

# 432 Class 01

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2026-01-13

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## Getting To These Slides

Our web site: <https://thomaselove.github.io/432-2026/>

- Note that this link is posted to the bottom of every slide.

Visit the [Calendar](#) at the top of the page, which will take you to the Class 01 README page.

- Slides for Class 01 linked at Class 01 README.
  - We'll look at the **HTML slides** during class.
  - We also provide the Quarto code, and a PDF version. All of the class materials are written in [Quarto](#) within RStudio.

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# Today's Agenda

1. Mechanics of the course
2. Why I write dates the way I do
3. Getting Organized
4. Building and Validating models for Penguin Bill Length
5. Setting Up [Lab 1](#)

# Course Mechanics

# Welcome to 432.

Just about everything is linked at  
<https://thomaseLove.github.io/432-2026>

- [Calendar](#)
  - final word on all deadlines, and links to each class and TA office hours.
- [Syllabus](#) (can download as PDF)
- [Course Notes](#) HTML and PDF

## Course Notes

- <https://thomaseLove.github.io/432-notes/> has 34 chapters (> 900 pages) to supplement this course.
- Review of Key Ideas from 431 in Chapters 3-5
  - Chapter 3: Comparing Means
  - Chapter 4: Comparing Rates/Proportions
  - Chapter 5: Fitting Linear Models (where we'll start)
- Walk-through of Two Large Examples (NHANES data in Chapters 1-2, BRFSS SMART data in Chapter 6)

# Also linked on our website

- [Software](#)
  - Updating / Installing R and RStudio, necessary R Packages
- Get Data (Code, Quarto templates) at [our 432-data page](#)
- Assignments (7 Labs, 2 Projects, 2 Quizzes)
- [Sources](#) (books, articles, videos, etc.)
- Key Links ([Canvas](#), Shared Google Drive, [Minute Papers](#))
- Contact Us (**431-help at case dot edu** + TA office hours + Me)

## Assignments

Every deliverable is listed in the [Calendar](#).

- [Welcome to 432 Survey](#) due tomorrow (2026-01-14) at Noon.
- Be sure you see the course in [Canvas](#), and the Shared Google Drive at your CWRU log-in. Thanks.

Assignments include two projects, seven labs, and two quizzes, plus some minute papers. Almost everything due on Wednesdays at noon.

# Two Projects

**Project A** (publicly available data: linear & logistic models)

- You'll need your data cleaned and in R by 2026-02-01
- Final Portfolio & (recorded) Presentation due 2026-02-25

**Project B** (use almost any data and build specific models)

1. Proposal is part of Lab 6: 2026-04-01
2. Presentation (in-person or Zoom) in late April
3. Portfolio (prepared using Quarto) due 2026-05-06

# Seven Labs

Seven labs, meant to be (generally) shorter than 431 Labs

1. **Lab 1** is due Wednesday 2026-01-21 at Noon.
2. **Lab 2** is due Wednesday 2026-01-28 at Noon.

**Lab 5** is about building or augmenting your website, and can be done now (or at any time), although it's not due until 2026-03-18.

- Everyone needs to do all seven labs - there is no “skipping” this term.

# Two Quizzes

- Quiz 1 due 2026-03-06 (Friday)
- Quiz 2 due 2026-04-24 (Friday)
  - Receive each Quiz one week in advance
  - Mostly multiple choice or short answer, via Google Form.

[Syllabus](#), Lab Instructions provide feedback details and grading approach for the semester.

## Getting Help

- 9 teaching assistants volunteering their time to help you.
- TAs hold Zoom Office Hours (every day but Wednesday) starting Friday 2026-01-16.
  - No office hours on MLK Day or during Spring Break.
- Or email us at **431-help** at **case dot edu**.
- I am also available after every class to chat.
- Email me at Thomas dot Love at case dot edu.

We WELCOME questions/comments/corrections/thoughts!

# Tools You Will Use in this Class

- **Course Website** (bottom of every slide) especially the [Calendar](#)
  - Each class has a README plus slides
- **R, RStudio and Quarto** for, well, everything
- [Canvas](#) for access to Zoom meetings and 432 recordings, submission of Labs and Project assignments

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# Tools You Will Use in this Class

- **Google Drive via CWRU** for forms (Surveys/Quizzes) and for feedback on assignments.
- **Zoom** for class sessions / recordings and TA office hours
- Jeff Leek's 80-page book [How To Be A Modern Scientist](#) which you'll finish reading by 2026-02-01.

A few source materials are **password-protected**. What is the password?

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An approximate answer to the right  
problem is worth a good deal more  
than an exact answer to an  
approximate problem.

— *John Tukey* —

AZ QUOTES

# Why I Write Dates The Way I Do



# How To Write Dates



## Data Organization in Spreadsheets (Broman & Woo)

- Create a data dictionary.
  - Jeff Leek has good thoughts on this in “How to Share Data with a Statistician” at <https://github.com/jtleek/datasharing>
  - Shannon Ellis and Jeff Leek’s preprint “How to Share data for Collaboration” touches on many of the same points at <https://peerj.com/preprints/3139v5.pdf>

# Sharing Data with a Statistician

We want:

1. The raw data.
2. A tidy data set.
3. A codebook describing each variable and its values in the tidy data set.
4. An explicit and exact recipe describing how you went from 1 to 2 and 3.

