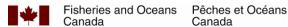
Proceedings of the Technical Expertise in Stock Assessment (TESA) national workshop on 'Best practices in age estimation', 31 January to 02 February 2023 in Moncton, New Brunswick

Daniel Ricard, Peter Comeau, Aaron Adamack, Jacob Burbank, Abby Daigle, Allan Debertin, Kim Emond, Tracey Loewen, Andrea Perreault, Gregory Puncher, Karen Robertson, Nicolas Rolland, Meredith Schofield, Andrew Smith, François-Étienne Sylvain and Stephen Wischniowski

Science Branch Gulf Region Fisheries and Oceans Canada Moncton, New Brunswick, E1C 5K4, Canada

2023

Canadian Technical Report of Fisheries and Aquatic Sciences ####





Canadian Technical Report of Fisheries and Aquatic Sciences

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Rapport technique canadien des sciences halieutiques et aquatiques

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PROCEEDINGS OF THE TECHNICAL EXPERTISE IN STOCK ASSESSMENT (TESA) 6 NATIONAL WORKSHOP ON BEST PRATICES IN AGE ESTIMATION, 31 JANUARY TO 02 FEBRUARY 2023 IN MONCTON, NEW BRUNSWICK

by 9

Daniel Ricard¹, Peter Comeau², Aaron Adamack³, Jacob Burbank¹, Abby Daigle¹, Allan Debertin⁴, Kim Emond⁵, Tracey Loewen⁶, Andrea Perreault³, Gregory Puncher², Karen Robertson¹, Nicolas Rolland¹, Meredith Schofield³, Andrew Smith⁵, Francois-Étienne Sylvain¹ and Stephen Wischniowski⁷

¹Gulf Fisheries Centre, Fisheries and Oceans Canada, 343 Université Avenue, Moncton, New Brunswick, E1C 9B6

²Bedford Institute of Oceanography, Fisheries and Oceans Canada, 1 Challenger Dr., Dartmouth, Nova Scotia, B2Y 4A2

³Nortwest Atlantic Fisheries Centre, Fisheries and Oceans Canada, 80 E White Hills Rd, St. John's, Newfoundland and Labrador, A1A 5J7

⁴Saint Andrews Biological Station, Fisheries and Oceans Canada, 125 Marine Science Dr., St. Andrews, New Brunswick, E5B 0E4

⁵Maurice Lamontagne Institute, Fisheries and Oceans Canada, 850 route de la Mer, Mont-Joli, Québec, QC G5H 3Z4

⁶Freshwater Institute, Fisheries and Oceans Canada, 501 University Crescent, Winnipeg, Manitoba, R3T 2N6

⁷Pacific Biological Station, Fisheries and Oceans Canada, 3190 Hammond Bay Road, Nanaimo, British Columbia, V9T 6N7

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57 ABSTRACT

Ricard, D., Comeau, P., Adamack, A., Burbank, J., Daigle, A., Debertin, A., Emond, K., Loewen,
T., Perreault, A., Puncher, G., Robertson, K., Rolland, N., Schofield, M., Smith, A., Sylvain,
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(TESA) national workshop on best pratices in age estimation, 31 January to 02 February 2023 in Moncton, New Brunswick. Can. Tech. Rep. Fish. Aquat. Sci. nnn: v+25 p.

A three-day workshop was held in Moncton, New Brunswick, from January 31 to February 02 2023. The aim of the workshop was to examine and discuss best practices in the use of structures for age estimation. A summary of the different presentations is given along with a brief overview of discussions that took place during the workshop. A total of 43 recommendations are formulated to provide guidance for practitioners involved in the collection of otoliths and other hard parts, and in analyses of age estimates.

69 **RÉSUMÉ**

Ricard, D., Comeau, P., Adamack, A., Burbank, J., Daigle, A., Debertin, A., Emond, K., Loewen,
T., Perreault, A., Puncher, G., Robertson, K., Rolland, N., Schofield, M., Smith, A., Sylvain,
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Un atelier de trois jours s'est tenu à Moncton, au Nouveau-Brunswick, du 31 janvier au 2 février 2023. Le but de l'atelier était d'examiner et de discuter des meilleures pratiques en matière d'utilisation de structures pour l'estimation de l'âge. Un résumé des différentes présentations est donné ainsi qu'un bref aperçu des discussions qui ont eu lieu au cours de l'atelier. Un total de 43 recommandations sont formulées pour guider les praticiens impliqués dans la collecte d'otolithes et d'autres parties dures, et dans l'analyse des estimations d'âge.

1 Introduction

- The purpose of the Technical Expertise in Stock Assessment (TESA) program is to promote stock assessment excellence through organizing national activities that contribute to the development of expertise in stock assessment across Fisheries and Oceans Canada (DFO).
- TESA was created in 2009, in response to a loss of stock assessment expertise in DFO owing to retirements. Each year, TESA organises three or four training courses and one or two topical workshops on stock assessment issues. TESA is a national program with one or two representatives in each region for more details see the GCpedia website (only accessible from the DFO network).
- This report provides a written record of the TESA workshop on "Best practices in age estimation" that was held in Moncton, NB, from January 31 to February 02 2023. The list of participants can be found in Table 2 and all resources related to the workshop were stored in a git repository maintained by TESA.

4 1.1 Motivations

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The use of an age-based population model is often lauded as the "gold standard" in stock assessment. Age-based models are favored over simpler population models because of their ability to convey biological realism and to provide a more nuanced understanding of population dynamics, including the effects of fishing on harvested populations. As the name implies, an age-based model requires information about the age of individuals in the population, which provides scientists with a foundation for analyzing the demographics of a population as it evolves over time. Age estimates, in conjunction with other observations, can inform us on the growth of individuals, the age-at-maturation of individuals, the age structure of a population and on other vital factors necessary to understand variations in numbers and biomass.

All DFO regions collect structures from marine organisms that are used in estimating their age.
The procedures associated with collecting, cataloguing, storing, and obtaining age estimates of samples vary from lab to lab. It is important to follow best practices when estimating age from hard structures and not all personnel are necessarily aware of these practices.

In any given year, DFO collects otoliths for a number of stocks, and processes these structures to obtain age estimates. It is estimated that tens of thousands of otoliths and scales are collected yearly in scientific activities at DFO. For example, thousands of otoliths are collected during annual mutli-species scientific surveys conducted by different DFO regions. Additional otoliths are collected in commercial port sampling activities and others come from fisheries observers.

