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HUMAN THERAPEUTIC AGENTS

Abstract

Human therapeutic treatment compositions comprising a curcumin component, a harmine component, and an isovanillin component, preferably all three in combination. The agents are synergistically effective for treatment of human conditions, especially human cancers, such as pancreatic cancers.

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Background/Summary

CROSS-REFERENCE TO RELATED APPLICATIONS [0001] This application is a continuation of U.S. application Ser. No. 18/629,507, filed Apr. 8, 2024, which is a continuation of U.S. application Ser. No. 17/589,060, filed Jan. 31, 2022 (issued as U.S. Pat. No. 11,951,099 on Apr. 9, 2024), which is a continuation of U.S. application Ser. No. 17/000,164 filed Aug. 21, 2020 (issued as U.S. Pat. No. 11,266,634 on Mar. 8, 2022), which is a continuation of U.S. application Ser. No. 16/724,979 filed Dec. 23, 2019 (issued as U.S. Pat. No. 10,751,330 on Aug. 25, 2020), which is a continuation of U.S. application Ser. No. 16/541,665 filed Aug. 15, 2019 (issued as U.S. Pat. No. 10,576,067 on Mar. 3, 2020), which is a continuation of US application SN 16,213,774 filed Dec. 7, 2018 (issued as U.S. Pat. No. 10,471,049 on Nov. 12, 2019), which is a continuation of U.S. application Ser. No. 15/826,101 filed Nov. 29, 2017, which is a continuation of U.S. application Ser. No. 15/337,987 filed Oct. 28, 2016 (issued as U.S. Pat. No. 9,907,786 on Mar. 6, 2018), which is a continuation-in-part of PCT application SN PCT/US2015/055968 filed Oct. 16, 2015, which is a continuation-in-part of U.S. utility application Ser. No. 14/721,011 filed May 26, 2015 (issued as U.S. Pat. No. 9,402,834 on Aug. 2, 2016), and which claims the benefit of U.S. Provisional Applications, Ser. 62/184,051 filed Jun. 24, 2015, Ser. 62/161,090 filed May 13, 2015, and Ser. 62/066,686 filed Oct. 21, 2014. U.S. application Ser. No. 15/337,987 filed Oct. 28, 2016, is also a continuation of PCT Application SN PCT/IB2016/000723 filed Apr. 20, 2016, which is a continuation-in-part of U.S. utility application Ser. No. 14/721,011 filed May 26, 2015 (issued as U.S. Pat. No. 9,402,834 on Aug. 2, 2016), and which claims the benefit of U.S. Provisional Application Ser. 62/184,051 filed Jun. 24, 2015, and Ser. 62/161,090 filed May 13, 2015. Each of the above provisional, non-provisional, and PCT applications is incorporated by reference herein in its entirety.

BACKGROUND OF THE INVENTION

Field of the Invention

[0002] The present invention relates to combination chemotherapeutics for treatment of humans, and especially for the treatment of human cancers, and corresponding methods for the treatment of humans suffering from cancers or other maladies. The invention further provides dosage forms and regimens for administration to human patients, and methods of formulating and administering such dosage forms to yield improvements in treatment outcomes. More particularly, the invention is concerned with the administration of specific chemotherapeutic dosage forms (e.g., liquid mixtures, capsules, pills, or tablets) containing one or more curcumin component(s), harmine component(s), and isovanillin component(s), and sub-combinations thereof.

Description of Related Art

[0003] Cancer is a generic term for a large group of diseases that can affect any part of the body. Other terms used are malignant tumors and neoplasms. One defining feature of cancer is the rapid creation of abnormal cells that grow beyond their usual boundaries, and which can then invade adjoining parts of the body and spread to other organs. This process is referred to as metastasis. Metastases are the major cause of death from cancer.

[0004] The transformation from a normal cell into a tumor cell is a multistage process, typically a progression from a pre-cancerous lesion to malignant tumors. These changes are the result of the interaction between a person's genetic factors and three categories of external agents, including:

[0005] physical carcinogens, such as ultraviolet and ionizing radiation [0006] chemical carcinogens, such as asbestos, components of tobacco smoke, aflatoxin (a food contaminant) and arsenic (a drinking water contaminant) [0007] biological carcinogens, such as infections from certain viruses, bacteria or parasites.

[0008] Some examples of infections associated with certain cancers: [0009] Viruses: hepatitis B and liver cancer, Human Papilloma Virus (HPV) and cervical cancer, and human immunodeficiency virus (HIV) and Kaposi sarcoma. [0010] Bacteria: *Helicobacter pylori* and stomach cancer. [0011] Parasites: schistosomiasis and bladder cancer.

[0012] Aging is another fundamental factor for the development of cancer. The incidence of cancer rises dramatically with age, most likely due to a buildup of risks for specific cancers that increase with age. The overall risk accumulation is combined with the tendency for cellular repair mechanisms to be less effective as a person grows older.

[0013] Tobacco use, alcohol use, low fruit and vegetable intake, and chronic infections from hepatitis B (HBV), hepatitis C virus (HCV) and some types of Human Papilloma Virus (HPV) are leading risk factors for cancer in low- and middle-income countries. Cervical cancer, which is caused by HPV, is a leading cause of cancer death among women in low-income countries. In high-income countries, tobacco use, alcohol use, and being overweight or obese are major risk factors for cancer.

[0014] The most common cancer treatment modalities are surgery, chemotherapy, and radiation treatments. All of these techniques have significant drawbacks in terms of side effects and patient discomfort. For example, chemotherapy may result in significant decreases in white blood cell count (neutropenia), red blood cell count (anemia), and platelet count (thrombocytopenia). This can result in pain, diarrhea, constipation, mouth sores, hair loss, nausea, and vomiting.

[0015] Biological therapy (sometimes called immunotherapy, biotherapy, or biological response modifier therapy) is a relatively new addition to the family of cancer treatments. Biological therapies use the body's immune system, either directly or indirectly, to fight cancer or to lessen the side effects that may be caused by some cancer treatments.

[0016] During chemotherapies involving multiple-drug treatments, adverse drug events are common, and indeed toxicities related to drug-drug interactions are one of the leading causes of hospitalizations in the US. Obach, R. S. "Drug-Drug Interactions: An Important Negative Attribute in Drugs." *Drugs Today* 39.5 (2003): 308-338. In fact, in any single-month period, one-fifth of all surveyed adults in the USA reported an adverse drug response. Hakkarainen, K. M. et al.

"Prevalence and Perceived Preventability of Self-Reported Adverse Drug Events—A Population-

Based Survey of 7,099 Adults.” *PLoS One* 8.9 (2013): e73166. A large-scale study of adults aged 57-85 found that 29% were taking more than five prescription medications and nearly 5% were at risk of major adverse drug-drug interactions. In the field of oncology, a review of over 400 cancer patients determined that 77% were taking drugs that were considered to have a moderately severe potential for adverse drug interactions, and 9% had major adverse drug interactions. Ghalib, M. S. et al. “Alterations of Chemotherapeutic Pharmacokinetic Profiles by Drug-Drug Interactions.” *Expert Opin. Drug Metabl. Toxicol* 5.2 (2009): 109-130.

[0017] Such interactions are a global health problem, and the WHO has determined that negative drug interactions are leading causes of morbidity and mortality around the world, with up to 7% of all hospitalizations in the US due to negative drug interactions. A recent survey of a single hospital shows that 83% of hospitalized patients were prescribed drug combinations with the potential to cause adverse reactions. Patel, P. S. et al. “A Study of Potential Adverse Drug-Drug Interactions Among Prescribed Drugs in a Medicine Outpatient Department of a Tertiary Care Teaching Hospital.” *J. Basic Clin. Pharm.* 5.2 (2014): 44-48.

[0018] Examples of famous negative drug interactions include the development of rhabdomyolysis, a severe muscle disease, when taking Simvastatin with Amiodarone. As a result, the FDA introduced a warning on the drug label about the interaction. The calcium channel blocker Mibefradif, taken for high blood pressure, was removed from the market because of the harmful interaction with drugs that work on the electrical activity of the heart.

[0019] Cancer cells are cells that, by definition, grow and divide without normal limitations. The unrestricted cell growth results in tumors, comprised of a variety of cell types. Treatments to fight cancer are frequently successful in killing the typical, differentiated cancer cells that form the majority of a solid tumor, otherwise known as the bulk cells. However even with the best treatment, the cancer may return a few months to years later (Prince, M. E. et al., “Cancer stem cells in head and neck squamous cell cancer.” *J. Clin. Oncol.* 26.17 (2008):2871-2875). For example, recurrence is frequently the case for pancreatic and head and neck cancer. It is now hypothesized that one of the key factors in the recurrence rate for cancers is the presence of cancer stem cells.

[0020] Cancer stem cells were not identified until the late 1990s and show two important properties of stem cells: 1) cancer stem cells can self-renew and, 2) cancer stem cells can differentiate into any other cell type (Bandhavkar, S. “Cancer stem cells: a metastasizing menace.” *Cancer Med.* (2016) doi:10.1002/cam4.629; Dick, J. E. “Stem cell concepts renew cancer research.” *Blood.* 112 (2008):4793-4807). While they make up only a small percentage of the total number of cells in a tumor, they compromise a unique category of cancer cells that are more likely to be resistant to chemotherapy or radiation therapy. In fact, it is now believed that the majority of cells in tumors are not cancer-causing and cannot initiate new tumors (Bandhavkar). Only cancer stem cells appear to be tumor-initiators (Visvader, J. E. et al. “Cancer stem cells: Current status and evolving complexities.” *Cell Stem Cell.* 10 (2012):717-728). Cancer stem cells have been shown to coordinate tumor cell growth, metastases (migration and invasion), and drug resistance (Cammarota, F. et al. “Mesenchymal stem/stromal cells in stromal evolution and cancer progression.” *Stem Cells Int.* (2016):4824573). These cancer stem cells behave differently than non-cancerous stem cells in the person (Cammarota et al.), and have been described as the “roots of aggressive tumors for which we have no effective treatment” (Doherty, M. R. et al. “Cancer stem cell plasticity drives therapeutic resistance.” *Cancers* 8,8 (2016) doi:10.3390). In general stem cells are naturally resistant to chemotherapies and radiation therapy (Diehn, M. et al. “Cancer stem cells and radiotherapy: new insights into tumor radioresistance.” *J. Natl. Cancer Inst.* 98 (2016):1775-1757; Mery, B. et al. “Targeting head and neck tumoral stem cells: From biological aspects to therapeutic perspectives.” *World J Stem Cells* 8.1 (2016):13-21), because they have chemical pumps that remove the chemotherapies out of the cells, thus they can have no effect on the stem cells. They also have a slow rate of turnover, and most radiation and chemotherapies are designed to only kill cells that are rapidly dividing, such as the majority of the cells within the tumor. These

characteristics explain, on a large scale, how stem cells associated with cancer are resistant to chemotherapy.

[0021] Current anticancer therapies may directly cause cancer cells to die or just inhibit their growth (Bandhavkar). If the anticancer therapy fails to target and remove the cancer stem cells, then relapse and drug resistance ensues. Selectively targeting and eliminating cancer stem cells would theoretically treat the primary tumors and halt any chance of recurrence (Mery et al.). Yet, the ability to kill cancer stem cells is currently considered a significant clinical challenge. Some recent evidence suggests that traditional chemotherapies can even induce the generation of new stem cells within tumors potentially making the cancer return faster (Doherty et. al).

[0022] Identification of the regulatory mechanisms and signaling pathways involved in cancer stem cells (CSCs) will help in designing novel agents to target this refractory cell population in pancreatic cancers. Cancer stem cells are capable of self-renewal and generating tumors resembling the primary tumor (Ponnuram, S. et al. "Quinomycin A targets Notch signaling pathway in pancreatic cancer stem cells." *Oncotarget* 7.3 (2015):3217-3232). The sphere-forming assays have been widely used to identify stem cells based on their reported capacity to evaluate self-renewal and differentiation.

[0023] U.S. Pat. No. 8,039,025 describes cancer treatments in the form of extracts of *Arum palaestinum* Boiss, supplemented with individual amounts of β -sitosterol, isovanillin, and linoleic acid, and this patent is incorporated by reference herein in its entirety.

[0024] Despite the immense amount of worldwide research and efforts to stem the tide of cancer and its side effects, the disease in its many manifestations continues to be a huge problem. Therefore, any new cancer treatment having a curative affect and/or the ability to ameliorate cancer symptoms and improve the lifestyle of patients is highly significant and important.

SUMMARY OF THE INVENTION

[0025] The present invention provides improved chemotherapeutics for treatment of humans, and especially in the treatment of human cancers, with novel combinations of compounds, which are useful against a wide variety of different cancers with minimal or nonexistent adverse side effects. Generally speaking, the chemotherapeutics of the invention comprise respective quantities of at least two of, curcumin component(s), harmine component(s), and isovanillin component(s). The preferred chemotherapeutics include all three of these components, but sub-combinations thereof are also useful, i.e., therapeutics comprising curcumin component(s) and harmine component(s), curcumin component(s) and isovanillin component(s), and harmine component(s) and isovanillin component(s). In particularly preferred embodiments, the active components of the compositions (i.e., those having a significant therapeutic effect) consist essentially of curcumin, harmine, and isovanillin components in the case of three-component compositions, and consist essentially of two of the three components in the case of the two-component compositions. Preferably, the at least one curcumin component comprises curcumin, the at least one harmine component comprises harmine, and the at least one isovanillin component comprises isovanillin or vanillin.

[0026] The invention also provides new methods for treatment of cancers by administration of appropriate quantities of the anti-cancer compositions hereof. Hence, the compositions are particularly designed for use in the treatment of cancers, and the compositions can be used for the manufacture of medicaments of anti-cancer therapeutic applications. In addition, the invention provides pharmaceutical compositions for the treatment of cancers comprising administering therapeutically effective amounts of the new compositions, prepared by processes known per se, with a pharmaceutically acceptable carrier.

[0027] Curcumin (CAS #458-37-7) is a diaryl heptanoid, and has the molecular formula $C_{21}H_{20}O_6$. Curcumin occurs as a part of a curcuminoid plant extract containing curcumin, demethoxycurcumin, and bis-demethoxycurcumin. One commercially available effective curcuminoid is sold as "Curcumin from *Curcuma Longa* (Turmeric)," which contains greater than 65% by weight curcumin, as determined by HPLC analysis.

[0028] Harmine (CAS #442-51-3) is a methoxy methyl pyrido indole belonging to the O-carboline family of compounds, and has the molecular formula C₁₃H₁₂N₂O. Harmine occurs in a number of different plants native to the Middle East and South America.

[0029] Isovanillin (CAS #621-59-0) is a phenolic aldehyde vanillin isomer, and has the molecular formula C₈H₈O₃.

[0030] A “chemotherapeutic” or “chemotherapeutic agent” as used herein refers to the combinations of chemical compounds described herein as useful in the treatment of human conditions, especially human cancers. Chemotherapeutics may be cytostatic, selectively toxic or destructive of cancerous tissue and/or cells, but also include indiscriminately cytotoxic compounds used in cancer treatments.

[0031] The combination therapeutic agents of the invention have been found to be effective in the treatment of a broad spectrum of human cancers, and also to other conditions, such as elevated PSA counts in men. The broad scope of utility with the agents of the invention is in itself highly unusual. However, this feature, together with the nonexistent or minimal side effects induced by the agents, represents a startling development in the art.

Description

BRIEF DESCRIPTION OF THE DRAWINGS

[0032] FIG. 1 is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof in inducing the death of two different types of human head and neck cancer cells, as described in Example 1;

[0033] FIG. 2A is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof GZ17-6.02 in inducing the death of pediatric leukemia cells, as described in Example 2;

[0034] FIG. 2B is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof GZ17-6.02 in inducing the death of pediatric osteosarcoma cells, as described in Example 2;

[0035] FIG. 3A is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof GZ17-6.02 in inducing the death of lymphoma cells, as described in Example 3;

[0036] FIG. 3B is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof GZ17-6.02 in inducing the death of lung cancer cells, as described in Example 3;

[0037] FIG. 4A is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof GZ17-6.02 in inducing the death of ovarian cancer cells, as described in Example 4;

[0038] FIG. 4B is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof in inducing the death of prostate cancer cells, as described in Example 4;

[0039] FIG. 5A is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof in inducing the death of human breast cancer cells, as described in Example 5;

[0040] FIG. 5B is a graph of cell number versus dosage amounts of GZ17-6.02, illustrating the effect thereof in inducing the death of pancreatic cancer cells, as described in Example 5;

[0041] FIG. 6 is a graph of cell number versus doses of GZ17-6.02 illustrating the relative effects thereof in inducing cell death in prostate cancer and ovarian cancer cells, as compared with non-cancerous fibroblasts, as described in Example 6;

[0042] FIG. 7A is a graph illustrating the effect of GZ17-6.02 in preventing migration of head and neck cancer cells, as described in Example 7;

[0043] FIG. 7B is a graph illustrating the effect of GZ17-6.02 in preventing invasion of head and neck cancer cells, as described in Example 7;

[0044] FIG. 8 is a graph of normalized cell number versus increasing doxorubicin doses alone and with the addition of two different concentrations of GZ17-6.02, as described in Example 8;

[0045] FIG. **9A** is a graph illustrating extent of apoptosis in a control test, as described in Example 9, and illustrating about 19.8% cell death via apoptosis;

[0046] FIG. **9B** is a graph illustrating extent of apoptosis in a test identical to the control test of FIG. **9A**, but including GZ17-6.02, as described in Example 9, and illustrating about 48.2% cell death via apoptosis;

[0047] FIG. **10A** is a graph illustrating that caspase 3 and 7 concentrations increased in response to GZ17-6.02, but caspase 9 levels did not change in response to GZ17-6.02 in lung cancer cells, as described in Example 10;

[0048] FIG. **10B** is a graph illustrating that caspase 6 concentrations increased in response to GZ17-6.02 in lung cancer cells, as described in Example 10;

[0049] FIG. **10C** is a graph illustrating that ATP levels, as a marker of mitochondrial toxicity, were not increased by GZ17-6.02 in lung cancer cells, as described in Example 10;

[0050] FIG. **11A** is a graph illustrating the mechanisms of ovarian cancer cells death by application of GZ17-6.02, as explained in Example 11;

[0051] FIG. **11B** is a graph illustrating the mechanisms of ovarian cancer cells death by application of GZ17-6.02, as explained in Example 11;

[0052] FIG. **11C** is a graph illustrating the mechanisms of ovarian cancer cells death by application of GZ17-6.02, as explained in Example 11;

[0053] FIG. **11D** is a graph illustrating the mechanisms of ovarian cancer cells death by application of GZ17-6.02, as explained in Example 11;

[0054] FIG. **11E** is a graph illustrating the mechanisms of ovarian cancer cells death by application of GZ17-6.02, as explained in Example 11;

[0055] FIG. **12A** is a graph illustrating the mechanisms of osteosarcoma cells death by application of GZ17-6.02, as explained in Example 12;

[0056] FIG. **12B** is a graph illustrating the mechanisms of osteosarcoma cells death by application of GZ17-6.02, as explained in Example 12;

[0057] FIG. **12C** is a graph illustrating the mechanisms of osteosarcoma cells death by application of GZ17-6.02, as explained in Example 12;

[0058] FIG. **12D** is a graph illustrating the mechanisms of osteosarcoma cells death by application of GZ17-6.02, as explained in Example 12;

[0059] FIG. **13** is a graph illustrating the mechanisms of human head and neck cancer cells death by application of GZ17-6.02, as explained in Example 13;

[0060] FIG. **14A** is a control quantitative dot-blot, which measures the relative amount of proteins, known to be involved in cell proliferation described in Example 14, where the proteins were epidermal growth factor receptor (EGFR), extracellular-signal-regulated kinase (ERK1/2), the catalytic subunit of AMP-activated kinase (AMPK α 2), β -catenin, and Chk-2.