## Data Organization in Spreadsheets: Be Consistent

- Consistent codes for categorical variables.
  - Either “M” or “Male” but not both at the same time.
  - Make it clear enough to reduce dependence on a codebook.
  - No spaces or special characters other than \_ in category names.

# Data Organization in Spreadsheets: Be Consistent

- Consistent fixed codes for missing values.
  - NA is the most convenient R choice.
- Consistent variable names
  - In R, I'll use `clean_names` from the `janitor` package to turn everything into snake\_case.
  - In R, start your variable names with letters. No spaces, no special characters other than `_`.

# Data Organization in Spreadsheets: Be Consistent

- Consistent subject / record identifiers
  - And if you're building a .csv in Excel, don't use ID as the name of that identifier.
- Consistent data layouts across multiple files.

# What Goes in a Cell?

- Make your data a rectangle.
  - Each row represents a record (sometimes a subject).
  - Each column represents a variable.
  - First column is a unique identifier for each record.
- No empty cells.
- One Thing in each cell.
- No calculations in the raw data
- No font colors and no highlighting

## Use consistent, strong file names.

Jenny Bryan's advice on "Naming Things" hold up well. There's a full presentation at [SpeakerDeck](#).

Good file names:

- are machine readable (easy to search, easy to extract info from names)
- are human readable (name contains content information, so it's easy to figure out what something is based on its name)

# from Jenny Bryan's “Naming Things” slides...

Good file names:

- play well with default ordering (something numeric first, left padded with zeros as needed, use ISO 8601 standard for dates)

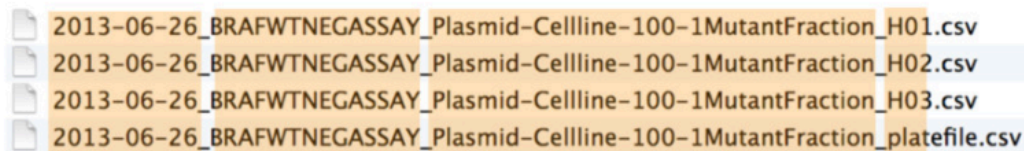
Avoid: spaces, punctuation, accented characters, case sensitivity

## from Jenny Bryan...

# Jenny Bryan: Deliberate Use of Delimiters

Deliberately use delimiters to make things easy to compute on and make it easy to recover meta-data from the filenames.

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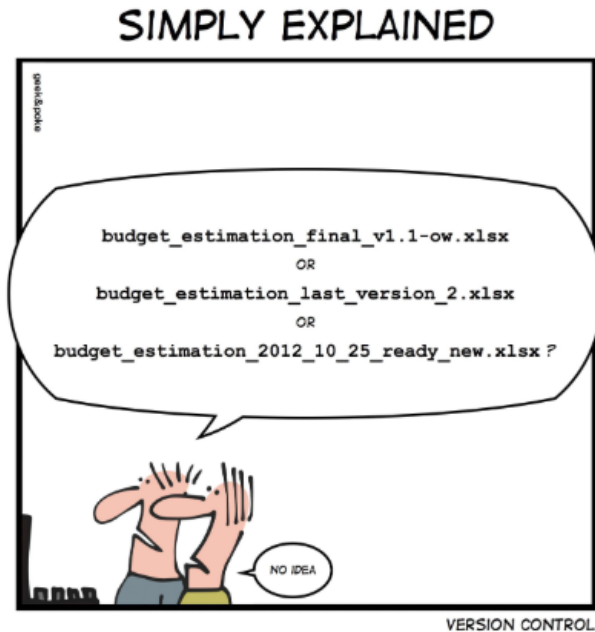
2013-06-26\_BRAFWTNEGASSAY\_Plasmid-Cellline-100-1MutantFraction\_H01.csv  
2013-06-26\_BRAFWTNEGASSAY\_Plasmid-Cellline-100-1MutantFraction\_H02.csv  
2013-06-26\_BRAFWTNEGASSAY\_Plasmid-Cellline-100-1MutantFraction\_H03.csv  
2013-06-26\_BRAFWTNEGASSAY\_Plasmid-Cellline-100-1MutantFraction\_platefile.csv

```
> flist <- list.files(pattern = "Plasmid") %>% head
> stringr::str_split_fixed(flist, "[_\\.]", 5)
      [,1]      [,2]      [,3]      [,4] [,5]
[1,] "2013-06-26" "BRAFWTNEGASSAY" "Plasmid-Cellline-100-1MutantFraction" "A01" "csv"
[2,] "2013-06-26" "BRAFWTNEGASSAY" "Plasmid-Cellline-100-1MutantFraction" "A02" "csv"
[3,] "2013-06-26" "BRAFWTNEGASSAY" "Plasmid-Cellline-100-1MutantFraction" "A03" "csv"
[4,] "2013-06-26" "BRAFWTNEGASSAY" "Plasmid-Cellline-100-1MutantFraction" "B01" "csv"
[5,] "2013-06-26" "BRAFWTNEGASSAY" "Plasmid-Cellline-100-1MutantFraction" "B02" "csv"
[6,] "2013-06-26" "BRAFWTNEGASSAY" "Plasmid-Cellline-100-1MutantFraction" "B03" "csv"
```

“\_” underscore used to delimit units of meta-data I want later

“-” hyphen used to delimit words so my eyes don’t bleed

# Goal: Avoid this...



Idea from Jen Simmons and John Albin Wilkins during episode #40 of "Web Ahead" about Git:  
<http://5by5.tv/webahead/40>

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## Get organized

### Be organized

do this as you go, not "tomorrow"

but also don't fret over past mistakes  
raise the bar for *new* work

Don't spend a lot of time bemoaning or cleaning up past ills.  
Strive to improve this sort of thing going forward.

