# Ovarian Cancer Histotypes: Report of Statistical Findings

Derek Chiu

June 20, 2025

# Table of contents

Pi	rerace	e	O
1	Intr	roduction	7
2	Met	thods	8
	2.1	Pre-Processing	8
		2.1.1 Case Selection	8
		2.1.2 Quality Control	8
		2.1.3 Housekeeping Genes Normalization	9
		2.1.4 Between CodeSet and Site Normalization	9
		2.1.5 Final Processing	10
	2.2	Classifiers	11
		2.2.1 Resampling of Training Set	12
		2.2.2 Hyperparameter Tuning	12
			12
			13
	2.3		13
		• 9	14
	2.4	66 6 6	15
			16
	2.5	00 0 0	17
			17
			19
	2.6		19
	2.7	66 6	19
		*	21
		2.1.1 variable importance	
3	Dist	tributions	23
	3.1	Histotype Distribution	23
	3.2	Cohort Distribution	25
	3.3	Quality Control	25
		3.3.1 Failed Samples	25
		3.3.2 %GD vs. SNR	27
	3.4		29
1	Res	valta.	33
4	4.1		აა 34
	4.1		
		v	34 36
		U	30 38
		1	38 40
		4 1 4 P 1-000PP	411

	4.1.5	Balanced Accuracy	42
	4.1.6	Kappa	44
4.2	Rank .	Aggregation	45
	4.2.1	Across Classes	46
	4.2.2	Across Metrics	49
	4.2.3	Top Workflows	49
4.3	Optim	al Gene Sets	53
	4.3.1	Sequential Algorithm	53
	4.3.2	SMOTE-Random Forest	56
	4.3.3	Two-Step	60
4.4	Test S	et Performance	63
	4.4.1	Confirmation Set	65
	4.4.2	Validation Set	73
Refere	nces		79

# List of Figures

2.1	Venn diagram of common and unique gene targets covered by each CodeSet 1	0
2.2	Cohorts Selection	1
2.3	Visualization of Subsampling Techniques	3
2.4	Two-Step Algorithm	4
2.5	Aggregating Predictions for Two-Step Algorithm	5
2.6	Sequential Algorithm	6
2.7	Aggregating Predictions for Sequential Algorithm	7
3.1	% Genes Detected vs. Signal to Noise Ratio	7
3.2	% Genes Detected vs. Signal to Noise Ratio (Zoomed)	8
3.3	Random1-Normalized CS1 vs. CS3 Gene Expression	9
3.4	Random1-Normalized CS2 vs. CS3 Gene Expression	0
3.5	HKgenes-Normalized CS1 vs. CS3 Gene Expression	1
3.6	HKgenes-Normalized CS2 vs. CS3 Gene Expression	2
4.1	Training Set Mean Accuracy	5
4.2	Training Set Mean Sensitivity	7
4.3	Training Set Mean Specificity	9
4.4	Training Set Mean F1-Score	1
4.5	Training Set Mean Balanced Accuracy	3
4.6	Training Set Mean Kappa	5
4.7	Top 5 Workflow Per-Class Evaluation Metrics by Metric	
4.8	Top 5 Workflow Per-Class Evaluation Metrics by Metric	2
4.9	Gene Optimization for Sequential Classifier	3
4.10	· · · · · · · · · · · · · · · · · · ·	6
4.11	Gene Optimization for Two-Step Classifier	0
	Entropy vs. Predicted Probability in Confirmation Set 6	6
	Gene Optimized Workflows Per-Class Metrics in Confirmation Set 6	6
	Confusion Matrices for Confirmation Set Models	
	ROC Curves for Sequential Full Model in Confirmation Set 6	
	ROC Curves for Sequential, Optimal Model in Confirmation Set 6	9
	ROC Curves for SMOTE-Random Forest, Full Set Model in Confirmation Set 7	
	ROC Curves for SMOTE-Random Forest, Optimal Set Model in Confirmation Set . 7	1
4.19	ROC Curves for Two-Step Full Model in Confirmation Set	2
4.20	ROC Curves for Two-Step Optimal Model in Confirmation Set	3
4.21	SMOTE-Random Forest Per-Class Metrics in Validation Set	4
	Confusion Matrix for Validation Set Model	4
4.23	ROC Curves for SMOTE-Random Forest, Optimal Set Model in Validation Set $$ 7	5
	Volcano Plots of Validation Set Predictions	7
4 25	Subtype Prediction Summary among Predicted HGSC Samples 7	8

# List of Tables

2.1	Gene Distribution	20
3.1	Histotype Distribution in Training Set by Processing Stage	23
3.2		24
3.3	Pre-QC Cohort Distribution by CodeSet	25
3.4	Quality Control Summary	26
3.5		32
4.1	Training Set Mean Accuracy	34
4.2	Training Set Mean Sensitivity	36
4.3	Training Set Mean Specificity	38
4.4	Training Set Mean F1-Score	40
4.5	Training Set Mean Balanced Accuracy	42
4.6	Training Set Mean Kappa	44
4.7	F1-Score Rank Aggregation Summary	46
4.8	Balanced Accuracy Rank Aggregation Summary	47
4.9	Kappa Rank Aggregation Summary	48
4.10	Rank Aggregation Comparison of Metrics Used	49
4.11	Top 5 Workflows from Final Rank Aggregation	49
4.12	Top Workflow Per-Class Evaluation Metrics	50
4.13	Top Workflow Per-Class Evaluation Metrics and Ranks	52
4.14	Gene Profile of Optimal Set in Sequential Algorithm	54
4.15	Gene Profile of Optimal Set in SMOTE-Random Forest Workflow	57
4.16	Gene Profile of Optimal Set in Two-Step Workflow	60
4.17	Evaluation Metrics on Confirmation Set Models	65
4.18	Evaluation Metrics on Validation Set Model, SMOTE-Random Forest, Optimal Set .	73
4.19	Clinicopath characteristics between correct and incorrect predictions of ENOC cases	76

# Preface

This report of statistical findings describes the classification of ovarian cancer histotypes using data from NanoString CodeSets.

Marina Pavanello conducted the initial exploratory data analysis, Cathy Tang implemented class imbalance techniques, Derek Chiu conducted the normalization and statistical analysis, and Lauren Tindale and Aline Talhouk are the project leads.

## 1 Introduction

Ovarian cancer has five major histotypes: high-grade serous carcinoma (HGSC), low-grade serous carcinoma (LGSC), endometrioid carcinoma (ENOC), mucinous carcinoma (MUC), and clear cell carcinoma (CCOC). A common problem with classifying these histotypes is that there is a class imbalance issue. HGSC dominates the distribution, commonly accounting for 70% of cases in many patient cohorts, while the other four histotypes are spread over the rest of the cases. Subsampling methods like up-sampling, down-sampling, and SMOTE can be used to mitigate this problem.

The supervised learning is performed under a consensus framework: we consider various classification algorithms and use evaluation metrics like accuracy, F1-score, and Kappa, to inform the decision of which methods to carry forward for prediction in confirmation and validation sets.

## 2 Methods

## 2.1 Pre-Processing

#### 2.1.1 Case Selection

Prior to pre-processing, samples were split into a training, a confirmation, and a validation set.

- Training
  - CS1: OOU, OOUE, VOA, MAYO, MTL
  - CS2: OOU, OOUE, VOA, MAYO, OVAR3, OVAR11, JAPAN, MTL, POOL-CTRL
  - CS3: OOU, OOUE, VOA, POOL-1, POOL-2, POOL-3
- Confirmation:
  - CS3: TNCO
- Validation:
  - CS3: DOVE4

#### 2.1.2 Quality Control

Before normalization, we calculated several quality control measures and excluded samples that failed to achieve sample quality in one or more of these measures.

- Linearity of positive control genes: If the R-squared from a linear model of positive controls and their concentrations is less than 0.95 or missing, then the sample is flagged.
- Imaging quality: The sample is flagged if the field of view percentage is less than 75%.
- Positive Control flag: We consider the two smallest positive controls at concentrations 0.5 and 1. If these two controls are less than the lower limit of detection (defined as two standard deviations below the mean of the negative control expression), or if the mean negative control expression is 0, the sample is flagged.
- The signal-to-noise ratio or percent of genes detected: These two measures are defined as the ratio of the average housekeeping gene expression over the upper limit of detection, defined as two standard deviations above the mean of the negative control expression (or 0 if this limit is less than 0.001), and the proportion of endogenous genes with expression greater than the upper limit of detection. These measures are flagged if they are below a prespecified threshold, which is determined visually by considering their bivariate distribution in a scatterplot. In this case, we used 100 for the SNR threshold and 50% for the threshold for genes detected. Note: these thresholds were determined by examining the relationship in Section 3.3.2.

#### 2.1.3 Housekeeping Genes Normalization

The full training set (n=1257) comprised of data from three CodeSets (CS) 1, 2, and 3. Data normalization removes technical variation from high-throughput platforms to improve the validity of comparative analyses.

Each CodeSet was first normalized to housekeeping genes: ACTB, RPL19, POLR1B, SDHA, and PGK1. Housekeeping genes encode proteins responsible for basic cell function and have consistent expression in all cells. All expression values were log2 transformed. Normalization to housekeeping genes corrects the viable RNA from each sample. This is achieved by subtracting the average log (base 2)-transformed expression of the housekeeping genes from the log (base 2)-transformed expression of each gene:

$$log_2({\rm endogenous\ gene\ expression}) - {\rm average}(log_2({\rm housekeeping\ gene\ expression})) = {\rm relative\ expression})$$

#### 2.1.4 Between CodeSet and Site Normalization

To normalize between CodeSets, we randomly selected five specimens, one from each histotype, among specimens repeated in all three CodeSets. This formed the reference set (Random 1). We selected only one sample from each histotype to use as few samples as possible for normalization and retain the rest for analysis.

A reference-based approach (Talhouk et al. (2016)) was used to normalize CS1 to CS3 and CS2 to CS3 across their common genes:

$$X-Norm_{CS1} = X_{CS1} + \bar{R}_{CS3} - \bar{R}_{CS1}X-Norm_{CS2} = X_{CS2} + \bar{R}_{CS3} - \bar{R}_{CS2}$$
 (2.2)

Samples in CS3 were processed at three different locations; we also had to normalize for "site" in this CodeSet. Finally, the CS3 expression samples were included in the training set without further normalization:

$$\text{X-Norm}_{\text{CS3-USC}} = X_{\text{CS3-USC}} + \bar{R}_{\text{CS3-VAN}} - \bar{R}_{\text{CS3-USC}} \\ \text{X-Norm}_{\text{CS3-AOC}} = X_{\text{CS3-AOC}} + \bar{R}_{\text{CS3-VAN}} - \bar{R}_{\text{CS3-AOC}} \\ (2.3)$$

Finally, the CS3 expression samples were included in the training set without further normalization. The initial training set is assembled by combining all four of the previously mentioned normalized datasets along with the two CS3 expression subsets not used in normalization:

$$\begin{aligned} \text{Training Set} &= \text{X-Norm}_{\text{CS1}} + \text{X-Norm}_{\text{CS2}} + \text{X-Norm}_{\text{CS3-USC}} + \text{X-Norm}_{\text{CS3-AOC}} + \text{X-Norm}_{\text{CS3}} + \text{X-Norm}_{\text{CS3-VAN}} \\ &= \text{X-Norm}_{\text{CS1}} + \text{X-Norm}_{\text{CS2}} + \text{X-Norm}_{\text{CS3}} \end{aligned}$$



Figure 2.1: Venn diagram of common and unique gene targets covered by each CodeSet

#### 2.1.5 Final Processing

We map ovarian histotypes to all remaining samples and keep the major histotypes for building the predictive model: high-grade serous carcinoma (HGSC), clear cell ovarian carcinoma (CCOC), endometrioid ovarian carcinoma (ENOC), low-grade serous carcinoma (LGSC), mucinous carcinoma (MUC).

Duplicate cases (two samples with the same ottaID) were removed before generating the final training set to use for fitting the classification models. All CS3 cases were preferred over CS1

and CS2, and CS3-Vancouver cases were preferred over CS3-AOC and CS3-USC when selecting duplicates.

The final training set used only genes that were common across all three CodeSets.

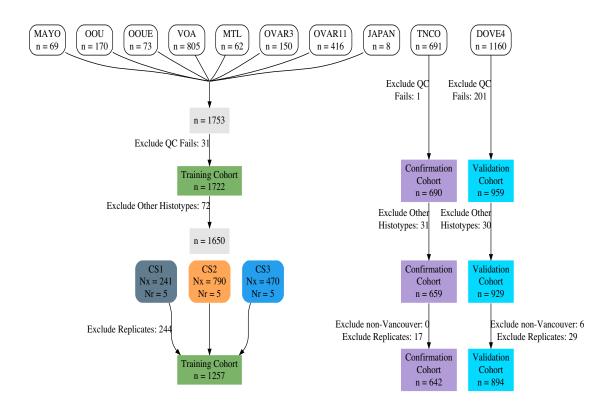


Figure 2.2: Cohorts Selection

#### 2.2 Classifiers

We use 4 classification algorithms in the supervised learning framework for the Training Set. The pipeline was run using SLURM batch jobs submitted to a partition on a CentOS 7 server. All resampling techniques, pre-processing, model specification, hyperparameter tuning, and evaluation metrics were implemented using the tidymodels suite of packages. The classifiers we used are:

- Random Forest (rf)
- Support Vector Machine (svm)
- XGBoost (xgb)
- Regularized Multinomial Regression (mr)

### 2.2.1 Resampling of Training Set

We used a nested cross-validation design to assess each classifier while also performing hyperparameter tuning. An outer 5-fold CV stratified by histotype was used together with an inner 5-fold CV with 2 repeats stratified by histotype. This design was chosen such that the test sets of the inner resamples would still have a reasonable number of samples belonging to the smallest minority class.

The outer resampling method cannot be the bootstrap, because the inner training and inner test sets will likely contain the same samples as a result of sampling with replacement in the outer training set. This phenomenon might result in inflated performance as some observations are used both to train and evaluate the hyperparameter tuning in the inner loop.

#### 2.2.2 Hyperparameter Tuning

The following specifications for each classifier were used for tuning hyperparameters:

- rf and xgb: The number of trees were fixed at 500. Other hyperparameters were tuned across 10 randomly selected points in a latin hypercube design.
- svm: Both the cost and sigma hyperparameters were tuned across 10 randomly selected points in a latin hypercube design. We tuned the cost parameter in the range [1, 8]. The range for tuning the sigma parameter was obtained from the 10% and 90% quantiles of the estimation using the kernlab::sigest() function.
- mr: We generated 10 randomly selected points in a latin hypercube design for the penalty (lambda) parameter. Then, we generated 10 evenly spaced points in [0, 1] for the mixture (alpha) parameter in the regularized multinomial regression model. These two sets of 10 points were crossed to generate a tuning grid of 100 points.

