bacteria with antibiotic resistance are called "drug resistant bacteria." In addition, drug-resistant bacteria show multidrug resistance in which, for example, only 10% and 50% of them can be regulated with penicillin and methicillin, respectively.

[0005] Antibiotic resistance has been found over a broad spectrum of Gram-positive pathogenic bacteria including Staphylococcus, Streptococcus, Entracoccus, Bacillus and Listeria. Of them, MRSA (Methicillin-Resistant Staphylococcus Aureus), VRSA (Vancomycin-Resistant Staphylococcus Aureus), MRE (multiple-drug resistant enterococci) and VRE (Vancomycin-Resistant enterococci) attract social interest because they are more dangerous.

[0006] For example, MRSA is treated with vancomycin, more potent than methicillin, but some show resistance to the drug, as well. Because such resistant pathogens are poor in susceptibility to drugs, it is often necessary to apply a large dose of antibiotics which is, however, noxious to the patient. In recent years, the increasing spread of drug resistance by *S. aureus* has been recognized as serious threat to the health of mankind. Therefore, there is a pressing need for a novel

antibiotic that works through a new mechanism that confers a potent therapeutic effect even at a small dose thereof on an infection of the drug-resistant *S. aureus*.

[0007] Classically, new antibiotics have been developed by chemically modifying bioactive compounds produced by microorganisms. With the great progress of genomic and drug discovery techniques such as high-throughput screening, many new drug targets are secured from which various drug candidates have been separated. The time and cost which it takes for such new drug candidates to undergo clinical trials and to acquire the approval of the FDA is currently acting as the main hindrance to the development of novel drugs. Hence, if found, new uses of commercially available drugs which have passed clinical trials and have had their safety verified in vivo will provide the advantage of significantly reducing the time and cost required to bring novel drugs to market.

[0008] Given this background, the present inventors have searched for finding a drug that works in a manner different from already known antibiotic mechanisms in the art in order to overcome the problem with multidrug resistance and, in addition, for finding a compound that exhibits antibiotic activity from among drugs currently used in clinics in order to make it easier to develop a novel antibiotic agent.

DISCLOSURE

Technical Problem

[0009] The present invention is intended to provide a novel antibiotic destructive of or inhibitory of the growth or metabolism of Gram-positive bacteria which does not confer drug resistance on the bacteria but is effective on drug resistant bacteria, and to reduce the time and cost taken for the development of a novel antibiotic by taking advantage of a preexisting drug currently used in clinics. More particularly, the present invention provides an antibiotic composition with potent inhibitory activity against Gram-positive bacteria, comprising flufenamic acid.

Technical Solution

[0010] The present invention newly discovers the use of flufenamic acid, currently used for anti-inflammation, as a novel antibiotic in the treatment of bacterial infections. More particularly, the present invention provides an antibiotic composition comprising flufenamic acid, a derivative thereof, an isomer thereof, or a salt thereof as an active ingredient, and various uses thereof, that is, a pharmaceutical or cosmetic composition with antibiotic activity.

[0011] In addition, the present invention is directed to an aldo-keto reductase, present in Gram-positive bacteria, which can be utilized as a target protein of an antibiotic for destroying or inhibiting the growth or metabolism of Gram-positive bacteria

[0012] Leading to the present invention, intensive and thorough research was done regarding an antibiotic which has a working mechanism that is different from classical antibiotics. This research resulted in the finding that flufenamic acid, currently used as an analgesic, can inhibit the growth and metabolism of Gram-positive bacteria through a novel antibiotic mechanism.

[0013] Below, a detailed description will be given of the present invention.

[0014] The present invention addresses an antibiotic composition comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt as an active ingredient.

[0015] In the present invention, flufenamic acid is used to regulate the growth or metabolism of Gram-positive bacteria, that is, it has potent inhibitory activity against Gram-positive bacteria

[0016] No special limitations are imparted to species of the Gram-positive bacteria against which the antibiotic composition of the present invention exhibits inhibitory activity. Preferred examples include *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Bacillus subtilis*, *Listeria monocytogenes*, and *Enterococcus faecium*. More preferred is mithicillinresistant *Staphylococcus aureus* (MRSA).

[0017] Particularly, the present invention relates to an antibiotic composition with potent destructive and inhibitory activity against drug-resistant bacteria, comprising flufenamic acid as an active ingredient. Included among the drug-resistant bacteria are MRSA (Methicillin-Resistant Staphylococcus aureus), VRSA (Vancomycin-Resistant Staphylococcus Aureus), MRE (multiple-drug resistant enterococci) and VRE (Vancomycin-Resistant enterococci).

