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(54) **CLEANING COMPOSITIONS AND USES
THEREOF**

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Related U.S. Application Data

ABSTRACT

(62) Division of application No. 18/342,030, filed on Jun.
27, 2023, now Pat. No. 12,319,895, which is a division
of application No. 17/188,017, filed on Mar. 1,
2021, now Pat. No. 11,739,287, which is a division of
application No. 16/500,424, filed on Oct. 3, 2019,

The present invention relates to compositions such as cleaning
compositions comprising a mix of enzymes. The invention
further relates, use of compositions comprising such
enzymes in cleaning processes.

Specification includes a Sequence Listing.

CLEANING COMPOSITIONS AND USES THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a division of U.S. application Ser. No. 18/342,030 filed on Jun. 27, 2023, now pending, which is a division of U.S. application Ser. No. 17/188,017 filed Mar. 1, 2021, now U.S. Pat. No. 11,739,287, which is a division of U.S. application Ser. No. 16/500,424 filed Oct. 3, 2019, now U.S. Pat. No. 10,968,416, which is a 35 U.S.C. 371 national application of international application no. PCT/EP2018/056730 filed Mar. 16, 2018, which claims priority under 35 U.S.C. 119 of European application no. 17165343.9 filed Apr. 6, 2017. The disclosure of each application is fully incorporated herein by reference.

REFERENCE TO A SEQUENCE LISTING

[0002] This application contains a Sequence Listing in computer readable form, which is hereby incorporated by reference in its entirety. The electronic sequence listing was created on May 14, 2025, is named SQ.xml, and is 96,964 bytes in size.

BACKGROUND OF THE INVENTION

[0003] The present invention relates to compositions such as cleaning compositions comprising a mix of enzymes. The invention further relates, use of compositions comprising such enzymes in cleaning processes and/or for deep cleaning of biofilm soiling, methods for removal or reduction of biofilm related soiling.

DESCRIPTION OF THE RELATED ART

[0004] Enzymes have been used in detergents for decades. Usually a cocktail of various enzymes is added to detergent compositions. The enzyme cocktail often comprises various enzymes, wherein each enzyme targets its specific substrate, e.g., amylases are active towards starch stains, proteases on protein stains and so forth. Textiles surface and hard surfaces, such as dishes or the inner space of a laundry machine enduring a number of wash cycles, become soiled with many different types of soiling which may compose of proteins, grease, starch etc. One type of soiling may be organic matter, such as biofilm, EPS, etc. Organic matter composes different molecules such as polysaccharides, extracellular DNA (eDNA), and proteins. Some organic matter composes an extracellular polymeric matrix, which may be sticky or glueing, which when present on textile, attracts soils and may cause redeposition or backstaining of soil resulting in a greying of the textile. Additionally, organic matters such as biofilms often cause malodor issue as various malodor molecules can be adhered by the polysaccharides, extracellular DNA (eDNA), and proteins in the complex extracellular matrix and be slowly released out to cause consumer noticeable malodor issue. There is still a need for cleaning compositions, which effectively prevent, reduce or remove components of organic stains, e.g., biofilm, an effect described in the present application as "deep cleaning". The present invention provides new compositions fulfilling such need.

SUMMARY OF THE INVENTION

[0005] A first aspect of the present invention, relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase. Another aspect of the invention relates to a cleaning composition comprising at least 0.001 ppm DNase and at least 0.001 ppm carbohydrase and a cleaning component, wherein the cleaning component is selected from

[0006] a. 0.1 to 15 wt. %, e.g., from about 1% to about 40% of at least one a surfactant;

[0007] b. 0.5 to 20 wt. %, e.g., from about 5% to about 50% of at least one builder; and

[0008] c. 0.01 to 10 wt. %, e.g., from about 1% to about 20% of at least one bleach component.

[0009] The invention further relates to the use of a composition for deep cleaning of an item, wherein the item is a textile or a surface. The invention further relates to the use of a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase. The invention further relates to a method of formulating a cleaning composition comprising adding a DNase, a carbohydrase and at least one cleaning component. The invention further relates to a kit intended for deep cleaning, wherein the kit comprises a solution of an enzyme mixture comprising a DNase and a carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase. The invention further relates to a method of deep cleaning an item, comprising the steps of: a) contacting the item with a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase; and b) optionally rinsing the item, wherein the item is preferably a textile. The invention further relates to a method of deep cleaning of an item, comprising the steps of: a) contacting the item with a solution comprising an enzyme mixture comprising a DNase and a carbohydrase and optionally a protease; and a cleaning component, wherein the cleaning component is selected from 0.1 to 15 wt. %, e.g., from about 1% to about 40% of at least one a surfactant; 0.5 to 20 wt. %, e.g., from about 5% to about 50% of at least one builder; and 0.01 to 10 wt. %, e.g., from about 1% to about 20% of at least one bleach component; and b) optionally rinsing the item, wherein the item is preferably a textile.

DETAILED DESCRIPTION OF THE INVENTION

[0010] Various enzymes are applied in cleaning processes each targeting specific types of soiling such as protein, starch and grease soiling. Enzymes are now standard ingredients in detergents for laundry and dish wash. The effectiveness of these commercial enzymes provides detergents which removes much of the soiling. However, organic matters such as EPS (extracellular polymeric substance) comprised in much biofilm constitute a challenging type of staining due to the complex nature of such organic matters. None of the commercially available cleaning compositions effectively remove or reduce EPS and/or biofilm related stains. Biofilm may be produced when a group of microorganisms' cells stick to each other or stick to a surface, such as a textile, dishware or hard surface or another kind of

surface. These adherent cells are frequently embedded within a self-produced matrix of extracellular polymeric substance (EPS), which constitute 50% to 90% of the biofilm's total organic matter. EPS is mostly composed of polysaccharides (exopolysaccharides) and proteins, but include other macro-molecules such as eDNA, lipids and other organic substances. Organic matter like biofilm may be sticky or glueing, which when present on textile, may give rise to redeposition or backstaining of soil resulting in a greying of the textile. Another drawback of organic matter, e.g., biofilm is the malodor as various malodor related molecules are often associated with organic matter, e.g., biofilm. Further, when dirty laundry items are washed together with less dirty laundry items the dirt present in the wash liquor tend to stick to organic matter, e.g., biofilm or biofilm components as a result, hereof the laundry item is more "soiled" after wash than before wash. This is effect may also be termed re-deposition.

[0011] The compositions of the invention comprise a blend of DNase and a carbohydراse, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase and effectively reduce or remove organic components, such as protein and DNA from surfaces such as textiles and hard surfaces, e.g., dishes.

[0012] The compositions of the invention comprise a blend of DNase and a carbohydراse, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase and effectively reduce or limit redeposition when applied in, e.g., laundry process.

[0013] The compositions of the invention comprise a blend of DNase and a carbohydراse, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase and effectively reduce or limit malodor of, e.g., textiles or hard surfaces such as dishes.

[0014] The compositions of the invention comprise a blend of DNase and a carbohydراse, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase and improve whiteness of textile.

[0015] A composition of the invention is preferably a cleaning composition, the composition of the invention comprises at least one DNase and at least one a carbohydراse, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase. Examples of useful DNases and carbohydراses are mentioned below in the sections "Polypeptides having DNase activity" and "Polypeptides having mannanase, cellulase, xylanase or amylase activity" respectively.

Polypeptides Having DNase Activity

[0016] The term "DNase" means a polypeptide with DNase (deoxyribonuclease) activity that catalyzes the hydrolytic cleavage of phosphodiester linkages in a DNA backbone, thus degrading DNA. Exodeoxyribonuclease cut or cleaves residues at the end of the DNA back bone where endo-deoxyribonucleases cleaves or cut within the DNA backbone. A DNase may cleave only double-stranded DNA or may cleave double stranded and single stranded DNA. The term "DNases" and the expression "a polypeptide with DNase activity" are used interchangeably throughout the application. For purposes of the present invention, DNase activity is determined according to the procedure described in the Assay I.

[0017] Preferably, the DNase is selected from any of the enzyme classes E.C.3.1, preferably E.C.3.1.21, e.g., such as

E.C.3.1.21.X, where X=1, 2, 3, 4, 5, 6, 7, 8 or 9, or, e.g., Deoxyribonuclease I, Deoxyribonuclease IV, Type I site-specific deoxyribonuclease, Type II site-specific deoxyribonuclease, Type III site-specific deoxyribonuclease, CC-pre-ferring endo-deoxyribonuclease, Deoxyribonuclease V, T(4) deoxyribonuclease II, T(4) deoxyribonuclease IV or E.C.3.1.22.Y where Y=1, 2, 4 or 5, e.g., Deoxyribonuclease II, *Aspergillus* deoxyribonuclease K(1), Crossover junction endo-deoxyribonuclease, Deoxyribonuclease X.

