

# 432 Class 04

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## Today's Agenda

- The HELP study
- Using tools from `rms` to fit:
  - linear models with `ols()`
  - logistic models with `lrm()`

# Today's R Setup

```
1 knitr::opts_chunk$set(comment = NA)
2
3 library(janitor)
4 library(naniar)
5 library(broom); library(gt); library(patchwork)
6
7 library(haven)           ## for zapping labels
8 library(mosaic)          ## auto-loads mosaicData - data source
9 library(GGally)          ## for scatterplot matrix
10 library(rsample)
11 library(yardstick)
12
13 library(rms)             ## auto-loads Hmisc
14 library(easystats)
15 library(tidyverse)
16
17 theme_set(theme_bw())
```

# Data from the HELP study

# New Data (The HELP study)

Today's main data set comes from the Health Evaluation and Linkage to Primary Care trial, and is stored as `HELPrct` in the `mosaicData` package.

HELP was a clinical trial of adult inpatients recruited from a detoxification unit. Patients with no primary care physician were randomized to receive a multidisciplinary assessment and a brief motivational intervention or usual care, with the goal of linking them to primary medical care.

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## Key Variables for Today

Variable	Description
<code>id</code>	subject identifier (note: $n = 453$ subjects)
<code>cesd</code>	Center for Epidemiologic Studies Depression measure (scale is 0-60; higher scores indicate more depressive symptoms)
<code>age</code>	subject age (in years)
<code>sex</code>	female ( $n = 107$ ) or male ( $n = 346$ )
<code>subst</code>	primary substance of abuse (alcohol, cocaine or heroin)
<code>mcs</code>	SF-36 Mental Component Score (lower = worse status)
<code>pcs</code>	SF-36 Physical Component Score (lower = worse status)
<code>pss_fr</code>	perceived social support by friends (higher = more support)

- All measures from baseline during the subjects' detoxification stay.
- More data and details at <https://nhorton.people.amherst.edu/help/>.

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# help\_rct data load

```
1 help_rct <- tibble(mosaicData::HELPrct) |>
2   select(id, cesd, age, sex, subst = substance, mcs, pcs, pss_fr) |>
3   mutate(across(where(is.character), as_factor)) |>
4   mutate(id = as.character(id))
5
6 help_rct
```

# A tibble: 453 × 8

	id	cesd	age	sex	subst	mcs	pcs	pss_fr
	<chr>	<int>	<int>	<fct>	<fct>	<dbl>	<dbl>	<int>
1	1	49	37	male	cocaine	25.1	58.4	0
2	2	30	37	male	alcohol	26.7	36.0	1
3	3	39	26	male	heroin	6.76	74.8	13
4	4	15	39	female	heroin	44.0	61.9	11
5	5	39	32	male	cocaine	21.7	37.3	10
6	6	6	47	female	cocaine	55.5	46.5	5
7	7	52	49	female	cocaine	21.8	24.5	1
8	8	32	28	male	alcohol	9.16	65.1	4
9	9	50	50	female	alcohol	22.0	38.3	5
10	10	46	39	male	heroin	36.1	22.6	0

# i 443 more rows

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## What the data look like in help\_rct

Note the labels.

```
1 str(help_rct)
```

```
tibble [453 × 8] (S3: tbl_df/tbl/data.frame)
 $ id      : chr [1:453] "1" "2" "3" "4" ...
 $ cesd    : int [1:453] 49 30 39 15 39 6 52 32 50 46 ...
 ..- attr(*, "label")= chr "CESD at baseline"
 $ age     : int [1:453] 37 37 26 39 32 47 49 28 50 39 ...
 ..- attr(*, "label")= chr "age (years)"
 $ sex     : Factor w/ 2 levels "female","male": 2 2 2 1 2 1 1 2 1 2 ...
 ..- attr(*, "label")= chr "sex"
 $ subst   : Factor w/ 3 levels "alcohol","cocaine",...: 2 1 3 3 2 2 2 1 1 3 ...
 ..- attr(*, "label")= chr "primary substance of abuse"
 $ mcs     : num [1:453] 25.11 26.67 6.76 43.97 21.68 ...
 ..- attr(*, "label")= chr "SF-36 Mental Component Score"
 $ pcs     : num [1:453] 58.4 36 74.8 61.9 37.3 ...
 ..- attr(*, "label")= chr "SF-36 Physical Component Score"
```

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# Getting rid of the labels

Suppose I don't want the labels for some reason...

```
1 help1 <- help_rct |> zap_label()
2 data_codebook(help1 |> select(-id))
```

select(help1, -id) (453 rows and 7 variables, 7 shown)

ID	Name	Type	Missings	Values	N
1	cesd	integer	0 (0.0%)	[1, 60]	453
2	age	integer	0 (0.0%)	[19, 60]	453
3	sex	categorical	0 (0.0%)	female male	107 (23.6%) 346 (76.4%)
4	subst	categorical	0 (0.0%)	alcohol cocaine heroin	177 (39.1%) 152 (33.6%) 124 (27.4%)

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## Quantitative Summaries

```
1 df_stats(~ cesd + age + mcs + pcs + pss_fr, data = help1) |>
2   gt() |>
3   fmt_number(min:max, decimals = 1) |>
4   fmt_number(mean:sd, decimals = 2) |>
5   tab_options(table.font.size = 24) |>
6   opt_stylize(style = 1, color = "blue")
```

response	min	Q1	median	Q3	max	mean	sd	n	missing
cesd	1.0	25.0	34.0	41.0	60.0	32.85	12.51	453	0
age	19.0	30.0	35.0	40.0	60.0	35.65	7.71	453	0
mcs	6.8	21.7	28.6	40.9	62.2	31.68	12.84	453	0
pcs	14.1	40.4	48.9	57.0	74.8	48.05	10.78	453	0
pss_fr	0.0	3.0	7.0	10.0	14.0	6.71	4.00	453	0

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# help1 categorical variables

```
1 help1 |> tabyl(sex, subst) |>
2   adorn_totals(where = c("row", "col")) |>
3   adorn_percentages(denominator = "row") |>
4   adorn_pct_formatting() |>
5   adorn_ns(position = "front") |>
6   adorn_title(placement = "combined") |>
7   gt() |> tab_options(table.font.size = 24) |>
8   opt_stylize(style = 2, color = "green")
```

sex/subst	alcohol	cocaine	heroin	Total
female	36 (33.6%)	41 (38.3%)	30 (28.0%)	107 (100.0%)
male	141 (40.8%)	111 (32.1%)	94 (27.2%)	346 (100.0%)
Total	177 (39.1%)	152 (33.6%)	124 (27.4%)	453 (100.0%)

## Our quantitative outcome

- The CES-D is a 20-item measure that asks people to rate how often over the past week they experienced symptoms associated with depression, such as restless sleep, poor appetite, and feeling lonely.
  - Each item is rated on a 0-3 scale, and then summed, so possible scores range from 0 to 60.
  - Higher scores indicate more symptoms (or more frequent symptoms.)
- A version of the CES-D scale is available [here as a PDF](#).