The hyperparameter combination that resulted in the highest average F1-score across the inner training sets was selected for each classifier to use as the model for assessing prediction performance in the outer training loop.

#### 2.2.3 Subsampling

Here are the specifications of the subsampling methods used to handle class imbalance:

- None: No subsampling is performed
- Down-sampling: All levels except the minority class are sampled down to the same frequency as the minority class
- Up-sampling: All levels except the majority class are sampled up to the same frequency as the majority class
- SMOTE: All levels except the majority class have synthetic data generated until they have the same frequency as the majority class
- Hybrid: All levels except the majority class have synthetic data generated up to 50% of the frequency of the majority class, then the majority class is sampled down to the same frequency as the rest.

The figure below helps visualize how the distribution of classes changes when we apply subsampling techniques to handle class imbalance:

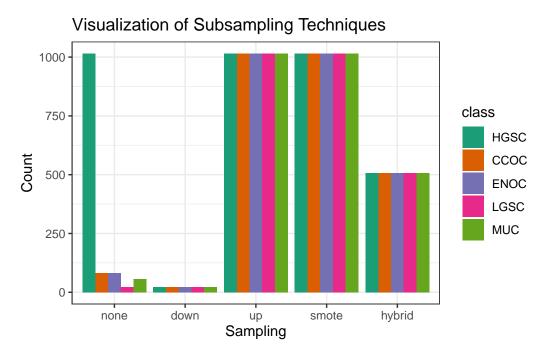


Figure 2.3: Visualization of Subsampling Techniques

#### 2.2.4 Workflows

The 4 algorithms and 5 subsampling methods are crossed to create 20 different classification workflows. For example, the hybrid\_xgb workflow is a classifier that first pre-processes a training set by applying a hybrid subsampling method, and then proceeds to use the XGBoost algorithm to classify ovarian histotypes.

## 2.3 Two-Step Algorithm

The HGSC histotype comprises of approximately 80% of cases among ovarian carcinoma patients, while the remaining 20% of cases are relatively, evenly distributed among ENOC, CCOC, LGSC, and MUC histotypes. We can implement a two-step algorithm as such:

- Step 1: use binary classification for HGSC vs. non-HGSC
- Step 2: use multinomial classification for the remaining non-HGSC classes

Let

$$\begin{split} X_k &= \text{Training data with k classes} \\ C_k &= \text{Class with highest } F_1 \text{ score from training } X_k \\ W_k &= \text{Workflow associated with } C_k \end{split} \tag{2.5}$$

Figure 2.4 shows how the two-step algorithm works:



Figure 2.4: Two-Step Algorithm

### 2.3.1 Aggregating Predictions

The aggregation for two-step predictions is quite straightforward:

- 1. Predict HGSC vs. non-HGSC
- 2. Among all non-HGSC cases, predict CCOC vs. LGSC vs. MUC vs. ENOC



Figure 2.5: Aggregating Predictions for Two-Step Algorithm

### 2.4 Sequential Algorithm

Instead of training on k classes simultaneously using multinomial classifiers, we can use a sequential algorithm that performs k-1 one-vs-all binary classifications iteratively to obtain a final prediction of all cases. At each step in the sequence, we classify one class vs. all other classes, where the classes that make up the "other" class are those not equal to the current "one" class and excluding all "one" classes from previous steps. For example, if the "one" class in step 1 was HGSC, the "other" classes would include CCOC, ENOC, LGSC, and MUC. If the "one" class in step 2 was CCOC, the "other" classes include ENOC, LGSC, and MUC.

The order of classes and workflows to use at each step in the sequential algorithm must be determined using a retraining procedure. After removing the data associated with a particular class, we retrain using the remaining data using multinomial classifiers as described before. The class and workflow to use for the next step in the sequence is selected based on the best per-class evaluation metric value (e.g. F1-score).

Figure 2.6 illustrates how the sequential algorithm works for K=5, using ovarian histotypes as an example for the classes.



Figure 2.6: Sequential Algorithm

The subsampling method used in the first step of the sequential algorithm is used in all subsequent steps in order to maintain data pre-processing consistency. As a result, we are only comparing classification algorithms within one subsampling method across the entire sequential algorithm.

#### 2.4.1 Aggregating Predictions

We have to aggregate the one-vs-all predictions from each of the sequential algorithm workflows in order to obtain a final class prediction on a holdout test set. Each sequential workflow has to be assessed on every sample to ensure that cases classified into the "all" class from a previous step of the sequence are eventually assigned a predicted class. For example, say that based on certain class-specific metrics we determined that the order of classes in the sequential algorithm was to predict HGSC vs. non-HGSC, CCOC vs. non-CCOC, LGSC vs. non-LGSC, and then MUC vs. ENOC. Figure 2.7 illustrates how the final predictions are assigned:



Figure 2.7: Aggregating Predictions for Sequential Algorithm

#### 2.5 Performance Evaluation

#### 2.5.1 Class Metrics

We use the accuracy, sensitivity, specificity, F1-score, kappa, balanced accuracy, and geometric mean, as class metrics to measure both training and test performance between different workflows. Multiclass extensions of these metrics can be calculated except for F1-score, where we use macro-averaging to obtain an overall metric. Class-specific metrics are calculated by recoding classes into one-vs-all categories for each class.

#### **2.5.1.1** Accuracy

The accuracy is defined as the proportion of correct predictions out of all cases:

$$accuracy = \frac{TP}{TP + FP + FN + TN} \tag{2.6}$$

#### 2.5.1.2 Sensitivity

Sensitivity is the proportional of correctly predicted positive cases, out of all cases that were truly positive

sensitivity = 
$$\frac{TP}{TP + FN}$$
 (2.7)

#### 2.5.1.3 Specificity

Specificity is the proportional of correctly predicted negative cases, out of all cases that were truly negative.

specificity = 
$$\frac{TN}{TN + FP}$$
 (2.8)

#### 2.5.1.4 F1-Score

The F-measure can be thought of as a harmonic mean between precision and recall:

$$F_{meas} = \frac{(1+\beta^2) \times precision \times recall}{(\beta^2 \times precision) + recall}$$
 (2.9)

The  $\beta$  value can be adjusted to place more weight upon precision or recall. The most common value is  $\beta$  is 1, which is also commonly known as the F1-score. A multiclass extension doesn't exist for the F1-score, so we use macro-averaging to calculate this metric when there are more than two classes. For example, with k classes, the macro-averaged F1-score is equal to:

$$F_{1_{macro}} = \frac{1}{k} \sum_{i=1}^{k} F_{1_i} \tag{2.10}$$

where each  $F_{1i}$  is the F1-score computed frrom recoding classes into k=i vs.  $k\neq i$ .

In situations where there is not at least one predicted case for each of the classes (e.g. for a poor classifier),  $F_{1i}$  is undefined because the per-class precision of class i is undefined. Those  $F_{1i}$  terms are removed from the  $F_{1macro}$  equation and the resulting value may be inflated. Interpreting the F1-score in such a case would be misleading.

#### 2.5.1.5 Balanced Accuracy

Balanced accuracy is the arithmetic mean of sensitivity and specificity.

Balanced Accuracy = 
$$\frac{\text{Sensitivity} + \text{Specificity}}{2}$$
 (2.11)

#### 2.5.1.6 Kappa

Kappa is the defined as:

$$kappa = \frac{p_0 - p_e}{1 - p_e} \tag{2.12}$$

where  $p_0$  is the observed agreement among raters and  $p_e$  is the hypothetical probability of agreement due to random chance.

#### 2.5.2 AUC

The area under the receiver operating curve (AUC) is calculated by adding up the area under the curve formed by plotting sensitivity vs. 1 - specificity. The Hand-till method is used as a multiclass extension for the AUC.

We did not use AUC to measure class-specific training set performance because combining predicted probabilities in a one-vs-all fashion might be potentially misleading. The sum of probabilities that add up to the "other" class is not equivalent to the predicted probability of the "other" class when using a multiclass classifier.

Instead, we only reported ROC curves and their associated AUCs for test set performance among the highest ranked algorithms.

## 2.6 Rank Aggregation

To select the best algorithm, we implemented a two-stage rank aggregation procedure using the Genetic Algorithm. First, we ranked all workflows based on per-class F1-scores, balanced accuracy, and kappa to see which workflows performed well in predicting all five histotypes. Then, we took the ranks from these three performance metrics and performed a second run of rank aggregation. The top 5 workflows were determined from the final rank aggregation result.

## 2.7 Gene Optimization

We want to discover an optimal set of genes for the classifiers while including specific genes from other studies such as PrOTYPE and SPOT. A total of 72 genes are used in the classifier training set.

There are 16 genes in the classifier set that overlap with the PrOTYPE classifier: COL11A1, CD74, CD2, TIMP3, LUM, CYTIP, COL3A1, THBS2, TCF7L1, HMGA2, FN1, POSTN, COL1A2, COL5A2, PDZK1IP1, FBN1.

There are also 13 genes in the classifier set that overlap with the SPOT signature: HIF1A, CXCL10, DUSP4, SOX17, MITF, CDKN3, BRCA2, CEACAM5, ANXA4, SERPINE1, TCF7L1, CRABP2, DNAJC9.

We obtain a total of 28 genes from the union of PrOTYPE and SPOT genes that we want to include in the final classifier, regardless of model performance. We then incrementally add genes one at a time from the remaining 44 candidate genes based on a variable importance rank to the set of 28 base genes and recalculate performance metrics. The number of genes at which the performance peaks or starts to plateau may indicate an optimal gene set model for us to compare with the full set model.

Here is the breakdown of genes used and whether they belong to the PrOTYPE and/or SPOT sets:

Table 2.1: Gene Distribution

Genes	PrOTYPE	SPOT
TCF7L1	V	v
COL11A1	V	
CD74	V	
CD2	V	
TIMP3	v	
LUM	v	
CYTIP	v	
COL3A1	v	
THBS2	v	
HMGA2	v	
FN1	v	
POSTN	v	
COL1A2	v	
COL5A2	v	
PDZK1IP1	v	
FBN1	v	
HIF1A		$\mathbf{V}$
CXCL10		$\mathbf{V}$
DUSP4		$\mathbf{V}$
SOX17		$\mathbf{V}$
MITF		V
CDKN3		V
BRCA2		$\mathbf{V}$
CEACAM5		$\mathbf{V}$
ANXA4		$\mathbf{V}$
SERPINE1		$\mathbf{V}$
CRABP2		$\mathbf{V}$
DNAJC9		V
C10orf116		
GAD1		
TPX2		
KGFLP2		
EGFL6		
KLK7		
PBX1		

LIN28B

TFF3

MUC5B

FUT3

STC1

BCL2

PAX8

GCNT3

GPR64

ADCYAP1R1

**IGKC** 

BRCA1

IGJ

TFF1

MET

CYP2C18

CYP4B1

SLC3A1

EPAS1

HNF1B

IL6

ATP5G3

DKK4

SENP8

CAPN2

C1orf173

CPNE8

IGFBP1

WT1

TP53

SEMA6A

SERPINA5

ZBED1

TSPAN8

SCGB1D2

LGALS4

MAP1LC3A

#### 2.7.1 Variable Importance

Variable importance is calculated using either a model-based approach if it is available, or a permutation-based VI score otherwise. The variable importance scores are averaged across the outer training folds, and then ranked from highest to lowest.

For the sequential and two-step classifiers, we calculate an overall VI rank by taking the cumulative union of genes at each variable importance rank across all sequences, until all genes have been included.

The variable importance measures are:

- Random Forest: impurity measure (Gini index)
- XGBoost: gain (fractional contribution of each feature to the model based on the total gain of the corresponding features's splits)
- SVM: permutation based p-values
- Multinomial regression: absolute value of estimated coefficients at cross-validated lambda value

# 3 Distributions

## 3.1 Histotype Distribution

Table 3.1: Histotype Distribution in Training Set by Processing Stage

Variable	Levels	CC1	CCO	CC2	Total
		CS1	CS2	CS3	Total
Selected	Cohorts HGSC	128 (44%)	655 (73%)	1808 (73%)	2591 (71%)
		. ,			
	CCOC	48 (16%)	62 (7%)	164 (7%)	274 (7%)
	ENOC	60 (20%)	49 (5%)	250 (10%)	359 (10%)
Histotype	MUC	17 (6%)	58 (6%)	68 (3%)	143 (4%)
	LGSC	19~(6%)	20~(2%)	36 (1%)	75~(2%)
	Other	22 (7%)	59 (7%)	151~(6%)	232~(6%)
Total	N (%)	294 (8%)	903~(25%)	2477~(67%)	$3674\ (100\%)$
$\overline{\mathbf{QC}}$					
-	HGSC	122~(43%)	$641\ (73\%)$	1676~(74%)	2439~(71%)
	CCOC	48 (17%)	62 (7%)	158 (7%)	268 (8%)
	ENOC	60 (21%)	47 (5%)	213 (9%)	320 (9%)
Histotype	MUC	16 (6%)	56 (6%)	65 (3%)	137 (4%)
	LGSC	18 (6%)	20 (2%)	36 (2%)	74 (2%)
	Other	22 (8%)	56 (6%)	125 (5%)	203 (6%)
Total	N (%)	286 (8%)	882 (26%)	2273 (66%)	3441 (100%)
Main His	totypes				
	HGSC	122~(46%)	641~(78%)	1676~(78%)	2439~(75%)
	CCOC	48 (18%)	62 (8%)	158 (7%)	268 (8%)
TT: 4	ENOC	60 (23%)	47 (6%)	213 (10%)	320 (10%)
Histotype	MUC	16 (6%)	56 (7%)	65 (3%)	137 (4%)
	LGSC	18 (7%)	20 (2%)	36 (2%)	74 (2%)
Total	N (%)	264 (8%)	826 (26%)	2148 (66%)	3238 (100%)
Removed	Duplica	ites			
	HGSC	118 (48%)	623~(78%)	1578~(78%)	2319~(76%)

