By Natalie Yoh (https://github.com/TallyYoh; tallyyoh@gmail.com) Cite as - Yoh, N., Kingston, T., McArthur, E., Aylen, O.E., Huang, J.C.C., Jinggong, E.R., Khan, F.A.A., Lee, B.P.Y.H., Mitchell, S.L.M., Bicknell, J.E., and Struebig, M.J. (2022). A machine learning framework to classify Southeast Asian echolocating bats, Ecological Indicators, 136. doi:10.1016/j.ecolind.2022.108696 This script applies the Borneo Bat Classifier (BBC) machine learning classifier to collated bat call parameter measurements from Borneo and assigns relevant labels The output includes: Pulse measurements Predicted classification labels (call type/sonotype/species) Confidence of classification labels Script to locate files based on labels & confidence on your desktop & reorder them for manual verification Software used to create classifier • R v3.6.3 Kaleidoscope v5.1.9g (Wildlife Acoustics, 2019) Adobe Audition v12.1.5 (Adobe Creative Cloud) Contributors Tigga Kingston (Department of Biological Sciences at Texas Tech University and the Southeast Asian Bat Conservation Research Unit, Lubbock, Texas, United Stated of America) Joe Chun-Chia Huang (Taiwan Forestry Research Institute, Taipei, Taiwan) • Ellen McArthur (Faculty of Resource Science and Technology, Universiti Malaysia Sarawak) • Benjamin P.Y.-H. Lee (Wildlife Management Division, National Parks Board, Singapore) • Faisal Ali Anwarali Khan (Faculty of Resource Science and Technology, Universiti Malaysia Sarawak) Emy Ritta Jinggong (Faculty of Resource Science and Technology, Universiti Malaysia Sarawak) • Oliver E. Aylen (, Department of Zoology, University of Otago, Otago, New Zealand) • Simon L. Mitchell (DICE, School of Anthropology and Conservation, University of Kent, Canterbury, United Kingdom) • Jake Bicknell (DICE, School of Anthropology and Conservation, University of Kent, Canterbury, United Kingdom) Matthew Struebig (DICE, School of Anthropology and Conservation, University of Kent, Canterbury, United Kingdom) Our aim is continuously test and update this tool as new reference data becomes available. Therefore, we would greatly appreciate users sharing any issues they find, particularly if this relates to species' IDs. Thank you! Updates from v1.0: • The call parameter data used in the v2.0 models is no longer scaled to ensure predictions are made on "true" numerical values The naming error for the FMqCF sonotypes has been corrected 1. PREPARE ENVIRONMENT Set memory size for Jupyter/R kernels In []: | memory.size() memory.limit(size=56000) Load packages Specify where your package directory In []: Dir_packages <-"C:/Users/Documents/R/win-library" Load packages In []: # For extracting call parameters library (bioacoustics) library(caret) # For supervised machine learning library(dplyr) # For data manipulation/selection library(gdata) # For data manipulation/selection # For progress bar library(pbapply) Specify user directories for importing & exporting !!! Before running, please ensure there is a back up copy of your raw files !!! Script includes moving files directly in file location Dir_clean_files_WAV = File location for 5 second calls (including all subfolders) • Dir_user_inputs = File location for csv inputs (e.g. threshold info) Dir_user_outputs = File location for data outputs Dir_classifier_models = File location for importing classifier models Dir_files_AutoID_WAV = File location for WAV files to be manually verified In []: <-"F:/Data_wav5sec_IDs_auto/CSVexports"</pre> 2. LOAD MODELS Set working directory to folder where models are stored In []: setwd(Dir classifier models) Load models Load stage 1 model to call type In []: model S1 type <- readRDS("model Type 1000 v2.0.rds")</pre> Load stage 2 model - to CF species model_S2_CF <- readRDS("model_CF_1000_v2.0.rds")</pre> Load stage 3 model - to FMqCF sonotype model S3 FMqCF <- readRDS("model FMqCF 2000 v2.0.rds")</pre> Load threshold reference information Set working directory & import csv In []: setwd(Dir user inputs) Data thres <-read.csv("ThresholdValues.csv")</pre> 3. IMPORT & EXTRACT CALL PARAMETERS Extracts call parameters from WAV files for classification using the Bioacoustic.R package WAV files for import should first have been subset to 5 second fragments to quantify a bat pass & be filtered for noise in Kaliedoscope or other sound analysis software. See Yoh et al. (2021) for more information Select file directories for where files are stored. This will perform extractions in two batches In []: files P1 <- dir(Dir clean files WAV, recursive = TRUE, full.names = TRUE, pattern = "[.]wav\$")</pre> Filter files for those identified as noise in Kaliedoscope In []: # convert to dataframe files P1 <-as.data.frame(files P1)</pre> # remove files listed as "noise" files_P1_crop <-as.character(files_P1[!grep1("NOISE", files_P1\$files_P1),])</pre> **Detect & extract pulse measurements** Extractions conducted using the Bioacoustics.R package threshold function (https://rdrr.io/cran/bioacoustics/) Extractions can be performed for time expansion 1 or 10 as necessary (use "time_exp = 10" if necessary) <- setNames(pblapply(files P1_crop, threshold detection, time exp = 1, threshold = 4, SNR thr = 4, FFT_size = 512,), basename(files_P1_crop)) Collate measurements Remove filenames where no values were extracted (e.g. only noise) In []: <- TDP1[lapply(TDP1, function(x) length(x\$data)) > 0] Keep the extracted features and merge in a single data frame for further analysis Data WAV raw <- do.call("rbind", c(lapply(TDP1, function(x) x\$data\$event data), list(stringsAsFactors = FALSE))) Remove file extention from filenames In []: Data WAV raw\$filename <-sub(pattern = "(.*)\\...*\$", replacement = "\\1", basename(Data WAV raw\$filename)) head (Data WAV raw) Include filename location information Extract filename from file locations In []: <-sub(pattern = "(.*)\\..*\$", replacement = "\\1", basename(files P1 crop))</pre> filename Create dataframe with full file location & filename In []: FileLoc <-data.frame(FileLoc=totalfileloc, filename=filename)</pre> Add to main dataframe Data WAV raw <-merge(Data WAV raw, FileLoc, by="filename") Clean & export pulse measurements Rename columns - Include/remove additionals where applicable In []: "freq_knee", "fc", "freq_bw_knee_fc", "bin_max_amp", "pc_freq_max_amp", "pc_freq_max", "pc_freq_min", "pc_knee", "temp_bw_knee_fc", "slope", "kalman_slope", "curve_neg", "curve_pos_start", "curve_pos_end", "mid offset" ,"snr", "hd", "smoothness", "FileLoc") Export raw call parameters In []: setwd(Dir user outputs) write.csv(Data WAV raw, file="Data Callparameters unclassified.csv", na = "NA") Create row ID for tracking pulses Data WAV raw\$ID <-as.vector(1:nrow(Data WAV raw))</pre> Isolate call parameter data Note - In previous versions, the call parameter data was scaled at this point. This step is no longer necessary and has been removed In []: Data CallValues Scaled <-as.data.frame(subset(Data WAV raw, select = -c(Filename, FileLoc, starting time, ID))) Select row information Data RowInfo <-subset(Data WAV raw, select = c(ID, Filename, FileLoc, starting time))</pre> Recombine In []: Data WAV scaled <-droplevels(cbind(Data_RowInfo, Data_CallValues_Scaled))</pre> 4. PERFORM CLASSIFICATIONS - STAGE 1 Predict the call type of each file using the first machine learning model Run predictions Run prediction without confidence values In []: predictionsResultsType <-predict(model_S1_type, Data_CallValues_Scaled)</pre> Run prediction with confidence values In []: PredictionResultsTypeProb <-predict(model S1 type, Data CallValues Scaled, type = "prob")</pre> PredictionResultsTypeProb\$ID <-as.vector(Data WAV raw\$ID)</pre> PredictionResultsTypeProb\$ID <-as.factor(as.character(PredictionResultsTypeProb\$ID))</pre> Combine predictions with confidence values In []: PredictionResultsTypeCombined <-cbind(PredictionResultsTypeProb, predictionsResultsType)</pre> Combine with file information PredictionsFinalStage1 <-merge(PredictionResultsTypeCombined, Data WAV scaled, by="ID")</pre> **Export stage 1 predictions** In []: | setwd(Dir user outputs) write.csv(PredictionsFinalStage1, file="Data PredictionsStage1.csv", na = "NA") Summarise results Summarises the pulse predictions to call type identification to the file/bat pass level Convert to factor for grouping In []: PredictionsFinalStage1\$ID <-as.factor(PredictionsFinalStage1\$ID)</pre> Create vectors for grouping columns In []: <- c("FM", "CF", "FMqCF", "QCF") cols sp cols ID <- c("ID", "predictionsResultsType")</pre> cols Files <- c("Filename", "predictionsResultsType")</pre> Isolate the confidence of the predicted species into new column Temp S1 Max ID <- PredictionsFinalStage1 %>% group by (across (all of (cols ID))) %>% mutate(MaxByID = max(c(FM, CF, FMqCF, QCF), na.rm = T))Find the pulse of highest confidence within each file for each species RES S1 summary <- Temp S1 Max ID %>% group by(across(all of(cols Files))) %>% summarise(MaxbyFile = max(MaxByID, na.rm = T)) Rename columns names(RES S1 summary)<-c("Filename","S1 Prediction","S1 Accuracy")</pre> Isolate files for manual verification & create library The following steps if for users who are only using the stage 1 classifications. Skip to stage 2 (section 5) if you are using stage 2/3 classifications to sonotype/species Selects WAV files which do not reach the necessary confidence threshold using their original filepathways and copies them into a new filepathway based on ID prediction & confidence threshold. Determine which files need manual verification Rename column levels to match colnames(Data_thres) <- c("Prediction", "Threshold")</pre> Merge confidence threshold information with the predictions data In []: RES S1 summary <- merge(RES S1 summary, Data thres, by = "Prediction", keep.all=TRUE)</pre> Create threshold level column RES S1 summary\$ThresLevel <-"" Ensure accuracy column is numeric RES_S1_summary\$Accuracy <-as.numeric(RES_S1_summary\$Accuracy) Remove predictions below 60% confidence RES S1 summary <-RES S1 summary [RES S1 summary \$Accuracy > 0.59,] Loop to determine which files met the necessarily confidence threshold In []: for (y in 1:nrow(RES S1 summary)) { if((RES S1 summary\$Accuracy[y]*100) == RES S1 summary\$Threshold[y]) { RES S1 summary\$ThresLevel[y] <- "Met"</pre> else if ((RES S1 summary\$Accuracy[y]*100) > RES S1 summary\$Threshold[y]) { RES S1 summary\$ThresLevel[y] <- "Met" }</pre> else if ((RES S1 summary\$Accuracy[y]*100) < RES S1 summary\$Threshold[y]) { RES total sum\$ThresLevel[y] <- "Not Met" }</pre> Filter data for files which did not meet the confidence threshold In []: DF NotMet <-filter(RES S1 summary, (ThresLevel=="Not Met"))</pre> Remove repeated files so a file is only manually checked once DF NotMet unique <- DF NotMet[!duplicated(DF NotMet['filename']),]</pre> Save data outputs setwd(Dir_user_outputs) write.csv(DF_NotMet_unique, file="DF_NotMet_unique.csv", na = "NA") write.csv(RES_S1_summary, file="Data_PredictionsSummary max.csv", na = "NA") !!! THE FOLLOWING CODE WILL MOVE FILES DIRECTORY ON YOUR COMPUTER !!! !!! ENSURE IT IS WORKING CORRECTLY USING A TEST FILE/BACK UP YOUR DATA BEFORE PROCEEDING !!! Specify ID levels In []: Lvls stageType <-levels(as.factor(RES S1 summary\$Prediction))</pre> Not reversible: Loop to create new folder pathway and copy WAV files - user needs to update pathway below In []: for (S in 1:length(Lvls stageType)) { # Specify prediction level TYPE <-Lvls_stageType[S] # Filter data for target species & confidence threshold RES_total_target <-filter(DF_NotMet_unique_cleaned, Prediction ==TYPE)</pre> # Create filename vector including file locations Sp file list <-as.character(RES total target\$FileLoc) # Create output folder for ID level setwd(Dir files AutoID WAV) newdir <-paste0(TYPE,"_", "ThresholdNotMet")</pre> dir.create(newdir) # Create directory in R to Species specific folder Dir_temp <-paste0("F:/Data_wav5sec_IDs_auto/",newdir) # **** NEEDS UPDATING BY USER **** # Go back to input WAV files directory setwd(Dir_clean_files_WAV2012) # Move each individual file to new directory for (F in 1:length(Sp_file_list)){ # Select file FILE <-Sp_file_list[F] # copy files file.copy(FILE, Dir temp) # Progress bar print(c("Loop", S, "from", length(Lvls_stageType), "File", F, "from", length(Sp_file_list))) ----- End for users only classifying to call type ------5. PERFORM CLASSIFICATIONS - STAGE 2 Split data based on the predictions from stage 1 Remove predictions below 60% confidence In []: Temp_S1_Max_ID <-Temp_S1_Max_ID[Temp_S1_Max_ID\$MaxByID > 0.59,] Divide into Type specific datasets based on predictions In []: Stage1 FM <-Temp_S1_Max_ID[Temp_S1_Max_ID\$predictionsResultsType=="FM",]</pre> Stage1_QCF <-Temp_S1_Max_ID[Temp_S1_Max_ID\$predictionsResultsType=="QCF",]</pre> Stage1_FMqCF <-Temp_S1_Max_ID[Temp_S1_Max_ID\$predictionsResultsType=="FMqCF",] <-Temp S1 Max ID[Temp S1 Max ID\$predictionsResultsType=="CF",]</pre> Stage1 CF For species which were identified as "CF" (constant-frequency) conduct a second classification stage using the second machine learning model which prioritises maximum frequency **Prepare CF data** Remove identifying information from CF data Data CallValues CF noID <-Stage1_CF[,c("duration","freq_max_amp" , "freq_max",</pre> "freq_min", "bandwidth", "freq_start", "freq_center", "freq_end", "freq_knee", "fc", "freq_bw_knee_fc", "bin_max_amp", "pc_freq_max_amp", "pc_freq_max", "pc_freq_min", "pc_knee", "temp_bw_knee_fc", "slope", "kalman_slope", "curve_neg", "curve_pos_start", "curve_pos_end", "mid_offset" , "snr", "hd", "smoothness")] Filter for complete cases In []: Data CallValues CF noID <-Data CallValues CF noID[complete.cases(Data CallValues CF noID),]</pre> Data CallValues CF noID <-drop.levels(Data CallValues CF noID)</pre> Run predictions Run prediction without confidence values In []: predictionsResultsCF <-predict(model S2 CF, Data CallValues CF noID)</pre> Run prediction without confidence <-predict(model S2 CF, Data CallValues CF noID, type = "prob")</pre> PredictionResultsProb CF <-Stage1 CF\$ID PredictionResultsProb_CF\$ID Combine predictions with confidence values In []: <-cbind(PredictionResultsProb CF, predictionsResultsCF)</pre> PredictionResultsCombined CF Combine with file information In []: PredictionsFinalStage2 <-merge(PredictionResultsCombined CF, Stage1 CF, by="ID")</pre> **Export stage 2 predictions** In []: setwd(Dir user outputs) write.csv(PredictionsFinalStage2, file="Data PredictionsStage2.csv", na = "NA") Run confidence thresholds Creates table to see which files meet the confidence thresholds necessary for file structure later Convert ID to factor for grouping In []: PredictionsFinalStage2\$ID <-as.factor(PredictionsFinalStage2\$ID)</pre> Identify which CF are present in the data In []: Levels CF <-levels(as.factor(PredictionsFinalStage2\$predictionsResultsCF))</pre> Create vector for grouping species (user needs to edit depending on the species listed in Levels_CF) cols sp "CF_Rbor" , "CF_Rcre", "CF_Rluc" , "CF_Rphi", "CF_Rsed", "CF_Rtri") Create vectors for grouping In []: <- c("ID", "predictionsResultsCF")</pre> cols ID cols Files <- c("Filename", "predictionsResultsCF")</pre> Isolate the confidence of the predicted species into new column (user needs to edit depending on the species listed in Levels_CF) In []: Temp S2 Max ID <- PredictionsFinalStage2 %>% group