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(54) CPG SINGLE STRAND DEOXYNUCLEOTIDES FOR USE AS ADJUVANT

(76) Inventors: Liying Wang, Beijing (CN); Musheng Bao, Beijing (CN); Yongli Yu, Beijing

Correspondence Address: KNOBBE MARTENS OLSON & BEAR LLP 2040 MAIN STREET FOURTEENTH FLOOR **IRVINE, CA 92614 (US)**

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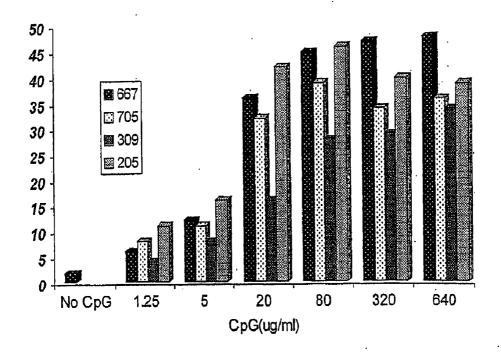
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ABSTRACT (57)

The present invention provides an adjuvant, which includes at least one single strand deoxynucleotide containing a CpG dinucleotide. The single strand deoxynucleotide comprises one or more CpG dinucleotides. When used in combination with rabies vaccine, HBV vaccine or other vaccines, the adjuvant can significantly improve the immune effect of the vaccine.



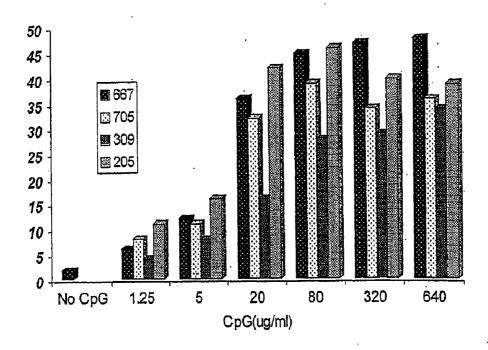


Fig.1

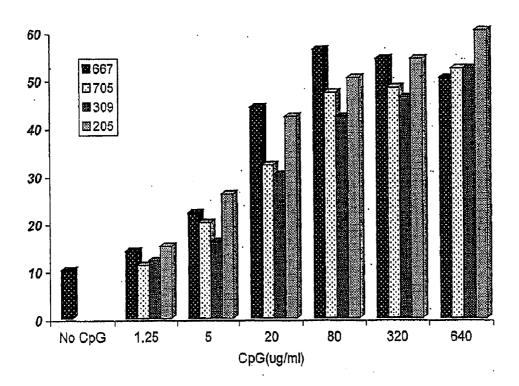
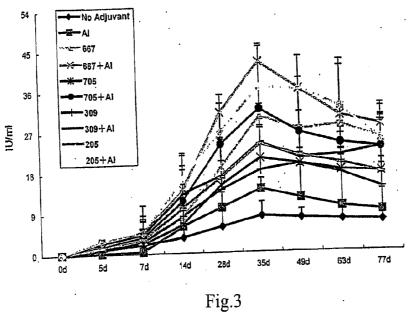


Fig.2



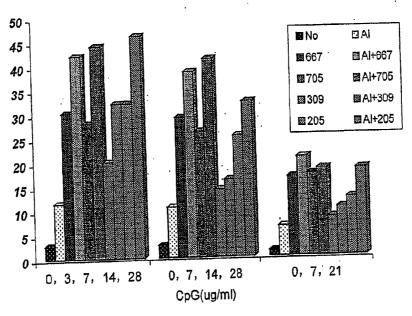


Fig.4

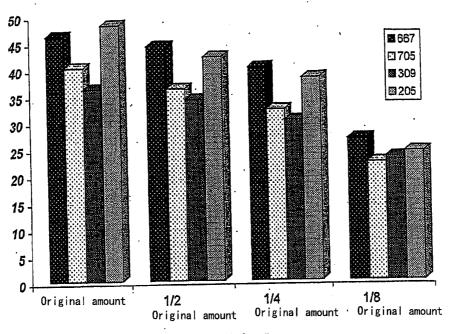


Fig.5

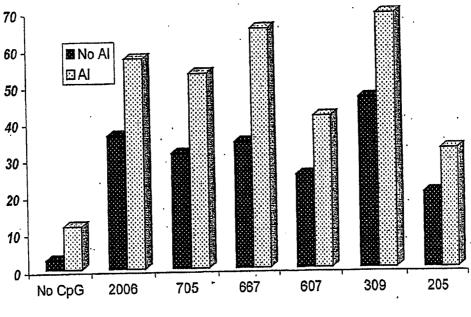
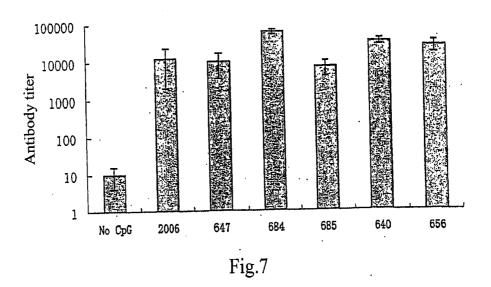
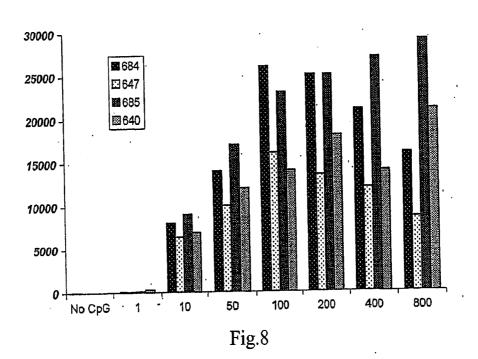


Fig.6





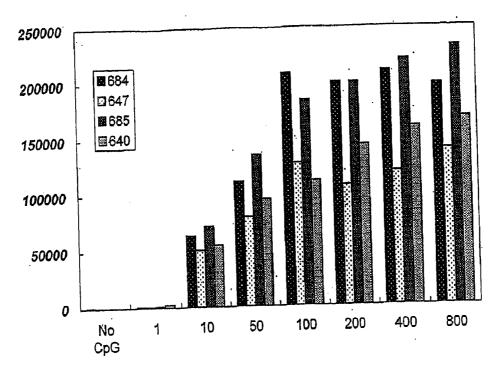
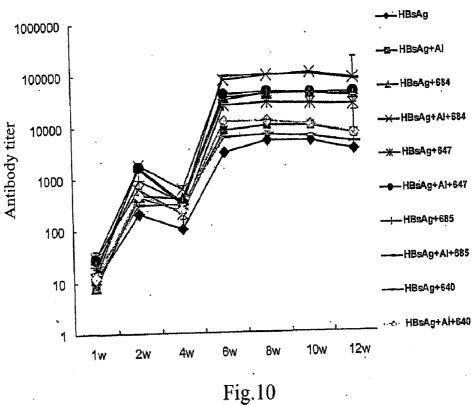


Fig.9



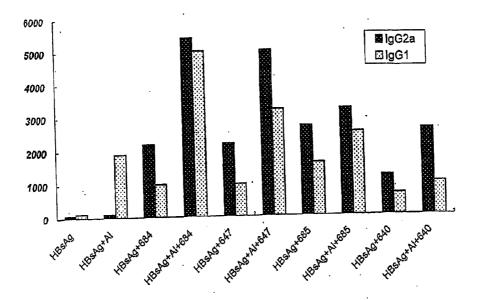


Fig.11

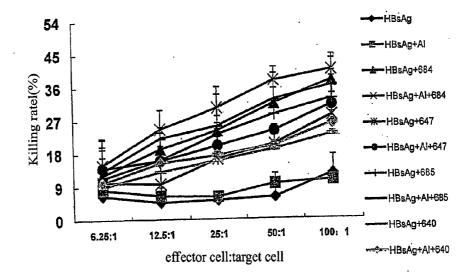
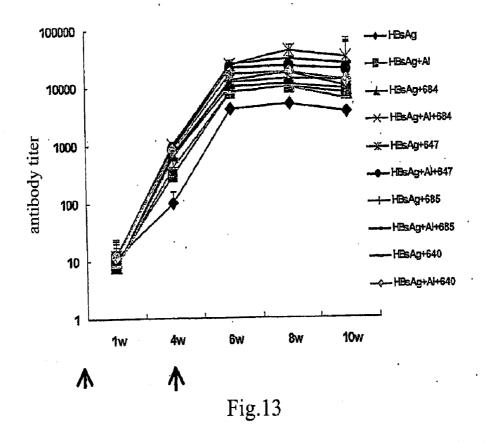


Fig.12



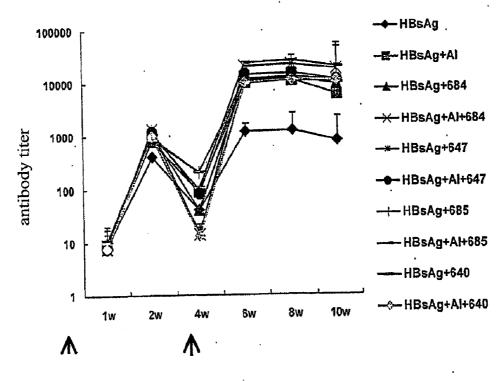


Fig.14

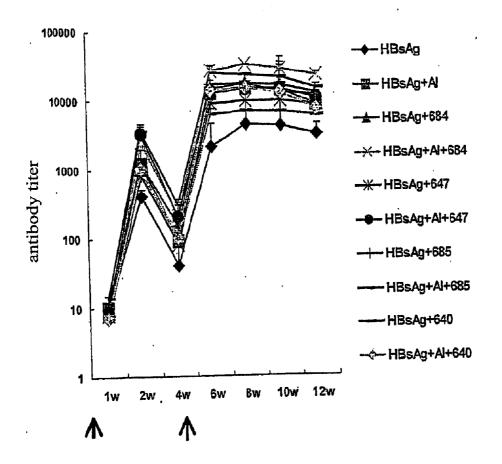


Fig.15

CPG SINGLE STRAND DEOXYNUCLEOTIDES FOR USE AS ADJUVANT

FIELD OF THE INVENTION

[0001] The present invention relates to a single strand deoxynucleotide containing CpG dinucleotides, and particularly relates to a single strand deoxynucleotide containing CpG dinucleotides as an adjuvant in hepatitis B virus (HBV) vaccine or rabies vaccine. The present invention further relates to sequences of the single strand deoxynucleotide containing CpG dinucleotides.

BACKGROUND OF THE INVENTION

[0002] CpG ODN is a type of oligodeoxynucleotides centered with non-methylated cytosine and guanine nucleotide (CpG). Typically, the CpG is flanked by base sequences in the following manner: 5'PurPurCGPyrPyr 3', i.e., 2 purines at its 5's end and 2 pyrimidines at its 3' end. (G Mutwiri, R. Pontarollo, S. BaBIUK. Biological activity of immunostimulatory CpG ODN motif in domestic animals. Veterinary Immunopathology, 2003, 91: 89-103). Studies indicate CpG ODN can activate a variety of immune effector cells, in which non-methylated CpG dinucleotides is believed to be of importance to the immunological activity of the CpG ODN. The DNAs of bacteria, viruses, and invertebrates possess immunological activation function because they have non-methylated CPG ODN sequences. The DNAs of vertebrates, in contrast, do not possess the function because their CpGs are methylated. Immune response against exogenous DNA is elicited by the immune system of the body through the recognition of the unmethylated CpG (YiAK, Klinman D M, Martin T L, et al. Rapid immune activation by CpG motifs in bacterial DNA. Systemic induction of IL-6 transcription through an antioxidantsensitive pathway. Immunol., 1996, 157(12):5394-402).

[0003] Rabies, also referred to as hydrophobia, takes place in over 60 countries all over the world. (Wolfgang Haupt. Rabies—risk of exposure and current trends in prevention of human cases. Vaccine 1999 (17): 1742-1749). The incidence of the disease in Southeast Asia countries is higher than in other regions of the world. In mainland China, the incidence is between 0.4/100,000-1.58/100,000. Recently, the incidence of rabies grew up rapidly in many countries including China (David W Dreesen. A global review of rabies vaccines for human use. Vaccine, 1997, 15, suppl s2-s6. D. Zienius, J. Bagdonas, A. Dranseika. Epidemiological situation of rabies in Lithuania from 1990 to 2000. *Veterinary Microbiology*, 2003(93):91-100.), with a death rate of nearly 100%.

[0004] Generally, human is infected by rabies virus through for instance, animal such as dog bite, where the animal carries rabies virus. In China, 80%-90% human rabies cases are caused by infected dogs. Human rabies is an acute infectious disease characterized by the invasion of central nervous system (Alan C. Jackson, William H. Wunner. Detection of Rabies Virus Genomic RNA and mRNA in Mouse and Human Brains by Using In Situ Hybridization. *Journal of Virology*, 1991, 65(6):2839-2844.), the clinical manifestations of which include hydrophobia, anxiety, fear of wind, pharynx spasm, progressing paralysis, etc.

[0005] Rabies virus belongs to Rhabdoviridae family, with a size of about 75×180 nm. It is a single minus strand RNA encapsulated by protein capsid, the surface of which is

covered by a lipoprotein envelope. The envelope further contains glycoprotein spikes. Rabies virus has immunogenicity, which can not only induce neutral antibodies but also cause RBC aggregation in animals such as chicken and goose. Rabies virus can be simply inactivated by UV radiation, quaternary ammonium compounds, iodine, potassium permanganate, alcohol, formaldehyde, etc. Heating at 100° C. for 2 minutes can also inactivate the virus. Rabies virus is tolerant to low temperatures. It can survive under –70° C. or –4° C. (in lyophilized form) for years.