[0061] FIG. **14B** is a quantitative dot-block similar to that described in FIG. **14A**, but illustrating the amounts of the test proteins upon application of GZ17-6.02, as described in Example 14;

[0062] FIG. **15A** is a graph depicting the results of two independent scientists, each carrying out an identical induced cell death test with GZ17-6.02 on ovarian cancer cells, as described in Example 15;

[0063] FIG. **15B** is a graph depicting the results of two independent scientists, each carrying out an identical induced cell death test with GZ17-6.02 on lung cancer cells, as described in Example 4; and

[0064] FIG. **16A** is a comparative graph of tumor volume versus days after cancer cell inoculation in mice, between control inoculations (ethanol vehicle) and test inoculations containing the vehicle and GZ17-6.02, illustrating the dramatic reduction in tumor volumes in the test mice, as explained in Example 16;

[0065] FIG. **16B** is a comparative graph of contralateral tumor volume versus days after cancer cell inoculation in mice, indicating a systemic effect of use of GZ17-6.02, as explained in Example 16;

[0066] FIG. 16C is a graph of fractional tumor volume versus days after treatment wherein one group of mice was implanted with human head and neck tumor cells using an implantation vehicle, and a control group of mice was implanted with only the vehicle, in order to determine tumor volume over time, as set forth in Example 16;

[0067] FIG. 17A is a graph of cell number versus dosage amounts of GZ17-6.02 (open circles) versus a combined product including $\frac{1}{3}$ by weight of each GZ17-6.02 component (filled circles), tested on ovarian cancer cells, as described in Example 4;

[0068] FIG. 17B is a graph of cell number versus dosage amounts of GZ17-6.02 (open circles) versus a combined product including $\frac{1}{3}$ by weight of each GZ17-6.02 component (filled circles), tested on lung cancer cells, as described in Example 3;

[0069] FIG. 17C is a graph of cell number versus dosage amounts of GZ17-6.02 (open circles) versus a combined product including $\frac{1}{3}$ by weight of each GZ17-6.02 component (filled circles), tested on prostate cancer cells, as described in Example 4;

[0070] FIG. 18A is a graph of percent ovarian cancer cell death versus different component combinations of GZ17-6.02, illustrating results using isovanillin alone, and two-component products respectively including isovanillin plus curcumin, and isovanillin plus harmine, where the isovanillin concentration was held constant throughout;

[0071] FIG. 18B is a graph of percent lung cancer cell death versus different component combinations of GZ17-6.02, illustrating results using isovanillin alone, and two-component products respectively including isovanillin plus curcumin, and isovanillin plus harmine, where the isovanillin concentration was held constant throughout;

[0072] FIG. 18C is a graph of percent prostate cancer cell death versus different component combinations of GZ17-6.02, illustrating results using isovanillin alone, and two-component products respectively including isovanillin plus curcumin, and isovanillin plus harmine, where the isovanillin concentration was held constant throughout;

[0073] FIG. 18D is a graph of percent lymphoma cancer cell death versus different component combinations of GZ17-6.02, illustrating results using isovanillin alone, and two-component products respectively including isovanillin plus curcumin, and isovanillin plus harmine, where the isovanillin concentration was held constant throughout;

[0074] FIG. 19A is a graph of percent ovarian cancer cell death versus different component combinations of GZ17-6.02, illustrating results using curcumin alone, and two-component products respectively including curcumin plus isovanillin, and curcumin plus harmine, where the curcumin concentration was held constant throughout;

[0075] FIG. 19B is a graph of percent lung cancer cell death versus different component combinations of GZ17-6.02, illustrating results using curcumin alone, and two-component products respectively including curcumin plus isovanillin, and curcumin plus harmine, where the curcumin concentration was held constant throughout;

[0076] FIG. 19C is a graph of percent prostate cancer cell death versus different component combinations of GZ17-6.02, illustrating results using curcumin alone, and two-component products respectively including curcumin plus isovanillin, and curcumin plus harmine, where the curcumin concentration was held constant throughout;

[0077] FIG. 19D is a graph of percent lymphoma cancer cell death versus different component combinations of GZ17-6.02, illustrating results using curcumin alone, and two-component products respectively including curcumin plus isovanillin, and curcumin plus harmine, where the curcumin concentration was held constant throughout;

[0078] FIG. 20A is a graph of percent ovarian cancer cell death versus different component combinations of GZ17-6.02, illustrating results using harmine alone, and two-component products respectively including harmine plus isovanillin, and harmine plus curcumin, where the harmine concentration was held constant throughout;

[0079] FIG. 20B is a graph of percent lung cancer cell death versus different component

combinations of GZ17-6.02, illustrating results using harmine alone, and two-component products respectively including harmine plus isovanillin, and harmine plus curcumin, where the harmine concentration was held constant throughout;

[0080] FIG. **20C** is a graph of percent prostate cancer cell death versus different component combinations of GZ17-6.02, illustrating results using harmine alone, and two-component products respectively including harmine plus isovanillin, and harmine plus curcumin, where the harmine concentration was held constant throughout;

[0081] FIG. **20D** is a graph of percent lymphoma cancer cell death versus different component combinations of GZ17-6.02, illustrating results using harmine alone, and two-component products respectively including harmine plus isovanillin, and harmine plus curcumin, where the harmine concentration was held constant throughout;

[0082] FIG. **21A** is a graph of lymphoma cancer cell lethality using GZ17-6.02 at a dosage rate of 12 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0083] FIG. **21B** is a graph of lymphoma cancer cell lethality using GZ17-6.02 at a dosage rate of 24 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0084] FIG. **21C** is a graph of lymphoma cancer cell lethality using GZ17-6.02 at a dosage rate of 48 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0085] FIG. **21D** is a graph of lymphoma cancer cell lethality using GZ17-6.02 at a dosage rate of 96 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0086] FIG. **22A** is a graph of ovarian cancer cell lethality using GZ17-6.02 at a dosage rate of 12 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0087] FIG. **22B** is a graph of ovarian cancer cell lethality using GZ17-6.02 at a dosage rate of 24 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0088] FIG. **22C** is a graph of breast cancer cell lethality using GZ17-6.02 at a dosage rate of 24 $\mu\text{g/mL}$, and using the three components of GZ17-6.02 individually at the concentration present in GZ17-6.02, and further illustrating the theoretical additive effect of the three components versus GZ17-6.02;

[0089] FIG. **23A** is a graph of lung cancer cell number versus increasing dosage amounts of vanillin alone, as described in Example 1;

[0090] FIG. **23B** is a graph of lung cancer cell number versus increasing dosage amounts of isovanillic acid alone, as described in Example 1;

[0091] FIG. **23C** is a graph of lung cancer cell number versus increasing dosage amounts of 0-vanillin alone, as described in Example 1;

[0092] FIG. **23D** is a graph of lung cancer cell number versus increasing dosage amounts of isovanillyl alcohol alone, as described in Example 1;

[0093] FIG. **23E** is a graph of ovarian cancer cell number versus increasing dosage amounts of vanillin alone, as described in Example 1;

[0094] FIG. **23F** is a graph of ovarian cancer cell number versus increasing dosage amounts of

isovanillic acid alone, as described in Example 1;

[0095] FIG. 23G is a graph of ovarian cancer cell number versus increasing dosage amounts of 0-vanillin alone, as described in Example 1;

[0096] FIG. 23H is a graph of ovarian cancer cell number versus increasing dosage amounts of isovanillyl alcohol alone, as described in Example 1;

[0097] FIG. 23I is a graph of prostate cancer cell number versus increasing dosage amounts of vanillin alone, as described in Example 1;

[0098] FIG. 23J is a graph of prostate cancer cell number versus increasing dosage amounts of isovanillic acid alone, as described in Example 1;

[0099] FIG. 23K is a graph of prostate cancer cell number versus increasing dosage amounts of 0-vanillin alone, as described in Example 1;

[0100] FIG. 23L is a graph of prostate cancer cell number versus increasing dosage amounts of isovanillyl alcohol alone, as described in Example 1;

[0101] FIG. 24A is a graph of lung cancer cell number versus increasing dosage amounts of harmaline alone, as described in Example 1;

[0102] FIG. 24B is a graph of lung cancer cell number versus increasing dosage amounts of tetrahydro-harmine alone, as described in Example 1;

[0103] FIG. 24C is a graph of lung cancer cell number versus increasing dosage amounts of harmol hydrochloride alone, as described in Example 1;

[0104] FIG. 24D is a graph of lung cancer cell number versus increasing dosage amounts of harmalol hydrochloride dihydrate alone, as described in Example 1;

[0105] FIG. 24E is a graph of lung cancer cell number versus increasing dosage amounts of harmane alone, as described in Example 1;

[0106] FIG. 24F is a graph of ovarian cancer cell number versus increasing dosage amounts of harmaline alone, as described in Example 1;

[0107] FIG. 24G is a graph of ovarian cancer cell number versus increasing dosage amounts of tetrahydro-harmine alone, as described in Example 1;

[0108] FIG. 24H is a graph of ovarian cancer cell number versus increasing dosage amounts of harmol hydrochloride alone, as described in Example 1;

[0109] FIG. 24I is a graph of ovarian cancer cell number versus increasing dosage amounts of harmalol hydrochloride dihydrate alone, as described in Example 1;

[0110] FIG. 24J is a graph of ovarian cancer cell number versus increasing dosage amounts of harmane alone, as described in Example 1;

[0111] FIG. 24K is a graph of prostate cancer cell number versus increasing dosage amounts of harmaline alone, as described in Example 1;

[0112] FIG. 24L is a graph of prostate cancer cell number versus increasing dosage amounts of tetrahydro-harmine alone, as described in Example 1;

[0113] FIG. 24M is a graph of prostate cancer cell number versus increasing dosage amounts of harmol hydrochloride alone, as described in Example 1;

[0114] FIG. 24N is a graph of prostate cancer cell number versus increasing dosage amounts of harmalol hydrochloride dihydrate alone, as described in Example 1;

[0115] FIG. 24O is a graph of prostate cancer cell number versus increasing dosage amounts of harmane alone, as described in Example 1;

[0116] FIG. 25A is a graph of lung cancer cell number versus increasing dosage amounts of bisdemethoxy curcumin alone, as described in Example 1;

[0117] FIG. 25B is a graph of ovarian cancer cell number versus increasing dosage amounts of bisdemethoxy curcumin alone, as described in Example 1;

[0118] FIG. 25C is a graph of prostate cancer cell number versus increasing dosage amounts of bisdemethoxy curcumin alone, as described in Example 1;

[0119] FIG. 26 is a graph of ovarian cancer cell number versus dosage amounts of GZ17-6.02

where the product was stored at varying temperatures over two months, confirming that the product has long-term stability;

[0120] FIG. **27** is a graph of ovarian cancer cell number versus dosage amounts of GZ17-6.02, where the GZ17-6.02 was subjected to a series of successive freeze/thaw cycles, confirming that the product has excellent freeze/thaw stability;

[0121] FIG. **28A** is a graph of cell number versus dosage amounts of GZ17-8.02, illustrating the effect thereof in inducing the death of two different types ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0122] FIG. **28B** is a graph of cell number versus dosage amounts of GZ17-8.03, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0123] FIG. **28C** is a graph of cell number versus dosage amounts of GZ17-8.04, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0124] FIG. **28D** is a graph of cell number versus dosage amounts of GZ17-8.05, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0125] FIG. **28E** is a graph of cell number versus dosage amounts of GZ17-8.06, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0126] FIG. **28F** is a graph of cell number versus dosage amounts of GZ17-8.07, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0127] FIG. **28G** is a graph of cell number versus dosage amounts of GZ17-8.08, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0128] FIG. **28H** is a graph of cell number versus dosage amounts of GZ17-8.09, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0129] FIG. **28I** is a graph of cell number versus dosage amounts of GZ17-8.10, illustrating the effect thereof in inducing the death of ovarian cancer, lung cancer, prostate cancer, pancreatic cancer and fibroblast cells, as described in Example 28;

[0130] FIG. **29A** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of ovarian cancer;

[0131] FIG. **29B** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of lung cancer;

[0132] FIG. **29C** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of prostate cancer;

[0133] FIG. **29D** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of head and neck cancer;

[0134] FIG. **29E** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of breast cancer;

[0135] FIG. **29F** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of leukemia;

[0136] FIG. **29G** is a graph of cell number versus dosage amounts of GZ17-8.11, illustrating the effect thereof in inducing the death of lymphoma;

[0137] FIG. **30A** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of ovarian cancer;

[0138] FIG. **30B** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of lung cancer;

[0139] FIG. **30C** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of prostate cancer;

[0140] FIG. **30D** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of head and neck cancer;

[0141] FIG. **30E** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of breast cancer;

[0142] FIG. **30F** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of leukemia;

[0143] FIG. **30G** is a graph of cell number versus dosage amounts of GZ17-8.12, illustrating the effect thereof in inducing the death of lymphoma;

[0144] FIG. **31A** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of ovarian cancer;

[0145] FIG. **31B** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of lung cancer;

[0146] FIG. **31C** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of prostate cancer;

[0147] FIG. **31D** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of head and neck cancer;

[0148] FIG. **31E** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of breast cancer;

[0149] FIG. **31F** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of leukemia;

[0150] FIG. **31G** is a graph of cell number versus dosage amounts of GZ17-8.13, illustrating the effect thereof in inducing the death of lymphoma;

[0151] FIG. **32A** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of ovarian cancer;

[0152] FIG. **32B** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of lung cancer;

[0153] FIG. **32C** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of prostate cancer;

[0154] FIG. **32D** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of head and neck cancer;

[0155] FIG. **32E** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of breast cancer;

[0156] FIG. **32F** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of leukemia;

[0157] FIG. **32G** is a graph of cell number versus dosage amounts of GZ17-8.14, illustrating the effect thereof in inducing the death of lymphoma;

[0158] FIG. **33A** is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of ovarian cancer;

[0159] FIG. **33B** is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of lung cancer;

[0160] FIG. **33C** is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of prostate cancer;

[0161] FIG. **33D** is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of head and neck cancer;

[0162] FIG. **33E** is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of breast cancer;

[0163] FIG. **33F** is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of leukemia;

[0164] FIG. 33G is a graph of cell number versus dosage amounts of GZ17-8.15, illustrating the effect thereof in inducing the death of lymphoma;

[0165] FIG. 34A is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of ovarian cancer;

[0166] FIG. 34B is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of lung cancer;

[0167] FIG. 34C is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of prostate cancer;

[0168] FIG. 34D is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of head and neck cancer;

[0169] FIG. 34E is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of breast cancer;

[0170] FIG. 34F is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of leukemia;

[0171] FIG. 34G is a graph of cell number versus dosage amounts of GZ17-8.16, illustrating the effect thereof in inducing the death of lymphoma;

[0172] FIG. 35A is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of ovarian cancer;

[0173] FIG. 35B is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of lung cancer;

[0174] FIG. 35C is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of prostate cancer;

[0175] FIG. 35D is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of head and neck cancer;

[0176] FIG. 35E is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of breast cancer;

[0177] FIG. 35F is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of leukemia;

[0178] FIG. 35G is a graph of cell number versus dosage amounts of GZ17-8.17, illustrating the effect thereof in inducing the death of lymphoma;

[0179] FIG. 36A is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of ovarian cancer;

[0180] FIG. 36B is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of lung cancer;

[0181] FIG. 36C is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of prostate cancer;

[0182] FIG. 36D is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of head and neck cancer;

[0183] FIG. 36E is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of breast cancer;

[0184] FIG. 36F is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of leukemia;

[0185] FIG. 36G is a graph of cell number versus dosage amounts of GZ17-8.18, illustrating the effect thereof in inducing the death of lymphoma;

[0186] FIG. 37A is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of ovarian cancer;

[0187] FIG. 37B is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of lung cancer;

[0188] FIG. 37C is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of prostate cancer;

[0189] FIG. 37D is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of head and neck cancer;

[0190] FIG. 37E is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of breast cancer;

[0191] FIG. 37F is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of leukemia;

[0192] FIG. 37G is a graph of cell number versus dosage amounts of GZ17-8.19, illustrating the effect thereof in inducing the death of lymphoma;

[0193] FIG. 38A is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of ovarian cancer;

[0194] FIG. 38B is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of lung cancer;

[0195] FIG. 38C is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of prostate cancer;

[0196] FIG. 38D is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of head and neck cancer;

[0197] FIG. 38E is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of breast cancer;

[0198] FIG. 38F is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of leukemia;

[0199] FIG. 38G is a graph of cell number versus dosage amounts of GZ17-8.20, illustrating the effect thereof in inducing the death of lymphoma;

[0200] FIG. 39A is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of ovarian cancer;

[0201] FIG. 39B is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of lung cancer;

[0202] FIG. 39C is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of prostate cancer;

[0203] FIG. 39D is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of head and neck cancer;

[0204] FIG. 39E is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of breast cancer;

[0205] FIG. 39F is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of leukemia;

[0206] FIG. 39G is a graph of cell number versus dosage amounts of GZ17-8.21, illustrating the effect thereof in inducing the death of lymphoma;

[0207] FIG. 40A is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of ovarian cancer;

[0208] FIG. 40B is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of lung cancer;

[0209] FIG. 40C is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of prostate cancer;

[0210] FIG. 40D is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of head and neck cancer;

[0211] FIG. 40E is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of breast cancer;

[0212] FIG. 40F is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of leukemia;

[0213] FIG. 40G is a graph of cell number versus dosage amounts of GZ17-8.22, illustrating the effect thereof in inducing the death of lymphoma;

[0214] FIG. **41A** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of ovarian cancer;

[0215] FIG. **41B** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of lung cancer;

[0216] FIG. **41C** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of prostate cancer;

[0217] FIG. **41D** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of head and neck cancer;

[0218] FIG. **41E** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of breast cancer;

[0219] FIG. **41F** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of leukemia;

[0220] FIG. **41G** is a graph of cell number versus dosage amounts of GZ17-8.23, illustrating the effect thereof in inducing the death of lymphoma;

[0221] FIG. **42A** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of ovarian cancer;

[0222] FIG. **42B** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of lung cancer;

[0223] FIG. **42C** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of prostate cancer;

[0224] FIG. **42D** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of head and neck cancer;

[0225] FIG. **42E** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of breast cancer;

[0226] FIG. **42F** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of leukemia;

[0227] FIG. **42G** is a graph of cell number versus dosage amounts of GZ17-8.24, illustrating the effect thereof in inducing the death of lymphoma;

[0228] FIG. **43A** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of ovarian cancer;

[0229] FIG. **43B** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of lung cancer;

[0230] FIG. **43C** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of prostate cancer;

[0231] FIG. **43D** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of head and neck cancer;

[0232] FIG. **43E** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of breast cancer;

[0233] FIG. **43F** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of leukemia;

[0234] FIG. **43G** is a graph of cell number versus dosage amounts of GZ17-8.25, illustrating the effect thereof in inducing the death of lymphoma;

[0235] FIG. **44A** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of ovarian cancer;

[0236] FIG. **44B** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of lung cancer;

[0237] FIG. **44C** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of prostate cancer;

[0238] FIG. **44D** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of head and neck cancer;

[0239] FIG. **44E** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of breast cancer;

[0240] FIG. **44F** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of leukemia;

[0241] FIG. **44G** is a graph of cell number versus dosage amounts of GZ17-8.26, illustrating the effect thereof in inducing the death of lymphoma;

[0242] FIG. **45A** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of ovarian cancer;

[0243] FIG. **45B** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of lung cancer;

[0244] FIG. **45C** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of prostate cancer;

[0245] FIG. **45D** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of head and neck cancer;

[0246] FIG. **45E** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of breast cancer;

[0247] FIG. **45F** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of leukemia;

[0248] FIG. **45G** is a graph of cell number versus dosage amounts of GZ17-8.27, illustrating the effect thereof in inducing the death of lymphoma;

[0249] FIG. **46A** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of ovarian cancer;

[0250] FIG. **46B** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of lung cancer;

[0251] FIG. **46C** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of prostate cancer;

[0252] FIG. **46D** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of head and neck cancer;

[0253] FIG. **46E** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of breast cancer;

[0254] FIG. **46F** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of leukemia;

[0255] FIG. **46G** is a graph of cell number versus dosage amounts of GZ17-8.28, illustrating the effect thereof in inducing the death of lymphoma;

[0256] FIG. **47A** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of ovarian cancer;

[0257] FIG. **47B** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of lung cancer;

[0258] FIG. **47C** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of prostate cancer;

[0259] FIG. **47D** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of head and neck cancer;

[0260] FIG. **47E** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of breast cancer;

[0261] FIG. **47F** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of leukemia;

[0262] FIG. **47G** is a graph of cell number versus dosage amounts of GZ17-8.29, illustrating the effect thereof in inducing the death of lymphoma;

[0263] FIG. **48A** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of ovarian cancer;

[0264] FIG. **48B** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of lung cancer;

[0265] FIG. **48C** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of prostate cancer;

[0266] FIG. **48D** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of head and neck cancer;

[0267] FIG. **48E** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of breast cancer;

[0268] FIG. **48F** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of leukemia;

[0269] FIG. **48G** is a graph of cell number versus dosage amounts of GZ17-8.30, illustrating the effect thereof in inducing the death of lymphoma;

[0270] FIG. **49A** is a graph of cell number versus dosage amounts of GZ17-8.31, illustrating the effect thereof in inducing the death of ovarian cancer;

[0271] FIG. **49B** is a graph of cell number versus dosage amounts of GZ17-8.31, illustrating the effect thereof in inducing the death of lung cancer;

[0272] FIG. **49C** is a graph of cell number versus dosage amounts of GZ17-8.31, illustrating the effect thereof in inducing the death of lymphoma;

[0273] FIG. **49D** is a graph of cell number versus dosage amounts of GZ17-8.31, illustrating the effect thereof in inducing the death of leukemia;

[0274] FIG. **50A** is a graph of cell number versus dosage amounts of GZ17-8.32, illustrating the effect thereof in inducing the death of ovarian cancer;

[0275] FIG. **50B** is a graph of cell number versus dosage amounts of GZ17-8.32, illustrating the effect thereof in inducing the death of lung cancer;

[0276] FIG. **50C** is a graph of cell number versus dosage amounts of GZ17-8.32, illustrating the effect thereof in inducing the death of lymphoma;

[0277] FIG. **50D** is a graph of cell number versus dosage amounts of GZ17-8.32, illustrating the effect thereof in inducing the death of leukemia;

[0278] FIG. **51A** is a graph of cell number versus dosage amounts of GZ17-8.33, illustrating the effect thereof in inducing the death of ovarian cancer;

[0279] FIG. **51B** is a graph of cell number versus dosage amounts of GZ17-8.33, illustrating the effect thereof in inducing the death of lung cancer;

[0280] FIG. **51C** is a graph of cell number versus dosage amounts of GZ17-8.33, illustrating the effect thereof in inducing the death of lymphoma;

[0281] FIG. **51D** is a graph of cell number versus dosage amounts of GZ17-8.33, illustrating the effect thereof in inducing the death of leukemia;