# A cutoff for CES-D: Our binary outcome

- Scores of 16 or higher on the CES-D scale are sometimes taken to indicate that a person is at risk for clinical depression.

```
1 help1 <- help1 |> mutate(cesd_hi = factor(as.numeric(cesd >= 16)))
2
3 help1 |> tabyl(cesd_hi) |> adorn_pct_formatting()
```

cesd_hi	n	percent
0	46	10.2%
1	407	89.8%

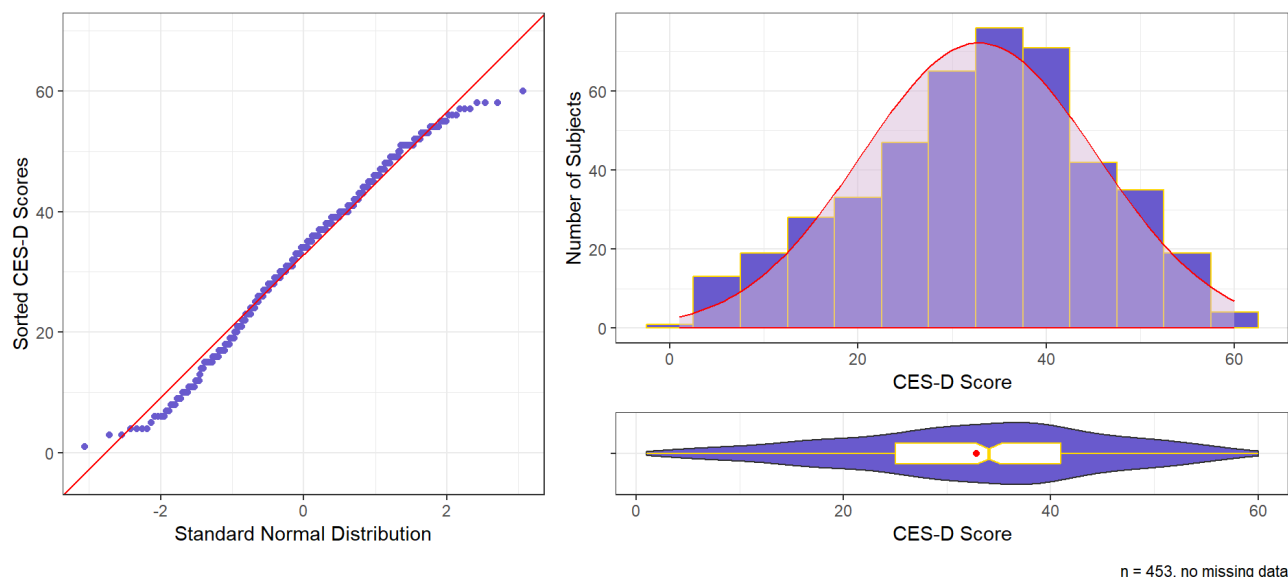
## Quantitative Outcome (CES-D)

```
1 p1 <- ggplot(help1, aes(sample = cesd)) +
2   geom_qq(col = "slateblue") + geom_qq_line(col = "red") +
3   theme(aspect.ratio = 1) +
4     labs(y = "Sorted CES-D Scores",
5          x = "Standard Normal Distribution")
6
7 bw = 5 # I tried a couple of things - this worked best for me with these data
8
9 p2 <- ggplot(help1, aes(x = cesd)) +
10   geom_histogram(binwidth = bw, fill = "slateblue", col = "gold") +
11   stat_function(fun = function(x)
12     dnorm(x, mean = mean(help1$cesd), sd = sd(help1$cesd)) *
13     length(help1$cesd) * bw,
14     geom = "area", alpha = 0.5, fill = "thistle", col = "red") +
15   labs(y = "Number of Subjects", x = "CES-D Score")
16
17 p3 <- ggplot(help1, aes(x = cesd, y = "")) +
18   geom_violin(fill = "slateblue") +
```

# Quantitative Outcome (CES-D)

CES-D Depression Scores from help1 data

Higher CES-D scores indicate more severe depressive symptoms



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## Describing CES-D (1/2)

```
1 describe(help1$cesd) ## describe comes from the Hmisc package
```

help1\$cesd

n	missing	distinct	Info	Mean	pMedian	Gmd	.05
453	0	58	0.999	32.85	33	14.23	10.0
.10	.25	.50	.75	.90	.95		
15.2	25.0	34.0	41.0	49.0	52.4		

lowest : 1 3 4 5 6, highest: 55 56 57 58 60

- **Info** = variable's information, between 0 and 1: the higher the **Info**, the more continuous the variable is (the fewer ties there are.)
- **pMedian** = Hodges-Lehman one-sample estimator of the pseudo-median. Median of all possible pairs of values.

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# Describing our outcome (2/2)

```
1 describe(help1$cesd)
```

```
help1$cesd
      n missing distinct      Info      Mean  pMedian      Gmd      .05
453      0       58      0.999     32.85      33      14.23     10.0
.10     .25     .50      .75      .90      .95
15.2    25.0    34.0     41.0     49.0     52.4
```

```
lowest : 1 3 4 5 6, highest: 55 56 57 58 60
```

- **Gmd** = Gini's mean difference, a robust measure of variation. If you select two subjects at random many times, the mean **cesd** difference will be 14.23 points.

