

Full RCode

#LSM3241 CA1 R Code

#install Bioconductor

```
if (!requireNamespace("BiocManager"))  
  install.packages("BiocManager")  
BiocManager::install()
```

#install Bioconductor packages

```
BiocManager::install('foo')
```

#install relevant packages needed

```
if (!requireNamespace("BiocManager", quietly = TRUE))  
  install.packages("BiocManager")  
BiocManager::install("GEOquery", version = "3.8")
```

```
BiocManager::install("affy")  
BiocManager::install("limma")  
BiocManager::install("hgu133plus2.db")  
BiocManager::install("org.Hs.eg.db")
```

#load packages installed

```
library(affy)  
library(limma)  
library(hgu133plus2.db)  
library(org.Hs.eg.db)
```

#check if packages are loaded

```
sessionInfo()
```

#Calling GSE50697

```
library(GEOquery)  
#downloading the gse file  
gse <- getGEO('GSE50697', GSEMatrix = FALSE)
```

#retrieving whole GSE

```
names(GSMList(gse))
```

```
[1] "GSM1226581" "GSM1226582" "GSM1226583" "GSM1226584" "GSM1226585"  
"GSM1226586"
```

#Getting the raw data for the series using GEOquery

filePaths <- getGEOSuppFiles('GSE50697')

#generating a vector containing the name of all the CEL files.

list.celfiles('GSE50697_RAW')

```
[1] "GSM1226581_S1100958.MDA.01_HG-U133_Plus_2_.CEL.gz"
[2] "GSM1226582_S1100958.MDA.02_HG-U133_Plus_2_.CEL.gz"
[3] "GSM1226583_S1100958.MDA.03_HG-U133_Plus_2_.CEL.gz"
[4] "GSM1226584_S1100958.MDA.04_HG-U133_Plus_2_.CEL.gz"
[5] "GSM1226585_S1100958.MDA.05_HG-U133_Plus_2_.CEL.gz"
[6] "GSM1226586_S1100958.MDA.06_HG-U133_Plus_2_.CEL.gz"
```

#finding info from meta data of gse

names(Meta(gse))

[1] "contact_address"	"contact_city"	"contact_country"
[4] "contact_department"	"contact_email"	"contact_institute"
[7] "contact_laboratory"	"contact_name"	"contact_phone"
[10] "contact_state"	"contact_zip/postal_code"	"contributor"
[13] "email"	"geo_accession"	"institute"
[16] "last_update_date"	"name"	"overall_design"
[19] "platform_id"	"platform_taxid"	"pubmed_id"
[22] "relation"	"sample_id"	"sample_taxid"
[25] "status"	"submission_date"	"summary"
[28] "supplementary_file"	"title"	"type"
[31] "web_link"		

#creating GSM object names

gsm <- GSMList(gse)[[1]]

#find information we need from the metadata of gsm

names(Meta(gsm))

[1] "channel_count"	"characteristics_ch1"	"contact_address"
[4] "contact_city"	"contact_country"	"contact_department"
[7] "contact_email"	"contact_institute"	"contact_laboratory"
[10] "contact_name"	"contact_phone"	"contact_state"
[13] "contact_zip/postal_code"	"data_processing"	"data_row_count"
[16] "extract_protocol_ch1"	"geo_accession"	"hyb_protocol"
[19] "label_ch1"	"label_protocol_ch1"	"last_update_date"
[22] "molecule_ch1"	"organism_ch1"	"platform_id"

[25] "scan_protocol"	"series_id"	"source_name_ch1"
[28] "status"	"submission_date"	"supplementary_file"
[31] "taxid_ch1"	"title"	"type"

#extract metadata of gsm not in gse

names(Meta(gsm))[!(names(Meta(gsm)) %in% names(Meta(gse)))]

[1] "channel_count"	"characteristics_ch1"	"data_processing"
[4] "data_row_count"	"extract_protocol_ch1"	"hyb_protocol"
[7] "label_ch1"	"label_protocol_ch1"	"molecule_ch1"
[10] "organism_ch1"	"scan_protocol"	"series_id"
[13] "source_name_ch1"	"taxid_ch1"	

Meta(gsm)[!(names(Meta(gsm)) %in% names(Meta(gse)))]

\$channel_count

[1] "1"

\$characteristics_ch1

[1] "cell line: SUM159" "treatment: control"

[3] "tissue: Claudin-low breast cancer"

\$data_processing

[1] "MAS 5.0"

\$data_row_count

[1] "54675"

\$extract_protocol_ch1

[1] "standard Affymetrix protocol"

