

RH: No effect of the K-Pg event on mammal disparity.

Mammalian morphological diversity does not increase in response to the Cretaceous-Paleogene mass extinction and the extinction of the (non-avian) dinosaurs.

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Abstract

Popular science accounts state that after the extinction of the non-avian dinosaurs at the Cretaceous-Paleogene (K-Pg) boundary 66 million years ago, mammals rapidly diversified to fill their empty ecological niches. However, evidence for this is mixed. Palaeontological analyses suggest that mammals radiated in response to the K-Pg extinction event, whereas neontological analyses suggest that mammals began to radiate before K-Pg and were not greatly affected by it. Here we aim to end this debate by looking at fossil and living taxa simultaneously.

We investigated the effect of the K-Pg extinction event on mammalian morphological diversity (disparity) using two Total Evidence tip-dated phylogenies of Mammaliaformes and Eutheria, containing both fossil and living taxa. Using a novel, continuous time-slicing method for measuring changes in disparity-through-time, we found no significant change in disparity before and after the K-Pg boundary, under either a gradual or punctuated model of evolution. This implies that the extinctions at the end of the Cretaceous did not affect mammalian morphological evolution. Our findings contradict the popular theory that the non-avian dinosaurs and other Mesozoic tetrapods were restricting mammalian evolution, and that their extinction liberated ecological niches for mammals to evolve into.

(Keywords: disparity, punctuated equilibrium, gradual evolution, time slicing, K-Pg)

INTRODUCTION

Throughout history, life on Earth has suffered a series of mass extinction events resulting in drastic declines in global biodiversity (e.g. Raup, 1979; Benton and Twitchett, 2003; Renne et al., 2013; Brusatte et al., 2015). The long-term effects of mass extinctions, however, are more varied (Erwin, 1998), and include species richness increases in some clades (Friedman, 2010) and declines in others (Benton, 1985), changes in morphological diversity (Ciampaglio et al., 2001; Ciampaglio, 2004; Korn et al., 2013) and shifts in ecological dominance (e.g. Brusatte et al., 2008b; Toljagic and Butler, 2013; Benson and Druckenmiller, 2014). These shifts are characterised by the decline of one clade that is replaced by a different unrelated clade with a similar ecological role (e.g. Brachiopoda and Bivalvia at the end Permian extinction; Liow et al. 2015 but see Payne et al. 2014). Shifts in ecological dominance are of particular interest because they are a fairly common pattern observed in the fossil record (e.g. Foraminifera; Coxall et al. 2006 ; Ichtyosauria; Thorne et al. 2011; Plesiosauria; Benson and Druckenmiller 2014) and are often linked to major macroevolutionary processes such as adaptive (Losos, 2010) or competitive (Brusatte et al., 2008b) radiations.

One classical example of a shift in ecological dominance is at the Cretaceous-Palaeogene (K-Pg) mass extinction 66 million years ago (Renne et al., 2013), where many terrestrial vertebrates (including the dominant non-avian dinosaur group; Archibald 2011; Renne et al. 2013; Brusatte et al. 2015) went extinct, allowing placental mammals to dominate the fauna (Archibald, 2011; Lovegrove et al., 2014). Some

43 authors suggest this reflects placental mammals filling the “empty” niches left after the
44 K-Pg extinction event (Archibald, 2011; O’Leary et al., 2013), others suggest it reflects a
45 release from predation and/or competition (Slater, 2013; Lovegrove et al., 2014).
46 However, evidence for the diversification of placental mammals being driven by the
47 K-Pg extinction event is mixed. Thorough analysis of the fossil record (e.g. Goswami
48 et al., 2011; O’Leary et al., 2013) supports the idea that placental mammals diversified
49 after the K-Pg extinction event as there are no undebated placental mammal fossils
50 before it and many afterwards (Archibald, 2011; Goswami et al., 2011; Slater, 2013;
51 O’Leary et al., 2013; Wilson, 2013; Brusatte et al., 2015). Conversely, evidence from
52 molecular data suggests that the diversification of placental mammals started prior to
53 the K-Pg extinction event without being drastically affected by it (e.g. Douady and
54 Douzery, 2003; Bininda-Emonds et al., 2007; Meredith et al., 2011; Stadler, 2011).
55 Therefore, whether the diversification of placental mammals began before the K-Pg
56 extinction event, or in response to the extinctions at K-Pg, is a matter of great debate
57 (dos Reis et al., 2012; O’Leary et al., 2013; Springer et al., 2013; O’Leary et al., 2013; dos
58 Reis et al., 2014).

59 There are two main reasons why there is still debate about the timing of the
60 diversification of placental mammals. Firstly, palaeontological and neontological data
61 show different patterns; palaeontological data generally suggest that placental
62 mammals diversified after K-Pg (e.g. O’Leary et al., 2013), whereas neontological data
63 suggest that K-Pg extinction event had little to no effect on mammalian diversification

(Bininda-Emonds et al., 2007; Meredith et al., 2011; Stadler, 2011). We can solve this issue by using both palaeontological and neontological data in our analyses. The Total Evidence method allows us to use cladistic data for both living and fossil taxa, along with molecular data for living taxa, to build phylogenies (Ronquist et al., 2012). This method can also be combined with the tip-dating method (Ronquist et al., 2012; Wood et al., 2013) to get more accurate estimates of diversification times for both fossil and living species (but see Arcila et al., 2015). Here we use two recent Total Evidence tip-dated phylogenies of mammals (Slater, 2013; Beck and Lee, 2014) to investigate palaeontological and neontological taxa simultaneously.

A second issue is that diversity can be defined in many different ways. In many studies it is measured as taxonomic diversity or species richness (Stadler, 2011; Meredith et al., 2011; O’Leary et al., 2013), but often the more interesting aspect of diversity is related to the ecological niches the species occupy (Wesley-Hunt, 2005; Brusatte et al., 2008b; Toljagic and Butler, 2013), particularly if we want to make hypotheses about macroevolutionary processes (Pearman et al., 2008; Olson and Arroyo-Santos, 2009; Losos, 2010; Glor, 2010; Benton, 2015). Sometimes taxonomic diversity is used as a proxy for other kinds of diversity, however, species richness can be decoupled from morphological diversity (e.g. Slater et al., 2010; Ruta et al., 2013; Hopkins, 2013), so it may not be the best proxy for ecological diversity. We can instead use morphological diversity, also known as disparity (e.g. Wills et al., 1994; Erwin, 2007; Hughes et al., 2013), as a way to quantify changes in mammalian morphology that

should relate to the ecology of the species. However some methods for measuring disparity are outdated and make inappropriate assumptions. Many methods for quantifying changes in morphological diversity were proposed > 20 years ago (Foote, 1994; Wills et al., 1994) and are sometimes used without modifications (e.g., Brusatte et al., 2008a,b; Cisneros and Ruta, 2010; Thorne et al., 2011; Prentice et al., 2011; Brusatte et al., 2012; Toljagic and Butler, 2013; Ruta et al., 2013; Benton et al., 2014; Benson and Druckenmiller, 2014). Additionally, previous methods are based on an underlying assumption that changes in disparity occur by punctuated evolution (e.g. Wesley-Hunt, 2005) which is not always the case (Hunt et al., 2015). Finally, most studies of disparity through time use unequal time units based on biostratigraphy (Brusatte et al., 2008b, 2012; Toljagic and Butler, 2013). This can be tautological as biostratigraphy is already based on changes in fossil assemblages and morphology through time. To deal with these issues, we propose an updated approach to test whether mammals diversified in response to the K-Pg event, using morphological disparity, measured as cladistic disparity (see Methods), as our proxy for diversity.

