

Ben_Linear_Regression

TJ

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Libraries

First we need to load in the data

```
#Summer data
top100phyvSum <- read.csv("data/NewSumWinData/POWOW_RawCounts_EcotypePhyla_Summer.csv") %>%
  column_to_rownames(., var = "Sample") %>%
  subset(., select = eMED4:Verrucomicrobia)
top100phyvSum.hell <- decostand(top100phyvSum, "hellinger") #Performing a hellinger transformation

#Winter data
top100phyvWin <- read.csv("data/NewSumWinData/POWOW_RawCounts_EcotypePhyla_Winter.csv") %>%
  column_to_rownames(., var = "Sample") %>%
  subset(., select = eMED4:Verrucomicrobia)
top100phyvWin.hell <- decostand(top100phyvWin, "hellinger") #Performing a hellinger transformation
```

Summer analysis

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9312lm_Sum_Intersept <- lm(eMIT9312 ~ 1, data = top100phyvSum.hell) #First we will define the intercept
eMIT9312lm_Sum <- lm(eMIT9312 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will
```

```
vif(eMIT9312lm_Sum)
```

##	Chloroflexi	Cyanobacteria
##	9.439458	3.903344
##	Deinococcus.Thermus	Euryarchaeota
##	1.639400	8.018580
##	Firmicutes	Fusobacteria
##	4.741412	1.609413
##	Gemmatimonadetes	Gracilibacteria

```
##          5.381990          2.023682
##          Lentisphaerae Marinimicrobia_SAR406_clade
##          2.010246          8.027217
##          PAUC34f          Planctomycetes
##          3.398654          2.102418
##          Proteobacteria          SHA.109
##          6.057502          1.313680
##          Thaumarchaeota          Verrucomicrobia
##          10.288416          2.990001
```

```
eMIT9312lm_Sum_both <- step(eMIT9312lm_Sum_Intersept, direction='both', scope=formula(eMIT9312lm_Sum),
```

```
eMIT9312lm_Sum_both$anova
```

```
##          Step Df    Deviance Resid. Df Resid. Dev      AIC
## 1              NA         NA         51  2.394479 -158.0601
## 2      + Proteobacteria -1 0.40757846         50  1.986901 -165.7627
## 3 + Deinococcus.Thermus -1 0.14579905         49  1.841102 -167.7257
## 4      + Planctomycetes -1 0.09743226         48  1.743669 -168.5531
```

```
eMIT9312lm_Sum_both$coefficients
```

```
##          (Intercept)      Proteobacteria Deinococcus.Thermus      Planctomycetes
##          0.3153202          -0.3806562          10.4842631          -0.6532889
```

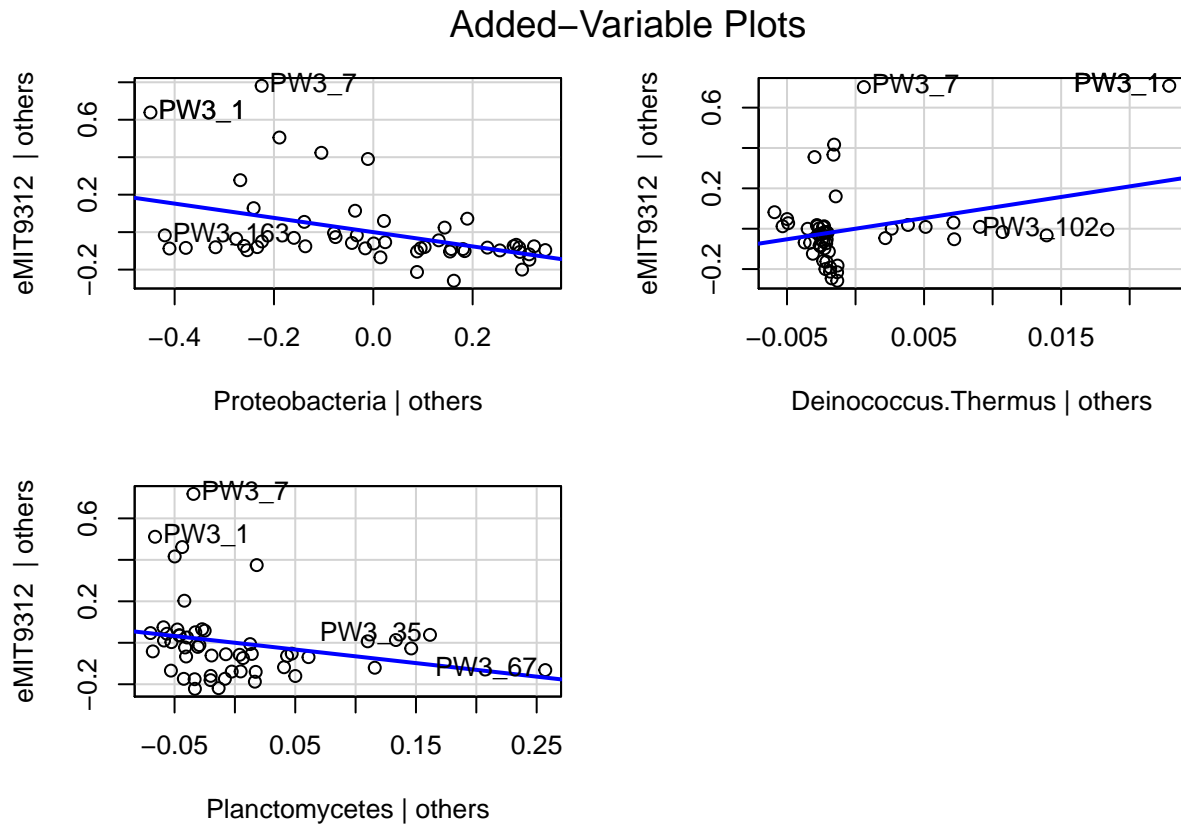
```
summary(eMIT9312lm_Sum_both)
```