More on the **Hmisc** package and **describe()** at [Frank Harrell's website](#)

## The **easystats** approach

```
1 describe_distribution(help1$cesd, iqr = FALSE, range = FALSE, ci = 0.90)
```

```
Mean | 90% CI (Mean) | SD | Skewness | Kurtosis | n | n_Missing
-----
32.85 | [32.11, 33.78] | 12.51 | -0.26 | -0.44 | 453 | 0
```

```
1 describe_distribution(help1$cesd,
2 centrality = "median", iqr = TRUE, quartiles = FALSE)
```

```
Median | MAD | IQR | Range | Skewness | Kurtosis | n | n_Missing
-----
34 | 11.86 | 16.50 | [1.00, 60.00] | -0.26 | -0.44 | 453 | 0
```

See [this link at the datawizard package](#) for more

# Scatterplot Matrix (code)

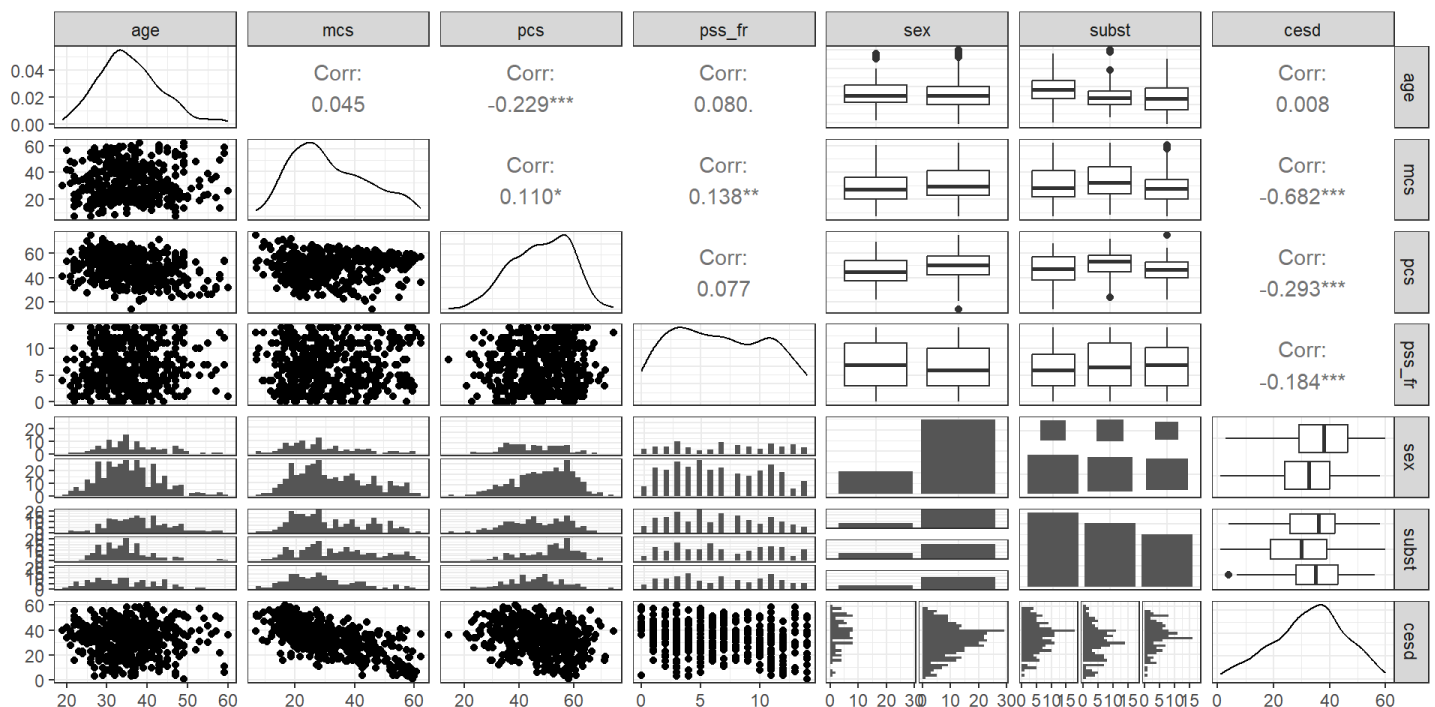
```
1 temp <- help1 |>
2   select(age, mcs, pcs, pss_fr, sex, subst, cesd)
3
4 ggpairs(temp) ## ggpairs from the GGally package
```

We place the outcome (**cesd**) last (result on next slide.)

## Saving the Data Set

```
1 write_rds(help1, "c04/data/help1.Rds")
```

# Scatterplot Matrix (result)



# Using `ols()` to fit a linear regression model

## Fitting using `ols()`

The `ols` function stands for ordinary least squares and comes from the `rms` package, by Frank Harrell and colleagues. Any model fit with `lm` can also be fit with `ols`.

- To predict `var_y` using `var_x` from the `my_tibble` data, we would use the following syntax:

```
1 dd <- datadist(my_tibble)
2 options(datadist = "dd")
3
4 model_name <- ols(var_y ~ var_x, data = my_tibble,
5                   x = TRUE, y = TRUE)
```

This leaves a few questions...

# What's the **datadist** stuff doing?

Before fitting an **ols** model to data from **my\_tibble**, use:

```
1 dd <- datadist(my_tibble)
2 options(datadist = "dd")
```

Run (the **datadist** code above) once before any models are fitted, storing the distribution summaries for all potential variables. Adjustment values are 0 for binary variables, the most frequent category (or optionally the first category level) for categorical (factor) variables, the middle level for ordered factor variables, and medians for continuous variables. (excerpt from **datadist** documentation)

## Why use **x = TRUE, y = TRUE**?

Once we've set up the summaries with **datadist**, we fit a model:

```
1 model_name <- ols(var_y ~ var_x, data = my_tibble,
2                   x = TRUE, y = TRUE)
```

- **ols** stores additional information beyond what **lm** does
- **x = TRUE** and **y = TRUE** save even more expanded information for building plots and summarizing fit.
- The defaults are **x = FALSE, y = FALSE**, but in 432, we'll want them saved.

# Using `ols` to fit a model

Let's try to predict our outcome (`cesd`) using `mcs` and `subst`

- Start with setting up the `datadist`
- Then fit the model, including `x = TRUE, y = TRUE`

```
1 dd <- datadist(help1)
2 options(datadist = "dd")
3
4 mod1 <- ols(cesd ~ mcs + subst, data = help1,
5             x = TRUE, y = TRUE)
```

## Contents of `mod1`?

```
1 mod1
```

Linear Regression Model

```
ols(formula = cesd ~ mcs + subst, data = help1, x = TRUE, y = TRUE)
```

		Model Likelihood		Discrimination	
		Ratio Test		Indexes	
Obs	453	LR chi2	295.10	R2	0.479
sigma	9.0657	d.f.	3	R2 adj	0.475
d.f.	449	Pr(> chi2)	0.0000	g	9.827

Residuals

	Min	1Q	Median	3Q	Max
	-25.43696	-6.74592	0.09334	6.16212	24.24842

# New elements in `ols`

For our `mod1`,

- Model Likelihood Ratio test output includes `LR chi2 = 295.10, d.f. = 3, Pr(> chi2) = 0.0000`

The log of the likelihood ratio, multiplied by -2, yields a test against a  $\chi^2$  distribution. Interpret this as a goodness-of-fit test that compares `mod1` to a null model with only an intercept term. In `ols` this is similar to a global (ANOVA) F test.