Here we measure the disparity of living and fossil mammals before or after K-Pg, using data taken from two previously published studies (Slater, 2013; Beck and Lee, 2014). Using a novel time-slicing approach, we produce fine-grained estimates of disparity through time under two different models of morphological character evolution (either gradual or punctuated). We also test whether mammals display significant changes in disparity between the end of the Cretaceous and throughout the

Cenozoic.

Until now, this question has only been investigated using data from North American Therian mammals (excluding Monotremata) and without formally testing the effect of the K-Pg extinction event (Wilson, 2013). To our knowledge, this study is the first to approach the debate about the effects of the K-Pg extinction event on mammalian evolution using Total Evidence phylogenies and by calculating disparity through time in a continuous way. We find no significant changes in mammalian disparity between the end of the Cretaceous and any time during the Paleocene. These results suggest that the extinction of non-avian dinosaurs and other terrestrial vertebrate clades at the end of the Cretaceous did not affect mammalian morphological evolution.

METHODS

Cladistic data and phylogenies

We used the cladistic morphological matrices and the Total Evidence tip-dated trees (Ronquist et al., 2012) from Slater (2013, 103 taxa with 446 morphological characters;) and Beck and Lee (2014, 102 taxa with 421 morphological characters). We chose these two datasets because they have a similar number of taxa and morphological characters. Slater (2013) ranges from 310 million years ago (Ma; Late Carboniferous) to the present and focuses on the clade Mammaliaformes at the family-level and is called hereafter the

Mammaliaformes dataset. Beck and Lee (2014) ranges from 170 Ma (Middle Jurassic) to the present and focuses on Eutheria at the genus-level and is called hereafter the Eutheria dataset. We used the first and last occurrences reported in Slater (2013) and Beck and Lee (2014) as the temporal range of each taxon in our analysis. Both phylogenies are illustrated in the supplementary material (see Fig S1 and S2). Both trees contain few taxa compared to the overall species richness of living and fossil mammals (Bininda-Emonds et al., 2007; Archibald, 2011). This is because Total Evidence trees need a lot of data, particularly morphological data for living taxa that can be hard to locate (Guillerme and Cooper, 2016). Therefore, most Total Evidence studies to date contain one or two orders of magnitude fewer taxa than phylogenies based solely on molecular data (e.g. thousands of taxa in Bininda-Emonds et al. 2007; Meredith et al. 2011 *vs.* hundreds in Ronquist et al. 2012; Slater 2013; Wood et al. 2013; Beck and Lee 2014).

Estimating ancestral character states

For both datasets we used the re-rooting method (Yang et al., 1996; Garland and Ives, 2000) to get Maximum Likelihood estimates of the ancestral states for each character at every node in the tree, using the `rerootingMethod` function from the R package `phytools` version 0.4-45 (Revell, 2012; R Core Team, 2015). Where there was missing character data for a taxon we followed the method of Lloyd (2015) and treated missing data as any possible observed state for each character. For example, if a character had two observed states (0 and 1) across all taxa, we attributed the multi-state “0&1” value

to the taxon with missing data, representing an equal probability of being either 0 or 1. This allows the ancestral node of a taxon with missing data to be estimated with no assumptions other than that the taxon has one of the observed character states. To prevent poor ancestral state reconstructions from biasing our results, especially when a lot of error is associated with the reconstruction, we only included ancestral state reconstructions with a scaled Likelihood ≥ 0.95 . Ancestral state reconstructions with scaled Likelihoods below this threshold were replaced by missing data ("?").

Building the cladisto-space

To explore variations in mammalian disparity through time (defined here as the variation in morphologies through time), we used a cladisto-space approach (e.g. Foote, 1994, 1996; Wesley-Hunt, 2005; Brusatte et al., 2008b; Friedman, 2010; Toljagic and Butler, 2013; Hughes et al., 2013). This approach is similar to constructing a morphospace based on continuous morphological data (e.g. Friedman, 2010), except a cladisto-space is an approximation of the morphospace based on cladistic data (i.e. the discrete morphological characters used to build a phylogenetic tree). Mathematically, a cladisto-space is an n dimensional object that summarises the cladistic distances between the taxa present in a cladistic matrix (see details below). Although empirically inter-taxon distances are the same in a morphospace or a cladisto-space (Foth et al., 2012; Hetherington et al., 2015), we prefer the term cladisto-space to make it clear that this space is estimated using cladistic data and not morphometric data and because both objects have slightly different properties. For example, because of its inherent

combinatory properties, a cladisto-space is a finite theoretical object limited by the product of the number of character states, whereas a morphospace is an infinite theoretical object. Thus a cladisto-space will be overloaded if the number of taxa is higher than the product of the number of character states, although this is rarely an issue with empirical data (our cladisto-spaces have maximal capacities of 1.9×10^{181} taxa for the Mammaliaformes dataset, i.e. 101 orders of magnitude more taxa than the number of particles in the universe; and 4.5×10^{159} taxa for the Eutheria dataset).

To estimate the cladisto-spaces for each of our datasets we first constructed pairwise distance matrices of length k , where k is the total number of tips and nodes in the datasets. For each dataset separately, we calculated the $k \times k$ distances using the Gower distance (Gower, 1971), i.e. the Euclidean distance between two taxa divided by the number of shared characters. This allows us to correct for distances between two taxa that share many characters and could be closer to each other than to taxa with fewer characters in common (i.e. because some pairs of taxa share more characters in common than others, they are more likely to be similar). For cladistic matrices, using this corrected distance is preferable to the raw Euclidean distance because of its ability to deal with discrete or/and ordinated characters as well as with missing data (Anderson and Friedman, 2012). However, the Gower distance cannot calculate distances when taxa have no overlapping data. Therefore, we used the TrimMorphDistMatrix function from the Claddis R package (Lloyd, 2015) to remove pairs of taxa with no cladistic characters in common. This led to us removing 11 taxa

from the Mammaliaformes dataset but none from the Eutheria dataset.

After calculating our distance matrices we transformed them using classical multidimensional scaling (MDS; Torgerson, 1965; Gower, 1966; Cailliez, 1983). This method (also referred to as PCO; e.g. Brusatte et al. 2015; or PCoA; e.g. Paradis et al. 2004) is an eigen decomposition of the distance matrix. Because we used Gower distances instead of raw Euclidean distances, negative eigenvalues can be calculated. To avoid this problem, we first transformed the distance matrices by applying the Cailliez correction (Cailliez, 1983) which adds a constant c^* to the values in a distance matrix (apart from the diagonal) so that all the Gower distances become Euclidean ($d_{Gower} + c^* = d_{Euclidean}$; Cailliez 1983). We were then able to extract n eigenvectors for each matrix (representing the n dimensions of the cladisto-space) where n is equal to $k - 2$, i.e. the number of taxa in the matrix (k) minus the last two eigenvectors that are always null after applying the Cailliez correction. Contrary to previous studies (e.g. Brusatte et al., 2008a; Cisneros and Ruta, 2010; Prentice et al., 2011; Anderson and Friedman, 2012; Hughes et al., 2013; Benton et al., 2014), we use all n dimensions of our cladisto-spaces and not a subsample representing the majority of the variance in the distance matrix (e.g. selecting only m dimensions that represent up to 90% of the variance in the distance matrix; Brusatte et al. 2008b; Toljagic and Butler 2013).

Note that our cladisto-spaces represent an ordination of all possible mammalian morphologies coded in each study through time. It is unlikely that all morphologies will co-occur at each time point, therefore, the disparity of the whole cladisto-space is

expected to be greater than the disparity at any specific point in time.