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# “Good Enough Practices”

1. Save the raw data.
2. Ensure that raw data is backed up more than once.
3. Create the data you wish to see in the world (the data you wish you had received.)
4. Create analysis-friendly, tidy data.
5. Record all of the steps used to process data.
6. Anticipate the need for multiple tables, and use a unique identifier for every record.

# Building and Validating Linear Prediction Models



# R Setup

```
1 knitr::opts_chunk$set(comment = NA)
2
3 library(janitor)
4
5 library(broom); library(car); library(GGally); library(glue)
6 library(gt); library(kableExtra); library(knitr); library(mosaic)
7 library(patchwork); library(rsample); library(palmerpenguins)
8
9 library(easystats)
10 library(tidyverse)
11
12 theme_set(theme_bw())
```

# Data Load

```
1 our_tibble <- penguins |>
2   select(species, sex, bill_length_mm) |>
3   drop_na()
4
5 our_tibble |> summary()
```

species	sex	bill_length_mm
Adelie :146	female:165	Min. :32.10
Chinstrap: 68	male :168	1st Qu.:39.50
Gentoo :119		Median :44.50
		Mean :43.99
		3rd Qu.:48.60
		Max. :59.60

- We're going to try to predict bill length using species and sex as predictors.

# Partition `our_tibble` into training/test samples

We will place 60% of the penguins in our training sample, and require that similar fractions of each species occur in our training and testing samples. We use functions from the `rsample` package here.

```
1 set.seed(20260113)
2 our_split <- initial_split(our_tibble, prop = 0.6, strata = species)
3 our_train <- training(our_split)
4 our_test <- testing(our_split)
```

We could have used `slice_sample()` as in the [Course Notes](#), too.

## Result of our partitioning

```
1 our_train |> tabyl(species) |> adorn_totals() |>
2   adorn_pct_formatting()
```

species	n	percent
Adelie	87	43.9%
Chinstrap	40	20.2%
Gentoo	71	35.9%
Total	198	100.0%

```
1 our_test |> tabyl(species) |> adorn_totals() |>
2   adorn_pct_formatting()
```

species	n	percent
Adelie	59	43.7%
Chinstrap	28	20.7%
Gentoo	48	35.6%
Total	135	100.0%

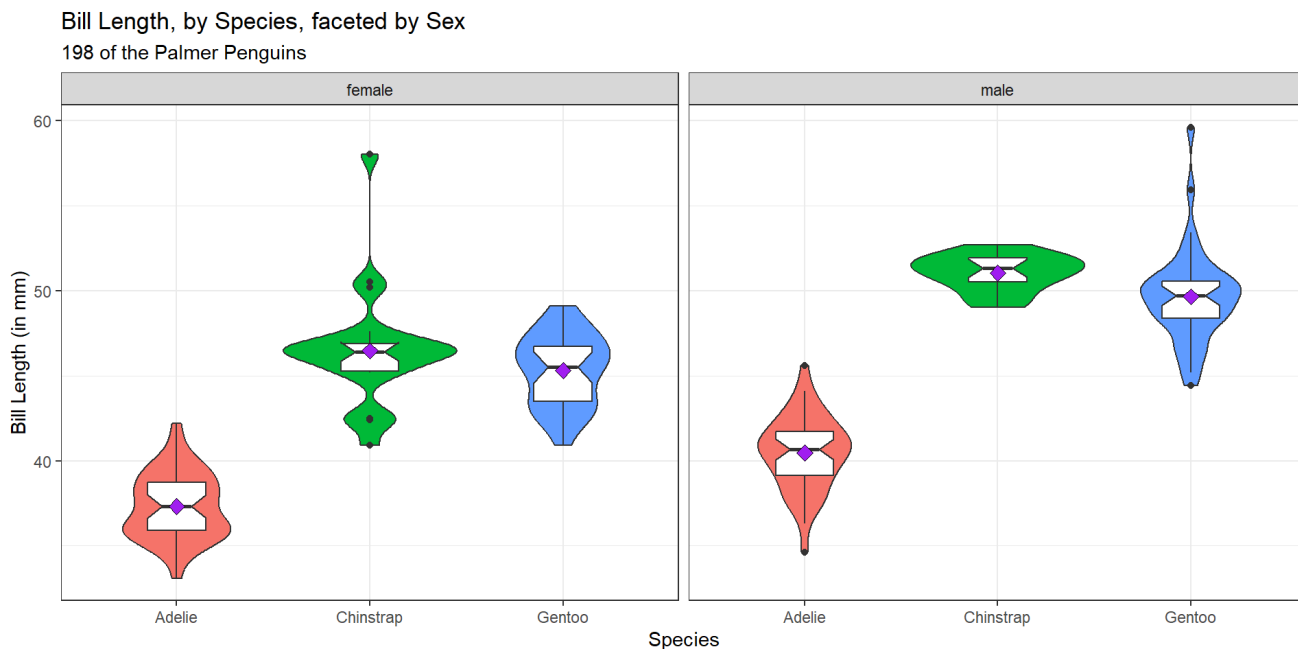
# What will this produce?

```
1 ggplot(data = our_train, aes(x = species, y = bill_length_mm)) +  
2   geom_violin(aes(fill = species)) +  
3   geom_boxplot(width = 0.3, notch = TRUE) +  
4   stat_summary(fill = "purple", fun = "mean", geom = "point",  
5               shape = 23, size = 3) +  
6   facet_wrap(~ sex) +  
7   guides(fill = "none") +  
8   labs(title = "Bill Length, by Species, faceted by Sex",  
9        subtitle = glue(nrow(our_train), " of the Palmer Penguins"),  
10       x = "Species", y = "Bill Length (in mm)")
```

