Ann. Zool. Fennici 22: 169-203, 1985

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The evolution and phylogenetic significance of the costal chaetotaxy in the Diptera

Walter Hackman & Rauno Väisänen

Hackman, W. & Väisänen, R. 1985: The evolution and phylogenetic significance of the costal chaetotaxy in the Diptera. — Ann. Zool. Fennici 22:169-203.

The pattern of the setae on the costal vein was investigated in 875 species belonging to 128 families of Diptera. The arrangement of the setae was classified into seven basic types but intermediate types and conspicuous variations also occur in this neglected taxonomic character.

In the "nematocerous" superfamilies the costal chaetotaxy usually consists of irregularly arranged hairs. In several groups of the "lower" Muscomorpha setae occur in two homomorphous rows. In the Schizophora types with a subdorsal row of spinulae and a subventral row of hairs are also commonly encountered. Again, in some families strong subventral spines occur. In the Muscoidea in the sense used here the basic plan seems to be two heteromorphous rows of hairs alternating with spinulae, the Scathophagidae being an exception, with irregularly inserted hairs, or hairs and spinulae. In the Oestroidea strikingly primitive types resembling those in the "lower" Muscomorpha are met.

The present data suggests that the costal chaetotaxy can be successfully used at different levels of Diptera systematics. The costal chaetotaxy can be used as a diagnostic character for several subfamilies and families, and in some extreme cases even for species of a single genus. This character complex also provides significant new material for studies of the macrosystematics of the order Diptera.

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1. Introduction

The delimitation of the numerous families of the Diptera and the elucidation of their interrelationships is notoriously difficult. Hitherto neglected characters, imaginal as well as larval, are therefore in need of study. One such somewhat neglected complex of characters is the arrangement of various types of setae on the costal vein of the wing. The aim of the present study is to describe these features, and to discuss the taxonomical consequences of the results.

As long ago as the 1950s Prof. Risto Tuomikoski (unpubl.) discovered that the chaetotaxy of the costal edge of the wing could be used in several cases as a family character in the acalyptrate Diptera. In Finnish material collected mainly by himself he separated six basic types of chaetotaxy in which different types of setae are combined in various ways, either irregularly or in more or less clearly defined rows. However, he never published his results, and when abandoning dipterology later in favour of other scientific interests he suggested the authors continue the work and apply it in connection with new literature on the systematics of the Diptera. The authors have extended the study of these overlooked taxonomic characters to the entire order of the Diptera and checked the world-wide material in the collection of the Zoological Museum at Helsinki University.

2. Material and methods

A study was made of dried specimens in the collections at the Zoological Museum of Helsinki University. 875 species belonging to 128 families (see Appendix 1) were examined. The species selected for the study were chosen to represent as many subfamilies, families and other higher taxa as possible. Species with an indistinct or isolated taxonomic position, or with spectacular morphological characters or unusual living habits, were also included. Species were also chosen to represent faunae from different zoogeographic regions. When a random sample of species from a taxon revealed dissimilarities, further material was examined.

The chaetotaxy of the frontal edge of the costal vein was studied in the long middle part of the costa, while the most proximal part, with a variable pattern (e.g. strong specialized spines near the costal breaks of some species, or irregularly arranged basal hairs in species with regular rows along most of the costal edge), and the distal part, with frequently reduced chaetotaxy, were omitted from the descriptions, unless otherwise stated. The "dorsal" and "ventral" rows mentioned in the text below refer to the rows of the frontal edge situated near the upper and lower surfaces of the wing.

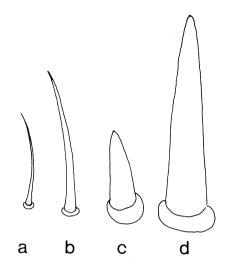


Fig. 1. Types of setae on the costal vein in the Diptera: a = hair, b = bristle, c = spinula, d = spine.

The chaetotaxy of the costa cannot be satisfactorily seen in a dorsal (or ventral) view of the wing and has therefore earlier been overlooked. In the case of larger-sized Diptera the chaetotaxy can be studied under an ordinary stereomicroscope with a strong lamp focused on the insect. Smaller Diptera were observed with a light microscope with fibre lights attached.

A few selected specimens were studied under the scanning electron microscope at the Department of Electron Microscopy of the University of Helsinki.

3. Types of setae and their arrangement on the costa

The setae (macrotrichia) are mostly confined to the costa in the "higher" Diptera, and they are hardly ever present on the wing membrane. The setae can be classified as follows (as proposed by Prof. R. Tuomikoski) (Fig. 1):

1. Hairs: gradually tapering to a very fine tip, and usually slightly curved towards the wing apex.

2. Bristles: similar to the hairs but longer and/or coarser. Intermediate types between these and hairs are commonly encountered.

- 3. Spinulae: stouter than the hairs and often also shorter owing to the attenuated tip, more abruptly tapering to an almost blunt apex, and typically appearing as a dense comb-like subdorsal row.
- 4. *Spines*: similar to spinulae, but longer and more widely spaced, often alternating with hairs in one ventral row.

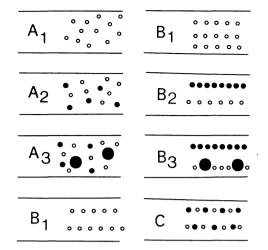


Fig. 2. Types of the costal chaetotaxy in the Diptera. — Open circle = hair or bristle, small dot = spinula, large dot = spine. The same symbols are used in Fig. 3.

5. Recurved sensory setae: very fine, solitary and widely spaced, with more or less erect base and recurved tip.

Comparable to the macrotrichia are the "pores" (campaniform sensillae), ring-like formations probably homologous to the sockets of macrotrichia. However, these hardly ever occur on the costa, though two of them may be found dorsally on the tip of R1 rather close to the costa. Only hairs, bristles, spinulae and spines are systematically considered below.

The arrangement of the setae on the costa can be classified into the following basic types (A and B types modified from Prof. R. Tuomikoski's proposal) as follows (Fig. 2):

- A setae irregularly arranged (or in several indistinct rows)
 - A₁ setae homomorphous (hairs or bristles)
 - A₂ setae dimorphous: hairs (or bristles) and spinulae
 - A₃ setae trimorphous: hairs, spinulae and spines
- B setae in two, seldom three, longitudinal rows, each row with homomorphous setae
 B₁ setae homomorphous (hairs or bristles)
 - B₂ setae dimorphous: a subdorsal row of spinulae and a subventral row of hairs or bristles

- B₃ setae trimorphous: a subdorsal row of spinulae, subventral rows of hairs (or bristles) and spines (sometimes hairs and spines almost in the same row
- C setae in two longitudinal rows, each row with dimorphous setae, hairs or bristles alternating with spinulae (additional hairs or bristles may occur).

The above classification is used in the text below with comments on any possible minor variation (e.g. additional dorsal and ventral rows of hairs). However, the study of this extensive material also revealed several additional types which are described separately.

4. Costal chaetotaxy in the superfamilies of the Diptera

4.1. The "nematocerous" superfamilies

As can be seen in Appendix 2 the species examined in the superfamilies Tipuloidea (Figs. 3a, 4), Blepharoceroidea, Psychodoidea, Tanyderoidea, Culicoidea, Thaumaleoidea, Anisopodoidea, Pachyneuroidea. Bibionoidea, Scatopsoidea, Mycetophiloidea and Cecidomyoidea all belong to type A₁ respecting their costal chaetotaxy. In this connection, it may be mentioned that Panorpa communis L., which was examined for comparison purposes as a representative of the Mecoptera, an insect order related to the Diptera, was also found to be of type A_1 (Fig. 5). It must be pointed out, however, that among the nematocerous families the setae of the costa are of a modified type in the Psychodidae (Psychodoidea; Figs. 6-7) and are represented by more or less narrow scales in the Chaoboridae and Culicidae (Culicoidea; Fig. 8). In Dixa borealis (Dixidae) and in the species examined of the genera Leia, Exechia and Dynatosoma (Mycetophilidae) there seems to be a tendency towards the formation of rows of hairs (i.e. of more than two rows).

In Nymphomyia (Nymphomyiidae) the costa is reduced and the macrotrichia of the front margin of the wing are of the same kind as those on the hind margin, being crossed pairs of long, fine hairs.

Among the Chironomoidea the single ceratopogonid species examined is of type A₁. In the Simuliidae, species of *Helodon* and *Prosimulium* are of type A₁, but species of



Fig. 21. The costal chaetotaxy of Lyciella decempunctata (Lauxaniidae). SEM.