[0006] Currently, there is no effective treatment for rabies infections. Up to now, rabies virus vaccine inoculation (hereafter referred to as rabies vaccine) and anti-rabies serum administration remain the major methods for preventing rabies infections. Presently in China, hamster renal cell vaccine is used as rabies vaccine. The subjects are administered 5 times intramuscularly by injection in a whole course, wherein each administration is taken on days 0, 3, 7, 14 and 30. For severe patients, they are administered 10 times in the whole course, i.e., one administration per day from the date of being bitten to 5 days after the bitten, and the rest of the administrations being taken on days 10, 14, 30 and 90. For people bitten by infected dogs, the average incidence rate is 15%-20%. In comparison, the incidence rate drops to 0.15% after taken a whole course administration

[0007] Hepatitis B is a liver disease caused by hepatitis B virus (HBV), which has severely affected the health of people around the globe. By 2004, the number of HBV patient has reached 400,000,000 worldwide (Lin K W, Kirchner J T. Hepatitis B. Am Fam Physician, 2004 Jan. 1, 69(1):75-82.), most of them are Asians. In China, HBV carriers account for about 10% of the entire population. Presently, the major method to prevent HBV propagation is by way of inoculating HbsAg genetic engineering vaccine, where Aluminum is used as an adjuvant. According to WTO reports, 1 billion dosages of HBV vaccine have been consumed since 1982, which take an important role in combating the disease. The mechanism of the vaccine is that it induces the body to produce/secrete protective antibody IgG1. Although the antibody can neutralize the viruses outside of the cells, it could not thoroughly eliminate latent HBVs inside the infected cells. Furthermore, 10% of the population are low responsive or even have no response to the vaccine. Therefore, what is needed now is to improve the immunocompetence of the present HBV vaccines or to develop new vaccines which can effectively eliminate the latent HBV inside the infected cells. A number of studies on the development of novel HBV genetic engineering vaccines have been carried out from different perspectives by researchers all over the world. Among them, an important topic relates to the finding of an effective adjuvant of HBV vaccine. In recent years, CpG ODN became one of the newly discovered immunological adjuvants proved to be effective.

[0008] The result of a variety of experimental studies indicates that CpG ODN can work with HBV vaccine synergistically to induce the production of specific antibodies and elicit CTL response in murine, human, and other primates (Weeranta R D, McCluskie M J, Xu Y. et al. CpG ODN is a novel non-toxic adjuvant which induces stronger immune responses than many conventional adjuvants. Vaccine, 2000, 18: 1755-62.). CpG ODN is considered as a safe and effective adjuvant for HBV vaccine.

[0009] In 1988, Davis et al. immunized BALB/c mice with HBsAg and CpG ODN1826 (as adjuvant). The results of HBsAb assay indicated that the HBsAbs produced by the co-administration of HBsAg and CpG ODN as adjuvant is 5 times higher than those produced by the co-administration of HBsAg and aluminum; the HBsAbs produced by the coadministration of HBsAg, and CpG ODN as well as aluminum as adjuvants is 35 times higher than those produced by the co-administration of HBsAg and aluminum; and in control group where only HBsAg was added, no HBsAb or very low lever HBsAb was observed. All these data shows as HBsAg adjuvants, CpG ODN functions better than aluminum in inducing HBsAb production, and CpG ODN further functions synergistically with aluminum. The results of ELISA and ⁵¹Cr killing test indicated the combination use of CpG ODN and HBsAg, or the combination use of CpG ODN, aluminum and HbsAg can elicit Th1 immune response in mice, resulting in the production of IgG2a HBsAb and accompanied by HBV-specific CTL response; while the combination use of aluminum and HBsAg mainly elicited Th2 immune response in mice, resulting in the production of IgG1 HBsAb and not accompanied by HBVspecific CTL response. The in vitro antibody staining result of cell surface molecule revealed that the mechanism CpG ODN used to enhance the immunological effectiveness of HBsAg is closely associated with mechanisms through which CpG induces APC to express co-stimulative molecules, and synergistically induces the class switch of the antibodies produced by B lymphocyte (Hartmann G. Weeratna R D, Ballas Z K, et al. Delineation of a CpG phosphorothioate oligodeoxynucleotide for activating primate immune responses in vitro and in vivo. Immunol, 2000, 164:1617-24). All the above-discussed results suggest CpG ODN is a promising adjuvant for HBV vaccine.

SUMMARY OF THE INVENTION

[0010] In one aspect, the present invention provides a single strand deoxynucleotide containing CpG, which can be used as an adjuvant for a vaccine.

[0011] The structure of the CpG ODN according to the present invention can be represented by formulas 1-5 as follows:

$$(G)_{n}(L)_{n}X_{1}X_{2}CGY_{1}Y_{2}(M)_{n}(G)_{n}$$
 1.

[0012] X₁=A, T, G; X₂=A, T; Y₁=A, T; Y₂=A, T, C; L, M=A, T, C, G; n is 0-6,

[0013] X_1 can be adenine, thymine, cytosine; X_2 can be adenine, thymine; Y_1 can be adenine, thymine; Y_2 can be adenine, thymine, guanine; L and M can be adenine, thymine, guanine and cytosine;

$$(G)_n(L)_n CG(XY)_n CG(M)_n(G)_n$$
 2.

[0014] X=A, T; Y=A, T; L, M=A, T, C, G; n is 0-6,

[0015] X can be adenine, thymine; Y can be adenine, thymine; L and M can be adenine, thymine, guanine and cytosine;

$$(TCG)_{n}(L)_{n}CG(M)_{n}(G)_{n}$$
3

[0016] L, M=A, T, C, G; n is 0-6,

[0017] L, M can be adenine, thymine, guanine and cytosine;

$$(TCG)_n(L)_nX_1X_2CG(M)_n$$
 4.

[0018] X_1 =A, T, G; X_2 =A, T; L, M=A, T, C, G; n is 0-6,

[0019] X_1 can be adenine, thymine, cytosine; X_2 can be adenine, thymine; L and M can be adenine, thymine, guanine and cytosine;

[0020] Based on the disclosure of the present invention, a skilled artisan will understand modifications can be made to the bases of the single strand deoxynucleotide, which include, but are not limited to sulpher modification, nonsulpher modification, partial sulpher modification, rare base modification (such as dI and dU), methylation modification, and other modifications where sulfhydryl, Aminolinker C6, Thiol-C6 S—S, etc. are used to couple to other substances. In addition, single strand deoxynucleotides comprising 2 or more CpG dinucleotides can function similarly to those comprising only one CpG to achieve the object of the present invention.

[0021] The single strand deoxynucleotides comprising CpG can be used in combination with other non-nucleic acid adjuvants to improve the immunological effect of a vaccine. The non-nucleic acid adjuvants include aluminum adjuvant, Freund's adjuvant, MPL, emulsions, etc.

[0022] In another aspect, the present invention provides a method for improving the immunogenicity of a vaccine, characterized by the combination use of the vaccine with the adjuvant according to the present invention, wherein the adjuvant comprises at least one single strand deoxynucleotide containing one or more CpG dinucleotides. The vaccine includes, but is not limited to rabies vaccine and HBV vaccine. The HBV vaccine includes but is not limited to hepatitis B virus blood-derived vaccine, hepatitis B virus genetic engineering protein vaccines, hepatitis B virus transgenic plant vaccine, HBV virus vector vaccine,

[0023] HBV bacterium vector vaccine and HBV DNA vaccine, of which HbsAg can be the major antigen component. Rabies vaccine includes, but is not limited to rabies virus blood-derived vaccine, rabies virus genetic engineering protein vaccines, rabies virus transgenic plant vaccine, rabies virus vector vaccine, rabies virus bacterium vector vaccine and rabies DNA vaccine. Alternatively, the adjuvant of the present invention can be used in combination with a non-nucleic acid adjuvant.

[0024] As examples, the combination use of a rabies vaccine with the adjuvant of the present invention not only significantly improved the immune response of the body to the vaccine, but also reduced the total administrations needed for the immunization; when CpG ODN is combined with HbsAg, it enhanced the immunogenicity of the HBV vaccine, quickly induced body reaction to the vaccine, elicited Th1 immune response, elongated the time limit of the immune response, reduced immune times, and improved the immunocompetence of unmatured or aged individuals. In summary, CpG ODN can be used as an effective adjuvant for HBV vaccine.

BRIEF DESCRIPTION OF THE DRAWINGS

[0025] FIG. 1 shows the effects of different dosages of a CpG ODN on antibody production stimulated by rabies vaccine.

[0026] FIG. 2 shows the effects of the combination use of aluminum adjuvant and different dosages of a CpG ODN on antibody production stimulated by rabies vaccine.

[0027] FIG. 3 shows the effect of CpG ODN on the speed of antibody production stimulated by rabies vaccine.

[0028] FIG. 4 shows the evaluation of whether CpG ODN, as an adjuvant, can reduce the immune times needed for the rabies vaccination.

[0029] FIG. 5 shows the evaluation of whether CpG ODN, as an adjuvant, can reduce the dosages needed for the rabies vaccination.

[0030] FIG. 6 shows the effects of different CpG ODNs on antibody production stimulated by rabies vaccine.

[0031] FIG. 7 shows the effects of different CpG ODNs on antibody production stimulated by HBsAg.

[0032] FIG. 8 shows the effects of different dosages of CpG ODN on antibody production stimulated by HBsAg.

[0033] FIG. 9 shows the effects of the combination use of aluminum adjuvant and different dosages of a CpG ODN on antibody production stimulated by HBsAg.

[0034] FIG. 10 shows the enhancement effect of the combination use of CpG ODN and an aluminum adjuvant on the immune effect of HBsAg.

[0035] FIG. 11 shows the comparison of the subtypes of the antibody produced by the stimulation of HbsAg in combination with different adjuvants.

[0036] FIG. 12 shows the effect of CpG ODN on HBV specific CTL induced by HBsAg.

[0037] FIG. 13 shows that CpG ODN enhanced the response of suckling mice to HBsAg.

[0038] FIG. 14 shows that CpG ODN enhanced the response of aged mice to HBsAg.

 $[0039]\ {\rm FIG.}\ 15$ shows that CpG ODN enhanced the response of rhesus to HBsAg.

PREFERRED EMBODIMENTS FOR CARRYING OUT THE INVENTION

[0040] The invention will be illustrated with reference to the following examples. However, a person of ordinary skill in the art will understood the examples are included only for illustrative purposes and are not intended to limit the scope of the invention. Therefore, it is intended that this invention be limited only by the scope of the appended claims. By referring to the following examples, these and other advantages will be apparent to a skill person in the relevant art.

EXAMPLE 1

Design of the Single Strand Deoxynucleotide Containing CpG

[0041] The sequences were designed as follows:

(1) $(G)n(L)n X_1X_2CGY_1Y_2(M)n (G)n$

(1) (G)M(B)M $x_1x_2 \in S_{1,2}(M)$ M (G)M $x_1 = A$, T, G; $x_2 = A$, T; $y_1 = A$, T; C, G; n is 0-6. 5'-ggggTCgTTCgTCgTTgggggg-3'	$Y_2 = A_{\prime}$ (SEQ				= A, T,
5'-ggggATAACgTTgCgggggg-3'	(SEQ	ID	NO:	2)	[143]
5'-ggggTgCAACgTTCAgggggg-3'	(SEQ	ID	NO:	3)	[402]
5'-ggggTCCTACgTAggAgggggg-3'	(SEQ	ID	NO:	4)	[123]
5'-ggggTCCATgACgTTCCTgAAgggggg-3'	(SEQ	ID	NO:	5)	[603]
5'-gggggACgTCgCCgggggggg-3'	(SEQ	ID	NO:	6)	[118]
5'-ggATCCgTACgCATgggggg-3'	(SEQ	ID	NO:	7)	[320]
5'-gggggAATCgATTCggggggg-3'	(SEQ	ID	NO:	8)	[154]
5'-gggATgCATCgATgCATCgggggg-3'	(SEQ	ID	NO:	9)	[464]
5'-ggTgCgACgTCgCAgggggg-3'	(SEQ	ID	NO:	10)	[471]
5'-gggACgTACgTCgggggg-3'	(SEQ	ID	NO:	11)	[390]
5'-gggggATCgACgTCgATCggggggg-3'	(SEQ	ID	NO:	12)	[322]
5'-ggCgATCgATCgATCgggggggg-3'	(SEQ	ID	NO:	13)	[333]
5'-ggggTCgATCgATCgAgggggg-3'	(SEQ	ID	NO:	14)	[113]
5'-ggTCgCgATCgCgAgggggg-3'	(SEQ	ID	NO:	15)	[307]
5'-ggGGTCAACGTTGAgggggG-3'	(SEQ	ID	NO:	16)	[156]
5'-gTCgTTTTCgTCgACgAATTgggggggg-3'	(SEQ	ID	NO:	17)	[222]
5'-gTCgTTATCgTTTTTCgTAgggggg-3'	(SEQ	ID	NO:	18)	[151]
5'-ggCgTTAACgACgggggg-3'	(SEQ	ID	NO:	19)	[288]
5'-gTCggCACgCgACgggggg-3'	(SEQ	ID	NO:	20)	[157]

5'-ggTgCgACgTCgCAgggggg-3'	(SEQ	ID	NO:	21)	[312]
5'-gTCTATTTTgTACgTACgTgggg-3'	(SEQ	ID	NO:	22)	[360]
5'-gACgTCgACgTCgACgTCAggggg-3'	(SEQ	ID	NO:	23)	[209]
5'-ggggTCgATCgTTgCTAgCgggggg-3'	(SEQ	ID	NO:	24)	[399]
5'-gggggACgTTATCgTATTgggggggg-3'	(SEQ	ID	NO:	25)	[600]
5'-ggggTCgTCgTTTgTCgTgTgTCgTTggggggg-3'	(SEQ	ID	NO:	26)	[408]
5'-ACgATCgATCgATCgggggg-3'	(SEQ	ID	NO:	27)	[304]
5'-AgACgTCTAACgTCggggg-3'	(SEQ	ID	NO:	28)	[301]
5'-ggggTgCTggCCgTCgTTgggggg-3'	(SEQ	ID	NO:	29)	[266]
5'-ggggTCgTTgCCgTCgggggg-3'	(SEQ	ID	NO:	30)	[248]
5'-ACCggTATCgATgCCggTgggggg-3'	(SEQ	ID	NO:	31)	[389]
5'-TTCgTTgCATCgATgCATCgTTgggggg-3'	(SEQ	ID	NO:	32)	[287]
<pre>(2) (G)n(L)nCG(XY)nCG(M)n(G)n X = A, T; Y = A, T; L, M = A, T, C, G; 5'-ggggACgATACgTCggggggg-3'</pre>	n is (SEQ			33)	[546]
5'-ggggACgATATCgATgggggg-3'	(SEQ	ID	NO:	34)	[1007]
5'-ggACgATCgATCgTgggggg-3'	(SEQ	ID	NO:	35)	[521]
5'-TCggggACgATCgTCgggggg-3'	(SEQ	ID	NO:	36)	[667]
5'-gggggATCgATATCgATCgggggg-3'	(SEQ	ID	NO:	37)	[576]
5'-ggATCgATCgATCgATgggggg-3'	(SEQ	ID	NO:	38)	[268]
5'-ggTgCATCgATCgATgCAgggggg-3'	(SEQ	ID	NO:	39)	[101]
5'-ggTgCATCgTACgATgCAgggggg-3'	(SEQ	ID	NO:	40)	[100]
5'-ggTgCgATCgATCgCAgggggg-3'	(SEQ	ID	NO:	41)	[134]
5'-gggggggTCgATCgATgggggg-3'	(SEQ	ID	NO:	42)	[519]
5'-ggggTCgTCgAACgTTgggggg-3'	(SEQ	ID	NO:	43)	[350]
5'-TgTCgTTCCTTgTCgTT-3'	(SEQ	ID	NO:	44)	[387]
5'-TTCgCTTCgCTTTCgCTTCgCTT-3'	(SEQ	ID	NO:	45)	[212]
5'-ACCgCCAAggAgAAgCCgCAggAggg-3'	(SEQ	ID	NO:	46)	[166]
5'-TACAACggCgAggAATACC-3'	(SEQ	ID	NO:	47)	[176]
5'-gTACAACggCgAggAATACCT-3'	(SEQ	ID	NO:	48)	[523]
5'-ACCgTCgTTgCCgTCggCCC-3'	(SEQ	ID	NO:	49)	[230]
5'-TgCTggCCgTCgTT-3'	(SEQ	ID	NO:	50)	[435]
5'-gTCggCACgCgACg-3'	(SEQ	ID	NO:	51)	[325]
5'-gTCggCACgCgACgCCCCC-3'	(SEQ	ID	NO:	52)	[523]
5'-TCCCgCTggACgTT-3'	(SEQ	ID	NO:	53)	[188]
5'-TTACCggTTAACgTTggCCggCC-3'	(SEQ	ID	NO:	54)	[403]
5'-ACCggTTAACgTTgTCCCCggggg-3'	(SEQ	ID	NO:	55)	[420]
5'-CgTTgACgATCgTCCCATggCggg-3'	(SEQ	ID	NO:	56)	[104]
5'-TCTgCggCCTTCgTCg-3'	(SEQ	ID	NO:	57)	[257]