[0018] Preferably, the polypeptide having DNase activity is obtained from a microorganism and the DNase is a microbial enzyme. The DNase is preferably of fungal or bacterial origin.

[0019] The DNase may be obtainable from *Bacillus* e.g., *Bacillus licheniformis*, *Bacillus subtilis*, *Bacillus* sp-62451, *Bacillus horikoshii*, *Bacillus* sp-62451, *Bacillus* sp-16840, *Bacillus* sp-62668, *Bacillus* sp-13395, *Bacillus horneckiae*, *Bacillus* sp-11238, *Bacillus cibi*, *Bacillus idriensis*, *Bacillus* sp-62520, *Bacillus* sp-16840, *Bacillus* sp-62668, *Bacillus algicola*, *Bacillus vietnamensis*, *Bacillus hwajinpoensis*, *Bacillus indicus*, *Bacillus marisflavi*, *Bacillus luciferensis*, *Bacillus* sp. SA2-6.

[0020] The DNase may also be obtained from any of the following *Pyrenophaetopsis* sp., *Vibrissa flavovirens*, *Setosphaeria rostrata*, *Endophragmiella valdina*, *Corynespora cassiicola*, *Paraphoma* sp. XZ1965, *Monilinia fructicola*, *Curvularia lunata*, *Penicillium reticulansporum*, *Penicillium quercetorum*, *Setophaeosphaeria* sp., *Alternaria*, *Alternaria* sp. XZ2545, *Trichoderma reesei*, *Chaetomium thermophilum*, *Scytalidium thermophilum*, *Metapochonia suchlasporia*, *Daldinia fissa*, *Acremonium* sp. XZ2007, *Acremonium* sp. XZ2414, *Acremonium dichromosporum*, *Sarocladium* sp. XZ2014, *Metarhizium* sp. HNA15-2, *Isaria tenuipes*, *Scytalidium circinatum*, *Metarhizium lepidiotae*, *Thermobispora bispora*, *Sporormia fimetaria*, *Pycnidioiphora* cf. *dispera*, Environmental sample D, Environmental sample O, *Clavicipitaceae* sp-70249, *Westerdykella* sp. AS85-2, *Humiocolopsis cephalosporioides*, *Neosartorya massa*, *Roussocella intermedia*, *Pleosporales*, *Phaeosphaeria* or *Didymosphaeria futilis*.

[0021] The DNases to be used in a composition of the invention preferable belong to the NUC1 group of DNases. The NUC1 group of DNases comprises polypeptides which in addition to having DNase activity, may comprise one or more of the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], or C[D/N]T[A/R]. One embodiment of the invention relates to a composition comprising a carbohydراse, selected from cellulases, amylases, mannanases and xylanases, and polypeptides having DNase activity, wherein the polypeptides comprises one or more of the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V] or C[D/N]T[A/R].

[0022] The DNases preferably comprises a NUC1_A domain [D/Q][I/V]DH. In addition to comprising any of the domain motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V] or C[D/N]T[A/R] the polypeptides having DNase activity, to be used in a composition of the invention, may comprise the NUC1_A domain and may share the common motif [D/Q][I/V]DH. One embodiment the invention relates to compositions comprising a carbohydراse, selected from cellulases, amylases, mannanases and xylanases, and polypeptides, which comprises one or more motifs selected from

the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R] and [D/Q][I/V]DH, wherein the polypeptides have DNase activity.

[0023] The DNases to be added to a composition of the invention preferably belong to the group of DNases comprised in the GYS-clade, which are group of DNases on the same branch of a phylogenetic tree having both structural and functional similarities. These NUC1 and/or NUC1_A DNases comprise the conservative motifs [D/M/L][S/T] GYSR[D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74) and share similar structural and functional properties. The DNases of the GYS-clade are preferably obtained from *Bacillus* genus.

[0024] One embodiment of the invention relates to a composition comprising a carbohydrate, selected from cellulases, amylases, mannanases and xylanases, a polypeptide of the GYS clade having DNase activity, optionally wherein the polypeptide comprise one or both of the motifs [D/M/L][S/T] GYSR[D/N](SEQ ID NO: 73), ASXNRSKG (SEQ ID NO: 74) and wherein the polypeptide is selected from the group of polypeptides:

[0025] a) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 1,

[0026] b) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 2,

[0027] c) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 3,

[0028] d) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 4,

[0029] e) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 5,

[0030] f) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 6,

[0031] g) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 7,

[0032] h) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 8,

[0033] i) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least

85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 9,

[0034] j) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 10,

[0035] k) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 11,

[0036] l) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 12,

[0037] m) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 13,

[0038] n) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 14,

[0039] o) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 15,

[0040] p) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 16,

[0041] q) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 17,

[0042] r) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 18,

[0043] s) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 19,

[0044] t) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 20,

[0045] u) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 21,

[0046] v) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 22,

[0047] w) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 23,

[0048] x) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 24, and

[0049] y) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 25.

[0050] Polypeptides having DNase activity and which comprise the GYS-clade motifs have shown particularly good deep cleaning properties, e.g., the DNases are particularly effective in removing or reducing DNA stains, e.g., associated with biofilm or dead cell debris, from an item such as a textile or a hard surface. In addition, these DNases are particularly effective in removing or reducing malodor, from an item such as a textile or a hard surface. Further, the GYS-clade DNases are particularly effective in preventing redeposition when laundering an item such as textile.

[0051] In one embodiment the DNases to be added in a composition of the invention preferably belong to the group of DNases comprised in the NAWK-clade, which are NUC1 and NUC1_A DNases, which may further comprise the conservative motifs [V/I]PL[S/A]NAWK (SEQ ID NO: 75) or NPQL (SEQ ID NO: 76).

[0052] One embodiment of the invention relates to a composition comprising a carbohydase selected from a cellulase, an amylase, a mannanase or a xylanase, and a polypeptide of the NAWK-clade having DNase activity, optionally wherein the polypeptide comprise one or both of the motifs [V/I]PL[S/A]NAWK (SEQ ID NO: 75) or NPQL (SEQ ID NO: 76) and wherein the polypeptide is selected from the group of polypeptides:

[0053] a) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 26,

[0054] b) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 27,

[0055] c) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 28,

[0056] d) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 29,

[0057] e) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 30,

[0058] f) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 31,

[0059] g) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 32,

[0060] h) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 33,

[0061] i) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 34,

[0062] j) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 35,

[0063] k) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 36,

[0064] l) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 37, and

[0065] m) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 38.

[0066] Polypeptides having DNase activity and which comprise the NAWK-clade motifs have shown particularly good deep cleaning properties, e.g., the DNases are particularly effective in removing or reducing DNA stains, e.g., associated with biofilm or dead cell debris, from an item such as a textile or a hard surface. In addition, these DNases are particularly effective in removing or reducing malodor, from an item such as a textile or a hard surface. Further, the NAWK-clade DNases are particularly effective in preventing redeposition when laundering an item such as textile.

[0067] The DNases to be added in a composition of the invention preferably belong to the group of DNases comprised in the KNAW-clade, which are NUC1 and NUC1_A DNases which may further comprise the conservative motifs P[Q/E]L[W/Y] or [K/H/E]NAW.

[0068] One embodiment of the invention relates to a composition comprising a carbohydase, selected from a cellulase, an amylase, a mannanase or a xylanase, and a polypeptide of the KNAW clade having DNase activity,

optionally wherein the polypeptide comprise one or both of the motifs P[Q/E]L[W/Y] or [K/H/E]NAW, and wherein the polypeptide is selected from the group of polypeptides:

[0069] a) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 39,

[0070] b) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 40,

[0071] c) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 41,

[0072] d) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 42,

[0073] e) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 43

[0074] f) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 44,

[0075] g) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 45,

[0076] h) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 46,

[0077] i) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 47,

[0078] j) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 48,

[0079] k) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 49,

[0080] l) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 50, and

[0081] m) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75% at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or at least 100% sequence identity to the polypeptide shown in SEQ ID NO: 51.

[0082] Polypeptides having DNase activity and which comprise the KNAW-clade motifs have shown particularly good deep cleaning properties, e.g., the DNases are particularly effective in removing or reducing DNA stains, e.g., associated with biofilm or dead cell debris, from an item such as a textile or a hard surface. In addition, these DNases are particularly effective in removing or reducing malodor, from an item such as a textile or a hard surface. Further, the KNAW-clade DNases are particularly effective in preventing redeposition when laundering an item such as textile.

[0083] In some embodiments, the present invention relates to compositions comprising a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and a polypeptide obtainable from *Bacillus*, e.g., obtainable from *Bacillus* sp-62451 and having a sequence identity to the polypeptide shown in SEQ ID NO: 1 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide shown in SEQ ID NO: 1.

