# vaRHC

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#### 1. Introduction

Variant classification is a manual complex long process that combines information of distinct nature. An accurate classification is necessary to ensure a proper genetic counselling and personalized risk estimation.

In 2015, the American College of Molecular Genetics and Genomics (ACMG) together with the Association of Molecular Pathologists published generic guidelines to standardize and provide an objective framework to evaluate variant pathogenicity in Mendelian disease. Later, specific guidelines have been published for some genes by collaborative groups.

Moreover, it has been demonstrated that the combination of criteria in ACMG/AMP guidelines is compatible with a quantitative Bayesian formulation (Tavtigian et al., 2018) and a naturally scaled point system has been further abstracted (Tavtigian et al., 2020). Additionally, CanVIG-UK consensus recommendations has proposed some limitations to overlapping criterion combination to avoid double counting of evidence (Garrett et al., 2021).

In the last 5 years, some programs have been developed with the aim to semi-automatize the process of variant classification. Most tools are based on ACMG-AMP general rules but others focus on a set of genes.

vaRHC has been developed to automate as much as possible the process of variant classification in hereditary cancer (HC). The aim is to streamline the work of biologists and avoid possible manual error following gene-specific guidelines for ATM, CDH1, CHEK2, MLH1, MSH2, MSH6, PMS2, PTEN and TP53 and the updated general ACMG rules for the remaining cancer susceptibility genes. The final classification is obtained according to Tatigian's natural scoring Bayesian-based metastructure (Tavtigian et al., 2020) but also considering the CanVIG-UK incompatibilities proposal. vaRHC gives the oportunity to export the output in a .xlsx file as a user-friendly way to examine and store the results allowing non-bioinformatic users to work with them, and even modify the file adding their considerations or information regarding the non-automatable criteria.

The current version of the package is based on the GRCh37 assembly of the human genome and works for single substitutions, deletions and insertions up to 25 bp, intronic variants and 5' or 3'-UTR variants 25 bp beyond the coding sequence.

### 2. Installation

vaRHC can be downloaded from GitHub using the remotes package:

```
if(!require("remotes", quietly = TRUE)) install.packages('remotes') ## Only the first time
library(remotes)
devtools::install_github("emunte/vaRHC")
```

Next, it has to be loaded into the workspace:

```
library("vaRHC")
```

#### 3. Main functions

The package consists of two main functions vaR() and vaRbatch().

3.1 vaR()

## 3.1.1 Input

**3.1.1.1 Parameters** Using a **gene** and a **variant** name (in coding DNA nomenclature) as input variables, vaR() gathers relevant information from different sources. Gene-specific ACMG/AMP guidelines are then applied in order to calculate whether the variant of interest meets different criteria. The output also provides an explanation of the reason for applying or rejecting each criterion. Additionally, it returns a final classification of the variant using Tavtigian Bayesian metastructure and also considering most of CanVIG-UK recommendations.

The following example shows how to call the vaR() function for the variant c.1137+1G>A in CDH1 gene.

```
eg.gene <- "CDH1"
eg.variant <- "c.1137+1G>A"

var.information <- vaR(gene = eg.gene, variant = eg.variant)</pre>
```

**3.1.1.2 Optional parameters** Optional parameters of vaR() are described below.

# 3.1.1.2.1 NM and CCDS

- NM: Accession number of the transcrit and mRNA from RefSeq. By default is NULL and vaRHC will consider the ones detailed above. Be careful if you use a different NM because the program has not been validated for it. If you provide a different NM, CCDS must also be provided. The transcript accession number is retrieved from RefSeq. By default, this function parameter is set to NULL. vaRHC will consider the NMs listed in the Supplementary Table 1. If a different NM is used than the ones described in the table, it must be noted that the program has not been validated yet for it. In this case, a CCDS ID is also required.
- CCDS: Consensus CDS ID retrieved from NCBI CCDS database (which can be found in the following link https://www.ncbi.nlm.nih.gov/projects/CCDS/CcdsBrowse.cgi). By default is set to NULL. vaRHC will consider the IDs detailed in the table above. Again, it must be noted that if a different CCDS ID is provided, the program has not been validated yet for it. In this case, NM accession number is required.

**3.1.1.2.2** gene.specific.df gene.specific.df: By default the parameter is set to NULL as it considers the gene-specific cutoffs, frequencies and additional parameters queried in the IDIBELL database as described in the paper vaRHC: an R package for semi-automation of variant classification in hereditary cancer genes according to ACMG/AMP and gene-specific ClinGen guidelines [Manuscript submitted for publication]. This table contains gene-specific information for applying the following criteria: BA1, BS1, BS2, PM2, predictors cut-off and BP7. If the user wants to modify any of these criteria or specify another additional gene, a table containing all these criteria can be loaded to the R environment, and set as the gene-specific.df parameter. A template of how the table needs to be can be downloaded from GitHub (https://github.com/emunte/vaRHC/blob/main/data/gene\_specific.txt) or it can be found at the package documentation. Column names cannot be modified. See below the default table and the explanation of each column:

gerichalde ABSBSESESE	Saturatis 2/d2 op hyddolololol	<b>op</b> yktyktedos		MANERICA SARAH SAR	<b>Bippppty</b>	
AT0M9500.00.500050ANAN	ANANAI e-NANANANAN	ANANANAN	VA< > 0.249733>>	0.50.5>	<= -	NANANANANANANA<⇒⇒1.10
	05				2.52.	.5
CDH91299000020000FL\$0SIS	LICENS MES INTERNAN AN AN AN	ANANANAN	JANANANANA<⇒	0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	05				2.52.	.5
CHE9920.00.0005400 NA	1e-5e-< >=6.6 <b>6</b> .66	NA	< >€.700700€	• 0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	$05 \ 05$				2.52.	.5
gende <b>99</b> 000.00.00 <b>0</b> 50FL <b>2</b> 0SS	NK8nMe <u>42Bae</u> aloh <u>y</u> Nc4NnAer	NA	< >=0.700700€>	0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	$05 \ 05$				2.52.	.5
MIQH9600.00.000000010 NA	homNø2enNaANANANA	NA	$NANANANA< \Rightarrow$	0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	05				2.52.	.5
MSH9500.00.00000010 NA	homNø2enNaANANANA	NA	$NANANANA< \Rightarrow$	• 0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	05				2.52.	.5
M <b>SH95</b> 00.00200022 NA	homNø2eh&AMANANA	NA	$NANANANA< \Rightarrow$	• 0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	05				2.52.	.5
PM <b>S25</b> 000.002080028 NA	homNø2enNaANANANA	NA	$NANANANA< \Rightarrow$	• 0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	05				2.52.	.5
PTMEA2000000000010FL20SS	MSMoMMealthealth.dander =	1 NANAN	JANANANANA<⇒	0.50.5>	<= -	$NANANANANANANA< \Rightarrow = 0.10$
	05				2.52.	.5
TP53999.00100037L20SS	<b>ILICENS_PERS</b> *DOS=6.6 <b>6</b> .66	NA	$NANANANA \Leftrightarrow$	0.50.5>	<= -	< >=0.16.16;⇒=1525<⇒=0.16
					2.52.	.5