\$hyb_protocol

[1] "standard Affymetrix protocol"

\$label_ch1

[1] "biotin"

\$label_protocol_ch1

[1] "standard Affymetrix protocol"

\$molecule_ch1

[1] "total RNA"

```
$organism_ch1  
[1] "Homo sapiens"
```

```
$scan_protocol  
[1] "standard Affymetrix protocol"
```

```
$series_id  
[1] "GSE50697"
```

```
$source_name_ch1  
[1] "SUM159"
```

```
$taxid_ch1  
[1] "9606"
```

#From the metadata information, we decided to use the following elements

#source_name_ch1 and characteristics_ch1

#getting sample growth conditions into a data frame

```
culture_medium <- function(gsm) {  
  Meta(gsm)[['characteristics_ch1']][2]  
}  
sapply(GSMList(gse),culture_medium)
```

GSM1226581	GSM1226582
"treatment: control"	"treatment: control"
GSM1226583	GSM1226584
"treatment: control"	"treatment: pBabe puro miR-203"
GSM1226585	GSM1226586
"treatment: pBabe puro miR-203"	"treatment: pBabe puro miR-203"

```
pd <- data.frame(culture=as.factor(sapply(GSMList(gse),culture_medium)))  
pd
```

	culture
GSM1226581	treatment: control
GSM1226582	treatment: control
GSM1226583	treatment: control
GSM1226584	treatment: pBabe puro miR-203
GSM1226585	treatment: pBabe puro miR-203
GSM1226586	treatment: pBabe puro miR-203

#simplifying our dataframe, convert columns to 2 values only

```
pd$culture <- as.factor(pd$culture)  
levels(pd$culture) <- c("control", "miR203")
```

#to enable the kable function, we need to install the knitr package

```
install.packages("knitr")
```

```
library(knitr)
```

```
kable(pd)
```

```
|      |culture |  
|:-----|:-----|  
|GSM1226581 |control |  
|GSM1226582 |control |  
|GSM1226583 |control |  
|GSM1226584 |miR203 |  
|GSM1226585 |miR203 |  
|GSM1226586 |miR203 |
```

#Reading in the CEL files with the phenoData

```
celfiles <- paste0('GSE50697_RAW/', list.celfiles('GSE50697_RAW/'),'.')
```

```
affydata <- read.affybatch(celfiles,phenoData = new("AnnotatedDataFrame",pd))
```

```
phenoData(affydata)
```

An object of class 'AnnotatedDataFrame'

sampleNames: GSM1226581 GSM1226582 ... GSM1226586 (6 total)