# New elements in `ols`

Under the  $R^2$  values, we have `g = 9.827`.

- This is the *g*-index, based on Gini's mean difference. If you randomly selected two of the subjects in the model, the average difference in predicted `cesd` will be 9.827.
- This can be compared to the Gini's mean difference for the original `cesd` values, from `describe`, which was `Gmd = 14.23`.

# Validate summaries from an `ols` fit

- The data used to fit the model provide an overly optimistic view of the quality of fit.
- We're interested here in assessing how well the model might work in new data, using a resampling approach.

## Validation Results

```
1 set.seed(432)
2 validate(mod1)
```

	index.orig	training	test	optimism	index.corrected	Lower	Upper
R-square	0.4787	0.4874	0.4737	0.0137	0.4650	0.3904	0.5302
MSE	81.4606	79.7851	82.2361	-2.4510	83.9116	75.1632	93.5270
g	9.8272	9.9133	9.8038	0.1095	9.7177	8.3704	10.7752
Intercept	0.0000	0.0000	0.2793	-0.2793	0.2793	-3.9317	5.1316
Slope	1.0000	1.0000	0.9894	0.0106	0.9894	0.8637	1.1075
n							
R-square	40						
MSE	40						
g	40						
Intercept	40						
Slope	40						

- `index.orig` for  $R^2$  is 0.4787. That's what we get from the data used to fit `mod1`.

# Resampling Validation for $R^2$

-	index.orig	training	test	optimism	index.corrected	n
$R^2$	0.4787	0.4874	0.4737	0.0137	0.4650	40

- With `validate` we create 40 (by default) bootstrapped resamples of the data and then split each of those into training and test samples.
  - For each of the 40 splits, R refits the model (same predictors) in the `training` sample to obtain  $R^2$ : mean across 40 splits is 0.4874
  - Check each model in its `test` sample: average  $R^2$  was 0.4737
- `optimism` = `training` result - `test` result = 0.0137
- `index.corrected` = `index.orig` - `optimism` = 0.4650

While our *nominal*  $R^2$  is 0.4787; correcting for optimism yields *validated*  $R^2$  of 0.4650, so we conclude that  $R^2 = 0.4650$  better estimates how `mod1` will perform in new data.

# Resampling Validation for MS(Error)

-	index.orig	training	test	optimism	index.corrected	n
MSE	81.4606	79.7851	82.2361	-2.4510	83.9116	40

- `index.orig` for MSE = 81.4606. That's what we get from the data used to fit `mod1`.
- For each of the 40 splits, R refits the model (same predictors) in the `training` sample to obtain MSE: mean across 40 splits is 79.7851
- Check each model in its `test` sample: average MSE was 82.2361
- `optimism` = `training` result - `test` result = -2.4510
- `index.corrected` = `index.orig` - `optimism` = 83.9116

While our *nominal* MSE is 81.4606 (so RMSE =  $\sqrt{81.4606} = 9.03$ ); correcting for optimism yields *validated* MSE of 83.9116 and validated RMSE =  $\sqrt{83.9116} = 9.16$ .

# ANOVA for `mod1` fit by `ols`

```
1 anova(mod1)
```

Analysis of Variance				Response: cesd	
Factor	d.f.	Partial SS	MS	F	P
mcs	1	31182.7237	31182.72373	379.42	<.0001
subst	2	968.7563	484.37816	5.89	0.003
REGRESSION	3	33886.8359	11295.61195	137.44	<.0001
ERROR	449	36901.6542	82.18631		

- This adds a line for the complete regression model (both terms) which can be helpful, but is otherwise the same as `anova()` after a fit using `lm()`.
- As with `lm`, this is a sequential ANOVA table, so if we had included `subst` in the model first, we'd get a different SS, MS, F and p for `mcs` and `subst`, but the same `REGRESSION` and `ERROR` results.

## summary for `mod1` fit by `ols`

```
1 summary(mod1, conf.int = 0.90)
```

Effects		Response : cesd				
Factor	Low	High	Diff.	Effect	S.E.	Lower 0.9
mcs	21.676	40.941	19.266	-12.6580	0.64984	-13.7290
subst - cocaine:alcohol	1.000	2.000	NA	-3.4440	1.00550	-5.1013
subst - heroin:alcohol	1.000	3.000	NA	-1.7791	1.06810	-3.5396

Upper 0.9  
-11.587000  
-1.786700  
-0.018654

- How do we interpret the `subst` effects estimated by this model?
  - Effect of `subst` being `cocaine` instead of `alcohol` on `ces_d` is -3.44 assuming no change in `mcs`, with 90% CI (-5.10, -1.79).
  - Effect of `subst` being `heroin` instead of `alcohol` on `ces_d` is -1.78 assuming no change in `mcs`, with 90% CI (-3.54, -0.02).

But what about the `mcs` effect?

# summary for `mod1` fit by `ols`

```
1 summary(mod1, conf.int = 0.90)
```

Effects	Response : cesd					
Factor	Low	High	Diff.	Effect	S.E.	Lower 0.9
mcs	21.676	40.941	19.266	-12.6580	0.64984	-13.7290
subst - cocaine:alcohol	1.000	2.000	NA	-3.4440	1.00550	-5.1013
subst - heroin:alcohol	1.000	3.000	NA	-1.7791	1.06810	-3.5396

Upper 0.9  
-11.587000  
-1.786700  
-0.018654

- Effect of `mcs`: -12.66 is the estimated change in `cesd` associated with a move from `mcs` = 21.68 (see `Low` value) to `mcs` = 40.94 (the `High` value) assuming no change in `subst`.
- `ols` chooses the `Low` and `High` values from the interquartile range.