- BA1: float number. Cut-off used to assign BA1 criterion.
- BS1: float number. Cut-off used to assign BS1 criterion.
- IC: float number. Interval of confidence used to assign BA1 and BS1 criterion
- alleles: int number. Minimum number of alleles needed in the subpopulation (it will only be considered when IC is not specified).
- BS2: int number. Minimum number of individuals that need to carry the variant to assign BS2.
- **BS2\_sup**: *int number*. Minimum number of individuals that need to carry the variant to assign BS2\_sup.
- **BS2\_db**: character Db to be queried for BS2 criterion. Please introduce one of the following options FLOSSIES, GNOMAD\_non\_cancer, GNOMAD\_non\_neuro or NA. .
- **status**: character Type of zigosity asked for BS2 criterion. Please introduce one of the following options homo\_healthy, hete\_healthy, NA.
- PM2: float number. Cut-off used to assign PM2 criterion.
- PM2\_sup: float number. Cut-off used to assign PM2\_supporting criterion.
- predictors section: Predictors shown are: nucleotide conservation(phylop, phastcons, gerp), protein level (revel, VEST4, provean, bayesDel\_noAF, agvgd, polyphen, MAPP, prior\_utah\_prot) and splicing predcitors (spliceai, trap). See references in the tool paper. For each predictor there are 4 columns:
  - op\_predictor\_ben: character Operator to use for benign cut-off. The possible options to introduce are <, >, =<, => or !=.

- op\_predictor\_pat : character Operator to use for pathogenic cut-off. The possible options to introduce are <, >, =<, => or !=.
- **predictor\_ben** : *float number*. Benign cut-off to use with this predictor
- **predictor\_pat** : *float number*. Pathogenic cut-off to use with this predictor

Be careful, changing a predictor cut-off will only be considered when the guidelines specify to use that predictor.

• **BP7\_splicing**: *character* Wheter the BP7 criteria is dependent on splicing prediction not being altered or not. Possible options are *dependent*, *indpendent*.

```
eg.gene <- "CDH1"
eg.variant <- "c.1137+1G>A"
data("gene_specific")
eg.gene.specific <- gene_specific
var.information <- vaR(gene = eg.gene, variant = eg.variant, gene.specific.df = eg.gene.specific)</pre>
```

**3.1.1.2.3** Connecting to webpages using javascript Insight database needs to be queried using RSelenium package. However, we have detected that some Institutes have unabled the possibility to connect to a remote server. The \*\*\*\*remotes\*\*\* parameter allows to decide if the user wants to allow this capability (TRUE) or not (FALSE). If it is set to FALSE the user will not collect information from insight database. The *browser* parameter is used to set which browser to start Rselenium server. By default is "firefox" (the recommended). If you do not have firefox installed try either "chrome" or "phantomjs" (but they have not been tested for this package).

### 3.1.1.2.4 SpliceAI related parameters

- *spliceai.program*: *Logical*. By default is FALSE, assuming that SpliceAI program is not installed in your computer. If this parameter is FALSE, the program will only classify substitutions and simple deletion variants considering a spliceAI distance of 1000 and will show masked results. If you want to classify other variants please install SpliceAI (https://pypi.org/project/spliceai/) and set the parameter to TRUE.
- *spliceai.referenc*: Path to the Reference genome hg19 fasta file. It can be downloaded from http://hgdownload.cse.ucsc.edu/goldenPath/hg19/bigZips/hg19.fa.gz. By default is NULL and it will only be taken into account if spliceai.program is set to TRUE.
- *spliceai.annotation*: Path to gene annotation file. By default it uses the file stored in package docs folder: "../docs/gencode.v38lift37.annotation.txt"
- *spliceai.distance*: *Integer*. Maximum distance between the variant and gained/lost splice site (default: 1000)
- *spliceai.masked*: Mask scores representing annotated acceptor/donor gain and unannotated acceptor/donor loss (default: 1, that is masked)

#### 3.1.1.2.5 Provean related parameters

- provean: Logical. By default is FALSE and it is assumed that provean program is not installed in your computer. Set to TRUE if you want to compute provean locally.
- provean.sh: Path to the provean.sh file. It will only be considered when provean is set to TRUE.

#### 3.1.1.2.6 Report related parameters

- excel.results: Logical. By default is FALSE and the excel file will not be created. If TRUE and excel file will be saved.
- path.original.file: If excel.results param is set to TRUE, path.original.file must contain the path to the excel template. By default is the template located in the package docs folder.

# 3.1.1.2.7 Output dir

- *output.dir*: By default is NULL and the output will be saved in the working directory. If yu want to save the output in another folder please enter here the path.
- **3.1.2 Output** The result is stored in a list but it can also be exported in a xlsx file. The list has to dimensions.
- **3.1.2.1 vaRinfo** Where all the information retrieved is stored. The names of the list elements are the following.

```
      names(var.information$vaRinfo)

      #> [1] "Variant.Info"
      "Variant.Info.other"

      #> [3] "variant.correction"
      "gene.specific.info"

      #> [5] "gnomAD"
      "flossies.db"

      #> [7] "clinVar"
      "predictors"

      #> [9] "codon.stop"
      "second.met"

      #> [11] "insight.info"
      "functional.assays"

      #> [13] "google.scholar.30.references"
      "cancer.hotspots"

      #> [15] "class.info"
```

 Variant.Info: a data.frame containing a summary of variant location, different nomenclatures and variant consequences.

```
#> 'data.frame':
                   1 obs. of 19 variables:
#> $ gene
                              : chr "CDH1"
#> $ NM
                              : chr "NM 004360.5"
#> $ initial.var
                             : chr "c.1137+1G>A"
#> $ variant
                              : chr "c.1137+1G>A"
#> $ protein
                              : chr "p.?"
#> $ genomic
                              : chr "NC_000016.9:g.68846167G>A"
#> $ chr
                              : chr "16"
#> $ start
                              : int 68846167
#> $ end
                              : int 68846167
#> $ ref
                              : chr "G"
                              : chr "A"
#> $ alt
#> $ strand
                              : int 1
#>
   $ exon_intron
                              :'data.frame':
                                               1 obs. of 1 variable:
#>
    ..$ exon: int 8
#> $ ensembl.id
                              : chr "ENST00000261769"
#> $ most.severe.consequence : chr "splice_donor_variant"
   $ most.severe.consequence.1: chr NA
#> $ most.severe.consequence.2: chr NA
#> $ CCDS
                              : chr "CCDS10869.1"
#> $ domain.info
                              : chr [1, 1:2] "Cadherin 3" "Cadherin"
```

