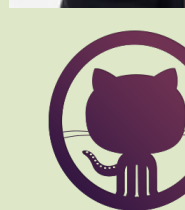# Identification of genomic regions carrying a causal mutation in unordered genomes

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I'm a predoctoral intern at The Sainsbury Laboratory doing Bioinformatics in Team MacLean.

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SNP_Distribution_method

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Motivation

EMS
Mutant

Mapping line

F1

F2

NGS

Contig assembly

SNP calling

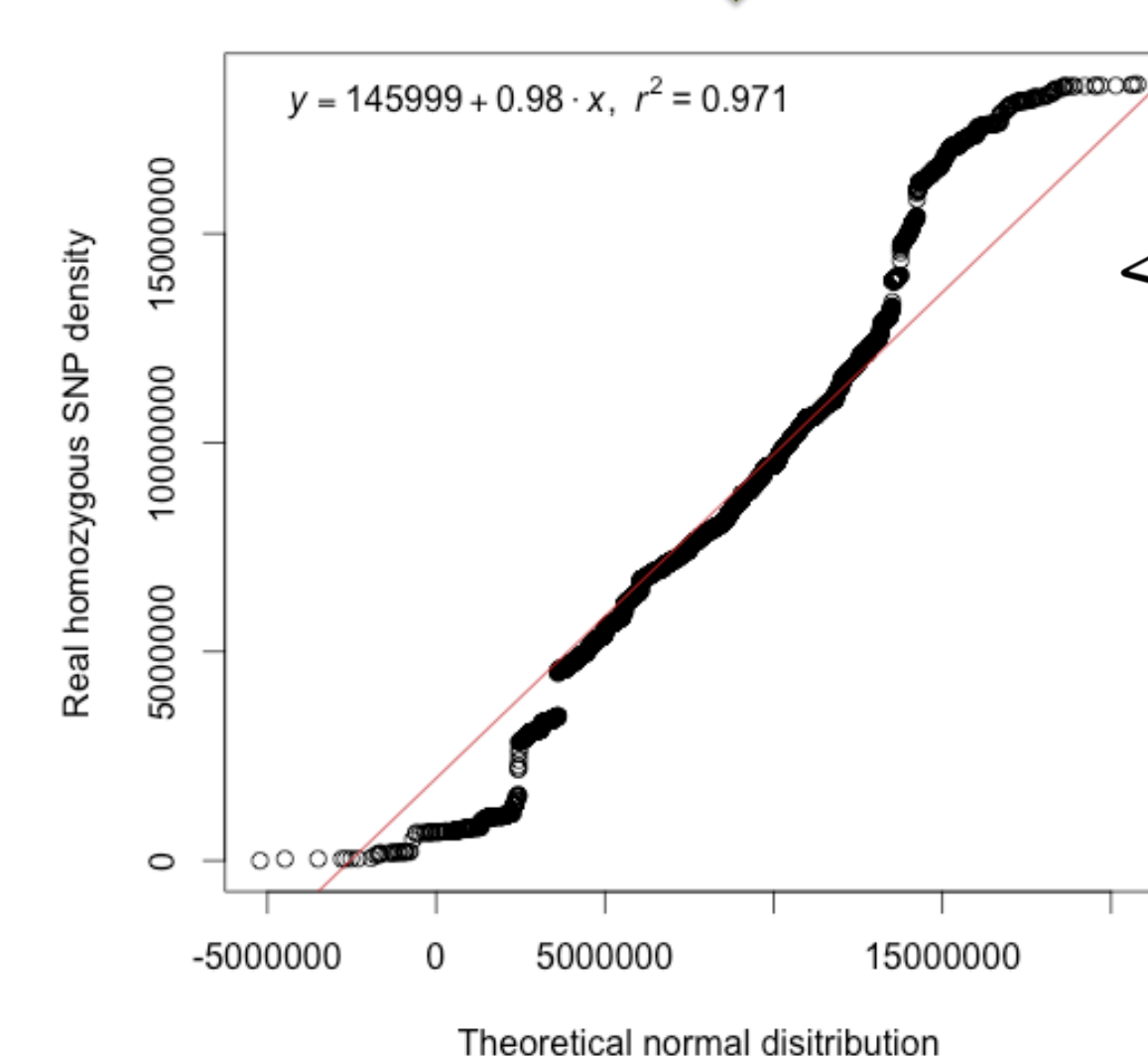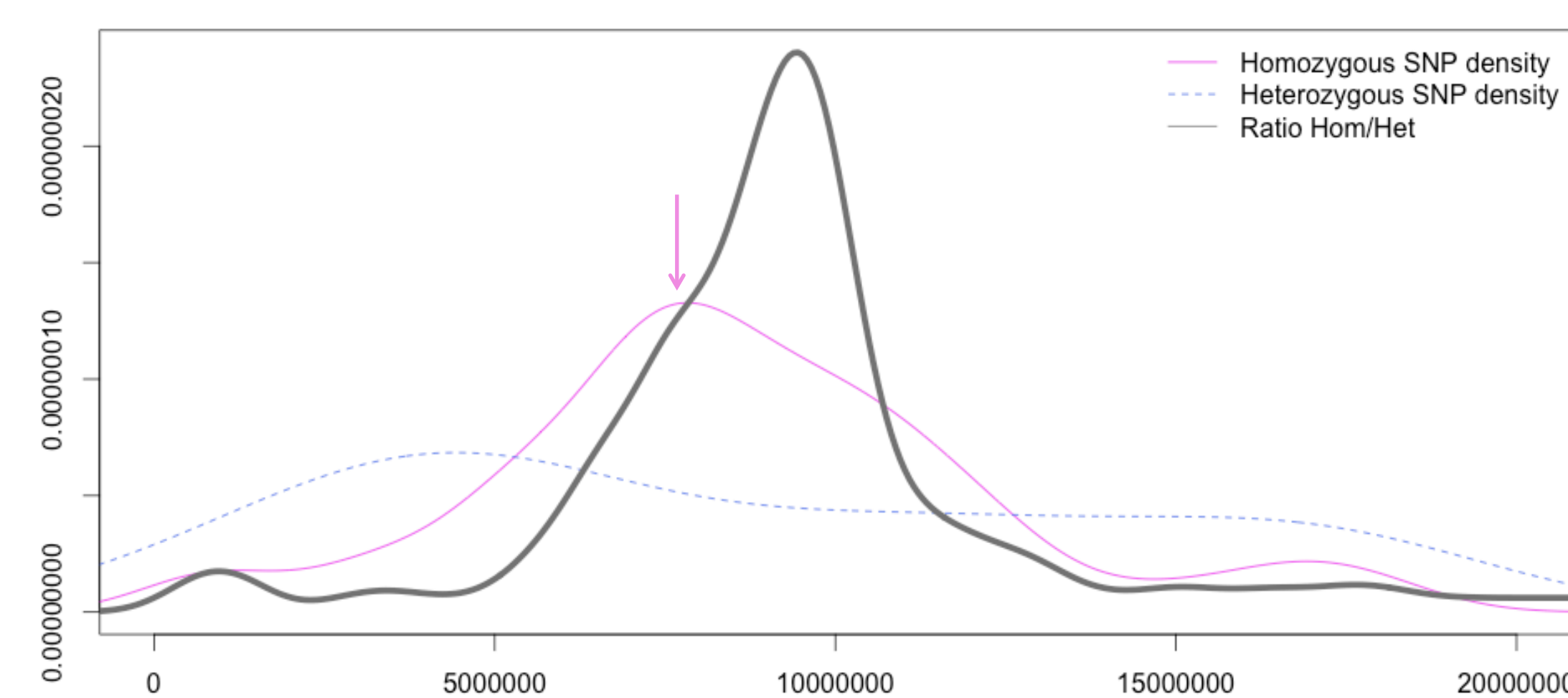Causative mutation identification without a reference genome

Forward genetic screens have been a fundamental strategy to find genes involved in biological pathways in model species. Mutagenized individuals with a phenotype of interest are isolated and a recombinant mapping population is created by back-crossing to the parental line or out-crossing to a polymorphic ecotype.