by (across (all of (cols ID))) %>% mutate(MaxByID = max(c(CF H140, CF Hate,CF Hbic,CF Hcer,CF Hcox , CF_Hdia,CF_Hgal, CF_Hlar, CF_Hrid, CF_Racu , CF Raff, CF Rbor, CF_Rcre, CF_Rluc , CF Rphi, CF Rsed, CF Rtri), na.rm = T)) Find the pulse of highest confidence within each file for each species RES S2 summary <- Temp S2 Max ID %>% group by(across(all of(cols Files))) %>% summarise(MaxbyFile = max(MaxByID, na.rm = T)) Rename columns In []: names (RES_S2_summary) <-c ("Filename", "S2_Prediction", "S2_Accuracy")</pre> 6. PERFORM CLASSIFICATIONS - STAGE 3 For species which were identified as "FMqCF" (frequency modulated quasi-constant frequency) conduct a second classification stage using the third machine learning model which prioritises call shape Prepare FMqCF data Remove identifying information from FMqCF data In []: Data CallValues FMqCF noID <-Stage1 FMqCF[,c("duration","freq max amp" , "freq max",</pre> "freq min", "bandwidth", "freq_start", "freq_center", "freq_end", "freq_knee", "fc", "freq bw knee fc", "bin max amp", "pc freq max amp", "pc_freq_max", "pc_freq_min", "pc_knee", "temp bw knee fc", "slope", "kalman slope", "curve_neg", "curve_pos_start", "curve_pos_end", "mid offset" ,"snr", "hd", "smoothness")] Filter for complete cases In []: Data CallValues FMqCF noID <-Data CallValues FMqCF noID[complete.cases(Data CallValues FMqCF noID),]</pre> Data CallValues FMqCF noID <-drop.levels(Data CallValues FMqCF noID)</pre> Run predictions Run predication without confidence predictionsResultsFMqCF <-predict(model S3 FMqCF, Data CallValues FMqCF noID)</pre> Run predictions with confidence In []: PredictionResultsProb FMqCF <-predict(model_S3_FMqCF, Data_CallValues_FMqCF_noID, type = "prob")</pre> PredictionResultsProb FMqCF\$ID <-Stage1 FMqCF\$ID Combine predictions with confidence values PredictionResultsCombined FMqCF <-cbind(PredictionResultsProb FMqCF, predictionsResultsFMqCF)</pre> Combine with file information In []: PredictionsFinalStage3 <-merge(PredictionResultsCombined FMqCF, Stage1 FMqCF, by="ID")</pre> **Export stage 3 predictions** setwd(Dir_user_outputs) write.csv(PredictionsFinalStage3, file="Data PredictionsStage3.csv", na = "NA") Run confidence thresholds Creates table to see which files meet the confidence thresholds necessary for file structure later Convert ID to factor for grouping In []: PredictionsFinalStage3\$ID <-as.factor(PredictionsFinalStage3\$ID)</pre> Identify which FMqCF are present in the data In []: Levels FMqCF <-levels(as.factor(PredictionsFinalStage3\$predictionsResultsFMqCF))</pre> Create vector for grouping species (user may need to edit depending on the species listed in Levels_FMqCF) <- c("FMqCF1" , "FMqCF2", "FMqCF3", "FMqCF4" , "FMqCF5", "LF" , "LF Acup")</pre> cols sp Create vectors for grouping In []: cols ID <- c("ID", "predictionsResultsFMqCF")</pre> cols Files <- c("Filename", "predictionsResultsFMqCF")</pre> Isolate the confidence of the predicted species into new column Temp S3 Max ID <- PredictionsFinalStage3 %>% group by (across (all of (cols ID))) %>% mutate(MaxByID = max(c(FMqCF1 , FMqCF2,FMqCF3,FMqCF4 , FMqCF5, LF , LF Acup) , na.rm = T))Find the pulse of highest confidence within each file for each species RES S3 summary <- Temp S3 Max ID %>% group_by(across(all_of(cols_Files))) %>% summarise(MaxbyFile = max(MaxByID, na.rm = T)) Rename columns In []: names(RES S3 summary)<-c("Filename","S3 Prediction","S3 Accuracy")</pre> 7. COMBINE FINAL PREDICTIONS Collate predictions from each classification stage & isolate files for manual verification Create summaries Select relevant columns In []: SummaryQCF <-Stage1 QCF[, c("ID","predictionsResultsType","MaxByID", "Filename", "FileLoc")]</pre> <-Stage1 FM[, c("ID", "predictionsResultsType", "MaxByID", "Filename", "FileLoc")]</pre> SummaryFM SummaryCF <-Temp S2 Max