-continue 5'-TAqTAACCqqTCCqqCqCCCCC-3'	d (SEQ ID NO: 58) [221]
5'-TTgCAgCgCTgCCggTggg-3'	(SEQ ID NO: 59) [611]
5'-CggCCCATCgAgggCgACggC-3'	(SEQ ID NO: 60) [378]
5'-TCATCgACTCTCgAgCgTTC-3'	(SEQ ID NO: 61) [599]
5'-ATCgTCgACTCTCgTgTTCTC-3'	(SEQ ID NO: 62) [201]
5'-TgCAgCTTgCTgCTTgCTgCTTC-3'	(SEQ ID NO: 63) [153]
5'-ggTgCgACgTCgCAgATgAT-3'	(SEQ ID NO: 64) [116]
5'-ggTCgAACgTTCgAgATgAT-3'	(SEQ ID NO: 65) [133]
5'-gggggCgTCgTTTTCgTCgACgAATT-3'	(SEQ ID NO: 66) [278]
5'-actcgagacgcccgttgatagctt-3'	(SEQ ID NO: 67) 355 [244]
5'-AACgTTggCgTCgACgTCAgCgCC-3'	(SEQ ID NO: 68) [623]
5'-gACgTCgACgTTgACgCT-3'	(SEQ ID NO: 69) [485]
5'-ggCgTTAACgTTAgCgCT-3'	(SEQ ID NO: 70) [579]
5'-AgCgCTAgCgCTgACgTT-3'	(SEQ ID NO: 71) [232]
5'-CTAgACgTTCAAgCgTT-3'	(SEQ ID NO: 72) [233]
5'-gACgATCgTCgACgATCgTC-3'	(SEQ ID NO: 73) [344]
5'-gTCgTTCgTAgTCgACTACgAgTT-3'	(SEQ ID NO: 74) [379]
5'-AAAAgACgTCgACgTCgACgTCTTTT-3'	(SEQ ID NO: 75) [489]
5'-TgCgACgATCgTCgCACgATCggAT-3'	(SEQ ID NO: 76) [479]
5'-TgCgACgTCgCACAgCgT-3'	(SEQ ID NO: 77) [492]
(3) (TCG)n(L)nCG (M)n(G)n L, M = A, T, C, G; n is 0-6 5'-TCgTTgCCgTCgg-3'	(SEQ ID NO: 78) [619]
5'-TCgTTgCCgTCggg-3'	(SEQ ID NO: 79) [577]
5'-TCgTTgCCgTCgggg-3'	(SEQ ID NO: 80) [533]
5'-TCgTTgCCgTCggggg-3'	(SEQ ID NO: 81) [537]
5'-TCgTTgCCgTCgggggg-3'	(SEQ ID NO: 82) [481]
5'-TCgTTgCCgTCggggggg-3'	(SEQ ID NO: 83) [177]
5'-TCgTTgCCgTCgggggggg-3'	(SEQ ID NO: 84) [111]
5'-TCgTTgCCgTCgggggggg-3'	(SEQ ID NO: 85) [105]
5'-TCgTCgggTgCATCgATgCAgggggg-3'	(SEQ ID NO: 86) [664]
5'-TCgTCgggTgCAACgTTgCAgggggg-3'	(SEQ ID NO: 87) [564]
5'-TCgTCgggTgCgTCgACgCAgggggg-3'	(SEQ ID NO: 88) [542]
5'-TCgTCgggTgCgATCgCAgggggg-3'	(SEQ ID NO: 89) [450]
5'-TCgTCgggTgCgACgATCgTCgCAgggggg-3'	(SEQ ID NO: 90) [465]
5'-TCgTCgTgCgACgTCgCAgggggg-3'	(SEQ ID NO: 91) [498]
5'-TCgTCgCAgAACgTTCTgggggg-3'	(SEQ ID NO: 92) [527]
5'-TCgTgCgACgTCgCAgggggg-3'	(SEQ ID NO: 93) [112]
5'-TCgTgCgACgATCgTCgCAgggggg-3'	(SEQ ID NO: 94) [178]
5'-TCgTATgCATCgATgCATAgggAgg-3'	(SEQ ID NO: 95) [410]

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5'-TCgTgCATCgATgCAgggggg-3'	(SEQ	ID	NO:	96)	[444]
5'-TCgAAACgTTTCgggggg-3'	(SEQ	ID	NO:	97)	[532]
5'-TCggACgATCgTCgggggg-3'	(SEQ	ID	NO:	98)	[598]
5'-TCgAgCgATCgCTCgAgggggg-3'	(SEQ	ID	NO:	99)	[555]
5'-TCgTCgCTTTgTCgTTgggg-3'	(SEQ	ID	NO:	100)	[418]
5'-TCgTCgTTTTgTCgTTgggg-3'	(SEQ	ID	NO:	101)	[208]
5'-TCgTCgggTgCgACgTCgCAgggggg-3'	(SEQ	ID	NO:	102)	[302]
5'-TCgTCgggTgCgACgATCgTCggggggg-3'	(SEQ	ID	NO:	103)	[290]
5'-TCgTCgTTTgCATCgATgCAggggggg-3'	(SEQ	ID	NO:	104)	[627]
5'-TCgTCgTTTTgACgATCgTCgggggg-3'	(SEQ	ID	NO:	105)	[500]
5'-TCgTTCggggTgCCg-3'	(SEQ	ID	NO:	106)	[103]
5'-TCgTTCggggTACCgATgggg-3'	(SEQ	ID	NO:	107)	[578]
5'-TCgTTgCgCTCCCATgCCgggggg-3'	(SEQ	ID	NO:	108)	[319]
5'-TCgTCgTTTCgTCgTTgggg-3'	(SEQ	ID	NO:	109)	[647]
5'-TCgTTgTCgTTTCgCTgCCggCggggg-3'	(SEQ	ID	NO:	110)	[417]
5'-TgCTTgggTggCAgCTgCCAgggggg-3'	(SEQ	ID	NO:	111)	[427]
5'-TgCTgCTTTgCTgCTTgggg-3'	(SEQ	ID	NO:	112)	[421]
5'-AACgTTCgACgTCgAACggggggg-3'	(SEQ	ID	NO:	113)	[453]
5'-AACgACgACgTTggggg-3'	(SEQ	ID	NO:	114)	[580]
(4) $(TCG)n(L)nX_1X_2CG$ (M)n $X_1 = A$, T, G; $X_2 = A$, T; L, M = A, T, The sequences were designed as follow	/s:				F 7
X_1 = A, T, G; X_2 = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3'	vs: (SEQ	ID	NO:	115)	[470]
X_1 = A, T, G; X_2 = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3'	rs: (SEQ (SEQ	ID	NO:	115) 116)	[502]
<pre>X₁ = A, T, G; X₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCgTCgTTTgCgTTTgCgTTGTCgTT-3'</pre>	vs: (SEQ (SEQ (SEQ	ID ID	NO: NO:	115) 116) 117)	[502] [601]
<pre>X₁ = A, T, G; X₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCGTAACGTTGTTTTTAACGTT-3' 5'-TCGTCGTATACGACGATCGTT-3' 5'-TCGTCGTTTGCGTTTGTCGTT-3'</pre>	VS: (SEQ (SEQ (SEQ (SEQ	ID ID ID	NO: NO: NO:	115) 116) 117) 118)	[502] [601] [625]
<pre>X₁ = A, T, G; X₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCgTCgTTTgCgTTTgCgTTGTCgTT-3'</pre>	VS: (SEQ (SEQ (SEQ (SEQ	ID ID ID	NO: NO: NO:	115) 116) 117) 118)	[502] [601]
<pre>X₁ = A, T, G; X₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCGTAACGTTGTTTTTAACGTT-3' 5'-TCGTCGTATACGACGATCGTT-3' 5'-TCGTCGTTTGCGTTTGTCGTT-3'</pre>	VS: (SEQ (SEQ (SEQ (SEQ	ID ID ID ID	NO: NO: NO: NO:	115) 116) 117) 118) 119)	[502] [601] [625]
<pre>X₁ = A, T, G; X₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCgTCgTTTgCgTTgTCgTT-3' 5'-TCCTgTCgTTTTgTCgTT-3' 5'-TCgTCgTTgTCgTTCgTT-3'</pre>	/s: (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID	NO: NO: NO: NO: NO:	115) 116) 117) 118) 119)	[502] [601] [625] [430]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCGTCgTTTTgCgTTgTCgTT-3' 5'-TCCTgTCgTTTTgTCgTT-3' 5'-TCgTCgTTgTCgTTCgCT-3' 5'-TCgTCgTTACCgATgACgTCgCCgT-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID ID	NO: NO: NO: NO: NO: NO:	115) 116) 117) 118) 119) 120)	[502] [601] [625] [430] [480]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCCTGTCgTTTTGCGTTGTCT-3' 5'-TCGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTGTCGTTCGCT-3' 5'-TCGTCGTTTACCGATGACGTCGCCGT-3' 5'-TCGTCGTTTTGCATCGATGCAGTCGTCGTT-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID ID	NO: NO: NO: NO: NO: NO: NO:	115) 116) 117) 118) 119) 120) 121)	[502] [601] [625] [430] [480] [108]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACGACGATCGTT-3' 5'-TCGTCGTTTTGCGTTGTCGTT-3' 5'-TCGTCGTTGTCGTTTGCGTT-3' 5'-TCGTCGTTGTCGTTCGCT-3' 5'-TCGTCGTTTGCATCGATGACGTCGCCGT-3' 5'-TCGTCGTTTGCATCGATGAGTCGTCGTT-3' 5'-TCGCCTCGTCGCCTTCGAGCG-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID ID	NO: NO: NO: NO: NO: NO: NO: NO: NO:	115) 116) 117) 118) 119) 120) 121) 122)	[502] [601] [625] [430] [480] [108]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTTTTGCGTTGTCGTT-3' 5'-TCCTGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTGTCGTTCGCT-3' 5'-TCGTCGTTTGCATCGATGACGTCGCCGT-3' 5'-TCGCCTCGTCGCCTTCGAGCG-3' 5'-TCGTTGTTGCGTTGCCTTCGAGCG-3' 5'-TCGTTGTTGCGTTGCCTTTGGATGCGT-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID ID ID	NO: NO: NO: NO: NO: NO: NO: NO: NO:	115) 116) 117) 118) 119) 120) 121) 122) 123)	[502] [601] [625] [430] [480] [108] [102]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCGTCgTTTTGCGTTGTCGTT-3' 5'-TCGTCgTTTTGTCGTT-3' 5'-TCGTCgTTTTGCGTTCGCT-3' 5'-TCGTCGTTTGCATCGATGCAGTCGTCGTT-3' 5'-TCGTCGTTTGCATCGATGCAGTCGTCGTT-3' 5'-TCGTCGTTGCGCTTCGAGCG-3' 5'-TCGTCGTGTGCGCTTCGAGCG-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID ID ID ID	NO: NO: NO: NO: NO: NO: NO: NO: NO:	115) 116) 117) 118) 119) 120) 121) 122) 123) 124)	[502] [601] [625] [430] [480] [108] [102] [406] [560]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACGACGATCGTT-3' 5'-TCGTCGTTTTGCGTTTGTCGTT-3' 5'-TCGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTTCGTTCGCT-3' 5'-TCGTCGTTTGCATCGATGACGTCGCCGT-3' 5'-TCGTCGTCTGCCCTTCGAGCG-3' 5'-TCGTCGTCGCCTTCGAGCG-3' 5'-TCGTCGTGGCGTGCCGTTGGT-3' 5'-TCGTCGTCGCGTGGCCGTTGGT-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID	NO:	115) 116) 117) 118) 119) 120) 121) 122) 123) 124) 125)	[502] [601] [625] [430] [480] [108] [102] [406] [560]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCCTTGTCgTTTTGTCGTT-3' 5'-TCGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTTGCGTTGCGT-3' 5'-TCGTCGTTTGCATCGATGCAGTCGTCGTT-3' 5'-TCGTCGTTGCGTTGCGTTGAGCG-3' 5'-TCGTCGTCGTCGCTTCGAGCG-3' 5'-TCGTCGTGGCGTGCGTTGGGT-3' 5'-TCGTCGTGGCGTGGCGTTGGGT-3' 5'-TCGTCGAGGGCGCGTTGGGT-3' 5'-TCGTCGCCGGTGGCGTGGTGGT-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID I	NO:	115) 116) 117) 118) 119) 120) 121) 122) 123) 124) 125) 126)	[502] [601] [625] [430] [480] [108] [102] [406] [560] [629]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCGTCgTTTTGCGTTTGTCGTT-3' 5'-TCGTCGTTTTGCGTTGCGT-3' 5'-TCGTCGTTTGCATCGATGACGTCGCGT-3' 5'-TCGTCGTTTGCATCGATGCAGTCGTCGTT-3' 5'-TCGTCGTTGCGTTGCGTT-3' 5'-TCGTCGTTGCGCTTCGAGCG-3' 5'-TCGTCGTCGCCGTGGCGTTGGGT-3' 5'-TCGTCGAGGGGCGGTGAC-3' 5'-TCGTCGCCGGTGGGGTGGTGGT-3' 5'-TCGTCGCCGGTGGGGTGGGTGG-3' 5'-TCGTCGCCGGTGGGGTGGGTGG-3'	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID	NO:	115) 116) 117) 118) 119) 120) 121) 122) 123) 124) 125) 126) 127) 128)	[502] [601] [625] [430] [480] [108] [102] [406] [560] [629] [440]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTTAACgTTGTTTTTAACGTT-3' 5'-TCGTCGTATACGACGATCGTT-3' 5'-TCGTCGTTTTGCGTTGTCGTT-3' 5'-TCGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTGCGTTGCGT-3' 5'-TCGTCGTTTGCATCGATGCAGTCGTCGTT-3' 5'-TCGTCGTTGCGCTTCGAGCG-3' 5'-TCGTCGTCGCCGTTGGAGCG-3' 5'-TCGTCGTGGCGTTGGAGCG-3' 5'-TCGTCGAGGGGCGGTGAC-3' 5'-TCGTCGCCGGTGGGGGTGAC-3' 5'-TCGTCGCCGGTGGGGGTGGTGGTG-3' 5'-TCGTCGCCGGTGGGGGGGGGGGGGGGGGGGGGGGGGGG	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID ID ID ID ID ID ID	NO:	115) 116) 117) 118) 119) 120) 121) 122) 123) 124) 125) 126) 127) 128)	[502] [601] [625] [430] [480] [108] [102] [406] [560] [629] [440] [207] [615]
X ₁ = A, T, G; X ₂ = A, T; L, M = A, T, The sequences were designed as follow 5'-TCgTAACgTTgTTTTTAACgTT-3' 5'-TCgTCgTATACgACgATCgTT-3' 5'-TCGTCgTTTTGCGTTGTT-3' 5'-TCCTGTCGTTTTGTCGTT-3' 5'-TCGTCGTTTTGCGTTGCGT-3' 5'-TCGTCGTTTGCATCGATGCAGTCGTCGTT-3' 5'-TCGTCGTTGCGCTTCGAGCG-3' 5'-TCGTCGTCGCCTTCGAGCG-3' 5'-TCGTCGTGGCGTGGCGTTGGT-3' 5'-TCGTCGAGGGCGGTGGCGTGGTGGT-3' 5'-TCGTCGCCGGTGGGCGTTGGGT-3' 5'-TCGTCGCCGGTGGGGGTGGTGGTG-3' 5'-TCGTCGCCGGTGGGGGGGGGGGGGGGGGGGGGGGGGGG	(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ	ID I	NO:	115) 116) 117) 118) 119) 120) 121) 122) 123) 124) 125) 126) 127) 128) 130)	[502] [601] [625] [430] [480] [108] [102] [406] [560] [629] [440] [207] [615] [610]