- Variant.Info.other: only for TP53 and CDK2NA other transcripts.
- variant.correction: output obtained by quering Mutalyzer.
- gene.specific.info: variant gene specificities used
- **gnomAD**: a list containing gnomAD v2.1.1 variant nomeclature, coverage and alleles information for non\_cancer and non\_neuro datasets. See below examples of how to obtain the gnomAD data

#### #nomenclature

var.information\$vaRinfo\$gnomAD\$nomenclature

#> [1] "16-68846167-G-A"

#coverage

var.information\$vaRinfo\$gnomAD\$coverage

#> \$exomes

#> [1] 92.4

#>

#> \$genomes

#> [1] 33.94

#information from exomes non cancer separated by subpopulations

knitr::kable(var.information\$vaRinfo\$gnomAD\$info\$exomes\$non.cancer\$subpopulations)

	AC	AN	nhomalt	AF	CI
non_cancer_nfe	0	102722	0	0	0
$non\_cancer\_fin$	0	21630	0	0	0
non_cancer_amr	0	34260	0	0	0
$non\_cancer\_afr$	0	14902	0	0	0
$non\_cancer\_sas$	0	30526	0	0	0
non_cancer_eas	0	17692	0	0	0
non_cancer_asj	0	9570	0	0	0
${\rm non\_cancer\_oth}$	0	5618	0	0	0

# #information from exomes + genomes non neuro overall frequency

knitr::kable(var.information\$vaRinfo\$gnomAD\$info\$exomes.genomes\$non.neuro\$overall)

rowname	AC	AN	nhomalt	AF	CI
	1	229382	0	4.4e-06	0

- flossies.db: a dataframe containing information obtained from FLOSSIES database.
- **clinVar**: a list containing information of the variant of interest and if it is a missense also information from other missense variants located at the same codon.
- **predictors**: a datafrane with all the predictors information stored. Only the predictors where use column is yes will be considered to calculate criteria.

	type	predictor	classific	catione	values posit	io <b>n</b> perate	or.b <b>Bhig</b> n	ıt. <b>of</b> ferate	or.pa <b>thb</b> g <b>ení</b> coff
Phylop	Nucleotide conserva- tion	Phylop	NA	no	9.31715NA	NA	NA	NA	NA
Phastcons	Nucleotide conserva- tion	Phastcons	NA	no	1.00000NA	NA	NA	NA	NA

	type	predictor	classifica	tiosae	value	s positi	operator	.bBHignut	.offerator.pa	af <b>hDgent</b> coff
Gerp	Nucleotide conserva- tion	Gerp	NA	no	5.720	00NA	NA	NA	NA	NA
Revel	Protein effect	Revel	NA	no	NA	NA	NA	NA	NA	NA
VEST4	Protein effect	VEST4	NA	no	NA	NA	<=	0.5	>	0.50
Provean	Protein effect	Provean	NA	no	NA	NA	>	-2.5	<=	-2.50
BayesDel_noA	FProtein effect	BayesDel_noA	FNA	no	0.141	00NA	NA	NA	NA	NA
aGVGD_zebra	afi <b>Eh</b> rotein effect	aGVGD_Zebra	afi <b>sia</b>	no	NA	NA	NA	NA	NA	NA
PolyPhen	Protein effect	PolyPhen	NA	no	NA	NA	NA	NA	NA	NA
MAPP	Protein effect	MAPP	NA	no	NA	NA	NA	NA	NA	NA
Prior_utah(M.	APPotPiR2) effect	Prior_utah(M.	ANA/PP2	)no	NA	NA	NA	NA	NA	NA
Prior_utah_sp	oli <b>splig<u>i</u>ng</b> feren Predictor	nderior_utah_sp	oli <b>NiA</b> g_ret	fe <b>ne</b> n	cŧNA	NA	NA	NA	NA	NA
Prior_utah_sp	oli <b>&amp;inlig</b> in <b>g</b> e_no Predictor	ovBrior_utah_sp	oli <b>MiA</b> g_de	_mov	νdNA	NA	NA	NA	NA	NA
SpliceAI- AcceptorGain	Splicing Predictor	SpliceAI- AcceptorGain	Benign	yes	0.010	0322	<=	0.15	>=	0.50
SpliceAI- AcceptorLoss	Splicing Predictor	SpliceAI- AcceptorLoss	Benign	yes	0.000	00 - 129	<=	0.15	>=	0.50
SpliceAI- DonorGain	Splicing Predictor	SpliceAI- DonorGain	Pathoge	niyes	0.560	00-84	<=	0.15	>=	0.50
SpliceAI- DonorLoss	Splicing Predictor	SpliceAI- DonorLoss	Pathoge	ni <u>w</u> es	0.990	00 -1	<=	0.15	>=	0.50
TraP	Splicing Predictor	TraP	NA	no	NA	NA	<=	0.459	>=	0.93

- codon.stop: a list with the following elements:
  - variant.exon: contains the exon where the variants is located and its coordinates.

transcript	exon	V1	V2	cStart	$\operatorname{cStop}$	cdna.var.pos
LRG_301t1	8	68846038	68846166	1009	1137	1

- **premature.ter.codon**: for frameshifts and nonsense variants it contains the exon where the stop codon is produced and its coordinates.
- length.transcript: total number of coding nucleotides that the transcript has
- porc.prot: percentatge of protein conserved
- canonical.skip.pred: only for canonical splice variants. It contains information from the predicted skipping variant, its consequence and percentage of protein conserved.

variant	protein	most.severe.consequence	porc.prot.splicing
c.1009_1137del	$p.(Ser 337\_Thr 379 del)$	$inframe\_deletion$	0.0524805

• exons: exon coordinates according to LRG transcript. In genomic and coding dna nomenclature.

