

# Saturation Binding Assay

Charles “Mitch” Boudreaux  
Talía Albert

November 3rd, 2023

## Contents

<b>Overview</b>	<b>1</b>
<b>Determining Protein Concentration</b>	<b>1</b>
<b>Hot Ligand Addition</b>	<b>2</b>
<b>Cold Ligand (Reference) Addition</b>	<b>2</b>
<b>Membrane (Protein) Addition</b>	<b>2</b>
<b>Filtering</b>	<b>3</b>
<b>Formulas</b>	<b>3</b>
Protein Concentration calculation . . . . .	3
3H Hot Ligand calculation . . . . .	3
Cold Ligand calculation . . . . .	3
Membrane Receptor calculation . . . . .	4
Determining Starting Concentration calculation . . . . .	4

## Overview

Final Volume in each well is **125 uL**

- 25 uL 3H Hot Ligand
- 25 uL Binding Buffer/Cold Ligand (Reference)
- 75 uL Membrane (Protein)

## Determining Protein Concentration

1. Resuspend pellet in 12 mL Lysis Buffer (10mM Tris + 5% Sucrose, pH 7.4):

- Use 1 mL Lysis Buffer to break up pellet
  - Fill to ~12 mL with Lysis Buffer
  - Pool pellets into one tube (if multiple pellets)
2. Perform Bradford Protein Concentration Assay:
    - Sample Preparation:
      - 10 uL Pellet Suspension + 790 uL dH2O + 200 uL Bradford Reagent
    - Blank Preparation:
      - 10 uL Lysis Buffer + 790 uL dH2O + 200 uL Bradford Reagent
    - Incubate @ RT 10 min
    - Measure absorbance @ 595 nm
    - Calculate protein concentration (Refer to Formulas section or Spreadsheet)

## Hot Ligand Addition

3. Prepare ~15 mL appropriate BB (Binding Buffer) w/BSA (~30 uL) in trough
4. In an empty 96-well shallow plate (Need one column):
  1. Add **330 uL** BB to well H and **165 uL** BB to well A-G
  2. Add 3H-Ligand to Well H (Refer to Formulas section or Spreadsheet)
  3. Perform a Serial Dilution (1:2) of **165 uL** up from well H to A
  4. Remove **25 uL** from Well A for radioactivity counts
  5. Using a multichannel pipettor, Transfer **25 uL** into 6 columns of the Drug Plate

## Cold Ligand (Reference) Addition

5. Add **25 uL** BB to first 3 columns
6. Prepare Cold Ligand (Reference):
  1. Usually, add **4 uL** Reference compound into **800 uL** BB into an eppendorf tube
  2. Using a single channel pipettor, add **25 uL** Reference Compound into the last 3 columns

## Membrane (Protein) Addition

7. Prepare ~4 mL membrane receptor:
  1. Add BB volume to new trough (Refer to Formulas section or Spreadsheet)
  2. Add protein to volume new trough (Refer to Formulas section or Spreadsheet)
  3. Using a multichannel pipettor, add **75 uL** protein dilution into all 6 columns

8. Incubate plates @ RT for 1 hour in drawer

## Filtering

## Formulas

### Protein Concentration calculation

Need:

- OD @ 595 nm

$$\text{Protein Concentration (ug/uL)} = \frac{\text{OD@595 nm} - 0.094}{0.503}$$

Where:

- 0.094 and 0.503 were determined from experimental procedure, provided by XP.

### 3H Hot Ligand calculation

Need:

- Starting Concentration (nM)
- 3H-Ligand Specific Activity (Ci/mmol)

$$\text{3H-Ligand Vol (uL)} = \frac{330 \text{ (uL)} * \text{Starting Concentration (nM)} * 5 * 1.2}{\text{Specific Activity (Ci/mmol)}^{-1} * 1000000}$$

Where:

- 330 uL is double the volume of 165 uL, (25 uL \* 6 wells \* 1.1 overage = 165 uL), so we can perform a serial dilution
- 5 is a Dilution Factor, the final volume in each well is 125 uL, we add 25 uL from the Hot-Ligand plate, (125/25=5)
- 1.2 is a 20% overage
- 1000000 is for unit conversion

### Cold Ligand calculation

Usually need **4 uL** Reference compound and **800 uL** BB

Need:

- Concentration of Cold Ligand (Reference) Stock (most are 10 mM)

$$\text{Reference Vol (uL)} = \frac{800 \text{ uL} * 10 \text{ uM Final Concentration} * 5}{10000 \text{ uM Starting Concentration}}$$

Where:

- 800 uL is approximate volume we need (8 Wells/Column \* 3 Columns \* 25 uL/Well \* 1.1 overage = 660), use 800 so pulling from reference is easier
- Final concentration of Cold Ligand (Reference) in each well is 10 uM
- 5 is a Dilution Factor, the final volume in each well is 125 uL, we add 25 uL from the Hot-Ligand plate, (125/25=5)
- 10000 uM or 10 mM is starting reference concentration, usually

## Membrane Receptor calculation

Need:

- Protein Concentration (ug/uL) (Found from “Determining Protein Concentration”)
- Protein/Well (ug) (XP will provide this)

$$\text{Volume of Protein (uL)} = \frac{\text{Protein/Well (ug)} * 4000 \text{ (uL)}}{\text{Protein Concentration (ug/uL)} * 75 \text{ (uL)}}$$

$$\text{Volume of BB (uL)} = 4000 \text{ (uL)} - \text{Volume of Protein (uL)}$$

Where:

- 75 uL is the volume that will be dispensed into each well
- 4000 uL is approximate volume we need (8 Wells/Column \* 6 Columns \* 75 uL/Well \* 1.1 overage = 3960 uL), round to 4000 uL for convenience

## Determining Starting Concentration calculation

After obtaining actual radioactive counts (dpm) starting concentration (concentration of <sup>3</sup>H-Ligand in Well H) can be determined.

Need:

- Specific Activity (Ci/mmol)
- Actual Counts (dpm)

$$\text{Starting Concentration (nM)} = \frac{\text{Actual Counts (dpm)} * 10^9 \text{ (nM/M)} * 2^7}{2.22 * 10^{12} \text{ (dpm/Ci)} * \text{Specific Activity (Ci/mmol)} * 0.125 \text{ (mL/Well)}}$$

Where:

- $10^9$  in a unit conversion to nM from M
- $2^7$  is for the 7 serial 1:2 dilutions
- $2.22 * 10^{12}$  dpm/Ci is a constant
- 0.125 mL/Well is the final volume in each well