

Ben_Linear_Regression

TJ

10/14/2021

Libraries

First we need to load in the data

```
#Summer data
top100phyvSum <- read.csv("data/NewSumWinData/POWOW_RawCounts_EcotypePhyla_Summer.csv") %>%
  column_to_rownames(., var = "Sample") %>%
  subset(., select = eMED4:Verrucomicrobia)
top100phyvSum.hell <- decostand(top100phyvSum, "hellinger") #Preforming a hellinger transfermation

#Winter data
top100phyvWin <- read.csv("data/NewSumWinData/POWOW_RawCounts_EcotypePhyla_Winter.csv") %>%
  column_to_rownames(., var = "Sample") %>%
  subset(., select = eMED4:Verrucomicrobia)
top100phyvWin.hell <- decostand(top100phyvWin, "hellinger") #Preforming a hellinger transfermation
```

Summer analysis

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9312lm_Sum_Intersept <- lm(eMIT9312 ~ 1, data = top100phyvSum.hell) #First we will define the intercept
eMIT9312lm_Sum <- lm(eMIT9312 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will fit the model
```

```
vif(eMIT9312lm_Sum)
```

##	Chloroflexi	Cyanobacteria
##	9.439458	3.903344
##	Deinococcus.Thermus	Euryarchaeota
##	1.639400	8.018580
##	Firmicutes	Fusobacteria
##	4.741412	1.609413
##	Gemmatimonadetes	Gracilibacteria

```

##          5.381990      2.023682
##Lentisphaerae Marinimicrobia_SAR406_clade
##          2.010246      8.027217
##PAUC34f           Planctomycetes
##          3.398654      2.102418
##Proteobacteria     SHA.109
##          6.057502      1.313680
##Thaumarchaeota    Verrucomicrobia
##          10.288416     2.990001

```

```
eMIT9312lm_Sum_both <- step(eMIT9312lm_Sum_Intersept, direction='both', scope=formula(eMIT9312lm_Sum), ...)
```

```
eMIT9312lm_Sum_both$anova
```

	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA	51	2.394479	-158.0601
## 2	+ Proteobacteria	-1	0.40757846	50	1.986901	-165.7627
## 3	+ Deinococcus.Thermus	-1	0.14579905	49	1.841102	-167.7257
## 4	+ Planctomycetes	-1	0.09743226	48	1.743669	-168.5531

```
eMIT9312lm_Sum_both$coefficients
```

	(Intercept)	Proteobacteria	Deinococcus.Thermus	Planctomycetes
##	0.3153202	-0.3806562	10.4842631	-0.6532889