```
##
## Call:
## lm(formula = eMIT9312 ~ Proteobacteria + Deinococcus.Thermus +
##     Planctomycetes, data = top100phyvSum.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.24368 -0.10140 -0.03093  0.03618  0.69555
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    0.31532    0.06936   4.546 3.71e-05 ***
## Proteobacteria -0.38066    0.11760  -3.237  0.00219 **
## Deinococcus.Thermus 10.48426    4.59146   2.283  0.02687 *
## Planctomycetes  -0.65329    0.39890  -1.638  0.10802
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1906 on 48 degrees of freedom
## Multiple R-squared:  0.2718, Adjusted R-squared:  0.2263
## F-statistic: 5.972 on 3 and 48 DF,  p-value: 0.001521
```

When comparing eMIT9312 against the Phylum in the summer samples, we found this ecotype has significant relationships with Proteobacteria ($p < .005$) and Deinococcus-Thermus ($p < .05$). Specifically we found a

0.4% decrease (± 0.11) in eMIT9312 abundance for every 1% increase in Proteobacteria and a 10% increase (± 4.6) in eMIT9312 abundance for every 1% increase in Deinococcus-Thermus abundance. Though we saw a decrease of .66 in eMIT9312 for every 1% increase in Planctomycetes, this decrease was not statistically significant ($p > .05$).

```
avPlots(eMIT9312lm_Sum_both)
```



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eNATL2Alm_Sum_Intersept <- lm(eNATL2A ~ 1, data = top100phyvSum.hell) #First we will define the intercept
eNATL2Alm_Sum <- lm(eNATL2A ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will
```

#Performing a stepwise in both directions to find the best model

```
eNATL2Alm_Sum_both <- step(eNATL2Alm_Sum_Intersept, direction='both', scope=formula(eNATL2Alm_Sum), tra
```

```
eNATL2Alm_Sum_both$anova
```

```
## Step Df Deviance Resid. Df Resid. Dev      AIC
## 1    NA      NA      51    2.742193 -151.0093
```

```
eNATL2Alm_Sum_both$coefficients
```

```
## (Intercept)
##      0.152451
```

```
summary(eNATL2Alm_Sum_both)
```

```
##
## Call:
## lm(formula = eNATL2A ~ 1, data = top100phyvSum.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.1525 -0.1502 -0.1426  0.1080  0.6204
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.15245    0.03216   4.741 1.75e-05 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2319 on 51 degrees of freedom
```

There is no sig correlation for eNATL2A

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9313lm_Sum_Intersept <- lm(eMIT9313 ~ 1, data = top100phyvSum.hell) #First we will define the intercept
eMIT9313lm_Sum <- lm(eMIT9313 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will
```

```
#Preforming a stepwise in both directions to find the best model
```

```
eMIT9313lm_Sum_both <- step(eMIT9313lm_Sum_Intersept, direction='both', scope=formula(eMIT9313lm_Sum),
```

```
eMIT9313lm_Sum_both$anova
```

```
##           Step Df   Deviance Resid.  Df Resid. Dev      AIC
## 1              NA      NA         51 0.06073271 -349.1309
## 2 + Chloroflexi -1 0.00295798      50 0.05777473 -349.7273
```

