**Università degli Studi di Padova – Department of Mathematics**

**Master’s Degree in Data Science**

**Structural Bioinformatics, A.Y. 2022/23**

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Classification of Contacts in Protein Structures

**1.** Training Set and Data Retrieval

The first step of the project was retrieving the data from each .tsv file from the features\_ring folder (1,807 PDBs in total) and storing it into a single DataFrame.

The data contained in each .tsv file consists in a tab-separated file available for each protein, in which each row represents a contact in the protein and each column represents a feature about that contact. The last column is the target label, specifying the interaction type.

|  |  |  |  |
| --- | --- | --- | --- |
| **Column position** | **Column name** | **Column meaning** | **Type of column** |
| 1 | **pdb\_id** |  |  |
| 2 | **s\_ch** | chain | source residue identifier |
| 3 | **s\_resi** | index |
| 4 | **s\_ins** | insertion code |
| 5 | **s\_resn** | name |
| 6 | **s\_ss8** | secondary structure 8 states (DSSP) | source residue features |
| 7 | **s\_rsa** | relative solvent accessibility |
| 8 | **s\_up** | half sphere exposure up |
| 9 | **s\_down** | half sphere exposure down |
| 10 | **s\_phi** | phi angle |
| 11 | **s\_psi** | psi angle |
| 12 | **s\_ss3** | secondary structure 3 states (from angles) |
| 13 | **s\_a1** | Atchley feature 1 |
| 14 | **s\_a2** | Atchley feature 2 |
| 15 | **s\_a3** | Atchley feature 3 |
| 16 | **s\_a4** | Atchley feature 4 |
| 17 | **s\_a5** | Atchley feature 5 |
| 18 | **t\_ch** | chain | target residue identifier |
| 19 | **t\_resi** | index |
| 20 | **t\_ins** | insertion code |
| 21 | **t\_resn** | name |
| 22 | **t\_ss8** | secondary structure 8 states (DSSP) | target residue features |
| 23 | **t\_rsa** | relative solvent accessibility |
| 24 | **t\_up** | half sphere exposure up |
| 25 | **t\_down** | half sphere exposure down |
| 26 | **t\_phi** | phi angle |
| 27 | **t\_psi** | psi angle |
| 28 | **t\_ss3** | secondary structure 3 states (from angles) |
| 29 | **t\_a1** | Atchley feature 1 |
| 30 | **t\_a2** | Atchley feature 2 |
| 31 | **t\_a3** | Atchley feature 3 |
| 32 | **t\_a4** | Atchley feature 4 |
| 33 | **t\_a5** | Atchley feature 5 |
| 34 | **Interaction** | interaction type |  |

Table 1.1: Training set description

|  |  |
| --- | --- |
| **Interaction Type** | **Count** |
| HBOND | 333,346 |
| VDW | 155,789 |
| PIPISTACK | 10,403 |
| IONIC | 9,068 |
| SSBOND | 866 |
| PICATION | 626 |
| *Unclassified* | 225,412 |

Table 1.2: Number of examples by bond type.

2. Data Preprocessing

The preprocessing pipeline starts with removing all samples where the label is unavailable (to explain how we are reinputting them).  Then, the missing values of each feature are replaced using the mean of the feature itself. (Numerical features?)

It was considered to perform best subset selection using Logistic Regression to determine which features are the most meaningful ones and the ones that are not influencing the decision much. However, all Atchley features are selected, so this step is skipped.

Scaling is then performed to standardize all the features to values between [0,1] to be then fed to the model.

The biggest criticality in the dataset is the heavy imbalance that is evident by looking at the number of contacts by interaction type. Notably, Hydrogen Bonds (HBOND) and Van der Waals contacts (VDW) are the most numerous, which makes them overrepresented in the training set. The remaining contact types, instead, are underrepresented. Training a model with such unbalanced datasets is sure to yield poor performance, especially when evaluating the model on new unseen data.

To mitigate this issue, oversampling the underrepresented classes is applied to encourage the model to better learn the minority interactions. It is to be noted, however, that altering the dataset, especially by oversampling, can yield overly optimistic results in the training performance, which don’t necessarily transfer to as good performances at inference time on new data. Therefore, the balancing process of the training set is performed carefully and conservatively.

Use of undersampeling lead to greatly diminished performance distinguishing the majority classes (hydrogen bonding and VDW interactions). For oversampling, SMOTE (Synthetic Minority Oversampling TEchnique) is used, which uses interpolation between samples to create new artificial data points. The sampling strategy is {IONIC: 20’000, PIPISTACK: 10’000, PICATION: 20’000, SSBOND: 10’000}

3. Model

3.1. Deep Neural Network

The model created to classify residue-residue contacts is a Deep Neural Network for multiclass classification (if ensemble approach goes through, this becomes binary classification using OvO). The Deep Learning library of choice is *Keras*, a very commonly used open-source library that acts as an interface for *TensorFlow*.