transcript	exon	V1	V2	cStart	cStop
LRG_301t1	1	68771195	68771366	-124	48
LRG_301t1	2	68772200	68772314	49	163
LRG_301t1	3	68835573	68835796	164	387
LRG_301t1	4	68842327	68842470	388	531
LRG_301t1	5	68842596	68842751	532	687
LRG_301t1	6	68844100	68844244	688	832
LRG_301t1	7	68845587	68845762	833	1008
LRG_301t1	8	68846038	68846166	1009	1137
LRG_301t1	9	68847216	68847398	1138	1320
$LRG\_301t1$	10	68849418	68849662	1321	1565
$LRG\_301t1$	11	68853183	68853328	1566	1711
$LRG\_301t1$	12	68855904	68856128	1712	1936
$LRG\_301t1$	13	68857302	68857529	1937	2164
$LRG\_301t1$	14	68862077	68862207	2165	2295
$LRG\_301t1$	15	68863557	68863700	2296	2439
$\rm LRG\_301t1$	16	68867193	68869444	2440	*2038

- second.met: only for start codon variants. It returns second metionine position and clinVar variants between first and second metionine.
- insight.info: information from INSIGHT database classifications and MMR Integrative Evaluation (http://www.insight-database.org/classifications/ and http://www.insight-database.org/classifications/mmr\_integrative\_eval.html)
- functional.assays: a list containing information from functional assays. Further details are explained in the tool article.
- cancer.hotspots: information obtained from Cancer Hotspots (http://www.insight-database.org/classifications/mmr\_integrative\_eval.html)
- 3.1.2.2 vaRclass All the assigned and denied criteria are stored in this list as well as final classification.

The list has the following items:

```
names(var.information$vaRclass)
#> [1] "final.classification" "final.criteria"
```

The final classification element contains:

- final.class: variant's final classification
- criteria.assingned: a vector containing all the criteria that are assigned to the variant
- sum.criteria: Tavtigian's Bayesian score
- $\bullet$   $\,$  discrep. reason: wheter there are discrepances or not and the reason.

The final criteria element contains: + Criteria res: a matrix with all the criteria assigned (1), denied (0), not applicable (NA) or not calculated (NC). + Criterion name + .message: reasoning for the criterion being assigned or denied.

	${\tt very\_strong}$	strong	moderate	supporting
PVS1	0	1	0	0
PS1	NA	NA	NA	NA
PS2	NC	NC	NC	NC
PS3	NC	NC	NC	NC
PS4	NC	NC	NC	NC
PM1	NA	NA	NA	NA
PM2	NA	NA	0	1
PM3	NC	NC	NC	NC
PM4	NA	NA	NC	NA
PM5	NC	NC	0	1
PM6	NC	NC	NC	NC
PP1	NC	NC	NC	NC
PP2	NA	NA	NA	NA
PP3	NA	NA	NA	NA
PP4	NC	NC	NC	NC
PP5	NA	NA	NA	NA
BA1	0	NA	NA	NA
BS1	NA	0	NA	NA
BS2	NC	0	NC	0
BS3	NC	NC	NC	NC
BS4	NC	NC	NC	NC
BP1	NA	NA	NA	NA
BP2	NA	NA	NA	0
BP3	NC	NC	NC	NC
BP4	NA	NA	NA	NA
BP5	NC	NC	NC	NC
BP6	NA	NA	NA	NA
BP7	NA	NA	NA	0
external	NC	NC	NC	NC

```
#> [1] "PVS1_strong is assigned according to site-specific recomendations in the splicing table (CDH1 G
#>
#> $PS1.message
#> [1] "PS1 does not apply for this variant."
```

#> \$PS3.message

#> \$PVS1.message

#> [1] "PS3 is not assigned because variant is not found in the automated functional studies. Please ch

#> \$BS3.message

#> [1] "BS3 is not assigned because variant is not found in the automated functional studies. Please ch #>

#> [1] "As PVS1 is assigned, PM1 is not calculated"

#> \$PM2.message

#> [1] "PM2\_supporting is assigned because the variant is observed in 4e-06 frequency which is < 9.9999

#> \$PM4.message

#> [1] "As PVS1 is assigned, PM4 is not calculated"

#>

#> \$PM5.message

```
#> [1] "PM5_supporting is assigned according to site-specific recommendations for canonical splicing va
#>
#> $PP2.message
#> [1] "PP2 does not apply for this variant or gene."
#> $PP3.message
#> [1] "PP3 is not calculated because PVS1 is met and co-usage is not permitted"
#> $BA1.message
#> [1] "Freq gnomAD all non-cancer v2.1.1 = 0.00037% (MCAF 99.99% = 0.00037%)..Max freq in afr subpopul
#> [1] "The location is well covered in gnomAD (at least in exomes) but the variant is found in AF < 0
#>
#> $BS2.message
#> [1] "BS2 is not assigned taking into account the db queried. Please check if there exist more."
#>
#> $BP1.message
#> [1] "Not automated criteria"
#> $BP2.message
#> [1] "BP2 is denied because the variant is not observed in homozygous state in gnomAD v2.1.1 non_canc
#>
#> $BP4.message
#> [1] "BP4 is not calculated because PVS1 is met and co-usage is not permitted."
#> $BP7.message
\# [1] "BP7 is denied bc.1399T>Cecause variants must be positioned at or beyond +7/-21."
#> $PP5.message
#> [1] "According to ClinGen PP5 should not be applied since variants in reputable sources are not alwa
#>
#> [1] "According to ClinGen BP6 should not be applied since variants in reputable sources are not alwa
#> $PS2.message
#> [1] "Not automated criteria"
#>
#> $PS4.message
#> [1] "Not automated criteria"
#> $PM3.message
#> [1] "Not automated criteria"
#>
#> $PM6.message
#> [1] "Not automated criteria"
#>
#> $PP1.message
#> [1] "Not automated criteria"
#> $PP4.message
#> [1] "Not automated criteria"
#>
```

#> \$BS4.message

```
#> [1] "Not automated criteria"
#>
#> $BP3.message
#> [1] "Not automated criteria"
#>
#>
$BP5.message
#> [1] "Not automated criteria"
```

3.1.2.3 Excel file The excel file has the following sheets:

- Classification summary: variant summary and criteria assigned.
- Evidence: justification of every evidence given to a variant.
- Frequency control: population dta from gnomAD v2.1.1 non\_cancer, non\_neuro, FLOSSIES database and cancer hotspots
- ClinVar: variant information from ClinVar.
- ClinVar variants: List of variants in ClinVar at the same codon(only for missense variants).
- ClinVar + variant name: One sheet per variant listed in the previous sheet containing all the information provided by ClinVar.
- Predictors: Variant scores for the predictors listed and its cut-offs.
- NMD: Information for loss of function variants due to frameshift, nonsense or canonical splice site alterations.
- Start Codon: for start codon variants: list of ClinVar variants upstream the second in-frame metionine
- Bibliography vaRCH: a string containing all possible variant nomenclature to put in a search engine such as google. It also returns the 30 first articles listed in google scholar. Be careful because Google scholar has a limited of searches per day. When this searches are overpasses an Error 429 is returned.