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# What will this produce?



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# Standing Break

## Model m1

```
1 m1 <- lm(bill_length_mm ~ species + sex, data = our_train)
2
3 anova(m1)
```

### Analysis of Variance Table

Response: bill\_length\_mm

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
species	2	4070.3	2035.16	365.42	< 2.2e-16 ***
sex	1	728.6	728.62	130.83	< 2.2e-16 ***
Residuals	194	1080.5	5.57		

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

# Model 1 coefficients

```
1 tidy(m1, conf.int = TRUE, conf.level = 0.90) |>
2   select(term, estimate, conf.low, conf.high)
```

# A tibble: 4 × 4

	term <chr>	estimate <dbl>	conf.low <dbl>	conf.high <dbl>
1	(Intercept)	37.0	36.5	37.5
2	speciesChinstrap	9.83	9.08	10.6
3	speciesGentoo	8.62	7.99	9.24
4	sexmale	3.85	3.30	4.41

```
1 model_parameters(m1, ci = 0.90)
```

Parameter	Coefficient	SE	90% CI	t(194)	p
(Intercept)	36.96	0.30	[36.45, 37.46]	121.18	< .001
species [Chinstrap]	9.83	0.45	[ 9.08, 10.57]	21.77	< .001
species [Gentoo]	8.62	0.38	[ 7.99,  9.24]	22.81	< .001
sex [male]	3.85	0.34	[ 3.30,  4.41]	11.44	< .001

## Interlude (Four Ways to Display Tables in Slides)

# Model 1 Parameters (version 1)

```
1 tidy(m1, conf.int = TRUE, conf.level = 0.90) |>
2   select(term, estimate, conf.low, conf.high)
```

# A tibble: 4 × 4

	term	estimate	conf.low	conf.high
	<chr>	<dbl>	<dbl>	<dbl>
1	(Intercept)	37.0	36.5	37.5
2	speciesChinstrap	9.83	9.08	10.6
3	speciesGentoo	8.62	7.99	9.24
4	sexmale	3.85	3.30	4.41

```
1 model_parameters(m1, ci = 0.90)
```

Parameter	Coefficient	SE	90% CI	t(194)	p
(Intercept)	36.96	0.30	[36.45, 37.46]	121.18	< .001
species [Chinstrap]	9.83	0.45	[ 9.08, 10.57]	21.77	< .001
species [Gentoo]	8.62	0.38	[ 7.99,  9.24]	22.81	< .001
sex [male]	3.85	0.34	[ 3.30,  4.41]	11.44	< .001

# Model 1 Parameters (version 2a)

```
1 tidy(m1, conf.int = TRUE, conf.level = 0.90) |>
2   select(term, estimate, conf.low, conf.high) |>
3   gt() |> fmt_number(decimals = 2) |>
4   tab_options(table.font.size = 24)
```

term	estimate	conf.low	conf.high
(Intercept)	36.96	36.45	37.46
speciesChinstrap	9.83	9.08	10.57
speciesGentoo	8.62	7.99	9.24
sexmale	3.85	3.30	4.41

# Model 1 Parameters (version 2b)

```
1 model_parameters(m1, ci = 0.90) |>
2   gt() |> fmt_number(columns = Coefficient:t, decimals = 2) |>
3   tab_options(table.font.size = 24)
```

Parameter	Coefficient	SE	CI	CI_low	CI_high	t	df_error	p
(Intercept)	36.96	0.30	0.90	36.45	37.46	121.18	194	8.692751e-185
speciesChinstrap	9.83	0.45	0.90	9.08	10.57	21.77	194	5.435642e-54
speciesGentoo	8.62	0.38	0.90	7.99	9.24	22.81	194	8.469781e-57
sexmale	3.85	0.34	0.90	3.30	4.41	11.44	194	1.731114e-23

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# Model 1 Parameters (version 3)

```
1 tidy(m1, conf.int = TRUE, conf.level = 0.90) |>
2   select(term, estimate, conf.low, conf.high) |>
3   kable(digits = 2) |> kable_styling(font_size = 24)
```

term	estimate	conf.low	conf.high
(Intercept)	36.96	36.45	37.46
speciesChinstrap	9.83	9.08	10.57
speciesGentoo	8.62	7.99	9.24
sexmale	3.85	3.30	4.41

```
1 model_parameters(m1, ci = 0.90) |>
2   kable(digits = 2) |> kable_styling(font_size = 24)
```