[0018] Any flufenamic acid, whether naturally occurring or artificially synthesized, may be used in the present invention. If artificially synthesized, flufenamic acid may be prepared from any material using any method. For example, flufenamic acid may be synthesized through substitution and fraction (Herbert O. House: Modern Synthetic Reactions, 2nd ED., The Benjamin/Cummings Publishing Co., 1972).

[0019] Instead of flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt may be used as an active ingredient of the composition according to the present invention.

[0020] It is naturally understood that all stereoisomers, diastereomers, enantiomers, and racemates of flufenamic acid fall within the scope of the present invention.

[0021] The salt of flufenamic acid useful in the present invention may be an acid salt or a base salt. By way of example, pharmaceutically acceptable salts of flufenamic acid include sodium, calcium and potassium salts for the hydroxy group and hydrobromide, sulfate, hydrogen sulfate, phosphate, hydrogen phosphate, dihydrogen phosphate, acetate, succinate, citrate, tartrate, lactate, mandelate, methanesulfonate (mesylate) and p-toluenesulfonate (tosylate) for the amino group, but are not limited thereto. They may be prepared using methods or techniques well known in the art. [0022] flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt act as an active ingredient in the antibiotic composition provided by the present invention, targeting the aldo-keto reductase of Grampositive bacteria.

[0023] As stated above, flufenamic acid inhibits the activity of aldo-keto reductase in Gram-positive bacteria. For example, the aldo-keto reductase which flufenamic acid target may be the enzymes present in *Staphylococcus aureus*, and may preferably be SA0658, SA1606 or SA2001.

[0024] In one embodiment of the present invention, the antibiotic composition may further comprise a conventional antibiotic against Gram-positive bacteria in addition to flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt thereof. So long as it is approved for use in the treatment of the infection of Gram-positive

bacteria, any conventional antibiotic may be employed in the composition of the present invention. Preferred is norfloxacin, oxacillin or cephalothin.

[0025] Also, the present invention addresses a Gram-positive bacterial aldo-keto reductase as a target protein of an antibiotic that is destructive or inhibitory of the growth or metabolism of Gram-positive bacteria.

[0026] The Gram-positive bacteria affected by the present invention are not limited specifically to species so long as they contain aldo-keto reductase therein. Preferred examples include Staphylococcus aureus, Staphylococcus epidermidis, Bacillus subtilis, Listeria monocytogenes and Enterococcus faecium. More preferred is methicillin-resistant S. aureus (MRSA).

[0027] In the present invention, the aldo-keto reductase functions as a target protein on which an inhibitory agent acting against the growth or metabolism of Gram-positive bacteria directly acts.

[0028] No particular limitations are imposed on the kinds of the inhibitory agent, whether natural or synthetic. The aldo-keto reductase which is regulated by the inhibitory agent may be an enzyme which is found in Gram-positive bacteria. [0029] For example, the aldo-keto reductase may be found in *Staphylococcus aureus* and preferably SA0658 with the amino acid sequence of SEQ ID NO: 1, SA1606 with the amino acid sequence of SEQ ID NO: 2 or SA2001 with the

[0030] Further, the present invention addressed an antibiotically active, pharmaceutical composition comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt as an active ingredient.

amino acid sequence of SEQ ID NO: 3.

[0031] Showing inhibitory activity against Gram-positive bacteria without side effects and toxicity in the body, the composition is useful for preventing and treating diseases caused by Gram-positive bacteria. No particular limitations are imparted on the type of inhibition exhibited against Grampositive bacteria. Preferably, it destroys or inhibits the growth or metabolism of Gram-positive bacteria.

[0032] The pharmaceutical composition of the present invention may be administered orally or parenterally to mammals such as rats, dogs, domestic animals, and humans. For example, it may be administered orally or injected intrarectally, intramuscularly or subcutaneously, or via the route of the dura mater within the uterine or via a cerebrovascular route. Preferable are parenteral routes, with greater preference for transdermal administration. Thus, the composition may be applied topically.

[0033] For practical use in the destruction or inhibition of growth or metabolism of Gram-positive bacteria, the composition in accordance with the present invention may be formulated into typical forms, for example, oral dosage forms such as tablets, capsules, troches, liquids, suspensions, etc., injection forms such as injectable solutions or suspensions, powder that can be reconstituted with distilled water for injection, and ready-to-use solutions, topical forms such as ointments, creams, lotions, liquids, sprays, etc.

