BATS OF BORNEO SEMI-AUTOMATED CLASSIFIER FOR **ECHOLOCATION CALLS (2021)** By Natalie Yoh (https://github.com/TallyYoh; tallyyoh@gmail.com) Cite as - Yoh, N., Kingston, T., McArthur, E., Aylen, O.E., Huang, J.C.C., Jinggong, E.R., Khan, F.A.A., Lee, B.P.Y.H., Mitchell, S.L.M., Bicknell, J.E., and Struebig, M.J. (2022). A machine learning framework to classify Southeast Asian echolocating bats, Ecological Indicators, 136. doi:10.1016/j.ecolind.2022.108696 This script applies the Borneo Bat Classifier (BBC) machine learning classifier to collated bat call parameter measurements from Borneo and assigns relevant labels The output includes: Pulse measurements Predicted classification labels (call type/sonotype/species) • Confidence of classification labels Script to locate files based on labels & confidence on your desktop & reorder them for manual verification Software used to create classifier • R v3.6.3 Kaleidoscope v5.1.9g (Wildlife Acoustics, 2019) Adobe Audition v12.1.5 (Adobe Creative Cloud) Contributors • Tigga Kingston (Department of Biological Sciences at Texas Tech University and the Southeast Asian Bat Conservation Research Unit, Lubbock, Texas, United Stated of America) Joe Chun-Chia Huang (Taiwan Forestry Research Institute, Taipei, Taiwan) Ellen McArthur (Faculty of Resource Science and Technology, Universiti Malaysia Sarawak) • Benjamin P.Y.-H. Lee (Wildlife Management Division, National Parks Board, Singapore) Faisal Ali Anwarali Khan (Faculty of Resource Science and Technology, Universiti Malaysia Sarawak) Emy Ritta Jinggong (Faculty of Resource Science and Technology, Universiti Malaysia Sarawak) Oliver E. Aylen (, Department of Zoology, University of Otago, Otago, New Zealand) Simon L. Mitchell (DICE, School of Anthropology and Conservation, University of Kent, Canterbury, United Kingdom) Jake Bicknell (DICE, School of Anthropology and Conservation, University of Kent, Canterbury, United Kingdom) • Matthew Struebig (DICE, School of Anthropology and Conservation, University of Kent, Canterbury, United Kingdom) Our aim is continuously test and update this tool as new reference data becomes available. Therefore, we would greatly appreciate users sharing any issues they find, particularly if this relates to species' IDs. Thank you! 1. PREPARE ENVIRONMENT Set memory size for Jupyter/R kernels memory.size() memory.limit(size=56000) Load packages Specify where your package directory <-"C:/Users/Documents/R/win-library" Dir packages setwd(Dir packages) Load packages library(bioacoustics) # For extracting call parameters # For supervised machine learning library(caret) library(dplyr) # For data manipulation/selection library (qdata) # For data manipulation/selection library (pbapply) # For progress bar Specify user directories for importing & exporting !!! Before running, please ensure there is a back up copy of your raw files !!! Script includes moving files directly in file location • Dir_clean_files_WAV = File location for 5 second calls (including all subfolders) • Dir_clean_files_WAVP1 = File location for 5 second calls (part 1 if WAV files across two subfolders*) • Dir_clean_files_WAVP2 = File location for 5 second calls (part 2 if WAV files across two subfolders*) Dir_user_inputs = File location for csv inputs (e.g. threshold info) Dir_user_outputs = File location for data outputs • Dir_classifier_models = File location for importing classifier models Dir_files_AutoID_WAV = File location for WAV files to be manually verified *This provides the infrastructure to perform the pulse measurements in 2+ batches Dir_clean_files_WAV

Dir_clean_files_WAVP1

Dir_clean_files_WAVP1

Dir_clean_files_WAVP2

Dir_user_inputs

Dir_user_outputs

Dir_classifier_models

Dir_files_AutoID_WAV

<-"F:/Data_wav5sec_clean/All_WAV/WAVP2"

<-"F:/R_inputs"

<-"F:/Data_wav5sec_IDs_auto/CSVexports"