[0282] FIG. **52A** is a graph of cell number versus dosage amounts of GZ17-8.34, illustrating the effect thereof in inducing the death of ovarian cancer;

[0283] FIG. **52B** is a graph of cell number versus dosage amounts of GZ17-8.34, illustrating the effect thereof in inducing the death of lung cancer;

[0284] FIG. **52C** is a graph of cell number versus dosage amounts of GZ17-8.34, illustrating the effect thereof in inducing the death of lymphoma;

[0285] FIG. **52D** is a graph of cell number versus dosage amounts of GZ17-8.34, illustrating the effect thereof in inducing the death of leukemia;

[0286] FIG. **53A** is a graph of cell number versus dosage amounts of GZ17-8.35, illustrating the effect thereof in inducing the death of ovarian cancer;

[0287] FIG. **53B** is a graph of cell number versus dosage amounts of GZ17-8.35, illustrating the effect thereof in inducing the death of lung cancer;

[0288] FIG. **53C** is a graph of cell number versus dosage amounts of GZ17-8.35, illustrating the effect thereof in inducing the death of lymphoma;

[illegible]

[illegible]

[illegible]

[illegible]

[0389] FIG. **78D** is a graph of cell number versus dosage amounts of GZ17-8.60, illustrating the effect thereof in inducing the death of leukemia;

[0390] FIG. **79A** is a graph of cell number versus dosage amounts of GZ17-8.61, illustrating the effect thereof in inducing the death of ovarian cancer;

[0391] FIG. **79B** is a graph of cell number versus dosage amounts of GZ17-8.61, illustrating the effect thereof in inducing the death of lung cancer;

[0392] FIG. **79C** is a graph of cell number versus dosage amounts of GZ17-8.61, illustrating the effect thereof in inducing the death of lymphoma;

[0393] FIG. **79D** is a graph of cell number versus dosage amounts of GZ17-8.61, illustrating the effect thereof in inducing the death of leukemia;

[0394] FIG. **80A** is a graph of cell number versus dosage amounts of GZ17-10.04, illustrating the effect thereof in inducing the death of ovarian cancer;

[0395] FIG. **80B** is a graph of cell number versus dosage amounts of GZ17-10.05, illustrating the effect thereof in inducing the death of ovarian cancer;

[0396] FIG. **80C** is a graph of cell number versus dosage amounts of GZ17-10.06, illustrating the effect thereof in inducing the death of ovarian cancer;

[0397] FIG. **80D** is a graph of cell number versus dosage amounts of GZ17-10.04, illustrating the effect thereof in inducing the death of lymphoma;

[0398] FIG. **80E** is a graph of cell number versus dosage amounts of GZ17-10.05, illustrating the effect thereof in inducing the death of lymphoma;

[0399] FIG. **80F** is a graph of cell number versus dosage amounts of GZ17-10.06, illustrating the effect thereof in inducing the death of lymphoma;

[0400] FIG. **80G** is a comparative bar graph illustrating the comparative lymphoma cell-killing effect of the individual components, GZ17-10.04-10.06, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0401] FIG. **80H** is a comparative bar graph illustrating the comparative lymphoma cell-killing effect of the individual components, GZ17-10.04 and 10.06, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0402] FIG. **80I** is a graph of cell number versus dosage amounts of GZ17-10.04, illustrating the effect thereof in inducing the death of leukemia;

[0403] FIG. **80J** is a graph of cell number versus dosage amounts of GZ17-10.05, illustrating the effect thereof in inducing the death of leukemia;

[0404] FIG. **80K** is a graph of cell number versus dosage amounts of GZ17-10.06, illustrating the effect thereof in inducing the death of leukemia;

[0405] FIG. **80L** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components, GZ17-10.04-10.06, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0406] FIG. **80M** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components, GZ17-10.04 and 10.06, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0407] FIG. **80N** is a graph of cell number versus dosage amounts of GZ17-10.04, illustrating the effect thereof in inducing the death of breast cancer;

[0408] FIG. **80O** is a graph of cell number versus dosage amounts of GZ17-10.05, illustrating the effect thereof in inducing the death of breast cancer;

[0409] FIG. **80P** is a graph of cell number versus dosage amounts of GZ17-10.06, illustrating the effect thereof in inducing the death of breast cancer;

[0410] FIG. **80Q** is a comparative bar graph illustrating the comparative breast cancer cell-killing effect of the individual components, GZ17-10.04-10.06, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0411] FIG. **80R** is a comparative bar graph illustrating the comparative breast cancer cell-killing effect of the individual components, GZ17-10.04 and 10.06, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0412] FIG. **81A** is a graph of cell number versus dosage amounts of GZ17-08.512, illustrating the effect thereof in inducing the death of ovarian cancer;

[0413] FIG. **81B** is a graph of cell number versus dosage amounts of GZ17-08.512, illustrating the effect thereof in inducing the death of lymphoma;

[0414] FIG. **81C** is a graph of cell number versus dosage amounts of GZ17-08.512, illustrating the effect thereof in inducing the death of head and neck cancer;

[0415] FIG. **81D** is a graph of cell number versus dosage amounts of GZ17-08.513, illustrating the effect thereof in inducing the death of ovarian cancer;

[0416] FIG. **81E** is a graph of cell number versus dosage amounts of GZ17-08.513, illustrating the effect thereof in inducing the death of lymphoma;

[0417] FIG. **81F** is a graph of cell number versus dosage amounts of GZ17-08.513, illustrating the effect thereof in inducing the death of head and neck cancer;

[0418] FIG. **81G** is a graph of cell number versus dosage amounts of GZ17-08.514, illustrating the effect thereof in inducing the death of ovarian cancer;

[0419] FIG. **81H** is a graph of cell number versus dosage amounts of GZ17-08.514, illustrating the effect thereof in inducing the death of lymphoma;

[0420] FIG. **81I** is a graph of cell number versus dosage amounts of GZ17-08.514, illustrating the effect thereof in inducing the death of head and neck cancer;

[0421] FIG. **81J** is a graph of cell number versus dosage amounts of GZ17-08.515, illustrating the effect thereof in inducing the death of ovarian cancer;

[0422] FIG. **81K** is a graph of cell number versus dosage amounts of GZ17-08.515, illustrating the effect thereof in inducing the death of lymphoma;

[0423] FIG. **81L** is a graph of cell number versus dosage amounts of GZ17-08.515, illustrating the effect thereof in inducing the death of head and neck cancer;

[0424] FIG. **81M** is a graph of cell number versus dosage amounts of GZ17-08.516, illustrating the effect thereof in inducing the death of ovarian cancer;

[0425] FIG. **81N** is a graph of cell number versus dosage amounts of GZ17-08.516, illustrating the effect thereof in inducing the death of lymphoma;

[0426] FIG. **81O** is a graph of cell number versus dosage amounts of GZ17-08.516, illustrating the effect thereof in inducing the death of head and neck cancer;

[0427] FIG. **81P** is a graph of cell number versus dosage amounts of GZ17-08.517, illustrating the effect thereof in inducing the death of ovarian cancer;

[0428] FIG. **81Q** is a graph of cell number versus dosage amounts of GZ17-08.517, illustrating the effect thereof in inducing the death of lymphoma;

[0429] FIG. **81R** is a graph of cell number versus dosage amounts of GZ17-08.517, illustrating the effect thereof in inducing the death of head and neck cancer;

[0430] FIG. **81S** is a graph of cell number versus dosage amounts of GZ17-08.518, illustrating the effect thereof in inducing the death of ovarian cancer;

[0431] FIG. **81T** is a graph of cell number versus dosage amounts of GZ17-08.518, illustrating the effect thereof in inducing the death of lymphoma;

[0432] FIG. **81U** is a graph of cell number versus dosage amounts of GZ17-08.518, illustrating the effect thereof in inducing the death of head and neck cancer;

[0433] FIG. **81V** is a graph of cell number versus dosage amounts of GZ17-08.519, illustrating the effect thereof in inducing the death of ovarian cancer;

[0434] FIG. **81W** is a graph of cell number versus dosage amounts of GZ17-08.519, illustrating the effect thereof in inducing the death of lymphoma;

[0435] FIG. **81X** is a graph of cell number versus dosage amounts of GZ17-08.519, illustrating the effect thereof in inducing the death of head and neck cancer;

[0436] FIG. **81Y** is a graph of cell number versus dosage amounts of GZ17-08.520, illustrating the effect thereof in inducing the death of ovarian cancer;

[0437] FIG. **81Z** is a graph of cell number versus dosage amounts of GZ17-08.520, illustrating the effect thereof in inducing the death of lymphoma;

[0438] FIG. **81AA** is a graph of cell number versus dosage amounts of GZ17-08.520, illustrating the effect thereof in inducing the death of head and neck cancer;

[0439] FIG. **81BB** is a graph of cell number versus dosage amounts of GZ17-08.521, illustrating the effect thereof in inducing the death of ovarian cancer;

[0440] FIG. **81CC** is a graph of cell number versus dosage amounts of GZ17-08.521, illustrating the effect thereof in inducing the death of lymphoma;

[0441] FIG. **81DD** is a graph of cell number versus dosage amounts of GZ17-08.521, illustrating the effect thereof in inducing the death of head and neck cancer;

[0442] FIG. **82-1** is a graph of cell number versus dosage amounts of GZ08.065, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0443] FIG. **82-2** is a graph of cell number versus dosage amounts of GZ08.066, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0444] FIG. **82-3** is a graph of cell number versus dosage amounts of GZ08.067, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0445] FIG. **82-4** is a graph of cell number versus dosage amounts of GZ08.068, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0446] FIG. **82-5** is a graph of cell number versus dosage amounts of GZ08.069, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0447] FIG. **82-6** is a graph of cell number versus dosage amounts of GZ08.070, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0448] FIG. **82-7** is a graph of cell number versus dosage amounts of GZ08.071, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0449] FIG. **82-8** is a graph of cell number versus dosage amounts of GZ08.072, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0450] FIG. **82-9** is a graph of cell number versus dosage amounts of GZ08.073, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0451] FIG. **82-10** is a graph of cell number versus dosage amounts of GZ08.074, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0452] FIG. **82-11** is a graph of cell number versus dosage amounts of GZ08.075, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0453] FIG. **82-12** is a graph of cell number versus dosage amounts of GZ08.076, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0454] FIG. **82-13** is a graph of cell number versus dosage amounts of GZ08.077, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0455] FIG. **82-14** is a graph of cell number versus dosage amounts of GZ08.078, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0456] FIG. **82-15** is a graph of cell number versus dosage amounts of GZ08.079, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[0457] FIG. **82-16** is a graph of cell number versus dosage amounts of GZ08.080, illustrating the effect thereof in inducing the death of lung cancer, as described in Example 82;

[illegible]

[illegible]

[illegible]

[0558] FIG. **82-117** is a graph of cell number versus dosage amounts of GZ08.099, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0559] FIG. **82-118** is a graph of cell number versus dosage amounts of GZ08.100, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0560] FIG. **82-119** is a graph of cell number versus dosage amounts of GZ08.101, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0561] FIG. **82-120** is a graph of cell number versus dosage amounts of GZ08.102, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0562] FIG. **82-121** is a graph of cell number versus dosage amounts of GZ08.103, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0563] FIG. **82-122** is a graph of cell number versus dosage amounts of GZ08.104, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0564] FIG. **82-123** is a graph of cell number versus dosage amounts of GZ08.105, illustrating the effect thereof in inducing the death of leukemia, as described in Example 82;

[0565] FIG. **82-124** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.065, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0566] FIG. **82-125** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.067, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0567] FIG. **82-126** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.068, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0568] FIG. **82-127** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.079, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0569] FIG. **82-128** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.080, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition;

[0570] FIG. **82-129** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.084, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0571] FIG. **82-130** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.085, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0572] FIG. **82-131** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.086, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0573] FIG. **82-132** is a comparative bar graph illustrating the comparative lung cancer cell-killing effect of the individual components of GZ08.087, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[illegible]

[0599] FIG. **82-158** is a comparative bar graph illustrating the comparative lymphoma cell-killing effect of the individual components of GZ08.099, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0600] FIG. **82-159** is a comparative bar graph illustrating the comparative lymphoma cell-killing effect of the individual components of GZ08.101, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0601] FIG. **82-160** is a comparative bar graph illustrating the comparative lymphoma cell-killing effect of the individual components of GZ08.105, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0602] FIG. **82-161** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.086, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0603] FIG. **82-162** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.087, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0604] FIG. **82-163** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.090, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0605] FIG. **82-164** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.094, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0606] FIG. **82-165** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.098, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0607] FIG. **82-166** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.104, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0608] FIG. **82-167** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components of GZ08.105, versus the theoretical additive effect of these components, and the actual effect thereof at the indicted concentration, demonstrating the synergism of the three-component composition, as described in Example 82;

[0609] FIG. **83A** is a graph of cell number versus dosage amounts of orthovanillin, illustrating the effect thereof in inducing the death of lymphoma, as described in Example 83;

[0610] FIG. **83B** is a graph of cell number versus dosage amounts of curcumin, illustrating the effect thereof in inducing the death of lymphoma, as described in Example 83;

[0611] FIG. **83C** is a graph of cell number versus dosage amounts of harmaline, illustrating the effect thereof in inducing the death of lymphoma, as described in Example 83;

[0612] FIG. **83D** is a comparative bar graph illustrating the comparative lymphoma cell-killing effect of the individual components, orthovanillin, curcumin, and harmaline, as shown in FIGS. **83A-83C**, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0613] FIG. **83E** is a graph of cell number versus dosage amounts of orthovanillin, illustrating the effect thereof in inducing the death of leukemia, as described in Example 83;

[0614] FIG. **83F** is a graph of cell number versus dosage amounts of curcumin, illustrating the effect thereof in inducing the death of leukemia, as described in Example 83;

[0615] FIG. **83G** is a graph of cell number versus dosage amounts of harmaline, illustrating the effect thereof in inducing the death of leukemia, as described in Example 83;

[0616] FIG. **83H** is a comparative bar graph illustrating the comparative leukemia cell-killing effect of the individual components, orthovanillin, curcumin, and harmaline, as shown in FIGS. **83E-83G**, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0617] FIG. **83I** is a graph of cell number versus dosage amounts of orthovanillin, illustrating the effect thereof in inducing the death of breast cancer, as described in Example 83;

[0618] FIG. **83J** is a graph of cell number versus dosage amounts of curcumin, illustrating the effect thereof in inducing the death of breast cancer, as described in Example 83;

[0619] FIG. **83K** is a graph of cell number versus dosage amounts of harmaline, illustrating the effect thereof in inducing the death of breast cancer, as described in Example 83;

[0620] FIG. **83L** is a comparative bar graph illustrating the comparative breast cancer cell-killing effect of the individual components, orthovanillin, curcumin, and harmaline, as shown in FIGS. **83I-83K**, versus the theoretical additive effect of these components, and the actual effect thereof, demonstrating the synergism of the three-component composition;

[0621] FIG. **84A** is series of photographs of pancreatic cancer spheres including a control, treatment of the spheres with GZ17-6.02 at levels of 0.5IC.sub.50 and IC.sub.50, illustrating the effect of GZ17-6.02 in reducing the number and size of pancreatic cancer spheres, as set forth in Example 84;

[0622] FIG. **84B** is a series of blots illustrating in vitro pancreatic cell treatment with GZ17-6.02 at levels of 0.5IC.sub.50 and IC.sub.50, and depicting decreases in cancer stem cell markers owing to the treatment with GZ17-6.02, as set forth in Example 84;

[0623] FIG. **84C** is a series of blots illustrating the treatment of mouse pancreatic cancer cells with GZ17-6.02, and depicting the decrease in cancer stem cell markers owing to the treatment with GZ17-6.02, as set forth in Example 84; and

[0624] FIG. **84D** is a possible pathway diagram illustrating the action of GZ17-6.02 on both cancer stem cells (CSC) and other cancer cells within a tumor, as set forth in Example 84.

DETAILED DESCRIPTION

[0625] The therapeutic agents of the invention are used in therapeutically effective amounts, i.e., amounts that will elicit the biological or medical response of a tissue, system, or subject that is being sought, and in particular to elicit some desired therapeutic effect against a variety of human diseases, and especially cancers; in the case of cancers, the agents operate by preventing and/or inhibiting proliferation and/or survival of cancerous cells, and/or by slowing the progression of cancers. Those skilled in the art recognize that an amount may be considered therapeutically effective even if the condition is not totally eradicated or prevented, but it or its symptoms and/or effects are improved or alleviated partially in the subject. Of course, the appropriate makeup of the agents hereof and dosing regimens using such agents will depend on the particular cancer being treated, the extent of the disease, and other factors related to the patient as determined by those skilled in the art. Hence, the terms “therapeutic” or “treat,” as used herein, refer to products or processes in accordance with the invention that are intended to produce a beneficial change in an existing condition (e.g., cancerous tissue, tumor size, metastases, etc.) of a subject, such as by reducing the severity of the clinical symptoms and/or effects of the condition, and/or reducing the duration of the symptoms/effects of a subject.

[0626] Additional ingredients may be included with the chemotherapeutic agents of the invention for administration to the subject. Such additional ingredients include, other active agents,

preservatives, buffering agents, salts, carriers, excipients, diluents, or other pharmaceutically-acceptable ingredients. The active agents that could be included in the compositions include antiviral, antibiotic, or other anticancer compounds.

[0627] The combined therapeutic agents of the invention preferably give synergistic results, which are entirely unexpected. Moreover, the lack of side effects when the agents are administered to patients is quite surprising and essentially unique. As used herein, the terms “combination” or “in combination” are intended to embrace compositions wherein the components are physically intermixed as dosage forms, and to situations where the individual components are separately administered to a subject over relatively short periods of time, which would have the same therapeutic effects as a single dosage form.

[0628] In use, a therapeutically effective amount of an agent in accordance with the invention is administered to a subject in need thereof. Such may comprise a single unit dosage or, more usually, periodic (e.g., daily) administration of lower dosages over time. Advantageously, administration of such therapeutically effective amounts achieves an unexpected therapeutic synergy. This means that the therapeutic two- or three-component compositions of the invention exhibit a joint action where one or more of the components supplements or enhances the action of at least one of the other components to produce an effect greater than that which may be obtained by use of individual components in equivalent quantities, or produce effects that could not be obtained with safe quantities of the other components, individually or in combination. Generally, one or more of the components working together produce an effect greater than the sum of their individual effects.

[0629] The dosages may be administered in any convenient manner, such as by oral, rectal, nasal, ophthalmic, parenteral (including intraperitoneal, gastrointestinal, intrathecal, intravenous, cutaneous (e.g., dermal patch), subcutaneous (e.g. injection or implant), or intramuscular) administrations. The dosage forms of the invention may be in the form of liquids, gels, suspensions, solutions, or solids (e.g., tablets, pills, or capsules). Moreover, therapeutically effective amounts of the agents of the invention may be co-administered with other chemotherapeutic agent(s), where the two products are administered substantially simultaneously or in any sequential manner.

[0630] Additional advantages of the various embodiments of the invention will be apparent to those skilled in the art upon review of the disclosure herein and the working examples below. It will be appreciated that the various embodiments described herein are not necessarily mutually exclusive unless otherwise indicated herein. For example, a feature described or depicted in one embodiment may also be included in other embodiments, but is not necessarily included. Thus, the present invention encompasses a variety of combinations and/or integrations of the specific embodiments described herein.

[0631] As used herein, the phrase “and/or,” when used in a list of two or more items, means that any one of the listed items can be employed by itself or any combination of two or more of the listed items can be employed. For example, if a composition is described as containing or excluding components A, B, and/or C, the composition can contain or exclude A alone; B alone; C alone; A and B in combination; A and C in combination; B and C in combination; or A, B, and C in combination.

[0632] The present description also uses numerical ranges to quantify certain parameters relating to various embodiments of the invention. It should be understood that when numerical ranges are provided, such ranges are to be construed as providing literal support for claim limitations that only recite the lower value of the range as well as claim limitations that only recite the upper value of the range. For example, a disclosed numerical range of about 10 to about 100 provides literal support for a claim reciting “greater than about 10” (with no upper bounds) and a claim reciting “less than about 100” (with no lower bounds).

[0633] In the ensuing discussion, the curcumin, harmine, and isovanillin component(s) will be individually described. In such discussions, where terms are used which specify or imply the

presence of carbon-carbon chains (e.g., alkyl, alkenyl, alkoxy, alkyl amine, alkenyl amine, aldehyde, carboxylate, or the like), these disclosures should be understood to refer to primary (straight), branched chain, or cyclic carbon chain groups. Moreover, unless indicated otherwise, reference to aryl groups means phenyl, substituted phenyl, naphthyl, substituted naphthyl; and heteroatom aryl groups refers to aryl groups containing a nitrogen, oxygen, boron, or sulfur atom, such as pyridine; heterocyclic groups refers to cyclic groups containing from 3-7 atoms, one or more of which is a nitrogen, oxygen, boron, or sulfur heteroatom; and amine refers to primary, secondary, tertiary, or quaternary amines.