The recombination frequency between the causal mutation and nearby genetic markers is low, so the alleles of these genetic markers will co-segregate with the phenotype-altering mutation while the remaining unlinked makers segregate randomly in the genome. Hence, allele distribution analysis can uncover these low recombinant regions to identify the location of the causal mutation.

Traditional genetic mapping is a work intensive and time-consuming process but recent advances in high-throughput sequencing (HTS) have greatly accelerated the identification of mutations underlying mutant phenotypes in forward genetic screens. In the last few years, researchers have developed user-friendly tools for mapping-by-sequencing, yet they are not applicable to **organisms with non-sequenced genomes.**
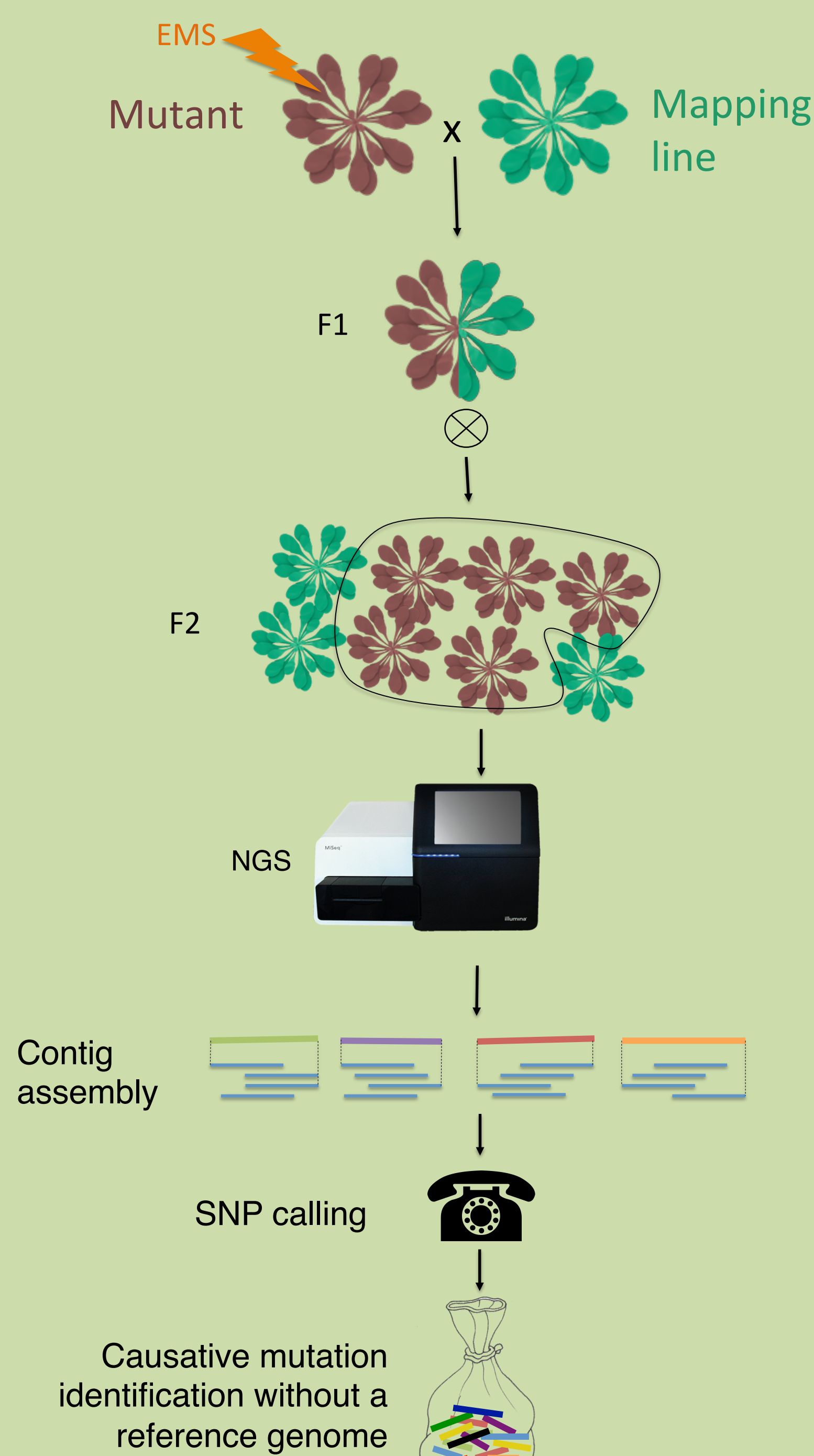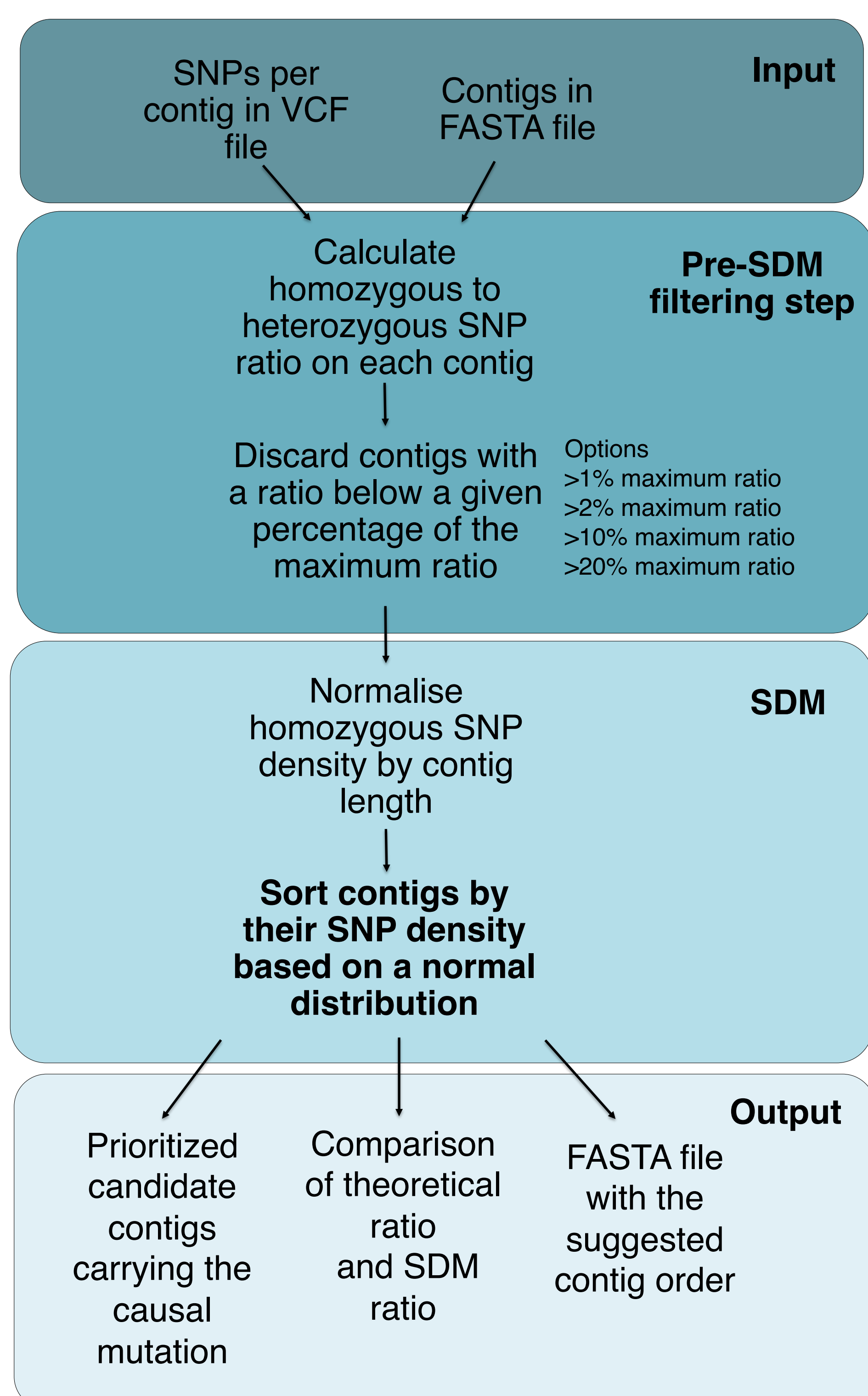
