



IL-6 in Inflammation, Immunity, and Disease

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Interleukin 6 (IL-6), promptly and transiently produced in response to infections and tissue injuries, contributes to host defense through the stimulation of acute phase responses, hematopoiesis, and immune reactions. Although its expression is strictly controlled by transcriptional and posttranscriptional mechanisms, dysregulated continual synthesis of IL-6 plays a pathological effect on chronic inflammation and autoimmunity. For this reason, tocilizumab, a humanized anti-IL-6 receptor antibody was developed. Various clinical trials have since shown the exceptional efficacy of tocilizumab, which resulted in its approval for the treatment of rheumatoid arthritis and juvenile idiopathic arthritis. Moreover, tocilizumab is expected to be effective for other intractable immune-mediated diseases. In this context, the mechanism for the continual synthesis of IL-6 needs to be elucidated to facilitate the development of more specific therapeutic approaches and analysis of the pathogenesis of specific diseases.

IL-6 is a soluble mediator with a pleiotropic effect on inflammation, immune response, and hematopoiesis. At first, distinct functions of IL-6 were studied and given distinct names based on their biological activity. For example, the name B-cell stimulatory factor 2 (BSF-2) was based on the ability to induce differentiation of activated B cells into antibody (Ab)-producing cells (Kishimoto 1985), the name hepatocyte-stimulating factor (HSF) on the effect of acute phase protein synthesis on hepatocytes,

the name hybridoma growth factor (HGF) on the enhancement of growth of fusion cells between plasma cells and myeloma cells, or the name interferon (IFN)- β 2 owing to its IFN antiviral activity. When the BSF-2 cDNA was successfully cloned in 1986 (Hirano et al. 1986), however, it was found that the molecules with different names studied by various groups were in fact identical, resulting in the single name IL-6 (Kishimoto 1989). Human IL-6 is made up of 212 amino acids, including a 28-amino-acid

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signal peptide, and its gene has been mapped to chromosome 7p21. Although the core protein is ~20 kDa, glycosylation accounts for the size of 21–26 kDa of natural IL-6.

BIOLOGICAL EFFECT OF IL-6 ON INFLAMMATION AND IMMUNITY

After IL-6 is synthesized in a local lesion in the initial stage of inflammation, it moves to the liver through the bloodstream, followed by the rapid induction of an extensive range of acute phase proteins such as C-reactive protein (CRP), serum amyloid A (SAA), fibrinogen, haptoglobin, and α 1-antichymotrypsin (Fig. 1) (Heinrich et al. 1990). On the other hand, IL-6 reduces the production of fibronectin, albumin, and transferrin. These biological effects on hepatocytes were at first studied as belonging to HSE. When high-level concentrations of SAA persist for a long time, it leads to a serious complication of several chronic inflammatory

diseases through the generation of amyloid A amyloidosis (Gillmore et al. 2001). This results in amyloid fibril deposition, which causes progressive deterioration in various organs. IL-6 is also involved in the regulation of serum iron and zinc levels via control of their transporters. As for serum iron, IL-6 induces hepcidin production, which blocks the action of iron transporter ferroportin 1 on gut and, thus, reduces serum iron levels (Nemeth et al. 2004). This means that the IL-6-hepcidin axis is responsible for hypoferrremia and anemia associated with chronic inflammation. IL-6 also enhances zinc importer ZIP14 expression on hepatocytes and so induces hypozincemia seen in inflammation (Liuzzi et al. 2005). When IL-6 reaches the bone marrow, it promotes megakaryocyte maturation, thus leading to the release of platelets (Ishibashi et al. 1989). These changes in acute phase protein levels and red blood cell and platelet counts are used for the evaluation of inflammatory severity in routine clinical laboratory examinations.

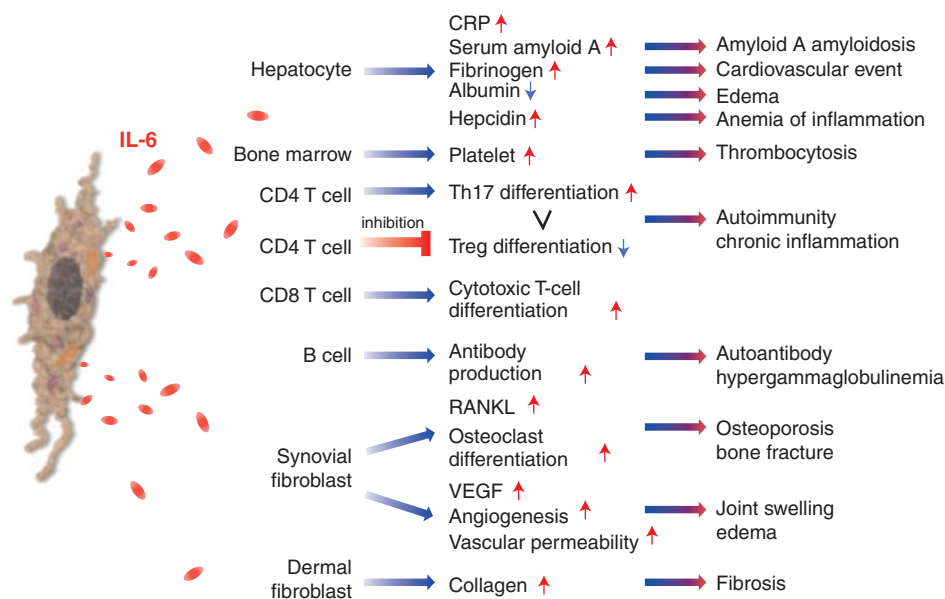


Figure 1. IL-6 in inflammation, immunity, and disease. IL-6 is a cytokine featuring pleiotropic activity; it induces synthesis of acute phase proteins such as CRP, serum amyloid A, fibrinogen, and hepcidin in hepatocytes, whereas it inhibits production of albumin. IL-6 also plays an important role on acquired immune response by stimulation of antibody production and of effector T-cell development. Moreover, IL-6 can promote differentiation or proliferation of several nonimmune cells. Because of the pleiotropic activity, dysregulated continual production of IL-6 leads to the onset or development of various diseases. Treg, regulatory T cell; RANKL, receptor activator of nuclear factor κ B (NF- κ B) ligand; VEGF, vascular endothelial growth factor.

