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(54) **BIODEGRADABLE POLY(PHOSPHOESTER) COMPOSITIONS FOR TREATING OVARIAN CANCER**

BIODEGRADIERBARE POLYPHOSPHOESTER-ZUSAMMENSETZUNGEN ZUR BEHANDLUNG VON OVARIALKREBS

POLY(PHOSPHOESTERS) BIODEGRADABLES POUR LE TRAITEMENT DU CANCER DES OVAIRES

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Description

BACKGROUND OF THE INVENTION

1. Field of the Invention

[0001] The present invention provides methods for treating ovarian cancer, in particular those pertaining to the extended release of an antineoplastic agent from biodegradable poly(phosphoester) compositions.

2. Description of the Related Art

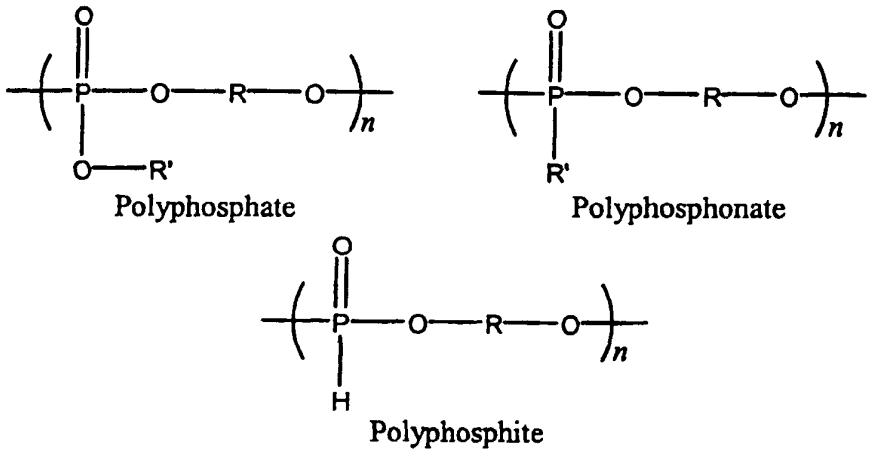
[0002] Antineoplastic agents, such as paclitaxel, have sometimes been used to treat ovarian cancer. For example, those in the art have attempted to administer paclitaxel in normal saline by infusion into the peritoneal cavity of women having ovarian cancer as a prolonged series of weekly treatments. Francis et al., "Phase I Feasibility and Pharmacologic Study of Weekly Intraperitoneal Paclitaxel: A Gynecologic Oncology Group Pilot Study", J. of Clinical Oncology. 13: 12, 2961-67 (1995). However, problems with multiple toxicities, such as abdominal pain, nausea, vomiting, leukopenia, and fatigue, are often encountered with the high fluid volumes and drug dosages required for efficacy with this approach. Further, the repeated dosing and attendant discomfort is often inconvenient and, sometimes, even unacceptable for patients.

[0003] Thus, there exists a need for a method of effecting the in vivo, controlled release of a variety of different antineoplastic agents into the peritoneum, whether they are small hydrophobic drugs, such as paclitaxel, or large and bulky bio-macromolecules, such as therapeutically useful proteins. Preferably, effective release of the antineoplastic agent should occur without requiring the presence of significant amounts of a physiologically acceptable fluid vehicle, such as normal saline or an organic solvent. There is also a continuing need for biodegradable polymer compositions that may provide extended release in such a way that trauma to the surrounding soft tissues can be minimized.

[0004] Biocompatible polymeric materials have been used in various therapeutic drug delivery and medical implant applications. If a medical implant is intended for use as a drug delivery or other controlled-release system, using a biodegradable polymeric carrier is one effective means to deliver the therapeutic agent locally and in a controlled fashion, see Langer et al., "Chemical and Physical Structures of Polymers as Carriers for Controlled Release of Bioactive Agents", J. Macro Science Rev. Macro. Chem. Phys., C23(1), 61-126 (1983). In this way, less total drug is required, and toxic side effects can be minimized.

[0005] Polymers have been used for some time as carriers of therapeutic agents to effect a localized and sustained release. See Leong et al., "Polymeric Controlled Drug Delivery", Advanced Drug Delivery Rev., 1:199-233 (1987); Langer, "New Methods of Drug Delivery", Science, 249:1527-33 (1990) and Chien et al., Novel Drug Delivery Systems (1982). Such delivery systems offer the potential of enhanced therapeutic efficacy and reduced overall toxicity. Examples of classes of synthetic polymers that have been studied as possible solid biodegradable materials include polyesters (Pitt et al., "Biodegradable Drug Delivery Systems Based on Aliphatic Polyesters: Applications to Contraceptives and Narcotic Antagonists", Controlled Release of Bioactive Materials, 19-44 (Richard Baker ed., 1980); poly(amino acids) and pseudo-poly(amino acids) (Pulapura et al. "Trends in the Development of Bioresorbable Polymers for Medical Applications", J. Biomaterials Appl., 6:1, 216-50 (1992); polyurethanes (Bruin et al., "Biodegradable Lysine Diisocyanate-based Poly(Glycolide-co-ε Caprolactone)-Urethane Network in Artificial Skin", Biomaterials, 11:4, 291-95 (1990); polyorthoesters (Heller et al., "Release of Norethindrone from Poly(Ortho Esters)", Polymer Engineering Sci 21:11, 727-31 (1981); and polyanhydrides (Leong et al., "Polyanhydrides for Controlled Release of Bioactive Agents", Biomaterials 7:5, 364-71 (1986).

[0006] Polymers having phosphate linkages, called poly(phosphates), poly(phosphonates) and poly(phosphites), are known. See Penczek et al., Handbook of Polymer Synthesis, Chapter 17: "Phosphorus-Containing Polymers", (Hans R. Kricheldorf ed., 1992). The respective structures of these three classes of compounds, each having a different side chain connected to the phosphorus atom, are as follows:



[0007] The versatility of these polymers comes from the versatility of the phosphorus atom, which is known for a multiplicity of reactions. Its bonding can involve the 3p orbitals or various 3s-3p hybrids; spd hybrids are also possible because of the accessible d orbitals. Thus, the physico-chemical properties of the poly(phosphoesters) can be readily changed by varying either the R or R' group. The biodegradability of the polymer is due primarily to the physiologically labile phosphoester bond in the backbone of the polymer. By manipulating the backbone or the side chain, a wide range of biodegradation rates are attainable.

[0008] An additional feature of poly(phosphoesters) is the availability of functional side groups. Because phosphorus can be pentavalent, drug molecules or other biologically active substances can be chemically linked to the polymer. For example, drugs with -O-carboxy groups maybe coupled to the phosphorus via a phosphoester bond, which is hydrolyzable. See. Leong, U.S. Patent Nos. 5,194,581 and 5,256,765. The P-O-C group in the backbone also lowers the glass transition temperatme of the polymer and, importantly, confers solubility in common organic solvents, which is desirable fin easy characterization and processing.

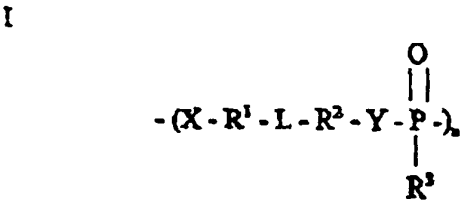
[0009] WO 98/44021 discloses biodegradable terephthalate polyester-poly(phosphate) compositions; WO 98/44020 discloses biodegradable compositions containing polymers chain-extended by phosphoesters; and International Application No. PCT/US98/09185 discloses biodegradable compositions comprising poly(cycloaliphatic phosphoester) compounds. However none of these disclosures suggests the specific use of biodegradable poly(phosphoester) compositions for treating ovarian cancer specifically.

[0010] Thus, there remains a need for new methods and materials for the difficult problem of successfully treating ovarian cancer with a minimum of discomfort, toxicities and prolonged, periodic re-dosing.

SUMMARY OF THE INVENTION

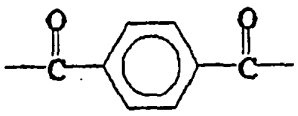
[0011] It has now been discovered that biodegradable polymer compositions comprising:

(a) at least one antineoplastic agent and
(b) a biodegradable polymer comprising the recurring monomeric units shown in formula I:



wherein X is -O- or -NR^4-, where R^4 is H or alkyl;
Y is -O-, -S- or -NR^4-;
each of R^1 and R^2 is a divalent organic moiety;
L is a divalent, branched or straight chain aliphatic group having 1-20 carbon atom, a cycloaliphatic group,

or a group having the formula:



;

R^3 is selected from the group consisting of H, alkyl, alkoxy, aryl, aryloxy, heterocyclic or heterocycloxy; and
n is about 5-5,000;

are suitable for intraperitoneal administration to treat a mammalian subject having ovarian cancer. These polymer compositions provide extended release of the antineoplastic agent within the peritoneum of the subject. Moreover, the polymer composition of the invention increases the median survival rate from the cancer by at least about 10%, as compared with the median survival rate obtained by administration of a composition comprising the same dosage of the antineoplastic agent without the biodegradable polymer of the invention.

[0012] Also disclosed is a solid article suitable for insertion into the peritoneum to treat a mammalian subject having ovarian cancer, the article comprising a biodegradable polymer composition comprising:

(a) at least one antineoplastic agent and
(b) a biodegradable polymer comprising the recurring monomeric units shown above in formula I.

[0013] A method is provided for treating a mammalian subject having ovarian cancer by the extended release of an antineoplastic agent, the method comprising the steps of:

(a) combining the antineoplastic agent with a biodegradable polymer having the recurring monomeric units shown above in formula I to form a composition; and
(b) inserting the composition in vivo into the peritoneum of the subject, such that the inserted composition is in at least partial contact with an ovarian cancer tumor,

wherein the median survival rate from the cancer is increased by at least about 10%, as compared with the median survival.rate obtained by administration of a composition comprising the same dosage of the antineoplastic agent without the biodegradable polymer.

[0014] The compositions can be used to deliver a wide variety of antineoplastic agents, for example, from hydrophobic drugs, such as paclitaxel, to large water-soluble macromolecules, such as proteins, over an extended period of time without necessitating significant volumes of a delivery fluid. The uses of the invention can thus be used to significantly increase the time period over which an effective dose of the antineoplastic agent is released and increases the survival time of subjects treated by the method to an unexpected degree. Further, the serious disease of ovarian cancer can be therapeutically managed with a minimum of side effects and without the unpleasantness and discomfort of a periodic series of parenteral treatments introducing significant amounts of fluid into the peritoneum.

BRIEF DESCRIPTION OF THE DRAWINGS

[0015]

Figure 1A shows the ^{1}H-NMR spectrum, and Figure 1B shows the ^{31}P-NMR spectrum for P(BHET-EOP/TC, 80/20).
Figure 2 shows the FT-IR spectrum for P(BHET-EOP/TC, 80/20).
Figure 3A shows the molecular weights and elemental analyses for P(BHET-EOP/TC, 80/20) and P(BHET-HOP/TC, 90/10), and Figure 3B shows the GPC chromatogram for P(BHET-EOP/TC, 80/20).
Figure 4A shows the DSC curve of P(BHET-EOP/TC, 80/20), and Figure 4B shows the DSC curve of P(BHET-EOP/TC, 50/50).
Figure 5A and 5B show the in vitro degradation data for P(BHET-EOP/TC, 80/20) and P(BHET-EOP/TC, 85/15).
Figure 6 shows the change in molecular weight of P(BHDPT-EOP) and P(BHDPT-EOP/TC) poly(phosphoesters) during in vitro degradation.
Figures 7A and 7B show the in vivo degradation of P(BHET-EOP/TC) in terms of weight or mass loss; Figure 7C shows the controlled delivery of hydrophobic small molecules, such as paclitaxel, from a p(BHET-EOP/TC, 80/20)

film.

Figure 8 shows an electron microscopic photograph of P(BHET-EOP/TC, 80/20) microspheres containing FITC-BSA.

Figure 9A shows the effect of loading level on the release kinetics of FITC-BSA from microspheres, and 9B shows the controlled delivery of hydrophobic small molecules, such as paclitaxel from a CHDM polymer.

Figure 10 shows the release of lidocaine from copolymer P(BHDPT-EOP/TC) microspheres.

Figure 11 shows the cytotoxicity of P(BHET-EOP/TC, 80/20) microspheres.

Figure 12 shows a toxicity assay plot of relative cell growth (%) vs. concentration of degraded polymer in a tissue-culture wall (mg/ml) for four separate polymers.

Figure 13 shows the cytotoxicity of P(BHET-EOP/TC, 80/20) microspheres.

Figure 14 shows the ^{1}H-NMR spectrum of a polymer of the invention, P(LAEG-EOP).

Figure 15 shows the ^{31}P-HMR spectrum of a polymer of the invention, P(LAEG-EOP).

Figure 16A and 16B show differential scanning calorimetry data for two polymers of the invention.

Figure 17 shows the results of a GPC analysis of a polymer of the invention in graphic form.

Figure 18 shows the change in Mw of two polymers of the invention after being exposed to air at room temperature for one month.

Figure 19 shows shelf stability data for a polymer of the invention at room temperature.

Figure 20A and 20B show the weight loss (20A) and the change in Mw (20B) for discs fabricated from two polymers of the invention over a period of eight days in PBS at 37°C.

Figure 21A and 21B show the weight loss (21A) and the change in Mw (21B) for discs fabricated from two polymers of the invention, in vitro.

Figure 22 shows biocompatibility data for polymers of the invention.

Figure 23 shows cytotoxicity data for microspheres of a polymer of the invention, P(LAEG-EOP).

Figure 24A shows the effect of fabrication method upon the release rate of microspheres of a polymer of the invention, and 24B shows the rate of release of lidocaine from microspheres of a polymer of the invention.

Figure 25(A) through 25(E) all show degradation and release data of p(DAPG-EOP) polymers *in vitro*.

Figure 26 shows the stracone of P(*trans*-CHDM-HOP) as determined by ^{31}P-NMR and ^{1}H-NMR.

Figure 27 shows the chromatogram and molecular weight distribution for P(cis-/trans-CHDM-HOP).

Figure 28A graphically represents the active energy as a function of frequency of P(*trans*- CHDM-HOP), and Figure 28B shows the corresponding viscosity.

Figure 29A shows HEK293 cells grown on a P(CHDM-HOP) surface after 72 hours of incubation, and Figure 29B shows HEK293 cells grown on a TCPS surface after 72 hours" incubation.

Figure 30 graphically represents the effect of the side chain structure an the in vitro degradation rate of three poly (phosphoesters) of the invention in phosphate buffer solution.

Figure 31 shows the release curves of the bio-macromolecule FITC-BSA from the polymer P(CHDM-HOP) at 33% loading.

Figure 32 graphically represents the in vitro release kinetics of FTTC-BSA as a function of a loading levels of 30%, 10% and 1 %.

Figure 33 graphically represents the in vitro effect of side chain structure on the protein release kinetics of FITC-BSA with a 10% loading level.

Figure 34 shows the release of low molecular weight drugs (doxorubicin, cisplatin, and 5-nuoroufacil) from P(CHDM-HOP).

Figure 35 shows the Calibration curves for the release of cisplatin and doxorubicin from a P(CHDM-HOP) matrix in tissue culture medium.

Figure 36 shows the distribution of tumor sizes in mice four weeks after tumor implantation in an in vivo melanoma tumor model.

Figure 37 shows the distribution of tumor sizes in mice six weeks after tumor implantation in an in vivo melanoma tumor model.

Figure 38 shows the percentage of survival as a function of time for four different treatment groups in an in vivo melanoma tumor model.

Figure 39 shows the release curves of two polymer compositions of the invention, one comprising the chemotherapeutic agent paclitaxel in the polymer P(CHDM-EOP) and the other comprising paclitaxel in the polymer P(CHDM-HOP).

Figure 40 shows the efficacy of paclitaxel in a solvent and paclitaxel in a p(DAPG-EOP) polymer in an ovarian cancer animal model (OVCAR3).

Figure 41 shows the efficacy of p(DAPG-EOP) containing paclitaxel in an OVCAR3 ovarian cancer animal model.

Figure 42 shows the efficacy of p(DAPG-EOP) containing paclitaxel in an OVCAR3 ovarian cancer animal model.

Figure 43 shows further efficacy data for p(DAPG-EOP) containing paclitaxel in an OVCAR3 ovarian cancer animal

model.

Figure 44 shows still further efficacy data for p(DAPG-EOP) containing paclitaxel in an OVCAR3 ovarian cancer animal model.

DETAILED DESCRIPTION OF THE INVENTION

Polymeric Compositions

[0016] As used herein, the expression "a mammalian subject" refers to any mammalian subject, such as mice, rats, guinea pigs, cats, dogs, human beings, cows, horses, sheep, or other livestock. The expression "a mammalian subject having ovarian cancer" includes, but is not limited to, subjects suffering from current symptoms of this disease; subjects who are recovering from other modes of treatment for the disease, such as surgery, chemotherapy, or other treatment; and subjects simply believed to be at greater than average risk for ovarian cancer, such as those who have at least partially recovered from the disease in the past or those subjects having a significant number of female relatives diagnosed as having or having had the disease.

[0017] As used herein, the term "treating" includes:

(i) preventing a disease, disorder or condition from occurring in an animal which may be predisposed to the disease, disorder and/or condition but has.not yet been diagnosed as having it;
(ii) inhibiting the disease, disorder or condition, i.e., arresting its development; and
(iii) relieving the disease, disorder or condition, i.e., causing regression of the disease, disorder and/or condition.

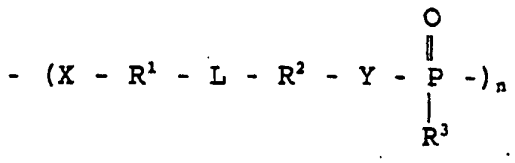
[0018] The term "aliphatic" refers to a linear, branched or cyclic alkane, alkene, or alkyne. Preferred linear or branched aliphatic groups in the poly(cycloaliphatic phosphoester) composition of the invention have from about 1 to 20 carbon atoms. Preferred cycloaliphatic groups may have one or more sites of unsaturation, i.e., double or triple bonds, but are not aromatic in nature.

[0019] As used herein, the term "aryl" refers to an unsaturated cyclic carbon compound with 4n+2 π electrons. As used herein, the term "heterocyclic" refers to a saturated or unsaturated ring compound having one or more atoms other than carbon in the ring, for example, nitrogen, oxygen or sulfur. "Heteroaryl" refers to a heterocyclic compound with 4n+2 electrons.

[0020] As used herein, the term "non-interfering substituent" means a substituent that does react with the monomers; does not catalyze, terminate or otherwise interfere with the polymerization reaction; and does not react with the resulting polymer chain through intra- or inter-molecular reactions.

[0021] The biodegradable and injectable polymer composition comprises a polymer having the recurring monomeric units shown in formula I:

I



wherein X is -O- or -NR^4-, where R^4 is H or alkyl, such as methyl, ethyl, 1,2-dimethylethyl, n-propyl, isopropyl, 2-methylpropyl, 2,2-dimethylpropyl or tert-butyl, n-pentyl, tert-pentyl, n-hexyl, n-heptyl and the like.

[0022] The group Y in formula I is -O-, -S- or -NR^4-, where R^4 is as defined above.

[0023] Each of R^1 and R^2 can be any divalent organic moiety, which may be either unsubstituted or substituted with one or more non-interfering substituents, so long as the moiety and its substituents do not interfere undesirably with the polymerization, copolymerization, or biodegradation reactions of the polymer. Specifically, each of R^1 and R^2 can be a branched or straight chain aliphatic group, preferably having about 1-20 carbon atoms. For example, R^1 and R^2 can be alkylene, such as methylene, ethylene, 1-methylethylene, 1,2-dimethylethylene, n-propylene, isopropylene, 2-methylpropylene, 2,2'-dimethylpropylene or tert-butylene, n-pentylene, tert-pentylene, n-hexylene, n-heptylene, n-octylene, n-nonylene, n-decylene, n-undecylene, n-dodecylene, and the like.

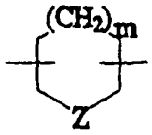
[0024] R^1 and R^2 can also be alkenylene, such as ethenylene, propenylene, 2-vinylpropenylene, n-butenylene, 3-ethenylbutylene, n-pentenylene, 4-(3-propenyl)hexylene, n-octenylene, 1-(4-butenyl)-3-methyldecylene, dodecenylene, 2-(3-propenyl)dodecylene, hexadecenylene, and the like. R^1 and R^2 can also be alkynylene, such as ethy-

nylene, propynylene, 3-(2-ethynyl)pentylene, n-hexynylene, octadecenynylene, 2-(2-propynyl)decylene, and the like.

[0025] R^1 and R^2 can also be an aliphatic group, such as an alkylene, alkenylene or alkynylene group, substituted with a non-interfering substituent, for example, a hydroxy, halogen or nitrogen group. Examples of such groups include, but are not limited to, 2-chloro-n-decylene, 1-hydroxy-3-ethenylbutylene, 2-propyl-6-nitro-10-dodecynylene and.the like.

[0026] Further, R^1 and R^2 can be a cycloaliphatic group, such as cyclopentylene, 2-methylcyclopentylene, cyclohexylene, cyclohexenylene and the like. Each of R^1 and R^2 can also be a divalent aromatic group, such as phenylene, benzylene, naphthalene, phenanthrenylene, and the like, or a divalent aromatic group substituted with a non-interfering substituent. Further each of R^1 and R^2 can be a divalent heterocyclic group, such as pyrrolylene, furanylene, thiophenylene, alkylene-pyrrolylene-alkylene, pyridylene, pyridinylene, pyrimidinylene and the like, or may be any of these substituted with a non-interfering substituent.

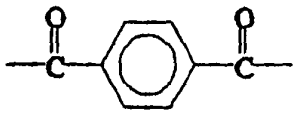
[0027] Preferably, R^1 and R^2 have from about 1-20 carbon atoms and are an alkylene group, a cycloaliphatic group, a phenylene group, or a divalent group having the formula:



wherein Z is oxygen, nitrogen, or sulfur, and m is 1 to 3. More preferably, each of R^1 and R^2 is a branched or straight chain alkylene group having from 1 to 7 carbon atoms. Most preferably, each of R^1 and R^2 is a methylene, ethylene group, n-propylene, 2-methylpropylene, or a 2,2'-dimethylpropylene group.

[0028] In one embodiment of the invention, either R^1, R^2 or both R^1 and R^2, can be an antineoplastic agent in a form capable of being released in a physiological environment. When the antineoplastic agent part of the poly(phosphoester) backbone in this way, it is released as the polymeric matrix formed by the composition of the invention degrades.

[0029] L in the polymer composition of the invention can be any divalent, branched or straight chain aliphatic group having 1-20 carbon atom, a cycloaliphatic group, or a group having the formula:



;

[0030] When L is a branched or straight chain alkylene group, it is preferably an alkylene group having from 1 to 7 carbon atoms, such as 2-methylmethylene or ethylene. When L is a cycloaliphatic group, it may be any divalent cycloaliphatic group so long as it does not interfere with the polymerization or biodegradation reactions of the polymer of the composition. Specific examples of useful unsubstituted and substituted cycloaliphatic L groups, include cycloalkylene groups such as cyclopentylene, 2-methylcyclopentylene, cyclohexylene, 2-chlorocyclohexylene, and the like; cycloalkenylene groups, such as cyclohexenylene; and cycloalkylene groups having fused or bridged additional ring structures on one or more sides, such as tetralinylene, decalinylene, and norpinanylene; or the like.

[0031] R^3 in the polymer composition is selected from the group consisting of H, alkyl, alkoxy, aryl, aryloxy, heterocyclic and heterocycloxy residues.

[0032] When R^3 is alkyl or alkoxy, it preferably contains about 1 to about 20 carbon atoms, even more preferably about 1 to about 15 carbon atoms and, most preferably about 1-7 carbon atoms. Examples of such groups include methyl, methoxy, ethyl, ethoxy, n-propyl, isopropoxy, n-butaxy, t-butyl, $-C_8H_{17}$; alkyl substituted with a non-interfering substituent, such as halogen, alkoxy or nitro;

[0033] When R^3 is aryl or the corresponding aryloxy group, it typically contains from about 5 to about 14 carbon atoms, preferably about 5 to 12 carbon atoms and, optionally, can contain one or more rings that are fused to each other. Examples of particularly suitable aromatic groups include phenyl, phenoxy, naphthyl, anthracenyl, phenanthrenyl and the like.

[0034] When R^3 is heterocyclic or heterocycloxy, it typically contains from about 5 to 14 ring atoms, preferably from about 5 to 12 ring atoms, and one or more heteroatoms. Examples of suitable heterocyclic groups include furan, thiophene, pyrrole, isopyrrole, 3-isopyrrole, pyrazole, 2-isoimidazole, 1,2,3-triazole, 1,2,4-triazole, oxazole, thiazole, iso-

thiazole, 1,2,3-oxadiazole, 1,2,4-oxadiazole, 1,2,5-oxadiazole, 1,3,4-oxadiazole, 1,2,3,4-oxatriazole, 1,2,3,5-oxatriazole, 1,2,3-dioxazole, 1,2,4-dioxazole, 1,3,2-dioxazole, 1,3,4-dioxazole, 1,2,5-oxatriazole, 1,3-oxathiole, 1,2-pyran, 1,4-pyran, 1,2-pyrone, 1,4-pyrone, 1,2-dioxin, 1,3-dioxin, pyridine, N-alkyl pyridinium, pyridazine, pyrimidine, pyrazine, 1,3,5-triazine, 1,2,4-triazine, 1,2,3-triazine, 1,2,4-oxazine, 1,3,2-oxazine, 1,3,5-oxazine, 1,4-oxazine, o-isoxazine, p-isoxazine, 1,2,5-oxathiazine, 1,2,6-oxathiazine, 1,4,2-oxadiazine, 1,3,5,2-oxadiazine, azepine, oxepin, thiepin, 1.2,4-diazepine, indene, isoindene, benzofuran, isobenzofuran, thionaphthene, isothionaphthene, indole, indolenine, 2-isobenzazole, 1,4-pyrindine, pyrando[3,4-b]-pyrrole, isoindazole, indoxazine, benzoxazole, anthranil, 1,2-benzopyran, 1,2-benzopyrone, 1,4-benzopyrone, 2,1-benzopyrone, 2,3-benzopyrone, quinoline, isoquinoline, 12,-benzodiazine, 1,3-benzodiazine, naphthpyridine, pyrido[3,4-b]-pyridine, pyrido [3,2-b]-pyridine, pyrido [4, 3-b] pyridine, 1,3,2-benzoxazine, 1,4,2-benzoxazine, 2,3,1-benzoxazine, 3,1,4-benzoxazine, 1,2-benzisoxazine, 1,4-benzisoxazine, carbazole, xanthrene, acridine, purine, and the like. Preferably, when R^3 is heterocyclic or heterocycloxy, it is selected from the group consisting of furan, pyridine, N-alkylpyridine, 1,2,3- and 1,2,4-triazoles, indene, anthracene and purine rings.

[0035] In a particularly preferred embodiment, R^3 is an alkyl group, an alkoxy group, a phenyl group, a phenoxy group, or a heterocycloxy group and, even more preferably, an alkoxy group having from 1 to 10 carbon atoms. Most preferably, R^3 is an ethoxy or hexyloxy group.

[0036] Alternatively, the side chain R^3 can be the antineoplastic agent or some other biologically active substance pendently attached to the polymer backbone, for example by ionic or covalent bonding. In this pendant system, the antineoplastic agent or other biologically active substance is released as the bond connecting R^3 with the phosphorous atom is cleaved under physiological conditions.

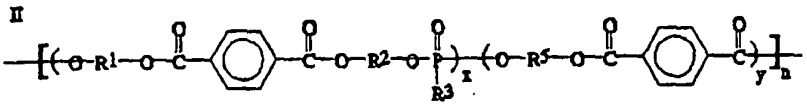
[0037] The number "n" can vary greatly depending on the biodegradability and the release characteristics desired in the polymer, but typically varies between about 5 and 1,000. Preferably, n is from about 5 to about 500 and, most preferably, is from about 5 to about 200.

[0038] When used in accordance with the uses of the invention, the polymer composition provides extended release of the antineoplastic agent into the peritoneum of a subject having ovarian cancer, preferably for a period greater than one week, more preferably for a period greater than two weeks. Even more preferably, this time extends for a period greater than about three weeks and, most preferably, is for a period greater than four weeks, for example, from four weeks to a year.

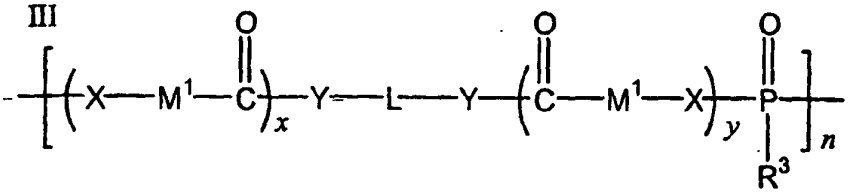
[0039] Further, use of the composition in accordance with the use of the invention increases the median survival rate from the cancer by at least about 10%, as compared with the median survival rate obtained by administration of a composition comprising the same dosage of antineoplastic agent without the biodegradable polymer of the invention. Preferably, the median survival rate is increased by at least about 20%, more preferably by at least 30% and, most preferably, by a factor of at least about 40%.

[0040] The polymer used in the invention is preferably selected from the group consisting of:

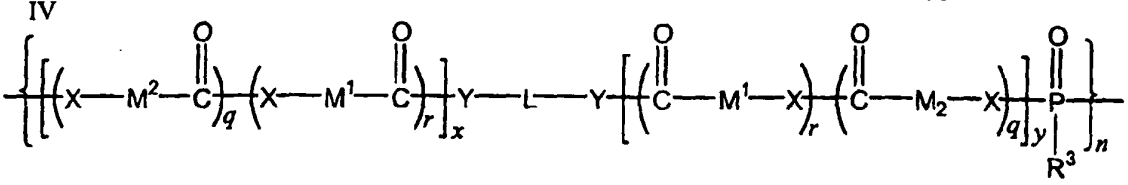
II



III

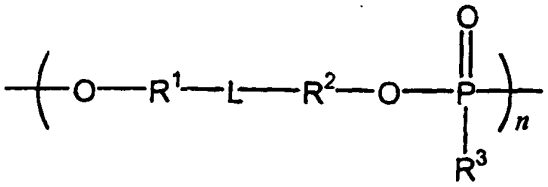


IV



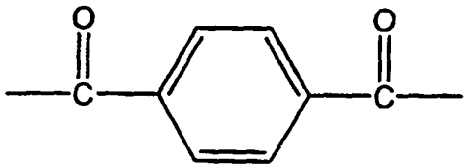
and

V



wherein R^1, R^2, R^3 and n are as defined above.

[0041] In polymers of formula II, R^5 is selected from the same groups as for R^1 and R^2, and L is preferably a group having the formula:



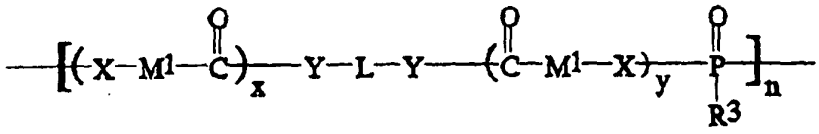
[0042] The molar ratio of x:y in formula II can vary greatly depending on the desired solubility of the polymer, the desired glass transition temperature (Tg), the desired stability of the polymer, the desired stiffness of the final polymers, and the biodegradability and the release characteristics desired in the polymer. However, the molar ratio of x:y typically varies between about 20:0 and 1:20. When y is 0, the polymer formed is a homopolymer. Preferably, however, the ratio x:y is from about 1:15 to about 15:1, more preferably, from about 10:1 to about 1:1.

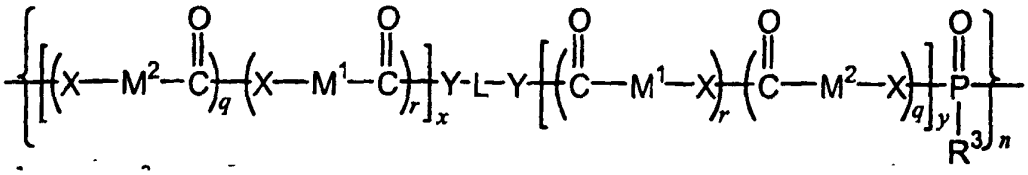
[0043] The most common way of controlling the molar ratio of x:y is to vary the feed ratio of the "x" portion to the "y" portion when synthesizing the polymer. Feed ratios can easily vary from 99: to 1:99, for example, 95:5, 90:10, 85:15, 80:20, 75:25, 70:30, 65:35, 60:40, 55:45, 50:50, 45:55, 20:80, 15:85, and the like. Preferably, the monomer feed ratio varies from about 90:10 to about 50:50, even more preferably from about 80:20 to about 50:50 and, most preferably, from about 80:20 to about 50:50.

[0044] Preferably, when the biodegradable polymer has formula II, R^3 is alkoxy, aryloxy or heterocycloxy; x is about 0.1 to 30, more preferably about 0.2 to 20, most preferably ≥1 (for example, about 2-20); and y is 2.

[0045] In preferred polymers of formula III and IV:

III





M^1 and M^2 are each independently (1) a branched or straight chain aliphatic group having from about 1-20 carbon atoms, even more preferably from about 1-7 carbon atoms; or (2) a branched or straight chain, oxy-, carboxy- or amino-aliphatic group having from about 1-20 carbon atoms, such as ethoxylene, 2-methylethoxylene, propoxylene, butoxylene, pentoxylene, dodecyloxylene, hexadecyloxylene, and the like;

each of x and y is about 1 to 1,000;

the molar ratio of x:y can vary greatly depending on the biodegradability and the release characteristics desired in the polymer but, typically, is about 1;

the molar ratio n:(x or y) can vary greatly depending on the biodegradability and the release characteristics desired in the polymer, but typically varies between about 200:1 and 1:200, preferably 100:1 and 1:100, more preferably from about 50:1 to about 1:50; and

the molar ratio q:r can also vary greatly depending on the biodegradability and the release characteristics desired in the polymer, but typically varies between about 1:200 and 200:1, preferably between about 1:150 to about 150:1 and, most preferably, between about 1:99 and 99:1.

[0046] In formula III, each of M^1 and L preferably has from 1 to 7 carbon atoms. More preferably, M^1 is an ethylene group or a methyl-substituted methylene group, and L is an ethylene group.

[0047] In formula IV, each of M^1 and M^2 is preferably a branched or straight chain alkylene or alkoxylene group, more preferably having from 1-20 carbon atoms. Even more preferably, at least one of M^1 and M^2 is an alkylene or alkoxylene group having a formula selected from the group consisting of $-(CH_2)_a-$, $-(CH_2)_a-O-$, and $-(CH_2)_a-O-(CH_2)_b-$, wherein each of a and b is 1-7.

[0048] When either M^1 and M^2 is a branched or straight chain, oxy-aliphatic group having from 1-20 carbon atoms it can also be, for example, a dioxyalkylene group such as such as dioxymethylene, dioxyethylene, 1,3-dioxypropylene, 2-methoxy-1,3-dioxypropylene, 1,3-dioxy-2-methylpropylene, dioxy-n-pentylene, dioxy-n-octadecylene, methoxylene-methoxylene, ethoxylene-methoxylene, ethoxylene-ethoxylene, ethoxylene-1-propoxylene, butoxylene-n-propoxylene, pentadecyloxylene-methoxylene, and the like. When M^1 and M^2 is a branched or straight chain, dioxo-aliphatic group, preferably it has the formula $-O-(CH_2)_a-O-$ or $-O-(CH_2)_a-O-(CH_2)_b-$, wherein each of a and b is from 1-7.

[0049] When either M^1 or M^2 is a branched or straight chain, carboxy-aliphatic group having from 1-20 carbon atoms, it can also be, for example, a divalent carboxylic acid ester such as the divalent radical corresponding to methyl formate, methyl acetate, ethyl acetate, n-propyl acetate, isopropyl acetate, n-butyl acetate, ethyl propionate, allyl propionate, t-butyl acrylate, n-butyl butyrate, vinyl chloroacetate, 2-methoxycarbonyl cyclohexanone, 2-acetoxycyclohexanone, and the like. When M^1 or M^2 is a branched or straight chain, carboxy-aliphatic group, it preferably has the formula -CHR'-CO-O-CHR"-, wherein R' and R" are each independently H, alkyl, alkoxy, aryl, aryloxy, heterocyclic or heterocycloxy.

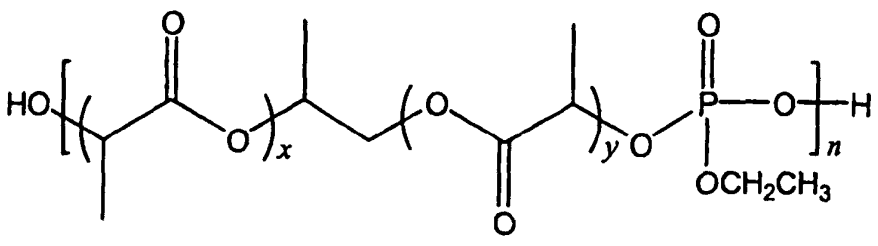
[0050] When either M^1 or M^2 is a branched or straight chain, amino-aliphatic group having from 1-20 carbon atoms, it can be a divalent amine such as $-CH_2NH-$, $-(CH_2)_2N-$, $-CH_2(C_2H_5)N-$, $-n-C_4H_9-NH-$, $-t-C_4H_9-NH-$, $-CH_2(C_3H_6)N-$, $-C_2H_5(C_3H_6)N-$, $-CH_2(C_8H_{17})N-$, and the like. When M^1 or M^2 is a branched or straight chain, amino-aliphatic group, it preferably has the formula $-(CH_2)a-NR'$ where R' is H or lower alkyl, and "a" is from 1 to 7.

[0051] Preferably, M^1 and/or M^2 is an alkylene group having the formula $-O-(CH_2)a-$ where a is 1 to 7 and, most preferably, is a divalent ethylene group. In another particularly preferred embodiment, M^1 and M^2 are n-pentylene and the divalent radical corresponding to methyl acetate respectively.

[0052] In a preferred embodiment, L in formulas III and IV is a branched or straight chain aliphatic group having from 1-20 carbon atoms, more preferably an alkylene group having from 1 to 7 carbon atoms, such as ethylene or methyl-substituted methylene.

[0053] In another particularly preferred polymer of formula IV, M^1 and M^2 are each an alkylene or alkoxylene group; L is an alkylene group; X is -O-; and R^3 is an alkoxy group. Most preferably, the biodegradable polymer used in the invention comprises the recurring monomeric units shown in formula VI:

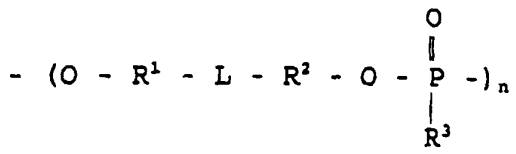
VI



wherein the molar ratio of x:y is about 1;
the molar ratio n:(x or y) is between about 200:1 and 1:200; and
n is about 5-5,000.

[0054] When the polymer used has formula V:

V



preferably, each of R^1 and R^2 is independently straight or branched aliphatic, such as a branched or straight chain alkylene group having from 1 to 7 carbon atoms, for example methylene or ethylene, either unsubstituted or substituted with one or more non-interfering substituents;

L is a divalent cycloaliphatic group, such as cyclohexylene, either unsubstituted or substituted with a non-interfering substituent;

R^3 is selected from the group consisting of H, alkyl, alkoxy, aryl, aryloxy, heterocyclic or heterocycloxy (preferably alkoxy such as ethoxy or hexyloxy); and

n is about 5-5,000, even more preferably 5 to 500.

[0055] The molecular weight of the polymer used in the invention can vary widely, depending on whether a rigid solid state (higher molecular weights) desirable, or whether a flowable or flexible state (lower molecular weights) is desired. Generally, however, weight-average molecular weights (Mw) typically vary from about 2,000 to about 400,000 daltons, preferably from about 2,000 to about 200,000 daltons and, even more preferably, from about 2,000 to 60,000 daltons. Most preferably, the Mw varies between about 10,0000 to 55,000. Number-average molecular weight (Mn) can also vary widely, but generally fall in the range of about 1,000 to about 200,000 daltons, preferably from about 1,000 to about 100,000 daltons and, even more preferably, from about 1,000 to about 50,000 daltons. Most preferably, Mn varies between about 8,000 and 45,000 daltons.