#### 3.1 vaRbatch()

**vaRbatch()** allows to perform vaR function in batch.

**3.1.1 Input** The input is a data frame containing gene and variant column. See and example below. This function only works for the NM described in 3.1.1.2.1 section.

```
#Example 1
data("example_input_vaRbatch")
example_input_vaRbatch[] <- lapply(example_input_vaRbatch, as.character) #convert to character
all <- vaRbatch( all.variants = example_input_vaRbatch, spliceai.program = TRUE, splieai.reference= "./z
#Example2
eg.variants <- data.frame(gene=c("ATM", "MSH6", "BRCA1"), variants = c("c.8420A>T", "c.1559G>A", "c.211.batch.results <- vaRbatch( all.variants = eg.variants, spliceai.program =FALSE, print.data.frame = FALSE.</pre>
```

**3.1.2 Output** It returns a list containing vaR() output for all variants. The user can choose to print a data frame (*print.data.frame*) to store classification results in a csv file. The function also returns a log file, with the detailed time execution and any error that may occur explained. The log file is stored in a log folder in the working directory.

# 4. Legal advice and privacy policy for users

Legal advice

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The user undertakes to indemnify and hold harmless the website for any damage, prejudice, penalty, fine, penalty, or compensation that may have to have the website.

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Furthermore, if you consider that you have not obtained satisfaction in the exercise of your rights, you will be able to file a complaint before the Catalan Data Protection Agency, and you will be able to find more information on the processing of your data from the IDIBELL to the following Privacy Policy.

# 5. Supplementary Tables

ST1

ensembltranscriptID	namegene	NM	NC	CCDS
ENST00000318602	A2M	NM_000014.6	NC_000012.11	CCDS44827.1
ENST00000642412	A4GALT	$NM\_017436.7$	$NC\_000022.10$	CCDS14041.1
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ENST00000374736	ABCA1	$NM\_005502.4$	$NC\_000009.11$	CCDS6762.1
ENST00000650372.1	ABCB11	$NM\_003742.4$	$NC\_000002.11$	CCDS46444.1
ENST00000265316.9	ABCB6	$NM\_005689.4$	$NC\_000002.11$	CCDS2436.1
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ENST00000645237.2	ABCC4	$NM\_005845.5$	$NC\_000013.10$	CCDS9474.1
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ENST00000218104.6	ABCD1	$NM\_000033.4$	$NC\_000023.10$	CCDS14728.1
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ENST00000405322.8	ABCG5	$NM_{-}022436.3$	$NC\_000002.11$	CCDS1814.1
ENST00000272286.4	ABCG8	$NM_{022437.3}$	$NC\_000002.11$	CCDS1815.1
ENST00000644371.2	ABHD5	$NM\_016006.6$	$NC\_000003.11$	CCDS2711.1
ENST00000318560.6	ABL1	$NM\_005157.6$	$NC\_000009.11$	CCDS35166.1
ENST00000611156	ABO	$NM_020469.3$	$NC\_000009.11$	-
ENST00000281182.9	ACAD8	$NM\_014384.2$	$NC\_000011.9$	CCDS8498.1
ENST00000370841.9	ACADM	$NM\_000016.5$	$NC\_000001.10$	CCDS668.1
ENST00000358776.7	ACADSB	$NM\_001609.3$	$NC\_000010.10$	CCDS7634.1
ENST00000265838.9	ACAT1	NM 000019.4	NC 000011.9	CCDS8339.1

ensembltranscriptID	namegene	NM	NC	CCDS
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ENST00000648477.1	ACP5	NM_001611.5	$NC\_000019.9$	CCDS12265.1
ENST00000366684.7	ACTA1	$NM_{-}001100.3$	$NC\_000001.10$	CCDS1578.1
ENST00000224784.10	ACTA2	$NM_{-}001613.2$	$NC\_000010.10$	CCDS7392.1
ENST00000646664.1	ACTB	NM_001101.3	$NC\_000007.13$	CCDS5341.1
ENST00000290378.6	ACTC1	$NM\_005159.4$	$NC\_000015.9$	CCDS10041.1
ENST00000394419.9	ACTN1	$NM\_001130004.1$	$NC\_000014.8$	CCDS45130.1
ENST00000366578.6	ACTN2	$NM_{-}001103.4$	NC_000001.10	CCDS1613.1
ENST00000388922.9	ACVRL1	$NM\_000020.2$	NC_000012.11	CCDS31804.1
ENST00000372874	ADA	$NM\_000022.2$	$NC\_000020.10$	CCDS13335.1
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ENST00000371929.7	ADAMTS13	$NM_{139025.4}$	NC_000009.11	CCDS6970.1
ENST00000368474.9	ADAR	NM_001111.5	NC_000001.10	CCDS1071.1
ENST00000297323.12	ADCY1	$NM_{-}021116.4$	NC_000007.13	CCDS34631.1
ENST00000405460.9	ADGRV1	$NM_032119.4$	NC_000005.9	CCDS47246.1
ENST00000241356.5	ADORA3	NM 000677.3	NC 000001.10	CCDS839.1
ENST00000280155.4	ADRA2A	NM_000681.4	NC_000010.10	CCDS7569.2
ENST00000269143.8	AFG3L2	NM 006796.2	NC 000018.9	CCDS11859.1
ENST00000649286.2	AGK	NM 018238.4	NC 000007.13	CCDS5865.1
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ENST00000279146	AIP	$NM_{003977.2}$	NC_000011.9	CCDS8168.1
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ENST00000320560.13	ANO6	NM 001025356.3	NC 000012.11	CCDS31782.1
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ENST00000255194.11	AP3B1	NM 003664.4	NC 000005.9	CCDS4041.1
ENST00000643116.3	AP3D1	NM 001261826.3	NC 000019.9	CCDS58638.1
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ENST00000673436.1	ATXN2	NM_001372574.1	NC_000012.11	-
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ENST00000674280.1	ATXN7	NM_001377405.1	NC_000003.11	CCDS43102.1
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ENST00000646375.1	AVPR2	NM_000054.7	NC_000023.10	CCDS14735.1
ENST00000307078.10	AXIN2	NM_004655.3	NC_000017.10	CCDS11662.1
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ENST00000379198.5	B3GALT6	NM_080605.4	NC_000001.10	CCDS13.1
ENST00000261499.10	B9D1	NM_015681.6	NC_000017.10	CCDS11205.1
ENST00000369085.8	BAG3	NM_004281.3	NC_000010.10	CCDS7615.1
ENST00000460680	BAP1	NM_004656.3	NC_000003.11	CCDS2853.1
ENST00000260947	BARD1	NM_000465.3	NC_000002.11	CCDS2397.1
ENST00000650064.2	BBS10	NM_024685.4	NC_000012.11	CCDS9014.2
ENST00000270233.12	BCAM	NM_005581.4	NC_000019.9	CCDS12644.1
ENST00000648566.1	BCL10	NM_003921.5	NC_00001.10	CCDS704.1
ENST00000333681.5	BCL2	NM_000633.3	NC_000018.9	CCDS11981.1

