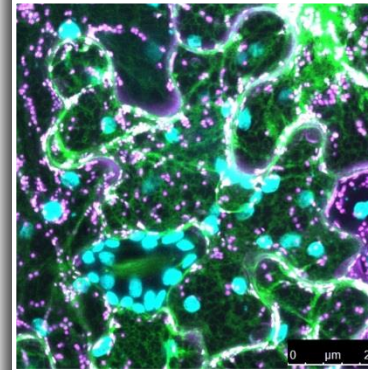


# Plant Endomembrane Trafficking

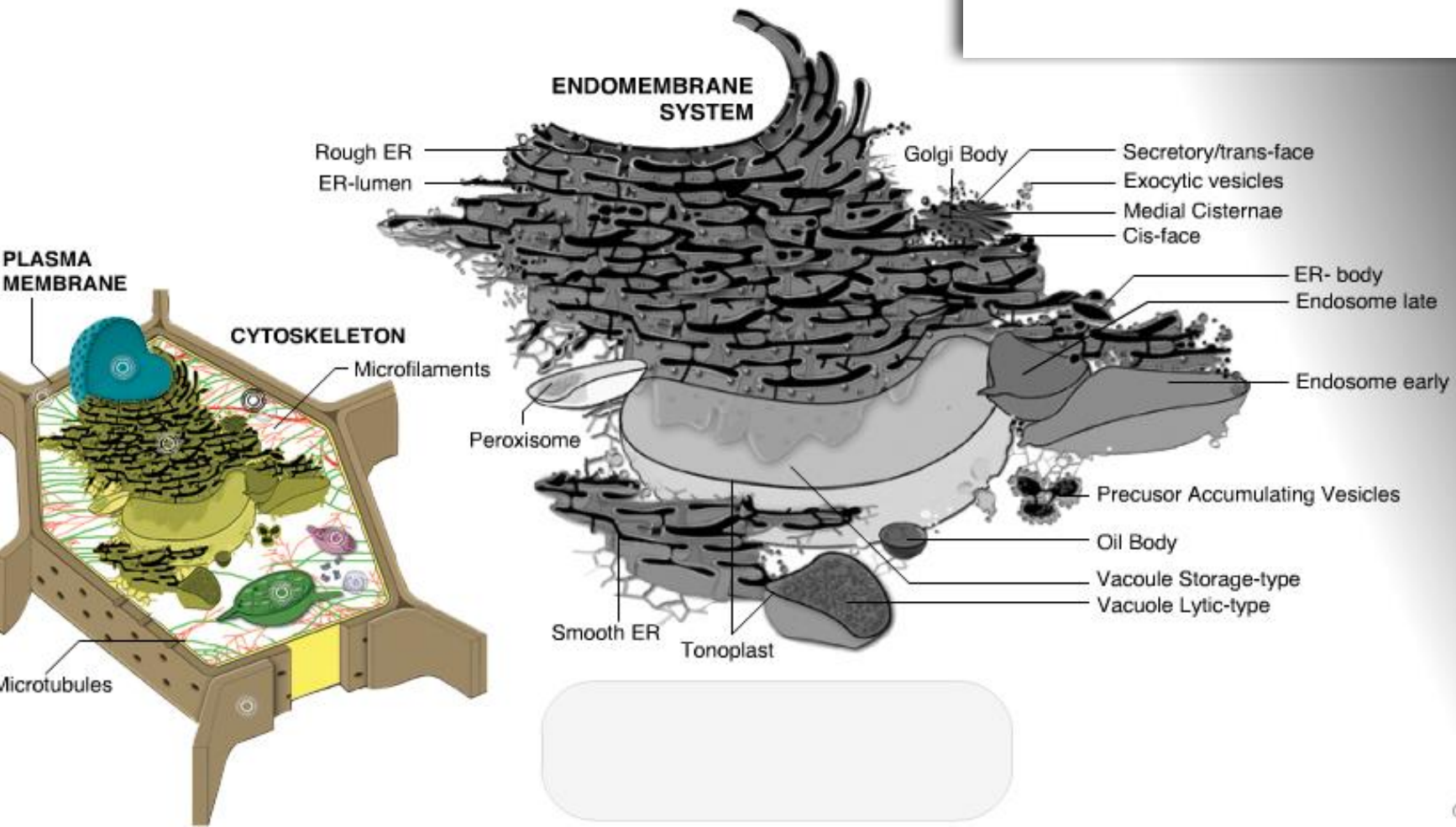
## Pathways and Regulators

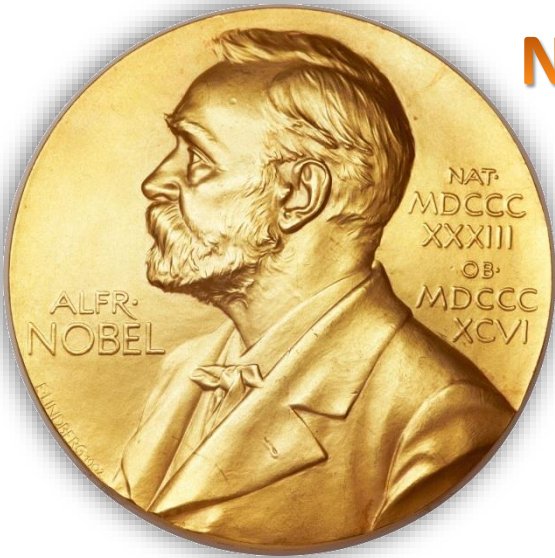


Summer School 2015

Robatzek lab  
Theoretical session

Michaela Kopischke





# Nobel Prize in Physiology or Medicine 2013

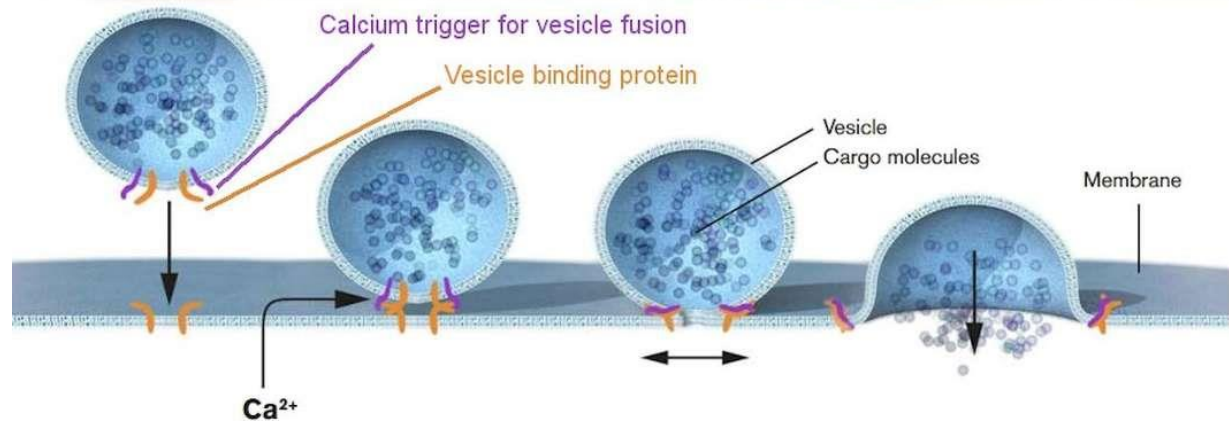
James E. Rothman  
Yale

Randy W. Schekman  
UC Berkeley

Thomas C. Südhof  
Stanford

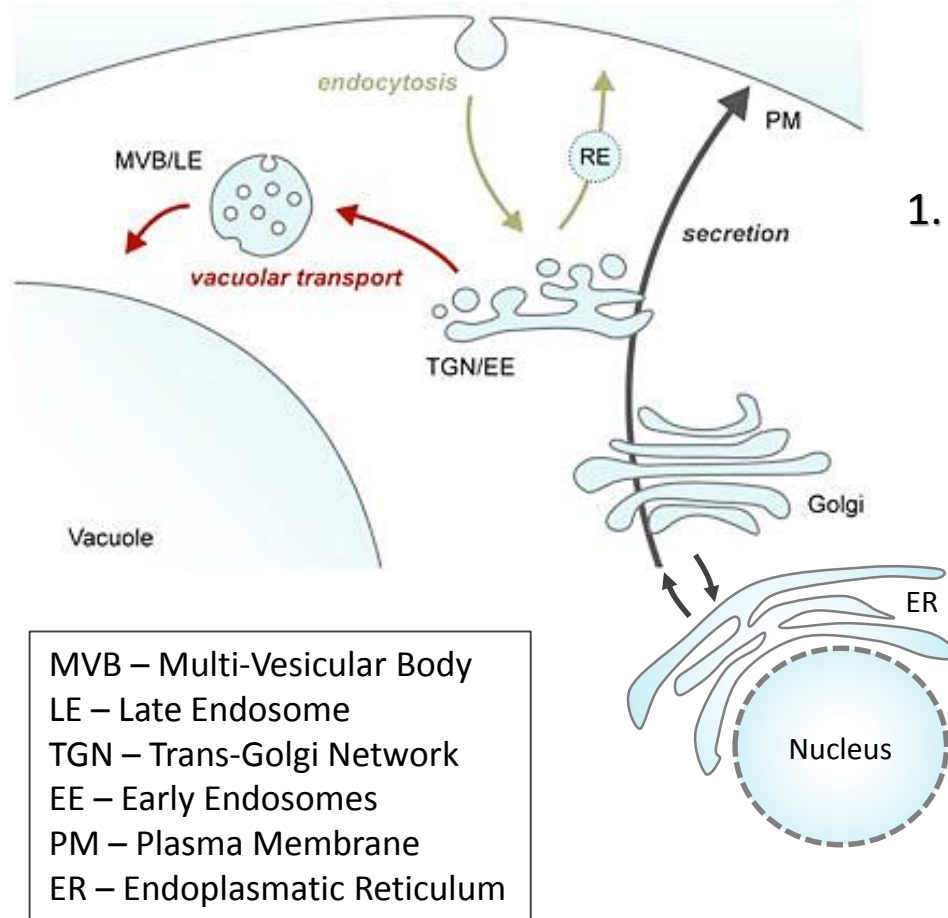


[...] James E. Rothman, Randy W. Schekman and Thomas C. Südhof have elucidated [...] fundamental processes in eukaryotic cells that [...] ensure that molecular cargo is correctly destined. [...] Vesicle transport and fusion operates, with the same general principles, in organisms as different as yeast and man. [...] Without this exquisitely precise organization, the cell would lapse into chaos.