[0034] So long as it is pharmaceutically accepted, any vehicle may be used for formulating the composition of the present invention. By way of example, the composition may be formulated with a binder, a suspension agent, a disintegrant, an excipient, a solubilizer, a dispersant, a stabilizer, a colorant, and/or a flavorant for oral dosage forms, or with a preservative, an appeaser, a solubilizer and/or a stabilizer for injection forms, or with a base, an excipient, a lubricant, a

preservative and/or a propellant for topical forms. The pharmaceutical formulations thus prepared may be administered orally or applied topically.

[0035] The effective dosage of the pharmaceutical composition in accordance with the present invention depends on various factors, including dosage form, the patient's age, weight, gender, state of health, diet, the time of administration, the route of administration, excretion rate, sensitivity, etc. In general, it may be administered in a single dose or in multiple doses per day at a daily dose ranging from 0.01 to 200 mg/day and preferably at a daily dose ranging from 5 to 50 mg/kg.

[0036] The antibiotically active, pharmaceutical composition comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt in according to the present invention may further comprise an antibiotic inhibitory of Gram-positive bacteria. Any antibiotic, if approved for use in inhibiting Gram-positive bacteria, may be used in the composition of the present invention. Preferred examples of the antibiotic include norfloxacin, oxacillin and cephalothin, but are not limited thereto.

[0037] In addition, the present invention addresses an antibiotically active, cosmetic composition comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt.

[0038] The cosmetic composition may further comprise an additive in addition to flufenamic acid as an active ingredient. The additive, if conventionally used, may be contained in the cosmetic composition. For example, a supplement and an excipient such as an antioxidant, a stabilizer, a solubilizer, vitamin, a colorant, a flavorant, etc. may be used. In the cosmetic composition, a transdermal delivery enhancer may be employed.

[0039] The composition of the present invention may contain flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt in an amount of from 0.0001 to 99 wt %, and preferably in an amount of from 0.001 to 90 wt %.

[0040] The cosmetic composition of the present invention may be prepared into any conventional formulation. For example, the cosmetic composition of the present invention may be formulated with a vehicle into cream, lotion, tonic, spray, aerosol, oil, solution, suspension, gel, ointment, emulsion, paste, or hair tonic. The pharmaceutical or cosmetic formulations may be prepared using any apparatus or method well known in the art (refer to Remington's Pharmaceutical Science, latest version).

[0041] In the composition of the present invention, a base may be properly combined with, for example, a cosmetically acceptable additive. So long as it is accepted in the art, any base may be used in the composition. Exemplary among them are distilled water, mineral water, ethanol, glycerin, squalene, 1,3-propyleneglycol, 1,3-butyleneglycol, castor oil, tsubaki oil, and liquid petrolatum. Examples of the additive include a surfactant, an emulsifier, a thickener, a preservative, an antioxidant, and a flavorant, but are not limited thereto. The surfactant may be cationic, anionic or non-ionic. Examples of the cationic surfactant include alkaline salts of higher fatty acids such as lauric acid, myristic acid, palmitic acid, stearic acid and oleic acid (e.g., sodium, potassium, ammonium, triethanolamine salt); esters of alkyl sulfonic acid such as sodium laurylsulfate, and triethanol amine laurylsulfonate; and alkyl ether sulfonate esters such as sodium polyoxyethylene laurylether sulfonate and triethanolamine polyoxyethylene laurylether sulfonate. Included among the non-ionic surfactants are polyoxyethylene alkyl ethers such as polyoxyethylene laurylether and polyoxyethylene oleylether; and alkanolamine such as coconut fatty acid diethanolamide and diethanolamide laurate. Stearyl trimethyl ammonium chloride, cetyl trimethyl ammonium chloride and stearyl bis(diethylalcohol)hydroxy ethyl ammonium chloride are examples of the anionic surfactants. Examples of the emulsifier include cetanol, stearyl alcohol and behenyl alcohol. Examples of the thickener include sodium alginate, methyl cellulose, carboxymethyl cellulose, polyvinyl alcohol and polyvinyl pyrrolidone.

[0042] Examples of the preservative include ethyl parahydroxybenzoate, butyl parahydroxybenzoate and benzalkonium chloride. Examples of the antioxidant include butyl hydroxytoluene, propyl galate and butyl hydroxyanisole. The flavorant may be perfume for general use and may be exemplified by citrus, lavender and floral.