```
summary(eMIT9312lm_Sum_both)
```

```

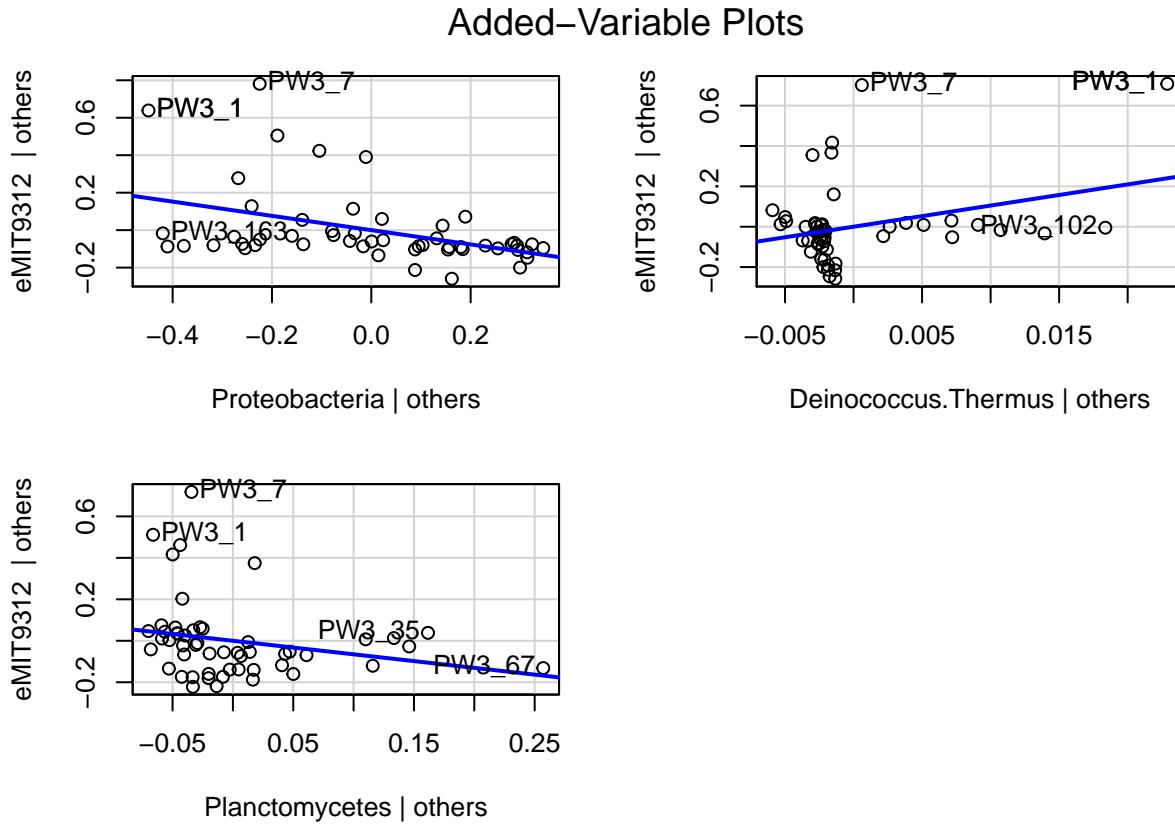
##
## Call:
## lm(formula = eMIT9312 ~ Proteobacteria + Deinococcus.Thermus +
##     Planctomycetes, data = top100phyvSum.hell)
##
## Residuals:
##       Min        1Q        Median        3Q        Max
## -0.24368 -0.10140 -0.03093  0.03618  0.69555
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 0.31532   0.06936  4.546 3.71e-05 ***
## Proteobacteria -0.38066   0.11760 -3.237 0.00219 **
## Deinococcus.Thermus 10.48426   4.59146  2.283 0.02687 *
## Planctomycetes -0.65329   0.39890 -1.638 0.10802
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1906 on 48 degrees of freedom
## Multiple R-squared:  0.2718, Adjusted R-squared:  0.2263
## F-statistic: 5.972 on 3 and 48 DF,  p-value: 0.001521

```

When comparing eMIT9312 against the Phylum in the summer samples, we found this ecotype has significant relationships with Proteobacteria ($p < .005$) and Deinococcus-Thermus ($p < .05$). Specifically we found a

0.4% decrease (± 0.11) in eMIT9312 abundance for every 1% increase in Proteobacteria and a 10% increase (± 4.6) in eMIT9312 abundance for every 1% increase in Deinococcus-Thermus abundance. Though we saw a decrease of .66 in eMIT9312 for every 1% increase in Planctomycetes, this decrease was not statistically significant ($p > .05$).

```
avPlots(eMIT9312lm_Sum_both)
```



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eNATL2Alm_Sum_Intersept <- lm(eNATL2A ~ 1, data = top100phyySum.hell) #First we will define the intercept
eNATL2Alm_Sum <- lm(eNATL2A ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyySum.hell) #Now I will add the variables
```

```
#Preforming a stepwise in both directions to find the best model
```

```
eNATL2Alm_Sum_both <- step(eNATL2Alm_Sum_Intersept, direction='both', scope=formula(eNATL2Alm_Sum), trace=TRUE)
```

```
eNATL2Alm_Sum_both$anova
```

##	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
##	1	NA	NA	51	2.742193	-151.0093

```

eNATL2Alm_Sum_both$coefficients

## (Intercept)
##      0.152451

summary(eNATL2Alm_Sum_both)

##
## Call:
## lm(formula = eNATL2A ~ 1, data = top100phyvSum.hell)
##
## Residuals:
##     Min      1Q  Median      3Q     Max 
## -0.1525 -0.1502 -0.1426  0.1080  0.6204 
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)    
## (Intercept) 0.15245   0.03216   4.741 1.75e-05 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2319 on 51 degrees of freedom

```

There is no sig correlation for eNATL2A

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```

eMIT9313lm_Sum_Intersept <- lm(eMIT9313 ~ 1, data = top100phyvSum.hell) #First we will define the intercept
eMIT9313lm_Sum <- lm(eMIT9313 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will add the other phyla

```

#Preforming a stepwise in both directions to find the best model

```

eMIT9313lm_Sum_both <- step(eMIT9313lm_Sum_Intersept, direction='both', scope=formula(eMIT9313lm_Sum), trace=1)

```

```
eMIT9313lm_Sum_both$anova
```

	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA	51	0.06073271	-349.1309
## 2 + Chloroflexi	-1	0.00295798		50	0.05777473	-349.7273

