RH: No effect of the K-Pg event on mammal disparity.

# Mammalian morphological diversity does not increase in response to the Cretaceous-Paleogene mass extinction and the extinction of the (non-avian) dinosaurs. Thomas

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1 Abstract

Popular science accounts state that after the extinction of the non-avian dinosaurs at the Cretaceous-Paleogene (K-Pg) boundary 66 million years ago, mammals rapidly diversified to fill their empty ecological niches. However, evidence for this is mixed. Palaeontological analyses suggest that mammals radiated in response to the K-Pg extinction event, whereas neontological analyses suggest that mammals began to radiate before K-Pg and were not greatly affected by it. Here we aim to end this debate by looking at fossil and living taxa simultaneously.

We investigated the effect of the K-Pg extinction event on mammalian morphological diversity (disparity) using two Total Evidence tip-dated phylogenies of Mammaliaformes and Eutheria, containing both fossil and living taxa. Using a novel, continuous time-slicing method for measuring changes in disparity-through-time, we found no significant change in disparity before and after the K-Pg boundary, under either a gradual or punctuated model of evolution. This implies that the extinctions at the end of the Cretaceous did not affect mammalian morphological evolution. Our findings contradict the popular theory that the non-avian dinosaurs and other Mesozoic tetrapods were restricting mammalian evolution, and that their extinction liberated ecological niches for mammals to evolve into.

(Keywords: disparity, punctuated equilibrium, gradual evolution, time slicing, K-Pg)

### Introduction

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Throughout history, life on Earth has suffered a series of mass extinction events resulting in drastic declines in global biodiversity (e.g. Raup, 1979; Benton and Twitchett, 2003; Renne et al., 2013; Brusatte et al., 2015). The long-term effects of mass extinctions, however, are more varied (Erwin, 1998), and include species richness increases in some clades (Friedman, 2010) and declines in others (Benton, 1985), changes in morphological diversity (Ciampaglio et al., 2001; Ciampaglio, 2004; Korn et al., 2013) and shifts in ecological dominance (e.g. Brusatte et al., 2008b; Toljagic and Butler, 2013; Benson and Druckenmiller, 2014). These shifts are characterized by the decline of one clade that is replaced by a different unrelated clade with a similar 31 ecological role (e.g. Brachiopoda and Bivalvia at the end Permian extinction; Liow et al. 2015 but see Payne et al. 2014). Shifts in ecological dominance are of particular interest 33 because they are a fairly common pattern observed in the fossil record (e.g. Foraminifera; Coxall et al. 2006 ; Ichtyosauria; Thorne et al. 2011; Plesiosauria; Benson 35 and Druckenmiller 2014) and are often linked to major macroevolutionary processes such as adaptive (Losos, 2010) or competitive (Brusatte et al., 2008b) radiations. 37 One classical example of a shift in ecological dominance is at the 38 Cretaceous-Palaeogene (K-Pg) mass extinction 66 million years ago (Renne et al., 2013), 39 where many terrestrial vertebrates (including the dominant non-avian dinosaur group; Archibald 2011; Renne et al. 2013; Brusatte et al. 2015) went extinct, allowing placental mammals to dominate the fauna (Archibald, 2011; Lovegrove et al., 2014). Some

authors suggest this reflects placental mammals filling the "empty" niches left after the K-Pg extinction event (Archibald, 2011; O'Leary et al., 2013), others suggest it reflects a release from predation and/or competition (Slater, 2013; Lovegrove et al., 2014). However, evidence for the diversification of placental mammals being driven by the K-Pg extinction event is mixed. Thorough analysis of the fossil record (e.g. Goswami et al., 2011; O'Leary et al., 2013) supports the idea that placental mammals diversified after the K-Pg extinction event as there are no undebated placental mammal fossils before it and many afterwards (Archibald, 2011; Goswami et al., 2011; Slater, 2013; O'Leary et al., 2013; Wilson, 2013; Brusatte et al., 2015). Conversely, evidence from 51 molecular data suggests that the diversification of placental mammals started prior to the K-Pg extinction event without being drastically affected by it (e.g. Douady and Douzery, 2003; Bininda-Emonds et al., 2007; Meredith et al., 2011; Stadler, 2011). Therefore, whether the diversification of placental mammals began before the K-Pg 55 extinction event, or in response to the extinctions at K-Pg, is a matter of great debate (dos Reis et al., 2012; O'Leary et al., 2013; Springer et al., 2013; OLeary et al., 2013; dos 57 Reis et al., 2014). 58 There are two main reasons why there is still debate about the timing of the 59 diversification of placental mammals. Firstly, palaeontological and neontological data show different patterns; palaeontological data generally suggest that placental

suggest that K-Pg extinction event had little to no effect on mammalian diversification

mammals diversified after K-Pg (e.g. O'Leary et al., 2013), whereas neontological data

(Bininda-Emonds et al., 2007; Meredith et al., 2011; Stadler, 2011). We can solve this issue by using both palaeontological and neontological data in our analyses. The Total Evidence method allows us to use cladistic data for both living and fossil taxa, along with molecular data for living taxa, to build phylogenies (Ronquist et al., 2012). This method can also be combined with the tip-dating method (Ronquist et al., 2012; Wood et al., 2013) to get more accurate estimates of diversification times for both fossil and living species (but see Arcila et al., 2015). Here we use two recent Total Evidence tip-dated phylogenies of mammals (Slater, 2013; Beck and Lee, 2014) to investigate palaeontological and neontological taxa simultaneously.

A second issue is that diversity can be defined in many different ways. In many
studies it is measured as taxonomic diversity or species richness (Stadler, 2011;

Meredith et al., 2011; O'Leary et al., 2013), but often the more interesting aspect of
diversity is related to the ecological niches the species occupy (Wesley-Hunt, 2005;

Brusatte et al., 2008b; Toljagic and Butler, 2013), particularly if we want to make
hypotheses about macroevolutionary processes (Pearman et al., 2008; Olson and
Arroyo-Santos, 2009; Losos, 2010; Glor, 2010; Benton, 2015). Sometimes taxonomic
diversity is used as a proxy for other kinds of diversity, however, species richness can
be decoupled from morphological diversity (e.g. Slater et al., 2010; Ruta et al., 2013;
Hopkins, 2013), so it may not be the best proxy for ecological diversity. We can instead
use morphological diversity, also known as disparity (e.g. Wills et al., 1994; Erwin, 2007;
Hughes et al., 2013), as a way to quantify changes in mammalian morphology that

should relate to the ecology of the species. However some methods for measuring disparity are outdated and make inappropriate assumptions. Many methods for quantifying changes in morphological diversity were proposed > 20 years ago (Foote, 1994; Wills et al., 1994) and are sometimes used without modifications (e.g., Brusatte et al., 2008a,b; Cisneros and Ruta, 2010; Thorne et al., 2011; Prentice et al., 2011; Brusatte et al., 2012; Toljagic and Butler, 2013; Ruta et al., 2013; Benton et al., 2014; Benson and Druckenmiller, 2014). Additionally, previous methods are based on an underlying 91 assumption that changes in disparity occur by punctuated evolution (e.g. Wesley-Hunt, 2005) which is not always the case (Hunt et al., 2015). Finally, most studies of disparity through time use unequal time units based on biostratigraphy (Brusatte et al., 2008b, 2012; Toljagic and Butler, 2013). This can be tautological as biostratigraphy is already based on changes in fossil assemblages and morphology through time. To deal with these issues, we propose an updated approach to test whether mammals diversified in 97 response to the K-Pg event, using morphological disparity, measured as cladistic disparity (see Methods), as our proxy for diversity. 99