[0634] As used herein, pharmaceutically acceptable salts with reference to the components means salts of the component compounds of the present invention which are pharmaceutically acceptable, i.e., salts which are useful in preparing pharmaceutical compositions that are generally safe, non-toxic, and neither biologically nor otherwise undesirable and are acceptable for human pharmaceutical use, and which possess the desired degree of pharmacological activity. Such pharmaceutically acceptable salts include acid addition salts formed with inorganic acids such as hydrochloric acid, hydrobromic acid, sulfuric acid, nitric acid, phosphoric acid, and the like; or with organic acids such as 1,2-ethanedisulfonic acid, 2-hydroxyethanesulfonic acid, 2-naphthalenesulfonic acid, 3-phenylpropionic acid, 4,4'-methylenebis(3-hydroxy-2-ene-1-carboxylic acid), 4-methylbicyclo[2.2.2]oct-2-ene-1-carboxylic acid, acetic acid, aliphatic mono- and dicarboxylic acids, aliphatic sulfuric acids, aromatic sulfuric acids, benzenesulfonic acid, benzoic acid, camphorsulfonic acid, carbonic acid, cinnamic acid, citric acid, cyclopentanepropionic acid, ethanesulfonic acid, fumaric acid, glucoheptonic acid, gluconic acid, glutamic acid, glycolic acid, heptanoic acid, hexanoic acid, hydroxynaphthoic acid, lactic acid, laurylsulfuric acid, maleic acid, malic acid, malonic acid, mandelic acid, methanesulfonic acid, muconic acid, o-(4-hydroxybenzoyl)benzoic acid, oxalic acid, p-chlorobenzenesulfonic acid, phenyl-substituted alkanolic acids, propionic acid, p-toluenesulfonic acid, pyruvic acid, salicylic acid, stearic acid, succinic acid, tartaric acid, tertiarybutylacetic acid, trimethylacetic acid, and the like.

Pharmaceutically acceptable salts also include base addition salts which may be formed when acidic protons present are capable of reacting with inorganic or organic bases. Acceptable inorganic bases include sodium hydroxide, sodium carbonate, potassium hydroxide, aluminum hydroxide and calcium hydroxide. Acceptable organic bases include ethanolamine, diethanolamine, triethanolamine, tromethamine, N-methylglucamine and the like. It should be recognized that the particular anion or cation forming a part of any salt of this invention is not critical, so long as the salt, as a whole, is pharmacologically acceptable. Additional examples of pharmaceutically acceptable salts and their methods of preparation and use are presented in *Handbook of Pharmaceutical Salts Properties, and Use*, P. H. Stahl & C. G. Wermuth eds., ISBN 978-3-90639-058-1 (2008).

[0635] As noted below, the curcumin, harmine, and isovanillin component(s) may be obtained as synthetic compounds of high purity, or from modified naturally occurring sources. In either case, however, it is preferred that the component(s) be purified to a level of at least about 50% by weight, more preferably at least about 70% by weight, still more preferably at least about 90% by weight, and most preferably at least about 98% by weight.

[0636] The individual discussions of the component(s) contain structural formulas. In order to be entirely clear, curcumin-based formulas are indicated "C-number," (not to be confused with carbon chain numbers, which are indicated as "Cnumber," without an intervening hyphen), harmine-based formulas are indicated as "H-number," and isovanillin-based formulas are indicated as "I-number."

[0637] Before discussing the individual components of the invention, it should be understood that use of unmodified, naturally occurring sources of the components is generally not appropriate or desirable, because these naturally occurring products contain relatively small amounts of the desired components and/or have potentially interfering compounds therein. For example, naturally occurring turmeric has only approximately 2-3% by weight curcumin therein and accordingly the

straightforward use of unmodified turmeric would not be suitable for the invention. In like manner, naturally occurring harmala seed contains only a very minor amount of harmine and such a product would also be inappropriate.

[0638] Thus, the preferred components of the invention are either synthetically derived or derived from one or more naturally occurring product(s) which have been significantly modified so as to contain at least about 25% by weight (more preferably at least about 50% by weight, and still more preferably about 70% by weight) of the desired component. As used herein, “synthetically derived” means that the component in question was synthesized using specific starting ingredients and one or more chemical and/or biological reactions to obtain substantially pure compounds. Modification of naturally occurring products may involve extractions, or any other physical or chemical steps to achieve the desired end product, e.g., harmine components may be obtained from treatment of harmala seed, or curcumin components may be obtained from treatment of turmeric.

[0639] For example, curcumin can be synthetically derived to a high degree of purity. Alternately, curcumin can be obtained by extraction or other treatment of naturally occurring turmeric so that the curcumin content of the modified turmeric has the above-noted levels of curcumin therein.

The Curcumin Component(s)

[0640] As used herein, “curcumin component(s)” shall mean curcumin, its metabolites and derivatives, isomers and tautomers thereof, esters, metal complexes (e.g., Cu, Fe, Zn, Pt, V), and pharmaceutically acceptable salts of any of the foregoing. Curcumin derivatives include both naturally occurring and synthetic derivatives, e.g., the spontaneous degradation products of curcumin, curcumin metabolites, and synthetic curcumin derivative compounds.

1. Curcumin.

[0641] Curcumin (diferuloylmethane, 1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) is a symmetrical diphenolic dienone, see structure C-I below. It exists in solution as an equilibrium mixture of the symmetrical dienone (diketo) and the keto-enol tautomer; the keto-enol form is strongly favored by intramolecular hydrogen bonding.

##STR00001##

Curcumin contains two aryl rings separated by an unsaturated 7-carbon linker having a symmetrical β -diketone group (as used herein, “ β -diketone” embraces both tautomeric forms, namely the diketo and enol forms). The aryl rings of curcumin contain a hydroxyl group in the para position and a methoxy group in the meta position.

2. Degradation Products of Curcumin.

[0642] It is known that, under certain pH and other conditions, curcumin will spontaneously form degradation products, and especially one or more of the following:

##STR00002## ##STR00003##

3. Curcumin Metabolites

[0643] It has been determined that curcumin is differently metabolized in vivo depending upon the route of administration, see, Shen et al. *The Pharmacology of Curcumin: Is it the Degradation Products?* Trends in Molecular Medicine, March 2012 Vol. 18, No. 2, incorporated by reference herein in its entirety. Thus, when curcumin is orally administered, the metabolites normally include one or more of the following:

##STR00004##

[0644] On the other hand, where the route of administration is intravenous/intraperitoneal, the metabolites generally include the following:

##STR00005##

[0645] Other naturally occurring curcumin derivatives include cyclocurcumin, bisdemethoxycurcumin, demethoxycurcumin, dihydrocurcumin, caffeic acid, cinnamic acid, isoeugenol, dibenzoylmethane, dehydrozingerone, capsaicin, [6]-gingerol, [6]-paradol, chlorogenic acid, yakuchinone A, oregonin, cassumuin A, and cassumuin B.

4. Synthetic Curcumin Derivatives.

[0646] Curcumin derivatives are expected to be beneficial for use in the treatment methods of the invention. The term “curcumin derivative” is used interchangeably with the term “curcumin analog” and “curcumin analogue” (alternative spelling) and includes, for example, curcumin derivatives, analogs, curcuminoids and chalcones. In one embodiment the curcumin derivative includes first and second aryl groups covalently attached by way of a linker or a linking group. In another embodiment, the second aryl group is absent, such that the curcumin derivative contains a first aryl group and the linker but no second aryl group at the distal end of the linker. Optionally, the first and/or second aryl group is a heteroaryl group. The first and second aryl groups may be independently substituted or unsubstituted.

[0647] Curcumin derivatives that exhibit improved pharmacokinetic properties and/or reduced toxicity are preferred. For example, curcumin derivatives that include heteroaryl groups and/or unsaturated linkers are expected to impart improved pharmacokinetic properties and/or reduced toxicity to the compounds, because they are expected to be less chemically reactive in vivo. One example of preferred curcumin derivatives includes those including one or two carbonyl groups in the linker region, including those derivatives that preserve the enone functionality of curcumin. Derivatives that include heteroaryl groups and/or unsaturated linkers are expected to be less likely to be degraded and/or form toxic adducts or intermediates under physiological conditions.

[0648] Thus, in one aspect, curcumin derivatives of the invention are generally encompassed by the formula:

Ar1-L-Ar2 C-12

where Ar1 and Ar2 are independently aryl groups, and L is a divalent linking group that includes between 3 and 7 backbone carbon atoms, wherein one or more of the backbone carbon atoms include a carbonyl or hydroxyl moiety.

a. Aryl Groups

[0649] Preferred aryl groups include phenyl, naphthyl, thienyl, pyridinium, and pyridyl groups.

[0650] Aryl groups Ar1 and Ar2 may be substituted or unsubstituted, and one or more of the ring carbons may be substituted with a heteroatom, and especially N, S, B, or O.

[0651] For example, in one embodiment of the invention, Ar1 can be an aryl group according to the formula:

##STR00006##

and Ar2 can be an aryl group according to the formula:

##STR00007##

where one or more of the aryl ring carbons of Ar1 and Ar2 may be independently substituted with a heteroatom selected from N, S, B, or O, and R1-R10 are independently selected from the group consisting of H, hydroxyl, halogen, amine, nitro, sulfonate, sulfoxide, thio, ester, carboxylate, amide, borate, C1-C4 boronate, C1-C8 alkyl, C2-C8 alkenyl, C1-C6 haloalkyl, C1-C6 alkoxy, C1-C6 amine, C2-C8 carboxyl, C2-C8 ester, C1-C4 aldehyde, and glucuronide groups; L is a divalent linker including from 3-7 backbone carbon atoms that form a chain connecting the Ar1, Ar2, and R11 groups as the case may be, where L includes at least one of a carbonyl or hydroxyl group. In further embodiments, Ar1 and Ar2 are phenyl groups; R1-R10 are independently selected from the group consisting of H, hydroxyl, halogen, amine, nitro, sulfonate, thio, borate, C1-C2 boronate, sulfoxide, C1-C4 alkyl, C1-C4 alkoxy, C1-C4 alkylamine, C2-C6 alkenylamine, C1-C6 acetoxy, C1-C4 carboxyl, with at least one of R1-R5 and R6-R10 being hydroxyl.

b. Divalent Linking Groups

[0652] The linker L is a spacer that preferably includes 3, 4, 5, 6 or 7 carbon atoms that form a linear carbon chain connecting the first and second aryl groups. The carbon atoms in the carbon chain that trace out shortest path between the first and optional second aryl groups are referred to herein as the “backbone” carbon atoms. The number of backbone carbon atoms is readily determined in straight chain alkyl groups. In linkers that include a cyclic alkyl group as a

constituent of the linear chain, the backbone carbon atoms include the least number of ring carbons possible. The number of backbone carbon atoms is used herein as a shorthand way to designate the length of the linker being used. For example, a 7-carbon linker is a divalent linker that includes 7 backbone carbon atoms.

[0653] Preferably at least one of the backbone carbon atoms is included in a carbonyl (C=O) or thio carbonyl (C=S) moiety. The linker may be substituted or unsubstituted. The linker may further be saturated or unsaturated. In a preferred embodiment, the linker contains an odd number of carbon atoms (i.e., 3, 5, or 7 carbon atoms), and at least one unsaturated carbon-carbon bond. In additional embodiments, the linker may include a hydroxyl moiety in place of, or in addition to, the at least one carbonyl moiety.

[0654] Curcumin derivatives of the invention include a linking group L that is preferably covalently attached at one end to aryl group Ar1. Optionally, the linking group L may also be covalently attached at the other end to a second aryl group, Ar2, which is selected independently from Ar1. The linking group L is a divalent linking group that preferably includes an alkylene or an alkenylene group having between 3 and 7 backbone carbon atoms, and more advantageously an odd number of backbone carbon atoms (i.e., 3, 5, or 7 carbon atoms). The linker also preferably has at least one carbonyl moiety, and may further include a hydroxyl moiety in place of, or in addition to, the at least one carbonyl moiety. The linking group may be substituted or unsubstituted, and may be saturated or unsaturated. Preferably, the linking group has a carbon-carbon double bond between the a and p carbons relative to Ar1 and/or Ar2 (e.g., see formulas C-1 and C-19 through C-33, which illustrate such a double bond. Still more preferably, the linking group includes conjugated double bonds. Table 1 shows compounds with 7-carbon linkers; Table 2 shows compounds with 5-carbon linkers; and Table 3 shows compounds with 3-carbon linkers.

[0655] A divalent linking group includes two carbons with unfilled valencies that provide valence points where a covalent bond can be formed to an adjacent alkyl or aryl group that also includes a carbon with an unfilled valency. Generally, a valence point is represented in a chemical formula by a bond that is shown as not being attached to another group (e.g., CH₃—, wherein — represents the valence point).

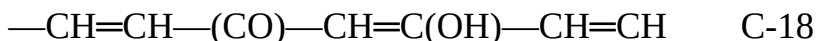
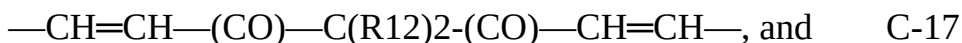
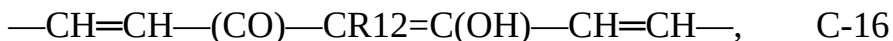
[0656] In embodiments wherein the curcumin derivative lacks the second aryl group Ar2, the distal valence point on the linking group can be filled with any substituent of interest, preferably a short chain alkyl group (e.g., C1-C6, more preferably C1-C4) or a hydrogen (H). Compounds lacking a second aryl group may be represented by formula:

Ar1-L-R11 C-15

R11 can be, for example, a heterocyclic group or an alkyl group, preferably an alkyl group having four or fewer carbon atoms, e.g., a methyl group. R11 can alternately be an amine, a hydroxyl, a hydrogen, nitro, sulfonate, sulfoxide, thio, ester, carboxylate, amide, borate, or a C1-C4 boronate.

i. Curcumin Derivatives Including 7-Carbon Linking Groups

[0657] In one embodiment of the invention, the curcumin derivatives include one or two aryl groups (Ar1 and optionally Ar2) and a linking group L that is a 7-carbon linking group (i.e., a linking group that includes 7 backbone carbon atoms). Preferably, the 7-carbon linking group includes at least one unsaturated carbon-carbon bond. Examples of 7-carbon linking groups include



where R₁₂ includes substituent alkyl, arylalkyl, or aryl groups comprising 10 carbon atoms or less. In some embodiments, R₁₂ may be a methyl, ethyl, or benzyl group. These linking groups are the

divalent forms of 4-alkyl-1,6 heptadiene-3,5-dione; 4,4-dialkyl-1,6 heptadiene-3,5-dione; and heptane-3,5-dione.

[0658] Table 1 shows a number of examples of curcumin derivatives that include a 7-carbon linker. The compounds shown contain two aryl rings separated by a 7-carbon linker having two carbonyls (or the equivalent keto-enol tautomer). In many, but not all, of the compounds, the linker is unsaturated. “Bn” refers to a benzyl group.

TABLE-US-00001 TABLE 1 7-Carbon Linker Analogs. [00008]

[00009] [00010] [00011] [00012] [00013] [00014] [00015] [00016] [00017] [00018] [00019] [00020] [00021] [00022] [00023] [00024] [00025] [00026] [00027] [00028] [00029] [00030]

ii. Curcumin Derivatives Including 5-Carbon Linking Groups

[0659] In a further embodiment of the invention, the curcumin derivatives include one or two aryl groups (Ar1 and optionally Ar2) that are linked by a linking group L that is a 5-carbon linking group (i.e., a linking group that includes 5 backbone carbon atoms). Preferably, the 5-carbon linking group includes at least one unsaturated carbon-carbon bond. Examples of 5-carbon linking groups include:

##STR00031##

[0660] These linking groups are the divalent forms of 1,4-pentadiene-3-one; pentan-3-one; pentan-3-ol, 2,6; bis(methylene)cyclohexanone; and 1,2,4,5-diepoxy pentan-3-one. As noted herein, curcumin derivatives may include a cyclic linking group. For example, 1-methyl-2,6-diphenyl-4-piperidone provides a compound with a 5-carbon linking group that is bridged by a tertiary amine to form a cyclic alkylene linking group including the heteroatom nitrogen.

[0661] Table 2 shows a number of examples of curcumin derivatives that include a 5-carbon linker. The compounds shown contain two aryl rings separated by a 5-carbon linker having a single carbonyl or hydroxyl. In many, but not all, of the compounds, the linker is unsaturated.

TABLE-US-00002 TABLE 2 5-Carbon Linker Analogs. [00032]














[00033] [00034] [00035] [00036] [00037] [00038] [00039] [00040] [00041] [00042] [00043] [00044] [00045] [00046] [00047] [00048] [00049] [00050] [00051] [00052] [00053] [00054] [00055] [00056] [00057] [00058] [00059] [00060] [00061] [00062] [00063] [00064] [00065] [00066] [00067] [00068] [00069] [00070] [00071] [00072] [00073] [00074] [00075]

iii. Curcumin Derivatives Including 3-Carbon Linking Groups

[0662] In a further embodiment of the invention, the curcumin derivatives include one or two aryl groups (Ar1 and optionally Ar2) that are linked by a linking group L that is a 3-carbon linking group (i.e., a linking group that includes 3 backbone carbon atoms). Preferably, the 3-carbon

linking group includes at least one unsaturated carbon-carbon bond. An example of a 3-carbon linking group is —CH=CH—(CO)— ; i.e., a divalent form of propenone.

[0663] Table 3 shows a number of examples of curcumin derivatives that include a 3-carbon linker. The compounds shown generally have an unsaturated 3-carbon linker having a single carbonyl. While most of the examples shown have two aryl groups separated by the linker, several of the embodiments include only a single aryl group. In the examples that include only a single aryl group, a methyl group is provided at the other end of the linking group. One compound includes the heteroatom N in place of one of the backbone carbon atoms; however, this is still considered a 3-C linker in that 3 atoms (C, N, and C) are present along the shortest bridge between the two aryl groups.

TABLE-US-00003 TABLE 3 3-Carbon Linker Analogs. [00076]  [00077]  [00078]  [00079]  [00080]  [00081]  [00082]  [00083]  [00084]  [00085]  [00086]  [00087]  [00088] 

iv. Additional Curcumin Derivatives

[0664] Curcumin derivatives of the invention may include a variety of linking groups and Ar groups while retaining the necessary activity. Accordingly, additional curcumin analogs are contemplated. These include curcumin analogs containing central methylene substituents such as ethyl, propyl, butyl, isopropyl and substituted benzyl groups according to the formula:

##STR00089##

Central Methylene Substituent Analogs

[0665] Additional analogs that are contemplated are those having a pyridine ring with and without a central methylene substituent on the 7-carbon linker such as those shown in the formulas:

##STR00090## [0666] each T, U, V is independently selected from N, CH, such that each Ar ring is a pyridine; $\text{R}_{14}=\text{CH.sub.2CH.sub.3}$, $\text{CH(CH.sub.3).sub.2}$, $\text{CH.sub.2CH.sub.2CH.sub.3}$, $\text{CH.sub.2CH.sub.2CH.sub.2CH.sub.3}$, $\text{CH.sub.2C.sub.6H.sub.4X}$; $\text{X}=\text{OH}$, OCH.sub.3 , N(CH.sub.3).sub.2 , CH.sub.3 Pyridine aryl ring analogs

[0667] Many curcumin analogs which have a 5-carbon linker possess significant activity. Additional active analogs in this series may contain substituents such as hydroxy and methoxy groups on the aryl rings. Examples of these analogs are shown in the formula:

##STR00091## [0668] each R_{15} is independently selected from CH.sub.2CH.sub.3 , $\text{CH(CH.sub.3).sub.2}$, $\text{CH.sub.2CH.sub.2CH.sub.3}$, $\text{CH.sub.2CH.sub.2CH.sub.2CH.sub.3}$

Aryl Substituent Analogs

[0669] Other analogs include heterocyclic moieties, which may be substituted or unsubstituted, as shown in the formulas:

##STR00092##

Heterocyclic Analogs

v. Synthetic Curcumin Derivatives Via Different Reaction Schemes

[0670] A large number of naturally occurring and synthetic curcumin derivatives, the latter explicated by reference to the methods of synthesis thereof, are disclosed in Anand et al.

“Biological Activities of Curcumin and Its Analogues (Congeners) Made By Man and Mother Nature.” *Biochemical Pharmacology* 76 (2008):1590-1611, which is incorporated by reference herein its entirety. Representative synthetic curcumin derivatives are set forth below.

##STR00093## ##STR00094## ##STR00095## ##STR00096## ##STR00097##

where R_{18} , R_{19} , and R_{20} are each independently selected from the group consisting of H, C1-C4 alkyl, and C1-C4 amine groups.

vi. Miscellaneous Curcumin Components

##STR00098## ##STR00099##

where R24 and R25 are independently selected from the group consisting of OH, C1-C4 alkoxy, and C1-C4 alkylcarbonyloxy; R26 and R27 are independently selected from the group consisting of H, OH, C1-C4 alkoxy, and C1-C4 alkylcarbonyloxy; R28 is selected from the group consisting of H, OH, and C1-C4 alkylcarbonyloxy; and R29 is selected from the group consisting of H and C1-C4 alkoxy.

##STR00100##

where R30, R31, R32, R33, R34, R35, and R36 are independently selected from the group consisting of H, OH, C1-C4 alkoxy, and C1-C4 alkylcarbonyloxy.

##STR00101##

where R37 and R38 are independently selected from the group consisting of methoxy and OH, and the center benzene ring may be substituted in the 1,3- or the 1,4-position with the acryloyl groups. [0671] Additionally, some or all of the carbonyl (C=O) moieties of formulae C-148 through C-162 may be substituted with thio carbonyl (C=S) moieties.

vii. Presently Preferred Curcumin Components

[0672] Based upon the structure activity relationships of curcumin and curcumin derivatives, curcumin components of formula C-13A are preferred:

##STR00102##

where R1-R5 are as previously defined, and the * denotes the valence point where the additional portion of the compounds are attached. That is, the preferred components have the moiety depicted in C-13A and additional portion exemplified by the foregoing disclosure, e.g., the remaining structure of the linker L and Ar2 or R11.