The logistical, financial and human resources associated with these otolith sampling programs are significant.

While the use of age-based models is common and the software available to implement such assessments are available and evolving, the manipulation of the input data that feeds into those models is often done in an ad-hoc fashion particular to the lab or agency in charge of the assessment. While these ad-hoc methods are most likely defensible and appropriate, the details

- that go into the computation of catch-at-age matrices are often poorly documented, making results hard to reproduce by someone outside a given lab or agency.
- This workshop was formulated to provided an opportunity for DFO scientists involved in age estimation, in analyses of age and length information, and in developing age-based assessments, to share ideas and develop better methods to obtain and use age estimates data in stock assessments.

125 1.2 Objectives

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The objectives of this workshop were to create a forum for discussion and for exchanging ideas among DFO scientists that use age-based models and that are involved in age estimation activities. In particular, the workshop provided:

- guidance on hard structures collection and on the sampling design for collecting hard structures
- guidance on age determination using hard structures, best practices for annuli validation, reference collections, age estimation technician calibration, archival of hard structures and data warehousing
- · guidance on digital imaging of hard structures
- guidance on going from length frequency samples and age-length keys to catch-at-age matrices that feed into age-based population models

137 **1.3 Format**

A hybrid meeting was held from January 31 2023 to February 02 2023. The list of participants and the attendance can be found in Table 2. A total of 76 participants attended the event (30 in person and 46 virtually), including two external experts who joined the meeting virtually. The workshop was attended by a wide variety of DFO personnel ranging from research scientists, to biologists, to fisheries technicians and also included two students and a professor from a local university.

2 Workshop activities and presentations

The workshop was facilitated by its two co-chairs and included a number of presentations from DFO personel and external experts (Table 1). Plenary presentations were followed by discussions between workshop participants on the topics at hand, with the main goal of exchanges being to openly share knowledge and to reach agreement on what constitutes "best practices" for age estimation using hard parts.

As part of the workshop planning, participants were asked to provide a summary of physical collections of hard structures and to identify the programs that were involved in the collection and analysis of ages using these structures. Given the large number of taxa for which age estimation materials are collected by different regions of Fisheries and Oceans Canada (Figure 2), the recommendations from this workshop involve programs spanning the whole country, and concern stocks in the Atlantic, Arctic and Pacific oceans, as well as a number of diadromous and freshwater species. The recommendations also apply to a number of structures, including ototliths, scales, vertebrae and mollusc shells (Table A.1).

2.1 Day 1 - Basics of age estimation

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The first day covered the basics of age estimation. The literature on research associated with otoliths is vast and varied. Peter Comeau gave a presentation on the history and realities of age estimation of marine organisms. His presentation highlighted the importance of age estimation in stock assessments. He also pointed out that there once was a stigma associated with jobs that involved age estimation, they were perceived as an entry-level position with a low retention rate, whereas the reality is that they require a unique skill set, considerable experience and are essential to scientific inquiries related to fish population dynamics.

Julie Coad Davies gave an overview of the age estimation activities that she is involved in, in her role as lab manager for DTU Aqua and also through her chairing the ICES Working Group on SmartDots Governance (WGSMART) and the ICES Working Group on Biological Parameters (WGBIOP). Julie highlighted the importance of clear communication between laboratories and the paramount role of exchanges of both physical otoliths and digital images to identify potential age estimation biases that may exist between laboratories. Similarly, the ICES working groups have in their mandate the formulation of shared methodologies and protocols and provide guidelines for how to best captures information on biological parameters and how to conduct otolith exchanges between laboratories (ICES).

Stephen Wischniowski shared his experience of running an age estimation laboratory (the Sclerochronology Lab at the Pacific Biological Station) and being involved in the Committee of Age Reading Experts (CARE). The demands for age estimation services from the lab far exceed its ability to process samples and a "quota" is imposed on the number of otoliths that the lab can process. The lab technicians are taught the intricacies associated with age estimation from all types of species and structures. Stephen estimates that it takes five years to train a technician to the point where they can reliably estimate ages and can teach others.

Sylvie Robichaud and Karen Robertson presented the procedures associated with age
estimation of Atlantic Herring in the Gulf Region. There are two spawning components that must
first be distinguished from each other before age estimates can be made. The morphology of
otoliths is used to distinguish between spawning components and the number of annuli and the
date of capture are then integrated into an age estimate. The otoliths come from a variety of
sources, ranging from research surveys to experimental fishing and from commercial fisheries.
The laboratory protocols used to process samples require the extraction of otoliths, which are
subsequently mounted in epoxy resin, photographed, and are then used for age estimation.

60 Kim Emond and Hélène Dionne presented the procedures associated with age estimation of

Atlantic Herring in the Québec Region. The otolith based techniques to decipher between spring spawning and fall spawning herring stocks of Atlantic Herring used by the Quebec region were explained and shown. Similarities were seen between approaches used by the technicians in the Gulf region and those in the Quebec region both for estimating Herring ages and spawning component assignment. The stock assessment for this stock relies on a variety of data sources and on a comprehensive sampling of otoliths. Ages determined from otolith's are essential to the stock assessment as they are a precursor for many important population model inputs such as the catch-at-age.

Tania Davignon-Burton gave a talk of reconciling expectations and realities in a production age estimation environment. There are often high expectations for estimating ages using otoliths, but the laboratory activities required to obtain age estimates mean that some prioritization must take place. In essence, it is not always quick and easy to age otoliths, it takes time and often demand outweighs technician time.

During the question period there was discussions pertaining to the importance of cleaning and storing hard structures appropriately. It was highlighted that clean otoliths are required for taking high quality pictures that are optimal for age estimation. Previously some labs had stored otoliths in glycerin solutions, however there was a consensus that it is better to store otoliths dry, this is better for otolith preservation, reading of older otoliths, otolith microchemistry and isotopes. Several participants agreed its best to clean otoliths immediately upon extraction, dry appropriately, and store dry, this is best for age estimation, microchemistry and isotopes. If otoliths are read with the help of solutions such as glycerin, after reading otoliths should be cleaned, dried and stored dry.