5'-TCgTgCAggCCAACgAggCCg-3'	(SEQ	ID	NO:	133) [631]
5'-TCgTTgCCgTCggCCC-3'	(SEQ	ID	NO:	134) [115]
5'-TCggCACgCgACgTgCTggCCgTCgTTTCC-3'	(SEQ	ID	NO:	135) [370]
5'-TcgTTgccgTcggccccccc-3'	(SEQ	ID	NO:	136) [309]
5'-TcgTTgccgTcggccccc-3'	(SEQ	ID	NO:	137) [506]
5'-TCgTTgCCgTCggCCCCC-3'	(SEQ	ID	NO:	138) [404]
5'-TCgTTgCCgTCggCCCC-3'	(SEQ	ID	NO:	139) [203]
5'-TcgTTgccgTcggcccccc-3'	(SEQ	ID	NO:	140) [501]
5'-TCgAggACAAgATTCTCgT-3'	(SEQ	ID	NO:	141) [305]
5'-TCggCACgCgACgTgCTggCCgTCgTT-3'	(SEQ	ID	NO:	142) [509]
5'-TCgTCgCgCCgTCACgggggg-3'	(SEQ	ID	NO:	143) [630]
5'-TCgTgTgCgTgCCgTTggg-3'	(SEQ	ID	NO:	144) [106]
5'-TCgTCgCCgTTgggCggg-3'	(SEQ	ID	NO:	145) [117]
5'-TCgTCgACgTCgTTgggCggg-3'	(SEQ	ID	NO:	146) [280]
5'-TCgCAgTTgTCgTAACgTTgggCggg-3'	(SEQ	ID	NO:	147) [205]
5'-TCgTCgTTggTATgTT-3'	(SEQ	ID	NO:	148) [613]
5'-TCgTCgTCgTCgTTgTCgTT-3'	(SEQ	ID	NO:	149) [306]
5'-TCgTCgTCgTCgTTgTCgTTgggg-3'	(SEQ	ID	NO:	150) [640]
5'-TCgTTCggggTgCCg-3'	(SEQ	ID	NO:	151) [409]
5'-TCgTTCggggTAACgATT-3'	(SEQ	ID	NO:	152) [508]
5'-TCgTTCggggTAACgTT-3'	(SEQ	ID	NO:	153) [540]
5'-TCgTTCggggTACCgAT-3'	(SEQ	ID	NO:	154) [401]
5'-TCgTACggCCgCCgTACggCggg-3'	(SEQ	ID	NO:	155) [607]
5'-TCgCgTCgACTCCCCTCgAgggg-3'	(SEQ	ID	NO:	156) [380]
5'-TCgTCgTCgACTCgTggTCggggg-3'	(SEQ	ID	NO:	157) [656]
5'-TCgggCgCCCgATCgggggg-3'	(SEQ	ID	NO:	158) [310]
5'-TCgTCggTCTTTCgAAATT-3'	(SEQ	ID	NO:	159) [109]
5'-TCgTgACgTCCTCgAgTT-3'	(SEQ	ID	NO:	160)[330]
5'-TCgTCTTTCgACTCgTTCTC-3'	(SEQ	ID	NO:	161) [605]
5'-TCgTCgTTTTgCgTTCTC-3'	(SEQ	ID	NO:	162) [504]
5'-TCgACTTTCgTCgTTCTgTT-3'	(SEQ	ID	NO:	163) [407]
5'-TCgTCgTTTCgTCgTTCTC-3'	(SEQ	ID	NO:	164) [550]
5'-TCgTCgTCgTCgTTgTCgTT-3'	(SEQ	ID	NO:	165) [612]
5'-TCgTTCTCgACTCgTTCTC-3'	(SEQ	ID	NO:	166) [277]
5'-TCgACgTTCgTCgTTCgTCgTTC-3'	(SEQ	ID	NO:	167) [684]
5'-TCgTCgACgTCgTTCgTTCTC-3'	(SEQ	ID	NO:	168) [685]
5'-TCgTgCgACgTCgCAgATgAT-3'	(SEQ	ID	NO:	169) [114]
5'-TCgTCgAgCgCTCgATCggAT-3'	(SEQ	ID	NO:	170) [211]

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5'-TCgTCgTTTCgTAgTCgTTgACg	r Cggg−3'	(SEQ	ID	NO:	171)	[204]
5'-TCgTCggACgTTTTCCgACgTTC	г-3'	(SEQ	ID	NO:	172)	[308]
5'-TCgTCgTTTTCgTCgTTTTTCgTC	gTT-3'	(SEQ	ID	NO:	173)	[340]
5'-TCgTCgTTTgTCgTgTgTCgTT-	3 '	(SEQ	ID	NO:	174)	[503]
5'-TCgTCgTTggTCggggTCgTTgg	ggTCgTT-3'	(SEQ	ID	NO:	175)	[405]
5'-TCgTCgTTTCgTCTCTCgTT-3'		(SEQ	ID	NO:	176)	[614]
5'-TCgTCgTTTTgCTgCgTCgTT-3	1	(SEQ	ID	NO:	177)	[505]
5'-TCgAgCgTTTTCgCTCgAATT-3	'	(SEQ	ID	NO:	178)	[530]
(5) Sequences containing T 5'-TTCgTCgTTTgATCgATgTTCgT		(SEQ	ID	NO:	179)	[507]
5'-TTCgTCgTTgTgATCgATggggg	g-3'	(SEQ	ID	NO:	180)	[210]
5'-TATCgATgTTTTCgTCgTCgTTg	ggggg-3'	(SEQ	ID	NO:	181)	[202]
5'-TCgACTTTCgTCgTTCTgTT-3'		(SEQ	ID	NO:	182)	[303]
5'-TCgTCgTTTCgTCgTTCTC-3'		(SEQ	ID	NO:	183)	[491]
5'-TCgACgTTCgTCgTTCgTCgTTC	-3'	(SEQ	ID	NO:	184)	[590]
5'-TCgTCgTTTTCgTCgTTTTCgTC	gTT-3'	(SEQ	ID	NO:	185)	[633]

[0042] Preferably, CpG ODNs according to the present invention have the following sequences:

CpG[647]/[205]: 5'-TCgTCgTTTCgTCgTTgggg-3'	(SEQ	ID	NO:	109)
CpG[640]/[309]: 5'-TCgTCgTCgTCgTTgTCgTTgggg-3'	(SEQ	ID	NO:	150)
CpG[684]/[667]: 5'-TCgACgTTCgTCgTTCgTCgTTC-3'	(SEQ	ID	NO:	167)
CpG[685]/[705]: 5'-TCgTCgACgTCgTTCgTTCTC-3'	(SEQ	ID	NO:	168)

[0043] In the following examples, CpG ODN sequences will be represented by the numbers in above brackets.

EXAMPLE 2

Synthesis of the Single Strand Deoxynucleotide Containing CpG

[0044] DNA fragment was synthesized by solid phase-phosphoramidite triestermethod. The method has been widely used in chemical synthesis of DNA due to its advantages of fast, high efficiency, etc.

[0045] Chemical synthesis of DNA is different from enzymatic synthesis of DNA. The latter is synthesized from 5' end to 3' end, while the former starts from 3' end. The synthesis steps are as follows:

[0046] 1. De-Protection

[0047] Trichloroacetic acid was used to remove protective group DMT conjugated to the nucleotides on Controlled Pore Glass (CPG) to obtain free 5' hydroxyl ends for being used in the following condensation reaction.

[0048] 2. Activation

[0049] The nucleotides units protected by phosphoramidite was mixed with tetrazolium activator and then loaded into a synthesis column to form an active intermediate of phosphoramiditetetrazole (the 3' end of which is activated, while the 5' end of which is still under DMT protection). The intermediate then reacted with the de-protected nucleotide on CPG in a condensation reaction.

[0050] 3. Linking

[0051] When contacted with the de-protected nucleotide on CPC; the intermediate reacted with its 5' hydroxyl group, and released tetrazole group in the condensation reaction. In such a way, one base was added to the synthesized oligonucleotide strand.

[0052] 4. Blocking

[0053] After the condensation reaction, acetylation is commonly used to block the hydroxyl end to prevent unreacted 5'hydroxyl group conjugated on CPG from further being extended in the following reactions. Generally, reactants used in acetylation are formulated by mixing acetic anhydride, N-methylimidazole, and the like.

[0054] 5. Oxidation

[0055] In the condensation reaction, a nucleotide bonded with the oligonucleotide on CPG through phosphate bond. Since phosphate bond is unstable, easy to be hydrolyzed by acids or bases, the tetrahydrofuran solution of iodine is conventionally used to transform phosphoramidite into phosphoric triester to produce stable oligonucleotides.

[0056] Through the five steps described above, a deoxynucleotide was bonded to the nucleotides on CPG. Likewise, after repeating all these five steps, a crude DNA fragment was obtained. The fragment was then subjected to postsynthesis processing such as cleavage, de-protection (gen-

erally, benzoyl protection is used for bases A and C; isobutyryl protection is used for base G; no protection is needed for base T; nitrile ethyl protection is used for phosphorous acid), purification (conventional methods such as HAP, PAGE, HPLC, C18, and OPC etc.) and quantification to produce oligonucleotide fragments qualified for experimental use.

[0057] Solid-phase oligonucleotide synthesis was carried out on a DNA synthesizer. After the protective groups were removed, the purity of the interest oligonucleotides produced by the method described above was very low, containing a large amount of impurities. Major impurities include benzoic acid ammonia and isobutyric acid ammonia formed by the removed protective groups and ammonia, nitrile ethyl from nitrile phosphate, and short chains produced during the synthesis. As a result, the content of the oligonucleotide in the crude product is only about 15%. Though the efficiency in every synthesis step is around 97%-98%, the add-up efficiency of the whole process is not high. As such, the content of the oligonucleotide of interest can not even reach 10%. Impurities such as salts and short chains in the crude product not only make the quantification inaccurate, but also influence the next reactions. Therefore, the oligonucleotides must be further purified. Polyacrylamide gel electrophoresis (PAGE) method is preferably used for the purification. The method can be conveniently used in most molecular biology laboratories and the products purified by the method can achieve relatively high purity. When cost becomes an issue, desalination can be used for some experiments with relatively lower requirements such as PCR.

[0058] Oligonucleotide fragment is quantified by OD_{260} value. In 1 ml standard quartz cuvette with a light path of 1 cm, a oligonucleotide solution with an absorbance of 1 under 260 nm wavelength is defined as 1 OD_{260} . Although the bases makeup in each specific oligonucleotide are not exactly the same, the weight of a oligonucleotide of 1 OD_{260} is about 33 μg .

EXAMPLE 3

Effects of Different Dosages of a CpG ODN on Antibody Production Stimulated by Rabies Vaccine

[0059] 1. Experimental animals: 200 white mice, with half males and half females, weighted from 18 to 22 g, aged from 6 to 8 weeks, and purchased from Beijing Weitonglihua Experimental Animal Ltd.

[0060] 2. Rabies vaccine: 1 ml/vial (containing 2.5 IU), purchased from Changehun Institute of Biological Product.

[0061] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0062] 4. Experimental groups: 8 mice for each group, with half males and half females.