Calculating disparity

Disparity can be estimated in many different ways (e.g. Wills et al., 1994; Ciampaglio, 2004; Thorne et al., 2011; Hopkins, 2013; Huang et al., 2015), however most studies estimate disparity using four metrics: the sum and products of ranges and variances, each of which gives a slightly different estimate of how the data fits within the cladisto-space (Foote, 1994; Wills et al., 1994; Brusatte et al., 2008a,b; Cisneros and Ruta, 2010; Thorne et al., 2011; Prentice et al., 2011; Brusatte et al., 2012; Toljagic and Butler, 2013; Ruta et al., 2013; Benton et al., 2014; Benson and Druckenmiller, 2014). Nonetheless, these methods suffer several methodological caveats. First, the range metrics are affected by the uneven sampling of the fossil record (Butler et al., 2012). Second, because we include all n dimensions in the analysis (see above), the products of ranges and variances will tend towards zero since the scores of the last dimension are usually really close to zero themselves. These features make using the sum and products of ranges and variances unfeasible in our study. Instead, we use a different metric that comes with no statistical assumptions for measuring the dispersion of the data in the cladisto-space: the median distance between tips and nodes and the centroid (similar but not equivalent to Wills et al. 1994; Korn et al. 2013; Huang et al. 2015) calculated as:

$$Disparity = median \sqrt{\sum (\mathbf{v}_n - Centroid_n)^2} \quad (1)$$

where:

$$Centroid_n = \frac{\sum(\mathbf{v}_n)}{k} \quad (2)$$

and \mathbf{v}_n is any of the n eigenvectors (i.e. any of the n dimensions of the cladisto-space), $Centroid_n$ is the mean value of the n^{th} eigenvector (equation 2) and k is the total number of tips and nodes. Note that we also calculated the sum and products of ranges and variances and refer to these results in the supplementary material (Fig S6 to S9).

Estimating disparity through time

Changes in disparity through time are generally investigated by calculating the disparity of taxa that occupy the cladisto-space during specific time intervals (e.g. Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; Hopkins, 2013; Benton et al., 2014; Benson and Druckenmiller, 2014). These time intervals are usually defined based on biostratigraphy (e.g. Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; Benton et al., 2014) but can also be arbitrarily chosen time periods of equal duration (Butler et al., 2012; Hopkins, 2013; Benson and Druckenmiller, 2014). However, this approach suffers from two main biases. First, if biostratigraphy is used to determine the time intervals, disparity may be distorted towards higher differences between time intervals because biostratigraphical periods are geologically defined based on differences in the morphology of fossils found in the different strata. Second, this approach assumes that all characters evolve following a punctuated equilibrium model, because disparity is only estimated once for each interval resulting in all

changes in disparity occurring between intervals, rather than also allowing for gradual changes within intervals (Hunt et al., 2015).

To address these issues, we used a “time-slicing” approach that considers subsets of taxa in the cladisto-space at specific equidistant points in time, as opposed to considering subsets of taxa between two points in time. This results in even-sampling of the cladisto-space across time and permits us to define the underlying model of character evolution (punctuated or gradual). In practice, time-slicing considers the disparity of any element present in the phylogeny (branches, nodes and tips) at any point in time. When the phylogenetic elements are nodes or tips, the eigenvector scores for the nodes (estimated using ancestral state reconstruction as described above) or tips are directly used for estimating disparity. When the phylogenetic elements are branches we chose the eigenvector score for the branch using one of two evolutionary models:

1. **Punctuated evolution.** This model selects the eigenvector score from either the ancestral node or the descendant node/tip of the branch regardless of the position of the slice along the branch. Similarly to the time interval approach, this reflects a model of punctuated evolution where changes in disparity occur either at the start or at the end of a branch over a relatively short time period and clades undergo long periods of stasis during their evolution (Gould and Eldredge, 1977; Hunt, 2007). We applied this model in three ways:

- (i) selecting the eigenvector score of the ancestral node of the branch (ACCTRAN).

(ii) selecting the eigenvector score of the descendant node/tip of the branch (DELTRAN).

(iii) randomly selecting either the eigenvector score of the ancestral node or the descendant node/tip of the branch (random).

Method (i) assumes that changes always occur early on the branch (accelerated transition, ACCTAN) and (ii) assumes that changes always occur later (delayed transition, DELTRAN). We prefer not to make either assumption so we report the results from (iii), although the ACCTAN and DELTRAN results are available in the Supplementary Information (Fig S6 to S9).

2. Gradual evolution. This model also selects the eigenvector score from either the ancestral node or the descendant node/tip of the branch, but the choice depends on the distance between the sampling time point and the end of the branch. If the sampling time point falls in the first half of the branch length the eigenvector score is taken from the ancestral node, conversely, if the sampling time point falls in the second half of the branch length the eigenvector score is taken from the descendant node/tip. This reflects a model of gradual evolution where changes in disparity are gradual and cumulative along the branch. Under this model, the gradual changes could be either directional or random, however, directional evolution have been empirically shown to be rare (only 5% of the time Hunt, 2007). We therefore considered that changes from a character state A to B were only dependent on the branch length.

We applied our time-slicing approach separately to the two cladisto-spaces calculated for the Mammaliaformes and Eutheria datasets, time-slicing the phylogeny every five million years from 170 Ma to the present resulting in 35 subsamples of the cladisto-space. For each subsample, we estimated its disparity assuming punctuated (ACCTRAN, DELTRAN and random) and gradual evolution as described above. To reduce the influence of outliers on our disparity estimates, we bootstrapped each disparity measurement by randomly resampling with replacement a new subsample of taxa from the observed taxa in the subsample 1000 times. We then calculated the median disparity value for each subsample along with the 50% and 95% confidence intervals. We also recorded the number of phylogenetic elements (nodes and tips) in each subsample as a proxy for taxonomic diversity. To compare our results to previous studies we also repeated our analyses using the time interval approach based on biostratigraphy (e.g. Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; Benton et al., 2014) using each geological stage from the Middle Jurassic to the present. We report the results of these analyses in the Supplementary Materials (Fig S6 to S9).

Testing the effects of the K-Pg extinction on mammalian disparity

If the K-Pg extinction event had a significant effect on mammalian disparity, we should see a significant difference between disparity at the end of the Cretaceous and disparity at the start of the Paleogene. To test this, we performed *t*-tests to look for differences in disparity between the time subsamples of interest (e.g. as used in Anderson and Friedman, 2012; Zelditch et al., 2012; Smith et al., 2014). We compared the last time

subsample before the K-Pg boundary (70 Ma) to the first subsample of the Paleocene (65 Ma) for both the Mammaliaformes and Eutheria datasets and using both the gradual and punctuated evolutionary models. Even though one million year after the K-Pg event (66 to 65 Ma) seems to be a rather short geological time frame, effects on mammalian evolution have been detected as early as half a million year after K-Pg (Wilson, 2013). However, the effect of extinction on a group's evolution might not be detectable directly after the event due to delays in recovery (e.g. Chen and Benton, 2012, estimated that ecosystems only fully recovered 8-9 Ma after the Permo-Triassic mass extinction). Therefore, we also tested whether there was a significant difference in disparity between the end of the Cretaceous (70 Ma) and all subsamples from the Paleocene (65, 60 and 55 Ma). Additionally, some authors argue that the major diversification event in mammals took place during the Paleocene-Eocene Thermal Maximum (PETM; ~ 56 Ma; Bininda-Emonds et al. 2007 but see Meredith et al. 2011 and Stadler 2011) with the extinctions at K-Pg providing the “empty” ecological space required for this diversification to occur. We therefore extended our comparisons between the last subsample of the Cretaceous (70 Ma) up to the late Eocene (35 Ma) to check for a delayed effect of the K-Pg extinction potentially allowing morphological diversification after the PETM. Because these analyses involved multiple comparisons, we used Bonferroni corrections (Holm, 1979) to ensure our significant results were robust to Type I error rate inflation.