# Endomembrane Trafficking Pathways

## 3. Endocytic Pathway



## 2. Vacuolar Transport Pathway

## 1. Secretory Pathway

# Plant Endosomal Compartments

## Endosome Functions:

- Trafficking of biosynthetic and endocytic cargo
- intermediate compartments for storage, sorting and organelle targeting of material within the cell
- Required for key plant processes:
  - Embry differentiation
  - Guard cell movement
  - Regulation of auxin transport
  - Gravitropism
  - Cell wall remodeling
  - Defense response against pathogens
  - Epidermis differentiation

# Plant Endosomal Compartments

## Endosome Functions:

- Trafficking of biosynthetic and endocytic cargo
- intermediate compartments for storage, sorting and organelle targeting of material within the cell
- Required for key plant processes
- Classification based on their main function: **early, recycling, intermediate and late endosomes**

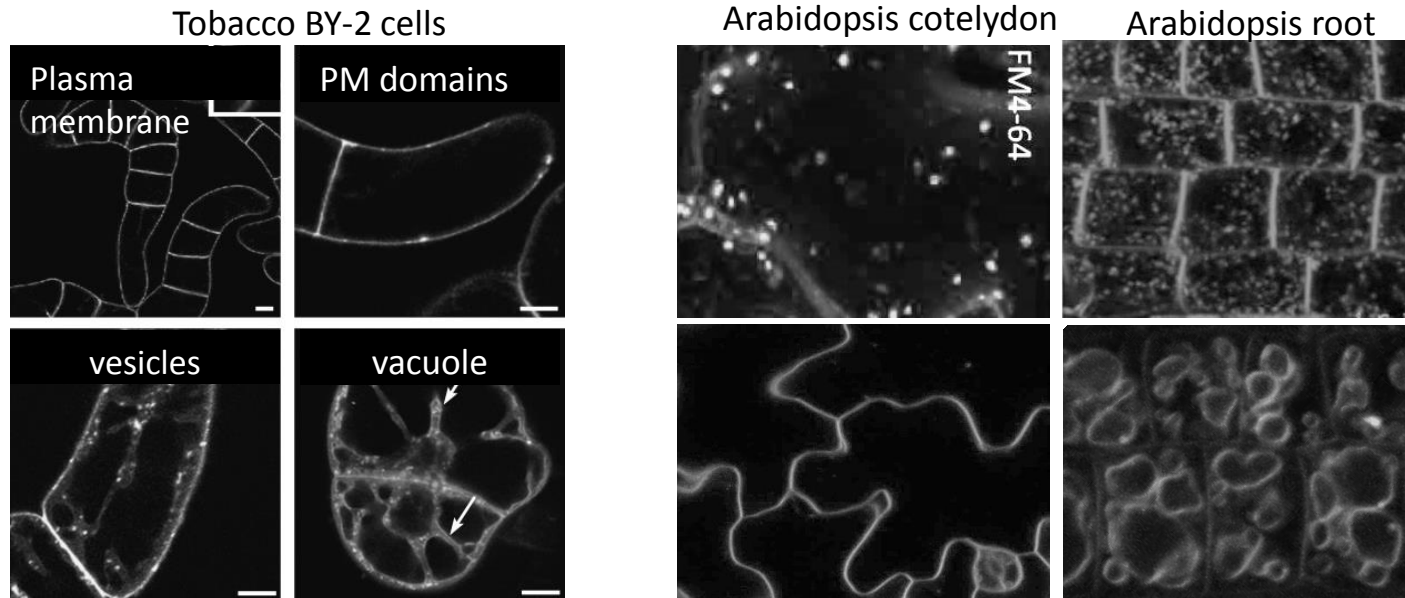


# Plant Endosomal Compartments

## Endosome Functions:

- Trafficking of biosynthetic and endocytic cargo
- intermediate compartments for storage, sorting and organelle targeting of material within the cell
- Required for key plant processes
- Classification based on their main function: **early, recycling, intermediate and late endosomes**

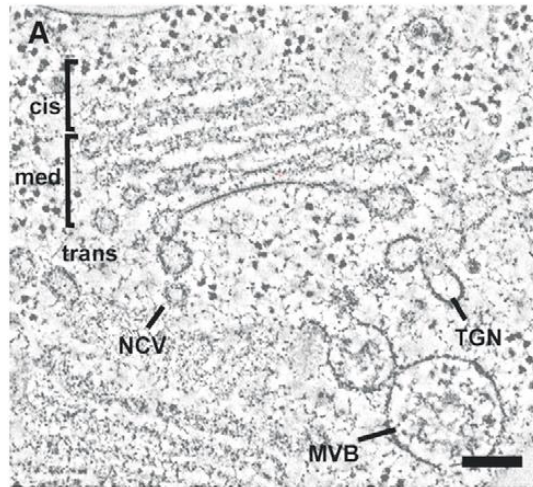
## Endocytic tracer FM4-64



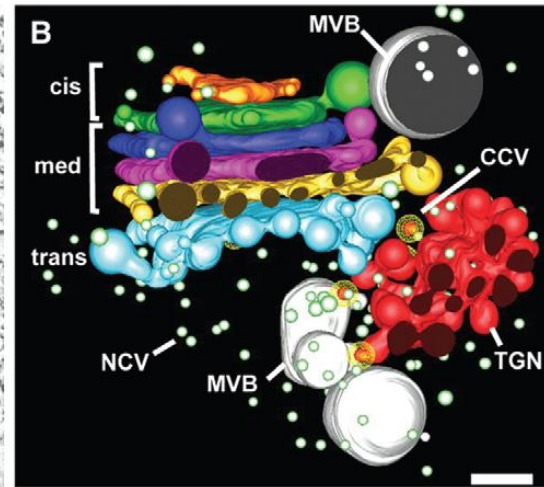
Excitation: 488 nm

Emission: 580 – 640 nm

# Plant Endosomal Compartments

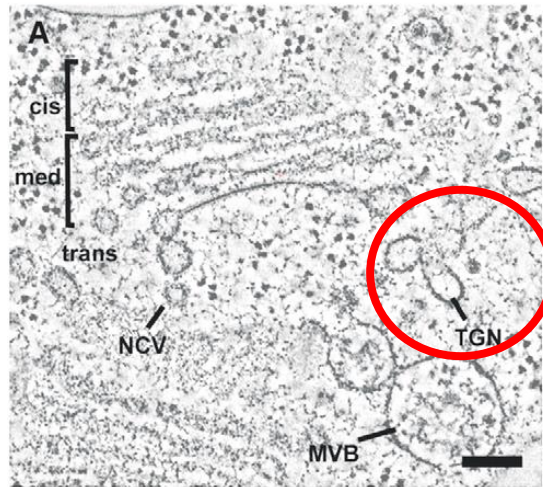


**Early Endosomes:  
Trans-Golgi-Network  
(TGN)**

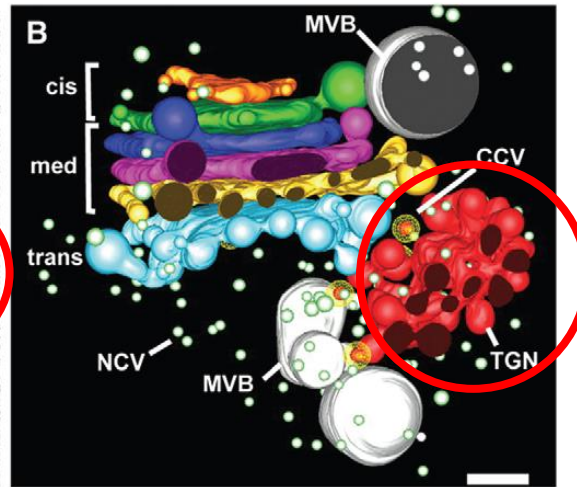


**Late Endosomes:  
Multi-Vesicular Bodies  
(MVBs)**

# Plant Endosomal Compartments



**Early Endosomes:  
Trans-Golgi-Network  
(TGN)**

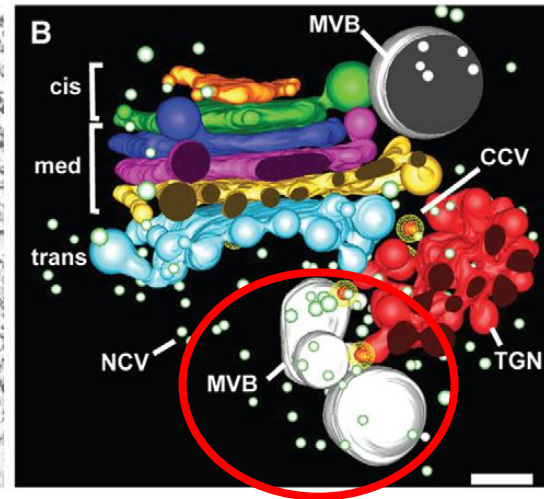


**Late Endosomes:  
Multi-Vesicular Bodies  
(MVBs)**

- 1<sup>st</sup> site for delivery of endocytosed cargo
- Rapidly labelled by FM4-64
- Central hub for biosynthetic and endocytic material
- Might contain specialised subdomains with distinct functions

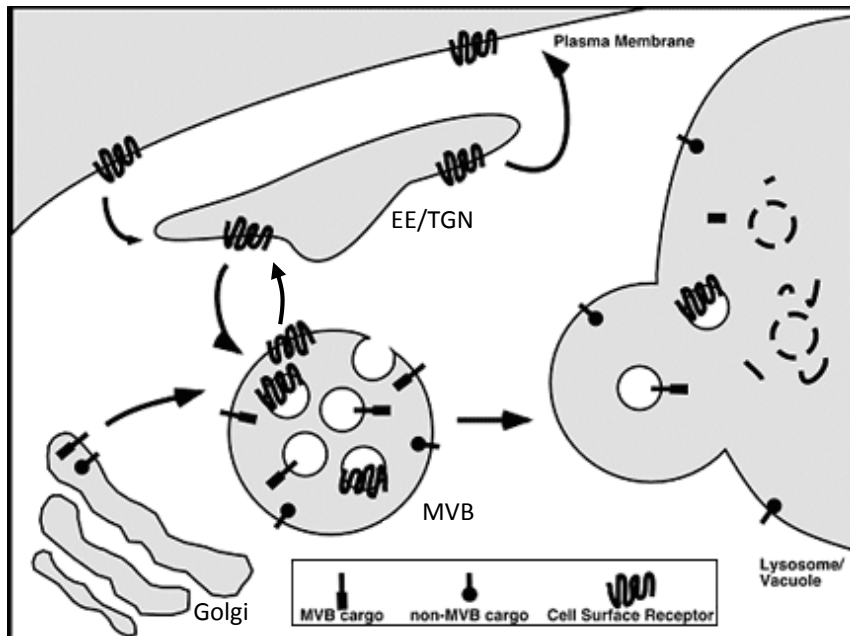


# Plant Endosomal Compartments



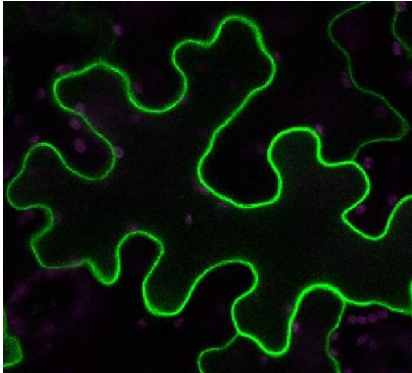
## Late Endosomes: Multi-Vesicular Bodies (MVBs)

- Multivesicular structure
- Cargo in outer MVB membrane → delivery to tonoplast membrane (biosynthetic cargo or lipids) or recycling to EE
- Cargo in intraluminal vesicles → delivery to vacuolar lumen, destined for degradation
- Intraluminal vesicles generated through action of endosomal sorting complex required for transport (ESCRT)