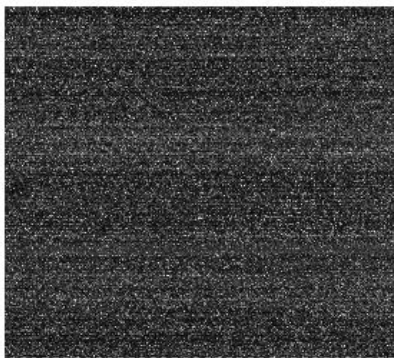
varLabels: culture

varMetadata: labelDescription

#pseudo images of chips

```
image(data[,1])
```

1226581_S1100958.MDA.01_HG-U133_Plus



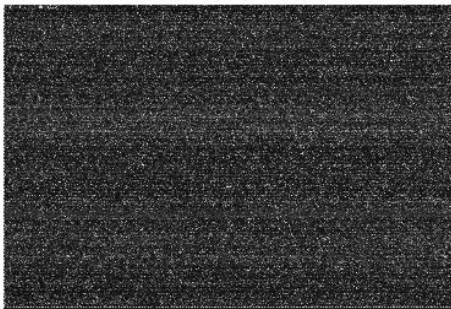
`image(data[,2])`

`226582_S1100958.MDA.02_HG-U133_PI`



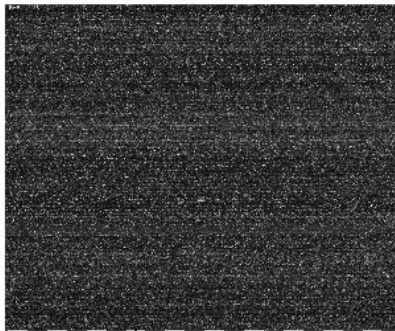
`image(data[,3])`

`GSM1226583_S1100958.MDA.03_HG-U133_Plus_2_(`



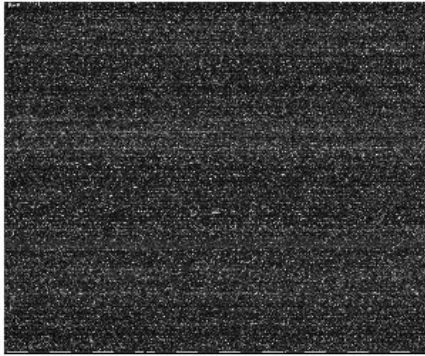
`image(data[,4])`

`M1226584_S1100958.MDA.04_HG-U133_Plus_`



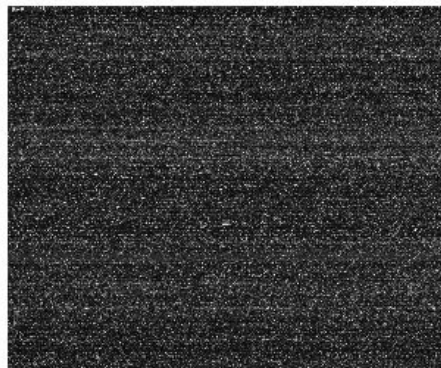
```
image(data[,5])
```

```
M1226585_S1100958.MDA.05_HG-U133_Plus_
```

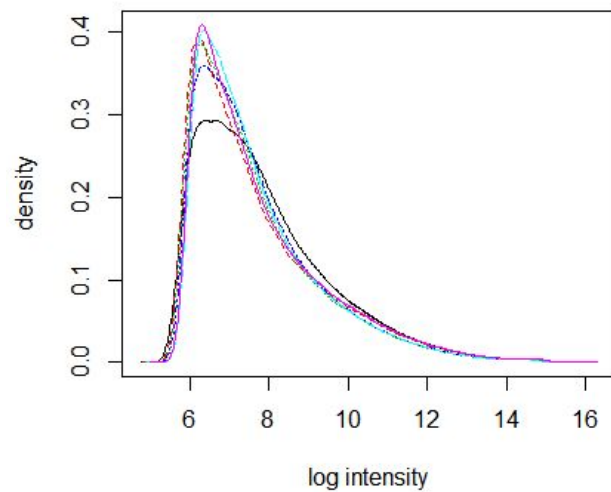


```
image(data[,6])
```

```
M1226586_S1100958.MDA.06_HG-U133_Plus_
```



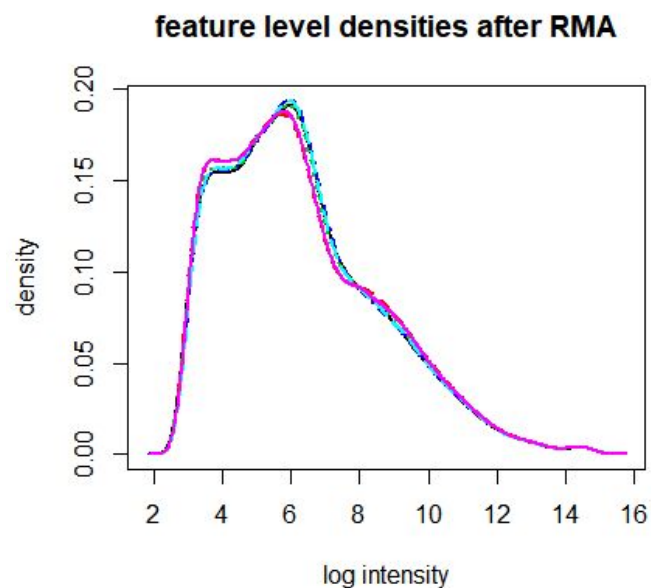
```
#CEL file densities before normalisation or background correction  
plotDensity.AffyBatch(affydata)
```



#perform RMA normalisation

```
eset <- rma(affydata)
```

```
plotDensity(exprs(eset),xlab='log intensity',main="feature level densities after  
RMA",lwd=2)
```



#phenotype data for each samples after rma is retained

```
pData(eset)
```

```
      culture
GSM1226581 control
GSM1226582 control
```


GSM1226583 control
GSM1226584 miR203
GSM1226585 miR203
GSM1226586 miR203

#generate model matrix

model <- model.matrix(~ 0 + eset\$culture)

#rename the model columns to correspond to the different growth conditions

colnames(model) <- levels(eset\$culture)

model

```
      control miR203
1         1     0
2         1     0
3         1     0
4         0     1
5         0     1
6         0     1
attr("assign")
[1] 1 1
attr("contrasts")
attr("contrasts")$`eset$culture`
[1] "contr.treatment"
```

#look at when the growth conditions differ, we create contrast

contrasts <- makeContrasts(control - miR203, levels=model)

contrasts

```
      Contrasts
Levels control - miR203
control          1
miR203          -1
```

#fit model and contrast matrix into a data

fit <- lmFit(eset, model)

fit

An object of class "MArrayLM"

\$coefficients

	control	miR203
1007_s_at	9.245338	9.408269
1053_at	9.586208	9.605038
117_at	6.404494	6.500503
121_at	8.496605	8.577745
1255_g_at	3.425654	3.360337

54670 more rows ...