Here we measure the disparity of living and fossil mammals before or after

K-Pg, using data taken from two previously published studies (Slater, 2013; Beck and

Lee, 2014). Using a novel time-slicing approach, we produce fine-grained estimates of

disparity through time under two different models of morphological character

evolution (either gradual or punctuated). We also test whether mammals display

significant changes in disparity between the end of the Cretaceous and throughout the

106 Cenozoic.

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Until now, this question has only been investigated using data from North 107 American Therian mammals (excluding Monotremata) and without formally testing the effect of the K-Pg extinction event (Wilson, 2013). To our knowledge, this study is the first to approach the debate about the effects of the K-Pg extinction event on 110 mammalian evolution using Total Evidence phylogenies and by calculating disparity through time in a continuous way. We find no significant changes in mammalian 112 disparity between the end of the Cretaceous and any time during the Paleocene. These 113 results suggest that the extinction of non-avian dinosaurs and other terrestrial 114 vertebrate clades at the end of the Cretaceous did not affect mammalian morphological 115 evolution. 116

Methods

# Cladistic data and phylogenies

We used the cladistic morphological matrices and the Total Evidence tip-dated trees

(Ronquist et al., 2012) from Slater (2013, 103 taxa with 446 morphological characters;)

and Beck and Lee (2014, 102 taxa with 421 morphological characters). We chose these

two datasets because they have a similar number of taxa and morphological characters.

Slater (2013) ranges from 310 million years ago (Ma; Late Carboniferous) to the present

and focuses on the clade Mammaliaformes at the family-level and is called hereafter the

Mammaliaformes dataset. Beck and Lee (2014) ranges from 170 Ma (Middle Jurassic) to the present and focuses on Eutheria at the genus-level and is called hereafter the 126 Eutheria dataset. We used the first and last occurrences reported in Slater (2013) and Beck and Lee (2014) as the temporal range of each taxon in our analysis. Both phylogenies are illustrated in the supplementary material (see Fig S1 and S2 @@@). 129 Both trees contain few taxa compared to the overall species richness of living and fossil 130 mammals (Bininda-Emonds et al., 2007; Archibald, 2011). This is because Total 131 Evidence trees need a lot of data, particularly morphological data for living taxa that 132 can be hard to locate (Guillerme and Cooper, In press). Therefore, most Total Evidence 133 studies to date contain one or two orders of magnitude fewer taxa than phylogenies 134 based solely on molecular data (e.g. thousands of taxa in Bininda-Emonds et al. 2007; 135 Meredith et al. 2011 vs. hundreds in Ronquist et al. 2012; Slater 2013; Wood et al. 2013; 136 Beck and Lee 2014).

# Estimating ancestral character states

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For both datasets we used the re-rooting method (Yang et al., 1996; Garland and Ives,
2000) to get Maximum Likelihood estimates of the ancestral states for each character at
every node in the tree, using the rerootingMethod function from the R package
phytools version 0.4-45 (Revell, 2012; R Core Team, 2015). Where there was missing
character data for a taxon we followed the method of Lloyd (2015) and treated missing
data as any possible observed state for each character. For example, if a character had
two observed states (0 and 1) across all taxa, we attributed the multi-state "0&1" value

to the taxon with missing data, representing an equal probability of being either 0 or 1.

This allows the ancestral node of a taxon with missing data to be estimated with no

assumptions other than that the taxon has one of the observed character states. To

prevent poor ancestral state reconstructions from biasing our results, especially when a

lot of error is associated with the reconstruction, we only included ancestral state

reconstructions with a scaled Likelihood  $\geq 0.95$ . Ancestral state reconstructions with

scaled Likelihoods below this threshold were replaced by missing data ("?").

### Building the cladisto-space

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To explore variations in mammalian disparity through time (defined here as the 154 variation in morphologies through time), we used a cladisto-space approach (e.g. Foote, 155 1994, 1996; Wesley-Hunt, 2005; Brusatte et al., 2008b; Friedman, 2010; Toljagic and 156 Butler, 2013; Hughes et al., 2013). This approach is similar to constructing a 157 morphospace based on continuous morphological data (e.g. Friedman, 2010), except a 158 cladisto-space is an approximation of the morphospace based on cladistic data (i.e. the discrete morphological characters used to build a phylogenetic tree). Mathematically, a cladisto-space is an *n* dimensional object that summarizes the cladistic distances between the taxa present in a cladistic matrix (see details below). Although empirically 162 inter-taxon distances are the same in a morphospace or a cladisto-space (Foth et al., 2012; Hetherington et al., 2015), we prefer the term cladisto-space to make it clear that this space is estimated using cladistic data and not morphometric data and because 165 both objects have slightly different properties. For example, because of its inherent

combinatory properties, a cladisto-space is a finite theoretical object limited by the product of the number of character states, whereas a morphospace is an infinite theoretical object. Thus a cladisto-space will be overloaded if the number of taxa is higher than the product of the number of character states, although this is rarely an issue with empirical data (our cladisto-spaces have maximal capacities of  $1.9 \times 10^{181}$  taxa for the Mammaliaformes dataset, i.e. 101 orders of magnitude more taxa than the number of particles in the universe; and  $4.5 \times 10^{159}$  taxa for the Eutheria dataset).

To estimate the cladisto-spaces for each of our datasets we first constructed 174 pairwise distance matrices of length k, where k is the total number of tips and nodes in 175 the datasets. For each dataset separately, we calculated the  $k \times k$  distances using the 176 Gower distance (Gower, 1971), i.e. the Euclidean distance between two taxa divided by 177 the number of shared characters. This allows us to correct for distances between two 178 taxa that share many characters and could be closer to each other than to taxa with 179 fewer characters in common (i.e. because some pairs of taxa share more characters in 180 common than others, they are more likely to be similar). For cladistic matrices, using 181 this corrected distance is preferable to the raw Euclidean distance because of its ability 182 to deal with discrete or/and ordinated characters as well as with missing data 183 (Anderson and Friedman, 2012). However, the Gower distance cannot calculate distances when taxa have no overlapping data. Therefore, we used the 185 TrimMorphDistMatrix function from the Claddis R package (Lloyd, 2015) to remove pairs of taxa with no cladistic characters in common. This led to us removing 11 taxa

from the Mammaliaformes dataset but none from the Eutheria dataset.

After calculating our distance matrices we transformed them using classical 189 multidimensional scaling (MDS; Torgerson, 1965; Gower, 1966; Cailliez, 1983). This method (also referred to as PCO; e.g. Brusatte et al. 2015; or PCoA; e.g. Paradis et al. 2004) is an eigen decomposition of the distance matrix. Because we used Gower 192 distances instead of raw Euclidean distances, negative eigenvalues can be calculated. To avoid this problem, we first transformed the distance matrices by applying the Cailliez 194 correction (Cailliez, 1983) which adds a constant  $c^*$  to the values in a distance matrix 195 (apart from the diagonal) so that all the Gower distances become Euclidean 196  $(d_{Gower} + c^* = d_{Euclidean};$  Cailliez 1983). We were then able to extract n eigenvectors for 197 each matrix (representing the *n* dimensions of the cladisto-space) where *n* is equal to 198 k-2, i.e. the number of taxa in the matrix (k) minus the last two eigenvectors that are 199 always null after applying the Cailliez correction. Contrary to previous studies (e.g. 200 Brusatte et al., 2008a; Cisneros and Ruta, 2010; Prentice et al., 2011; Anderson and 201 Friedman, 2012; Hughes et al., 2013; Benton et al., 2014), we use all n dimensions of our 202 cladisto-spaces and not a subsample representing the majority of the variance in the 203 distance matrix (e.g. selecting only *m* dimensions that represent up to 90% of the 204 variance in the distance matrix; Brusatte et al. 2008b; Toljagic and Butler 2013). 205 Note that our cladisto-spaces represent an ordination of all possible mammalian 206 morphologies coded in each study through time. It is unlikely that all morphologies will co-occur at each time point, therefore, the disparity of the whole cladisto-space is

expected to be greater than the disparity at any specific point in time.