ID[, c("ID", "predictionsResultsCF", "MaxByID", "Filename", "FileLoc")]</pre> ${\tt SummaryFMqCF}$ <-Temp S3 Max ID[, c("ID", "predictionsResultsFMqCF", "MaxByID", "Filename", "FileLoc")]</pre> Rename column names to match In []: colnames(SummaryQCF) <-c("ID","Prediction","Accuracy","Filename", "FileLoc")</pre> colnames(SummaryFMqCF) <-c("ID", "Prediction", "Accuracy", "Filename", "FileLoc")</pre> Join predictions In []: RES total raw <-rbind(SummaryQCF, SummaryFM, SummaryCF, SummaryFMqCF)</pre> Subset again for files above 60% confidence This will subset for files identified in stages 2 and 3 to sonotype/species In []: RES total raw <-RES total raw[RES total raw\$Accuracy>0.59,] **Export final predictions** In []: setwd(Dir user outputs) write.csv(RES total raw, file="Data PredictionsSummary.csv", na = "NA") Summarise results Summarises the pulse predictions to sonotype/species identification to the file/bat pass level Convert to factor for grouping In []: RES total raw\$ID <-as.factor(RES total raw\$ID)</pre> Create vectors for grouping columns In []: cols sp <- levels(as.factor(RES total raw\$Prediction))</pre> cols Files <- c("Filename", "Prediction")</pre> Find the pulse of highest confidence within each file for each species In []: RES_total_sum <- RES_total_raw %>% group_by(across(all_of(cols_Files))) %>% summarise(MaxbyFile = max(Accuracy, na.rm = T)) Rename columns names (RES total sum) <-c("filename", "Prediction", "Accuracy")</pre> Readd file location RES total sum <-merge(RES total sum, FileLoc, by="filename", all= FALSE) Isolate files for manual verification & create library The following steps if for users who are using stage 2/3 classifications. Selects WAV files which do not reach the necessary confidence threshold using their original filepathways and copies them into a new filepathway based on ID prediction & confidence threshold. Determine which files need manual verification Rename column levels to match colnames(Data thres) <- c("Prediction", "Threshold")</pre> Merge confidence threshold information with the predictions data In []: RES total sum <- merge(RES total sum, Data thres, by = "Prediction", keep.all=TRUE)</pre> Create threshold level column RES total sum\$ThresLevel <-"" Ensure accuracy column is numeric RES total sum\$Accuracy <-as.numeric(RES_total_sum\$Accuracy) Remove predictions below 60% confidence In []: RES total sum <-RES_total_sum[RES_total_sum\$Accuracy > 0.59,] Loop to determine which files met the necessarily confidence threshold for (y in 1:nrow(RES total sum)){ if((RES total sum\$Accuracy[y]*100) == RES total sum\$Threshold[y]) { RES total sum\$ThresLevel[y] <- "Met"</pre> else if ((RES total sum\$Accuracy[y]*100) > RES total sum\$Threshold[y]) { RES total sum\$ThresLevel[y] <- "Met" }</pre> else if ((RES total sum\$Accuracy[y]*100) < RES total sum\$Threshold[y]) {</pre> RES total sum\$ThresLevel[y] <- "Not Met" }</pre> Filter data for files which did not meet the confidence threshold In []: DF NotMet <-filter(RES total sum, (ThresLevel=="Not Met"))</pre> Remove repeated files so a file is only manually checked once DF NotMet unique <- DF NotMet[!duplicated(DF_NotMet['filename']),]</pre> Save data outputs In []: setwd(Dir user outputs) write.csv(DF NotMet unique, file="DF NotMet unique.csv", na = "NA") write.csv(RES total sum, file="Data PredictionsSummary max.csv", na = "NA") !!! THE FOLLOWING CODE WILL MOVE FILES DIRECTORY ON YOUR COMPUTER !!! !!! ENSURE IT IS WORKING CORRECTLY USING A TEST FILE/BACK UP YOUR DATA BEFORE PROCEEDING !!! Specify ID levels In []: Lvls stageAll <-levels(as.factor(RES total sum\$Prediction))</pre> Not reversible: Loop to create new folder pathway and copy WAV files - user needs to update pathway below

BATS OF BORNEO SEMI-AUTOMATED CLASSIFIER FOR

ECHOLOCATION CALLS (2021)

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