

User manual for QC

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4/06/2022

Here, we show how to use CellDestiny as a package using lentiviral barcoding data studied in https://github.com/TeamPerie/HadjAbed-et-al._2022.

In this script we visualise key QC steps of the data before proceeding to make comparisons between cDC1 and cDC2 dendritic cells subtypes in three mice.

Install the package and load libraries

```
library(devtools)
devtools::install_github("TeamPerie/CellDestiny", quiet = TRUE)
library(CellDestiny)
library(ggplot2)
```

Load data and give duplicat variable name

Like for the application format, the first step of the QC part of the package format is to load count and metadata matrices and give the name of the variable describing your *duplicates*. It corresponds to one of your metadata column name.

```
# set working directory
setwd(getwd())
# import files
count_matrix <- read.csv("../testData/LentiviralBarcodingData/QC_data/QC_duplicate_matrix_Mouse_Lung")
metadata <- read.csv("../testData/LentiviralBarcodingData/QC_data/QC_duplicate_matrix_Mouse_Lung_cDC")

metadata
```

```
##      type mouse duplicates
## 1 cDC1      2            a
## 2 cDC2      4            b
## 3              5
```

Here, it is “duplicates”.

```
# Common parameters
dup_var="duplicates"
dup_val=metadata$duplicates
```

Reformat matrix for QC

The first function to call is `ReformatQCmatrix()` that calcul correlations and transforms your count matrix in a way that fits `MakeDuplicatesMatrix()` or `MakeRepeatUseMatrix()` input matrix format.

```
qc_mat<-ReformatQCmatrix(count_matrix, metadata, dup_var, dup_val,
                          sampleNameFieldsep = "_", transformation = "arcsin")

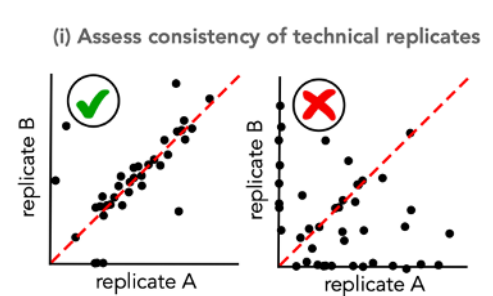
# Here, sampleNameFieldsep and transformation parameters are set to default
# ones.
# The transformation is applied to duplicat columns and saved in trans_dup1 and
# trans_dup2 column names as follow.

head(qc_mat)
```

```
##   Sample_names      Barcodes      a      b total_read type mouse  cor
## 1      cDC1_2 AACGTACAAC TCACA 3.101978 424.9833  428.0853 cDC1    2 0.94
## 2      cDC1_2 AACGTACAAC TCACA 3.101978 424.9833  428.0853 cDC1    2 0.94
## 3      cDC1_2 AACGTACAAC TCACA 3.101978 424.9833  428.0853 cDC1    2 0.94
## 4      cDC1_2 AACGTACAAC TCACA 3.101978 424.9833  428.0853 cDC1    2 0.94
## 5      cDC1_2 AACGTACAAC TCACA 3.101978 424.9833  428.0853 cDC1    2 0.94
## 6      cDC1_2 AACGTACAAC TCACA 3.101978 424.9833  428.0853 cDC1    2 0.94
##   trans_dup1 trans_dup2
## 1    1.850211    6.745199
## 2    1.850211    6.745199
## 3    1.850211    6.745199
## 4    1.850211    6.745199
## 5    1.850211    6.745199
## 6    1.850211    6.745199
```