Furthermore, IL-6 promotes specific differentiation of naïve CD4⁺ T cells, thus performing an important function in the linking of innate to acquired immune response. It has been shown that IL-6, in combination with transforming growth factor (TGF)- β , is indispensable for Th17 differentiation from naïve CD4⁺ T cells (Korn et al. 2009), but that IL-6 also inhibits TGF- β -induced Treg differentiation (Bettelli et al. 2006). Up-regulation of the Th17/Treg balance is considered to be responsible for the disruption of immunological tolerance, and is thus pathologically involved in the development of autoimmune and chronic inflammatory diseases (Kimura and Kishimoto 2010). It has been further shown that IL-6 also promotes T-follicular helper-cell differentiation as well as production of IL-21 (Ma et al. 2012), which regulates immunoglobulin (Ig) synthesis and IgG4 production in particular. IL-6 also induces the differentiation of CD8⁺ T cells into cytotoxic T cells (Okada et al. 1988). Under one of its previous names, BSF-2, IL-6 was found to be able to induce the differentiation of activated B cells into Ab-producing plasma cells, so that continuous oversynthesis of IL-6 results in hypergammaglobulinemia and autoantibody production.

IL-6 exerts various effects other than those on hepatocytes and lymphocytes and these are frequently detected in chronic inflammatory diseases (Kishimoto 1989; Hirano et al. 1990; Akira et al. 1993). One of these effects is that, when IL-6 is generated in bone marrow stromal cells, it stimulates the RANKL (Hashizume et al. 2008), which is indispensable for the differentiation and activation of osteoclasts (Kotake et al. 1996), and this leads to bone resorption and osteoporosis (Poli et al. 1994). IL-6 also induces excess production of VEGF, leading to enhanced angiogenesis and increased vascular permeability, which are pathological features of inflammatory lesions and are seen in, for example, synovial tissues of rheumatoid arthritis (RA) or edema of remitting seronegative symmetrical synovitis with pitting edema (RS3PE) syndrome (Nakahara et al. 2003; Hashizume et al. 2009). Finally, it has been reported that IL-6 aids keratinocyte proliferation (Grossman et al. 1989) or the generation of collagen in dermal

fibroblasts that may account for changes in the skin of patients with systemic sclerosis (Duncan and Berman 1991).

REGULATION OF IL-6 SYNTHESIS

IL-6 functions as a mediator for notification of the occurrence of some emergent event. IL-6 is generated in an infectious lesion and sends out a warning signal to the entire body. The signature of exogenous pathogens, known as pathogen-associated molecular patterns, is recognized in the infected lesion by pathogen-recognition receptors (PRRs) of immune cells such as monocytes and macrophages (Kumar et al. 2011). These PRRs comprise Toll-like receptors (TLRs), retinoic acid-inducible gene-1-like receptors, nucleotide-binding oligomerization domain-like receptors, and DNA receptors. They stimulate a range of signaling pathways including NF- κ B, and enhance the transcription of the mRNA of inflammatory cytokines such as IL-6, tumor necrosis factor (TNF)- α , and IL-1 β . TNF- α and IL-1 β also activate transcription factors to produce IL-6.

IL-6 also issues a warning signal in the event of tissue damage. Damage-associated molecular patterns (DAMPs), which are released from damaged or dying cells in noninfectious inflammations such as burn or trauma, directly or indirectly promote inflammation. During sterile surgical operations, an increase in serum IL-6 levels precedes elevation of body temperature and serum acute phase protein concentration (Nishimoto et al. 1989). DAMPs from injured cells contain a variety of molecules such as mitochondrial (mt) DNA, high mobility group box 1 (HMGB1), and S100 proteins (Bianchi 2007). Serum mtDNA levels in trauma patients are thousands of times higher than in controls and this elevation leads to TLR9 stimulation and NF- κ B activation (Zhang et al. 2010), whereas binding of HMGB1 to TLR2, TLR4, and the receptor of advanced glycation end products (RAGE) can promote inflammation. The S100 family of proteins comprises more than 25 members, some of which also interact with RAGE to evoke sterile inflammation (Sims et al. 2010).



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In addition to immune-mediated cells, mesenchymal cells, endothelial cells, fibroblasts, and many other cells are involved in the production of IL-6 in response to various stimuli (Akira et al. 1993). The fact that IL-6 issues a warning signal to indicate occurrence of an emergency accounts for the strict regulation of IL-6 synthesis both gene transcriptionally and posttranscriptionally. A number of transcription factors have been shown to regulate the IL-6 gene transcription (Fig. 2). The functional *cis*-regulatory elements in the human IL-6 gene 5' flanking region are found binding sites for NF- κ B, specificity protein 1 (SP1), nuclear factor IL-6 (NF-IL-6) (also known as CAAT/enhancer-binding protein β), activator protein 1 (AP-1),

and interferon regulatory factor 1 (Libermann and Baltimore 1990; Akira and Kishimoto 1992; Matsusaka et al. 1993). Activation of *cis*-regulatory elements by stimulation with IL-1, TNE, TLR-mediated signal, and forskolin lead to activation of the IL-6 promoter.