Parameter	Coefficient	SE	CI	CI_low	CI_high	t	df_error	p
(Intercept)	36.96	0.30	0.9	36.45	37.46	121.18	194	0
speciesChinstrap	9.83	0.45	0.9	9.08	10.57	21.77	194	0
speciesGentoo	8.62	0.38	0.9	7.99	9.24	22.81	194	0
sexmale	3.85	0.34	0.9	3.30	4.41	11.44	194	0

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# Model 1 Parameters (version 4)

```
1 tidy(m1, conf.int = TRUE, conf.level = 0.90) |>
2   select(term, estimate, conf.low, conf.high) |>
3   print_html(digits = 2)
```

term	estimate	conf.low	conf.high
(Intercept)	36.96	36.45	37.46
speciesChinstrap	9.83	9.08	10.57
speciesGentoo	8.62	7.99	9.24
sexmale	3.85	3.30	4.41

```
1 model_parameters(m1, ci = 0.90) |>
2   print_html(digits = 2, font_size = "60%")
```

Parameter	Coefficient	SE	90% CI	t(194)	p
(Intercept)	36.96	0.30	(36.45, 37.46)	121.18	< .001
species (Chinstrap)	9.83	0.45	(9.08, 10.57)	21.77	< .001
species (Gentoo)	8.62	0.38	(7.99, 9.24)	22.81	< .001
sex (male)	3.85	0.34	(3.30, 4.41)	11.44	< .001

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## Model 1 performance

```
1 model_performance(m1) |> gt() |>
2   fmt_number(decimals = 3) |> tab_options(table.font.size = 24)
```

AIC	AICc	BIC	R2	R2_adjusted	RMSE	Sigma
907.880	908.192	924.321	0.816	0.813	2.336	2.360

```
1 glance(m1) |> gt() |>
2   fmt_number(columns = -c("df", "df.residual", "nobs"), decimals = 3) |>
3   tab_options(table.font.size = 20)
```

r.squared	adj.r.squared	sigma	statistic	p.value	df	logLik	AIC	BIC	deviance	df.residual	nobs
0.816	0.813	2.360	287.223	0.000	3	-448.940	907.880	924.321	1,080.453	194	198



# Model m2

```
1 m2 <- lm(bill_length_mm ~ species, data = our_train)
2
3 ## anova(m2) yields p-value < 2.2e-16 (not shown here)
4
5 tidy(m2, conf.int = TRUE, conf.level = 0.90) |>
6   select(term, estimate, conf.low, conf.high) |>
7   kable(digits = 1)
```

term	estimate	conf.low	conf.high
(Intercept)	38.9	38.4	39.4
speciesChinstrap	9.6	8.6	10.6
speciesGentoo	8.8	8.0	9.6

## Comparison of Coefficients

```
1 compare_models(m1, m2)
```

Parameter	m1	m2
(Intercept)	36.96 (36.36, 37.56)	38.91 (38.26, 39.55)
species [Chinstrap]	9.83 ( 8.94, 10.72)	9.61 ( 8.46, 10.76)
species [Gentoo]	8.62 ( 7.87, 9.36)	8.84 ( 7.88, 9.80)
sex [male]	3.85 ( 3.19, 4.52)	
Observations	198	198

# In-Sample Comparison

```
1 bind_rows(glance(m1), glance(m2)) |>
2   mutate(model = c("m1", "m2")) |>
3   select(model, r2 = r.squared, adjr2 = adj.r.squared,
4           AIC, BIC, sigma, nobs) |>
5   kable(digits = c(0, 3, 3, 1, 1, 2, 0))
```

model	r2	adjr2	AIC	BIC	sigma	nobs
m1	0.816	0.813	907.9	924.3	2.36	198
m2	0.692	0.689	1007.9	1021.1	3.05	198

Which model has better in-sample performance?

## Comparing **m1** vs. **m2** performance

```
1 compare_performance(m1, m2)
```

# Comparison of Model Performance Indices

Name	Model	AIC (weights)	AICc (weights)	BIC (weights)	R2
m1	lm	907.9 (>.999)	908.2 (>.999)	924.3 (>.999)	0.816
m2	lm	1007.9 (<.001)	1008.1 (<.001)	1021.1 (<.001)	0.692

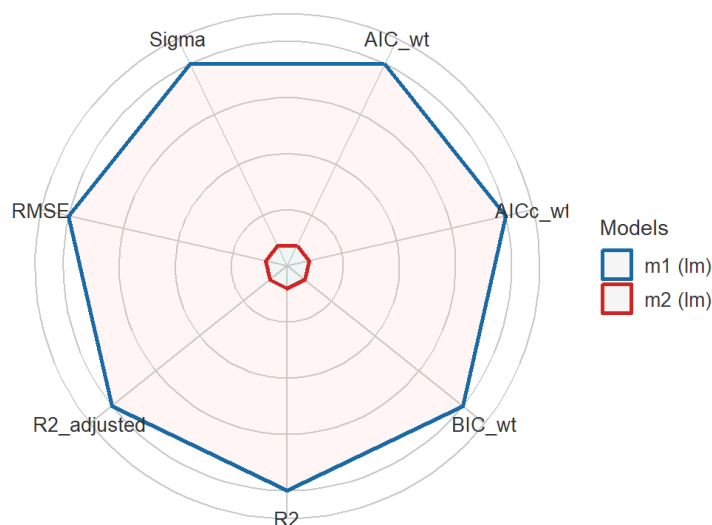
Name	R2 (adj.)	RMSE	Sigma
m1	0.813	2.336	2.360
m2	0.689	3.023	3.046

Which model has better in-sample performance?