4.19. Opomyzoidea

In this superfamily the species examined are mostly of type B₂ with minor variations. Species of the family Lonchaeidae belong to this type but in the most distal parts of the costa type B₁ may be represented (*Dasiops latifrons*). In some Clusiidae there are some ventral hairs in addition to the normal B₂ pattern (*Clusia flava*). Species of the family Acartophthalmidae are of type B₂, both rows being homomorphous and consisting of relatively poorly differentiated spinulae/hairs. The most conspicuous exception in the superfamily Opomyzoidea is the family Agromyzidae in which all the species belong to type B₁, the rows consisting of slender hairs only.

4.20. Asteioidea

All the families of the Asteioidea, judging from the relatively few species examined, belong in general to type B₁, but in the Anthomyzidae there is some variation. Anthomyza, Ischnomyia and Mumetopia have two rows of long fine hairs, but in the dorsal row there are also some slender spinulae among the hairs. In Stenomicra only hairs occur in the two rows.

4.21. Lauxanioidea

Species of the family Lauxaniidae distinctly belong to type B_2 (Figs. 3h, 21). The dorsal row of spinulae is dense and comb-like. Numerous Finnish species of various genera

examined by Tuomikoski (unpubl.) all exhibit this type of costal chaetotaxy. The Celyphidae species examined belong to type B₁. Species of the family Chamaemyiidae belong to a type intermediate between B₁ and B₂ the "spinulae" of the subdorsal row being not much thicker than the setae of the subventral row.

4.22. Drosophiloidea

With much hesitation we have considered this superfamily in a wider sense (cf. Hennig 1958, 1971, Griffiths 1972).

The Drosophilidae species are of type B₂, with a dense subdorsal row of spinulae. The Curtonotidae and Camillidae species belong to type B₃, with a subdorsal row of spinulae and a subventral row of hairs and some spines sparsely among these hairs. The Diastatidae represent type B₃, with subventral spines only in the basal third of the costa (spines entirely missing in *Odiniomorpha*).

Among the subfamily Psilopinae of the Ephydridae the small Atissa species belong to type B₁, the other species investigated to type B₂. In the Notiphilinae Notiphila and Ilythea belong to type B₂. Hydrellia griseola is almost of type B₁, having only a few spinulae in the subdorsal row. Dichaeta in the subdorsal row has spinulae of two sizes, the subventral row consisting of slender hairs only. The Parydrinae are of type B₁ or nearly so (Pelina has some subdorsal spinulae among the hairs). Among the Ephydrinae, Lamproscatella, Paracoenia (almost), Scatella and Setacera (with a somewhat irregular subventral-ventral row of hairs) are of type B₂, Philotelma approaching

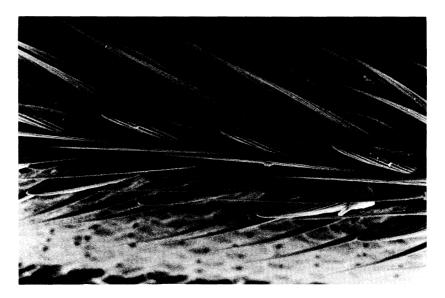


Fig. 22. The costal chaetotaxy of *Scathophaga stercoraria* (Scathophagidae). SEM.

B₁. Ephydra is of a special type. There are some longer spinulae among the subdorsal row of ordinary spinulae, and in the subventral row of hairs there are some widely spaced spines, thus representing a variant of type B₃.

Cryptochaetidae species belong to type B_1 or nearly so (*Cryptochaetum aenescens* with additional hairs in the apical part of the costa). Again, all the Carnidae, Tethinidae and Chloropidae (including the Siphonellopsidae sensu Nartshuk 1983; see also Andersson 1977) are of type B_1 .

The Milichiidae belong to type B₂ as do the Canaceidae species examined, but in the latter family the subforsal row of spinulae is sparse and less distinct. In *Canace actites* there is an additional median row of short hairs.

4.23. Muscoidea

This superfamily is considered here in a much stricter sense than in Griffiths (1972) or even McAlpine (1979), thus containing the families Scathophagidae, Anthomyiidae, Eginidae, Muscidae, Fanniidae, Calliphoridae, Sarcophagidae, Rhinophoridae, Phasiidae and Tachinidae.

The Scathophagidae are of types A₁, A₂ and their intermediates (sometimes also indistinct rows of spinulae may occur). In the Scathophaginae *Cochliarium* and *Gimnomera* (if

correctly classified) belong to type A_1 , and Norellia and Norellisoma approach (or represent) that type; the rest of the genera examined belong to type A_2 (Fig. 22). The Delininae seem to be predominantly of type A₁. In *Phrosia* there are sometimes a few spinulae among the irregularly arranged hairs (A_2) . In *Micropselapha* there is apically a tendency towards the formation of rows. Hexamitocera loxocerata represents a unique case among the Diptera material examined: a case of apparent sexual dimorphism in the costal chaetotaxy. In the 32 females examined the costal chaetotaxy is of type A₂ with many irregularly arranged spinulae among the hairs, whereas in the 24 males it is of type A_1 (Fig. 23).

Most of the Anthomyiidae species clearly belong to type C, with hairs or bristles usually alternating with spinulae in heteromorphous subdorsal and subventral rows. In Fucellia there is an additional sparse row of strong ventral spinulae. In Pegomya and Hylemya there are more or less irregularly arranged additional ventral setae. All these can easily be regarded as belonging to type C, the variation only being due to additional setae. More aberrant chaetotaxy patterns are found in the genera Monocrotogaster and Eustralomyia. These have a dorsal row of fine hairs, subdorsal and subventral rows of spinulae and a ventral row of fine hairs, and in Eustalomyia irregularly arranged ventral hairs as well.

which occurs in numerous genera of the other Sciomyzoidea and only in one genus of the Diopsidae among the Nothyboidea.

The Rhopalomeridae show the biseriate type of ordinary hairs similar to that of the Micropezoidea and most of the Nothyboidea, and the Sepsidae and Megamerinidae among the Sciomyzoidea. Although the Rhopalomeridae are very unlike the typical Sepsidae, they have a remarkable similarity with the Sepsidae in possessing quite similar metastigmatical bristles, an apomorphy which seems to be unknown in the other families of the so-called Acalyptrata.

We have followed J. F. McAlpine's (1979) example and included in the superfamily Heleomyzoidea the families Heleomyzidae, Trixoscelidae, Chyromyidae and, with some hesitation, the Sphaeroceridae. Moreover, we have also added here the families Borboropsidae and Rhinotoridae. Griffiths (1972) combined the Heteromyzidae with the Tanypezidae to form a prefamily Tanypezoinea in his superfamily Muscoidea. We have classified the "Heteromyzidae", however, as a subfamily of the Heleomyzidae, in which the genera of this taxon were included prior to 1972. Costal chaetotaxy type B_3 occurs both in the Heteromyzinae and in the other Heleomyzidae. The Sphaeroceridae, with their primitive types A_1 and B_1 , seem to have an enigmatic systematic position, but they obviously fit better into the Heleomyzoidea than into the Drosophiloidea s. lat.

Among the Heleomyzidae the genus *Orbellia* represents an exceptionally primitive type of costal chaetotaxy (A₁-A₂). It is not, however, possible to say if this is a plesiomorphous condition, or the result of a reduction when compared with the B₃ type of the other Heleomyzidae. It may be mentioned that *Orbellia* shows the most plesiomorphous conditions of the male postabdomen known among the Heleomyzidae: in most Heleomyzidae the seventh and eighth tergum vestiges have been lost, but both are well developed in *Orbellia* (Griffiths 1972). The possible relationship of *Orbellia* to the Borboropsidae should be studied.

Among the unplaced groups the genus *Notomyza* was classified in the Heleomyzidae by Malloch (1933) in spite of the lack of costal spines. Griffiths (1972) described the family Notomyzidae but did not place them in any superfamilies. The family Chiropteromyzidae was positioned by Griffiths in his Tephri-

toinea before the Mormotomyiidae (not examined; possibly fit better near the Muscoidea or Glossinoidea) and Cnemospathidae. costal chaetotaxy of the Chiropteromyzidae could indicate a relationship with the Heleomyzoidea. The genus Listromastax was placed by Griffiths (1972) in the Tethinidae but it does not well fit there. The costal chaetotaxy could indicate a relationship with the Heleomyzoidea. Borboroides is one of the unplaced genera referred to the Heleomyzidae (cf. Richards 1973) but does not appear to belong there. The genus Dichrochira has been tentatively placed by Hennig (1971) in the Heleomyzidae but being of type B₁, like Borboroides it hardly belongs there.