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### Calculating disparity

Disparity can be estimated in many different ways (e.g. Wills et al., 1994; Ciampaglio, 211 2004; Thorne et al., 2011; Hopkins, 2013; Huang et al., 2015), however most studies 212 estimate disparity using four metrics: the sum and products of ranges and variances, 213 each of which gives a slightly different estimate of how the data fits within the 214 cladisto-space (Foote, 1994; Wills et al., 1994; Brusatte et al., 2008a,b; Cisneros and Ruta, 215 2010; Thorne et al., 2011; Prentice et al., 2011; Brusatte et al., 2012; Toljagic and Butler, 216 2013; Ruta et al., 2013; Benton et al., 2014; Benson and Druckenmiller, 2014). Nonetheless, these methods suffer several methodological caveats. First, the range metrics are affected by the uneven sampling of the fossil record (Butler et al., 2012) Second, because we include all *n* dimensions in the analysis (see above), the products of ranges and variances will tend towards zero since the scores of the last dimension are 221 usually really close to zero themselves. These features make using the sum and 222 products of ranges and variances unfeasible in our study. Instead, we use a different 223 metric that comes with no statistical assumptions for measuring the dispersion of the 224 data in the cladisto-space: the median distance between tips and nodes and the 225 centroid (similar but not equivalent to Wills et al. 1994; Korn et al. 2013; Huang et al. 226 2015) calculated as:

Disparity = 
$$median \sqrt{\sum (\mathbf{v}_n - Centroid_n)^2}$$
 (1)

228 where:

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$$Centroid_n = \frac{\sum (\mathbf{v}_n)}{k} \tag{2}$$

and  $\mathbf{v}_n$  is any of the n eigenvectors (i.e. any of the n dimensions of the cladisto-space),

Centroid<sub>n</sub> is the mean value of the  $n^{th}$  eigenvector (equation 2) and k is the total number

of tips and nodes. Note that we also calculated the sum and products of ranges and

variances and refer to these results in the supplementary material (@@@).

### Estimating disparity through time

Changes in disparity through time are generally investigated by calculating the 234 disparity of taxa that occupy the cladisto-space during specific time intervals (e.g. 235 Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; Hopkins, 2013; 236 Benton et al., 2014; Benson and Druckenmiller, 2014). These time intervals are usually 237 defined based on biostratigraphy (e.g. Cisneros and Ruta, 2010; Prentice et al., 2011; 238 Hughes et al., 2013; Benton et al., 2014) but can also be arbitrarily chosen time periods 239 of equal duration (Butler et al., 2012; Hopkins, 2013; Benson and Druckenmiller, 2014). 240 However, this approach suffers from two main biases. First, if biostratigraphy is used to determine the time intervals, disparity may be distorted towards higher differences between time intervals because biostratigraphical periods are geologically defined based on differences in the morphology of fossils found in the different strata. Second, this approach assumes that all characters evolve following a punctuated equilibrium model, because disparity is only estimated once for each interval resulting in all

changes in disparity occurring between intervals, rather than also allowing for gradual
changes within intervals (Hunt et al., 2015).

To address these issues, we used a "time-slicing" approach that considers subsets of taxa in the cladisto-space at specific equidistant points in time, as opposed to considering subsets of taxa between two points in time. This results in even-sampling 251 of the cladisto-space across time and permits us to define the underlying model of character evolution (punctuated or gradual). In practice, time-slicing considers the 253 disparity of any element present in the phylogeny (branches, nodes and tips) at any point in time. When the phylogenetic elements are nodes or tips, the eigenvector scores 255 for the nodes (estimated using ancestral state reconstruction as described above) or tips 256 are directly used for estimating disparity. When the phylogenetic elements are branches 257 we chose the eigenvector score for the branch using one of two evolutionary models: 258

1. **Punctuated evolution.** This model selects the eigenvector score from either the ancestral node or the descendant node/tip of the branch regardless of the position of the slice along the branch. Similarly to the time interval approach, this reflects a model of punctuated evolution where changes in disparity occur either at the start or at the end of a branch over a relatively short time period and clades undergo long periods of stasis during their evolution (Gould and Eldredge, 1977; Hunt, 2007). We applied this model in three ways:

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(i) selecting the eigenvector score of the ancestral node of the branch (ACCTRAN).

- (ii) selecting the eigenvector score of the descendant node/tip of the branch (DELTRAN).
- (iii) randomly selecting either the eigenvector score of the ancestral node or the descendant node/tip of the branch (random).

Method (i) assumes that changes always occur early on the branch (accelerated transition, ACCTRAN) and (ii) assumes that changes always occur later (delayed transition, DELTRAN). We prefer not to make either assumption so we report the results from (iii), although the ACCTRAN and DELTRAN results are available in the Supplementary Information @@@.

2. Gradual evolution. This model also selects the eigenvector score from either the ancestral node or the descendant node/tip of the branch, but the choice depends on the distance between the sampling time point and the end of the branch. If the sampling time point falls in the first half of the branch length the eigenvector score is taken from the ancestral node, conversely, if the sampling time point falls in the second half of the branch length the eigenvector score is taken from the descendant node/tip. This reflects a model of gradual evolution where changes in disparity are gradual and cumulative along the branch. Under this model, the gradual changes could be either directional or random, however, directional evolution have been empirically shown to be rare (only 5% of the time Hunt, 2007). We therefore considered that changes from a character state A to B were only dependent on the branch length.

We applied our time-slicing approach separately to the two cladisto-spaces calculated for the Mammaliaformes and Eutheria datasets, time-slicing the phylogeny every five million years from 170 Ma to the present resulting in 35 subsamples of the cladisto-space. For each subsample, we estimated its disparity assuming punctuated (ACCTRAN, DELTRAN and random) and gradual evolution as described above. To 293 reduce the influence of outliers on our disparity estimates, we bootstrapped each disparity measurement by randomly resampling with replacement a new subsample of 295 taxa from the observed taxa in the subsample 1000 times. We then calculated the median disparity value for each subsample along with the 50% and 95% confidence 297 intervals. We also recorded the number of phylogenetic elements (nodes and tips) in 298 each subsample as a proxy for taxonomic diversity. To compare our results to previous 299 studies we also repeated our analyses using the time interval approach based on 300 biostratigraphy (e.g. Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; 301 Benton et al., 2014) using each geological stage from the Middle Jurassic to the present. 302 We report the results of these analyses in the Supplementary Materials (@@@). 303