A polymorphism at position -174 of the IL-6 promoter region is reportedly associated with systemic onset juvenile idiopathic arthritis (Fishman et al. 1998) and susceptibility to RA in Europeans (Lee et al. 2012). Stimulation with lipopolysaccharide (LPS) and IL-1 did not evoke any response in a reporter assay using -174 C construct. A -174 G construct, on the other hand, was found to promote transcription of the reporter gene, suggesting that a genetic back-

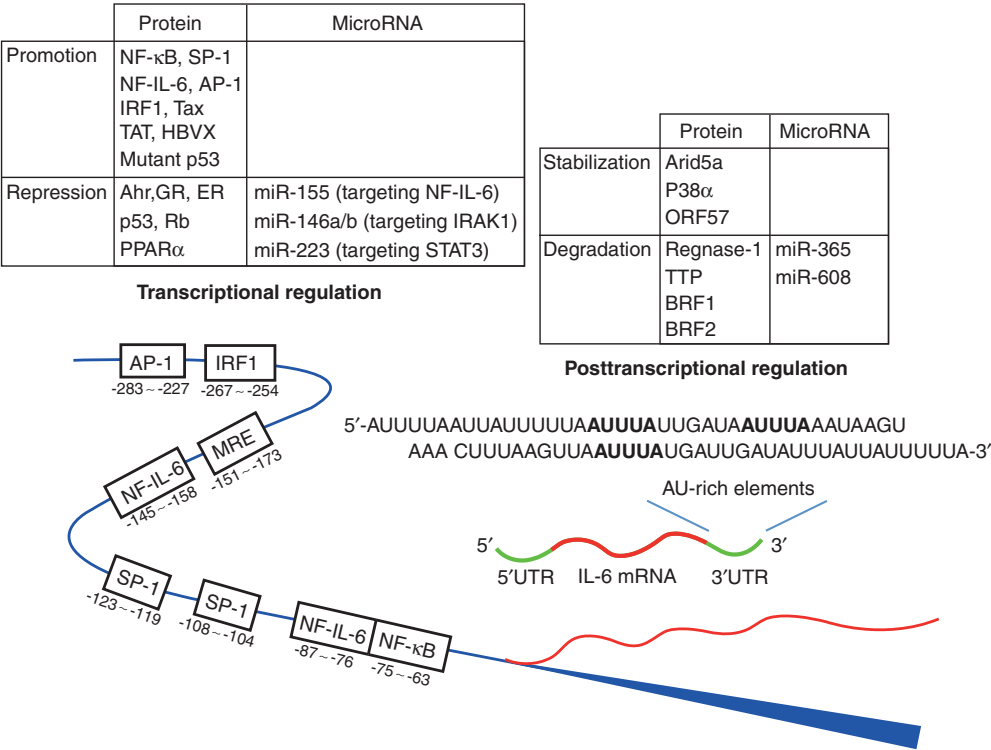


Figure 2. Transcriptional and posttranscriptional regulation of IL-6 gene. The expression and degradation of IL-6 mRNA is regulated transcriptionally and posttranscriptionally by several proteins and microRNAs. Activation of these proteins and microRNAs determines the fate of IL-6 mRNA. NF-IL-6, nuclear factor of IL-6; Tax, transactivator protein; TAT, transactivator of the transcription; HBVX, hepatitis B virus X protein; Ahr, aryl hydrocarbon receptor; GR, glucocorticoid receptor; ER, estrogen receptor; Rb, retinoblastoma; PPAR α , peroxisome proliferator-activated receptor α ; miR, microRNA; IRAK1, IL-1 receptor-associated kinase 1; STAT3, signal transducer and activator of transcription 3; ORE, open reading frame; TTP, tristetraprolin; BRF1, butyrate response factor 1.



ground of excess IL-6 production constitutes a risk factor for juvenile idiopathic arthritis and RA.

An interesting finding is that some viral products enhance the DNA-binding activity of NF- κ B and NF-IL-6, resulting in an increase in IL-6 mRNA transcription. An instance of this phenomenon is that interaction with NF- κ B of the Tax derived from the human T lymphotropic virus 1 enhances IL-6 production (Ballard et al. 1988; Leung and Nabel 1988). Another example is the enhancement of both NF- κ B and NF-IL-6 DNA-binding activity by the transactivator of the TAT protein of the human immunodeficiency virus 1 (Scala et al. 1994; Ambrosino et al. 1997). Moreover, it has been shown that DNA binding of NF-IL-6 can be enhanced by the human hepatitis B virus X protein (Mahe et al. 1991; Ohno et al. 1999).

On the other hand, some transcription factors suppress IL-6 expression. Peroxisome proliferator-activated receptors (PPARs) are ligand-activated transcription factors consisting of three subtypes: α , β , and γ . Among three PPARs, fibrates-activated PPAR α interacts with c-Jun and p65 NF- κ B subunits, which negatively regulate IL-6 transcription (Delerive et al. 1999). In addition, some hormone receptors have been identified as repressors of IL-6 expression. The increase in serum IL-6 after menopause or ovariectomy is reportedly associated with suppression of IL-6 expression by estrogen receptors (Jilka et al. 1992), whereas activation of the glucocorticoid receptor can repress IL-6 expression, and this is thought to be one of mechanisms responsible for the anti-inflammatory effects of corticosteroids (Ray and Prefontaine 1994). It has further been shown that retinoblastoma protein and p53 repress the IL-6 gene promoter, whereas it is up-regulated by mutant p53 (Santhanam et al. 1991).

In addition, some microRNAs directly or indirectly regulate transcription activity. Interaction of microRNA-155 with the 3' untranslated regions (UTR) of NF-IL-6 results in suppression of NF-IL-6 expression (He et al. 2009), whereas microRNA-146a/b and -223 indirectly suppress transcription of IL-6 by respectively targeting IL-1 receptor-associated kinase

1 and STAT3 (Chen et al. 2012; Zilahi et al. 2012).