# Plot for m1 vs. m2 (training)

```
1 plot(compare_performance(m1, m2))
```

Comparison of Model Indices



## Assessing Performance in Test Sample

```
1 m1_aug <- augment(m1, newdata = our_test)
2
3 m1_res <- m1_aug |>
4   summarize(val_R_sq = cor(bill_length_mm, .fitted)^2,
5             MAPE = mean(abs(.resid)),
6             RMSPE = sqrt(mean(.resid^2)),
7             max_Error = max(abs(.resid)))
8
9 m2_aug <- augment(m2, newdata = our_test)
10
11 m2_res <- m2_aug |>
12   summarize(val_R_sq = cor(bill_length_mm, .fitted)^2,
13             MAPE = mean(abs(.resid)),
14             RMSPE = sqrt(mean(.resid^2)),
15             max_Error = max(abs(.resid)))
```

# Test Sample Performance

```
1 bind_rows(m1_res, m2_res) |>
2 mutate(model = c("m1", "m2")) |>
3 relocate(model) |> kable(digits = c(0, 3, 2, 2, 1))
```

model	val_R_sq	MAPE	RMSPE	max_Error
m1	0.827	1.77	2.28	5.7
m2	0.725	2.34	2.88	7.4

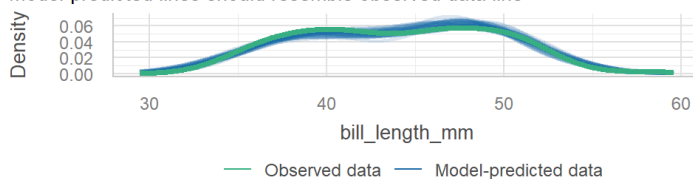
Which model predicts better in the test sample?