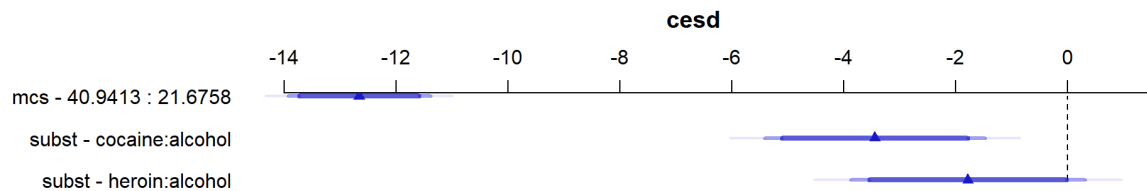
```
1 quantile(help1$mcs, c(0.25, 0.75))
```

```
25%      75%  
21.67575 40.94134
```

## Plot the summary to see effect sizes

- Goal: plot effect sizes for similar moves within predictor distributions.

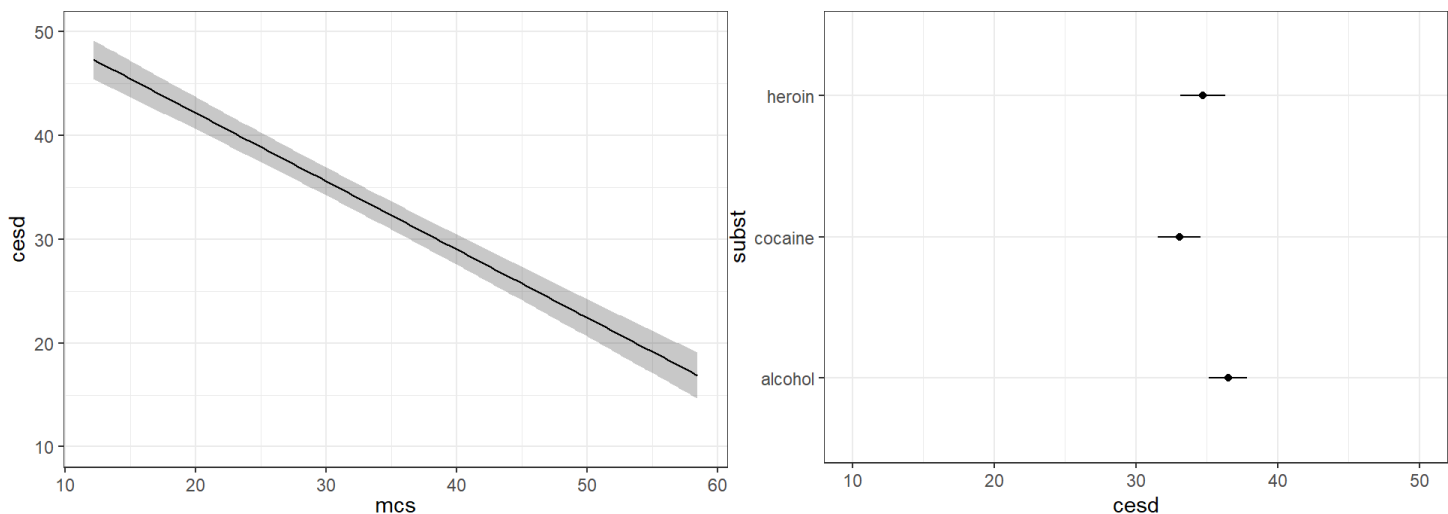
```
1 plot(summary(mod1))
```



- The triangles indicate the point estimate, augmented with confidence interval bars.
  - The 90% confidence intervals are plotted with the thickest bars.
  - The 95% CIs are then shown with thinner, more transparent bars.
  - Finally, the 99% CIs are shown as the longest, thinnest bars.

# Plot the individual effects?

```
1 ggplot(Predict(mod1, conf.int = 0.95), layout = c(1,2))
```

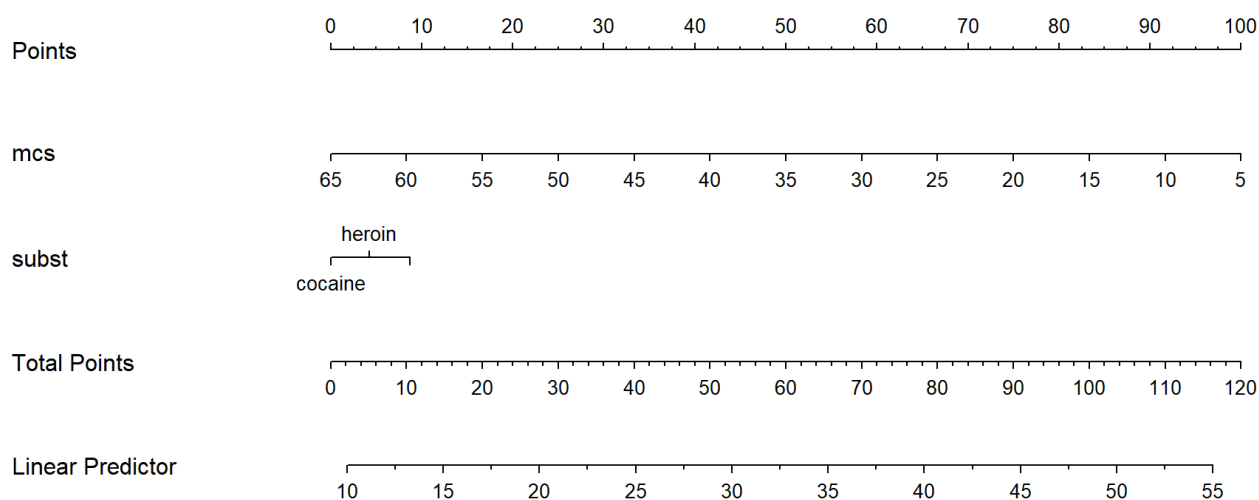


- At left, impact of changing `mcs` on `cesd` holding `subst` at its baseline (alcohol).
- At right, impact of changing `subst` on `cesd` holding `mcs` at its median (28.602417).
- Defaults: add 95% CI bands and layout tries for a square.

# Standing Break

# Build a nomogram for the `ols` fit

```
1 plot(nomogram(mod1))
```



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## Nomograms

For complex models (this model isn't actually very complex) it can be helpful to have a tool that will help you see the modeled effects in terms of their impact on the predicted outcome.

A *nomogram* is an established graphical tool for doing this.

