

# PCA And Sequence Alignment

Abdullah Taman

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```
library(tidyverse)
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.4      v readr      2.1.5
## v forcats    1.0.0      v stringr   1.5.1
## v ggplot2    3.4.4      v tibble    3.2.1
## v lubridate  1.9.3      v tidyr     1.3.1
## v purrr      1.0.2
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors
```

```
library(rentrez)
```

```
## Warning: package 'rentrez' was built under R version 4.3.3
```

```
library(seqinr)
```

```
## Warning: package 'seqinr' was built under R version 4.3.3
```

```
##
## Attaching package: 'seqinr'
##
## The following object is masked from 'package:dplyr':
##
##     count
```

```
library(Biostrings)
```

```
## Loading required package: BiocGenerics
##
## Attaching package: 'BiocGenerics'
##
## The following objects are masked from 'package:lubridate':
##
##     intersect, setdiff, union
##
## The following objects are masked from 'package:dplyr':
```

```

##
##   combine, intersect, setdiff, union
##
## The following objects are masked from 'package:stats':
##
##   IQR, mad, sd, var, xtabs
##
## The following objects are masked from 'package:base':
##
##   anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##   colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
##   get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,
##   match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,
##   Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort,
##   table, tapply, union, unique, unsplit, which.max, which.min
##
## Loading required package: S4Vectors
## Loading required package: stats4
##
## Attaching package: 'S4Vectors'
##
## The following objects are masked from 'package:lubridate':
##
##   second, second<-
##
## The following objects are masked from 'package:dplyr':
##
##   first, rename
##
## The following object is masked from 'package:tidyr':
##
##   expand
##
## The following object is masked from 'package:utils':
##
##   findMatches
##
## The following objects are masked from 'package:base':
##
##   expand.grid, I, unname
##
## Loading required package: IRanges
##
## Attaching package: 'IRanges'
##
## The following object is masked from 'package:lubridate':
##
##   %within%
##
## The following objects are masked from 'package:dplyr':
##
##   collapse, desc, slice
##
## The following object is masked from 'package:purrr':

```

```
##
##      reduce
##
## The following object is masked from 'package:grDevices':
##
##      windows
##
## Loading required package: XVector
##
## Attaching package: 'XVector'
##
## The following object is masked from 'package:purrr':
##
##      compact
##
## Loading required package: GenomeInfoDb
##
## Attaching package: 'Biostrings'
##
## The following object is masked from 'package:seqinr':
##
##      translate
##
## The following object is masked from 'package:base':
##
##      strsplit
```

```
library(ggplot2)
library("FactoMineR")
```

```
## Warning: package 'FactoMineR' was built under R version 4.3.3
```

```
library(ggcorrplot)
```

```
## Warning: package 'ggcorrplot' was built under R version 4.3.3
```

```
library('corrr')
```

```
## Warning: package 'corrr' was built under R version 4.3.3
```

## # Part One

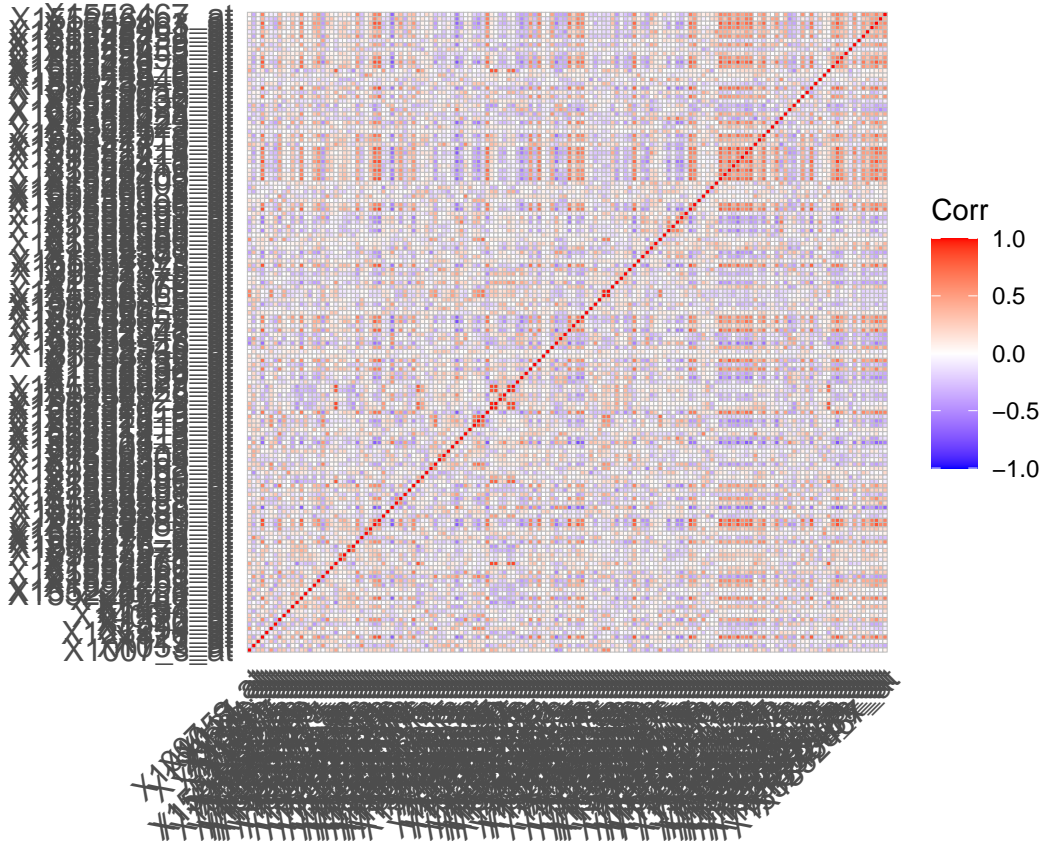
First performing the PCA