PRODUCTION AND FUNCTION OF IL-6 AND ARL HYDROCARBON RECEPTOR

Aryl hydrocarbon receptor (Ahr) not only affects IL-6 transcription, but also regulates innate and acquired immune response. Ahr, also known as the dioxin receptor, is a ligand-activated transcription factor that belongs to the basic helix-loop-helix PER-ARNT-SIM family (Burbach et al. 1992; Ema et al. 1992). Ahr is present in the cytoplasm, where it forms a complex with Ahr-interacting protein (Bell and Poland 2000). On binding with a ligand, Ahr moves to the nucleus and dimerizes with the Ahr nuclear translocator (Arnt). Within the nucleus, the Ahr/Arnt heterodimer then binds to the xenobiotic response element (XRE), which leads to various toxicological effects (Fujii-Kuriyama et al. 1994; Dragan and Schrenk 2000; Ohtake et al. 2003; Puga et al. 2005). Although the physiological ligands for Ahr are not well known, indoleamine 2,3-dioxygenase (IDO), which catalyzes tryptophan into kynurenine, is induced by Ahr signaling and kynurenine is one of the ligands of Ahr (Vogel et al. 2008; Jux et al. 2009).

An animal model of RA was used to show the essential role of Ahr in the induction of Th17 cells and Th17-dependent collagen-induced arthritis (CIA) (Kimura et al. 2008; Nakahama et al. 2011). Stimulation of naïve T cells with IL-6 plus TGF- β (Th17 cell-inducing condition) induced Ahr expression and deletion of the Ahr gene nullified the induction of Th17 cells.

Ahr interacted with and inhibited the activities of STAT1 or STAT5, which mediate the anti-inflammatory signals of IL-27 and IFN- γ , or IL-2, respectively (Harrington et al. 2005; Stumhofer et al. 2006; Laurence et al. 2007; Kimura et al. 2008), thus suppressing the inhibitory signals for the induction of Th17 cells. Retinoid-related orphan receptors (ROR) γ and α , which are activated by STAT3, are essential transcription factors for Th17-cell induction (Ivanov et al. 2006; Yang et al. 2008) and Ahr was

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found not to affect the ROR- γ and - α expression. In the Ahr gene-deficient mice, no arthritis developed in CIA. Moreover, with T-cell-specific deletion of the Ahr gene, no development of CIA was observed (Nakahama et al. 2011). These results clearly show that CIA is a T-cell-dependent disease and the presence of Ahr is essential for its development. In these Ahr-deficient mice, the number of Th17 cells decreased and that of Th1 cells increased but no significant changes were observed in Foxp3-expressing Treg cells.

Ahr also regulates Th17-cell induction through regulation of microRNAs. In our study, Ahr-induced microRNA-132/212 cluster under Th17 cell-inducing conditions and transfection of microRNA-212 into naïve T cells under these circumstances augmented the expression of IL-17-related genes such as IL-17A, IL-22, and IL-23R (Nakahama et al. 2013). One of the target genes of this microRNA is B-cell lymphoma 6, which is known as an inhibitor of Th17-cell induction (Yu et al. 2009). All of these findings show that Ahr accelerates inflammation through the enhancement of Th17-cell induction by several mechanisms.

Interestingly, Ahr showed a negative regulatory effect on peritoneal macrophages and bone marrow-derived dendritic cells (BMDC) (Nguyen et al. 2010). In the absence of Ahr, LPS-induced production of inflammatory cytokines such as IL-6, TNF, and IL-12 showed major increases in macrophages, indicating that Ahr negatively regulates inflammatory cytokine production. Ahr interacts with STAT1 and NF- κ B and the resultant complex of Ahr/STAT1 and NF- κ B leads to inhibition of the promoter activity of IL-6 and other inflammatory cytokines (Kimura et al. 2009). In BMDC, Ahr is required for the activation of IDO leading to kynurenine production because the deletion of Ahr in BMDC leads to loss of IL-10 and kynurenine production. Coculture of naïve T cells with Ahr-deficient BMDC in the presence of LPS resulted in reduction of Treg-cell induction, whereas addition of kynurenine rescued the induction of Treg cells by BMDC (Nguyen et al. 2010). These findings indicate that Ahr is required for the regulatory BMDC cells through the induction of IDO.

STABILIZATION AND DEGRADATION OF IL-6 MRNA (ARID5A AND REGNASE-1)

As for posttranscriptional regulation of cytokine expression, cytokine mRNA is controlled through both the 5' and 3'UTR (Chen and Shyu 1995; Anderson 2008). Initiation of mRNA translation is determined by the 5'UTR, and the stability of mRNA by the 3'UTR. IL-6 mRNA is regulated by modulation of AU-rich elements located in the 3'UTR region, whereas a number of RNA-binding proteins and microRNAs bind to the 3'UTRs and regulate the stability of IL-6 mRNA (Fig. 2). For example, IL-6 mRNA stabilization is promoted by mitogen-activated protein kinase (MAPK) p38 α via 3'UTRs of IL-6 (Zhao et al. 2008), and the stabilization of both viral and human IL-6 mRNA by the Kaposi's sarcoma-associated herpesvirus (KSHV) ORF-57 by competing with the binding of microRNA-1293 to the viral or of microRNA-608 to the human IL-6 mRNA (Kang et al. 2011). RNA-binding proteins, such as TTP and BRF1 and 2, on the other hand, promote IL-6 mRNA degradation (Palanisamy et al. 2012), whereas IL-6 mRNA levels are reduced by microRNAs such as microRNA-365 and -608 through direct interaction with IL-6 3'UTR (Kang et al. 2011; Xu et al. 2011).