Tracey Loewen and Rick Wastle presented their work on estimating ages of difficult-to-age species and otolith microchemistry techniques used for age estimation. They explained that their Otolith Ageing Lab at the Freshwater Institute in Winnipeg is responsible for age estimation of 32 species found in the Arctic, including but not limited to Arctic Char (Salvelinus alpinus), Dolly Varden (Salvelinus malma), Lake Trout (Salvelinus namaycush), Greenland Halibut (Reinhardtius hippoglossoides), Redfish (Sebastes) and Arctic Cod (Boreogadus saida). Otolith microchemistry techniques are used to support and validate age estimation technique, such as annual Zinc markers that align with growth rings on otoliths. Rick Wastle went in depth on approaches used to age Greenland Halibut in the Arctic as this species is difficult to age because of its slow growth. Despite the difficulty with estimating ages for the species, the lab persisted, testing a wide range of approaches before settling on what is perceived as the best and appropriate age estimation methods and these methods that they developed improve the understanding of growth for the species. They elaborated that Strontium Calcium (SrCI) marking was used to try and validate growth rings with moderate success, and concluded sectioning the left otolith bulge as well as longitudinal sectioning have advantages for estimating ages in this slow growing species.

Daniel Ricard presented the Standard Operating Procedures (SOP) document used in the Gulf Region. This document is meant as a central point for any person seeking information about age estimation in the Gulf Region. The SOP document contains detailed information the collection, preparation, age estimation and storage of age structures for fish species that are aged in the Gulf region. The summary of otolith collections held by the Gulf Region and available as OpenData records was also presented and suggested as a starting point for documenting

physical collections, including reference collections.

2.2 Day 2 - Digital imaging of ageing structures

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The second day of the workshop focused on topics related to the capture of digital images of 237 ageing structures, how to annotate, and common software to store and manipulate images. 238

The day started with a tour of the laboratory facilities used in age estimations at the Gulf 239 Fisheries Centre. The laboratories used for age estimation of Atlantic Cod, American Plaice, 240 Winter Flounder, Yellowtail Flounder and Atlantic Herring. The facilities used for imaging whole 241 otoliths mounted in epoxy and imaging of whole otoliths were visited and participants had the opportunity to ask questions and try the tools and software used by their Gulf Region colleagues. Participants also visited the Atlantic Cod, American Plaice, Winter Flounder, Yellowtail Flounder and Atlantic Herring otolith collections, had the opportunity to see the vast amounts of otoliths 245 stored at the Gulf Fisheries Centre and see how the ageing structures are soted and archeived. 246

A presentation about SmartDots from external expert Julie Coad Davies showed how this tool is being used for otolith exchanges between laboratories in Europe. The development of the SmartDots application is managed by ICES Working Group on SmartDots Governance (WGSMART) in close collaboration with the ICES Secretariat as well as national fisheries laboratories. There was tremendous interest among DFO employees to use SmatDots. Several people noted the value of being able to exchange digital otoliths and provide several annotations/reader interpretations on an otolith image and compare and contrast them. THis would also allow individuals have readers from other regions age an otolith to provide crossregion comparisons of age interpretation and would help house ageing structure interpretations in one common place.

Karen Robertson gave a presentation on taking good images of otoliths, which could be 257 transferable to other ageing structures. She showed how it is important to have a reliable 258 workspace with appropriate lighting and a microscope equipped with a digital camera. The image capture can be done in a number of software, including the one provided by the microscope 260 manufacturer, the Leica LAS X software suite. Karen also pointed out the importance of having a well-defined naming convention for images and a strict protocol for file storage. Image 262 processing is an important step to improve the readability of otolith images and a number of 263 filtering options are available. There were brief discussions on what the appropriate naming 264 convention should be and consensus among the group that this should be standardized when 265 possible. 266

David Fishman presented options for developing a storage solution for otolith/ageing structure images in the form of DFO Dots, which can be integrated with Smartdots. A discussion about DFO Dots followed. Participants felt that having access to a tool that helps in managing otolith 269 collections and curating reference collections would be beneficial to their work. There was general 270 support for devlopement of DFO Dots across participants. 271

A group discussion on the reporting features of *SmartDots* and how the tool can be used for 272 obtaining growth increments followed. Participants noted the value of having annotated imaged 273 otoliths and being able to readily generate quick reports in SmartDots to rapidly assess results of 275 age reading runs.

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Others, particularily those in production ageing labs noted capturing digital images of ageing structures adds time to the laboratory process used to obtain age estimates. It is unclear 277 whether the benefits of obtaining digital images of ageing structures outweighs the costs 278 associated with the additional time required. To address this it was suggested that as a starting 279 point, a reference collection should be imaged. Having a reference collection of imaged 280 otoliths/ageing structures allows researchers to document examples of what otoliths/ageing 281 structures of a particular age-class look like and to explain why classification of being a different 282 age would be incorrect. Reference collections are also useful for training new readers and for 283 providing continuity in ageing methodologies when there are discontinuities in an aging program 284 (e.g. times when there is no overlap between new and former readers). 285

Once digital images of ageing structures are available, an additional step is to annotate them. It was notes that it is probably impractical to annotate images in production ageing situations, but as a starting point the reference collection should be annotated in SmartDots. This provides an authoritative record that can be used to better document the reference collection. If steps are done to facilitate more rapid imagaing of otoliths and pontetially automated annotations, such approaches could eventually be integrated into the workflow of production ageing labs. However, such advances are far off and the automation of image annotation can have major drawbacks, as annotation and reading of age structures takes significant expertise and can include some subjective decisions on unclear growth increments.

2.3 Day 3 - Analysis of age estimates

The third day of the workshop focused on what is done with age estimates, namely the use of such data in fitting growth models and in computing catch-at-age matrices. Such applications of ageing data are often the backbone of stock assessment models, making their computation imperative for well informed estimates of population abundance and projections.

Daniel Ricard and Andrea Perreault gave presentations on growth models. Daniel Ricard 300 showed how growth models are typically fit and gave examples of developing growth models for 301 American Plaice. Daniel also showed how to effectively fit growth models for males and females 302 and highlighted the importance of weighting observations to provide a more true fit. Andrea 303 Perrault gave a talk on the importance of accounting for length-stratified sampling when fitting 304 growth models from survey data as length-stratified sampling can lead to biased estimates of 305 growth model parameters (Perreault et al. 2020). Andrea went over several potential methods 306 that can be used to account for length-stratified length sampling when fitting growth models and with simulations showed the method that leads to the least amount of bias. During the 308 question period Andrea pointed out the risks of overestimating Linf, which include overestimation 309 of biomass and one would think fish are bigger than they actually are, and this bias propegates 310 through to the biomass estimate. 311

Lisa Ailloud from NOAA NMFS gave a presentation about age-length keys. She discussed the distinction between forward and reverse age-length keys, and provided an alternative hybrid methodology that was previously applied to bluefin tuna (Ailloud et al. 2019; Ailloud and Hoenig 2019). She also described how to fill gaps in age-length keys when not enough information is

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Catch-at-age calculations presentation by Andrew Smith from the Quebec Region described
his experience with trying to replicate catch-at-age calculations that were previously done in a
bespoke software called *catch.exe*. An R package that was developed to compute catch-at-age
matrices was also presented (Ouellette-Plante et al. 2022). Use of the package and cautions of
trying to reproduce exact catch-at-age matrices from previous work that made several subjective
decisions was shared. The catchR package was presented and it was shown how the package
can reduce subjectivity in the computation of catch-at-age.