```
cancer <- read.csv("J:/champions work/datasets/BrainCancerMin.csv", header = TRUE)
?princomp()
```

```
## starting httpd help server ... done
```

Now we'll perform the PCA I faced an issue with the code, This error: Error in princomp.default(remove\_type, cor = TRUE, scores = TRUE) : 'princomp' can only be used with more units than variables

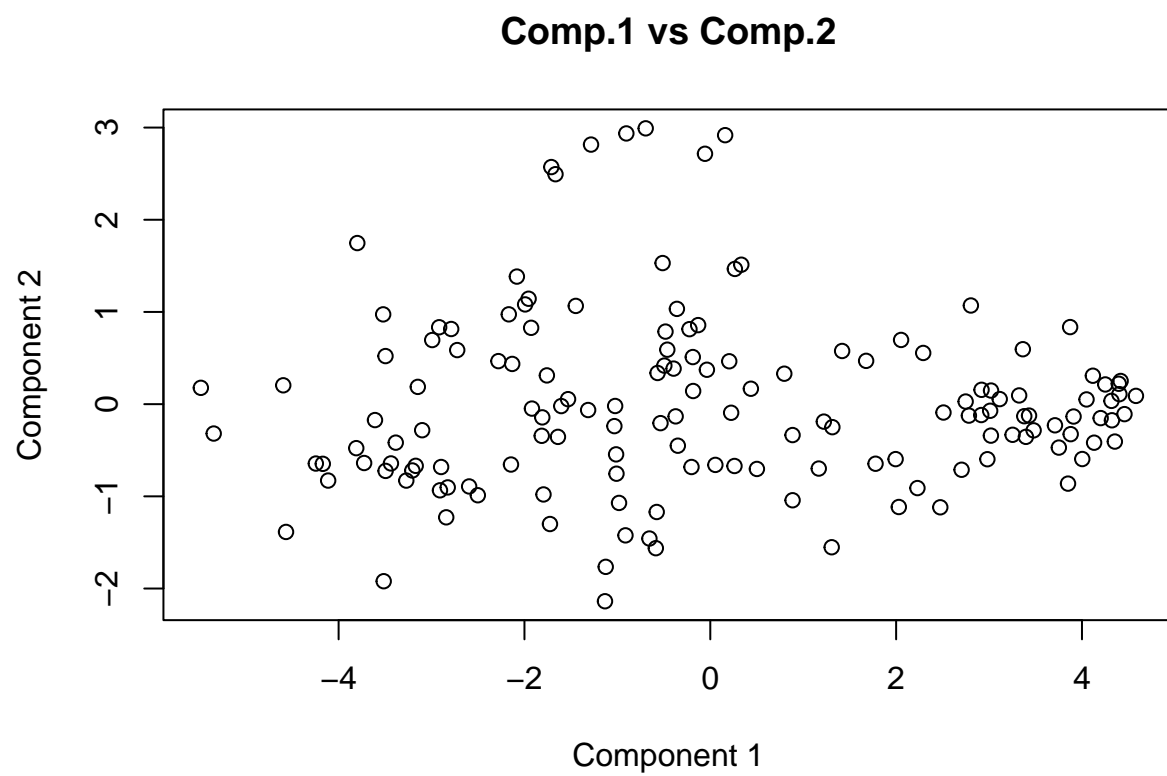
```
remove_type <- subset(cancer, select = c(3:ncol(cancer)))
data_normalized <- scale(remove_type)
corr_matrix <- cor(data_normalized)
ggcorrplot(corr_matrix)
```



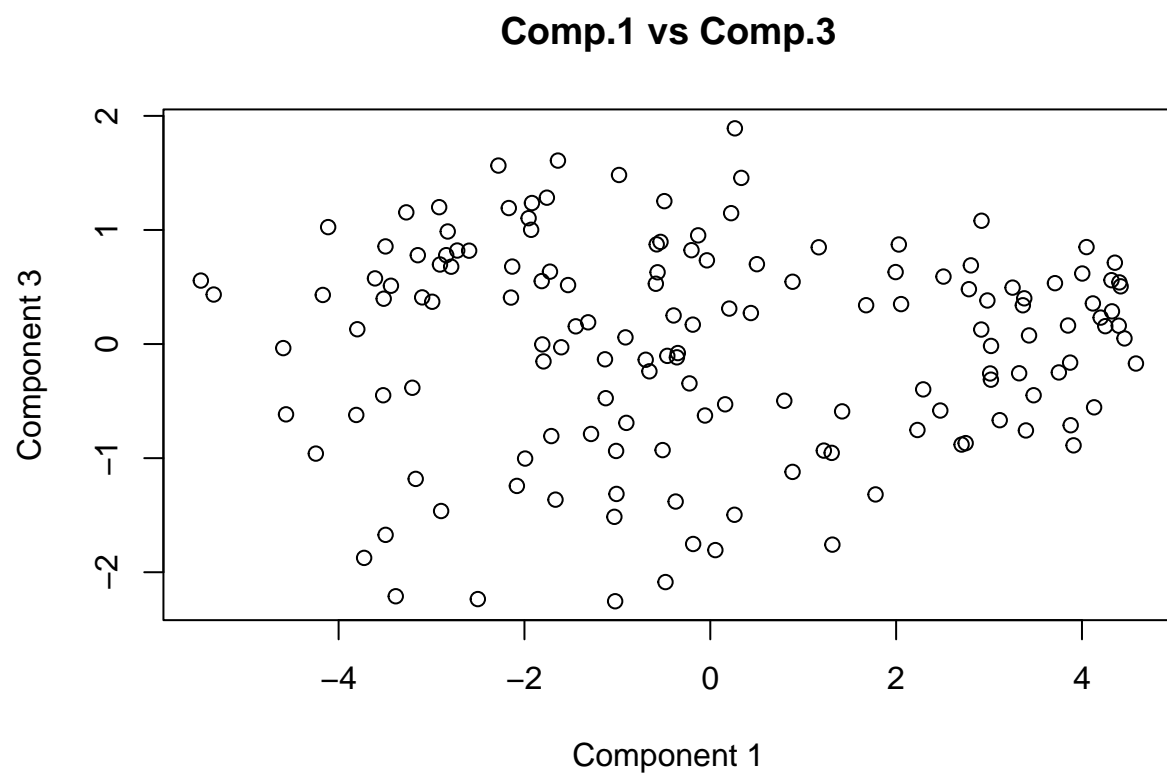
```
data.pca <- princomp(as.data.frame(corr_matrix))
#summary(data.pca)
```