SNP density plots revealed the homozygous SNP linkage around the causative mutation causing a high homozygous to heterozygous ratio signal where the mutation is located
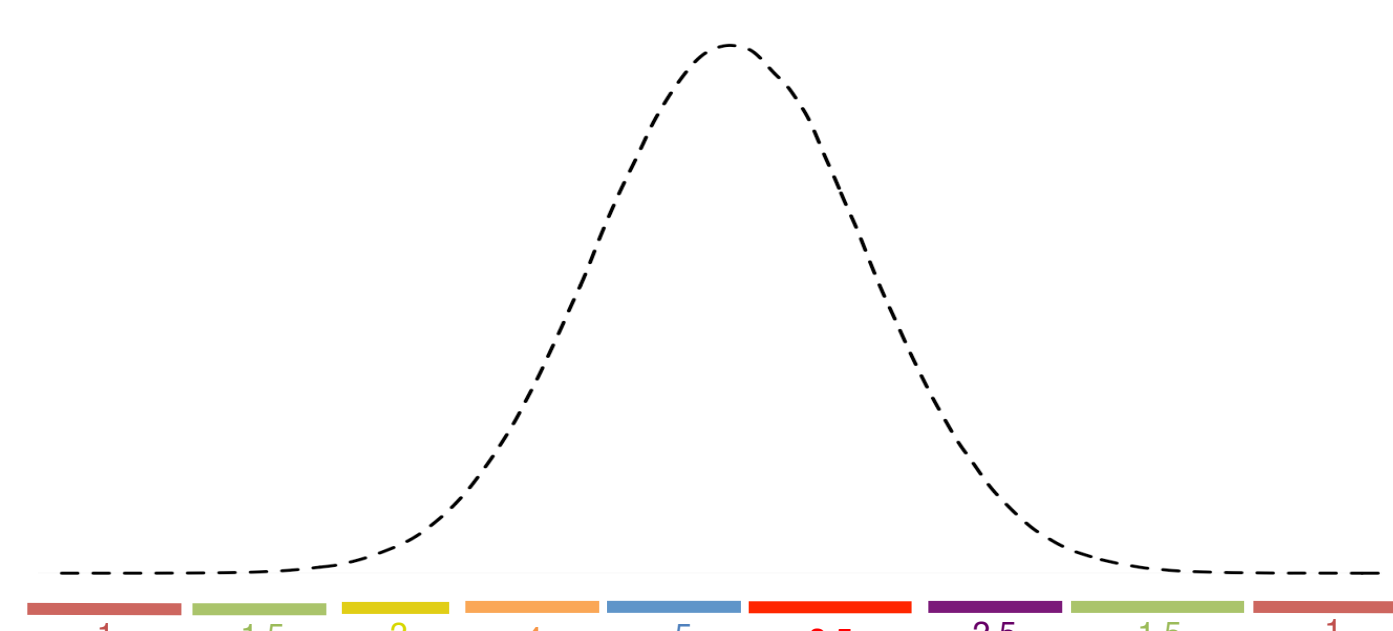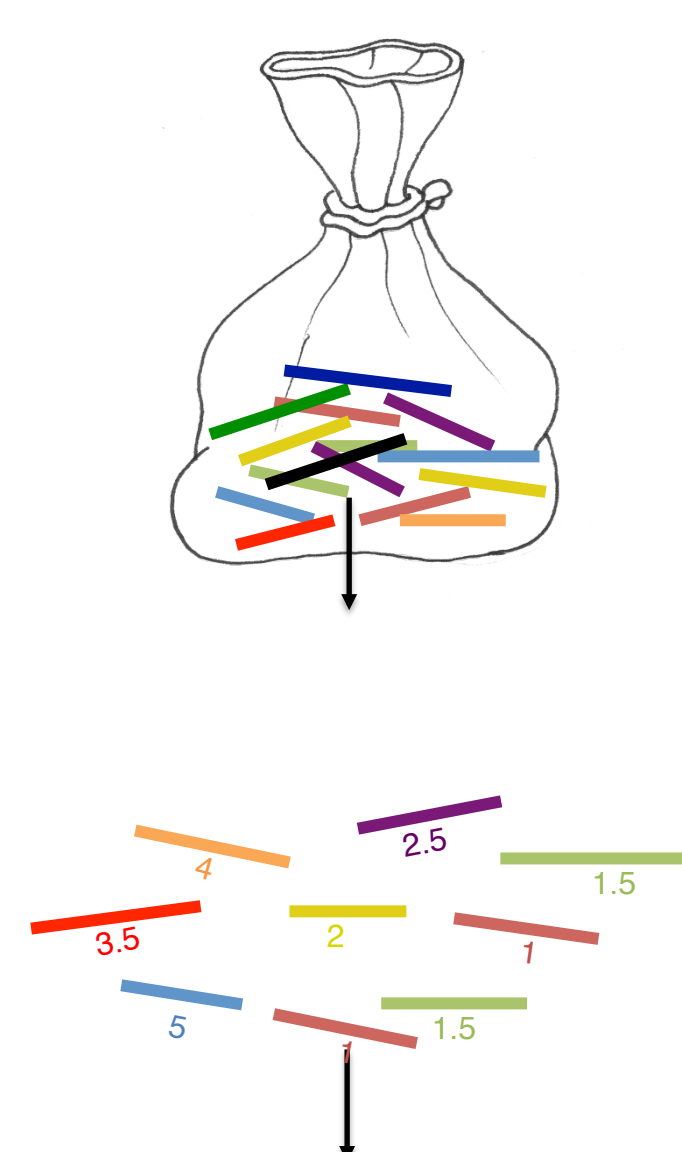