```
eMIT9313lm_Sum_both$coefficients
```

```
## (Intercept) Chloroflexi
##  0.01764023 -0.22909016
```

```
summary(eMIT9313lm_Sum_both)
```

```
##
## Call:
## lm(formula = eMIT9313 ~ Chloroflexi, data = top100phyvSum.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.017640 -0.016232 -0.009533 -0.001025  0.150683
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.017640   0.005875   3.003  0.00417 **
## Chloroflexi -0.229090   0.143183  -1.600  0.11590
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.03399 on 50 degrees of freedom
## Multiple R-squared:  0.0487, Adjusted R-squared:  0.02968
## F-statistic:  2.56 on 1 and 50 DF,  p-value: 0.1159
```

Only the intercept is sig here

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMED4lm_Sum_Intersept <- lm(eMED4 ~ 1, data = top100phyvSum.hell) #First we will define the intersept
eMED4lm_Sum <- lm(eMED4 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvSum.hell) #Now I will
```

```
#Preforming a stepwise in both directions to find the best model
eMED4lm_Sum_both <- step(eMED4lm_Sum_Intersept, direction='both', scope=formula(eMED4lm_Sum), trace=0)
```

```
eMED4lm_Sum_both$anova
```

##		Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA		51	6.705967	-104.5088
## 2	+ Proteobacteria	-1	3.6507605		50	3.055207	-143.3886
## 3	+ Deinococcus.Thermus	-1	0.3633326		49	2.691874	-147.9723
## 4	+ Euryarchaeota	-1	0.1121722		48	2.579702	-148.1856

```
eMED4lm_Sum_both$coefficients
```

##	(Intercept)	Proteobacteria	Deinococcus.Thermus	Euryarchaeota
##	1.0359735	-1.1104049	-12.2798426	-0.3903005

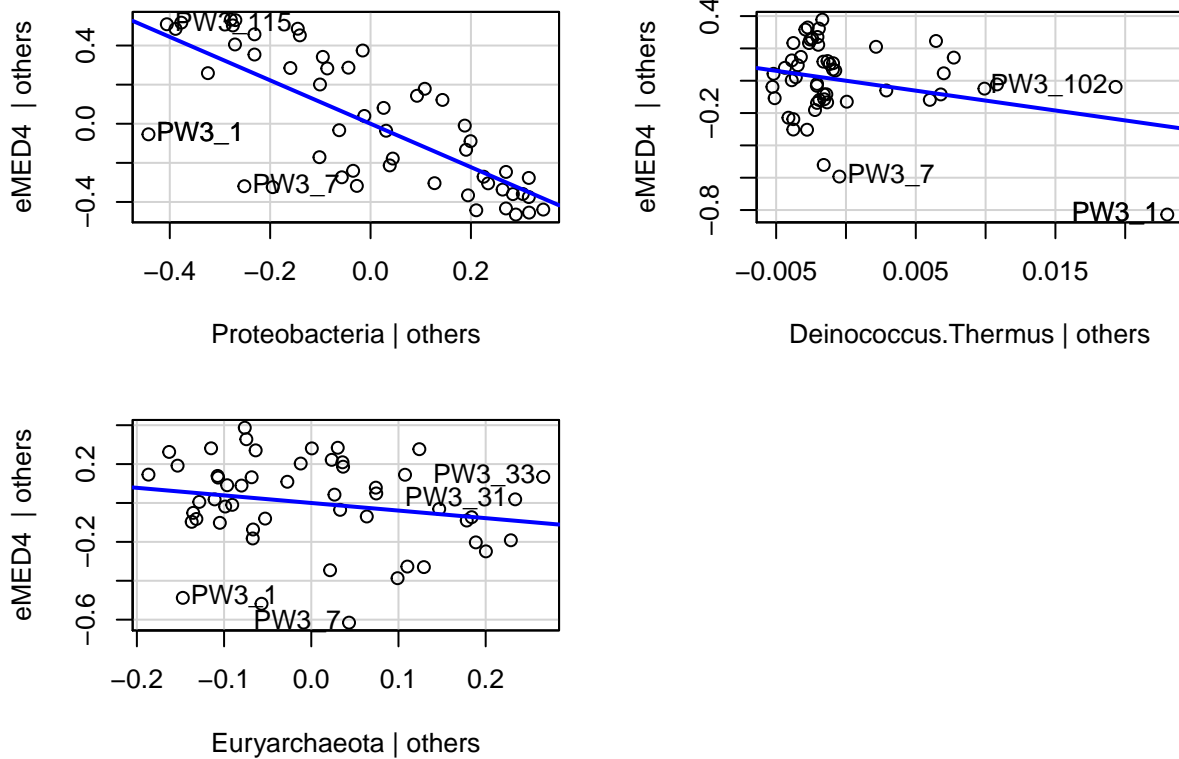
```
summary(eMED4lm_Sum_both)
```

```
##
## Call:
## lm(formula = eMED4 ~ Proteobacteria + Deinococcus.Thermus + Euryarchaeota,
##     data = top100phyvSum.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.59850 -0.13013  0.03972  0.18986  0.35708
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      1.03597    0.08509   12.176 2.75e-16 ***
## Proteobacteria    -1.11040    0.14299   -7.766 4.98e-10 ***
## Deinococcus.Thermus -12.27984    5.64451   -2.176  0.0345 *
## Euryarchaeota     -0.39030    0.27016   -1.445  0.1550
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2318 on 48 degrees of freedom
## Multiple R-squared:  0.6153, Adjusted R-squared:  0.5913
## F-statistic: 25.59 on 3 and 48 DF,  p-value: 4.917e-10
```