```
# Extract principal component scores
pc_scores <- data.pca$scores
```

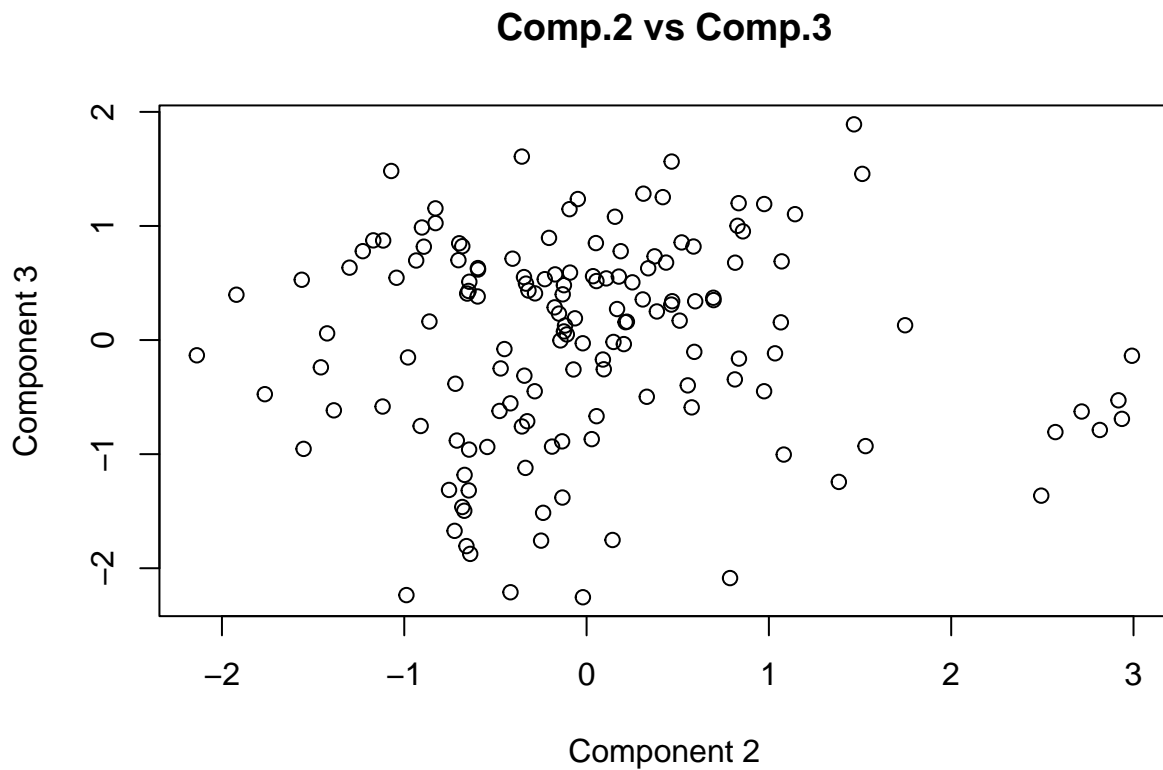
```
# Plot Comp.1 vs Comp.2
plot(pc_scores[,1], pc_scores[,2], xlab = "Component 1", ylab = "Component 2",
     main = "Comp.1 vs Comp.2")
```



```
# Plot Comp.1 vs Comp.3  
plot(pc_scores[,1], pc_scores[,3], xlab = "Component 1", ylab = "Component 3",  