[0673] The most preferred curcumin components are exemplified by formula C-12 where: [0674] Ar1 and Ar2 are each aryl groups, and especially phenyl, naphthyl, thienyl, pyridinium, and pyridyl groups, and most preferably phenyl groups, wherein all of the foregoing may be substituted or unsubstituted; [0675] L contains either 5 or 7 backbone carbon atoms and at least one of a carbonyl or hydroxyl group; [0676] at least one of R1-R5 and R6-R10 is hydroxyl.

[0677] Still more preferably, the curcumin components should have: [0678] the R2 and R7, or R3 and R7, substituents of formulas C-13 and C-14 as hydroxyl; [0679] the R3 and R8 substituents of formulas C-13 and C-14 as methoxy or ethoxy (most preferably methoxy); [0680] a β -diketone group in the linker L of formula C-12; and/or [0681] at least one, and preferably two, carbon-carbon double bonds in the linker L of formula C-12, where at least one of the double bonds is located between the a and p carbons relative to Ar1 and/or Ar2; [0682] where the curcumin component is not apocynin.

[0683] In other embodiments, the curcumin component is taken from compounds of the general formula

Ar1-L-Ar2 C-12

where Ar1 and Ar2 are independently selected from the group consisting phenyl and naphthyl groups, where the phenyl groups may be substituted with one or more substituents selected from the group consisting of OH, C1-C4 alkoxys (more preferably C1-C2 alkoxys), C1-C4 haloalkyls (more preferably C1-C2 haloalkyls), halo, and L has from 3-7 backbone carbon atoms including at least one carbonyl group therein.

[0684] Use has also been made of a commercially available, proprietary curcumin product sold under the designation "ResCu" by Davospharma, and which is asserted to be a more bioavailable form of standard curcumin. This commercial product has been tested in connection with the invention and found to be a suitable curcumin component.

The Harmine Component(s)

[0685] As used herein, "harmine component(s)" shall mean harmine, its metabolites and derivatives, isomers and tautomers thereof, and esters and pharmaceutically acceptable salts of any of the foregoing.

[0686] Harmine belongs to the family of β -carboline alkaloids, its chemical name is 7-methoxy-1-methyl 9H-pyrrole[3,4-b]indole, and its molecular formula is C₁₃H₁₂N₂O. The base structure of 3-carboline is shown in H-0.

##STR00103##

[0687] Harmine has a molecular weight of 212.25 and a melting point of 261° C. Harmine was originally isolated from *Peganum harmala*, which is widely used as a traditional herbal drug in the Middle East and North Africa. The chemical structure of harmine, 1-methyl-7-methoxy- β -carboline, is shown as follows:

##STR00104##

[0688] Harmine derivatives, which are variously substituted β -carbolines, are illustrated in the following formulas, which differ in that H-2 has a quaternary ammonium group at the 2-position, whereas H-3 has a tertiary nitrogen at the 2-position.

##STR00105##

With reference to formulas H-2 and H-3: [0689] Z₁ is hydrogen; a C₁-C₆ alkyl or haloalkyl, a C₂-C₆ alkenyl or haloalkenyl; an aryl group or an arylalkyl group, wherein the aryl group is optionally substituted at any position with halogen, nitro, hydroxyl, C₁-C₃ alkoxy, amino, sulfonate, sulfoxide, thio, ester, carboxylate, amide, borate, or C₁-C₄ boronate and wherein the alkyl group is selected from C₁-C₄ alkyl; or a heterocyclic group; [0690] Z₂ is hydrogen; a C₁-C₆ carboxyl, ester, carboxylate, acylamino, acyl halide, sulfonate, sulfoxide, thio, amide or alkoxycarbonyl group; an aryloxycarbonyl group; alkyl group optionally substituted with hydroxyl or alkoxycarbonyl; carbamate; acylhydrazine; or a heterocyclic oxycarbonyl group, where the heterocyclic portion contains from 3-7 atoms and a nitrogen, oxygen, boron, or sulfur heteroatom; [0691] Z₃ is hydrogen; a C₁-C₆ alkyl or haloalkyl; sulfonate, sulfoxide, thio, carboxylate, amide, C₂-C₆ alkenyl group; hydroxyl; a C₁-C₆ alkoxy group; a C₁-C₆ carboxylic ester group; an arylalkoxy group where the alkoxy portion contains from 1-6 carbon atoms; or a heterocyclic group containing from 3-7 atoms and a nitrogen, oxygen, boron, or sulfur heteroatom; [0692] Z₄ is hydrogen; a C₁-C₆ alkyl, haloalkyl; C₂-C₆ alkenyl group; a hydroxyalkyl group where the alkyl portion contains from 1-6 carbon atoms; an arylalkyl group wherein the aryl group is optionally substituted at any position with halogen, nitro, hydroxyl, C₁-C₃ alkoxy, borate, C₁-C₄ boronate, sulfonate, sulfoxide, thio, ester, carboxylate, amide or amino, and wherein the alkyl group is selected from C₁-C₄ alkyl group; an arylalkanone; or a heterocyclic group containing from 3-7 atoms and a nitrogen, oxygen, boron or sulfur heteroatom; [0693] Z₅ is hydrogen; a C₁-C₆ alkyl group; an aryl group substituted at any position with one or more of (1)-(2), where (1) is a C₁-C₄ alkyl group, and (2) is a C₁-C₆ carbonyl, hydroxycarbonyl, ester, sulfonate, sulfoxide, thio, ester, carboxylate, amide or amino group; arylalkyl where the alkyl group is a C₁-C₆ alkyl; 1-5 substituted arylalkyl; arylhydrocarbyl; arylcarboxyl; aryl ester group; arylamino group; or a heterocyclic group containing from 3-7 atoms and a nitrogen, oxygen, boron or sulfur heteroatom; X is a halogen; a sulfonic group, a sulfuric group, a nitric acid group or a carboxylate, in the compounds of the above formulas H-2 and H-3: [0694] Z₁ is preferably hydrogen, a C₁-C₄ alkyl group, or an arylalkyl group; more preferably hydrogen, or a C₁-C₂ alkyl group; and most preferably methyl; [0695] Z₂ is preferably hydrogen or a C₁-C₄ alkoxycarbonyl group; more preferably a C₁-C₂ alkoxycarbonyl group; and most preferably hydrogen; [0696] Z₃ is preferably hydrogen, hydroxyl, or a C₁-C₄ alkyloxy group; more preferably methoxy; [0697] Z₄ is preferably hydrogen, a C₁-C₄ alkyl group, a C₁-C₄ hydroxyalkyl group, or an optionally substituted arylalkyl group; more preferably hydrogen, a C₁-C₂ alkyl group, or a C₁-C₂ hydroxyalkyl group; still more preferably ethyl or benzyl; and most preferably hydrogen.

[0698] In certain embodiments where Z₁ is hydrogen, a C₁-C₄ alkyl group, or an arylalkyl group, Z₂ is hydrogen, hydroxyl, a C₁-C₄ carboxyl group, a C₁-C₄ ester group, a carboxylate group, a halogen, or a C₁-C₄ alkoxycarbonyl group; Z₃ is hydrogen, hydroxyl, or a C₁-C₄ alkoxy group; Z₄ is hydrogen, a C₁-C₂ alkyl group, a C₁-C₂ hydroxyalkyl group, or an optionally substituted

arylalkyl group; and Z5 is hydrogen, a C1-C6 alkyl group, or an optionally substituted aryl-alkyl group.

[0699] In certain embodiments where Z1 is hydrogen, Z2 is a C1-C2 alkoxy carbonyl group, Z3 is hydrogen, and Z4 is C1-C2 alkyl group, or an optionally substituted arylalkyl group.

[0700] In certain embodiments where Z1 is hydrogen, Z2 is ethoxycarbonyl, Z3 is hydrogen, and Z4 is ethyl or benzyl.

[0701] In certain embodiments where Z1 is methyl, Z2 is ethoxycarbonyl, Z3 is hydrogen, Z4 is pentafluorobenzyl, and Z5 is hydrogen.

[0702] In certain embodiments where Z1 is hydrogen, Z2 is hydrogen, Z3 is hydrogen, Z4 is benzyl, Z5 is benzyl, and X is bromine.

[0703] Certain preferred harmine components include β -carboline, tryptoline, pinoline, harmine, harmalol, harmalol hydrochloride hydrate, tetrahydroharmine, harmane, harmol, vasicine, and vasicinone.


[0704] Other harmine derivatives include:

##STR00106##

where Z1, Z2, and Z9 are set forth below, and X1 is a halogen preferably Cl, Br, and F or carboxylate anion.

[0705] Based upon the structure-activity relationships of harmine and harmine derivatives, preferred components are illustrated by formula H-6:



##STR00107##



where the three * denote valence points where respective additional atoms or portions of the components are attached to give complete compounds; the dotted lines in the right-hand six-membered ring of H-6 indicate that the ring may optionally be aromatic; Z1 and Z4 are independently H or a C1-C6 alkyl group, more preferably a C1-C4 alkyl group, and most preferably methyl for Z1 and hydrogen for Z4; and the remaining unoccupied positions on the six-membered rings (i.e., the left-hand phenyl group and the right-hand six-membered ring) are each independently selected from the group consisting of Z1, as previously defined. That is, the preferred harmine components have the moiety depicted in H-1 and additional portions exemplified by the foregoing disclosure. N⁺ is optionally substituted with Z5, as previously defined. In some cases, X^{sup.-} as previously defined is present. One of ordinary skill in the art would understand that  custom-character indicates a double or single bond is present. For example, where one of the -* substituents is OCH3 and is located at the No. 7 position as illustrated in formula H-1, the other -* substituents on the remaining unoccupied positions on the terminal 6-membered rings are all H, Z4 is H and Z1 is methyl, and Z5 and X^{sup.-} not existing, the resulting compound is harmine.

[0706] One class of preferred harmine components are shown in formula H-3, where Z3 is a methoxy or ethoxy group and Z4 is a benzyl group. A second preferred class of harmine components is shown in formula H-2, where Z1 is a methyl group, Z2 is hydrogen, Z3 is a benzyloxy group, Z4 and Z5 are benzyl groups.



TABLE-US-00004 Z6 Z7 Z8 [00108]  embedded image CH.sub.3CH.sub.2CH.sub.2CH.sub.2—



[00109]  embedded image CH.sub.3— [00110]  embedded image [00111]  embedded image



[00112]  embedded image [00113]  embedded image CH.sub.3CH.sub.2CH.sub.2CH.sub.2—




[00114]  embedded image [00115]  embedded image CH.sub.3CH.sub.2CH.sub.2CH.sub.2— H




— [00116]  embedded image [00117]  embedded image [00118]  embedded image [00119]



 embedded image CH.sub.3CH.sub.2CH.sub.2CH.sub.2— [00120]  embedded image [00121]


 embedded image CH.sub.3CH.sub.2CH.sub.2CH.sub.2— [00122]  embedded image [00123]

 embedded image CH.sub.3CH.sub.2CH.sub.2CH.sub.2— [00124]  embedded image [00125]

 embedded image [00126]  embedded image [00127]  embedded image [00128]

 embedded image [00129]  embedded image [00130]  embedded image [00131]

 embedded image [00132]  embedded image H— CH.sub.3CH.sub.2CH.sub.2CH.sub.2—

[00133]  embedded image

[0707] In other embodiments, preferred harmine components include compounds of the formula
##STR00134##

R39 is selected from the group consisting of H, C1-C4 alkoxys (more preferably C1-C2 alkoxys), O, and OH substituents, R40 is selected from the group consisting of H, C1-C4 alkyls (more preferably C1-C2 alkyls), C1-C4 haloalkyls (more preferably C1-C2 haloalkyls), C1-C4 alkoxys (more preferably C1-C2 alkoxys), nitrophenyls, C1-C4 organic acids (more preferably C1-C2 organic acids), C1-C4 alkylalcohols (more preferably C1-C2 alkylalcohols), and C1-C6 alkyl esters (more preferably C2-C4 alkyl esters), and R41 is selected from the group consisting of nothing, H, and nitrophenyls.

The Isovanillin Component(s)

[0708] As used herein, “isovanillin component(s)” refers to isovanillin, its metabolites and derivatives, isomers and tautomers thereof, and esters and pharmaceutically acceptable salts of any of the foregoing.

[0709] Isovanillin is a phenolic aldehyde having a hydroxyl group at the meta position and a methoxy group at the para position. Isovanillin is illustrated in the following structure:

##STR00135##

[0710] Isovanillin is metabolized in vivo to vanillin, which is the same as structure I-1, except that the hydroxyl and methoxy substituents are exchanged, i.e., in vanillin, the hydroxyl group is in the para position, and the methoxy group is in the meta position.

[0711] Useful derivatives of isovanillin have the following general formula:

##STR00136##

where at least one of Q1-Q6 is an alkoxy group and/or an aldehyde group, especially where the alkoxy and/or aldehyde groups contain from 1-6 carbon atoms; more preferably, where Q1 is an aldehyde, alcohol, amine, carbonyl, carboxylate, C1-C6 alkylhydroxy, ester, imidazole, nitro, sulfonate, sulfoxide, thio, amide, oxime, borate, or boronate, or semicarbazone group; the Q2-Q6 groups are independently selected from hydrogen, hydroxyl, halo, nitro, C1-C6 alkoxy, C1-C6 alkyl or alkenyl groups, with the proviso that at least one of the Q2-Q6 groups is an alkoxy group. In more preferred forms, the aldehyde, alcohol, amine, carbonyl, carboxylate, and ester groups should have a C1-C6 carbon chain length, the boronate is a C1-C4 boronate, and at least one of the Q2-Q6 groups is an alkoxy group (most preferably methoxy), and another is a hydroxyl group; advantageously, the alkoxy and hydroxyl groups are adjacent each other. In particularly preferred forms, the remainder of the Q2-Q6 groups, apart from the alkoxy and hydroxyl groups, are all H. Advantageously, the isovanillin components of the invention should include only one phenyl group and are free of any fused ring structures (as used herein, “fused ring structures” refer to structures such as found in naphthalene or anthracene where two rings share common atoms).

[0712] Some preferred isovanillin components are selected from isovanillin, vanillin, ethyl vanillin, ortho-vanillin, vanillic acid, isovanillic acid, vanillic alcohol, isovanillic alcohol, 6-bromine-5-hydroxy-4-methoxybenzaldehyde, 4-hydroxy-3,5-dimethoxybenzaldehyde, 4,5-dihydroxy-3-methoxybenzaldehyde, 5-hydroxy-4-methoxybenzaldehyde, 2-Benzyloxy-3-methoxybenzaldehyde, 2-(2-benzyloxy-3-methoxyphenyl)-1H-benzimidazole, N-1-(2-benzyloxy-3-methoxybenzyl)-2-(2-benzyloxy-3-methoxyphenyl)-1H-benzimidazole, and (S)-1-(2-benzyloxy-3-methoxyphenyl)-2, 2, 2-trichloroethyl benzenesulfonate (regarding the last four compounds, see Al-Mударis et al., *Anticancer Properties of Novel Synthetic Vanillin Derivatives* (2012)).

[0713] Certain imidazole derivatives (such as nitro-imidazoles) have been shown to have significant anticancer activity, see, e.g., Sharma et al., “Imidazole Derivatives Show Anticancer Potential by Inducing Apoptosis and Cellular Senescence,” *Med.Chem.Commun.* 2014, 5, 1751, incorporated by reference herein. Representative imidazole derivatives include:

##STR00137## ##STR00138##

Additional isovanillin derivatives of interest are:

##STR00139##

[0714] It will be appreciated that the aryl group substituents in formulas I-14-I18 can be at any desired position on the on the aryl rings, e.g., the —OH need not be adjacent and the heteroatom substituents may be at any desired location.

[0715] Based upon the structure-activity relationships of isovanillin and isovanillin derivatives, preferred components are illustrated by formula I-13:

##STR00140##

where the two * denote valence points where respective additional atoms or portions of the components are attached to form complete compounds, the remaining phenyl ring positions not taken by the two depicted moieties are independently R1 as previously defined with reference to formula C-13, and Q7 is H or a C1-C6 alkyl group, more preferably a C1-C4 alkyl group, and most preferably methyl. That is, the preferred isovanillin components have the moiety depicted in I-13 and additional portions exemplified by the foregoing disclosure. For example, where * is CHO, the substituent —O-Q7 is located at the para position, Q7 is a methyl group, and the —O—* group is located at the meta position, the * of the —O—* moiety is H, and the remaining unoccupied positions on the phenyl ring are all H, the resulting compound is isovanillin. In preferred forms, the isovanillin component is not apocynin.

[0716] In other embodiments, the isovanillin components are selected from the compounds of the formula

##STR00141##

where Q9 is selected from the group consisting of C1-C4 aldehydes (more preferably C1-C2 aldehydes), C1-C4 alkylalcohols (more preferably C1-C2 alkylalcohols), C1-C4 alkyl esters (more preferably C1-C2 alkyl esters), and C1-C4 organic acids (more preferably C1-C2 organic acids), Q10 is selected from the group consisting of OH, H, C1-C6 alkyl esters (more preferably C1-C4 alkyl esters), C1-C4 alkoxys (more preferably C1-C2 alkoxys), and benzyloxy, and Q11 is selected from the group consisting of H, C1-C4 alkoxys (more preferably C1-C2 alkoxys), and OH.

The Complete Compositions of the Invention

[0717] In the case of the preferred three-component compositions of the invention, made by combining individual quantities of normally highly purified curcumin component(s), harmine component(s), and isovanillin component(s), the as-added amounts should give weight ratios of about 10:1.7:0.85 (isovanillin component(s):harmine component(s):curcumin component(s)), but more broadly, the ratios are approximately 0.1-25:0.1-5:0.1-5 (isovanillin component(s):harmine component(s):curcumin component(s)). In this respect, it will be seen that the isovanillin component(s) of the preferred GZ17-6.02 product is/are the preponderant component(s) in the compositions on a weight basis, with the harmine and curcumin component(s) being present in lesser amounts on a weight basis. Generally, the isovanillin component(s) in the most preferred product should be present at a level at least three times (more preferably at least five times) greater than that of each of the harmine and curcumin component(s), again on a weight basis. However, the invention is not limited to such weight ratios. As noted in Example 17, an isovanillin component:harmine component:curcumin component weight ratio of 1:1:1 is also effective. In terms of amounts of the three components, the isovanillin component(s) should be present at a level of from about 25-85% by weight, the harmine component(s) should be present at a level of from about 7-50% by weight, and the curcumin component(s) should be present at a level of from about 5-40% by weight, based upon the total weight of the three components taken as 100% by weight.

[0718] In the case of two-component compositions in accordance with the invention made up of curcumin component(s) plus harmine component(s), the weight ratio of the curcumin component(s):harmine component(s) should range from about 0.01:1 to 10:1; and the weight amounts of the curcumin component(s) should range from about 20% to 75% by weight (more preferably from about 30-55% by weight, and most preferably about 45% by weight), and the weight amounts of the harmine component(s) should range from about 25% to 80% by weight (more preferably from about 45% to 70% by weight, and most preferably about 55% by weight),

based upon the total weight of the two components in the compositions taken as 100% by weight. Generally, the amount of the harmine component should be greater than the amount of the curcumin component in these two-component compositions.

[0719] Two-component compositions made up of isovanillin component(s) plus curcumin component(s) should have a weight ratio of the isovanillin component(s):curcumin component(s) ranging from about 0.5:1 to 25:1; and the weight amounts of the isovanillin component(s) should range from about 25% to 95% by weight (more preferably from about 75% to 95% by weight, and most preferably about 88% by weight), and the weight amounts of the curcumin component(s) should range from about 5% to 75% by weight (more preferably from about 5% to 25% by weight, and most preferably about 12% by weight), based upon the total weight of the two components in the compositions taken as 100% by weight.

[0720] Finally, two-component compositions made up of isovanillin component(s) plus harmine component(s) should have a weight ratio of the isovanillin component(s):harmine component(s) should range from about 0.5:1 to 15:1; and the weight amounts of the isovanillin component(s) should range from about 25% to 95% by weight (more preferably from about 75% to 95% by weight, and most preferably about 85% by weight), and the weight amounts of the harmine component(s) should range from about 5% to 75% by weight (more preferably from about 5% to 25% by weight, and most preferably about 15% by weight) by weight, based upon the total weight of the two components in the compositions taken as 100% by weight.

[0721] In this connection, it should be understood that the isovanillin component(s) of any of the foregoing two- or three-component compositions containing isovanillin component(s) may be made up of initially added isovanillin component(s) plus any degradation products of the other component(s) yielding products within the ambit of the isovanillin component(s); for example, it is known that under certain circumstances curcumin will spontaneously degrade to give quantities of vanillin, and, in such circumstances, the total amount of the isovanillin component(s) would be the initially added amounts together with these degradation products. More broadly, the ultimate amounts of the curcumin, harmine, and isovanillin component(s) in the compositions of the invention should be determined based upon the actual contents of the component(s) in question, regardless of whether these component(s) are derived from the originally added component(s) or as degradation products of some or all of these originally added component(s).