\$rank

[1] 2

\$assign

[1] 1 1

\$qr

\$qr

	control	miR203
1	-1.7320508	0.0000000
2	0.5773503	-1.7320508
3	0.5773503	0.0000000
4	0.0000000	0.5773503
5	0.0000000	0.5773503
6	0.0000000	0.5773503

attr("assign")
[1] 1 1
attr("contrasts")
attr("contrasts")\$`eset\$culture`
[1] "contr.treatment"

\$qraux

[1] 1.57735 1.00000

\$pivot

[1] 1 2

\$tol

[1] 1e-07

\$rank

[1] 2

\$df.residual

[1] 4 4 4 4 4

54670 more elements ...

\$sigma

1007_s_at 1053_at 117_at 121_at 1255_g_at
0.07378465 0.13302616 0.18946888 0.14112115 0.06639880
54670 more elements ...

\$cov.coefficients

	control	miR203
control	0.3333333	0.0000000
miR203	0.0000000	0.3333333

\$stdev.unscaled

	control	miR203
1007_s_at	0.5773503	0.5773503
1053_at	0.5773503	0.5773503
117_at	0.5773503	0.5773503
121_at	0.5773503	0.5773503
1255_g_at	0.5773503	0.5773503

54670 more rows ...

\$pivot

[1] 1 2

\$Amean

1007_s_at 1053_at 117_at 121_at 1255_g_at
9.326804 9.595623 6.452499 8.537175 3.392996
54670 more elements ...

\$method

[1] "ls"

\$design

	control	miR203
1	1	0
2	1	0
3	1	0

```

4    0    1
5    0    1
6    0    1
attr(,"assign")
[1] 1 1
attr(,"contrasts")
attr(,"contrasts")$`eset$culture`
[1] "contr.treatment"

```

```

fitted.contrast <- contrasts.fit(fit,contrasts)
fitted.contrast

```

An object of class "MArrayLM"

```

$coefficients
      Contrasts
      control - miR203
1007_s_at    -0.16293107
1053_at      -0.01883039
117_at       -0.09600894
121_at       -0.08113934
1255_g_at    0.06531688
54670 more rows ...

```

```

$rank
[1] 2

```

```

$assign
[1] 1 1

```

```

$qr
$qr
      control      miR203
1 -1.7320508  0.0000000
2  0.5773503 -1.7320508
3  0.5773503  0.0000000
4  0.0000000  0.5773503
5  0.0000000  0.5773503
6  0.0000000  0.5773503
attr(,"assign")
[1] 1 1
attr(,"contrasts")
attr(,"contrasts")$`eset$culture`
[1] "contr.treatment"

```

\$qraux
[1] 1.57735 1.00000

\$pivot
[1] 1 2

\$tol
[1] 1e-07

\$rank
[1] 2

\$df.residual
[1] 4 4 4 4 4
54670 more elements ...

\$sigma
1007_s_at 1053_at 117_at 121_at 1255_g_at
0.07378465 0.13302616 0.18946888 0.14112115 0.06639880
54670 more elements ...

\$cov.coefficients
Contrasts
Contrasts control - miR203
control - miR203 0.6666667

\$stdev.unscaled
Contrasts
control - miR203
1007_s_at 0.8164966
1053_at 0.8164966
117_at 0.8164966
121_at 0.8164966
1255_g_at 0.8164966
54670 more rows ...

\$Amean
1007_s_at 1053_at 117_at 121_at 1255_g_at
9.326804 9.595623 6.452499 8.537175 3.392996
54670 more elements ...

```
$method
```

```
[1] "ls"
```

```
$design
```

	control	miR203
1	1	0
2	1	0
3	1	0
4	0	1
5	0	1
6	0	1

```
attr("assign")
```

```
[1] 1 1
```

```
attr("contrasts")
```

```
attr("contrasts")$`reset$culture`
```

```
[1] "contr.treatment"
```

```
$contrasts
```

	Contrasts
Levels	control - miR203
control	1
miR203	-1

#calculate test statistics via performing eBayes correction

```
fitted.ebayes <- eBayes(fitted.contrast)
```

```
fitted.ebayes
```