     main = "Comp.1 vs Comp.3")
```



```
# Plot Comp.2 vs Comp.3  
plot(pc_scores[,2], pc_scores[,3], xlab = "Component 2", ylab = "Component 3",  
     main = "Comp.2 vs Comp.3")
```



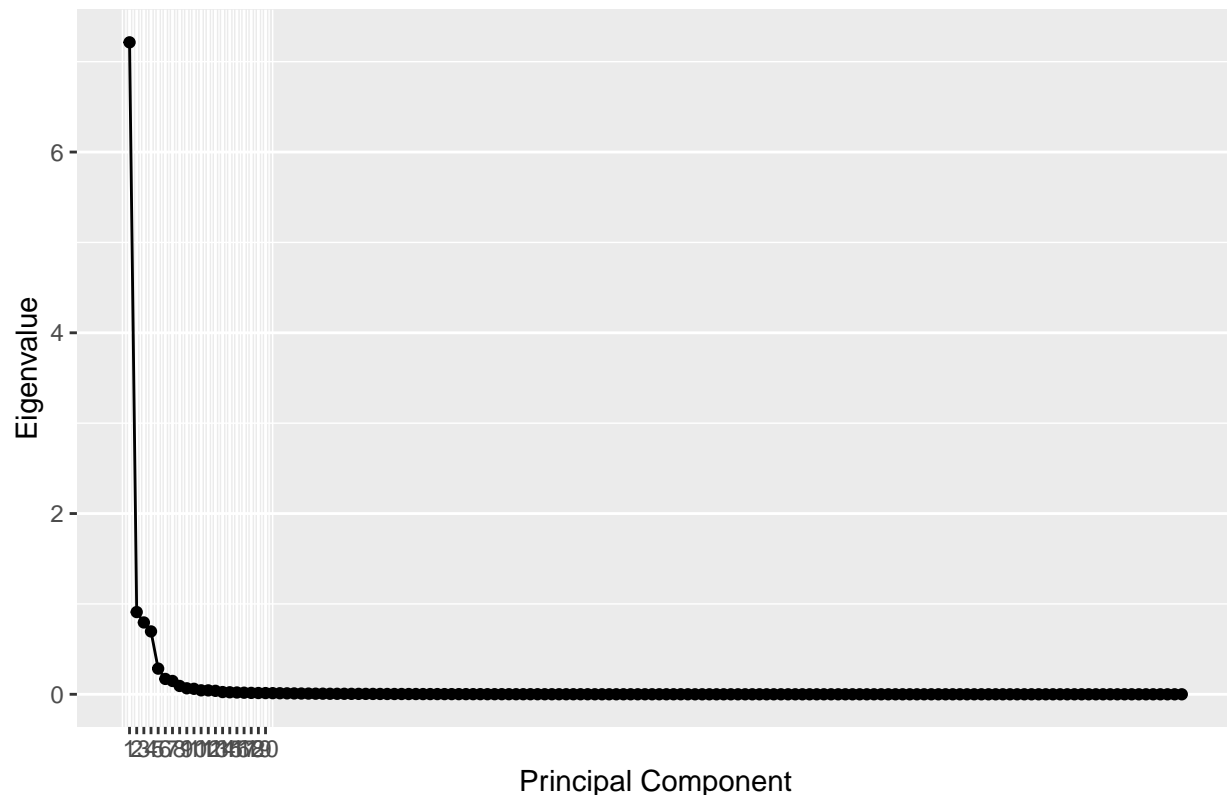
Component 1 vs Component 2 is the best, why ? You could see that the number of points that are overlapped is the least number in it.