```
eMIT9313lm_Sum_both$coefficients
```

```

## (Intercept) Chloroflexi
##      0.01764023 -0.22909016

```

```

summary(eMIT9313lm_Sum_both)

##
## Call:
## lm(formula = eMIT9313 ~ Chloroflexi, data = top100phyvSum.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.017640 -0.016232 -0.009533 -0.001025  0.150683
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 0.017640  0.005875  3.003  0.00417 **
## Chloroflexi -0.229090  0.143183 -1.600  0.11590
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.03399 on 50 degrees of freedom
## Multiple R-squared:  0.0487, Adjusted R-squared:  0.02968
## F-statistic:  2.56 on 1 and 50 DF,  p-value: 0.1159

```

Only the intercept is sig here

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```

eMED4lm_Sum_Intersept <- lm(eMED4 ~ 1, data = top100phyvSum.hell) #First we will define the intersept
eMED4lm_Sum <- lm(eMED4 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will

#Preforming a stepwise in both directions to find the best model
eMED4lm_Sum_both <- step(eMED4lm_Sum_Intersept, direction='both', scope=formula(eMED4lm_Sum), trace=0)

```

```
eMED4lm_Sum_both$anova
```

	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA	51	6.705967	-104.5088
## 2	+ Proteobacteria	-1	3.6507605	50	3.055207	-143.3886
## 3	+ Deinococcus.Thermus	-1	0.3633326	49	2.691874	-147.9723
## 4	+ Euryarchaeota	-1	0.1121722	48	2.579702	-148.1856

```
eMED4lm_Sum_both$coefficients
```

	(Intercept)	Proteobacteria	Deinococcus.Thermus	Euryarchaeota
##	1.0359735	-1.1104049	-12.2798426	-0.3903005

```

summary(eMED4lm_Sum_both)

##
## Call:
## lm(formula = eMED4 ~ Proteobacteria + Deinococcus.Thermus + Euryarchaeota,
##      data = top100phyvSum.hell)
##
## Residuals:
##       Min     1Q   Median     3Q    Max 
## -0.59850 -0.13013  0.03972  0.18986  0.35708
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)    
## (Intercept)  1.03597   0.08509 12.176 2.75e-16 ***
## Proteobacteria -1.11040   0.14299 -7.766 4.98e-10 ***
## Deinococcus.Thermus -12.27984   5.64451 -2.176   0.0345 *  
## Euryarchaeota   -0.39030   0.27016 -1.445   0.1550    
## ---    
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2318 on 48 degrees of freedom
## Multiple R-squared:  0.6153, Adjusted R-squared:  0.5913 
## F-statistic: 25.59 on 3 and 48 DF,  p-value: 4.917e-10