<-"F:/Data_wav5sec_IDs_auto" 2. LOAD MODELS Set working directory to folder where models are stored setwd(Dir classifier models) Load models Load stage 1 model to call type model S1 type <- readRDS("model Type 1000.rds")</pre> Load stage 2 model - to CF species model S2 CF <- readRDS("model CF 1000.rds")</pre> Load stage 3 model - to FMqCF sonotype model S3 FMqCF <- readRDS("model FMqCF 2000.rds")</pre> Load threshold reference information Set working directory & import csv setwd(Dir user inputs) Data thres <-read.csv("ThresholdValues.csv")</pre> 3. IMPORT & EXTRACT CALL PARAMETERS Extracts call parameters from WAV files for classification using the Bioacoustic.R package WAV files for import should first have been subset to 5 second fragments to quantify a bat pass & be filtered for noise in Kaliedoscope or other sound analysis software. See Yoh et al. (2021) for more information Select file directories for where files are stored. This will perform extractions in two batches files P1 <- dir(Dir clean files WAVP1, recursive = TRUE, full.names = TRUE, pattern = "[.]wav\$")</pre> files P2 <- dir(Dir clean files WAVP2, recursive = TRUE, full.names = TRUE, pattern = "[.]wav\$")</pre> Filter files for those identified as noise in Kaliedoscope # convert to dataframe files_P1 <-as.data.frame(files_P1)
files P2 <-as.data.frame(files_P2) files P2 <-as.data.frame(files P2)</pre> # remove files listed as "noise" files_P1_crop <-as.character(files_P1[!grep1("NOISE", files_P1\$files_P1),])</pre> files_P2_crop <-as.character(files_P2[!grep1("NOISE", files_P2\$files_P2),])</pre> **Detect & extract pulse measurements** Extractions conducted using the Bioacoustics.R package threshold function (https://rdrr.io/cran/bioacoustics/) Extractions can be performed for time expansion 1 or 10 as necessary (use "time_exp = 10" if necessary) <- setNames(TDP1 pblapply(files P1_crop, threshold detection, time exp = 1, threshold = 4,SNR thr = 4, FFT size = 512,), basename(files P1 crop)) TDP2 <- setNames(pblapply(files P2 crop, threshold detection, time exp = 1, threshold = 4,SNR thr = 4, FFT size = 512,), basename(files P2 crop)) Collate measurements Remove filenames where no values were extracted (e.g. only noise) TDP1 <- TDP1[lapply(TDP1, function(x) length(x\$data)) >] TDP2 <- TDP2[lapply(TDP2, function(x) length(x\$data)) >] Keep the extracted features and merge in a single data frame for further analysis Data WAV rawP1 <- do.call("rbind", c(lapply(TDP1, function(x) x\$data\$event data), list(stringsAsFactors = FALSE))) Data_WAV_rawP2 <- do.call("rbind", c(lapply(TDP2, function(x) x\$data\$event_data),</pre> list(stringsAsFactors = FALSE))) Row bind data Data WAV raw <-rbind(Data_WAV_rawP1, Data_WAV_rawP2)</pre> Remove file extention from filenames Data WAV raw\$filename <-sub(pattern = "(.*)\\...*\$", replacement = "\\1", basename(Data WAV raw\$filename)) head (Data WAV raw) Calculate number of audio events extracted <-nrow(Data WAV raw))</pre> (Rawcalls Include filename location information Combine file locations totalfileloc <-c(files P1 crop, files P2 crop) Extract filename only <-sub(pattern = "(.*)\\..*\$", replacement = "\\1", basename(totalfileloc))</pre> filename Create dataframe with full file location & filename <-data.frame(FileLoc=totalfileloc, filename=filename)</pre> FileLoc Add to main dataframe Data WAV raw <-merge(Data WAV raw, FileLoc, by="filename") Clean & export pulse measurements Rename columns - Include/remove additionals where applicable "freq_knee", "fc", "freq_bw_knee_fc", "bin_max_amp", "pc_freq_max_amp", "pc_freq_max", "pc_freq_min", "pc_knee", "temp_bw_knee_fc", "slope", "kalman_slope", "curve_neg", "curve_pos_start", "curve_pos_end", "mid_offset" ,"snr", "hd", "smoothness", "FileLoc") **Export