```
# Extract eigenvalues
eigenvalues <- data.pca$sdev^2

# Create data frame for plotting
scree_data <- data.frame(Component = 1:length(eigenvalues), Eigenvalue = eigenvalues)

# Plot scree plot
ggplot(scree_data, aes(x = Component, y = Eigenvalue)) +
  geom_point() +
  geom_line() +
  scale_x_continuous(breaks = seq(1, 20, by = 1)) + # Adjust x-axis breaks for better readability
  labs(x = "Principal Component", y = "Eigenvalue", title = "Scree Plot for First 20 Principal Components")
```

## Scree Plot for First 20 Principal Components



In multivariate statistics, a scree plot is a line plot of the eigenvalues of factors or principal components in an analysis.[1] The scree plot is used to determine the number of factors to retain in an exploratory factor analysis (FA) or principal components to keep in a principal component analysis (PCA). We could see that starting from the 6th eigen value the eigen values are so small that they don't affect anything in the results.

## # Part Two

```
diab <- read.csv("J:/champions work/datasets/diabetes_prediction_dataset.csv")
```

```
allele1 <- substr(diab$alleles, 1, 1) # First character of alleles
```

```
allele2 <- substr(diab$alleles, 2, 2) # Second character of alleles
```

```
allele_counts <- table(No_Diabetes = c(allele1, allele2), Diabetes = rep(diab$diabetes, 2))
```

```
# Convert table to data frame for easier manipulation
```

```
allele_counts_df <- as.data.frame(allele_counts)
```

```
# Print the resulting table
```

```
print(allele_counts_df)
```

```
##   No_Diabetes Diabetes  Freq
## 1           A         0 91660
## 2           C         0 91340
## 3           A         1  8499
## 4           C         1  8501
```



```

allele_counts <- matrix(c(91660, 8499, 91340, 8501), ncol = 2, byrow = TRUE)

# Perform Fisher's exact test
fisher_result <- fisher.test(allele_counts)

# Extract p-value
p_value <- fisher_result$p.value

# Report the p-value
print(p_value)

```

```
## [1] 0.816139
```

It's not significant at all, .81 is very big. Which means that the association is not significant.

```

# Extract BMI samples for each allele family
BMI_AA <- subset(diab, alleles == "AA")$bmi
BMI_AC <- subset(diab, alleles == "AC")$bmi
BMI_CC <- subset(diab, alleles == "CC")$bmi

# Perform t-test between BMI samples for AA and AC allele families
t_test_AA_AC <- t.test(BMI_AA, BMI_AC)

# Perform t-test between BMI samples for AA and CC allele families
t_test_AA_CC <- t.test(BMI_AA, BMI_CC)

# Perform t-test between BMI samples for AC and CC allele families
t_test_AC_CC <- t.test(BMI_AC, BMI_CC)

# Report p-values
p_value_AA_AC <- t_test_AA_AC$p.value
p_value_AA_CC <- t_test_AA_CC$p.value
p_value_AC_CC <- t_test_AC_CC$p.value