In the catalogue of afrotropical Diptera (Crosskey 1980) the superfamily Opomyzoidea includes the families Lonchaeidae, Piophilidae, Opomyzidae, Clusiidae, Odiniidae and Agromyzidae. However, we think that the Piophilidae fit better in the Otitoidea, where J. F. McAlpine placed them in his monograph (1977). We have also added here the Acartophthalmidae. In the Opomyzoidea type B₂ may form the general pattern. In such small flies as the Agromyzidae type B₁ may have secondarily developed from B₂.

Among the Asteioidea, which are predominantly of type B₁, the Anthomyzidae appear to constitute a slight exception, having some dorsal spinulae. The genus *Stenomicra* has been tentatively placed by Hennig (1971) in the Aulacigastridae but Griffiths (1972) retains it in the Anthomyzidae. Its type B₁ seems to lend support to Hennig's view.

The Lauxanioidea appears to be a relatively homogenous superfamily in terms of costal chaetotaxy. The Celyphidae, included by Griffiths (1972) in the Lauxaniidae, are retained here as a separate family, as was done by Miller (1980). Miller points out that the Celyphidae are lacking in the costal spinulae ("setulae") typical of the Lauxaniidae. The Lauxanioidea, with type B₂ as their general pattern (at least in the Lauxaniidae), and lacking the swinging mechanism of the aedeagus, might be related to the Opomyzoidea, where the swinging mechanism has also been lost in the case of two families (Lonchaeidae and Odinidae).

The families Curtonotidae, Drosophilidae, Camillidae, Diastatidae (including Griffiths' Campichaetidae and the genus *Diastata*, placed by him in the Ephydridae) and Ephydridae are the members of the Droso-

philoidea in a stricter sense (see Hennig 1958, 1971, Griffiths 1972). The family Cryptochaetidae is probably rather closely related to these families (D. K. McAlpine 1982). In a wider sense (see also Crosskey 1980, and further Speight 1969 and J. F. McAlpine 1979) the families Milichiidae, Carnidae, Braulidae Canaceidae (wingless), Tethinidae. and Chloropidae (including the Siphonellopsinae; cf. Nartshuk 1983) are included here. In addition, the Risidae and Tunisimviidae can be accommodated here (see Papp 1977, 1980). We admit that the Drosophiloidea in its widest sense is a taxon containing heterogenous elements. The Drosophiloidea of Hennig (1958, 1971) and Griffiths (1972) is most probably monophyletic and the type B₂ costal chaetotaxy obviously figures in their general plan. We have, however, considered it practical to include here other families as well: if they are removed, it is doubtful whether they form a monophyletic superfamily. The position of the Carnidae is unclear and a closer relationship with the Milichiidae is uncertain. The Tethinidae and Canaceidae are probably related to each other. It has not been demonstrated whether the Miliichiidae and Chloropidae (s. lat.) form a monophyletic entity. To the Milichiidae was also referred the genus Australimyza Harrison from Australia, New Zealand and some subantarctic islands (Harrison 1959), but Griffiths (1972) placed the genus in a family of its own, the Australimyzidae ("prefamily Australimyzoinea"), near the Nothyboidea. We have unfortunately not seen any material of this taxon. According to Hardy (1980), Australimyza belongs to the Xenasteiidae (Asteioidea).

6.4. The roots of the Calyptratae

Where the calyptrate Diptera have their origin in the system of the Schizophora is difficult to elucidate. The Scatophagidae have two types of costal chaetotaxy, A₁ and A₂. The type A₁ of most of the Delininae could be a primitive feature but there is also the possibility that it has arisen from the A₂ of the Scathophaginae by the loss of the sometimes very sparsely arranged spinulae. A well-developed swinging mechanism of the aedeagus was found by author Hackman in numerous species of the Scathophagidae and demonstrated from fresh material of *Pogonota barbata* (see Hackman 1956: Figs. 1-3). In all

the other families included here in the superfamily Muscoidea (s. str.) type C is a basic feature (a probable synapomorphy) and the different aberrant types can be easily derived from C. Thus, there is a considerable gap between the Scathophagidae and Anthomyiidae as regards costal chaetotaxy. The Scathophagidae were earlier included as a subfamily of the Anthomyiidae (see Roback 1951), but have recently been considered a separate family by most authors. It seems possible that the Scathophagidae represent the plesiomorphous state of the costal chaetotaxy in the Muscoidea.

The Oestroidea have surprisingly primitive looking types of costal chaetotaxy, viz. with a few exceptions variations on type A₁. It cannot be denied that in the oestroid families there are features of the costal chaetotaxy very similar to those of some large Asiloidea, Xylophagoidea and Tabanoidea (such as Pantophthalmidae and Mydidae): there are often upright hairs or setae near the dorsal margin of the costa, a bare subdorsal area and (median-)subventral-ventral irregularly inserted hairs or setae directed more or less along the costa.

Hennig (1971) pointed out that there is no doubt that the calyptrate Diptera form a monophyletic group and this opinion is also shared by Griffiths (1972), who includes the Oestroidea (as limited here) in the Tachinidae. Considering the types of costal chaetotaxy, there appears to be a considerable gap between the Muscoidea and Oestroidea. The gap is, however, bridged by the genus *Pharyngomyia* (Oestridae), and possibly there are more species with that type of costal chaetotaxy among the oestrid genera related to Pharyngomyia. According to Grunin (1966), Pharyngomyia belongs to the Cephenomyiinae, but its costal chaetotaxy would suggest a closer relationship with the Oestrinae. The seemingly primitive type A₁ of many Oestroidea has been found among the Muscoidea in the Scathophagidae (a dissimilar variation of A_1).

The type C costal chaetotaxy found in the genus *Pharyngomyia* may also be a result of evolution paralleling that of the Muscoidea. There even exists the possibility that the type C of the Oestridae (*Pharyngomyia*) could have developed from the type A₁ occurring in the Cuterebridae as well as in the "lower" Muscomorpha, and that the type C of the Muscoidea could have developed from the oestroid C. In this case the type C would have been subsequently transformed to the primi-

tive type A_2 and A_1 of the Scathophagidae. It seems to us unwarranted to derive the Oestroidea (with very primitive costal pattern) from the muscoid families. Especially the Cuterebridae appear to be primitive in several respects among the cyclorrhaphous Diptera: in their karyotypes (Boyes 1963), well-formed mouthparts resembling those of male tabanids, the presence of large alulae and two or three anal veins, and in the prothoracic spiracles of their puparia being the functional respiratory system (as in several "orthorrhaphous" flies), as well as in the paleontology of the family (Bennett in Zumpt 1957). The Cuterebridae have been derived from the Calliphoridae but their type of costal chaetotaxy does not support this view. Thus, in spite of their well-developed mouthparts they seem to fit better into the Oestroidea as limited here.

The Glossinidae, being of type A_2 , form together with the Hippoboscidae, Streblidae and the wingless Nycteribiidae the superfamily Glossinoidea. The Hippoboscidae and Streblidae are of types A_1 and B_1 but these types may have developed from A₂ by the loss of the spinulae. Hennig (1971) has pointed out that there is no doubt that the Glossinoidea form a monophyletic group, all of them producing larvae ready to pupate (see also Griffiths 1976). The swinging mechanism of the aedeagus occurs in the Glossinidae but is obviously secondarily lost in the other glossinid families. The relationship between the Glossinoidea and the other Calyptratae remains unclear.

6.5. Concluding remarks

It is apparently still premature to attempt to construct a dendrogram of the Schizophora, and there are even questions of the composition of superfamilies where the available data are undoubtedly inadequate. In the use of costal chaetotaxy in the classification of the Diptera it is often not possible to judge where a certain type represents an apomorphous or a plesiomorphous character state. In this extensive study comparatively few species of large families were examined and we hope this will encourage specialists to check their own groups more intensively and to study more details of the costal chaetotaxy. After the approximately two hundred years of scientific dipterology when the wing characters have always provided corner stones for classification the time would appear to be ripe for widening the one-sided approach and for viewing the wing from the side as well.

Acknowledgements. We are most grateful to Professor Risto Tuomikoski for encouragement and for the initial idea of studying this character. Dr S. Panelius and Professor M. Meinander are thanked for their valuable comments on the manuscript. Our joint project profited also from the comments on the Chironomidae by Dr B. Lindeberg. We are also indebted to Dr R. Rosengren for technical help in photography. R. Väisänen's studies were supported by a grant from the University of Helsinki. The names of the authors appear in alphabetical order.