[0084] In some embodiments, the present invention relates to compositions comprising a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and a polypeptide obtainable from *Bacillus*, e.g., obtainable from *Bacillus horikoshii* and having a sequence identity to the polypeptide shown in SEQ ID NO: 2 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide shown in SEQ ID NO: 2.

[0085] In some embodiments, the present invention relates to compositions comprising a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and a polypeptide obtainable from *Bacillus*, e.g., obtainable from *Bacillus* sp-62520 and having a sequence identity to the polypeptide shown in SEQ ID NO: 3 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide shown in SEQ ID NO: 3.

[0086] In some embodiments, the present invention relates to compositions comprising a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and a polypeptide obtainable from *Bacillus*, e.g., obtainable from

at least 98%, at least 99%, or 100% and which have DNase activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide shown in SEQ ID NO: 67.

[0150] In some embodiments, the present invention relates to compositions comprising a carbohydراse, selected from cellulases, amylases, mannanases and xylanases, and a polypeptide obtainable from *Trichoderma*, e.g., obtainable from *Trichoderma harzianum* having a sequence identity to the polypeptide shown in SEQ ID NO: 68 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide shown in SEQ ID NO: 68.

[0151] The DNases above may be combined with any of the carbohydrases below to form a blend to be added to a composition according to the invention.

[0152] Polypeptides having Carbohydراse activity (Carbohydrases) Carbohydراse is a protein/enzyme that catalyse carbohydrates to break down carbohydrates to, e.g., simple sugar such as monosaccharides. Thus, carbohydrases are any of a group of enzymes that promote hydrolysis of a carbohydrate. Starch hydrolyzing carbohydrases (e.g., amylases) work on, e.g., amylose and amylopectin and non-starch carbohydrases includes enzymes which hydrolyze polymers made up of carbon sugars, e.g., cellulases which will ultimately produce glucose when complete hydrolysis is achieved. Another example is lactase which hydrolyses lactose to glucose and galactose. Examples of carbohydrases includes amylases, cellulases and mannanases. The carbohydrases to be incorporated in a composition according to the invention is preferably selected from xylanases, cellulases, mannanases and amylases.

Polypeptides Having Mannanase Activity

[0153] The term “mannanase” is defined here as an enzyme that hydrolyses compounds known as mannans. The term “mannanase activity” is as an enzyme catalyzed hydrolysis of mannan, for purposes of the present invention, mannanase activity is determined according to the procedure described in the Assay II. Suitable mannanases include those of bacterial or fungal origin. Chemically or genetically modified mannanases are included. The mannanase may be an alkaline mannanase of Family 5 or 26. It may be a wild-type from *Bacillus* or *Humicola*, particularly *B. agaradhaerens*, *B. licheniformis*, *B. halodurans*, *B. clausii*, or *H. insolens*. Suitable mannanases are described in WO 99/64619. Commercially available mannanases are Mannaway (Novozymes A/S), and EFFECTENZ™ M1000 from Dupont. In one aspect, the present invention relates to a cleaning composition comprising at least one enzyme having mannanase activity, which may be obtained from a bacterial strain of the genus *Bacillus*. Preferably, a polypeptide selected from the group of polypeptides comprising the amino acid sequence shown in SEQ ID NO: 82. In one aspect the present invention relates to a cleaning composition comprising at least one enzyme classified in the EC 3.2.1.78 and which has mannanase activity.

[0154] Useful mannanases include polypeptides that are substantially homologous to the polypeptides shown in SEQ ID NO: 82 and species homologs (paralogs or orthologs) thereof.

[0155] The term “substantially homologous” is used herein to denote polypeptides having at least 60%, at least 65%, preferably at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90% more preferably at least 95%, more preferably at least 97%, even more preferably at least 98% sequence identity to the sequence.

[0156] In some embodiments, the present invention relates to compositions comprising a DNase and a polypeptide having a sequence identity to the polypeptide shown in SEQ ID NO: 82 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have mannanase activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide shown in SEQ ID NO: 82.

[0157] The mannanase suitable for a composition of the invention may, in addition to the enzyme core comprising a catalytically domain, also comprise a cellulose binding domain (CBD), the cellulose binding domain and enzyme core (the catalytically active domain) of the enzyme being operably linked.

[0158] In one aspect, the present invention relates to a composition comprising a mannanase and a DNase, wherein the mannanase is: i) polypeptide comprising an amino acid sequence as shown in SEQ ID NO: 82; or ii) or a polypeptide having a sequence identity of at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% to the amino acid sequence shown in SEQ ID NO: 82.

[0159] In one aspect, the present invention relates to a composition comprising a mannanase and a DNase, wherein the mannanase is selected from a polypeptide having at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82.

Polypeptides Having Cellulase Activity

[0160] The term “cellulase” is defined in the present context as an enzyme that hydrolyses cellulose. In a preferred embodiment of the invention, the cellulase is an endoglucanase. The term “cellulase activity” is defined herein as an enzyme catalyzed hydrolysis of 1,4-beta-D-glucosidic linkages in beta-1,4-glucan (cellulose). For purposes of the present invention, cellulase activity is determined using AZCL-HE-cellulose (from Megazyme) as the reaction substrate, as shown in Assay IV. Suitable cellulases include those of bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Suitable cellulases include cellulases from the genera *Bacillus*, *Pseudomonas*, *Humicola*, *Fusarium*, *Thielavia*, *Acremonium*, e.g., the fungal cellulases produced from *Humicola insolens*, *Myceliophthora thermophila* and *Fusarium oxysporum* disclosed in U.S. Pat. Nos. 4,435,307, 5,648,263, 5,691,178, 5,776,757 and WO 89/09259.

[0161] Especially suitable cellulases are the alkaline or neutral cellulases having color care benefits. Examples of such cellulases are cellulases described in EP 0495257, EP 0531372, WO 96/11262, WO 96/29397, WO 98/08940. Other examples are cellulase variants such as those

described in WO 94/07998, EP 0 531 315, U.S. Pat. Nos. 5,457,046, 5,686,593, 5,763,254, WO 95/24471, WO 98/12307 and WO 99/01544.

[0162] Other cellulases are endo-beta-1,4-glucanase enzyme having a sequence of at least 97% identity to the amino acid sequence of position 1 to position 773 of SEQ ID NO:2 of WO 02/99091 or a family 44 xyloglucanase, which a xyloglucanase enzyme having a sequence of at least 60% identity to positions 40-559 of SEQ ID NO: 2 of WO 01/62903.

[0163] Commercially available cellulases include Cel-luzyme™, and Carezyme™ (Novozymes A/S) Carezyme Premium™ (Novozymes A/S), Celluclean™ (Novozymes A/S), Celluclean Classic™ (Novozymes A/S), Cellusoft™ (Novozymes A/S), Whitezyme™ (Novozymes A/S), Clazinase™, and Puradax HA™ (Genencor International Inc.), and KAC-500(B)™ (Kao Corporation), Revitalenz™ 1000, Revitalenz™ 2000, Revitalenz™ 3000 (Dupont).

[0164] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprise the amino acid sequence of SEQ ID NO: 83. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 83.

[0165] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprise the amino acid sequence of SEQ ID NO: 84. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 84.

[0166] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprise the amino acid sequence of SEQ ID NO: 85. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 85.

[0167] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprise the amino acid sequence of SEQ ID NO: 86. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having cellulase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least

70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 86.

[0168] In the present invention, the cleaning composition can also comprise a cellulase, which is a xyloglucanase. The term "xyloglucanase activity" is defined herein as an enzyme catalyzed hydrolysis of xyloglucan, which is shown in Assay III. Xyloglucanase can comprise parent xyloglucanase and the variants thereof.

[0169] In one embodiment of the present invention, the xyloglucanase is a polypeptide comprising an amino acid sequence of SEQ ID NO: 87. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having xyloglucanase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 87.

Polypeptides Having Amylase Activity

[0170] An amylase is an enzyme that hydrolyses starch into sugars, for purposes of the present invention, amylase activity is determined according to the procedure described in the Assay V. Suitable amylases include alpha-amylases and/or a glucoamylases and may be of bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Amylases include, for example, alpha-amylases obtained from *Bacillus*, e.g., a special strain of *Bacillus licheniformis*, described in more detail in GB 1,296,839.

[0171] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprise the amino acid sequence of SEQ ID NO: 88. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 88.

[0172] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprise the amino acid sequence of SEQ ID NO: 89. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprises an amino acid sequence having at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, even most preferably at least 99% or 100% sequence identity to SEQ ID NO: 89.