It was recently found that a nuclease known as regulatory RNase-1 (regnase-1) (also known as Zc3h12a) plays a part in the destabilization of IL-6 mRNA, and that the relevant knockout mice spontaneously develop autoimmune diseases accompanied by splenomegaly and lymphadenopathy (Matsushita et al. 2009). The inhibitor of NF- κ B (I κ B) kinase (IKK) complex controls IL-6 mRNA stability by phosphorylating regnase-1 in response to IL-1R/TLR stimulation (Iwasaki et al. 2011). Phosphorylated regnase-1 underwent ubiquitination and degradation. Regnase-1 re-expressed in IL-1R/TLR-activated cells was found to feature delayed kinetics, and regnase-1 mRNA to be negatively regulated by regnase-1 itself via a stem-loop region present in the regnase-1 3'UTR. These findings show that IKK complex phosphorylates not only I κ B α , activating transcription, but also regnase-1, releasing the brake on IL-6 mRNA ex-

pression. Regnase-1 also regulates the mRNAs of a set of genes, including c-Rel, O \times 40, and IL-2 through cleavage of their 3'UTRs in T cells. T-cell receptor engagement then leads to cleavage of regnase-1, which frees T cells from regnase-mediated suppression, thus indicating that regnase-1 may play a crucial role in T-cell activation (Uehata et al. 2013).

We have recently identified a novel RNA-binding protein, AT-rich interactive domain-containing protein 5a (Arid5a), which binds to the 3'UTR of IL-6 mRNA, resulting in the selective stabilization of IL-6 but not of TNF- α or IL-12 mRNA (Masuda et al. 2013). Arid5a expression was found to be enhanced in macrophages in response to LPS, IL-1 β , and IL-6, and also to be induced under Th17-polarizing conditions in T cells. We also found that Arid5a gene deficiency inhibited elevation of IL-6 levels in LPS-injected mice and preferential Th17-cell development in experimental autoimmune encephalomyelitis. Moreover, Arid5a counteracted the destabilizing function of regnase-1 on IL-6 mRNA (Fig. 3), indicating that the balance between Arid5a and regnase-1 plays an important

role in IL-6 mRNA stability. All of these results suggest that posttranscriptional regulation of IL-6 mRNA by Arid5a and regnase-1 may play an important role in the expression of IL-6 and that the predominance of Arid5a over regnase-1 promotes inflammatory processes and possibly induces the development of autoimmune inflammatory diseases.

During the so-called "cytokine storm," a potentially fatal immune reaction induced by hyperactivation of T cells, a major boost in IL-6 production is observed but without comparable production of other inflammatory cytokines. A recent study showed that the cytokine storm induced by cancer immunotherapy using T-cell transfection was counteracted by the anti-IL-6 receptor antibody, tocilizumab (Grupp et al. 2013). Experimentally, inhalation by mice of peroxidized phospholipids induced a cytokine storm resulting from a greatly marked increase in the production of IL-6 but not TNF (Imai et al. 2008).

These results showing the IL-6-specific elevation without any effect on the other inflammatory cytokines strongly suggest the impor-

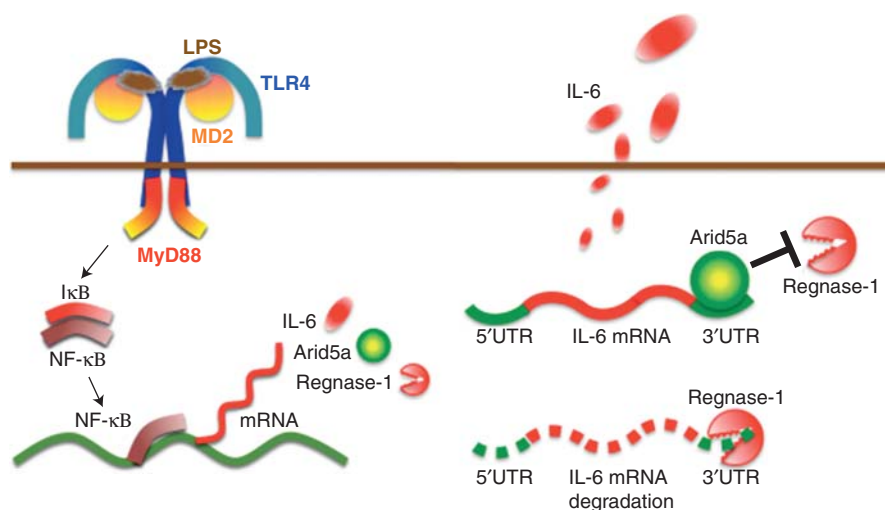


Figure 3. IL-6 synthesis and regulation of IL-6 mRNA stability by Arid5a. Pathogen-associated molecular patterns are recognized by pathogen-recognition receptors to induce proinflammatory cytokines; in this figure, TLR4 recognizes LPS and induces IL-6 mRNA via activation of the NF- κ B signaling pathway. Regnase-1 promotes IL-6 mRNA degradation, whereas Arid5a inhibits destabilizing effects of regnase-1. The balance between Arid5a and regnase-1 is important for the regulation of IL-6 mRNA. MD2, myeloid differentiation protein 2; MyD88, myeloid differentiation primary response 88; I κ B, inhibitor of NF- κ B.

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tance of posttranscriptional modification of IL-6 mRNA by Arid5a and regnase-1. The balance between Arid5a and regnase-1 may determine the pathological increase of IL-6 in various diseases including autoimmunity and even cytokine storm.