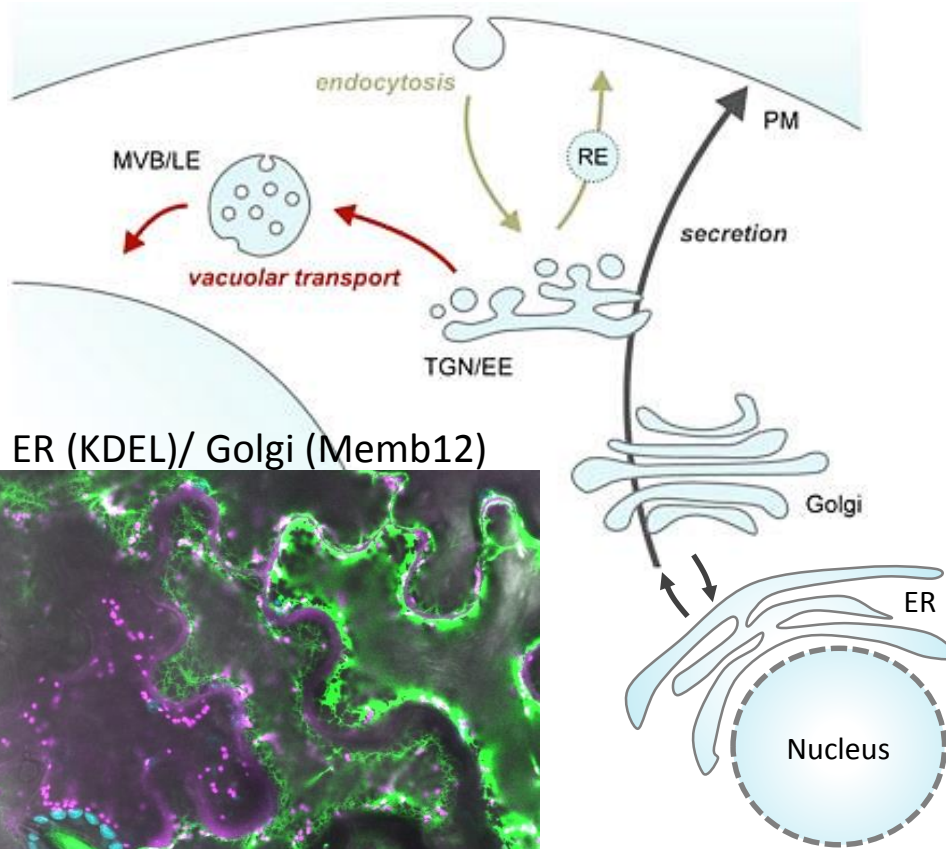
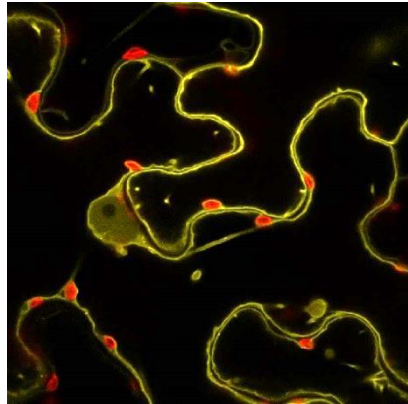


# Endomembrane Trafficking Pathways

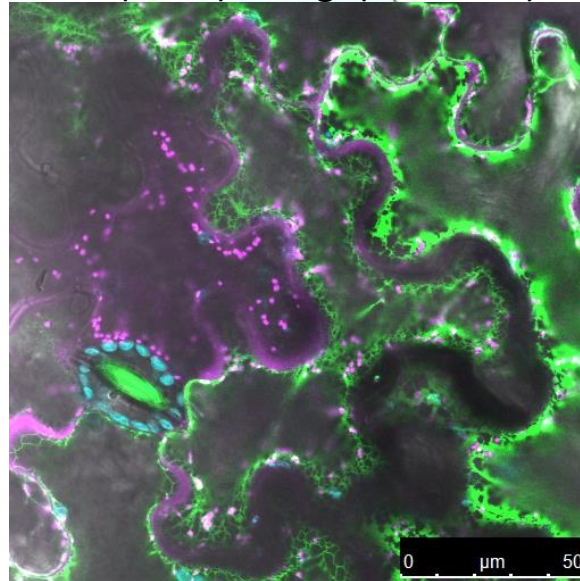
PM – FLS2



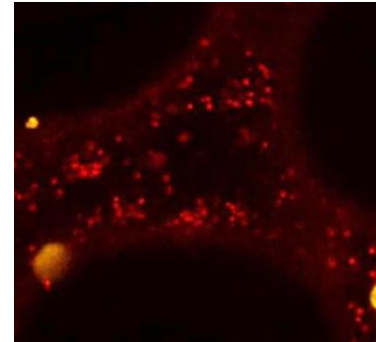
Vacuole – RabG3F



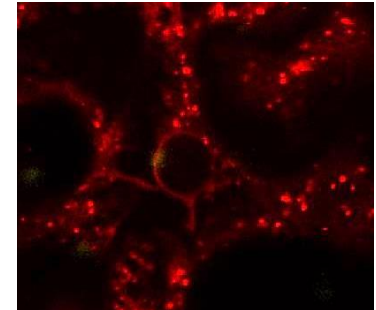
ER (KDEL)/ Golgi (Memb12)



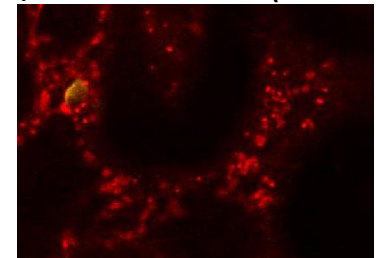
TGN – Vha-A1



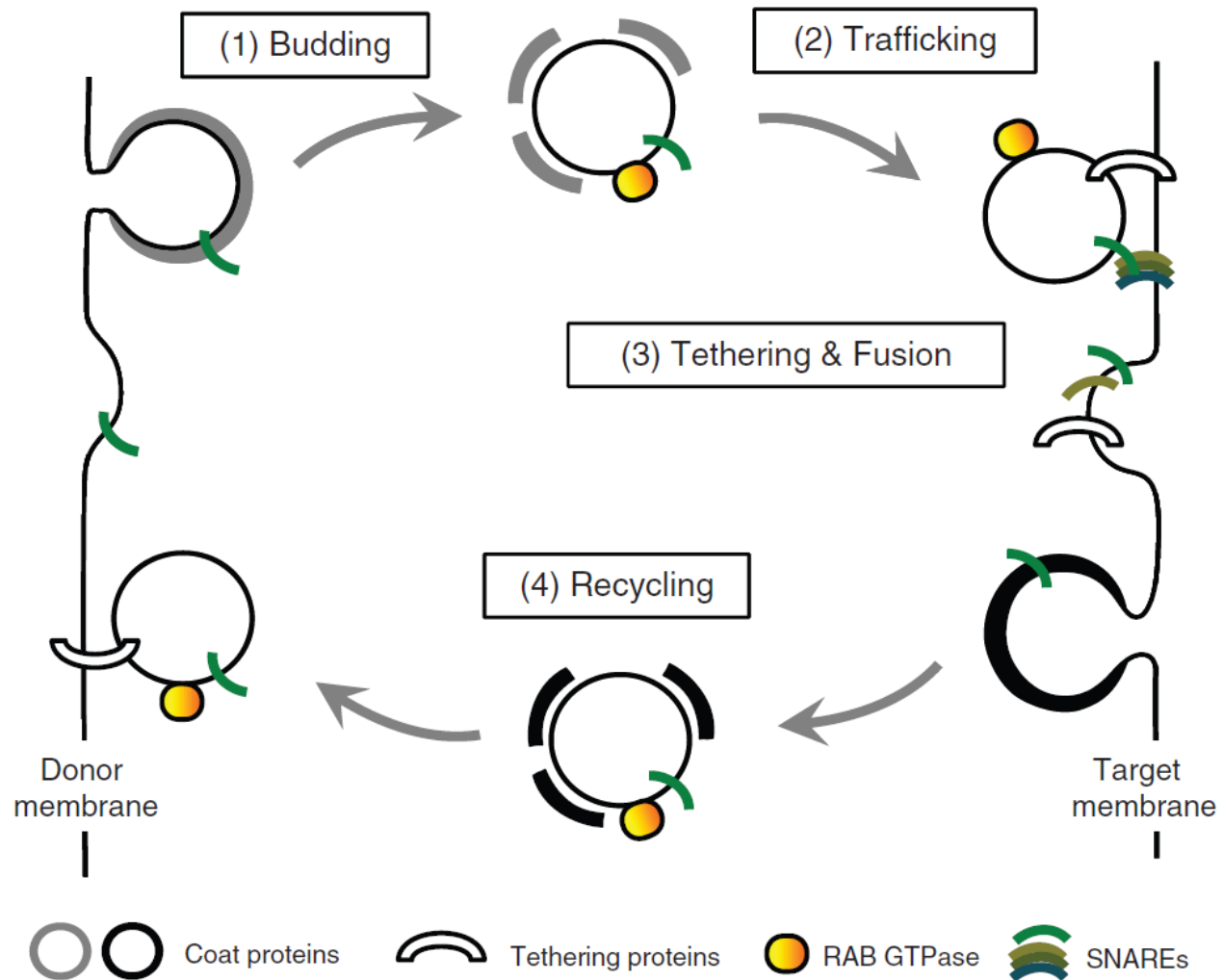
EE/LE – ARA7 (RabF2b)