Assess the frequency of repeat-use barcodes

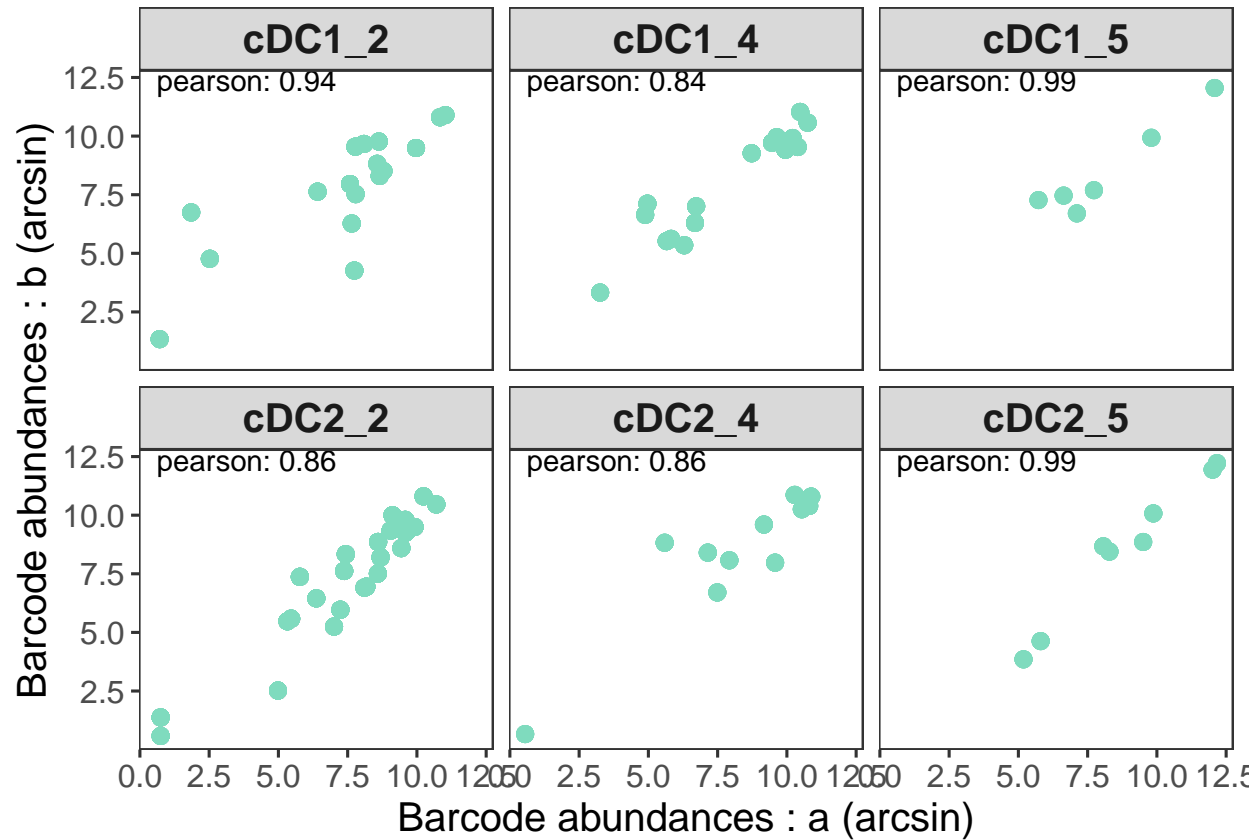
The integration of the same barcode into multiple cells, called repeat usage, is also an important QC metric that should be considered in a lineage tracing analysis pipeline, as a high incidence of repeat usage may lead to false lineage relationship assignments. The transfer of progenitors from the same transduction batch into at least two separate mice, followed by subsequent comparison of the barcodes recovered from those mice, can be used to estimate the frequency of repeat barcode use within one mouse.



We want to plot duplicates of all samples, not specific ones. To do so, we select all values from a variable (here variable “type” and its values : “cDC1” and “cDC2”).

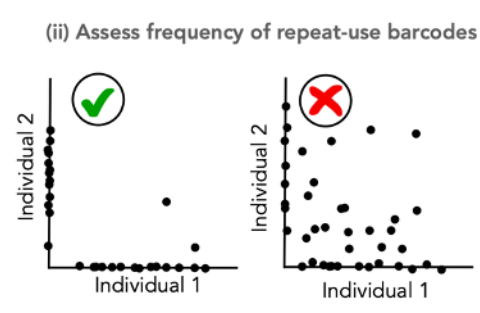
```
# parameter describing our cell types
list_var = c("type")
list_val = metadata$type

dup_mat<-MakeDuplicatesMatrix(matrix = qc_mat,
                              listVar = list_var, listVal = list_val,
                              metadata = metadata)
PlotDuplicates(dup_mat, dup_val, transformation = "arcsin")
```



Repeat Use checking

Here we assess the frequency of repeat use barcodes in the data. Repeat used barcodes are compared between individuals. Hence, fill out the variable name describing your individuals and all its values.