raw call parameters** setwd(Dir user outputs) write.csv(Data_WAV_raw, file="Data_Callparameters_unclassified.csv", na = "NA") Scale call parameters Create row ID for tracking pulses Data WAV raw\$ID <-as.vector(1:nrow(Data WAV raw))</pre> Scale call parameter data Data CallValues Scaled <-scale(as.data.frame(subset(Data WAV raw, select = -c(Filename, FileLoc, starting time, ID)), center = TRUE, scale = TRUE)) Select row information <-subset(Data WAV raw, select = c(ID, Filename, FileLoc, starting time))</pre> Data RowInfo Recombine Data WAV scaled <-droplevels(cbind(Data_RowInfo, Data_CallValues_Scaled))</pre> 4. PERFORM CLASSIFICATIONS - STAGE 1 Predict the call type of each file using the first machine learning model **Run predictions** Run prediction without confidence values predictionsResultsType <-predict(model S1 type, Data CallValues Scaled)</pre> Run prediction with confidence values PredictionResultsTypeProb <-predict(model S1 type, Data CallValues Scaled, type = "prob")</pre> PredictionResultsTypeProb\$ID <-as.vector(Data WAV raw\$ID)</pre> PredictionResultsTypeProb\$ID <-as.factor(as.character(PredictionResultsTypeProb\$ID))</pre> Combine predictions with confidence values PredictionResultsTypeCombined <-cbind(PredictionResultsTypeProb, predictionsResultsType)</pre> Combine with file information PredictionsFinalStage1 <-merge(PredictionResultsTypeCombined, Data WAV scaled, by="ID")</pre> **Export stage 1 predictions** setwd(Dir user outputs) write.csv(PredictionsFinalStage1, file="Data PredictionsStage1.csv", na = "NA") Run confidence thresholds Creates table to see which files meet the confidence thresholds necessary for file structure later Create vectors for loop PredictionsFinalStage1\$ID <-as.factor(PredictionsFinalStage1\$ID)</pre> codes <-levels(droplevels(PredictionsFinalStage1\$ID))</pre> Create empty dataframe RES Class firststage <-data.frame(matrix(ncol = 11, nrow =)) Loop to determine which thresholds the files achieved for call type for (i in 1:length(codes)) { # Select row no. code<-codes[i]</pre> # Select results/file step1<-PredictionsFinalStage1[PredictionsFinalStage1\$ID==code,]</pre> # list species / file step1\$predictionsResultsType <-as.factor(step1\$predictionsResultsType)</pre> <-c(levels(droplevels(step1\$predictionsResultsType)))</pre> step1\$predictionsResultsType <-as.character(step1\$predictionsResultsType)</pre> # Loop for different species within one recording for (y in 1:length(sps)){ #select a single species sp<-sps[y] <-step1[step1\$predictionsResultsType==sp,]</pre> stepsps <-select(stepsps,sp)</pre> stepsp maxprob <-max(stepsp) # Specify confidence thresholds if (maxprob>=0.6) R60<-as.character("done")</pre> if (maxprob<0.6) R60<-as.character("check")</pre> if (maxprob>=0.65) R65<-as.character("done")</pre> if (maxprob<0.65) R65<-as.character("check")</pre> if (maxprob>=0.7) R70<-as.character("done")</pre> if (maxprob<0.7) R70<-as.character("check") if (maxprob>=0.75) R75<-as.character("done")</pre> if (maxprob<0.75) R75<-as.character("check")</pre> if (maxprob>=0.8) R80<-as.character("done")</pre> if (maxprob<0.8) R80<-as.character("check") if (maxprob>=0.85) R85<-as.character("done")</pre> if (maxprob<0.85) R85<-as.character("check")</pre> if (maxprob>=0.9) R90<-as.character("done")</pre> if (maxprob<0.9) R90<-as.character("check")</pre> if (maxprob>=0.95) R95<-as.character("done")</pre> if (maxprob<0.95) R95<-as.character("check") temp <-as.character(c(code, sp, maxprob, R60, R65, R70, R75, R80, R85, R90, R95)) # Bind results to main table RES Class firststage <-rbind(RES Class firststage,temp)</pre> RES Class firststage *>* mutate if(is.factor, as.character) -> RES