ensembl transcript ID	namegene	NM	NC	CCDS
ENST00000615946.4	BCL2L11	NM_001204107.1	NC_000002.11	CCDS56132.1
ENST00000342274.8	BCOR	NM_001123383.1	$NC\_000023.10$	CCDS14250.1
ENST00000218147	BCORL1	NM 021946.4	NC 000023.10	CCDS14616.1
ENST00000305877.13	BCR	NM 004327.4	$\overline{NC} 000022.10$	CCDS13806.1
ENST00000392111.7	BCS1L	$\overline{NM} 004328.4$	$\overline{NC} 000002.11$	CCDS2419.1
ENST00000373886.8	BICC1	$\overline{NM} 001080512.3$	$\overline{NC} 000010.10$	CCDS31206.1
ENST00000316724.10	BIN1	$\overline{NM} 139343.3$	$\overline{NC} 000002.11$	CCDS2138.1
ENST00000263464.9	BIRC3	$\overline{NM} 001165.5$	NC 000011.9	CCDS8315.1
ENST00000355112.8	$\operatorname{BLM}$	NM 000057.2	$\overline{NC} 000015.9$	CCDS10363.1
ENST00000224337.10	BLNK	$\overline{NM} 013314.3$	$\overline{NC} 000010.10$	CCDS7446.1
ENST00000433642.3	BLOC1S3	$\overline{NM} 212550.4$	$\overline{NC} 000019.9$	CCDS12656.1
ENST00000220531.9	BLOC1S6	$\overline{NM} 012388.3$	$\overline{NC} 000015.9$	CCDS10126.1
ENST00000295379.2	BMP10	NM 014482.1	NC 000002.11	CCDS1890.1
ENST00000649409.2	BMPER	NM 001365308.1	NC 000007.13	CCDS5442.1
ENST00000372037	BMPR1A	NM 004329.2	NC 000010.10	CCDS7378.1
ENST00000374580.10	BMPR2	$\overline{NM} 001204.6$	$\overline{NC} 000002.11$	CCDS33361.1
ENST00000646891.1	$\operatorname{BRAF}$	NM 004333.4	NC 000007.13	CCDS5863.1
ENST00000357654	BRCA1	NM 007294.3	NC 000017.10	CCDS11453.1
ENST00000544455	BRCA2	NM 000059.3	NC 000013.10	CCDS9344.1
ENST00000259008	BRIP1	NM 032043.2	NC 000017.10	CCDS11631.1
ENST00000360796.10	BSCL2	NM 001122955.3	NC 000011.9	CCDS44627.1
ENST00000353555.9	BSG	NM 198589.3	NC 000019.9	CCDS12034.1
ENST00000651561.1	BSND	NM 057176.3	$\overline{NC} 000001.10$	CCDS602.1
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ENST00000308731.8	BTK	NM 000061.2	$\overline{NC} 000023.10$	CCDS14482.1
ENST00000287598	BUB1B	$\overline{NM} 001211.5$	$\overline{NC} 000015.9$	CCDS10053.1
ENST00000374642.8	C1QA	NM 015991.2	$\overline{NC} 000001.10$	CCDS226.1
ENST00000314933	C1QB	NM 000491.3	NC 000001.10	CCDS228.1
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ENST00000360817.10	C1S	$NM_001734.3$	NC_000012.11	CCDS31735.1
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ENST00000245907.11	C3	$NM\_000064.2$	$NC\_000019.9$	CCDS32883.1
ENST00000428956.7	C4A	$NM\_007293.2$	$NC\_000006.11$	CCDS47404.1
ENST00000435363.7	C4B	${\rm NM}\_001002029.3$	$NC\_000006.11$	CCDS47405.1
ENST00000367070.8	C4BPA	$NM\_000715.4$	$NC\_000001.10$	CCDS1477.1
ENST00000367078.8	C4BPB	$NM\_001017365.3$	$NC\_000001.10$	CCDS1476.1
ENST00000223642.3	C5	$NM\_001735.2$	$NC\_000009.11$	CCDS6826.1
ENST00000392122	C6	$NM\_000065.2$	$NC\_000005.9$	CCDS3936.1
ENST00000313164.10	C7	$NM\_000587.2$	$NC\_000005.9$	CCDS47201.1
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ENST00000381911.6	TNNI2	$NM\_003282.4$	$NC\_000011.9$	CCDS31333.1
ENST00000344887.10	TNNI3	$NM\_000363.4$	$NC\_000019.9$	CCDS42628.1
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ENST00000588981.6	TNNT1	$NM\_003283.6$	$NC\_000019.9$	CCDS12917.1
ENST00000656932.1	TNNT2	$NM\_001276345.2$	NC_000001.10	CCDS73003.1
ENST00000278317.11	TNNT3	$NM\_006757.4$	NC_000011.9	CCDS7727.1
ENST00000351698.5	TOR1A	$NM_{-}000113.3$	$NC\_000009.11$	CCDS6930.1
ENST00000269305	TP53	$NM\_000546.5$	$NC\_000017.10$	CCDS11118.1
ENST00000264731.8	TP63	$NM\_003722.5$	$NC\_000003.11$	CCDS3293.1
ENST00000396705.10	TPI1	$NM\_000365.6$	$NC\_000012.11$	CCDS8566.1
ENST00000403994.9	TPM1	$NM\_001018005.2$	$NC\_000015.9$	CCDS45273.1
ENST00000378292.9	TPM2	$NM_{213674.1}$	$NC\_000009.11$	CCDS6586.1
ENST00000323144.12	TPM3	$NM\_001043353.2$	$NC\_000001.10$	CCDS41401.1
ENST00000643579.2	TPM4	$NM\_003290.3$	$NC\_000019.9$	CCDS12338.1
ENST00000309983.5	TPMT	$NM\_000367.5$	$NC\_000006.11$	CCDS4543.1
ENST00000299427.12	TPP1	$NM\_000391.4$	NC_000011.9	CCDS7770.1
ENST00000376052.5	TPP2	$NM\_001330588.2$	$NC\_000013.10$	CCDS81777.1
ENST00000409012.6	TPRN	$NM\_001128228.3$	$NC\_000009.11$	CCDS56594.1
ENST00000392745.8	TRAF3	$NM_{145725.2}$	$NC\_000014.8$	CCDS9975.1
ENST00000368761.11	TRAF3IP2	$NM_{147686.4}$	NC_000006.11	CCDS5093.1
ENST00000648948.2	TRAPPC9	NM_031466.8	NC_000008.10	CCDS55278.1
ENST00000373113.8	TREM2	$NM\_018965.3$	NC_000006.11	CCDS4852.1
ENST00000625293.3	TREX1	$NM_033629.6$	NC_000003.11	CCDS2769.1
ENST00000450136.2	TRIM32	$NM_012210.3$	NC_000009.11	CCDS6817.1
ENST00000374272.4	TRIM63	$NM\_032588.3$	NC_000001.10	CCDS273.1
ENST00000251607.11	TRNT1	$NM_{182916.3}$	NC_000003.11	CCDS2561.2
ENST00000646667.1	TRPM7	$NM\_017672.6$	$NC\_000015.9$	CCDS42035.1
ENST00000261740.7	TRPV4	$NM_021625.4$	NC_000012.11	CCDS9134.1
ENST00000298552	TSC1	$NM\_000368.4$	NC_000009.11	CCDS6956.1
ENST00000219476	TSC2	$NM\_000548.3$	$NC\_000016.9$	CCDS10458.1
ENST00000298171.7	TSHR	$NM\_000369.2$	$NC\_000014.8$	CCDS9872.1
ENST00000368608	TSPYL1	$NM\_003309.3$	NC_000006.11	CCDS34518.1
ENST00000358746.7	TTC37	$NM\_014639.3$	$NC\_000005.9$	CCDS4072.1
ENST00000319190.11	TTC7A	$NM\_020458.4$	NC_000002.11	CCDS33193.1
ENST00000589042	TTN	$NM\_001267550.1$	NC_000002.11	CCDS59435.1
ENST00000237014.8	TTR	NM_000371.3	NC_000018.9	CCDS11899.1