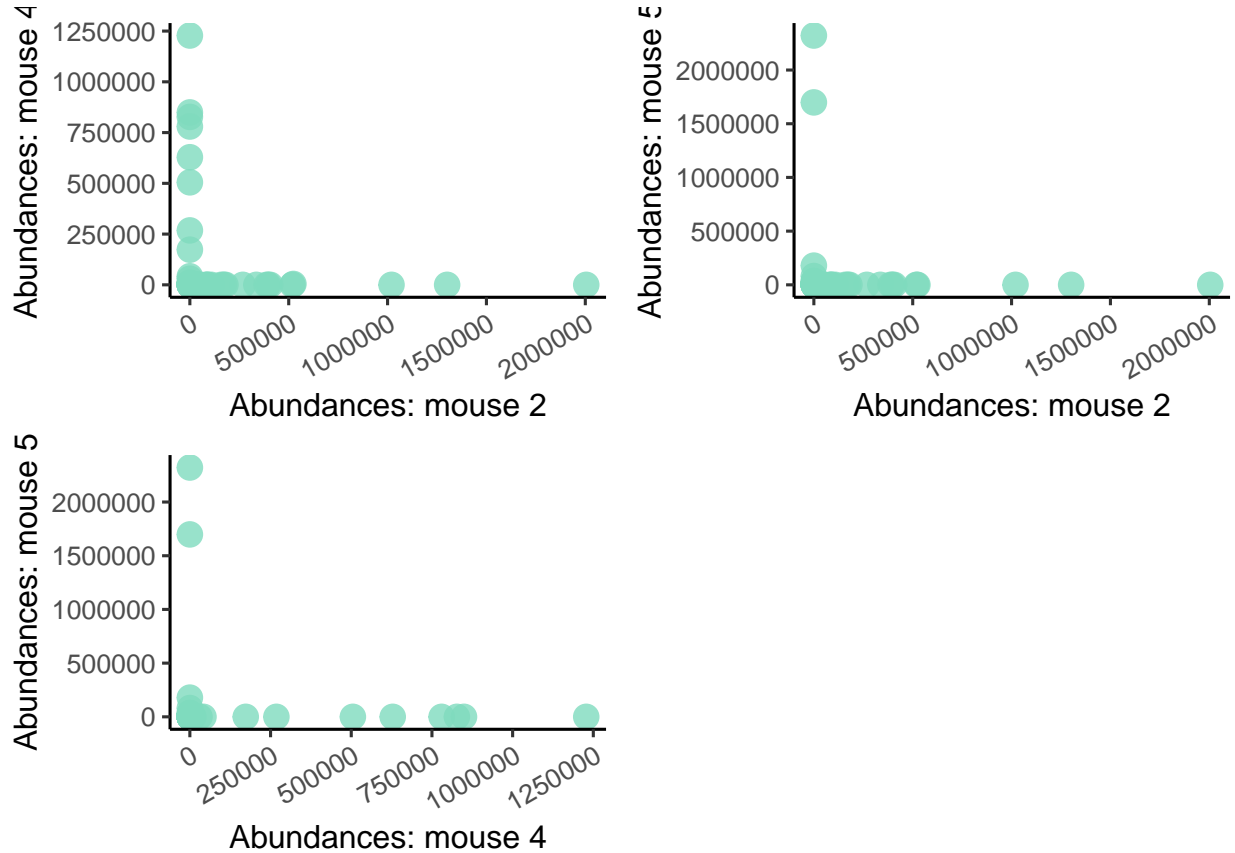


```

# parameter describing our cell types
list_var = c("type")
list_val = metadata$type
#parameters
indiv_var="mouse"
indiv_val=metadata$mouse

ru_mat<-MakeRepeatUseMatrix(qc_mat, indiv_var, indiv_val)
PlotRepeatUse(ru_mat, indiv_var, textSize = 12)

```



Both duplicates and repeat use checking are ok. We can now go further in the biological analysis. Open `2.User_manual_for_analysis.html` file.