Homozygous SNPs are normally distributed around the causal mutation!

Modelling

Model genomes are useful to help us developing our method and identifying its limitations. By using idealised SNP distributions, we can predict where a mutation is going to be located and estimate the deviation of SDM from this expected position. A normal distribution was used for the homozygous SNPs while heterozygous SNPs followed a uniform distribution. We created different model genomes based on *Arabidopsis thaliana* chromosome 1. We tested the effect of genome length and contig size on SDM performance.
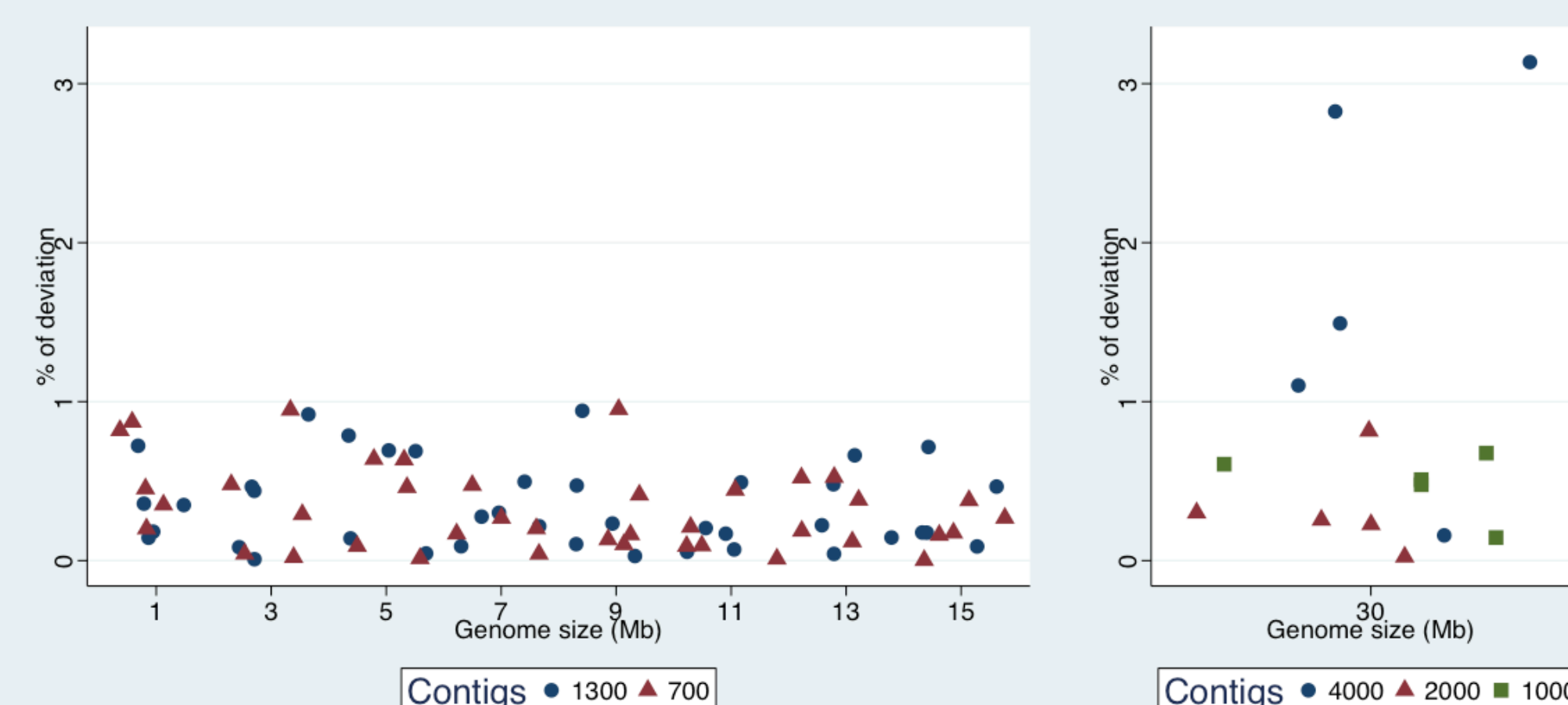
We define the homozygous to heterozygous SNP ratio on contig n as:

$$Ratio_n = \frac{(\sum Hom) + 1}{(\sum Het) + 1}$$

Contigs that are located furtheraway from the causal mutation have a constant homozygous SNP density due to recombination. The low ratio in these regions is used as a filter to focus on the genomic region where the mutation is likely to be found. Contigs with a ratio falling below a given percentage of the maximum ratio will be discarded.