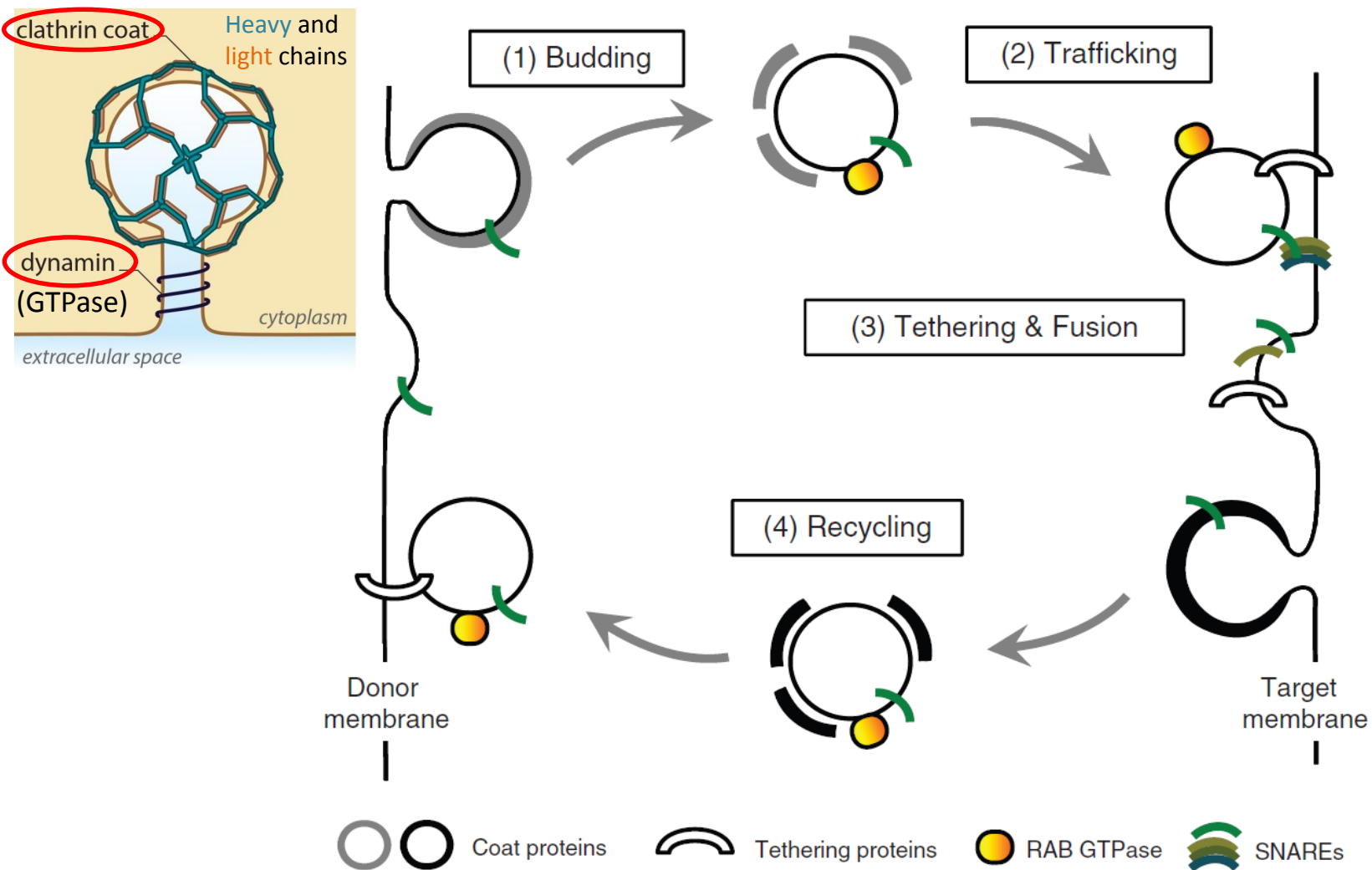
LE/MVB – ARA6 (RabF1)



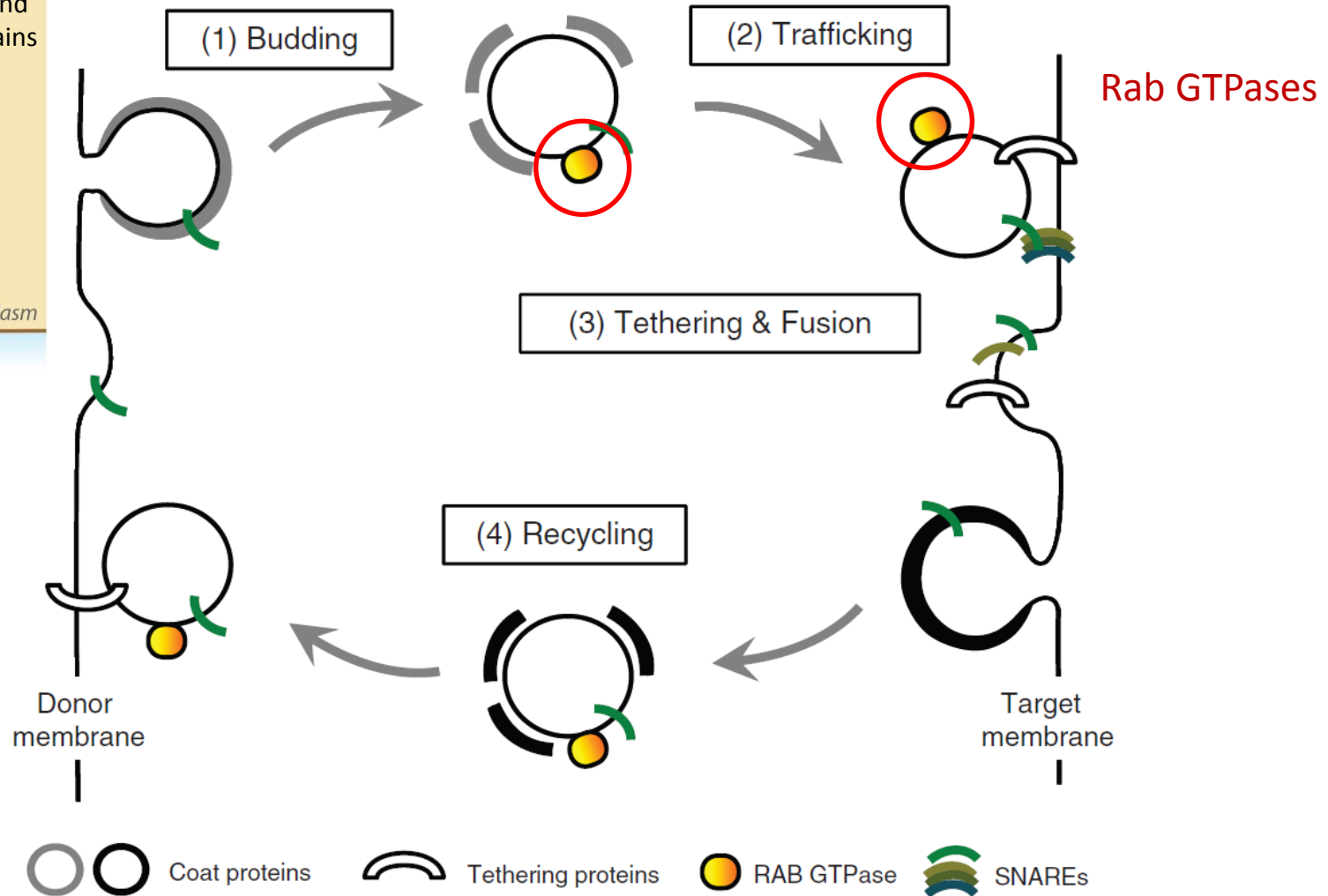
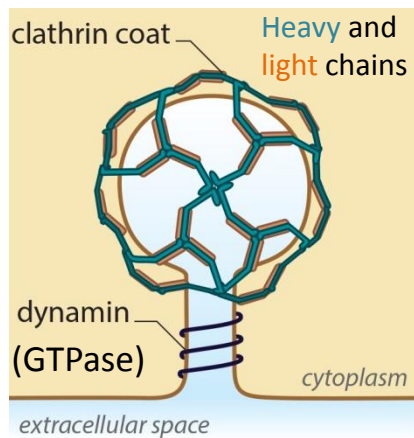
# General Membrane Trafficking Machinery