[0056] A preferred method to determine molecular weight is by gel permeation chromatography ("GPC"), e.g., mixed bed columns, CH_2Cl_2 solvent, light scattering detector, and off-line dn/dc.

[0057] The glass transition temperature (Tg) of the polymer used in the invention can vary widely depending upon the degree of branching in R^1 and R^2, the relative proportion of phosphorous-containing monomer used to make the polymer, and the like. When the article of the invention is a rigid solid, the Tg is preferably within the range of from about -10°C to about 80°C, even more preferably between about 0 and 50°C and, most preferably between about 25°C to about 35°C.

[0058] In other embodiments, the Tg is preferably low enough to keep the composition of the invention flowable at body temperature. Then, the glass transition temperature of the polymer used in the invention is preferably about 0 to about 37°C, more preferably from about 0 to about 25°C.

[0059] The biodegradable polymer used in the invention is preferably sufficiently pure to be biocompatible itself and remains biocompatible upon biodegradation. By "biocompatible", it is meant that the biodegradation products or the polymer itself are non-toxic and result in only minimal tissue irritation when injected or placed into intimate contact with vasculated tissues. The requirement for biocompatibility is more easily accomplished because the presence of an organic solvent is not required in the polymer composition of the invention.

[0060] However, the polymer used in the invention is preferably soluble in one or more common organic solvents for ease of synthesis, purification and handling. Common organic solvents include such solvents as ethanol; chloroform, dichloromethane (dimethylene chloride), acetone, ethyl acetate, DMAC, N-methyl pyrrolidone, dimethylformamide, and dimethylsulfoxide. The polymer is preferably soluble in at least one of the above solvents.

[0061] The polymer can also comprise additional biocompatible monomeric units so long as they do not interfere with the biodegradable characteristics and the desirable flow characteristics of the invention. Such additional monomeric units may offer even greater flexibility in designing the precise release profile desired for targeted drug delivery or the precise rate of biodegradability desired for other applications. When such additional monomeric units are used, however, they should be used in small enough amounts to insure the production of a biodegradable copolymer having the desired physical characteristics, such as rigidity, viscosity, flowability, flexibility or a particular morphology.

[0062] Examples of such additional biocompatible monomers include the recurring units found in other poly(phosphoesters), poly(lactides), poly(glycolides), poly(caprolactones), poly(anhydrides), poly(amides), poly(urethanes), poly (esteramides), poly(orthoesters), poly(dioxanones), poly(acetals), poly(ketals), poly(carbonates), poly(orthocarbonates), poly(phosphazenes), poly(hydroxybutyrates), poly(hydroxyvalerates), poly(alkylene oxalates), poly(alkylene succinates), poly(malic acids), poly(amino acids), poly(vinylpyrrolidone), poly(ethylene glycol), poly-(hydroxycellulose), chitin, chitosan, and copolymers, terpolymers, or combinations or mixtures of the above materials.

[0063] When additional monomeric units are used, those which have a lower degree of crystallization and are more hydrophobic are preferred. Especially preferred recurring units with the desired physical characteristics are those derived from poly(lactides), poly(caprolactones), and copolymers of these with glycolide, in which there are more amorphous regions.

General Synthesis of Phosphoester Polymers

[0064] The most common general reaction in preparing poly-(phosphates) is a dehydrochlorination between a phosphorodichloridate and a diol according to the following equation:

Most poly(phosphonates) are also obtained by condensation between appropriately substituted dichlorides and diols.

[0065] Poly(phosphites) have been prepared from glycols in a two-step condensation reaction. A 20% molar excess of a dimethylphosphite is used to react with the glycol, followed by the removal of the methoxyphosphonyl end groups in the oligomers by high temperature.

[0066] An advantage of melt polycondensation is that it avoids the use of solvents and large amounts of other additives, thus making purification more straightforward. It can also provide polymers of reasonably high molecular weight. Somewhat rigorous conditions, however, are often required and can lead to chain acidolysis (or hydrolysis if water is present). Unwanted, thermally-induced side reactions, such as crosslinking reactions, can also occur if the polymer backbone is susceptible to hydrogen atom abstraction or oxidation with subsequent macroradical recombination.

[0067] To minimize these side reactions, the polymerization can also be carried out in solution. Solution polycondensation requires that both the prepolymer and the phosphorus component be soluble in a common solvent. Typically, a chlorinated organic solvent is used, such as chloroform, dichloromethane, or dichloroethane.

[0068] A solution polymerization is preferably run in the presence of equimolar amounts of the reactants and a stoichiometric amount of an acid acceptor, usually a tertiary amine such as pyridine or triethylamine. Reaction times tend to be longer with solution polymerization than with melt polymerization. However, because overall milder reaction conditions can be used, side reactions are minimized, and more sensitive functional groups can be incorporated into the polymer. Moreover, attainment of high molecular weights is less likely with solution polymerization.

[0069] Interfacial polycondensation can be used when high reaction rates are desired. The mild conditions used minimize side reactions, and there is no need for stoichiometric equivalence between the diol and dichloridate starting materials as in solution methods. However, hydrolysis of the acid chloride may occur in the alkaline aqueous phase. Sensitive dichloridates that have some solubility in water are generally subject to hydrolysis rather than polymerization. Phase transfer catalysts, such as crown ethers or tertiary ammonium chloride, can be used to bring the ionized diol to the interface to facilitate the polycondensation reaction. The yield and molecular weight of the resulting polymer after interfacial polycondensation are affected by reaction time, molar ratio of the monomers, volume ratio of the immiscible solvents, the type of acid acceptor, and the type and concentration of the phase transfer catalyst.

[0070] The purpose of the polymerization reaction is to form a polymer comprising (i) divalent organic recurring units and (ii) phosphoester recurring units. The result can be a homopolymer, a relatively homogeneous copolymer, or a block copolymer that has a somewhat heterogeneous microcrystalline structure. Any one of these three embodiments is well-suited for use as a controlled release medium.

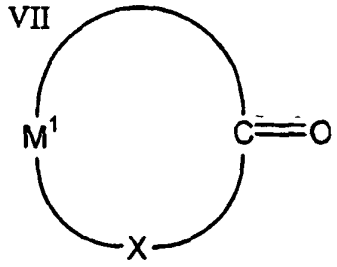
[0071] While the process may be in bulk, in solution, by interfacial polycondensation, or any other convenient method of polymerization, preferably, the process takes place under solution conditions. Particularly useful solvents include methylene chloride, chloroform, tetrahydrofuran, dimethyl formamide, dimethyl sulfoxide, toluene, or any of a wide variety of other inert organic solvents.

[0072] Particularly when solution polymerization reaction is used, an acid acceptor is advantageously present during the polymerization reaction. A particularly suitable class of acid acceptor comprises tertiary amines, such as pyridine, trimethylamine, triethylamine, substituted anilines and substituted aminopyridines. The most preferred acid acceptor is the substituted aminopyridine 4-dimethylaminopyridine ("DMAP").

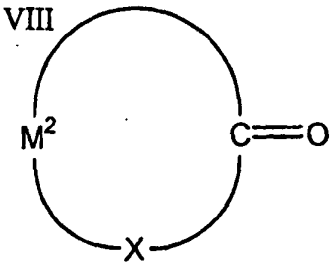
[0073] For example, the biodegradable polymer of formula III or IV is made by a process comprising the steps of:

(a) reacting at least one heterocyclic ring compound having formula VII, VIII or IX:

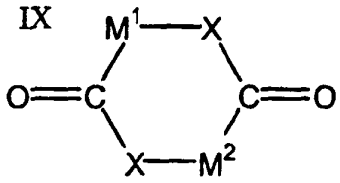
VII



VIII



IX

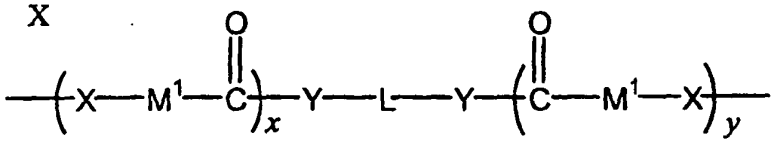


wherein M^1, M^2 and X are as defined above, with an initiator having the formula:

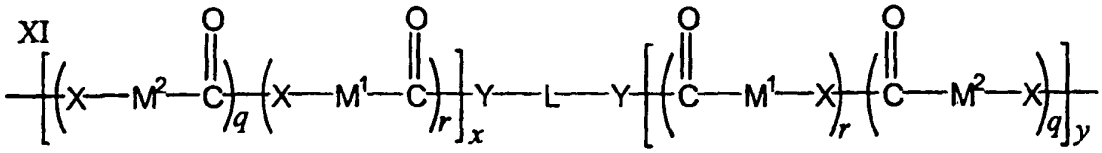
H-Y-L-Y-H,

wherein Y and L are as defined as above, to form a prepolymer of formula X or XI, shown below:

X

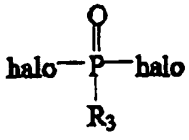


XI



wherein X, M^1, M^2, Y, L, R, x, y, q and r are as defined above; and
(b) further reacting the prepolymer with a phosphorodihalidate of formula XII:

XII



where "halo" is Br, Cl or I; and R^3 is as defined above, to form a polymer of formula III or IV.

[0074] The function of the first reaction step (a) is to use the initiator to open the ring of the heterocyclic ring compound of formula VII, VIII or IX. Examples of useful heterocyclic compounds of formula VII, VIII or IX include caprolactones, caprolactams, amino acid anhydrides such as glycine anhydride, cycloalkylene carbonates, dioxanones, glycolids, lactides and the like.

[0075] When the compound has formula III, only one heterocyclic ring compound of formula VII, which contains M^1, may be used to prepare the prepolymer in step (a). When the compound of the invention has formula IV, then a combination of a heterocyclic compound of formula VII, which contains M^1, and a heterocyclic compound of formula VIII, which contains M^2 may be used in step (a). Alternatively, when the compound of the invention has formula IV, a single heterocyclic compound of formula IX, which contains both M^1 and M^2 can be used in step (a).

[0076] Examples of suitable initiators include a wide variety of compounds having at least two active hydrogens (H-Y-L-Y-H) where L is a linking group and is defined above, and Y can be -O-, -S- or $-NR^4$, where R^4 is as defined above. The linking group L is can be a straight chain group, e.g., alkylene, but it may also be substituted with one or more additional active-hydrogen-containing groups. For example, L may be a straight chain alkylene group substituted with one or more additional alkyl groups, each bearing a activated hydrogen moiety, such as -OH, -SH, or NH_2. In this way, various branched polymers can be prepared using the branched active hydrogen initiators to design the resulting polymer such that it has the desired properties. However, when branched polymers are reacted with acid chlorides, cross-linked polymers will result.

[0077] The reaction step (a) can take place at widely varying temperatures, depending upon the solvent used, the molecular weight desired, the susceptibility of the reactants to form side reactions, and the presence of a catalyst. Preferably, however, the reaction step (a) takes place at a temperature from about 0 to about +235°C for melt conditions. Somewhat lower temperatures may be possible with the use of either a cationic or anionic catalyst.

[0078] While the reaction step (a) may be in bulk, in solution, by interfacial polycondensation, or any other convenient method of polymerization, preferably, the reaction step (a) takes place under melt conditions.

[0079] Examples of particularly useful prepolymers of formula X include:

(i) OH-terminated prepolymer derived from polycaprolactone

$$H\text{-}[\text{-}O(CH_2)_5\text{-}CO\text{-}]_x\text{-}O\text{-}CH_2\text{-}CH_2\text{-}O\text{-}[\text{-}CO\text{-}(CH_2)_5\text{-}O\text{-}]_y\text{-}H\ ;$$

(ii) NH-terminated prepolymer derived from polycaprolactam (Nylon 6)

$$H\text{-}[\text{-}NH\text{-}(CH_2)_5\text{-}CO\text{-}]_x\text{-}NH\text{-}CH_2\text{-}CH_2\text{-}NH\text{-}[\text{-}CO\text{-}(CH_2)_5\text{-}NH\text{-}]_y\text{-}H\ ;$$

(iii) OH-terminated prepolymer derived from polylactide

$$H\text{-}[\text{-}OCH(CH_3)\text{-}CO\text{-}]_x\text{-}O\text{-}CH_2\text{-}CH_2\text{-}O\text{-}[\text{-}CO\text{-}CH(CH_3)\text{-}O\text{-}]_y\text{-}H\ ;$$

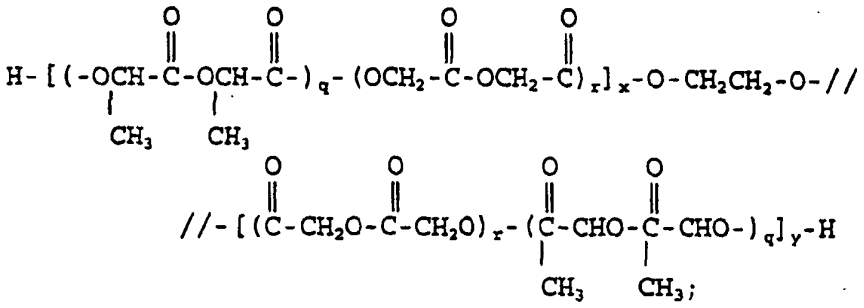
and

(iv) OH-terminated prepolymer derived from polytrimethylene carbonate

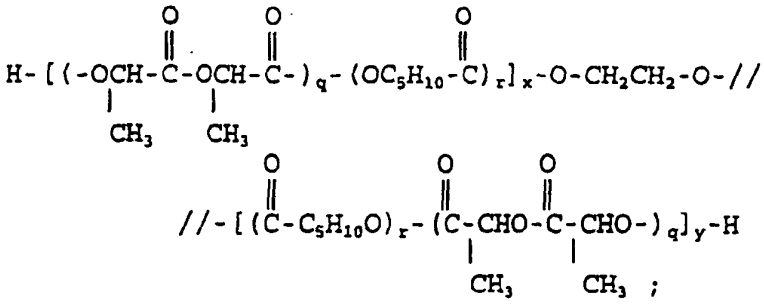
$$H\text{-}\ [\text{-}O(CH_2)_3\text{-}O\text{-}CO\text{-}]_x\text{-}O\text{-}CH_2\text{-}CH_2\text{-}O\text{-}[\text{-}CO\text{-}O\text{-}(CH_2)_3\text{-}O\text{-}]_y\text{-}H$$

[0080] Examples of particularly useful prepolymers of formula XI include:

(i) OH-terminated copolymer derived from lactide and glycolide:

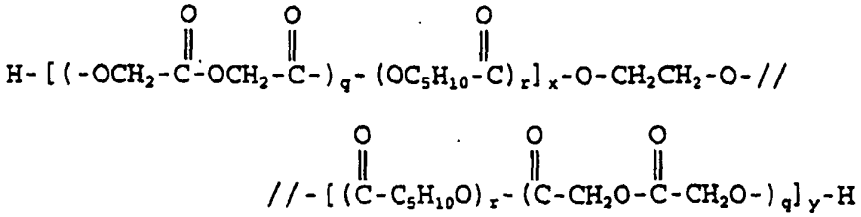


(ii) OH-terminated copolymer derived from lactide and caprolactone:



and

(iii) OH-terminated copolymer derived from glycolide and caprolactone:



[0081] The purpose of the polymerization of step (b) is to form a polymer comprising (i) the prepolymer produced as a result of step (a) and (ii) interconnecting phosphorylated units: The result can be a block copolymer having a microcrystalline structure that is particularly well-suited to use as a controlled release medium.

[0082] The polymerization step (b) usually takes place at a slightly lower temperature than the temperature of step (a), but also may vary widely, depending upon the type of polymerization reaction used, the presence of one or more catalysts, the molecular weight desired, and the susceptibility of the reactants to undesirable side reaction. When melt conditions are used, the temperature may vary from about 0-150°C. However, when the polymerization step (b) is carried out in a solution polymerization reaction, it typically takes place at a temperature between about -40 and 100°C.

Antineoplastic Agent

[0083] Generally speaking, the antineoplastic agent can vary widely depending upon the pharmacological strategy selected for inhibiting, destroying, or preventing the ovarian cancer. The antineoplastic agent may be described as a single entity or a combination of entities. The compositions, articles and methods are designed to be used with antineoplastic agents having high water-solubility, as well as those having low water-solubility, to produce a delivery system that has controlled release rates.

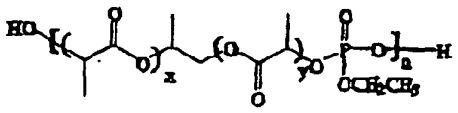
[0084] The term antineoplastic agent includes, without limitation, alkylating agents, such as carboplatin and cisplatin; nitrogen mustard alkylating agents; nitrosourea alkylating agents, such as carmustine (BCNU); antimetabolites, such as methotrexate; purine analog antimetabolites; pyrimidine analog antimetabolites, such as fluorouracil (5-FU) and gemcitabine; hormonal antineoplastics, such as goserelin, leuprolide, and tamoxifen; natural antineoplastics, such as aldesleukin, interleukin-2, docetaxel, etoposide (VP-16), interferon alfa, paclitaxel, and tretinoin (ATRA); antibiotic natural antineoplastics, such as bleomycin, dactinomycin, daunorubicin, doxorubicin, and mitomycin; and vinca alkaloid natural antineoplastics, such as vinblastine and vincriatine. Preferably, the antineoplastic agent is selected from the group consisting of paclitaxel, BCNU, carboplatin and cisplatin. Most preferably, the antineoplastic agent is paclitaxel.

[0085] Various forms of the antineoplastic agents and/or other biologically active agents may be used. These include, without limitation, such forms as uncharged molecules, molecular complexes, salts, ethers, esters, amides, and the like, which are biologically activated when implanted, injected or otherwise placed into the body.

[0086] In a particularly preferred embodiment, a biodegradable polymer composition suitable for intraperitoneal administration to treat a mammalian subject having ovarian cancer comprises:

(a) paclitaxel and
(b) a biodegradable polymer comprising the recurring monomeric units shown in formula VI:

VI



wherein the molar ratio of x:y is about 1;
the molar ratio n: (x or y) is between about 200:1 and 1:200; and
a is about 5-5,000.

Biodegradation and Release Characteristics

[0087] Biodegradable polymers differ from non-biodegradable polymers in that they can be degraded during in vivo therapy. This generally involves breaking down the polymer into its monomeric subunits. In principle, the ultimate hydrolytic breakdown products of the polymer used in the invention are a cycloaliphatic diol, an aliphatic alcohol and phosphate. All of these degradation products are potentially non-toxic. However, the intermediate oligomeric products of the hydrolysis may have different properties. Thus, the toxicology of a biodegradable polymer intended for insertion into the body, even one synthesized from apparently innocuous monomeric structures, is typically determined after one or more toxicity analyses.

[0088] There are many different ways of testing for toxicity and/or.biocompatibility known to those of ordinary skill in the art. A typical in vitro toxicity assay, however, would be performed with live carcinoma cells, such as GT3TKB tumor cells, in the following manner:

Two hundred microliters of various concentrations of the degraded polymer products are placed in 96-well tissue culture plates seeded with human gastric carcinoma cells (GT3TKB) at 10^4/well density. The degraded polymer products are incubated with the GT3TKB cells for 48 hours. The results of the assay can be plotted as % relative growth vs. concentration of degraded polymer in the tissue-culture well.

Polymers can also be evaluated by well-known in vivo biocompatibility tests, such as by subcutaneous implantation or injection in rats to confirm that the systems hydrolyze without significant levels of irritation or inflammation at the insertion site.

[0089] The polymer of formula I is usually characterized by a biodegradation rate that is controlled at least in part as a function of hydrolysis of the phosphoester bond of the polymer. Other factors are also important. For example, the lifetime of a biodegradable polymer in vivo also depends upon its molecular weight, crystallinity, biostability, and the degree of crosslinking. In general, the greater the molecular weight, the higher the degree of crystallinity, and the greater the biostability, the slower biodegradation will be. In addition, the rate of degradation of the polymer can be further controlled by choosing a side chain of differing lengths. Accordingly, degradation times can very widely, preferably from less than a day to several months.

[0090] Accordingly, the structure of the side chain can influence the release behavior of compositions comprising a biologically active substance. For example, it is expected that conversion of the phosphate side chain to a more lipophilic, more hydrophobic or bulky group would slow down the degradation process. Thus, release is usually faster from polymer compositions with.a small aliphatic group side chain than with a bulky aromatic side chain.

[0091] The expression "extended release", as used herein, includes, without limitation various forms of release, such as controlled release, timed release, sustained release, delayed release, long acing, and pulsatile delivery, immediate release that occurs with various rates. The ability to obtain extended release, controlled release, timed release, sustained release, delayed release, long acting, pulsatile delivery or immediate release is performed using well-known procedures and techniques available to the ordinarily skilled artisan. None of these specific techniques or procedures constitute an inventive aspect of this invention.

Polymer Compositions

[0092] The antineoplastic agents are used in amounts that are therapeutically effective, which varies widely depending largely on the particular antineoplastic agent being used. The amount of antineoplastic agent incorporated into the

composition also depends upon the desired release profile, the concentration of the agent required for a biological effect, and the length of time that the biologically active substance has to be released for treatment. Preferably, the biologically active substance is blended with the polymer matrix of the invention at different loading levels, preferably at room temperature and without the need for an organic solvent.

[0093] There is no critical upper limit on the amount of antineoplastic agent incorporated except for that of an acceptable solution or dispersion viscosity to maintain the physical characteristics desired for the composition. The lower limit of the antineoplastic agent incorporated into the delivery system is dependent upon the activity of the drug and the length of time needed for treatment. Thus, the amount of the antineoplastic agent should not be so small that it fails to produce the desired physiological effect, nor so large that the antineoplastic agent is released in an uncontrollable manner.

[0094] Typically, within these limits, amounts of the antineoplastic agent from about 1% up to about 65% can be incorporated into the present delivery systems. However, lesser amounts may be used to achieve efficacious levels of treatment for antineoplastic agent that are particularly potent.

[0095] In addition, the polymer composition can also comprise blends of the polymer of the invention with other biocompatible polymers or copolymers, so long as the additional polymers or copolymers do not interfere undesirably with the biodegradable or mechanical characteristics of the composition. Blends of the polymer of the invention with such other polymers may offer even greater flexibility in designing the precise release profile desired for targeted drug delivery or the precise rate of biodegradability desired. Examples of such additional biocompatible polymers include other poly(phosphoesters), poly(carbonates), poly(esters), poly(orthoesters), poly(amides), poly(urethanes), poly(imino-carbonates), and poly(anhydrides).

[0096] Pharmaceutically acceptable polymeric carriers may also comprise a wide range of additional materials. Without being limited thereto, such materials may include diluents, binders and adhesives, lubricants, disintegrants, colorants, bulking agents, flavorings, sweeteners, and miscellaneous materials such as buffers and adsorbents, in order to prepare a particular medicated composition, with the condition that none of these additional materials will interfere with the biocompatibility, biodegradability and physical state desired of the polymer compositions of the invention.

[0097] For delivery of an antineoplastic agent or some other biologically active substance, the agent or substance is added to the polymer composition. The agent or substance is either dissolved to form a homogeneous solution of reasonably constant concentration in the polymer composition, or dispersed to form a suspension or dispersion within the polymer composition at a desired level of "loading" (grams of biologically active substance per grams of total composition including the biologically active substance, usually expressed as a percentage).

[0098] While it is possible that the biodegradable polymer or the biologically active agent may be dissolved in a small quantity of a solvent that is non-toxic to more efficiently produce an amorphous, monolithic distribution or a fine dispersion of the biologically active agent in the flexible or flowable composition, it is an advantage of the invention that, in a preferred embodiment, no solvent is needed to form a flowable composition. Moreover, the use of solvents is preferably avoided because, once a polymer composition containing solvent is placed totally or partially within the body, the solvent dissipates or diffuses away from the polymer and must be processed and eliminated by the body, placing an extra burden on the body's clearance ability at a time when the illness (and/or other treatments for the illness) may have already deleteriously affected it.

[0099] However, when a solvent is used to facilitate mixing or to maintain the flowability of the polymer composition of the invention, it should be non-toxic, otherwise biocompatible, and should be used in minimal amounts. Solvents that are toxic clearly should not be used in any material to be placed even partially within a living body. Such a solvent also must not cause tissue irritation or necrosis at the site of administration.

[0100] Examples of suitable biocompatible solvents, when used, include N-methyl-2-pyrrolidone, 2-pyrrolidone, ethanol, propylene glycol, acetone, methyl acetate, ethyl acetate, methyl ethyl ketone, dimethylformamide, dimethyl sulfoxide, tetrahydrofuran, caprolactam, dimethyl-sulfoxide, oleic acid, or 1-dodecylazacycloheptan-2-one. Preferred solvents include N-methyl-2-pyrrolidone, 2-pyrrolidone, dimethyl sulfoxide, and acetone because of their solvating ability and their biocompatibility.

[0101] The polymer composition may be a flexible or flowable material. By "flowable" is meant the ability to assume, over time, the shape of the space containing it at body temperature. This includes, for example, liquid compositions that are capable of being sprayed into a site; injected with a manually operated syringe fitted with, for example, a 23-gauge needle; or delivered through a catheter.

[0102] Also included by the term "flowable", however, are highly viscous, "gel-like" materials at room temperature that may be delivered to the desired site by pouring, squeezing from a tube, or being injected with any one of the commercially available power injection devices that provide injection pressures greater than would be exerted by manual means alone for highly viscous, but still flowable, materials. When the polymer used is itself flowable, the polymer composition of the invention, even when viscous, need not include a biocompatible solvent to be flowable, although trace or residual amounts of biocompatible solvents may still be present. The degree of viscosity of the polymer can be adjusted by the molecular weight of the polymer, as well as by mixing the cis- and trans-isomers of the cyclohexane

dimethanol in the backbone of the polymer.

[0103] The polymer composition can be administered by a variety of routes. For example, if flowable, it can be injected to form, after injection, a temporary biomechanical barrier to coat or encapsulate internal organs or tissues. The polymer composition of the invention can also be used to produce coatings for solid implantable devices.

[0104] However, most importantly, the polymer composition provides controllable and effective release of the antineoplastic agent over time, even in the case of large bio-macromolecules.

Implants and Delivery Systems

[0105] In its simplest form, a biodegradable polymer delivery system consists of a solution or dispersion of an antineoplastic agent in a polymer matrix having an unstable (biodegradable) bond incorporated into the polymer backbone. In a particularly preferred embodiment, a solid article comprising the composition of the invention is to be inserted within the peritoneum by implantation, injection, laparoscopy or otherwise being placed within the peritoneum of the subject being treated, for example, during or after the surgical removal of visibly cancerous tissue.

[0106] The antineoplastic agent of the composition and the polymer may form a homogeneous matrix, or the biologically active substance may be encapsulated in some way within the polymer. For example, the biologically active substance may be first encapsulated in a microsphere and then combined with the polymer in such a way that at least a portion of the microsphere structure is maintained. Alternatively, the biologically active substance may be sufficiently immiscible in the polymer of the invention that it is dispersed as small droplets, rather than being dissolved, in the polymer.

[0107] As a structural medical device, the polymer compositions of the inventions provide a wide variety of physical forms having specific chemical, physical and mechanical properties suitable for insertion into the peritoneum, in addition to being a composition that degrades in vivo into non-toxic residues.

[0108] Biodegradable drug delivery articles can be prepared in several ways. The polymer can be melt processed using conventional extrusion or injection molding techniques, or these products can be prepared by dissolving in an appropriate solvent, followed by formation of the device, and subsequent removal of the solvent by evaporation or extraction, e.g., by spray drying. By these methods, the polymers may be formed into articles of almost any size or shape desired, for example, implantable solid discs or wafers or injectable rods, microspheres, or other microparticles. Typical medical articles also include such as implants as laminates for degradable fabric or coatings to be placed on other implant devices.

[0109] The antineoplastic agent is typically released from the polymeric matrix at least as quickly as the matrix biodegrades in vivo. With some antineoplastic agents, the agent will be released only after the polymer has been degraded to a point where a non-diffusing substance has been exposed to bodily fluids. As the polymer begins to degrade, the biologically active substance that was completely surrounded by the polymer matrix begins to be liberated.

[0110] However, with this mechanism, a long peptide chain that is physically entangled in a rigid solid implant structure may tend to degrade along with the matrix and break off from the remainder of the peptide chain, thereby releasing incomplete fragments of molecules. When the polymer compositions of the invention are designed to be flexible, however, the polymer will typically degrade after the peptide or protein has been released in part. In a particularly preferred mechanism, when a peptide chain is being released from the composition of the invention, the composition remains flexible and allows a large-molecule protein to, at least partially, diffuse through the polymeric matrix prior to its own or the polymer's biodegradation.

[0111] The initial release rate of proteins from the compositions is therefore generally diffusion-controlled through channels in the matrix structure, the rate of which is inversely proportional to the molecular weight of the protein. Once polymer degradation begins, however, the protein remaining in the matrix may also be released by the forces of erosion.

[0112] The biodegradable amorphous matrices typically contain polymer chains that are associated with other chains. These associations can be created by a simple entanglement of polymer chains within the matrix, as opposed to hydrogen bonding or Van der Vaals interactions or between crystalline regions of the polymer or interactions that are ionic in nature. Alternatively, the synthesis of block copolymers or the blending of two different polymers can be used to create viscous, "putty-like" materials with a wide variation in physical and mechanical properties.

[0113] In a particularly preferred embodiment, the composition is sufficiently flowable to be injected into the body. It is particularly important that the injected composition result in minimal tissue irritation after injection or otherwise being inserted into the peritoneal cavity.

[0114] In one embodiment, the polymer composition of the invention is used to form a soft, drug-delivery "depot" that can be administered as a liquid, for example, by injection, but which remains sufficiently viscous to maintain the drug within the localized area around the injection site. The degradation time of the depot so formed can be varied from several days to a year or more, depending upon the polymer selected and its molecular weight. By using a polymer composition in flowable form, even the need to make an incision can be eliminated. In any event, the flexible or flowable delivery "depot" will adjust to the shape of the space it occupies within the body with a minimum of trauma to surrounding

tissues.

[0115] When the polymer composition is flexible or flowable, it can be placed anywhere within the body, including a cavity such as the peritoneum, sprayed onto or poured into open wounds, or used as a site delivery system during surgery. When flowable, the composition of the invention can also be used to act as a temporary barrier in preventing the direct adhesion of different types of tissue to each other, for example, after abdominal surgery, due to its ability to encapsulate tissues, organs and prosthetic devices.

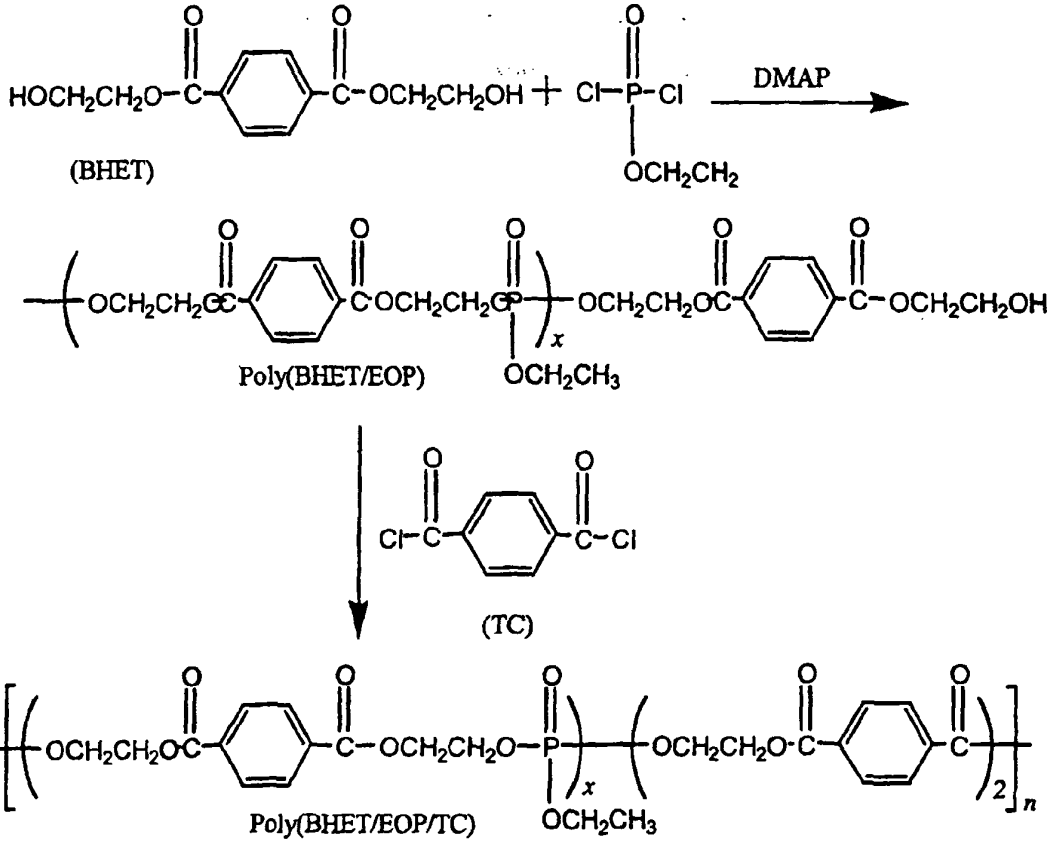
[0116] Once inserted, the polymer composition should remain in at least partial contact with a biological fluid, such as blood, internal organ secretions, mucous membranes, and the like. The implanted or injected composition will release the antineoplastic agent contained within its matrix at a controlled rate until the substance is depleted, following the general rules for diffusion or dissolution from a rigid, flexible or flowable biodegradable polymeric matrix.

[0117] The following examples are illustrative of preferred embodiments of the invention and are not to be construed as limiting the invention thereto. All polymer molecular weights are average molecular weights. All percentages are based on the percent by weight of the final delivery system or formulation being prepared, unless otherwise indicated, and all totals equal 100% by weight.

EXAMPLES

Example 1: Synthesis of Copolymer P (BHET-EOP/TC, 80/20)

[0118]



[0119] Under an argon stream, 10 g of 1,4-bis(hydroxyethyl) terephthalate (BHET), 9.61 g of 4-dimethylaminopyridine (DMAP), and 70 mL of methylene chloride were placed in a 250 mL flask equipped with a funnel. The solution in the flask was cooled down to -40°C with stirring, and a solution of 5.13 g of ethyl phosphorodichloridate (EOP) (distilled before use) in 20 mL of methylene chloride was added dropwise through the funnel. After addition was complete, the mixture was stirred at room temperature for four hours to form the homopolymer BHET-EOP.

[0120] A solution of 1.60 g of terephthaloyl chloride (TC) (obtained from Aldrich Chemical Company and recrystallized with hexane before use) in 20 mL of methylene chloride was then added drop by drop. The temperature was brought up to about 45-50°C gradually, and the reaction mixture was kept refluxing overnight to complete the copolymerization

of the homopolymer P(BHET-EOP) with the additional monomer TC to form the copolymer P(HHET-EOP/TC).

[0121] The solvent was then evaporated, and the residue was redissolved in about 100-200 mL of chloroform. The chloroform solution was washed with a saturated NaCl solution three times, dried over anhydrous Na_2SO_4, and quenched into ether. The resulting precipitate was redissolved in chloroform and quenched again into ether. The resulting tough, off-white solid precipitate was filtered off and dried under vacuum. Yield 82%.

[0122] The structure of P(BHET-EOP/TC, 80/20) was ascertained by ^{1}H-NMR, ^{31}p -NMR and FT-IR spectra, as shown in Figures 1 and 2. The structure was also confirmed by elemental analysis, which correlated closely with theoretical ratios. The results of the elemental analysis are shown in Figure 3.

[0123] The molecular weight of P(BHET-EOP/TC, 80/20) was first measured by gel permeation chromatography (GPC) with polystyrene as the calibration standard. The resulting graph established a weight average molecular weight (Mw) of about 6100 and a number average molecular weight (Mn) of about 2200, as shown in Figure 4. Vapor pressure osmometry ("VPO") for this copolymer gave an Mn value of about 7900. The results of these molecular weight studies are also shown in Figure 3.

Example 2: Feed Ratio Variations of P(BHET-EOP/TC)

[0124] A series of other P(BHET-EOP/TC) copolymers of the invention were prepared by following the procedure described above in Example 1 except that the feed ratio of the EOP to TC used during the initial polymerization step and copolymerization step respectively was varied. The results are shown below in Table 1. From the feed ratio of EOP/TC, the value of "x" from the formula shown below can be calculated. For example, in P(BHET-EOP/TC, 80/20) prepared above in Example 1, x is 8.

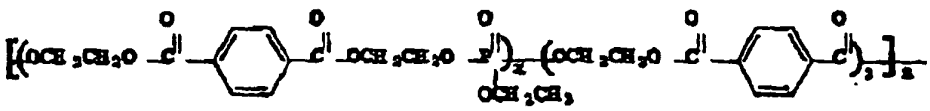


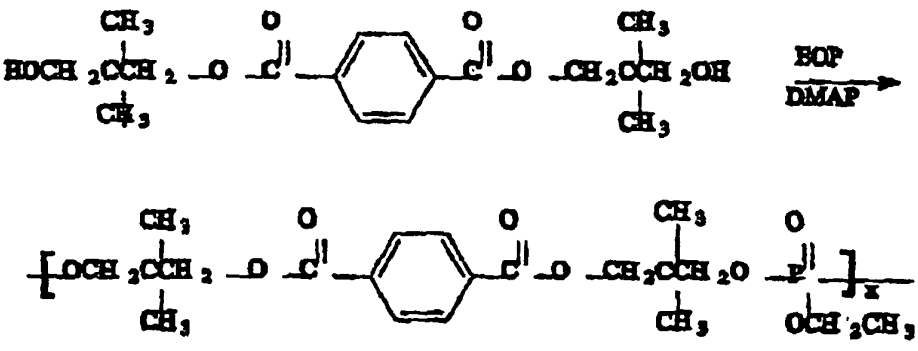
Table 1

Variation of Feed Ratio of EOP to TC in P(BHET-EOP/TC)						
Feed Ratio of EOP/TC*	100/0	95/5	90/10	85/15	80/20	50:50
"x"	-	38	18	11.4	8	2

*Feed ratio of ethyl phosphorodichloridate to terephthaloyl chloride.

Example 3: Synthesis and Isolation of the Homonolymer P(BHDPT-EOP)

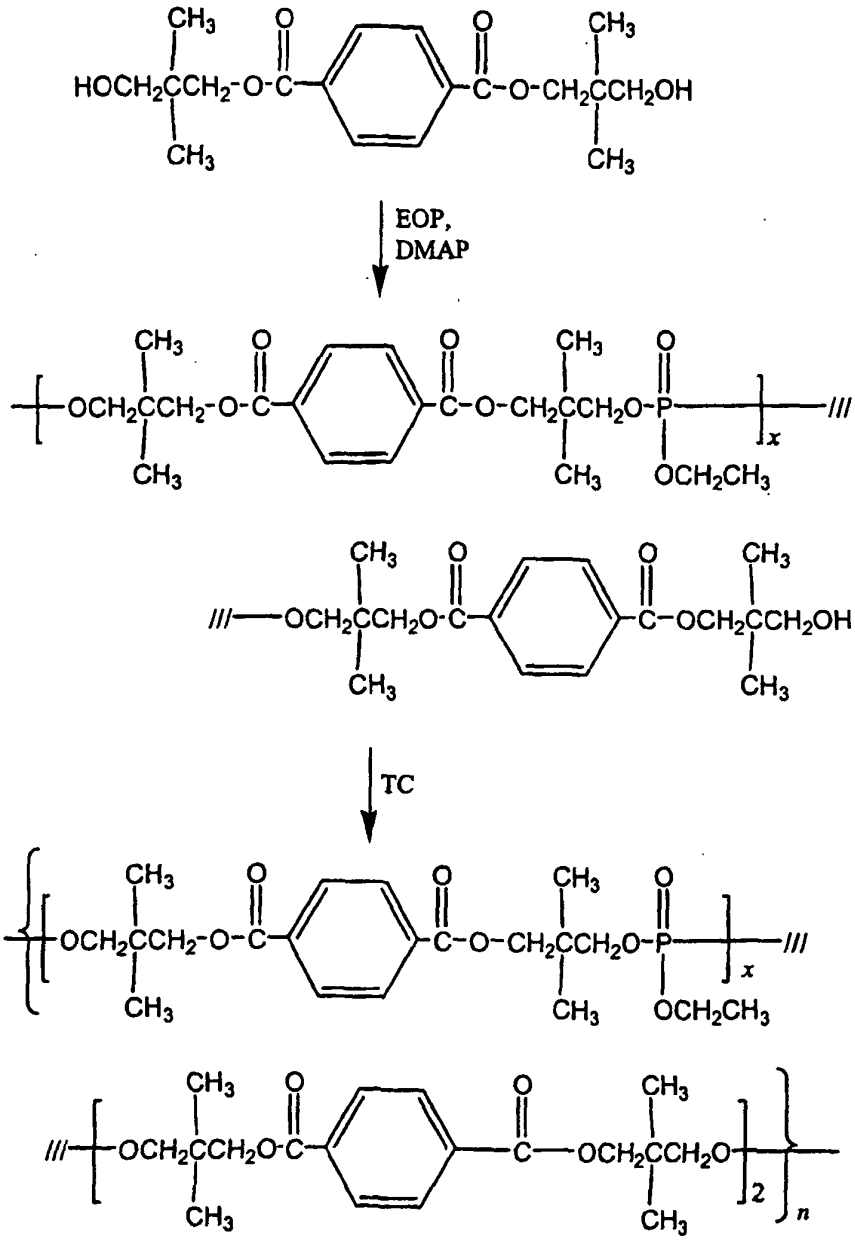
[0125]



[0126] The BHDPT monomer and the acid acceptor 4-dimethylaminopyridine (DMAP) were dissolved in methylene chloride. The resulting solution was chilled to -70 °C using a dry ice/acetone bath, and an equal molar amount of ethyl phosphorodichloridate (EOP) was slowly added. The reaction mixture was then heated and refluxed overnight. The salt formed in the polymerization was removed by filtration. The remaining polymer solution (filtrate) was washed with a saturated NaCl solution three times, and the homopolymer was precipitated in diethyl ether.