Kim Emond from the Quebec Region presented the methodologies used in the herring stock assessment to compute catch-at-age matrices from commercial landings and from hydroacoustic surveys. Once again, Kim highlighted the need to reduce subjectivity in computations of catch-at-age, to document decisions and be consitent with decision making across years.

Christopher Corriveau and Ellie Weise from Dalhousie University presented their work on using DNA methylation to estimate ages. The presentation provided an overview of this new methodology and highlied a novel potential way forward for ageing that could provide less reader subjectivity if refined and calibrated appropriately.

A group discussion finally took place to draft the recommendations that appears in section 5. The recommendations cover a variety of subjects, ranging from practical recommendations for age estimation to institutional recommendations that could foster the development of a community of age readers within DFO.

3 Remarks from external experts

Remarks from Julie Coad Davies from DTU Aqua:

The workshop has been an excellent forum for those working with age reading and the age data resulting from their work. With such a variety of expertise across labs and regions under the DFO it is important that a community is formed to facilitate knowledge sharing on best practices. There are opportunities for age reading calibration across labs who are age reading the same stocks, cooperation on updating reference collections and knowledge sharing on image acquisition and method testing.

Future plans to have a DFO version of SmartDots will facilitate this and will support an overall improvement in the quality of the age data coming from the age reading labs and how the data is subsequently used in the stock assessment process. Forming a working group who meet annually will foster the communication required to sustain the community.

Remarks from Lisa Ailloud from NOAA NMFS:

The 2023 TESA best practices in ageing workshop was a very successful meeting. It provided a forum for experts from different labs and regions to share

experiences, pain points and new developments in fish age determination and modeling. Age and growth play an important role in stock assessment and any biases in these input quantities can ultimately affect the correct evaluation of stock status and management advice. It is therefore essential to identify the potential sources of bias and take the necessary steps to mitigate them. Discussions carried out during the workshop helped advance this goal. Having a wide range of expertise present helped broaden the discussions and bridge the gap between the various stages of data and model development.

As participants shared their individual experiences, advances and setbacks, it became clear that many of the issues raised were common across labs and relevant to the group as a whole. This type of forum where solutions can be worked out and shared among experts is an important step towards homogenizing protocols and increasing overall efficiency. Participants clearly showed an interest in understanding past practices and improving upon them. Discussions around making use of automation and open science principles to increase transparency in the process were very encouraging. New technological developments were also shared with the group. Preliminary results for their application appear very promising.

4 Discussion

Over and over during the workshop, it was emphasized that while otoliths are the most common structure used for estimating ages, other ageing structures are also used, and the best practices used for otoliths are applicable to other structures.

lt was discussed how age estimation using ageing structures is more than just "counting the rings" and requires a wealth of experience in order to interpret the patterns observed on ageing structures.

The workshop attendees identified the need to develop and foster a community of DFO scientists whose tasks involve age estimation. As such, the workshop provided a starting point in establishing this community.

5 Recommendations

The working group formulated the following recommendations for how to deal with physical collections of structures used for age estimation.

1. Institutional

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- (a) Foster a community of DFO scientists whose mandated tasks include sampling of ageing structures, age estimation using ageing structures or analysis of age estimates data.
- (b) Emulate the Pacific Ocean's Committee of Age Reading Experts ([CARE]https://care.psmfc.org/) for the Atlantic and Arctic Oceans.

- (c) Creation of a DFO working group that meets regularly to ensure that age estimations practices in different labs follow shared best practices.
- (d) Provide support for inter-regional ageing structure exchanges and secondary reader testing.
- (e) Achieve Canadian representation on appropriate ICES working groups related to quality assurance of age estimation as input for stock assessment (WGBIOP) or those developing the SmartDots platform (WGSMART).
- (f) Recognise the fact that the skills required to obtain unbiased age estimates from ageing structures are unique and take time and dedication to acquire.

2. Ageing structure cataloguing, storage and inventory

- (a) Ensure that physical collections of ageing structures follow the DFO "Policy on Collection, Storage, Management and Use of Physical Samples for Science Research".
- (b) Ageing structures should be stored in an environment that minimizes degradation and that maintains readability.
- (c) Ageing structures removed from an individual should be uniquely identified.
- (d) Clearly label ageing structures so they can be traced back to their collection, implement good bookkeeping of your ageing structures.
- (e) An electronic inventory of the physical ageing structures available should be documented and updated regularly.
- (f) A subset of ageing structures should be preserved in their unaltered state for future unforeseen usage (i.e. not in resin or glycerin).

3. Reference collection

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- (a) Actively curate reference collections so that old ageing structures that have lost their readability are replaced by new ageing structures, this includes renewing ageing structures.
- (b) Ensure that the age structures in the reference collection are representative of what will be available to age estimation technicians (the structures should cover wide spatial and temporal ranges, both annual and inter annual, all ages used in the stock assessment and both easy and difficult to read ageing structures,...).
- (c) Ensure reference collections are updated with samples from recent years.
- (d) In addition to a physical reference collection, develop its digital equivalent by taking images of the ageing structures.
- (e) Ideally, also annotate your digital reference collection using SmartDots.
- (f) Strive to have a validation study to ascertain the periodicity of ageing patterns.

4. Laboratory operations

(a) Strive to obtain unbiased age estimates by regularly performing age reader calibrations and by carrying out regular quality-checks for each species/stock (both within and across laboratories).