# Print p-values
print(p_value_AA_AC)

```

```
## [1] 0.5125191
```

```
print(p_value_AA_CC)
```

```
## [1] 0.8228364
```

```
print(p_value_AC_CC)
```

```
## [1] 0.6691533
```

It's clear that there is no real significant difference between the family of alleles when it's related to BMI phenotype.

## # Part Three

There are two mismatches.

Descriptions

Graphic Summary

Alignments

Dot Plot

Sequences producing significant alignments

Download Select columns Show 100

☒ select all 1 sequences selected

GraphicsMSA Viewer

Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<input checked="" type="checkbox"/> None provided		174	174	100%	2e-49	98.00%	100	Query_2135223

Descriptions

Graphic Summary

Alignments

Dot Plot

hover to see the title click to show alignments

Alignment Scores < 40 40 - 50 50 - 80 80 - 200 >= 200

1 sequences selected

Distribution of the top 1 Blast Hits on 1 subject sequences

Descriptions

Graphic Summary

Alignments

Dot Plot

Alignment view Pairwise CDS feature Restore defaults Download

1 sequences selected

Download Graphics

Next Previous Descriptions

Sequence ID: Query\_2135223 Length: 100 Number of Matches: 1

Range 1: 1 to 100 Graphics

Next Match Previous Match

Score	Expect	Identities	Gaps	Strand
174 bits(94)	2e-49	98/100(98%)	0/100(0%)	Plus/Plus

Query 1

G G G C A G G A G C C A G G G C T G G G C A T A A A A G T C A G G G C A G A G C C A T C T A T T G C T T A C A T T T G C

60

Sbjct 1

G G G C A G G A G C C A G G G C T G G G C A T A A A A G T C A G G G C A G A G C C A T C T A T T G C T T A C A C T T G C

60

Query 61

T T C T G A C A C A A C T G T G T T C A C T A G C A A C C T C A A A C A G A C A

100

Sbjct 61

T T C T G A C A C A A C T G T G T T C A C G A G C A A C C T C A A A C A G A C A

100

Descriptions

Graphic Summary

Alignments

Dot Plot

Plot of lcl|Query\_2135221 vs lcl|Query\_2135223

Descriptions

Graphic Summary

Alignments

Dot Plot

```
seq1 <- entrez_fetch(db = "nucleotide", id = "NG_050578.1", rettype = "fasta")
seq2 <- entrez_fetch(db = "nucleotide", id = "X03562.1", rettype = "fasta")
seq1
```

```
## [1] ">NG_050578.1 Homo sapiens INS-IGF2 readthrough (INS-IGF2), RefSeqGene on chromosome 11\nGAGGTGC
```

```
seq1_lines <- unlist(strsplit(seq1, "\n"))
seq1_lines <- seq1_lines[!grepl(">", seq1_lines)]
seq1_clean <- paste(seq1_lines, collapse = "")

seq2_lines <- unlist(strsplit(seq2, "\n"))
seq2_lines <- seq2_lines[!grepl(">", seq2_lines)]
seq2_clean <- paste(seq2_lines, collapse = "")

seq1 <- DNASTringSet(seq1_clean)
seq2 <- DNASTringSet(seq2_clean)

print(seq1)
```

```
## DNASTringSet object of length 1:
##      width seq
## [1] 39098 GAGGTGCGGATCCTGGGCGGCCAGGAAGGTCTC...CCATCCCTCCACTCATCCATCCACTCATCCCTC
```

```
print(seq2)
```

```
## DNASTringSet object of length 1:
##      width seq
## [1] 8837 CCCAACCCGCGCACAGCGGGCACTGGTTTCGGG...TCTCCCTTCTCACGGGAATTTTCAGGGTAAACT
```

```
freq_seq1 <- alphabetFrequency(seq1)
freq_seq2 <- alphabetFrequency(seq2)
print(freq_seq1)
```

```
##      A      C      G      T M R W S Y K V H D B N - + .
## [1,] 7355 12386 11660 7697 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
```

```
print(freq_seq2)
```

```
##      A      C      G      T M R W S Y K V H D B N - + .
## [1,] 1388 3037 2697 1685 0 0 0 0 0 0 0 0 0 0 0 30 0 0 0
```

```
seq1_has_gaps_ambiguous <- any(names(freq_seq1) %in% c("-", "N"))
seq2_has_gaps_ambiguous <- any(names(freq_seq2) %in% c("-", "N"))
print(seq1_has_gaps_ambiguous)
```

```
## [1] FALSE
```

```
print(seq2_has_gaps_ambiguous)
```

```
## [1] FALSE
```

this means that the two sequences are clean

```
seq1_has_gaps_ambiguous <- any(grepl("[-N]", seq1))
seq2_has_gaps_ambiguous <- any(grepl("[-N]", seq2))
```

```
# 2. Remove gaps and ambiguous bases from sequences
```

```
seq1_cleaned <- gsub("[-N]", "", seq1)
seq2_cleaned <- gsub("[-N]", "", seq2)
seq1_length_before <- nchar(seq1)
seq2_length_before <- nchar(seq2)
seq1_length_after <- nchar(seq1_cleaned)
seq2_length_after <- nchar(seq2_cleaned)
print(paste("Sequence 1 length before cleaning:", seq1_length_before))
```

```
## [1] "Sequence 1 length before cleaning: 39098"
```

```
print(paste("Sequence 2 length before cleaning:", seq2_length_before))
```

```
## [1] "Sequence 2 length before cleaning: 8837"
```

```
print(paste("Sequence 1 length after cleaning:", seq1_length_after))
```

```
## [1] "Sequence 1 length after cleaning: 39098"
```

```
print(paste("Sequence 2 length after cleaning:", seq2_length_after))
```

```
## [1] "Sequence 2 length after cleaning: 8807"
```

For confirmation, now we're 100% percent that they were clean. We'll work with the seq1, seq2.