Testing the effects of the K-Pg extinction on mammalian disparity

If the K-Pg extinction event had a significant effect on mammalian disparity, we should

see a significant difference between disparity at the end of the Cretaceous and disparity

at the start of the Paleogene. To test this, we performed *t*-tests to look for differences in

disparity between the time subsamples of interest (e.g. as used in Anderson and

Friedman, 2012; Zelditch et al., 2012; Smith et al., 2014). We compared the last time

subsample before the K-Pg boundary (70 Ma) to the first subsample of the Paleocene (65 Ma) for both the Mammaliaformes and Eutheria datasets and using both the 311 gradual and punctuated evolutionary models. Even though one million year after the K-Pg event (66 to 65 Ma) seems to be a rather short geological time frame, effects on mammalian evolution have been detected as early as half a million year after K-Pg 314 (Wilson, 2013). However, the effect of extinction on a group's evolution might not be 315 detectable directly after the event due to delays in recovery (e.g. Chen and Benton, 316 2012, estimated that ecosystems only fully recovered 8-9 Ma after the Permo-Triassic 317 mass extinction). Therefore, we also tested whether there was a significant difference in 318 disparity between the end of the Cretaceous (70 Ma) and all subsamples from the 319 Paleocene (65, 60 and 55 Ma). Additionally, some authors argue that the major 320 diversification event in mammals took place during the Paleocene-Eocene Thermal 321 Maximum (PETM;  $\sim$  56 Ma; Bininda-Emonds et al. 2007 but see Meredith et al. 2011 322 and Stadler 2011) with the extinctions at K-Pg providing the "empty" ecological space 323 required for this diversification to occur. We therefore extended our comparisons 324 between the last subsample of the Cretaceous (70 Ma) up to the late Eocene (35 Ma) to 325 check for a delayed effect of the K-Pg extinction potentially allowing morphological 326 diversification after the PETM. Because these analyses involved multiple comparisons, we used Bonferonni corrections (Holm, 1979) to ensure our significant results were 328 robust to Type I error rate inflation.

Finally, disparity may be higher in subsamples with more phylogenetic elements

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simply because there are more taxa represented. To test whether this influenced our results, we repeated the *t*-tests using the rarefied Mammaliaformes and Eutheria disparities. In the Mammaliaformes, the minimum number of taxa in each subsample from 170 Ma to present was eight. In the Eutheria, the minimum number of taxa in each subsample was three, however, from 150 Ma until the present, the minimum number of taxa is eight. To make both datasets comparable, we used eight as a minimum number of taxa for the rarefied bootstrap measurements, therefore in the Eutheria we ignored the subsample between 170 and 150 Ma that only contains three taxa.

RESULTS

Disparity in the Mammaliaformes reaches a plateau during the Middle Triassic around 340 240 Ma, and fluctuates slightly around this during the rest of the Mesozoic and the 341 Cenozoic (Fig 1 and Fig @@@ supplementary). The number of tips and nodes in each 342 time subsample (a proxy for taxonomic richness), however, show a more idiosyncratic 343 pattern with a steady increase until the Middle Jurassic around 170 Ma, Fig @@@ 344 supplementary) followed by random fluctuations during the rest of the Mesozoic and the Cenozoic (Fig 1). Disparity in the Eutheria reaches a plateau at the end of the Jurassic around 150 Ma, whereas the number of tips and nodes increases up to the K-Pg boundary and then decreases throughout the Cenozoic (Fig 1). For both Mammalia formes and Eutheria the same patterns in changes in disparity appear in the rarefied analyses (Fig 1 and @@rar@@sup). In both datasets the two evolutionary 350 models (gradual or punctuated) also yield similar results (Fig 1). 351

We found no significant differences in disparity between the last subsample of the Cretaceous (70 Ma) and the first subsample of the Paleogene (65 Ma; Table 1), using 353 both datasets and under both evolutionary models. We also found no significant differences in disparity between the last subsample of the Cretaceous (70 Ma) and any subsamples of the Paleocene and Eocene in Mammaliaformes under both evolutionary models and in Eutheria under a gradual evolutionary model (Table 1). However, in Eutheria under the punctuated evolutionary model, we found a small significant 358 difference (after applying Bonferonni corrections) in disparity between the last 359 subsample of the Cretaceous (70 Ma) and the subsamples at 45 Ma (an increase in 360 disparity of 0.17; Table 1). However, this result is not significant in the rarefied analyses 361 (Tables supplementary@@@). Otherwise the results of the rarefied analyses are the 362 same as when using the complete datasets. 363

### **Discussion**

Previous authors have suggested that the K-Pg extinction event released mammals from ecological pressures such as competition and predation, allowing them to radiate into newly available ecological niches (Archibald, 2011; O'Leary et al., 2013; Lovegrove et al., 2014; Slater, 2013). However, we did not detect any significant changes in mammalian disparity before and after K-Pg in either Mammaliaformes or Eutheria, under a model of punctuated or gradual evolution. Additionally, we tested whether the absence of a detectable effect might be due to a lag effect, with the effect only becoming obvious

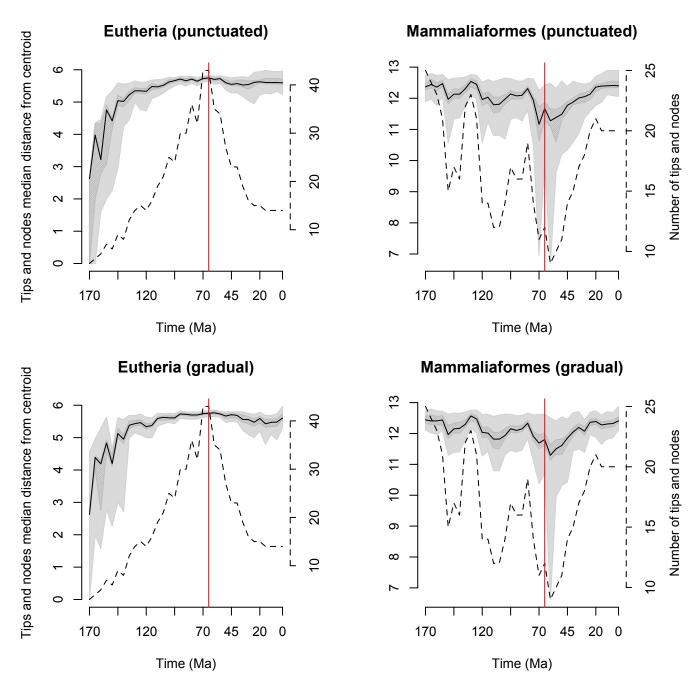


Figure 1: Disparity through time in Eutheria and Mammaliaformes calculated using a model of punctuated (upper panels) or gradual (lower panels) evolution. The x axis represents time in millions of years before the present (Ma). The y axis represents disparity, measured as the median distance between the centroid of the ordinated space and the tips/nodes in each time subsample. The solid black lines show the mean disparity estimated from 1000 bootstrapped pseudoreplicates and confidence intervals (CI) are represented by the grey polygons (50% CI in dark grey and 95% CI in light grey). The dashed line and the right hand axis represents the number of tips/nodes in each time slice. The red vertical line indicates the Cretaceous-Paleogene (K-Pg) boundary (66 Ma). Note that scale bars differ among panels.

later in the Paleocene. Even when accounting for such a lag effect, we did not detect
any significant effect of the K-Pg extinction event on mammalian disparity. Our results
imply that mammals did not diversify morphologically in response to the K-Pg
extinction event. Instead, their diversification appears to have begun before the end of
the Cretaceous (Fig 1, Table 1 and see Meredith et al., 2011; dos Reis et al., 2014; Close
et al., 2015; Lee and Beck, 2015).