## Checking **m1** (see next 3 slides)

```
1 check_model(m1, detrend = FALSE)
```

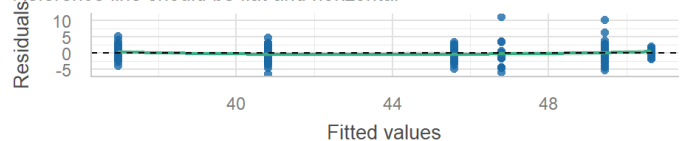
### Posterior Predictive Check

Model-predicted lines should resemble observed data line



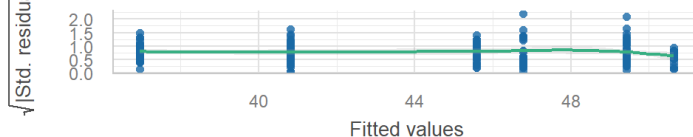
### Linearity

Reference line should be flat and horizontal



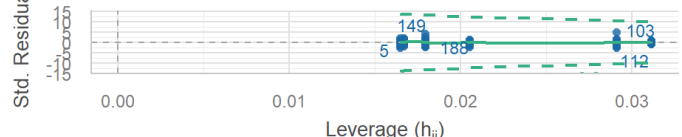
### Homogeneity of Variance

Reference line should be flat and horizontal



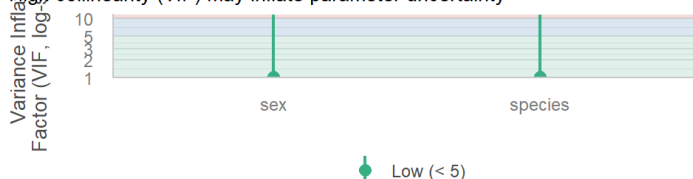
### Influential Observations

Points should be inside the contour lines



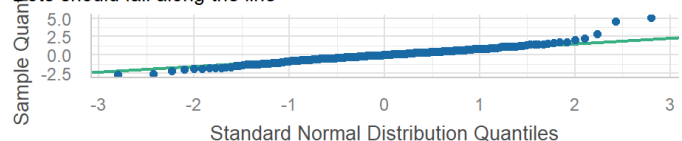
### Collinearity

High collinearity (VIF) may inflate parameter uncertainty



### Normality of Residuals

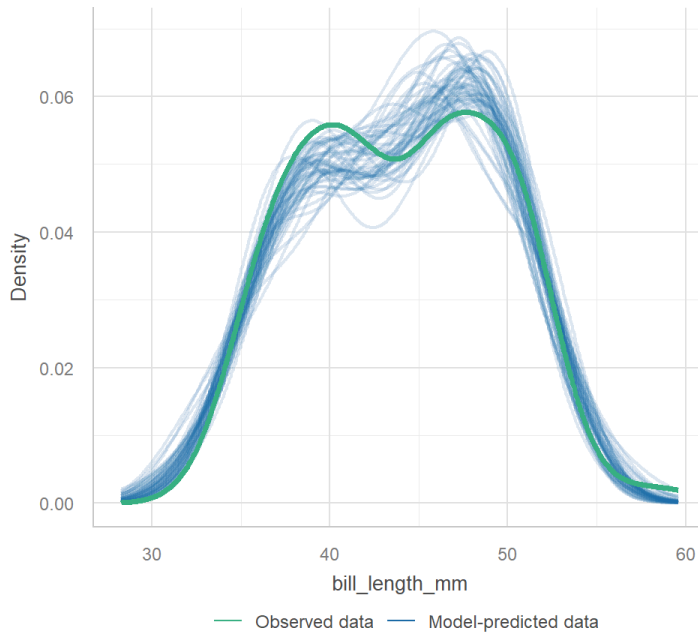
Points should fall along the line



# check\_model(m1): first 2 plots

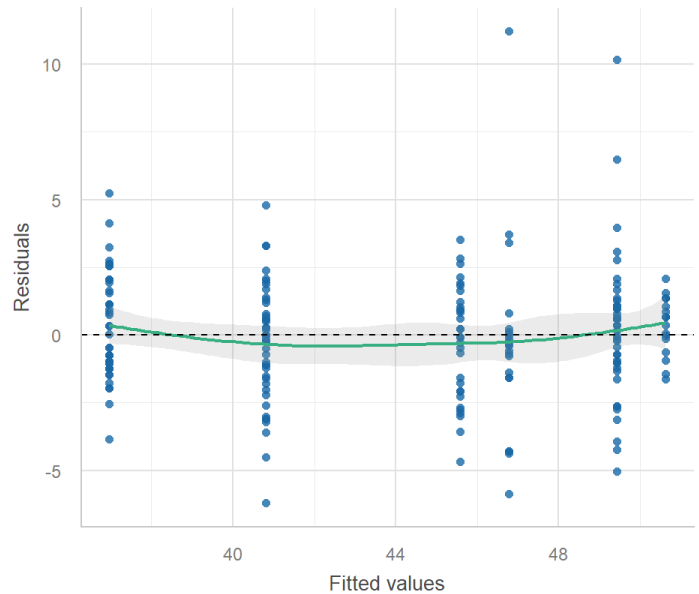
## Posterior Predictive Check

Model-predicted lines should resemble observed data line



## Linearity

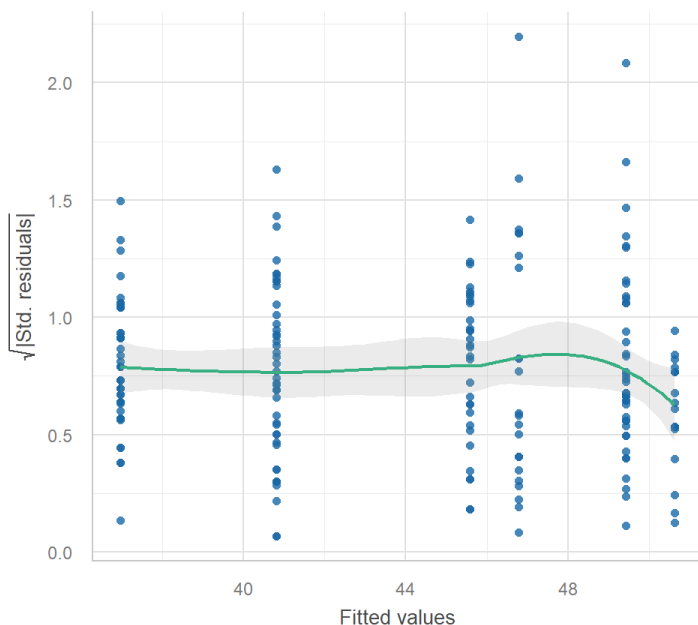
Reference line should be flat and horizontal