[0043] Various factors including the patient's sex, age and health state, the severity of disease, etc., must be considered in determining the quantity of a dose and the number of doses of the composition according to the present invention.

[0044] The antibiotically active, cosmetic composition comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt in according to the present invention may further comprise an antibiotic inhibitory of Gram-positive bacteria. Any antibiotic, if approved for use in inhibiting Gram-positive bacteria, may be used in the composition of the present invention. Preferred examples of the antibiotic include norfloxacin, oxacillin and cephalothin, but are not limited thereto.

Advantageous Effects

[0045] Because the inhibition mechanism used is different from those of conventional antibiotics, flufenamic acid can exert inhibitory activity effectively on bacteria resistant to conventional antibiotics. This novel antibiotic may be used alone or in combination with a conventional antibiotic. In the latter case, high therapeutic effects can be obtained at a low dose, with the concomitant reduction of side effects and toxicity. Because its intrinsic medical efficacy is newly discovered, the antibiotic of the present invention, which has been used as an anti-inflammatory drug, does not need to undergo separate clinical trials for the FDA's approval, thus remarkably reducing the time and cost of drug development and having a great advantage in market entry.

DESCRIPTION OF DRAWINGS

[0046] FIG. 1 shows a chemical structure of flufenamic

[0047] FIG. 2 shows inhibitory concentrations of flufe-namic acid against *Staphylococcus aureus*.

[0048] FIG. 3 shows inhibitory concentrations of flufenamic acid against *Staphylococcus epidermidis*, *Bacillus* subtilis, *Listeria monocytogenes* and *Enterococcus faecium*.

[0049] FIG. 4 shows inhibitory concentration of flufenamic acid used in combination with Norfloxacin, oxacillin, and cephalothin against *Staphylococcus aureus*.

[0050] FIG. 5 shows comparison of sequence alignments of AKR1C3, SA0658, SA1606, SA2001, and 2,5-diketo-D-gluconate reductase A.

[0051] FIG. 6 shows three-dimensional structures and essential residues of AKR1C3, SA0658, SA1606, and SA2001.

[0052] FIG. 7 shows purification of recombinant proteins (SA1606 and SA2001) for examining the function of SaAKRs

[0053] FIG. 8 shows binding affinity of FLF for SA1606 and SA2001.

[0054] FIG. 9 shows enzymatic activities of AKR on methylglyoxal in the presence and absence of FLF, confirming the inhibitory activity of FLF against SaAKRs.

MODE FOR INVENTION

[0055] The present invention is further defined in the following Examples. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions. Thus, various modifications of the invention in addition to those shown and described herein will be apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims.

EXAMPLES

Example 1

Bacterial Strain

[0056] CCARM3080, a clinically identified species of methicillin-resistant *S. aureus* (MRSA), was purchased from the Culture Collection of antibiotic Resistant Microbes (www.ccarm.or.kr) and used for screening compounds. *Listeria monocytogenes* (KCTC3586), *Staphylococcus epidermidis* (KCTC1917) and *Bacillus subtilis* (KCTC2217) were obtained from the Korean Collection for Type Cultures (http://kctc.kribb.re.kr). *Enterococcus faecium* was granted from Dr. Ko, K. S. Sungkyunkwan University.

Example 2

Identification of Novel Compounds

[0057] To screen compounds having inhibitory activity against bacteria, MRSA was 200-fold diluted in 0.2 mL of LB in 96-well microplates and cultured overnight at 37° C. with agitation at 180 RPM. Prestwick chemical library was added at a final concentration of 100 $\mu g/mL$ per well. The microplates were incubated at 37° C. for 24 hours with agitation at 180 RPM and the optical density at 600 nm was measured. [0058] Of the candidate compounds, flufenamic acid (FLF), shown in FIG. 1, was identified to be the most inhibitory of the bacteria.

Example 3

Assay for Inhibition against Methicillin-Resistant Staphylococcus aureus

[0059] FLF was assayed for inhibitory activity against Methicillin-resistant *staphylococcus aureus* (MRSA). In this regard, Methicillin-resistant *Staphylococcus aureus* (MRSA)

was cultured overnight and 200-fold diluted in 2 mL of LB containing various concentrations of FLF. After additional incubation at 37° C. for 8 hours, cell densities were measured by recoding OD at 600 nm.