- (b) Develop and publish Standard Operating Procedures for each lab.
 - (c) Develop standardized protocols to validate age estimates (add to your regional SOP).
 - (d) Favour age estimation in a "blind" setting, where no prior knowledge (except date of capture) is used when interpreting patterns in ageing structures, don't let outside information influence your age estimates.
 - (e) Establish a quality-control process to identify mistakes made during data entry (e.g. typos).
 - (f) Document the uncertainty associated with age estimates by means of a standardized quality assurance scale.
 - (g) Generate an age-error matrix as part of your standard procedures.
 - (h) Have at least two age readers for any given stock, for redundancy.

5. Training

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- (a) Develop a document that details the training steps for new agers, and/or add it to your regional SOP.
- (b) Establish procedures that integrate current practices (e.g. labels that have been used for many year) into improved practices.

6. Digital imaging of ageing structures

- (a) Ensure that a correctly calibrated scale bar is present in the image so that the image scale in pixels per mm can be determined.
- (b) The file naming convention to use when taking images of ageing structures should uniquely identify each ageing structure, should contain the image number (in cases where more than one picture is taken for each ageing structure) and also contain the resolution of the image (in pixels per mm).
- (c) When practical, promote the capture of digital images of ageing structures.
- (d) Digital images (JPEG format) of ageing structures should be stored in enterprise-level infrastructure where proper backups are in place.
- (e) A DFO-led app to store and retrieve digital images of ageing structures should be implemented to facilitate the management of these images and to provide integration with the SmartDots software.

7. Analysis of age estimates

- (a) When computing catch-at-age matrices, the sampling design used in the collection of data should be accounted for.
- (b) Analyses that compute catch-at-age matrices from length samples and age estimates should strive to be fully documented and reproducible.
- (c) Contemporary procedures that reduce bias in age structure estimates should be used.
- (d) Assessment methods would ideally incorporate ageing errors into the assessment framework.

- 8. Preparing for changes associated with a warming climate
 - (a) Rates of growths are likely to change, so are the patterns associated with age estimation. So regularly revisit the pattern in the growth structures recognition used for estimating ages to identify periods of weaker or stronger growth and/or changes in the timing of annuli formation.
 - (b) Continue inter-regional communication and organise events that promote collaborations.
 - (c) Northward migration of species will shift stock boundaries and change the species that will require science advice.
 - (d) Establish a process so that "cheat sheets" and guides used for training agers, and the associated laboratory processes for age estimation are updated regularly.
- 9. Research and development

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- (a) Pursue further studies in sclerochronology that will support the identification of changes in patterns used in estimating ages.
- (b) Investigate the trade-offs involved between taking images and annotating them versus just estimating a single age-length pair.

These recommendations were formulated at the workshop and further revised by all authors.
They represent procedures that should be followed by all DFO personnel involved in age
estimations and also contain suggestions for fostering a community of age readers within DFO.

6 Acknowledgements

The authors would like to acknowledge all the workshop participants for their attendance and their active participation in discussions. Joeleen Savoie helped with workshop organisation and logistics. Lisa Leblanc from the ASEC was extremely generous with her time and hospitality during the workshop. We thank the Gulf Region publication coordinator Jeffery Clements for his help in handling this report submission.

7 Tables

Table 1. List of presentations given at the TESA workshop on best practices in age estimation.

Presenter(s)	Presentation title	Link to slides
Day 1		
Peter Comeau	Fish age determination - Some of the basics	Power Point file
Julie Davies		PDF file
Tracey Loewen and Rick Wastle	Otolith microchemistry, difficult- to-age marine species, element marking in otoliths	PDF file
Stephen Wischniowski Daniel Ricard		PDF file
Kim Emond and Hélène Dionne	Age determination of Atlantic Herring in the Québec region	Power Point file
Sylvie Robichaud and Karen Robertson	Age determination of Atlantic Herring in the Gulf region	Power Point file
Tania Davignon-Burton	Reconciling dreams, expectations and reality in a production ageing environment	Power Point file
Day 2		
Julie Davies	SmartDots – a tool created by the users for the users	PDF file
Karen Robertson, Isabelle Forest and Sylvie Robichaud	Taking good pictures of otoliths, and annotating them in SmartDots	Power Point file
Day 3		
Andrea Perreault	Impacts of ignoring length-stratified sampling design	Power Point file
Lisa Ailloud	Analyses of ageing data / A general theory of age-length keys	Power Point file
Kim Emond and Hélène Dionne	Catch-at-age of commercial herring landings and numbers-at-age from acoustics surveys	PDF file
Chris Corriveau and Ellie Weise	Developing aging clocks for fish using DNA methylation	Power Point file

Table 2. Alphabetical list of participants (by last name) to the TESA worskhop on best practices in age estimation. Participants from Fisheries and Oceans Canada (DFO) are identified by their region (NL is the Newfoundland and Labrador Region, MAR is the Maritimes Region, GUL is the Gulf Region, QUE is the Quebec Region, OP is Ontario and Prairie Region, ARC is the Arctic Region and PAC is the Pacific Region).

Name	Affiliation	Attendance	Day 1	Day 2	Day 3
Aaron Adamack	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Lisa Ailloud (external	NOAA NMFS	Virtual			\checkmark
expert)					
Laura Alsip	DFO - ARC	Virtual	\checkmark	\checkmark	\checkmark
Kelly Antaya	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Mark Billard	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Jacob Burbank	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Lauren Burke	DFO - OP	Virtual	\checkmark	\checkmark	\checkmark
Barbara Campbell	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Karalea Cantera	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Lynn Collier	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Peter Comeau (co-chair)	DFO - MAR	In person	\checkmark	\checkmark	\checkmark
Chelsea Cooke	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Christopher Corriveau	Dalhousie University	In person	\checkmark	\checkmark	\checkmark
Abby Daigle	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Andrew Darcy	DFO - GLF	In person	\checkmark	\checkmark	
Guillaume Dauphin	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Julie Davies (external	DTU Aqua	Virtual	\checkmark	\checkmark	\checkmark
expert)					
Tania Davignon-Burton	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Allan Debertin	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Nell den Heyer	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Mathieu Desgagnés	DFO - QUE	Virtual	\checkmark	\checkmark	\checkmark
Hélène Dionne	DFO - QUE	In person	\checkmark	\checkmark	\checkmark
Dwight Drover	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Kim Emond	DFO - QUE	In person	\checkmark	\checkmark	\checkmark
Gillian Forbes	DFO - NL	In person	\checkmark	\checkmark	\checkmark
Isabelle Forest	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Danni Harper	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Sarah Hawkshaw	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Victoria Healey	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Erin Herder	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Kendra Holt	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Matthew Horsman	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Mary-Jane Hudson	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Samantha Hudson	DFO - GLF	Virtual	\checkmark	\checkmark	\checkmark
Yeongha Jung	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Kelly Kraska	DFO - MAR	In person	\checkmark	\checkmark	\checkmark
Madeline Lavery	DFO - PAC	Virtual	\checkmark	\checkmark	✓