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Appendices

Appendix 1. A list of the dipterous species examined. Finland is abbreviated to F.

TIPULOIDEA

Tipulidae: Tipulinae: Nephrotoma crocata (L.) (F), N. pratensis (L.) (F), Tipula maxima Poda (F), T. nubeculosa Meigen (F), T. scripta Meigen (F). — Cylindrotominae: Cylindrotoma distinctissima (Meigen) (F). — Limoniinae: Limonia quadrimaculata (L.) (F), Pedicia rivosa (L.) (F).

Trichoceridae: Trichocera lutea Becker (Spitzbergen), T. maculipennis (Siebke) (F), T. regelationis (L.) (F).

NYMPHOMYOIDEA

Nymphomyiidae: Nymphomyia alba Tokunaga (Japan).

BLEPHAROCEROIDEA

Blepharoceridae: Liponeura cinerascens Loew (Italian Alps).

PSYCHODOIDEA

Psychodidae: Psychodinae: Clytocerus ocellaris (Meigen) (F), Psychoda alternata Say (F), Satchelliella mutua (Eaton) (F), Ulomyia fuliginosa (Meigen) (F).

TANYDEROIDEA

Ptychopteridae: Bittacomorphella jonesi (Johnson) (USA: Michigan), Ptychoptera contaminata (L.) (F), P. paludosa Meigen (F).

CULICOIDEA

Dixidae: Dixa borealis Martini (F), D. sp. (Burma), Dixella aestivalis (Meigen) (F).

Chaoboridae: Chaoborus chrystallinus (De Geer) (F). Culicidae: Aedes annulipes Meigen (F), A. communis (Harris) (F); Melanostomini: Melanostoma dubium (Zetterstedt) (F); Bacchini: Baccha elongata (Fabricius) (F): Chrysotoxini: Chrysotoxum arcuatum (L.) (F): Paragini: Paragus tibialis (Fallén) (F). — Milesiinae: Pipizini: Pipiza austriaca Meigen (F), P. festiva Meigen (F), Pipizella virens (Fabricius) (F), Triglyphus formosanus Shiraki (Burma); Cheilosini: Chamaesyrphus lusitanicus Mik (F), Cheilosia gagatea Loew (Germany), C. vicina (Zetterstedt) (F), Ferdinandea cuprea (Scopoli) (F), Pelecocera tricincta Meigen (F); Brachyopini: Brachyopa cinerea Wahlberg (F), B. pilosa Collin (F); Volucellini: Graptomyza microdon Osten Sacken (Philippines), Volucella inanis (L.) (F); Merodontini: Eumerus ruficornis (Meigen) (F), Merodon clavipes (Fabricius) (Greece), M. equestris (Fabricius) (F); Ceriodini: Ceriana conopsoides (L.) (F); Eristalini: Eristalis abusivus Collin (F); Milesini: Sphecomyia vespiformis Gorski (F), Spilomyia diophthalma (L.) (F); Xvlotini: Xvlota sylvarum (L.) (F), X. tarda Meigen (F), Chalcosyrphus eumerus (Loew) (USSR: European NW); Sericomyiini: Sericomyia lapponica (L.) (Norway), S. silentis (Harris) (USSR: European NW). - Microdontinae: Microdontini: Microdon devius (L.) (F), M. eggeri Mik (F).

CONOPOIDEA

Conopidae: Conopinae: Brachyglossum coronatum Rondani (Czechoslovakia), Conops bakeri Kröber (Philippines), C. excisus Wiedemann (USA: Kansas), C. flavifrons Meigen (Hungary), Physocephala rufipes (Fabricius) (Germany, FRG). — Zodioninae: Zodion americanum Wiedemann (Canada: Quebec), Z. cinereus (Fabricius) (Hungary), Z. fulvifrons Say (USA: Kansas), Z. sp. (Brazil), Thecophora fulvipes (Robineau-Desvoidy) (Germany, GDR). — Stylogastrinae: Stylogaster neglecta Williston (USA: New York). — Dalmanniinae: Dalmannia marginata (Meigen) (Spain), D. vitiosa Coquillett (USA: Illinois). — Myopinae: Melanosoma bicolor (Meigen) (Czechoslovakia), M. buccata (L.) (Germany, GDR), M. morio (Meigen) (Spain), Sicus ferrugineus (L.) (F).

MICROPEZOIDEA

Cypselosomatidae: Aureomyza ignipennis Frey (Burma), Formicosepsis sp. (Burma), Rhinopomyzella nigrimana Hennig (Jamaica).

Neriidae: Telostylinae: Chaetonerius inermis (Schiner) (Burma), Telostylus maccus Osten Sacken (Philippines).

— Neriinae: Longina abdominalis (Wiedemann) (Paraguay), Nerius plurivittatus Bigot (Brazil).

Micropezidae: Taeniapterinae: Glyphodera mantis Enderlein (Sierra Leone), Grallipeza imbecilla (Enderlein) (Brazil), Taeniaptera strigata (Enderlein) (Brazil). — Calobatinae: Calobata petronella (L.) (F). — Micropezinae: Micropeza corregiolata (L.) (F).

NOTHYBOIDEA

Nothybidae: Nothybus biguttatus van der Wulp (Indonesia), N. kempi (Brunetti) (Laos), N. longithorax Rondani (Indonesia), N. triguttatus Bezzi (Phillipines).

Tanypezidae: Tanypezinae: Neotanypeza nigripalpis Hendel (Brazil), N. sp. (Brazil), Tanypeza luteipennis Knab & Shannon (Canada: Quebec). — Strongylophthalmyiinae: Strongylophthalmyia curvinervis Frey (Burma), S. punctum (Burma).

Psilidae: Chyliza nobilis Frey (Burma), Loxocera ichneumonea (L.) (F), Psila audoini (Zetterstedt) (F), P. gracilis Meigen (Hungary), P. merdaria Collin (F).

Diopsidae: Centrioncus prodiosis Speiser (Zimbabwe), Eurydiopsis subnotata Westwood (Philippines), Diopsis indica Westwood (Indonesia), D. thoracica Westwood (Tanzania), Teleopsis notatrix Osten Sacken (Philippines)

Teratomyzidae: Teratomyza sp. (Burma).

Periscelididae: Microperiscelis annulata (Fallén) (F), Periscelis annulipes Loew (F), P. nigra (Zetterstedt) (F).

Somatiidae: Somatia sophiston Steyskal (Trinidad), S. xanthomelas Schiner (Brazil).

OTITOIDEA

Otitidae: Otitinae: Delphinia picta (Fabricius) (USA: Pennsylvania), Dorvcera maculipennis Meigen (Greece: Lesbos), Herina tristis (Meigen) (Atlas Mts.), Meliera obscuripes Loew (F), Otites formosa Panzer (Hungary), Seioptera vibrans (L.) (F), Tetanops ferdinandi Frey (USSR: Siberia), T. maroccana Frey (Morocco), T. sintenisi Becker (F). — Pterocallinae: Neomyennis appendiculata (Hendel) (Paraguay), Pterocalla sp. (Brazil), Terpnomyia sp. (Paraguay). — Ulidiinae: Euphara coerulea (Macquart) (Brazil), Euxesta eluta Loew (Brazil), Homalocephala bimaculata Wahlberg (F), Physiphora aenea (Fabricius) (Taiwan), P. africana (Hendel) (East Africa), P. demandata (Fabricius) (Cape Verde Isl.), P. longicornis (Hendel) (Taiwan), P. smaragdina (Loew) (Cape Verde Isl.), Timia klugi Hendel (USSR), Ulidia apicalis Meigen (Atlas Mts.), U. erythrophthalma Meigen (Morocco).

Platystomatidae: Trapherinae: Aglaioptera incomparabilis Frey (Burma), Lule stellata Enderlein (Cameroon), Phasiamyia metallica Walker (Indonesia), Piara chrysoptera Frey (Chapa, ? in Burma), Poecilotraphera taeniata (Macquart) (Malaysia), Xiria obliqua Osten Sacken (Philippines). - Platystomatinae: Achias australis Malloch (Australia), A. latidens Walker (New Guinea), Antineura stolata Osten Sacken (Philippines), Brommophila caffra (Macquart) (S Africa), Elassogaster anteapicalis Hendel (Burma), Euprosopia chalybea Frey (Philippines), E. grahami Malloch (Japan), E. trivittata Bezzi (Philippines), Loxoneura pictipennis (Walker) (Burma), Peltacanthina simillima Hendel (Malawi), Platystoma lugubre Robineau-Desvoidy European W), P. seminationis L. (F), Rivellia syngenesiae (Fabricius) (F), Xenaspis pictipennis (Walker) (India). — Plastotephritinae: Plastotephritis gratiosa Enderlein (W Africa). — Scholastinae: Naupoda platessa Osten Sacken (Philippines), Pterogenia valida Bezzi (Philippines), Zygaenula paradoxa Doleschall (Philippines).