An object of class "MArrayLM"

```
$coefficients
```

	Contrasts
	control - miR203
1007_s_at	-0.16293107
1053_at	-0.01883039
117_at	-0.09600894
121_at	-0.08113934
1255_g_at	0.06531688
54670 more rows ...	

```
$rank
```

```
[1] 2
```

\$assign

[1] 1 1

\$qr

\$qr

	control	miR203
1	-1.7320508	0.0000000
2	0.5773503	-1.7320508
3	0.5773503	0.0000000
4	0.0000000	0.5773503
5	0.0000000	0.5773503
6	0.0000000	0.5773503

attr("assign")

[1] 1 1

attr("contrasts")

attr("contrasts")\$`eset\$culture`

[1] "contr.treatment"

\$qraux

[1] 1.57735 1.00000

\$pivot

[1] 1 2

\$tol

[1] 1e-07

\$rank

[1] 2

\$df.residual

[1] 4 4 4 4 4

54670 more elements ...

\$sigma

1007_s_at	1053_at	117_at	121_at	1255_g_at
0.07378465	0.13302616	0.18946888	0.14112115	0.06639880

54670 more elements ...

\$cov.coefficients

```

Contrasts
Contrasts      control - miR203
control - miR203      0.6666667

```

```
$stdev.unscaled
```

```

Contrasts
      control - miR203
1007_s_at      0.8164966
1053_at      0.8164966
117_at      0.8164966
121_at      0.8164966
1255_g_at      0.8164966
54670 more rows ...

```

```
$Amean
```

```

1007_s_at 1053_at 117_at 121_at 1255_g_at
9.326804 9.595623 6.452499 8.537175 3.392996
54670 more elements ...

```

```
$method
```

```
[1] "ls"
```

```
$design
```

```
control miR203
```

```

1  1  0
2  1  0
3  1  0
4  0  1
5  0  1
6  0  1

```

```
attr("assign")
```

```
[1] 1 1
```

```
attr("contrasts")
```

```
attr("contrasts")$`eset$culture`
```

```
[1] "contr.treatment"
```

```
$contrasts
```

```

Contrasts
Levels      control - miR203
control      1
miR203      -1

```


\$df.prior
[1] 5.163163

\$s2.prior
[1] 0.01315895

\$var.prior
[1] 51.55685

\$proportion
[1] 0.01

\$s2.post
1007_s_at 1053_at 117_at 121_at 1255_g_at
0.009791214 0.015139491 0.023085437 0.016108249 0.009339242
54670 more elements ...

\$t
Contrasts
control - miR203
1007_s_at -2.0166534
1053_at -0.1874344
117_at -0.7739061
121_at -0.7829841
1255_g_at 0.8277806
54670 more rows ...

\$df.total
[1] 9.163163 9.163163 9.163163 9.163163 9.163163
54670 more elements ...

\$p.value
Contrasts
control - miR203
1007_s_at 0.07396289
1053_at 0.85540486
117_at 0.45848725
121_at 0.45339651
1255_g_at 0.42882619
54670 more rows ...

\$lods
Contrasts

```

                control - miR203
1007_s_at      -4.937866
1053_at        -6.756422
117_at         -6.458114
121_at         -6.450863
1255_g_at      -6.414005
54670 more rows ...

```

```

$F
[1] 4.06689075 0.03513165 0.59893067 0.61306414 0.68522080
54670 more elements ...

```

```

$F.p.value
[1] 0.07396289 0.85540486 0.45848725 0.45339651 0.42882619
54670 more elements ...

```

#extracting differentially expressed genes
topTable(fitted.ebayes)

```

                logFC    AveExpr    t        P.Value
209719_x_at -2.096174  9.668246 -27.98136 3.449900e-10
210413_x_at -1.984099  9.649669 -25.77291 7.265063e-10
201721_s_at  2.254152  9.710288  25.72106 7.398668e-10
211906_s_at -2.143763  9.071134 -24.70101 1.066850e-09
206172_at   -1.789371  7.706033 -24.07788 1.343931e-09
209720_s_at -2.432950  8.729486 -23.12911 1.931967e-09
211756_at   -3.067195  4.671276 -21.91533 3.140852e-09
202949_s_at -1.502988 10.092484 -20.84188 4.935440e-09
202007_at    3.494914  5.646599  20.62198 5.429179e-09
206002_at    1.704752  8.907492  20.45564 5.839018e-09
                adj.P.Val    B
209719_x_at 1.348407e-05 12.13796
210413_x_at 1.348407e-05 11.73165
201721_s_at 1.348407e-05 11.72129
211906_s_at 1.458251e-05 11.50898
206172_at   1.469588e-05 11.37091
209720_s_at 1.760505e-05 11.14745
211756_at   2.453230e-05 10.83610
202949_s_at 3.192483e-05 10.53422
202007_at   3.192483e-05 10.46905
206002_at   3.192483e-05 10.41895

