# Package 'STged'

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Title STged: Gene expression deconvolution for spatial transcriptomic data

Type Package

**Description** 

Version 0.1.0			
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<b>Description</b> STged is a gene expression deconvolution mathod. STged employs graph-guided spatial correlations and prior-guided gene expression similarities within a non-negative least-squares regression framework.			
<b>Depends</b> R (>= $4.2.1$ )			
Imports MASS, Matrix, nnls, reticulate			
Suggests knitr, rmarkdown			
VignetteBuilder knitr			
License GPL(>= 2)			
Encoding UTF-8			
LazyData true			
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This function focuses on cleaning scRNA-seq and SRT datasets.

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## Usage

```
create_group_exp(sc_exp, sc_label)
```

#### **Arguments**

sc\_exp scRNA-seq matrix, genes \* cells. The format should be raw-counts. The matrix

need include gene names and cell names.

sc\_label cell type information. The cells are need be divided into multiple category.

# **Examples**

```
data("PDAC")
refmu = create_group_exp(sc_exp,sc_label)
```

data\_process

This function focuses on cleaning scRNA-seq and SRT datasets.

#### **Description**

This function focuses on cleaning scRNA-seq and SRT datasets.

#### Usage

```
data_process(
    sc_exp,
    sc_label,
    spot_exp,
    spot_loc,
    gene_det_in_min_cells_per = 0.01,
    expression_threshold = 0,
    nUMI = 100,
    verbose = FALSE,
    depthscale = 1,
    clean.only = FALSE
)
```

cess.

# Arguments

sc_exp	scRNA-seq matrix, genes * cells. The format should be raw-counts. The matrix need include gene names and cell names.		
sc_label	cell type information. The cells are need be divided into multiple category.		
spot_exp	SRT gene expression matrix, genes * spots. The format should be raw counts. The matrix need include gene names and spot names.		
spot_loc	coordinate matrix, spots $*$ coordinates. The matrix need include spot names and coordinate name $(x, y)$ .		
gene_det_in_min_cells_per			
	a floor variable. minimum percent of genes that need to be detected in a cell.		
expression_threshold			
	a floor variable. Threshold to consider a gene expressed.		
nUMI	a floor variable. minimum of read count that need to be detected in a cell or spot.		
verbose	a logical variable that defines whether to print the processing flow of data pro-		

dis\_weight 3

#### Value

a list includes processed scRNA-seq matrix, cell type, stRNA-seq matrix.

### **Examples**

```
data("PDAC")
datax = data_process(sc_exp = sc_exp, sc_label = sc_label, spot_exp = spot_exp, spot_loc = spot_loc)
```

dis\_weight

This function focuses on cleaning scRNA-seq and SRT datasets.

### **Description**

This function focuses on cleaning scRNA-seq and SRT datasets.

# Usage

```
dis_weight(
   spot_loc = spot_loc,
   spot_exp = spot_exp,
   k = 6,
   quantile_prob_bandwidth = 1/3,
   method = "Hex",
   coord_type = "grid"
)
```

# Arguments

coord\_type

spot\_loc coordinate matrix, spots \* coordinates. The matrix need include spot names and coordinate name (x, y).

spot\_exp stRNA-seq matrix, genes \* spots. The format should be raw-counts. The matrix need include gene names and spot names.

k number of neighbor spots for construct spatial neighboring graph, details refer to Squidpy.

quantile\_prob\_bandwidth selection of bandwidth of the spatial kernel of each spot.

method the method used to construct spatial neighboring graph, details refer to Squidpy.

grid coordinates, details refer to Squidpy.

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**PDAC** 

Input data of the STged

### **Description**

Input data of the STged

#### Usage

data(PDAC)

#### **Format**

a list of matrices contains SRT gene expression matrix with coordinates information and corresponding cell type proportion information, and annotated scRNA-seq data as cell type reference (bottom).

# **Examples**

data(PDAC)

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STged: Gene expression deconvolution for spatial transcriptomic data

### Description

STged is a gene expression deconvolution mathod. STged employs graph-guided spatial correlations and prior-guided gene expression similarities within a non-negative least-squares regression framework.

# Usage

```
model.est = STged(sc_exp = sc_exp, sc_label = sc_label, spot_exp = spot_exp, spot_loc = spot_loc
```

# Arguments

sc_exp	scRNA-seq matrix, genes * cells. The format should be raw-counts. The matrix need include gene names and cell names.		
sc_label	cell type information. The cell need be divided into multiple categories.		
spot_exp	stRNA-seq matrix, genes * spots. The format should be raw-counts. The matrix need include gene names and spot names.		
spot_loc	coordinate matrix, spots $\ast$ coordinates. The matrix need include spot names and coordinate name $(x,y)$ .		
beta	cell type proportion matrix, spots * cell types.		
<pre>gene_det_in_min_cells_per</pre>			
	a floor variable. minimum percent # of genes that need to be detected in a cell.		
expression_threshold			
	a floor variable. Threshold to consider a gene expressed.		

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nUMI a floor variable. minimum # of read count that need to be detected in a cell or

spot.

verbose a logical variable that defines whether to print the processing flow of data pro-

ess.

clean.only a logical variable that defines whether to normalize data with log transform.

python\_env the path of python environment.

truncate a logical variable, that defines whether to truncate the gene expression data for

both scRNA-seq and SRT.

qt Winsorize expression values to prevent outliers. Values below this quantile and

above 1-this quantile will be set to the quantile value.

knei number of neighbor spots for construct spatial neighboring graph, details refer

to Squidpy.

methodL the method used to construct spatial neighboring graph, details refer to Squidpy.

coord\_type grid coordinates, details refer to Squidpy.

quantile\_prob\_bandwidth

selection of bandwidth of the spatial kernel of each spot.

lambda1 tuning parameter to balance the graph regularization term. If the tuning param-

eter is set to NULL, then, we will adopt the value in our algorithm.

lambda2 tuning parameter to balance the prior regularization term. If the tuning parame-

ter is set to NULL, then, we will adopt the value in our algorithm.

cutoff a cutoff value of cell type proportion.

rho the penalty parameter in the ADMM algorithm.

rho.incr the increase step parameter for varying penalty parameter rho.

rho.max the maximum value of rho.

maxiter a positive integer represents the maximum number of updating algorithm. De-

fault setting is 100.

epsilon a parameter represents the stop criterion.

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