We did, however, detect a small, yet significant, increase in disparity during the
Eocene (45 Ma) under a punctuated evolutionary model using the Eutheria dataset.

This might be due to a long lag effect of  $\sim$ 21 Ma after K-Pg. Note however, that this is 380 double the lag time observed in other mass extinctions (Chen and Benton, 2012). 381 Therefore, it may be more likely to be attributed to a lag effect of the Palaeocene-Eocene 382 Thermal Maximum (PETM;  $\sim$ 11 Ma afterwards; Bininda-Emonds et al., 2007). 383 However, this significant increase in disparity is only detected at 45 Ma but not 384 afterwards (which would be expected if the increase was due to an evolutionary 385 radiation) and is not seen under the gradual evolution model. This indicates that it is 386 more likely due to differences in the evolutionary models rather than an actual increase 387 in disparity. The 45 Ma time slice samples the long branch ( $\sim$ 50 Ma) leading to 388 Lepidictis (33.9 to 33.3 Ma) that branches with its closest relative Gypsonictops (66.8 to 66 Ma) in the early Upper Cretaceous  $\sim$ 90 Ma (see Fig S2 @@@). Therefore, in this time-slice under the gradual evolution model, the data for *Lepidictis* is always sampled, but under the punctuated evolution model the algorithm can also randomly sample the

data from its ancestor in the early Upper Cretaceous (see methods). This may inflate
differences compared to other slices. Incidentally, this increase can also be linked to the
number of tips and nodes used in the comparison (43 versus 23 tips and nodes at
respectively 70 and 45 Ma), because the increase is not significant in the rarefied
analysis (see supplementary results @@@ with only eight tips and nodes). Given these
caveats we believe that no strong conclusions can be drawn from the increase in
disparity during the Eocene.

Our results differ from a previous study that found an increase in disparity in 400 North American Theria as soon as  $\sim$ 0.5 Ma after K-Pg (Wilson, 2013). These differences 401 are likely to be related to several methodological differences between the present study 402 and the previous one (Wilson, 2013). Firstly, Wilson (2013) only measures disparity at a 403 regional scale (North America) and proposes that the observed increases in disparity 404 are linked to the immigration of new species into the study localities. This strongly 405 implies that disparity was higher on a global scale. Secondly, most of the debate on 406 mammalian diversification around the K-Pg boundary seems to be linked to the 407 conflicting signal between palaeontological and neontological data (Meredith et al. 2011 408 vs. O'Leary et al. 2013 but see dos Reis et al. 2014). Therefore, an effect of the K-Pg 409 extinction event might be detectable only when using just palaeontological data. In this 410 study, however, we use Total Evidence tip-dated trees based on both palaeontological 411 and neontological data (Slater, 2013; Beck and Lee, 2014), which may account for the differences between our study and that of Wilson (2013) who used only fossil data.

Interestingly, however, our results also differ from Slater (2013), the source of data for the Mammaliaformes dataset. Slater (2013) found support for a shift in the mode of body mass evolution (from an Ornstein-Uhlenbeck to a Brownian Motion model) directly after K-Pg suggesting a release in competition pressure or new niches becoming availabile for mammals in the early Paleocene. Our studies may show 418 different results due to the difference between changes observed in one continuous life-history trait (body mass; Slater, 2013) versus changes in an aggregate of 446 discrete 420 morphological traits (the cladistic characters) in the present analysis. Body mass and 421 disparity might be decoupled in a similar way to taxonomic diversity and disparity 422 (e.g. Slater et al., 2010; Ruta et al., 2013; Hopkins, 2013) because the latter does not rely 423 on size but rather on discrete morphological features. It is not unlikely that mammalian 424 disparity (sensu baüplan; e.g. Hughes et al., 2013) increased rapidly early in their 425 evolutionary history and then remained constant (Fig 1; Close et al., 2015; Lee and 426 Beck, 2015) while body mass variation continued to increase, especially after K-Pg 427 (Slater, 2013). Note, however, that our methods did not investigate changes in body 428 mass across the K-Pg boundary so they do not allow us to test this hypothesis. We 429 remain confident in our results because we recovered the same pattern from two 430 independent datasets (Slater, 2013; Beck and Lee, 2014). 431

There are several caveats to consider when interpreting our results. Firstly, both our datasets are limited taxonomically. They do not represent all known mammalian taxa, especially during the Neogene (23–2.58 Ma) where there are no fossil taxa in

boundary and not during the whole Cenozoic. Besides, this might not cause a serious
underestimation of disparity, at least for the Mammaliaformes, because their diversity
peaked during the late Cretaceous (Campanian; 72.1–83.6 Ma; Newham et al., 2014) and
mammalian diversification rates declined throughout the Cenozoic (Raia et al., 2012).
Therefore, an effect of the K-Pg boundary would be more likely to be detected during
the Paleogene when mammalian diversity was highest, so we do not believe that
increasing taxon sampling would greatly alter our conclusions.

Secondly, testing for significant changes in disparity through time is problematic. 443 The disparity of each subsample is necessarily dependent on disparity in the previous subsamples. For example, the tips and nodes used to estimate disparity are linked by 445 common evolutionary history, therefore two tips or nodes sharing a close ancestor are 446 more likely to have similar morphological features than more distantly related tips and 447 nodes. Similarly, when looking at disparity through time, different subsamples are 448 related by time, therefore, two subsamples closely together in time are more likely to 449 have the same disparity value than more distant subsamples. Additionally, because 450 disparity is a single value summarizing morphological disparity, its variance and mean 451 were calculated by bootstrapping, thus the variances and means used in our t-tests are 452 calculated from non-independent pseudoreplicates rather than true replicates. A 453 second caveat arising from using bootstraps is that using a large number of pseudoreplicates is likely to inflate Type I error rates. Currently, however, this method

is still widely used in disparity analyses for lack of a better alternative (e.g. Anderson and Friedman, 2012; Zelditch et al., 2012; Smith et al., 2014).

458

Methodological improvements for measuring disparity through time Our results may differ from previous studies because of our specific methodological choices. Throughout this paper, we propose several incremental changes to the classical 460 ways of measuring disparity. Firstly we used all the axes of the cladisto-space, as 461 opposed to previous studies that selected a subsample of the cladisto-space arguing 462 that the m first axes usually contain most of the dataset's variance (e.g Brusatte et al., 463 2008a; Cisneros and Ruta, 2010; Prentice et al., 2011; Anderson and Friedman, 2012; 464 Hughes et al., 2013; Benton et al., 2014). We argue that even if the last dimensions of the 465 cladisto-space contain a trivial amount of variance, there is no statistical justification for 466 excluding them. However, by doing so, we included dimensions of the cladisto-space 467 with near zero variance and range (the last dimension's variance was  $2 \times 10^{-14}$  and  $1.15 \times 10^{-15}$  and the range was  $7.31 \times 10^{-7}$  and  $3.33 \times 10^{-7}$  for respectively the Mammaliaformes and Eutheria datasets). An alternative method avoids this problem by simply not ordinating the data and using the raw distance matrix (e.g. Benson and Druckenmiller, 2014; Close et al., 2015). However, in both this method and our method, 472 the calculation of the products of ranges and variances is impossible.