IL-6 RECEPTOR-MEDIATED SIGNALING SYSTEM

The IL-6 receptor–signaling system is made up of two receptor chains and downstream signaling molecules (Kishimoto et al. 1992). The IL-6 receptor (IL-6R) constitutes the IL-6-binding chain, which occurs in two forms, 80 kDa transmembrane and 50–55 kDa–soluble IL-6R (sIL-6R), whereas 130 kDa gp130 constitutes the signal-transducing chain. Both of these proteins belong to the cytokine receptor family with a Trp-Ser-X-Trp-Ser motif (Yamasaki et al. 1988; Hibi et al. 1990). sIL-6R without the cytoplasmic region is present in human serum and after IL-6 binding to sIL-6R; the resultant complex induces the IL-6 signal on gp130-expressing cells (Narazaki et al. 1993). The pleiotropic effect of IL-6 on various cells derives from the broad range of gp130 expression observed on cells (Taga and Kishimoto 1997). After binding

of IL-6 to IL-6R, the IL-6/IL-6R complex in turn induces homodimerization of gp130 and triggers a downstream signal cascade (Fig. 4) (Murakami et al. 1993). The activated IL-6 receptor complex is generated in the form of a hexameric structure comprising two molecules each of IL-6, IL-6R, and gp130 (Boulanger et al. 2003). Of these components, IL-6R is a unique binding-receptor for IL-6, whereas the signal-transducing chain gp130 is shared by members of the IL-6 family of cytokines, that is, leukemia inhibitory factor, oncostatin M, ciliary neurotrophic factor, IL-11, cardiotrophin 1, cardiotrophin-like cytokine, IL-27, and IL-35. Although all of these cytokines thus bind to their specific binding receptors, they use the same gp130 for their signals (Kishimoto et al. 1994, 1995). The only exception is virus-encoded IL-6, which is the product of KSHV (also known as human herpesvirus 8), and directly binds to and activates gp130 (Aoki et al. 2001). The mechanism that the IL-6 cytokine family members use to employ the common signal transducer makes it clear why members of the IL-6 family show functional redundancy. The molecular elucidation of the IL-6–signaling system finally solved the long-standing mystery of why cytokines featured pleiotropy and redundancy.

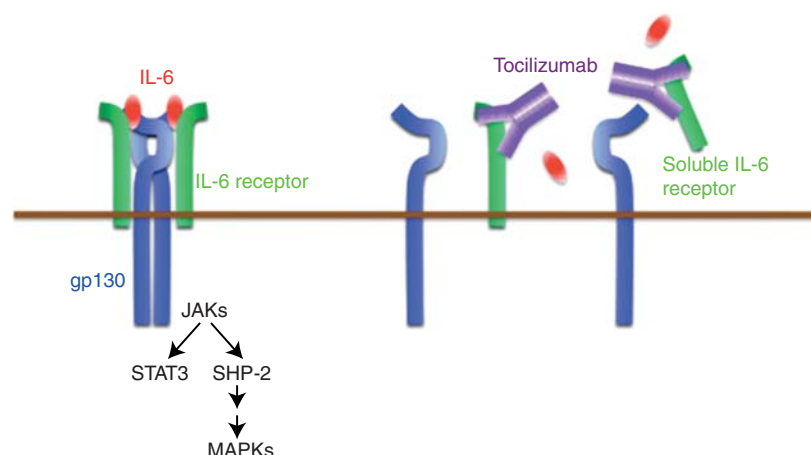


Figure 4. IL-6 receptor system and IL-6 blocker, a humanized anti-IL-6 receptor antibody tocilizumab. IL-6 binds to soluble and transmembrane IL-6R and the complex, then induces homodimerization of gp130, leading to activation of the signaling system. A humanized anti-IL-6R antibody, tocilizumab, blocks IL-6-mediated signaling pathway by its inhibition of IL-6 binding to both receptors. JAKs, Janus kinases; SHP-2, SH2-domain containing protein tyrosine phosphatase-2.

Activation of gp130 in turn triggers activation of downstream signaling molecules, that is, the Janus kinase (JAK)-STAT3 pathway and the JAK-SHP-2-mitogen-activated protein (MAP) kinase pathway. The regulation of various sets of IL-6 responsive genes, including acute phase proteins, is accounted for by the transcription factor STAT3, which also induces the suppressor of cytokine signaling 1 (SOCS1) and SOCS3, which share the SH2-domain. In this context, SOCS1 binds to tyrosine-phosphorylated JAK (Naka et al. 1997), whereas SOCS3 binds to tyrosine-phosphorylated gp130 (Schmitz et al. 2000) to stop IL-6 signaling by means of a negative feedback loop.

IL-6 AND DISEASE

An immediate and transient expression of IL-6 is generated in response to environmental stress factors such as infections and tissue injuries. This expression triggers an alarm signal and activates host defense mechanisms against stress. Removal of the source of stress from the host is followed by cessation of IL-6-mediated activation of the signal-transduction cascade by negative regulatory systems such as ligand-induced internalization and degradation of gp130 and recruitment of SOCS (Naka et al. 1997), as well as degradation of IL-6 mRNA by regnase-1 leading to termination of IL-6 production. However, dysregulated and persistent IL-6 production of mostly unknown etiology, one of which may be the unbalance between Arid5a and regnase-1, in certain cell populations leads to the development of various diseases. This association of IL-6 with disease development was first shown in a case of cardiac myxoma. The culture of fluid obtained from the myxoma tissues of a patient who presented with fever, polyarthritis with positivity for antinuclear factor, elevated CRP level, and hypergammaglobulinemia, contained a large quantity of IL-6, which suggested that IL-6 may contribute to chronic inflammation and autoimmunity (Hirano et al. 1987). Subsequent studies have shown that dysregulation of IL-6 production occurs in the synovial cells of RA (Hirano et al. 1988), swollen lymph nodes of Castleman's disease

(Yoshizaki et al. 1989), myeloma cells (Kawano et al. 1988), and peripheral blood cells or involved tissues in various other autoimmune and chronic inflammatory diseases and even malignant cells in cancers (Nishimoto et al. 1989, 2005).