When comparing eMED4 against the Phylum in the summary samples, we found this ecotype has significant relationships with Proteobacteria ($p < .001$) and Deinococcus-Thermus ($p < .05$). Specifically we found a 1.1% decrease (± 0.14) in eMED4 abundance for every 1% increase in Proteobacteria and a 12% decrease (± 5.6) in eMED4 abundance for every 1% increase in Deinococcus-Thermus abundance. Though we saw a decrease of .4% in eMED4 for every 1% increase in Euryarchaeota, this decrease was not statistically significant ($p > .05$).

```
avPlots(eMED4lm_Sum_both)
```

Added-Variable Plots



Winter analysis

Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9312lm_Win_Intersept <- lm(eMIT9312 ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eMIT9312lm_Win <- lm(eMIT9312 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin.hell)
```

```
vif(eMIT9312lm_Win)
```

```
##          Chloroflexi          Cyanobacteria
##          7.281146          4.581259
##    Deinococcus.Thermus    Euryarchaeota
##          3.314307          15.599366
##          Firmicutes          Fusobacteria
##          9.626650          1.884679
##    Gemmatimonadetes    Gracilibacteria
##          3.660177          4.701867
##    Lentisphaerae Marinimicrobia_SAR406_clade
##          3.977338          26.827255
```

```
##          PAUC34f          Planctomycetes
##          4.937170          3.616936
##          Proteobacteria          SHA.109
##          5.094804          3.066283
##          Thaumarchaeota          Verrucomicrobia
##          4.332598          8.003607
```

```
eMIT9312lm_Win_both <- step(eMIT9312lm_Win_Intersept, direction='both', scope=formula(eMIT9312lm_Win),
```

```
eMIT9312lm_Win_both$anova
```

```
##          Step Df Deviance Resid. Df Resid. Dev      AIC
## 1          NA      NA      42    5.222196 -88.65613
## 2 + Proteobacteria -1 2.622288      41    2.599908 -116.64614
## 3   + Chloroflexi -1 0.438386      40    2.161522 -122.58667
```