Secondly, we used median distance between tips and nodes to centroid as a 474 disparity metric, rather than the classical sums and products of ranges and variances 475 (Wills et al., 1994). This metric is not affected by problems with using the last

dimensions of the cladisto-space (see above). Also, it has several other advantages over
other metrics. For example, it measures directly the median spread of the taxa in the
cladisto-space unlike the sum and products of ranges and variances that measure the
size of the cladisto-space dimensions (Wills et al., 1994). Additionally, it comes with no
statistical caveats unlike the sums or products of variances that should also include
covariances between axes to correctly assess the exact size of the cladisto-space (even
though the covariance term is usually close to o because of the eigen decomposition).

Finally, we used a time-slicing method instead of binning the data into time

Finally, we used a time-slicing method instead of binning the data into time intervals (e.g in: Cisneros and Ruta, 2010; Prentice et al., 2011; Hughes et al., 2013; 485 Hopkins, 2013; Benton et al., 2014; Benson and Druckenmiller, 2014) thus allowing us to 486 avoid two caveats of using the time intervals approach. Because time intervals are often 487 based on biostratigraphy, which is in turn based on notable differences in fossil fauna 488 and flora, this method is likely to artificially emphasize disparity differences among 489 time intervals. It is also possible to use arbitrary time bins of equal duration rather than 490 biostratigraphy (Butler et al., 2012; Hopkins, 2013; Benson and Druckenmiller, 2014), 491 but both approaches make the underlying assumption that disparity changes in a 492 punctuated manner, i.e. changes occur only between time intervals. However, gradual 493 evolution has been shown to be relatively common in the fossil record (Hunt, 2007; Hunt et al., 2015), so this assumption is unfounded. Our approach allowed us to fit different evolutionary models to our data - either assuming punctuated or gradual evolution. This is an improvement on previous approaches but could be improved

further by implementing other common but more complex models for example, a combined stasis and random walk (Hunt et al., 2015) or models based on morphological rates rather than just branch lengths.

501 Conclusion

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514

Evidence for whether mammals diversified before or after the K-Pg boundary is mixed

(Meredith et al., 2011; O'Leary et al., 2013; dos Reis et al., 2014; Beck and Lee, 2014),

and appears to be related to the kind of data used (fossils or living species) and how

the analyses were conducted. Using both fossil and living taxa, and investigating

morphological disparity through time rather than taxonomic diversity, we find no

direct effect of the K-Pg extinction event on the diversity of mammals. We therefore

suggest that, contrary to popular belief, the extinction of many terrestrial vertebrates

including the non-avian dinosaurs 66 million years ago, did not significantly affect the

evolution of mammals throughout the Cenozoic.

## Data availability and reproducibility

Data are available on Dryad or Figshare. Code for reproducing the analyses is available on GitHub (github.com/TGuillerme/SpatioTemporal\_Disparity).

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521 References

Anderson, P. S. and M. Friedman. 2012. Using cladistic characters to predict functional variety: experiments using early gnathostomes. Journal of Vertebrate Paleontology 32:1254–1270.

Archibald, J. D. 2011. Extinction and radiation: how the fall of dinosaurs led to the rise of mammals. JHU Press.

Arcila, D., R. A. Pyron, J. C. Tyler, G. Ort, and R. Betancur-R. 2015. An evaluation of fossil tip-dating versus node-age calibrations in tetraodontiform fishes (teleostei:

Percomorphaceae). Molecular Phylogenetics and Evolution 82, Part A:131 – 145.

Beck, R. M. and M. S. Lee. 2014. Ancient dates or accelerated rates? Morphological clocks and the antiquity of placental mammals. Proceedings of the Royal Society B:

Biological Sciences 281:1–10.

- Benson, R. B. J. and P. S. Druckenmiller. 2014. Faunal turnover of marine tetrapods
  during the Jurassic-Cretaceous transition. Biological Reviews 89:1–23.
- Benton, M. J. 1985. Mass extinction among non-marine tetrapods. Nature 316:811–814.
- Benton, M. J. 2015. Exploring macroevolution using modern and fossil data.
- Proceedings of the Royal Society of London B: Biological Sciences 282.
- Benton, M. J., J. Forth, and M. C. Langer. 2014. Models for the rise of the dinosaurs.
- Current biology: CB 24:R87–R95.
- Benton, M. J. and R. J. Twitchett. 2003. How to kill (almost) all life: the end-permian

  extinction event. Trends in Ecology and Evolution 18:358 365.
- Bininda-Emonds, O. R., M. Cardillo, K. E. Jones, R. D. MacPhee, R. M. Beck, R. Grenyer,
- S. A. Price, R. A. Vos, J. L. Gittleman, and A. Purvis. 2007. The delayed rise of
- present-day mammals. Nature 446:507–512.
- Brusatte, S., R. J. Butler, A. Prieto-Márquez, and M. A. Norell. 2012. Dinosaur
- morphological diversity and the end-Cretaceous extinction. Nature Communications
- 3:804-804.
- Brusatte, S. L., M. J. Benton, M. Ruta, and G. T. Lloyd. 2008a. The first 50Łmyr of
- dinosaur evolution: macroevolutionary pattern and morphological disparity. Biology
- 550 Letters 4:733–736.

- Brusatte, S. L., M. J. Benton, M. Ruta, and G. T. Lloyd. 2008b. Superiority, competition, and opportunism in the evolutionary radiation of dinosaurs. Science 321:1485–1488.
- Brusatte, S. L., R. J. Butler, P. M. Barrett, M. T. Carrano, D. C. Evans, G. T. Lloyd, P. D.
- Mannion, M. A. Norell, D. J. Peppe, P. Upchurch, and T. E. Williamson. 2015. The
  extinction of the dinosaurs. Biological Reviews 90:628–642.
- Butler, R. J., S. L. Brusatte, B. Andres, and R. B. J. Benson. 2012. How do geological
   sampling biases affect studies of morphological evolution in deep time? a case study
   of pterosaur (reptilia: Archosauria) disparity. Evolution 66:147–162.
- <sup>559</sup> Cailliez, F. 1983. The analytical solution of the additive constant problem.
- Psychometrika 48:305–308.
- Chen, Z.-Q. and M. J. Benton. 2012. The timing and pattern of biotic recovery following
  the end-permian mass extinction. Nature Geoscience 5:375–383.
- Ciampaglio, C. N. 2004. Measuring changes in articulate brachiopod morphology

  before and after the permian mass extinction event: do developmental constraints

  limit morphological innovation? Evolution & Development 6:260–274.
- Ciampaglio, C. N., M. Kemp, and D. W. McShea. 2001. Detecting changes in
   morphospace occupation patterns in the fossil record: characterization and analysis
   of measures of disparity. Paleobiology 27:695–715.
- Cisneros, J. C. and M. Ruta. 2010. Morphological diversity and biogeography of

- procolophonids (amniota: Parareptilia). Journal of Systematic Palaeontology 8:607–625.
- <sup>572</sup> Close, R., M. Friedman, G. Lloyd, and R. Benson. 2015. Evidence for a mid-jurassic adaptive radiation in mammals. Current Biology .
- <sup>574</sup> Coxall, H. K., S. D'Hondt, and J. C. Zachos. 2006. Pelagic evolution and environmental recovery after the cretaceous-paleogene mass extinction. Geology 34:297–300.
- dos Reis, M., P. C. J. Donoghue, and Z. Yang. 2014. Neither phylogenomic nor
  palaeontological data support a palaeogene origin of placental mammals. Biology
  Letters 10.
- dos Reis, M., J. Inoue, M. Hasegawa, R. J. Asher, P. C. J. Donoghue, and Z. Yang. 2012.
- Phylogenomic datasets provide both precision and accuracy in estimating the
- timescale of placental mammal phylogeny. Proceedings of the Royal Society of
- London B: Biological Sciences .
- Douady, C. J. and E. J. Douzery. 2003. Molecular estimation of eulipotyphlan divergence times and the evolution of insectivora. Molecular Phylogenetics and Evolution 28:285 – 296.
- Erwin, D. H. 1998. The end and the beginning: recoveries from mass extinctions. Trends in Ecology and Evolution 13:344 – 349.
- Erwin, D. H. 2007. Disparity: Morphological pattern and developmental context.
- Palaeontology 50:57-73.