[0060] As can be seen in FIG. 2 and Table 1, the growth of MRSA was 90% inhibited after incubation with 40 µg/mL FLF for 8 hours (FIG. 2) and the MIC of FLF against MRSA was found to be 64 µg/mL when measured after 24 hours of incubation (Table 1).

Example 4

Assay for Inhibition against Staphylococcus epidermidis, Bacillussubtilis, Listeria monocytogenes, and Enterococcus faecium

[0061] FLF was examined for inhibitory activity against various bacteria. Staphylococcus epidermidis, Bacillus subtilis, Listeria monocytogenes and Enterococcus faecium were cultured overnight and 200-fold diluted in 2 mL of LB containing various concentrations of FLF. Cell densities were determined by measuring the OD at 600 nm after incubation at 37° C. for 8 hours. The MICs of FLF were determined using a broth dilution method. In brief, bacteria were inoculated into LB containing serially diluted concentrations of FLF in 96-well microplates. After 24 hours of incubation, bacterial growth was examined. Antibacterial activity of FLF was tested over various Gram-positive bacteria including Staphylococcus epidermidis, Bacillus subtilis, Listeria monocytogenes, and Enterococcus faecium.

TABLE 1

Bacteria	MICs (μg/ml)
S. aureus MSSA	64
S. aureus MRSA	64
S. epidermidis	64
B. subtilis	32
L. monocytogenes	32
E. faecium	32

[0062] As shown in FIG. 3 and Table 1, Staphylococcus epidermidis, Bacillus subtilis, Listeria monocytogenes, and Enterococcus faecium were susceptible to FLF. Their growth was 90% inhibited after incubation with 20~40 μ g/mL FLF for 8 hours (FIG. 3). The MICs of FLF against these bacteria were found to range from 32 to 64 μ g/mL when measured after 24 hours of incubation (Table 1).

Example 5

Synergistic Effect of FLF with Other Antibiotics

[0063] To examine the synergistic effect of FLF with other antibiotics, Methicillin-resistant <code>Staphylococcus aureus</code> (MRSA) was cultured overnight and 200-fold diluted in 2 mL of LB containing 20 µg/ml FLF together with 0, 10, 20, 30, or 40 µg/ml Norfloxacin, Oxacillin or Cephalocin. Cell densities were determined by measuring OD at 600 nm after incubation at 37° C. for 8 hours.

TABLE 2

MICs of Antibiotics against MRSA in the Presence of Flufenamic Acid

		I	MICs (μg/ml)	
Antibiotics (mg/ml)	No FLF	10 μg/ml FLF	20 μg/ml FLF	30μg/ml FLF	40 μg/ml FLF
Norfloxacin	128	64	32	16	8
Oxacillin	>256	256	128	32	8
Cephalothin	128	128	64	16	4

[0064] As can be seen in Table 2 and FIG. 4, the MIC of Norfloxacin, a representative quinolone antibiotic targeting DNA gyrase, was reduced from 128 to 32 μ g/ml in the presence of 20 μ g/ml FLF (FIG. 4A). Similarly, FLF allows MRSA to be far more susceptible to oxacillin (FIG. 4B) and cephalothin (FIG. 4C), both β -lactam antibiotics targeting the biosynthesis of the cell wall. When used in combination with classical antibiotics, FLF synergistically inhibited MRSA infection and thus showed effective therapeutic effects (Table 2).

Example 6

Analysis of Bacterial Target Protein of Flufenamic Acid Using Bioinformation

[0065] In order to identify the bacterial protein target of FLF, human AKR1C3, known as a protein to be associated with FLF, was compared with a library or database of sequences using the BLAST bioinformatics program.

[0066] As shown in FIG. 5, a Blast search identified ORFs—SA0658, SA1606, and SA2001 as being comparable with the sequence of human AKR1C3. The AKR analogs were found to share sequence homology of 34, 36 and 31%, respectively, with AKR1C3. In addition, these three proteins showed sequence homology of 41~45% with 2,5-diketo-Dgluconate reductase A (DkgA), one member of the AKR family. To examine whether FLF is associated with the ORFs (SA0658, SA1606, and SA2001) identified in S. aureus MRSA, DkgA of E. coli the three-dimensional structure of which had been established was used as a template for remodeling the three-dimensional structures of the proteins of interest. Primary modeling was performed with DS MODELLER, followed by optimization using the protocol of Discovery Studio (Accelrys, Inc). The models thus built were qualitatively checked using PROCHECK. Sequence alignments among AKR1C3, SA0658, SA1606, SA2001, and DkgA were established using the CLUSTALW program with a default parameter.