[0722] As indicated previously, levels of dosing using the compositions of the invention is quite variable owing to factors such as the patient's age, patient's physical condition, the type of condition(s) being treated (e.g., specific cancer(s)), and the severity of the conditions. In general, however, regardless of the dosage form or route of administration employed, such as liquid solutions or suspensions, capsules, pills, or tablets, via oral, parenteral, or injection, the compositions should be dosed of from about 5 to 2000 mg per day, and more usually from about 100-800 mg per day. Such dosages may be based on a single administration per day, but more usually multiple administrations per day.

[0723] Finally, the invention also embraces compositions and methods where the individual components are provided and administered separately to subjects, so long as the therapeutic effects of the invention are substantially preserved.

Examples 1-28

[0724] The following Examples set forth preferred therapeutic agents and methods in accordance with the invention, but it is to be understood that these examples are given by way of illustration only, and nothing therein should be taken as a limitation upon the overall scope of the invention. A number of the Examples set forth various tests using the preferred drug of the invention, GZ17-6.02, which is sometimes referred to as GZ17Syn-6.02.

[0725] The GZ17-6.02 product of the Examples was made by dispersing quantities of solid synthetic isovanillin (771 mg, 98% by weight purity), synthetic harmine (130.3 mg, 99% by weight purity), and a commercially available curcumin product derived by the treatment of turmeric (98.7

mg, containing 99.76% by weight curcuminoids, namely 71.38% curcumin, 15.68% demethoxycurcumin, and 12.70% bisdemethoxycurcumin), in a 1 mL ethanol at a weight ratio of 771:130.3:98.7 (isovanillin:harmin:curcumin product) in ethanol followed by sonication of the dispersion. Aliquots of this stock solution were then used to create the different GZ17-6.02 dilutions using stock media of the cells in question.

[0726] The two-component products described in Example 23 were made in the same fashion as the GZ17-6.02 agent, and the weight ratios of the two components were as set forth immediately above, e.g., the isovanillin/harmin sub-combinations contained a weight ratio of 771:130.3, and so forth.

Example 1

[0727] In this example, the preferred GZ17-6.02 product was tested with two different human head and neck cancers (HN5 and OSC19), in order to determine the extent of cell death induced by the product.

Methods

[0728] The respective cells were individually cultured in a growth medium prepared using RPMI-1640 medium containing 11.1 mM D-glucose with 10% fetal bovine serum, 10 mM HEPES, 2 mM L-glutamine, 1 mM sodium pyruvate, 0.05 mM 2-mercaptoethanol, and Antibiotic-Antimycotic. These cells were maintained in T75 tissue culture flasks in a humidified incubator at 37° C. and 5% CO₂. The media were changed on a third day after cell plating, and the cells were passaged on day 5 by trypsinization.

Formation of Cancer Spheroids

[0729] Custom-made micromolds with 100 μ m diameter wells were loaded with the cells (U.S. Pat. No. 8,735,154, incorporated by reference herein). The media were changed every day by partial replacement. Cell aggregates were allowed to form in the micromolds for 72 hours and then were transferred to a 100 mm non-treated tissue culture dishes for 3 additional days. The spheroids were passed through 70 μ m and 100 μ m cell strainers (#3431751 and #43152, Corning, Tewksbury, MA) and maintained in HEPES Balanced Salt Solution comprised of: 20 mM HEPES, 114 mM NaCl, 4.7 mM KCl, 1.2 mM KH₂PO₄, 1.16 mM MgSO₄, 2.5 mM CaCl₂, 25.5 mM NaHCO₃ and 0.2% bovine serum albumin (fatty acid free), pH 7.2.

Testing of GZ17-6.02 on Head and Neck Cancer Spheroids

[0730] Individual wells of a 96 well plate were manually loaded with 15-20 cancer spheroids each, and exposed to selected doses of GZ17-6.02. Each trial was run with at least 4 replicates at each dose. After a 24 hour exposure to the selected dosages of GZ17-6.02, PrestoBlue (Life Technologies, Inc) was added to each well and fluorescence readings were taken 4-6 hours later with an excitation wavelength of 485 nm and an emission wavelength of 560 nm, using a microplate reader (Enspire Multimode, PerkinElmer). Results were averaged following background subtraction and normalized to untreated cell controls.

[0731] FIG. 1 demonstrated that GZ17-6.02 dramatically kills HN5 and OSC19 cancer cells in a dose-dependent manner. The X axis is the increasing dose of GZ17-6.02 and the Y axis is the average number of live cells in each well at the end of a 24 hour exposure to the stated dose of GZ17-6.02.

Example 2

[0732] In this example, GZ17-6.02 was found to induce significant cancer cell death in human pediatric leukemia cells and pediatric osteosarcoma in a dose-dependent manner.

[0733] Jurkat leukemia cells were grown in suspension in media (RPMI supplemented with 10% FBS), maintained at approximately 500,000 cells/mL. The cells were plated in 96-well plates, and each well was exposed to a selected dose of GZ17-6.02 for 24 hours (a minimum of 4 replicates for each dosage). These cells were not treated to generate spheroids, but were directly plated onto the well plates. After a 24 hour exposure to the selected dosages of GZ17-6.02, PrestoBlue (Life Technologies, Inc) was added to each well and fluorescence readings were taken 4-6 hours later

with an excitation wavelength of 485 nm and an emission wavelength of 560 nm, using a microplate reader (Enspire Multimode, PerkinElmer). Results were averaged following background subtraction and normalized to untreated cell controls.

[0734] Human osteosarcoma cells (HOS) also had a significant dose-dependent increase in cell death when exposed to GZ17-6.02 for 24 hours. The methods to create and test osteosarcoma spheroids were the same as the methods described in Example 1.

[0735] As illustrated in FIGS. 2A and 2B, increasing doses of GZ17-6.02 induced significant cell death in both leukemia and osteosarcoma cell types tested.

Example 3

[0736] In this example, the effect of increasing doses of GZ17-6.02 in killing human lymphoma cells (mo205) and human lung cancer cells (H358) was tested.

[0737] The lymphomas treatment methods used were identical to those described in Example 2 relative to the pediatric leukemia cells, whereas the lung cancer treatment method was the same as described in Example 1.

[0738] FIGS. 3A and 3B set forth the results of these tests, and demonstrate the effectiveness of GZ17-6.02 in inducing lymphoma and lung cancer cell death.

Example 4

[0739] In this example, the effect of increasing doses of GZ17-6.02 in killing human ovarian cancer cells (A1847) and human prostate cancer cells (22rv1) was tested.

[0740] The cells were treated and tested as set forth in Example 1.

[0741] This test confirmed that both types of cells experienced dose-dependent death when exposed to GZ17-6.02, see FIGS. 4A and 4B.

Example 5

[0742] In this example, the effect of increasing doses of GZ17-6.02 in killing human breast cancer cells (du4475) and human pancreatic cancer cells (panc-1) was tested.

[0743] The cells were treated and tested as set forth in Example 1, except that a different growth medium was used, namely Dulbecco's Modified Eagle Medium (DMEM) with 10% fetal bovine serum, and 1% penicillin/streptomycin mix (Sigma-Aldrich).

[0744] This test confirmed that both types of cells experienced dose-dependent death when exposed to GZ17-6.02, see FIGS. 5A and 5B.

[0745] As is evident from Examples 1-5, ten different cancer types were tested with GZ17-6.02 and they all were sensitive to the therapeutic agent. In most of the cancers (head and neck cancer (OSC19), leukemia, osteosarcoma (bone cancer), lymphoma, lung cancer, ovarian, prostate, breast, and pancreatic cancer), the compound killed nearly all of the cells in the dish at a dose of 3.13-6.25 mg/mL. This array of cancer types represents a substantial portion of human cancers. Moreover, the results confirm that the therapeutic agent is effective in both solid tumor cancers and blood cancers.

Example 6

[0746] In this test, non-cancerous integumental (dermal) fibroblasts (hgf-1) were tested with GZ17-6.02 and compared with prostate cancer (22rv1) and ovarian cancer cells (A1847), to determine the effect of GZ17-6.02 on the non-cancerous cells versus the prostate cancer and ovarian cancer cells.

[0747] The fibroblasts were treated as follows: The cells were grown to confluence in the DMEM medium of Example 5, and then placed in 96-well plates where they adhered to the bottom of the plates. Each well was then exposed to a selected dose of GZ17-6.02 for 24 hours (a minimum of 4 replicates each), and tested according to Example 1. The prostate cancer and ovarian cancer results were taken from the previous Example 4.

[0748] As illustrated in FIG. 6, the cancer cells begin to die at lower GZ17-6.02 doses than the non-cancerous fibroblast cells, demonstrating that GZ17-6.02 is more toxic to cancer cells versus the non-cancerous fibroblasts.

Example 7

[0749] Migration of cells away from a primary tumor, followed by invasion of those tumor cells

first into the blood vessels and later out of the blood vessels and into other tissues, are the two steps essential for metastases to form from a primary tumor. The cells were grown in the DMEM media of Example 5.

[0750] Head and neck cancer cells (OSC19) were tested using a commercially available migration assay kit, Cultrex 96 Well Migration Assay (R&D Systems). The cell movement from the upper compartment to the lower compartment was measured as representative of cell migration. A negative control migration was also measured, using the GZ17-6.02 vehicle, namely a dilution of ethanol in the growth media to the same concentration as GZ17-6.02 in the growth media.

[0751] The head and neck cancer cells were also assayed for migration using a Cultrex Cell Invasion Assay (R&D Systems). The ability of the head and neck cancer cells to invade a collagen matrix was measured by counting cells that exited the upper chamber and passed through a collagen-surrounded lower chamber. Similarly, a negative control for invasion was also measured, using the same ethanol vehicle employed in the migration assay.

[0752] As illustrated in FIG. 7A, the GZ17-6.02 significantly inhibited migration so that almost no head and neck cancer cells could migrate from the site of origin. Likewise, GZ17-6.02 significantly inhibited invasion (FIG. 7B) by more than 60%.

Example 8

[0753] In this test, the synergism of GZ17-6.02 with doxorubicin (a common chemotherapy drug) was tested on ovarian cancer cells. In this test, increasing doses of doxorubicin were administered to ovarian cancer spheroids and cell deaths were measured, as described in Example 1. The protocol involved testing doxorubicin alone at various dosages (FIG. 8, doxorubicin alone—closed circles), with comparative tests using the various dosages of doxorubicin in combination with either 0.2 mg/mL (FIG. 8 open circles) or 0.4 mg/mL (FIG. 8 closed triangles) GZ17-6.02. Additionally, the induced cell death results from use of 0.2 mg/mL GZ17-6.02 alone (FIG. 8, line A) and 0.4 mg/mL GZ17-6.02 alone (FIG. 8, line B) were determined.

[0754] As is evident from FIG. 8, even at the lowest dose of doxorubicin (0.002 μ M), the addition of GZ17-6.02 at 0.2 and 0.4 mg/mL increased cell death significantly below the level of either compound alone. At low doses of doxorubicin alone (up to 0.158 μ M), doxorubicin was ineffective. GZ17-6.02 administered alone at a dose of 0.2 mg/mL resulted in cell death at a level represented by the upper horizontal dotted line in FIG. 8. Moreover, GZ17-6.02 administered alone at a dose of 0.4 mg/mL resulted in cell death at a level represented by the lower horizontal dotted line in FIG. 8. Thus, even at these low dosages, addition of GZ17-6.02 significantly increased cell death (see data points under both of the horizontal dotted lines in FIG. 8). This demonstrates the synergy of the combination of GZ17-6.02 and doxorubicin, even at low dosages of both GZ17-6.02 and doxorubicin. This trend is confirmed at higher dosages of doxorubicin, where the combined products were always more effective than doxorubicin alone.

Example 9

[0755] There are multiple general ways in which cancer (blood or solid tumors) can be blocked by chemotherapy. One is to actually induce death in the cancer cells (apoptosis), and a second is to block the ability of cells to receive nutrients, basically starving them to death (necrosis). This test demonstrates that at the ED.sub.50 of GZ17-6.02 on head and neck cancer cells (OSC19) caused cell death via apoptosis. In the test, head and neck cancer cells were stained with an apoptotic fluorophore and sorted by flow cytometry, which allows a single cell to be counted and measured for fluorescence simultaneously. This histograms in FIGS. 9A and 9B illustrate the shift in fluorescence intensity to a lower level, indicating apoptotic cell death. While the control (FIG. 9A, head and neck cancer cells without GZ17-6.02) had little apoptotic cell death (19.8%), 48.2% of the cells exposed to GZ17-6.02 died of apoptosis (FIG. 9B). This indicates one type of mechanism of action of GZ17-6.02.

Example 10

[0756] Caspase proteins are intracellular proteins involved in the apoptotic programmed cell death

pathway. In instances where live lung cancer cell number decrease while a caspase protein increases, it can be inferred by association that the caspase was activated, and was thus associated with the molecular pathway of the cell death.

[0757] In these tests, the lung cancer cells (H358) were assayed using caspase testing kits (Promega Caspase-Glo 3/7, -Glo 2, -Glo 6, -Glo 8, and -Glo 9). Sample lung cancer cells were homogenized on ice in the provided preparation buffer and agitated on a plate shaker at low speeds (around 300 rpm). The supernatants were exposed to the provided assay solutions and read on a microplate reader for luminescent output (Enspire Multimode, PerkinElmer).

[0758] The test results indicated that caspases 3 and 7 (FIG. 10A, filled triangles) increased dramatically in association with cell death (filled circles). In addition, results indicated that caspase 6 (FIG. 10B, open circles) increased in association with cell death (filled circles). In contrast, caspase 9 was not activated (FIG. 10A, open circles). This implies a receptor-mediated cell death pathway and not a mitochondrial cell death pathway. In order to confirm this, an ATP assay (CellTiter-Glo, Promega) was conducted in which toxicity of the mitochondria were measured at the same time as cell death, based upon ATP concentrations. If a mitochondrial toxicity were responsible for cell death, it would happen at lower doses or prior in time to the cell death. FIG. 10C confirms that there was no significant mitochondrial toxicity (filled circles) while there was cell death (open circles).

Example 11

[0759] The procedures of Example 10 were followed, with testing of GZ17-6.02 on ovarian cancer cells (A1847). Caspases 3 and 7 were activated at low doses of GZ17-6.02, indicating a receptor-mediated cell death (FIG. 11A, filled circles). Caspase 6 was active at low doses of GZ17-6.02 (FIG. 11B, filled circles). However, caspase 8 was activated, which typically signals cell death via the mitochondria (FIGS. 11C, filled circles) and caspase 9 was also activated (FIG. 11D, filled circles). In order to confirm the mitochondrial cell death pathway, mitochondrial toxicity was monitored in comparison to cell death (FIG. 11E, filled circles), indicating that the mitochondrial toxicity was at least partially involved in the subsequent cell death.

Example 12

[0760] The procedures of Example 10 were followed, with testing of GZ17-6.02 on osteosarcoma cells (HOS). These tests indicated that levels of caspases 3 and 7 were significantly higher when exposed to doses of GZ17-6.02 over 0.4 mg/mL before or during cell death (FIG. 12A, filled circles). Caspase 6 was activated at high doses of GZ17-6.02 (FIG. 12B, filled circles). In contrast, caspase 2 (FIG. 12C, filled circles) and caspase 9 (FIG. 12D, filled circles) showed no activation that would indicate that they were involved in the cell death pathways induced by GZ17-6.02.

Example 13

[0761] The procedures of Example 10 were followed, with testing of GZ17-6.02 on human head and neck cancer cells (OSC19). FIG. 13 (filled circles) indicates that there is no mitochondrial toxicity involved in GZ17-6.02 induced cell death in the head and neck cancer cells.

Example 14

[0762] In addition to directly killing cancer cells, chemotherapeutic agents can work by blocking the rapid proliferation of cancer cells, thus eventually halting the progression of the cancer.

[0763] In this test, a multiplex dot-based Western assay for proliferative proteins was used to measure the relative amounts of proteins known to be involved in cell proliferation signaling, in both a control test (no GZ17-6.02) and an inactivation test (with GZ17-6.02, at the ED.sub.50 for head and neck cancer cells).

[0764] Following a 24 hour exposure to GZ17-6.02 or vehicle head and neck cancer cells (OSC19) were homogenized and the lysate loaded onto the Human Phospho-Kinase Array Kit (R&D Systems) using the following procedure. The antibody-attached membranes were incubated overnight at 4° C. with the respective cell lysates, followed by repeated washing and incubation in the detection antibodies for 24 hours. After subsequent repeated washes, streptavidin-TRP was

applied for 30 minutes. Membranes were washed and exposed to film after applying chemiluminescence reagent mix.

[0765] Cellular proteins specific to a pathway of cell proliferation were tested in duplicate (resulting in 2 side-by-side dots for each protein in FIG. 14A). The darker the dot, the more relative protein was present in the homogenate. As shown in FIG. 14B, cell proliferation proteins showed a dramatic reduction in amount when exposed to the head and neck cancer cell ED.sub.50 dose of GZ17-6.02. The proteins showing the greatest reduction included epidermal growth factor receptor (block a compared to block a'), extracellular-signal-regulated kinase (block b compared to block b'), the catalytic subunit of AMP-activated kinase (block c compared to block c'), 0-catenin (block d compared to block d'), STAT2 (block e compared to block e') and Chk-2 (block f compared to block f').

Example 15

[0766] In this example, two independent scientists conducted identical tests with ovarian cancer and lung cancer cells (A1847 and H358) using the techniques described in Example 1, in order to determine induced cell death upon application of GZ17-6.02. The nearly identical results (filled circles versus open circles) confirm the reliability of the testing as a determination of GZ17-6.02 effectiveness. FIG. 15A sets forth the ovarian cancer results, whereas FIG. 15B gives the lung cancer cell results.

Example 16

[0767] Six-week old FOXN1 mice were inoculated with human head and neck cancer cells (OSC19) bilaterally into the flank region, in order to induce the formation of palpable tumors. The bilateral tumors were measured at one week after cell injection, and had each grown to an average volume of approximately 9 mm.sup.3. Half of the mice were injected daily with 15 mg/kg body weight GZ17-6.02 stock solution into the right side tumors, starting on day 7 after inoculation of the tumor cells. The other half of the mice were injected with the same volume of the ethanol carrier of the stock solution into the right side tumors. Thus, each mouse had 2 palpable tumors, one on either flank, but only one side was injected with GZ17-6.02. Every 2-4 days, the tumors on both sides of each mouse were measured with vernier calipers in 2 perpendicular dimensions and the volumes calculated.

[0768] FIG. 16A shows that GZ17-6.02 dramatically halted tumor growth in the treated mice, so that by 3 weeks there was a distinct and statistically significant difference between the vehicle-treated controls and the GZ17-6.02 treated mice. Further, the trend in the GZ17-6.02 treated mice was that the tumors began to decrease in volume from day 21 (although the decrease was not statistically significant) from tumor size at day 28. FIG. 16A shows the decline in tumor volume in the tumors that were directly injected with GZ17-6.02. FIG. 16B shows the halt in tumor growth that occurred in the non-injected tumors on the contralateral side of the neck of the treated mice. FIG. 16B demonstrates that GZ17-6.02 has a systemic anticancer effect, i.e., the contralateral tumor size decrease without direct injection of GZ17-6.02 indicates that the agent traveled through the bloodstream of the mice.

[0769] Throughout the study, the animals showed no signs of complications, distress, or toxicity. Thus, none of the over 30 mice in the study exhibited any observable adverse reactions. Daily observation of the mice documented that none suffered any weight loss, new tumor formation, loss of appetite, change in fur appearance or grooming behavior, or change in activity owing to lethargy. Moreover, there were no gross abnormalities of any of the internal organs of the mice upon necropsy. It was thus concluded that there were no drug-drug interactions, which are common with multiple-drug anti-cancer compositions.

[0770] In another mice study, a head and neck tumor was surgically removed from a human patient, and approximate 35 mg-portions of the tumor were implanted in a first randomized group of ten nude-FOXN1 mice using a 5% ethanol in saline vehicle via oral gavage. A second randomized group of ten mice was treated only with the vehicle as a control, in the same manner as the first

group. The first mice group was treated with 30 mg/kg/day doses of GZ17-6.02 five days/week, and the doses were increased to 50 mg/kg/day during the second week of treatment. The two groups of mice were treated for a total of three weeks, and tumor volumes were measured twice a week using a Vernier caliper. The results of this study are set forth in FIG. 16C, which illustrates that the fractional tumor volumes relative to the pretreatment tumor size were reduced in the first group of mice, while the control mice exhibited gradually increasing tumor burdens.

Example 17

[0771] In this Example, a different ratio of isovanillin, harmine, and curcumin was used, as compared with GZ17-6.02, namely $\frac{1}{3}$ by weight of each component. This formulation was prepared by mixing the three GZ17-6.02 components in 1 mL of ethanol to obtain a secondary stock solution. FIG. 17A shows that the original stock solution (filled circles) and the secondary stock solution (open circles) are bioequivalent in terms of ovarian cancer cell (A1847) kill rate. FIGS. 17B and 17C show the same general effect for lung cancer cells (H358) and prostate cancer cells (22rv1).

Example 18

[0772] In this Example, a known ovarian cancer cell kill rate for GZ17-6.02 was selected, and the amount of isovanillin in this dosage (0.78 mg/mL) was tested against GZ17-6.02. Additionally, sub-combinations of isovanillin:curcumin and isovanillin:harmine were tested, where the concentrations of the two ingredients were identical to those in the GZ17-6.02. Thus, the concentration of the isovanillin:curcumin product was 0.78 mg/mL isovanillin and 0.13 mg/mL curcumin, and the concentration of the isovanillin:harmine product was 0.78 mg/mL isovanillin and 0.099 mg/mL harmine. FIGS. 18A-18D illustrate these results with ovarian cancer cells (A1847), lung cancer cells (H358), prostate cancer cells (22rv1), and lymphoma cells (M0205). As is evident, the two-component products each exhibited greater kill rates as compared with isovanillin alone.