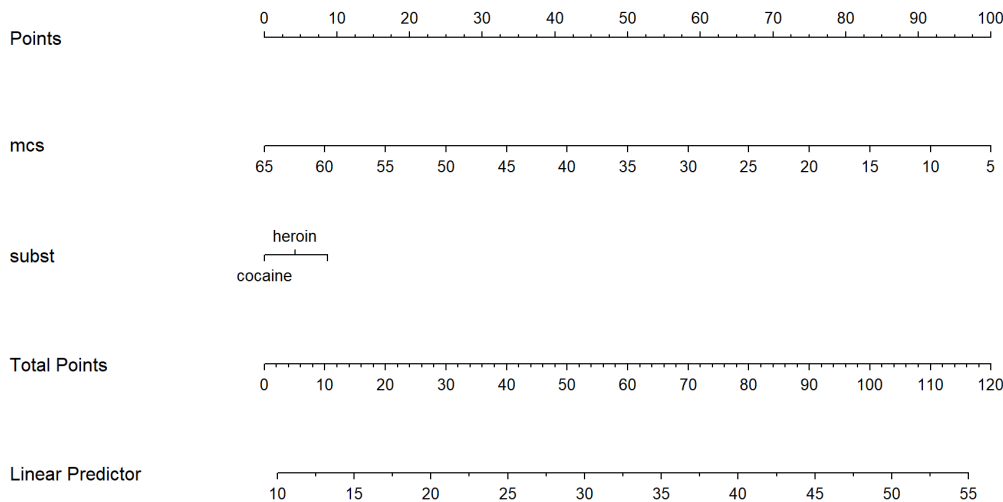
- Find the value of each predictor on its provided line, and identify the “points” for that predictor by drawing a vertical line up to the “Points”.
- Then sum up the points over all predictors to obtain “Total Points”.
- Draw a vertical line down from the “Total Points” to the “Linear Predictor” to get the predicted `cesd` for this subject.

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# Using the nomogram for **mod1**

Predicted **cesd** if **mcs** = 35 and **subst** = heroin?



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## Actual Prediction for this subject...

- The **predict** function for our **ols** fit provides fitted values.

```
1 predict(mod1, newdata = tibble(mcs = 35, subst = "heroin"))
```

1

30.52766

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# Using `lrm()` to fit a logistic regression model

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## Fitting using `lrm()`

The `lrm()` function stands for logistic regression model and also comes from the `rms` package. Let's predict our binary outcome (`cesd_hi`) using `mcs` and `subst`.

- Start with setting up the `datadist` Then fit model, including `x = TRUE, y = TRUE`

```
1 dd <- datadist(help1)
2 options(datadist = "dd")
3
4 mod2 <- lrm(cesd_hi ~ mcs + subst, data = help1, x = TRUE, y = TRUE)
```

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# Contents of `mod2`?

1 `mod2`

Logistic Regression Model

```
lrm(formula = cesd_hi ~ mcs + subst, data = help1, x = TRUE,  
     y = TRUE)
```

		Model	Likelihood	Discrimination	Rank	Discrim.	
			Ratio Test	Indexes		Indexes	
Obs	453	LR chi2	134.24	R2	0.533	C	0.938
0	46	d.f.	3	R2(3,453)	0.252	Dxy	0.875
1	407	Pr(> chi2)	<0.0001	R2(3,124)	0.653	gamma	0.875
max  deriv	6e-06			Brier	0.056	tau-a	0.160

	Coef	S.E.	Wald Z	Pr(> Z )
Intercept	10.5778	1.2429	8.51	<0.0001

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## New elements in `lrm`

For our `mod2`,

- Model Likelihood Ratio test output includes `LR chi2 = 134.24, d.f. = 3, Pr(> chi2) <0.0001`

Again, the log of the likelihood ratio, multiplied by -2, yields a test against a  $\chi^2$  distribution. Interpret this as a goodness-of-fit test that compares `mod2` to a null model with only an intercept term.

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# Discrimination Indexes in `lrm()`

`R2 = 0.533`, `R2(3,453) = 0.252`, `R2(3,124) = 0.653`,  
`Brier = 0.056`

The `R2` value is the *Nagelkerke*  $R^2$ , which is another pseudo- $R^2$  measure that provides a correction to the Cox-Snell  $R^2$  so that the maximum value is 1.

- Other  $R^2$ s [are detailed here](#)

# Discrimination Indexes in `lrm()`

`R2 = 0.533`, `R2(3,453) = 0.252`, `R2(3,124) = 0.653`,  
`Brier = 0.056`

The `Brier` score is the mean squared error between predictions and actual (1/0) observations. The lower the score (closer to 0), the better the model's predictions are calibrated. It's not really useful on its own, but helps when comparing models.

# Rank Discrimination Indexes in `lrm()`

$C = 0.938$ ,  $D_{xy} = 0.875$ ,  $\gamma = 0.875$ ,  $\tau\text{-}a = 0.160$

- $C$  is the  $C$  statistic, the area under the ROC curve
- $D_{xy}$  is Somers'  $d$ , and note that  $C = 0.5 + (D_{xy}/2)$
- $\gamma$  is the Goodman-Kruskal  $\gamma$  statistic
- $\tau\text{-}a$  is the Kendall  $\tau$  statistic (version a)

## Validate summaries from an `lrm` fit

- Can we validate summary statistics by resampling?

```
1 set.seed(432432)
2 validate(mod2)
```

	index.orig	training	test	optimism	index.corrected	Lower	Upper	n
Dxy	0.8751	0.8825	0.8707	0.0118	0.8634	0.8139	0.9271	40
R2	0.5326	0.5421	0.5247	0.0174	0.5152	0.4313	0.6253	40
Intercept	0.0000	0.0000	0.0069	-0.0069	0.0069	-0.5537	0.6074	40
Slope	1.0000	1.0000	0.9619	0.0381	0.9619	0.6776	1.2723	40
E <sub>max</sub>	0.0000	0.0000	0.0582	-0.0582	0.0582	-0.0216	0.1823	40
D	0.2941	0.2988	0.2891	0.0097	0.2844	0.2113	0.3697	40
U	-0.0044	-0.0044	0.0001	-0.0045	0.0001	-0.0079	0.0163	40
Q	0.2985	0.3032	0.2890	0.0142	0.2843	0.2048	0.3666	40
B	0.0560	0.0548	0.0571	-0.0022	0.0583	0.0419	0.0736	40
g	2.7444	2.8543	2.7041	0.1502	2.5942	1.9223	3.2205	40
gp	0.1577	0.1573	0.1569	0.0004	0.1574	0.1197	0.1921	40

# Resampling Validation after `lrm()`

	– index.orig	training	test	optimism	index.corrected	n
Dxy	0.8751	0.8825	0.8707	0.0118	0.8634	40
R2	0.5326	0.5421	0.5247	0.0174	0.5152	40

- Dxy = Somers' d, and the area under the ROC curve is  $C = 0.5 + (Dxy/2)$
- Our original Dxy = 0.8751, implying  $C = 0.9376$
- Our validated Dxy = 0.8634, so validated  $C = 0.5 + (0.8634/2) = 0.9317$
- While our *nominal*  $R^2$  is 0.5326; correcting for optimism yields *validated*  $R^2$  of 0.5152.