	CCOC	45 (18%)	56 (7%)	146~(7%)	247 (8%)
TT:	ENOC	56 (23%)	43 (5%)	200 (10%)	299 (10%)
Histotype	MUC	13 (5%)	54 (7%)	55 (3%)	122 (4%)
	LGSC	14 (6%)	19 (2%)	32 (2%)	65 (2%)
Total	N (%)	246 (8%)	795 (26%)	2011 (66%)	3052 (100%)
Normaliz	ed and I	Recombined	l		
	HGSC	117 (49%)	622~(79%)	454~(97%)	1193 (79%)
	CCOC	44 (18%)	55 (7%)	4 (1%)	103 (7%)
TT: 4 4	ENOC	55~(23%)	42 (5%)	4 (1%)	101 (7%)
Histotype	MUC	12 (5%)	53 (7%)	4 (1%)	69 (5%)
	LGSC	13 (5%)	18 (2%)	4 (1%)	35 (2%)
Total	N (%)	241 (16%)	790 (53%)	470 (31%)	1501 (100%)
Removed	Replica	tes			
	HGSC	9~(12%)	552~(78%)	454~(97%)	1015~(81%)
	ENOC	38 (49%)	40 (6%)	4 (1%)	82 (7%)
TT: 4 4	CCOC	24 (31%)	53 (7%)	4 (1%)	81 (6%)
Histotype	MUC	3 (4%)	50 (7%)	4 (1%)	57 (5%)
	LGSC	3 (4%)	15 (2%)	4 (1%)	22 (2%)
Total	N (%)	77 (6%)	710 (56%)	470 (37%)	1257 (100%)

Table 3.2: Histotype Distribution in Training, Confirmation, and Validation Sets

Levels	Training	Confirmation	Validation
HGSC	1015 (81%)	424~(66%)	699 (78%)
CCOC	81 (6%)	72 (11%)	69 (8%)
ENOC	82 (7%)	107 (17%)	88 (10%)
MUC	57 (5%)	27 (4%)	23 (3%)
LGSC	22~(2%)	12 (2%)	15 (2%)
N (%)	1257~(45%)	642 (23%)	894 (32%)
	HGSC CCOC ENOC MUC LGSC	HGSC 1015 (81%) CCOC 81 (6%) ENOC 82 (7%) MUC 57 (5%) LGSC 22 (2%)	HGSC 1015 (81%) 424 (66%)  CCOC 81 (6%) 72 (11%)  ENOC 82 (7%) 107 (17%)  MUC 57 (5%) 27 (4%)  LGSC 22 (2%) 12 (2%)

### 3.2 Cohort Distribution

Table 3.3: Pre-QC Cohort Distribution by CodeSet

$\mathbf{CodeSet}$	$\mathbf{CS1}$ $N = 294$	$\mathbf{CS2}$ $N = 903$	$\mathbf{CS3}$ $N = 2,477$
Cohort			
OOU	108 (37%)	$43 \ (4.8\%)$	19~(0.8%)
OOUE	32 (11%)	30 (3.3%)	11 (0.4%)
VOA	145 (49%)	122 (14%)	538 (22%)
OVAR3	0 (0%)	150 (17%)	0 (0%)
OVAR11	0 (0%)	416 (46%)	0 (0%)
MAYO	6(2.0%)	63 (7.0%)	0 (0%)
DOVE4	0 (0%)	0 (0%)	1,160 (47%)
TNCO	0 (0%)	0 (0%)	691 (28%)
$\operatorname{MTL}$	3 (1.0%)	59 (6.5%)	0 (0%)
JAPAN	0 (0%)	8 (0.9%)	0 (0%)
POOL-CTRL	0 (0%)	12(1.3%)	0 (0%)
POOL-1	0 (0%)	0 (0%)	$31\ (1.3\%)$
POOL-2	0 (0%)	0 (0%)	14~(0.6%)
POOL-3	0 (0%)	0 (0%)	13 (0.5%)
1 (0-1)			

<sup>&</sup>lt;sup>1</sup> n (%)

## 3.3 Quality Control

### 3.3.1 Failed Samples

We use an aggregated QCFlag that considers a sample to have failed QC if any of the following QC conditions are flagged:

- Linearity
- Imaging
- Smallest Positive Control
- Normality

Table 3.4: Quality Control Summary

Quality Control Floor	CS1	CS2	CS3
Quality Control Flag	N = 294	N = 903	N = 2,477
Linearity			
Failed	0~(0%)	4~(0.4%)	0 (0%)
Passed	$294 \ (100\%)$	899 (100%)	$2,477 \ (100\%)$
Imaging			
Failed	3~(1.0%)	0~(0%)	4~(0.2%)
Passed	291~(99%)	903~(100%)	$2,473 \ (100\%)$
Smallest Positive Control			
Failed	0 (0%)	2 (0.2%)	0 (0%)
Passed	$294\ (100\%)$	901 (100%)	$2,477 \ (100\%)$
Normality			
Failed	5~(1.7%)	19 (2.1%)	200~(8.1%)
Passed	289~(98%)	884~(98%)	2,277 (92%)
Overall QC			
Failed	8~(2.7%)	$21\ (2.3\%)$	204~(8.2%)
Passed	286~(97%)	882 (98%)	2,273 (92%)

<sup>&</sup>lt;sup>1</sup> n (%)

### 3.3.2 %GD vs. SNR

## % Genes Detected vs. Signal-to-Noise Ratio



Figure 3.1: % Genes Detected vs. Signal to Noise Ratio

## % Genes Detected vs. Signal-to-Noise Ratio (Zoomed)



Figure 3.2: % Genes Detected vs. Signal to Noise Ratio (Zoomed)

## 3.4 Pairwise Gene Expression

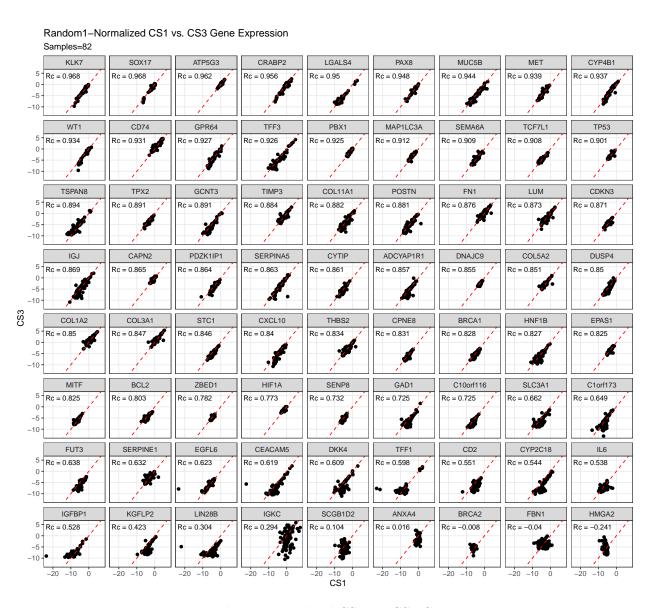


Figure 3.3: Random1-Normalized CS1 vs. CS3 Gene Expression

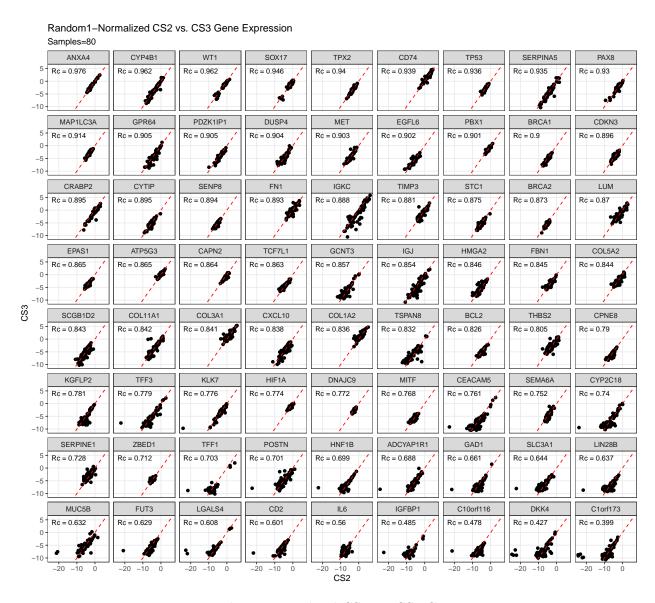


Figure 3.4: Random1-Normalized CS2 vs. CS3 Gene Expression



Figure 3.5: HKgenes-Normalized CS1 vs. CS3 Gene Expression

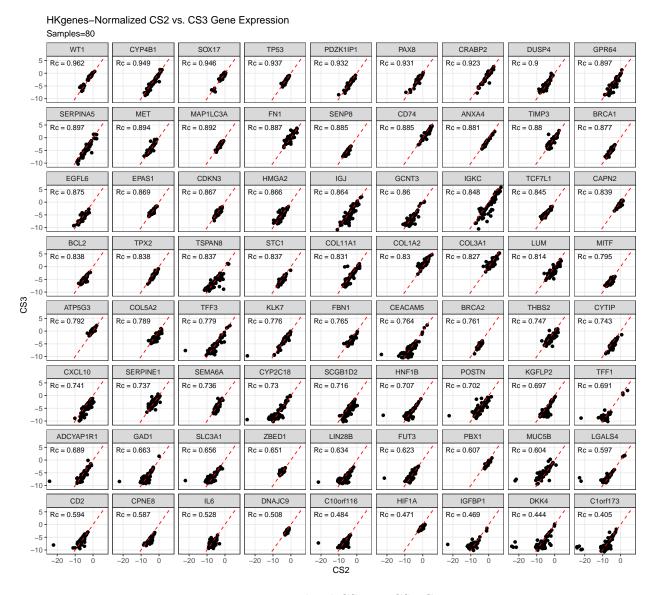


Figure 3.6: HKgenes-Normalized CS2 vs. CS3 Gene Expression

Table 3.5: Wilcoxon signed rank test of gene correlations between normalization methods

Correlation	Housekeeping Genes $N = 72^{1}$	Random1 $N = 72^1$	$\mathbf{p} ext{-}\mathbf{value}^2$
CS1 vs. CS3	$0.84\ (0.72,\ 0.90)$	$0.85 \ (0.64, \ 0.90)$	0.160
CS2 vs. CS3	$0.79 \ (0.69, \ 0.88)$	$0.84 \ (0.73, \ 0.90)$	< 0.001

<sup>&</sup>lt;sup>1</sup>Median (Q1, Q3)

 $<sup>^2\</sup>mathrm{Wilcoxon}$  signed rank test with continuity correction

# 4 Results

We summarize cross-validated training performance of class metrics in the training set. The accuracy, F1-score, and kappa, are the metrics of interest. Workflows are ordered by their mean estimates across the outer folds of the nested CV for each metric.

## 4.1 Training Set

## 4.1.1 Accuracy

Table 4.1: Training Set Mean Accuracy

		Histotypes					
Subsampling	Algorithms	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	rf	0.912	0.935	0.982	0.949	0.982	0.975
	svm	0.925	0.945	0.979	0.962	0.985	0.98
none	xgb	0.81	0.811	0.937	0.935	0.982	0.955
	mr	0.809	0.811	0.934	0.936	0.982	0.955
	rf	0.824	0.873	0.977	0.928	0.92	0.95
	svm	0.803	0.839	0.977	0.905	0.915	0.97
down	xgb	0.694	0.758	0.928	0.921	0.839	0.942
	mr	0.841	0.873	0.979	0.934	0.928	0.967
	rf	0.928	0.958	0.982	0.957	0.983	0.976
	svm	0.916	0.944	0.979	0.955	0.978	0.977
up	xgb	0.923	0.953	0.981	0.958	0.982	0.972
	mr	0.886	0.924	0.977	0.94	0.967	0.963
	$\operatorname{rf}$	0.928	0.955	0.983	0.959	0.982	0.976
	svm	0.916	0.947	0.973	0.953	0.982	0.976
smote	xgb	0.927	0.957	0.98	0.959	0.985	0.972
	mr	0.901	0.935	0.982	0.949	0.969	0.967
	rf	0.917	0.95	0.976	0.953	0.981	0.975
	svm	0.916	0.943	0.979	0.953	0.979	0.977
hybrid	xgb	0.925	0.954	0.982	0.959	0.983	0.972
	mr	0.893	0.927	0.979	0.947	0.964	0.968

#### **Training Set Mean Accuracy** Overall 1.0 + 0.8 0.6 0.4 HGSC 1.0 0.8 0.6 0.4 Algorithms ccoc rf 1.0 svm 0.8 xgb 0.6 mr Accuracy 0.1 **ENOC** Subsampling none 0.8 down 0.6 up 0.4 smote LGSC hybrid 1.0 0.8 0.6 0.4 0.8 0.6 "JP whid too morid sun snote tall none tob 18 to 19 smote mi mbid m UP INT none mi HOWN MI down th Dan't, SALL Workflow

Figure 4.1: Training Set Mean Accuracy

## 4.1.2 Sensitivity

Table 4.2: Training Set Mean Sensitivity

		Histotypes					
Subsampling	Algorithms	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	rf	0.579	0.994	0.79	0.393	0	0.718
	svm	0.674	0.989	0.724	0.642	0.302	0.714
none	xgb	0.208	1	0.04	0	0	0
	mr	0.207	1	0.013	0.022	0	0
down	rf	0.742	0.854	0.886	0.441	0.783	0.743
	svm	0.81	0.808	0.822	0.681	0.95	0.786
	xgb	0.693	0.701	0.873	0.4	0.855	0.636
	mr	0.815	0.851	0.861	0.689	0.855	0.822
	rf	0.687	0.987	0.785	0.648	0.262	0.753
up	svm	0.751	0.962	0.786	0.69	0.548	0.77
	xgb	0.761	0.967	0.819	0.633	0.548	0.839
	$\operatorname{mr}$	0.766	0.922	0.81	0.671	0.648	0.776
	rf	0.712	0.979	0.833	0.646	0.312	0.788
smote	svm	0.744	0.967	0.74	0.646	0.598	0.77
	xgb	0.79	0.965	0.846	0.63	0.655	0.856
	mr	0.776	0.935	0.833	0.691	0.626	0.794
	rf	0.737	0.964	0.808	0.648	0.462	0.803
	svm	0.751	0.963	0.74	0.699	0.598	0.754
hybrid	xgb	0.79	0.964	0.846	0.646	0.655	0.839
	mr	0.796	0.924	0.833	0.657	0.755	0.81