Finally, disparity may be higher in subsamples with more phylogenetic elements

simply because there are more taxa represented. To test whether this influenced our results, we repeated the *t*-tests using the rarefied Mammaliaformes and Eutheria disparities. In the Mammaliaformes, the minimum number of taxa in each subsample from 170 Ma to present was eight. In the Eutheria, the minimum number of taxa in each subsample was three, however, from 150 Ma until the present, the minimum number of taxa is eight. To make both datasets comparable, we used eight as a minimum number of taxa for the rarefied bootstrap measurements, therefore in the Eutheria we ignored the subsample between 170 and 150 Ma that only contains three taxa.

RESULTS

Disparity in the Mammaliaformes reaches a plateau during the Middle Triassic around 240 Ma, and fluctuates slightly around this during the rest of the Mesozoic and the Cenozoic (Fig 1 and Fig S5). The number of tips and nodes in each time subsample (a proxy for taxonomic richness), however, show a more idiosyncratic pattern with a steady increase until the Middle Jurassic around 170 Ma (Fig S5) followed by random fluctuations during the rest of the Mesozoic and the Cenozoic (Fig 1). Disparity in the Eutheria reaches a plateau at the end of the Jurassic around 150 Ma, whereas the number of tips and nodes increases up to the K-Pg boundary and then decreases throughout the Cenozoic (Fig 1). For both Mammaliaformes and Eutheria the same patterns in changes in disparity appear in the rarefied analyses (Fig 1 and S3). In both datasets the two evolutionary models (gradual or punctuated) also yield similar results (Fig 1).

We found no significant differences in disparity between the last subsample of the Cretaceous (70 Ma) and the first subsample of the Paleogene (65 Ma; Table 1), using both datasets and under both evolutionary models. We also found no significant differences in disparity between the last subsample of the Cretaceous (70 Ma) and any subsamples of the Paleocene and Eocene in Mammaliaformes under both evolutionary models and in Eutheria under a gradual evolutionary model (Table 1). However, in Eutheria under the punctuated evolutionary model, we found a small significant difference (after applying Bonferonni corrections) in disparity between the last subsample of the Cretaceous (70 Ma) and the subsamples at 45 Ma (an increase in disparity of 0.17; Table 1). However, this result is not significant in the rarefied analyses (Table S1). Otherwise the results of the rarefied analyses are the same as when using the complete datasets.

DISCUSSION

Previous authors have suggested that the K-Pg extinction event released mammals from ecological pressures such as competition and predation, allowing them to radiate into newly available ecological niches (Archibald, 2011; O'Leary et al., 2013; Lovegrove et al., 2014; Slater, 2013). However, we did not detect any significant changes in mammalian disparity before and after K-Pg in either Mammaliaformes or Eutheria, under a model of punctuated or gradual evolution. Additionally, we tested whether the absence of a detectable effect might be due to a lag effect, with the effect only becoming obvious

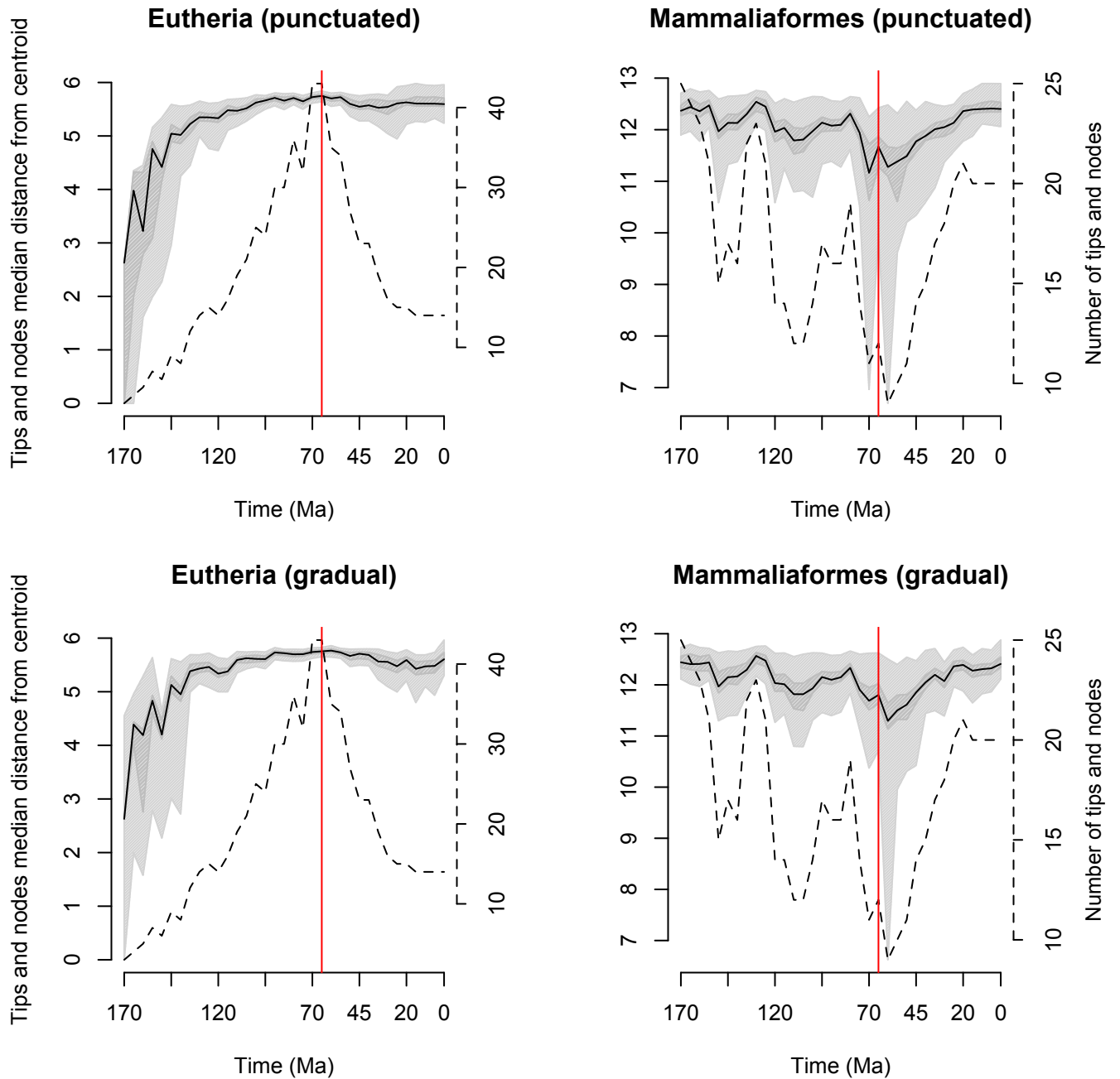


Figure 1: Disparity through time in Eutheria and Mammaliaformes calculated using a model of punctuated (upper panels) or gradual (lower panels) evolution. The x axis represents time in millions of years before the present (Ma). The y axis represents disparity, measured as the median distance between the centroid of the ordinated space and the tips/nodes in each time subsample. The solid black lines show the mean disparity estimated from 1000 bootstrapped pseudoreplicates and confidence intervals (CI) are represented by the grey polygons (50% CI in dark grey and 95% CI in light grey). The dashed line and the right hand axis represents the number of tips/nodes in each time slice. The red vertical line indicates the Cretaceous-Paleogene (K-Pg) boundary (66 Ma). Note that scale bars differ among panels.

later in the Paleocene. Even when accounting for such a lag effect, we did not detect any significant effect of the K-Pg extinction event on mammalian disparity. Our results imply that mammals did not diversify morphologically in response to the K-Pg extinction event. Instead, their diversification appears to have begun before the end of the Cretaceous (Fig 1, Table 1 and see Meredith et al., 2011; dos Reis et al., 2014; Close et al., 2015; Lee and Beck, 2015).