[0173] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprise the amino acid sequence

of SEQ ID NO: 90. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprises an amino acid sequence at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99%, identity SEQ ID NO: 90.

[0174] In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprise the amino acid sequence of SEQ ID NO: 91. In one aspect of the present invention, the cleaning composition comprise a DNase and a polypeptide having amylase activity, which comprises an amino acid sequence at least 60%, at least 65%, at least 70%, more preferably at least 75%, more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, more preferably at least 95%, more preferably at least 96%, even more preferably at least 97%, most preferably at least 98%, or even most preferably at least 99%, identity SEQ ID NO: 91.

[0175] Additional amylases include amylases comprising the polypeptide shown in SEQ ID NO: 2 in WO 95/10603 or variants having 90% sequence identity to SEQ ID NO: 3 thereof. Preferred variants are described in WO 94/02597, WO 94/18314, WO 97/43424 and SEQ ID NO: 4 of WO 99/019467, such as variants with substitutions in one or more of the following positions: 15, 23, 105, 106, 124, 128, 133, 154, 156, 178, 179, 181, 188, 190, 197, 201, 202, 207, 208, 209, 211, 243, 264, 304, 305, 391, 408, and 444. Different suitable amylases include amylases having SEQ ID NO: 6 in WO 02/010355 or variants thereof having 90% sequence identity to SEQ ID NO: 6. Preferred variants of SEQ ID NO: 6 are those having a deletion in positions 181 and 182 and a substitution in position 193. Other amylases which are suitable are hybrid alpha-amylase comprising residues 1-33 of the alpha-amylase derived from *B. amyloliquefaciens* shown in SEQ ID NO: 6 of WO 2006/066594 and residues 36-483 of the *B. licheniformis* alpha-amylase shown in SEQ ID NO: 4 of WO 2006/066594 or variants having 90% sequence identity thereof. Preferred variants of this hybrid alpha-amylase are those having a substitution, a deletion or an insertion in one of more of the following positions: 48, 49, 107, 156, 181, 190, 197, 201, 209 and 264. Most preferred variants of the hybrid alpha-amylase comprising residues 1-33 of the alpha-amylase derived from *B. amyloliquefaciens* shown in SEQ ID NO: 6 of WO 2006/066594 and residues 36-483 of SEQ ID NO: 4 are those having the substitutions:

[0176] M197T;

[0177] H156Y+A181T+N190F+A209V+Q264S; or

[0178] G48A+T49I+G107A+H156Y+A181T+N190F+I201F+A209V+Q264S.

[0179] Further amylases which are suitable are amylases having SEQ ID NO: 6 in WO 99/19467 or variants thereof having 90% sequence identity to SEQ ID NO: 6. Preferred variants of SEQ ID NO: 6 are those having a substitution, a deletion or an insertion in one or more of the following positions: R181, G182, H183, G184, N195, I206, E212, E216 and K269. Particularly preferred amylases are those having deletion in positions R181 and G182, or positions H183 and G184. Additional amylases which can be used are

those having SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 2 or SEQ ID NO: 7 of WO 96/23873 or variants thereof having 90% sequence identity to SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3 or SEQ ID NO: 7. Preferred variants of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3 or SEQ ID NO: 7 are those having a substitution, a deletion or an insertion in one or more of the following positions: 140, 181, 182, 183, 184, 195, 206, 212, 243, 260, 269, 304 and 476, using SEQ ID 2 of WO 96/23873 for numbering. More preferred variants are those having a deletion in two positions selected from 181, 182, 183 and 184, such as 181 and 182, 182 and 183, or positions 183 and 184. Most preferred amylase variants of SEQ ID NO: 1, SEQ ID NO: 2 or SEQ ID NO: 7 are those having a deletion in positions 183 and 184 and a substitution in one or more of positions 140, 195, 206, 243, 260, 304 and 476.

[0180] Other amylases which can be used are amylases having SEQ ID NO: 2 of WO 2008/153815, SEQ ID NO: 10 in WO 01/66712 or variants thereof having 90% sequence identity to SEQ ID NO: 2 of WO 2008/153815 or 90% sequence identity to SEQ ID NO: 10 in WO 01/66712. Preferred variants of SEQ ID NO: 10 in WO 01/66712 are those having a substitution, a deletion or an insertion in one of more of the following positions: 176, 177, 178, 179, 190, 201, 207, 211 and 264. Further suitable amylases are amylases having SEQ ID NO: 2 of WO 2009/061380 or variants having 90% sequence identity to SEQ ID NO: 2 thereof. Preferred variants of SEQ ID NO: 2 are those having a truncation of the C-terminus and/or a substitution, a deletion or an insertion in one of more of the following positions: Q87, Q98, S125, N128, T131, T165, K178, R180, S181, T182, G183, M201, F202, N225, S243, N272, N282, Y305, R309, D319, Q320, Q359, K444 and G475. More preferred variants of SEQ ID NO: 2 are those having the substitution in one of more of the following positions: Q87E,R, Q98R, S125A, N128C, T131 I, T165I, K178L, T182G, M201L, F202Y, N225E,R, N272E,R, S243Q,A,E,D, Y305R, R309A, Q320R, Q359E, K444E and G475K and/or deletion in position R180 and/or S181 or of T182 and/or G183. Most preferred amylase variants of SEQ ID NO: 2 are those having the substitutions:

[0181] N128C+K178L+T182G+Y305R+G475K;

[0182] N128C+K178L+T182G+F202Y+Y305R+D319T+G475K;

[0183] S125A+N128C+K178L+T182G+Y305R+G475K; or

[0184] S125A+N128C+T131I+T165I+K178L+T182G+Y305R+G475K wherein the variants are C-terminally truncated and optionally further comprises a substitution at position 243 and/or a deletion at position 180 and/or position 181.

[0185] Further suitable amylases are amylases having SEQ ID NO: 1 of WO 2013/184577 or variants having 90% sequence identity to SEQ ID NO: 1 thereof. Preferred variants of SEQ ID NO: 1 are those having a substitution, a deletion or an insertion in one of more of the following positions: K176, R178, G179, T180, G181, E187, N192, M199, I203, S241, R458, T459, D460, G476 and G477. More preferred variants of SEQ ID NO: 1 are those having the substitution in one of more of the following positions: K176L, E187P, N192FYH, M199L, I203YF, S241QADN, R458N, T459S, D460T, G476K and G477K and/or deletion

in position R178 and/or S179 or of T180 and/or G181. Most preferred amylase variants of SEQ ID NO: 1 are those having the substitutions:

[0186] E187P+I203Y+G476K

[0187] E187P+I203Y+R458N+T459S+D460T+G476K,

wherein the variants optionally further comprise a substitution at position 241 and/or a deletion at position 178 and/or position 179.

[0188] Further suitable amylases are amylases having SEQ ID NO: 1 of WO 2010/104675 or variants having 90% sequence identity to SEQ ID NO: 1 thereof. Preferred variants of SEQ ID NO: 1 are those having a substitution, a deletion or an insertion in one of more of the following positions: N21, D97, V128 K177, R179, S180, I181, G182, M200, L204, E242, G477 and G478. More preferred variants of SEQ ID NO: 1 are those having the substitution in one of more of the following positions: N21D, D97N, V128I, K177L, M200L, L204YF, E242QA, G477K and G478K and/or deletion in position R179 and/or S180 or of I181 and/or G182. Most preferred amylase variants of SEQ ID NO: 1 are those having the substitutions:

[0189] N21 D+D97N+V128I

wherein the variants optionally further comprise a substitution at position 200 and/or a deletion at position 180 and/or position 181. Other suitable amylases are the alpha-amylase comprising the polypeptide sequence shown in SEQ ID NO: 12 in WO 01/66712 or a variant having at least 90% sequence identity to SEQ ID NO: 12. Preferred amylase variants are those having a substitution, a deletion or an insertion in one of more of the following positions of SEQ ID NO: 12 in WO 01/66712: R28, R118, N174; R181, G182, D183, G184, G186, W189, N195, M202, Y298, N299, K302, S303, N306, R310, N314; R320, H324, E345, Y396, R400, W439, R444, N445, K446, Q449, R458, N471, N484. Preferred amylases include variants having a deletion of D183 and G184 and having the substitutions R118K, N195F, R320K and R458K, and a variant additionally having substitutions in one or more position selected from the group: M9, G149, G182, G186, M202, T257, Y295, N299, M323, E345 and A339, most preferred a variant that additionally has substitutions in all these positions. Other examples are amylase variants such as those described in WO 2011/098531, WO 2013/001078 and WO 2013/001087.

[0190] Commercially available amylases are DuramylTM, TermamylTM, FungamylTM, StainzymeTM, Stainzyme PlusTM, NatalaseTM, Liquozyme X and BANTM (from Novozymes A/S), and RapidaseTM, PurastarTM/EffectenzTM, Powerase, Preferenz S1000, Preferenz S100 and Preferenz S110 (from Genencor International Inc./DuPont).