```
eMIT9312lm_Win_both$coefficients
```

```
##      (Intercept) Proteobacteria Chloroflexi
##      0.9885717      -1.2591248      -2.2176446
```

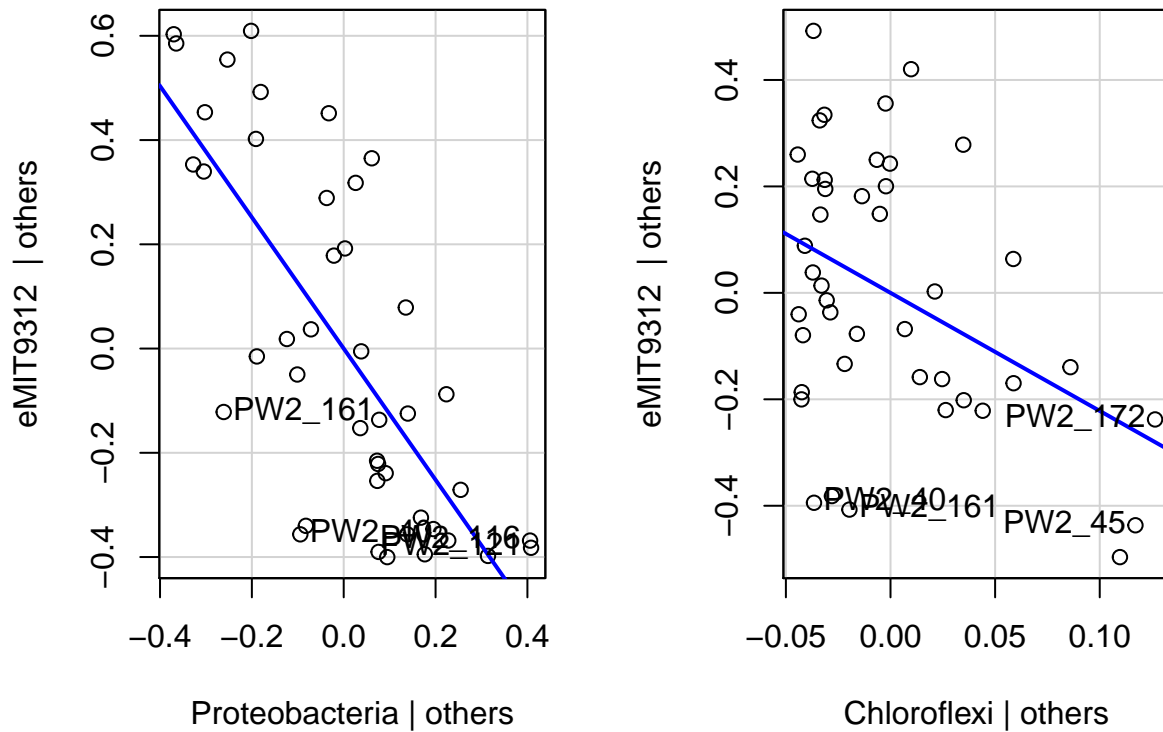
```
summary(eMIT9312lm_Win_both)
```

```
##
## Call:
## lm(formula = eMIT9312 ~ Proteobacteria + Chloroflexi, data = top100phyvWin.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.47548 -0.13240 -0.03934  0.15643  0.44203
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    0.98857    0.09644  10.251 9.40e-13 ***
## Proteobacteria -1.25912    0.17868  -7.047 1.61e-08 ***
## Chloroflexi    -2.21764    0.77860  -2.848 0.00691 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.2325 on 40 degrees of freedom
## Multiple R-squared:  0.5861, Adjusted R-squared:  0.5654
## F-statistic: 28.32 on 2 and 40 DF,  p-value: 2.178e-08
```

When comparing eMIT9312 against the Phylum in the winter samples, we found this ecotype has significant relationships with Proteobacteria ($p < .001$) and Chloroflexi ($p < .001$). Specifically we found a 1% decrease (± 0.2) in eMIT9312 abundance for every 1% increase in Proteobacteria and a 2% decrease ($\pm .8$) in eMIT9312 abundance for every 1% increase in Chloroflexi abundance.

```
avPlots(eMIT9312lm_Win_both)
```


Added-Variable Plots



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eNATL2Alm_Win_Intersept <- lm(eNATL2A ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eNATL2Alm_Win <- lm(eNATL2A ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin)
```

```
#Performing a stepwise in both directions to find the best model
eNATL2Alm_Win_both <- step(eNATL2Alm_Win_Intersept, direction='both', scope=formula(eNATL2Alm_Win), trace=TRUE)
```

```
eNATL2Alm_Win_both$anova
```

##	Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1	NA	NA		42	1.0863957	-156.1684
## 2	+ Lentisphaerae	-1	0.20703655	41	0.8793592	-163.2598
## 3	+ SHA.109	-1	0.08435733	40	0.7950018	-165.5963
## 4	+ Euryarchaeota	-1	0.14629648	39	0.6487054	-172.3410
## 5	+ Gemmatimonadetes	-1	0.02998499	38	0.6187204	-172.3760
## 6	+ Chloroflexi	-1	0.06308949	37	0.5556309	-175.0006

```
eNATL2Alm_Win_both$coefficients
```

```
##      (Intercept)    Lentisphaerae      SHA.109    Euryarchaeota
##      0.1238786      3.4909464      -14.5247285      0.4720197
## Gemmatimonadetes    Chloroflexi
##      -6.7064609      1.4658046
```