Pyrgotidae: Apyrgota scioidea Hendel (Laos), Lachnostylia sp. (Colombia), Leptopyrgota sp. (Brazil), Pyrgota undata Wiedemann (USA: Pennsylvania), P.

valida (Harris) (USA: Pennsylvania). Tephritidae: Dacinae: Callantra longicornis (Wiedemann) (Indonesia), Dacus furcatus Wiedemann (S Africa), D. tau (Walker) (Philippines), D. umbrosus (Fabricius) (Phillippines). - Urophorinae: Hypenidium nowacki Strobl (Cyprus), Myopites nigrescens Becker (Canary Isl.), Parahypenidium polyfasciatum (Miyake) (Japan), Urophora solstitialis (L.) (F). — Adraminae: Adrama determinata (Walker) (Burma), Meracanthomyia kotiensis Kapoor (Burma), Munromyia nudiseta Bezzi (S Africa), Pseudosophira bakeri Malloch (Philippines). — Euphrantinae: Euphranta connexa (Fabricius) (F). — Trypetinae: Bezzi (India), Enicoptera Anoplomus flexuosus sumatrana Hering (Indonesia), Orellia falcata (Scopoli) (USSR: Turkestan), Terellia serratulae (L.) (France), Trypeta zoe (Meigen) (F). — Ceratitinae: Ceratitis capitata (Wiedemann) (F. introduced with oranges). — Acanthoneurinae: Acanthoneura trigona Matsumura (Japan), Diarrhegma modestum (Fabricius) (Philippines), Hexacinia pellucens Hardy (Philippines), lunifera (Philippines), T. maculipennis (Westwood) (Indonesia), T. ostensackeni Hardy (Philippines), Ortalotrypeta isshikii Matsumura (Japan), Rioxa sexmaculata (van der Wulp) (Philippines), Xarnuta leucotelus Walker (Philippines). — Aciurinae: Aciura coryli Rossi (Yugoslavia), Oxyaciura tibialis Robineau-Desvoidy (Cape Verde Isl.), Spathulina tristis Loew (Canary Isl.). — Tephritinae: Plastensini: Plastensina sp. (Philippines); Euarestini: Euaresta comma Banks (USA: Texas); Tephrellini: Tephrella caloptera Loew (USSR: Dauria); Tephritini: Campiglossa borealis (Portschinsky) (F), C. grandinata Rondani (Yugoslavia), Camaromyia bullans Wiedemann (? Poland), Oedosphenella canariensis (Macquart) (Canary Isl.), Paroxyna martii Becker (Canary Isl.); Xyphosini: Xyphosia miliaria (Schrank) (F).

Tachiniscidae: Tachinisca cyaneiventris Hendel (Colombia).

Richardiidae: Automola atomaria (Wiedemann) (Bolivia), Hemixantha pulchripennis Hendel (Brazil), Richardia podagrica (Fabricius) (Brazil), Sepsidosoma sp. (Brazil).

Pallopteridae: Heloparia bicolor Walker (Patagonia), Palloptera (Palloptera) arcuata (Fabricius) (Italy), P. (P.) formosa Frey (F), P. (P.) terminalis Loew (Alaska), P. (P.) ustulata Fallén (F), P. (Temnosira) saltuum (L.) (F), P. (Toxoneura) muliebris (Harris) (Corsica), P. (Alasia) ambusta (Meigen) (F), P. (A.) canypta Czerny (Germany, GDR), P. (A.) aphippium Zetterstedt (F), P. (A.) laetabilis Loew (F), P. (A.) septentrionalis Czerny (USSR: European NW), P. (A.) superba Loew (USA: Illinois), P. (A.) trimacula Meigen (F), P. (A.) umbellatarum (Fabricius) (F), P. (A.) usta Meigen (F).

Eurygnathomyiidae: Eurygnathomyia bicolor (Zettersted) (F).

Piophilidae: Neottiophilinae: Neottiophilum praeustum Meigen (Germany, GDR). — Piophilinae: Amphipogon flavus (Zetterstedt) (F), Piophila casei (L.) (F), Mycetaulus bipunctatus (Fallén) (F).

SCIOMYZÓIDEA

Coelopidae: Coelopa frigida (Fabricius) (F), C. pilipes Haliday (Spain), Malacomyia sciomyzina (Haliday) (Canary Isl.), Heterocheila buccata (Fallen) (Sweden).

Dryomyzidae: Dryomyza anilis Fallén (F), D. flaveola (Fabricius) (F), D. formosa Wiedemann (Japan), Helcomyza ustulata Curtis (Germany, GDR).

Sciomyzidae: Phaeomyiinae: Pelidnoptera fuscipennis (Meigen) (F). — Sciomyzinae: Antichaeta analis (Meigen) (F), A. atriseta (Loew) (F), Atrichomelina pubera (Loew) (Canada: Quebec), Ditaenia seticosta Steyskal (USA: Michigan), Pherbellia ventralis (Fallén) (F), Pteromicra angustipennis (Staeger) (F), Sciomyza dryomyzina Zetterstedt (F), S. simplex (Fallén) (F), Tetanocera arrogans (Meigen) (F), Tetanura pallidiventris Fallén (F), Sepedon spinipes (Scopoli) (F), S. sphegeus (Fabricius) (F).

Sepsidae: Australosepsis frontalis (Walker) (Philippines), Orygma luctuosa Meigen (Norway), Saltella spondylii (Schrank) (F), Sepsis punctum (Fabricius) (F), Themira annulipes (USSR: European NW), Toxopoda contracta (Walker) (Burma).

Rhopalomeridae: Rhopalomera clavipes (Fabricius) (Brazil), R. femorata (Fabricius) (Brazil), Willistoniella pleuropunctata (Wiedemann) (Bolivia).

Megamerinidae: Megamerina loxocerina (Fallén) (F), Texara dioctrioides Walker (Taiwan).

HELEOMYZOIDEA

Heleomyzidae: Heteromyzinae: Heteromyza atricornis (Meigen) (Netherlands), H. oculata Fallén (Germany, GDR), Tephrochlamys flavipes (Zetterstedt) (F), T. ufiventris (Meigen) (F), T. tarsalis (Zetterstedt) (Germany). — Suillinae: Suillia laevifrons (Loew) (F), S. humilis (Meigen) (Germany, FRG), S. ustulata (Meigen)

(Italy), S. similis (Meigen) (Netherlands), Allophyla laevis Loew (Canada: Ontario). — Heleomyzinae: Diplogeomyza media D. K. McAlpine (Australia: New South Wales), Heleomyza serrata (L.) (F), Oecothea fenestralis (Fallén) (F), Orbellia nivicola Frey (F), O. tokyoensis Czerny (Japan), Scoliocentra nigrinervis (Wahlgren) (F), S. villosa (Meigen) (England).

Borboropsidae: Borboropsis puberula (Zetterstedt)

Trixoscelidae: Trixoscelis laeta (Becker) (Morocco), T. sabulicola Frey (Canary Isl.).

Chyromyiidae: Chyromyia flava (L.) (F), C. oppidana (Scopoli) (F).

Sphaeroceridae: Copromyzinae: Copromyza (Crumomyia) notabilis Collin (F), C. (Fungobia) nitida (Meigen) (F), C. (Alloborborus) pallifrons Fallén (F), C. (Borborillus) fumipennis (Stenhammar) (F), C. (Olinea) atra (Meigen) (F), C. (Gymnometopina) clunicrus (Duda) (Tanzania). — Sphaerocerinae: Ischiolepta vaporariorum (Haliday) (F), Lotobia pallidiventris (Meigen) (F), Sphaerocera curvipes Latreille (F). - Leptocerinae: Ceroptera rufitarsis Meigen (Morocco), Chaetopodella curzoni (Richards) (Zimbabwe), Kimosina empirica Hutton (Germany, GDR), Leptocera (Leptocera) fontinalis (Fallén) (F), L. (Opacifrons) coxata (Stenhammar) (Azores), L. (O.) humida (Haliday) (Austria), L. silvatica (Meigen) (F), Poecilosomella angulata (Thomson) (Cape Verde Isl.), P. giraffa (Richards) (Nigeria), Thoracochaeta zosterae (Haliday) (USSR: Far East).