[0067] As can be seen in FIG. 6, analysis of the crystal structure of human AKR1C3 complexed with FLF exhibited that FLF binds to the active site of AKR1C3. The inhibitory activity of FLF is due to the occupation of the oxyanion hole of AKR1C3 having a carboxylic group by FLF through hydrogen bonds with His117 and Tyr55 (FIG. 6). Each of the three AKR analogs SA0658, SA1606 and SA2001, identified in *S. aureus*, has four essential residues Asp50, Tyr55, Lys84, and His117, which are highly conserved in the AKR family and play an important role in enzymatic activity.

[0068] Analysis results showed that SaAKRs were significantly similar to human ARK1C3 in entire structure (FIG. 6A) as well as active site structure (FIG. 6B) and that the docking of FLF into the active site was successfully achieved (FIG. 6C). In this docking model, the oxygen atoms of the

carboxylic acid group form hydrogen bonds with Tyr49 and His111, allowing FLF to occupy the oxyanion hole of the active site. Consequently, the three AKR analogs SA0658, SA1606 and SA2001, identified in *S. aureus*, were observed to have the same structure and working mechanism as those of ARK.

Example 7

Identification of the Binding of FLF to Target Proteins SA1606 and SA2001 by Gene Cloning and Protein Purification

[0069] To examine the practical binding affinity of FLF for *S. aureus* protein targets (SA0658, SA1606, SA2001) and inhibitory activity against *S. aureus*, the targets were cloned. In the case of SA0658, although it was cloned and expressed, experiments could not be completed because the expressed protein precipitated.

[0070] Genes of SA1606 (GenBank number: BAB42874) and SA2001 (GenBank number: BAB43292) were amplified using PCR with the genomic DNA of S. aureus N315 serving as a template. PCR primers were 5'-GCGCGCGCCATG-GAGGTT AAAACATTTTAT-3' and 5'-CGGCGCCTC-GAGTCCTTCAAAAGTTTTTGG-3' for SA1606, and 5'-GCGCGCGCCATGGATCATATTGAAATAAGT-3' and 5'-CGGCGCCTCGAGTTCTCTGCTTGTAAGTGC-3' for SA2001. NcoI and XhoI restriction sites were incorporated into the forward primer sequences and the reverse primer sequences, respectively. The PCR products were digested with NcoI and XhoI and inserted into NcoI/XhoI-digested pET28a vectors (Novagen). The DNAs thus ligated were transformed into E. coli BL21 DE3 (Invitrogen). Coding sequences of the cloned genes SA1606 and SA2001 were confirmed by nucleotide sequencing. Recombinant SA1606 and SA2001 with histidine tag at the C-terminus were expressed in E. coli and then loaded into Hi-Trap chelating columns (GE Healthcare) equilibrated with buffer A (20 mM HEPES, pH 7.5, 50 mM imadazole and 500 mM NaCl). The bound proteins were eluted with a linear gradient of 0.01-0.5 M imidazole in buffer A. Active fractions were concentrated and gel filtrated using a Superdex 200 column (GE Healthcare) in a buffer containing 20 mM HEPES (pH 7.5) and 2 mM DTT. The purified proteins are as shown in FIG. 7.

Example 8

Evaluation of Binding Affinity of FLF for Target Proteins by Isothermal Titration calorimetry

[0071] Binding affinity for SA1606 and SA2001 of FLF was measured using VP-ITC Micro calorimeter (MicroCal Inc). All of the proteins and FLF were prepared in 10 mM HEPES, pH 7.0 and 3% DMSO, followed by removing gas from the samples. In each titration experiment, 8 μl of FLF was injected into 1.5 mL of sample cells containing the proteins 20~30 times at regular intervals of 4 min. Thereafter, the affinity of FLF for the proteins were calculated on the basis of the heat generated upon the binding of FLF to the proteins. The data was analyzed with ORIGIN software (MicroCal Inc).

[0072] As is apparent form the data of FIG. 8, dissociation constants (Kd) between FLF and SA1606 and between FLF and SA2001 were 14.5 μ M and 38.1 μ M, respectively, indi-

cating that FLF can practically bind to SA1606 and SA2001 and that the protein targets of FLF in *S. aureus* MRSA are SA1606 and SA2001.