Example 4: Synthesis of Copolymer P(BHDPT-EOP/TC)

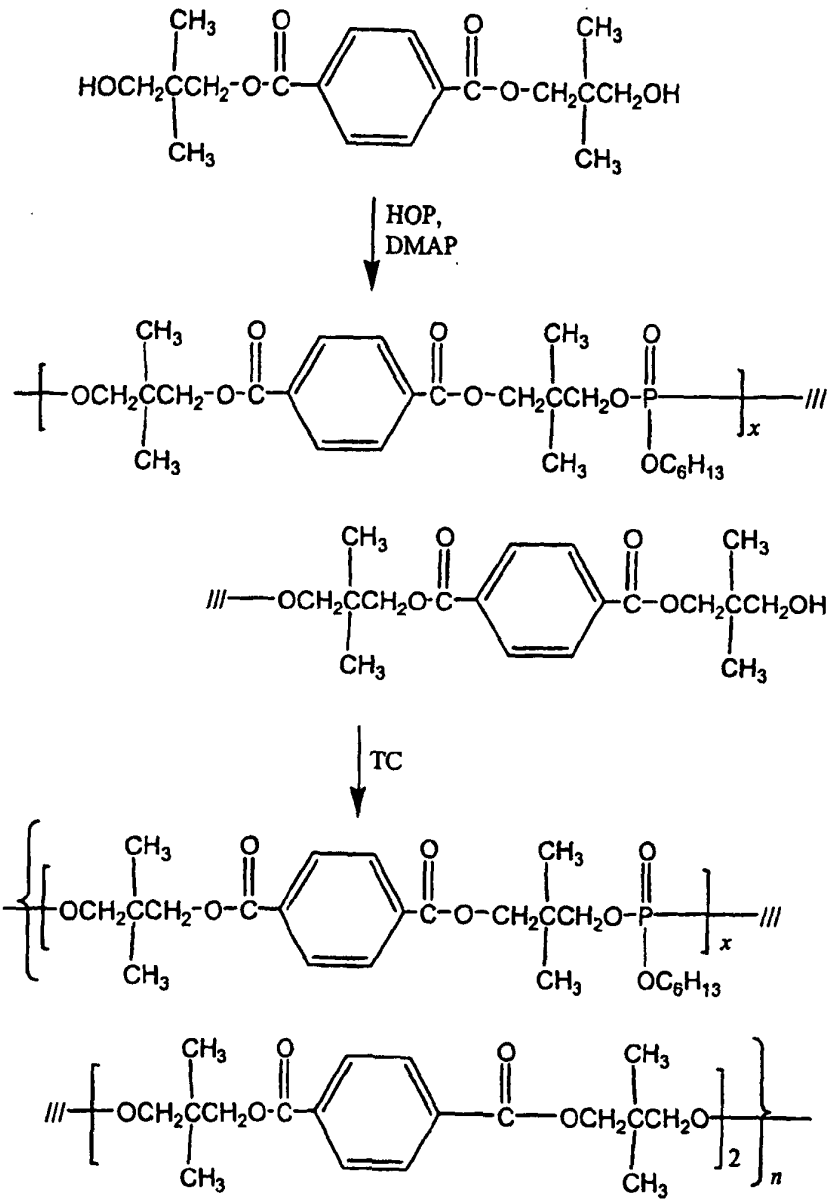
[0127]



[0128] Copolymers of P(BHDPT-EOP) with TC were synthesized by the two-step solution copolymerization shown above. After the reaction between BHDPT and EOP had proceeded at room temperature for one hour, the reaction flask was cooled in a dry ice/acetone bath. An appropriate amount of TC (the number of moles of TC and EOP combined equaled the number of moles of BHDPT) was slowly added to the flask. The reaction mixture was then heated and refluxed overnight. The salt formed in the polymerization was removed by filtration. The remaining copolymer solution (filtrate) was washed with a saturated NaCl solution three times, and the copolymer was precipitated out in diethyl ether.

Example 5: Synthesis of Poly(phosphoester) P(BHDPT-HOP/TC)

[0129]

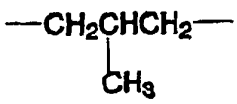


[0130] Copolymers of P(BHDPT-HOP) with TC were synthesized by a two-step solution polymerization. After the reaction between BHDPT and HOP had proceeded at room temperature for one hour, the reaction flask was cooled in a dry ice/acetone bath. An appropriate amount of TC (the number of moles of TC and HOP combined equaled the number of moles of BHDPT) was slowly added to the flask. The reaction mixture was then heated and refluxed overnight. The salt formed during the copolymerization was removed by filtration. The remaining copolymer solution (filtrate) was washed with a saturated NaCl solution three times, and the copolymer was precipitated out in diethyl ether.

Example 6: Other Diol Variations

[0131] Diol terephthalates that are structurally related to that of BHET and BHDPT were synthesized by reacting TC with either n-propylenediol or 2-methylpropylenediol, the structures of which are shown below, to form the corresponding diol terephthalate.

$$—CH_2CH_2CH_2—$$



These diol terephthalates were then reacted with EOP to form the corresponding homopolymers. The homopolymers so formed were then used to produce the copolymers of the invention in a second reaction with TC, as described above in Example 4.

Example 7: Glass Transition Temperatures for P(BHET-EOP/TC) copolymers

[0132] By differential scanning calorimetry (DSC), the glass transition temperatures (Tg's) of P(BHET-EOP/TC, 80/20) and P(BHET-EOP/TC, 50/50) were determined to be 24.5°C and 62.2°C respectively. Figure 4 shows the DSC curves for these two polymers. The Tg's of four additional P(BHET-EOP/TC) copolymers of differing EOP/TC feed ratios were determined, and the results were tabulated, as shown below in Table 2:

Table 2

Glass Transition Temperatures (Tg's) of (BHET-EOP/TC) Polymers						
Ratio of EOP/TC*	100/0	95/5	90/10	85/15	80/20	50:50
Tg (°C)	19.1	20.7	21.2	29.8	24.5	62.2

*Feed ratio of ethyl phosphorodichloridate to terephthaloyl chloride

The Tg increased as the proportion of EOP decreased and the proportion of TC increased.

Example 8: Glass Transition Temperatures for P(HHDPT-EOP/TC) Copolymers

[0133] A study of the influence of an increasing proportion of terephthaloyl chloride (TC) on the Tg's of P(BHDPT-EOP/TC)polymers was also conducted. The results are shown below in Table 3.

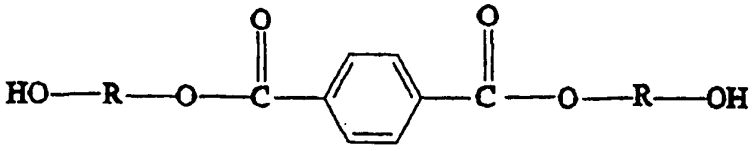
Table 3

Influence of EOP/TC Ratio on the Tg of P(BHDPT-EOP/TC)	
Molar ratio (BHDPT/EOP/TC) *	Tg (°C)
100:100:0	14
100:100:0	19
100:90:10	16
100:85:15	24
100:80:20	23
100:75:25	33
100:75:25	49
100:50:50	43

*The total molar amount of TC and EOP equaled the molar amount of BHDPT.

Example 9: Glass Transition Temperatures for Various R Groups

[0134] A study was also conducted showing the effect on glass transition temperature (Tg) for copolymers made from the following series of diols having varying R groups:

II 

where R is $-CH_2CH_2-$; $-CH_2CH_2CH_2-$; $-CH_2CH(CH_3)CH_2-$; and $-CH_2CH(CH_3)_2CH_2-$. The results are shown below in Table 4:

Table 4
Influence of the Changing "R" Group on Tg of Polymer

"R" Group	Structure	Tg (°C)
ethylene	$-CH_2CH_2-$	14-19
n-propylene	$-CH_2CH_2CH_2-$	-15
2-methyl-propylene	$-CH_2CH(CH_3)CH_2-$	11
2,2'-dimethyl-propylene	$-CH_2C(CH_3)_2CH_2-$	19

As shown in Table 4, the Tg increased as the size and the degree of branching of the R group increased. In addition, the polymers changed in physical state as the Tg changed. Specifically, as Tg increased, the polymers changed from rubbery to fine powders.

Example 10: Solubilities of the Polymers of the Invention

[0135] The solubility in organic solvents was determined for the homopolymer P(BHET-EOP, 100/0) and for the following block copolymers:

P(BHET-EOP/TC, 95/5),
P (BHET-EOP/TC, 90/10),
P(BHET-EOP/TC, 85/15),
P (HHET-EOP/TC, 80/20)-, and
P(BHET-EOP/TC, 50/50).

The organic solvents used for the test were chloroform, methylene chloride, N-methylpyrrolidone (NMP), dimethylfor-

mamide (DMF) and dimethylsulfoxide (DMSO). The results of these solubility tests are summarized below in Table 5.

TABLE 5

Polymer	$CHCl_3$	CH_2Cl_2	NMP	DMF	DMSO
P(BHET-EOP, 100/0)	Easily soluble	Easily soluble	Good solubility	Good solubility	Good solubility
P(BHET-EOP/TC, 95/5)	Easily soluble	Easily soluble	Good solubility	Good solubility	Good solubility
P(BHET-EOP/TC, 90/10)	Easily soluble	Easily soluble	Good solubility	Good solubility	Good solubility
P(BHET-EOP/TC, 85/15)	Relatively soluble	Relatively soluble	Good solubility	Good solubility	Good solubility
P(BHET-EOP/TC, 80/20)	Relatively soluble	Relatively soluble	Good solubility	Good solubility	Good solubility
P(BHET-EOP/TC, 50/50)	Not soluble	Not soluble	Soluble with heating	Soluble with heating	Soluble with heating

The results showed that the solubility of these polymers in organic solvents increased as the EOP/TC ratio increased.

Example 11: Viscosities of the Polymers

[0136] The intrinsic viscosities of a series of P(BHET-EOP/TC) polymers of varying feed ratios were measured in chloroform (CH_3Cl) at 40°C in a Ubbelohde viscometer. The results are shown below in Table 6.

Table 6

Intrinsic Viscosities of P(BHET-EOP/TC) Polymers						
Ratio of EOP/TC*	100/0	95/5	90/10	85/15	80/20	50: 50
[η] (dL/g)	.081	.089	.148	.146	0.180	N.D. †

*Feed ratio of ethyl phosphorodichloridate to terephthaloyl chloride.

†The intrinsic viscosity of P(BHET-EOP/TC, 50/50) was not determined because it was not soluble in chloroform.

Example 12: In vitro Degradation

[0137] Films of P(BHET-EOP/TC, 80/20) and P(BHET-EOP/TC, 85/15) were made by solution casting methods and were dried under vacuum for two days. Discs 1 mm in thickness and 6 mm in diameter were cut from these film sheets. Three discs of each copolymer were placed in 4 mL of phosphate buffer saline (PBS) (0.1M, pH 7.4) at 37°C. The discs were taken out of the PBS at different points in time, washed with distilled water, and dried overnight.

[0138] The samples were analyzed for change in molecular weight and weight loss over time, as shown in Figures 7A and 7B. The weight average molecular weight of P(BHET-EOP/TC, 80/20) decreased about 20% in three days. After 18 days, the P(BHET-EOP/TC, 85/15) and P(BHET-EOP/TC, 80/20) discs had lost about 40% and 20% in mass respectively.

[0139] This data demonstrated the feasibility of fine-tuning the degradation rate of the copolymers and confirmed that the copolymers became more hydrolytically labile as the phosphate component (EOP) was increased.

[0140] The same process was repeated for the P(BHDPT-EOP) copolymers having different feed ratios of EOP to TC. Figure 6 is a graphic representation of the degree of degradation, as measured by change in molecular weight, over time for the homopolymer P(BHDPT-EOP) and the following block copolymers:

P(BHDPT-EOP/TC, 85/15),
P(BHDPT-EOP/TC, 75/25), and
P (HHDPT-EOP/TC, 50/50).

Example 13: In vivo Degradation of P(BHET-EOP/TC) Copolymer and Paclitaxel Release in vitro

[0141] Figures 7A and 7B shows the in vivo degradation of P(BHET-EOP/TC, 80/20), as measured by weight loss. Figure 7C shows paclitaxel release from film in vitro.

Example 14: In vitro Biocompatability/Cytotoxicity of P(BHET-EOP/TC, 80/20)

[0142] The cytotoxicity of P(BHET-EOP/TC, 80/20) copolymer was assessed by culturing human embryonic kidney (HEK) cells on a cover slip that had been coated with the copolymer. P (BHET-EOP/TC, 80/20). As a control, HEK cells were also cultured on a coverslip coated with TCPS. The cells cultured on the copolymer-coated cover slip exhibited normal morphology at all times and proliferated significantly in 72 days, as compared to a considerably lower amount when identical HEK cells were cultured on TCPS.

Example 15: In vivo Biocompatibility of P(BHET-EOP/TC. 80/20)

[0143] A 100 mg polymer wafer was formed from P(BHET-EOP/TC, 80/20) and, as a reference, a copolymer of lactic and glycolic acid (75/25, "PLGA") known to be biocompatible. These wafers were inserted between muscle layers of the right limb of adult SPF Sprague-Dawley rats under anesthesia. The wafers were retrieved at specific times, and the surrounding tissues were prepared for histopathological analysis by a certified pathologist using the following scoring:

Score	Level of Irritation
0	No Irritation
0 - 200	Slight Irritation
200 - 400	Mild Irritation
400 - 600	Moderate Irritation
More than 600	Severe Irritation

The results of the histopathological analysis are shown below in Table 7.

Table 7

Inflammatory Response at Site of Implantation (i.m.)						
Polymer	3 Days	7 Days	14 Days	1 Mo.	2 Mos.	3 Mos.
P(BHET-EOP/TC, 80/20)	151	116	163	98	60	35
PLGA (75/25)	148	98	137	105	94	43

The phosphoester copolymer P(BHET-EOP/TC, 80/20) was shown to have an acceptable biocompatability similar to that exhibited by the PLGA reference wafer.

Example 16: Preparation of P(BHET-EOP/TC, 80/20) Microspheres Encapsulating FITC-BSA

[0144] Microspheres were prepared via a double-emulsion/solvent-extraction method using FITC-labeled bovine serum albumin (FITC-BSA) as a model protein drug. One hundred μL of an FITC-BSA solution (10 mg/mL) were added to a solution of 100 mg of P(BHET-EOP/TC, 80/20) in 1 mL of methylene chloride, and emulsified via sonication for 15 seconds on ice. The resulting emulsion was immediately poured into 5 mL of a vortexing aqueous solution of 1% polyvinyl alcohol (PVA) and 5% NaCl. The vortexing was maintained for one minute. The resulting emulsion was poured into 20 mL of an aqueous solution of 0.3% PVA and 5% NaCl, which was being stirred vigorously. Twenty-five mL of a 2% isopropanol solution was added, and the mixture was kept stirring for one hour to ensure complete extraction. The resulting microspheres were collected via centrifugation at 3000 X g, washed three times with water, and lyophilized. Empty microspheres were prepared in the same way except that water was used as the inner aqueous phase.

[0145] These preparation conditions had been optimized for increased encapsulation efficiency, improved microsphere morphology, and minimal burst release. The resulting microspheres were mostly between 5 and 20 μm in diameter and exhibited a smooth surface morphology. Figure 8 shows the size and smoothness of the microspheres, as demonstrated by electron microscopy.

[0146] The loading level of FITC-BSA was determined by assaying for FITC after hydrolyzing the microspheres in a

0.5 N NaOH solution overnight. Loading levels were determined by comparison with a standard curve, which had been generated by making a series of FITC-BSA solutions in 0.5 N NaOH. Protein loading levels of 1.5, 14.1 and 22.8 wt. % were readily obtained.

[0147] The encapsulation efficiency of FITC-BSA by the microspheres was determined at different loading levels by comparing the quantity of FITC-BSA entrapped with the initial amount in solution via fluorometry. As shown below in Table 8, encapsulation efficiencies of 84.6 and 99.6% were obtained. These results showed that encapsulation efficiencies of 70-90% would be readily obtainable.

Table 8

Encapsulation Efficiency and Loading Level of FITC-BSA in P(BHET-EOP/TC, 80/20)		
Loading (%)	High Loading (22.8%)	Low Loading (1.5%)
Encapsulation Efficiency (%)	99.6	84.6

In addition, it was determined by observation with confocal fluorescence microscopy that the encapsulated FITC-BSA was distributed uniformly within the microspheres.

Example 17: Preparation of P(BHDPT-EOP/TC, 50/50) Microspheres Containing Lidocaine

[0148] An aqueous solution of 0.5% w/v polyvinyl alcohol (PVA) was prepared in a 600 mL beaker by combining 1.35 g of PVA with 270 mL of deionized water. The solution was stirred for one hour and filtered. A copolymer/drug solution was prepared by combining 900 mg of P(BHDPT-EOP/TC, 50/50) copolymer and 100 mg of lidocaine in 9 mL of methylene chloride and vortex-mixing.

[0149] While the PVA solution was being stirred at 800 rpm with an overhead mixer, the polymer/drug mixture was added dropwise. The combination was stirred for one and a half hours. The microspheres thus formed were then filtered, washed with deionized water, and lyophilized overnight. The experiment yielded 625 mg of microspheres loaded with 3.7% w/w lidocaine.

[0150] Lidocaine-containing microspheres were also prepared from P(BHDPT-HOP/TC, 50/50) by the same process. This experiment yielded 676 mg of microspheres loaded with 5.3% w/w lidocaine.

Example 18: In vitro Release Kinetics of Microepheres Prepared from P(BHET-EOP/TC, 80/20) Copolymers

[0151] Five mg of P(BHBT-EOP/TC, 80/20) microspheres containing FITC-BSA were suspended in one mL of phosphate buffer saline (PBS) at pH 7.4 and placed into a shaker heated to a temperature of 37°C. At various points in time, the suspension was spun..at 3000.X g for 10 minutes, and 500 μl samples of the supernatant fluid were withdrawn and replaced with fresh PBS. The release of FITC-BSA from the microspheres was followed by measuring the fluorescence intensity of the withdrawn samples at 519 nm.

[0152] Scaling up, 50 mg of P(BHET-EOP/TC, 80/20) microspheres were suspended in vials containing 10 mL of phosphate buffer saline (PBS). The vials were heated in an incubator to a temperature of 37°C and shaken at 220 rpm. Samples of the supernatant were withdrawn and replaced at various points in time, and the amount of FITC-BSA released into the samples was analyzed by spectrophotometry at 492 nm.

[0153] The results indicated that over 80% of the encapsulated FITC-BSA was.released within the first two days, with an additional amount of about 5% being released after 10 days in PBS at 37°C. The release kinetics of FITC-BSA from P(BHET-EOP/TC, 80/20) microspheres at different loading levels are shown in Figure 9A.

Example 19: In vitro Release Kinetics of Microspheres Prepared from P(BHDPTEOP/TC. 50/50) Copolymers

[0154] Approximately 10 mg of P(BHDPT-EOP/TC, 50/50) microspheres loaded with lidocaine were placed in PBS (0.1 M, pH 7.4) at 37°C on a shaker. Samples of the incubation solution were withdrawn periodically, and the amount of lidocaine released into the samples was assayed by HPLC. Figure 10 shows the resulting release kinetics.

[0155] The same process was followed for microspheres prepared from P(BHDPT-HOP/TC, 50/50). Figure 10 also shows the release kinetics of lidocaine from these microspheres.

Example 20: In vitro Cytotoxicity Assay of Copolymer on Cells

[0156] P(BHET-EOP/TC, 80/20) microspheres were added to 96-well tissue culture plates at different concentrations. The wells were then seeded with human gastric carcinoma cells (GT3TKB) at a density of 10^4 cells/well. The cells

were incubated with the microspheres for 48 hours at 37°C. The resulting cell proliferation rate was analyzed by MTT assay and plotted as % relative growth vs. concentration of copolymer microspheres in the tissue culture well. The results are shown in Figure 13.

Example 21: Toxicity Assay of Polymer-Degradation products on GT3TKB Tumor Cells

[0157] About 100-150 mg of each of the following polymers were degraded separately in 20 mL of 1M NaOH at 37°C for 1-2 days:

PLLA (Mw = 14,000)
P(BHET-EOP)
PCPP:SA (20:80)
Poly(L-lysine) (Mw = 88,000)

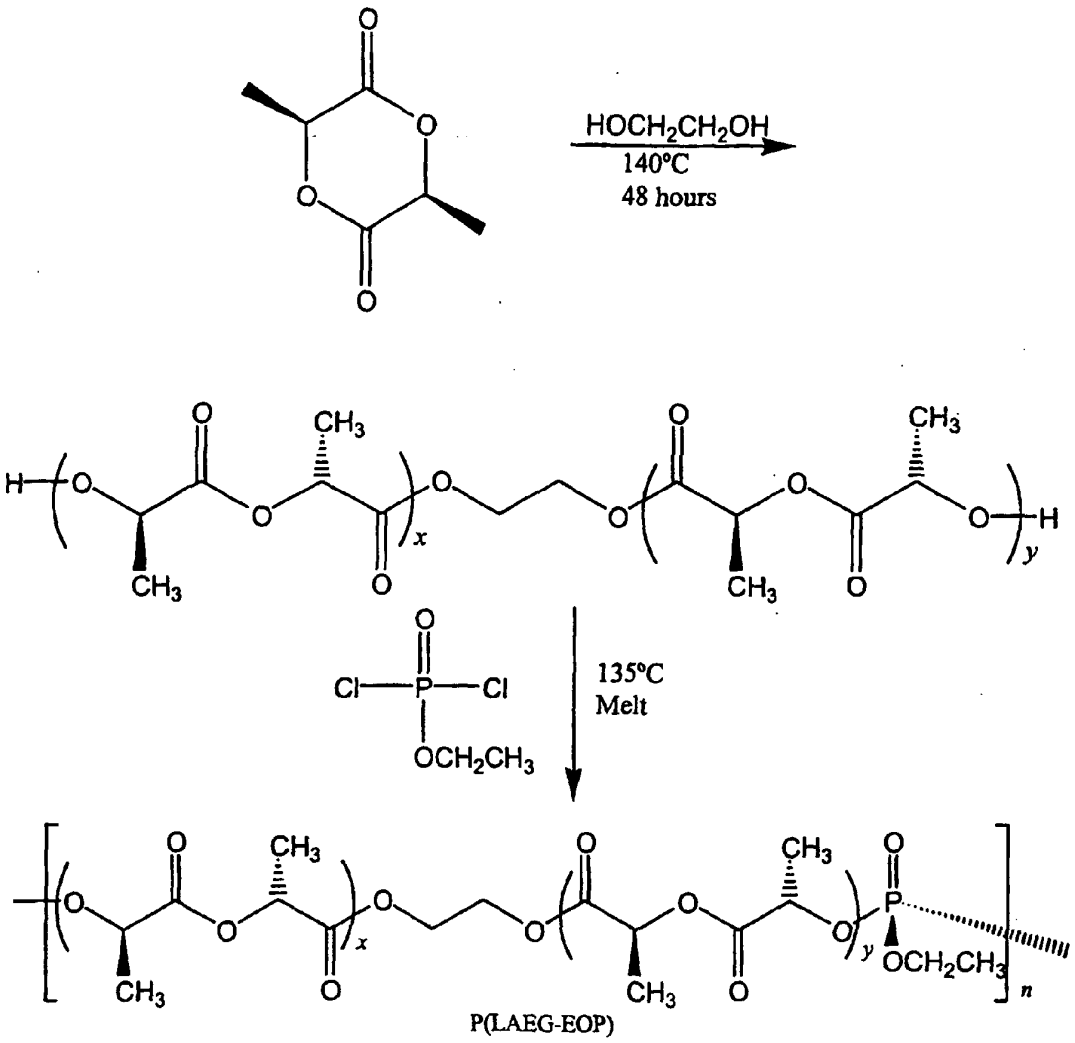
Complete degradation was observed for all of the polymers. The solution was then neutralized with 20 mL of 1M HCl.

[0158] About 200 μL of various concentrations of the degraded polymer products were placed in 96-well tissue culture plates and seeded with human gastric carcinoma cells (GT3TKB) at a density of 10^4/well. The degraded polymer products were incubated with the GT3TKB cells for 48 hours. The results of the assay were plotted as % relative growth vs. concentration of degraded polymer in the tissue-culture well and are shown in Figure 13.

[0159] An additional toxicity assay was conducted with microspheres prepared from the monomer BHET and from the homopolymer BHET-EOP, and compared with microspheres prepared from LA and PLLA. The results of the assay were plotted as % relative growth vs. concentration of the polymers or microspheres in a tissue-culture cell and are shown in Figure 12.

Example 22: Synthesis of Poly(L-lactide-co-ethylphosphate) [Poly (LAEG-EOP)]

[0160]



[0161] 20 g (0.139 mole of (3S)-cis-3,6-dimethyl-1,4-dioxane-2,5-dione (L-lactide) (obtained from Aldrich Chemical Company, recrystallized with ethyl acetate, sublimed, and recrystallized with ethyl acetate again) and 0.432g (6.94 mmole) of ethylene glycol (99.8%, anhydrous, from Aldrich) were placed in a 250 mL roundbottomed flask flushed with dried argon. The flask was closed under vacuum and placed in an oven heated to 140°C. The flask was kept at this temperature for about 48 hours with occasional shaking.

[0162] The flask was then filled with dried argon and placed in oil bath heated to 135°C. Under an argon stream, 1.13 g of ethyl phosphorodichloridate was added with stirring. After one hour of stirring, a low vacuum (about 20mm Hg) was applied to the system, and it was allowed to stand overnight. One hour before work-up, a high vacuum was applied. After cooling, the polymer was dissolved in 200 mL of chloroform and quenched into one liter of ether twice to an off-white precipitate, which was dried under vacuum.

[0163] It was confirmed by NMR spectroscopy that the polymer obtained was the desired product, poly(L-lactide-co-ethyl-phosphate) [P(LAEG-EOP)], as shown in Figures 14 and 15.

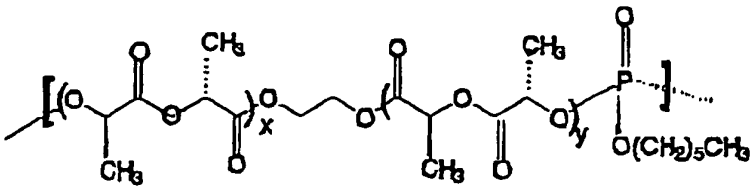
Example 23 : Properties of P (LAEC-EOP)

[0164] A P(LAEG-BOP) polymer where (x or y)/n = 10:1 was prepared as described above in Example 22. The resulting poly(phosphoester-co-ester) polymer was analyzed by GPC using polystyrene as a standard, and the resulting graph established an Mw of 33,000 and an Mn of 4800, as shown in Figure 17.

[0165] The viscosity was measured in chloroform (CH_3Cl) at 40°C and determined to be 0.315 dL/g. The polymer was. soluble in ethyl acetate, acetone, acetonitrile, chloroform, dichloromethane, tetrahydrofuran, N-methylpyrrolidone, dimethylformamide, and dimethyl sulfoxide. The polymer formed a brittle film, and the Tg was determined by DSC to be 51.5°C, as shown in Figures 16A and 16B.

Example 24: Synthesis of Poly(L-lactide-co-hexylphosphate) [Poly(LAEG-HOP)]

[0166] A second poly(L-lactide-phosphate) having the following structure:



was also prepared by the method described in Example 22, except that hexyl phosphorodichloridate ("HOP") was substituted for EOP (ethyl phosphorodichloridate).

Example 25: Properties of P (LAEG-EOP) and P(LAEG-HOP)

[0167] The weight-average molecular weight (Mw) of the phosphoester-co-ester polymer of Example 22, P (LAEG-EOP), and the polymer of Example 24, P(LAEG-HOP), were first determined by gel permeation chromatography (GPC) with polystyrene as the calibration standard, as shown in Figure 18. Samples of each were then allowed to remain exposed to room temperature air to test for ambient, unprotected storage capability. After one month, the Mw was again determined for each polymer. The results (plotted in Figure 18) showed that, while the Mw for p(LAEG-EOP) was reduced by about one-third after a month of unprotected ambient conditions, the Mw for p(LAEG-HOP) remained fairly constant, even showing a slight increase. See also Figure 19.

[0168] Discs for degradation studies were then fabricated from each polymer by compression molding at 50°C and a pressure of 200 MPa. The discs were 4 mm in diameter. 1.5 mm in thickness, and 40 mg in weight. The degradation studies were conducted by placing the discs in 4 mL of 0.1M PBS (pH 7.4) at 37°C. Duplicate samples were removed at different time points up to eight days, washed with distilled water, and dried under vacuum overnight. Samples were analyzed for weight loss and molecular weight change (GPC), and the results are shown in Figures 20A, 20B, 21A and 21B. Both polymers, P(LAEG-EOP) and P(LAEG-HOP), demonstrated favorable degradation profiles.

Example 26: In vivo Biocompatibility of P(LAEG-EOP)

[0169] A 100 mg polymer wafer was formed) from P(LAEG-EOP) and, as a reference, a copolymer of lactic and glycolic acid ["PLGA (RG755) "] known to be biocompatible. These wafers were inserted between muscle layers of the right limb.of adult SPP Sprague-Dawley rats under anesthesia. The wafers were retrieved at specific times, and the surrounding tissues were prepared for histopathological analysis by a certified pathologist using the following scoring:

Score	Level of Irritation
0	No irritation
0 - 200	Slight Irritation
200 - 400	Mild Irrigation
400 - 600	Moderate Irritation
More than 600	Severe Irritation

The results of the histopathological analysis are shown below in Table 9.

Table 9

Inflammatory Response at Site of Implantation (i.m.)						
Polymer	3 Days	7 Days	14 Days	1 Mo.	2 Mos.	3 Mos.
P(LAEG-EOP)	130	123	180	198	106	99
PLGA. (RG755)	148	98	137	105	94	43

See also Figure 22. The phosphoester copolymer P(LAEG-EOP) was shown to have an acceptable biocompatability similar to that exhibited by the PLGA reference wafer.

[0170] Similar tests were done after intramuscular injection of microspheres into male S-D rats, tabulating implant site macrophage counts, as well as irritation scores, as shown below:

	3 Day		7 Day		14 Day		31 Day	
Polymer	#	Irritation	#	Irritation	#	Irritation	#	Irritation
p(BHET-EOP/TC) 80/20	247	Mild	298	Mild	196	S1.	32	S1.
p (HHET-EOP/TC) 82.5/ 17.5	445	Mod.	498	Mod.	406	Mod.	38	S1.
p(BHET-EOP/TC) 85/15	161	S1.	374	Mild	586*	Mod.	274	Mild
p(CHDM-HOP)	399	Mild	169	S1.	762	Sev.	607	Sev.
p(BHET-EOP/TC) 90/10	206	Mild	476	Mod.	557	Mod.	72	S1.
P(DAPG-EOP) 1:10	360	Mild	323	Mild	569	Mod.	96	S1.
PLGA (RG755)	419	Mod.	331	Mod.	219	Mild	150	S1.
Control (no polymer)	219	Mild	-	-	-	-	-	-

= Mean count

* Only two animals present in this group.

[0171] still further tests were done after subcutaneous injection into male S-D rats, tabulating implant site macrophage counts, as well as irritation scores, as shown below:

	7 Day		14 Day		31 Day	
Group	#	Irrita tion	#	Irrita tion	#	Irrita tion
Vehicle only (0.7 ml) (n=3)	0	-	0	-	0	-
Acetic Acid (0.7 ml) (n=3)	208	Mild	166	S1.	20	S1.
p(dl) Lactic Acid (89 g/kg) (0.7ml) (n=3)	302	Mild	37	S1.	0	-
p(DAPG-HOP) (89 mg/kg) (0.7 ml) (n=6)	355	Mild	192'	S1.	101	S1.
p(CHDM-HOP) (89 mg/kg) (0.7 ml) (n=6)	652	Sev.	352	Mild	633	Sev.
P(BHET-EOP/TC) (89 mg/kg) (0.7 ml) (n=6)	325	Mild	423	Mod.	197	S1.
Vehicle (2.0 ml) (n=3)	65	S1.	0	-	0	-
Acetic Acid (2.0 ml) (n=3)	267	Mild	334	Mild	32	S1.
p(dl) Lactic Acid (267 g/kg) (2.0ml) (n=3)	85	S1.	18	S1.	279	Mild
p(DAPG-HOP) (267 mg/kg) (2.0 ml) (n=6)	386	Mild	273	Mild	279	Mild
p(CHDM-HOP) (267 mg/kg) (2.0 ml)(n=6)	471	Mod.	599	Mod.	618	Sev.
P(BHET-EOP/TC) (267 mg/kg) (2.0 ml) (n=6)	292	Mild	327	Mild	178	Sl.
# = Mean count						

Example 27: Preparation of Copolymer Microspheres Containing FITC-BSA with 10% Theoretical Loading Level

[0172] One hundred mL of FITC-BSA solution (100 mg/mL dissolved in water) was added to a solution of 100 mg of P(LAEG-EOP) in 1 mL of methylene chloride, and emulsified via sonication for 15 seconds on ice. The resulting emulsion was immediately poured into 5 mL of vortexing a 1% solution of polyvinyl alcohol (PVA) in 5% NaCl, and vortexing was maintained for one minute. The emulsion thus formed was then poured into 20 mL of a 0.3% PVA solution in 5% NaCl, which was being stirred vigorously. Twenty five mL of a 2% solution of isopropanol was added, and the mixture was kept stirring for one hour to ensure complete extraction. The resulting microspheres were collected via centrifugation at 3000 X g, washed 3 times with water, and freeze dried.

[0173] Different formulations of microspheres were made by using as the second aqueous phase a 5% NaCl solution or a 5% NaCl solution also containing 1% PEG 8000. Yet another technique was used in evaporating the solvent by stirring the mixture overnight, thus forming microspheres by solvent evaporation.

Example 28: Estimation of Encapsulation Efficiency and Loading Level

[0174] The loading level of FITC-BSA was determined by assaying for FITC after hydrolyzing the microspheres with 0.5 N NaOH overnight. The amount of FITC-BSA was compared with a standard curve that had been generated by making a series of FITC-BSA solutions in 0.5 N NaOH. The encapsulation efficiency of the microspheres was determined by comparing the quantity of FITC-BSA entrapped with the initial amount in solution via fluorometry. The encapsulation efficiency (%) and loading level (%) of FITC-BSA are shown in Table 10 below.

Table 10

Encapsulation Efficiency and Loading Level of FITC-BSA		
Loading (%)	High Loading (24.98%)	Low Loading (1.5%)
Encapsulation Efficiency (%)	98.10	91.70

Example 29: Cytotoxicity of the Copolymer

[0175] Microspheres containing P(LAEG-EOP) were added to 96-well tissue culture plates at different concentrations. Human gastric carcinoma cells (GT3TKB) were then seeded at a rate of 10^4 cells/well. The cells were then incubated with the microspheres in the wells for 48 hours at 37°C. The cell proliferation rate was analyzed by MTT assay, and the results were plotted as % relative growth vs. concentration of copolymer microspheres in the tissue culture well, as shown in Figure 23.

Example 30: Effect of fabrication method on the Release of FITC-BSA from Microspheres

[0176] Fifty mg of microspheres of a polymer of the invention were suspended in vials containing 10 mL of PBS, and the vials were shaken in an incubator at 37°C and at a rate of 220 rpm. The supernatant fluid was replaced at various time points, and the amount of FITC-BSA released was analyzed by spectrophotometry at 492 nm. The results were plotted as % cumulative release of FITC-BSA from the microspheres vs. time in hours, as shown in Figure 24A.

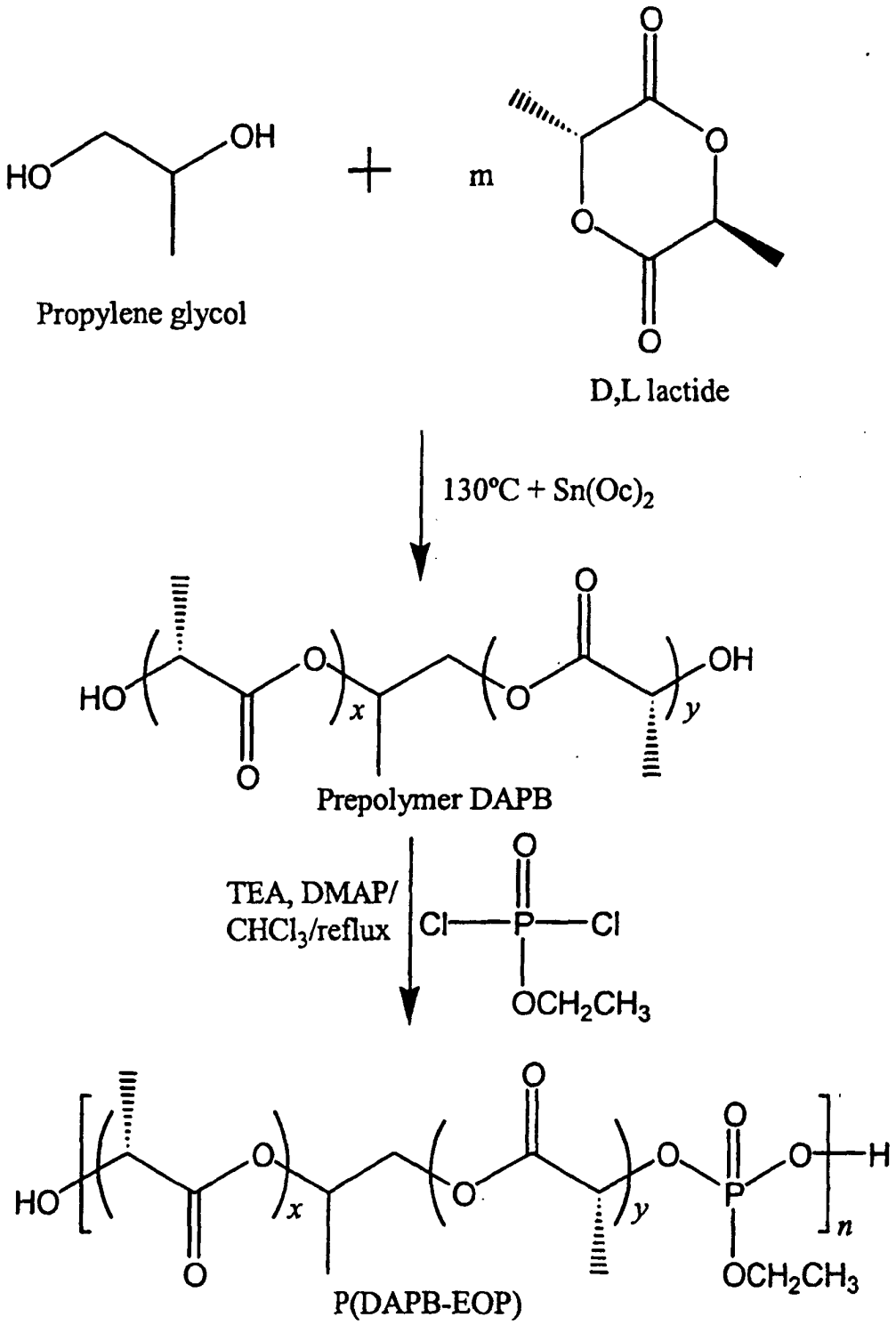
Example 31: Preparation of P(LAEG-EOP) Microspheres Containing Lidocaine Using Polyvinyl Alcohol as the Non-Solvent Phase

[0177] A solution of 0.5% w/v polyvinyl alcohol (PVA) in deionized water solution was prepared in a 600 mL beaker by combining 1.05 g of PVA with 210 mL of deionized water. The solution was stirred for one hour and filtered. A polymer/drug solution was prepared by combining 630 mg of polymer and 70 mg of lidocaine in 7 mL of methylene chloride and mixing by vortex. The PVA solution was mixed at 500 rpm with an overhead mixer, and the polymer/drug solution was added dropwise. - After 30 minutes of mixing, 200 mL of cold deionized water was added to the stirring PVA solution. The resulting mixture was stirred for a total of 3.5 hours. The microspheres formed were filtered off, washed with deionized water, and lyophilized overnight.