Moreover, the pathological role of IL-6 in disease development has been shown in numerous animal models of diseases as well as the fact that IL-6 blockade by means of gene knockout or administration of anti-IL-6 or anti-IL-6R Ab can result in the preventive or therapeutic suppression of disease development. For example, IL-6 blockade resulted in a noticeable reduction in susceptibility to Castleman's disease-like symptoms in IL-6 transgenic mice (Katsume et al. 2002). Similar effects were observed in models of RA (Alonzi et al. 1998; Ohshima et al. 1998; Fujimoto et al. 2008), systemic lupus erythematosus (Mihara et al. 1998), systemic sclerosis (Kitaba et al. 2012), inflammatory myopathies (Okiyama et al. 2009), experimental autoimmune uveoretinitis (Haruta et al. 2011), experimental autoimmune encephalomyelitis (Serada et al. 2008), and many other diseases.

IL-6 TARGETING AS STRATEGY FOR TREATMENT OF IMMUNE-MEDIATED DISEASES

In view of the range of biological activities of IL-6 and its pathological role in various diseases described above, it was anticipated that IL-6 targeting would constitute a novel treatment strategy for various immune-mediated diseases. The development of tocilizumab was a direct result of this hypothesis. Tocilizumab is a humanized anti-IL-6R monoclonal Ab of the IgG1 class that was generated by grafting the complementarity determining regions of a mouse antihuman IL-6R Ab onto human IgG1 (Sato et al. 1993), and it blocks IL-6-mediated signal transduction by inhibiting IL-6 binding to transmembrane and soluble IL-6R (Fig. 4). The outstanding efficacy, tolerability, and safety of tocilizumab were verified in numerous worldwide clinical trials initiated in the late 1990s. This has resulted in the approval of this biologic for the treatment of RA in more than 100 countries (Tanaka et al. 2013),



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as well as for systemic and polyarticular juvenile idiopathic arthritis (Yokota et al. 2008, 2012; De Benedetti et al. 2012) and Castleman's disease (Nishimoto et al. 2005) in several countries. Although other biologics including TNF inhibitors, T-cell stimulator blocker, B-cell depletory, or IL-1R antagonist are currently used for RA, tocilizumab has proved its superior efficacy as monotherapy for moderate-to-severe active RA (Tanaka and Kishimoto 2011; Emery et al. 2013) and is recommended as a first-line biologic. Moreover, the outstanding efficacy of tocilizumab for systemic juvenile idiopathic arthritis has led to the recognition of the start of a new era in the treatment of this disease, which was long considered to be one of the most intractable juvenile diseases (Sandborg and Mellins 2012).

Furthermore, there are strong indications based on favorable results detailed in numerous recent case reports, series, and pilot studies of the off-label application of tocilizumab that it can be used for the treatment of various immune-

mediated diseases (Tanaka and Kishimoto 2012; Tanaka et al. 2012). These comprise autoimmune, chronic inflammatory, autoinflammatory, and other diseases. The first group includes systemic sclerosis, inflammatory myopathies, large vessel vasculitis, systemic lupus erythematosus, relapsing polychondritis, autoimmune hemolytic anemia, acquired hemophilia A, neuromyelitis optica, and Cogan's syndrome. The second group includes adult-onset Still's disease, amyloid A amyloidosis, polymyalgia rheumatica, RS3PE, Behcet's disease, uveitis, Crohn's disease, graft-versus-host disease, pulmonary arterial hypertension, and IgG4-related diseases. The third group comprises such autoinflammatory diseases as TNF-receptor-associated periodic syndrome and chronic inflammatory neurological cutaneous articular syndrome, whereas other diseases include atherosclerosis, type 2 diabetes mellitus, atopic dermatitis, scitica, and cancer-associated cachexia. Clinical trials are in progress to identify additional indications for tocilizumab (Table 1).

Table 1. Ongoing clinical trials of tocilizumab

Targeted diseases	Status	Identifier
Diabetes mellitus (type 2), obesity	Phase 2	NCT01073826
Graves' ophthalmopathy	Phase 3	NCT01297699
Cardiovascular disease in RA (vs. etanercept)	Phase 4	NCT01331837
Polymyalgia rheumatica	Phase 2	NCT01396317
	Phase 2	NCT01713842
Giant-cell arteritis	Phase 2	NCT01450137
	Phase 3	NCT01791153
Steroid refractory acute GVHD	Phase 1/2	NCT01475162
After HSCT	Phase 2	NCT01757197
Non-ST elevation myocardial infarction	Phase 2	NCT01491074
Noninfectious uveitis	Phase 1/2	NCT01717170
Systemic sclerosis	Phase 2/3	NCT01532869
Transplant rates awaiting kidney transplantation	Phase 1/2	NCT01594424
JIA-associated uveitis	Phase 1/2	NCT01603355
Recurrent ovarian cancer	Phase 1/2	NCT01637532
Behcet's syndrome	Phase 2	NCT01693653
Schizophrenia	Phase 1	NCT01696929
Erdheim–Chester disease	Phase 2	NCT01727206
Primary Sjogren's syndrome	Phase 2/3	NCT01782235
Fibrous dysplasia of bone	Phase 2	NCT01791842

Registered with ClinicalTrials.gov for diseases other than rheumatoid arthritis, juvenile idiopathic arthritis, and Castleman's disease.

GVHD, graft-versus-host disease; HSCT, hematopoietic stem cell transplant; JIA, juvenile idiopathic arthritis.