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ENST00000217133.2	TUBB1	NM 030773.3	NC 000020.10	CCDS13475.1
ENST00000400521.7	TXNRD2	NM 006440.3	NC 000022.10	CCDS42981.1
ENST00000525621.6	TYK2	NM 003331.4	$\overline{NC} 000019.9$	CCDS12236.1
ENST00000395680.6	TYMP	$\overline{NM} 001257988.1$	$\overline{NC} 000022.10$	CCDS14096.1
ENST00000323274.15	TYMS	$\overline{NM} 001071.2$	$\overline{NC} 000018.9$	CCDS11821.1
ENST00000262629.9	TYROBP	NM 003332.3	$\overline{NC} 000019.9$	CCDS12482.1
ENST00000380276.6	U2AF1	$\overline{NM} 001025203.1$	$\overline{NC} 000021.8$	CCDS33574.1
ENST00000450554	U2AF2	$\overline{NM} 007279.2$	$\overline{NC} 000019.9$	CCDS12933.1
ENST00000646651.1	UBE2T	$\overline{NM} 014176.4$	$\overline{NC} 000001.10$	CCDS1425.1
ENST00000438097	UBE3A	NM 130838.1	$\overline{NC} 000015.9$	CCDS32177.1
ENST00000338222	UBQLN2	NM 013444.3	NC 000023.10	CCDS14374.1
ENST00000305208.10	UGT1A1	NM 000463.2	NC 000002.11	CCDS2510.1
ENST00000335765.9	UNC119	$\overline{NM} 005148.3$	$\overline{NC} 000017.10$	CCDS11233.1
ENST00000207549.9	UNC13D	$\overline{NM} 199242.2$	$\overline{NC} 000017.10$	CCDS11730.1
ENST00000227471.7	UNC93B1	NM 030930.2	$\overline{NC} 000011.9$	CCDS73334.1
ENST00000242576.7	UNG	NM 080911.1	NC 000012.11	CCDS9124.1
ENST00000246337.9	UROD	$\overline{NM} 000374.5$	$\overline{NC} 000001.10$	CCDS518.1
ENST00000368797.10	UROS	$\overline{NM} 000375.3$	$\overline{NC} 000010.10$	CCDS7648.1
ENST00000219281.8	USB1	$\overline{NM} 024598.3$	$\overline{NC} 000016.9$	CCDS10791.1
ENST00000614341.5	USH1G	NM 173477.5	$\overline{NC} 000017.10$	CCDS32725.1
ENST00000372429.8	USP20	$\overline{NM} 001110303.4$	NC 000009.11	CCDS43892.1
ENST00000475243.6	VAPB	$\overline{NM} 004738.4$	$\overline{NC} 000020.10$	CCDS33498.1
ENST00000211998.10	VCL	$NM_{014000.2}$	$NC\_000010.10$	CCDS7341.1
ENST00000358901.11	VCP	$NM_{-}007126.3$	NC_000009.11	CCDS6573.1
ENST00000256474	VHL	$NM\_000551.3$	NC_000003.11	CCDS2597.1
ENST00000557658.6	VIPAS39	$NM\_001193315.2$	$NC\_000014.8$	CCDS9862.1
ENST00000394975.3	VKORC1	$NM_024006.6$	$NC\_000023.10$	CCDS10703.1
ENST00000330374.7	VMA21	$NM\_001017980.3$	NC_000008.10	CCDS35430.1
ENST00000358544.7	VPS13B	$NM_017890.4$	NC_000001.10	CCDS6280.1
ENST00000620676.6	VPS13D	$NM_015378.4$	$NC\_000015.9$	CCDS30588.1
ENST00000333371.8	VPS33B	NM_018668.4	NC_000001.10	CCDS10369.1
ENST00000644510.2	VPS45	$NM\_007259.5$	NC_000014.8	CCDS944.1
ENST00000261405.10	VWF	$NM\_000552.5$	NC_000012.11	CCDS8539.1
ENST00000376701.5	WAS	$NM\_000377.2$	NC_000023.10	CCDS14303.1
ENST00000448612.6	WDR27	$NM_{182552.5}$	NC_000006.11	CCDS47520.2
ENST00000226760.5	WFS1	NM_006005.3	NC_000004.11	CCDS3386.1
ENST00000362057.4	WHRN	NM_015404.4	NC_000009.11	CCDS6806.1
ENST00000359761.7	WIPF1	NM_001077269.1	NC_000002.11	CCDS2260.1
ENST00000315939.11	WNK1	NM_018979.4	NC_000012.11	CCDS8506.1
ENST00000316024.9	WRAP53	NM_018081.2	NC_000017.10	CCDS11119.1
ENST00000298139.7	WRN	NM_000553.4	NC_000008.10	CCDS6082.1
ENST00000332351	WT1	NM_024426.3	NC_000011.9	CCDS7878.2
ENST00000381174.10	XG	NM_175569.3	NC_000023.10	CCDS14120.1
ENST00000371199.8	XIAP	NM_001167.3	NC_000023.10	CCDS14606.1
ENST00000378616.5	XK	NM_021083.4	NC_000023.10	CCDS14241.1
ENST00000375128.5	XPA	NM_000380.3	NC_000009.11	CCDS6729.1
ENST00000285021	XPC	NM_004628.4	NC_000003.11	CCDS46763.1
ENST00000262887.10	XRCC1	NM_006297.2	NC_000019.9	CCDS12624.1
ENST00000359321.2	XRCC2	NM_005431.2	NC_000007.13	CCDS5933.1
ENST00000373477.9	YARS1	NM_003680.3	NC_000001.10	CCDS368.1
ENST00000264972.10	ZAP70	NM_014707.2	NC_000002.11	CCDS33254.1
ENST00000230122.4	ZBTB24	NM_014797.2	NC_000006.11	CCDS34509.1