```

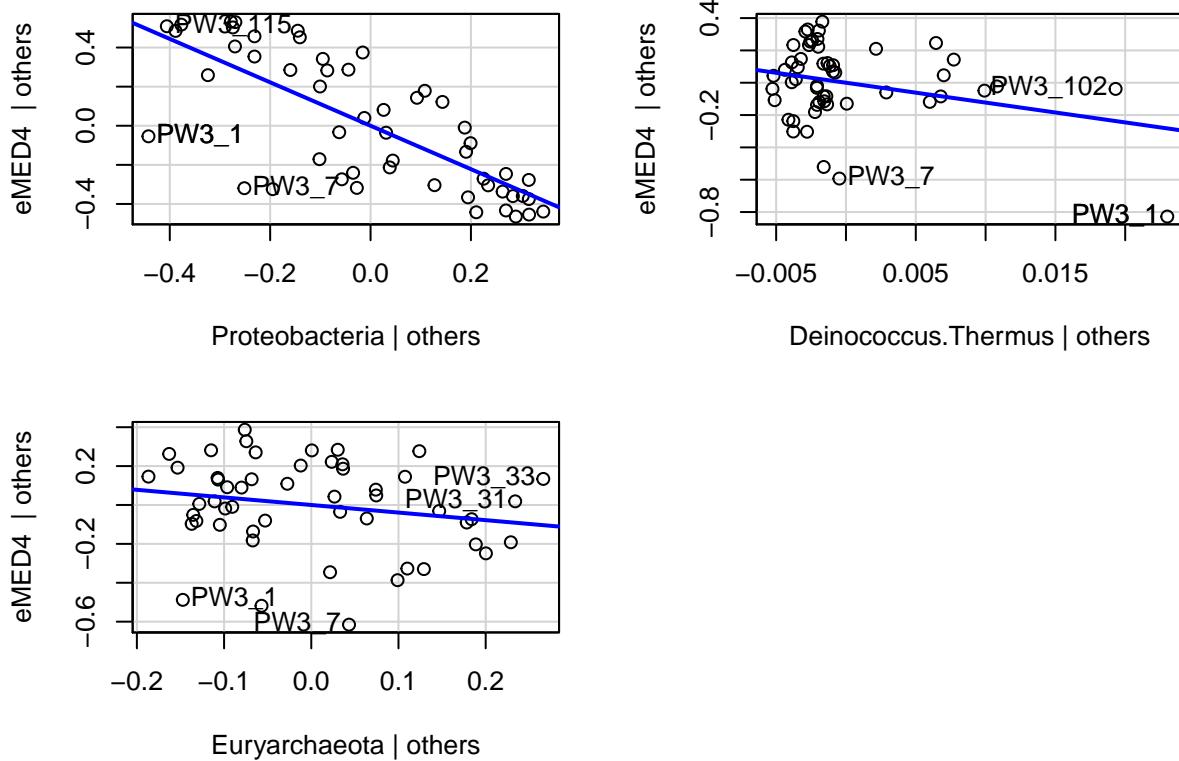
When comparing eMED4 against the Phylum in the summary samples, we found this ecotype has significant relationships with Proteobacteria ($p < .001$) and Deinococcus-Thermus ($p < .05$). Specifically we found a 1.1% decrease (± 0.14) in eMED4 abundance for every 1% increase in Proteobacteria and a 12% decrease (± 5.6) in eMED4 abundance for every 1% increase in Deinococcus-Thermus abundance. Though we saw a decrease of .4% in eMED4 for every 1% increase in Euryarchaeota, this decrease was not statistically significant ($p > .05$).

```

avPlots(eMED4lm_Sum_both)

```

Added-Variable Plots



Winter analysis

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9312lm_Win_Intercept <- lm(eMIT9312 ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eMIT9312lm_Win <- lm(eMIT9312 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin.hell)
```

```
vif(eMIT9312lm_Win)
```

##	Chloroflexi	Cyanobacteria
##	7.281146	4.581259
##	Deinococcus.Thermus	Euryarchaeota
##	3.314307	15.599366
##	Firmicutes	Fusobacteria
##	9.626650	1.884679
##	Gemmatimonadetes	Gracilibacteria
##	3.660177	4.701867
##	Lentisphaerae	Marinimicrobia_SAR406_clade
##	3.977338	26.827255

```

##          PAUC34f           Planctomycetes
##          4.937170          3.616936
##          Proteobacteria      SHA.109
##          5.094804          3.066283
##          Thaumarchaeota     Verrucomicrobia
##          4.332598          8.003607

eMIT9312lm_Win_both <- step(eMIT9312lm_Win_Intersept, direction='both', scope=formula(eMIT9312lm_Win), ...)

eMIT9312lm_Win_both$anova

##          Step Df Deviance Resid. Df Resid. Dev      AIC
## 1             NA      NA      42  5.222196 -88.65613
## 2 + Proteobacteria -1 2.622288      41  2.599908 -116.64614
## 3   + Chloroflexi  -1 0.438386      40  2.161522 -122.58667

eMIT9312lm_Win_both$coefficients

##      (Intercept) Proteobacteria  Chloroflexi
##      0.9885717    -1.2591248    -2.2176446

summary(eMIT9312lm_Win_both)