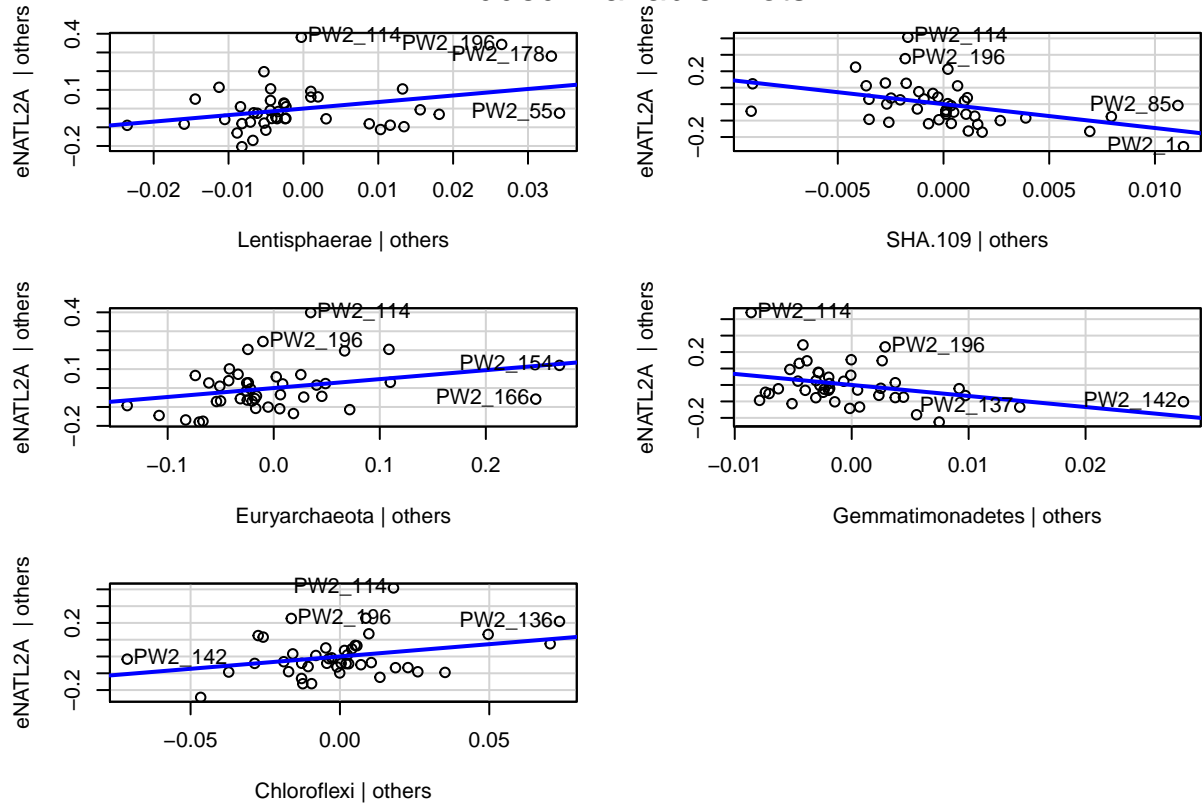
```
summary(eNATL2Alm_Win_both)
```

```
##
## Call:
## lm(formula = eNATL2A ~ Lentisphaerae + SHA.109 + Euryarchaeota +
##      Gemmatimonadetes + Chloroflexi, data = top100phyvWin.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.17516 -0.06311 -0.02168  0.05683  0.38218
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    0.1239     0.0305   4.062 0.000243 ***
## Lentisphaerae    3.4910     1.5603   2.237 0.031372 *
## SHA.109        -14.5247     4.7546  -3.055 0.004161 **
## Euryarchaeota     0.4720     0.2445   1.931 0.061185 .
## Gemmatimonadetes -6.7065     2.8527  -2.351 0.024165 *
## Chloroflexi      1.4658     0.7151   2.050 0.047534 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1225 on 37 degrees of freedom
## Multiple R-squared:  0.4886, Adjusted R-squared:  0.4194
## F-statistic: 7.069 on 5 and 37 DF,  p-value: 1e-04
```

When comparing eNATL2A against the Phylum in the winter samples, we found this ecotype has significant relationships with Lentisphaerae ($p < .05$), the uncultured phylum SHA.109 ($p < .005$), Gemmatimonadetes ($p < .05$), and Chloroflexi ($p < .05$). Specifically we found a 3.4% increase (± 1.5) in eNATL2A abundance for every 1% increase in Lentisphaerae, a 15% decrease (± 5) in eNATL2A abundance for every 1% increase in SHA.109 abundance, a 7% (± 3) decrease in eNATL2A abundance for every 1% increase in Gemmatimonadetes, and a 2% increase in eNATL2A abundance for every 1% increase in Chloroflexi. While we found a .5% increase in eNATL2A for every 1% increase in Euryarchaeota, this was not statistically significant ($p > .05$)

```
avPlots(eNATL2Alm_Win_both)
```

Added-Variable Plots



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMIT9313lm_Win_Intersept <- lm(eMIT9313 ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eMIT9313lm_Win <- lm(eMIT9313 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin.hell)
```

#Performing a stepwise in both directions to find the best model