We did, however, detect a small, yet significant, increase in disparity during the Eocene (45 Ma) under a punctuated evolutionary model using the Eutheria dataset. This might be due to a long lag effect of ~ 21 Ma after K-Pg. Note however, that this is double the lag time observed in other mass extinctions (Chen and Benton, 2012). Therefore, it may be more likely to be attributed to a lag effect of the Palaeocene-Eocene Thermal Maximum (PETM; ~ 11 Ma afterwards; Bininda-Emonds et al., 2007). However, this significant increase in disparity is only detected at 45 Ma but not afterwards (which would be expected if the increase was due to an evolutionary radiation) and is not seen under the gradual evolution model. This indicates that it is more likely due to differences in the evolutionary models rather than an actual increase in disparity. The 45 Ma subsample samples the long branch (~ 50 Ma) leading to *Lepidictis* (33.9 to 33.3 Ma) that branches with its closest relative *Gypsonictops* (66.8 to 66 Ma) in the early Upper Cretaceous ~ 90 Ma (see Fig S2). Therefore, in this time-slice under the gradual evolution model, the data for *Lepidictis* is always sampled, but under the punctuated evolution model the algorithm can also randomly sample the data from

its ancestor in the early Upper Cretaceous (see methods). This may inflate differences compared to other slices. Incidentally, this increase can also be linked to the number of tips and nodes used in the comparison (43 versus 23 tips and nodes at respectively 70 and 45 Ma), because the increase is not significant in the rarefied analysis (see Fig S3 with only eight tips and nodes). Given these caveats we believe that no strong conclusions can be drawn from the increase in disparity during the Eocene.

Our results differ from a previous study that found an increase in disparity in North American Theria as soon as ~ 0.5 Ma after K-Pg (Wilson, 2013). These differences are likely to be related to several methodological differences between the present study and the previous one (Wilson, 2013). Firstly, Wilson (2013) only measures disparity at a regional scale (North America) and proposes that the observed increases in disparity are linked to the immigration of new species into the study localities. This strongly implies that disparity was higher on a global scale. Secondly, most of the debate on mammalian diversification around the K-Pg boundary seems to be linked to the conflicting signal between palaeontological and neontological data (Meredith et al. 2011 *vs.* O'Leary et al. 2013 but see dos Reis et al. 2014). Therefore, an effect of the K-Pg extinction event might be detectable only when using just palaeontological data. In this study, however, we use Total Evidence tip-dated trees based on both palaeontological and neontological data (Slater, 2013; Beck and Lee, 2014), which may account for the differences between our study and that of Wilson (2013) who used only fossil data.

Interestingly, however, our results also differ from Slater (2013), the source of

data for the Mammaliaformes dataset. Slater (2013) found support for a shift in the mode of body mass evolution (from an Ornstein–Uhlenbeck to a Brownian Motion model) directly after K-Pg suggesting a release in competition pressure or new niches becoming available for mammals in the early Paleocene. Our studies may show different results due to the difference between changes observed in one continuous life-history trait (body mass; Slater, 2013) versus changes in an aggregate of 446 discrete morphological traits (the cladistic characters) in the present analysis. Body mass and disparity might be decoupled in a similar way to taxonomic diversity and disparity (e.g. Slater et al., 2010; Ruta et al., 2013; Hopkins, 2013) because the latter does not rely on size but rather on discrete morphological features. It is not unlikely that mammalian disparity increased rapidly early in their evolutionary history and then remained constant (Fig 1; Close et al., 2015; Lee and Beck, 2015) while body mass variation continued to increase, especially after K-Pg (Slater, 2013). Note, however, that our methods did not investigate changes in body mass across the K-Pg boundary so they do not allow us to test this hypothesis. We remain confident in our results because we recovered the same pattern from two independent datasets (Slater, 2013; Beck and Lee, 2014).

There are several caveats to consider when interpreting our results. Firstly, both our datasets are limited taxonomically. They do not represent all known mammalian taxa, especially during the Neogene (23–2.58 Ma) where there are no fossil taxa in either dataset. Our study, however, focuses on changes in disparity around the K-Pg

boundary and not during the whole Cenozoic. Besides, this might not cause a serious underestimation of disparity, at least for the Mammaliaformes, because their diversity peaked during the late Cretaceous (Campanian; 72.1–83.6 Ma; Newham et al., 2014) and mammalian diversification rates declined throughout the Cenozoic (Raia et al., 2012). Therefore, an effect of the K-Pg boundary would be more likely to be detected during the Paleogene when mammalian diversity was highest, so we do not believe that increasing taxon sampling would greatly alter our conclusions.

Secondly, testing for significant changes in disparity through time is problematic. The disparity of each subsample can be dependent on disparity in the previous subsamples. For example, the tips and nodes used to estimate disparity are linked by common evolutionary history, therefore two tips or nodes sharing a close ancestor are more likely to have similar morphological features than more distantly related tips and nodes. Similarly, when looking at disparity through time, different subsamples are related by time, therefore, two subsamples closely together in time are more likely to have the same disparity value than more distant subsamples. Additionally, because disparity is a single value summarizing morphological disparity, its variance and mean were calculated by bootstrapping, thus the variances and means used in our *t*-tests are calculated from non-independent pseudoreplicates rather than true replicates. A second caveat arising from using bootstraps is that using a large number of pseudoreplicates is likely to inflate Type I error rates. Currently, however, this method is still widely used in disparity analyses for lack of a better alternative (e.g. Anderson

and Friedman, 2012; Zelditch et al., 2012; Smith et al., 2014).

Methodological improvements for measuring disparity through time

Our results may differ from previous studies because of our specific methodological choices. Throughout this paper, we propose several incremental changes to the classical ways of measuring disparity. Firstly we used all the axes of the cladisto-space, as opposed to previous studies that selected a subsample of the cladisto-space arguing that the m first axes usually contain most of the dataset's variance (e.g Brusatte et al., 2008a; Cisneros and Ruta, 2010; Prentice et al., 2011; Anderson and Friedman, 2012; Hughes et al., 2013; Benton et al., 2014). We argue that even if the last dimensions of the cladisto-space contain a trivial amount of variance, there is no statistical justification for excluding them. However, by doing so, we included dimensions of the cladisto-space with near zero variance and range (the last dimension's variance was 2×10^{-14} and 1.15×10^{-15} and the range was 7.31×10^{-7} and 3.33×10^{-7} for respectively the Mammaliaformes and Eutheria datasets). An alternative method avoids this problem by simply not ordinating the data and using the raw distance matrix (e.g. Benson and Druckenmiller, 2014; Close et al., 2015). However, in both this method and our method, the calculation of the products of ranges and variances is impossible.

Secondly, we used median distance between tips and nodes to centroid as a disparity metric, rather than the classical sums and products of ranges and variances (Wills et al., 1994). This metric is not affected by problems with using the last dimensions of the cladisto-space (see above). Also, it has several other advantages over

other metrics. For example, it measures directly the median spread of the taxa in the cladisto-space unlike the sum and products of ranges and variances that measure the size of the cladisto-space dimensions (Wills et al., 1994). Additionally, it comes with no statistical caveats unlike the sums or products of variances that should also include covariances between axes to correctly assess the exact size of the cladisto-space (even though the covariance term is usually close to 0 because of the eigen decomposition; Gower, 1966).