[0178] Microspheres loaded with 4.2% w/w lidocaine were thus obtained. Approximately 10 mg of microspheres were placed in a phosphate buffer saline (0.1M, pH 7.4) at 37°C on a shaker and sampled regularly. The results were plotted as % lidocaine released vs. time in days, as shown in Figure 24B.

Example 32: Synthesis of P(DAPG-EOP)

[0179] The d,l racemic mixture of poly (L-lactide-co-propyl-phosphate) ["P(DAPG-EOP)"] was prepared as follows:



The product was obtained as a white solid soluble in organic solvents. Depending on reaction conditions, different intrinsic viscosities and different molecular weights were obtained, as shown below in summary form:

Base(s)	Reaction Time/Temp	Eq $EOPCl_2$	Mw	IV
3.0 eq Reillex	18hrs/ reflux	1.05	--	0.06
3.0 eq Reillex	40hrs/ reflux	1.05	--	0.06
3.0 eq Reillex & 0.1% (w/w) DMAP	18hrs/ reflux	1.05	--	0.08
3.0 eq Reillex	18hrs/ reflux	1.00	--	0.06
2.5 eq TEA; 0.5 eq DMAP	15mins/ room temp.	1.05	--	0.42
2.5 eq TEA; 0.5 eq DMAP	18hrs/ reflux	1.05	--	0.27
2.5 eq TEA; 0.5 eq DMAP	about 2.5 days/ reflux	1.05	--	0.39
2.5 eq TEA; 0.1 eq DMAP	1 h/4°C; 2 h/room temp.	1.01	--	0.06
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.01	91,100	0.47
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.01	95,900 (Mn 44,200; Mw/Mn 2.2)	0.42
1.1 eq DMAP	1 h/4°C; 2 h/room temp.	1.01	--	0.08
1.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.01	--	0.23
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 17 h/room temp.	1.00	28,400	0.25
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.00	26,800 (Mn 12,900; Mw/Mn 2.1)	0.23
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.01	14,700	0.16
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.01 -	32,200 (Mn 13,000; Mw/Mn 2.5)	0.32
3.0 eq DMAP	1 h/4°C; 2 h/room temp.	1.00	--	0.20
2.5 eq TEA; 0.5 eq DMAP	1 h/4°C; 2 h/room temp.	1.00	--	0.22

Example 33: Preparation of P (DAEG-EOP) Microspheres With Lidocaine Using Silicon Oil as the Non-solvent Phase

[0180] Two percent sorbitan-trioleate, which is commercially available from Aldrich under the tradename Span-85, in silicon oil was prepared in a 400 mL beaker by combining 3 mL of Span-85 with 150 mL of silicone oil and mixing with an overhead stirrer set at 500 rpm. A d, l racemic mixture of poly(L-lactide-co-ethylphosphate) P(DAEG-EOP) polymer/drug solution was prepared by dissolving 400 mg of the polymer prepared above in Example 33, and 100 mg of lidocaine in 4.5 mL of methylene chloride. The resulting polymer/drug solution was added dropwise to the silicone oil/span mixture with stirring. The mixture was stirred for an hour and 15 minutes. The microspheres thus formed were filtered off and washed with petroleum ether to remove the silicone oil/span mixture, and lyophilized overnight.

[0181] 450 mg of microspheres loaded with 7.6% w/w lidocaine were thus obtained. Approximately 10 mg of microspheres were placed in phosphate buffer saline (0.1M, pH 7.4) at 37°C on a shaker and sampled regularly. The results were plotted as % lidocaine released vs. time in days.

[0182] Similar data for P(DAPG-EOP) microspheres containing paclitaxel was obtained, as shown in Figure 25A, 25B, 25C, 25D, 25E.

Example 34: Biocompatibility of Poly (phosphoester) Microspheres in Mouse Peritoneal Cavity

[0183] The biocompatibility of biodegradable poly(ghosphoester) microspheres of the invention was tested as follows:

[0184] Three 30 mg/mL samples of lyophilized poly(L-lactide-co-ethyl-phosphate) microspheres were prepared, the first having diameters greater than 75 microns, the second having diameters within the range of 75-125 microns, and the third having diameters within the range of 125-250 microns. Each sample was injected intra-peritoneally into a group of 18 female CD-1 mice having a starting body weight of 25 g.' Animals in each group were weighed, sacrificed, and necropsied at 2, 7 and 14 days, and at 1, 2 and 3 months. Any lesions detected during the necropsy were graded

on a scale of 0 to 4, with 0 indicating no response to treatment and 4 indicating a severe response to treatment.

[0185] Inflammatory lesions were observed to be restricted to an association with the microspheres on peritoneal surfaces or within fat tissue, and were compatible with foreign body isolation and encapsulation. Focal to multifocal supportive peritoneal steatitis with mesothelial hyperplasia was observed at 2-7 days, but gradually resolved by macrophage infiltration, the formation of inflammatory giant cells, and fibrous encapsulation of the microspheres at later sacrifices. Occasional adherence of microspheres to the liver and diaphragm, with associated inflammatory reaction, was also seen. Lesions related to microspheres were not seen within abdominal or thoracic organs. Microspheres, which were detected throughout the duration of the study, appeared transparent at early sacrifices but, at later times, developed crystalline material internally. No effects on body growth were observed. The peritoneal reaction was observed to be limited to areas directly adjacent to the microspheres with no apparent deleterious effects on major thoracic or abdominal organs.

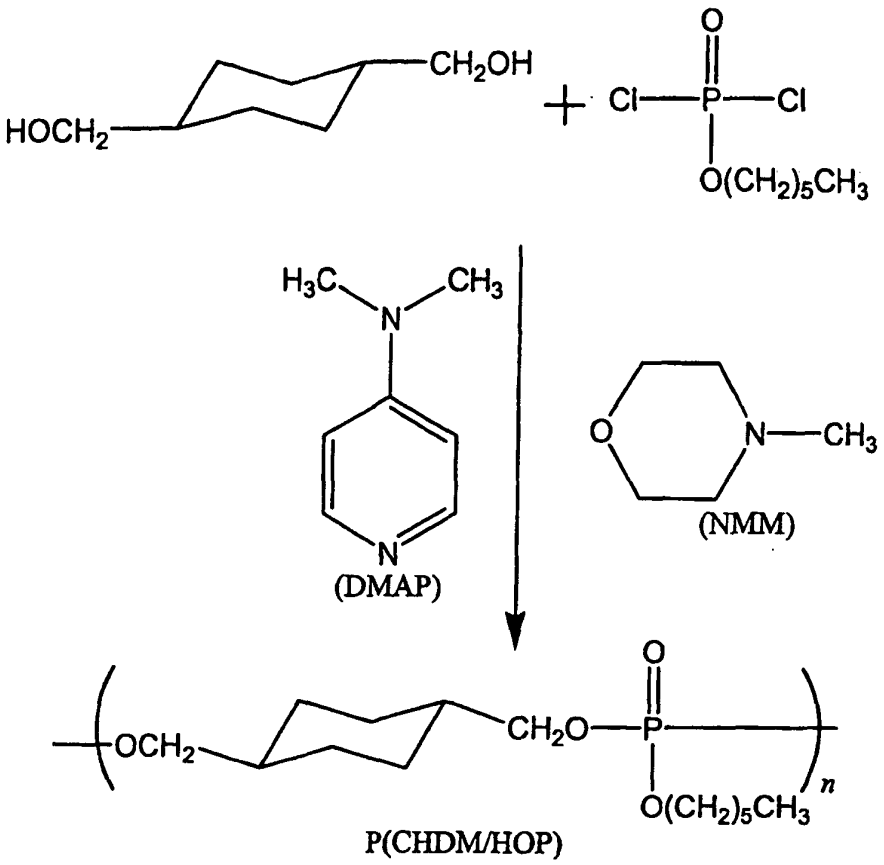
[0186] Similar intraperitoneal injection of DAPG-EOP into male and female S-D rats gave the following results:

Dose Level	Test Material	Initial No. in Test		Cumulative Mortality[a]	
(mg/ kg)		M	F	M	F
0	10% Dextran 40 in 0.9% Saline	25	25	0	0
30	DAPG-EOP	25	25	1	0
100	DAPG-EOP	25	25	0	0
300	DAPG-EOP	25	25	0	0

[a]Represents animals found dead or sacrificed in moribund condition during study period. M = Male; F = Female

Example 35: Synthesis of the Poly(phosphoester) P(trans-CHDM-HOP)

[0187]



[0188] Under an argon stream, 10 g of trans-1,4-cyclohexane dimethanol (CHDM), 1.794 g of 4-dimethylaminopyridine (DMAP), 15.25 ml (14.03 g) of N-methyl morpholine (NMM), and 50 ml of methylene chloride, were transferred into a 250 ml flask equipped with a funnel. The solution in the flask was cooled down to -15°C with stirring, and a solution of 15.19 g of hexyl phosphorodichloridate (HOP) in 30 ml of methylene chloride was added through the funnel. The temperature of the reaction mixture was raised to the boiling point gradually and maintained at reflux temperature overnight.

[0189] The reaction mixture was filtered, and the filtrate was evaporated to dryness. The residue was re-dissolved in 100 ml of chloroform. This solution was washed with 0.1 M solution of a mixture of HCl and NaCl, dried over anhydrous Na_2SO_4, and quenched into 500 ml of ether. The resulting flowable precipitate was collected and dried under vacuum to form a clear pale yellow gelatinous polymer with the flow characteristics of a viscous syrup. The yield for this polymer was 70-80%. The structure of P(trans-CHDM-HOP) was ascertained by ^{31}P-NMR and ^{1}H-NMR spectra, as shown in Figure 27, and by FT-IR spectra. The molecular weights (Mw=8584; Mn=3076) were determined by gel permeation chromatography (GPC), as shown in Figure 27A, 27B and 27C using polystyrene as a calibration standard.

Example 36: Synthesis of the Poly(Phosphoester) P(cis & trans - CHDM-HOP)

[0190] Poly (phosphoester) P(cis/trans-1,4-cyclohexanedimethanol hexyl phosphate) was prepared by following the procedure described above in Example 34 except that a mixture of cis- and trans-1,4-cyclohexanedimethanol was used as the starting material. As expected, the product cis - /trans -P (CHDM-HOP) was less viscous than the trans isomer obtained in Example 34.

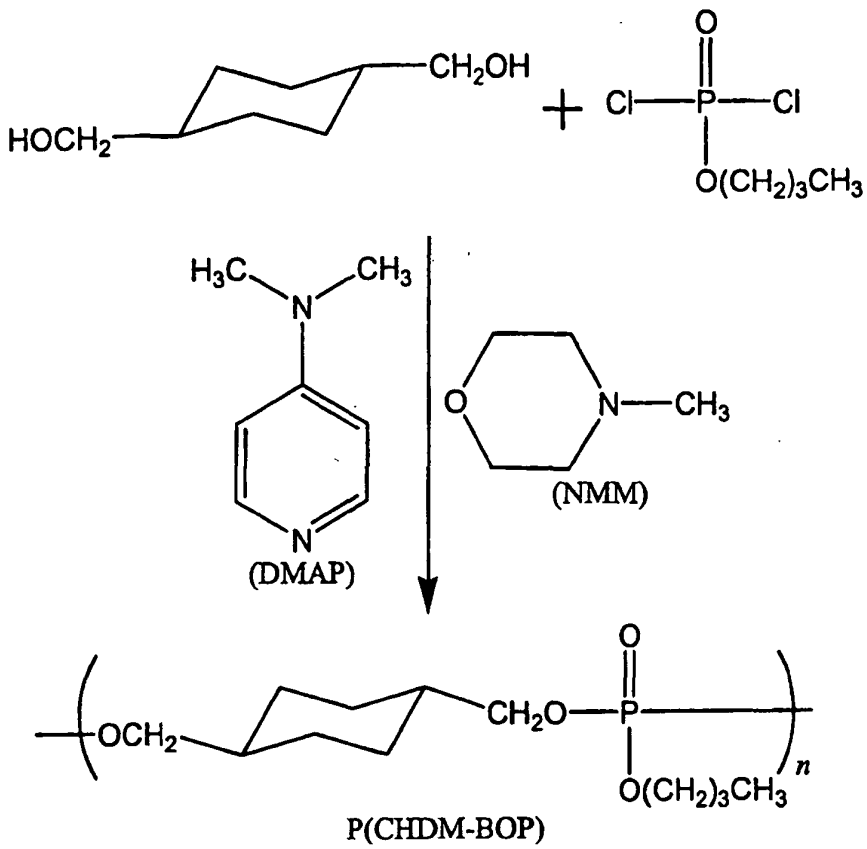
Example 37: Synthesis of Low Molecular Weight P(CHDM-HOP)

[0191] Under an argon stream, 10 g of trans-1,4-cyclohexane dimethanol (CHDM), 15.25 mL (14.03 g) of N-methyl morpholine (NMM), and 50 mL of methylene chloride were transferred into a 250 mL flash equipped with a funnel. The solution in the flask was cooled down to -40°C with stirring. A solution of 15..19 g of hexyl phosphoro-dichloridate (HOP) in 20 mL of methylene chloride was added through the funnel, and an additional 10 mL of methylene chloride was used to flush through the funnel. The mixture was then brought up to room temperature gradually and kept stirring for four hours.

[0192] The reaction mixture was filtered, and the filtrate was evaporated to dryness. The residue was re-dissolved in 100 ml of chloroform. This solution was washed with 0.5 M mixture of HCl-NaCl solution, washed with saturated NaCl solution, dried over anhydrous Na_2SO_4, and quenched into a 1:5 ether-petroleum mixture. The resulting oily precipitate was collected and dried under vacuum to form a clear, pale yellow viscous material. The structure of the product was confirmed by ^{1}H-NMR, ^{31}P-NMR and FT-IR spectra.

Example 38: Synthesis of the Poly(phosphoester) P(trans-CHDM-BOP)

[0193]



P(CHDM-BOP)

[0194] Under an argon stream, 10 g of trans-1,4-cyclohexane dimethanol (CHDM), 0.424 g (5%) of 4-dimethylamino-pyridine (DMAP), 15.25 mL (14.03 g) of N-methyl morpholine (NMM) and 50 mL of methylene chloride were transferred into a 250 mL flask equipped with a funnel. The solution in the flask was cooled down to -40°C with stirring. A solution of 13.24 g of butyl phosphoro-dichloridate (BOP) in 20 mL of methylene chloride was added through the funnel, with an additional 10 mL of methylene chloride being used to flush through the funnel. The mixture was heated to the boiling point gradually, and kept refluxing for four hours. The reaction mixture was filtered, and the filtrate was evaporated to dryness, taking care to keep the temperature below 60°C. The residue was redissolved in 100 mL of chloroform. The solution formed was washed with 0.5 M of HCl-NaCl solution and saturated NaCl solution, dried over anhydrous Na2SO4, and quenched into a 1:5 ether-petroleum mixture. The resulting oily precipitate was collected and dried under vacuum to produce a clear, pale yellow viscous material.

Example 39: Rheological Properties of P(trans-CHDM-HOP)

[0195] P(trans-CHDM-HOP) remained in a flowable gel-like state at room temperature. The polymer exhibited a steady viscosity of 327Pa·s at 25°C (shown in Figure 28B), and a flowing active energy of 67.5 KJ/mol (shown in Figure 28A).

Example 40: In Vitro Cytotoxicity of P(trans-CHDM-HOP)

[0196] Cover slips were coated with P (trans-CHDM-HOP) by a spin coating method. The coated coverslips were then dried and sterilized by UV irradiation overnight under a hood. A P (trans-CHDM-HOP)-coated cover slip was placed at the bottom of each well of a 6-well plate. $5x10^5$ HEK293 (human embryonic kidney) cells were plated into each well and cultured for 72 hours at 37°C. The resulting cell morphology was examined, using tissue culture polystyrene (TCPS) as a positive control. The cells growing on the P(CHDM-HOP) surface proliferated at a slightly slower rate, as shown by Figures 29A and 29B However, the morphology of cells grown on the polymer surface was similar

to the morphology of cells grown on the TCPS surface.

Example 41: In Vitro Degradation of P(CHDM-Alkyl Phosphates)

[0197] Each of the following poly(phosphate)s was prepared as described above:

Table 11

Polymer	Side Chain
P(CHDM-HOP)	-O-hexyl group
P(CHDM-BOP)	-O-bucyl group
P(CHDM-EOP)	-O-ethyl group

A sample of 50 mg of each polymer was incubated in 5 mL of 0.1 M, pH 7.4 phosphate buffer saline (PBS) at 37°C. At various points in time, the supernatant was poured out, and the polymer samples were washed three times with distilled water. The polymer samples were then extracted with chloroform, and the chloroform solution was evaporated to dryness. The residue was analyzed for weight loss by comparing with the original 50 mg. sample. Figure 30 graphically represents the effect of the side chain structure on the in vitro degradation rate of poly(phosphates) in PBS.

Example 42: In Vitro Release Profile of Protein by P(CHDM-HOP)

[0198] The polymer P(CHDH-HOP) was blended with the protein FITC-BSA (bovine serum albumin, a protein, tagged with the fluorescent label PITC; "FITC-HSA") at a 2:1 (w/w) ratio (33% loading). Measured amounts (66 mg or 104 mg) of the polymer-protein blend were placed into 10 ml of PBS (0.1M, pH 7.4), a phosphate buffer. At regular intervals (roughly every day), the samples were centrifuged, the supernatant buffer was removed and subjected to absorption spectroscopy (501 nm), and fresh amounts of buffer were added to the samples. The resulting release curve, plotting the cumulative percentage of FITC-BSA released versus time, is graphically represented in Figure 31. The loading level of the protein in both cases was 33 weight %.

Example 43: In vitro Protein Release Profile At Various Loading Levels

[0199] FITC-BSA was blended with P) (CHDM-HOP) at different loading levels (1%, 10% and 30%) at room temperature until the mixture formed a homogenous paste. 60 mg of the protein-loaded polymer paste was placed in 6 mL of 0.1 M phosphate buffer and constantly shaken at 37°C. At various time points, samples were centrifuged, and the supernatant was replaced with fresh buffer. The released FITC-BSA in the supernatant was measured by UV spectrophotometry at 501 nm. Figure 6 graphically represents the in vitro release kinetics of FITC-BSA as a function of loading level.

Example 44: Effect of Side Chain Structure on In vitro Protein Release Kinetics of FITC-BSA

[0200] The following three polymers were prepared as described above:

P(CHDM-EOP)
P(CHPM-BOP) and
P(CHDM-HOP)

FITC-BSA was blended with each polymer at a 10% loading level at room temperature to form a homogenous paste. 60 mg of the protein-loaded polymer paste was placed in 6 mL of 0.1 M phosphate buffer with constant shaking at 37°C. At various time points, samples were centrifuged, and the supernatant was replaced with fresh buffer. The released FITC-BSA in the supernatant was measured by UV spectrophotometry at 501 nm. Figure 32 graphically represents the in vitro effect of side chain variations on the protein release kinetics of FITC-BSA at 10% loading level.

Example 45: In Vitro Small Molecular Weight Drug Release from P(CHDM-HOP)

[0201] A P(CHDM-HOP) paste containing doxorubicin, cisplatin, or 5-fluorouracil, was prepared by blending 100 mg of P(CHDM-HOP) with 1 mg of the desired drug at room temperature, respectively. An aliquot of 60 mg of the drug-loaded paste was placed in 6 mL of 0.1 M phosphate buffer at 37°C with constant shaking, with three samples being

done for each drug being tested. At various time points, the supernatant was replaced with fresh buffer solution. The levels of doxorubicin and 5-fluorouracil in the supernatant were quantified by UV spectrophotometry at 484 nm and 280 nm, respectively. The cisplatin level was measured with an atomic absorbance spectrophotometer. Figure 34 shows the release of these low molecular weight drugs from P(CHDM-HOP).

[0202] Figure 9B shows the release of hydrophobic small molecules, such as paclitaxel, from p(CHDM-HOP).

Example 46: In Vitro Release Profile of Doxorubicin and Cisplatin from P·(CHDM-HOP)

[0203] A paste was made by blending 300 mg of P(CHDM-HOP) with 6 mg of doxorubicin and 6 mg of cisplatin at room temperature to form a uniform dispersion. A sample of 100 mg of the paste was incubated in 10 mL of phosphate buffer (pH 7.4) at 37°C with shaking. At different time points, samples were centrifuged, 9 mL of the supernatant were withdrawn and replaced with fresh buffer. The withdrawn supernatant was assayed spectrophocometrically at 484 nm to determine the amount of doxorubicin released into the withdrawn supernatant, and the cisplatin release was measured by atomic absorbance spectrophotometer. Figure 35. graphically represents the simultaneous release of cisplatin and doxorubicin from P(CHDM-HOP).

Example 47: In Vivo Biocompatibility of P trans-CHDH-HOP)

[0204] The polymer P(trans-CHDM-HOP) was synthesized as described in Example 1. To facilitate injection, ethyl alcohol was added to the polymer at levels of 10% and 20% by volume to reduce the viscosity. Samples of 25 μL of the polymer alone, 25 μL of the polymer containing 10% alcohol, and 25 μL of the polymer containing 20% alcohol, were injected into the back muscles of Sprague Dawley rats. Tissues at the injection sites were harvested at either three or thirteen days post-injection, processed for paraffin histology, stained with hematoxylin, eosin dye and analyzed. Medical-grade silicon oil was injected into the control group rats.

[0205] Histological examination of the back muscle sections of the rats injected with the polymer diluted with ethanol showed no acute inflammatory response. The level of macrophage presence was comparable to that of the control group, which had been injected with medical-grade silicon oil, and neutrophils were not present in any of the samples taken on either the third or thirteenth day.

Example 48: Controlled Delivery of Interleukin-2 and Doxorubicin from P(CHDM-HOP) in an In Vivo Tumor Model

[0206] Lyophilized interleukin-2 ("IL-2") was purchased from Chiron, mouse Interferon-γ ("mIFN-γ") was obtained from Boehringer Mannheim, and doxorubicin hydrochloride ("DOX") was obtained from Sigma. C57BL/6 mice, 6-8 weeks of age, were obtained from Charles River. The aggressive melanoma cell line B16/F10 was used to cause tumors in the mice, and the cells were maintained by weekly passages. The polymer P(CHDM-HOP) was synthesized as described in Example 35.

[0207] Mice were randomly allocated into groups as shown below in Table 12. The day of tumor injection with cells of the melanoma cell line was denoted as Day 0. Each mouse received a subcutaneous injection of 50 μl (10^5) tumor cells in phosphate buffer saline (PBS) in the left flank. On Day 3 or Day 7, the tumor-bearing mice were selectively injected in the right flank with one of the following formulations: (1) a bolus of IL-2, (2) a bolus of DOX, (3) a polymer paste of IL-2, (4) a polymer paste of DOX, (5) a polymer paste containing both IL-2 and DOX, or (6) a polymer paste containing both IL-2 and mIFN-γ). A control group and negative control group received no further injections on Day 3 or Day 7.

[0208] The bolus preparation of either IL-2 or DOX was prepared by dissolving an appropriate amount of IL-2 or DOX in 50 μl of isotonic solution just prior to the injection. The polymer paste formulations of either IL-2, DOX, a mixture of both IL-2 and DOX, or a mixture of IL-2 and mIFN-γ, were prepared by blending 50 μl of sterilized P(CHDM-HOP) with the drug(s) until homogeneous.

Table 12:

Allocation of Groups of Mice for In Vivo Tumor Model			
Group	Number of Mice	Day of Injection	Formulation
Control	5	--	Nothing
Negative Control	5	--	Nothing
Bolus IL-2	8	3	0.8 X 10^6 IU
Bolus DOX	8	3	0.5 mg

Table 12: (continued)

Allocation of Groups of Mice for In Vivo Tumor Model			
Group	Number of Mice	Day of Injection	Formulation
Bolus DOX	8	7	0.5 mg
Paste IL-2	10	3	0.8 X 10^6 IU
Paste IL-2	10	7	0.8 X 10^6 IU
Paste DOX	10	3	0.5 mg
Paste DOX	10	7	0.5 mg
Paste (IL-2 + DOX)	10	3	0.8 X 10^6 IU + 0.5 mg
Paste (IL-2 + DOX)	10	7	0.8 X 10^6 IU + 0.5 mg
Paste (IL-2 + mIFN-γ)	10	3	10^6 IU

[0209] On Day 28 and Day 42 of tumor growth, the tumor sizes of the various mice were measured. The results are shown below in Table 13, which shows the numerical data for the growth of tumor volumes on Day 28 and Day 42 and the number of mice who survived the experiment per drug grouping. Tumor volume was calculated as half the product of the length and the square of the width, in accordance with the procedure of Osieka et al., 1981.

Table 13:

CHDM-HOP Polymer as Carrier for Cytokine and Drug Delivery in Melanoma Model			
Group	Initial Number of Mice	Tumor Volume (mm^3±SEM*) After Tumor Injection	
		28 days	42 days
		Number of Mice Survived	
Control	5	No tumor	No tumor
Negative Control	5	2458 ± 1070.7	5656
		4	1
Bolus IL-2 (3d)	8	1946 ± 505.6	3282 ± 1403.3
		8	4
Bolus Dox (3d)	8	1218.9 ± 304.1	3942.5 ± 1818
		8	5
Bolus Dox (7d)	8	1661.2 ± 301.8	4394.3 ± 741.3
		8	3
Paste IL-2 (3d)	10	934.1 ± 230	3183 ± 1223.4
		10	5
Paste IL-2 (7d)	10	2709.8 ± 397.3	10491 ± 2485.5
		10	3
Paste Dox (3d)	10	1410 ± 475.3	4648.9 ± 1202.2
		8	7
Paste Dox (7d)	10	1480 ± 287	3915 ± 1739.7
		9	4
Paste (IL-2 + DOX) (3d)	10	657.3 ± 248.9	3362.8 ± 1120.1
		8	7

* Standard Error of the Mean

Table 13: (continued)

CHDM-HOP Polymer as Carrier for Cytokine and Drug Delivery in Melanoma Model			
Group	Initial Number of Mice	Tumor Volume (mm^3±SEM*) After Tumor Injection	
		28 days	42 days
		Number of Mice Survived	
Paste (IL-2 + DOX) (7d)	10	857.2 ± 243.6	3449.8 ± 1285.9
		8	5
Paste (IL-2 + mIFN-γ) (3d)	10	1217.9 ± 168.4	4469.8 ± 2018.7
		9	4

* Standard Error of the Mean

Based on these measurements, the distribution of tumors sizes were graphically represented in Figure 36 or Day 28 (four weeks after tumor implantation) and in Figure 37 for Day 42 (six weeks after tumor implantation). The graphs were subdivided into plots according to the different treatments given to the tumor-bearing mice.

[0210] The results on Day 28 showed that, in comparison with the control group (tumor without treatment) and the bolus injection of IL-2, the group of mice receiving a polymer/IL-2 paste injection successfully delayed the tumor's growth. However, for the group of mice not receiving a polymer/IL-2 paste injection until Day 7, the tumor had already become of substantial size by Day 7 and, accordingly, a significant reduction in tumor size was not observed.

[0211] Excellent tumor reduction was obtained with the combination of IL-2 and DOX. The average size of a tumor treated with an injection of a polymer paste containing both IL-2 and DOX was significantly smaller than the tumor in the control group. Specifically, the average tumor size for mice receiving the IL-2 and DOX/polymer paste on Day 3 was 657.3 mm^3, as opposed to 2458 mm^3 for the control group. Even when treatment was relayed until Day 7 of tumor growth, a therapeutic effect could still be seen with the polymer paste formulation containing both IL-2 and DOX.

[0212] The results on Day 42 of tumor growth also confirmed that the Day 3 injection of polymer paste containing both IL-2 and DOX gave the best result in delaying tumor growth. The combined therapy of IL-2 and DOX in a polymer paste of the invention resulted in the occurrence of smaller sized tumors in more of the test animals. According to the distribution data shown in Figure 15, there were four mice bearing tumors of less than 1000 mm^3 in the case of the combined IL-2 and DOX polymer paste therapy, as compared with only one mouse inside that range for the polymer paste injection of DOX alone. It was also clear that IL-2 alone did not provide the most desirable effect, as evaluated on the 42nd day of tumor growth. Despite the good distribution of small tumor sizes on the 28th day, the long-time survival data appeared to be adversely affected, not only by the progression of tumor growth at that point, but also by the lack of continued, controlled delivery of IL-2 over a longer time period. with the polymer paste formulation of both IL-2 and DOX, the polymer degraded slowly, allowing a gradual decrease in the diffusion rate of the therapeutic agent over time.

[0213] However, because of the significant difference of the distribution in tumor sizes inside each group, the average tumor size as seen in Table 13 did not provide a complete picture. A fuller appreciation of the significance of the treatments of the invention can be gained by comparing data from the distribution graph of Figure 37 which shows the dispersity in tumor sizes six weeks after tumor implantation, with the survival curve shown in Figure 38, which shows the massive.death of mice in all groups before the Day 42 measurement, except for the groups of animals that had received the 3rd day injection of paste containing either DOX alone or the combination of IL-2 and DOX. Thus, the data, taken as a whole, shows that the combined therapy of IL-2 and box in the paste both significantly delayed tumor growth and extended life.

[0214] Early deaths about 3-4 days after the injections of the DOX-containing polymer paste were thought to be due, at least in part, to the toxic effect of DOX causing the deaths of the weaker animals. Corresponding injections of bolus DOX did not produce early death, probably because of the rapid distribution and clearance from the body of the bolus-injected DOX.

Example 49: Incorporating Paclitaxel into P(CRDM-HOP) or PICHDM-EOP)

[0215] 100 mg of each of the polymers p(CHDM-HOP) and p(CHDM-EOP) was dissolved in ethanol at a concentration of about 50%. After the polymer was completely dissolved, 5 mg of paclitaxel powder (a chemotherapeutic drug) was added to the solution and stirred until the powder was completely dissolved. This solution was then poured into ice water to precipitate the polymer composition. The resulting suspension was centrifuged, decanted, and lyophilized

overnight, to obtain a viscous gelatinous product.

Example 50: In Vitro Release of Paclitaxel from PCCRDM-HOP) and P(CHDM-EOP)

[0216] In a 1.7 mL plastic micro centrifuge tube, S mg of both of the paclitaxel polymer formulations of Example 20 to be tested was incubated with 1 mL of a buffer mixture of 90% PBS and 20% PEG 400 at 37°C. Four samples of each formulation to be tested were prepared. At specific time points, approximately every day, the PBS: PEG buffer was poured off for paclitaxel analysis by HPLC, and fresh buffer was added to the microcentrifuge tube. The release study was terminated at day 26, at which point the remaining paclitaxel in the polymer was extracted with a solvent to do a mass balance on paclitaxel.

[0217] The resulting release curves for the release of paclitaxel from both polymers are shown in Figure 39. The total paclitaxel recovery was 65% for the P(CHDM-HOP) formulation and 75% for the P(CHDM-EOP) formulation.

Example 51: In Vitro Release of Paclitaxel from P(DAEG-EOP)

[0218] P(DAPO-EOP) microspheres were prepared by a solvent evaporation method, using ethyl acetate as the solvent and 0.5% PVA in water as a non-solvent. The resulting microspheres are spherical in shape with particle sizes ranging from about 20-150 μm, most preferably 20-50 μm.

[0219] The in vitro release of paclitaxel from the microspheres was carried out in PBS (pH 7.4) at 37°C. To maintain a sink condition, an octanol layer was placed on top of the PBS to continuously extract the released paclitaxel. The released paclitaxel was quantified using an HPLC method, and the in vitro mass loss of the polymer was obtained by a gravimetric method. The in vitro release of paclitaxel from the microspheres was slow and continuous with concomitant polymer mass loss, as shown in Figure 25A.

Example 52: In Vivo Release of Paclitaxel from P(DAPG-EOP)

[0220] P(DAPG-EOP) microspheres were prepared as described above in Example 52, and the In vivo release of paclitaxel from the microspheres was studied on nude mice. Plasma was collected from each of the test animals at 1, 14 and 28 days after injection, and paclitaxel concentration was analyzed by HPLC with MS-MS detection. For efficacy studies, test animals received intraperitoneal injections of a human ovarian cancer cell line OVCAR3 obtained from carrier animals. P(DAFC-EOP) microspheres incorporating paclitaxel or paclitaxel without the biodegradable polymer were also given intraperitoneally at one day post cell injection. The survival of the animals was also monitored.

[0221] Following a single intraperitoneal administration of the microspheres, a sustained level of paclitaxel in plasma was obtained for at least 28 days, as shown below in Table 14:

Table 14

Paclitaxel Plasma Concentration		
	Paclitaxel Concentration (ng/ml)	
	Paclitaxel in Microspheres (125 mg/kg)	Paclitaxel W/O polymer (120 mg/kg)
1 day	38.98±7.53	357.67±136.39
14 days	4.50±1.21	Animal died
28 days	3.98±0.99	Animal died

when a comparable dose of paclitaxel was given intraperitoneally, the nude mice could not tolerate the dose due to the toxicity.

[0222] The biodegradable polymer microsphere delivery system was surprisingly effective in treating ovarian cancer in the animal model OVCAR3. As shown in Figure 43, superior efficacy was obtained, as compared with paclitaxel without the biodegradable polymer.

Example 53: Median Survival Data for P(DAPC-EOP) Paclitaxel

[0223] P(DAPG-EOP) microspheres containing 10 mg/kg or 40 mg/kg paclitaxel were injected into the peritoneums of test animals having ovarian cancer. other test animals were injected with paclitaxel in an organic solvent, commercially available under the trade name Taxol, at the same dosage levels. The test animals were monitored, and median survival times were noted. The results are summarized below:

Material Administered	Median Survival
Control	23 days
Taxol, 10 mg/kg	64 days
Taxol, 40 mg/kg	67 days
Paclitaxel in microspheres, 10 mg/kgr	69 days
Paclitaxel in microspheres, 40 mg/kg	115 days

[0224] These results are represented graphically in Figure 43 and indicate an unexpectedly large increase in median survival for the test animals given the paclitaxel in the form of biodegradable microspheres.

[0225] A comparison of a different set of dosage levels gave the following similar data:

Material Administered	Median Survival
Control	30 days
Taxol, 40 mg/kg	77 days
Faclitaxel in microspheres, 4 mg/kg	83 days
Paclitaxel in microspheres, 10 mg/kg	95 days
Paclitaxel in microspheres, 40 mg/kg	>110 days

[0226] These results are represented graphically in Figure 41 and confirm the unexpectedly large increase in median survival for the test animals given the paclitaxel in the form of biodegradable microspheres. Additional graphical representations of this data are provided by Figures 40 and 42.

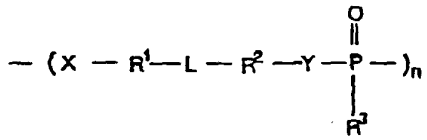
Claims

1. Use of a polymer composition for the manufacture of a medicament for the treatment of ovarian cancer, said polymer composition comprising:

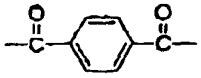
(a) at least one antineoplastic agent and

(b) a biodegradable polymer comprising the recurring monomeric units shown in formula I:

I



wherein X is -O- or -NR^4-, where R^4 is H or alkyl;
Y is -O-, -S- or -NR^4-;
each of R^1 and R^2 is a divalent organic moiety;
L is a divalent, branched or straight chain aliphatic group having 1-20 carbon atom, a cycloaliphatic group, or a group having the formula:



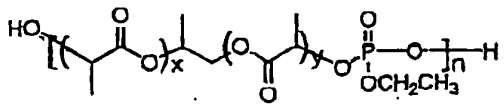
R^3 is selected from the group consisting of H, alkyl, alkoxy, aryl, aryloxy, heterocyclic or heterocycloxy; and
n is 5-5,000;
wherein the polymer composition is suitable to be inserted in vivo into the peritoneum of said subject such that the inserted composition is in at least partial contact with an ovarian cancer tumor.

2. Use of a biodegradable the polymer composition for the manufacture of a medicament for the treatment of ovarian cancer, said polymer composition comprising:

(a) paclitaxel; and

(b) the biodegradable polymer shown in formula- VI:

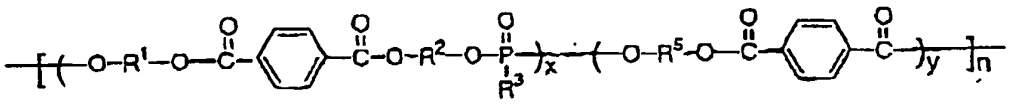
VI



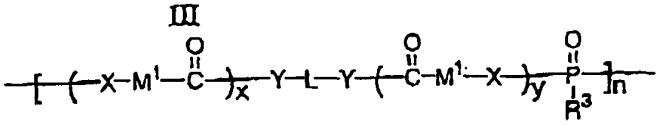
wherein the molar ratio of x:y is about 1;
the molar ratio n:(x or y) is between 200 1 and 1:200; and
n is 5-5,000;
wherein said polymer composition is suitable to be inserted in vivo into the peritoneum of said subject such that the inserted composition is in at least partial contact with on ovarian cancer tumor.

3. The use of claim 1, wherein said polymer is selected from the group consisting of:

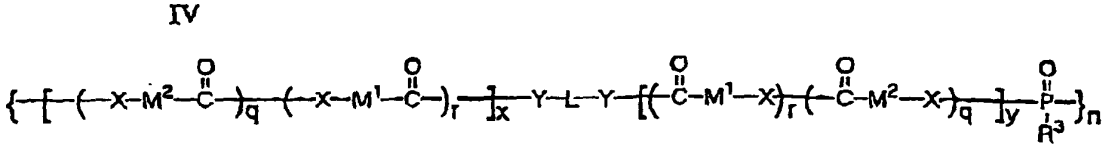
II



III

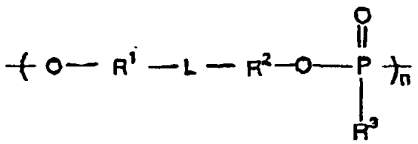


IV



and

V



wherein;
R^5 is a divalent organic moiety,
M^1 and M^2 are each Independently (1) a branched or straight chain aliphatic group having from 1-20 carbon atoms; or (2) a branched or straight chain, oxy-, carboxy- or amino-aliphatic group having from 1-20 carbon atoms;
the molar ratio of x:y is about 1;
the molar ratio n:(x or y) is between 200:1 and 1:200; and
the molar ratio q:r is between 1 :99 and 99:1.

4. The use of any one of the preceding claims, wherein the polymer composition is capable of releasing at least one antineoplastic agent in a controlled fashion at the site of a ovarian cancer tumor or the tissue surrounding the site of an ovarian cancer tumor after administration to a mammal.

5. The use of claim 4, wherein the cancer tumor has been removed from the site of an ovarian cancer tumor.

6. The use of any one of the preceding claims, wherein at least one antineoplastic agent comprises paclitaxel.

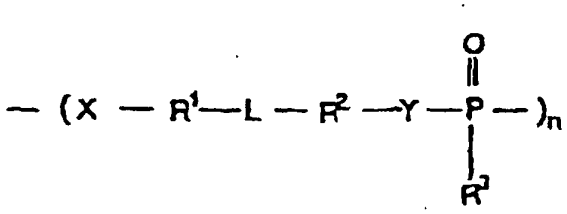
7. The use of any one of the preceding claims, wherein the polymer composition provides extended release of at least one antineoplastic agent for at least about four weeks.