Compositions

[0191] The invention relates to compositions, preferably cleaning compositions comprising a DNase and a carbohydase in combination with one or more additional cleaning composition components.

[0192] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydase and a cleaning component, wherein the carbohydase is a cellulase, an amylase, a mannanase or a xylanase. The carbohydase may be any of the cellulases, amylases, mannanases or xylanases mentioned under the heading "Polypeptides having cellulase, amylase, mannanase or xylanase activity" respectively.

[0193] As shown in the examples of the present invention carbohydases such as cellulases act synergistically with the DNase in reduction, and removal of biofilm or components hereof. Biofilm is a complex structure comprising, the target substrate, e.g., the DNA may be embedded in the biofilm structure and it's believed that when the DNases and carbohydases are acting together, the DNA components are more effectively dispersed or removed. It is thus advantageous to formulate DNases with carbohydases such as cellulases, amylases, mannanases and xylanases in cleaning compositions, e.g., for deep cleaning. One aspect of the invention relates to a method of formulating a cleaning composition comprising adding a DNase, at least one carbohydase and a cleaning component, wherein the carbohydase is a cellulase, an amylase, a mannanase or a xylanase. The invention further relates to a kit intended for deep cleaning, wherein the kit comprises a solution of an enzyme mixture comprising a DNase and a carbohydase, wherein the carbohydase is a cellulase, an amylase, a mannanase or a xylanase.

[0194] In one aspect of the invention the carbohydase is a cellulase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydase and a cleaning component, wherein the carbohydase is a cellulase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydase and a cleaning component, wherein the carbohydase is a cellulase, preferably selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 86.

[0195] In one aspect of the invention the carbohydase is an amylase.

[0196] In one aspect of the invention the carbohydase is an amylase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydase and a cleaning component, wherein the carbohydase is an amylase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydase and a cleaning component, wherein the carbohydase is an amylase, preferably selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91.

least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0197] In one aspect of the invention the carbohydrase is a mannanase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydrase and a cleaning component, wherein the carbohydrase is a mannanase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydrase and a cleaning component, wherein the carbohydrase is a mannanase, preferably a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82.

[0198] In one aspect of the invention the carbohydrase is a xylanase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydrase and a cleaning component, wherein the carbohydrase is a xylanase. In one aspect, the invention relates to a cleaning composition comprising a DNase, a carbohydrase and a cleaning component, wherein the carbohydrase is a xylanase, preferably a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87.

[0199] The DNases to be formulated together with the carbohydrases or to be used together with the carbohydrases should be compatible with cleaning components. DNases are at present not standard ingredients in cleaning compositions. However, the applicant has identified DNases suitable for use in cleaning compositions, e.g., in WO 2017/060475, WO 2014/087011, WO 2015/155350 and WO 2015/155351. These applications also mentioned that DNases may be formulated with other enzymes, e.g., carbohydrases. However, none of these applications indicate that the DNases may have synergy with, e.g., cellulases. Enzymes, such as DNases should not only be compatible with the cleaning components, the DNases should also be compatible with other enzymes which may be present in a typical cleaning composition. Surprisingly, it was found that carbohydrases such as cellulases and DNases not only are compatible but even act synergistically in respect of biofilm reduction and removal, e.g., in deep cleaning.

[0200] Particularly useful DNases may be those of microbial origin. One embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase and at least one cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase is microbial, preferably obtained from bacteria or fungi. In one embodiment, the cleaning composition comprise a DNase from bacteria. One embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase and at least one cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase is obtained from *Bacillus*, preferably *Bacillus cibi*, *Bacillus horikoshii*, *Bacillus licheniformis*, *Bacillus subtilis*, *Bacillus horneckiae*, *Bacillus idriensis*, *Bacillus algicola*, *Bacillus vietnamensis*, *Bacillus hwajinpoensis*, *Bacillus indicus*, *Bacillus marisflavi* or *Bacillus luciferensis*.

[0201] As mentioned above the DNases to be used in a composition of the invention preferable belong to the NUC1

group of DNases. The NUC1 group of DNases may comprise one or more of the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], or C[D/N]T[A/R]. One embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and at least one cleaning component, wherein the DNase comprises one or more of the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V] or C[D/N]T[A/R]. The DNases preferably additionally comprises a NUC1_A domain [D/Q][I/V]DH.

[0202] One embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and at least one cleaning component, wherein the DNase comprises one or more motifs selected from the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R] and [D/Q][I/V]DH.

[0203] One preferred embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and at least one cleaning component, wherein the DNase comprises two or more motifs selected from the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R] and [D/Q][I/V]DH.

[0204] One preferred embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and at least one cleaning component, wherein the DNase comprises three or more motifs selected from the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R] and [D/Q][I/V]DH.

[0205] One preferred embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and at least one cleaning component, wherein the DNase comprises four or more motifs selected from the motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R] and [D/Q][I/V]DH.

[0206] One preferred embodiment of the invention relates to a cleaning composition comprising a DNase, a carbohydrase, selected from cellulases, amylases, mannanases and xylanases, and at least one cleaning component, wherein the DNase comprises all five motifs [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R] and [D/Q][I/V]DH.

[0207] The DNases to be added to a composition of the invention preferably belong to the group of DNases comprised in the GYS-clade, which are NUC1 and NUC1_A DNases further comprising the conservative motifs [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74) and which share similar structural and functional properties. The DNases of the GYS-clade are preferably obtained from *Bacillus* genus.

[0208] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase comprises one or both of the motif(s) [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74).

[0209] In a particularly preferred embodiment the *Bacillus* DNase comprises one or both of the motif(s) [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74). In another particularly preferred embodiment the DNase comprises one or both of the motif(s) [D/M/L][S/T]

GYSR[D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74) and is obtained from *Bacillus cibi*. In yet another preferred embodiment the DNase comprises the amino acid sequence shown in SEQ ID NO: 13 or DNases closely related hereto.

[0210] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 13.

[0211] Other preferred DNases include those comprising the amino acid sequence shown in SEQ ID Nos: 65 and 66.

[0212] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 65.

[0213] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 66.

[0214] The DNase may also preferably be fungal. Particularly preferred are DNases obtained from *Aspergillus* in particular, *Aspergillus oryzae*.

[0215] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 67.

[0216] Other particularly preferred are DNases obtained from *Trichoderma* in particular, *Trichoderma harzianum*.

[0217] One embodiment of the invention relates to a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0218] One embodiment relates to a cleaning composition comprising a *Bacillus* DNase, a cellulase and at least one cleaning component, wherein the cellulase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at

least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 86.

[0219] One embodiment relates to a cleaning composition comprising a *Bacillus* DNase, an amylase and at least one cleaning component, wherein the amylase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0220] One embodiment relates to a cleaning composition comprising a *Bacillus* DNase, a mannanase and at least one cleaning component, wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82.

[0221] One embodiment relates to a cleaning composition comprising a *Bacillus* DNase, a xylanase and at least one cleaning component, wherein the xylanase is selected from preferably a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87.

[0222] One embodiment relates to a cleaning composition comprising a DNase, a cellulase and at least one cleaning component, wherein the cellulase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 86 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87.

at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 86 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0239] One embodiment relates to a cleaning composition comprising a DNase, an amylase and at least one cleaning component, wherein the amylase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0240] One embodiment relates to a cleaning composition comprising a DNase, a mannanase and at least one cleaning component, wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0241] One embodiment relates to a cleaning composition comprising a DNase, a xylanase and at least one cleaning component, wherein the xylanase is selected from preferably a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0242] One embodiment of the invention relates to a composition comprising

[0243] a) at least 0.001 ppm, e.g., 0.1 ppm or 1 ppm of at least one polypeptide having DNase activity, wherein the DNase is selected for the group consisting of:

[0244] i) a DNase comprising one or more of the motif(s) [T/D/S][G/N]PQL, [F/L/Y/I]A[N/R]D[L/I/P/V], C[D/N]T[A/R];

[0245] ii) a DNase comprising the motif [D/Q][I/V]DH;

[0246] iii) a DNase comprising one or both of the motif(s) [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74);

[0247] iv) a DNase comprising one or both of the motifs [V/I]PL[S/A]NAWK (SEQ ID NO: 75) or NPQL (SEQ ID NO: 76);

[0248] v) a DNase comprising one or both of the motifs P[Q/E]L[W/Y] or [K/H/E]NAW;

[0249] vi) a DNase selected from: a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 1, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 2, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 3, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 4, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 5, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 6, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 7, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 8, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 9, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 10, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 11, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 12, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 13, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 14, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 15, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 16, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 17, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 18, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 19, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 20, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 21, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 22, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 23, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 24, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 25, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 26, a polypeptide having at least 80% sequence identity to the poly-

peptide shown in SEQ ID NO: 59, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 60, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 61, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 62, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 63, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 64, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 65, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 66, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 67, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 68; and

[0250] b) at least 0.001 ppm, e.g., 0.1 ppm or 1 ppm of one or more carbohydrase, wherein the carbohydrase is selected from the group consisting of:

[0251] i. a cellulase selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 86;

[0252] ii. a xylanase selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87;

[0253] iii. a mannanase selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82;

[0254] iv. an amylase selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least

98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91; and

[0255] c) at least one additional component, e.g., cleaning component, preferably selected from surfactants, builders, bleach components, polymers and dispersing agents.