Finally, we used a time-slicing method instead of binning the data into time intervals (e.g in: Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; Hopkins, 2013; Benton et al., 2014; Benson and Druckenmiller, 2014) thus allowing us to avoid two caveats of using the time intervals approach. Because time intervals are often based on biostratigraphy, which is in turn based on notable differences in fossil fauna and flora, this method is likely to artificially emphasise disparity differences among time intervals. It is also possible to use arbitrary time bins of equal duration rather than biostratigraphy (Butler et al., 2012; Hopkins, 2013; Benson and Druckenmiller, 2014), but both approaches make the underlying assumption that disparity changes in a punctuated manner, i.e. changes occur only between time intervals. However, gradual evolution has been shown to be relatively common in the fossil record (Hunt, 2007; Hunt et al., 2015), so this assumption is unfounded. Our approach allowed us to fit different evolutionary models to our data - either assuming punctuated or gradual evolution. This is an improvement on previous approaches but could be improved

further by implementing other common but more complex models for example, a combined stasis and random walk (Hunt et al., 2015) or models based on morphological rates rather than just branch lengths.

Conclusion

Evidence for whether mammals diversified before or after the K-Pg boundary is mixed (Meredith et al., 2011; O’Leary et al., 2013; dos Reis et al., 2014; Beck and Lee, 2014), and appears to be related to the kind of data used (fossils or living species) and how the analyses were conducted. Using both fossil and living taxa, and investigating morphological disparity through time rather than taxonomic diversity, we find no direct effect of the K-Pg extinction event on the diversity of mammals. We therefore suggest that, contrary to popular belief, the extinction of many terrestrial vertebrates including the non-avian dinosaurs 66 million years ago, did not significantly affect the evolution of mammals throughout the Cenozoic.

DATA AVAILABILITY AND REPRODUCIBILITY

Data are available on Dryad or Figshare. Code for reproducing the analyses is available on GitHub (github.com/TGuillerme/SpatioTemporal_Disparity).

ACKNOWLEDGMENTS

Thanks to Graeme Lloyd, Andrew Jackson, Gavin Thomas, Graham Slater and Sive
Finlay.

FUNDING

This work was funded by a European Commission CORDIS Seventh Framework
Programme (FP7) Marie Curie CIG grant (proposal number: 321696).

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References

Anderson, P. S. and M. Friedman. 2012. Using cladistic characters to predict functional
variety: experiments using early gnathostomes. *Journal of Vertebrate Paleontology*
32:1254–1270.

Archibald, J. D. 2011. Extinction and radiation: how the fall of dinosaurs led to the rise
of mammals. JHU Press.

Arcila, D., R. A. Pyron, J. C. Tyler, G. Ort, and R. Betancur-R. 2015. An evaluation of
fossil tip-dating versus node-age calibrations in tetraodontiform fishes (teleostei:
Percomorphaceae). *Molecular Phylogenetics and Evolution* 82, Part A:131 – 145.

Beck, R. M. and M. S. Lee. 2014. Ancient dates or accelerated rates? Morphological
clocks and the antiquity of placental mammals. *Proceedings of the Royal Society B:*
Biological Sciences 281:1–10.

- 533 Benson, R. B. J. and P. S. Druckenmiller. 2014. Faunal turnover of marine tetrapods
534 during the Jurassic-Cretaceous transition. *Biological Reviews* 89:1–23.
- 535 Benton, M. J. 1985. Mass extinction among non-marine tetrapods. *Nature* 316:811–814.
- 536 Benton, M. J. 2015. Exploring macroevolution using modern and fossil data.
537 *Proceedings of the Royal Society of London B: Biological Sciences* 282.
- 538 Benton, M. J., J. Forth, and M. C. Langer. 2014. Models for the rise of the dinosaurs.
539 *Current biology* : CB 24:R87–R95.
- 540 Benton, M. J. and R. J. Twitchett. 2003. How to kill (almost) all life: the end-permian
541 extinction event. *Trends in Ecology and Evolution* 18:358 – 365.
- 542 Bininda-Emonds, O. R., M. Cardillo, K. E. Jones, R. D. MacPhee, R. M. Beck, R. Grenyer,
543 S. A. Price, R. A. Vos, J. L. Gittleman, and A. Purvis. 2007. The delayed rise of
544 present-day mammals. *Nature* 446:507–512.
- 545 Brusatte, S., R. J. Butler, A. Prieto-Márquez, and M. A. Norell. 2012. Dinosaur
546 morphological diversity and the end-Cretaceous extinction. *Nature Communications*
547 3:804–804.
- 548 Brusatte, S. L., M. J. Benton, M. Ruta, and G. T. Lloyd. 2008a. The first 50 Myr of
549 dinosaur evolution: macroevolutionary pattern and morphological disparity. *Biology*
550 *Letters* 4:733–736.

- 551 Brusatte, S. L., M. J. Benton, M. Ruta, and G. T. Lloyd. 2008b. Superiority, competition,
552 and opportunism in the evolutionary radiation of dinosaurs. *Science* 321:1485–1488.
- 553 Brusatte, S. L., R. J. Butler, P. M. Barrett, M. T. Carrano, D. C. Evans, G. T. Lloyd, P. D.
554 Mannion, M. A. Norell, D. J. Peppe, P. Upchurch, and T. E. Williamson. 2015. The
555 extinction of the dinosaurs. *Biological Reviews* 90:628–642.
- 556 Butler, R. J., S. L. Brusatte, B. Andres, and R. B. J. Benson. 2012. How do geological
557 sampling biases affect studies of morphological evolution in deep time? a case study
558 of pterosaur (reptilia: Archosauria) disparity. *Evolution* 66:147–162.
- 559 Cailliez, F. 1983. The analytical solution of the additive constant problem.
560 *Psychometrika* 48:305–308.
- 561 Chen, Z.-Q. and M. J. Benton. 2012. The timing and pattern of biotic recovery following
562 the end-permian mass extinction. *Nature Geoscience* 5:375–383.
- 563 Ciampaglio, C. N. 2004. Measuring changes in articulate brachiopod morphology
564 before and after the permian mass extinction event: do developmental constraints
565 limit morphological innovation? *Evolution & Development* 6:260–274.
- 566 Ciampaglio, C. N., M. Kemp, and D. W. McShea. 2001. Detecting changes in
567 morphospace occupation patterns in the fossil record: characterization and analysis
568 of measures of disparity. *Paleobiology* 27:695–715.
- 569 Cisneros, J. C. and M. Ruta. 2010. Morphological diversity and biogeography of

procolophonids (amniota: Parareptilia). *Journal of Systematic Palaeontology*
8:607–625.

Close, R., M. Friedman, G. Lloyd, and R. Benson. 2015. Evidence for a mid-jurassic
adaptive radiation in mammals. *Current Biology* .

Coxall, H. K., S. D'Hondt, and J. C. Zachos. 2006. Pelagic evolution and environmental
recovery after the cretaceous-paleogene mass extinction. *Geology* 34:297–300.

dos Reis, M., P. C. J. Donoghue, and Z. Yang. 2014. Neither phylogenomic nor
palaeontological data support a palaeogene origin of placental mammals. *Biology*
Letters 10.

dos Reis, M., J. Inoue, M. Hasegawa, R. J. Asher, P. C. J. Donoghue, and Z. Yang. 2012.
Phylogenomic datasets provide both precision and accuracy in estimating the
timescale of placental mammal phylogeny. *Proceedings of the Royal Society of*
London B: Biological Sciences .