Patentansprüche

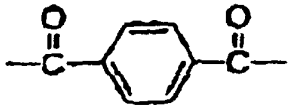
1. Verwendung einer Polymerzusammensetzung zur Herstellung eines Medikaments für die Behandlung von Ovarialkrebs, wobei die Polymerzusammensetzung umfasst:

(a) mindestens ein antineoplastisches Mittel; und
(b) ein bioabbaubares Polymer, das die sich wiederholenden Monomereinheiten umfasst, die in Formel I dargestellt sind:

I



wobei
X -O- oder -NR^4- ist, wobei R^4 H oder Alkyl ist;
Y -O-, -S- oder -NR^4- ist;
jedes von R^1 und R^2 eine zweiwertige organische Komponente ist;
L eine zweiwertige, verzweigte oder geradkettige aliphatische Gruppe mit 1-20 Kohlenstoffatomen, eine cycloaliphatische Gruppe oder eine Gruppe mit der formel:

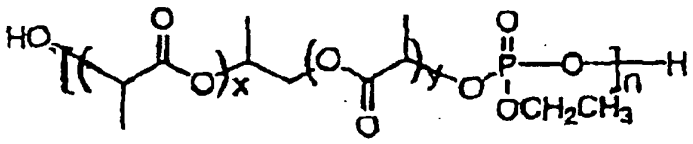


ist;
R^3 aus der Gruppe bestehend aus H, Alkyl, Alkoxy, Aryl, Aryloxy, einem Heterocyclus oder Heterocycloxy ausgewählt wird; und
n 5-5.000 ist;
wobei die Polymerzusammensetzung dazu geeignet ist, in vivo so in das Peritoneum der Patientin eingesetzt zu werden, dass die eingesetzte Zusammensetzung in zumindest teilweisem Kontakt mit einem Ovarialkrebs-Tumor steht.

2. Verwendung einer bioabbaubaren Polymerzusammensetzung zur Herstellung eines Medikaments für die Behandlung von Ovarialkrebs, wobei die Polymerzusammensetzung umfasst:

(a) Paclitaxel; und
(b) das bioabbaubare Polymer, das in Formel VI dargestellt ist:

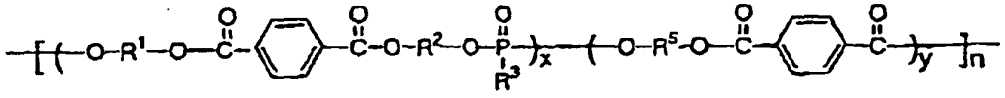
VI



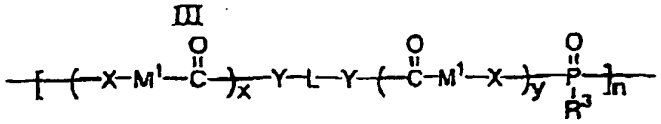
wobei
das Molverhältnis x:y ungefähr 1 beträgt;
das Molverhältnis n:(x oder y) zwischen 200:1 und 1:200 beträgt; und
n 5-5.000 ist;
wobei die Polymerzusammensetzung dazu geeignet ist, in vivo so in das Peritoneum der Patientin eingesetzt zu werden, dass die eingesetzte Zusammensetzung in zumindest teilweisem Kontakt mit einem Ovarialkrebs-Tumor steht.

3. Verwendung nach Anspruch 1, wobei das Polymer aus der Gruppe bestehend aus:

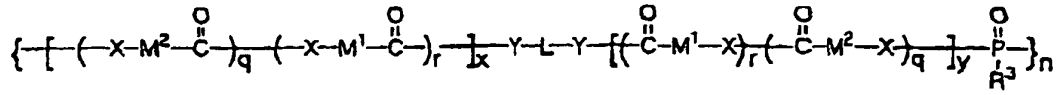
II



III

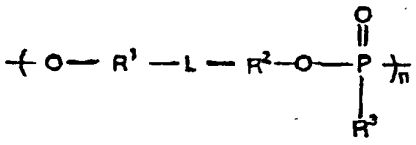


IV



und

V



ausgewählt wird;
wobei:

R^5 eine zweiwertige organische Komponente ist;
M^1 und M^2 jeweils unabhängig (1) eine verzweigte oder geradkettige aliphatische Gruppe mit 1-20 Kohlenstoffatomen sind; oder (2) eine verzweigte oder geradkettige, Oxy-, Carboxy- oder Amino-aliphatische Gruppe mit 1-20 Kohlenstoffatomen sind;
das Molverhältnis x:y ungefähr 1 beträgt;
das Molverhältnis n:(x oder y) zwischen 200:1 und 1:200 beträgt; und
das Molverhältnis q:r zwischen 1:99 und 99:1 beträgt.

4. Verwendung nach irgendeinem der vorangehenden Ansprüche, wobei die Polymerzusammensetzung in der Lage ist, nach Verabreichung an ein Säugetier mindestens ein antineoplastisches Mittel in kontrollierter Weise an der Stelle eines Ovarialkrebs-Tumors oder an dem die Stelle eines Ovarialkrebs-Tumors umgebenden Gewebes freizusetzen.

5. Verwendung nach Anspruch 4, wobei der Krebstumor von der Stelle eines Ovarialkrebs-Tumors entfernt wurde.

6. Verwendung nach irgendeinem der vorangehenden Ansprüche, wobei mindestens ein antineoplastisches Mittel Paclitaxel umfasst.

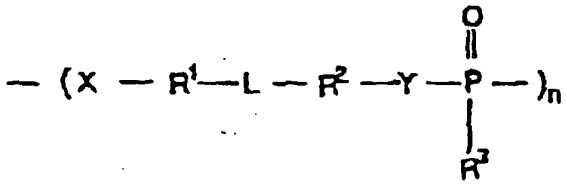
7. Verwendung nach irgendeinem der vorangehenden Ansprüche, wobei die Polymerzusammensetzung eine verlängerte Freisetzung von mindestens einem antineoplastischen Mittel für mindestens ungefähr vier Wochen bereitstellt.

Revendications

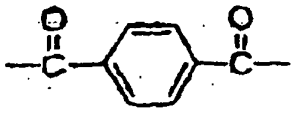
1. Utilisation d'une composition polymère pour la fabrication d'un médicament destiné au traitement du cancer des ovaires, ladite composition polymère comprenant :

(a) au moins un agent antinéoplasique et
(b) un polymère biodégradable comprenant les motifs monomères récurrents indiqués dans la formule 1 :

I



dans laquelle X représente -O- ou -NR^4-, où R^4 est un atome de H ou un groupe alkyle;
Y représente -O-, -S- ou -NR^4-,
chacun de R^1 et de R^2 est une fraction organique bivalente;
L est un groupe aliphatique bivalent, ramifié ou à chaîne droite, ayant de 1 à 20 atomes de carbone, un groupe cycloaliphatique ou un groupe ayant pour formule :

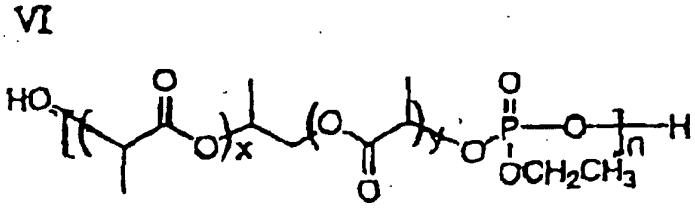


R^3 est choisi dans le groupe formé par un atome de H, un groupe alkyle, alcoxy, aryle, aryloxy, hétérocyclique ou hétérocyclo-oxy; et
n vaut de 5 à 5000;
dans laquelle la composition polymère convient à l'insertion *in vivo* dans le péritoine dudit sujet de sorte que la composition insérée est au moins en contact partiel avec une tumeur de cancer des ovaires.

2. Utilisation d'une composition polymère biodégradable pour la fabrication d'un médicament destiné au traitement du cancer des ovaires, ladite composition polymère comprenant :

(a) du paclitaxel; et
(b) le polymère biodégradable indiqué dans la formule VI :

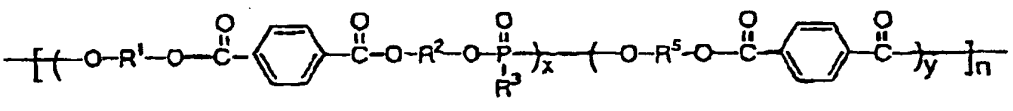
VI



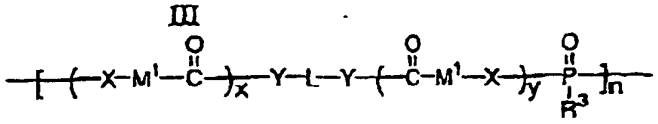
dans laquelle le rapport molaire x:y est d'environ 1;
le rapport molaire n:(x ou y) est compris entre 200:1 et 1:200; et
n vaut de 5 à 5000;
dans laquelle ladite composition polymère convient à l'insertion *in vivo* dans le péritoine dudit sujet de sorte que la composition insérée est au moins en contact partiel avec une tumeur de cancer des ovaires.

3. Utilisation selon la revendication 1, dans laquelle ledit polymère est choisi dans le groupe formé par :

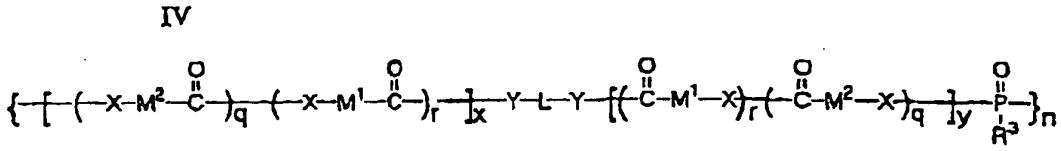
II



III

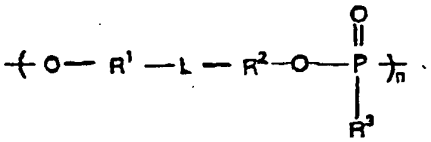


IV



et

V



dans lesquelles :

R^5 est une fraction organique bivalente,
M^1 et M^2 représentent chacun indépendamment (1) un groupe aliphatique ramifié ou à chaîne droite ayant de 1-20 atomes de carbones ; ou bien (2) un groupe oxy-, carboxy-, ou amino-aliphatique; ramifié où à chaîne droite, ayant de 1-20 atomes de carbone;
le rapport molaire x:y est d'environ 1:
le rapport molaire n: (x ou y) est compris entre 200:1 et 1:200; et
le rapport molaire q:r est compris entre 1:99 et 99:1.

4. Utilisation selon l'une quelconque des revendications précédentes, dans laquelle la composition polymère est susceptible de libérer au moins un agent anti-néoplasique d'une façon contrôlée au niveau du site d'une tumeur de cancer des ovaires ou du tissu entourant le site d'une tumeur de cancer des ovaires, après administration à un mammifère.

5. Utilisation selon la revendication 4, dans laquelle la tumeur cancéreuse a été retirée du site d'une tumeur de cancer des ovaires.

6. Utilisation selon l'une quelconque des revendications précédentes, dans laquelle au moins un agent antinéoplasique comprend le paclitaxel.

7. Utilisation selon l'une quelconque des revendications précédentes, dans laquelle la composition polymère offre une libération étendue d'au moins un agent antinéoplasique pendant au moins environ quatre semaines.

80/20-B, DMSO
OBSERVE H1
FREQUENCY 399.952 MHz
SPECTRAL WIDTH 5000.0 Hz
ACQUISITION TIME 3.744 sec
RELAXATION DELAY 0.000 sec
PULSE WIDTH 7.0μsec
AMBIENT TEMPERATURE
NO. REPETITIONS 12
DOUBLE PRECISION ACQUISITION
DATA PROCESSING
FT SIZE 65536
TOTAL ACQUISITION TIME 1 minute

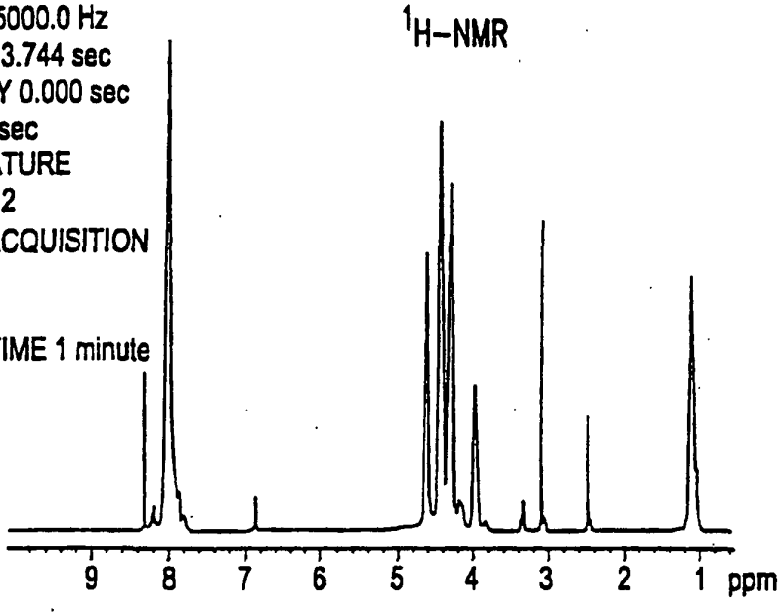


FIG.1A

80/20-B,BB, DMSO
OBSERVE P31
FREQUENCY 161.903 MHz
SPECTRAL WIDTH 16000.0 Hz
ACQUISITION TIME 0.800 sec
RELAXATION DELAY 0.000 sec
PULSE WIDTH 10.0μsec
TEMPERATURE 37.0°C
NO. REPETITIONS 77
DECOUPLE H1
HIGH POWER 33
DECOUPLER CONTINUOUSLY ON
WALTZ16 MODULATED
DOUBLE PRECISION ACQUISITION
DATA PROCESSING
LINE BROADENING 15.0Hz
FT SIZE 32768
TOTAL ACQUISITION TIME 1 minute

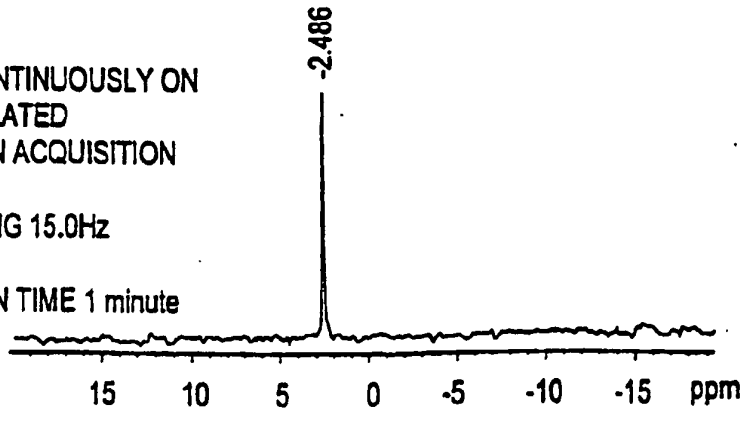


FIG.1B

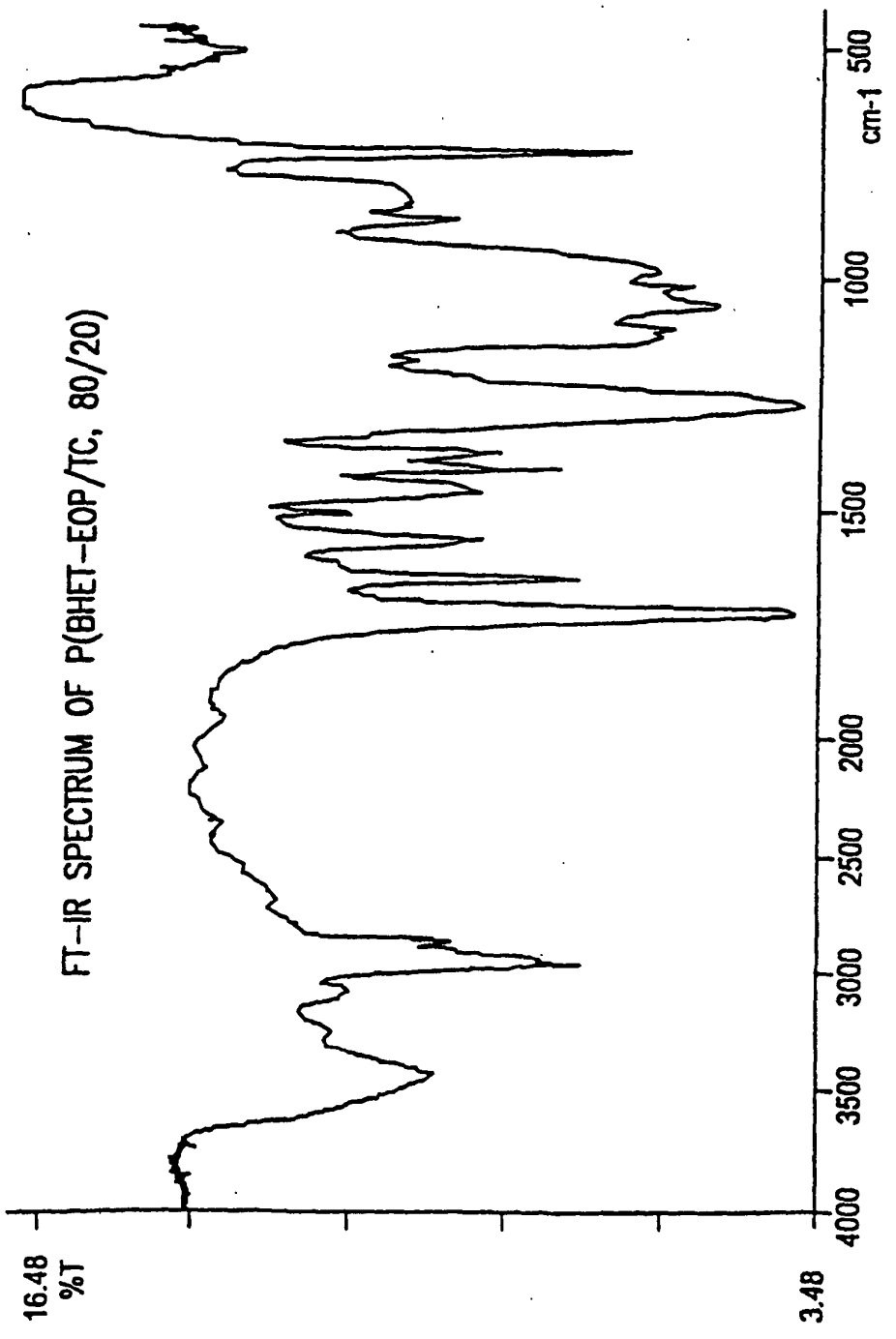

FT-IR SPECTRUM OF P(BHET-EOP/TC, 80/20)
16.48
%T
3.48
4000
3500
3000
2500
2000
1500
1000
500
cm-1

FIG.2

MOLECULAR WEIGHT AND ELEMENTAL ANALYSIS

Polymer	Mn (VPO)	Mw/Mn (GPC)	Elemental Analysis; Found (Theory)		
			C (%)	H (%)	P (%)
P(BHET-EOP/TC, 80/20)	7918	6119 /2219	49.61 (51.82)	4.95 (4.81)	6.24 (7.03)
P(BHET-HOP/TC, 90/10)	6364	4201 /1587	51.67 (53.11)	5.99 (5.90)	6.26 (6.77)

FIG.3A

GPC CHROMATOGRAM FOR P(BHET-EOP/TC, 80/20)

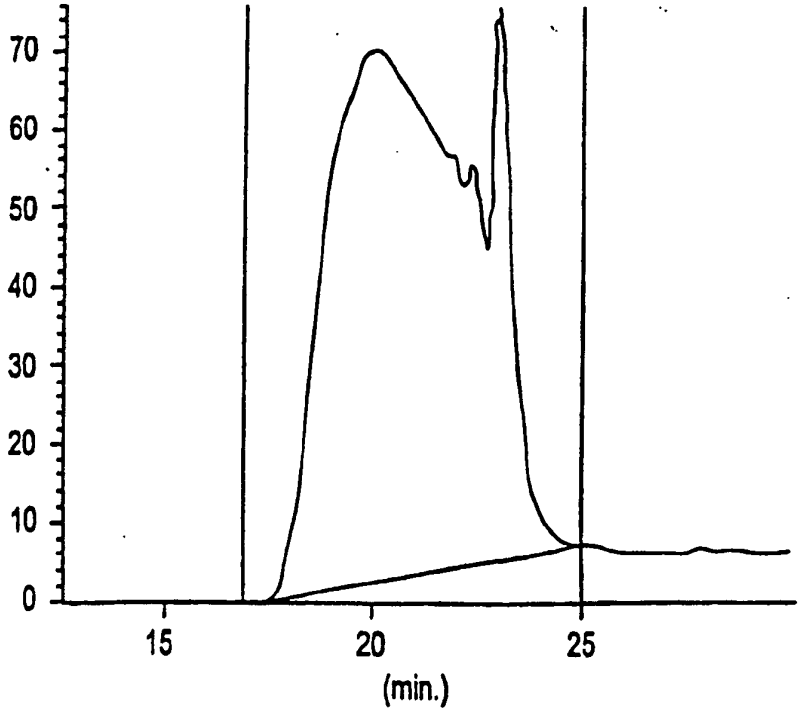


FIG.3B

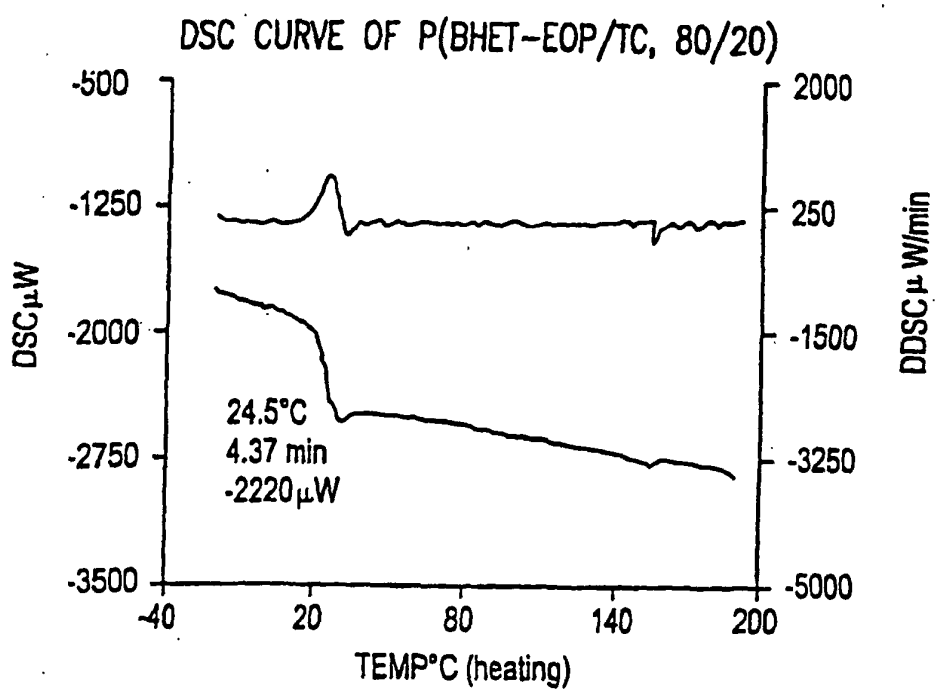

DSC CURVE OF P(BHET-EOP/TC, 80/20)
-500
-1250
-2000
-2750
-3500
2000
250
-1500
-3250
-5000
DSCμW
DDSC μ W/min
24.5°C
4.37 min
-2220μW
-40
20
80
140
200
TEMP°C (heating)

FIG.4A

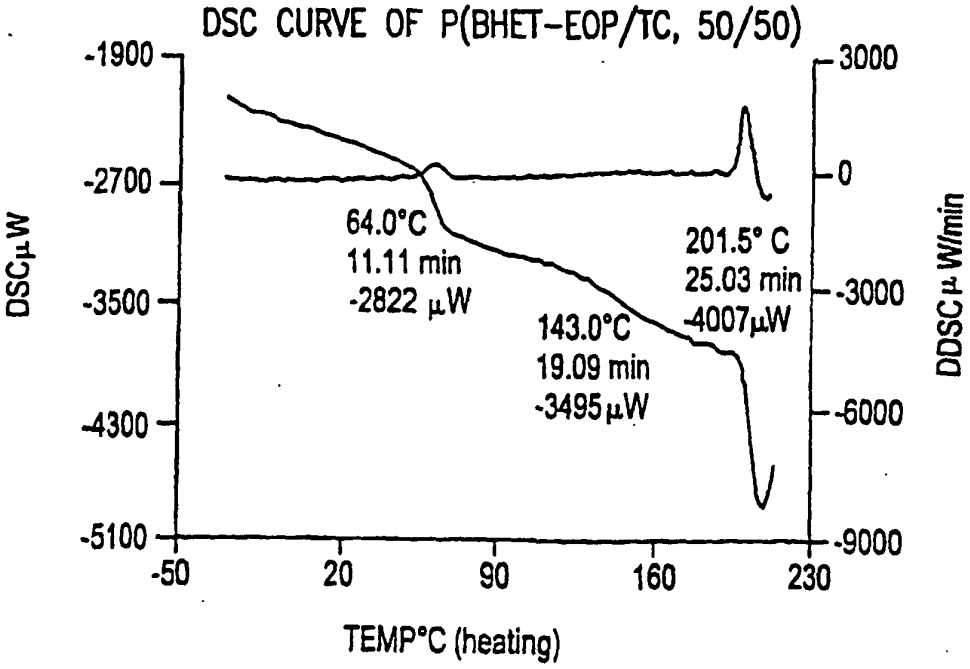

DSC CURVE OF P(BHET-EOP/TC, 50/50)
-1900
-2700
-3500
-4300
-5100
3000
0
-3000
-6000
-9000
DSCμW
DDSC μ W/min
64.0°C
11.11 min
-2822 μW
143.0°C
19.09 min
-3495μW
201.5° C
25.03 min
-4007μW
-50
20
90
160
230
TEMP°C (heating)

FIG.4B

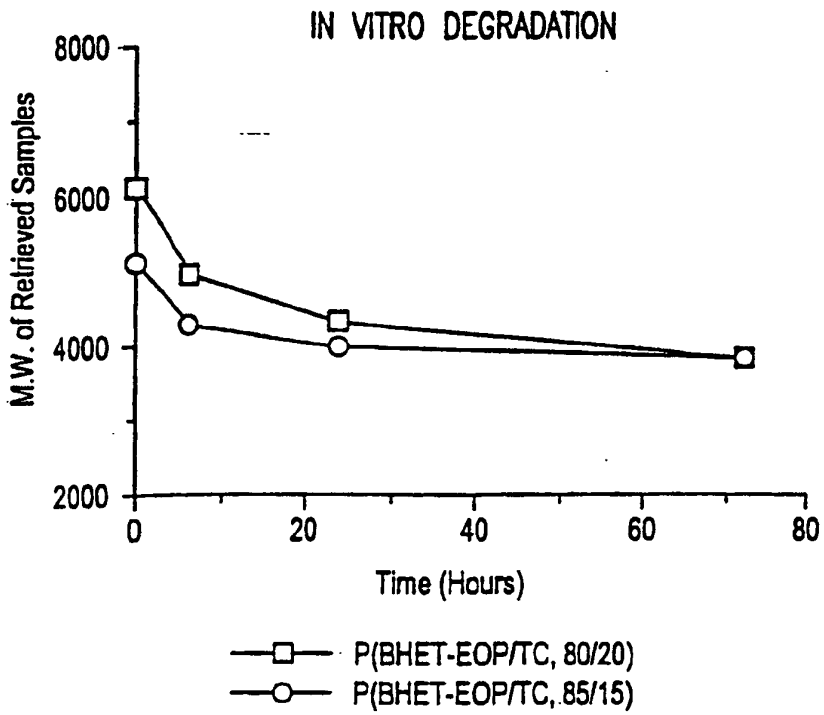

IN VITRO DEGRADATION
M.W. of Retrieved Samples
8000
6000
4000
2000
0
20
40
60
80
Time (Hours)
P(BHET-EOP/TC, 80/20)
P(BHET-EOP/TC, 85/15)

FIG.5A

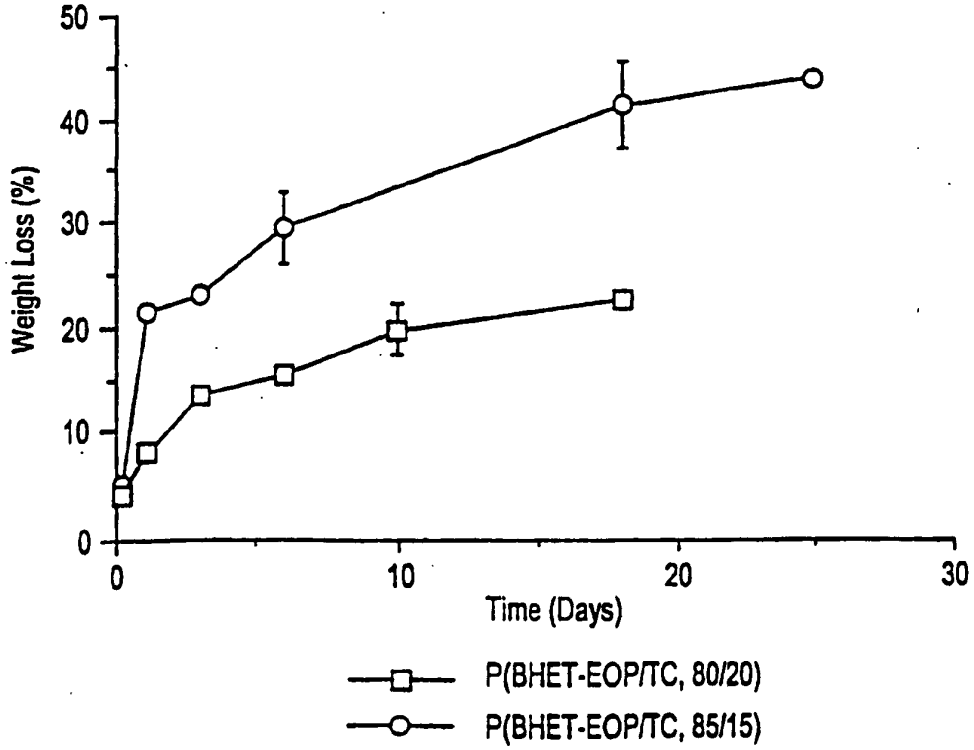

Weight Loss (%)
50
40
30
20
10
0
0
10
20
30
Time (Days)
P(BHET-EOP/TC, 80/20)
P(BHET-EOP/TC, 85/15)

FIG.5B

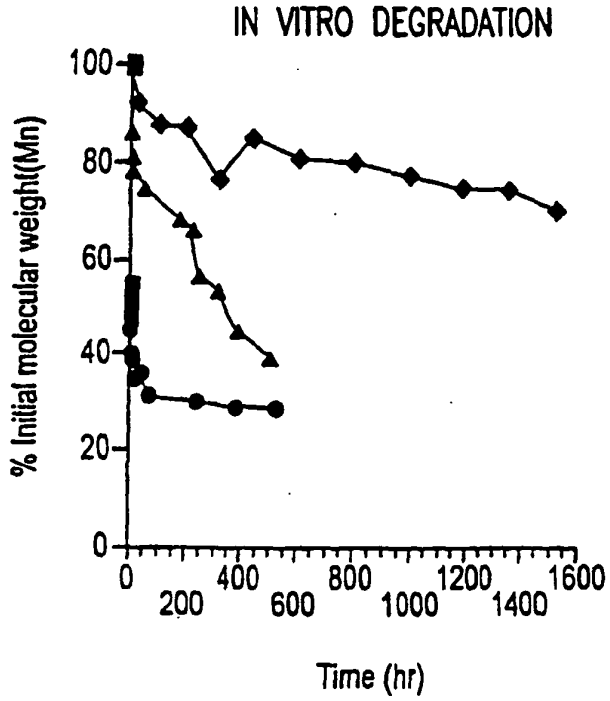

IN VITRO DEGRADATION
% Initial molecular weight(Mn)
100
80
60
40
20
0
0
200
400
600
800
1000
1200
1400
1600
Time (hr)
p(BHDPT-EOP)
p(BHDPT-EOP/TC, 85/15)
p(BHDPT-EOP/TC, 75/25)
p(BHDPT-EOP/TC, 50/50)

FIG.6

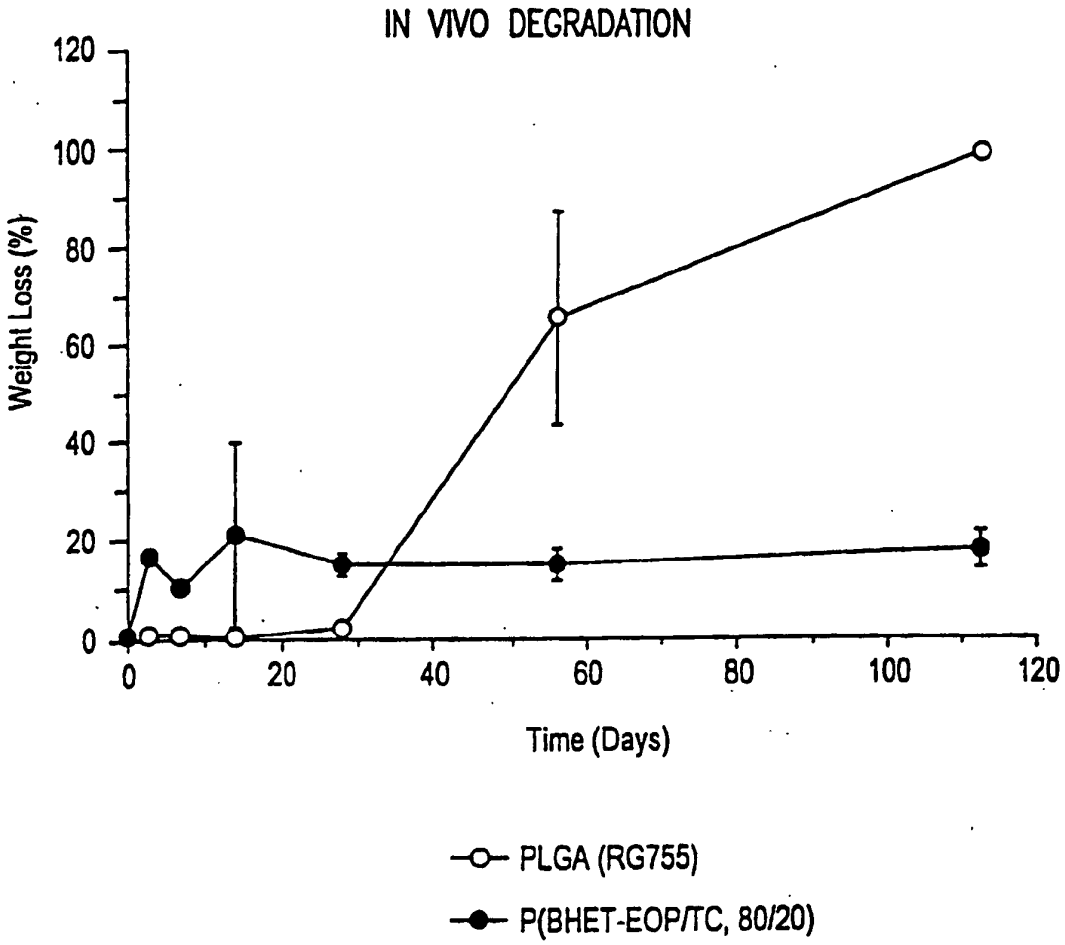

IN VIVO DEGRADATION
Weight Loss (%)
120
100
80
60
40
20
0
0
20
40
60
80
100
120
Time (Days)
PLGA (RG755)
P(BHET-EOP/TC, 80/20)