# General Membrane Trafficking Machinery

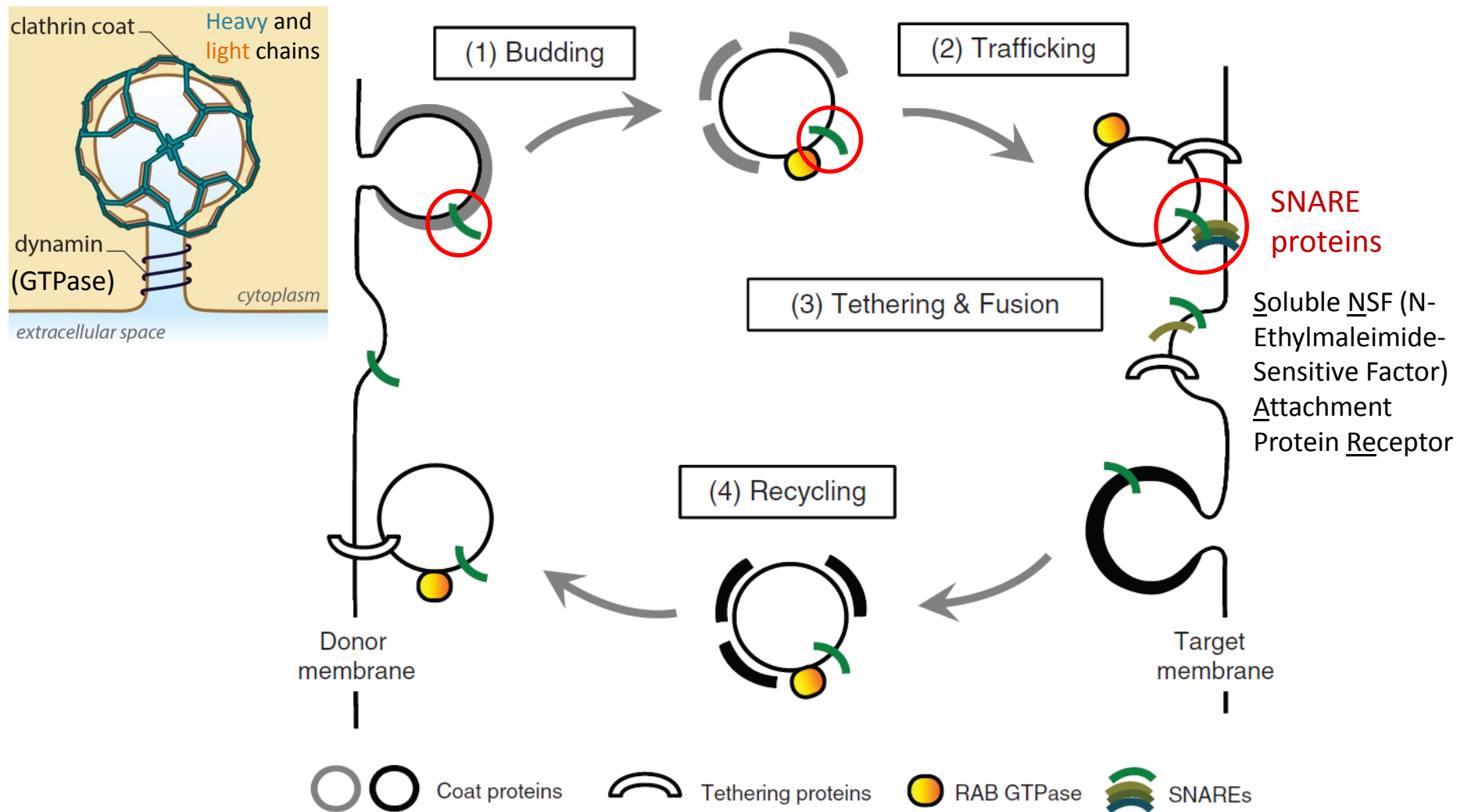


# General Membrane Trafficking Machinery

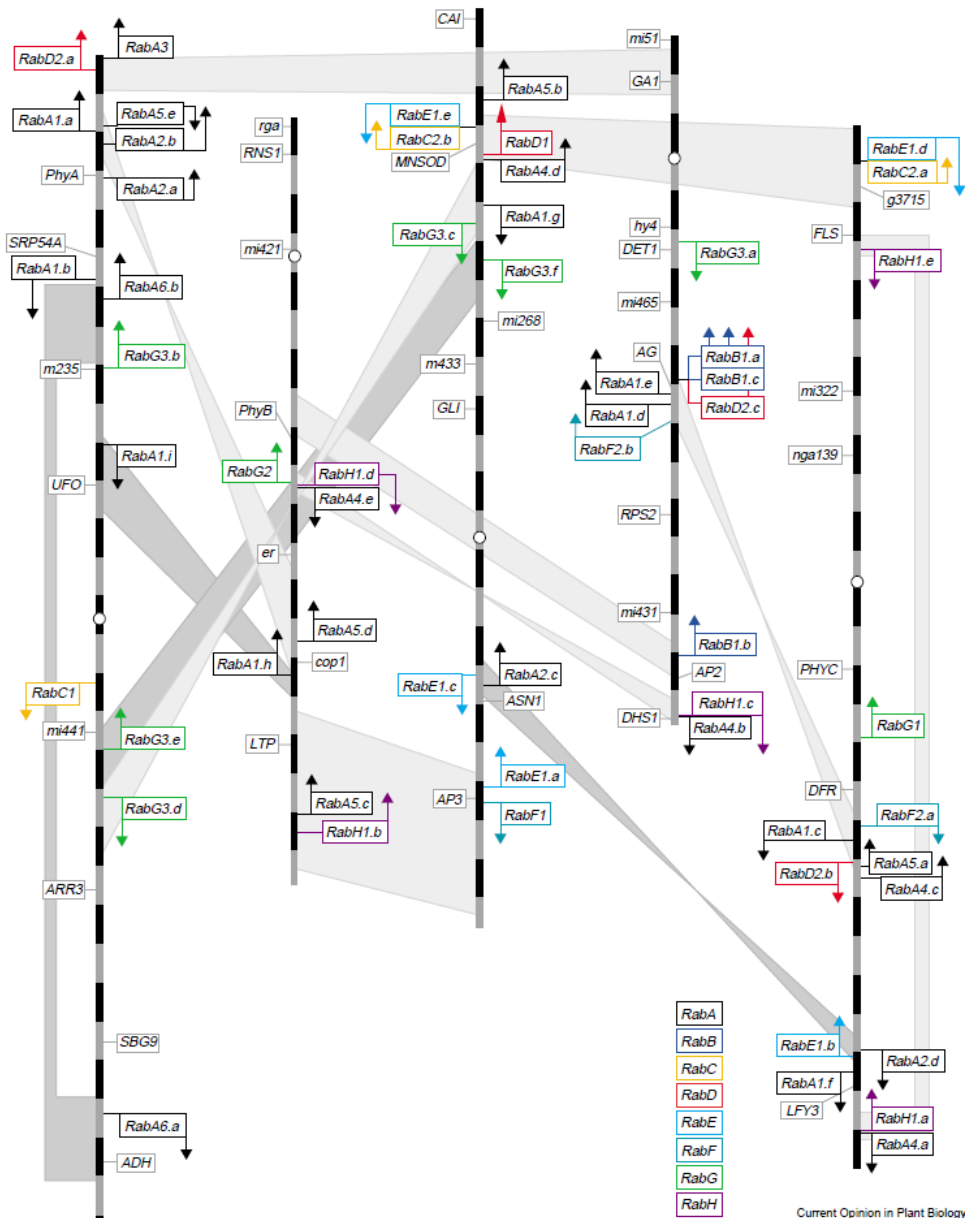




# General Membrane Trafficking Machinery



## Small Rab GTPases



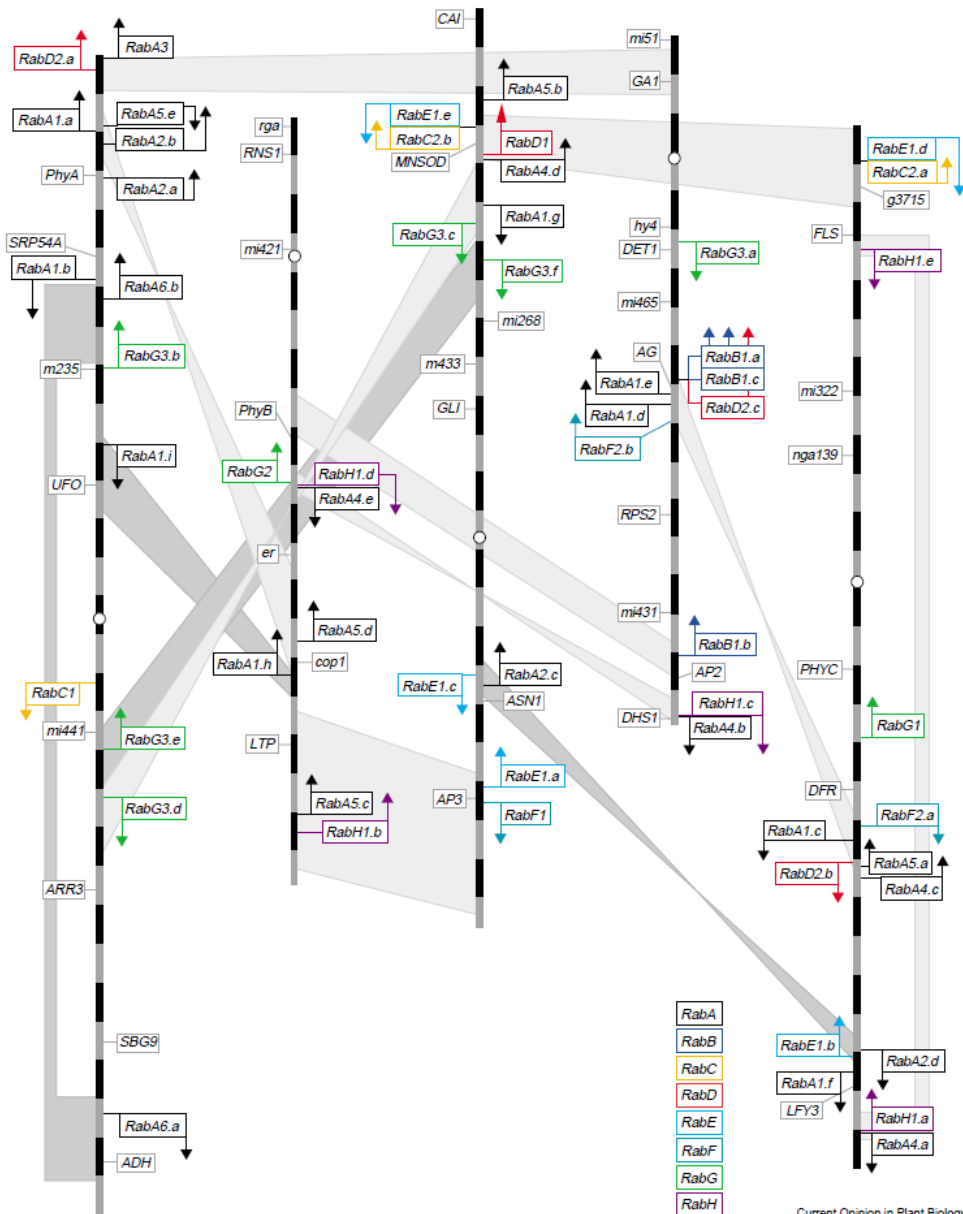
- Rab = “rat brain” proteins, low  $M_w$
- Largest family of small GTPases (57 Arabidopsis, 52 Rice)
- Grouped into 8 classes (RabA-H):

Rab1/RabD	Rab2/RabB
Rab5/RabF	Rab6/RabH
Rab7/RabG	Rab8/RabE
Rab11/RabA	Rab18/RabC

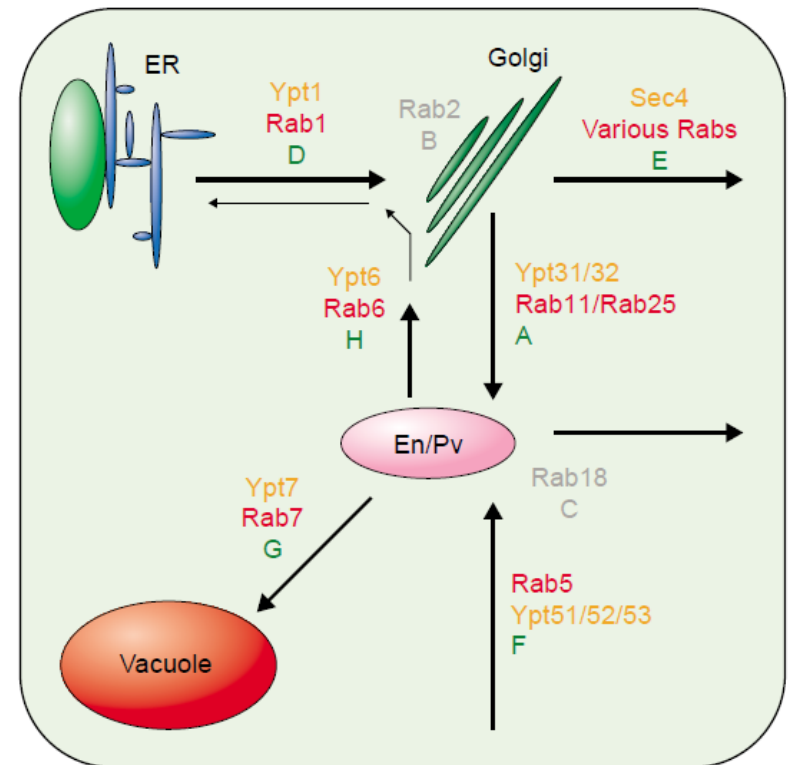
# Small Rab GTPases

- Rab = “rat brain” proteins, low  $M_w$
- Largest family of small GTPases (57 Arabidopsis, 52 Rice)
- Grouped into 8 classes (RabA-H):
 

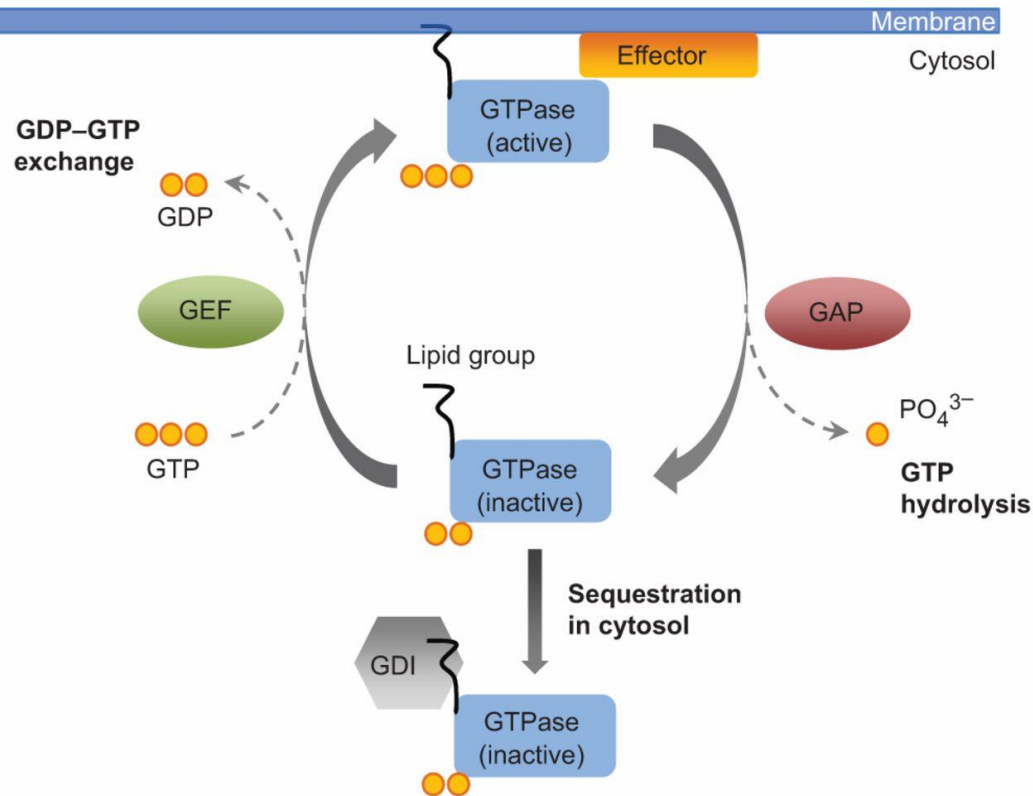
Rab1/RabD	Rab2/RabB
Rab5/RabF	Rab6/RabH
Rab7/RabG	Rab8/RabE
Rab11/RabA	Rab18/RabC