Class firststage # Show progress print(c("Loop ", i, "from", length(codes))) Rename columns names(RES Class firststage)<-c("ID", "Prediction", "S1 Accuracy", "S1 R60",</pre> "S1 R65", "S1 R70", "S1 R75", "S1 R80", "S1 R85", "S1 R90", "S1 R95") Re-add call parameter/file info RES_Class_firststage <-merge(RES_Class_firststage, PredictionsFinalStage1, by="ID") (i) Isolate files for manual verification & create library The following series (i) if for users who are only using the stage 1 classifications. Skip the following series (i) if you are using stage 2/3 classifications to sonotype/species Selects WAV files which do not reach the necessary confidence threshold using their original filepathways and copies them into a new filepathway based on ID prediction & confidence threshold. !!! THE FOLLOWING CODE WILL MOVE FILES DIRECTORY ON YOUR COMPUTER !!! !!! ENSURE IT IS WORKING CORRECTLY USING A TEST FILE/BACK UP YOUR DATA BEFORE PROCEEDING !!! Summarise pulses to bat passes Determines the highest accuracy recorded for each ID for each file to quantify bat activity Create empty dataframe RES firststage_sum <-data.frame(matrix(ncol=ncol(RES_Class_firststage),nrow=0))</pre> Rename columns colnames(RES firststage sum) <-colnames(RES Class firststage)</pre> Create vector for filenames AllFiles <-levels(as.factor(RES Class firststage\$Filename))</pre> Summarise bat passes by the highest accuracy pulse per file per species for (a in 1:length(AllFiles)) { # Select file <-AllFiles[a] FileLabel # Subset main data for file TempDFByfile <-RES Class firststage[RES Class firststage\$Filename==FileLabel,]</pre> # Create vector for the no. of species within said file TempSpVector <-levels(as.factor(TempDFByfile\$Prediction))</pre> # For each species within said file find the highest accuracy for (b in 1:length(TempSpVector)){ # Select species TempSp <-TempSpVector[b]</pre> # Subset filename for pulses of that species only <-TempDFByfile[TempDFByfile\$Prediction == TempSp,]</pre> # Find highest accuracy value obtained <-which.max(TempDFSp\$S1 Accuracy)</pre> # Select row for pulse of highest accuracy <-TempDFSp[MaxAcc,]</pre> TempRow # Rowbind into main datatable RES firststage sum <-rbind(RES firststage sum, TempRow) print(c("Loop", a, "from", length(AllFiles))) Select files for manual verification (i) Add data threshold information colnames(Data thres) <- c("Prediction", "Threshold")</pre> <- merge(RES firststage sum, Data thres, by = "Prediction", keep.all=TRUE)</pre> RES firststage sum2 RES firststage sum2\$ThresLevel <-"" RES firststage sum2\$\$1 Accuracy <-as.numeric(RES firststage sum2\$\$1 Accuracy) # make sure column is numeric for (y in 1:nrow(RES firststage sum2)) { $if((RES_firststage_sum2\$S1_Accuracy[y]*100) == RES_firststage_sum2\$Threshold[y])$ { RES firststage sum2\$ThresLevel[y] <- "Met"</pre> else if ((RES firststage sum2\$S1 Accuracy[y]*100) > RES firststage sum2\$Threshold[y]) { RES firststage sum2\$ThresLevel[y] <- "Met" }</pre> else if ((RES firststage sum2\$S1 Accuracy[y]*100) < RES firststage sum2\$Threshold[y]) { RES firststage sum2\$ThresLevel[y] <- "Not Met" }</pre> (ii) Specify ID levels Lvls stage1 <-levels(as.factor(RES firststage sum2\$Prediction)) (iii) Remove duplicates # Filter data for confidence threshold DF NotMet <-filter(RES firststage sum2, (ThresLevel=="Not Met"))</pre> # Remove repeated files so a file is only manually checked once DF NotMet unique <- DF NotMet[!duplicated(DF NotMet['Filename']),]</pre> (iv) Not reversible: Loop to create new folder pathway and copy WAV files - user needs to update pathway below for (S in 1:length(Lvls stage1)){ # Specify prediction level TYPE <-Lvls_stage1[S] # Filter data for target call type & confidence threshold RES_firststage_target <-filter(DF_NotMet_unique, (ThresLevel=="Not Met") & (Prediction ==TYPE))</pre> # Create filename vector including file locations Ty_file_list <-as.character(RES_firststage target\$FileLoc)</pre> # Remove duplicates (shouldn't remove any values) Ty_file_list <-Ty_file_list[!duplicated(Ty_file_list)]</pre> # Create output folder for ID level setwd(Dir_files_AutoID_WAV) newdir <-paste0(TYPE," ", "ThresholdNotMet")</pre> dir.create(newdir) # Create directory in R to call type specific folder Dir temp <-paste0("F:/Data wav5sec IDs auto/", newdir) # **** NEEDS UPDATING BY THE USER **** # Go back to input WAV files directory setwd(Dir_clean_files_WAV) # Move each individual file to new directory for (F in 1:length(Ty_file_list)){ # Select file FILE <-Ty_file_list[F] # copy files file.copy(FILE, Dir_temp) # Progress bar print(c("Loop", S, "from", length(Lvls_stage1))) ----- End for users only classifying to call type ------Split data based on call type predictions for stage 2 Divide into Type specific datasets based on predictions Stage1_FM <-RES_Class_firststage[RES_Class_firststage\$Prediction=="FM",] Stage1 QCF <-RES Class_firststage[RES_Class_firststage\$Prediction=="QCF",] Stage1_FMqCF <-RES_Class_firststage[RES_Class_firststage\$Prediction=="FMqCF",]</pre> Stage1_CF <-RES_Class_firststage[RES_Class_firststage\$Prediction=="CF",]</pre> 5. PERFORM CLASSIFICATIONS - STAGE 2 For species which were identified as "CF" (constant-frequency) conduct a second classification stage using the second machine learning model which prioritises maximum frequency Remove Identifying information from CF data Data_CallValues_CF_noID <-Stage1_CF[,c("duration", "freq_max_amp" , "freq_max", "freq_min", "bandwidth", "freq_start", "freq_center", "freq_end" "freq_knee", "fc", "freq_bw_knee_fc", "bin_max_amp", "pc freq max amp", "pc_freq_max", "pc_freq_min", "pc knee", "temp bw knee fc", "slope", "kalman slope", "curve_neg", "curve_pos_start", "curve_pos_end", "mid_offset", "snr", "hd", "smoothness")] Data CallValues CF noID <-Data CallValues CF noID[complete.cases(Data CallValues CF noID),]</pre> Data CallValues CF noID <-drop.levels(Data CallValues CF noID)</pre> Run predictions Run prediction without confidence values predictionsResultsCF <-predict(model_S2_CF, Data_CallValues_CF_noID)</pre> Run prediction with confidence values <-predict(model_S2_CF, Data_CallValues_CF_noID, type = "prob")</pre> PredictionResultsProb CF PredictionResultsProb CF\$ID <-Stage1 CF\$ID Combine predictions with confidence values PredictionResultsCombined CF <-cbind(PredictionResultsProb CF, predictionsResultsCF)</pre> Combine with file information PredictionsFinalStage2 <-merge(PredictionResultsCombined CF, Stage1 CF, by="ID")</pre> Export stage 2 predictions setwd(Dir user_outputs) write.csv(PredictionsFinalStage2, file="Data PredictionsStage2.csv", na = "NA") Run confidence thresholds Creates table to see which files meet the confidence thresholds necessary for file structure later Create vectors for loop PredictionsFinalStage2\$ID <-as.factor(PredictionsFinalStage2\$ID)</pre> <-levels(droplevels(PredictionsFinalStage2\$ID))</pre> codes Create empty dataframe RES Class secondstage <-data.frame(matrix(ncol = 11, nrow =)) Loop to determine which thresholds the files achieved for CF species for (i in 1:length(codes)){ # Select row no. code<-codes[i]</pre> # Select results/file step1<-PredictionsFinalStage2[PredictionsFinalStage2\$ID==code,]</pre> # list species / file step1\$predictionsResultsCF <-as.factor(step1\$predictionsResultsCF)</pre> <-c(levels(droplevels(step1\$predictionsResultsCF)))</pre> step1\$predictionsResultsCF <-as.character(step1\$predictionsResultsCF)</pre> # Loop