* 2 ml tubes and 15 µl/mg feces get the numbers a bit higher
* 2 ml tubes get rid of the squeeze problem
* Think about the best (most thorough) homogenization that is easily feasible
  + Basically decided: use 15 µl Buffer per mg feces but NB: you can max have then 120 mg aliquots in a 2ml tube because you need to add 1800 µl Buffer!
* Using Filter or spin:
  + Filter gives slightly or really higher numbers (might be because you concentrated the solution by losing volume)
  + Still open question: If you decide for filter, make sure the amount of volume you lose is equal between samples.
* Big Problem, the unstained samples:
  + Contamination from previously run stained samples:
    - Arguing for:
    - Arguing against:
      * Buffer samples is almost without events even after stained samples
  + Temperature: something becomes autofluorescent when the cells warm up
    - Arguing for:
      * It seems the longer you wait the worse the problem gets
    - Arguing against:
  + Test options:
    - Run unstained samples after fridge (should be clean), take plate, run again unstained samples 40 min later (without having had stained samples on)
    - Warm up to 37 degrees, does it get worse
    - Run stained, then unstained, should be dirty, then clean the machine like Lau suggested, run again unstained, should be clean now if contamination
    - Run Buffer without DAPI
* Other big problem: The balance:
  + Please don’t accept uncertainty of more than 1-2 mg