```

#Get a limited number of probesets

```
ps <- rownames(topTable(fitted.ebayes))
ps
```

```
[1] "209719_x_at" "210413_x_at" "201721_s_at" "211906_s_at"
[5] "206172_at"   "209720_s_at" "211756_at"   "202949_s_at"
[9] "202007_at"   "206002_at"
```

#The AnnotationDbi interface

#look at available columns for our chip

```
columns(hgu133plus2.db)
```

```
[1] "ACCNUM"      "ALIAS"       "ENSEMBL"     "ENSEMBLPROT"
[5] "ENSEMBLTRANS" "ENTREZID"    "ENZYME"      "EVIDENCE"
[9] "EVIDENCEALL"  "GENENAME"    "GO"          "GOALL"
[13] "IPI"         "MAP"         "OMIM"        "ONTOLOGY"
[17] "ONTOLOGYALL"  "PATH"        "PFAM"        "PMID"
[21] "PROBEID"     "PROSITE"     "REFSEQ"      "SYMBOL"
[25] "UCSCKG"      "UNIGENE"     "UNIPROT"
```

#Look at which that can be used as keys

```
keytypes(hgu133plus2.db)
```

```
[1] "ACCNUM"      "ALIAS"       "ENSEMBL"     "ENSEMBLPROT"
[5] "ENSEMBLTRANS" "ENTREZID"    "ENZYME"      "EVIDENCE"
[9] "EVIDENCEALL"  "GENENAME"    "GO"          "GOALL"
[13] "IPI"         "MAP"         "OMIM"        "ONTOLOGY"
[17] "ONTOLOGYALL"  "PATH"        "PFAM"        "PMID"
[21] "PROBEID"     "PROSITE"     "REFSEQ"      "SYMBOL"
[25] "UCSCKG"      "UNIGENE"     "UNIPROT"
```

#realise all can be used as keys, choose "PROBEID" as most suitable

```
head(keys(hgu133plus2.db,keytype="PROBEID"))
```

```
[1] "1007_s_at" "1053_at"  "117_at"   "121_at"   "1255_g_at"
[6] "1294_at"
```

```
AnnotationDbi::select(hgu133plus2.db,ps,c("SYMBOL","ENTREZID","GENENAME"),keyty
pe="PROBEID")
```

	PROBEID	SYMBOL	ENTREZID
1	209719_x_at	SERPINB3	6317
2	210413_x_at	SERPINB4	6318
3	210413_x_at	SERPINB3	6317

4	201721_s_at	LAPTM5	7805
5	211906_s_at	SERPINB4	6318
6	206172_at	IL13RA2	3598
7	209720_s_at	SERPINB3	6317
8	211756_at	PTHLH	5744
9	202949_s_at	FHL2	2274
10	202007_at	NID1	4811
11	206002_at	ADGRG2	10149

GENENAME

1	serpin family B member 3
2	serpin family B member 4
3	serpin family B member 3
4	lysosomal protein transmembrane 5
5	serpin family B member 4
6	interleukin 13 receptor subunit alpha 2
7	serpin family B member 3
8	parathyroid hormone like hormone
9	four and a half LIM domains 2
10	nidogen 1
11	adhesion G protein-coupled receptor G2

#Restrict to upregulated genes

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differentially_expressed <- topTable(fitted.ebayes,number=Inf,p.value=0.1,lfc=1)
differentially_expressed
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210413_x_at	-1.984099	9.649669	-25.772906	7.265063e-10
201721_s_at	2.254152	9.710288	25.721059	7.398668e-10
211906_s_at	-2.143763	9.071134	-24.701007	1.066850e-09
206172_at	-1.789371	7.706033	-24.077878	1.343931e-09
209720_s_at	-2.432950	8.729486	-23.129114	1.931967e-09
211756_at	-3.067195	4.671276	-21.915333	3.140852e-09
202949_s_at	-1.502988	10.092484	-20.841881	4.935440e-09
202007_at	3.494914	5.646599	20.621979	5.429179e-09
206002_at	1.704752	8.907492	20.455638	5.839018e-09
228946_at	2.850568	7.433498	19.353438	9.598769e-09
203296_s_at	1.436208	7.128925	18.624289	1.353915e-08
200953_s_at	2.114548	6.413324	18.518165	1.424938e-08
201681_s_at	1.343568	10.848433	18.048727	1.792645e-08
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214051_at	1.266167	6.486634	16.511752	3.963213e-08