FIG.7A

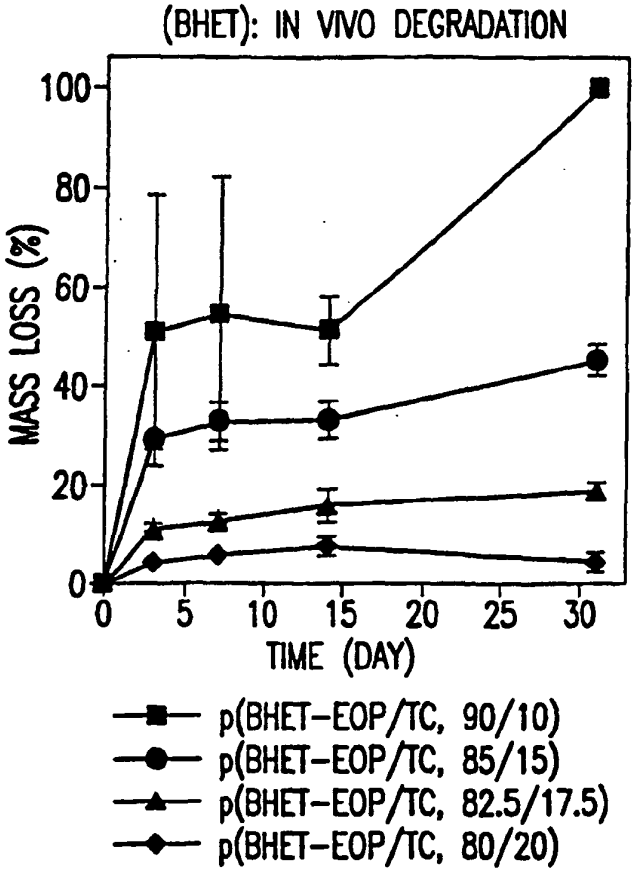


FIG. 7B

P(BHET-EOP/TC 80/20): CONTROLLED DELIVERY OF HYDROPHOBIC SMALL MOLECULES

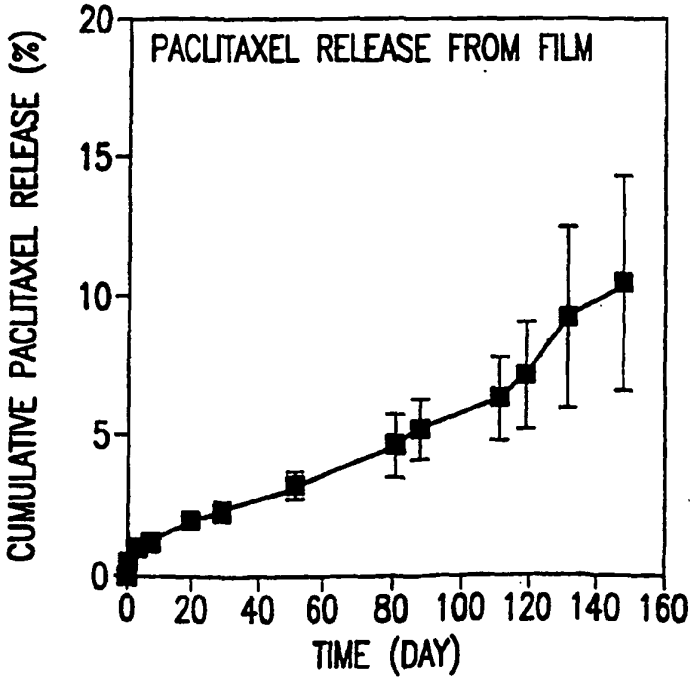


FIG. 7C

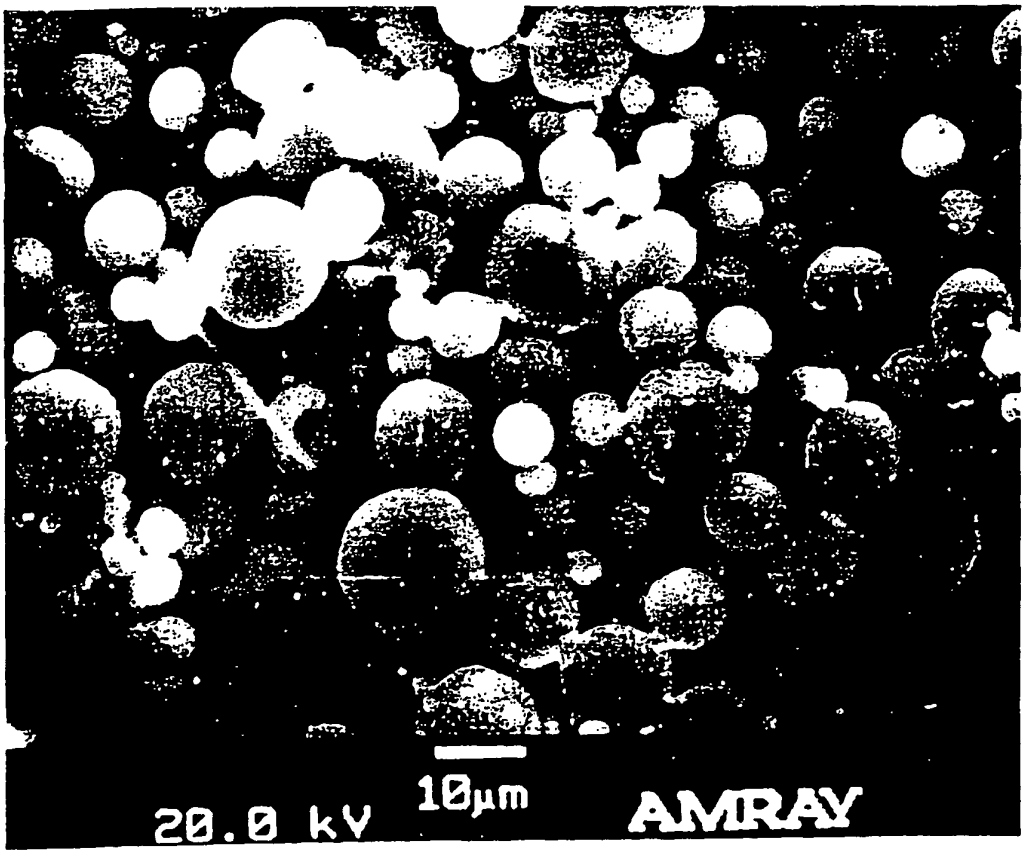


FIG.8

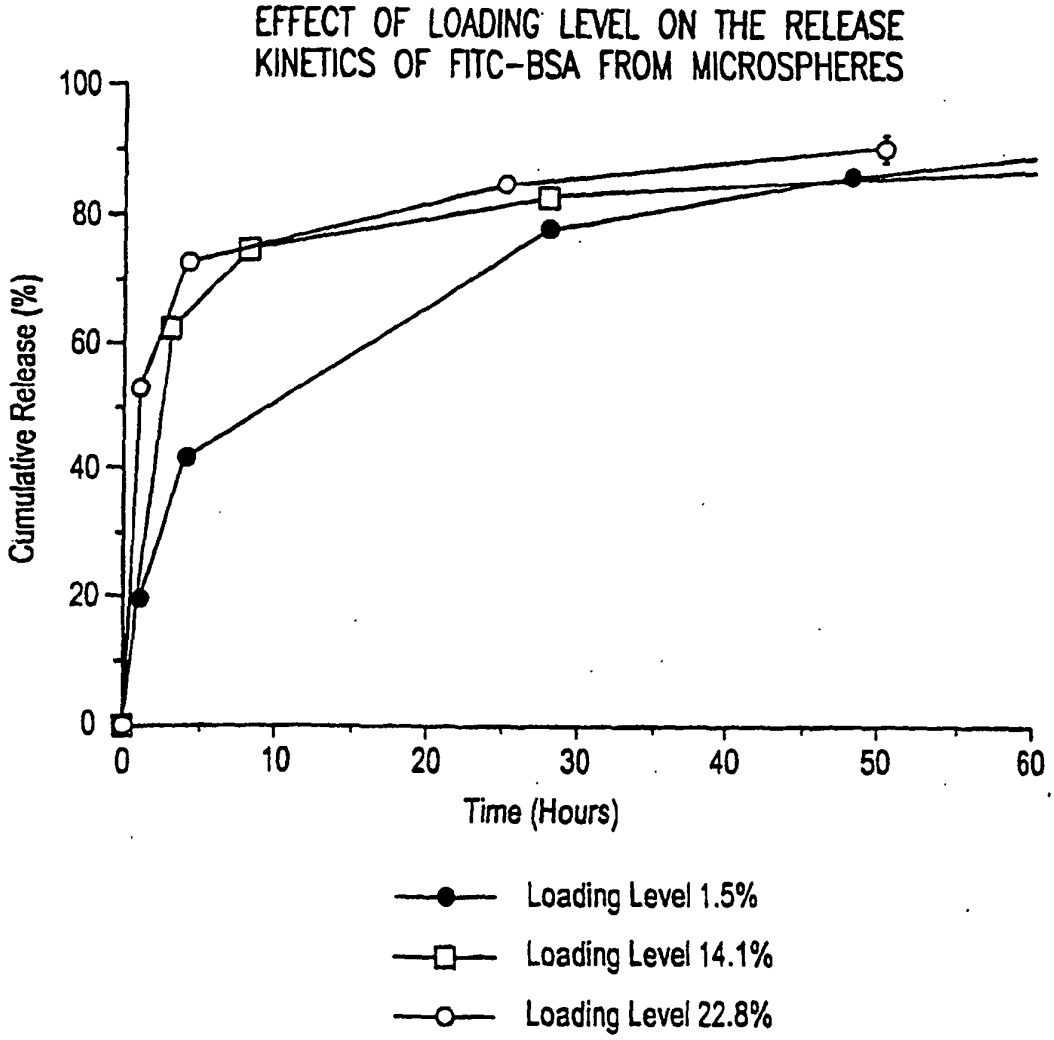

EFFECT OF LOADING LEVEL ON THE RELEASE KINETICS OF FITC-BSA FROM MICROSPHERES
Cumulative Release (%)
100
80
60
40
20
0
0
10
20
30
40
50
60
Time (Hours)
Loading Level 1.5%
Loading Level 14.1%
Loading Level 22.8%

FIG.9A

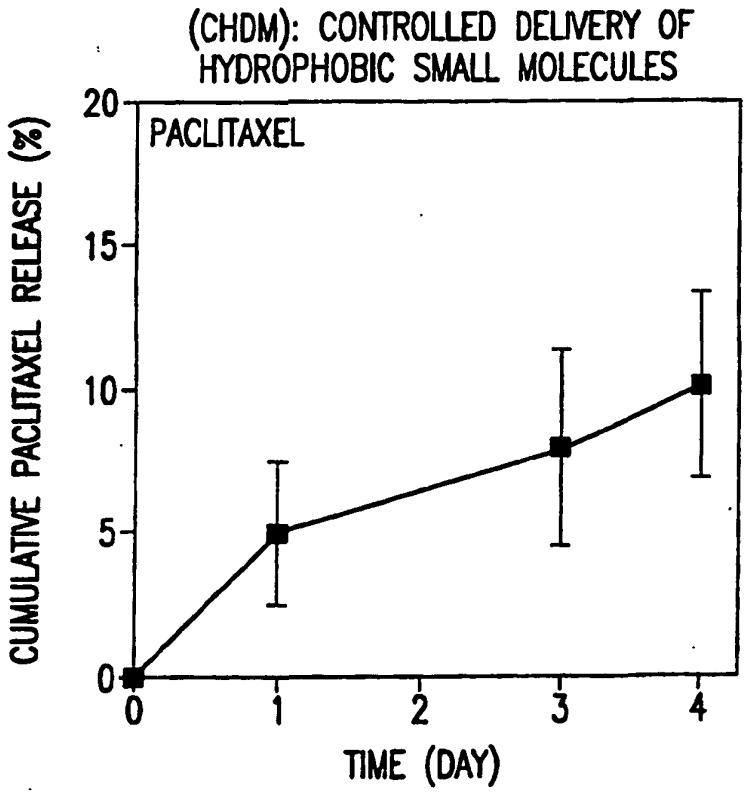

(CHDM): CONTROLLED DELIVERY OF HYDROPHOBIC SMALL MOLECULES
PACLITAXEL
CUMULATIVE PACLITAXEL RELEASE (%)
20
15
10
5
0
0
1
2
3
4
TIME (DAY)

FIG.9B

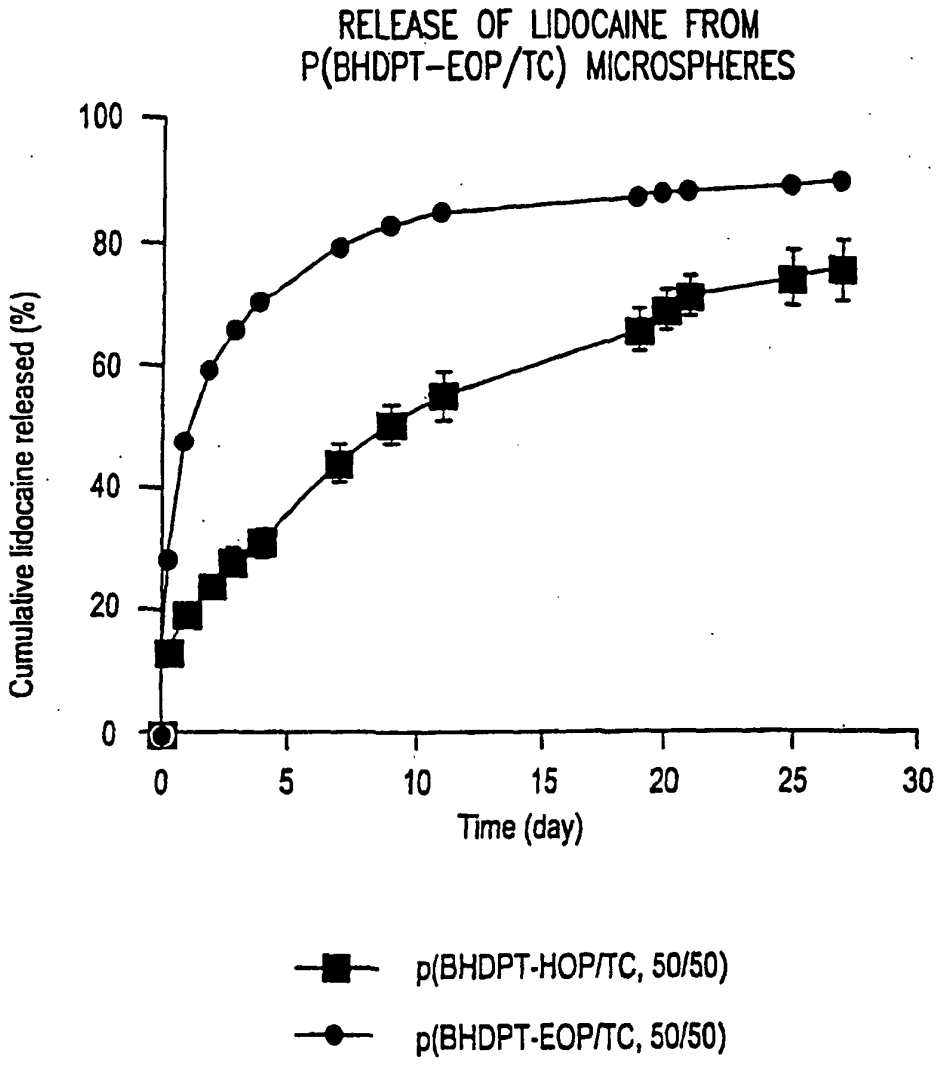

RELEASE OF LIDOCAINE FROM
P(BHDPT-EOP/TC) MICROSPHERES
Cumulative lidocaine released (%)
100
80
60
40
20
0
0
5
10
15
20
25
30
Time (day)
p(BHDPT-HOP/TC, 50/50)
p(BHDPT-EOP/TC, 50/50)

FIG.10

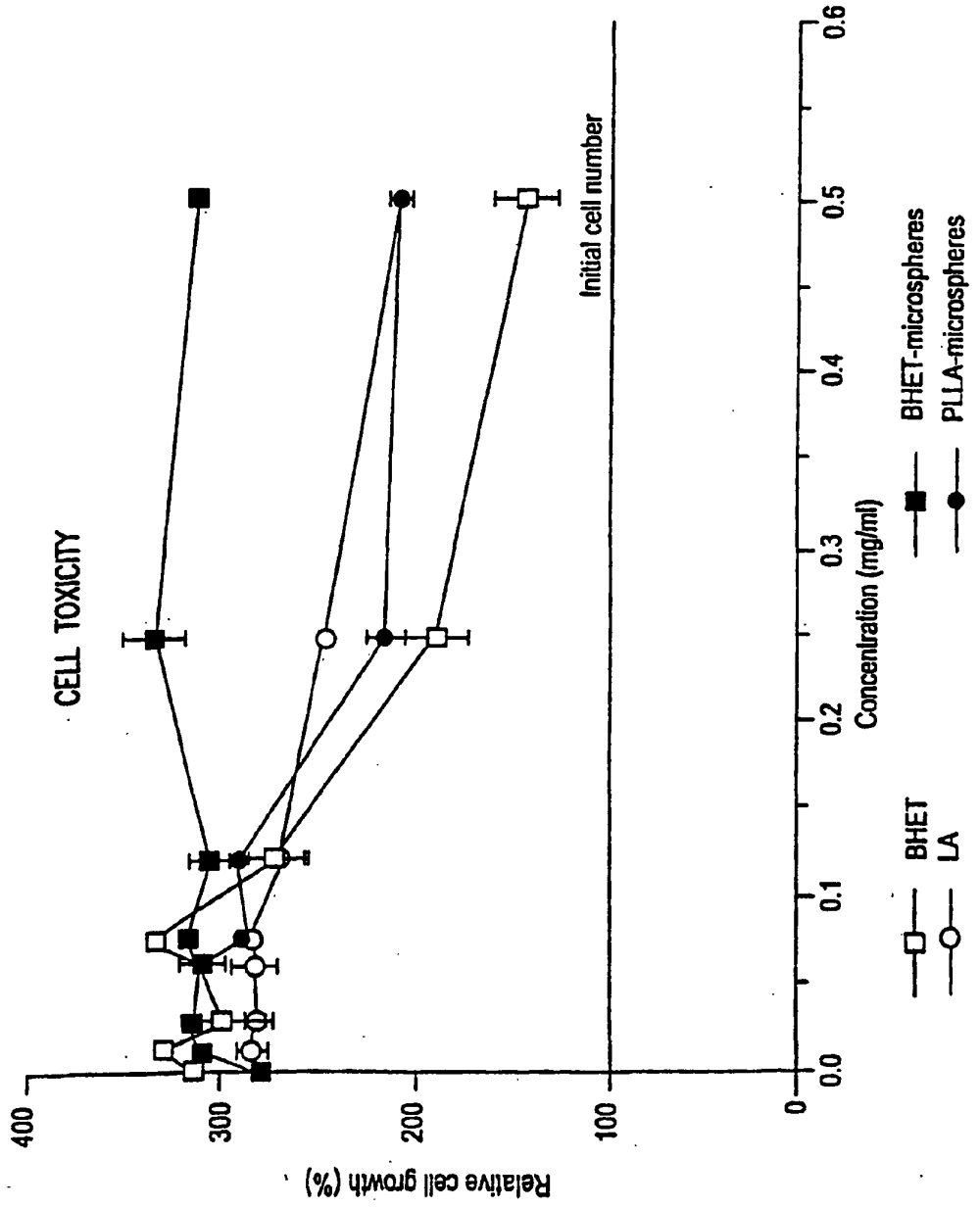

CELL TOXICITY
Relative cell growth (%)
400
300
200
100
0
Initial cell number
0.0
0.1
0.2
0.3
0.4
0.5
0.6
Concentration (mg/ml)
BHET
LA
BHET-microspheres
PLLA-microspheres

FIG.11

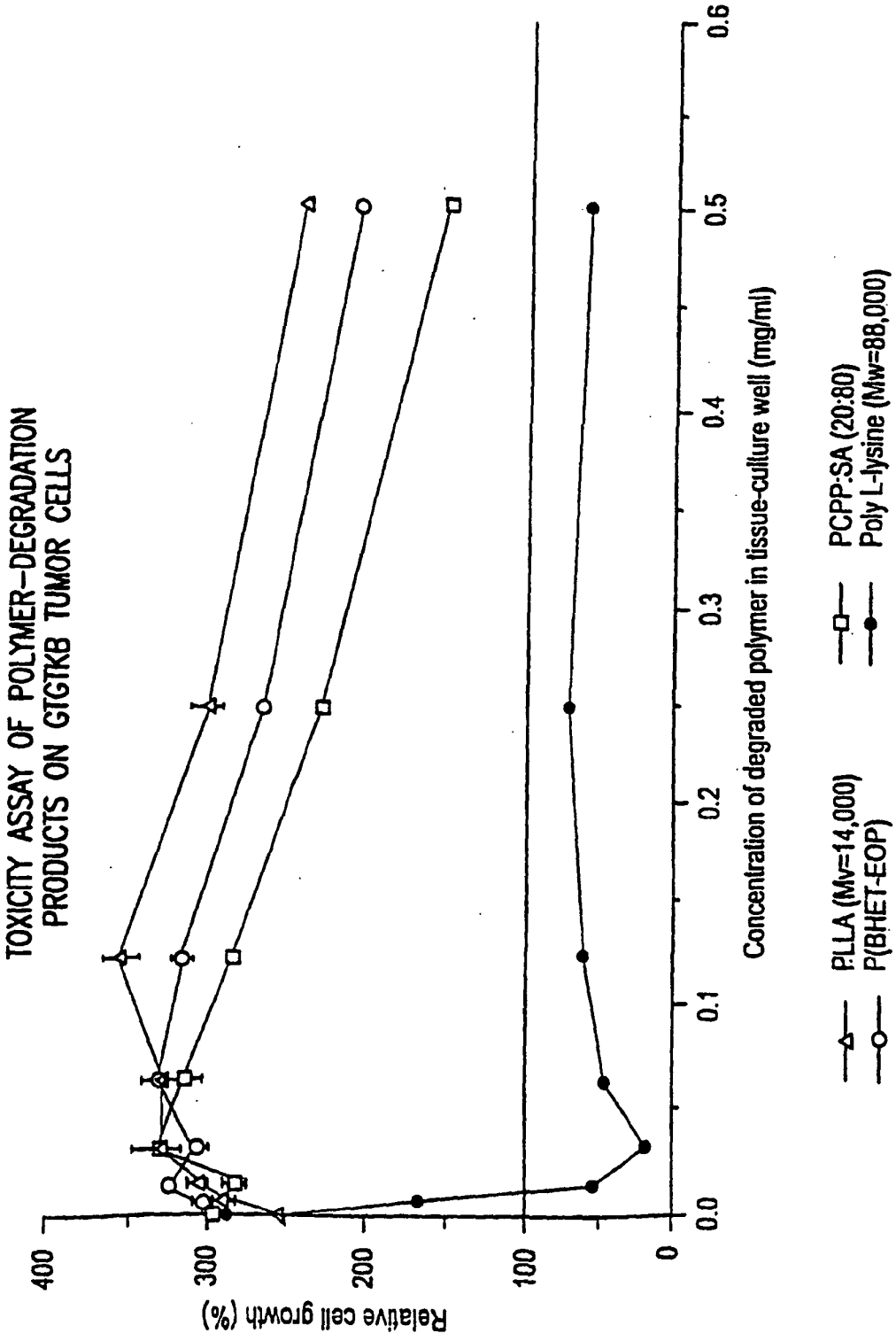

TOXICITY ASSAY OF POLYMER-DEGRADATION PRODUCTS ON GTGTKB TUMOR CELLS
Relative cell growth (%)
400
300
200
100
0
0.0
0.1
0.2
0.3
0.4
0.5
0.6
Concentration of degraded polymer in tissue-culture well (mg/ml)
PLLA (Mv=14,000)
P(BHET-EOP)
PCPP:SA (20:80)
Poly L-lysine (Mw=88,000)

FIG.12

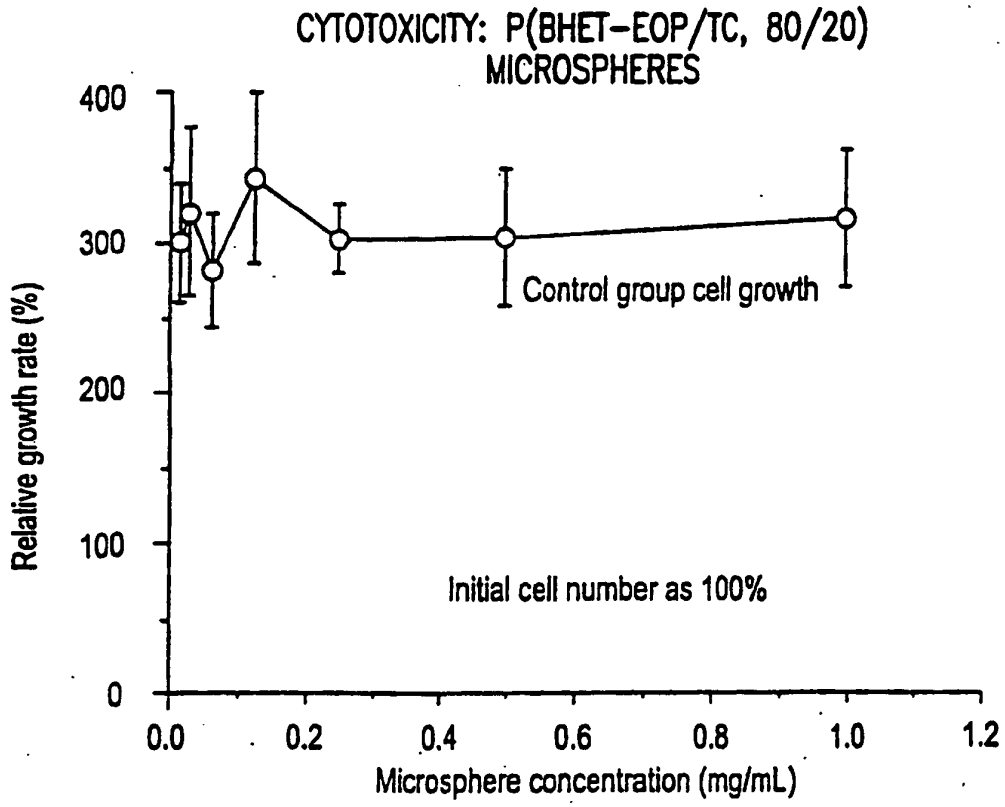

CYTOTOXICITY: P(BHET-EOP/TC, 80/20)
MICROSPHERES
Relative growth rate (%)
400
300
200
100
0
Control group cell growth
Initial cell number as 100%
0.0
0.2
0.4
0.6
0.8
1.0
1.2
Microsphere concentration (mg/mL)

FIG.13

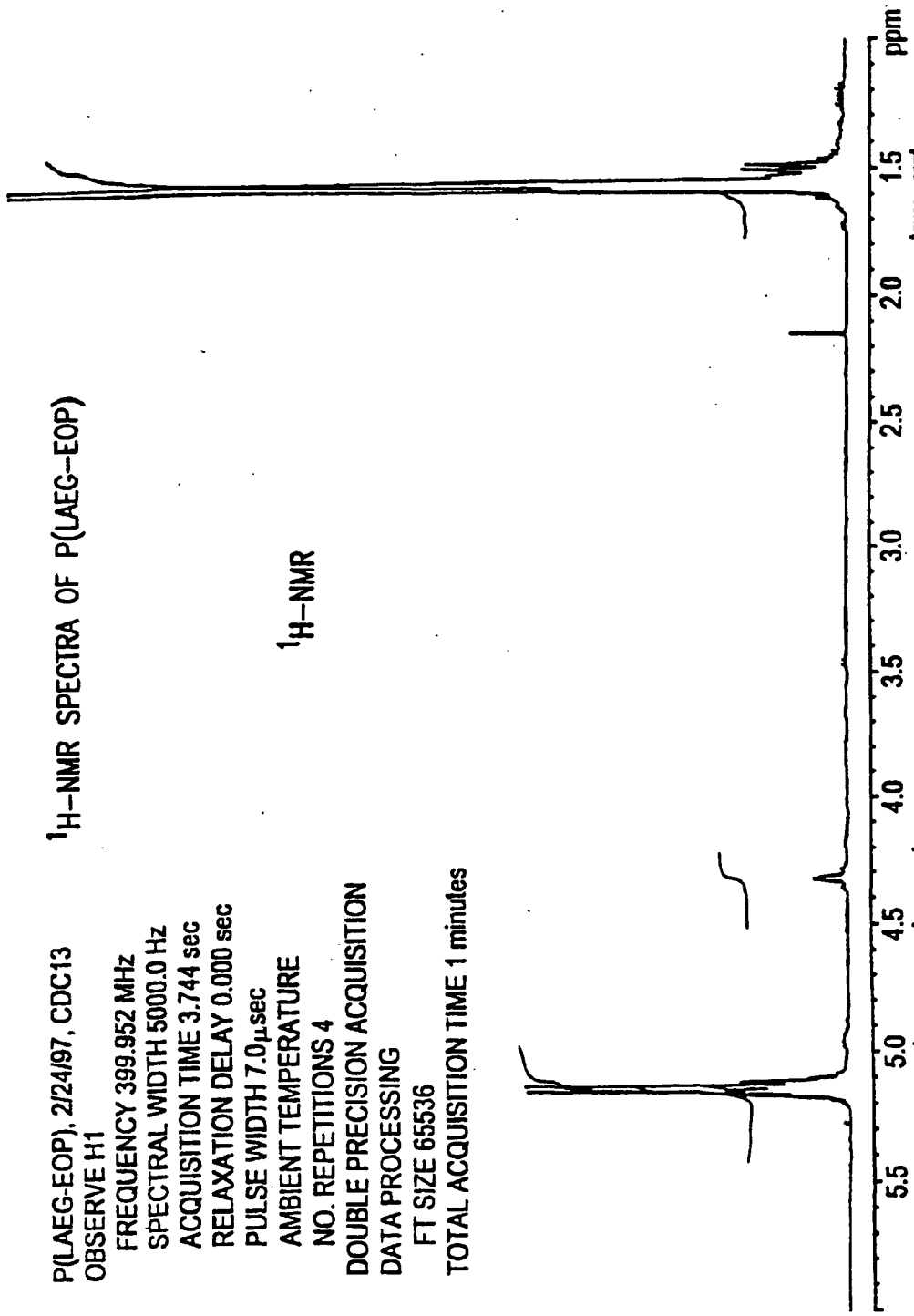

P(LAEG-EOP), 2/24/97, CDC13
OBSERVE H1
FREQUENCY 399.952 MHz
SPECTRAL WIDTH 5000.0 Hz
ACQUISITION TIME 3.744 sec
RELAXATION DELAY 0.000 sec
PULSE WIDTH 7.0 μsec
AMBIENT TEMPERATURE
NO. REPETITIONS 4
DOUBLE PRECISION ACQUISITION
DATA PROCESSING
FT SIZE 65536
TOTAL ACQUISITION TIME 1 minutes
1H-NMR SPECTRA OF P(LAEG-EOP)
1H-NMR
5.5
5.0
4.5
4.0
3.5
3.0
2.5
2.0
1.5
ppm
6.59
0.78
20.19

FIG.14

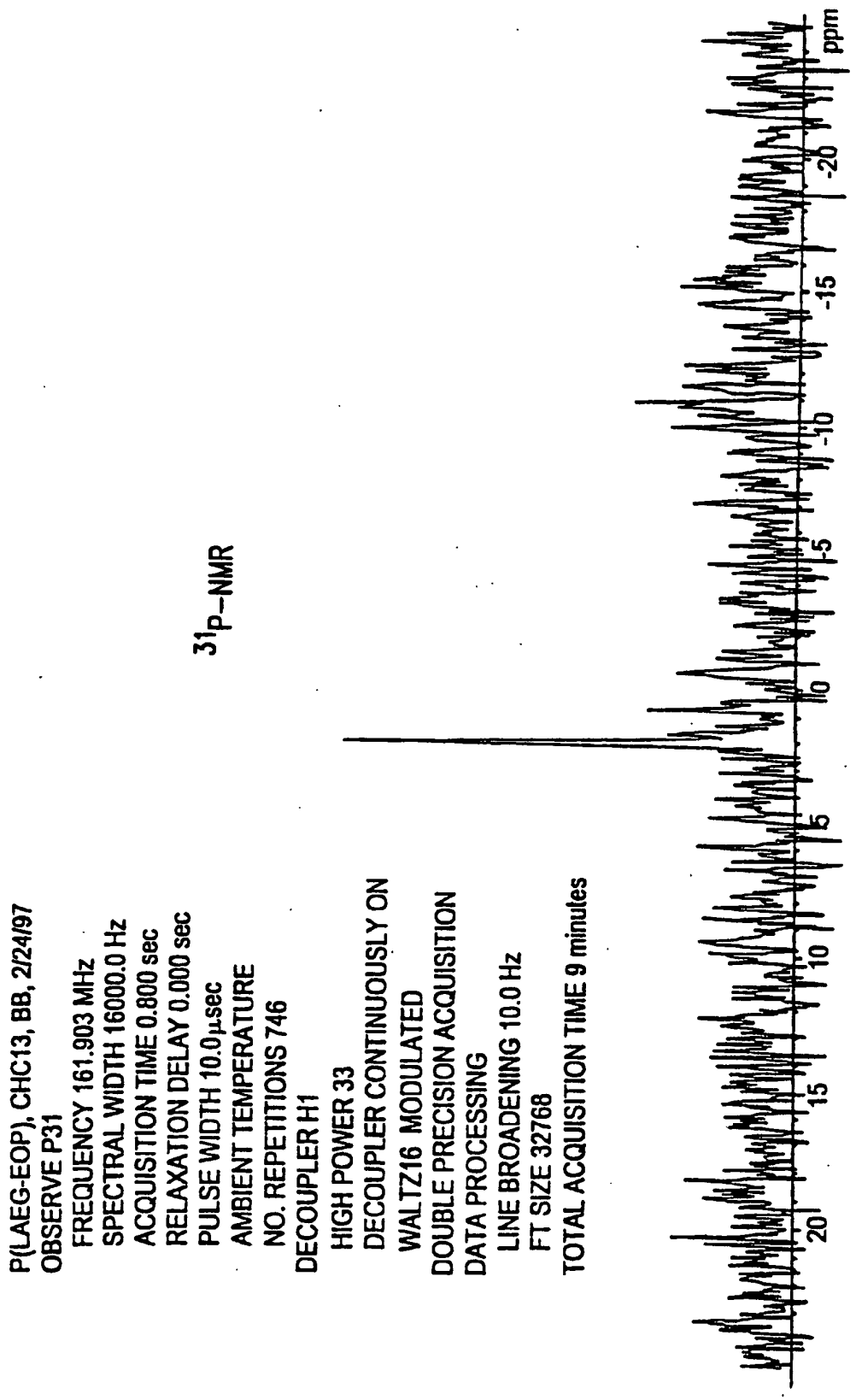

P(LAEG-EOP), CHC13, BB, 2/24/97
OBSERVE P31
FREQUENCY 161.903 MHz
SPECTRAL WIDTH 16000.0 Hz
ACQUISITION TIME 0.800 sec
RELAXATION DELAY 0.000 sec
PULSE WIDTH 10.0 µsec
AMBIENT TEMPERATURE
NO. REPETITIONS 746
DECOUPLER H1
HIGH POWER 33
DECOUPLER CONTINUOUSLY ON
WALTZ16 MODULATED
DOUBLE PRECISION ACQUISITION
DATA PROCESSING
LINE BROADENING 10.0 Hz
FT SIZE 32768
TOTAL ACQUISITION TIME 9 minutes
31P-NMR
20
15
10
5
0
-5
-10
-15
-20
ppm

FIG.15

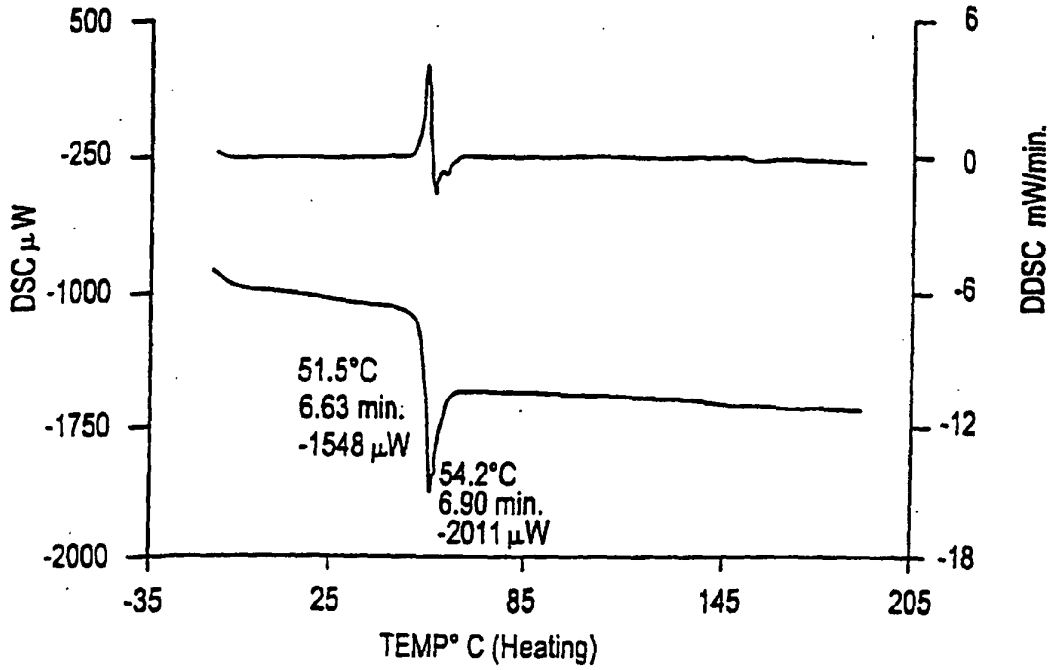


FIG.16A

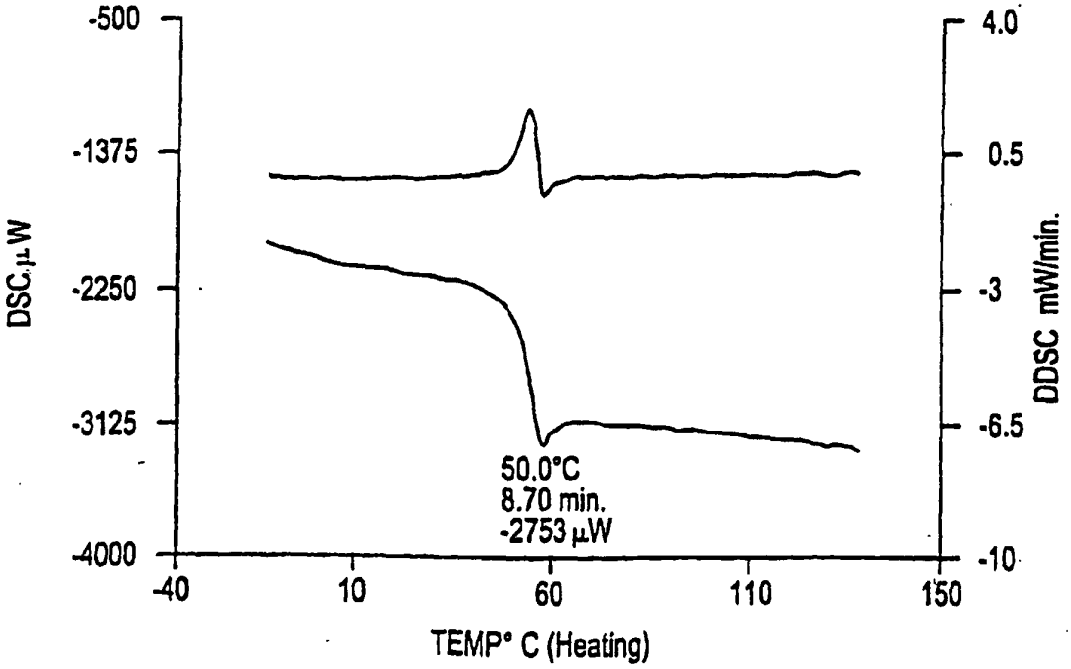
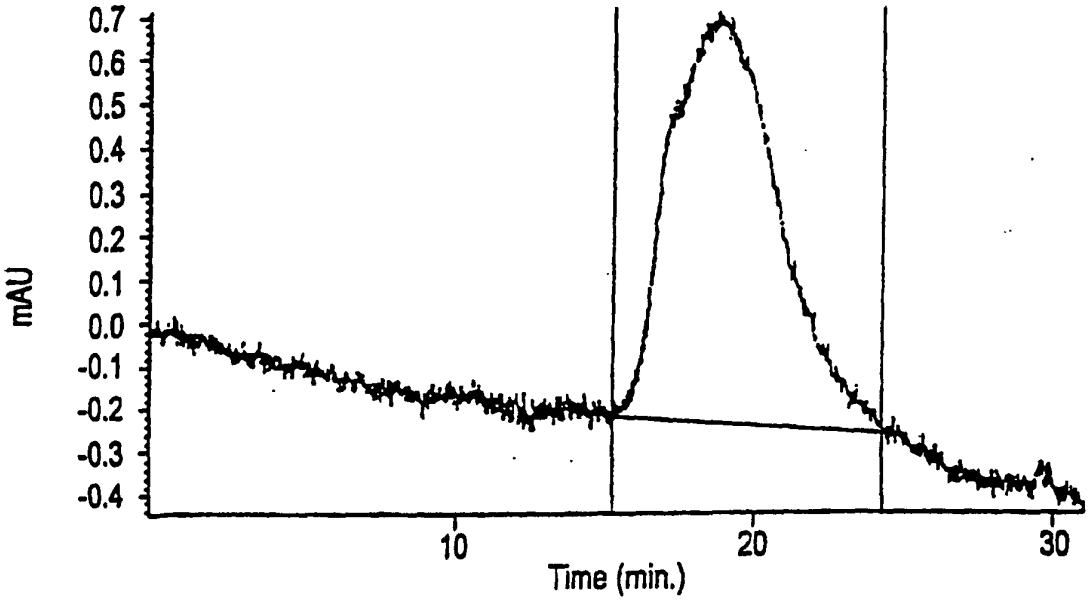


FIG.16B



Mn= 4752	Mw/Mn= 6.9421	MV= 0
Mw= 32991	Mz/Mw= 3.3116	Int. Visc.= 0.00000
Mz=109253	M(z+1)= 218140	SL Slope= -0.00745
Mp= 13123	M(Z+2)= 335155	BL Slope= -0.10995

FIG.17

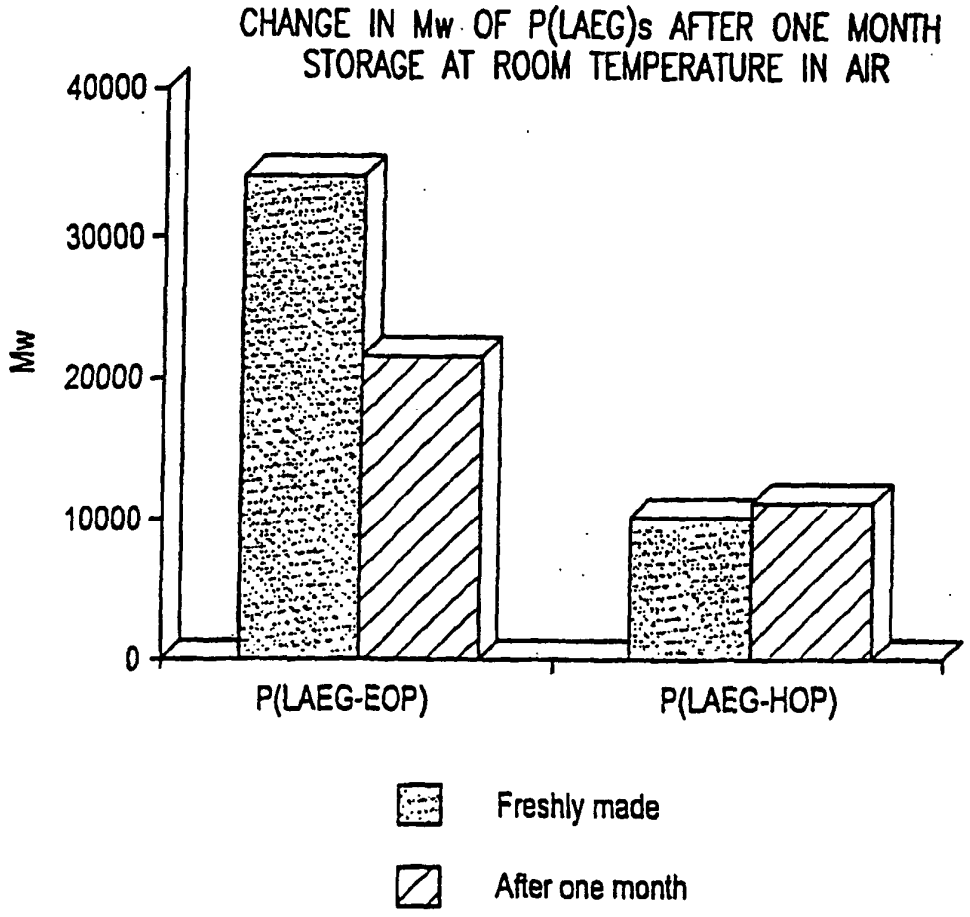

CHANGE IN Mw OF P(LAEG)s AFTER ONE MONTH STORAGE AT ROOM TEMPERATURE IN AIR
Mw
40000
30000
20000
10000
0
P(LAEG-EOP)
P(LAEG-HOP)
Freshly made
After one month