UNPLACED FAMILIES AND GENERA

Notomyzidae: Notomyza edwardsi Malloch (Tierra del Fuego).

Chiropteromyzidae: Chiropteromyza wegelii Frey (F), Neossos marylandica Malloch (Canada: Quebec), N. nitidicola (Frey) (F).

Genus Borboroides: B. atra Malloch (Australia).

Genus Listromastax: L. littorea Enderlein (Crozet Isl.).
Genus Dichrochira: D. sanguiniceps Wiedemann (Paraguay).

OPOMYZOIDEA

Lonchaeidae: Dasiops latifrons (Meigen) (Hungary), D. procera Morge (F), Lamprolonchaea aurea (Macquart) (Canary Isl.), Lonchaea zetterstedti Becker (F).

Opomyzidae: Anomalochaeta guttipennis (Zetterstedt) (F), Geomyza advena Frey (Japan), G. tripunctata (Fallén) (F), Opomyza germinationis (L.) (Switzerland).

Clusiidae: Clusiodinae: Clusiodes nigrifrons Frey (Burma), Hendelia orientalis (Frey) (Burma). — Clusiinae: Clusia flava (Meigen) (F), C. sexlineata Frey (Burma), Heteromeringia malaisei Frey (Burma).

Acartophthalmidae: Acartophthalmus bicolor Oldenburg (F), A. nigrinus (Zetterstedt) (F).

Odiniidae: Neoalticomerus formosus (Loew) (F), Odinia boletina (Zetterstedt) (F), O. ornata (Zetterstedt) (F), Traginops irrorata (Coquillett) (USA: Massachusetts). ASTEIOIDEA

Agromyzidae: Eucolocerinae: Eucolocera bicolor Loew (Sweden). — Agromyzinae: Agromyza albipennis Meigen (USSR: Kamtchatka), Dizygomyza morosa Meigen (Austria). — Phytomyzinae: Cerodonta denticornis (Panzer) (Azores), Napomyza lateralis Fallén (Azores), Phytomyza nigripennis Fallén (F).

Aulacigastridae: Aulacigaster leucopeza (Meigen) (F), Cyamops nebulosus Melander (USA: W Virginia).

Anthomyzidae: Anthomyza albimana Meigen (Denmark), A. dissors Collin (Germany, GDR), Ischnomyia albicosta Walker (Canada: Quebec), Mumetopia occipitalis Melander (USA), Stenomicra sp.

(Brazil).

Asteiidae: Asteia multipunctata Sabrosky (USA), A. plaumanni Sabrosky (Brazil), Leiomyza laevigata (Meigen) (Germany, GDR).

Neurochaetidae: Neurochaeta inversa D. K. McAlpine

(Australia).

LAUXANIOIDEA

Lauxaniidae: Asilostoma sp. (Brazil), Homoneura bispina (Loew) (USA: Illinois), Lauxania cylindricornis (Fabricius) (Austria), Lyciella decempunctata (Fallén) (F), Neohomoneura orientalis (Wiedemann) (Indonesia), Physogenia variegata Loew (Brazil), Poichilus fasciatus Frey (Philippines), Xenochaetina ferruginosa Hendel (Brazil).

Celyphidae: Celyphus difficilis Malloch (Taiwan), C. hyacinthus Bigot (Laos).

Chamaemyiidae: Acrometopia wahlbergi (Zetterstedt) (Germany, FRG), Chamaemyia flavipalpis (Haliday) (Germany, FRG), Leucopis scutellaris Frey (Canary Isl.), Paroctiphila inconstans Becker (Spain), Pseudodinia varipes (Coquillett) (Canada: British Columbia).

DROSOPHILOIDEA

Curtonotidae: Curtonotum anus Meigen (USSR: Far East), Cyrtona consobrina Hackman (S Africa).

Drosophilidae: Steganinae: Stegana coleoptrata (Scopoli) (F), S. sp. (Sierra Leone). — Drosophilinae: Amiota alboguttata (Wahlberg) (F), A. sp. (variegata group) (Japan), Drosophila (Idiomyia) obscuripes (Grimshaw) (Hawaii), D. (Sophophora) melanogaster Meigen (Azores), Leucophenga abbreviata (de Meijere) (Philippines), Scaptomyza flava (Fallén) (Azores).

Camillidae: Camilla acutipennis Loew (Germany, GDR), C. atripes Duda (F), C. glabra (Fallén) (F), C.

glabrata Collin (F).

Diastatidae: Campichaeta griseola (Zetterstedt) (F), C. obscuripennis (Meigen) (Poland), Diastata nebulosa (Fallén) (Germany, GDR), Odiniomorpha sp. (Brazil).

Ephydridae: Psilopinae: Atissa kairensis Becker (Cape Verde Isl.), A. limosina Becker (F), Athyroglossa glabra (Meigen) (F), Discomyza incurva (Fallén) (F), Discocerina obscurella (Fallén) (F), Mosillus subsultans (Fabricius) (F), Psilopa flavipalpis Becker (Cape Verde Isl.). — Notiphilinae: Notiphila uliginosa Haliday (F), Hydrellia griseola (Fallén) (F), Ilythea spilota (Curtis) (F), Dichaeta caudata (Fallén) (F). — Parydrinae: Euraeniotum zuttipenne (Stenhammar) (F), Hyadina nitida (Macquart) (F), Lytogaster abdominalis (Stenhammar) (F), Ochtera mantis (De Geer) (F), Parydra pusilla (Meigen) (F), Pelina aenea (Fallén) (F). — Ephydrinae: Ephydra riparia Fallén (F), E. scholtzi (Becker) (F), Lamproscatella sibilans (Haliday) (F), Scatella subguttata (Meigen) (F), Setacera aurata (Stenhammar) (F).

Cryptochaetidae: Cryptochaetum aenescens de Meijere (Burma), C. icerya Williston (Australia).

Milichiidae: Desmometopa sordida (Fallén) (F), Leptometopa latipes (Meigen) (F), Madiza glabra Fallén (F), Milichia speciosa Meigen (Hungary), Neophyllomyza acyglossa (Villeneuve) (F), Pholemyia leucozona Bilimek (Paraguay).

Carnidae: Carnus haemapterus Nitzsch (F), Meoneura neglecta Collin (F).

Tethinidae: Pelomyiella mallochi (Sturtevant) (Austria), Tethina albipila Hendel (Canary Isl.), T. illota Haliday (Sweden).

Canaceidae: Canace actites Mathis (Canary Isl.), C. nasica Haliday (Canary Isl.), Dinomyia ranula Loew (Germany, FRG), Procanace grisescens Hendel (Taiwan).

Chloropidae: Rhodesiellinae: Dactylothyrea infumata de Meijere (Phillipines). — Oscinellinae: Anatrichus erinaceus Loew (Africa), Lipara lucens Meigen (F). — Chloropinae: Camarota curvinervis (Latreille) (Atlas Mts.), Formosina lucens (de Meijere) (Philippines), Parectecephala longicornis (Zetterstedt) (Sweden), Platycephala planifrons (Fabricius) (F). — Siphonellopsinae: Lasiopleura lutea de Meijere (New Guinea), L. ornatifrons de Meijere (Philippines), Parahippelates fuscipleuris Becker (New Guinea).

MUSCOIDEA

Scathophagidae: Scathophaginae: Norellisoma spinimanum (Fallén) (USSR: Estonian SSR), Norellia spinipes (Meigen) (England), Cleigastra apicalis (Meigen) (F), Gonarcticus abdominalis (Zetterstedt) (F), Hydromyza livens (Fabricius) (F), Megaphthalmoides unilineatus (Zetterstedt) (F), Nanna tibiella (Zetterstedt) (F), Scathophaga stercoraria (L.) (F), Cochliarium albipilum (Zetterstedt) (F), Gimnomera tarsea (Fallén) (F), Cordilura aberrans Becker (F), Cosmetopus dentimanus (Zetterstedt) (F), Staegeria kunzei (Zetterstedt) (F), Acanthocnema glaucescens (Loew) (F), Microprosopa haemorrhoidalis (Meigen) (F), Okeniella dasyprocta (Loew) (F). — Delininae: Hexamitocera loxocerata (Fallén) (F), Leptopa filiformis Zetterstedt (F), Parallelomma vittatum (Meigen) (F), Phrosia albilabris (Fabricius) (USSR: European NW), Micropselapha filiformis (Zetterstedt) (F).