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227609_at -1.245758 7.672937 -14.484025 1.264433e-07
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adj.P.Val B

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209839_at 3.586247e-03 3.0775943
207173_x_at 3.826645e-03 2.9324569
211959_at 3.998774e-03 2.8595595
222662_at 4.290230e-03 2.7236000
1569157_s_at 4.404616e-03 2.6756776
224367_at 4.907541e-03 2.4765306

```

205830_at 5.000643e-03 2.4525107
212706_at 5.101094e-03 2.4149637
205535_s_at 5.580465e-03 2.2430129
210355_at 5.586227e-03 2.2378631
205532_s_at 5.808908e-03 2.1780336
1558501_at 5.837822e-03 2.1496409
201147_s_at 6.099969e-03 2.0657510
1555673_at 6.197900e-03 2.0430339
235561_at 6.275685e-03 2.0162314
202920_at 6.724926e-03 1.8978896
243009_at 6.913504e-03 1.8528277
220030_at 8.990789e-03 1.3947082
206300_s_at 9.250357e-03 1.3480368
205533_s_at 9.416580e-03 1.3192648
220301_at 1.041079e-02 1.1586786
1554131_at 1.420825e-02 0.6515165
[ reached 'max' /getOption("max.print") -- omitted 5 rows ]

```

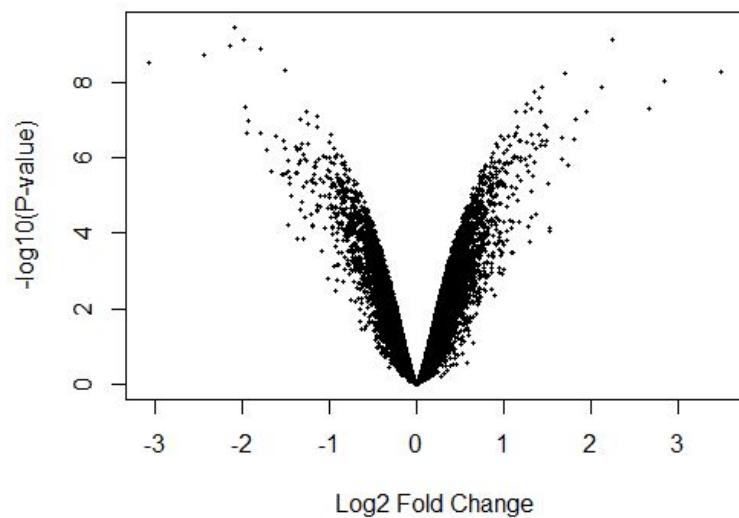
```

upregulated <- differentially_expressed[differentially_expressed$logFC > 0,]
genes_of_interest <- AnnotationDbi::select(hgu133plus2.db,
                                           keys=rownames(upregulated),
                                           columns=c("SYMBOL", "ENTREZID", "GENENAME"),
                                           keytype="PROBEID")

genes_of_interest
#save data into a csv file
write.csv(genes_of_interest, 'genes_of_interest.csv')

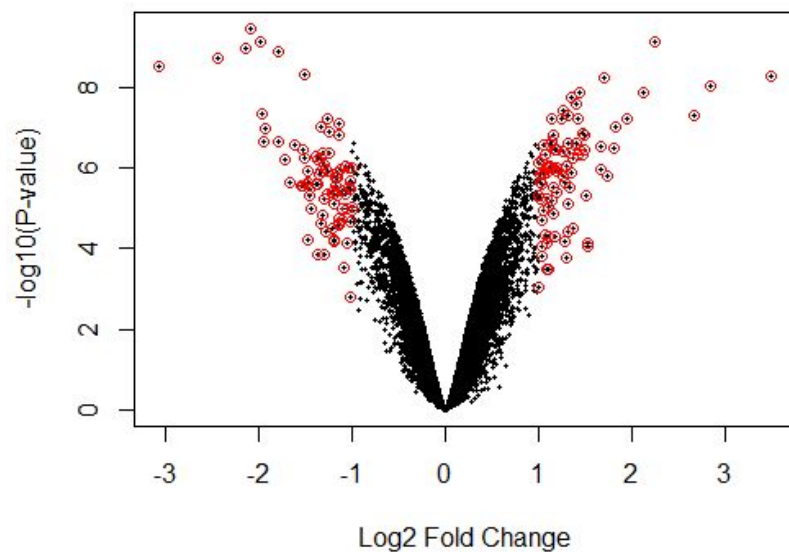
#volcanoplots
volcanoplot(fitted.ebayes)