for different species within one recording for (y in 1:length(sps)){ #select a single species sp<-sps[y] <-step1[step1\$predictionsResultsCF==sp,]</pre> stepsps stepsp <-select(stepsps,sp)</pre> maxprob <-max(stepsp)</pre> # Specify confidence thresholds if (maxprob>=0.6) R60<-as.character("done")</pre> if (maxprob<0.6) R60<-as.character("check") if (maxprob>=0.65) R65<-as.character("done")</pre> if (maxprob<0.65) R65<-as.character("check") if (maxprob>=0.7) R70<-as.character("done")</pre> if (maxprob<0.7) R70<-as.character("check") if (maxprob>=0.75) R75<-as.character("done")</pre> if (maxprob<0.75) R75<-as.character("check") if (maxprob>=0.8) R80<-as.character("done")</pre> if (maxprob<0.8) R80<-as.character("check") if (maxprob>=0.85) R85<-as.character("done")</pre> if (maxprob<0.85) R85<-as.character("check") if (maxprob>=0.9) R90<-as.character("done")</pre> if (maxprob<0.9) R90<-as.character("check") if (maxprob>=0.95) R95<-as.character("done") if (maxprob<0.95) R95<-as.character("check") temp <-as.character(c(code, sp, maxprob, R60, R65, R70, R75, R80, R85, R90, R95)) # Bind results to main table RES Class secondstage <-rbind(RES Class secondstage,temp)</pre> RES Class secondstage *>* mutate if(is.factor, as.character) -> RES Class secondstage # Show progress print(c("Loop ", i, "from", length(codes))) Rename columns names(RES Class secondstage) <- c("ID", "S2 Prediction", "S2 Accuracy", "S2 R60", "S2 R65", "S2 R70", "S2 R75", "S2 R80", "S2 R85", "S2 R90", "S2 R95") Re-add call parameter/file info RES Class secondstage <-merge (RES Class secondstage, PredictionsFinalStage2, by="ID") 5. PERFORM CLASSIFICATIONS - STAGE 3 For species which were identified as "FMqCF" (frequency modulated quasi-constant frequency) conduct a second classification stage using the third machine learning model which prioritises call shape Remove Identifying information from FMqCF data <-Stage1_FMqCF[,c("duration","freq_max_amp" , "freq_max", "freq_min",</pre> Data CallValues FMqCF noID "bandwidth", "freq_start", "freq_center", "freq_end", "freq knee", "fc", "freq bw knee fc", "bin max amp", "pc freq max amp", "pc freq max", "pc freq min", "pc knee", "temp bw knee fc", "slope", "kalman slope", "curve_neg", "curve_pos_start", "curve_pos_end", "mid_offset" ,"snr", "hd", "smoothness")] Data CallValues FMqCF noID <-Data CallValues FMqCF noID[complete.cases(Data CallValues FMqCF noID),]</pre> Data CallValues FMqCF noID <-drop.levels(Data CallValues FMqCF noID)</pre> Run predictions Run prediction without confidence values <-predict(model S3 FMqCF, Data CallValues FMqCF noID)</pre> predictionsResultsFMqCF Run predictions with confidences PredictionResultsProb FMqCF <-predict(model S3 FMqCF, Data CallValues FMqCF noID, type = "prob")</pre> PredictionResultsProb FMqCF\$ID <-Stage1 FMqCF\$ID Combine predictions with confidence values PredictionResultsCombined FMqCF <-cbind(PredictionResultsProb FMqCF, predictionsResultsFMqCF)</pre> Combine with file information PredictionsFinalStage3 <-merge(PredictionResultsCombined FMqCF, Stage1 FMqCF, by="ID")</pre> Export stage 3 predictions setwd(Dir user outputs) write.csv(PredictionsFinalStage3, file="Data PredictionsStage3.csv", na = "NA") Run confidence thresholds Creates table to see which files meet the confidence thresholds necessary for file structure later Create vectors for loop PredictionsFinalStage3\$ID <-as.factor(PredictionsFinalStage3\$ID)</pre> <-levels(droplevels(PredictionsFinalStage3\$ID))</pre> Create empty dataframe RES Class thirdstage <-data.frame(matrix(ncol = 11, nrow =)) Loop to determine which thresholds the files achieved for FMCF sonotypes/species for (i in 1:length(codes)){ # Select row