Current Opinion in Plant Biology

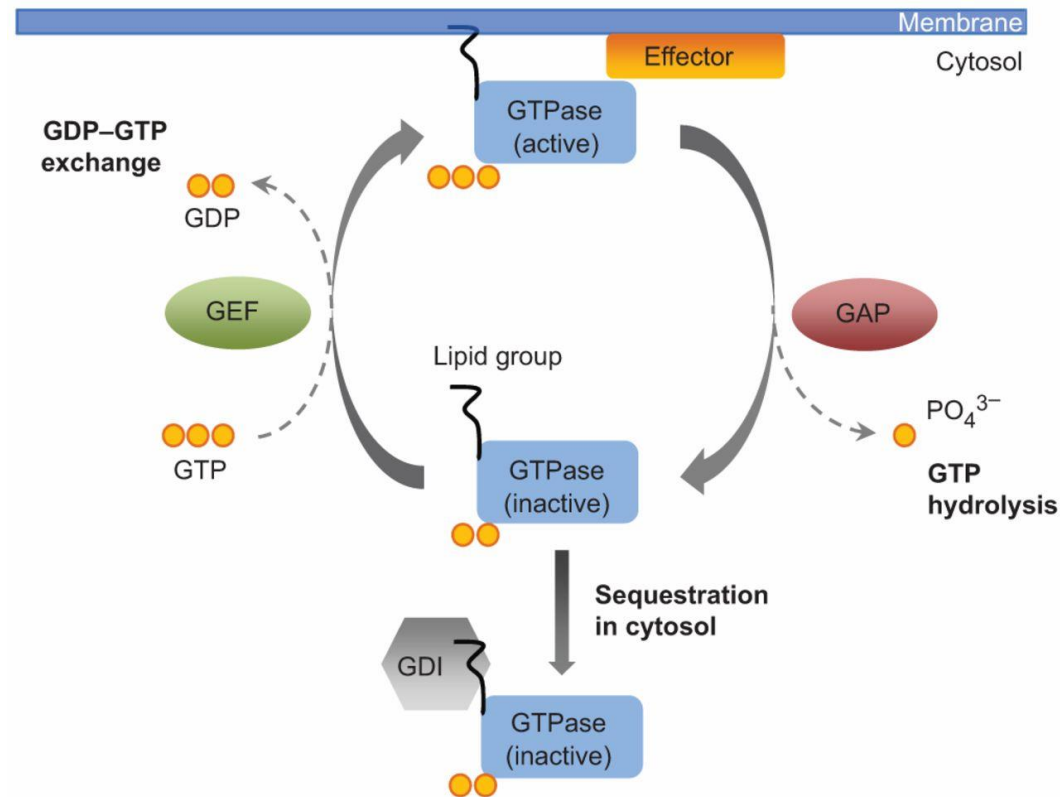


# GDP-GTP exchange cycle of small GTPases

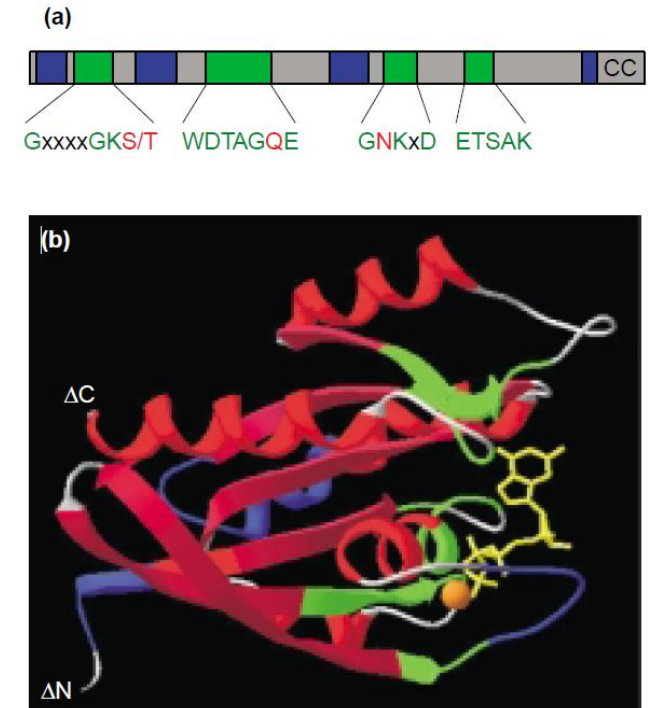


GAP – GTPase-activating proteins  
GEF – guanine exchange factor  
GDI – GDP-displacement inhibitor

# GDP-GTP exchange cycle of small GTPases



GAP – GTPase-activating proteins  
 GEF – guanine exchange factor  
 GDI – GDP-displacement inhibitor

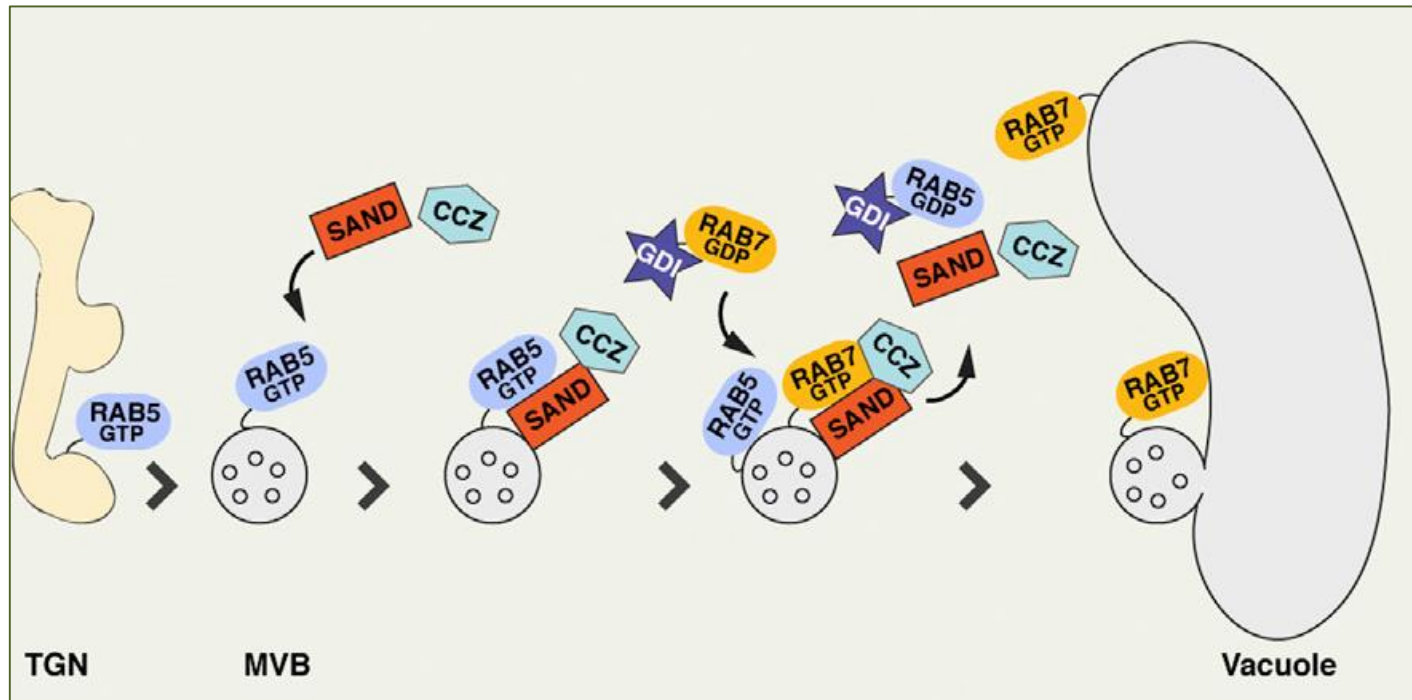


**Dominant negative (DN):** constitutively GDP-bound or inhibited GTP hydrolysis

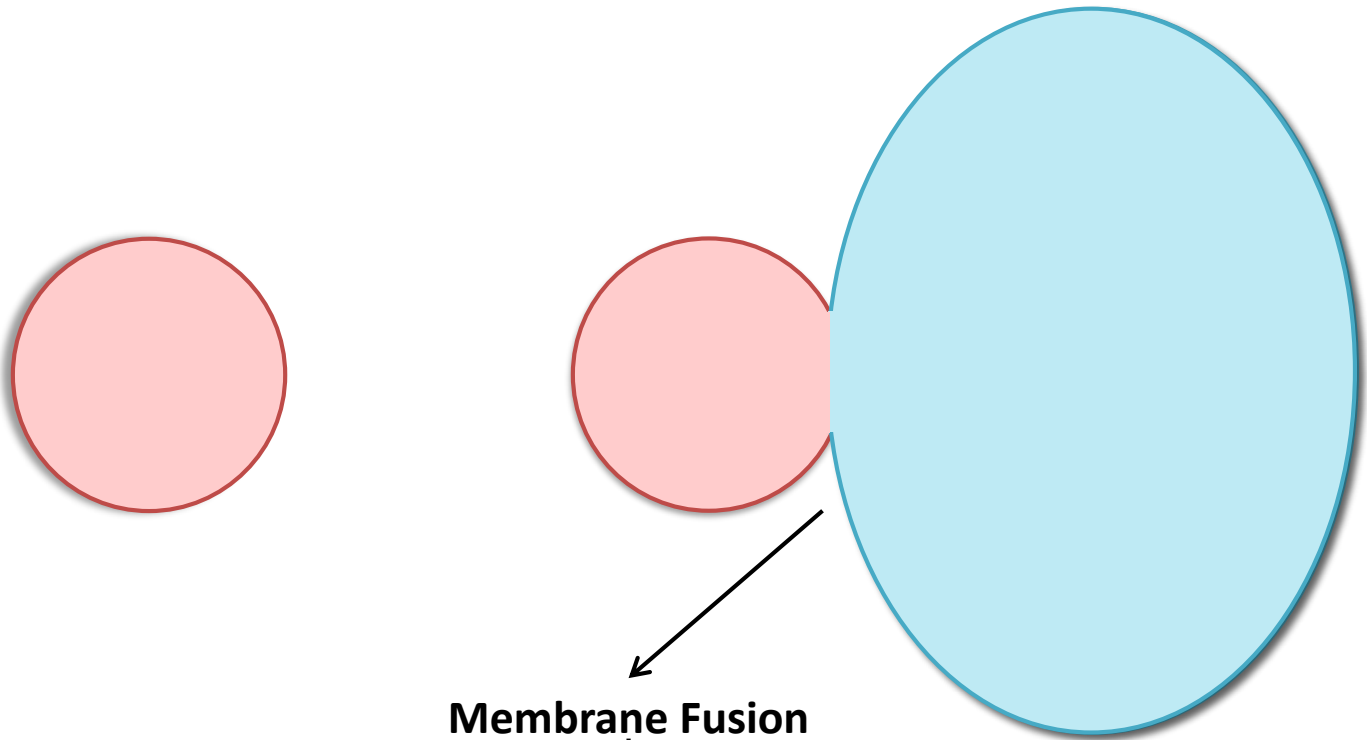
**Constitutively active (CA):** constitutively GTP-bound