[0063] rabies vaccine

[**0064**] rabies vaccine+1.25 μg CpG667

[0065] rabies vaccine+1.25 μg CpG705

[0066] rabies vaccine+1.25 µg CpG309

[0067] rabies vaccine+1.25 μg CpG205

[0068] rabies vaccine+5 μg CpG667

[0069] rabies vaccine+5 μg CpG705

[0070] rabies vaccine+5 μg CpG309

[0071] rabies vaccine+5 µg CpG205

[0072] rabies vaccine+20 µg CpG667

[0073] rabies vaccine+20 µg CpG705

[0074] rabies vaccine+20 µg CpG309

[0075] rabies vaccine+20 µg CpG205

[0076] rabies vaccine+80 µg CpG667

[0077] rabies vaccine+80 μg CpG705

[0078] rabies vaccine+80 µg CpG309

[0079] rabies vaccine+80 µg CpG205

[0080] rabies vaccine+320 μg CpG667

[0081] rabies vaccine+320 μg CpG705

[0082] rabies vaccine+320 μg CpG309

[0083] rabies vaccine+320 μg CpG205

[0084] rabies vaccine+640 μg CpG667 [0085] rabies vaccine+640 μg CpG705

[0086] rabies vaccine+640 µg CpG309

[0087] rabies vaccine+640 µg CpG205

[0088] 5. CpGODN formulation: 50 μ l PBS was used to dissolve different dosages of CpGODN to prepare CpGODN solutions with different concentrations.

[0089] 6. White mice inoculation: On days 0, 3, 7, 14 and 28, rabies vaccine or rabies vaccine+CpG ODN as listed above in experimental groups were respectively administrated to white mice via intraperitoneal injection, wherein the dosage of rabies vaccine is 0.5 ml/mouse.

[0090] 7. Experimental groups: 8 mice for each group, with half males and half females.

[0091] A volume of 0.5 ml was used in the inoculation.

[0092] 8. Rabies vaccine antibody assay: On day 35, blood was drawn from the caudal vein of the white mice. The blood was assayed with Quick Rabies vaccine Fluorescence Foci Inhibitory test (RFFIT) (Yan jiaxin, Li chengping, Zhu jiahong, et al. The establishment of a quick rabies vaccine fluorescence foci inhibitory experimental method to detect the neutral antibody of rabies virus. *Journal of China Biological Product*, 1998, 11 (2): 93-96; Ministry of Health of PRC. Chinese Biological Product Regulations (Department 1), Beijing: Chinese Population Publishing House, 1996, 201.) to detect rabies virus-specific antibodies in serum. The blood was drawn from caudal vein of the mice two days prior to the inoculation, and the serum obtained was used as negative control.

[0093] 9. Results: With the increase of the dosage of CpG ODN, the level of rabies vaccine-specific antibodies in mice sera also increased (See FIG. 1).

[0094] 10. Conclusion: CpG ODN can significantly improve the production of rabies vaccine-specific antibodies

in white mice elicited by rabies vaccine, which indicates CpG ODN can be used as an effective adjuvant for a rabies vaccine

EXAMPLE 4

Effects of the Combination Use of Aluminum Adjuvant and Different Dosages of a CpG ODN on Antibody Production Stimulated by Rabies Vaccine

[0095] 1. Experimental animals: 200 white mice, with half males and half females, weighted from 18 to 22 g, aged from 6 to 8 weeks, and purchased from Beijing Weitonglihua Experimental Animal Ltd.

[0096] 2. Rabies vaccine: 1 ml/vial (containing 2.5 IU), purchased from Changchun Institute of Biological Product.

[0097] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0098] 4. Experimental groups: 8 mice for each group, with half males and half females.

[0099] rabies vaccine

[0100] rabies vaccine+A1 adjuvant (purchased from Changchun Institute of Biological Product)

[0101] rabies vaccine+1.25 μg CpG667+A1 adjuvant

[0102] rabies vaccine+1.25 µg CpG705+A1 adjuvant

[0103] rabies vaccine+1.25 µg CpG309+A1 adjuvant

[0104] rabies vaccine+1.25 µg CpG205+A1 adjuvant

[0105] rabies vaccine+5 μg CpG667+A1 adjuvant

[0106] rabies vaccine+5 µg CpG705+A1 adjuvant

[0107] rabies vaccine+5 µg CpG309+A1 adjuvant

[0108] rabies vaccine+5 µg CpG205+A1 adjuvant

[0109] rabies vaccine+20 µg CpG667+A1 adjuvant

[0110] rabies vaccine+20 µg CpG705+A1 adjuvant

[0111] rabies vaccine+20 μg CpG309+A1 adjuvant

[0112] rabies vaccine+20 μg CpG205+A1 adjuvant

[0113] rabies vaccine+80 μg CpG667+A1 adjuvant

[0114] rabies vaccine+80 μg CpG705+A1 adjuvant

[0115] rabies vaccine+80 µg CpG309+A1 adjuvant

[0116] rabies vaccine+80 µg CpG205+A1 adjuvant

[0117] rabies vaccine+320 μg CpG667+A1 adjuvant

[0118] rabies vaccine+320 μg CpG705+A1 adjuvant

[0119] rabies vaccine+320 μg CpG309+A1 adjuvant

[0120] rabies vaccine+320 µg CpG205+A1 adjuvant

[0121] rabies vaccine+640 g CpG667+A1 adjuvant

[0122] rabies vaccine+640 μg CpG705+A1 adjuvant

[0123] rabies vaccine+640 µg CpG309+A1 adjuvant

[0124] rabies vaccine+640 μg CpG205+A1 adjuvant

[0125] 5. CpGODN formulation: 50 μ l PBS was used to dissolve different dosages of a CpGODN to prepare CpGODN solutions with different concentrations.

[0126] 6. White mice inoculation: On days 0, 3, 7, 14 and 28, rabies vaccine+A1 adjuvant or rabies vaccine+A1 adjuvant+CpG ODN as listed above in experimental groups were respectively administrated to white mice via intraperitoneal injection. The dosage of rabies vaccine used is 0.5 ml, and the final concentration of the A1 adjuvant is 0.5 mg/ml.

[0127] 7. Rabies vaccine antibody assay: On day 35, blood was drawn from the caudal vein of the white mice. The serum was assayed with Quick Rabies vaccine Fluorescence Foci Inhibitory test (RFFIT) to detect the titers of rabies virus-specific antibodies in serum. The blood was drawn from caudal vein of the mice two days prior to the inoculation, and the serum obtained was used as negative control.

[0128] 8. Results: With the increase of the dosage of CpG ODN, the level of rabies vaccine-specific antibodies in mice sera also increased. FIG. 2 shows the effects of the combination use of aluminum adjuvant and different dosages of CPG ODN on antibody production stimulated by rabies vaccine.

[0129] 9. Conclusion: CpG ODN and A1 adjuvant can significantly improve the production of rabies vaccine-specific antibodies in white mice elicited by rabies vaccine, which indicates the combination of CpG ODN and A1 can be used effectively as a potent adjuvant for a rabies vaccine.

EXAMPLE 5

Effect of CPG ODN on the Speed of Antibody Production Stimulated by Rabies Vaccine

[0130] 1. Experimental animals: 80 white mice, with half males and half females, weighted from 18 to 22 g, aged from 6 to 8 weeks, and purchased from Beijing Weitonglihua Experimental Animal Ltd.

[0131] 2. Rabies vaccine: 1 ml/vial (containing 2.5 IU), purchased from Changehun Institute of Biological Product.

[0132] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0133] 4. Experimental groups: 8 mice for each group, with half males and half females.

[0134] rabies vaccine

[0135] rabies vaccine+A1 adjuvant

[0136] rabies vaccine+80 µg CpG667

[0137] rabies vaccine+80 µg CpG667+A1 adjuvant

[0138] rabies vaccine+80 µg CpG705

[0139] rabies vaccine+80 µg CpG705+A1 adjuvant

[0140] rabies vaccine+80 fig CpG309

[0141] rabies vaccine+80 µg CpG309+A1 adjuvant

[0142] rabies vaccine+80 µg CpG205

[0143] rabies vaccine+80 μg CpG205+A1 adjuvant

[0144] All the above rabies vaccines and CpGODNs were dissolved in PBS.

[0145] 5. White mice inoculation: On days 0, 3, 7, 14 and 28, rabies vaccine or rabies vaccine+A1 adjuvant or rabies vaccine+A1 adjuvant+CpG ODN as listed above in experimental groups were respectively administrated to white mice

via intraperitoneal injection. The dosage of rabies vaccine used is 0.5 ml/mouse, and the final concentration of the A1 adjuvant is 0.5 mg/ml.

[0146] 6.Rabies vaccine antibody assay: On days 0, 5, 7, 14, 28, 35, 49, 63, and 77, blood was drawn from the caudal vein of the white mice, and the serum was separated from the blood. The serum was assayed with Quick Rabies vaccine Fluorescence Foci Inhibitory test (RFFIT) to evaluate the titers of the rabies vaccine antibodies produced. The blood was drawn from caudal vein of the mice two days prior to the inoculation, and the serum obtained was used as negative control.

[0147] 7. Results: When CpGODN and A1 adjuvant were used in combination, the titer of the antibodies produced at any specific time was higher than those of the antibodies produced where rabies vaccine, A1 adjuvant, or CpG ODN was used alone (See FIG. 3).

[0148] 8. Conclusion: The combination use of CpG ODN and A1 adjuvant can expediate the appearance of the rabies virus-specific antibody in mice immunized with rabies vaccine.

EXAMPLE 6

Evaluation of Whether CpG ODN, as an Adjuvant for a Rabies Vaccine, can Reduce the Immune Times Needed for a Rabies Vaccination

[0149] 1. Experimental animals: 80 white mice, with half males and half females, weighted from 18 to 22 g, aged from 6 to 8 weeks, and purchased from Beijing Weitonglihua Experimental Animal Ltd.

[0150] 2. Rabies vaccine: 1 ml/vial (containing 2.5 IU), purchased from Changchun Institute of Biological Product.

[0151] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0152] 4. Experimental groups: a total of 10 groups, with 8 mice for each group, half males and half females.

[0153] rabies vaccine

[0154] rabies vaccine+A1 adjuvant

[0155] rabies vaccine+80 μg CpG667

[0156] rabies vaccine+80 µg CpG667+A1 adjuvant

[0157] rabies vaccine+80 μg CpG705

[0158] rabies vaccine+80 μg CpG705+A1 adjuvant

[0159] rabies vaccine+80 µg CpG309

[0160] rabies vaccine+80 μg CpG309+A1 adjuvant

[0161] rabies vaccine+80 µg CpG205

[0162] rabies vaccine+80 μg CpG205+A1 adjuvant

[0163] All the above rabies vaccines and CpGODNs were dissolved in PBS.

[0164] 5. Immunization: The mice were immunized according to different groups. 0.5 ml of rabies vaccine was used to inoculate the mice via intraperitoneal injection. The immune times were set to 5, 4, and 3. When the immune times was set to 5, the mice was inoculated on days 0, 3, 7, 14, and 28, respectively. When the immune times was set to

4, the mice was inoculated on days 0, 7, 14, and 28, respectively. When the immune times was set to 3, the mice was inoculated on days 0, 7, and 21, respectively. Rabies vaccine or rabies vaccine+A1 adjuvant or rabies vaccine+A1 adjuvant+CpG ODN as listed above in different group were respectively administrated to white mice via intraperitoneal injection. The final concentration of the A1 adjuvant is 0.5 mg/ml.

[0165] 6. Rabies vaccine antibody assay: Blood was drawn from the caudal vein of the white mice, and the serum was separated from the blood 7 days after the final inoculation. The serum was assayed with Quick Rabies vaccine Fluorescence Foci Inhibitory test (RFFIT) to test the titers of the rabies vaccine antibodies produced. The blood was drawn from caudal vein of the mice two days prior to the inoculation, and the serum obtained was used as negative control.

[0166] 7. Results: The combination use of CpGODN and A1 adjuvant can produce relatively high levels of rabies virus-specific antibodies in mice immunized with rabies vaccine by 3, 4 or 5 times (See FIG. 4).

[0167] 8. Conclusion: The combination use of CpGODN and A1 adjuvant can produce a relatively high level of rabies virus-specific antibodies in mice immunized with rabies vaccine by 3 times.

EXAMPLE 7

Evaluation of Whether CpG ODN, as an Adjuvant for a Rabies Vaccine, can Reduce the Dosages Needed for the Rabies Vaccination

[0168] 1. Experimental animals 128 white mice, with half males and half females, weighted from 18 to 22 g, aged from 6 to 8 weeks, and purchased from Beijing Weitonglihua Experimental Animal Ltd.

[0169] 2. Rabies vaccine: 1 ml/vial (containing 2.5 IU), purchased from Changchun Institute of Biological Product.

[0170] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0171] 4. Experimental groups: 8 mice for each group, with half males and half females.

[0172] rabies vaccine+80 µg CpG705+A1 adjuvant

[0173] $^{1}\!\!/_{2}$ rabies vaccine+80 µg CpG705+A1 adjuvant

[0174] ¹/₄ rabies vaccine+80 μg CpG705+A1 adjuvant

[0175] ¹/₈ rabies vaccine+80 μg CpG705+A1 adjuvant

[0176] rabies vaccine+80 g CpG667+A1 adjuvant

[0177] ½ rabies vaccine+80 g CpG667+A1 adjuvant

[0178] ¹/₄ rabies vaccine+80 μg CpG667+A1 adjuvant

[0179] ¹/₈ rabies vaccine+80 μg CpG667+A1 adjuvant

[0180] rabies vaccine+80 µg CpG309+A1 adjuvant

[0181] ½ rabies vaccine+80 μg CpG309+A1 adjuvant

[0182] ¹/₄ rabies vaccine+80 µg CpG309+A1 adjuvant

[0102] 74 lables vaccine+80 µg CpO309+A1 adjuvani

[0183] 1/8 rabies vaccine+80 μg CpG309+A1 adjuvant

[0184] rabies vaccine+80 µg CpG205+A1 adjuvant

[0185] ½ rabies vaccine+80 μg CpG205+A1 adjuvant

[0186] ¹/₄ rabies vaccine+80 µg CpG205+A1 adjuvant

[0187] ½ rabies vaccine+80 μg CpG205+A1 adjuvant

[0188] All the above rabies vaccines and CpGODNs were dissolved in PBS.

[0189] 5. White mice inoculation: On days 0, 3, 7, 14 and 21, white mice were immunized according to different groups. 0.5 ml/mouse of rabies vaccine was inoculated to the mice via intraperitoneal injection. The final concentration of the A1 adjuvant is 0.5 mg/ml.

[0190] 6. Rabies vaccine antibody assay: On day 28, blood was drawn from the caudal vein of the white mice, and the serum was separated from the blood. The serum was assayed with Quick Rabies vaccine Fluorescence Foci Inhibitory test (RFFIT) to evaluate the titers of the rabies vaccine antibodies produced. The blood was drawn from caudal vein of the mice two days prior to the inoculation, and the serum obtained was used as negative control.