- Foote, M. 1994. Morphological disparity in ordovician-devonian crinoids and the early saturation of morphological space. Paleobiology 20:320–344.
- Foote, M. 1996. Ecological controls on the evolutionary recovery of post-paleozoic crinoids. Science 274:1492–1495.
- Foth, C., S. Brusatte, and R. Butler. 2012. Do different disparity proxies converge on a common signal? insights from the cranial morphometrics and evolutionary history of pterosauria (diapsida: Archosauria). Journal of evolutionary biology 25:904–915.
- Friedman, M. 2010. Explosive morphological diversification of spiny-finned teleost fishes in the aftermath of the end-Cretaceous extinction. Proceedings of the Royal Society B: Biological Sciences 277:1675–1683.
- Garland, J., Theodore and A. R. Ives. 2000. Using the past to predict the present:
- Confidence intervals for regression equations in phylogenetic comparative methods.
- The American Naturalist 155:346–364.
- Glor, R. E. 2010. Phylogenetic insights on adaptive radiation. Annual Review of
   Ecology, Evolution, and Systematics 41:251–270.
- Goswami, A., G. V. Prasad, P. Upchurch, D. M. Boyer, E. R. Seiffert, O. Verma,
- E. Gheerbrant, and J. J. Flynn. 2011. A radiation of arboreal basal eutherian mammals
- beginning in the late cretaceous of india. Proceedings of the National Academy of
- Sciences of the United States of America 108:16333–16338.

- Gould, S. J. and N. Eldredge. 1977. Punctuated equilibria: The tempo and mode of
  evolution reconsidered. Paleobiology 3:pp. 115–151.
- Gower, J. C. 1966. Some distance properties of latent root and vector methods used in multivariate analysis. Biometrika 53:325–338.
- 613 Gower, J. C. 1971. A general coefficient of similarity and some of its properties.
- Biometrics 27:pp. 857–871.
- Guillerme, T. and N. Cooper. In press. Effects of missing data on topological inference using a total evidence approach,. Molecular Phylogenetics and Evolution .
- Hetherington, A. J., E. Sherratt, M. Ruta, M. Wilkinson, B. Deline, and P. C. Donoghue.
- 2015. Do cladistic and morphometric data capture common patterns of
- morphological disparity? Palaeontology 58:393–399.
- Holm, S. 1979. A simple sequentially rejective multiple test procedure. Scandinavian journal of statistics Pages 65–70.
- Hopkins, M. 2013. Decoupling of taxonomic diversity and morphological disparity
  during decline of the cambrian trilobite family pterocephaliidae. Journal of
  Evolutionary Biology 26:1665–1676.
- Huang, S., K. Roy, and D. Jablonski. 2015. Origins, bottlenecks, and present-day diversity: Patterns of morphospace occupation in marine bivalves. Evolution .
- Hughes, M., S. Gerber, and M. A. Wills. 2013. Clades reach highest morphological

- disparity early in their evolution. Proceedings of the National Academy of Sciences
  110:13875–13879.
- Hunt, G. 2007. The relative importance of directional change, random walks, and stasis in the evolution of fossil lineages. Proceedings of the National Academy of Sciences
  104:18404–18408.
- Hunt, G., M. J. Hopkins, and S. Lidgard. 2015. Simple versus complex models of trait
  evolution and stasis as a response to environmental change. Proceedings of the
  National Academy of Sciences 112:4885–4890.
- Korn, D., M. J. Hopkins, and S. A. Walton. 2013. Extinction spacea method for the quantification and classification of changes in morphospace across extinction boundaries. Evolution 67:2795–2810.
- Lee, M. and R. Beck. 2015. Mammalian evolution: A jurassic spark. Current Biology
  25:R759 R761.
- Liow, L. H., T. Reitan, and P. G. Harnik. 2015. Ecological interactions on
  macroevolutionary time scales: clams and brachiopods are more than ships that pass
  in the night. Ecology Letters Pages n/a–n/a.
- Lloyd, G. T. 2015. Claddis: Measuring Morphological Diversity and Evolutionary

  Tempo. R package version 0.1.
- Losos, J. B. 2010. Adaptive radiation, ecological opportunity, and evolutionary determinism. The American Naturalist 175:pp. 623–639.

- Lovegrove, B. G., K. D. Lobban, and D. L. Levesque. 2014. Mammal survival at the
- cretaceous-palaeogene boundary: metabolic homeostasis in prolonged tropical
- hibernation in tenrecs. Proceedings of the Royal Society of London B: Biological
- Sciences 281.
- Meredith, R., J. Janečka, J. Gatesy, O. Ryder, C. Fisher, E. Teeling, A. Goodbla, E. Eizirik,
- T. L. Simão, T. Stadler, D. Rabosky, R. Honeycutt, J. Flynn, C. Ingram, C. Steiner,
- T. Williams, T. Robinson, B. Angela, M. Westerman, N. Ayoub, M. Springer, and
- W. Murphy. 2011. Impacts of the Cretaceous terrestrial revolution and KPg extinction
- on mammal diversification. Science 334:521–524.
- Newham, E., R. Benson, P. Upchurch, and A. Goswami. 2014. Mesozoic mammaliaform
- diversity: The effect of sampling corrections on reconstructions of evolutionary
- dynamics. Palaeogeography, Palaeoclimatology, Palaeoecology 412:32 44.
- O'Leary, M. A., J. I. Bloch, J. J. Flynn, T. J. Gaudin, A. Giallombardo, N. P. Giannini, S. L.
- Goldberg, B. P. Kraatz, Z.-X. Luo, J. Meng, X. Ni, M. J. Novacek, F. A. Perini, Z. S.
- Randall, G. W. Rougier, E. J. Sargis, M. T. Silcox, N. B. Simmons, M. Spaulding, P. M.
- Velazco, M. Weksler, J. R. Wible, and A. L. Cirranello. 2013. The placental mammal
- ancestor and the postK-Pg radiation of placentals. Science 339:662–667.
- Olson, M. E. and A. Arroyo-Santos. 2009. Thinking in continua: beyond the adaptive
- radiation metaphor. BioEssays 31:1337–1346.
- OLeary, M. A., J. I. Bloch, J. J. Flynn, T. J. Gaudin, A. Giallombardo, N. P. Giannini, S. L.