Name	Affiliation	Attendance	Day 1	Day 2	Day 3
Michael Legge	DFO - OP	In person	✓	√	\checkmark
Marc Legresley	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Lingbo Li	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Tracey Loewen	DFO - OP	In person	\checkmark	\checkmark	\checkmark
Ellen MacEachern	DFO - MAR	In person	\checkmark	\checkmark	\checkmark
Colin MacFarlane	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Brendan K Malley	DFO - ARC	Virtual	\checkmark	\checkmark	\checkmark
Kiana Matwichuk	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Mackenzie Mazur	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Judy McArthur	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Kelsey McGee	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Jessie Mcintyre	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Liz Miller	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Maya Miller	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Kirby Morrill	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
George Nau	DFO - MAR	Virtual	\checkmark	\checkmark	\checkmark
Andrea Perreault	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Hannah Polaczek	DFO - NL	Virtual	\checkmark	\checkmark	\checkmark
Gregory Puncher	DFO - MAR	In person	\checkmark	\checkmark	\checkmark
Catriona	DFO - MAR	, Virtual	\checkmark	\checkmark	\checkmark
Regnier-McKellar					
Daniel Ricard (co-chair)	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Kierstyn Rideout	DFO - NL	In person	\checkmark	\checkmark	\checkmark
Karen Robertson	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Sylvie Robichaud	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Nicolas Rolland	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Chelsea Rothkop	DFO - PAC	Virtual	\checkmark	\checkmark	\checkmark
Daniel Ruzzante	Dalhousie University	Virtual			\checkmark
Meredith Schofield	DFO - NL	In person	\checkmark	\checkmark	\checkmark
Andrew Smith	DFO - QUE	Virtual	\checkmark	\checkmark	\checkmark
Jolene Sutton	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
François-Étienne Sylvain	DFO - GLF	In person	\checkmark	\checkmark	\checkmark
Jaime Thomson	DFO - NL	In person	✓	✓	✓
François Turcotte	DFO - GLF	Virtual			✓
Audrey Ty	DFO - PAC	Virtual	\checkmark	\checkmark	✓
Kari Underhill	DFO - GLF	In person	✓	✓	✓
Lenore J Vandenbyllaardt	DFO - ARC	Virtual	· ✓	√	· ✓
Rick J Wastle	DFO - ARC	Virtual	· ✓	√	↓
Emily Way-Nee	DFO - MAR	Virtual	√	√	↓
Ellie Weise	Dalhousie University	In person	√	,	↓
Gabrielle Wilson	DFO - MAR	Virtual	√	√	↓
Stephen Wischniowski	DFO - PAC	Virtual	√	√	↓

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Name	Affiliation	Attendance	Day 1	Day 2	Day 3
Emily Yungwirth	DFO - PAC	Virtual	✓	✓	√

8 Figures

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Figure 1. Day 1 of the TESA workshop "best practices in age estimation" held at the Atlantic Science Enterprise Center in Moncton, NB on 31 January, 01-02 February 2023.

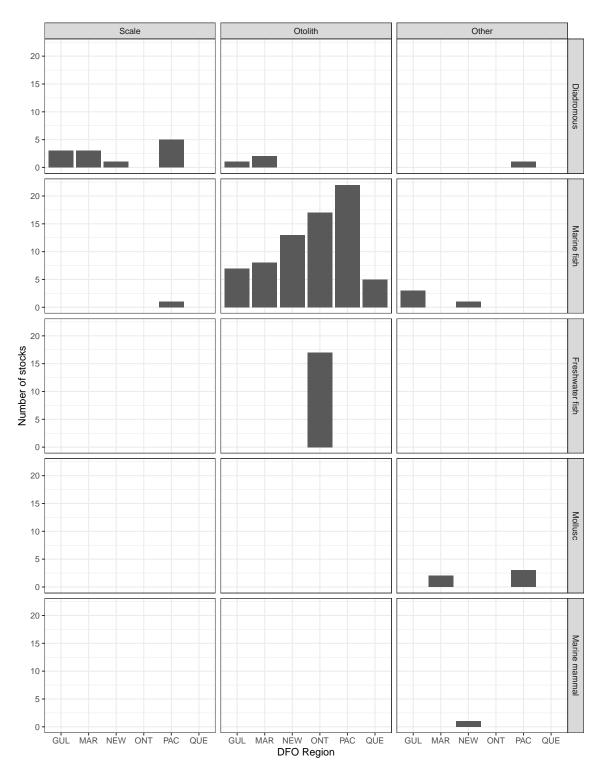


Figure 2. Number of stocks for which ageing materials are collected and analysed by the different regions of Fisheries and Oceans Canada. The list of collections used to generate this figure appear in Table A.1.

9 References

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APPENDIX A List of DFO age structure collections

As part of the planning for the workshop, participants were asked to provide an overview of age structures that are collected as part of scientific activities in different regions of DFO. The list appearing in Table A.1 shows the different collections that exist at DFO.

Table A.1. Fisheries and Oceans Canada (DFO) regions (NL is the Newfoundland and Labrador Region, MAR is the Maritimes Region, GUL is the Gulf Region, QUE is the Quebec Region, OP is Ontario and Prairie Region, ARC is the Arctic Region and PAC is the Pacific Region). SFA is Salmon Fishing Area, NAFO is the Northwest Atlantic Fisheries Organization and PMFC is the Pacific Marine Fisheries Commission