Example 19

[0773] The procedures of Example 18 were followed, except that curcumin was tested alone versus GZ17-6.02, together with curcumin:isovanillin and curcumin:harmine two-component products. In this test, the curcumin was present at a level of 0.78 mg/mL and the concentrations in the curcumin:isovanillin product were 0.78 mg/mL curcumin and 3.59 mg/mL isovanillin, and in the curcumin:harmine product the concentrations were 0.78 mg/mL curcumin and 0.59 mg/mL harmine. FIGS. 19A-19D illustrate the results of this test, and confirm that the two-component products give better results than curcumin alone.

Example 20

[0774] The procedures of Example 18 were followed, except that harmine was tested alone versus GZ17-6.02, together with harmine:isovanillin and harmine:curcumin two-component products. In this test, the harmine was present at a level of 0.78 mg/mL and the concentrations in the harmine:isovanillin product were 0.78 mg/mL harmine and 6.09 mg/mL isovanillin, and in the harmine:curcumin product the concentrations were 0.78 mg/mL curcumin and 1.03 mg/mL curcumin. FIGS. 20A-20D illustrate the results of this test, and confirm that the two-component products give better results than harmine alone.

Example 21

[0775] In this example, the individual components of GZ17-6.02 were tested against lymphoma cancer cells (M0205) at the component concentrations found in GZ17-6.02, at dosage rates of 12, 24, 48, and 96 μ g/mL. The theoretical additive effect (black bar) of the 3 components was also calculated in each case and shown versus the actual test results found using GZ17-6.02. FIGS. 21A-21D illustrate the results of these tests, and confirm that, at the tested dosages, the GZ17-6.02 product had a greater effect than the individual components and the theoretical additive effect thereof, thus establishing synergistic effects. In this regard, those skilled in the art understand that dosages of anticancer products actually administered to patients depend on a number of factors,

including age, weight, sex, physical conditions, types of cancers, and stages of cancers. By the same token, it should also be understood that at some dosage levels and/or specific concentrations of components, the results of FIGS. **21A-21D** may not be duplicated. These same considerations apply to the tests set forth in Examples 22 and 23 below. In most cases, however, compositions giving therapeutic synergies at the selected dosages are preferred.

Example 22

[0776] In this example, the individual components of GZ17-6.02 were tested against ovarian cancer cells (A1847) and breast cancer cells (du4475) at the component concentrations found in GZ17-6.02, at dosage rates of 12 and 24 $\mu\text{g/mL}$. The theoretical additive effect (black bar) of the 3 components was also calculated in each case and shown versus the actual test results found using GZ17-6.02. FIGS. **22A** and **22B** illustrate the results of these ovarian cancer tests, and confirm that, at the tested dosages, the GZ17-6.02 product had a greater effect than the individual components and the theoretical additive effect thereof. In an additional test at 48 $\mu\text{g/mL}$ dosage rate, this effect was not observed. FIG. **22C** sets forth the results using breast cancer cells.

Example 23

[0777] In this example, increasing concentrations of different isovanillin derivatives or metabolites were tested using lung cancer cells (H358), ovarian cancer cells (A1847), and prostate cancer cells (22rv1), to determine the anticancer effects of the derivatives/metabolites, using the techniques of Example 1. Each of the derivatives/metabolites (vanillin, isovanillic acid, 0-vanillin, and isovanillyl alcohol) demonstrated anticancer effects (see FIGS. **23A-23L**).

Example 24

[0778] In this example, increasing concentrations of different harmine derivatives or metabolites were tested using lung cancer cells (H358), ovarian cancer cells (A1847), and prostate cancer cells (22rv1), to determine the anticancer effects of the derivatives/metabolites, using the techniques of Example 1. Each of the derivatives/metabolites (harmaline, tetrahydro-harmine, harmole hydrochloride, harmalol hydrochloride, and harmane) demonstrated anticancer effects (see FIGS. **24A-24O**).

Example 25

[0779] In this example, increasing concentrations of a curcumin derivative, bisdemethoxy curcumin, were tested using lung cancer cells (H358), ovarian cancer cells (A1847), and prostate cancer cells (22rv1), to determine the anticancer effects of the derivative, using the techniques of Example 1. The results confirmed the anticancer effects of this derivative (see FIGS. **25A-25C**).

Example 26

[0780] In this test, the preferred GZ17-6.02 product was stored at various temperatures over a two-month period, and then tested against ovarian cancer cells (A1847). The results are set forth in FIG. **26**, which illustrates that at storage temperatures of -20°C . and 4°C ., the product maintained its potency.

Example 27

[0781] In this test, the GZ17-6.02 product was subjected to successive freeze/thaw cycles. In each cycle, the product was frozen at -20°C ., followed by allowing the product temperature to equilibrate at room temperature. At the end of each cycle, the product was tested against ovarian cancer cells (A1847) using the methods of Example 1. A total of 10 successive freeze/thaw cycles were performed on the same sample. FIG. **27** illustrates that there was no significant change in the efficacy of the GZ17-6.02 against ovarian cancer cells.

Example 28

[0782] In this series of tests, GZ17-8.02 (which is identical with the preferred GZ17-6.02 composition), and several different combinations of curcumin, harmine, and isovanillin derivatives were tested against ovarian cancer (A1847), lung cancer (H358), prostate cancer (22rv1), pancreatic cancer, and fibroblast cells (hgf-1), in order to confirm that such derivative combinations were effective. In each case, the weight ratio of the isovanillin component:curcumin

component:harmin component was 7.7:1:1.3, and the compositions were prepared as described above and the tests were carried out as described in Examples 1, 2, and 6. The compositions are identified below, including the respective graphs giving the results of the tests: [0783] GZI7-8.02 (FIG. 28A) [0784] GZI7-8.03 contained vanillin, turmeric-derived curcumin, and harmine (FIG. 28B) [0785] GZI7-8.04 contained orthovanillin, turmeric-derived curcumin, and harmine, and (FIG. 28C) [0786] GZI7-8.05 contained isovanillyl alcohol, turmeric-derived curcumin, and harmine (FIG. 28D) [0787] GZI7-8.06 contained isovanillic acid, turmeric-derived curcumin, and harmine (FIG. 28E) [0788] GZI7-8.07 contained isovanillin, turmeric-derived curcumin, and harmaline (FIG. 28F) [0789] GZI7-8.08 contained isovanillin, turmeric-derived curcumin, and harmine (FIG. 28G) [0790] GZI7-8.09 contained isovanillin, 100% synthetic curcumin, and harmine (FIG. 28H) [0791] GZI7-8.10 contained isovanillin, bisdemethoxy curcumin, and harmine (FIG. 28I)

Examples 29-79

[0792] In each of the following examples, respective compositions having different combinations of isovanillin, curcumin, and harmine components were tested as set forth in Examples 1-6, against cancer cells to determine the anti-cancer effectiveness thereof. The specific cancer cell lines used in each case were those previously described. In all cases, the weight ratio of the isovanillin component:curcumin component:harmin component was 7.7:1:1.3, and the compositions were prepared as described above in connection with the preparation of GZ17-6.02. A separate graph is provided for each test, which identifies the cancer cells tested.

[0793] The following Table sets forth the Example numbers, composition designation, components, and corresponding Figure numbers for this series of tests. Where “curcumin” is specified, this refers to turmeric-derived curcumin, and where “curcumin (syn)” is specified, this refers to essentially pure synthetically-derived curcumin.

TABLE-US-00005	Example	Isovanillin	Curcumin	Harmine	FIG. No.	Composition	Component
	29	GZ17-08.11	isovanillin	bisdemethoxy harmaline	29	A-G	curcumin
	30	GZ17-08.12	isovanillin	bisdemethoxy harmine	30	A-G	curcumin
	31	GZ17-08.13	isovanillin	bisdemethoxy harmaline	31	A-G	curcumin
	32	GZ17-08.14	isovanillin	bisdemethoxy harmol	32	A-G	curcumin
	33	GZ17-08.15	vanillin	bisdemethoxy harmine	33	A-G	curcumin
	34	GZ17-08.16	vanillin	bisdemethoxy harmaline	34	A-G	curcumin
	35	GZ17-08.17	vanillin	bisdemethoxy harmine	35	A-G	curcumin
	36	GZ17-08.18	vanillin	bisdemethoxy harmaline	36	A-G	curcumin
	37	GZ17-08.19	vanillin	bisdemethoxy harmol	37	A-G	curcumin
	38	GZ17-08.20	isovanillin	curcumin (syn)	38	A-G	harmol
	39	GZ17-08.21	isovanillin	curcumin (syn)	39	A-G	harmol
	40	GZ17-08.22	vanillin	curcumin (syn)	40	A-G	harmaline
	41	GZ17-08.23	vanillin	curcumin (syn)	41	A-G	harmine
	42	GZ17-08.24	vanillin	curcumin (syn)	42	A-G	harmol
	43	GZ17-08.25	vanillin	curcumin (syn)	43	A-G	harmol
	44	GZ17-08.26	orthovanillin	bisdemethoxy harmine	44	A-G	curcumin
	45	GZ17-08.27	orthovanillin	bisdemethoxy harmaline	45	A-G	curcumin
	46	GZ17-08.28	orthovanillin	bisdemethoxy harmine	46	A-G	curcumin
	47	GZ17-08.29	orthovanillin	bisdemethoxy harmaline	47	A-G	curcumin
	48	GZ17-08.30	orthovanillin	bisdemethoxy harmol	48	A-G	curcumin
	49	GZ17-08.31	isovanillyl	bisdemethoxy harmine	49	A-D	alcohol curcumin
	50	GZ17-08.32	isovanillyl	bisdemethoxy harmaline	50	A-D	alcohol curcumin
	51	GZ17-08.33	isovanillyl	bisdemethoxy harmine	51	A-D	alcohol curcumin
	52	GZ17-08.34	isovanillyl	bisdemethoxy harmaline	52	A-D	alcohol curcumin
	53	GZ17-08.35	isovanillyl	bisdemethoxy harmol	53	A-D	alcohol curcumin
	54	GZ17-08.36	orthovanillin	curcumin (syn)	54	A-D	harmaline
	55	GZ17-08.37	orthovanillin	curcumin (syn)	55	A-D	harmine
	56	GZ17-08.38	orthovanillin	curcumin (syn)	56	A-D	harmol
	57	GZ17-08.39	orthovanillin	curcumin (syn)	57	A-D	harmol
	58	GZ17-08.40	isovanillyl	curcumin (syn)	58	A-D	alcohol
	59	GZ17-08.41	isovanillyl	curcumin (syn)	59	A-D	alcohol
	60	GZ17-08.42	isovanillyl	curcumin (syn)	60	A-D	alcohol
	61	GZ17-08.43	isovanillyl	curcumin (syn)	61	A-D	alcohol
	62	GZ17-08.44	isovanillic acid	bisdemethoxy harmine	62	A-D	curcumin
	63	GZ17-08.45	isovanillic acid	bisdemethoxy harmaline	63	A-D	

curcumin 64 GZ17-08.46 isovanillic acid bisdemethoxy harmane 64 A-D curcumin 65 GZ17-08.47 isovanillic acid bisdemethoxy harmalol 65 A-D curcumin 66 GZ17-08.48 isovanillic acid bisdemethoxy harmol 66 A-D curcumin 67 GZ17-08.49 isovanillic acid curcumin (syn) harmaline 67 A-D 68 GZ17-08.50 isovanillic acid curcumin (syn) harmane 68 A-D 69 GZ17-08.51 isovanillic acid curcumin (syn) harmalol 69 A-D 70 GZ17-08.52 isovanillic acid curcumin (syn) harmol 70 A-D 71 GZ17-08.53 5-bromovanillin curcumin harmine 71 A-D 72 GZ17-08.54 2-bromo-3-curcumin harmine 72 A-D hydroxy-4- methoxy benzaldehyde 73 GZ17-08.55 2-iodo-3- curcumin harmine 73 A-D hydroxy-4- methoxy benzaldehyde 74 GZ17-08.56 isovanillin (1E,4E)-1,5-bis(3,5- harmine 74 A-D dimethoxyphenyl)- 1,4-pentadien-3-one 75 GZ17-08.57 isovanillin cardamonin harmine 75 A-D 76 GZ17-08.58 isovanillin 2'-hydroxy-3,4,4',5'- harmine 76 A-D tetramethoxychalcone 77 GZ17-08.59 isovanillin 2,6'-2'-dihydroxy-4', harmine 77 A-D dimethoxychalcone 78 GZ17-08.60 isovanillin (1E,4E)-1,5-Bis(2- harmine 78 A-D Hydroxyphenyl)-1,4- pentadien-3-one 79 GZ17-08.61 isovanillin curcumin 1,2,3,4- 79 A-D tetrahy- droharmane- 3-carboxylic acid

[0794] These test results confirm that all of the compositions were effective anti-cancer agents.

Example 80

[0795] In this series of tests, the following compositions were prepared:

TABLE-US-00006 Isovanillin Curcumin Harmine Composition Component Component
 Component GZ17-10.00 orthovanillin curcumin harmaline GZ17-10.01 orthovanillin — harmaline
 GZ17-10.02 orthovanillin curcumin — GZ17-10.03 — curcumin harmaline GZ17-10.04
 orthovanillin — — GZ17-10.05 — curcumin — GZ17-10.06 — — harmaline

GZ17-10.00 is identical with the above-described GZ17-6.02, having the identical amounts of the components and method of preparation. The two-component compositions have the same relative ratios, namely 7.7:1.3 for GZ17-10.01, 7.7:1 for GZ17-10.02, and 1:1.3 for GZ17-10.03. The single-component compositions contain the same amount of component as used in the three- and two-component compositions.

[0796] The respective compositions of this Example were tested against different cancer cell lines, as previously identified, using the methods of Examples 1-6, as set forth in the accompanying relevant graphs, **80A-80R**. Six of these (FIGS. **80G**, H, L, M, Q, and R) are comparative bar graphs setting forth the theoretical additive effect of the components, and the results obtained using the complete formulations, thereby demonstrating the synergistic effects of the compositions of the invention.

[0797] These test results confirm that all of the compositions were effective anti-cancer agents.

Example 81

[0798] In this series of tests, the following compositions were prepared:

TABLE-US-00007 Isovanillin Component Harmine Component Composition (ratio component = 7.7) (ratio component = 1.3) GZ17-08.512 isovanillin harmine GZ17-08.513 isovanillin harmaline
 GZ17-08.514 isovanillin harmane GZ17-08.515 isovanillin harmalol GZ17-08.516 isovanillin harmol GZ17-08.517 vanillin harmine GZ17-08.518 vanillin harmaline GZ17-08.519 vanillin harmane GZ17-08.520 vanillin harmalol GZ17-08.521 vanillin harmol

[0799] The respective compositions of this Example were tested against different cancer cell lines, as previously identified, using the methods of Examples 1-6, as set forth in the accompanying relevant graphs, **81A-81DD**.

[0800] These test results confirm that all of the compositions were effective anti-cancer agents.

Example 82

[0801] In each of the following examples, respective compositions having different combinations of isovanillin, curcumin, and harmine components were tested as set forth in Examples 1-3, against lung cancer, lymphoma, and leukemia cells to determine the anti-cancer effectiveness thereof. The compositions were prepared by individually mixing the listed components in 3 mL of ethanol followed by adding the so-mixed components together to form the resultant compositions. A

separate graph is provided for each test, which identifies the cancer cells tested. Specifically, FIGS. **82-1** through **82-41** set forth the results of the lung cancer (LC) tests using the compositions, FIGS. **82-42** through **82-82** give the results for the lymphoma (LY) tests, and FIGS. **82-83** through **82-123** give the results of the leukemia (LK) tests.

[0802] The first Table below sets forth the identities of the isovanillin components i01-i15, the harmine components h01-h13, and the curcumin components c01-c13, used in these tests. The second Table below sets forth the multiple-component compositions tested, GZ08.065-GZ08.105, for lung cancer, lymphoma, and leukemia cells and the respective Figure numbers associated with each such composition and cell line. The recitation “curcumin (syn)” refers to essentially pure synthetically-derived curcumin.

[0803] All of the second Table compositions exhibited anti-cancer activity against lung cancer, lymphoma, and leukemia cells. In addition, FIGS. **82-124** through **82-167** are comparative bar graphs confirming the synergism found in exemplary compositions of this Example, at various concentration levels, against lung cancer, lymphoma, and leukemia cells.

TABLE-US-00008

Code	Compound	CAS#	MW	i01	o-anisaldehyde	135-02-4	136.15	i02	isovanillin oxime	51673-94-0	167.166	i03	ethyl vanillin	121-32-4	166.18	i04	vanillin isobutyrate	20665-85-4	222.24	i05	veratraldehyde	120-14-9	166.18	i06	5-nitrovanillin	6635-20-7	197.14	i07	vanillin acetate	881-68-5	194.18	i08	3-benzyloxy-4-methoxybenzaldehyde	6346-05-0	242.27	i09	3-hydroxy-5-methoxybenzaldehyde	672-13-9	152.15	i10	methyl isovanillate	6702-50-7	182.18	i11	acetovanillone (apocynin)	498-02-2	166.17	i12	2-hydroxy-4-methoxybenzaldehyde	673-22-3	152.15	i13	trans-ferulic acid	537-98-4	194.18	i14	3-hydroxy-4-methoxycinnamic acid	537-73-5	194.18	i15	caffeic acid	331-39-5	180.16	h01	norharmane	244-63-3	168.19	h02	6-methoxyharmalan	3589-73-9	214.26	h03	bromo harmine	na	372.06	h04	2-methyl harmine	21236-68-0	227.28	h05	4,9-dihydro-3H-beta-carbolin-1-yl methyl ether	na	200.24	h06	1-(4-nitrophenyl)-2,3,4,9-tetrahydro-1H-hydrochloride	3380-77-6	329.79	h07	1,2,3,4-tetrahydro-9H-pyrido[3,4-b]indole	16502-01-5	172.23	{THbC}	h08	1,2,3,4-tetrahydro-beta-carboline-1-carboxylic acid	na	216.24	h09	6-Methoxy-1,2,3,4-tetrahydro-9H-pyrido[3,4-b]indole	20315-68-8	202.25	h10	3-hydroxymethyl-b-carboline	65474-79-5	198.22	h11	2,3,4,5-tetrahydro-8-methoxy-1H-pyrido[4,3-b]indole	na	202.25	h12	6-Methoxy-1,2,3,4-tetrahydro-9H-pyrido[3,4-b]indole-1-carboxylic acid	17952-63-5	246.26	h13	ethyl b-carboline-3-carboxylate	74214-62-3	240.26	c01	tetrahydro curcumin	36062-04-1	372.41	c02	demethyl curcumin	149732-51-4	354.35	c03	demethoxy curcumin	22608-11-3	338.35	c04	1,3-diphenyl-2-propanone	102-04-5	210.27	c05	caffeic acid phenethyl ester	104594-70-9	284.31	c06	(1E,4E)-1,5-bis[3,5-bis(methoxymethoxy)phenyl]-1,4-pentadiene-3-one	917813-62-8	474.51	c07	1,7-di(1-naphthyl)-2,6-heptanedione	na	380.49	c08	trans,trans-1,5-Bis[4-(trifluoromethyl)phenyl]-1,4-pentadien-3-one	103836-71-1	370.29	c09	1,5-dibenzoylpentane	28861-22-5	280.36	c10	(2E,5E)-2,5-dibenzylidenecyclopentanone	895-80-7	260.33	c11	2,6-bis(4-fluorobenzal)cyclohexanone	62085-74-9	310.34	c12	(1E,4E)-1,5-bis(4-fluorophenyl)-1,4-pentadien-3-one	na	354.4	c13	FLLL31	52328-97-9	424.49
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TABLE-US-00009

Isovanillin	Harmine	Curcumin	Component	Component	Component	Total	LC
LY	LK	(176 mg/3 mL)	(30 mg/3 mL)	(23 mg/3 mL)	(mg/ Total Fig. Fig. Fig. Comp. EtOH)	EtOH)	EtOH)
08.065	i01	harmine	curcumin (syn)	76.3	498.8	82-1	82-42 82-83
08.066	i02	harmine	curcumin (syn)	76.3	418.8	82-2	82-43 82-84
08.067	i03	harmine	curcumin (syn)	76.3	331.9	82-4	82-45 82-86
08.069	i05	harmine	curcumin (syn)	76.3	420.9	82-5	82-46 82-87
08.070	i06	harmine	curcumin (syn)	76.3	365.5	82-6	82-47 82-88
08.071	i07	harmine	curcumin (syn)	76.3	370.0	82-7	82-48 82-89
08.072	i08	harmine	curcumin (syn)	76.3	310.1	82-8	82-49 82-90
08.073	i09	harmine	curcumin (syn)	76.3	389.9	82-10	82-51 82-92
08.075	i11	harmine	curcumin (syn)	76.3	421.0	82-11	82-52 82-93
08.076	i12	harmine	curcumin (syn)	76.3	453.5	82-12	82-53 82-94
08.077	i13	harmine	curcumin (syn)	76.3	370.0	82-13	

82-54 82-95 08.078 i14 harmine curcumin (syn) 76.3 370.0 82-14 82-55 82-96 08.079 i15 harmine curcumin (syn) 76.3 393.5 82-15 82-56 82-97 08.080 isovanillin h01 curcumin (syn) 76.3 465.9 82-16 82-57 82-98 08.081 isovanillin h02 curcumin (syn) 76.3 453.1 82-17 82-58 82-99 08.082 isovanillin h03 curcumin (syn) 76.3 433.3 82-18 82-59 82-100 08.083 isovanillin h04 curcumin (syn) 76.3 450.4 82-19 82-60 82-101 08.084 isovanillin h05 curcumin (syn) 76.3 456.3 82-20 82-61 82-102 08.085 isovanillin h06 curcumin (syn) 76.3 436.7 82-21 82-62 82-103 08.086 isovanillin h07 curcumin (syn) 76.3 464.5 82-22 82-63 82-104 08.087 isovanillin h08 curcumin (syn) 76.3 452.6 82-23 82-64 82-105 08.088 isovanillin h09 curcumin (syn) 76.3 455.8 82-24 82-65 82-106 08.089 isovanillin h10 curcumin (syn) 76.3 456.8 82-25 82-66 82-107 08.090 isovanillin h11 curcumin (syn) 76.3 455.8 82-26 82-67 82-108 08.091 isovanillin h12 curcumin (syn) 76.3 447.0 82-27 82-68 82-109 08.092 isovanillin h13 curcumin (syn) 76.3 448.0 82-28 82-69 82-110 08.093 isovanillin harmine c01 76.3 453.3 82-29 82-70 82-111 08.094 isovanillin harmine c02 76.3 454.3 82-30 82-71 82-112 08.095 isovanillin harmine c03 76.3 455.4 82-31 82-72 82-113 08.096 isovanillin harmine c04 76.3 469.2 82-32 82-73 82-114 08.097 isovanillin harmine c05 76.3 459.7 82-33 82-74 82-115 08.098 isovanillin harmine c06 76.3 448.9 82-34 82-75 82-116 08.099 isovanillin harmine c07 76.3 452.8 82-35 82-76 82-117 08.100 isovanillin harmine c08 76.3 453.4 82-36 82-77 82-118 08.101 isovanillin harmine c09 76.3 460.0 82-37 82-78 82-119 08.102 isovanillin harmine c10 76.3 462.1 82-38 82-79 82-120 08.103 isovanillin harmine c11 76.3 457.4 82-39 82-80 82-121 08.104 isovanillin harmine c12 76.3 454.3 82-40 82-81 82-122 08.105 isovanillin harmine c13 76.3 450.8 82-41 82-82 82-123

[0804] All of the foregoing tests confirmed significant anti-cancer activity against lung cancer cells, H358, lymphoma cells, M0205, and leukemia cells, jurkat E6-1. Accordingly, all combinations of i01-i15, h01-h13, and c01-c13, whether three-component or two-component, would exhibit such anti-cancer activity. Moreover, as shown above, the combinations of harmine and curcumin with i01-15, isovanillin and curcumin with h01-h13, and isovanillin and harmine with c01-c13 all have useful anti-cancer activity.