## ANOVA for `mod2` fit by `lrm`

```
1 anova(mod2)
```

Wald Statistics

Response: cesd\_hi

Factor	Chi-Square	d.f.	P
mcs	58.43	1	<.0001
subst	10.04	2	0.0066
TOTAL	62.30	3	<.0001

- Again, this is a sequential ANOVA table, so if we had included `subst` in the model first, we'd get a different Chi-Square, and p for `mcs` and `subst`, but the same **TOTAL** result.

# summary for `mod2` fit by `lrm`

```
1 summary(mod2, conf.int = 0.90)
```

Effects	Response : cesd_hi					
Factor	Low	High	Diff.	Effect	S.E.	Lower 0.9
mcs	21.676	40.941	19.266	-3.460400	0.45270	-4.20500
Odds Ratio	21.676	40.941	19.266	0.031417	NA	0.01492
subst - cocaine:alcohol	1.000	2.000	NA	-1.502500	0.48114	-2.29390
Odds Ratio	1.000	2.000	NA	0.222580	NA	0.10087
subst - heroin:alcohol	1.000	3.000	NA	-1.269500	0.59788	-2.25290
Odds Ratio	1.000	3.000	NA	0.280980	NA	0.10509
Upper 0.9						
	-2.715800					
	0.066152					
	-0.711070					
	0.491120					

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# summary for `mod2` fit by `lrm`

Factor	Low	High	Diff.	Effect	S.E.	Lower 0.9	Upper 0.9
mcs	21.676	40.941	19.266	-3.46040	0.4527	-4.2050	-2.71580
Odds Ratio	21.676	40.941	19.266	0.03142	NA	0.0149	0.06615

- Odds of `cesd_hi` are 0.03 times as high for a subject with `mcs` = 40.94 (**High**) as compared to a subject with `mcs` = 21.68 (**Low**) assuming no change in `subst`.
- `ols` chooses the **Low** and **High** values from the interquartile range.

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# summary for `mod2` fit by `lrm`

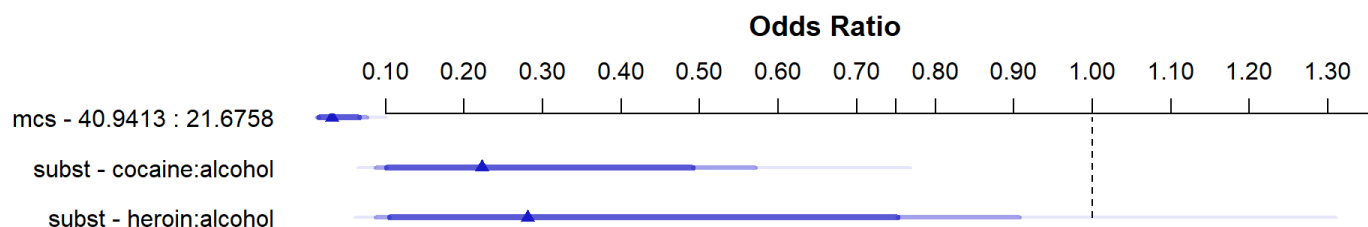
Factor	Low	High	Diff	Effect	S.E.	Lower 0.9	Upper 0.9
subst - cocaine:alcohol	1	2	NA	-1.5025	0.4811	-2.2939	-0.71107
Odds Ratio	1	2	NA	0.2226	NA	0.1009	0.49112
subst - heroin:alcohol	1	3	NA	-1.2695	0.5979	-2.2529	-0.28607
Odds Ratio	1	3	NA	0.2810	NA	0.1051	0.75121

- Effect of `subst` being `cocaine` instead of `alcohol` on `cesd_hi` is an Odds Ratio of 0.22 (0.10, 0.49), assuming no change in `mcs`.
- Effect of `subst` being `heroin` instead of `alcohol` on `cesd_hi` is an Odds Ratio of 0.28 (0.11, 0.75), assuming no change in `mcs`.

## Plot the summary to see effect sizes

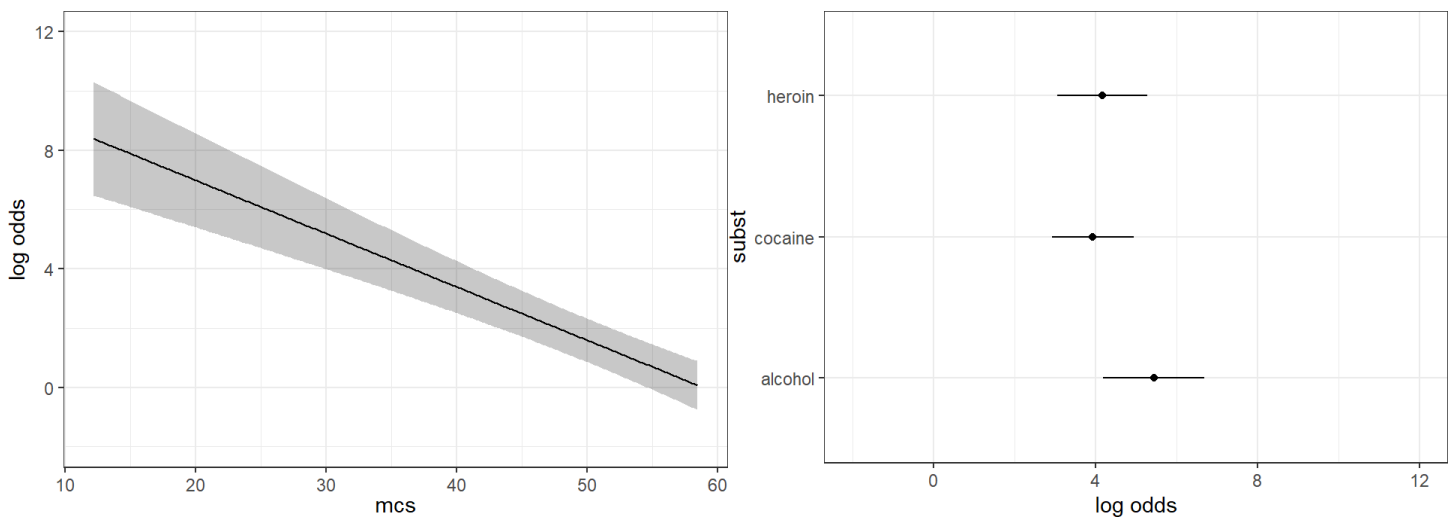
- Goal: plot effect sizes for similar moves within predictor distributions.

```
1 plot(summary(mod2))
```



# Plot the individual effects?

```
1 ggplot(Predict(mod2, conf.int = 0.95), layout = c(1,2))
```



- At left, impact of changing `mcs` on `cesd` holding `subst` at its baseline (alcohol).
- At right, impact of changing `subst` on `cesd` holding `mcs` at its median (28.602417).
- Defaults: add 95% CI bands and layout tries for a square.