# Rab5 to Rab7 conversion on vacuolar trafficking pathway



# “SNARE – Ware”



**Membrane Fusion**

energetically unfavorable

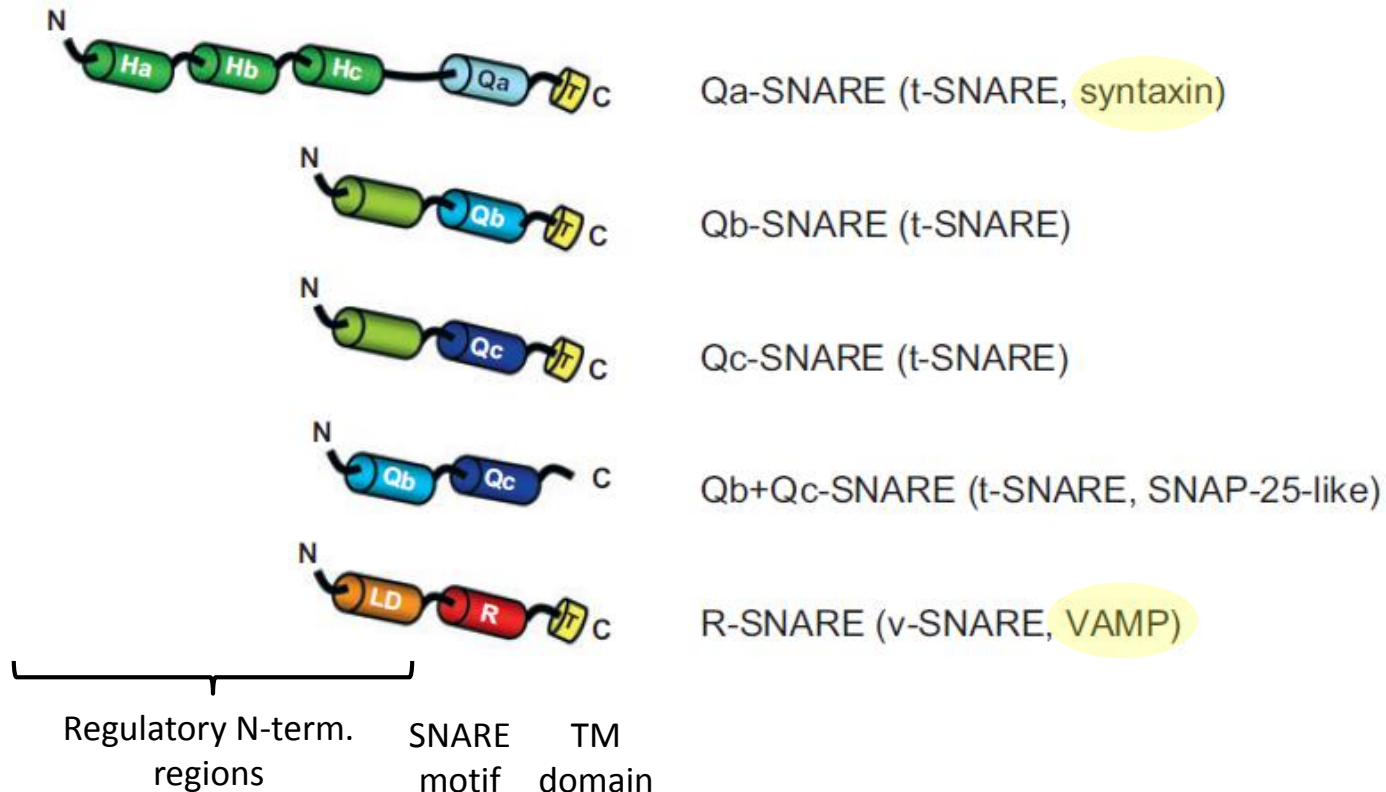
**SNAREs as mediator**

# “SNARE – Ware”

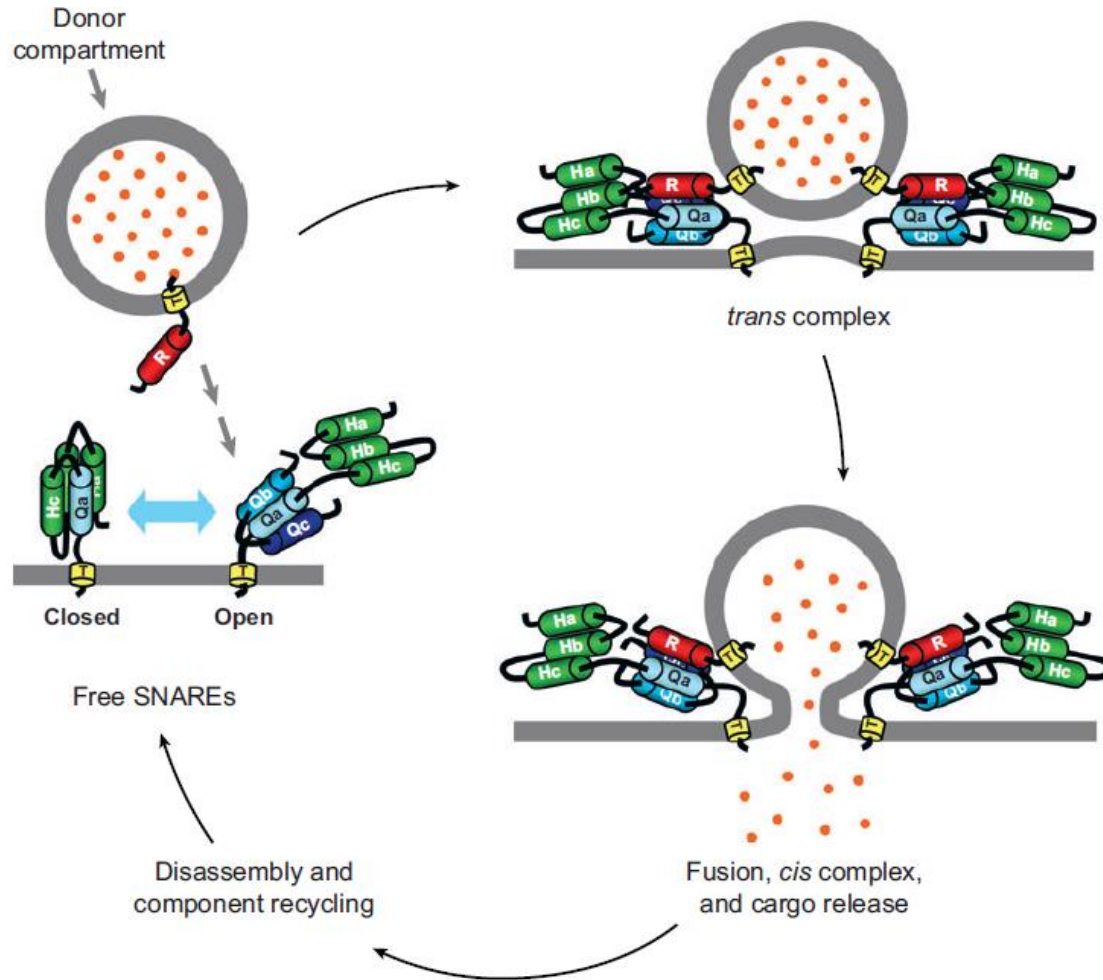
Functional classification: **t**-SNARE – at **t**arget membrane  
**v**-SNARE – at **v**esicle membrane

Structural classification:

200 – 400 aa

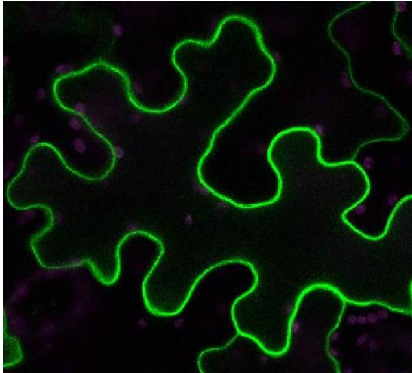


# "SNARE – Ware"

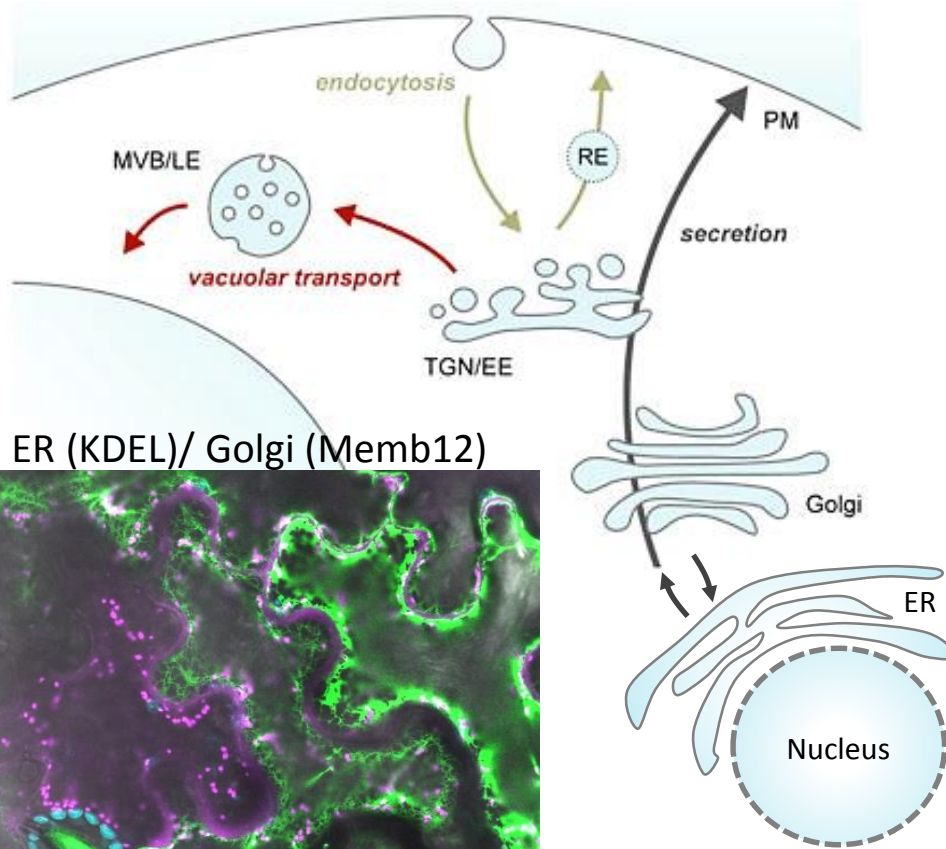
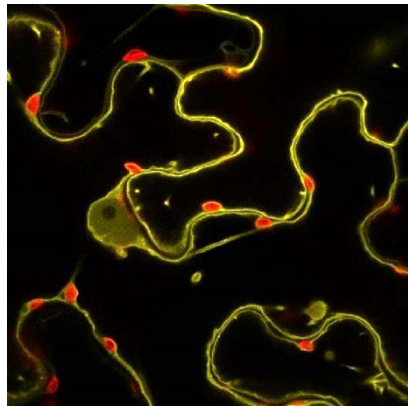