[0805] Moreover, as noted previously, the compounds i01-i15, h01-h13, and c01-c13 may be individually and independently in the form of the corresponding esters, metal complexes, pharmaceutically acceptable salts, and mixtures thereof. That is, e.g., a given curcumin component may be in the form of an ester, complex, or salt, independently of the form of the remaining components of the composition.

Example 83

[0806] In another series of tests, compositions were prepared containing: (1) orthovanillin+curcumin+harmaline; and (2) single-component compositions containing orthovanillin, curcumin, and harmaline, respectively. The three-component composition had a weight ratio of orthovanillin:curcumin:harmaline of 771:130.3:98.7, and the single-component compositions contained varying amounts of orthovanillin, curcumin, and harmaline.

[0807] These compositions were prepared by mixing together quantities of solid synthetic orthovanillin (99% by weight purity), synthetic harmaline (92% by weight purity), and a commercial turmeric product (ResCu) in ethanol, and allowing the mixtures to react for a period of 24 hours.

[0808] These compositions were tested against lymphoma (M0205), leukemia (jurkat E6-1), and breast cancer (du4475) cell lines, using the assays described in Examples 1-3.

[0809] The results of the lymphoma tests are set forth in FIGS. **83A-83D**. These tests confirmed that the combination of orthovanillin+curcumin+harmaline (FIG. **83D**) exhibited synergistic results as compared with the individual tests of orthovanillin, curcumin, and harmaline (FIGS. **83A-83C**).

[0810] In like manner, the leukemia tests (FIGS. **83E-83H**) confirmed that the combination of orthovanillin+curcumin+harmaline (FIG. **83H**) exhibited synergistic results as compared with the individual tests of orthovanillin, curcumin, and harmaline (FIGS. **83E-83G**).

[0811] To a similar effect, the breast cancer tests (FIGS. **83I-83L**) confirmed that the combination

of orthovanillin+curcumin+harmaline (FIG. 83L) exhibited synergistic results as compared with the individual tests of orthovanillin, curcumin, and harmaline (FIGS. 83I-83K).

Example 84

[0812] In this Example, GZ17-6.02 was tested with pancreatic cancer cells (52-007) to determine whether it had an effect on cancer stem cell proteins using the techniques described in Examples 1-3, and it was found that the treatment decreased the number and size of the pancreatic cancer spheres at both the IC_{sub}.50 concentration and a concentration of half that amount (see FIG. 84A). In additional tests (FIG. 84B), treated cells exhibited a dose-dependent decline in doublecortin calmodulin-like kinase 1 (Dclk-1), a microtubule-associated protein that serves as a marker for intestinal and pancreatic cancer stem cells (Mwangi, S. M. et al. "DCAMKL-1: a new horizon for pancreatic progenitor identification." *Am J Phys-Gastro Liver Physiol* 299 (2010):G301-302), and also differentiates between tumor stem cells and normal stem cells (Nakanishi, Y. et al. "Dclk1 distinguishes between tumor and normal stem cells in the intestine." *Nature Gen* 45 (2013):98-103). Epithelial cell adhesion molecule (EpCAM) is another cancer stem cell marker and it also decreased in level with GZ17-6.02 exposure. Likewise, the leucine-rich-repeat containing G-protein-coupled receptor 5 (LGR5) is another cancer stem cell marker and its levels decreased with the GZ17-6.02 treatment (FIG. 84B).

[0813] When tumors from treated mice were analyzed, the tumors also showed a clear reduction in EPCAM, DCLK1, LGR5, and SOX9 (FIG. 84C). GZ17-6.02 is thus seen to block pancreatic spheroid formation and cancer stem cell marker expression, both in cancer cells grown and treated in vitro and in tumor samples from mice treated with GZ17-6.02.

[0814] GZ17-6.02 significantly reduced tumorigenesis both with in vitro and in vivo pancreatic cancer cell models, partially via a route that inhibits the presence of cancer stem cells. In this regard, it appears that GZ17-6.02 has several different mechanisms of action resulting in decreased tumor growth and inhibition of metastases. FIG. 84D illustrates a possible pathway for the action of GZ17-6.02 on both cancer stem cells (CSC) and on the other cancer cells within the tumor. With respect to the cancer stem cells, the GZ17-6.02 decreased the common biomarkers of cancer stem cells (Dclk1, LGR5, EpCam, and Sox9). It apparently did this by acting through the Sonic Hedgehog pathway (FIG. 84D, SHH). Simultaneously, GZ17-6.02 blocked the bulk tumor cells in two ways by directly inducing apoptosis via the caspase pathway, as measured by the BAX/Bcl2 ratio, and through the EGFR pathway. All three of these mechanisms led to reduced tumor size, inhibition of metastasis and decline in the presence of cancer stem cells. Thus, the compositions of the invention are effective for killing or inhibiting the growth of cancer stem cells.

Example 85

[0815] The purpose of this study was to determine the effect of GZ17-6.02 on the regression of growth and the inhibition of metastases in a pancreatic cancer mouse model. Twelve immunocompromised (nude) mice were treated injected with MiaPaCa-2 cells (a pancreatic cancer cell line) that had been genetically transformed to express TdTomato and luciferin for in vivo tumor imaging. After tumors reached a measurable size (approximately 2 weeks), half of the mice were randomly assigned to the experimental group and the other half to the control group.

[0816] The experimental group was treated with GZ17-6.02 at a daily dose of 100 mg/kg body weight. The drug administration was oral, dissolved in peptamen. Control mice were fed the vehicle (peptamen) alone. Once a week mice were lightly anesthetized in order to use IVIS imaging to detect the tumor sizes. On day 5, there was no statistical difference in the amount of fluorescence measured from the control (placebo-fed) and GZ17-6.02-fed mice. However, by day 20, there was a statistically significant decrease in the amount of fluorescence, indicating a decreased total tumor burden, in the mice fed GZ17-6.02, as compared with the control. In addition, the placebo-treated mice had dramatic levels of peritoneal ascites that were not present in the GZ17-6.02 group.

[0817] At the end of 21 days of treatment with GZ17-6.02 or the placebo, the mice were sacrificed and tumor samples removed from the pancreas, liver, and lungs. The primary pancreatic tumors

were weighed. The placebo-fed animals had pancreatic tumors that averaged 2.4 times statistically larger than the GZ17-6.02 fed mice.

Example 86

[0818] A 63-year-old male patient having a Prostate-Specific Antigen (PSA) count of 8.2 began a treatment regimen using a dosage form of GZ17-6.02. Specifically, solid GZ17-6.02 prepared by evaporating the ethanol from the previously described GZ17-6.02 agent was dispersed in water in equal weight amounts, e.g., 5 grams solid GZ17-6.02 in 5 grams water. This dosage form was taken three times daily for six weeks, with each dose being four fluid ounces of the 50%:50% dispersion. At the end of six weeks, the patient's PSA count had dropped to 2.1, and all prostate and urological tests were normal. The treating physician had no explanation for the decline in PSA. The patient experienced no observable adverse reactions to the treatment with GZ17-6.02.

[0819] Although not wishing to be bound by any theory of operation, the inventors believe that the therapeutic agents of the invention ameliorate a number of conditions or illnesses, and especially reduce and/or eliminate cancer and/or the symptoms thereof by augmenting or stimulating the patients' immune systems. In this sense, the invention is believed to be a form of biological therapy. As such, it is considered that the invention is applicable to virtually all cancers, such as the following: Acute Lymphoblastic Leukemia, Adult; Acute Lymphoblastic Leukemia, Childhood; Acute Myeloid Leukemia, Adult; Acute Myeloid Leukemia, Childhood; Adrenocortical Carcinoma; Adrenocortical Carcinoma, Childhood; Adolescents, Cancer in; AIDS-Related Cancers; AIDS-Related Lymphoma; Anal Cancer; Appendix Cancer; Astrocytomas, Childhood; Atypical Teratoid/Rhabdoid Tumor, Childhood, Central Nervous System; Basal Cell Carcinoma; Bile Duct Cancer, Extrahepatic; Bladder Cancer; Bladder Cancer, Childhood; Bone Cancer, Osteosarcoma and Malignant Fibrous Histiocytoma; Brain Stem Glioma, Childhood; Brain Tumor, Adult; Brain Tumor, Brain Stem Glioma, Childhood; Brain Tumor, Central Nervous System Atypical Teratoid/Rhabdoid Tumor, Childhood; Brain Tumor, Central Nervous System Embryonal Tumors, Childhood; Brain Tumor, Astrocytomas, Childhood; Brain Tumor, Craniopharyngioma, Childhood; Brain Tumor, Ependymoblastoma, Childhood; Brain Tumor, Ependymoma, Childhood; Brain Tumor, Medulloblastoma, Childhood; Brain Tumor, Medulloepithelioma, Childhood; Brain Tumor, Pineal Parenchymal Tumors of Intermediate Differentiation, Childhood; Brain Tumor, Supratentorial Primitive Neuroectodermal Tumors and Pineoblastoma, Childhood; Brain and Spinal Cord Tumors, Childhood (Other); Breast Cancer; Breast Cancer and Pregnancy; Breast Cancer, Childhood; Breast Cancer, Male; Bronchial Tumors, Childhood; Burkitt Lymphoma; Carcinoid Tumor, Childhood; Carcinoid Tumor, Gastrointestinal; Carcinoma of Unknown Primary; Central Nervous System Atypical Teratoid/Rhabdoid Tumor, Childhood; Central Nervous System Embryonal Tumors, Childhood; Central Nervous System (CNS) Lymphoma, Primary; Cervical Cancer; Cervical Cancer, Childhood; Childhood Cancers; Chordoma, Childhood; Chronic Lymphocytic Leukemia; Chronic Myelogenous Leukemia; Chronic Myeloproliferative Disorders; Colon Cancer; Colorectal Cancer, Childhood; Craniopharyngioma, Childhood; Cutaneous T-Cell Lymphoma; Embryonal Tumors, Central Nervous System, Childhood; Endometrial Cancer; Ependymoblastoma, Childhood; Ependymoma, Childhood; Esophageal Cancer; Esophageal Cancer, Childhood; Esthesioneuroblastoma, Childhood; Ewing Sarcoma Family of Tumors; Extracranial Germ Cell Tumor, Childhood; Extragonadal Germ Cell Tumor; Extrahepatic Bile Duct Cancer; Eye Cancer, Intraocular Melanoma; Eye Cancer, Retinoblastoma; Gallbladder Cancer; Gastric (Stomach) Cancer; Gastric (Stomach) Cancer, Childhood; Gastrointestinal Carcinoid Tumor; Gastrointestinal Stromal Tumor (GIST); Gastrointestinal Stromal Cell Tumor, Childhood; Germ Cell Tumor, Extracranial, Childhood; Germ Cell Tumor, Extragonadal; Germ Cell Tumor, Ovarian; Gestational Trophoblastic Tumor; Glioma, Adult; Glioma, Childhood Brain Stem; Hairy Cell Leukemia; Head and Neck Cancer; Heart Cancer, Childhood; Hepatocellular (Liver) Cancer, Adult (Primary); Hepatocellular (Liver) Cancer, Childhood (Primary); Histiocytosis, Langerhans Cell; Hodgkin Lymphoma, Adult; Hodgkin Lymphoma, Childhood; Hypopharyngeal Cancer;

Intraocular Melanoma; Islet Cell Tumors (Endocrine Pancreas); Kaposi Sarcoma; Kidney (Renal Cell) Cancer; Kidney Cancer, Childhood; Langerhans Cell Histiocytosis; Laryngeal Cancer; Laryngeal Cancer, Childhood; Leukemia, Acute Lymphoblastic, Adult; Leukemia, Acute Lymphoblastic, Childhood; Leukemia, Acute Myeloid, Adult; Leukemia, Acute Myeloid, Childhood; Leukemia, Chronic Lymphocytic; Leukemia, Chronic Myelogenous; Leukemia, Hairy Cell; Lip and Oral Cavity Cancer; Liver Cancer, Adult (Primary); Liver Cancer, Childhood (Primary); Lung Cancer, Non-Small Cell; Lung Cancer, Small Cell; Lymphoma, AIDS-Related; Lymphoma, Burkitt; Lymphoma, Cutaneous T-Cell; Lymphoma, Hodgkin, Adult; Lymphoma, Hodgkin, Childhood; Lymphoma, Non-Hodgkin, Adult; Lymphoma, Non-Hodgkin, Childhood; Lymphoma, Primary Central Nervous System (CNS); Macroglobulinemia, Waldenstrom; Malignant Fibrous Histiocytoma of Bone and Osteosarcoma; Medulloblastoma, Childhood; Medulloepithelioma, Childhood; Melanoma; Melanoma, Intraocular (Eye); Merkel Cell Carcinoma; Mesothelioma, Adult Malignant; Mesothelioma, Childhood; Metastatic Squamous Neck Cancer with Occult Primary; Mouth Cancer; Multiple Endocrine Neoplasia Syndromes, Childhood; Multiple Myeloma/Plasma Cell Neoplasm; Mycosis Fungoides; Myelodysplastic Syndromes; Myelodysplastic/Myeloproliferative Neoplasms; Myelogenous Leukemia, Chronic; Myeloid Leukemia, Adult Acute; Myeloid Leukemia, Childhood Acute; Myeloma, Multiple; Myeloproliferative Disorders, Chronic; Nasal Cavity and Paranasal Sinus Cancer; Nasopharyngeal Cancer; Nasopharyngeal Cancer, Childhood; Neuroblastoma; Non-Hodgkin Lymphoma, Adult; Non-Hodgkin Lymphoma, Childhood; Non-Small Cell Lung Cancer; Oral Cancer, Childhood; Oral Cavity Cancer, Lip and; Oropharyngeal Cancer; Osteosarcoma and Malignant Fibrous Histiocytoma of Bone; Ovarian Cancer, Childhood; Ovarian Epithelial Cancer; Ovarian Germ Cell Tumor; Ovarian Low Malignant Potential Tumor; Pancreatic Cancer; Pancreatic Cancer, Childhood; Pancreatic Cancer, Islet Cell Tumors; Papillomatosis, Childhood; Paranasal Sinus and Nasal Cavity Cancer; Parathyroid Cancer; Penile Cancer; Pharyngeal Cancer; Pineal Parenchymal Tumors of Intermediate Differentiation, Childhood; Pineoblastoma and Supratentorial Primitive Neuroectodermal Tumors, Childhood; Pituitary Tumor; Plasma Cell Neoplasm/Multiple Myeloma; Pleuropulmonary Blastoma, Childhood; Pregnancy and Breast Cancer; Primary Central Nervous System (CNS) Lymphoma; Prostate Cancer; Rectal Cancer; Renal Cell (Kidney) Cancer; Renal Pelvis and Ureter, Transitional Cell Cancer; Respiratory Tract Cancer with Chromosome 15 Changes; Retinoblastoma; Rhabdomyosarcoma, Childhood; Salivary Gland Cancer; Salivary Gland Cancer, Childhood; Sarcoma, Ewing Sarcoma Family of Tumors; Sarcoma, Kaposi; Sarcoma, Soft Tissue, Adult; Sarcoma, Soft Tissue, Childhood; Sarcoma, Uterine; Sezary Syndrome; Skin Cancer (Nonmelanoma); Skin Cancer, Childhood; Skin Cancer (Melanoma); Skin Carcinoma, Merkel Cell; Small Cell Lung Cancer; Small Intestine Cancer; Soft Tissue Sarcoma, Adult; Soft Tissue Sarcoma, Childhood; Squamous Cell Carcinoma; Squamous Neck Cancer with Occult Primary, Metastatic; Stomach (Gastric) Cancer; Stomach (Gastric) Cancer, Childhood; Supratentorial Primitive Neuroectodermal Tumors, Childhood; T-Cell Lymphoma, Cutaneous; Testicular Cancer; Testicular Cancer, Childhood; Throat Cancer; Thymoma and Thymic Carcinoma; Thymoma and Thymic Carcinoma, Childhood; Thyroid Cancer; Thyroid Cancer, Childhood; Transitional Cell Cancer of the Renal Pelvis and Ureter; Trophoblastic Tumor, Gestational; Unknown Primary Site, Carcinoma of, Adult; Unknown Primary Site, Cancer of, Childhood; Unusual Cancers of Childhood; Ureter and Renal Pelvis, Transitional Cell Cancer; Urethral Cancer; Uterine Cancer, Endometrial; Uterine Sarcoma; Vaginal Cancer; Vaginal Cancer, Childhood; Vulvar Cancer; Waldenstrom Macroglobulinemia; Wilms Tumor; Women's Cancers.

Claims

1. A method of treating a patient suffering from a pancreatic cancer or tumor, comprising the step of administering to the patient an anticancer composition comprising a combination of

- synergistically effective amounts of at least one curcumin component comprising curcumin, at least one harmine component comprising harmine, and at least one isovanillin component comprising isovanillin, wherein the components of the composition are different.
- 2.** The method of claim 1, wherein said curcumin is synthetically derived or derived from naturally occurring product(s) containing said curcumin which have been treated so that the curcumin is present at a level of at least about 25% by weight in the treated, curcumin-containing naturally occurring product(s), said harmine is synthetically derived or derived from naturally occurring product(s) containing said harmine which have been treated so that the harmine is present at a level of at least about 25% by weight in the treated, harmine-containing naturally occurring product(s), and said isovanillin is synthetically derived or derived from naturally occurring product(s) containing said isovanillin which have been treated so that the isovanillin is present at a level of at least about 25% by weight in the treated, isovanillin-containing naturally occurring product(s).
- 3.** The method of claim 1, wherein said curcumin, said harmine, and said isovanillin are each at least about 50% pure.
- 4.** The method of claim 1, wherein said isovanillin is present in an amount greater than the amounts of the harmine and the curcumin.
- 5.** The method of claim 1, wherein the ratio of said at least one curcumin component, said at least one harmine component, and said at least one isovanillin component is from about 0.1-5:0.1-5:0.1-25.
- 6.** The method of claim 1, wherein the at least one isovanillin component is present at a level of from about 25-85% by weight, the at least one harmine component is present at a level of from about 7-50% by weight, and the at least one curcumin component is present at a level of from about 5-40% by weight, based upon the total weight of the three components taken as 100% by weight.
- 7.** The method of claim 1, wherein said at least one curcumin component consists essentially of curcumin, said at least one harmine component consists essentially of harmine, and said at least one isovanillin component consists essentially of isovanillin.
- 8.** The method of claim 1, wherein said pancreatic cancer is selected from the group consisting of exocrine pancreatic cancer or islet cell tumor.
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