#### **Training Set Mean Sensitivity** Overall 1.00 0.75 0.50 0.25 0.00 **HGSC** 1.00 0.75 0.50 0.25 Algorithms 0.00 rf 1.00 svm 0.75 xgb 0.50 0.25 mr Sensitivity 0.00 **ENOC** Subsampling 1.00 0.75 none 0.50 down 0.25 up 0.00 smote LGSC hybrid 1.00 0.75 0.50 0.25 0.00 MUC 1.00 0.75 0.50 0.25 Jin to but by 0.00 Thorid syn · World A NB SWU JP M down it smote it in tan ship none it hous house int BOWN THE TOP THE BOWN WHITH IN Workflow

Figure 4.2: Training Set Mean Sensitivity

## 4.1.3 Specificity

Table 4.3: Training Set Mean Specificity

				Н	listotypes		
Subsampling	Algorithms	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	rf	0.933	0.694	0.996	0.987	1	0.988
	svm	0.947	0.765	0.996	0.984	0.997	0.993
none	xgb	0.803	0.016	0.999	1	1	1
	mr	0.804	0.021	0.997	1	1	1
	rf	0.956	0.954	0.983	0.962	0.922	0.96
	svm	0.954	0.971	0.987	0.919	0.914	0.979
down	xgb	0.932	0.974	0.932	0.96	0.838	0.957
	mr	0.961	0.962	0.987	0.95	0.93	0.975
	rf	0.96	0.84	0.996	0.978	0.997	0.988
	svm	0.962	0.874	0.991	0.974	0.985	0.988
up	xgb	0.967	0.897	0.991	0.98	0.99	0.979
	mr	0.966	0.935	0.988	0.959	0.973	0.973
	$\operatorname{rf}$	0.963	0.861	0.993	0.98	0.994	0.986
	svm	0.96	0.863	0.989	0.974	0.99	0.987
smote	xgb	0.972	0.921	0.989	0.981	0.99	0.978
	mr	0.968	0.933	0.991	0.967	0.975	0.976
	rf	0.966	0.893	0.987	0.974	0.991	0.983
	svm	0.96	0.862	0.995	0.97	0.986	0.988
hybrid	xgb	0.97	0.91	0.991	0.981	0.989	0.979
	mr	0.967	0.938	0.989	0.967	0.968	0.977

#### **Training Set Mean Specificity** Overall 1.00 0.75 0.50 0.25 0.00 **HGSC** 1.00 0.75 0.50 0.25 Algorithms 0.00 CCOC rf 1.00 svm 0.75 xgb 0.50 0.25 mr Specificity 0.00 1.00 1.00 0.75 **ENOC** Subsampling 0.75 none 0.50 down 0.25 up 0.00 smote LGSC hybrid 1.00 0.75 0.50 0.25 0.00 1.00 0.75 0.50 0.25 one snote ! tan love zhu 0.00 down sun Thorid tab hybrid sym JP Sym smore tolo mybrid A smote mi Two id mi John Mi down th down tob 18 top UP MT. Workflow

Figure 4.3: Training Set Mean Specificity

## 4.1.4 F1-Score

Table 4.4: Training Set Mean F1-Score

				Н	listotypes		
Subsampling	Algorithms	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	rf	0.752	0.961	0.848	0.487	NaN	0.713
	svm	0.723	0.967	0.8	0.673	0.413	0.762
none	xgb	0.749	0.895	0.167	NaN	NaN	NaN
	mr	0.569	0.895	0.042	0.2	NaN	NaN
	rf	0.605	0.916	0.832	0.433	0.27	0.574
	svm	0.635	0.89	0.82	0.478	0.292	0.698
down	xgb	0.511	0.798	0.661	0.425	0.197	0.497
	mr	0.661	0.915	0.844	0.563	0.293	0.692
	rf	0.736	0.974	0.846	0.652	0.392	0.734
	svm	0.729	0.965	0.822	0.661	0.448	0.751
up	xgb	0.736	0.971	0.84	0.648	0.489	0.73
	mr	0.683	0.952	0.815	0.59	0.403	0.657
	rf	0.747	0.972	0.858	0.663	0.421	0.742
	svm	0.73	0.967	0.779	0.637	0.521	0.745
smote	xgb	0.755	0.973	0.84	0.654	0.576	0.733
	mr	0.708	0.959	0.848	0.633	0.417	0.682
	rf	0.718	0.968	0.809	0.632	0.449	0.732
	svm	0.731	0.965	0.813	0.65	0.482	0.746
hybrid	xgb	0.753	0.971	0.852	0.659	0.55	0.729
	mr	0.703	0.953	0.832	0.615	0.422	0.695

#### **Training Set Mean F1-Score** Overall 1.00 0.75 0.50 0.25 0.00 HGSC 1.00 0.75 0.50 0.25 Algorithms 0.00 CCOC rf 1.00 svm 0.75 xgb 0.50 0.25 mr F1-Score 0.00 **ENOC** Subsampling 1.00 0.75 none 0.50 down 0.25 up 0.00 smote **LGSC** hybrid 1.00 0.75 0.50 0.25 0.00 MUC 1.00 0.75 0.50 0.25 note it top sur 0.00 JP Syll ur while sun Twhild told UP TOP hybrid A hybrid mr snote mi HOWN THE 18 j ND Jul down it your tope by

Figure 4.4: Training Set Mean F1-Score

Workflow

## 4.1.5 Balanced Accuracy

Table 4.5: Training Set Mean Balanced Accuracy

				Н	listotypes		
Subsampling	Algorithms	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	rf	0.756	0.844	0.893	0.69	0.5	0.853
	svm	0.811	0.877	0.86	0.813	0.65	0.854
none	xgb	0.506	0.508	0.52	0.5	0.5	0.5
	mr	0.505	0.511	0.505	0.511	0.5	0.5
	rf	0.849	0.904	0.934	0.702	0.852	0.852
	svm	0.882	0.89	0.905	0.8	0.932	0.883
down	xgb	0.813	0.838	0.902	0.68	0.846	0.796
	mr	0.888	0.906	0.924	0.819	0.892	0.898
	rf	0.823	0.913	0.891	0.813	0.629	0.87
	svm	0.857	0.918	0.889	0.832	0.767	0.879
up	xgb	0.864	0.932	0.905	0.806	0.769	0.909
	mr	0.866	0.928	0.899	0.815	0.81	0.875
	$\operatorname{rf}$	0.837	0.92	0.913	0.813	0.653	0.887
	svm	0.852	0.915	0.864	0.81	0.794	0.878
smote	xgb	0.881	0.943	0.917	0.806	0.823	0.917
	mr	0.872	0.934	0.912	0.829	0.801	0.885
	rf	0.851	0.929	0.897	0.811	0.726	0.893
	svm	0.856	0.913	0.867	0.835	0.792	0.871
hybrid	xgb	0.88	0.937	0.919	0.814	0.822	0.909
	mr	0.882	0.931	0.911	0.812	0.861	0.893

#### **Training Set Mean Balanced Accuracy** Overall 1.0 0.9 -8.0 0.7 0.6 0.5 **HGSC** 1.0 0.9 8.0 0.7 0.6 Algorithms 0.5 ccoc rf 1.0 svm 0.9 0.8 0.7 0.6 0.5 xgb **Balanced Accuracy** mr ENOC Subsampling 1.0 · 0.9 · 0.8 · 0.7 · none down 0.6 up 0.5 smote **LGSC** hybrid 1.0 0.9 -0.8 -0.7 0.6 0.5 MUC 1.0 0.9 0.8 0.7 0.6 Boundard top 0.5 Thorid syn whild m mybrid A JP SVM snote syn none sum down sum JR TOP Smote mi UP MI smote A down tob none it none mi none tob 18/

Figure 4.5: Training Set Mean Balanced Accuracy

Workflow

## 4.1.6 Kappa

Table 4.6: Training Set Mean Kappa

				Н	listotypes		
Subsampling	Algorithms	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	rf	0.7	0.768	0.839	0.463	0	0.7
	svm	0.754	0.808	0.789	0.653	0.407	0.752
none	xgb	0.023	0.026	0.062	0	0	0
	mr	0.025	0.034	0.019	0.039	0	0
	rf	0.582	0.663	0.82	0.395	0.249	0.55
	svm	0.565	0.602	0.807	0.432	0.271	0.682
down	xgb	0.447	0.501	0.628	0.308	0.171	0.469
	mr	0.623	0.663	0.833	0.529	0.273	0.675
	rf	0.778	0.861	0.837	0.629	0.308	0.722
	svm	0.754	0.822	0.81	0.637	0.437	0.739
up	xgb	0.773	0.85	0.83	0.625	0.481	0.716
	mr	0.695	0.777	0.802	0.558	0.389	0.638
	rf	0.78	0.856	0.849	0.641	0.33	0.73
	svm	0.749	0.829	0.764	0.612	0.512	0.733
smote	xgb	0.788	0.866	0.83	0.632	0.569	0.719
	mr	0.727	0.803	0.838	0.606	0.404	0.665
	rf	0.759	0.844	0.797	0.607	0.44	0.719
	svm	0.751	0.82	0.802	0.625	0.472	0.734
hybrid	xgb	0.782	0.854	0.842	0.638	0.543	0.715
	mr	0.711	0.784	0.821	0.586	0.408	0.679

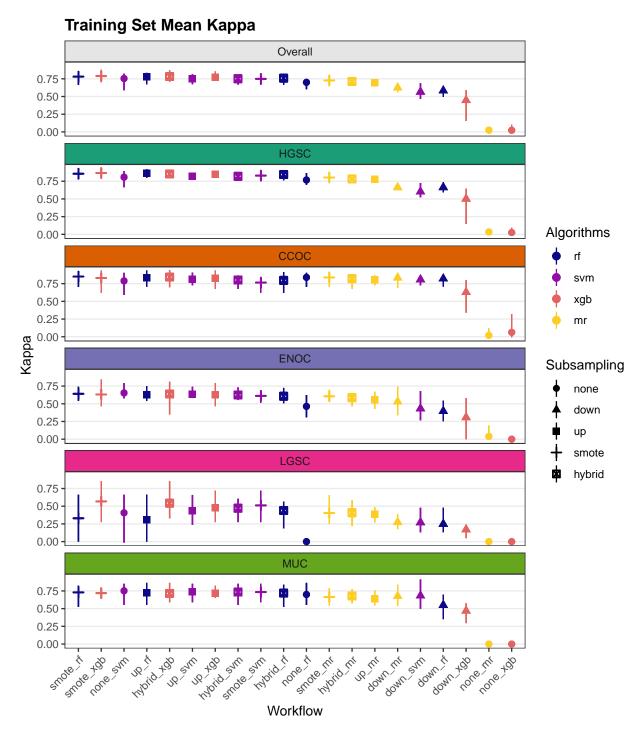


Figure 4.6: Training Set Mean Kappa

## 4.2 Rank Aggregation

Multi-step methods:

• sequential: sequential algorithm sequence of subsampling methods and algorithms used are:

- HGSC vs. non-HGSC using upsubsampling and random forest
- CCOC vs. non-CCOC using SMOTE subsampling and XGBoost
- ENOC vs. non-ENOC using hybrid subsampling and support vector machine
- LGSC vs. MUC using hybrid subsampling and random forest
- two\_step: two-step algorithm sequence of subsampling methods and algorithms used are:
  - HGSC vs. non-HGSC using SMOTE subsampling and random forest
  - CCOC vs. ENOC vs. MUC vs. LGSC using hybrid subsampling and support vector machine

We conduct rank aggregation using a two-stage nested appraoch:

- 1. First we rank aggregate the per-class metrics for F1-score, balanced accuracy and kappa.
- 2. Then we take the aggregated lists from the three metrics and perform a final rank aggregation.
- 3. The top workflows from the final rank aggregation are used for gene optimization in the confirmation set

#### 4.2.1 Across Classes

#### 4.2.1.1 F1-Score

Table 4.7: F1-Score Rank Aggregation Summary

Workflow	Rank	HGSC ∳	ccoc	ENOC ⊕	LGSC ♦	MUC ♦
All	All	All	All	All	All	All
sequential	1	0.97	0.891	0.852	0.92	0.963
two_step	2	0.969	0.865	0.738	0.782	0.864
smote_rf	3	0.972	0.858	0.663	0.421	0.742
hybrid_xgb	4	0.971	0.852	0.659	0.55	0.729
smote_xgb	5	0.973	0.84	0.654	0.576	0.733
up_rf	6	0.974	0.846	0.652	0.392	0.734
hybrid_svm	7	0.965	0.813	0.65	0.482	0.746
up_svm	8	0.965	0.822	0.661	0.448	0.751
smote_svm	9	0.967	0.779	0.637	0.521	0.745
up_xgb	10	0.971	0.84	0.648	0.489	0.73
hybrid_rf	11	0.968	0.809	0.632	0.449	0.732
none_svm	12	0.967	0.8	0.673	0.413	0.762
smote_mr	13	0.959	0.848	0.633	0.417	0.682
hybrid_mr	14	0.953	0.832	0.615	0.422	0.695
up_mr	15	0.952	0.815	0.59	0.403	0.657
down_mr	16	0.915	0.844	0.563	0.293	0.692
down_svm	17	0.89	0.82	0.478	0.292	0.698
down_rf	18	0.916	0.832	0.433	0.27	0.574
down_xgb	19	0.798	0.661	0.425	0.197	0.497

## 4.2.1.2 Balanced Accuracy

Table 4.8: Balanced Accuracy Rank Aggregation Summary

Workflow	Rank 🔷	HGSC ∳	ccoc +	ENOC ∳	LGSC ∳	MUC ∳
All	All	All	All	All	All	All
sequential	1	0.913	0.913	0.858	0.953	0.953
smote_xgb	2	0.943	0.917	0.806	0.823	0.917
hybrid_xgb	3	0.937	0.919	0.814	0.822	0.909
smote_mr	4	0.934	0.912	0.829	0.801	0.885
down_mr	5	0.906	0.924	0.819	0.892	0.898
two_step	6	0.919	0.893	0.819	0.924	0.908
up_xgb	7	0.932	0.905	0.806	0.769	0.909
hybrid_mr	8	0.931	0.911	0.812	0.861	0.893
smote_rf	9	0.92	0.913	0.813	0.653	0.887
up_mr	10	0.928	0.899	0.815	0.81	0.875
down_svm	11	0.89	0.905	0.8	0.932	0.883
up_svm	12	0.918	0.889	0.832	0.767	0.879
hybrid_rf	13	0.929	0.897	0.811	0.726	0.893
smote_svm	14	0.915	0.864	0.81	0.794	0.878
hybrid_svm	15	0.913	0.867	0.835	0.792	0.871
up_rf	16	0.913	0.891	0.813	0.629	0.87
down_rf	17	0.904	0.934	0.702	0.852	0.852
none_svm	18	0.877	0.86	0.813	0.65	0.854
none_rf	19	0.844	0.893	0.69	0.5	0.853
down_xgb	20	0.838	0.902	0.68	0.846	0.796
none_mr	21	0.511	0.505	0.511	0.5	0.5
none_xgb	22	0.508	0.52	0.5	0.5	0.5