no. code<-codes[i]</pre> # Select results/file step1<-PredictionsFinalStage3[PredictionsFinalStage3\$ID==code,]</pre> # list species / file step1\$predictionsResultsFMqCF <-as.factor(step1\$predictionsResultsFMqCF)</pre> <-c(levels(droplevels(step1\$predictionsResultsFMqCF)))</pre> step1\$predictionsResultsFMqCF <-as.character(step1\$predictionsResultsFMqCF)</pre> # Loop for different species within one recording for (y in 1:length(sps)){ #select a single species sp<-sps[y] stepsps <-step1[step1\$predictionsResultsFMqCF==sp,]</pre> <-select(stepsps,sp)</pre> stepsp maxprob <-max(stepsp)</pre> # Specify confidence thresholds if (maxprob>=0.6) R60<-as.character("done")</pre> if (maxprob<0.6) R60<-as.character("check") if (maxprob>=0.65) R65<-as.character("done")</pre> if (maxprob<0.65) R65<-as.character("check") if (maxprob>=0.7) R70<-as.character("done")</pre> if (maxprob<0.7) R70<-as.character("check") if (maxprob>=0.75) R75<-as.character("done")</pre> if (maxprob<0.75) R75<-as.character("check") if (maxprob>=0.8) R80<-as.character("done")</pre> if (maxprob<0.8) R80<-as.character("check") if (maxprob>=0.85) R85<-as.character("done")</pre> if (maxprob<0.85) R85<-as.character("check") if (maxprob>=0.9) R90<-as.character("done")</pre> if (maxprob<0.9) R90<-as.character("check") if (maxprob>=0.95) R95<-as.character("done")</pre> if (maxprob<0.95) R95<-as.character("check") temp <-as.character(c(code, sp, maxprob, R60, R65, R70, R75, R80, R85, R90, R95)) # Bind results to main table RES Class thirdstage <-rbind(RES Class thirdstage, temp)</pre> RES Class thirdstage %>% mutate if(is.factor, as.character) -> RES Class thirdstage # Show progress print(c("Loop ", i, "from", length(codes))) Rename columns names(RES Class thirdstage) <- c("ID", "S3 Prediction", "S3 Accuracy", "S3 R60", "S3 R65", "S3 R70", "S3 R75", "S3 R80", "S3 R85", "S3 R90", "S3 R95") Re-add call parameter/file info RES Class thirdstage <-merge(RES Class thirdstage, PredictionsFinalStage3, by="ID") 6. COMBINE FINAL PREDICTIONS Collate predictions from each classification stage & isolate files for manual verification **Create summaries** Select relevant columns <-Stage1 QCF[, c("ID", "Prediction", "S1 Accuracy", "S1 R60",</pre> ${\tt SummaryQCF}$ "S1_R65", "S1_R70", "S1_R75", "S1_R80", "S1_R85", "S1 R90", "S1 R95", "Filename", "FileLoc")] <-Stage1 FM[, c("ID", "Prediction", "S1 Accuracy", "S1 R60",</pre> ${\tt SummaryFM}$ "S1 R65", "S1 R70", "S1 R75", "S1 R80", "S1 R85", "S1 R90", "S1 R95", "Filename", "FileLoc")] <-RES Class secondstage[, c("ID", "S2 Prediction", "S2 Accuracy",</pre> SummaryCF "S2_R60", "S2_R65", "S2_R70", "S2_R75", "S2_R80", "S2 R85", "S2 R90", "S2 R95", "Filename", "FileLoc")] ${\tt SummaryFMqCF}$ <-RES Class thirdstage[, c("ID", "S3 Prediction", "S3 Accuracy",</pre> "S3 R60", "S3 R65", "S3 R70", "S3 R75", "S3 R80", "S3 R85", "S3 R90", "S3 R95", "Filename", "FileLoc")] Rename columns to match

```
colnames(SummaryQCF)
                            <-c("ID", "Prediction", "Accuracy", "R60", "R65", "R70", "R75", "R80",</pre>
                                 "R85", "R90", "R95", "Filename", "FileLoc")
                            <-c("ID", "Prediction", "Accuracy", "R60", "R65", "R70", "R75", "R80",
colnames(SummaryFM)
                                "R85", "R90", "R95", "Filename", "FileLoc")
                            <-c("ID", "Prediction", "Accuracy", "R60", "R65", "R70", "R75", "R80",
colnames(SummaryCF)
                                 "R85", "R90", "R95", "Filename", "FileLoc")
                            <-c("ID", "Prediction", "Accuracy", "R60", "R65", "R70", "R75", "R80",
colnames(SummaryFMqCF)
                                 "R85", "R90", "R95", "Filename", "FileLoc")
```