```
run_pairwise_alignment <- function(seq1, seq2) {
  # Run Pairwise Local Alignment
  alignment <- pairwiseAlignment(seq1, seq2, substitutionMatrix = "BLOSUM62", gapOpening = -10, gapExtension = -1)

  # Extract alignment score
  alignment_score <- score(alignment)

  # Calculate width of each sequence before and after alignment
  seq1_width_before <- nchar(seq1)
  seq2_width_before <- nchar(seq2)
  seq1_width_after <- nchar(pattern(alignment))
  seq2_width_after <- nchar(subject(alignment))

  # Return results
  return(list(

```

```

    alignment_score = alignment_score,
    seq1_width_before = seq1_width_before,
    seq2_width_before = seq2_width_before,
    seq1_width_after = seq1_width_after,
    seq2_width_after = seq2_width_after,
    alignment = alignment
  ))
}

```

```
alignment_result <- run_pairwise_alignment(seq1, seq2)
```

```

# Report results
print("Alignment Score:")

```

```
## [1] "Alignment Score:"
```

```
print(alignment_result$alignment_score)
```

```
## [1] 25861
```

```
print("Width of Sequences Before Alignment:")
```

```
## [1] "Width of Sequences Before Alignment:"
```

```
print(paste("Sequence 1:", alignment_result$seq1_width_before))
```

```
## [1] "Sequence 1: 39098"
```

```
print(paste("Sequence 2:", alignment_result$seq2_width_before))
```

```
## [1] "Sequence 2: 8837"
```

```
print("Width of Sequences After Alignment:")
```

```
## [1] "Width of Sequences After Alignment:"
```

```
print(paste("Sequence 1:", alignment_result$seq1_width_after))
```

```
## [1] "Sequence 1: 8920"
```

```
print(paste("Sequence 2:", alignment_result$seq2_width_after))
```

```
## [1] "Sequence 2: 8920"
```

```
print(nmismatch(alignment_result$alignment))
```

```
## [1] 98
```

```
print(mismatchTable(alignment_result$alignment))
```

##	PatternId	PatternStart	PatternEnd	PatternSubstring	SubjectStart	SubjectEnd
## 1	1	25041	25041	C	58	58
## 2	1	25042	25042	C	59	59
## 3	1	25221	25221	T	238	238
## 4	1	25222	25222	T	239	239
## 5	1	25449	25449	T	466	466
## 6	1	25727	25727	G	733	733
## 7	1	25759	25759	G	765	765
## 8	1	25780	25780	A	785	785
## 9	1	25794	25794	G	799	799
## 10	1	26198	26198	G	1200	1200
## 11	1	26262	26262	C	1264	1264
## 12	1	26263	26263	G	1265	1265
## 13	1	26533	26533	A	1535	1535
## 14	1	26755	26755	G	1756	1756
## 15	1	28038	28038	C	3036	3036
## 16	1	28184	28184	C	3178	3178
## 17	1	28185	28185	T	3179	3179
## 18	1	28186	28186	G	3180	3180
## 19	1	28187	28187	G	3181	3181
## 20	1	28188	28188	C	3182	3182
## 21	1	28189	28189	G	3183	3183
## 22	1	28190	28190	G	3184	3184
## 23	1	28191	28191	C	3185	3185
## 24	1	28192	28192	G	3186	3186
## 25	1	28193	28193	G	3187	3187
## 26	1	28194	28194	A	3188	3188
## 27	1	28195	28195	G	3189	3189
## 28	1	28198	28198	G	3190	3190
## 29	1	28199	28199	G	3191	3191
## 30	1	28200	28200	G	3192	3192
## 31	1	28201	28201	G	3193	3193
## 32	1	28202	28202	G	3194	3194
## 33	1	28203	28203	T	3195	