Douady, C. J. and E. J. Douzery. 2003. Molecular estimation of eulipotyphlan divergence
times and the evolution of insectivora. *Molecular Phylogenetics and Evolution* 28:285
– 296.

Erwin, D. H. 1998. The end and the beginning: recoveries from mass extinctions. *Trends*
in Ecology and Evolution 13:344 – 349.

Erwin, D. H. 2007. Disparity: Morphological pattern and developmental context.
Palaeontology 50:57–73.

- 590 Foote, M. 1994. Morphological disparity in ordovician-devonian crinoids and the early
591 saturation of morphological space. *Paleobiology* 20:320–344.
- 592 Foote, M. 1996. Ecological controls on the evolutionary recovery of post-paleozoic
593 crinoids. *Science* 274:1492–1495.
- 594 Foth, C., S. Brusatte, and R. Butler. 2012. Do different disparity proxies converge on a
595 common signal? insights from the cranial morphometrics and evolutionary history of
596 pterosauria (diapsida: Archosauria). *Journal of evolutionary biology* 25:904–915.
- 597 Friedman, M. 2010. Explosive morphological diversification of spiny-finned teleost
598 fishes in the aftermath of the end-Cretaceous extinction. *Proceedings of the Royal
599 Society B: Biological Sciences* 277:1675–1683.
- 600 Garland, J., Theodore and A. R. Ives. 2000. Using the past to predict the present:
601 Confidence intervals for regression equations in phylogenetic comparative methods.
602 *The American Naturalist* 155:346–364.
- 603 Glor, R. E. 2010. Phylogenetic insights on adaptive radiation. *Annual Review of
604 Ecology, Evolution, and Systematics* 41:251–270.
- 605 Goswami, A., G. V. Prasad, P. Upchurch, D. M. Boyer, E. R. Seiffert, O. Verma,
606 E. Gheerbrant, and J. J. Flynn. 2011. A radiation of arboreal basal eutherian mammals
607 beginning in the late cretaceous of india. *Proceedings of the National Academy of
608 Sciences of the United States of America* 108:16333–16338.

609 Gould, S. J. and N. Eldredge. 1977. Punctuated equilibria: The tempo and mode of
610 evolution reconsidered. *Paleobiology* 3:pp. 115–151.

611 Gower, J. C. 1966. Some distance properties of latent root and vector methods used in
612 multivariate analysis. *Biometrika* 53:325–338.

613 Gower, J. C. 1971. A general coefficient of similarity and some of its properties.
614 *Biometrics* 27:pp. 857–871.

615 Guillerme, T. and N. Cooper. 2016. Effects of missing data on topological inference
616 using a total evidence approach. *Molecular Phylogenetics and Evolution* 94, Part
617 A:146 – 158.

618 Hetherington, A. J., E. Sherratt, M. Ruta, M. Wilkinson, B. Deline, and P. C. Donoghue.
619 2015. Do cladistic and morphometric data capture common patterns of
620 morphological disparity? *Palaeontology* 58:393–399.

621 Holm, S. 1979. A simple sequentially rejective multiple test procedure. *Scandinavian*
622 *journal of statistics* Pages 65–70.

623 Hopkins, M. 2013. Decoupling of taxonomic diversity and morphological disparity
624 during decline of the cambrian trilobite family pterocephaliidae. *Journal of*
625 *Evolutionary Biology* 26:1665–1676.

626 Huang, S., K. Roy, and D. Jablonski. 2015. Origins, bottlenecks, and present-day
627 diversity: Patterns of morphospace occupation in marine bivalves. *Evolution* .

628 Hughes, M., S. Gerber, and M. A. Wills. 2013. Clades reach highest morphological
 629 disparity early in their evolution. *Proceedings of the National Academy of Sciences*
 630 110:13875–13879.

631 Hunt, G. 2007. The relative importance of directional change, random walks, and stasis
 632 in the evolution of fossil lineages. *Proceedings of the National Academy of Sciences*
 633 104:18404–18408.

634 Hunt, G., M. J. Hopkins, and S. Lidgard. 2015. Simple versus complex models of trait
 635 evolution and stasis as a response to environmental change. *Proceedings of the*
 636 *National Academy of Sciences* 112:4885–4890.

637 Korn, D., M. J. Hopkins, and S. A. Walton. 2013. Extinction spacea method for the
 638 quantification and classification of changes in morphospace across extinction
 639 boundaries. *Evolution* 67:2795–2810.

640 Lee, M. and R. Beck. 2015. Mammalian evolution: A jurassic spark. *Current Biology*
 641 25:R759 – R761.

642 Liow, L. H., T. Reitan, and P. G. Harnik. 2015. Ecological interactions on
 643 macroevolutionary time scales: clams and brachiopods are more than ships that pass
 644 in the night. *Ecology Letters* Pages n/a–n/a.

645 Lloyd, G. T. 2015. Claddis: Measuring Morphological Diversity and Evolutionary
 646 Tempo. R package version 0.1.

- 647 Losos, J. B. 2010. Adaptive radiation, ecological opportunity, and evolutionary
648 determinism. *The American Naturalist* 175:pp. 623–639.
- 649 Lovegrove, B. G., K. D. Lobban, and D. L. Levesque. 2014. Mammal survival at the
650 cretaceous–palaeogene boundary: metabolic homeostasis in prolonged tropical
651 hibernation in tenrecs. *Proceedings of the Royal Society of London B: Biological*
652 *Sciences* 281.
- 653 Meredith, R., J. Janečka, J. Gatesy, O. Ryder, C. Fisher, E. Teeling, A. Goodbla, E. Eizirik,
654 T. L. Simão, T. Stadler, D. Rabosky, R. Honeycutt, J. Flynn, C. Ingram, C. Steiner,
655 T. Williams, T. Robinson, B. Angela, M. Westerman, N. Ayoub, M. Springer, and
656 W. Murphy. 2011. Impacts of the Cretaceous terrestrial revolution and KPg extinction
657 on mammal diversification. *Science* 334:521–524.
- 658 Newham, E., R. Benson, P. Upchurch, and A. Goswami. 2014. Mesozoic mammaliaform
659 diversity: The effect of sampling corrections on reconstructions of evolutionary
660 dynamics. *Palaeogeography, Palaeoclimatology, Palaeoecology* 412:32 – 44.
- 661 O’Leary, M. A., J. I. Bloch, J. J. Flynn, T. J. Gaudin, A. Giallombardo, N. P. Giannini, S. L.
662 Goldberg, B. P. Kraatz, Z.-X. Luo, J. Meng, X. Ni, M. J. Novacek, F. A. Perini, Z. S.
663 Randall, G. W. Rougier, E. J. Sargis, M. T. Silcox, N. B. Simmons, M. Spaulding, P. M.
664 Velazco, M. Weksler, J. R. Wible, and A. L. Cirranello. 2013. The placental mammal
665 ancestor and the postK-Pg radiation of placentals. *Science* 339:662–667.

666 Olson, M. E. and A. Arroyo-Santos. 2009. Thinking in continua: beyond the adaptive
 667 radiation metaphor. *BioEssays* 31:1337–1346.

668 OLeary, M. A., J. I. Bloch, J. J. Flynn, T. J. Gaudin, A. Giallombardo, N. P. Giannini, S. L.
 669 Goldberg, B. P. Kraatz, Z.-X. Luo, J. Meng, X. Ni, M. J. Novacek, F. A. Perini,
 670 Z. Randall, G. W. Rougier, E. J. Sargis, M. T. Silcox, N. B. Simmons, M. Spaulding,
 671 P. M. Velazco, M. Weksler, J. R. Wible, and A. L. Cirranello. 2013. Response to
 672 comment on the placental mammal ancestor and the postk-pg radiation of placentals.
 673 *Science* 341:613.