[0191] 7. Results: The combination use of CpG and reduced amount of rabies vaccine can still elicit a relatively high level of rabies virus-specific antibody production, indicating CpG ODN can lower the dosage of the rabies vaccine (See FIG. 5).

[0192] 8. Conclusion: CpG ODN can lower the dosage of the rabies vaccine.

EXAMPLE 8

Effects of Different CpG ODNs on Antibody Production Stimulated by Rabies Vaccine

[0193] 1. Experimental animals: 112 white mice, with half males and half females, weighted from 18 to 22 g, aged from 6 to 8 weeks, and purchased from Beijing Weitonglihua Experimental Animal Ltd.

[0194] 2. Rabies vaccine: 1 ml/vial (containing 2.5 IU), purchased from Changchun Institute of Biological Product.

[0195] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0196] 4. Control sequence:

[0197] 5'-TCgTCgTTTTgTCgTTTTgTcgTT-3'[2006]

[0198] 5. Experimental groups: 8 mice for each group, with half males and half females.

[0199] rabies vaccine

[0200] rabies vaccine+A1 adjuvant

[0201] rabies vaccine+80 μg CpG667

[0202] rabies vaccine+80 μg CpG667+A1 adjuvant

[0203] rabies vaccine+80 µg CpG705

[0204] rabies vaccine+80 μg CpG705+A1 adjuvant

[0205] rabies vaccine+80 μg CpG309

[0206] rabies vaccine+80 μg CpG309+A1 adjuvant

[0207] rabies vaccine+80 μg CpG205

[0208] rabies vaccine+80 µg CpG205+A1 adjuvant

[0209] rabies vaccine+80 μg CpG607

[0210] rabies vaccine+80 μg CpG607+A1 adjuvant

[**0211**] rabies vaccine+80 μg CpG2006

[0212] rabies vaccine+80 μg CpG2006+A1 adjuvant

[0213] All the above rabies vaccine and CpG ODN were dissolved in PBS.

[0214] 6. White mice inoculation: On days 0, 3, 7, 14 and 21, white mice were immunized according to different groups. 0.5 ml/mouse of rabies vaccine was inoculated to the mice via intraperitoneal injection. The final concentration of the A1 adjuvant is 0.5 mg/ml.

[0215] 7. Rabies vaccine antibody assay: On day 35, blood was drawn from the caudal vein of the white mice, and the serum was separated from the blood. The serum was assayed with Quick Rabies vaccine Fluorescence Foci Inhibitory test (RFFIT) to detect the titers of rabies virus-specific antibodies in serum. The blood was drawn from caudal vein of the mice two days prior to the inoculation, and the serum obtained was used as negative control.

[0216] 8. Results: The effect of antibody production elicited by rabies vaccine+CpG667 or rabies vaccine+CpG309 is significantly superior to that elicited by rabies vaccine+CpG2006 (See FIG. 6).

[0217] 9. Conclusion: CpGODN667 and CpGODN309 can significantly improve the immune effect of rabies vaccine

EXAMPLE 9

HBV Antibody Assay by ELISA

I. Regents

[0218] 1. HBsAg (containing no A1 adjuvant, Beijing Institute of Biological Product) vaccine formulation: 1 mg lyophilized HBsAg protein powder was dissolved into 1 ml PBS to prepare stock solution (1 mg/ml).

[0219] 2. HRP-Horse-anti-mouse secondary antibody: Beijing Dingguo Biotechnology Ltd.

[0220] 3. PBS: 1000 ml

NaCl KCl Na ₂ HPO ₄ •12H ₂ O	8 g (Beijing Chemical Plant) 0.2 g (Beijing Chemical Plant) 2.9 g (Beijing Chemical Plant)
KH ₂ PO4	0.2 g (Beijing Chemical Plant)

[0221] After dissolved in 800 ml ultra pure water, the pH of the resultant solution was adjusted to 7.2-7.4 by HCl or NaOH, and then made up to a volume of 1000 ml.

[0222] 4. Coating solution 1100 ml

PBS	80 ml
50% glutaraldehyde	1.6 ml (Beijing Chemical Plant)

[0223] After sufficiently dissolved, the resultant solution was made up to a volume of 100 ml with PBS.

[0224] 5. Washing solution: 500 ml

PBS	400 ml
Tween20	0.5 ml (Beijing Chemical Plant)
NaCl	14.625 g (Beijing Chemical Plant)

[0225] After sufficiently dissolved, the resultant solution was made up to a volume of 500 ml with PBS.

[0226] 6. Blocking solution 1100 ml

	ml g (Beijing Dingguo Biotechnology Ltd.) g (Beijing Dingguo Biotechnology Ltd.)
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[0227] After sufficiently dissolved, the solution was made up to a volume of 100 ml with PBS, followed by adding 0.05 g of sodium azide.

[0228] 7. Sample diluent: 1000 ml

Tris NaCl	2.42 g (Beijing Chemical Plant) 8.77 g (Beijing Chemical Plant)	

[0229] After dissolved in 800 ml ultra pure water, the pH of the resultant solution was adjusted to 7.1 with HCl, followed by the addition of

BSA	1 g
Tween20	0.5 ml

[0230] ultra pure water was used to make up the solution to a volume of 1000 ml.

[0231] 8. Substrate solution

[0232] Solution A:

citric acid	19.2 g (Beijing Chemical Plant)

[0233] After dissolved in 800 ml ultra pure water, the resultant solution was made up to a volume of 1000 ml with the ultra pure water.

[0234] Solution B:

Na ₂ HPO ₄ •12H ₂ O	71.7 g (Beijing Chemical Plant)

[0235] After dissolved in 800 ml ultra pure water, the resultant solution was made up to a volume of 1000 ml with the ultra pure water.

[0236] Substrate Solution:

Solution A Solution B	47.276 ml 50 ml	
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[0237] Above volumes of Solution A and Solution B were taken respectively and mixed, followed by filtration through a $0.22 \mu m$ filter to remove bacteria.

[0238] 9. Stop solution: 100 ml

Concentrated H ₂ SO ₄	20 ml (Beijing Chemical Plant)

[0239] slowly added into 80 ml ultra pure water while stirring.

II. Methods

[0240] 1. Coating: 100 μ l HBsAg (1 mg/ml) was added into 10 ml coating solution, i.e., the HBsAg was diluted to a final concentration of 10 μ g/ml by the coating solution. Then, 100 μ l diluted HBsAg was added into each well of an ELISA plate, which was then left stand overnight under a temperature of 40 \square .

[0241] 2. Washing: The next day, the liquid left in the wells of the plate was removed completely, and 300 μ l washing solution was added into each well. Before removing the washing solution, the plate was left stand for 3 minute at room temperature. Finally, the plate was dried by tapping on an absorbent paper. Same washing procedure was repeated for 3 times.

[0242] 3. Blocking 300 μ l blocking solution was added into the wells of the ELISA plate, and was left stand for 2 hs at room temperature.

[0243] 4. Adding samples to be tested: Same washing procedures described in step 2 were applied. Samples to be tested were diluted with sample diluent into different concentrations before being added. 100 μ l/welldiluted sample was added into wells in duplicate and was left stand for 2 hs at room temperature.

[0244] 5. Adding HRP-horse-anti-mouse secondary antibody: Same washing procedures described in step 2 were applied. HRP-horse-anti-mouse secondary antibody was diluted with sample diluent (1:1000). 100 µl/well diluted HRP-horse-anti-mouse secondary antibody was added into wells and was left stand for 2 hs in darkness at room temperature.

[0245] 6. Adding substrate solution: Same washing procedures described in step 2 were applied. $100~\mu l$ freshly made substrate solution was added into each well, and left stand for 20 minutes in darkness at room temperature.

[0246] 7. Adding stop solution: The ELISA plate was incubated for 20 minutes, after which 50 μl stop solution was added into each well.

[0247] 8. ELISA assay (A492 nm): ELISA assay (A492 nm) was carried out within 5 minutes after the addition of the stop solution.

[0248] 9. Determination of a positive value: A positive value is defined where the OD value of a sample/the OD value of the negative control is 2 or more.

EXAMPLE 10

Comparison of the Effects of Different CpG ODNs on Antibody Production Stimulated by HBsAg

I. Animals and Regents

[0249] 1. Animals: BALB/c mouse, female, aged from 6-8 weeks (available from Beijing Weitonglihua Experimental Animal Ltd.).

[0250] 2. HBsAg: containing no A1 adjuvant, purchased from Changchun Institute of Biological Product.

[0251] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0252] 4. CpGODN formulation 100 μ g CpG ODN was dissolved into 50 μ l PBS to prepare application solution.

[0253] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 µl CpG application solution and 1 µl HBsAg application solution were first mixed thoroughly and then placed on ice for 10 minutes before the injection.

[0254] 6. Control CpGODN sequence:

5'-TCgTCgTTTTgTCgTTTTgTcgTT-3' [2006]

II. Methods

[0255] 1. Mice grouping 10 mice/group (the dosage of HBV vaccine is 1 μ g/mouse, and the content of A1 adjuvant is 25 μ g/ml)

[**0256**] 1) HBsAg group

[**0257**] 2) HBsAg+2006(100 µg) group

[**0258**] 3) HBsAg+647(100 μg) group

[**0259**] 4) HBsAg+684(100 µg) group

[**0260**] 5) HBsAg+685(100 µg) group

[**0261**] 6) HBsAg+640(100 μg) group

[**0262**] 7) HBsAg+656(100 µg) group

[0263] 2. The effects of different CPG ODNs on antibody production stimulated by HBsAg: The mice were inoculated through tibialis anterior with HBsAg application solution and application solutions of CpG ODNs with different sequences. Blood was drawn from the caudal vein of the mice three days prior to the inoculation (negative serum) and four weeks after the inoculation, immediately followed by adding 2 µl of sodium heparin to each 10 µl of the blood for the purpose of anticoagulation (Shanghai Zhixin Chemical Ltd. 0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 4° C. The plasma in the upper layer was collected and then stored at -20° C. ELISA was used to assay HbsAb titer (As shown in Example 3). The result of the assay indicates when used as adjuvants, all the CpGODNs with different sequences can improve the HbsAb titers produced by mice, though to different extents. Compared with using HBsAg vaccine alone, HBsAg+CpG can significantly elicit mice to produce a relatively high level of HBsAb. The result of the variance analysis showed a significant difference (P<0.05). The comparison of antibody titers among different groups is shown in FIG. 7.

EXAMPLE 11

The Effects of Different Dosages of CpG ODN on Antibody Production Stimulated by HBsAg

I. Animals and Regents

[0264] 1. Animals: BALB/c mouse, female, aged from 6-8 weeks (available from Beijing Weitonglihua Experimental Animal Ltd.).

[0265] 2. HBsAg: containing no A1 adjuvant, purchased from Beijing Institute of Biological Product.

[0266] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0267] 4. CpGODN formulation: different dosages of CpGODN were dissolved into 50 μ l PBS to prepare CpG ODN solution with corresponding concentrations.

[0268] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 μ l CpG ODN solution and 1 μ l HBsAg application solution were first mixed thoroughly and then placed on ice for 10 minutes before the injection (the dosage of HBV vaccine is 1 μ g/mouse, and the content of A1 adjuvant is 25 μ g/ml).

II. Methods

[0269] 1. Mice grouping 10 mice/group (the dosage of HBV vaccine is 1 μ g/mouse, and the content of A1 adjuvant is 25 μ g/ml)

[0270] HBsAg group

[0271] HBsAg+CpG(684)(1 µg) group

[**0272**] HBsAg+CpG(647)(1 μg) group

[0273] HBsAg+CpG(685)(1 µg) group

[0274] $HBsAg+CpG(640)(1 \mu g)$ group

[0275] HBsAg+CpG(684)(10 μg) group

[0276] HBsAg+CpG(647)(10 µg) group

[0277] HBsAg+CpG(685)(10 μg) group

[**0278**] HBsAg+CpG(640)(10 μg) group [**0279**] HBsAg+CpG(684)(50 μg) group

[**0280**] HBsAg+CpG(647)(50 μg) group

[**0281**] HBsAg+CpG(685)(50 μg) group

[**0282**] HBsAg+CpG(640)(50 μg) group

[0283] HBsAg+CpG(684)(100 µg) group

[0284] HBsAg+CpG(647)(100 μ g) group

[**0285**] HBsAg+CpG(685)(100 µg) group

[0286] HBsAg+CpG(640)(100 µg) group

[0287] HBsAg+CpG(684)(200 μg) group

[**0288**] HBsAg+CpG(647)(200 µg) group

[0289] HBsAg+CpG(685)(200 μ g) group

[0290] HBsAg+CpG(640)(200 µg) group

[**0291**] HBsAg+CpG(684)(400 μg) group

[**0292**] HBsAg+CpG(647)(400 µg) group

[0293] HBsAg+CpG(685)(400 µg) group

[0294] HBsAg+CpG(640)(400 μ g) group

[0295] HBsAg+CpG(684)(800 μg) group

 $\textbf{[0296]} \quad HBsAg+CpG(647)(800~\mu g)~group$

[0297] HBsAg+CpG(685)(800 µg) group

[**0298**] HBsAg+CpG(640)(800 μg) group

[0299] 2. The effects of different dosages of CPG ODN on antibody production stimulated by HBsAg: The mice were inoculated through tibialis anterior with HBsAg and different dosages of CpG ODN. Blood was drawn from the caudal vein of the mice three days prior to the inoculation (negative serum) and four weeks after the inoculation, immediately followed by adding 2 µl of sodium heparin to each 10 pt of the blood for the purpose of anticoagulation (0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 40. The plasma in the upper layer was collected and then stored at -20° C. ELISA was used to assay HbsAb titer (As shown in Example 3). The result of the assay indicates, though to different extents, the CpGODN in all the dosages can improve the HbsAb titers produced by mice. When compared with using HBsAg alone, the CpGODN can significantly improved the HBsAb titer produce by mouse, so long as the dosage of CpG ODN used is equal to or more than 10 µg (P<0.05). The comparison of antibody titers among different groups is shown in FIG. 8.

EXAMPLE 12

The Effects of the Combination Use of Aluminum Adjuvant and Different Dosages of CpG ODN on Antibody Production Stimulated by HBsAg

I. Animals and Regents

[0300] 1. Animals: BALB/c mouse, female, aged from 6-8 weeks (available from Beijing Weitonglihua Experimental Animal Ltd.).

[0301] 2. HBsAg: containing A1 adjuvant (25 mg A1³⁺/ mg HBsAg), purchased from Beijing Institute of Biological Product.

[0302] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0303] 4. CpGODN formulation: different dosages of CpG ODN were dissolved into 50 μ l PBS to prepare CpG ODN solution with corresponding concentrations.