Example 9

Enzymatic Assay for Inhibitory Activity of FLF Against Protein Targets

[0073] Aldo-keto reductase (AKR) activity was measured to examine whether FLF inhibits the enzymatic activities of SA1606 and SA2001 on the basis of the data of Example 8 showing the binding of FLF to SA16006 and SA2001. On the whole, bacterial AKR catalyzes the reduction of various chemicals with NADPH. Hence, the enzymatic activity of AKR can be determined by monitoring the absorbance of NADPH at 340 nm. As is evidenced in Example 6, SA1606 and SA2001 have structural and functional similarity with the human aldo-keto reductase AKR1C3 and the bacterial aldoketo reductase DkgA. Thus, the enzymatic activity of DkgA was measured to analyze the inhibitory activity of FLF. The reaction for evaluating enzymatic activity was comprised of 0.5 mM NADPH, 5 mM methylglyoxal and 10 μM aldo-keto reductase in 50 mM sodium phosphate buffer, pH 7.0. Absorbance at 340 nm of the reaction was measured for 10 min to calculate the consumption of NADPH.

[0074] As can be seen in FIG. 9, SA1606 and SA2001 reduced methylglyoxal with NADPH (yellow lines), but the enzymatic activity was inhibited in the presence of FLF (brown lines), demonstrating that FLF is bound directly to the active sites of the enzymes SA1606 and SA2001, thus inhibiting the enzymatic activity of the *S. aureus* aldo-keto reductase.

FORMULATION EXAMPLES

[0075] A better understanding of the present invention may be obtained through the following formulation examples of the antibiotically active, pharmaceutical or cosmetic composition comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt which are set forth to illustrate, but are not to be construed as the limit of the present invention.

EXAMPLES OF COSMETIC FORMULATION

Formulation Example 1

Skin Lotion (Content: wt %)

 [0076]
 The composition: 0.001

 [0077]
 Glycerin: 3.0

 [0078]
 Butylene glycol: 2.0

 [0079]
 Propylene glycol: 2.0

 [0080]
 Carboxyvinyl polymer: 0.1

 [0081]
 Ethanol: 10.0

 [0082]
 Triethanolamine: 0.1

[0083] Preservative, pigment, flavorant, distilled water: q.s

[0084] Total: 100.0

Formulation Example 2

Nutrient Cream (Content: wt %)

[0085] The composition: 0.001 [0086] Beeswax: 10.0 [0087] Polysorbate 60: 1.5 [0088] Sorbitan sesquiolate: 0.5[0089] Liquid paraffin: 10.0[0090] Squalane: 5.0

[0091] Caprilic/capric triglyceride: 5.0

[0092] Triethanolamine 0.2

[0093] Preservative, pigment, flavorant, distilled water: q.s.

[0094] Total: 100.0

Formulation Example 3

Hair Soap (Content: wt %)

[0095] The composition: 0.001 [0096] Titanium dioxide: 0.2 [0097] Polyethylene glycole: 0.8 [0098] Glycerin: 0.5

[0099] Ethylene diaminetetraacetic acid: 0.05

[0100] Sodium: 1.0 [0101] Pigment: q.s. [0102] Soap flavor: q.s.

 $\begin{tabular}{ll} [0103] & Cosmetic soap base (water content 13, wt parts): q.s. \\ \end{tabular}$

[0104] Total: 100.0

EXAMPLES OF PHARMACEUTICAL FORMULATION

Formulation Example 1

Ointment

[0105] The composition: 3 g [0106] Dexpanthenol: 1.5 g [0107] Stearic acid: 1.0 g [0108] Liquid paraffin: 5.0 g [0109] Spermaceti: 4.0 g [0110] Cetanol: 3.0 g

[0111] Propylene glycol: 13.0 g [0112] Triethanolamine: 1.5 g

[0113] Dibutylhydroxytoluene: 0.025 g [0114] Ethyl benzoate: 0.0225 g

[0115] Propyl benzoate: 0.015 g

[0116] Polysorbate: 0.1 g [0117] Purified water: 65.0 g

[0118] An ointment was prepared from these ingredients according to a conventional method.

[0119] The ointment was applied at a dose of 0.5~1 g twice a day constantly for 3~6 months to the lesion.

Formulation Example 2

Lotion

[0120] The composition: 1.0 g [0121] Dexpanthenol: 1.5 g [0122] Glycerin: 0.6 g

[0123] Hydroxypropylcellulose: 0.085 g

[0124] Beegum: 0.0255 g [0125] Purified water: q.s.

[0126] A lotion was prepared from these ingredient according to a conventional method.

[0127] The lotion was applied at a dose of 0.1~1 mL twice a day constantly for 3~6 months to the lesion.