## 4.2.1.3 Kappa

Table 4.9: Kappa Rank Aggregation Summary

Workflow	Rank	♦ HGSC ♦	ccoc ∳	ENOC 🔷	LGSC ∳	MUC ∳
All	All	All	All	All	All	All
sequential		1 0.842	0.839	0.715	0.884	0.884
smote_rf		2 0.856	0.849	0.641	0.33	0.73
smote_xgb		3 0.866	0.83	0.632	0.569	0.719
hybrid_xgb		4 0.854	0.842	0.638	0.543	0.715
two_step		5 0.833	0.796	0.632	0.758	0.818
up_svm		6 0.822	0.81	0.637	0.437	0.739
up_xgb		7 0.85	0.83	0.625	0.481	0.716
up_rf		8 0.861	0.837	0.629	0.308	0.722
smote_svm		9 0.829	0.764	0.612	0.512	0.733
hybrid_svm		0 0.82	0.802	0.625	0.472	0.734
hybrid_rf		1 0.844	0.797	0.607	0.44	0.719
none_svm		2 0.808	0.789	0.653	0.407	0.752
smote_mr		3 0.803	0.838	0.606	0.404	0.665
hybrid_mr		4 0.784	0.821	0.586	0.408	0.679
up_mr		5 0.777	0.802	0.558	0.389	0.638
down_mr		6 0.663	0.833	0.529	0.273	0.675
none_rf		7 0.768	0.839	0.463	0	0.7
down_svm		8 0.602	0.807	0.432	0.271	0.682
down_rf		9 0.663	0.82	0.395	0.249	0.55
down_xgb		20 0.501	0.628	0.308	0.171	0.469
none_mr		21 0.034	0.019	0.039	0	0
none_xgb		22 0.026	0.062	0	0	0

## 4.2.2 Across Metrics

Table 4.10: Rank Aggregation Comparison of Metrics Used

Rank	F1	Balanced Accuracy	Kappa
1	sequential	sequential	sequential
2	$two\_step$	$smote\_xgb$	$\operatorname{smote\_rf}$
3	$smote\_rf$	hybrid_xgb	$smote\_xgb$
4	hybrid_xgb	$smote\_mr$	hybrid_xgb
5	$smote\_xgb$	$\operatorname{down\_mr}$	$two\_step$
6	up_rf	$two\_step$	$up\_svm$
7	$hybrid\_svm$	up_xgb	$up\_xgb$
8	$up\_svm$	hybrid_mr	up_rf
9	$smote\_svm$	$\operatorname{smote\_rf}$	$smote\_svm$
10	up_xgb	up_mr	$hybrid\_svm$
11	hybrid_rf	$down\_svm$	hybrid_rf
12	${\rm none\_svm}$	up_svm	${\rm none\_svm}$
13	$smote\_mr$	hybrid_rf	$smote\_mr$
14	$hybrid\_mr$	$smote\_svm$	$hybrid\_mr$
15	$up\_mr$	$hybrid\_svm$	$up\_mr$
16	$\operatorname{down\_mr}$	up_rf	$\operatorname{down\_mr}$
17	$\operatorname{down}\operatorname{\underline{\hspace{1em}}}\operatorname{sym}$	$\operatorname{down\_rf}$	$\mathrm{none}\_\mathrm{rf}$
18	$\operatorname{down}$ _rf	$none\_svm$	$\operatorname{down}\operatorname{\underline{\hspace{1em}}}\operatorname{sym}$
19	$\operatorname{down}$ _xgb	$\mathrm{none}\_\mathrm{rf}$	$\operatorname{down}$ _rf
20	NA	$\operatorname{down}$ _xgb	$\operatorname{down}$ _xgb
21	NA	$none\_mr$	$none\_mr$
22	NA	none_xgb	none_xgb

Table 4.11: Top 5 Workflows from Final Rank Aggregation

Rank	Workflow
1 2	sequential smote rf
3	$\operatorname{smote} \underline{\hspace{0.1cm}} \operatorname{xgb}$
$\frac{4}{5}$	hybrid_xgb two_step

## 4.2.3 Top Workflows

We look at the per-class evaluation metrics of the top 5 workflows.

Table 4.12: Top Workflow Per-Class Evaluation Metrics

				Histotypes		
Metric	Workflow	HGSC	CCOC	ENOC	LGSC	MUC
	sequential	0.951 (0.94, 0.964)	0.929 (0.875, 0.96)	0.857 (0.781, 0.935)	0.95 (0.867, 1)	0.95 (0.867, 1)
	smote_rf	0.955 (0.936, 0.98)	0.983 (0.972, 0.988)	0.959 (0.94, 0.972)	0.982 (0.972, 0.992)	0.976 (0.96, 0.984)
	smote_xgb	0.957 (0.936, 0.98)	0.98 (0.96, 0.992)	0.959 (0.937, 0.976)	0.985 (0.976, 0.992)	0.972 (0.968, 0.98)
Accuracy	hybrid_xgb	0.954 (0.936, 0.968)	0.982 (0.968, 0.992)	0.959 (0.925, 0.972)	0.983 (0.972, 0.992)	0.972 (0.964, 0.988)
	two_step	0.949 (0.924, 0.964)	0.909 (0.826, 0.957)	0.848 (0.783, 0.936)	0.957 (0.935, 0.978)	0.931 (0.891, 0.957)
	sequential	0.975 (0.961, 0.99)	0.863 (0.75, 0.941)	0.817 (0.75, 0.938)	0.96 (0.8, 1)	0.947 (0.818, 1)
	smote_rf	0.979 (0.969, 0.986)	0.833 (0.6, 0.944)	0.646 (0.556, 0.75)	0.312 (0, 0.667)	0.788 (0.462, 0.909)
a	smote_xgb	0.965 (0.951, 0.986)	0.846 (0.6, 0.944)	0.63 (0.444, 0.818)	$0.655 \ (0.25, \ 0.857)$	0.856 (0.615, 1)
Sensitivity	hybrid_xgb	0.964 (0.956, 0.972)	0.846 (0.667, 0.944)	0.646 (0.333, 0.773)	0.655 (0.25, 0.857)	0.839 (0.538, 1)
	two_step	0.967 (0.95, 0.98)	0.839 (0.688, 0.933)	0.754 (0.583, 1)	0.883 (0.667, 1)	0.856 (0.786, 1)
	sequential	0.851 (0.833, 0.875)	0.963 (0.938, 1)	0.899 (0.812, 0.938)	0.947 (0.818, 1)	0.96 (0.8, 1)
	smote_rf	0.861 (0.776, 0.946)	0.993 (0.987, 0.996)	0.98 (0.966, 0.988)	0.994 (0.984, 1)	0.986 (0.979, 0.996)
G 10.1	smote_xgb	0.921 (0.857, 0.965)	0.989 (0.983, 1)	0.981 (0.97, 0.991)	0.99 (0.98, 0.996)	0.978 (0.971, 0.988)
Specificity	hybrid_xgb	0.91 (0.837, 0.947)	0.991 (0.987, 0.996)	0.981 (0.97, 0.991)	0.989 (0.976, 0.996)	0.979 (0.967, 0.992)
	two_step	0.871 (0.766, 0.957)	0.947 (0.9, 1)	0.884 (0.812, 1)	0.966 (0.921, 1)	0.96 (0.917, 1)
	sequential	0.97 (0.963, 0.978)	0.891 (0.8, 0.941)	0.852 (0.774, 0.938)	0.92 (0.8, 1)	0.963 (0.9, 1)
	smote_rf	0.972 (0.959, 0.988)	0.858 (0.72, 0.919)	0.663 (0.571, 0.762)	0.421 (0.222, 0.667)	0.742 (0.545, 0.833)
P4 G	smote_xgb	0.973 (0.96, 0.988)	0.84 (0.643, 0.941)	0.654 (0.5, 0.857)	0.576 (0.286, 0.857)	0.733 (0.667, 0.783)
F1-Score	hybrid_xgb	0.971 (0.96, 0.981)	0.852 (0.714, 0.944)	0.659 (0.387, 0.829)	0.55 (0.333, 0.857)	0.729 (0.609, 0.87)
	two_step	0.969 (0.954, 0.978)	0.865 (0.733, 0.941)	0.738 (0.615, 0.897)	0.782 (0.667, 0.842)	0.864 (0.762, 0.917)
	sequential	0.913 (0.899, 0.93)	0.913 (0.844, 0.955)	0.858 (0.781, 0.935)	0.953 (0.9, 1)	0.953 (0.9, 1)
	smote_rf	0.92 (0.878, 0.966)	0.913 (0.798, 0.968)	0.813 (0.772, 0.864)	0.653 (0.5, 0.831)	0.887 (0.724, 0.946)
D.1. 1.4	smote_xgb	0.943 (0.906, 0.97)	0.917 (0.792, 0.964)	0.806 (0.709, 0.905)	0.823 (0.621, 0.927)	0.917 (0.801, 0.986)
Balanced Accuracy	hybrid_xgb	0.937 (0.899, 0.959)	0.919 (0.827, 0.97)	0.814 (0.652, 0.882)	0.822 (0.623, 0.927)	0.909 (0.763, 0.988)
	two_step	0.919 (0.863, 0.954)	0.893 (0.794, 0.951)	0.819 (0.745, 0.956)	0.924 (0.833, 0.989)	0.908 (0.858, 0.971)
	sequential	0.842 (0.805, 0.881)	0.839 (0.71, 0.911)	0.715 (0.562, 0.871)	0.884 (0.706, 1)	0.884 (0.706, 1)
	smote_rf	0.856 (0.799, 0.922)	0.849 (0.706, 0.912)	0.641 (0.539, 0.74)	0.33 (0, 0.663)	$0.73\ (0.525,\ 0.825)$
**	smote_xgb	0.866 (0.8, 0.922)	0.83 (0.622, 0.937)	0.632 (0.467, 0.844)	0.569 (0.276, 0.853)	0.719 (0.65, 0.772)
Kappa	hybrid_xgb	0.854 (0.797, 0.9)	0.842 (0.697, 0.94)	0.638 (0.348, 0.814)	0.543 (0.326, 0.853)	0.715 (0.59, 0.863)
	two_step	0.833 (0.745, 0.883)	0.796 (0.605, 0.908)	0.632 (0.465, 0.851)	0.758 (0.647, 0.802)	0.818 (0.692, 0.888)

## Top 5 Workflow Per-Class Evaluation Metrics by Metric

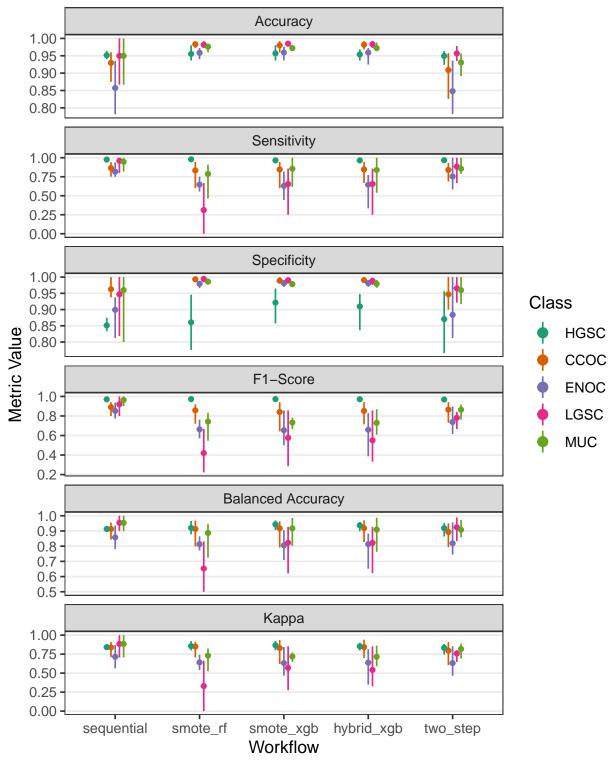


Figure 4.7: Top 5 Workflow Per-Class Evaluation Metrics by Metric

Table 4.13: Top Workflow Per-Class Evaluation Metrics and Ranks

Workflow	Rank	HGSC	CCOC	ENOC	LGSC	MUC
F1-Score						
sequential	1	0.970	0.891	0.852	0.920	0.963
$two\_step$	2	0.969	0.865	0.738	0.782	0.864
$smote\_rf$	3	0.972	0.858	0.663	0.421	0.742
hybrid_xgb	4	0.971	0.852	0.659	0.550	0.729
$smote\_xgb$	5	0.973	0.840	0.654	0.576	0.733
Balanced Acc	curacy					
sequential	1	0.913	0.913	0.858	0.953	0.953
$smote\_xgb$	2	0.943	0.917	0.806	0.823	0.917
hybrid_xgb	3	0.937	0.919	0.814	0.822	0.909
$two\_step$	6	0.919	0.893	0.819	0.924	0.908
$smote\_rf$	9	0.920	0.913	0.813	0.653	0.887
Kappa						
sequential	1	0.842	0.839	0.715	0.884	0.884
$smote\_rf$	2	0.856	0.849	0.641	0.330	0.730
$smote\_xgb$	3	0.866	0.830	0.632	0.569	0.719
hybrid_xgb	4	0.854	0.842	0.638	0.543	0.715
two_step	5	0.833	0.796	0.632	0.758	0.818

Top 5 Workflow Per-Class Evaluation Metrics by Metric

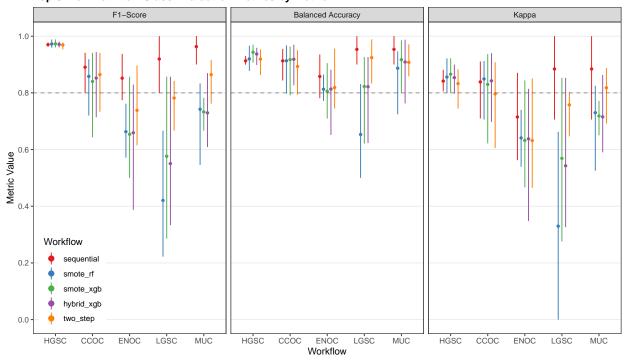


Figure 4.8: Top 5 Workflow Per-Class Evaluation Metrics by Metric

Misclassified cases from a previous step of the sequence of classifiers are not included in subsequent steps of the training set CV folds. Thus, we cannot piece together the test set predictions from the sequential and two-step algorithms to obtain overall metrics.