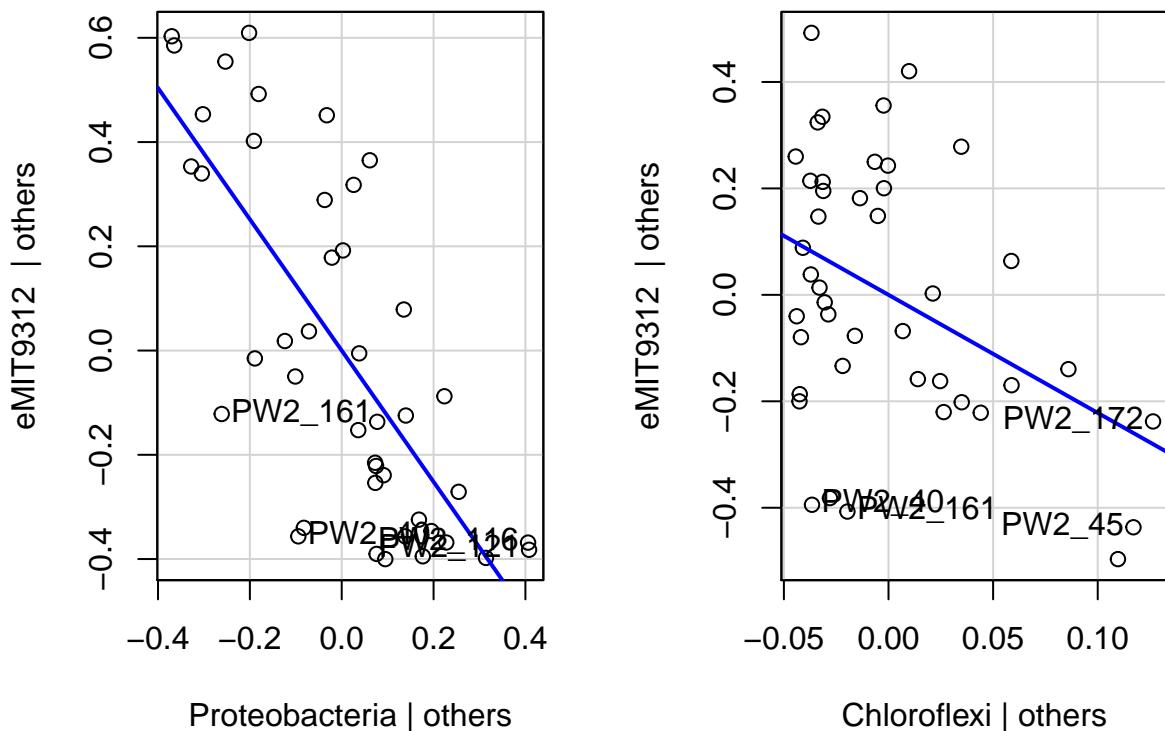
## 
## Call:
## lm(formula = eMIT9312 ~ Proteobacteria + Chloroflexi, data = top100phyvWin.hell)
## 
## Residuals:
##      Min       1Q       Median      3Q      Max
## -0.47548 -0.13240 -0.03934  0.15643  0.44203
## 
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.98857    0.09644 10.251 9.40e-13 ***
## Proteobacteria -1.25912    0.17868 -7.047 1.61e-08 ***
## Chloroflexi    -2.21764    0.77860 -2.848  0.00691 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## 
## Residual standard error: 0.2325 on 40 degrees of freedom
## Multiple R-squared:  0.5861, Adjusted R-squared:  0.5654
## F-statistic: 28.32 on 2 and 40 DF,  p-value: 2.178e-08

```

When comparing eMIT9312 against the Phylum in the winter samples, we found this ecotype has significant relationships with Proteobacteria ($p < .001$) and Chloroflexi ($p < .001$). Specifically we found a 1% decrease (± 0.2) in eMIT9312 abundance for every 1% increase in Proteobacteria and a 2% decrease ($\pm .8$) in eMIT9312 abundance for every 1% increase in Chloroflexi abundance.

```
avPlots(eMIT9312lm_Win_both)
```

Added-Variable Plots



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eNATL2Alm_Win_Intercept <- lm(eNATL2A ~ 1, data = top100phyyWin.hell) #First we will define the intercept
eNATL2Alm_Win <- lm(eNATL2A ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyyWin.hell)
```

#Preforming a stepwise in both directions to find the best model

```
eNATL2Alm_Win_both <- step(eNATL2Alm_Win_Intercept, direction='both', scope=formula(eNATL2Alm_Win), trace=TRUE)
```

```
eNATL2Alm_Win_both$anova
```