# check\_model(m1): next 2 plots

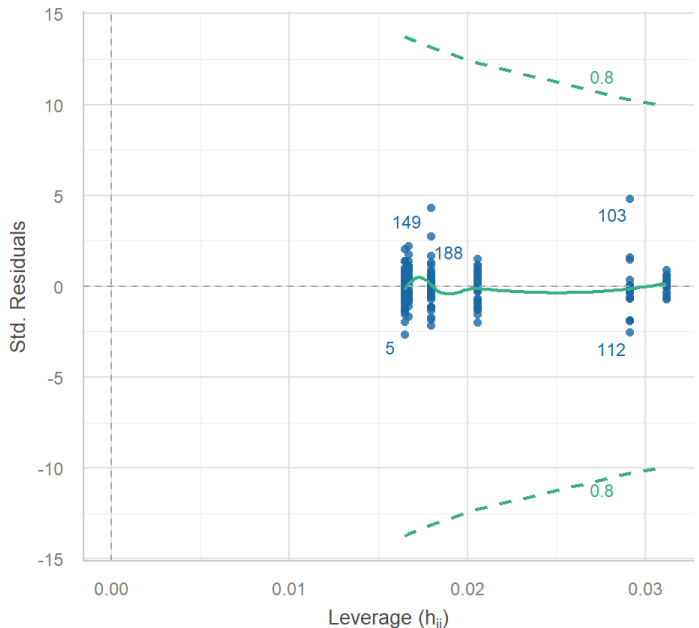
## Homogeneity of Variance

Reference line should be flat and horizontal



## Influential Observations

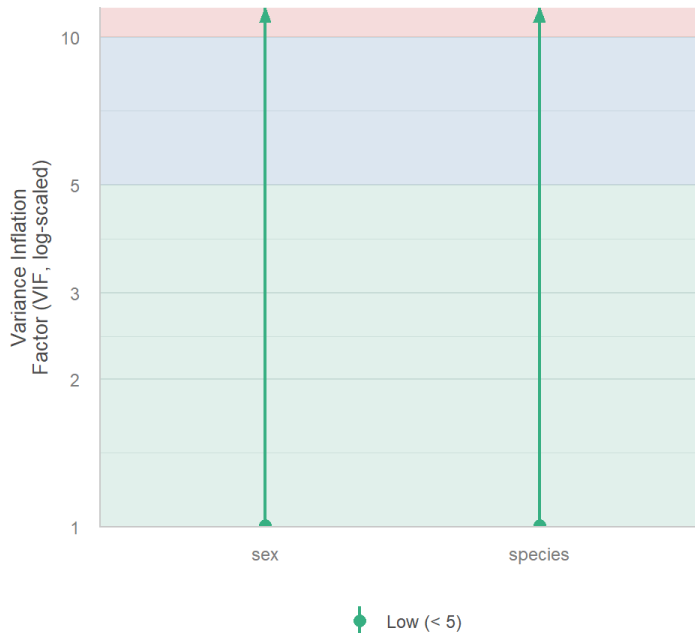
Points should be inside the contour lines



# check\_model(m1): final 2 plots

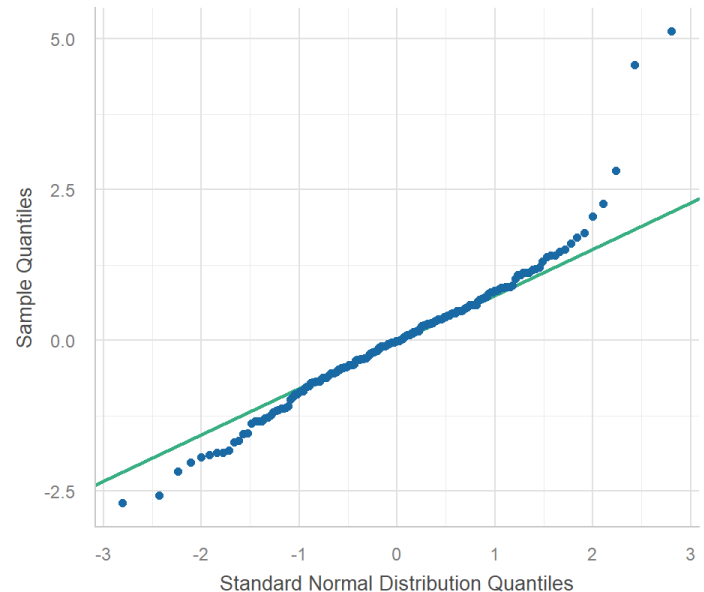
## Collinearity

High collinearity (VIF) may inflate parameter uncertainty



## Normality of Residuals

Dots should fall along the line



## What we did in this example...

1. R packages, usual commands, ingest the data.
2. Look at what we have and ensure it makes sense. (DTDP)
3. Partition the data into a training sample and a test sample.
4. Run a two-way ANOVA model (called `m1`) in the training sample; evaluate the quality of fit.
5. Run a one-way ANOVA model (called `m2`) in the training sample; evaluate the quality of fit.

# What we did in this example...

6. Use augment to predict from each model into the test sample; summarize and compare predictive quality.
7. Choose between the models and evaluate assumptions for our choice.

# Setting Up Lab 1, due 2025-01-21 at Noon

# Lab 1 Question 1

I provide some County Health Rankings data for 30 variables and 3054 counties included in the CHR 2024 report. You will filter the data down to the 88 counties in Ohio, and check for missing values.

Then you will create a visualization involving information from three different variables (from a list of 15) using R and Quarto.

There is a [Quarto template for Lab 1](#), in addition to the data set.

# Lab 1 Question 2

Create a linear regression model to predict **obesity** as a function of **food\_env**, adjusting for **unemployment** (all of these are quantitative variables.)

- Specify and fit the model, interpret **food\_env** coefficient and its confidence interval carefully.
- Evaluate quality of model in terms of adherence to regression assumptions via **check\_model()**.
- Build a nice table comparing your model to a simple regression for **obesity** using only **food\_env**, then reflect on your findings.



# For Next Time...

1. If you're not registered with SIS, do so, for PQHS/CRSP/MPHP 432.
2. Check that you see the course on [Canvas](#) and that you see the Shared Google Drive when logged into Google via CWRU.
3. Review the website and [Calendar](#), and skim the [Syllabus](#) and [Course Notes](#).
4. Welcome to 432 Survey at <https://bit.ly/432-2026-welcome-survey> by noon tomorrow (2026-01-14.)

# For Next Time...

5. Buy Jeff Leek's [How to be a Modern Scientist](#) and read it by the end of January.
6. Get started installing or updating the [software](#) you need for the course.
7. Get started on [Lab 1](#), due Wednesday 2026-01-21 at Noon.