Region	Common name	Scientific name	Stock	Type of structure	Years collected
GUL	Atlantic Cod	Gadus morhua	NAFO 4T	otolith	1971 to 2022
GUL	White Hake	Urophycis tenuis	NAFO 4T	otolith	1971 to 2022
GUL	American Plaice	Hippoglossoides platessoides	NAFO 4T	otolith	1971 to 2022
GUL	Winter Flounder	Pseudopleuronectes americanus	NAFO 4T	otolith	1971 to 2022
GUL	Yellowtail Flounder	Limande ferruginea	NAFO 4T	otolith	1971 to 2022
GUL	Witch Flounder	Glyptocephalus cynoglossus	NAFO 4RST	otolith	1971 to 2022
GUL	Winter Skate	Leucoraja ocellata	NAFO 4T	vertebrae	2004 to 2007
GUL	Smooth Skate	Malacoraja senta	NAFO 4T	vertebrae	2004 to 2017
GUL	Thorny Skate	Amblyraja radiata	NAFO 4T	vertebrae	2004 to 2013
GUL	Atlantic Herring	Clupea harengus	NAFO 4T	otolith	1965 to 2022
GUL	Alewife	Alosa pseudoharengus	SFA 16, 18	scale	1983 to 2022
GUL	Atlantic Salmon	Salmo salar	SFA 15, 16, 18	scale	1973 to 2022
GUL	Blueback Herring	Alosa aestivalis	SFA 18	otolith	2021 and 2022
GUL	Striped Bass	Morone saxatilis	SFA 16, 18	scale	1995 to 2022
MAR	Atlantic Cod	Gadus morhua	NAFO 5Z	otolith	1970 to 2022
MAR	Atlantic Cod	Gadus morhua	NAFO 4X	otolith	1970 to 2022
MAR	Atlantic Herring	Clupea harengus	NAFO 4VWX	otolith	1954 to 2022
MAR	Haddock	Melanogrammus aeglefinus	NAFO 5Z	otolith	1970 to 2022
MAR	Haddock	Melanogrammus aeglefinus	NAFO 4X	otolith	1970 to 2022
MAR	Pollock	Pollachius pollachius	NAFO 4X	otolith	1970 to 2022
MAR	Silver Hake	Brosme brosme	NAFO 4VWX	otolith	1975 to 2022

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Region	Common name	Scientific name	Stock	Type of structure	Years collected
MAR	Atlantic Halibut	Hippoglossus	NAFO	otolith	1995 to 2022
		hippoglosus	3NOPs4VWX5Zc		
MAR	Arctic Surf Clam	Mactromeris polynyma		shell	??
MAR	Sea Scallop	Placopecten		shell	??
		magellanicus			
MAR	Atlantic Salmon	Salmo salar		scale	1950s to 2022
MAR	Alewife	Alosa pseudoharengus		scale	1970s to 2022
MAR	Blueback Herring	Alosa aestivalis		scale	1970s to 2022
MAR	Striped Bass	Morone saxatilis		otolith / Scales	1980s to 2022
MAR	American Eel	Anguilla rostrata		otolith	1980 to 2022
QUE	Atlantic Herring	Clupea harengus	NAFO 4RS	otolith	4R: 1965 to 2022,
					4S: 1985 to 2022
QUE	Capelin	Mallotus sp.	NAFO 4RST	otolith	1984 to 2022
QUE	Atlantic Mackerel	Scomber scombrus	NAFO 3-5	otolith	1973 to 2022
QUE	Atlantic Halibut	Hippoglossus hippoglossus	NAFO 4RST	otolith	1990 to 2022
QUE	Atlantic Cod	Gadus morhua	NAFO 3Pn4RS	otolith	1974 to 2022
PAC	Turbot/Arrowtooth flounder	Atheresthes stomias	PMFC 3CD, 5ABCDE	otolith	1980 to 2022
PAC	Petrale Sole	Eopsetta jordani	PMFC 3CD, 5ABCDE	otolith	1964 to 2022
PAC	Dover Sole	Microstomus pacificus	PMFC 3CD, 5ABCDE	otolith	1979 to 2022
PAC	Rougheye Rockfish	Sebastes aleutianus	PMFC 3CD, 5ABCDE	otolith	1978 to 2022
PAC	Pacific Ocean Perch	Sebastes alutus	PMFC 3CD, 5ABCDE	otolith	1973 to 2022
PAC	Redbanded Rockfish	Sebastes babcocki	PMFC 3CD, 5ABCDE	otolith	1986 to 2022
PAC	Shortraker Rockfish	Sebastes borealis	PMFC 3CD, 5ABCDE	otolith	1980 to 2022
PAC	Silvergray Rockfish	Sebastes brevispinis	PMFC 3CD, 4B, 5ABCDE	otolith	1973 to 2022
PAC	Copper Rockfish	Sebastes caurinus	PMFC 3CD, 4B, 5ABCDE	otolith	1984 to 2022

Region	Common name	Scientific name	Stock	Type of structure	Years collected
PAC	Widow Rockfish	Sebastes entomelas	PMFC 3CD, 4B, 5ABCDE	otolith	1979 to 2022
PAC	Yellowtail Rockfish	Sebastes flavidus	PMFC 3CD, 4B, 5ABCDE	otolith	1977 to 2022
PAC	Bocaccio	Sebastes paucispinis	PMFC 3CD, 5ABCDE	otolith	1978 to 2022
PAC	Canary Rockfish	Sebastes pinniger	PMFC 3CD, 4B, 5ABCDE	otolith	1973 to 2022
PAC	Redstripe Rockfish	Sebastes proriger	PMFC 3CD, 5ABCDE	otolith	1977 to 2022
PAC	Yellowmouth Rockfish	Sebastes reedi	PMFC 3CD, 5ABCDE	otolith	1977 to 2022
PAC	Yelloweye Rockfish	Sebastes ruberrimus	PMFC 3CD, 4B, 5ABCDE	otolith	1979 to 2022
PAC	Quillback Eockfish	Sebastes maliger	PMFC 3CD, 4B, 5ABCDE	otolith	1979 to 2022
PAC	Black Spotted Rockfish	Sebastes melanostictus	PMFC 3CD, 5ABCDE	otolith	1966 to 2022
PAC	Sablefish	Anoplopoma fimbria	PMFC 3ACD,5ABCE	otolith	1965 to 2022
PAC	Lingcod	Ophiodon elongatus	PMFC 3CD,5ABCDE	finray/otolith	1965 to 2022
PAC	Hake	Merluccius productus	PMFC 3CD,4B,5ABCE	otolith	1985 to 2022
PAC	Eulachon	Thaleichthys pacificus	selective collections	otolith	2015 to 2022
PAC	Herring	Clupea harengus pallasi	2E,2W,3-8,10, 14,17,19,23-27 SA6	scale	1992 to 2022
PAC	Chum	Oncorhynchus keta	2-29, 110-130, WCVI, Yukon	scale	1900 to 2022
PAC	Coho	Oncorhynchus kisutch	2-20, 110-130	scale	1901 to 2022
PAC	Steelhead	Oncorhynchus mykiss	incidental	scale	1902 to 2022
PAC	Sockeye/Kokanee	Oncorhynchus nerka	1-29,120,130, WCVI	scale	1903 to 2022
PAC	Chinook	Oncorhynchus tshawytscha	2-29,110-130, WCVI, Yukon	scale	1904 to 2022
PAC	Abalone	Holiotus kamtschatkana	Research	shell	2000 to 2022