[0304] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 µl CpG

ODN solution and 1 μ l HBsAg (25 mg Al³⁺/mg HBsAg) were first mixed thoroughly and then placed on ice for 10 minutes before the injection (the dosage of HBV vaccine is 1 μ g/mouse, and the content of Al adjuvant is 25 μ g/ml).

II. Method:

[0305] 1. Mice grouping 10 mice/group

[0306] 1) HBsAg (25 mg A1³⁺/mg HBsAg) group

[**0307**] 2) HBsAg (25 mg A1³⁺/mg HBsAg)+CpG(684)(1 μg) group

[0308] 3) HBsAg (25 mg $A1^{3+}$ /mg HBsAg)+CpG(647)(11 g) group

[0309] 4) HBsAg (25 mg $A1^{3+}/mg$ HBsAg)+CpG(685)(1 μ g) group

[0310]~ 5) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+CpG(640)(1 $\mu g)$ group

[**0311**] 6) HBsAg (25 mg A1³⁺/mg HBsAg)+CpG(684)(10 μg) group

[0312] 7) HBsAg (25 mg A1³⁺/mg HBsAg)+CpG(647)(10 μ g) group

[**0313**] 8) HBsAg (25 mg A1³⁺/mg HBsAg)+СрG(685)(10 µg) group

[**0314**] 9) HBsAg (25 mg A1³⁺/mg HBsAg)+CpG(640)(10 µg) group

[0315] 10) HBsAg (25 mg A1³+/mg HBsAg)+ CpG(684)(50 μ g) group

[0316] 11) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(647)(50 µg) group

[0317] 12) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(685)(50 µg) group

[0318] 13) HBsAg (25 mg A1³⁺/mg HBsAg)+ CpG(640)(50 µg) group

[0319] 14) HBsAg (25 mg A1³⁺/mg HBsAg)+ CpG(684)(100 µg) group [0320] 15) HBsAg (25 mg A1³⁺/mg HBsAg)+

CpG(647)(100 μg) group
[0321] 16) HBsAg (25 mg A1³⁺/mg HBsAg)+

CpG(685)(100 μ g) group [0322] 17) HBsAg (25 mg A1³⁺/mg HBsAg)+

CpG(640)(100 μg) group
[0323] 18) HBsAg (25 mg A1³⁺/mg HBsAg)+

[0323] 18) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(684)(200 µg) group

[0324] 19) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(647)(200 µg) group

[0325] 20) HBsAg (25 mg $A1^{3+}/mg$ HBsAg)+ CpG(685)(200 μ g) group

[0326] 21) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(640)(200 µg) group

[0327] 22) HBsAg (25 mg $A1^{3+}/mg$ HBsAg)+ CpG(684)(400 μ g) group

[0328] 23) HBsAg (25 mg $A1^{3+}/mg$ HBsAg)+ CpG(647)(400 μ g) group

[0329] 24) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(685)(400 µg) group

[0330] 25) HBsAg (25 mg $A1^{3+}/mg$ HBsAg)+ CpG(640)(400 μ g) group

[0331] 26) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(684)(800 µg) group

[0332] 27) HBsAg (25 mg $A1^{3+}/mg$ HBsAg)+ CpG(647)(800 μ g) group

[0333] 28) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(685)(800 µg) group

[0334] 29) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(640)(800 µg) group

[0335] 2. The combination use of aluminum adjuvant and different dosages of CPG ODN on antibody production stimulated by HBsAg: The mice were inoculated through tibialis anterior with HBsAg and different dosages of CpG ODN. Blood was drawn from the caudal vein of the mice three days prior to the inoculation (negative serum) and four weeks after the inoculation, immediately followed by adding 2 μl of sodium heparin to each 10 μl of the blood for the purpose of anticoagulation (0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 4° C. The plasma in the upper layer was collected and then stored at -20° C. ELISA was used to assay HbsAb titer (As shown in Example 3). The result of the assay indicates, though to different extents, the CpGODN in all the dosages can improve the HbsAb titers produced by mice. Compared with the combination of HBsAg and A1 adjuvant (25 mg A1³⁺/ mg HBsAg), all the combination use of CpGODN and HBsAg (containing no A1 adjuvant) can significantly improved the HBsAb titer produce by mouse (P<0.05), so long as the dosage of the CpGODN used is equal to or more than 10 µg. The comparison of antibody titers among different groups is shown in FIG. 9.

EXAMPLE 13

The Enhancement Effect of the Combination Use of CpG ODN and A1 Adjuvant on the Immune Effect of HBsAg

I. Animals and Regents

[0336] 1. Animals: BALB/c mouse, female, aged from 6-8 weeks (available from Beijing Weitonglihua Experimental Animal Ltd.).

[0337] 2. HBsAg (containing A1 adjuvant), HBsAg (containing no A1 adjuvant), purchased from Beijing Institute of Biological Product.

[0338] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0339] 4. CpGODN formulation 100 µg CpG ODN was dissolved into 50 µl PBS to prepare application solution.

[0340] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 μ l CpG ODN application solution and 1 μ l HBsAg (containing or

not containing A1 adjuvant) were first mixed thoroughly and then placed on ice for 10 minutes before the injection (the dosage of HBV vaccine is 1 μ g/mouse, and the content of A1 adjuvant is 25 μ g/ml).

II. Methods

[0341] 1. Mice grouping: 10 mice/group

[0342] HBsAg group

[0343] HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg) group

[0344] 3) HBsAg+CpG (684) (100 µg) group

[0345] 4) HBsAg+CpG (647) (100 µg) group

[0346] 5) HBsAg+CpG (685) (100 µg) group

[0347] 6) HBsAg+CpG (640) (100 µg) group

[0348] 7) HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg)+CpG (684) (100 μg) group

[**0349**] 8) HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg)+CpG (647) (100 μg) group

[0350] 9) HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg)+CpG (685) (100 µg) group

[0351] 10) HBsAg containing A1 adjuvant (25 mg A1³⁺/ mg HBsAg)+CpG (640) (100 µg) group

[0352] 2. The enhancement effect of the combination use of CpG ODN and A1 adjuvant on the immune effect of HBsAg: The experimental mice were divided into four groups, i.e., HBsAg group, HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg) group, HBsAg+CpGODN group, and HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg)+CpGODN group. The mice were inoculated according to different groups through tibialis anterior. Blood was drawn from the caudal vein of the mice three days prior to the inoculation (negative serum), immediately followed by adding 2 µl of sodium heparin to each 10 µl of the blood for the purpose of anticoagulation (0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 4° C. The plasma in the upper layer was collected and then stored at -20° C. The mice were boosted 4 weeks after the initial inoculation (week 0). Blood was drawn from the caudal vein of the mice 1 week, 2 weeks, 4 weeks, 6 weeks, 8 weeks, 10 weeks and 12 weeks after the inoculation. Plasma was separated from the blood and ELISA was used to assay HbsAb titer (As shown in Example 3). The result of the assay indicates, when compared with HBsAg group or HBsAg containing A1 adjuvant (25 mg A1³⁺/mg HBsAg) group, the use of CpGODN alone or in combination with A1 adjuvant both can improve the HBsAb titers. All the results of the variance analyses showed significant differences (P<0.05). The HBsAb titer is the highest when CpGODN is used in combination with A1 adjuvant. The comparison of antibody titers among different groups is shown in FIG. 10.

EXAMPLE 14

The Comparison of the Subtypes of the Antibody Produced by the Stimulation of HbsAg and Different Adjuvants

I. Animals and Regents

[0353] 1. Animals: BALB/c mouse, female, aged from 6-8 weeks (available from Beijing Weitonglihua Experimental Animal Ltd.).

[0354] 2. HBsAg (containing A1 adjuvant), HBsAg (containing no A1 adjuvant), purchased from Beijing Institute of Biological Product.

[0355] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0356] 4. CpGODN formulation: 100 µg CpG ODN was dissolved into 50 µl PBS to prepare application solution.

[0357] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 μ l CpG ODN application solution and 1 μ l HBsAg (containing or not containing A1 adjuvant) application solution were first mixed thoroughly and then placed on ice for 10 minutes before the injection.

[0358] HBsAg (containing no A1 adjuvant, available from Beijing Institute of Biological Product) formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution.

[0359] 6. HRP labeled goat-anti-mouse IgG2a and IgG1: product of Serotec Co.

[0360] 7. Regents formulation:

[0361] PBS: 1000 ml

NaCl	8 g (Beijing Chemical Plant)
KCl	0.2 g (Beijing Chemical Plant)
Na_2HPO_4 •12 H_2O	2.9 g (Beijing Chemical Plant)
KH ₂ PO4	0.2 g (Beijing Chemical Plant)

[0362] After sufficiently dissolved in 800 ml ultra pure water, the resultant solution was adjusted to a pH of 7.2-7.4 with HCl or NaOH, and made up to a volume of 1000 ml.

[0363] Coating solution: 100 ml

PBS	80 ml
50% glutaraldehyde	1.6 ml (Beijing Chemical Plant)

[0364] After sufficiently dissolved, the resultant solution was made up by PBS to a volume of 100 ml.

[0365] Washing solution: 500 ml

PBS	400 ml
Tween20	0.5 ml (Beijing Chemical Plant)
NaCl	14.625 g (Beijing Chemical Plant)

[0366] After sufficiently dissolved, the resultant solution was made up by PBS to a volume of 500 ml.

[0367] Block solution 1100 ml

PBS	80 ml
Skimmed milk	5 g (Beijing Dingguo Biotechnology Ltd.)
BSA	1 g (Beijing Dingguo Biotechnology Ltd.)

[0368] After sufficiently dissolved, the resultant solution was made up by PBS to a volume of 100 ml, followed by the addition of 0.05 g sodium azide.

[0369] Sample diluent 1000 ml

NaCl 8.77 g (Beijing Chemical Plant)	Tris NaCl	2.42 g (Beijing Chemical Plant) 8.77 g (Beijing Chemical Plant)
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[0370] After sufficiently dissolved in 800 ml ultra pure water, the resultant solution was adjusted to a pH of 7.1 with HCl, followed by the addition of

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[0371] made up to a volume of 1000 ml with ultra pure water.

[0372] Substrate Solution:

[0373] Solution A:

	Citric acid 19.2 g (Beijing Chemical Plant)
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[0374] After sufficiently dissolved in 800 ml ultra pure water, the resultant solution was made up to a volume of 1000 ml by ultra pure water.

[0375] Solution B:

Na ₂ HPO ₄ •12H ₂ O	71.7 g (Beijing Chemical Plant)

[0376] After sufficiently dissolved in 800 ml ultra pure water, the resultant solution was made up to a volume of 1000 ml by ultra pure water.

[0377] Substrate Solution:

	Solution A Solution B	47.276 ml 50 ml	
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[0378] Above volumes of Solution A and Solution B were taken respectively and mixed, followed by filtration through a 0.22 µm filter to remove bacteria.

[0379] Stop solution 100 ml

Concentrated H2SO4

20 ml (Beijing Chemical Plant)

[0380] slowly added into 80 ml ultra pure water while stirring.

Mice Immunization

[0381] 1. Mice grouping

[0382] 1) HBsAg group

[0383] 2) HBsAg+CpG(684)(100 μg) group

[0384] 3) HBsAg+CpG(647)(100 µg) group

[0385] 4) HBsAg+CpG(685)(100 µg) group

[0386] 5) HBsAg+CpG(640)(100 µg) group

[0387] 6) HBsAg (25 mg A1³⁺/mg HBsAg)+ CpG(684)(100 μg) group

[0388] 7) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(647)(100 μ g) group

[0389] 8) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(685)(100 µg) group

[0390] 9) HBsAg (25 mg A1 $^{3+}$ /mg HBsAg)+ CpG(640)(100 µg) group

[0391] 2. Mice immunization: On week 0 and week 4, mice were inoculated according to different groups through tibialis anterior. Blood was drawn from the caudal vein of the mice three days prior to the inoculation (negative serum), immediately followed by adding 2 μ l of sodium heparin to each 10 μ l of the blood for the purpose of anticoagulation (0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 4° C. The plasma in the upper layer was collected and then stored at -20° C. (the dosage of HBV vaccine is 1 μ g/mouse, and the content of A1 adjuvant is 25 μ g/ml)

II. Methods

[0392] 1. Coating 100 µl HBsAg (1 mg/ml) was added into 10 ml coating solution. The diluted HBsAg was added into each well of an ELISA plate, which was then left stand overnight under a temperature of 4° C.

[0393] 2. Washing: The next day, the liquid left in the wells of the plate was removed completely, and 300 μ l washing solution was added into each well. Before removing the washing solution, the plate was left stand for 3 minute at room temperature. Finally, the plate was dried by tapping on an absorbent paper. Same washing procedure was repeated for 3 times.

[0394] 3. Blocking: $300 \mu l$ blocking solution was added into the wells of the ELISA plate, and was left stand for 2 hs at room temperature.

[0395] 4. Adding samples to be tested: Same washing procedures described in step 2 were applied. Samples to be tested were diluted into different concentrations with sample

diluent before being added. $100\,\mu l$ diluted sample was added into wells in duplicate and was left stand for 2 hs at room temperature.

[0396] 5. Adding HRP labeled goat-anti-mouse IgG2a and IgG1: Same washing procedures described in step 2 were applied. HRP labeled goat-anti-mouse IgG2a and IgG1 was diluted with sample diluent (1:1000). 100 µl diluted HRP labeled goat-anti-mouse IgG2a and IgG1 was added into wells and was left stand for 2 hs in darkness at room temperature.

[0397] 6. Adding substrate solution: Same washing procedures described in step 2 were applied. 100 µl freshly made substrate solution was added into each well, and left stand for 20 minutes in darkness at room temperature.

[0398] 7. Adding stop solution: The ELISA plate was incubated for 20 minutes, after which 50 μ l stop solution was added into each well.

[0399] 8. ELISA assay (A492 nm): ELISA assay (A492 nm) was carried out within 5 minutes after the addition of the stop solution.

[0400] 9. Determination of a positive value: A positive value is defined where the OD value of a sample/the OD value of the negative control is 2 or more.

[0401] Compared with A1 adjuvant, the use of CpG as adjuvant produced a higher level of IgG2a antibody. The result of variance analysis (See FIG. 11) showed significant differences (P<0.05).