CONCLUDING REMARKS AND FUTURE PROSPECTS

The first report of the existence of soluble factors for the enhancement of IgG and IgE Ab responses was published by Kishimoto and Ishizaka in 1973, and it took 13 years until the actual cloning of the IL-6 gene (Hirano et al. 1986). After this success, the fundamental research progressed rapidly and the whole picture of the IL-6 signaling system was completed in the early 1990s (Kishimoto et al. 1994). In parallel with this development, the pathological involvement of IL-6 in various diseases was also established (Nishimoto et al. 1989, 2005; Yoshizaki et al. 1989). It was found that if free serum concentration of tocilizumab is maintained at more than

1 $\mu\text{g/ml}$, CRP remains negative (Nishimoto et al. 2008), indicating that IL-6 plays a major role in the induction of CRP expression and that IL-6 may be involved in the development of almost all chronic inflammatory diseases with CRP elevation. Clinical trials of tocilizumab started in the late 1990s and this humanized monoclonal Ab was approved for the treatment of Castleman's disease in 2005 in Japan, nearly 20 years after the successful molecular cloning of the IL-6 gene (Fig. 5) (Kishimoto 2005). During the following years, tocilizumab has become a first-line biologic for the treatment of moderate-to-severe active RA and the only approved first-line biologic for systemic juvenile idiopathic arthritis. It is anticipated that during the next decade this IL-6 blocker will be widely used for

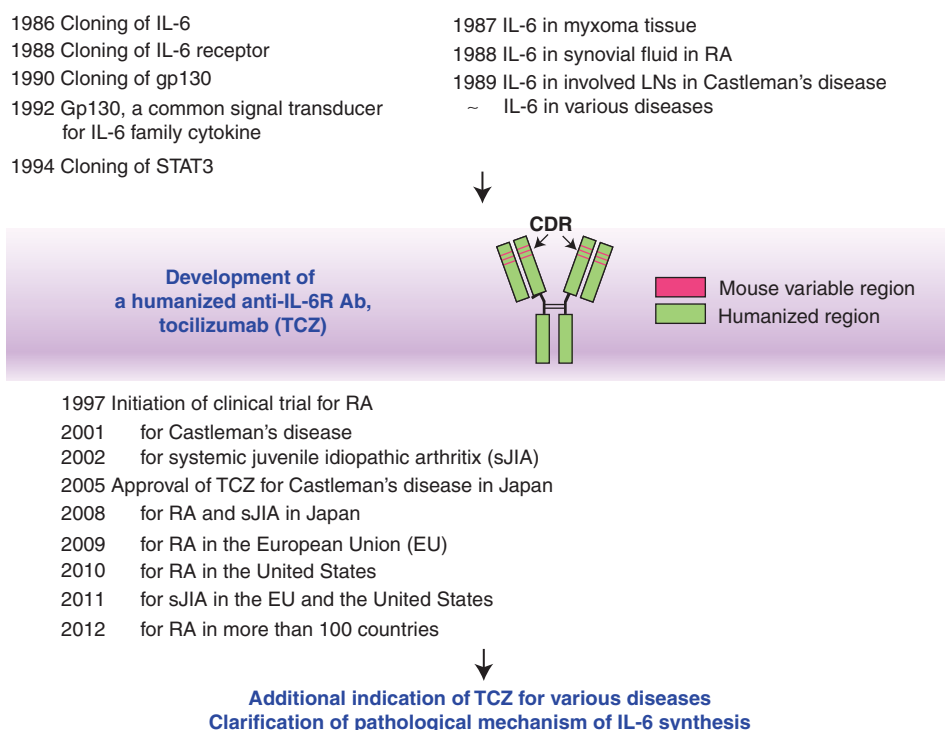


Figure 5. Major discoveries in IL-6-related research and establishment of IL-6 targeting strategy for immune-mediated diseases. Basic research regarding IL-6 clarified the molecular basis of the characteristics of cytokine, redundancy, and pleiotropy, whereas clinical research revealed its pathological significance in disease development. These findings led to concept that IL-6 targeting would constitute a novel therapeutic strategy for immune-mediated diseases and indeed, tocilizumab, a humanized anti-IL-6R antibody became an innovative biologic for the treatment of intractable diseases such as RA, systemic juvenile idiopathic arthritis (sJIA), and Castleman's disease. It will be expected that this strategy would be widely applicable for other immune-mediated diseases. LNs, lymph nodes; TCZ, tocilizumab; CDR, complementarity determining region.



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the treatment of various as-yet intractable diseases including cytokine storm and its application will overcome the refractoriness of such diseases.

To achieve this goal, however, there are several hurdles to overcome. First, additional clinical trials will be needed to evaluate the efficacy and safety of tocilizumab for various diseases. The second, and also important, hurdle is to clarify the mechanisms that render tocilizumab efficacious for phenotypically different diseases. In the case of amyloid A amyloidosis, the ameliorative effect of tocilizumab can be explained by its potent suppression of serum amyloid A protein synthesis (Tanaka et al. 2011). In the case of RA, it has been shown that tocilizumab treatment led to improvement in systemic and joint inflammatory markers (Garnero et al. 2010; Terpos et al. 2011; Kanbe et al. 2012), but it remained to be determined whether it can rectify fundamental immunological abnormalities, such as autoantibody production or imbalance of effector CD4⁺ T-cell subsets (Tanaka 2013). However, recent preliminary results indicate that tocilizumab treatment can correct the imbalance between Th17 and Treg in peripheral blood CD4⁺ T cells (Samson et al. 2012; Pesce et al. 2013). Moreover, it was shown that the treatment caused a reduction in the level of serum IgG4-class anticyclic citrullinated peptide Ab in RA (Carbone et al. 2013). Anti-aquaporin 4 (AQP4) Ab plays a pathological role in neuromyelitis optica, and tocilizumab treatment was found to improve clinical symptoms and reduce serum anti-AQP4 Ab titers, perhaps by inhibiting cell survival of the plasmablasts secreting this Ab (Chihara et al. 2011; Araki et al. 2013). If IL-6 blockade can actually correct these immunological abnormalities, it will in fact be possible to use tocilizumab for the treatment of a wide variety of immune-mediated diseases.

Finally, the mystery remains why IL-6 is persistently expressed in distinct cell populations in various diseases. Accurate and detailed analyses of proteins such as Arid5a and regnase-1 and of microRNAs that regulate IL-6 synthesis will be helpful for solving this mystery, whereas clarification of the mechanism(s) involved will facilitate the identification of more specific target

molecules and investigations into the pathogenesis of specific diseases.

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