The first step to be carried out is the encoding of the label of each sample into an identity vector, using a common practice called “*One-Hot Encoding*”. This ensures that each class is uniquely identified and independent of the others. It helps the neural network to better understand the categorical nature of the data and prevents any ordinal relationship assumptions between the classes.

*One-Hot Encoding* is also performed to then set the number of neurons of the output layer to be equal to the number of classes in the dataset, in order to train each output neuron to determine the probability P(*Ci*| data), with ∑ P(*Ci* | data) = 1. Therefore, the output layer represents the probability distribution of a contact being of a certain type. (again, if ensemble is used, this has to be revised, as well as the output csv)

Although the model is a relatively small Deep Neural Network with fully connected layers, overfitting is mitigated using L2 Regularization (Weight Decay) on each Dense layer, and Early Stopping monitoring the loss progression over the epochs. These two techniques provide better generalization performance. The use of random dropout of neurons after each hidden layer was also tested, without any meaningful improvement on final test performance.

After much experimentation with a single model, it was decided to use Ensemble Methods, implementing a *One vs. One* (*OvO*) approach, which assumes that grouping together multiple weaker models can yield better performance by leveraging the strengths of each predictor. Therefore, multiple models are trained to discriminate between pair of classes. The final prediction of the ensemble is decided through majority voting.

3.2. Model Details and Hyperparameters

|  |  |
| --- | --- |
| **Hyperparameters** | |
| Batch Size | 16’000 |
| Weight Initialization | Xavier (GlorotNormal) |
| Loss Function | Categorical Cross-Entropy |
| Optimizer | Adam |
| Hidden Layers: Activation Function | ReLU |
| Output Layer: Activation Function | Softmax |
|  |  |

Table 3.2.1: Model’s hyperparameters (Keras)

|  |  |  |
| --- | --- | --- |
| **Layer Type** | **Output Shape** | **Param #** |
| Dense | 128 |  |
| Dense | 128 |  |
| Dense | 128 |  |
| Dense | 128 |  |
| Dense | 128 |  |
| # Total Parameters |  | 174,918 |

Table 3.2.2: Model’s architecture (Keras)

4. Results

4.1 Performance

Testing data consisted of 51,608 examples, or 10% of unbalanced data. As demonstrated by the accompanying metrics, the model preforms better that the provided naive Bayes classifier achieving a total accuracy of 71%, ROC AUC of 0.79, and a Matthews Correlation Coefficient of 0.38.

A graph of a training and validation loss

Description automatically generated A graph with red and blue lines

Description automatically generated

Test performance by Interaction type

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | precision | recall | f1-score | support |
| HBOND | 0.77 | 0.85 | 0.81 | 33656 |
| IONIC | 0.46 | 0.26 | 0.33 | 916 |
| PICATION | 0.05 | 0.60 | 0.09 | 63 |
| PIPISTACK | 0.39 | 0.99 | 0.56 | 1050 |
| SSBOND | 0.43 | 1.00 | 0.60 | 88 |
| VDW | 0.61 | 0.41 | 0.49 | 15735 |
| macro avg | 0.45 | 0.68 | 0.48 | 51508 |
| weighted avg | 0.71 | 0.70 | 0.70 |

Insert here:

* Loss plot over epochs (both training and validation)
* AUC curve (both training and validation)
* Precision-Recall Barplots (both training and validation)
* Average statistics at test time? Actual metrics evaluated by professor?

4.2 Issues

In attempt to address poor precision on minority classes, many alternative approaches were tested. Including ensemble methods like one-vs-one, one-vs-all, and model stacking; different architectures; regularizers; and balancing. However, it was found that ordinary deep neural nets had the best compromise between minority class and majority class precision. Models that showed improvements in one of these areas consistently suffered in the other.

Implementing dataset balancing greatly improved model recall on minority classes. This would indicate that the model is effectively learning information about the classes. However, these classes still suffer from poor precision with resampling likely attributed to the relatively extremely low unique examples not allowing for a good representation of the class to be learned. The model additionally showed great sensitivity to undersampling, quickly leading to diminished ability to distinguish hydrogen bond and VDW interactions, the largest influence on total accuracy. The final model relied primarily on having the minority classes oversampled. However, oversampling additionally quickly lead to poor generalization on minority classes due to the extremely low number of unique examples making learning a good representation of the classes difficult.

5. Usage

To predict the contacts of a new PDB file using the pretrained model, run the following command in your terminal, using arguments:

python3 contact\_net.py --inference --pdb your\_pdb\_id

To perform retraining or the model, instead, run:

python3 contact\_net.py --train

The full documentation of the software is available in the GitHub repository at the following [link](https://github.com/TannerAGraves/StructBioInfo/blob/main/docs/documentation.md).