## 4.3 Optimal Gene Sets

#### 4.3.1 Sequential Algorithm

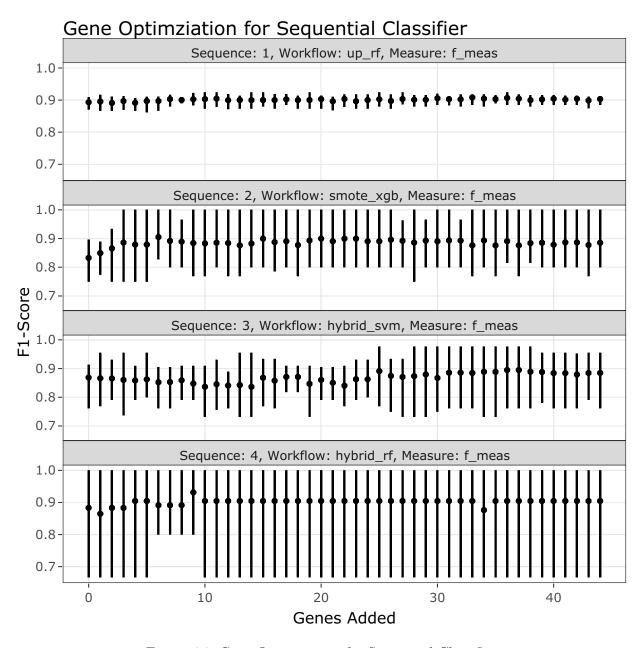


Figure 4.9: Gene Optimization for Sequential Classifier

In the sequential algorithm, all sequences have relatively flat average F1-scores across the number of genes added. However, we can observe in sequence 4, the F1-score is highest when we reach 9 genes added, hence the optimal number of genes used will be n=28+9=37 The added genes are: CYP2C18, HNF1B, EGFL6, TFF3, IL6, CYP4B1, LGALS4, SLC3A1 and IGFBP1.

Table 4.14: Gene Profile of Optimal Set in Sequential Algorithm

Set	Genes	PrOTYPE	SPOT	Optimal Set	Candidate Rank
	COL11A1	V		(*)	
	CD74	V		(*)	
	$\overline{\mathrm{CD2}}$	V		(*)	
	TIMP3	V		(*)	
	LUM	V		(*)	
	CYTIP	V		(*)	
	COL3A1	V		(*)	
	THBS2	V		(*)	
	TCF7L1	V	V	(*)	
	HMGA2	V		(*)	
	FN1	V		(*)	
	POSTN	V		(*)	
	COL1A2	V		(*)	
	COL5A2	V		(*)	
	PDZK1IP1	V		(*)	
	FBN1	V		(*)	
	HIF1A		V	(*)	
Base	CXCL10		V	(*)	
	DUSP4		V	(*)	
	SOX17		V	(*)	
	MITF		V	(*)	
	CDKN3		V	(*)	
	BRCA2		V	(*)	
	CEACAM5		V	(*)	
	ANXA4		v	(*)	
	SERPINE1		V	(*)	
	CRABP2		V	(*)	
	DNAJC9		v	(*)	

HNF1B (*) 2 EGFL6 (*) 3 TFF3 (*) 4 IL6 (*) 5 CYP4B1 (*) 6 LGALS4 (*) 7 SLC3A1 (*) 8 IGFBP1 (*) 9 WT1 (x) 10 MUC5B (x) 11 TFF1 (x) 12 GPR64 (x) 13 TP53 (x) 14 BRCA1 (x) 15 MET (x) 16 FUT3 (x) 17 CPNE8 (x) 18 TPX2 (x) 19 PBX1 (x) 20 EPAS1 (x) 21 SCGB1D2 (x) 22 KLK7 (x) 23 SEMA6A (x) 24 DKK4 (x) 25 CAPN2 (x) 29 GCNT3 (x) 31 SERPINA5 (x) 32 Clorf173 (x) 32 Clorf173 (x) 33 PAX8 (x) 33 PAX8 (x) 34	CYP2C18	(*)	1
TFF3 (*) 4  IL6 (*) 5  CYP4B1 (*) 6  LGALS4 (*) 7  SLC3A1 (*) 8  IGFBP1 (*) 9  WT1 (x) 10  MUC5B (x) 11  TFF1 (x) 12  GPR64 (x) 13  TP53 (x) 14  BRCA1 (x) 15  MET (x) 16  FUT3 (x) 17  CPNE8 (x) 18  TPX2 (x) 19  PBX1 (x) 20  EPAS1 (x) 21  SCGB1D2 (x) 22  KLK7 (x) 23  SEMA6A (x) 24  DKK4 (x) 25  CAPN2 (x) 29  GCNT3 (x) 31  SERPINA5 (x) 31  SERPINA5 (x) 32  CIorf173 (x) 31  SERPINA5 (x) 32  CIorf173 (x) 32  CIorf173 (x) 32  CIorf173 (x) 32  CIorf173 (x) 32	HNF1B	(*)	2
IL6	EGFL6	(*)	3
CYP4B1       (*)       6         LGALS4       (*)       7         SLC3A1       (*)       8         IGFBP1       (*)       9         WT1       (x)       10         MUC5B       (x)       11         TFF1       (x)       12         GPR64       (x)       13         TP53       (x)       14         BRCA1       (x)       15         MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5	TFF3	(*)	4
LGALS4	IL6	(*)	5
SLC3A1       (*)       8         IGFBP1       (*)       9         WT1       (x)       10         MUC5B       (x)       11         TFF1       (x)       12         GPR64       (x)       13         TP53       (x)       14         BRCA1       (x)       15         MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	CYP4B1	(*)	6
IGFBP1       (*)       9         WT1       (x)       10         MUC5B       (x)       11         TFF1       (x)       12         GPR64       (x)       13         TP53       (x)       14         BRCA1       (x)       15         MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         Clorf173       (x)       33	LGALS4	(*)	7
WT1       (x)       10         MUC5B       (x)       11         TFF1       (x)       12         GPR64       (x)       13         TP53       (x)       14         BRCA1       (x)       15         MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         Clorf173       (x)       33	SLC3A1	(*)	8
MUC5B       (x)       11         TFF1       (x)       12         GPR64       (x)       13         TP53       (x)       14         BRCA1       (x)       15         MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         Clorf173       (x)       33	IGFBP1	(*)	9
TFF1 (x) 12  GPR64 (x) 13  TP53 (x) 14  BRCA1 (x) 15  MET (x) 16  FUT3 (x) 17  CPNE8 (x) 18  TPX2 (x) 19  PBX1 (x) 20  EPAS1 (x) 21  SCGB1D2 (x) 22  KLK7 (x) 23  SEMA6A (x) 24  DKK4 (x) 25  CAPN2 (x) 26  GAD1 (x) 27  STC1 (x) 28  IGJ (x) 29  GCNT3 (x) 30  TSPAN8 (x) 31  SERPINA5 (x) 32  Clorf173 (x) 33	WT1	(x)	10
GPR64       (x)       13         TP53       (x)       14         BRCA1       (x)       15         MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         Clorf173       (x)       33	MUC5B	(x)	11
TP53 (x) 14  BRCA1 (x) 15  MET (x) 16  FUT3 (x) 17  CPNE8 (x) 18  TPX2 (x) 19  PBX1 (x) 20  EPAS1 (x) 21  SCGB1D2 (x) 22  KLK7 (x) 23  SEMA6A (x) 24  DKK4 (x) 25  CAPN2 (x) 26  GAD1 (x) 27  STC1 (x) 28  IGJ (x) 29  GCNT3 (x) 31  SERPINA5 (x) 32  Clorf173 (x) 33	TFF1	(x)	12
BRCA1         (x)         15           MET         (x)         16           FUT3         (x)         17           CPNE8         (x)         18           TPX2         (x)         19           PBX1         (x)         20           EPAS1         (x)         21           SCGB1D2         (x)         22           KLK7         (x)         23           SEMA6A         (x)         24           DKK4         (x)         25           CAPN2         (x)         26           GAD1         (x)         27           STC1         (x)         28           IGJ         (x)         29           GCNT3         (x)         30           TSPAN8         (x)         31           SERPINA5         (x)         32           C1orf173         (x)         33	GPR64	(x)	13
MET       (x)       16         FUT3       (x)       17         CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	TP53	(x)	14
FUT3 (x) 17  CPNE8 (x) 18  TPX2 (x) 19  PBX1 (x) 20  EPAS1 (x) 21  SCGB1D2 (x) 22  KLK7 (x) 23  SEMA6A (x) 24  DKK4 (x) 25  CAPN2 (x) 26  GAD1 (x) 27  STC1 (x) 28  IGJ (x) 29  GCNT3 (x) 30  TSPAN8 (x) 31  SERPINA5 (x) 32  Clorf173 (x) 33	BRCA1	(x)	15
CPNE8       (x)       18         TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	MET	(x)	16
TPX2       (x)       19         PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         Clorf173       (x)       33	FUT3	(x)	17
PBX1       (x)       20         EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	CPNE8	(x)	18
EPAS1       (x)       21         SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	TPX2	(x)	19
SCGB1D2       (x)       22         KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	PBX1	(x)	20
KLK7       (x)       23         SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         Clorf173       (x)       33	EPAS1	(x)	21
SEMA6A       (x)       24         DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	SCGB1D2	(x)	22
DKK4       (x)       25         CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	KLK7	(x)	23
CAPN2       (x)       26         GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	SEMA6A	(x)	24
GAD1       (x)       27         STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	DKK4	(x)	25
STC1       (x)       28         IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	CAPN2	(x)	26
IGJ       (x)       29         GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	GAD1	(x)	27
GCNT3       (x)       30         TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	STC1	(x)	28
TSPAN8       (x)       31         SERPINA5       (x)       32         C1orf173       (x)       33	IGJ	(x)	29
SERPINA5         (x)         32           C1orf173         (x)         33	GCNT3	(x)	30
C1orf173 (x) 33	TSPAN8	(x)	31
	SERPINA5	(x)	32
PAX8 (x) 34	C1orf173	(x)	33
	PAX8	(x)	34

LIN28B	(x)	35
ZBED1	(x)	36
ATP5G3	(x)	37
BCL2	(x)	38
KGFLP2	(x)	39
IGKC	(x)	40
SENP8	(x)	41
MAP1LC3A	(x)	42
C10orf116	(x)	43
ADCYAP1R1	(x)	44

#### 4.3.2 SMOTE-Random Forest



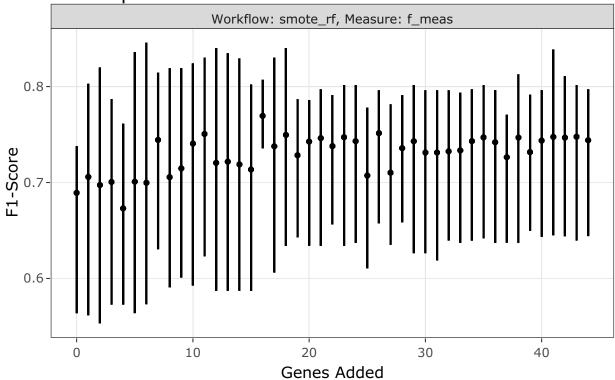


Figure 4.10: Gene Optimization for SMOTE-Random Forest Classifier

In the SMOTE-Random Forest classifier, the mean F1-score is highest when we reach 16 genes added, hence the optimal number of genes used will be n=28+16=44 The added genes are: HNF1B, TFF3, TPX2, SLC3A1, CYP2C18, TFF1, WT1, KLK7, IGFBP1, LGALS4, GAD1, GCNT3, C1orf173, CAPN2, FUT3 and DKK4.

Table 4.15: Gene Profile of Optimal Set in SMOTE-Random Forest Workflow

Set	Genes	PrOTYPE	SPOT	Optimal Set	Candidate Rank
	COL11A1	V		(*)	
	CD74	V		(*)	
	$\overline{\mathrm{CD2}}$	V		(*)	
	TIMP3	V		(*)	
	LUM	V		(*)	
	CYTIP	V		(*)	
	COL3A1	V		(*)	
	THBS2	V		(*)	
	TCF7L1	V	V	(*)	
	HMGA2	V		(*)	
	FN1	V		(*)	
	POSTN	V		(*)	
	COL1A2	V		(*)	
	COL5A2	V		(*)	
	PDZK1IP1	V		(*)	
	FBN1	V		(*)	
	HIF1A		V	(*)	
Base	CXCL10		V	(*)	
	DUSP4		V	(*)	
	SOX17		V	(*)	
	MITF		V	(*)	
	CDKN3		V	(*)	
	BRCA2		V	(*)	
	CEACAM5		V	(*)	
	ANXA4		V	(*)	
	SERPINE1		V	(*)	
	CRABP2		V	(*)	
	DNAJC9		V	(*)	
	HNF1B			(*)	1
	TFF3			(*)	2
	TPX2			(*)	3
	SLC3A1			(*)	4