```
eMIT9313lm_Win_both <- step(eMIT9313lm_Win_Intersept, direction='both', scope=formula(eMIT9313lm_Win), direction='both')
```

```
eMIT9313lm_Win_both$anova
```

##		Step	Df	Deviance	Resid. Df	Resid. Dev	AIC
## 1		NA	NA		42	0.09969080	-258.8759
## 2	+ Lentisphaerae	-1	0.015145467		41	0.08454533	-263.9617
## 3	+ Chloroflexi	-1	0.006593508		40	0.07795182	-265.4532
## 4	+ Deinococcus.Thermus	-1	0.010578864		39	0.06737296	-269.7246
## 5	+ Marinimicrobia_SAR406_clade	-1	0.010032686		38	0.05734027	-274.6579
## 6	+ Euryarchaeota	-1	0.007122087		37	0.05021818	-278.3609

```
eMIT9313lm_Win_both$coefficients
```

```
##              (Intercept)              Lentisphaerae
##              0.0101600              1.4998094
##              Chloroflexi              Deinococcus.Thermus
##              1.0958169              -3.0503470
## Marinimicrobia_SAR406_clade              Euryarchaeota
##              -1.1138399              0.2947599
```

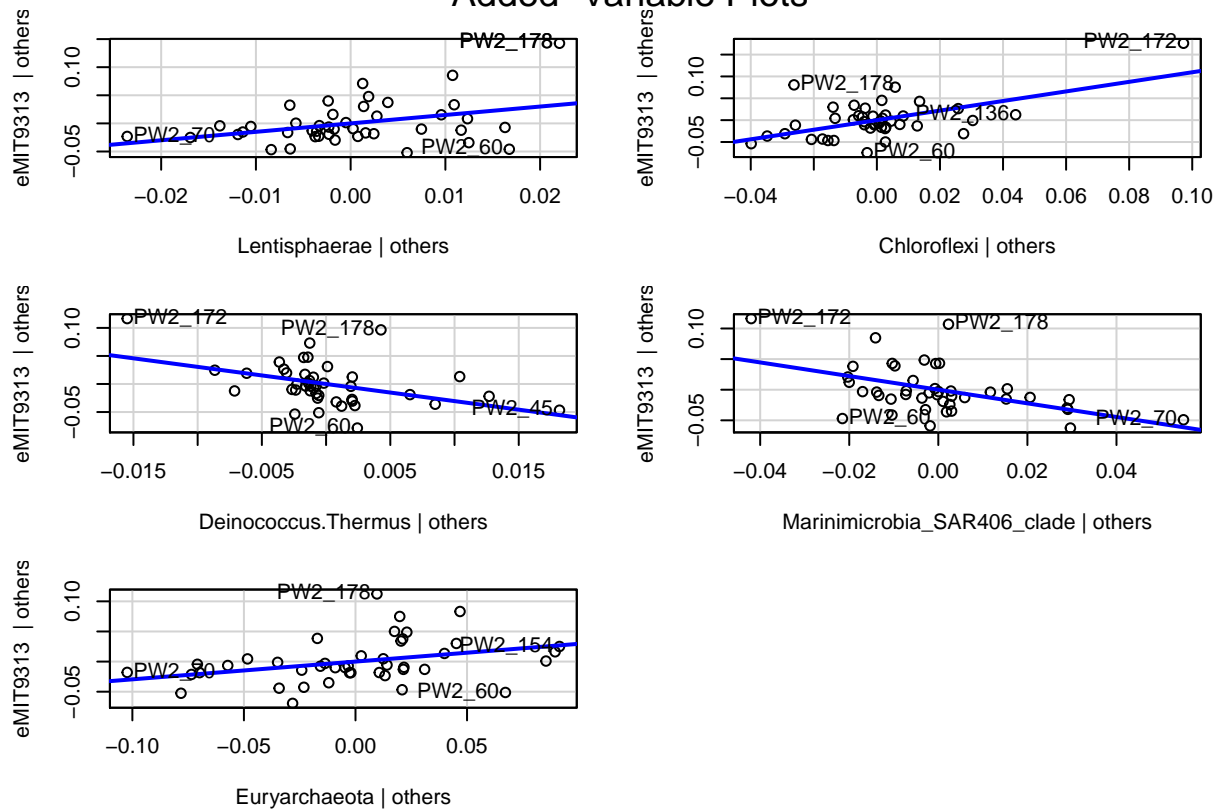
```
summary(eMIT9313lm_Win_both)
```