Anthomyiidae: Acrostilpna atricauda (Zetterstedt) (F), Chiastochaeta trollii (Zetterstedt) (F), Craspedochaeta angulata (Tiensuu) (F), Egle minuta (Meigen) (F), Fucellia fucorum (Fallén) (F), F. tergina (Zetterstedt) (F), Hylemya nigrimana (Meigen) (F), Monocrotogaster unicolor Ringdahl (F), Myopina myopina (Fallén) (F), Nupedia infirma (Meigen (F), Pegomya fulgens (Meigen) (F), P. geniculata (Bouché) (F), Eustalomyia festiva (Zetterstedt) (F), Leucophora cinerea Robineu-Desvoidy

(F), Delia nuda (Strobl) (F).

Eginidae: Egina ocypterata Meigen (Czechoslovakia), Xenotachina pallida Malloch (Philippines).

Muscidae: Achanthipterinae: Achanthiptera rohrelliformis (Robineau-Desvoidy) (F). — Muscinae: Mesembrina mystacea (L.) (F), Morellia nigrisquama Malloch (Burma), Musca domestica L. (Canary Isl.), Orthellia cornicina (Fabricius) (F), Drymeia hamata (Fallén) (F), Ophyra leucostoma (Wiedemann) (F), Pogonomyia tetra (Wiedemann) (F). — Phaoniinae: Phaonia morio (Zetterstedt) (F). — Coenosiinae: Coenosia octopunctata (Zetterstedt) (F). — Stomoxyinae: Stomoxys calcitrans (L.) (Germany).

Fanniidae: Fannia genualis (Stein) (F), F. canicularis (L.) (F), Coelomyia mollissima Haliday (F).

Calliphoridae: Calliphorinae: Bengalia jejuna (Fabricius) (Sri Lanka), Calliphora vomitoria (L.) (Japan), Catepicephala splendens (Macquart) (Philippines), Chrysomyia albiceps (Wiedemann) (Cape Verde Isl.), C. macellaria (Fabricius) (USA: California), C. megacephala (Fabricius) (Philippines), C. regalis Robineau-Desvoidy (S Africa), Lucilia caesar (L.) (F), Phormia terraenovae Robineau-Desvoidy (Mongolia), Pollenia rudis (Fabricius) (F). — Rhiniinae: Rhinia apicalis Wiedemann (Canary Isl.), Rhynchomyia speciosa Loew (Cyprus), Stomorrhina lunata (Fabricius) (Cape Verde Isl.).

Sarcophagidae: Araba stelviana (Brauer & Bergenstamm) (Switzerland), Helicophagella melanura (Meigen) (F), Hilarella hilarella (Zetterstedt) (F), Pechia praeceps (Wiedemann) (S. Domingo), Pierretia clathrata (Meigen) (F), Sarcophaga albiceps Meigen (Japan), Synorbitomyia insularis Verves (Philippines), Thyrsocnema incisilobata (Pandellé).

Rhinophoridae: Angioneura fimbriata (Meigen) (F), Anthracomya melanoptera (Fallén) (F), Melonomya nana (Meigen) (F), Melanophora roralis (L.) (Azores), Rhinophora lepida (Meigen) (France).

Phasiidae: Alophora subcoleoptrata (L.) (F), Clara dimidiata Brauer & Bergenstamm (S Africa), Clytiomya pellucens (Fallén) (F), Cylindromyia interrupta (Meigen) (F), C. pusilla (Meigen) (F), Gymnosoma fuliginosum Robineau-Desvoidy (Canada: Quebec), G. rotundatum (L.) (F), Opesia cana (Meigen) (USSR: European NW), Ormia bilimeci Brues (Mexico), Perigymnosoma rubidum (Mesnil) (Burma), Phasia crassipennis (Fabricius) (France), Subclytia rotundiventris (Fallén) (F).

Tachinidae: Dufourinae: Frerea gagatea Robineau-Desvoidy (F), Dufouria nigrita (Fallén) (F), Rondania dimidiata (Meigen) (F), Anthomyiopsis nigrisquamata (Zetterstedt) (F). — Dexiinae: Billaea fortis (Rondani) (F), Cholomyia inaequalis Bigot (Paraguay), Rutilia atribasis (Walker) (Australia), Dexa vacua (Fallén) (F). ruficauda (Zetterstedt) Tachininae: Lypha Macquartia dispar (Fallén) (F), Solieria inanis (Fallén) (F), Tachina fera (L.) (F), Voria ruralis (Fallén) (F). — Goniinae: Carcelia excisa (Fallén) (F), Onychogonia Salmacia bimaculata flaviceps (Zetterstedt) (F), Wiedemann (Canary Isl.).

OESTROIDEA

Cuterebridae: Cuterebra americana Macquart (Mexico),

C. sp. (Brazil: Uperaba), C. sp. (Brazil: Espirido Sta. Theresa), C. sp. (Paraguay), Dermatobia hominis (Pallas) (Paraguay).

Oestridae: Oestrinae: Oestrus ovis (L.) (Paraguay). — Cephenomyiinae: Cephenomyia stimulator Clark (Austria), C. trompe (Modeer) (Sweden), Pharyngomyia picta (Meigen) (Czechoslovakia). — Hypodermatinae: Hypoderma bovis (L.) (F), H. lineatum (de Villiers) (F), Oedemagena tarandi (L.) (F).

Gasterophilidae: Gasterophilus haemorrhoidalis (L.) (F), G. intestinalis (De Geer) (F), G. nasalis (L.) (F), G. inermis Brauer (C. Europe), G. pecorum Fabricius (Austria).

GLOSSINOIDEA

Glossinidae: Glossina longipennis Corti (E Africa), G. morsitans Westwood (Ghana), G. palpalis Robineau-Desvoidy (Cameroon), G. tachinoides Westwood (Nigeria).

Hippoboscidae: Hippobosca camelina Leach (Morocco), H. equina (L.) (F), Lipoptena cervi (L.) (F), Olfersia fumipennis (Sahlberg) (F), Ornithomyia chloropus Bergroth (F), Stenopteryx hirundinis (L.) (F).

Streblidae: Brachytarsina alluaudi (Falcoz) (Sudan), Trichobius major Coquillett (USA: Florida).

Appendix 2. The main types of costal chaetotaxy in the families examined (families not examined given in parentheses). Number of genera/species studied given after family name. Aberrant types are marked with asterisks. $AB_1 = \text{transitional}$ between A_1 and B_1 . R = reduced.

SUPERFAMILY Family				Ch	aeto	taxy				SUPERFAMILY Family				Ch	aetot	axy			
	A ₁	A ₂	A ₃	AB ₁	B ₁	B ₂	B ₃	С	R		$\overline{A_1}$	\mathbf{A}_2	A ₃	$\overline{AB_1}$	\mathbf{B}_1	B ₂	B ₃	С	R
TIPULOIDEA										MYCETOPHILOIDE	A								
Tipulidae 4/7	\mathbf{A}_{1}									Bolitophilidae 1/1	\mathbf{A}_{1}								
Trichoceridae 1/3	\mathbf{A}_{1}									Ditomviidae 1/1	\mathbf{A}_1								
NYMPHOMYOIDEA										Diadocidiidae 1/1	\mathbf{A}_{1}								
Nymphomyiidae 1/1									R	Keroplatidae 3/3	\mathbf{A}_{1}								
BLEPHAROCEROID	EA									Mycetophilidae 8/9	\mathbf{A}_{1}			$(AB_1$)				
Blepharoceridae 1/1	Aı									Sciaridae 2/2	$\mathbf{A_1}$,				
(Deuterophlebiidae)										CECIDOMYOIDEA									
PSYCHODOIDEA										Cecidomyiidae 1/1	\mathbf{A}_{1}								
Psychodidae 4/4	A1*									XYLOPHAGOIDEA	•								
TANYDEROIDEA	-									Coenomyiidae 3/3	$\mathbf{A_1}$								
(Tanyderidae)										Rhagionidae 11/27	Aı			AB_1	Bı				
Ptychopteridae 2/3	\mathbf{A}_{1}									Pelecorhynchidae 1/3					-•				
CULICOIDEA										Pantoph-									
Dixidae 2/3	$\mathbf{A}_{\mathbf{I}}$			(AB)					thalmidae 2/3	\mathbf{A}_1								
Chaoboridae 1/1	A ₁ *			` '	,					Rhaciceridae 3/3	A ₁								
Culicidae 3/4	Aı*									Xylophagidae 1/2	A ₁								
CHIRONOMOIDEA										(? Glutopidae)	,								
Ceratopogonidae 1/1	Αı									STRATIOMYOIDEA									
Simuliidae 5/9	\mathbf{A}_{1}	\mathbf{A}_2								Xylomyidae 1/2	$\mathbf{A}_{\mathbf{I}}$								
Chironomidae 5/5	\mathbf{A}_1	-							(R)	Stratiomyidae 18/19	A			ABı	B				R
THAUMALEOIDEA									()	TABANOIDEA									
Thaumaleidae 1/1	Αı									Tabanidae 6/8	\mathbf{A}_{1}								
PACHYNEUROIDEA	٠.									Athericidae 1/2	Aı								
Pachyneuridae 3/3	Aı									NEMESTRINOIDEA									
(Axymyiidae)										Nemestrinidae 4/6	\mathbf{A}_1								(R)
(Perissomatidae)										Acroceridae 6/6	(A_1)								R
ANISOPODOIDEA										Bombyliidae 14/15	Aı			AB_1	Вı				
Anisopodidae 3/5	Aı									(? Hilamomorphidae)									
BIBIONOIDEA										ASILOIDEA	′								
Bibionidae 4/5	Aı									Asilidae 25/34	\mathbf{A}_{1}			AB_1	\mathbf{B}_1				
SCATOPSOIDEA										Leptogastridae 4/7					B ₁				
Scatopsidae 2/2	Αı									Therevidae 8/11	\mathbf{A}_{1}				B ₁				
Synneuridae 1/1	Aı									Scenopinidae 1/3					B ₁				
Canthyloscelidae 1/1	-									Mydidae 5/7	\mathbf{A}_1				(B ₁ *	١			