Assay of CTL Specific Killing

[0402] (1) Preparation of effector cell 12 weeks after vaccination, the spleen of the mice was removed under sterile condition and placed into a dish containing IMDM (Gibcol Co.) supplemented with 10% calf serum. The spleen was gently ground with a frosted glass plate, and then filtered through a filter screen having 200 meshes. 3-5 ml RBC lysis solution (139.6 mmol/L NH₄Cl, 16.96 mmol/L Tris, pH was adjusted to 7.2 with 1 mol/HCl) was added to the filtered fluid, and the resultant was left stand for 10 minutes. The spleen cells were washed twice with normal saline before being counted. The spleen cells was adjusted to a concentration of 3×10⁷/ml with IMDM containing 10% calf serum. Mitomycin (Sigma, formulated with IMDM containing no serum to a concentration of 500 µg/ml) was added into P815 cells (ATCC) transfected with HbsAg gene to a final concentration of 50 µg/ml. The resultant was incubated at 37° C. for 2 hs and then washed 3 times with normal saline. 1×10⁶ P815 cells were co-cultured with the spleen cells for 5 days at 37° C. in 5% CO₂, after which the spleen cells were collected as effector cells.

[0403] (2) 51 Cr labeled target cells: 100 μ l 51 Cr (Perkin Elmer life Science) was added into 1×10^6 P815 cells transfected with HbsAg gene. The resultant was incubated at 37° C. for 1 h, during which the P815 cells were gently stirred every 5-10 minutes. The P815 cells were used as target cells after being washed three times with IMDM containing 10% calf serum.

[0404] (3) CTL killing test: The effector cells were diluted into different concentrations with IMDM containing 10% calf serum, into which ⁵¹Cr labeled P815 cells were added to a final ratio of effector cells: target cells of 100:1~12.5:1.

After incubated at 37° C. for 4 hs, the supernatant of the cells was collected and tested for c.p.m value.

[0405] (4) The result indicated as an adjuvant, CpGODN can significantly improve HBV-specific CTL activity produced by mice stimulated with HbsAg. Compared with A1 adjuvant+HbsAg, CpGODN+HbsAg elicited a stronger HBV-specific CTL activity (P<0.05) (See FIG. 12).

EXAMPLE 15

CpG ODN Enhanced the Response of Suckling Mice to HBV Vaccine

I. Animals and Regents

[0406] 1. Animals: BALB/c suckling mouse, female, aged from 6-8 days (available from Beijing Weitonglihua Experimental Animal Ltd.).

[0407] 2. HBsAg: containing A1 adjuvant (25 mg A1^{3+/} mg HBsAg) or containing no A1 adjuvant, purchased from Beijing Institute of Biological Product.

[0408] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0409] 4. CpGODN formulation 100 µg CpG ODN were dissolved into 10 µl PBS to prepare application solutions.

[0410] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 10 μ l CpG ODN application solution and 1 μ l HBsAg (containing A1 adjuvant or not containing A1 adjuvant) application solution were first mixed thoroughly and then placed on ice for 10 minutes before the injection.

II. Methods

[0411] 1. Suckling mice grouping 110 suckling mice/group

[**0412**] 1) HBsAg group

[0413] 2) HBsAg+A1 adjuvant group

[**0414**] 3) HBsAg+CpG (684) (100 μg) group

[**0415**] 4) HBsAg+CpG (647) (100 μg) group

[**0416**] 5) HBsAg+CpG (685) (100 µg) group

[**0417**] 6) HBsAg+CpG (640) (100 μg) group

[**0418**] 7) HBsAg+CpG(684) (100 μg)+A1 adjuvant group

[0419] 8) HBsAg+CpG(647) (100 μg)+A1 adjuvant group

[0420] 9) HBsAg+CpG(685) (100 µg)+A1 adjuvant group

[0421] 10) HBsAg+CpG(640) (100 μ g)+A1 adjuvant group

[0422] 2. Test of whether CpG ODN can enhance the response of suckling mice to HBV vaccines: The experimental suckling mice were divided into four groups, i.e., HBsAg group, HBsAg+A1 adjuvant group, HBsAg+Cp-GODN group, and HBsAg+A1 adjuvant+CpGODN group. The suckling mice were inoculated according to different groups through tibialis anterior. The mice were boosted 4 weeks after the initial inoculation (week 0). Blood was drawn 1 week, 4 weeks, 6 weeks, 8 weeks and 10 weeks after the inoculation, immediately followed by adding 2 μl

of sodium heparin to each 10 µl of the blood for the purpose of anticoagulation (0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 4° C. The plasma in the upper layer was collected and then stored at -20° C. Plasma was separated from the blood and ELISA was used to assay HbsAb titer (As shown in Example 3). The result of the assay indicates, when compared with HBsAg group or HBsAg (25 mg A13+/mg HBsAg) group, the use of CpGODN alone or in combination with A1 adjuvant both can improve the HBsAb titers in suckling mice. All the results of the variance analyses showed significant differences (P<0.05). The HBsAb titer produced by the suckling mice is the highest when CpGODN is used in combination with A1 adjuvant. The comparison of antibody titers among different groups is shown in FIG. 13. The test result of this example indicated CpG ODN can elicit relatively stronger humoral immunity response from individuals having weak humoral immunity response.

EXAMPLE 16

CpG ODN Enhanced the Response of Aged Mice to HBV Vaccine

I. Animals and Regents

[0423] 1. Animals: BALB/c mouse, female, aged from 20-24 months (Beijing Weitonglihua Experimental Animal Ltd.).

[0424] 2. HBsAg: containing A1 adjuvant (25 mg A1^{3+/} mg HBsAg) or containing no A1 adjuvant, purchased from Beijing Institute of Biological Product.

[0425] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0426] 4. CpGODN formulation 100 μg CpG ODN were dissolved into 50 μl PBS to prepare application solutions.

[0427] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 μ l CpG ODN application solution and 1 μ l HBsAg (containing A1 adjuvant or not containing A1 adjuvant) were first mixed thoroughly and then placed on ice for 10 minutes before the injection (the dosage of HBV vaccine is 1 μ g/mouse, and the content of A1 adjuvant is 25 μ g/ml).

II. Methods

[0428] 1. Mice grouping: 10 mice/group

[**0429**] 1) HBsAg group

[0430] 2) HBsAg+A1 adjuvant group

[0431] 3) HBsAg+CpG(684) (100 μg) group

[**0432**] 4) HBsAg+CpG(647) (100 μg) group

[0433] 5) HBsAg+CpG (685) (100 μ g) group

[**0434**] 6) HBsAg+CpG (640) (100 μg) group

[0435] 7) HBsAg+CpG(684) (100 μg)+A1 adjuvant group

[0436] 8) HBsAg+CpG(647) (100 µg)+A1 adjuvant group

[0437] 9) HBsAg+CpG(685) (100 µg)+A1 adjuvant group

[0438] 10) HBsAg+CpG(640) (100 μ g)+A1 adjuvant group

[0439] 2. Antibody assay: The aged mice were divided into four groups, i.e., HBsAg group, HBsAg+A1 adjuvant group, HBsAg+CpGODN group, and HBsAg+A1 adjuvant+ CpGODN group. The aged mice were inoculated according to different groups through tibialis anterior. The mice were boosted 4 weeks after the initial inoculation (week 0). Blood was drawn 3 days prior to the inoculation (negative serum), immediately followed by adding 2 µl of sodium heparin to each 10 µl of the blood for the purpose of anticoagulation (0.2 g heparin was weighted and dissolved in 100 ml ddH₂O to obtain a concentration of 0.2%, and the resultant solution was sterilized under a pressure of 15 pounds for 20 minutes). The blood sample was centrifuged at 4,000 rpm for 20 minutes under 4° C. The plasma in the upper layer was collected and then stored at -20° C. Blood was drawn 2 weeks, 4 weeks, 6 weeks, 8 weeks, 10 weeks and 12 weeks after the inoculation. Plasma was separated from the blood and ELISA was used to assay HbsAb titer (As shown in Example 3). The result of the assay indicates, when compared with HBsAg group or HBsAg (25 mg A1³⁺/mg HBsAg) group, the use of CpGODN alone or in combination with A1 adjuvant both can improve the HBsAb titers in aged mice. All the results of the variance analyses showed significant differences (P<0.05). The HBsAb titer produced by the aged mice is the highest when CpGODN is used in combination with A1 adjuvant. The comparison of antibody titers among different groups is shown in FIG. 14. The test result of this example indicated CpG ODN can elicit relatively stronger humoral immunity response from individuals having weak humoral immunity response.

EXAMPLE 17

CpG ODN Enhanced the Response of Rhesus to HBV Vaccine

I. Animals and Regents

[0440] 1. Animals rhesus, 2-4 kg, female (Beijing Weitonglihua Experimental Animal Ltd.).

[0441] 2. HBsAg: containing A1 adjuvant (25 mg A1³⁺/ mg HBsAg) or containing no A1 adjuvant, purchased from Beijing Institute of Biological Product.

[0442] 3. CpGODN: synthesized by Shanghai Shenggong Biotechnology Service Ltd.

[0443] 4. CpGODN formulation 100 µg CpG ODN were dissolved into 50 µl PBS to prepare application solutions.

[0444] 5. HBsAg formulation: 1 mg lyophilized HBsAg protein powders were dissolved into 1 ml PBS to prepare application solution. For intramuscular injection, 50 µl CpG ODN application solution and 1 µl HBsAg (containing A1 adjuvant or not containing A1 adjuvant) application solution were first mixed thoroughly and then placed on ice for 10 minutes before the injection (the dosage of HBV vaccine is 1 µg/rhesus, and the content of A1 adjuvant is 25 µg/ml).

II. Method

[0445] 1. Rhesuses grouping:

[0446] 1) HBsAg(10 µg)+A1 adjuvant group

[0447] 2) HBsAg(10 μg)+CpG (684) (1000 μg) group

[0448] 3) HBsAg (10 µg)+CpG (647) (1000 µg) group

[0449] 4) HBsAg (10 μ g)+CpG (685) (1000 μ g) group

[0450] 5) HBsAg (10 µg)+CpG (640) (1000 µg) group

[0451] 6) HBsAg(10 μ g)+CpG (684) (1000 μ g)+A1 adjuvant group

[0452] 7) HBsAg (10 μ g)+CpG (647) (1000 μ g)+A1 adjuvant group

[0453] 8) HBsAg (10 μ g)+CpG (685) (1000 μ g)+A1 adjuvant group

[**0454**] 9) HBsAg(10 µg)+CpG (640) (1000 µg)+A1 adjuvant group

[0455] 2. Antibody assay: Blood was drawn from the rhesuses before inoculation, and negative plasma was isolated from the blood. The rhesuses were inoculated according to different groups through deltoid. The rhesuses were boosted 4 weeks after the initial inoculation (week 0). Blood was drawn 2 weeks, 4 weeks, 6 weeks, 8 weeks, 10 weeks and 12 weeks after the inoculation. Plasma was separated form the blood and ELISA was used to assay HbsAb titer. The result of the assay indicates, when compared with HBsAg group or HBsAg (25 mg A1³⁺/mg HBsAg) group, the use of CpGODN alone or in combination with A1 adjuvant both can improve the HBsAb titers in rhesuses. All the results of the variance analyses showed significant differences (P<0.05). The HBsAb titer produced by the rhesuses is the highest when CpGODN is used in combination with A1 adjuvant. The comparison of antibody titers among different groups is shown in FIG. 15.

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26

- 1. A single strand deoxynucleotide, comprising one or more CpG dinucleotides.
- 2. The single strand deoxynucleotide of claim 1, wherein the deoxynucleotide is represented by one of the following formulas:
 - (i) (G)_n(L)_nX₁X₂CGY₁Y₂(M)_n(G)_n, wherein X₁=A, T or G; X₂=A or T; Y₁=A or T; Y₂=A, T or C; L, M=A, T, C or G; n is 0-6;
 - (ii) $(G)_n(L)_nCG(XY)_nCG(M)_n(G)_n$, wherein X=A or T; Y=A or T; L, M=A, T, C or G; n is 0-6;
 - (iii) (TCG)_n(L)_nCG(M)_n(G)_n, wherein L, M=A, T, C or G; n is 0-6;
 - (iv) (TCG)_n(L)_nX₁X₂CG(M)_n, wherein X₁=A, T or G; X₂=A or T; L, M=A, T, C or G; n is 0-6; or
 - (v) sequences containing TTCGTCG.
- 3. The single strand deoxynucleotide of claim 2, comprising any one of SEQ ID NO: 1 to SEQ ID NO: 185.
- **4**. The single strand deoxynucleotide of claim 3, comprising any one of SEQ ID NO: 109, 150, 167 or 168.
- 5. The single strand deoxynucleotide of claim 1, wherein bases of the deoxynucleotide are modified by one or more of the following modifications: non-sulpher modification, sulpher modification, partial sulpher modification, rare base modification, methylation modification, and other modifications where sulfhydryl, Aminolinker C6, Thiol-C6 S—S, etc. are used to couple to other substances.
- **6.** A vaccine adjuvant comprising the single strand deoxynucleotide of claim 1.
- 7. The vaccine adjuvant of claim 6, wherein the single strand deoxynucleotide is used alone or in combination with a non-nucleic acid adjuvant including aluminum adjuvant, Freund's adjuvant, MPL, or emulsion.
- **8**. The vaccine adjuvant of claim 6, wherein the single strand deoxynucleotide is admixed with or chemical coupled to a vaccine, or the single strand deoxynucleotide is cloned into a DNA vaccine.

- 9. The vaccine adjuvant of claim 8, wherein the vaccine is selected from the group consisting of hepatitis B virus blood-derived vaccine, hepatitis B virus genetic engineering protein vaccines, HBV virus vector vaccine, hepatitis B virus bacterium vector vaccine, hepatitis B virus transgenic plant vaccine, rabies virus blood-derived vaccine, rabies virus genetic engineering protein vaccines, rabies virus vector vaccine, rabies virus bacterium vector vaccine, and rabies virus transgenic plant vaccine, and the DNA vaccine is selected from the group consisting of hepatitis B virus DNA vaccine and rabies DNA vaccine.
- 10. A method of improving immunogenicity of a vaccine, comprising coadministering said vaccine to a subject with at least one single strand deoxynucleotide of claim 1 and a vaccine.
- 11. The method of claim 10, further comprising administering non-nucleic acid adjuvants.
 - 12. The method of claim 10, wherein the subject is human.
- 13. The method of claim 10, wherein the subject is a mammal.
- 14. A method for preventing virus infection and a disease related to virus infection comprising administering to a subject in need thereof the single strand deoxynucleotide of claim 1.
- **15**. The method of claim 14, wherein the single strand deoxynucleotide is administered alone or in combination with a vaccine for preventing virus infection.
- **16**. The method of claim 15, wherein said administering is intramuscular, subcutaneous, mucosal, gastrointestinal, intravenous, or intraperitoneal.
- 17. The single strand deoxynucleotide of claim 5, wherein said rare base modification is dI or dU.

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