##	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA	42	1.0863957	-156.1684
## 2	+ Lentisphaerae	-1	0.20703655	41	0.8793592	-163.2598
## 3	+ SHA.109	-1	0.08435733	40	0.7950018	-165.5963
## 4	+ Euryarchaeota	-1	0.14629648	39	0.6487054	-172.3410
## 5	+ Gemmatimonadetes	-1	0.02998499	38	0.6187204	-172.3760
## 6	+ Chloroflexi	-1	0.06308949	37	0.5556309	-175.0006

```

eNATL2Alm_Win_both$coefficients

##          (Intercept)    Lentisphaerae      SHA.109   Euryarchaeota
##          0.1238786     3.4909464     -14.5247285     0.4720197
## Gemmatimonadetes    Chloroflexi
##          -6.7064609     1.4658046

summary(eNATL2Alm_Win_both)

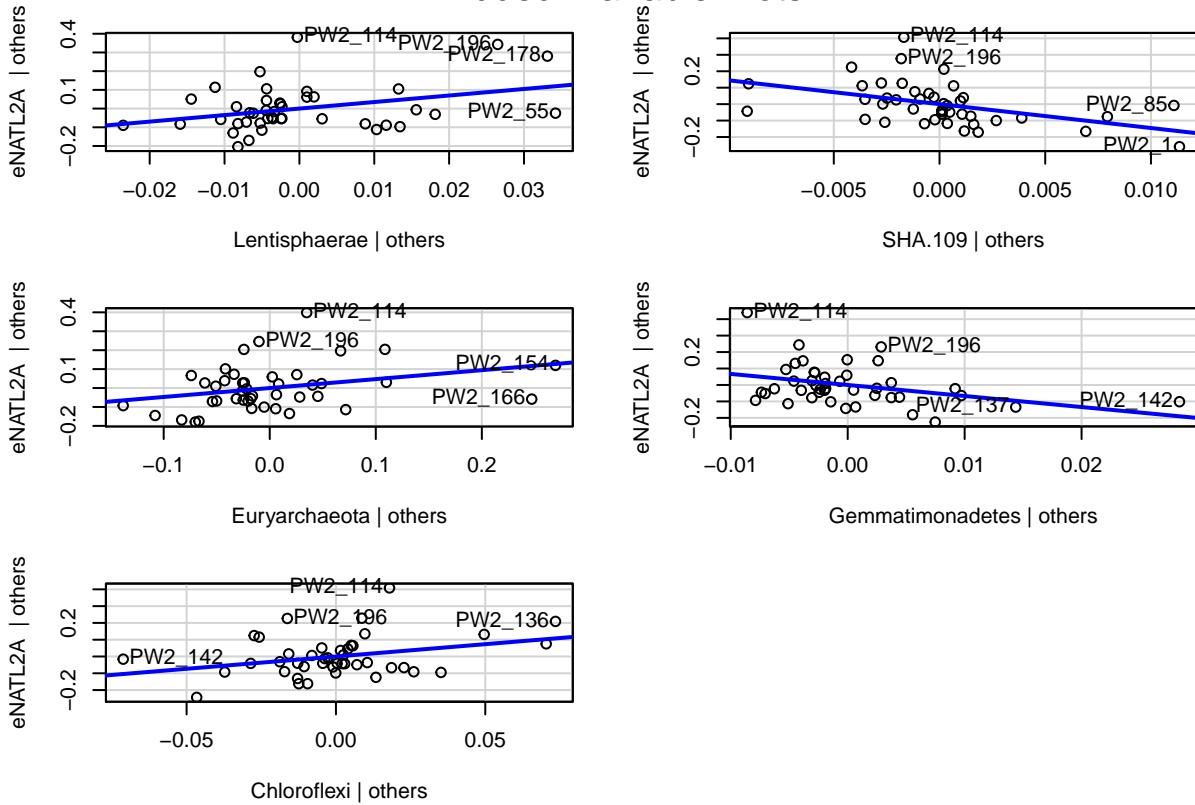
##
## Call:
## lm(formula = eNATL2A ~ Lentisphaerae + SHA.109 + Euryarchaeota +
##       Gemmatimonadetes + Chloroflexi, data = top100phyvWin.hell)
##
## Residuals:
##      Min        1Q     Median        3Q       Max
## -0.17516 -0.06311 -0.02168  0.05683  0.38218
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.1239    0.0305  4.062 0.000243 ***
## Lentisphaerae 3.4910    1.5603  2.237 0.031372 *
## SHA.109     -14.5247   4.7546 -3.055 0.004161 **
## Euryarchaeota 0.4720    0.2445  1.931 0.061185 .
## Gemmatimonadetes -6.7065   2.8527 -2.351 0.024165 *
## Chloroflexi    1.4658    0.7151  2.050 0.047534 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1225 on 37 degrees of freedom
## Multiple R-squared:  0.4886, Adjusted R-squared:  0.4194
## F-statistic: 7.069 on 5 and 37 DF,  p-value: 1e-04

```

When comparing eNATL2A against the Phylum in the winter samples, we found this ecotype has significant relationships with Lentisphaerae ($p < .05$), the uncultured phylum SHA.109 ($p < .005$), Gemmatimonadetes ($p < .05$), and Chloroflexi ($p < .05$). Specifically we found a 3.4% increase (± 1.5) in eNATL2A abundance for every 1% increase in Lentisphaerae, a 15% decrease (± 5) in eNATL2A abundance for every 1% increase in SHA.109 abundance, a 7% (± 3) decrease in eNATL2A abundance for every 1% increase in Gemmatimonadetes, and a 2% increase in eNATL2A abundance for every 1% increase in Chloroflexi. While we found a .5% increase in eNATL2A for every 1% increase in Euryarchaeota, this was not statistically significant ($p > .05$)

```
avPlots(eNATL2Alm_Win_both)
```