TFF1 (*) 6 WT1 (*) 7 KLK7 (*) 8 IGFBP1 (*) 9 LGALS4 (*) 10 GAD1 (*) 11 GCNT3 (*) 12 C1orf173 (*) 13 CAPN2 (*) 14 FUT3 (*) 15 DKK4 (*) 16 C10orf116 (x) 17 MUC5B (x) 18 MET (x) 19 GPR64 (x) 20 IGKC (x) 21 PAX8 (x) 22 ATP5G3 (x) 23 CPNE8 (x) 24 PBX1 (x) 25 IL6 (x) 27 KGFLP2 (x) 28 EGFL6 (x) 29 SEMA6A (x) 30 CYP4B1 (x) 31 STC1 (x) 34 LIN28B (x) 36 SERPINA5 (x) 36 SERPINA5 (x) 37 SCGB1D2 (x) 38	CYP2C18	(*)	5
KLK7	TFF1	(*)	6
IGFBP1       (*)       9         LGALS4       (*)       10         GAD1       (*)       11         GCNT3       (*)       12         Clorf173       (*)       13         CAPN2       (*)       14         FUT3       (*)       15         DKK4       (*)       16         C10orf116       (x)       17         MUC5B       (x)       18         MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       25         IL6       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       34         LIN28B       (x)       35         TSPAN8	WT1	(*)	7
LGALS4       (*)       10         GAD1       (*)       11         GCNT3       (*)       12         Clorf173       (*)       13         CAPN2       (*)       14         FUT3       (*)       15         DKK4       (*)       16         C10orf116       (x)       17         MUC5B       (x)       18         MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       36         SERPINA5 <td>KLK7</td> <td>(*)</td> <td>8</td>	KLK7	(*)	8
GAD1 (*) 11 GCNT3 (*) 12 C1orf173 (*) 13 CAPN2 (*) 14 FUT3 (*) 15 DKK4 (*) 16 C10orf116 (x) 17 MUC5B (x) 18 MET (x) 19 GPR64 (x) 20 IGKC (x) 21 PAX8 (x) 22 ATP5G3 (x) 23 CPNE8 (x) 24 PBX1 (x) 25 IL6 (x) 26 TP53 (x) 27 KGFLP2 (x) 28 EGFL6 (x) 29 SEMA6A (x) 30 CYP4B1 (x) 31 STC1 (x) 32 EPAS1 (x) 33 BRCA1 (x) 36 SERPINA5 (x) 36 SERPINA5 (x) 36 SERPINA5	IGFBP1	(*)	9
GCNT3  Clorf173  (*) 12  Clorf173  (*) 13  CAPN2  (*) 14  FUT3  (*) 15  DKK4  (*) 16  Cloorf116  (x) 17  MUC5B  MET  (x) 19  GPR64  (x) 20  IGKC  (x) 21  PAX8  (x) 22  ATP5G3  (x) 23  CPNE8  (x) 24  PBX1  (x) 25  IL6  (x) 26  TP53  (x) 27  KGFLP2  (x) 28  EGFL6  (x) 29  SEMA6A  (x) 30  CYP4B1  (x) 31  STC1  (x) 32  EPAS1  (x) 33  BRCA1  LIN28B  (x) 12  (*) 14  (*) 15  Id  (*) 15  Id  (*) 16  (*) 26  TP5AN8  (x) 37	LGALS4	(*)	10
C1orf173       (*)       13         CAPN2       (*)       14         FUT3       (*)       15         DKK4       (*)       16         C10orf116       (x)       17         MUC5B       (x)       18         MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	GAD1	(*)	11
CAPN2 (*) 14 FUT3 (*) 15 DKK4 (*) 16 C10orf116 (x) 17 MUC5B (x) 18 MET (x) 19 GPR64 (x) 20 IGKC (x) 21 PAX8 (x) 22 ATP5G3 (x) 23 CPNE8 (x) 24 PBX1 (x) 25 IL6 (x) 26 TP53 (x) 27 KGFLP2 (x) 28 EGFL6 (x) 29 SEMA6A (x) 30 CYP4B1 (x) 31 STC1 (x) 32 EPAS1 (x) 33 BRCA1 (x) 36 SERPINA5 (x) 36 SERPINA5	GCNT3	(*)	12
FUT3    (*)   15	C1orf173	(*)	13
DKK4       (*)       16         C10orf116       (x)       17         MUC5B       (x)       18         MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	CAPN2	(*)	14
C10orf116       (x)       17         MUC5B       (x)       18         MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	FUT3	(*)	15
MUC5B       (x)       18         MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	DKK4	(*)	16
MET       (x)       19         GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	C10orf116	(x)	17
GPR64       (x)       20         IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	MUC5B	(x)	18
IGKC       (x)       21         PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	MET	(x)	19
PAX8       (x)       22         ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	GPR64	(x)	20
ATP5G3       (x)       23         CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	IGKC	(x)	21
CPNE8       (x)       24         PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	PAX8	(x)	22
PBX1       (x)       25         IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	ATP5G3	(x)	23
IL6       (x)       26         TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	CPNE8	(x)	24
TP53       (x)       27         KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	PBX1	(x)	25
KGFLP2       (x)       28         EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	IL6	(x)	26
EGFL6       (x)       29         SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	TP53	(x)	27
SEMA6A       (x)       30         CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	KGFLP2	(x)	28
CYP4B1       (x)       31         STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	EGFL6	(x)	29
STC1       (x)       32         EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	SEMA6A	(x)	30
EPAS1       (x)       33         BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	CYP4B1	(x)	31
BRCA1       (x)       34         LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	STC1	(x)	32
LIN28B       (x)       35         TSPAN8       (x)       36         SERPINA5       (x)       37	EPAS1	(x)	33
TSPAN8         (x)         36           SERPINA5         (x)         37	BRCA1	(x)	34
SERPINA5 (x) 37	LIN28B	(x)	35
	TSPAN8	(x)	36
SCGB1D2 (x) 38	SERPINA5	(x)	37
	SCGB1D2	(x)	38

BCL2	(x)	39
ZBED1	(x)	40
ADCYAP1R1	(x)	41
IGJ	(x)	42
SENP8	(x)	43
MAP1LC3A	(x)	44

## 4.3.3 Two-Step

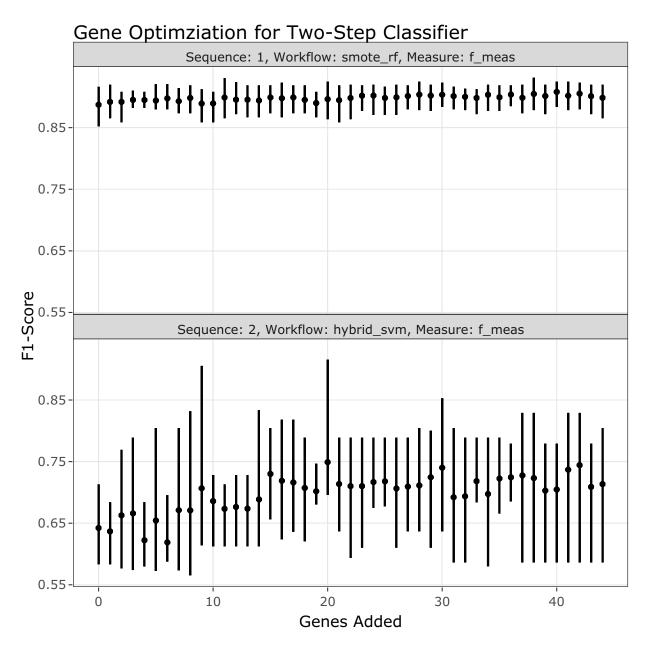


Figure 4.11: Gene Optimization for Two-Step Classifier

Table 4.16: Gene Profile of Optimal Set in Two-Step Workflow

Set	Genes	PrOTYPE	SPOT	Optimal Set	Candidate Rank
	COL11A1	V		(*)	
	CD74	V		(*)	
	$\overline{\mathrm{CD2}}$	V		(*)	

	-				
	TIMP3	V		(*)	
	LUM	V		(*)	
	CYTIP	V		(*)	
	COL3A1	V		(*)	
	THBS2	V		(*)	
	TCF7L1	v	v	(*)	
	HMGA2	v		(*)	
	FN1	v		(*)	
	POSTN	v		(*)	
	COL1A2	V		(*)	
	COL5A2	V		(*)	
	PDZK1IP1	V		(*)	
	FBN1	V		(*)	
	HIF1A		V	(*)	
Base	CXCL10		v	(*)	
Dase	DUSP4		v	(*)	
	SOX17		V	(*)	
	MITF		V	(*)	
	CDKN3		v	(*)	
	BRCA2		v	(*)	
	CEACAM5		v	(*)	
	ANXA4		v	(*)	
	SERPINE1		V	(*)	
	CRABP2		v	(*)	
	DNAJC9		v	(*)	
	CYP2C18			(*)	1
	MUC5B			(*)	2
	HNF1B			(*)	3
	IL6			(*)	4
	SLC3A1			(*)	5
	EGFL6			(*)	6
	WT1			(*)	7
	ZBED1			(*)	8
	MET			(*)	9

SENP8	(*)	10
KLK7	(*)	11
TFF3	(*)	12
CPNE8	(*)	13
STC1	(*)	14
GAD1	(*)	15
LIN28B	(x)	16
IGJ	(x)	17
DKK4	(x)	18
EPAS1	(x)	19
GCNT3	(x)	20
SCGB1D2	(x)	21
CYP4B1	(x)	22
C1orf173	(x)	23
IGFBP1	(x)	24
TPX2	(x)	25
SEMA6A	(x)	26
ATP5G3	(x)	27
SERPINA5	(x)	28
FUT3	(x)	29
C10orf116	(x)	30
KGFLP2	(x)	31
ADCYAP1R1	(x)	32
TP53	(x)	33
PBX1	(x)	34
GPR64	(x)	35
LGALS4	(x)	36
CAPN2	(x)	37
BCL2	(x)	38
MAP1LC3A	(x)	39
TSPAN8	(x)	40
TFF1	(x)	41
PAX8	(x)	42
BRCA1	(x)	43

## 4.4 Test Set Performance

Now we'd like to see how our best methods perform in the confirmation and validation sets. The class-specific F1-scores will be used.

The top 2 methods are the sequential and SMOTE-Random Forest classifiers. We can test 2 additional methods by using either the full set of genes or the optimal set of genes for both of these classifiers.

## 4.4.1 Confirmation Set

Table 4.17: Evaluation Metrics on Confirmation Set Models

		Histotypes					
Method	Metric	Overall	HGSC	CCOC	ENOC	LGSC	MUC
	Accuracy	0.829	0.861	0.964	0.888	0.975	0.969
	Sensitivity	0.591	0.950	0.861	0.467	0.083	0.593
	Specificity	0.923	0.688	0.977	0.972	0.992	0.985
Sequential, Full Set	F1-Score	0.610	0.901	0.844	0.581	0.111	0.615
,	Balanced Accuracy	0.757	0.819	0.919	0.720	0.538	0.789
	Kappa	0.646	0.674	0.823	0.521	0.100	0.599
	Accuracy	0.816	0.852	0.963	0.875	0.970	0.972
	Sensitivity	0.554	0.955	0.875	0.383	0.000	0.556
	Specificity	0.916	0.651	0.974	0.974	0.989	0.990
Sequential, Optimal Set	F1-Score	0.573	0.895	0.840	0.506	0.000	0.625
· , · ·	Balanced Accuracy	0.735	0.803	0.924	0.679	0.494	0.773
	Kappa	0.614	0.648	0.819	0.443	-0.014	0.611
	Accuracy	0.841	0.864	0.972	0.896	0.977	0.974
	Sensitivity	0.647	0.960	0.861	0.458	0.250	0.704
	Specificity	0.925	0.679	0.986	0.983	0.990	0.985
SMOTE-Random Forest, Full Set	F1-Score	0.669	0.903	0.873	0.594	0.286	0.691
,	Balanced Accuracy	0.786	0.819	0.924	0.721	0.620	0.845
	Kappa	0.669	0.679	0.857	0.540	0.274	0.677
	Accuracy	0.838	0.868	0.967	0.896	0.980	0.966
	Sensitivity	0.659	0.948	0.861	0.486	0.333	0.667
	Specificity	0.928	0.711	0.981	0.978	0.992	0.979
SMOTE-Random Forest, Optimal Set	F1-Score	0.674	0.904	0.855	0.608	0.381	0.621
, .	Balanced Accuracy	0.794	0.830	0.921	0.732	0.663	0.823
	Kappa	0.669	0.691	0.837	0.552	0.371	0.603
	Accuracy	0.835	0.861	0.966	0.891	0.972	0.980
	Sensitivity	0.651	0.941	0.875	0.486	0.250	0.704
	Specificity	0.927	0.706	0.977	0.972	0.986	0.992
Two-Step, Full Set	F1-Score	0.669	0.900	0.851	0.598	0.250	0.745
- /	Balanced Accuracy	0.789	0.824	0.926	0.729	0.618	0.848
	Kappa	0.664	0.677	0.832	0.538	0.236	0.735
	Accuracy	0.843	0.866	0.967	0.900	0.975	0.977
	Sensitivity	0.639	0.953	0.875	0.495	0.167	0.704
	Specificity	0.927	0.697	0.979	0.981	0.990	0.989
Two-Step, Optimal Set	F1-Score	0.660	0.904	0.857	0.624	0.200	0.717
	Balanced Accuracy	0.783	0.825	0.927	0.738	0.579	0.846
	Kappa	0.676	0.685	0.839	0.570	0.188	0.705

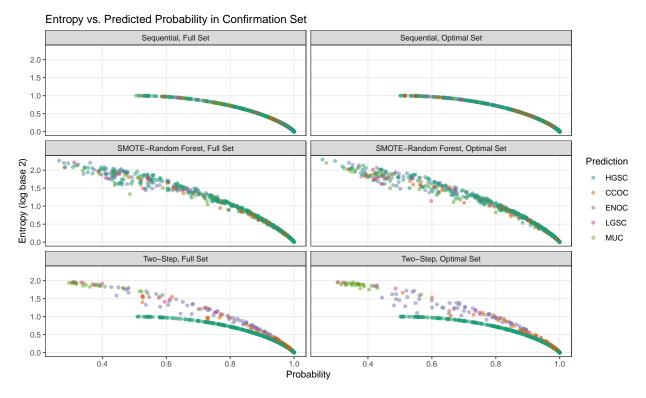


Figure 4.12: Entropy vs. Predicted Probability in Confirmation Set

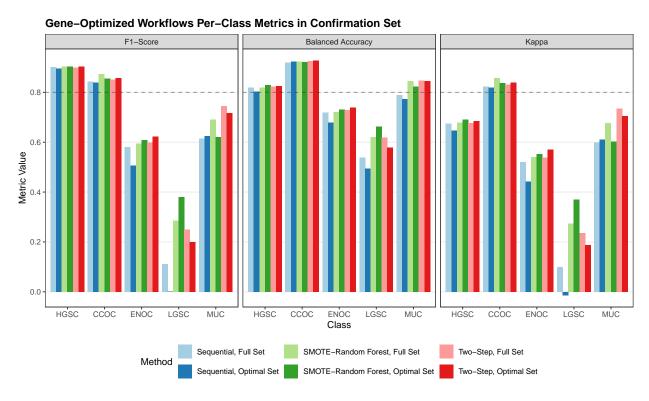


Figure 4.13: Gene Optimized Workflows Per-Class Metrics in Confirmation Set

#### Confusion Matrices for Confirmation Set Models В Α Sequential, Full Set Sequential, Optimal Set **HGSC HGSC** CCOC CCOC Prediction Prediction **ENOC ENOC** LGSC **LGSC** MUC MUC LGSC LGSC **HGSC** CCOC **ENOC** MUC **HGSC** CCOC **ENOC** MUC Truth Truth С D SMOTE-Random Forest, Full Set SMOTE-Random Forest, Optimal Set **HGSC HGSC** CCOC CCOC Prediction SONS FINANCE FOR PROPERTY PR Prediction **ENOC LGSC LGSC** MUC MUC **HGSC** CCOC **ENOC LGSC** MUC **HGSC** CCOC **ENOC** LGSC MUC Truth Truth F Е Two-Step, Full Set Two-Step, Optimal Set **HGSC HGSC**