# Endomembrane Marker Proteins

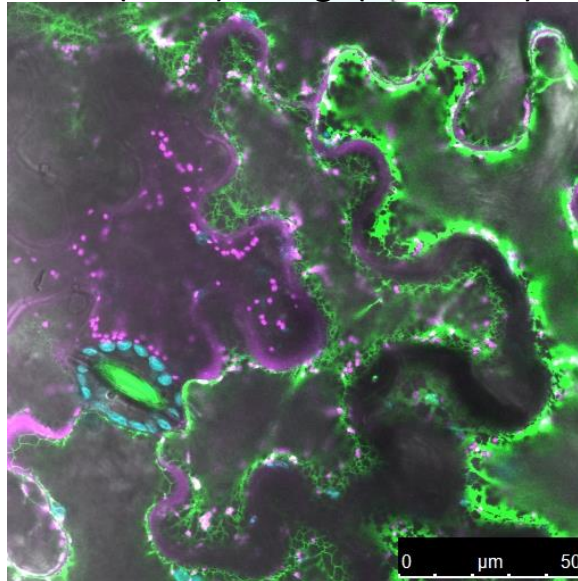
PM – FLS2



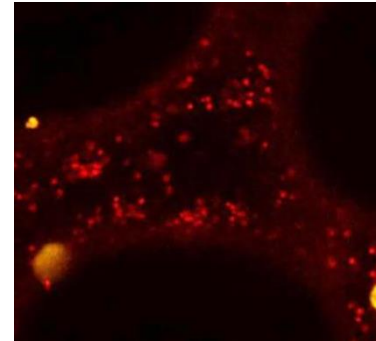
Vacuole – RabG3F



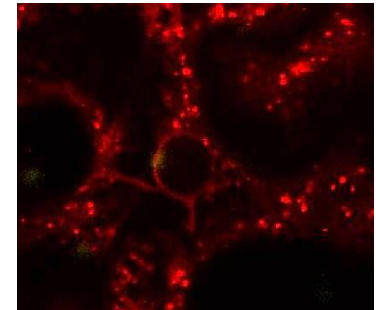
ER (KDEL)/ Golgi (Memb12)



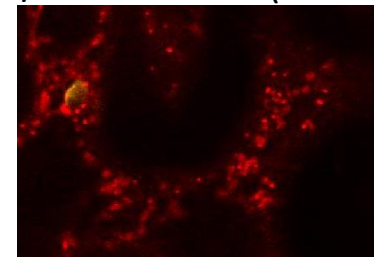
TGN – Vha-A1



EE/LE – ARA7 (RabF2b)



LE/MVB – ARA6 (RabF1)





# Endomembrane Marker Proteins

**ARA7** (Rab5/ RabF2b) –  
TGN/MVB

**ARA6** (Rab5/ RabF1) –  
MVB/ tonoplast

**RabG3F** (Rab7) –  
tonoplast

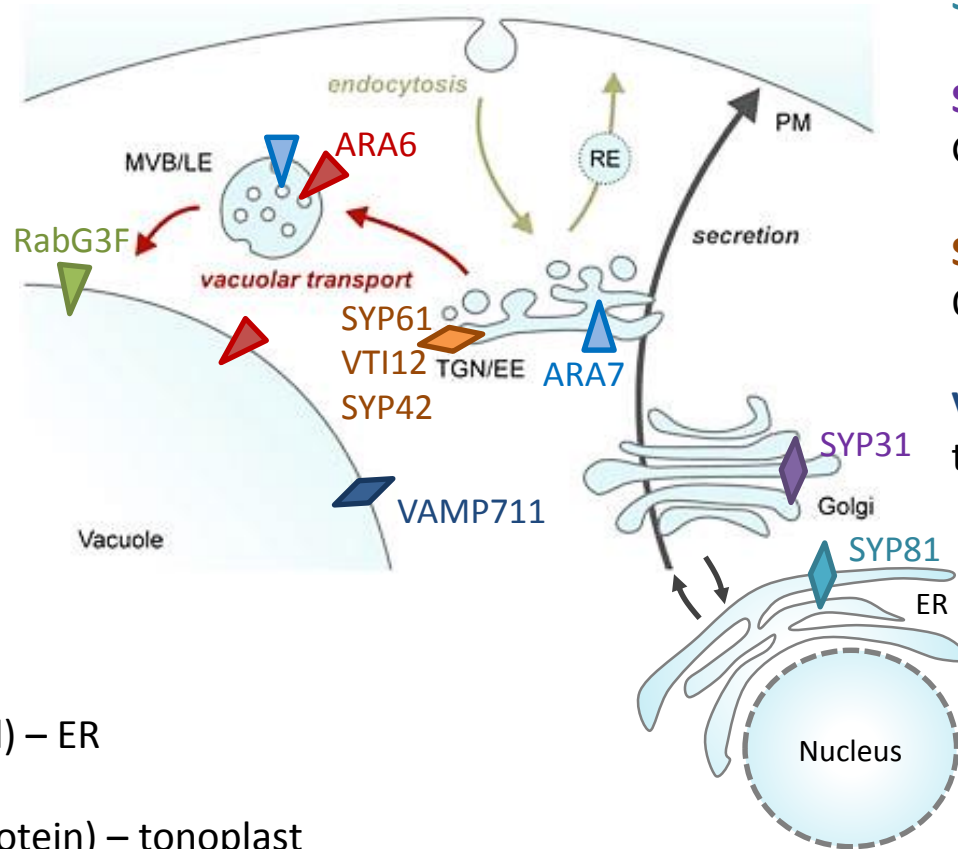
## Organelle Marker:

HDEL (ER-retention signal) – ER

Mannosidase – Golgi

TIP (tonoplast intrinsic protein) – tonoplast

**FM4-64** – endocytic tracer



**SYP81** – Qa-SNARE – ER

**SYP31** – Qa-SNARE –  
Golgi

**SYP61/ SYP42/ VTI12** –  
Q-SNAREs – TGN

**VAMP711** – R-SNARE –  
tonoplast

# Endomembrane Marker Proteins

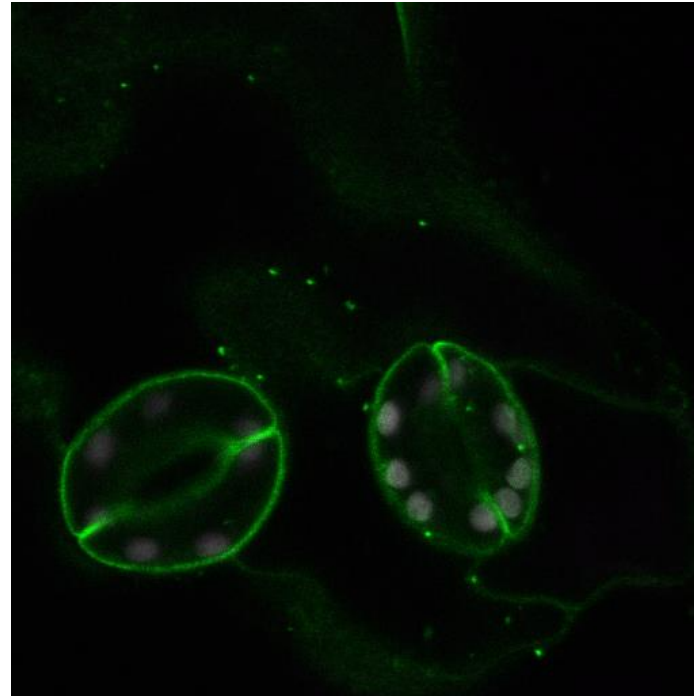
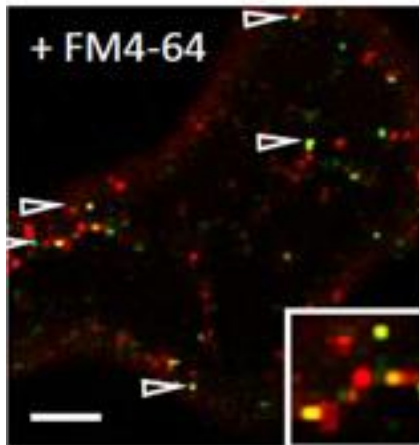
Markers:

Compartment	Construct	Reference
ER: Endoplasmic reticulum	<u>Sno<sup>a</sup></u> (EGFP)	Cutler et al., 2000
Golgi	Memb12-mCherry	Geldner et al., 2009
	ST-YFP	Zheng et al., 2004
TGN/EE: Trans-Golgi Network/Early Endosome	VHAa1-mRFP/-GFP	Dettmer et al., 2006
	CFP/mRFP-SYP61	Drakakaki et al., 2012; Robert et al., 2008
	YFP-Rab-A2	Chow et al., 2008
	VHA-a1-GFP/RFP	Dettmer et al., 2006
	SYP41-mRFP	Dettmer et al., 2006
	YFP-SCAMP1	Lam et al., 2007
	Ara7/RabF2b-mRFP	Ueda et al., 2004; Ebine et al., 2011
MVB/PVC/LE: Multivesicular body, prevacuolar compartment or late endosome	ARA6/RabF1-mRFP	Ueda et al., 2004; Ebine et al., 2011
Vacuole	VAMP711-YFP	Geldner et al., 2009
PM: Plasma Membrane	NPSN12	Geldner et al., 2009
Nucleoplasm	mRFP-H2B	He et al., 2015
Endocytic tracer that internalizes via endocytosis from the plasma membrane to the <u>tonoplast</u> .	FM4-64	Bolte et al., 2004; Jelínková et al., 2010

# Practical: Tracing the Transport Route of FLS2

FLS2-GFP is internalised into vesicles upon flg22-treatment

FLS2-GFP vesicles are labelled by FM4-64



What is the nature of FLS2-GFP positive endosomes?

→ Co-localisation with:

SYP61 (TGN)

ARA7 (TGN/MVB)

ARA6 (MVB/tonoplast)