- Goldberg, B. P. Kraatz, Z.-X. Luo, J. Meng, X. Ni, M. J. Novacek, F. A. Perini,
- Z. Randall, G. W. Rougier, E. J. Sargis, M. T. Silcox, N. B. Simmons, M. Spaulding,
- P. M. Velazco, M. Weksler, J. R. Wible, and A. L. Cirranello. 2013. Response to
- comment on the placental mammal ancestor and the postk-pg radiation of placentals.
- 672 Science 341:613.
- Paradis, E., J. Claude, and K. Strimmer. 2004. APE: analyses of phylogenetics and
- evolution in R language. Bioinformatics 20:289–290.
- Payne, J. L., N. A. Heim, M. L. Knope, and C. R. McClain. 2014. Metabolic dominance
- of bivalves predates brachiopod diversity decline by more than 150 million years.
- Proceedings of the Royal Society B: Biological Sciences 281.
- Pearman, P. B., A. Guisan, O. Broennimann, and C. F. Randin. 2008. Niche dynamics in
- space and time. Trends in Ecology and Evolution 23:149 158.
- Prentice, K. C., P. Ruta, and M. J. Benton. 2011. Evolution of morphological disparity in
- pterosaurs. Journal of Systematic Palaeontology 9:337–353.
- R Core Team. 2015. R: a language and environment for statistical computing. R
- Foundation for Statistical Computing Vienna, Austria.
- Raia, P., F. Carotenuto, F. Passaro, P. Piras, D. Fulgione, L. Werdelin, J. Saarinen, and
- M. Fortelius. 2012. Rapid action in the palaeogene, the relationship between
- phenotypic and taxonomic diversification in coenozoic mammals. Proceedings of the
- Royal Society of London B: Biological Sciences 280.

- Raup, D. M. 1979. Size of the permo-triassic bottleneck and its evolutionary implications. Science 206:217–218.
- Renne, P. R., A. L. Deino, F. J. Hilgen, K. F. Kuiper, D. F. Mark, W. S. Mitchell, L. E.
- Morgan, R. Mundil, and J. Smit. 2013. Time scales of critical events around the
- 692 Cretaceous-Paleogene boundary. Science 339:684–687.
- Revell, L. J. 2012. phytools: An R package for phylogenetic comparative biology (and other things). Methods in Ecology and Evolution 3:217–223.
- Ronquist, F., S. Klopfstein, L. Vilhelmsen, S. Schulmeister, D. Murray, and A. Rasnitsyn.
- 2012. A total-evidence approach to dating with fossils, applied to the early radiation
- of the Hymenoptera. Systematic Biology 61:973–999.
- Ruta, M., K. D. Angielczyk, J. Fröbisch, and M. J. Benton. 2013. Decoupling of
- morphological disparity and taxic diversity during the adaptive radiation of
- anomodont therapsids. Proceedings of the Royal Society of London B: Biological
- <sup>701</sup> Sciences 280.
- Slater, G. J. 2013. Phylogenetic evidence for a shift in the mode of mammalian body size
- evolution at the cretaceous-palaeogene boundary. Methods in Ecology and Evolution
- 704 **4:734-744**.
- Slater, G. J., S. A. Price, F. Santini, and M. E. Alfaro. 2010. Diversity versus disparity and
- the radiation of modern cetaceans. Proceedings of the Royal Society of London B:
- Biological Sciences .

- Smith, A. J., M. V. Rosario, T. P. Eiting, and E. R. Dumont. 2014. Joined at the hip:
- Linked characters and the problem of missing data in studies of disparity. Evolution
- 710 68:2386–2400.
- Springer, M. S., R. W. Meredith, E. C. Teeling, and W. J. Murphy. 2013. Technical
- comment on the placental mammal ancestor and the postk-pg radiation of placentals.
- 713 Science 341:613.
- Stadler, T. 2011. Mammalian phylogeny reveals recent diversification rate shifts.
- Proceedings of the National Academy of Sciences 108:6187–6192.
- Thorne, P. M., M. Ruta, and M. J. Benton. 2011. Resetting the evolution of marine
- reptiles at the Triassic-Jurassic boundary. Proceedings of the National Academy of
- Sciences 108:8339–8344.
- Toljagic, O. and R. J. Butler. 2013. Triassic-Jurassic mass extinction as trigger for the
- mesozoic radiation of crocodylomorphs. Biology Letters 9.
- Torgerson, W. S. 1965. Multidimensional scaling of similarity. Psychometrika
- 722 30:379-393.
- Wesley-Hunt, G. D. 2005. The morphological diversification of carnivores in north
- america. Paleobiology 31:35–55.
- Wills, M. A., D. E. G. Briggs, and R. A. Fortey. 1994. Disparity as an evolutionary index:
- A comparison of cambrian and recent arthropods. Paleobiology 20:93–130.

- Wilson, G. P. 2013. Mammals across the k/pg boundary in northeastern montana, u.s.a.:
- dental morphology and body-size patterns reveal extinction selectivity and
- immigrant-fueled ecospace filling. Paleobiology 39:429–469.
- Wood, H. M., N. J. Matzke, R. G. Gillespie, and C. E. Griswold. 2013. Treating fossils as
- terminal taxa in divergence time estimation reveals ancient vicariance patterns in the
- palpimanoid spiders. Systematic Biology 62:264–284.
- Yang, Z., S. Kumar, and M. Nei. 1996. A new method of inference of ancestral
- nucleotide and amino acid sequences. Genetics 141:1641–50.
- Zelditch, M. L., D. L. Swiderski, and H. D. Sheets. 2012. Geometric morphometrics for
- biologists: a primer. Academic Press.

**Table 1:** Results of *t*-tests comparing disparity at the last subsample of the Cretaceous (70 Ma) to subsamples of the Paleocene and Eocene, under both gradual and punctuated evolutionary models, in Mammaliaformes and Eutheria. Difference = mean difference in disparity between the two subsamples being compared; df = degrees of freedom; p value = original p value prior to Bonferonni correction. Significant differences (after applying Bonferonni corrections for multiple comparisons) are highlighted in bold.

Subsamples	Gradual evolution model				Punctuated evolution model			
compared	difference	df	t	p value	difference	df	t	p value
Mammaliaformes								
70 vs. 65	-0.420	21	-0.808	0.428	-0.030	21	-0.058	0.954
70 vs. 60	0.030	18	0.046	0.964	0.210	18	0.379	0.709
70 vs. 55	0.010	19	0.021	0.983	0.110	19	0.225	0.824
70 vs. 50	-0.260	20	-0.456	0.653	0.030	20	0.060	0.953
70 vs. 45	-0.430	23	-0.869	0.394	0.060	23	0.132	0.896
70 <i>vs.</i> 40	-0.620	24	-1.388	0.178	-0.410	24	-1.031	0.313
70 vs. 35	-0.730	26	-1.742	0.093	-0.340	26	-0.861	0.397
Eutheria								
70 vs. 65	-0.020	84	-0.503	0.616	0.010	84	0.288	0.774
70 <i>vs.</i> 60	0.030	76	0.617	0.539	0.080	76	1.693	0.095
70 vs. 55	0.030	75	0.519	0.605	0.030	75	0.699	0.486
70 vs. 50	0.130	68	2.101	0.039 <sup>1</sup>	0.080	68	1.458	0.149
70 <i>vs.</i> 45	0.190	64	2.679	0.009 <sup>1</sup>	0.170	64	2.730	0.006 <sup>2</sup>
70 <i>vs.</i> 40	0.160	64	2.249	0.0281	0.130	64	2.084	0.041 <sup>1</sup>
70 vs. 35	0.190	60	2.358	0.022	0.120	60	1.893	0.063

<sup>&</sup>lt;sup>1</sup>p value is non-significant after applying Bonferonni correction; <sup>2</sup>p value is **0.048** after applying