Combine tables

```
RES total raw
                    <-rbind(SummaryQCF, SummaryFM, SummaryCF, SummaryFMqCF)</pre>
```

Subset for files above 60% confidence

```
RES_total_raw
                     <-RES_total_raw[RES_total_raw$R60=="done",]</pre>
```

Export complete predictions

```
setwd(Dir user outputs)
write.csv(RES total raw, file="Data PredictionsSummary.csv", na = "NA")
```

Determines the highest accuracy recorded for each ID for each file to quantify bat activity

Summarise pulses to bat passes

Convert ID to factor for grouping

<-as.factor(RES_total_raw\$ID)</pre>

RES_total_raw\$ID

```
Create vectors for grouping columns
```

<- levels(as.factor(RES total raw\$Prediction))</pre> cols sp

```
cols Files <- c("Filename", "Prediction")</pre>
Find the pulse of highest confidence within each file for each species
```

RES_total_sum <- RES_total_raw %>% group by(across(all of(cols Files))) %>%

names (RES total sum) <-c("filename", "Prediction", "Accuracy")</pre>

```
summarise(MaxbyFile = max(Accuracy, na.rm = T))
Rename columns
```

Readd file location information

```
head (FileLoc)
```

Add to main dataframe

```
RES total sum <-merge(RES total sum, FileLoc, by="filename", all= FALSE)
```

setwd(Dir_user_outputs)

Export summary predictions

```
write.csv(RES_total_sum, file="Data_PredictionsSummary_max.csv", na = "NA")
(ii) Isolate files for manual verification & create library
```

The following series (ii) if for users who use all three classification stages. If you used series (i) the following series will overwrite those files

colnames(Data thres) <- c("Prediction", "Threshold")</pre>

Selects WAV files which do not reach the necessary confidence threshold using their original filepathways and copies them into a new filepathway based on ID prediction & confidence threshold.

!!! THE FOLLOWING CODE WILL MOVE FILES DIRECTORY ON YOUR COMPUTER - ENSURE IT IS WORKING CORRECTLY USING A TEST FILE/BACK UP YOUR DATA BEFORE PROCEEDING !!!

(i) Merge with threshold information

```
<- merge(RES total sum, Data thres, by = "Prediction", keep.all=TRUE)</pre>
RES total sum$ThresLevel <-""
RES total sum$Accuracy <-as.numeric(RES total sum$Accuracy)
for (y in 1:nrow(RES total sum)){
  if((RES total sum$Accuracy[y]*100) == RES total sum$Threshold[y]) {
    RES total sum$ThresLevel[y] <- "Met"</pre>
  else if ((RES total sum$Accuracy[y]*100) > RES total sum$Threshold[y]) {
    RES total sum$ThresLevel[y] <- "Met" }</pre>
  else if ((RES total sum$Accuracy[y]*100) < RES total sum$Threshold[y]) {
    RES total sum$ThresLevel[y] <- "Not Met" }</pre>
```

Lvls stageAll

(ii) Specify ID levels

```
(iii) Remove duplicate levels
```

Filter data for confidence threshold

```
<-filter(RES_total_sum, (ThresLevel=="Not Met"))</pre>
 DF_NotMet
 # Remove repeated files so a file is only manually checked once
 DF_NotMet_unique <- DF_NotMet[!duplicated(DF_NotMet['filename']),]</pre>
(iv) Not reversible: Loop to create new folder pathway and copy WAV files - user needs to update pathway below
```

for (S in 1:length(Lvls_stageAll)) {

<-levels(as.factor(RES_total_sum\$Prediction))</pre>

```
# Specify prediction level
                 <-Lvls stageAll[S]
# Filter data for target species & confidence threshold
RES_total_target <-filter(DF_NotMet_unique, Prediction ==SPECIES)</pre>
# Create filename vector including file locations
Sp_file_list <-as.character(RES_total_target$FileLoc)</pre>
# Remove duplicates (shouldn't remove any values)
Sp_file_list <-Sp_file_list[!duplicated(Sp_file_list)]</pre>
# Create output folder for ID level
setwd(Dir files AutoID WAV)
                  <-paste0(SPECIES,"_", "ThresholdNotMet")</pre>
newdir
dir.create(newdir)
# Create directory in R to Species specific folder
                 <-paste0("F:/Data_wav5sec_IDs_auto/",newdir) # **** NEEDS UPDATING BY USER ****</pre>
Dir_temp
# Go back to input WAV files directory
setwd(Dir_clean_files_WAV)
# Move each individual file to new directory
for (F in 1:length(Sp_file_list)){
  # Select file
 FILE <-Sp_file_list[F]
  # copy files
  file.copy(FILE, Dir_temp)
# Progress bar
print(c("Loop", S, "from", length(Lvls_stageAll)))
```