SUPERFAMILY Family				Cha	etot	axy					SUPERFAMILY				Ch	aetot	axy									
	A ₁	A ₂	A ₃	AB ₁	B ₁	B ₂	B ₃	С	I		Family	A ₁	A ₂	A ₃	AB ₁	B ₁	B ₂	B ₃	С	R						
	A ₁									_	Chiroptero-	<u>-</u>														
EMPIDOIDEA											myzidae 2/3							\mathbf{B}_3								
	\mathbf{B}_1										(Mormotomyiidae)															
Atelestidae 2/2					\mathbf{B}_1						(Cnemospathidae)															
Empididae 8/14					\mathbf{B}_1	B_2 *					Borboroides 1/1					\mathbf{B}_1										
Microphoridae 1/1					\mathbf{B}_1						Listromastax 1/1		$2A_1$ *													
Dolichopo-											Dichrochira 1/1					\mathbf{B}_1										
didae 12/22	_				\mathbf{B}_1	\mathbf{B}_2 *					OPOMYZOIDEA															
LONCHOPTEROIDE.	A										Lonchaeidae 3/4						\mathbf{B}_2									
Lonchopteridae 1/4						\mathbf{B}_2 *					Opomyzidae 3/4						\mathbf{B}_2									
PHOROIDEA					n						Clusiidae 4/5						\mathbf{B}_2									
Phoridae 10/13					\mathbf{B}_1						Acartoph-						ъ									
(Irinomyiidae) (Sciadoceridae)											thalmidae 1/2 Odiniidae 3/4						B ₂									
PLATYPEZOIDEA .											Agromyzidae 6/6					\mathbf{B}_1	\mathbf{B}_2									
Platypezidae 6/9					\mathbf{B}_1			С			ASTEIOIDEA					Di										
SYRPHOIDEA					Di			-			Aulacigastridae 2/2					\mathbf{B}_1										
Pipunculidae 7/13	\mathbf{A}_{1}			AB_1	B.						Anthomyzidae 4/5					B ₁	(\mathbf{B}_2)									
Syrphidae 36/47	Aı			AB ₁							Asteiidae 2/3					B ₁	(12)									
CONOPOIDEA					-•						Neurochaetidae 1/1					B ₁										
Conopidae 9/17	\mathbf{A}_1			AB_1	\mathbf{B}_1						(Xenasteiidae)															
MICROPEZOIDEA											LAUXANIOIDEA															
Cypselo-											Lauxaniidae 8/8						\mathbf{B}_2									
somatidae 3/3					\mathbf{B}_1						Celyphidae 1/2					\mathbf{B}_1										
Neriidae 4/4					\mathbf{B}_{1}						Chamaemyiidae 5/5					B_1-I	32									
Micropezidae 5/5					\mathbf{B}_1						(Eurychoromyiidae)															
NOTHYBOIDEA											DROSOPHILOIDEA															
Nothybidae 1/4								C			Curtonotidae 2/2							\mathbf{B}_3								
Tanypezidae 3/5				$(\mathbf{AB_1})$							Drosophilidae 5/8						\mathbf{B}_2									
Psilidae 3/5					\mathbf{B}_1						Camillidae 1/4							\mathbf{B}_3								
Diopsidae 4/5					\mathbf{B}_1						Diastatidae 3/4					_	\mathbf{B}_2	\mathbf{B}_3								
(Syringogastridae)											Ephydridae 20/23					B ₁	\mathbf{B}_2	B ₃ *								
Teratomyzidae 1/1					Bı						Cryptochaetidae 1/2					\mathbf{B}_1										
Periscelididae 2/3					B ₁						Milichiidae 6/6					n	\mathbf{B}_2									
Somatiidae 1/2 OTITOIDEA					\mathbf{B}_1						Carnidae 2/2					\mathbf{B}_{1}			, .	1						
Otitidae 16/23					\mathbf{B}_1	\mathbf{B}_2		С			(Braulidae) Tethinidae 2/3					ъ			(WII	ngless)						
Platysto-					Di	D ₂		C			Canaceidae 3/4					\mathbf{B}_1	\mathbf{B}_2									
matidae 20/24	Aı			(AB ₁)	R.						Chloropidae 9/10					\mathbf{B}_1	D2									
Pyrgotidae 4/5	Aı			AB_1	, D						(Risidae)					D										
Tephritidae 35/40	Aı			AB ₁	Bı	B2*					(? Tunisimyiidae)															
Tachiniscidae 1/1						B ₂ *					MUSCOIDEA															
F.ichardiidae 4/4					\mathbf{B}_1						Scatho-															
Pallopteridae 2/16		(A_2)			\mathbf{B}_{1}	\mathbf{B}_2					phagidae 22/22	A	1 A2	•												
Eurygnatho-											Anthomyiidae 13/15								C*							
myiidae 1/1			A3*								Eginidae 2/2								C*							
Piophilidae 4/4			A3*			\mathbf{B}_2					Muscidae 11/11								C							
SCIOMYZOIDEA											Fanniidae 2/3								C							
Coelopidae 3/4					\mathbf{B}_{1}	\mathbf{B}_2					Calliphoridae 10/13								C							
Dryomyzidae 2/4						B ₂ *	\mathbf{B}_3				Sarcophagidae 8/8								C							
Sciomyzidae 10/13				AB_1		B ₂ *					Rhinophoridae 5/5								C*							
(Helosciomyzidae)					_						Phasiidae 10/12								C*							
Sepsidae 6/6					B ₁	B ₂ *					Tachinidae 16/16	(A	1 A2)					С							
Rhopalomeridae 2/3					Bı						OESTROIDEA															
Megamerinidae 2/2					\mathbf{B}_1						Cuterebridae 2/4	A				n .			(0.4)							
(Cremifaniidae) HELEOMYZOIDEA											Oestridae 5/7	A				B ₁ *			(C*))						
Heleomyzidae 9/17	\mathbf{A}_1	\mathbf{A}_{2}					B ₃				Gasterophilidae 1/5 GLOSSINOIDEA	A	1			(B ₁ *	,									
Borboropsidae 1/1	Λij	A ₂					13				Glossinidae 1/4		\mathbf{A}_2	*												
(Rhinotoridae)		1112									Hippoboscidae 5/6	A			ΔD	R.										
Trixoscelidae 1/2							B ₃				(Nycteribiidae)	А	ı		AB	\mathbf{B}_1			/	ngless)						
Chyromyiidae 1/2						B ₂ *	203				Streblidae 2/2	A	,						(WI	igicss)						
Sphaeroceridae 13/19				AB_1	R.	-52					Suconade 4/4	л	1													
UNPLACED FAMILII					١,						Total number:															
											128 families 655/8'	75														
AND GENERA																										

Received 30.V.1984, revised 6.XI.1984 Printed 11.VII.1985