[0256] The carbohydrazase and DNase may be included in the cleaning composition of the present invention at a level of from 0.01 to 1000 ppm, from 1 ppm to 1000 ppm, from 10 ppm to 1000 ppm, from 50 ppm to 1000 ppm, from 100 ppm to 1000 ppm, from 150 ppm to 1000 ppm, from 200 ppm to 1000 ppm, from 250 ppm to 1000 ppm, from 250 ppm to 750 ppm, from 250 ppm to 500 ppm. The DNases above may be combined with carbohydrazases to form a blend to be added to the wash liquor solution according to the invention. The concentration of the DNase in the wash liquor solution is typically in the range of wash liquor from 0.00001 ppm to 10 ppm, from 0.00002 ppm to 10 ppm, from 0.0001 ppm to 10 ppm, from 0.0002 ppm to 10 ppm, from 0.001 ppm to 10 ppm, from 0.002 ppm to 10 ppm, from 0.01 ppm to 10 ppm, from 0.02 ppm to 10 ppm, 0.1 ppm to 10 ppm, from 0.2 ppm to 10 ppm, from 0.5 ppm to 5 ppm. The concentration of the carbohydrazases in the wash liquor solution is typically in the range of wash liquor from 0.00001 ppm to 10 ppm, from 0.00002 ppm to 10 ppm, from 0.0001 ppm to 10 ppm, from 0.0002 ppm to 10 ppm, from 0.001 ppm to 10 ppm, from 0.002 ppm to 10 ppm, from 0.01 ppm to 10 ppm, from 0.02 ppm to 10 ppm, 0.1 ppm to 10 ppm, from 0.2 ppm to 10 ppm, from 0.5 ppm to 5 ppm. The DNases may be combined with any of the carbohydrazases below to form a blend to be added to a composition according to the invention.

[0257] One embodiment relates to a cleaning composition comprising a DNase, a carbohydrazase and at least one cleaning component, wherein the amount of DNase in the composition is from 0.01 to 1000 ppm and the amount of carbohydrazase is from 0.01 to 1000 ppm.

[0258] The invention relates to cleaning compositions comprising an enzyme combination of the present invention in combination with one or more additional cleaning composition component(s). The choice of additional components is within the skill of the artisan and includes conventional ingredients, including the exemplary non-limiting components set forth below.

[0259] The choice of cleaning components may include, for textile care, the consideration of the type of textile to be cleaned, the type and/or degree of soiling, the temperature at which cleaning is to take place, and the formulation of the detergent product. Although components mentioned below are categorized by general header according to a particular functionality, this is not to be construed as a limitation, as a component may comprise additional functionalities as will be appreciated by the skilled artisan.

Surfactants

[0260] The detergent composition may comprise one or more surfactants, which may be anionic and/or cationic and/or non-ionic and/or semi-polar and/or zwitterionic, or a mixture thereof. In a particular embodiment, the detergent composition includes a mixture of one or more nonionic surfactants and one or more anionic surfactants. The surfactant(s) is typically present at a level of from about 0.1% to 60% by weight, such as about 0.1% to about 15%, such as about 1% to about 40%, or about 3% to about 20%, or about

3% to about 10%. The surfactant(s) is chosen based on the desired cleaning application, and may include any conventional surfactant(s) known in the art. When included therein the detergent will usually contain from about 1% to about 40% by weight of an anionic surfactant, such as from about 5% to about 30%, including from about 5% to about 15%, or from about 15% to about 20%, or from about 20% to about 25% of an anionic surfactant. Non-limiting examples of anionic surfactants include sulfates and sulfonates, in particular, linear alkylbenzenesulfonates (LAS), isomers of LAS, branched alkylbenzenesulfonates (BABS), phenylkanesulfonates, alpha-olefinsulfonates (AOS), olefin sulfonates, alkene sulfonates, alkane-2,3-diyldisulfates, hydroxyalkanesulfonates and disulfonates, alkyl sulfates (AS) such as sodium dodecyl sulfate (SDS), fatty alcohol sulfates (FAS), primary alcohol sulfates (PAS), alcohol ethersulfates (AES or AEOS or FES, also known as alcohol ethoxysulfates or fatty alcohol ether sulfates), secondary alkanesulfonates (SAS), paraffin sulfonates (PS), ester sulfonates, sulfonated fatty acid glycerol esters, alpha-sulfo fatty acid methyl esters (alpha-SFMe or SES) including methyl ester sulfonate (MES), alkyl- or alkenylsuccinic acid, dodecenyl/tetradecenyl succinic acid (DTSA), fatty acid derivatives of amino acids, diesters and monoesters of sulfo-succinic acid or salt of fatty acids (soap), and combinations thereof.

[0261] When included therein the detergent will usually contain from about 1% to about 40% by weight of a cationic surfactant, for example from about 0.5% to about 30%, in particular from about 1% to about 20%, from about 3% to about 10%, such as from about 3% to about 5%, from about 8% to about 12% or from about 10% to about 12%. Non-limiting examples of cationic surfactants include alkyldimethylethanolamine quat (ADMEAQ), cetyltrimethylammonium bromide (CTAB), dimethyldistearylammnonium chloride (DSDMAC), and alkylbenzyldimethylammonium, alkyl quaternary ammonium compounds, alkoxyLATED quaternary ammonium (AQA) compounds, ester quats, and combinations thereof.

[0262] When included therein the detergent will usually contain from about 0.2% to about 40% by weight of a nonionic surfactant, for example from about 0.5% to about 30%, in particular from about 1% to about 20%, from about 3% to about 10%, such as from about 3% to about 5%, from about 8% to about 12%, or from about 10% to about 12%. Non-limiting examples of nonionic surfactants include alcohol ethoxylates (AE or AEO), alcohol propoxylates, propoxylated fatty alcohols (PFA), alkoxyLATED fatty acid alkyl esters, such as ethoxylated and/or propoxylated fatty acid alkyl esters, alkylphenol ethoxylates (APE), nonylphenol ethoxylates (NPE), alkylpolyglycosides (APG), alkoxyLATED amines, fatty acid monoethanolamides (FAM), fatty acid diethanolamides (FADA), ethoxylated fatty acid monoethanolamides (EFAM), propoxylated fatty acid monoethanolamides (PFAM), polyhydroxyalkyl fatty acid amides, or N-acyl N-alkyl derivatives of glucosamine (glucamides, GA, or fatty acid glucamides, FAGA), as well as products available under the trade names SPAN and TWEEN, and combinations thereof. When included therein the detergent will usually contain from about 0.01% to about 10% by weight of a semipolar surfactant. Non-limiting examples of semipolar surfactants include amine oxides (AO) such as alkyldimethylamineoxide, N-(coco alkyl)-N,N-dimethylamine oxide and N-(tallow-alkyl)-N,N-bis(2-hydroxyethyl)

amine oxide, and combinations thereof. When included therein the detergent will usually contain from about 0.01% to about 10% by weight of a zwitterionic surfactant. Non-limiting examples of zwitterionic surfactants include betaines such as alkyldimethylbetaines, sulfobetaines, and combinations thereof.

Builders and Co-Builders

[0263] The detergent composition may contain about 0-65% by weight, such as about 5% to about 50%, such as from about 0.5 to about 20% of a detergent builder or co-builder, or a mixture thereof. In a dish wash detergent, the level of builder is typically 40-65%, particularly 50-65%. The builder and/or co-builder may particularly be a chelating agent that forms water-soluble complexes with Ca and Mg. Any builder and/or co-builder known in the art for use in cleaning detergents may be utilized. Non-limiting examples of builders include zeolites, diphosphates (pyrophosphates), triphosphates such as sodium tripolyphosphate (STP or STPP), carbonates such as sodium carbonate, soluble silicates such as sodium metasilicate, layered silicates (e.g., SKS-6 from Hoechst), ethanolamines such as 2-aminoethan-1-ol (MEA), diethanolamine (DEA, also known as 2,2'-iminodiethan-1-ol), triethanolamine (TEA, also known as 2,2',2"-nitrilotriethan-1-ol), and (carboxymethyl)inulin (CMI), and combinations thereof.