Added-Variable Plots



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9313lm_Win_Intercept <- lm(eMIT9313 ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eMIT9313lm_Win <- lm(eMIT9313 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin.hell)
```

#Preforming a stepwise in both directions to find the best model

```
eMIT9313lm_Win_both <- step(eMIT9313lm_Win_Intercept, direction='both', scope=formula(eMIT9313lm_Win), ...)
```

```
eMIT9313lm_Win_both$anova
```

##	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA	42	0.09969080	-258.8759
## 2	+ Lentisphaerae	-1	0.015145467	41	0.08454533	-263.9617
## 3	+ Chloroflexi	-1	0.006593508	40	0.07795182	-265.4532
## 4	+ Deinococcus.Thermus	-1	0.010578864	39	0.06737296	-269.7246
## 5	+ Marinimicrobia_SAR406_clade	-1	0.010032686	38	0.05734027	-274.6579
## 6	+ Euryarchaeota	-1	0.007122087	37	0.05021818	-278.3609

```
eMIT9313lm_Win_both$coefficients
```

```
##              (Intercept)          Lentisphaerae
##                  0.0101600      1.4998094
##          Chloroflexi        Deinococcus.Thermus
##                  1.0958169     -3.0503470
## Marinimicrobia_SAR406_clade    Euryarchaeota
##                  -1.1138399      0.2947599

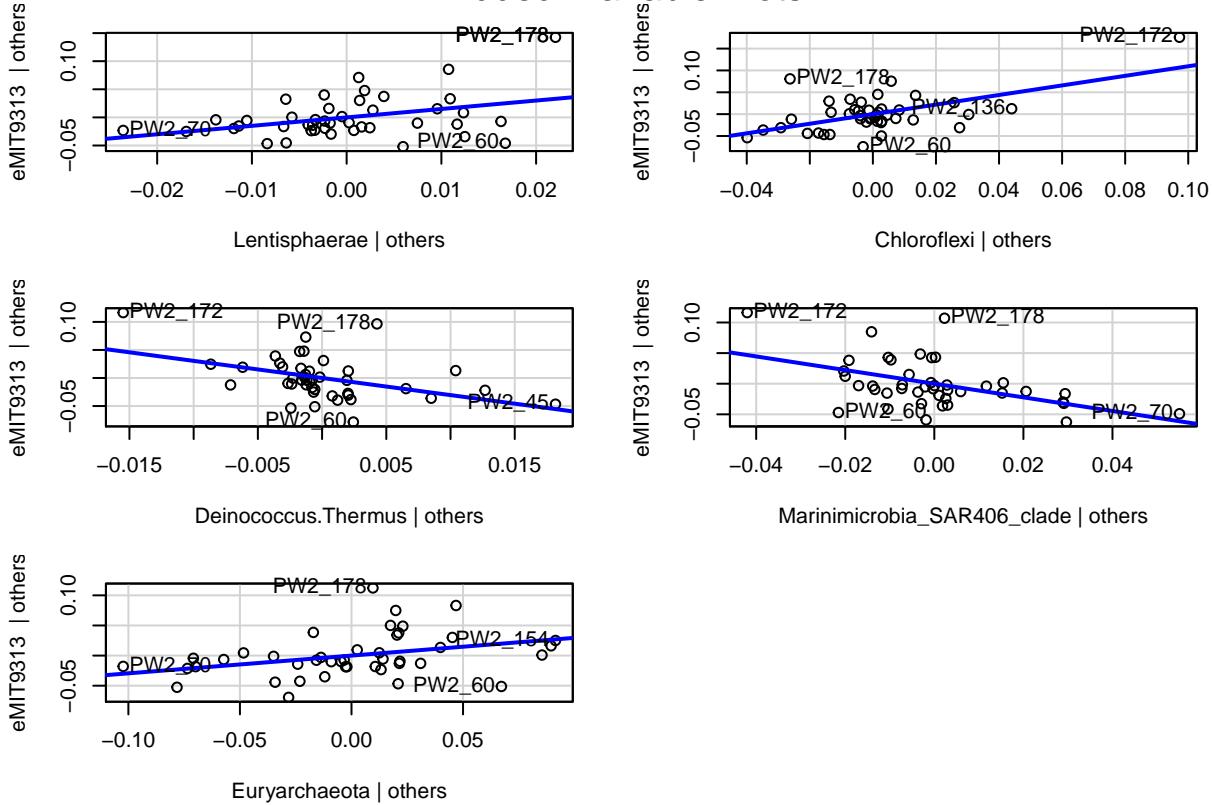
summary(eMIT9313lm_Win_both)