FIG.18

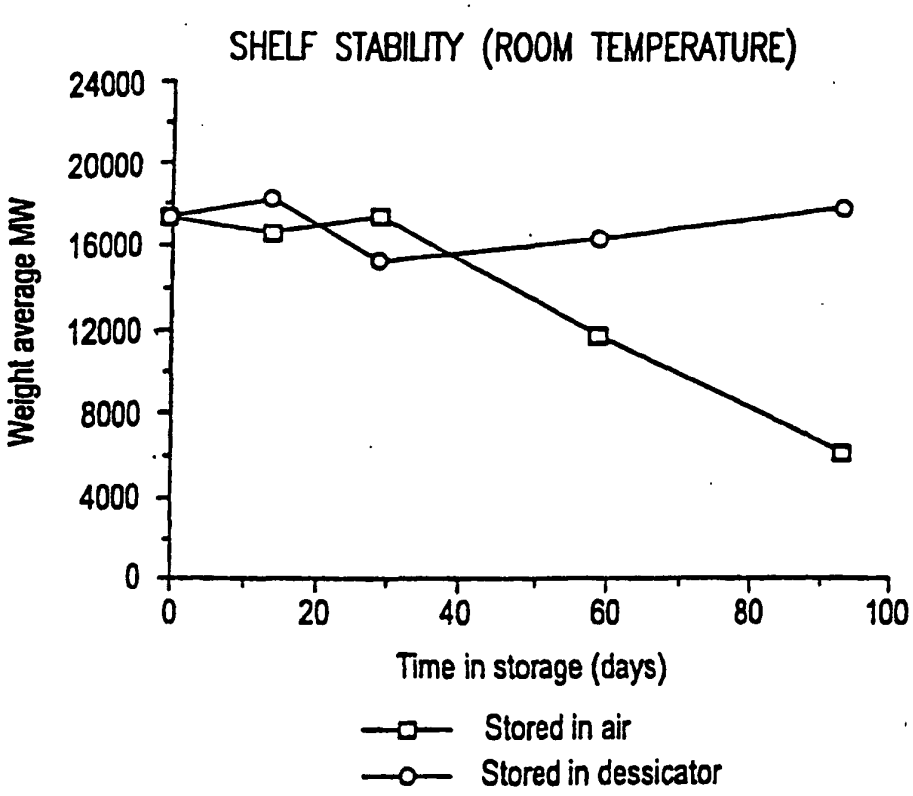

SHELF STABILITY (ROOM TEMPERATURE)
Weight average MW
24000
20000
16000
12000
8000
4000
0
0
20
40
60
80
100
Time in storage (days)
Stored in air
Stored in dessicator

FIG.19

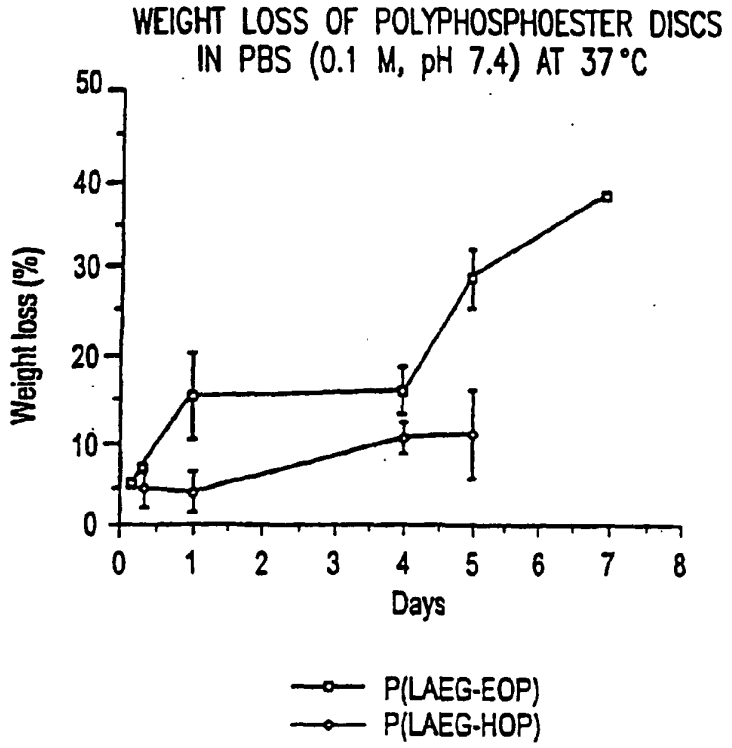


FIG.20A

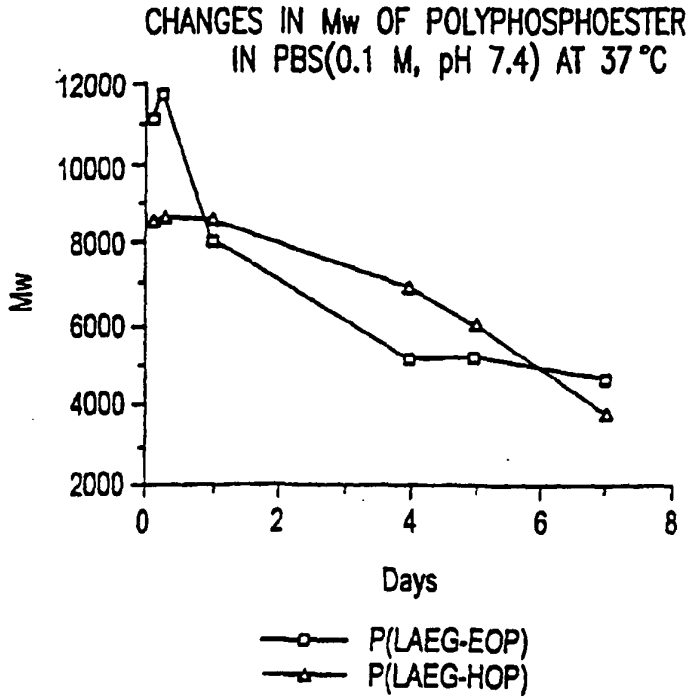


FIG.20B

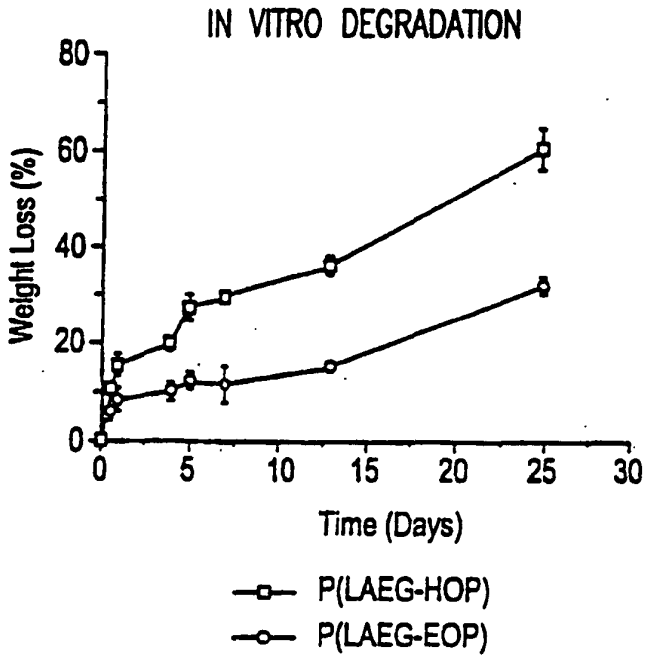


FIG.21A

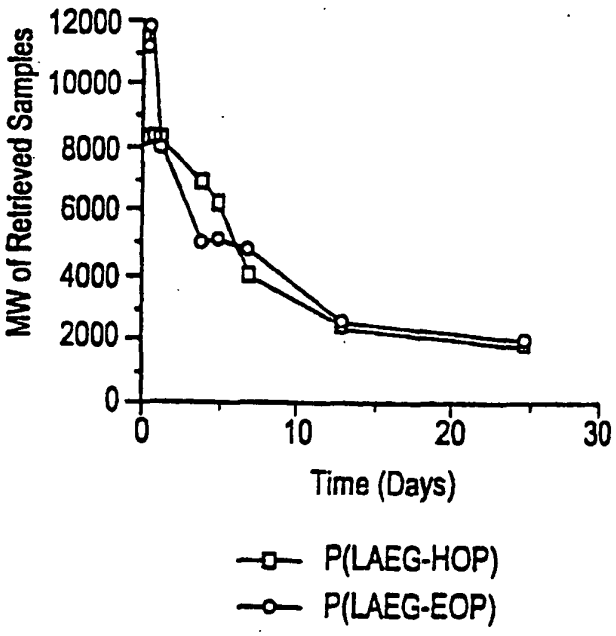


FIG.21B

BIOCOMPATIBILITY:
HISTOPATHOLOGICAL ANALYSIS

Inflammatory Response at the Site of Implantation (i.m.)						
Polymer	3 D	7 D	14 D	1 M	2 M	4 M
P(LAEG-EOP)	SI (130)	SI (123)	SI (180)	SI (198)	SI (106)	SI (99)
PLGA(RG755)	SI (148)	SI (98)	SI (137)	SI (105)	SI (94)	SI (43)

Score:	No Irritation (0)	Slight Irritation (0-200)	Mild Irritation (200-400)	Moderate Irritation (400-600)	Severe Irritation (>600)

FIG.22

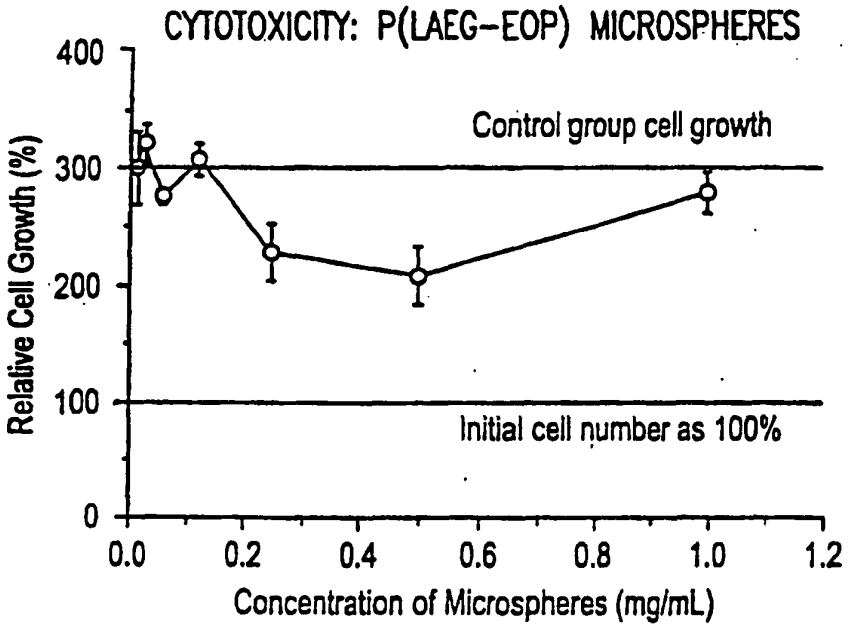

CYTOTOXICITY: P(LAEG-EOP) MICROSPHERES
Relative Cell Growth (%)
400
300
200
100
0
Control group cell growth
Initial cell number as 100%
0.0
0.2
0.4
0.6
0.8
1.0
1.2
Concentration of Microspheres (mg/mL)

FIG.23

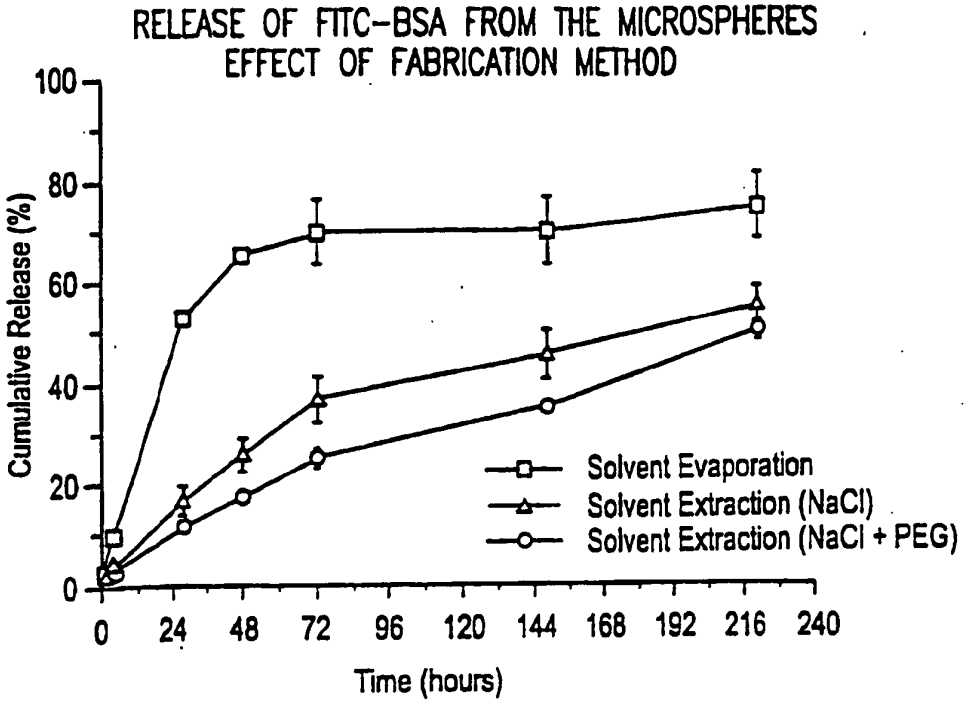


FIG.24A

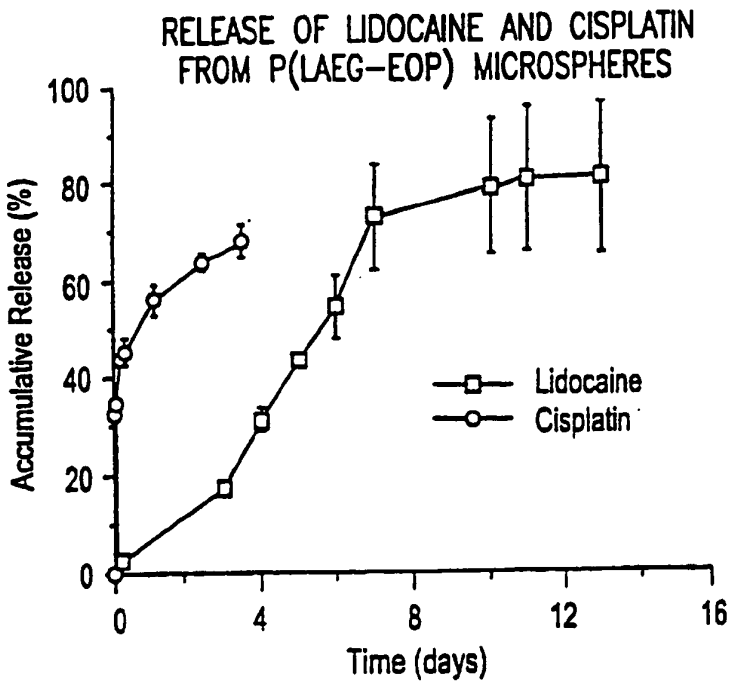


FIG.24B

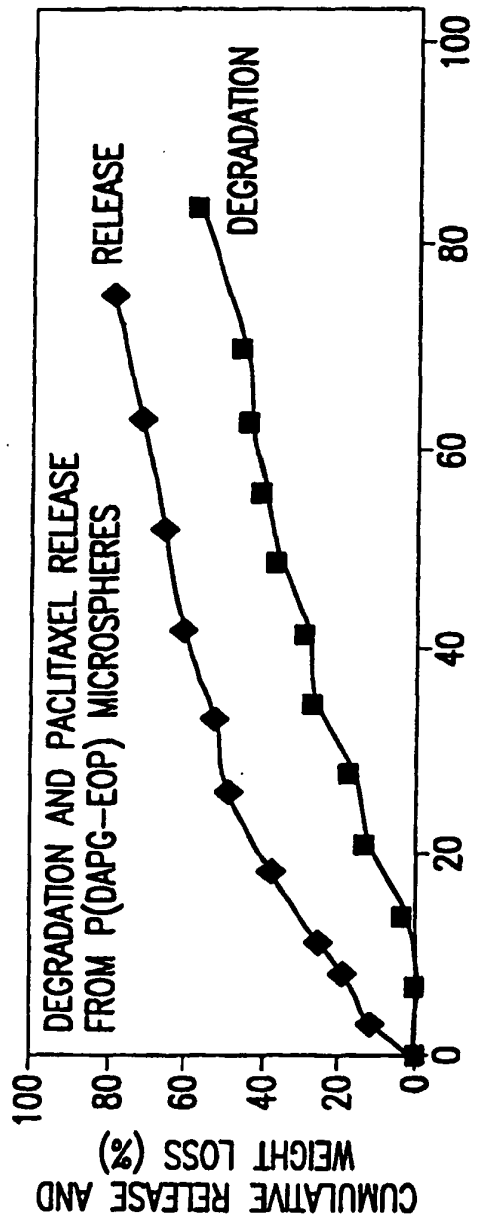

DEGRADATION AND PACLITAXEL RELEASE
FROM P(DAPG-EOP) MICROSPHERES
RELEASE
DEGRADATION
CUMULATIVE RELEASE AND
WEIGHT LOSS (%)
TIME (DAY)
0
20
40
60
80
100

FIG.25A

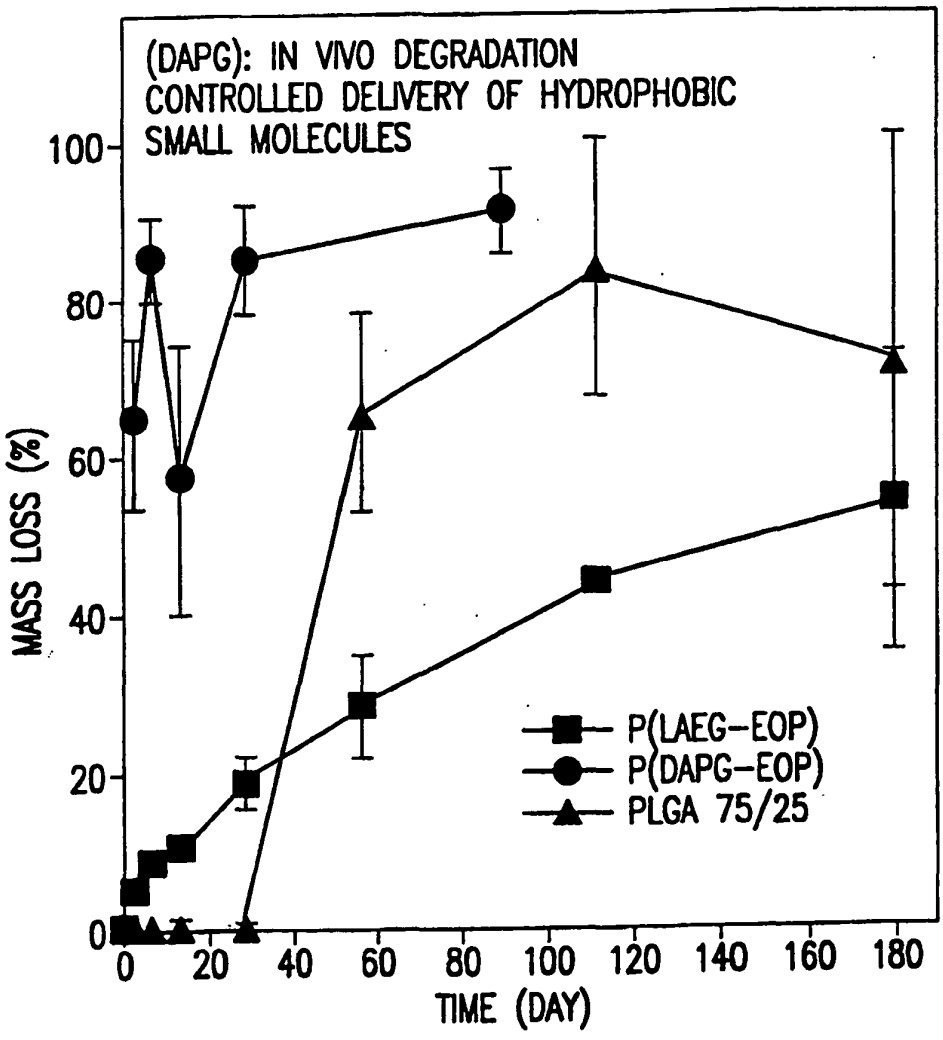

(DAPG): IN VIVO DEGRADATION
CONTROLLED DELIVERY OF HYDROPHOBIC
SMALL MOLECULES
MASS LOSS (%)
TIME (DAY)
P(LAEG-EOP)
P(DAPG-EOP)
PLGA 75/25
0
20
40
60
80
100
120
140
160
180

FIG.25B

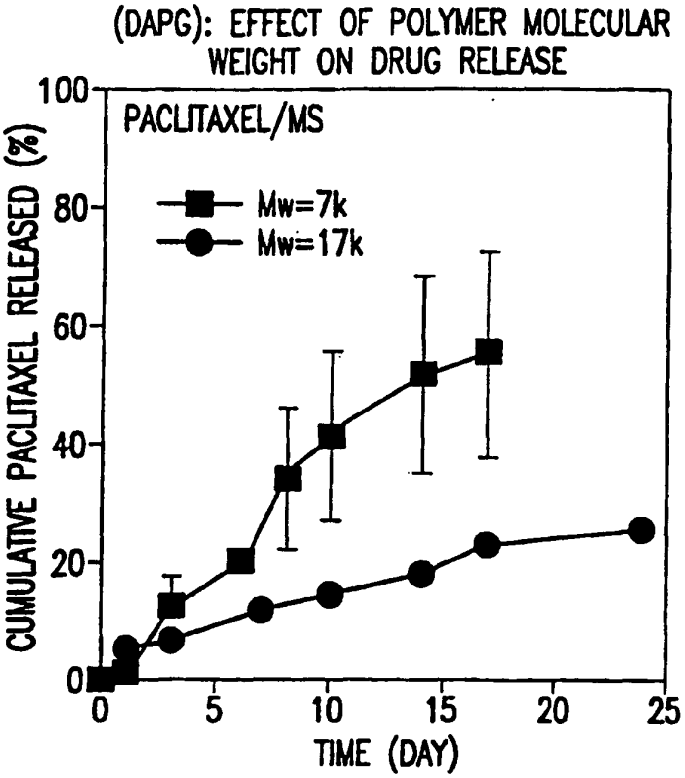


FIG.25C

PLEASE NOTE: Mw IS RELATIVE TO POLYSTYRENE STANDARDS (GPC, RI DETECTOR)

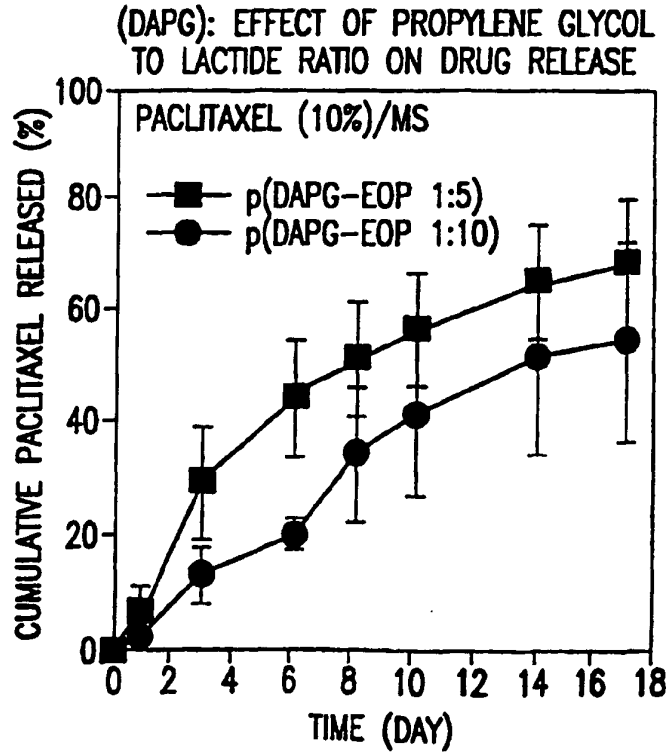


FIG.25D

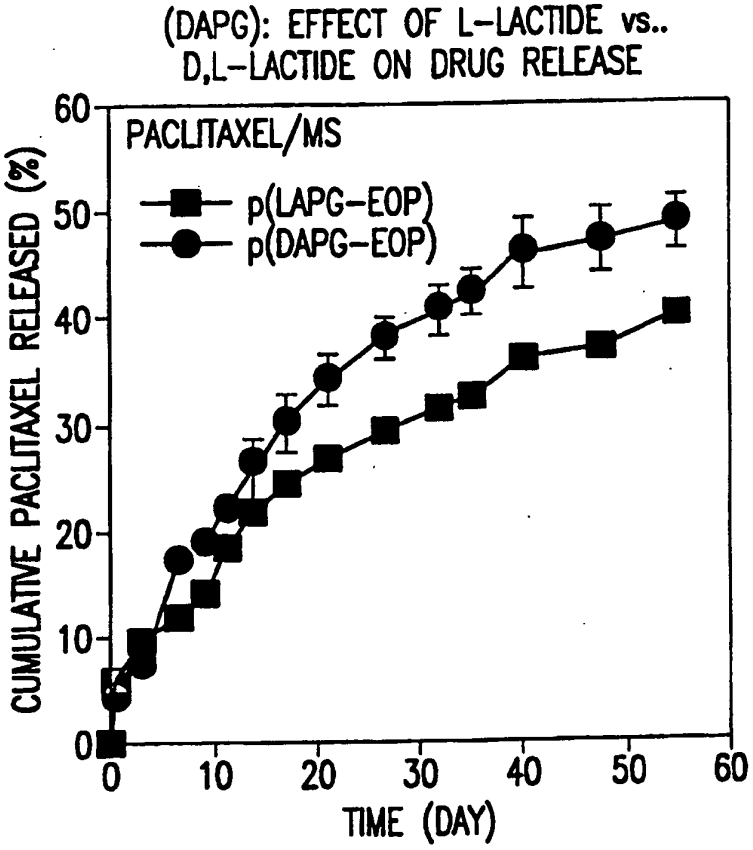

(DAPG): EFFECT OF L-LACTIDE vs..
D,L-LACTIDE ON DRUG RELEASE
PACLITAXEL/MS
p(LAPG-EOP)
p(DAPG-EOP)
CUMULATIVE PACLITAXEL RELEASED (%)
TIME (DAY)
0
10
20
30
40
50
60

FIG.25E

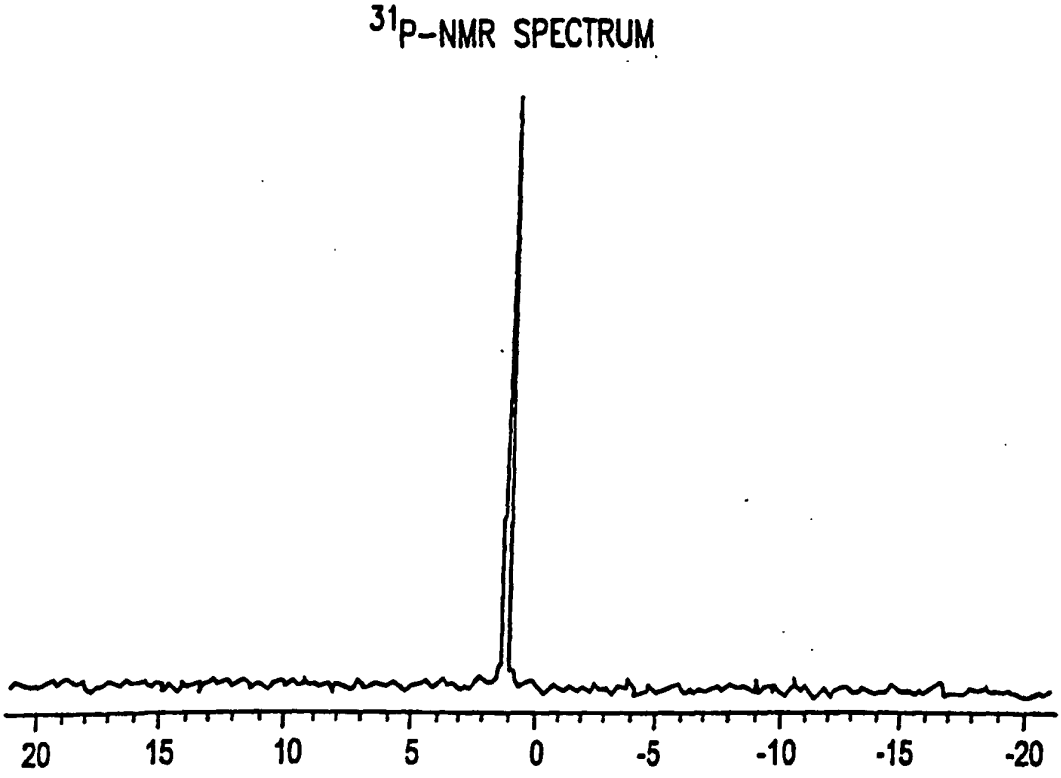


FIG. 26A

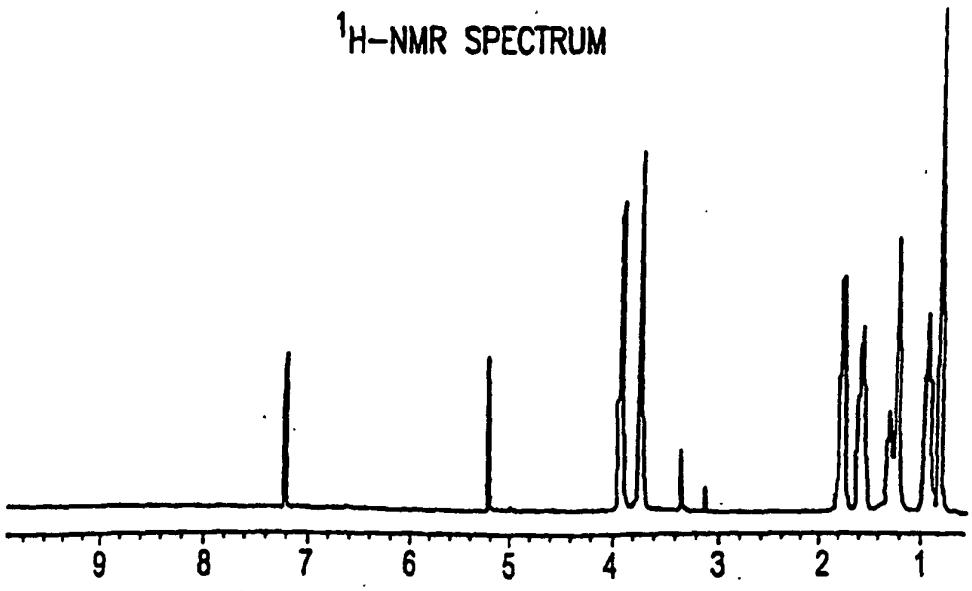
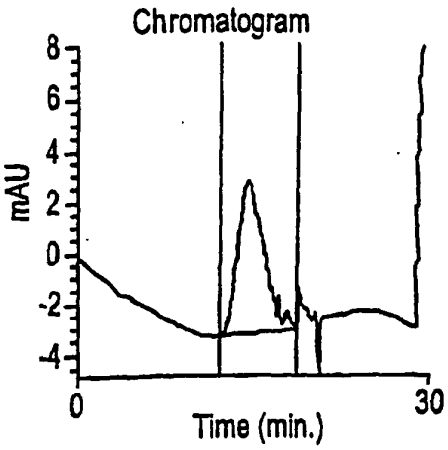
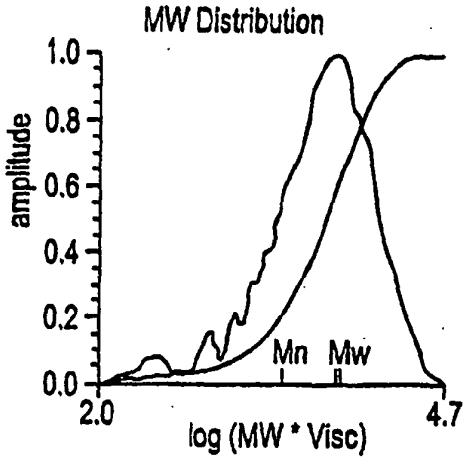


FIG. 26B



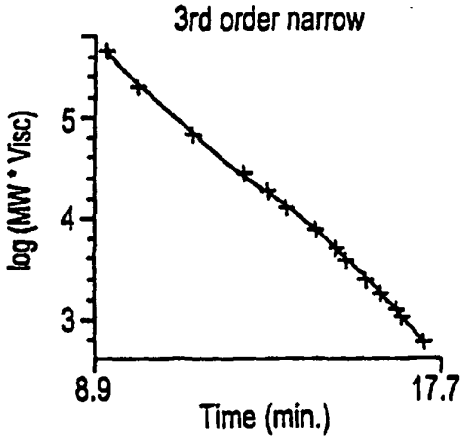
Mn=3076 Mp=8092
Mw=8584 Mv=0
Mz=14427 0=2.7910

FIG.27A



Mn=3076 Mp=8092
Mw=8584 Mv=0
Mz=14427 0=2.7910

FIG.27B



Mn=3076 Mp=8092
Mw=8584 Mv=0
Mz=14427 0=2.7910

FIG.27C

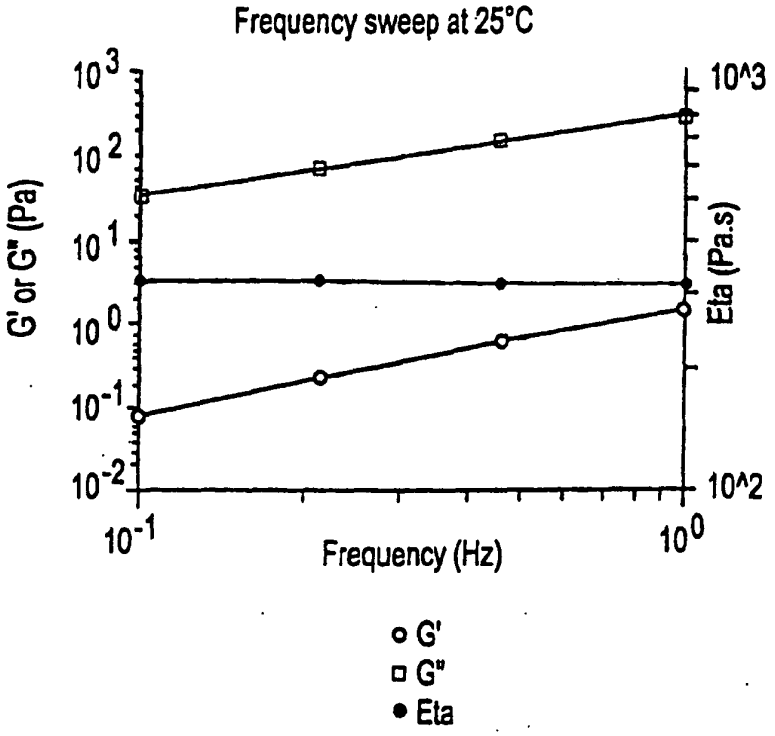


FIG.28A

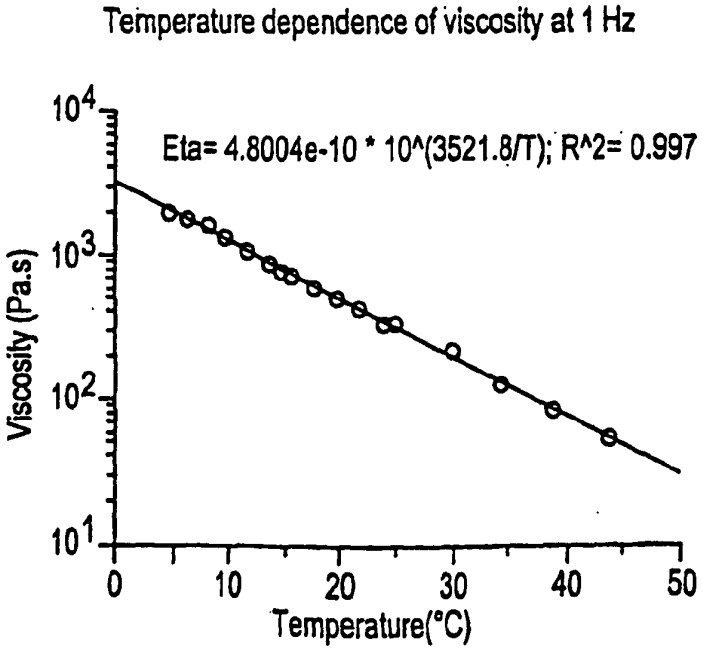


FIG.28B

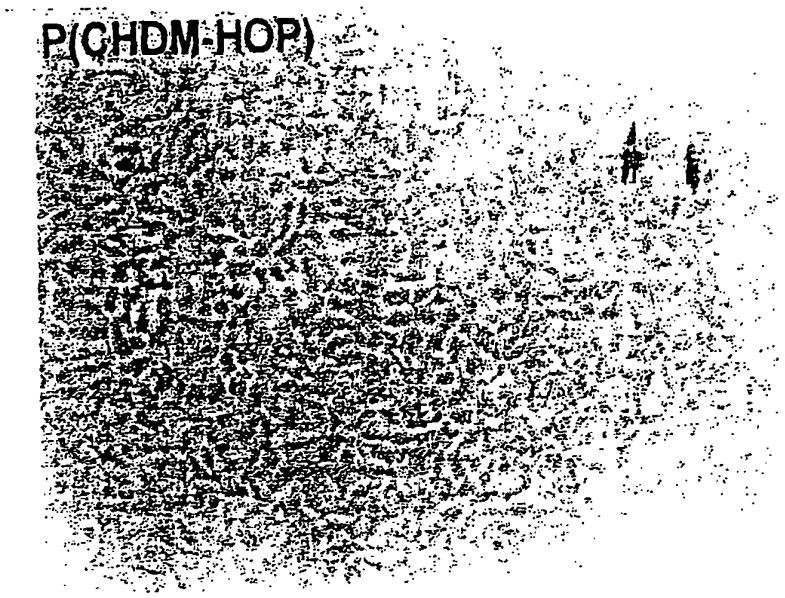


FIG.29A

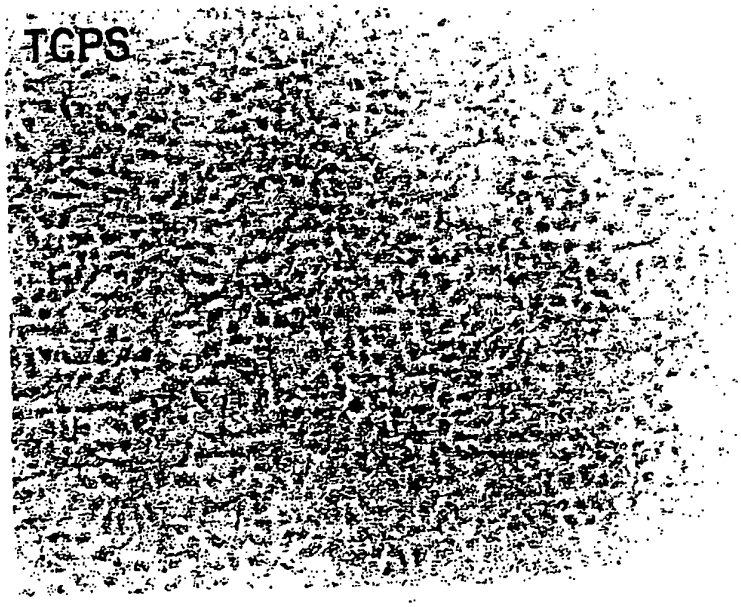
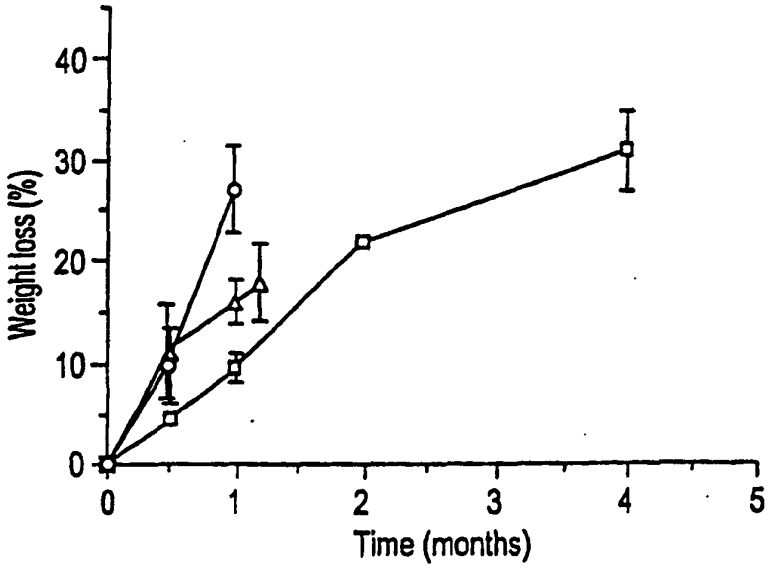