SNP Distribution Method

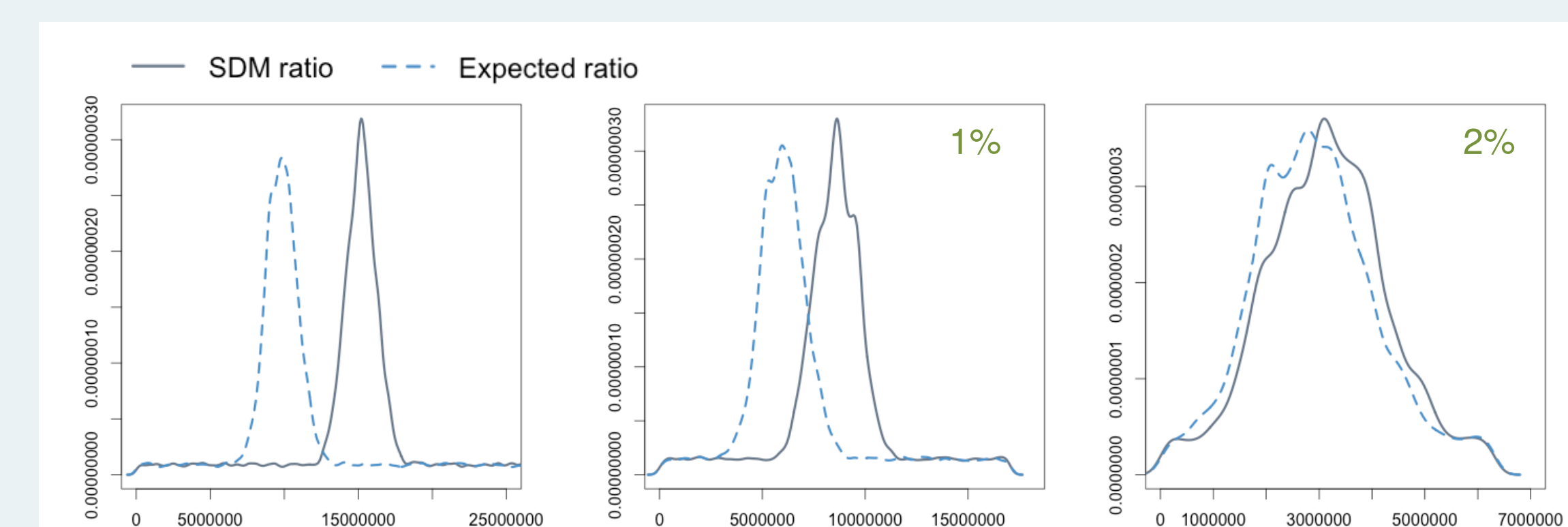|	
SNPs per contig in VCF file / Contigs in FASTA file	**Input**
Calculate homozygous to heterozygous SNP ratio on each contig	**Pre-SDM filtering step**
Discard contigs with a ratio below a given percentage of the maximum ratio	Options: >1% maximum ratio, >2% maximum ratio, >10% maximum ratio, >20% maximum ratio
Normalise homozygous SNP density by contig length / **Sort contigs by their SNP density based on a normal distribution**	**SDM**
Prioritized candidate contigs carrying the causal mutation / Comparison of theoretical ratio and SDM ratio / FASTA file with the suggested contig order	**Output**

The SNP Distribution Method (SDM) is an approach for fast causative mutant identification based on a simple reference-free contig assembly that allows the detection of candidate causative SNPs. Instead of relying on a genome comparison, it focuses on the SNP linkage around acausal mutation and analyses the SNP distribution to identify the chromosome area where the putative mutated gene is located.

Take home

✓ **Forward genetic screens** are very useful to identify genes responsible for particular phenotypes.

✓ Homozygous SNPs are **normally distributed** in the mutant genome of back-crossed and out-crossed individuals. We defined a theoretical SNP distribution used by SDM to identify the genomic region where the causative mutation is located.

✓ SDM does not rely on previously known genetic markers and can be used on extremely **fragmented genome assemblies**, even down to the level of long reads.