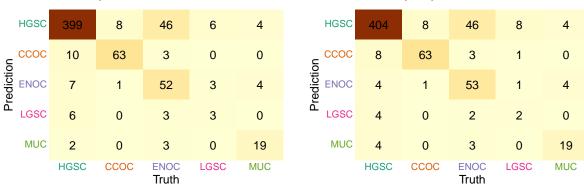


Figure 4.14: Confusion Matrices for Confirmation Set Models

#### 4.4.1.1 Sequential, Full

## ROC Curves for Sequential, Full Set Model in Confirmation Set

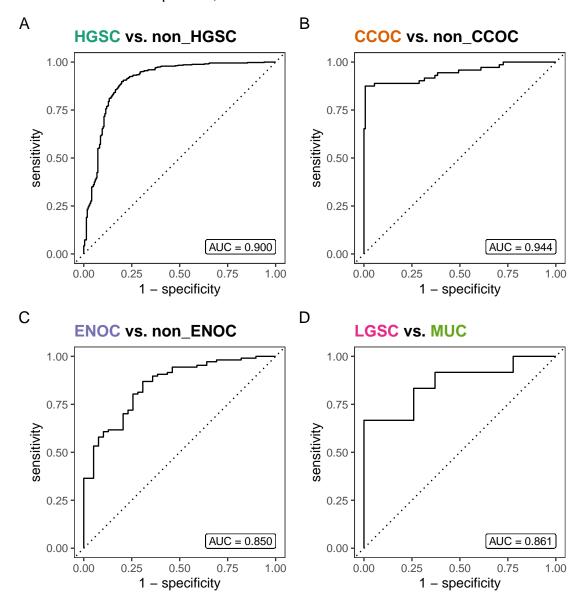


Figure 4.15: ROC Curves for Sequential Full Model in Confirmation Set

#### 4.4.1.2 Sequential, Optimal

ROC Curves for Sequential, Optimal Set Model in Confirmation Set

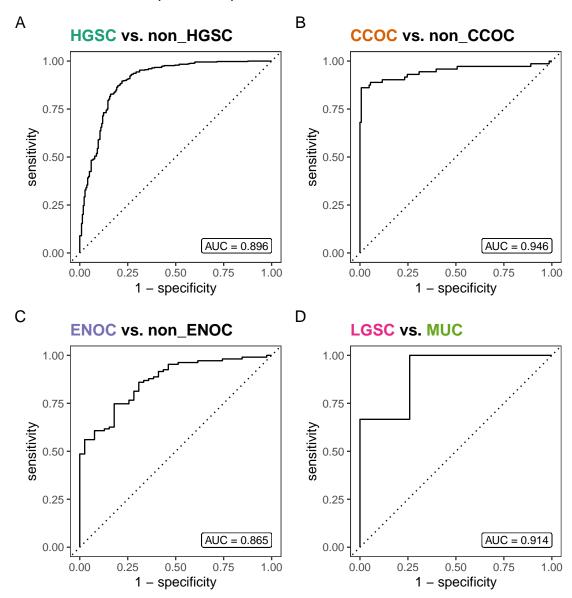


Figure 4.16: ROC Curves for Sequential, Optimal Model in Confirmation Set

## 4.4.1.3 SMOTE-Random Forest, Full

# ROC Curve for SMOTE-Random Forest, Full Set Model in Confirmation Set AUC = 0.890

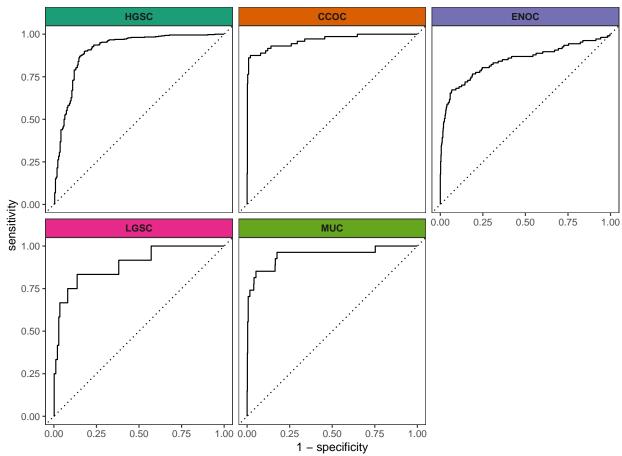


Figure 4.17: ROC Curves for SMOTE-Random Forest, Full Set Model in Confirmation Set

## 4.4.1.4 SMOTE-Random Forest, Optimal

0.75

0.50

0.25

0.00

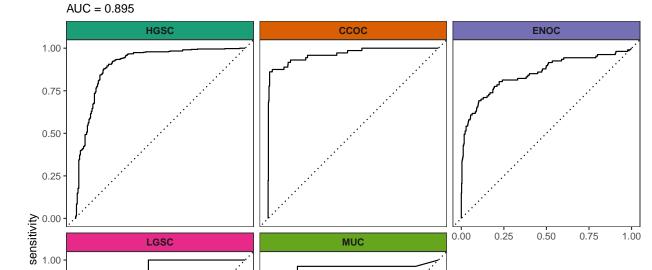
0.00

0.25

0.50

0.75

1.00 0.00



ROC Curve for SMOTE-Random Forest, Optimal Set Model in Confirmation Set

Figure 4.18: ROC Curves for SMOTE-Random Forest, Optimal Set Model in Confirmation Set

0.50

1 - specificity

0.25

0.75

1.00

## 4.4.1.5 Two-Step, Full

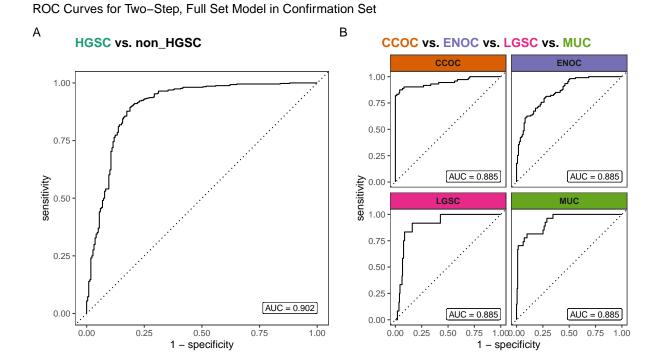
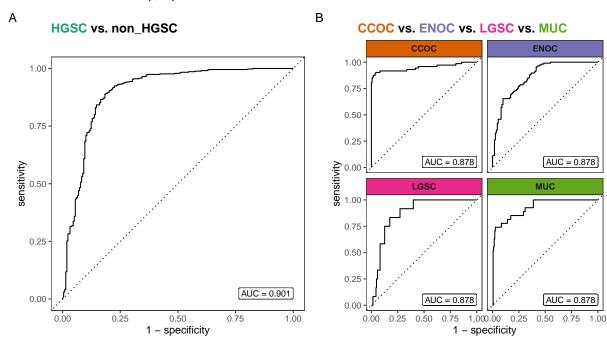


Figure 4.19: ROC Curves for Two-Step Full Model in Confirmation Set

#### 4.4.1.6 Two-Step, Optimal



ROC Curves for Two-Step, Optimal Set Model in Confirmation Set

Figure 4.20: ROC Curves for Two-Step Optimal Model in Confirmation Set

## 4.4.2 Validation Set

Table 4.18: Evaluation Metrics on Validation Set Model, SMOTE-Random Forest, Optimal Set

		Histotypes				
Metric	Overall	HGSC	CCOC	ENOC	LGSC	MUC
Accuracy	0.889	0.907	0.970	0.946	0.977	0.979
Sensitivity	0.781	0.917	0.971	0.682	0.467	0.870
Specificity	0.957	0.872	0.970	0.975	0.985	0.982
F1-Score	0.713	0.939	0.832	0.714	0.400	0.678
Balanced Accuracy	0.869	0.894	0.970	0.829	0.726	0.926
Kappa	0.723	0.743	0.816	0.685	0.388	0.668

#### **SMOTE-Random Forest Per-Class Metrics in Validation Set**

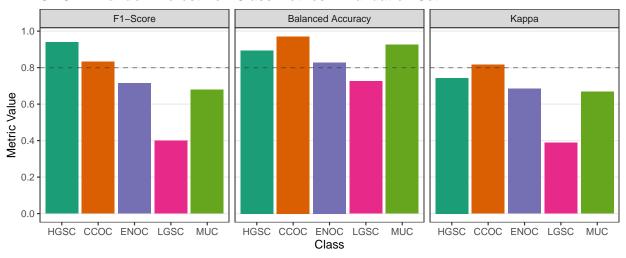


Figure 4.21: SMOTE-Random Forest Per-Class Metrics in Validation Set

# Confusion Matrix for Validation Set Model SMOTE-Random Forest, Optimal Set

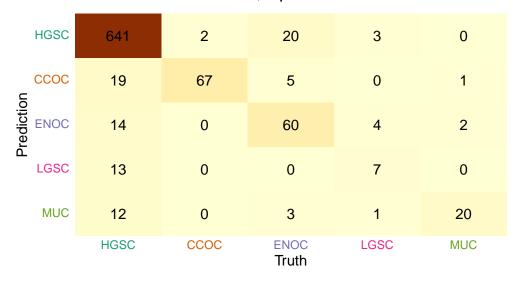
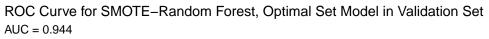


Figure 4.22: Confusion Matrix for Validation Set Model



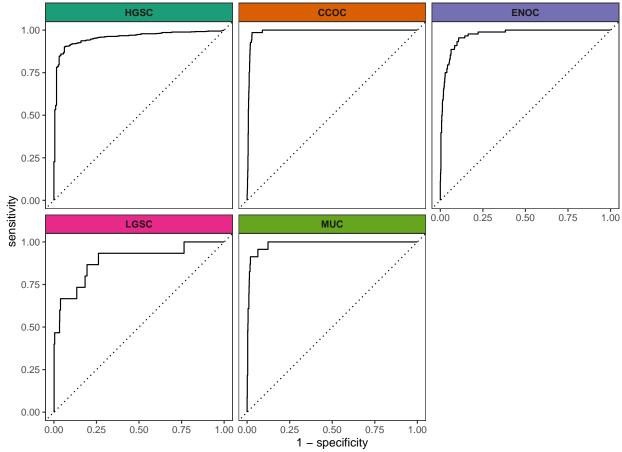


Figure 4.23: ROC Curves for SMOTE-Random Forest, Optimal Set Model in Validation Set

Table 4.19: Clinicopath characteristics between correct and incorrect predictions of ENOC cases

Characteristic	Predicted ENOC Correctly $N = 60^{1}$	Missed ENOC $N = 28^1$	$\mathbf{p} ext{-}\mathbf{value}^2$
Age at diagnosis	53 (46, 62)	58 (52, 64)	0.095
Tumour grade			< 0.001
low grade	47 (96%)	12~(52%)	
high grade	2(4.1%)	11 (48%)	
Unknown	11	5	
FIGO tumour stage			0.012
I	47 (80%)	15 (54%)	
II-IV	12 (20%)	13 (46%)	
Unknown	1	0	
Race			> 0.9
white	54 (92%)	22 (92%)	
non-white	5 (8.5%)	2(8.3%)	
Unknown	1	$\stackrel{\cdot}{4}$	
ARID1A			>0.9
absent/subclonal	11 (18%)	5 (18%)	
present	49 (82%)	23 (82%)	

<sup>&</sup>lt;sup>1</sup>Median (Q1, Q3); n (%)

<sup>&</sup>lt;sup>2</sup>Wilcoxon rank sum test; Fisher's exact test; Pearson's Chi-squared test

#### Volcano Plots of Validation Set Predictions

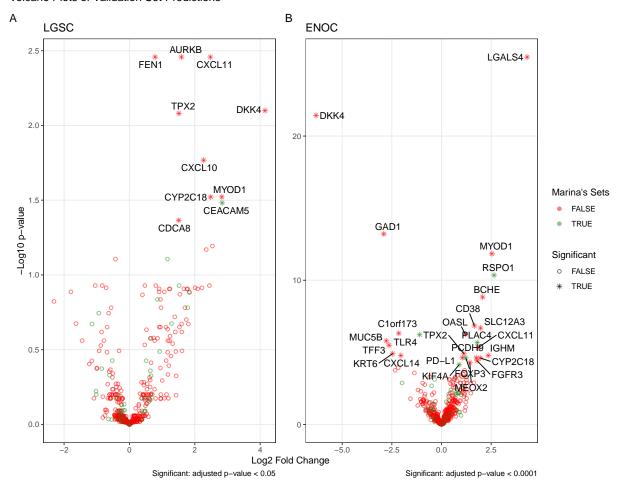


Figure 4.24: Volcano Plots of Validation Set Predictions

#### **Subtype Prediction Summary among Predicted HGSC Samples**

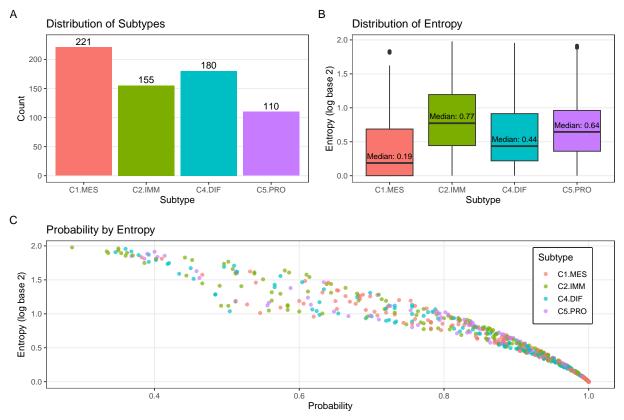


Figure 4.25: Subtype Prediction Summary among Predicted HGSC Samples

# References

Talhouk, Aline, Stefan Kommoss, Robertson Mackenzie, Martin Cheung, Samuel Leung, Derek S. Chiu, Steve E. Kalloger, et al. 2016. "Single-Patient Molecular Testing with NanoString nCounter Data Using a Reference-Based Strategy for Batch Effect Correction." Edited by Benjamin Haibe-Kains. *PLOS ONE* 11 (4): e0153844. https://doi.org/10.1371/journal.pone. 0153844.