##
## Call:
## lm(formula = eMIT9313 ~ Lentisphaerae + Chloroflexi + Deinococcus.Thermus +
##     Marinimicrobia_SAR406_clade + Euryarchaeota, data = top100phyvWin.hell)
##
## Residuals:
##       Min     1Q   Median     3Q    Max 
## -0.071085 -0.020302 -0.003054  0.014299  0.109504 
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)    
## (Intercept) 0.010160  0.009119  1.114 0.272415    
## Lentisphaerae 1.499809  0.608108  2.466 0.018406 *  
## Chloroflexi   1.095817  0.253925  4.316 0.000114 *** 
## Deinococcus.Thermus -3.050347  1.043942 -2.922 0.005899 ** 
## Marinimicrobia_SAR406_clade -1.113840  0.330429 -3.371 0.001765 ** 
## Euryarchaeota  0.294760  0.128675  2.291 0.027774 *  
## --- 
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.03684 on 37 degrees of freedom
## Multiple R-squared:  0.4963, Adjusted R-squared:  0.4282 
## F-statistic:  7.29 on 5 and 37 DF,  p-value: 7.716e-05
```

When comparing eMIT9313 against the Phylum in the winter samples, we found this ecotype has significant relationships with Lentisphaerae ($p < .05$), the uncultured phylum Chloroflexi ($p < .0005$), Deinococcus-Thermus ($p < .01$), Marinimicrobia_SAR406_clade ($p < .005$), and Euryarchaeota ($p < .05$). Specifically we found a 1.5% increase ($\pm .6$) in eMIT9313 abundance for every 1% increase in Lentisphaerae, a 1.1% increase ($\pm .25$) in eMIT9313 abundance for every 1% increase in Chloroflexi abundance, a 3% (± 1) decrease in eMIT9313 abundance for every 1% increase in Deinococcus-Thermus, a 1.1% ($\pm .33$) decrease in eMIT9313 abundance for every 1% increase in Marinimicrobia_SAR406_clade, and a .3% ($\pm .13$) increase in eMIT9313 abundance for every 1% increase in Euryarchaeota.

```
avPlots(eMIT9313lm_Win_both)
```

Added-Variable Plots



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMED4lm_Win_Intercept <- lm(eMED4 ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eMED4lm_Win <- lm(eMED4 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin.hell)
```

#Preforming a stepwise in both directions to find the best model

```
eMED4lm_Win_both <- step(eMED4lm_Win_Intercept, direction='both', scope=formula(eMED4lm_Win), trace=0)
```

```
eMED4lm_Win_both$anova
```

##	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA	42	2.124159	-127.3364
## 2 + Proteobacteria	-1	0.5683472		41	1.555811	-138.7257
## 3 + Planctomycetes	-1	0.1313164		40	1.424495	-140.5175

```
eMED4lm_Win_both$coefficients
```

##	(Intercept)	Proteobacteria	Planctomycetes
##	0.4911182	-0.6401957	0.6483462

```

summary(eMED4lm_Win_both)

##
## Call:
## lm(formula = eMED4 ~ Proteobacteria + Planctomycetes, data = top100phyvWin.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max 
## -0.31781 -0.12590 -0.01619  0.12732  0.45972 
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)    
## (Intercept)  0.49112   0.08053   6.099 3.44e-07 ***
## Proteobacteria -0.64020   0.14843  -4.313 0.000102 ***
## Planctomycetes  0.64835   0.33764   1.920 0.061976 .  
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1887 on 40 degrees of freedom
## Multiple R-squared:  0.3294, Adjusted R-squared:  0.2959 
## F-statistic: 9.823 on 2 and 40 DF,  p-value: 0.0003384

```

When comparing eMED4 against the Phylum in the winter samples, we found this ecotype has significant relationships with Proteobacteria ($p < .05$)

Specifically we found a .6% decrease ($\pm .1$) in eMED4 abundance for every 1% increase in Proteobacteria. While we did find a .6% increase in eMED4 abundance for every 1% increase in Planctomycetes, this was not statistically significant ($p > .05$)

```
avPlots(eMED4lm_Win_both)
```

Added-Variable Plots