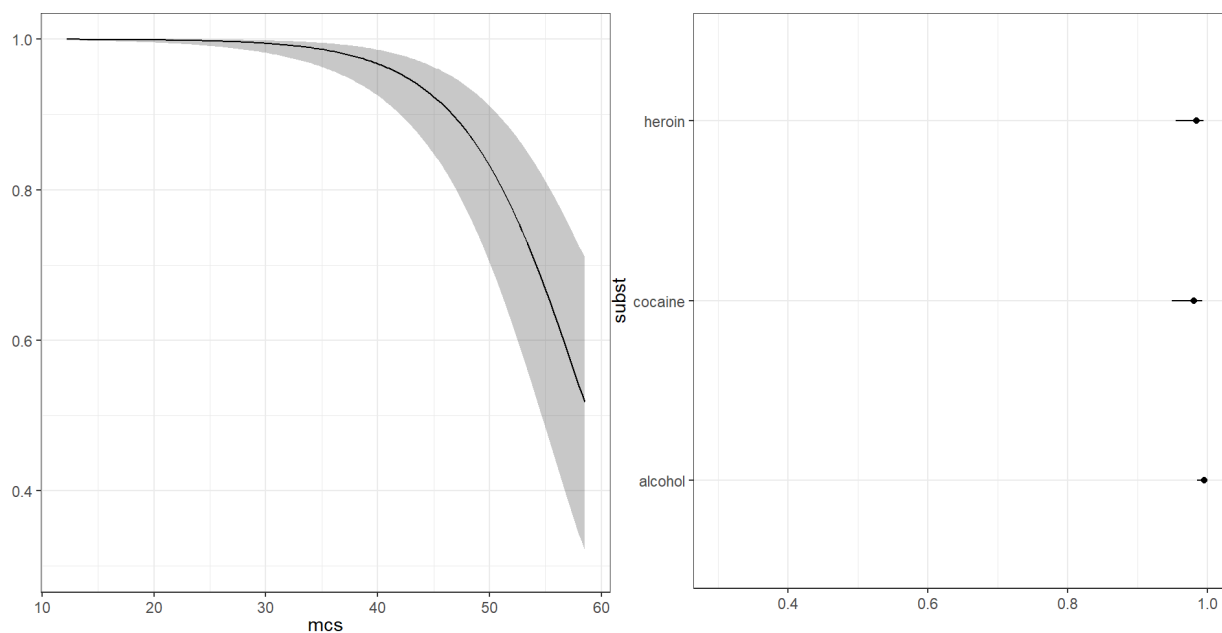
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# Plot on probability scale?

Add `fun = plogis`.

```
1 ggplot(Predict(mod2, conf.int = 0.95, fun = plogis), layout = c(1,2))
```

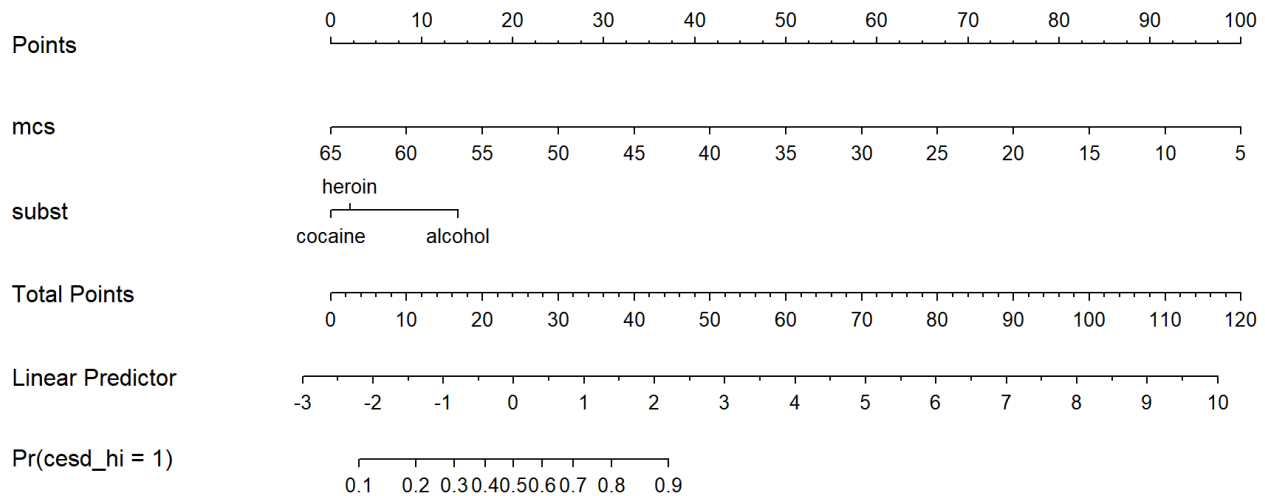


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# Build a nomogram for the `ols` fit

```
1 plot(nomogram(mod2, fun = plogis, funlabel = 'Pr(cesd_hi = 1)'))
```



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## Making a Prediction...

- The `predict` function for our `lrm()` fit provides fitted values, either on the log odds scale...

```
1 predict(mod2, newdata = tibble(mcs = 35, subst = "heroin"), type = "lp")
```

1

3.021763

- or on the probability scale ...

```
1 predict(mod2, newdata = tibble(mcs = 35, subst = "heroin"), type = "fitted")
```

1

0.9535477

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# Getting more good stuff

- Anything you can fit with `ols()` can also be fit with `lm()`, so you have access to everything in `lm()` as well, like `check_model()`, etc.
- Same goes for `glm(..., family = binomial(link = "logit"))` and `lrm()`.

## Coming Soon

- Fitting more complex linear and logistic regression models
  - Adding non-linearity in the predictors through interactions, polynomials and splines
  - Spending degrees of freedom and the Spearman  $\rho^2$  plot