674 Paradis, E., J. Claude, and K. Strimmer. 2004. APE: analyses of phylogenetics and
 675 evolution in R language. *Bioinformatics* 20:289–290.

676 Payne, J. L., N. A. Heim, M. L. Knope, and C. R. McClain. 2014. Metabolic dominance
 677 of bivalves predates brachiopod diversity decline by more than 150 million years.
 678 *Proceedings of the Royal Society B: Biological Sciences* 281.

679 Pearman, P. B., A. Guisan, O. Broennimann, and C. F. Randin. 2008. Niche dynamics in
 680 space and time. *Trends in Ecology and Evolution* 23:149 – 158.

681 Prentice, K. C., P. Ruta, and M. J. Benton. 2011. Evolution of morphological disparity in
 682 pterosaurs. *Journal of Systematic Palaeontology* 9:337–353.

683 R Core Team. 2015. R: a language and environment for statistical computing. R
 684 Foundation for Statistical Computing Vienna, Austria.

- 685 Raia, P., F. Carotenuto, F. Passaro, P. Piras, D. Fulgione, L. Werdelin, J. Saarinen, and
686 M. Fortelius. 2012. Rapid action in the palaeogene, the relationship between
687 phenotypic and taxonomic diversification in coenozoic mammals. Proceedings of the
688 Royal Society of London B: Biological Sciences 280.
- 689 Raup, D. M. 1979. Size of the permo-triassic bottleneck and its evolutionary
690 implications. Science 206:217–218.
- 691 Renne, P. R., A. L. Deino, F. J. Hilgen, K. F. Kuiper, D. F. Mark, W. S. Mitchell, L. E.
692 Morgan, R. Mundil, and J. Smit. 2013. Time scales of critical events around the
693 Cretaceous-Paleogene boundary. Science 339:684–687.
- 694 Revell, L. J. 2012. phytools: An R package for phylogenetic comparative biology (and
695 other things). Methods in Ecology and Evolution 3:217–223.
- 696 Ronquist, F., S. Klopstein, L. Vilhelmsen, S. Schulmeister, D. Murray, and A. Rasnitsyn.
697 2012. A total-evidence approach to dating with fossils, applied to the early radiation
698 of the Hymenoptera. Systematic Biology 61:973–999.
- 699 Ruta, M., K. D. Angielczyk, J. Fröbisch, and M. J. Benton. 2013. Decoupling of
700 morphological disparity and taxic diversity during the adaptive radiation of
701 anomodont therapsids. Proceedings of the Royal Society of London B: Biological
702 Sciences 280.
- 703 Slater, G. J. 2013. Phylogenetic evidence for a shift in the mode of mammalian body size

evolution at the cretaceous-palaeogene boundary. *Methods in Ecology and Evolution*
4:734–744.

Slater, G. J., S. A. Price, F. Santini, and M. E. Alfaro. 2010. Diversity versus disparity and
the radiation of modern cetaceans. *Proceedings of the Royal Society of London B:
Biological Sciences* .

Smith, A. J., M. V. Rosario, T. P. Eiting, and E. R. Dumont. 2014. Joined at the hip:
Linked characters and the problem of missing data in studies of disparity. *Evolution*
68:2386–2400.

Springer, M. S., R. W. Meredith, E. C. Teeling, and W. J. Murphy. 2013. Technical
comment on the placental mammal ancestor and the postk-pg radiation of placentals.
Science 341:613.

Stadler, T. 2011. Mammalian phylogeny reveals recent diversification rate shifts.
Proceedings of the National Academy of Sciences 108:6187–6192.

Thorne, P. M., M. Ruta, and M. J. Benton. 2011. Resetting the evolution of marine
reptiles at the Triassic-Jurassic boundary. *Proceedings of the National Academy of
Sciences* 108:8339–8344.

Toljagic, O. and R. J. Butler. 2013. Triassic-Jurassic mass extinction as trigger for the
mesozoic radiation of crocodylomorphs. *Biology Letters* 9.

Torgerson, W. S. 1965. Multidimensional scaling of similarity. *Psychometrika*
30:379–393.

- 724 Wesley-Hunt, G. D. 2005. The morphological diversification of carnivores in north
725 america. *Paleobiology* 31:35–55.
- 726 Wills, M. A., D. E. G. Briggs, and R. A. Fortey. 1994. Disparity as an evolutionary index:
727 A comparison of cambrian and recent arthropods. *Paleobiology* 20:93–130.
- 728 Wilson, G. P. 2013. Mammals across the k/pg boundary in northeastern montana, u.s.a.:
729 dental morphology and body-size patterns reveal extinction selectivity and
730 immigrant-fueled ecospace filling. *Paleobiology* 39:429–469.
- 731 Wood, H. M., N. J. Matzke, R. G. Gillespie, and C. E. Griswold. 2013. Treating fossils as
732 terminal taxa in divergence time estimation reveals ancient vicariance patterns in the
733 palpimanoid spiders. *Systematic Biology* 62:264–284.
- 734 Yang, Z., S. Kumar, and M. Nei. 1996. A new method of inference of ancestral
735 nucleotide and amino acid sequences. *Genetics* 141:1641–50.
- 736 Zelditch, M. L., D. L. Swiderski, and H. D. Sheets. 2012. Geometric morphometrics for
737 biologists: a primer. Academic Press.

Table 1: Results of *t*-tests comparing disparity at the last subsample of the Cretaceous (70 Ma) to subsamples of the Paleocene and Eocene, under both gradual and punctuated evolutionary models, in Mammaliaformes and Eutheria. Difference = mean difference in disparity between the two subsamples being compared; df = degrees of freedom; p value = original p value prior to Bonferonni correction. Significant differences (after applying Bonferonni corrections for multiple comparisons) are highlighted in bold.

Subsamples compared	Gradual evolution model				Punctuated evolution model			
	difference	df	t	p value	difference	df	t	p value
Mammaliaformes								
70 vs. 65	-0.420	21	-0.808	0.428	-0.030	21	-0.058	0.954
70 vs. 60	0.030	18	0.046	0.964	0.210	18	0.379	0.709
70 vs. 55	0.010	19	0.021	0.983	0.110	19	0.225	0.824
70 vs. 50	-0.260	20	-0.456	0.653	0.030	20	0.060	0.953
70 vs. 45	-0.430	23	-0.869	0.394	0.060	23	0.132	0.896
70 vs. 40	-0.620	24	-1.388	0.178	-0.410	24	-1.031	0.313
70 vs. 35	-0.730	26	-1.742	0.093	-0.340	26	-0.861	0.397
Eutheria								
70 vs. 65	-0.020	84	-0.503	0.616	0.010	84	0.288	0.774
70 vs. 60	0.030	76	0.617	0.539	0.080	76	1.693	0.095
70 vs. 55	0.030	75	0.519	0.605	0.030	75	0.699	0.486
70 vs. 50	0.130	68	2.101	0.039 ¹	0.080	68	1.458	0.149
70 vs. 45	0.190	64	2.679	0.009 ¹	0.170	64	2.730	0.006²
70 vs. 40	0.160	64	2.249	0.028 ¹	0.130	64	2.084	0.041 ¹
70 vs. 35	0.190	60	2.358	0.022 ¹	0.120	60	1.893	0.063

¹p value is non-significant after applying Bonferonni correction; ²p value is **0.048** after applying

Bonferonni⁴⁰ correction.