```



```
interesting_genes <- topTable(fitted.ebayes,number=Inf,p.value = 0.1,lfc=1)
volcanoplot(fitted.ebayes, main=sprintf("%d features pass our
cutoffs",nrow(interesting_genes)))
```

171 features pass our cutoffs



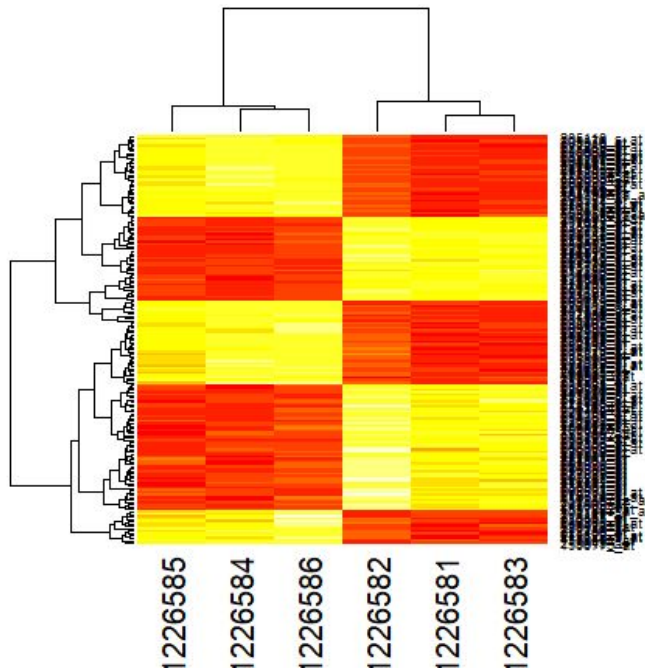
```
points(interesting_genes[['logFC']], -log10(interesting_genes[['P.Value']]), col='red')
```

#heatmaps

#normalise expression value

```
eset_of_interest <- eset[rownames(interesting_genes),]
```

```
heatmap(exprs(eset_of_interest))
```



#fix and beautify heatmap

```
install.packages("RColorBrewer")
```

```
library(RColorBrewer)
```

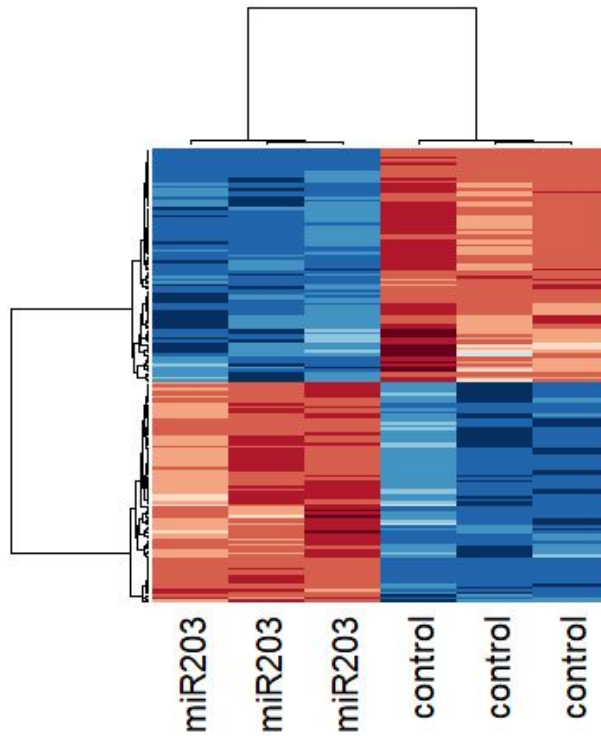
```
eset_of_interest <- eset[rownames(interesting_genes),]
```

```
heatmap(exprs(eset_of_interest),
```

```
  labCol=eset$culture,labRow=NA,
```

```
  col    = rev(brewer.pal(10, "RdBu")),
```

```
  distfun = function(x) as.dist(1-cor(t(x))))
```



#Results

```
recommendations <- AnnotationDbi::select(hgu133plus2.db,
                                         keys=rownames(interesting_genes),
                                         columns=c("SYMBOL","ENTREZID","GENENAME"),
                                         keytype="PROBEID")
write.csv(recommendations,'recommendations.csv')
#shown in appendix below
```