# **AWSEM tutorial**

#### Requirements

Linux

VMD 1.9.3 [http://www.ks.uiuc.edu/]

Grace [http://plasma-gate.weizmann.] or GNUplot (sudo apt-get gnuplot grace)
Python (sudo apt-get python python-pip – sudo pip install biopython – sudo pip install numpy)

LAMMPS and AWSEM (attached file – read README.md file for installation)

1) Untar the programs file we sent you.

```
tar -xvf 'lammps-awsem.tar.bz2'
```

Or download a zip from <a href="https://github.com/Tatotavella/Workshop-3\_PBATEOTW\_Chile">https://github.com/Tatotavella/Workshop-3\_PBATEOTW\_Chile</a>

Go to lammps-awsem/src and write:

```
make clean
make all
```

Check if a Imp\_serial file appears in lammps-awsem/bin

2) Go to Tools folder and run

```
dir = lammps-awsem
cd $dir/tools
cd lammps-awsem/tools
./install_tools.sh
```

3) Make a folder to run the examples **inside lammps-awsem** folder.

```
mkdir monomer (you have to be in lammps-awsem folder!!)
cp ../bin/lmp_serial ./monomer/ (copy executable to folder)
```

### 4) Monomer structure prediction

- a) Go to PDB and download the protein with the code 1R69 (https://www.rcsb.org/structure/1r69)
- b) Copy 1r69.pdb inside monomer directory
- c) Run the script to generate initial LAMMPS files (from within monomer folder):

```
../tools/PdbCoords2Lammps.sh 1r69 1r69
```

d) File copying:

```
cp $dir/parameters/* .
cp $dir/monomer_backup_files/unfold.restart .
```

```
cp $dir/monomer_backup_files/1r69.mem .
```

python \$dir/src/awsemmd-CGpH/create\_project\_tools/GetCACADistancesFile.py 1r69
rnative.da t

python \$dir/src/awsemmd-CGpH/create\_project\_tools/GetCACoordinatesFromPDB.py 1r69
nativecoords.dat

Go to JPRED homepage: <a href="http://www.compbio.dundee.ac.uk/www-jpred/">http://www.compbio.dundee.ac.uk/www-jpred/</a>

Feed the sequence from fasta file into JPRED. Choose to continue carrying out a Jpred prediction. When the prediction is complete, look at the prediction in "ViewSimple"

Copy the JPRED prediction into a new text file, called IDjpred Call command to generate ssweight file

```
python $dir/create_project_tools/GenSswight.py IDjpred ssweight
```

#### Edit 1r69.in file:

Go to the read data line and write:

```
read_restart unfolded.restart.1r69
```

Go to the velocity attribute and write:

```
velocity 800.0 XXXXXXX
```

where in the "XXXXXX" you have to write any number of 7 digits (This is the random seed!)

Go to the fix 1 line and replace by:

```
fix 1 all nvt temp 800.0 200.0 100.0
```

Under the fix 2 add the calculation of extra variables:

```
qw alpha_carbons qwolynes rnative.dat 2 0.15
compute
variable
           qw equal c_qw
compute
          rg alpha_carbons gyration
variable
           rg equal c_rg
compute go alpha_carbons gonuchic cutoff 12.0 (continues next line)
       nativecoords.dat 1.2
variable
           qo equal c_qo
compute
          tc beta_atoms totalcontacts 6.5 2
variable
          tc equal c_tc
variable
          S_T equal step
variable
          E_P equal pe
```

## variable T\_E equal temp

## And under this add the file writing (in one line):

$$\label{eq:fix} fix \quad 4 \mbox{ all print 1000 "$\{S_T\} $\{T_E\} $\{qw\} $\{rg\} $\{qo\} $\{tc\} $\{E_P\}$" file extra_variables.dat screen no title "# Tstep Temp Qw Rg Qo Tc Energy"$$

Got to the thermo line and write:

thermo 1000

Go to the dump line and write:

dump
 1 all atom 1000 dump.lammpstrj

Go to the run line and write:

run 1000000

## e) Run your simulation!

 $./lmp\_serial < 1r69.in$ 

- 5) Follow <a href="https://github.com/adavtyan/awsemmd/wiki">https://github.com/adavtyan/awsemmd/wiki</a> for dimer prediction or monomer prediction steps.
- 7) You can try with a protein of your own interests.
- 8) You can follow AMH-Go calculations.
- 9) Try to repeat calculations with the fragment memory style.