Distributions of the dipteran genera *Drosophila* and *Scaptomyza* in Australia in relation to resource utilization

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ABSTRACT. The two Australian *Scaptomyza* species, especially the endemic *S. australis*, have broader limits for resource utilization as measured by temperature and relative humidity than do endemic *Drosophila* species. This has enabled *S. australis* in particular to spread to both hot arid and cold high upland regions of the continent where no *Drosphila* are found.

Introduction

Australian Drosophila species have a maximum range for resource utilization of about from 12-26° C (or 14° C), with many endemic species possessing somewhat smaller ranges (Parsons, 1975, 1977). Ranges of <16°C are characteristic of aquatic animals (Andrewartha & Birch, 1974) living in places where differences between maximum and minimum temperatures are small. Terrestrial or amphibious animals, however, live in places where temperatures are more variable, with ranges of usually > 20°C. The narrow Drosophila range appears to be explained by the fact that at least most members of the genus exploit the moist microbial degradation products of plants (Throckmorton, 1975), and thus occupy niches with some analogies to the aquatic environment.

The genus Scaptomyza is closely related to Drosophila and indeed in Hawaii flies are found with characters intermediate between those of the two genera (Carson et al., 1970). Scaptomyza species can, however, exist in more extreme habitats than Drosophila species (Throckmorton, 1975), so that in Hawaii where there are major radiations in both genera, the scaptomyzoids are often found in drier and more exposed habitats of high light intensity, although they also broadly overlap into drosophiloid habits; in addition Hawaiian scaptomyzoids are found in cooler regions

than drosophiloids. In South America, the southernmost endemic *Drosphila* species occurs at about 44° S, while endemic *Scaptomyza* are found in the vicinity of Punta Arenas at 53° 40′ S (Brncic & Dobzhansky, 1957). *Scaptomyza* species are also common in the Arctic (Basden, 1956).

From the above observations it is tempting to generalize that *Scaptomyza* may occur in more severe habitats than *Drosophila* both at cool and warm extremes. This paper reviews the situation in respect of the Australian species.

Methods and species

Flies were collected by sweeping foliage with a deep net (0.39 m diam.), permitting the precise characterization of preferred microhabitats. In a few cases flies were collected directly into vials from fern fronds where they were motionless. Temperature and humidities were taken with a Bacharach Hygrometer. The Victorian and Tasmanian habitats surveyed (Fig. 1) are the tree fern/sedge temperature zone habitats described in Parsons & Bock (1977). The South Australian habitats have plants with fleshy leaves, and flowers characteristic of the arid zone flora in these regions. The endemic Drosophila species, with all of the subgenus Scaptodrosophila, are described in Bock (1976), with the exception of two



FIG. 1. Map of southeastern Australia showing localities in Table 1 from which flies were collected.

TABLE 1. Microhabitats of *Drosophila* spp. and *S. australis* for various collections (see Fig. 1) in southern Australia with temperatures and relative humidities at 1 m from permanent moisture

Collection No.	Temperature (°C)	Relative humidity (%)	Distance from permanent moisture	
			Drosophila	S. australis
2	25	62	1 m	15 m
30	15	87	3 m	Open forest
34	15.5	95	3 m	Open forest
38	20	73	2 m	2 m
42	18	82	2 m	4 m
96	17	40		1 m
97	23	28		1 m
98	23.5	29	_	1 m
99	26	29		1 m
123	14	90		Off ferns
132	17	74	2 m	3 m
134	16	83	2 m	2 m

species in Parsons & Bock (1977); and the two Australian *Scaptomyza* species are redescribed in Bock (1977). All of these species hardly ever come to conventional fermented baits.

Results

Approximate distances from permanent moisture are given in Table 1, with temperatures and relative humidities at about 1 m from

permanent moisture, for twelve collections in southeastern Australia in which Scaptomyza australis was found. The lowest humidities (R.H. $\leq 40\%$), which are also associated with quite high temperatures, are from the Flinders Range collections (96-99), where S. australis only was found. The humidities of the latter collection sites are lower than those at any sites where Drosophila species have been detected, and indeed out of over 150 southern Australian collections from which one or more Drosophila species were recovered, made during the last 2 years, the relative humidity was < 60% only six times and never < 50%. For collections 2, 30, 34, 42 and 132, S. australis was further, or considerably further, from permanent moisture than Drosophila ('open forest' may of course include rivers and creeks, but the Scaptomyza recovered from these collections were found at considerable distances from permanent moisture), and not unexpectedly there are collections (38, 134) where no differentiation could be made. It is

quite clear, however, that where collections were made at the same site, S. australis showed a marked tendency to be less dependent on permanent moisture than Drosophila, while the converse situation never occurred. It should be noted that S. australis is frequently collected off flowering plants, indicating possible differences in resource utilization; however, collections 96-99 show that S. australis can clearly exist in physical environments far more extreme than Drosophila.

S. australis only was found for collection 123 at a temperature of 14°C when flies were knocked off fern fronds. This is a temperature at which Drosophila can only be swept with difficulty. Indeed at higher temperatures Drosophila but not S. australis were readily swept at this site; three such collections yielded a total of 102 Drosophila of five species, compared with five S. australis for collection 123. These findings suggest that S. australis may be active at lower temperatures than *Drosophila* species.