[0264] The detergent composition may also contain 0-50% by weight, such as about 5% to about 30%, of a detergent co-builder. The detergent composition may include a co-builder alone, or in combination with a builder, for example a zeolite builder. Non-limiting examples of co-builders include homopolymers of polyacrylates or copolymers thereof, such as poly(acrylic acid) (PAA) or copoly(acrylic acid/maleic acid) (PAA/PMA). Further non-limiting examples include citrate, chelators such as aminocarboxylates, aminopolycarboxylates and phosphonates, and alkyl- or alkenylsuccinic acid. Additional specific examples include 2,2',2"-nitrilotriacetic acid (NTA), ethylenediaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTPA), iminodisuccinic acid (IDS), ethylenediamine-N,N'-disuccinic acid (EDDS), methylglycinediacetic acid (MGDA), glutamic acid-N,N-diacetic acid (GLDA), 1-hydroxyethane-1,1-diphosphonic acid (HEDP), ethylenediaminetetra(methyleneephosphonic acid) (EDTPMPA), diethylenetriaminepentakis(methyleneephosphonic acid) (DTMPA or DTPMPA), N-(2-hydroxyethyl)iminodiacetic acid (EDG), aspartic acid-N-monoacetic acid (ASMA), aspartic acid-N,N-diacetic acid (ASDA), aspartic acid-N-monopropionic acid (ASMP), iminodisuccinic acid (IDA), N-(2-sulfomethyl)-aspartic acid (SMAS), N-(2-sulfoethyl)-aspartic acid (SEAS), N-(2-sulfomethyl)-glutamic acid (SMGL), N-(2-sulfoethyl)-glutamic acid (SEGL), N-methyliminodiacetic acid (MIDA), α -alanine-N,N-diacetic acid (α -ALDA), serine-N,N-diacetic acid (SEDA), isoserine-N, N-diacetic acid (ISDA), phenylalanine-N,N-diacetic acid (PHDA), anthranilic acid-N,N-diacetic acid (ANDA), sulfanilic acid-N,N-diacetic acid (SLDA), taurine-N,N-diacetic acid (TUDA) and sulfomethyl-N,N-diacetic acid (SMDA), N-(2-hydroxyethyl)ethylenediamine-N,N',N"-triacetic acid (HEDTA), diethanolglycine (DEG), diethylenetriamine penta(methyleneephosphonic acid) (DTPMP), aminotris(methyleneephosphonic acid) (ATMP), and combinations and salts

thereof. Further exemplary builders and/or co-builders are described in, e.g., WO 2009/102854, U.S. Pat. No. 5,977,053.

Bleaching Systems

[0265] The detergent may contain 0-30% by weight, such as about 1% to about 20%, such as from about 0.01 to about 10 wt. % of a bleaching system. Any bleaching system comprising components known in the art for use in cleaning detergents may be utilized. Suitable bleaching system components include sources of hydrogen peroxide; sources of peracids; and bleach catalysts or boosters.

Sources of Hydrogen Peroxide:

[0266] Suitable sources of hydrogen peroxide are inorganic persalts, including alkali metal salts such as sodium percarbonate and sodium perborates (usually mono- or tetrahydrate), and hydrogen peroxide-urea (1/1).

Sources of Peracids:

[0267] Peracids may be (a) incorporated directly as pre-formed peracids or (b) formed in situ in the wash liquor from hydrogen peroxide and a bleach activator (perhydrolysis) or (c) formed in situ in the wash liquor from hydrogen peroxide and a perhydrolase and a suitable substrate for the latter, e.g., an ester.

[0268] a) Suitable preformed peracids include, but are not limited to, peroxycarboxylic acids such as peroxybenzoic acid and its ring-substituted derivatives, peroxy- α -naphthoic acid, peroxyphthalic acid, peroxylauric acid, peroxystearic acid, ϵ -phthalimidoperoxycaproic acid [phthalimidoperoxyhexanoic acid (PAP)], and o-carboxybenzamidoperoxycaproic acid; aliphatic and aromatic diperoxydicarboxylic acids such as diperoxydodecanedioic acid, diperoxyazelaic acid, diperoxysebacic acid, diperoxybrassylic acid, 2-decyldiperoxybutanedioic acid, and diperoxyphthalic, -isophthalic and -terephthalic acids; perimidic acids; peroxymonosulfuric acid; peroxydisulfuric acid; peroxyphosphoric acid; peroxysilicic acid; and mixtures of said compounds. It is understood that the peracids mentioned may in some cases be best added as suitable salts, such as alkali metal salts (e.g., Oxone®) or alkaline earth-metal salts.

[0269] b) Suitable bleach activators include those belonging to the class of esters, amides, imides, nitriles or anhydrides and, where applicable, salts thereof. Suitable examples are tetraacetyl ethylenediamine (TAED), sodium 4-[(3,5,5-trimethylhexanoyl)oxy]benzene-1-sulfonate (ISONOBS), sodium 4-(dodecanoyloxy)benzene-1-sulfonate (LOBS), sodium 4-(decanoxyloxy)benzene-1-sulfonate, 4-(decanoxyloxy)benzoic acid (DOBA), sodium 4-(nonanoyloxy)benzene-1-sulfonate (NOBS), and/or those disclosed in WO 98/17767. A particular family of bleach activators of interest was disclosed in EP 624154 and particularly preferred in that family is acetyl triethyl citrate (ATC). ATC or a short chain triglyceride like triacetin has the advantage that they are environmentally friendly. Furthermore, acetyl triethyl citrate and triacetin have good hydrolytical stability in the product upon storage and are efficient bleach activators. Finally, ATC is multi-

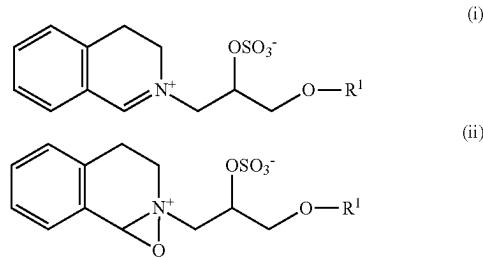
functional, as the citrate released in the perhydrolysis reaction may function as a builder.

Bleach Catalysts and Boosters

[0270] The bleaching system may also include a bleach catalyst or booster.

[0271] Some non-limiting examples of bleach catalysts that may be used in the compositions of the present invention include manganese oxalate, manganese acetate, manganese-collagen, cobalt-amine catalysts and manganese triazacyclononane (MnTACN) catalysts; particularly preferred are complexes of manganese with 1,4,7-trimethyl-1,4,7-triazacyclononane (Me₃-TACN) or 1,2,4,7-tetramethyl-1,4,7-triazacyclononane (Me₄-TACN), in particular Me₃-TACN, such as the dinuclear manganese complex [(Me₃-TACN)Mn(O)₃Mn(Me₃-TACN)](PF₆)₂, and [2,2',2"-nitrilotris(ethane-1,2-diylazanylidene-κN-methanylidene)triphenolato-κ₃O]manganese(III). The bleach catalysts may also be other metal compounds; such as iron or cobalt complexes.

[0272] In some embodiments, where a source of a peracid is included, an organic bleach catalyst or bleach booster may be used having one of the following formulae:



(iii) and mixtures thereof; wherein each R¹ is independently a branched alkyl group containing from 9 to 24 carbons or linear alkyl group containing from 11 to 24 carbons, preferably each R¹ is independently a branched alkyl group containing from 9 to 18 carbons or linear alkyl group containing from 11 to 18 carbons, more preferably each R¹ is independently selected from the group consisting of 2-propylheptyl, 2-butyloctyl, 2-pentylnonyl, 2-hexyldecyl, dodecyl, tetradecyl, hexadecyl, octadecyl, isononyl, isodecyl, isotridecyl and isopentadecyl.

[0273] Other exemplary bleaching systems are described, e.g., in WO 2007/087258, WO 2007/087244, WO 2007/087259, EP 1867708 (Vitamin K) and WO 2007/087242. Suitable photobleaches may for example be sulfonated zinc or aluminium phthalocyanines.

Metal Care Agents

[0274] Metal care agents may prevent or reduce the tarnishing, corrosion or oxidation of metals, including aluminum, stainless steel and non-ferrous metals, such as silver and copper. Suitable examples include one or more of the following:

[0275] (a) benzatriazoles, including benzotriazole or bis-benzotriazole and substituted derivatives thereof. Benzotriazole derivatives are those compounds in which the available substitution sites on the aromatic ring are partially or completely substituted. Suitable

substituents include linear or branch-chain C1-C20-alkyl groups (e.g., C1-C20-alkyl groups) and hydroxyl, thio, phenyl or halogen such as fluorine, chlorine, bromine and iodine.