```
##
## Call:
## lm(formula = eMIT9313 ~ Lentisphaerae + Chloroflexi + Deinococcus.Thermus +
##     Marinimicrobia_SAR406_clade + Euryarchaeota, data = top100phyvWin.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.071085 -0.020302 -0.003054  0.014299  0.109504
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      0.010160   0.009119   1.114 0.272415
## Lentisphaerae      1.499809   0.608108   2.466 0.018406 *
## Chloroflexi       1.095817   0.253925   4.316 0.000114 ***
## Deinococcus.Thermus -3.050347   1.043942  -2.922 0.005899 **
## Marinimicrobia_SAR406_clade -1.113840   0.330429  -3.371 0.001765 **
## Euryarchaeota      0.294760   0.128675   2.291 0.027774 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.03684 on 37 degrees of freedom
## Multiple R-squared:  0.4963, Adjusted R-squared:  0.4282
## F-statistic:  7.29 on 5 and 37 DF,  p-value: 7.716e-05
```

When comparing eMIT9313 against the Phylum in the winter samples, we found this ecotype has significant relationships with Lentisphaerae ($p < .05$), the uncultured phylum Chloroflexi ($p < .0005$), Deinococcus-Thermus ($p < .01$), Marinimicrobia_SAR406_clade ($p < .005$), and Euryarchaeota ($p < .05$). Specifically we found a 1.5% increase ($\pm .6$) in eMIT9313 abundance for every 1% increase in Lentisphaerae, a 1.1% increase ($\pm .25$) in eMIT9313 abundance for every 1% increase in Chloroflexi abundance, a 3% (± 1) decrease in eMIT9313 abundance for every 1% increase in Deinococcus-Thermus, a 1.1% ($\pm .33$) decrease in eMIT9313 abundance for every 1% increase in Marinimicrobia_SAR406_clade, and a .3% ($\pm .13$) increase in eMIT9313 abundance for every 1% increase in Euryarchaeota.

```
avPlots(eMIT9313lm_Win_both)
```

Added-Variable Plots



Now I want to evaluate the correlations between eMIT9312 and the top100 phylum

```
eMED4lm_Win_Intersept <- lm(eMED4 ~ 1, data = top100phyvWin.hell) #First we will define the intercept
eMED4lm_Win <- lm(eMED4 ~ Chloroflexi + Cyanobacteria + Deinococcus.Thermus + Euryarchaeota +
  Firmicutes + Fusobacteria + Gemmatimonadetes + Gracilibacteria +
  Lentisphaerae + Marinimicrobia_SAR406_clade + PAUC34f + Planctomycetes +
  Proteobacteria + SHA.109 + Thaumarchaeota + Verrucomicrobia, data = top100phyvWin.hell)
```

```
#Performing a stepwise in both directions to find the best model
eMED4lm_Win_both <- step(eMED4lm_Win_Intersept, direction='both', scope=formula(eMED4lm_Win), trace=0)
```

```
eMED4lm_Win_both$anova
```

```
##           Step Df  Deviance Resid. Df Resid. Dev      AIC
## 1              NA      NA         42   2.124159 -127.3364
## 2 + Proteobacteria -1 0.5683472      41   1.555811 -138.7257
## 3 + Planctomycetes -1 0.1313164      40   1.424495 -140.5175
```

```
eMED4lm_Win_both$coefficients
```

```
##      (Intercept) Proteobacteria Planctomycetes
##      0.4911182    -0.6401957      0.6483462
```

```
summary(eMED4lm_Win_both)
```

```
##
## Call:
## lm(formula = eMED4 ~ Proteobacteria + Planctomycetes, data = top100phyvWin.hell)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.31781 -0.12590 -0.01619  0.12732  0.45972
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    0.49112    0.08053   6.099 3.44e-07 ***
## Proteobacteria -0.64020    0.14843  -4.313 0.000102 ***
## Planctomycetes  0.64835    0.33764   1.920 0.061976 .
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.1887 on 40 degrees of freedom
## Multiple R-squared:  0.3294, Adjusted R-squared:  0.2959
## F-statistic: 9.823 on 2 and 40 DF,  p-value: 0.0003384
```

When comparing eMED4 against the Phylum in the winter samples, we found this ecotype has significant relationships with Proteobacteria ($p < .05$)

Specifically we found a .6% decrease ($\pm .1$) in eMED4 abundance for every 1% increase in Proteobacteria. While we did find a .6% increase in eMED4 abundance for every 1% increase in Planctomycetes, this was not statistically significant ($p > .05$)

```
avPlots(eMED4lm_Win_both)
```

Added-Variable Plots