FIG.29B



P(CHDM-HOP)
P(CHDM-BOP)
P(CHDM-EOP)

FIG.30

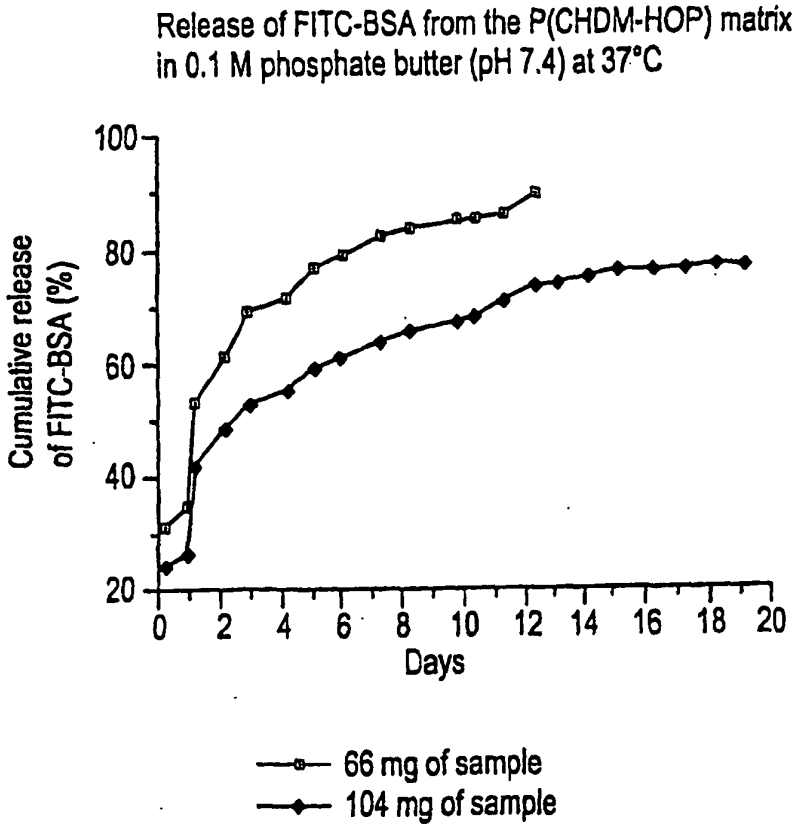

Release of FITC-BSA from the P(CHDM-HOP) matrix
in 0.1 M phosphate butter (pH 7.4) at 37°C
Cumulative release
of FITC-BSA (%)
100
80
60
40
20
0
2
4
6
8
10
12
14
16
18
20
Days
66 mg of sample
104 mg of sample

FIG.31

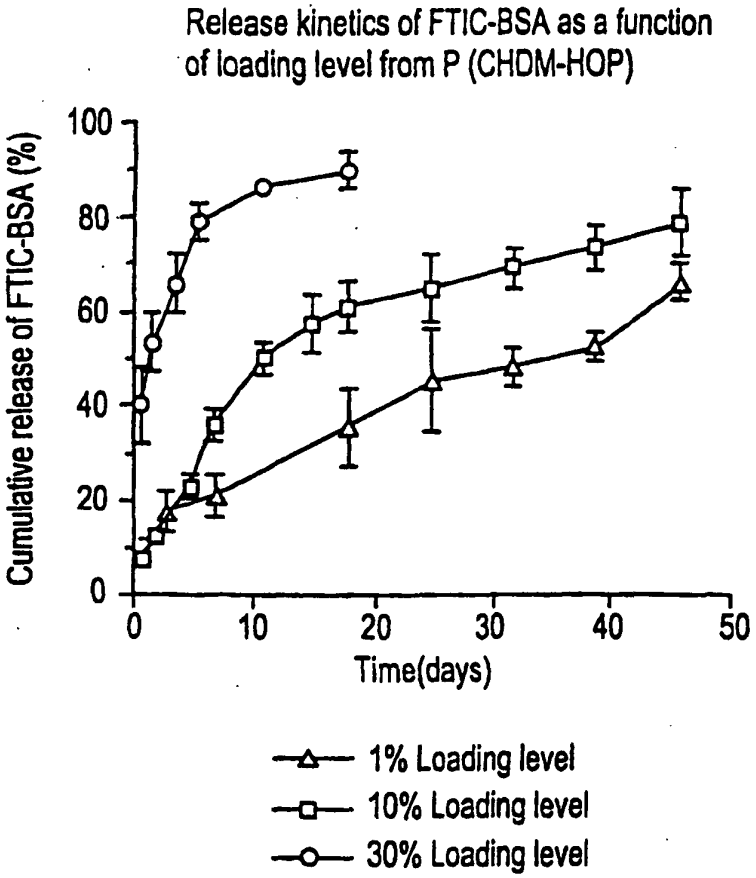

Release kinetics of FTIC-BSA as a function of loading level from P (CHDM-HOP)
Cumulative release of FTIC-BSA (%)
100
80
60
40
20
0
0
10
20
30
40
50
Time(days)
1% Loading level
10% Loading level
30% Loading level

FIG.32

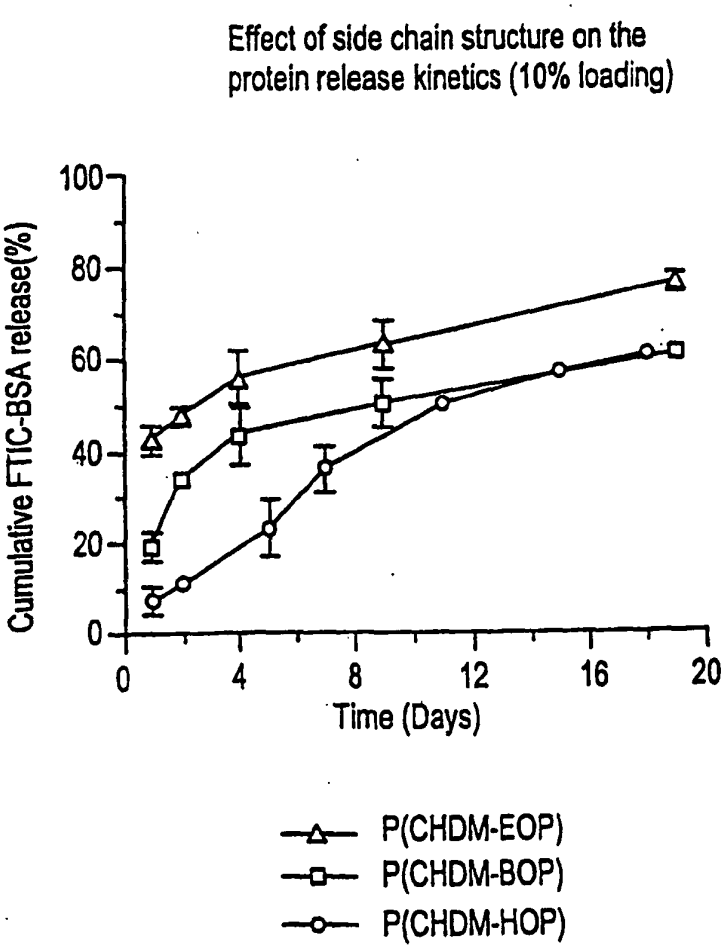

Effect of side chain structure on the protein release kinetics (10% loading)
Cumulative FTIC-BSA release(%)
100
80
60
40
20
0
0
4
8
12
16
20
Time (Days)
P(CHDM-EOP)
P(CHDM-BOP)
P(CHDM-HOP)

FIG.33

Release of low molecular weight drug from P(CHDM-HOP).

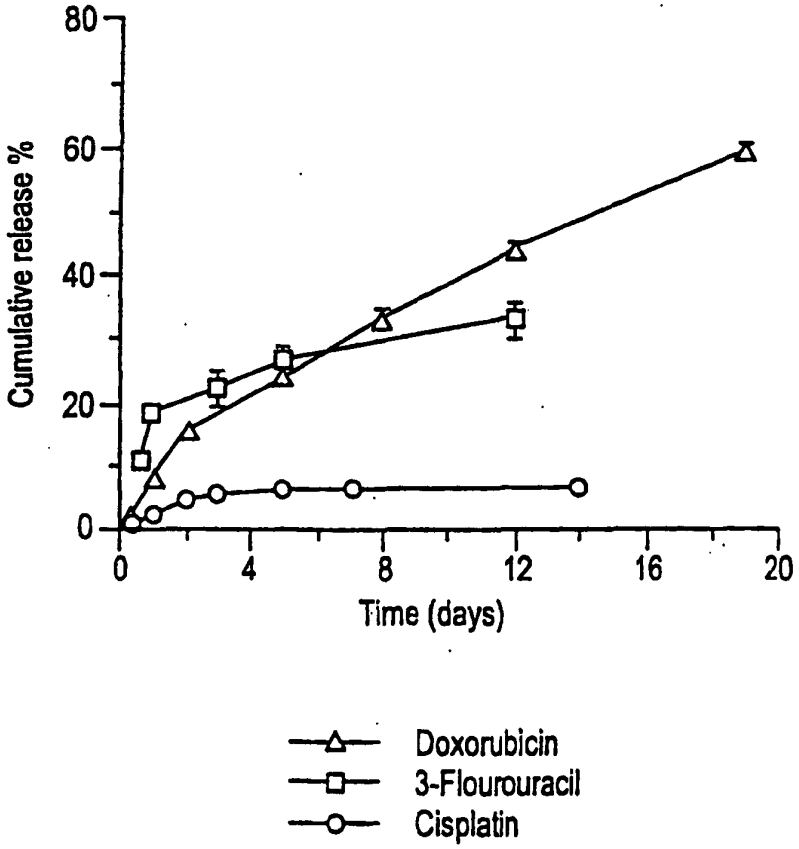


FIG.34

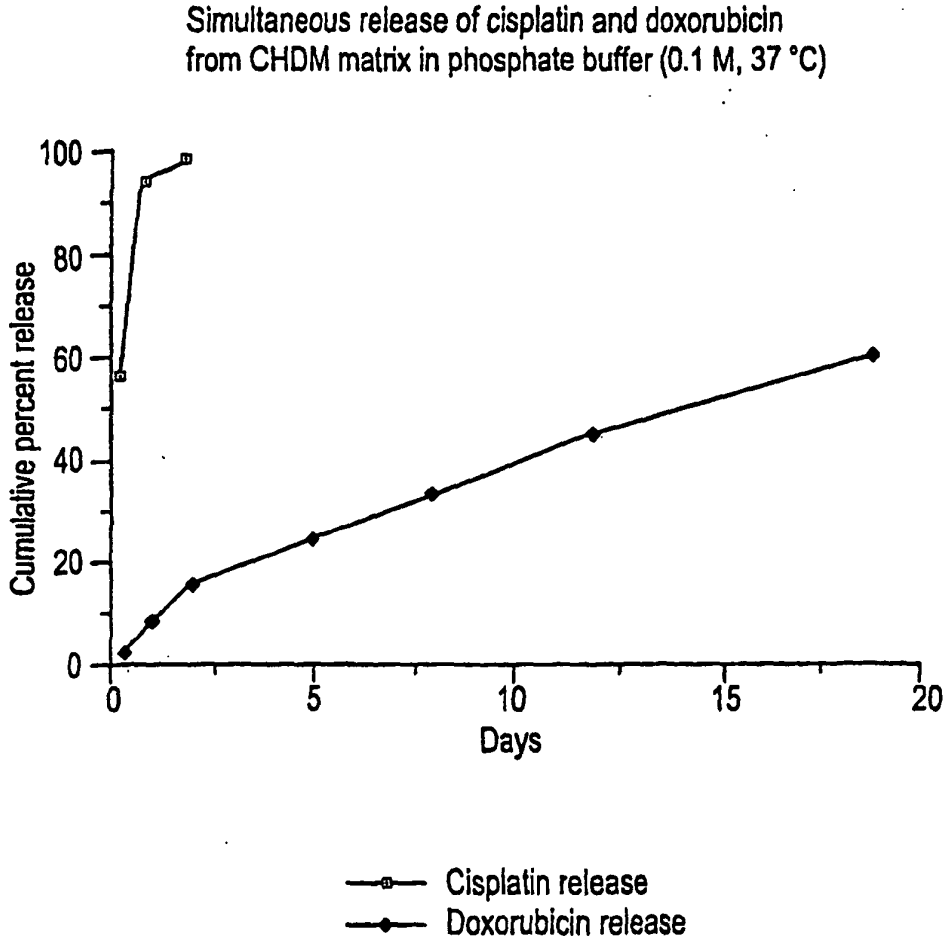

Simultaneous release of cisplatin and doxorubicin
from CHDM matrix in phosphate buffer (0.1 M, 37 °C)
Cumulative percent release
100
80
60
40
20
0
0
5
10
15
20
Days
Cisplatin release
Doxorubicin release

FIG.35

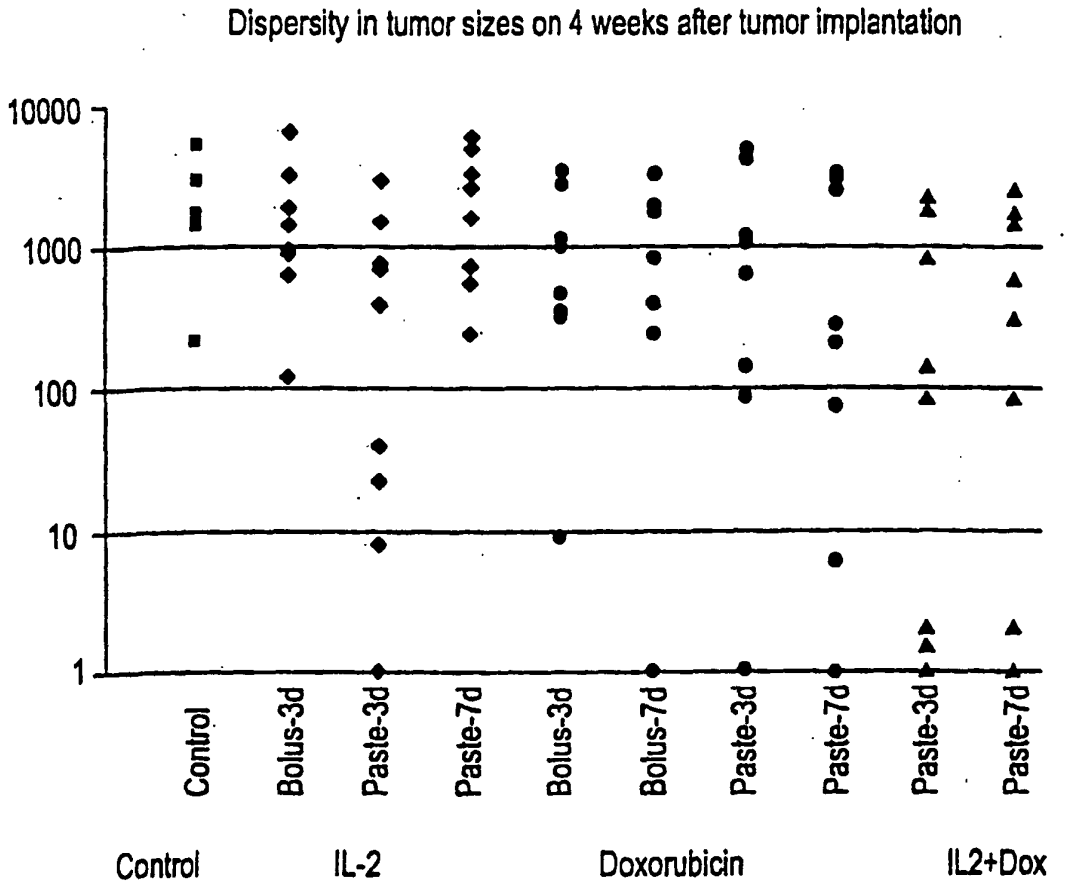

Dispersity in tumor sizes on 4 weeks after tumor implantation
10000
1000
100
10
1
Control
Bolus-3d
Paste-3d
Paste-7d
Bolus-3d
Bolus-7d
Paste-3d
Paste-7d
Paste-3d
Paste-7d
Control
IL-2
Doxorubicin
IL2+Dox

FIG.36

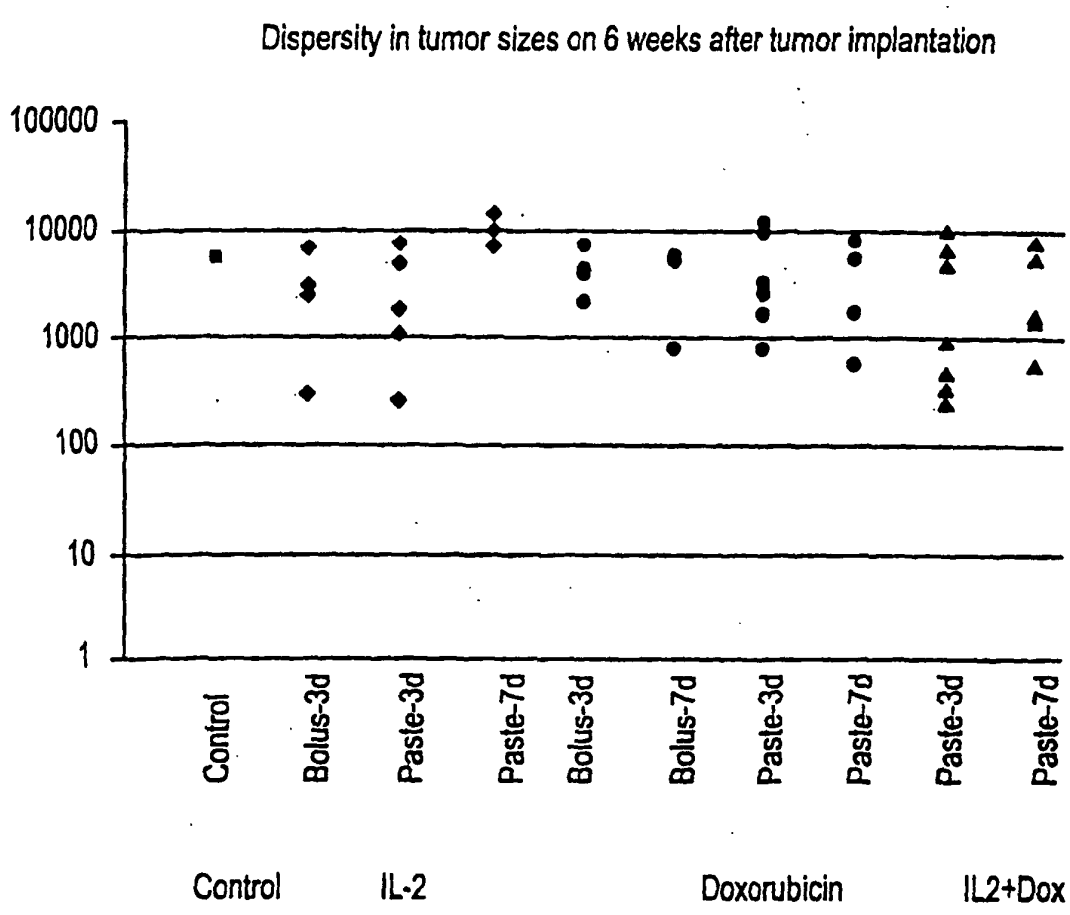

Dispersity in tumor sizes on 6 weeks after tumor implantation
100000
10000
1000
100
10
1
Control
Bolus-3d
Paste-3d
Paste-7d
Bolus-3d
Bolus-7d
Paste-3d
Paste-7d
Paste-3d
Paste-7d
Control
IL-2
Doxorubicin
IL2+Dox

FIG.37

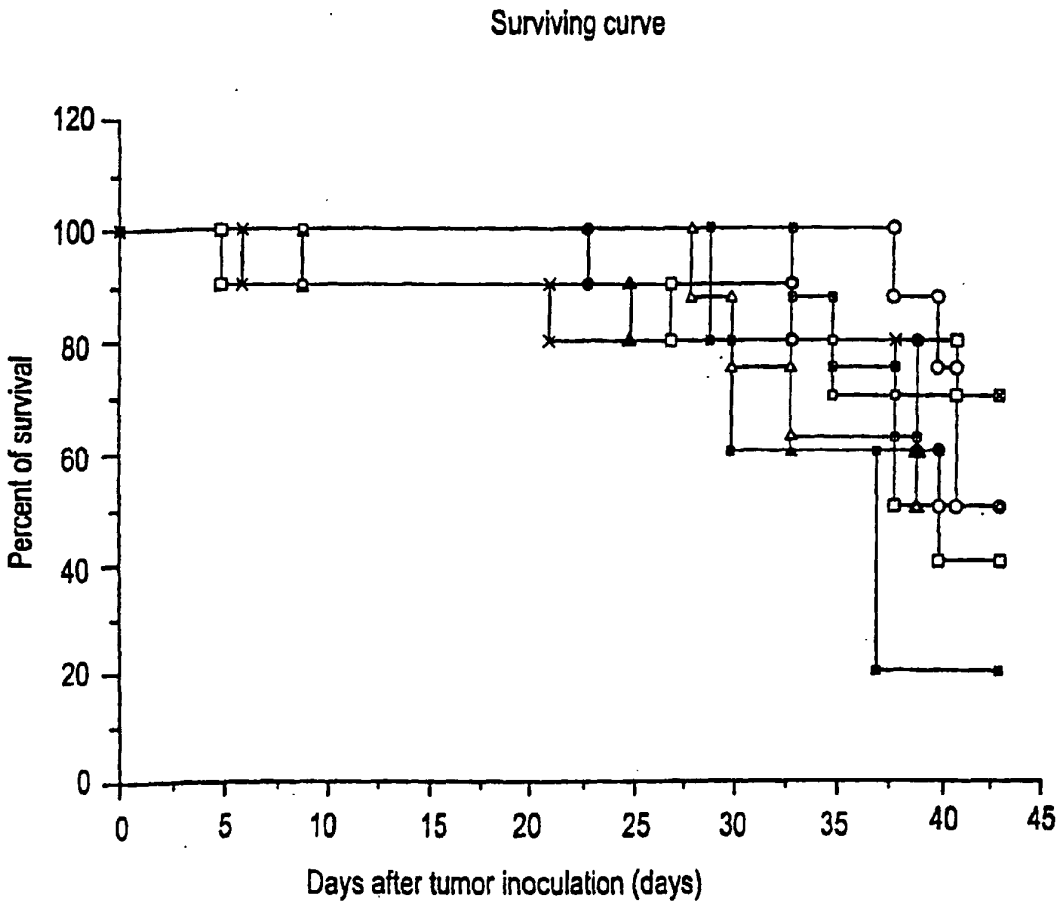

Surviving curve
Percent of survival
120
100
80
60
40
20
0
0
5
10
15
20
25
30
35
40
45
Days after tumor inoculation (days)
paste IL-2+IFN (3d)
paste IL-2+DOX (3d)
paste IL-2+DOX (7d)
paste DOX 3(d)
paste DOX 7(d)
bolus DOX (3d)
bolus DOX (7d)
bolus IL-2 (3d)
control (v)

FIG.38

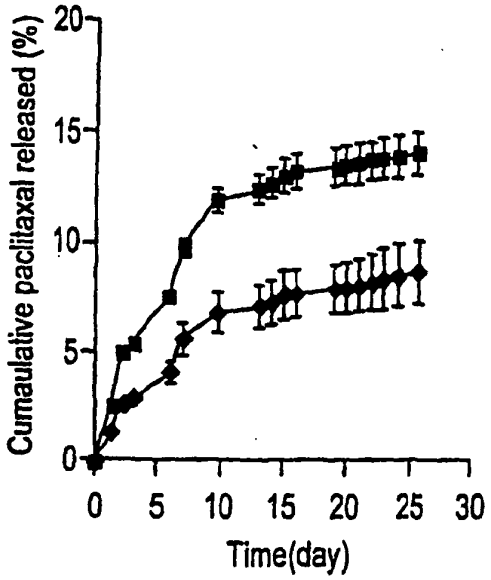

Cumaulative paclitaxal released (%)
20
15
10
5
0
0 5 10 15 20 25 30
Time(day)
p(CHDM-EOP)
p(CHDM-HOP)

FIG.39

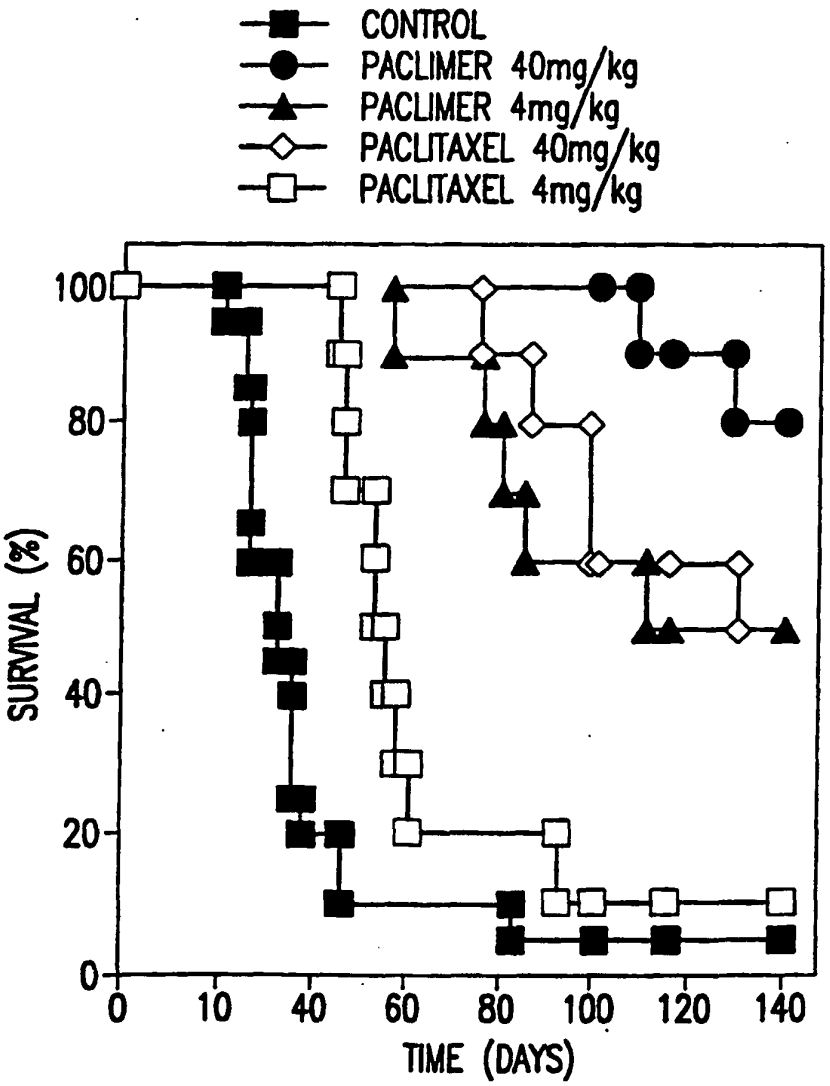

CONTROL
PACLIMER 40mg/kg
PACLIMER 4mg/kg
PACLITAXEL 40mg/kg
PACLITAXEL 4mg/kg
SURVIVAL (%)
100
80
60
40
20
0
0
10
40
60
80
100
120
140
TIME (DAYS)

FIG.40

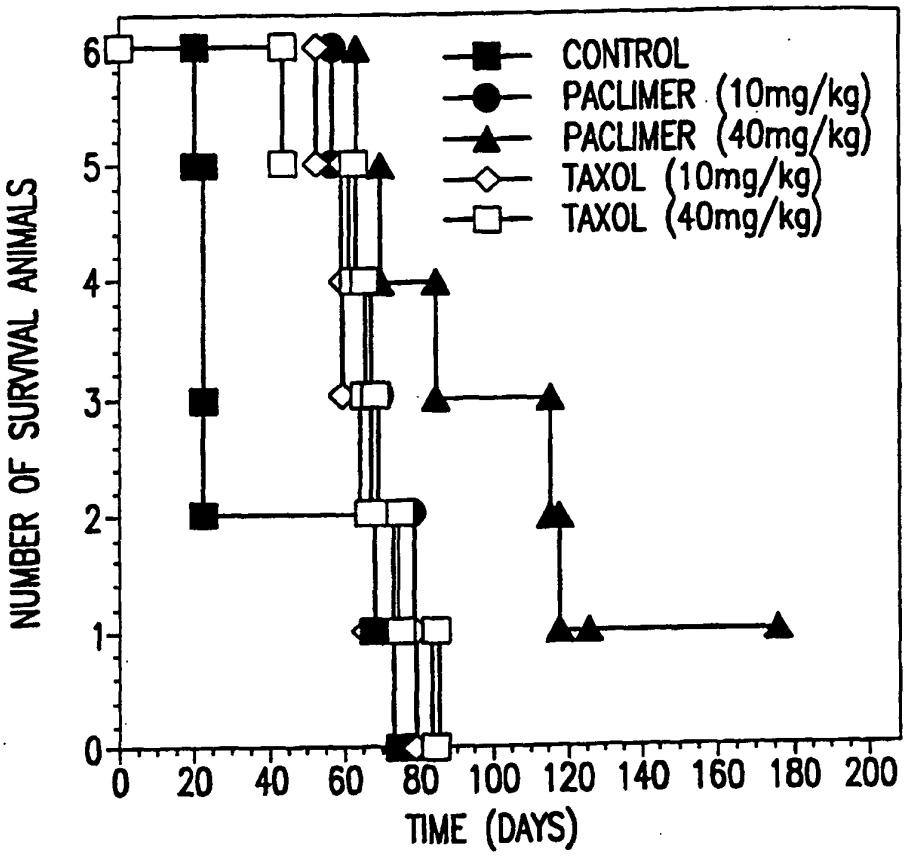

NUMBER OF SURVIVAL ANIMALS
TIME (DAYS)
CONTROL
PACLIMER (10mg/kg)
PACLIMER (40mg/kg)
TAXOL (10mg/kg)
TAXOL (40mg/kg)
0
1
2
3
4
5
6
0
20
40
60
80
100
120
140
160
180
200

FIG.41

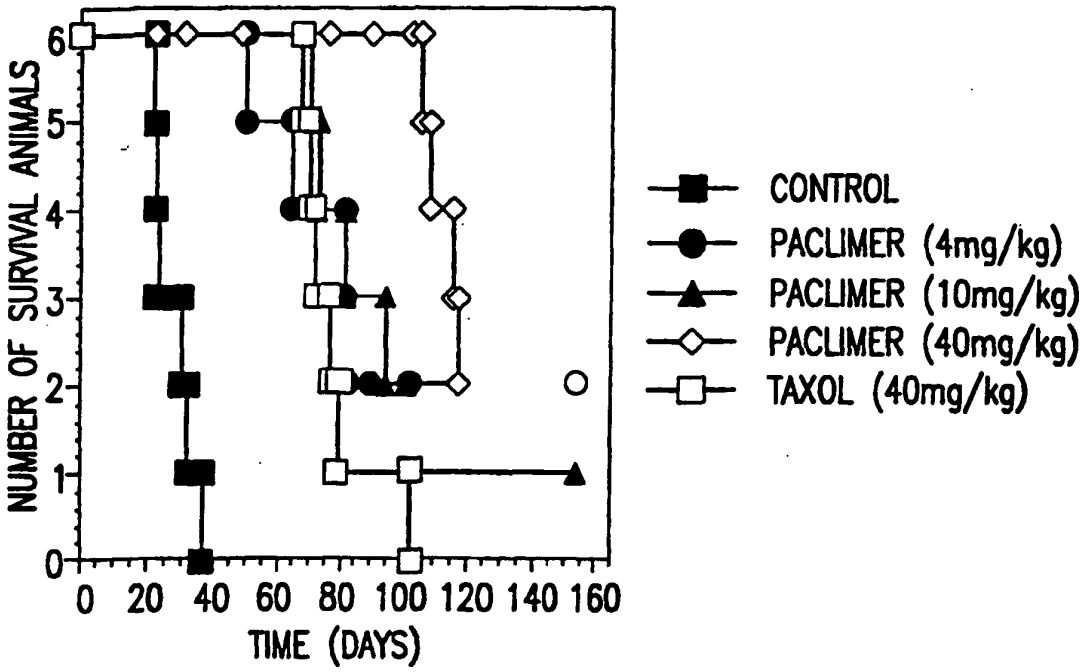

NUMBER OF SURVIVAL ANIMALS
0
1
2
3
4
5
6
0 20 40 60 80 100 120 140 160
TIME (DAYS)
CONTROL
PACLIMER (4mg/kg)
PACLIMER (10mg/kg)
PACLIMER (40mg/kg)
TAXOL (40mg/kg)

FIG. 42

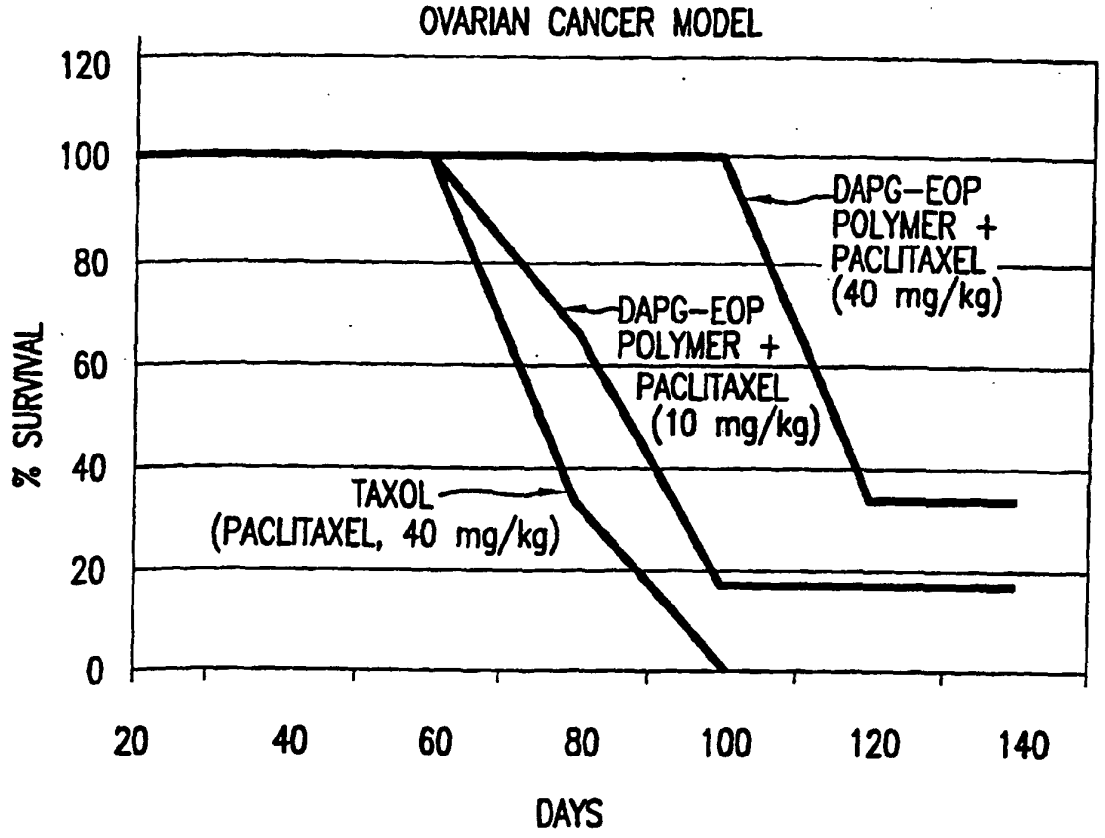

OVARIAN CANCER MODEL
120
100
80
60
40
20
0
% SURVIVAL
20
40
60
80
100
120
140
DAYS
DAPG-EOP POLYMER + PACLITAXEL (40 mg/kg)
DAPG-EOP POLYMER + PACLITAXEL (10 mg/kg)
TAXOL (PACLITAXEL, 40 mg/kg)

FIG. 43

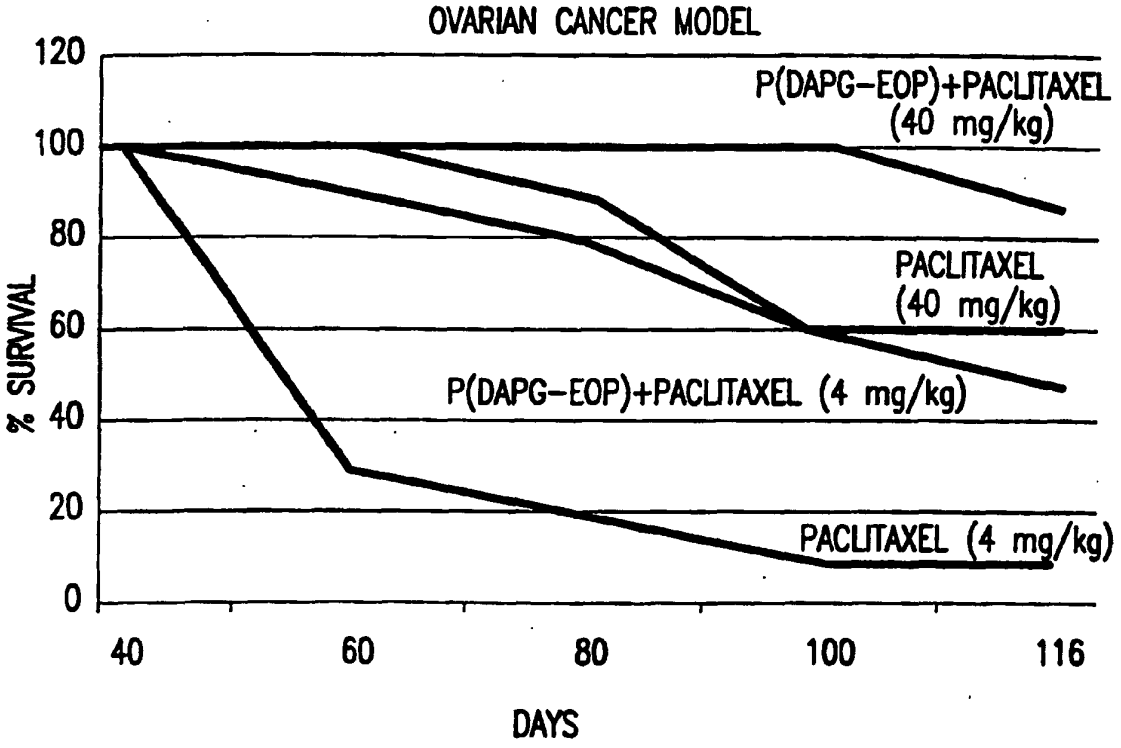

OVARIAN CANCER MODEL
120
100
80
60
40
20
0
% SURVIVAL
40
60
80
100
116
DAYS
P(DAPG-EOP)+PACLITAXEL (40 mg/kg)
PACLITAXEL (40 mg/kg)
P(DAPG-EOP)+PACLITAXEL (4 mg/kg)
PACLITAXEL (4 mg/kg)

FIG. 44