3195
## 34	1	28204	28204	G	3196	3196
## 35	1	28205	28205	G	3197	3197
## 36	1	28206	28206	G	3198	3198
## 37	1	28207	28207	G	3199	3199
## 38	1	28208	28208	T	3200	3200
## 39	1	28209	28209	G	3201	3201
## 40	1	28210	28210	G	3202	3202
## 41	1	28211	28211	G	3203	3203
## 42	1	28213	28213	G	3205	3205
## 43	1	28218	28218	G	3210	3210
## 44	1	28249	28249	C	3217	3217
## 45	1	28383	28383	A	3341	3341

## 46	1	28426	28426	T	3386	3386
## 47	1	28483	28483	T	3443	3443
## 48	1	28626	28626	A	3588	3588
## 49	1	28627	28627	G	3589	3589
## 50	1	28628	28628	T	3590	3590
## 51	1	28629	28629	G	3591	3591
## 52	1	28634	28634	G	3596	3596
## 53	1	28638	28638	G	3600	3600
## 54	1	28695	28695	C	3661	3661
## 55	1	28829	28829	A	3793	3793
## 56	1	28856	28856	G	3819	3819
## 57	1	28930	28930	C	3891	3891
## 58	1	29345	29345	T	4309	4309
## 59	1	29346	29346	T	4310	4310
## 60	1	29467	29467	A	4431	4431
## 61	1	29510	29510	C	4478	4478
## 62	1	29511	29511	T	4479	4479
## 63	1	29647	29647	T	4614	4614
## 64	1	29662	29662	G	4629	4629
## 65	1	29728	29728	G	4694	4694
## 66	1	29834	29834	C	4798	4798
## 67	1	30198	30198	A	5160	5160
## 68	1	30210	30210	G	5172	5172
## 69	1	30212	30212	A	5174	5174
## 70	1	30213	30213	T	5175	5175
## 71	1	30284	30284	G	5245	5245
## 72	1	30396	30396	G	5356	5356
## 73	1	30853	30853	T	5813	5813
## 74	1	30905	30905	G	5866	5866
## 75	1	30910	30910	A	5871	5871
## 76	1	31090	31090	C	6051	6051
## 77	1	31091	31091	T	6052	6052
## 78	1	31092	31092	C	6053	6053
## 79	1	31227	31227	C	6188	6188
## 80	1	31587	31587	C	6548	6548
## 81	1	31897	31897	T	6855	6855
## 82	1	32135	32135	T	7096	7096
## 83	1	32165	32165	G	7126	7126
## 84	1	32323	32323	G	7282	7282
## 85	1	32325	32325	C	7284	7284
## 86	1	32342	32342	T	7301	7301
## 87	1	32395	32395	A	7354	7354
## 88	1	32434	32434	T	7393	7393
## 89	1	32473	32473	C	7432	7432
## 90	1	32474	32474	C	7433	7433
## 91	1	32475	32475	C	7434	7434
## 92	1	32476	32476	C	7435	7435
## 93	1	32477	32477	C	7436	7436
## 94	1	32481	32481	A	7440	7440
## 95	1	32501	32501	A	7461	7461
## 96	1	33806	33806	A	8765	8765
## 97	1	33835	33835	T	8794	8794
## 98	1	33836	33836	G	8795	8795
##	SubjectSubstring					

## 1	G
## 2	T
## 3	N
## 4	N
## 5	C
## 6	N
## 7	A
## 8	G
## 9	A
## 10	A
## 11	G
## 12	C
## 13	G
## 14	C
## 15	G
## 16	N
## 17	N
## 18	N
## 19	N
## 20	N
## 21	N
## 22	N
## 23	N
## 24	N
## 25	N
## 26	N
## 27	N
## 28	N
## 29	N
## 30	N
## 31	N
## 32	N
## 33	N
## 34	N
## 35	N
## 36	N
## 37	N
## 38	N
## 39	N
## 40	N
## 41	N
## 42	A
## 43	C
## 44	A
## 45	G
## 46	C
## 47	C
## 48	C
## 49	A
## 50	G
## 51	T
## 52	A
## 53	T
## 54	A



## 55	T
## 56	T
## 57	T
## 58	C
## 59	G
## 60	G
## 61	T
## 62	C
## 63	A
## 64	A
## 65	A
## 66	T
## 67	G
## 68	T
## 69	G
## 70	A
## 71	A
## 72	C
## 73	A
## 74	T
## 75	T
## 76	T
## 77	C
## 78	T
## 79	T
## 80	A
## 81	C
## 82	C
## 83	C
## 84	A
## 85	T
## 86	C
## 87	G
## 88	N
## 89	G
## 90	G
## 91	G
## 92	G
## 93	G
## 94	G
## 95	G
## 96	G
## 97	G
## 98	T