Formulation Example 3

Capsule

[0128] The composition: 10 g [0129] Crystalline cellulose: 3 g [0130] Lactose: 14.8 g

[0131] Magnesium stearate: 0.2 g

[0132] These ingredients were mixed and loaded into gelatin capsules according to a conventional method to form capsules.

Formulation Example 4

Injection

[0133] The composition: 10 g [0134] Mannitol: 180 g

[0135] Sterile water for injection: 2974 g

[0136] Na₂HPO₄.12H₂O: 26 g [0137] These ingredients were dissolved to form a total volume of 2 liters according to a conventional method.

Formulation Example 5

Liquid

[0138] The composition: 20 mg [0139]Isomerose: 10 g

[0140]Mannitol: 5 g [0141]Purified water: q.s.

[0142] Using a conventional method, the ingredients were added to distilled water to which lemon flavor was then added in a suitable amount. After the ingredients were mixed, distilled water was added to bring the total volume to 100 mL, which was then loaded into a brown bottle and sterilized.

SEQUENCE LISTING

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Asp	Glu	Tyr	Leu	Glu 325	His	His	His	His	His 330	His					

- 1. An antibiotically active composition, comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt as an active ingredient.
- 2. The antibiotically active composition of claim 1, functioning to destroy or inhibit growth or metabolism of Grampositive bacteria.
- 3. The antibiotically active composition of claim 2, wherein the Gram-positive bacteria is selected from the group consisting of *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Bacillus subtilis*, *Listeria monocytogenes*, and *Enterococcus faecium*.
- **4**. The antibiotically active composition of claim **2**, wherein the active ingredient targets Gram-positive bacterial aldo-keto reductase.
- **5**. The antibiotically active composition of claim **4**, wherein the reductase is selected from the group consisting of SA0658, SA1606, and SA2001, which are *Staphylococcal* aldo-keto reductases.
- The antibiotically active composition of claim 1, functioning to destroy or inhibit growth or metabolism of antibiotic-resistant bacteria.
- 7. The antibiotically active composition of claim 6, wherein the antibiotic-resistant bacteria is selected from the group consisting of MRSA (Methicillin-Resistant *Staphylococcus aureus*), VRSA (Vancomycin-Resistant *Staphylococcus aureus*), MRE (multiple-drug resistant enterococci), and VRE (Vancomycin-Resistant enterococci).
- **8.** An aldo-keto reductase, present in Gram-positive bacteria, as a target protein of an antibiotic functioning to destroy or inhibit growth or metabolism of Gram-positive bacteria.
- 9. The aldo-keto reductase of claim 8, wherein the Grampositive bacteria is *Staphylococcus aureus*.
- 10. The aldo-keto reductase of claim 9, having an amino acid sequence of SEQ ID NO: 1 (SA0658), serving as a target

- of an antibiotic destructive or inhibitory of growth or metabolism of *Staphylococcus aureus*.
- 11. The aldo-keto reductase of claim 9, having an amino acid sequence of SEQ ID NO: 2 (SA1606), serving as a target of an antibiotic destructive or inhibitory of growth or metabolism of *Staphylococcus aureus*.
- 12. The aldo-keto reductase of claim 9, having an amino acid sequence of SEQ ID NO: 3 (SA2001), serving as a target of an antibiotic destructive or inhibitory of growth or metabolism of *Staphylococcus aureus*.
- 13. An antibiotically active, pharmaceutical composition, comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt as an active ingredient.
- **14**. The antibiotically active, pharmaceutical composition of claim **13**, further comprising an antibiotic.
- 15. The antibiotically active, pharmaceutical composition of claim 14, wherein the antibiotic is selected from the group consisting of norfloxacin, oxacillin, and cephalothin.
- 16. An antibiotically active, cosmetic composition, comprising flufenamic acid, flufenamic acid derivative, flufenamic acid isomer, or flufenamic acid salt as an active ingredient.
- 17. The antibiotically active, cosmetic composition of claim 16, being in a form selected from the group consisting of hand wash, face wash, cleansing agent, shampoo, rinse, soap, lotion, essence, cream, wax, nutrient pack, a dyeing agent, a waving agent, and spray.
- **18**. The antibiotically active, cosmetic composition of claim **16**, further comprising an antibiotic.
- 19. The antibiotically active, cosmetic composition of claim 18, wherein the antibiotic is selected from the group consisting of norfloxacin, oxacillin, and cephalothin.

* * * * *