ensembltranscriptID	namegene	NM	NC	CCDS
ENST00000374423.9	ZDBF2	NM_020923.3	NC_000002.11	CCDS46501.1
ENST00000376335.8 ENST00000372759.4	ZIC2 ZMPSTE24	NM_007129.5 NM_005857.3	NC_000013.10 NC_000001.10	CCDS9495.1 CCDS449.1
ENST00000253144.13	ZNF331	$NM\_018555.6$	NC_000019.9	CCDS33102.1
ENST00000301744.7 ENST00000433976.7	ZNF597 ZNF778	NM_152457.3 NM 001201407.1	NC_000016.9 NC_000016.9	CCDS10505.1 CCDS73928.1
ENST00000433970.7 ENST00000307771.8	ZRSR2	NM_005089.3	NC_000016.9 NC_000023.10	CCDS13928.1 CCDS14172.1

#### 6. Session info

```
sessionInfo()
#> R version 4.2.1 (2022-06-23)
#> Platform: x86_64-pc-linux-gnu (64-bit)
#> Running under: Ubuntu 20.04.4 LTS
#>
#> Matrix products: default
#> BLAS: /usr/lib/x86_64-linux-gnu/blas/libblas.so.3.9.0
#> LAPACK: /usr/lib/x86_64-linux-gnu/lapack/liblapack.so.3.9.0
#>
#> locale:
#> [1] LC_CTYPE=en_US.UTF-8
                                  LC_NUMERIC=C
#> [3] LC_TIME=en_GB.UTF-8
                                  LC_COLLATE=en_US.UTF-8
#> [5] LC_MONETARY=en_GB.UTF-8
                                LC_MESSAGES=en_US.UTF-8
#> [7] LC_PAPER=en_GB.UTF-8
                                  LC_NAME = C
#> [9] LC_ADDRESS=C
                                   LC_TELEPHONE=C
#> [11] LC_MEASUREMENT=en_GB.UTF-8 LC_IDENTIFICATION=C
#> attached base packages:
                graphics grDevices utils
#> [1] stats
                                               datasets methods
#>
#> other attached packages:
#> [1] vaRHC_0.0.0.9000 dplyr_1.0.10
#> loaded via a namespace (and not attached):
#> [1] Rcpp_1.0.8.3
                                   lattice\_0.20-45
#> [3] binman_0.1.2
                                    Rsamtools_2.12.0
#> [5] Biostrings_2.64.0
                                   assertthat\_0.2.1
#> [7] digest_0.6.29
                                    utf8_1.2.2
#> [9] R6_2.5.1
                                    GenomeInfoDb_1.32.2
#> [11] stats4_4.2.1
                                    evaluate_0.15
#> [13] highr_0.9
                                    httr_1.4.2
#> [15] pillar_1.7.0
                                    zlibbioc_1.42.0
#> [17] rlang_1.0.4
                                    rstudioapi\_0.13
#> [19] S4Vectors_0.34.0
                                    Matrix_1.4-1
#> [21] rmarkdown_2.13
                                    RMySQL_0.10.23
#> [23] BiocParallel 1.30.3
                                    RSelenium 1.7.7
#> [25] wdman_0.2.5
                                    stringr_1.4.0
#> [27] RCurl_1.98-1.7
                                    DelayedArray_0.22.0
#> [29] compiler_4.2.1
                                    rtracklayer_1.56.1
#> [31] xfun_0.30
                                    pkgconfig_2.0.3
```

```
#> [33] askpass_1.1
                                   BiocGenerics_0.42.0
#> [35] htmltools_0.5.2
                                    openssl_2.0.0
#> [37] tidyselect_1.1.2
                                   SummarizedExperiment\_1.26.1
#> [39] tibble_3.1.6
                                   GenomeInfoDbData\_1.2.8
#> [41] IRanges_2.30.0
                                    codetools_0.2-18
#> [43] matrixStats_0.62.0
                                   XML_3.99-0.10
#> [45] fansi_1.0.3
                                   crayon_1.5.1
#> [47] GenomicAlignments_1.32.1
                                   bitops_1.0-7
#> [49] grid_4.2.1
                                    jsonlite_1.8.0
#> [51] lifecycle_1.0.1
                                   DBI_1.1.2
#> [53] semver_0.2.0
                                   magrittr_2.0.3
#> [55] cli_3.4.1
                                   stringi_1.7.6
#> [57] XVector_0.36.0
                                   xml2_1.3.3
#> [59] ellipsis_0.3.2
                                   generics_0.1.2
#> [61] vctrs_0.4.1
                                   rjson_0.2.21
#> [63] restfulr_0.0.15
                                  tools\_4.2.1
#> [65] Biobase_2.56.0
                                   glue_1.6.2
#> [67] purrr_0.3.4
                                   MatrixGenerics_1.8.1
#> [69] parallel_4.2.1
                                   fastmap_1.1.0
#> [71] yaml_2.3.5
                                   GenomicRanges_1.48.0
#> [73] caTools_1.18.2
                                   knitr_1.38
#> [75] BiocIO_1.6.0
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