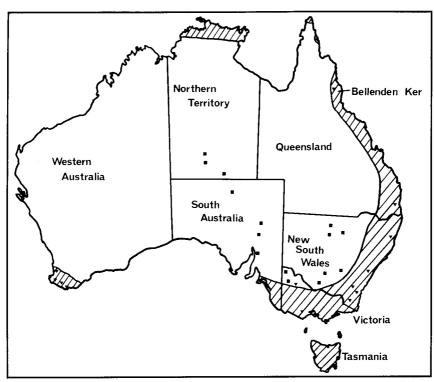


FIG. 2. Map of Australia showing the approximate known distributions of Drosophila (shaded). All known localities where S. pallida has been found are indicated. S. australis has been found in almost all of the Drosophila areas, and further specific localities for S. australis are given outside these areas. ■ S. Australis; A S. pallida.

baits and fifty-six were swept, but S. pallida

Australian distribution data

was not found.

Localities where endemic Drosophila and Scaptomyza species have been collected over a total period of several decades are given in the references listed in the methods section. These are plotted, along with the localities indicated in Fig. 1, in Fig. 2. The shaded Drosophila region corresponds to the wetter sections of Burbidge's (1960) temperate and tropical floristic zones, a finding which is not unexpected, as with few exceptions endemic Drosophila occur mainly in rain forests and permanently damp undisturbed habitats within dry sclerophyll forests, grassy forests and woodlands; the latter habitats account for the extension of Drosophila outside rain forests into a number of southern habitats, including southwest Western Australia. There is no such extension away from rain forests in the north, presumably because of the high temperature/ desiccation stresses of these regions. The drier inland and northern regions would exceed the postulated upper limit of 26°C for Drosophila resource utilization for long periods of the year, while this is not so in the south.

S. australis occurs throughout the Drosophila regions but, in addition, is also found in much more arid regions, as shown by the records in Fig. 2; this finding might be predicted from collections 96-99 in Table 1. S. australis therefore extends well into Burbidge's (1960) Eremaean floristic zone, which covers the whole of arid Australia. S. pallida, on the other hand, has not to date been

detected in the Eremaean floristic zone. A morphological difference between the two species of possible ecological significance is that the egg guides of S. australis are weakly sclerotized, while those of S. pallida are small but strongly sclerotized (Bock, 1977). S. australis, especially in arid regions, is often to be found on the fleshy flowers and leaves characteristic of these regions, suggesting that its larvae might be miners of soft plant parts, and indeed S. australis larvae are known to mine the soft fleshy leaves of chick-weed (Stellaria media) (Bock, 1977). In contrast, S. pallida larvae may be miners of some of the less fleshy (sclerophyll) leaves which are widespread in the Australian flora outside tropical rain forests (Webb, 1959), as at the summit of Mt Bellenden Ker; such an environment would be harsh indeed in the Eremaean zone.

At the cool end of the temperate range, there are records of S. australis at altitudes of just above 1600 m in the uplands of southern New South Wales; for D. fuscithorax, which occurs at the highest altitudes for Drosophila in this region, there are no records above 1500 m (Bock, 1976, 1977). Considering latitudinal extremes, both species have been found at sea level at Catamaran (collection 134 - Table 1), at the extreme south of Tasmania; the upland collections do, however, indicate the likelihood that S. australis may be tolerant of cooler temperatures than Drosophila. S. pallida has been collected in upland habitats of southern New South Wales, although not as high as S. australis; the total number of known specimens of S. pallida listed in Bock (1977) is, however, only twenty for the entire continent, so that the records are inadequate for firm conclusions to be drawn. The Mt Bellenden Ker data do, however, suggest the likelihood that S. pallida is more cold-tolerant than Drosophila.

Discussion and conclusions

From the foregoing data, it appears valid to conclude that in a continent with an extreme diversity of habitats *Scaptomyza* species, especially *S. australis*, occur in more extreme habitats than *Drosophila*. To what extent the range of 12–26°C for resource utilization determined for *Drosophila* is extended by

Scaptomyza is difficult to say; indeed the temperature range may not be greatly extended, although the effectiveness of resource utilization at the extremes is probably increased relative to that of Drosophila. There is no doubt, however, that S. australis is tolerant of much lower humidities than is any endemic Drosophila species.

The Australian endemic S. australis occurs on both sides of the Nullarbor Plain which separates the temperate zones of southwestern and southeastern Australia (S. pallida is a cosmopolitan species). D. fuscithorax, a species tolerant of greater environmental extremes than other related Drosophila species, is the only endemic Drosophila species previously recorded on both sides of the Nullarbor, although one individual of a second endemic species (D. collessi) was recently found in the southwest. D. fuscithorax would probably have maintained cross-Nullarbor links during recent arid times longer than other related southern species which, while all belonging to the inornata species group, are differentiated into closely related southwestern and southeastern forms (Bock, 1976; Parsons & Bock, 1977). By comparison, the lack of differentiation of S. australis, given that it exploits arid zone plant resources, is to be expected. Indeed detailed collections across the Nullarbor Plain for Scaptomyza, which to our knowledge have not been carried out, may well provide specimens.

In conclusion Scaptomyza australis, the only endemic member of its genus in Australia, is demonstrably more resistant to environmental extremes than are the Drosophila species and it has therefore been able to occupy territory far outside the Drosophila ranges. S. pallida, a cosmopolitan species, appears to be more resistant at least to cold than the Drosophila species but data for pallida are insufficient as yet to warrant further conclusions about this. The findings for S. australis concur with those for Scaptomyza species in other parts of the world.

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