Region	Common name	Scientific name	Stock	Type of structure	Years collected
PAC	Geoduck	Panopea abrupta	NC, CC, HG, SOG, WCVI	shell	2003 to 2020
PAC	Manila clam	Tapes philippinarum	Research	shell	1995 to 2022
PAC	Rocky Mountain Riged Mussel	Gonidea angulata	Okanogan lake CAN	shell	2019 to 2022
NEW	Capelin	Mallotus villosus	NAFO 2J3KL	otolith	1978 to 2022
NEW	Atlantic Herring	Clupea harengus	NAFO 3KLPs	otolith	1965 to 2022
NEW	Atlantic Cod	Gadus morhua	NAFO 2J3KL, 3NO, 3Ps	otolith	1950s to 2022
NEW	American Plaice	Hippoglossoides platessoides	NAFO 2J3KL, 3NO, 3Ps	otolith	1978 to 2022
NEW	Greenland Halibut	Reinhardtius hippoglossoides	NAFO 2+3KLMNO	otolith	1978 to 2022
NEW	White Hake	Urophycis tenuis	NAFO 3PLNO	otolith	1984 to 2019
NEW	Atlantic Salmon	Salmo salar	NAFO 2HJ, 3KLMNO, 3Ps, 4R	scale	1975 to 2023
NEW	Harp seal	Pagophilus groenlandicus	Northwest Atlantic	canine tooth	1979 to 2022
NEW NEW	Wolffish (3 Species) Sand Lance	Anarhichadidae spp. Ammodytes spp.	NAFO 2J3KLNOP	otolith otolith	2001 to 2006
NEW	Redfish	Sebastes spp.	NAFO 2HJ3K, 3LN, 3O	otolith	1978 to 2022
NEW	Yellowtail Flounder	Limande ferruginea	NAFO 3LNO	otolith	1978 to 2022
NEW	Witch Flounder	Glyptocephalus cynoglossus	NAFO 2J3KL, 3NO, 3Ps	otolith	1978 to 2022
NEW	Atlantic Halibut	Hippoglossus hippoglossus	NAFO 3NOPs4VWX5Zc	otolith	1978 to 2022
NEW	Haddock	Melanogrammus aeglefinus	NAFO 2HJ3K, 3LNO, 3Ps	otolith	1978 to 2022
NEW	Skates (Various)	Rajidae spp.	NAFO 2J3KLNOP	vertebrae	2004 to 2022

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Region	Common name	Scientific name	Stock	Type of structure	Years collected
ONT	Broad Whitefish	Coregonus nasus	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Lake Cisco	Coregonus artedi	Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Arctic Cisco	Coregonus autumnalis	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Least Cisco	Coregonus sardinella	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Inconnu	Stenodus leucichthys	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Arctic Grayling	Thymallus arcticus	Various Locations Arctic Canada	otoliths and fin clips	1970s to 2022
ONT	Round Whitefish	Prosopium cylindrecium	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Arctic Char	Salvelinus alpinus	Canada and international locations	otoliths and fin clips	1970s to 2022
ONT	Lake Trout	Salvelinus namaycush	Canada	otoliths and fin clips	1970s to 2022
ONT	Bull Trout	Salvelinus confluentus	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Dolly Varden Char	Salvelinus malma	Western Canadian Arctic	otoliths and fin clips	1970s to 2022
ONT	Burbot	Lota lota	Canada	otoliths	1970s to 2022
ONT	Walleye	Sander vitreum	Southern Canada	otoliths, dorsal spines	1970s to 2022
ONT	Greenland Halibut	Reinhardtius hippoglossoides	NAFO Div. 0	otoliths	1996 to 2022
ONT	Northern Pike	Esox lucius	Canada	otoliths and cliethra	1970s to 2022
ONT	Arctic Flounder	Liopsetta glacialis	Canadian Arctic	otoliths	2012 to 2022
ONT	Starry Flounder	Platichthys stellatus	Canadian Arctic	otoliths	2012 to 2022
ONT	Slimy Sculpin	Cottus cognatus	Canadian Arctic	otoliths	2012 to 2022

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Region	Common name	Scientific name	Stock	Type of structure	Years collected
ONT	Fourhorn Sculpin	Myoxocephalus quadricornis	Canadian Arctic	otoliths	2012 to 2022
ONT	Arctic Staghorn Sculpin	Gymnocanthus tricuspis	Canadian Arctic	otoliths	2012 to 2022
ONT	Grubby Sculpin	Myoxocephalus aenaeus	Canadian Arctic	otoliths	2012 to 2022
ONT	Longnose Sucker	Catostomus catostomus	Canadian Arctic	otoliths	2012 to 2022
ONT	Pacific Herring	Clupea pallasii	Canadian Arctic	otoliths	2012 to 2022
ONT	Bering Wolffish	Anarhichas orientalis	Western Canadian Arctic	otoliths	2012 to 2022
ONT	Capelin	Mallotus villosus	Canadian Arctic	otoliths	2012 to 2022
ONT	Pacific Sand Lance	Ammodytes personatus	Canadian Arctic	otoliths	2012 to 2022
ONT	Banded Gunnel	Ammodytes personatus	Canadian Arctic	otoliths	2012 to 2022
ONT	Greenland Cod	Gadus ogac	Western Canadian Arctic	otoliths	2012 to 2022
ONT	Saffron Cod	Eleginus gracilis	Canadian Arctic	otoliths	2012 to 2022
ONT	Arctic Cod	Boreogadus saida	Canadian Arctic	otoliths	2012 to 2022
ONT	Polar Cod	Arctogadus glacialis	Canadian Arctic	otoliths	2012 to 2022
ONT	Channel Catfish	Ictalurus punctatus	Manitoba	otoliths	2022
ONT	Freshwater Drum	Aplodinotus grunniens	Unknown	otoliths	
ONT	Deepwater Redfish	Sebastes mentella	NAFO 0B	otoliths	2019 to 2022