[0276] (b) metal salts and complexes chosen from the group consisting of zinc, manganese, titanium, zirconium, hafnium, vanadium, cobalt, gallium and cerium salts and/or complexes, the metals being in one of the oxidation states II, III, IV, V or VI. In one aspect, suitable metal salts and/or metal complexes may be chosen from the group consisting of Mn(II) sulphate, Mn(II) citrate, Mn(II) stearate, Mn(II) acetylacetone, K⁺TiF₆ (e.g., K₂TiF₆), K⁺ZrF₆ (e.g., K₂ZrF₆), CoSO₄, Co(NOs)₂ and Ce(NO₃)₃, zinc salts, for example zinc sulphate, hydrozincite or zinc acetate.

[0277] (c) silicates, including sodium or potassium silicate, sodium disilicate, sodium metasilicate, crystalline phyllosilicate and mixtures thereof.

[0278] Further suitable organic and inorganic redox-active substances that act as silver/copper corrosion inhibitors are disclosed in WO 94/26860 and WO 94/26859. Preferably the composition of the invention comprises from 0.1 to 5% by weight of the composition of a metal care agent, preferably the metal care agent is a zinc salt.

Hydrotopes

[0279] The detergent may contain 0-10% by weight, for example 0-5% by weight, such as about 0.5 to about 5%, or about 3% to about 5%, of a hydrotrope. Any hydrotrope known in the art for use in detergents may be utilized. Non-limiting examples of hydrotopes include sodium benzenesulfonate, sodium p-toluene sulfonate (STS), sodium xylene sulfonate (SXS), sodium cumene sulfonate (SCS), sodium cymene sulfonate, amine oxides, alcohols and polyglycolethers, sodium hydroxynaphthoate, sodium hydroxynaphthalene sulfonate, sodium ethylhexyl sulfate, and combinations thereof.

Polymers

[0280] The detergent may contain 0-10% by weight, such as 0.5-5%, 2-5%, 0.5-2% or 0.2-1% of a polymer. Any polymer known in the art for use in detergents may be utilized. The polymer may function as a co-builder as mentioned above, or may provide antiredeposition, fiber protection, soil release, dye transfer inhibition, grease cleaning and/or anti-foaming properties. Some polymers may have more than one of the above-mentioned properties and/or more than one of the below-mentioned motifs. Exemplary polymers include (carboxymethyl)cellulose (CMC), poly(vinyl alcohol) (PVA), poly(vinylpyrrolidone) (PVP), poly(ethyleneglycol) or poly(ethylene oxide) (PEG), ethoxylated poly(ethyleneimine), carboxymethyl inulin (CMI), and polycarboxylates such as PAA, PAA/PMA, poly-aspartic acid, and lauryl methacrylate/acrylic acid copolymers, hydrophobically modified CMC (HM-CMC) and silicones, copolymers of terephthalic acid and oligomeric glycols, copolymers of poly(ethylene terephthalate) and poly(oxyethylene terephthalate) (PET-POET), PVP, poly(vinylimidazole) (PVI), poly(vinylpyridine-N-oxide) (PVPO or PVPNO) and polyvinylpyrrolidone-vinylimidazole (PVPVI). Suitable examples include PVP-K15, PVP-K30, Chromabond S-400, Chromabond S-403E and Chromabond S-100 from Ashland Aqualon, and Sokalan®

HP 165, Sokalan® HP 50 (Dispersing agent), Sokalan® HP 53 (Dispersing agent), Sokalan® HP 59 (Dispersing agent), Sokalan® HP 56 (dye transfer inhibitor), Sokalan® HP 66K (dye transfer inhibitor) from BASF. Further exemplary polymers include sulfonated polycarboxylates, polyethylene oxide and polypropylene oxide (PEO-PPO) and diquaternium ethoxy sulfate. Other exemplary polymers are disclosed in, e.g., WO 2006/130575. Salts of the above-mentioned polymers are also contemplated. Particularly preferred polymer is ethoxylated homopolymer Sokalan® HP 20 from BASF.

Fabric Hueing Agents

[0281] The detergent compositions of the present invention may also include fabric hueing agents such as dyes or pigments, which when formulated in detergent compositions can deposit onto a fabric when said fabric is contacted with a wash liquor comprising said detergent compositions and thus altering the tint of said fabric through absorption/reflection of visible light. Fluorescent whitening agents emit at least some visible light. In contrast, fabric hueing agents alter the tint of a surface as they absorb at least a portion of the visible light spectrum. Suitable fabric hueing agents include dyes and dye-clay conjugates, and may also include pigments. Suitable dyes include small molecule dyes and polymeric dyes. Suitable small molecule dyes include small molecule dyes selected from the group consisting of dyes falling into the Colour Index (C.I.) classifications of Direct Blue, Direct Red, Direct Violet, Acid Blue, Acid Red, Acid Violet, Basic Blue, Basic Violet and Basic Red, or mixtures thereof, for example as described in WO 2005/003274, WO 2005/003275, WO 2005/003276 and EP 1876226 (hereby incorporated by reference). The detergent composition preferably comprises from about 0.00003 wt. % to about 0.2 wt. %, from about 0.00008 wt. % to about 0.05 wt. %, or even from about 0.0001 wt. % to about 0.04 wt. % fabric hueing agent. The composition may comprise from 0.0001 wt. % to 0.2 wt. % fabric hueing agent, this may be especially preferred when the composition is in the form of a unit dose pouch. Suitable hueing agents are also disclosed in, e.g., WO 2007/087257 and WO 2007/087243.

Enzymes

[0282] The detergent additive as well as the detergent composition may comprise one or more additional enzymes such as one or more protease, lipase, cutinase, pectinase, arabinase, galactanase, xylanase, oxidase, e.g., a laccase, and/or peroxidase.

[0283] In general, the properties of the selected enzyme(s) should be compatible with the selected detergent, (i.e., pH-optimum, compatibility with other enzymatic and non-enzymatic ingredients, etc.), and the enzyme(s) should be present in effective amounts.

Proteases

[0284] The term “protease activity” means a proteolytic activity (EC 3.4). Proteases usably in cleaning compositions of the present invention are mainly endopeptidases (EC 3.4.21). There are several protease activity types: The three main activity types are: trypsin-like where there is cleavage of amide substrates following Arg or Lys at P1, chy-

motrypsin-like where cleavage occurs following one of the hydrophobic amino acids at P1, and elastase-like with cleavage following an Ala at P1.

[0285] The most widely used proteases in the detergent industry such as laundry and dish wash are the serine proteases. Serine proteases is a subgroup of proteases characterised by having a serine in the active site, which forms a covalent adduct with the substrate. Serine proteases are characterized by having two active site amino acid residues apart from the serine, namely a histidine residue and an aspartic acid residue. Subtilase refer to a sub-group of serine protease according to Siezen et al., 1991, *Protein Engng.* 4: 719-737 and Siezen et al., 1997, *Protein Science* 6: 501-523. The subtilases may be divided into 6 sub-divisions, i.e., the Subtilisin family, the Thermitase family, the Proteinase K family, the Lantibiotic peptidase family, the Kexin family and the Pyrolysin family.

[0286] Suitable proteases for the compositions of the invention include those of bacterial, fungal, plant, viral or animal origin, e.g., vegetable or microbial origin. Microbial origin is preferred. Chemically modified or protein engineered mutants are included. It may be an alkaline protease, such as a serine protease or a metalloprotease. A serine protease may for example be of the S1 family, such as trypsin, or the S8 family such as subtilisin. A metalloproteases protease may for example be a thermolysin from, e.g., family M4 or other metalloprotease such as those from M5, M7 or M8 families.

[0287] Examples of subtilases are those derived from *Bacillus* such as *Bacillus lentus*, *Bacillus alkalophilus*, *Bacillus subtilis*, *Bacillus amyloliquefaciens*, *Bacillus pumilus* and *Bacillus gibsonii* described in: U.S. Pat. No. 7,262,042 and WO 2009/021867, and subtilisin *lentus*, subtilisin Novo, subtilisin Carlsberg, *Bacillus licheniformis*, subtilisin BPN', subtilisin 309, subtilisin 147 and subtilisin 168 described in WO 89/06279 and protease PD138 described in (WO 93/18140). Other useful proteases may be those described in WO 92/175177, WO 01/16285, WO 02/26024 and WO 02/16547. Examples of trypsin-like proteases are trypsin (e.g., of porcine or bovine origin) and the *Fusarium* protease described in WO 89/06270, WO 94/25583 and WO 2005/040372, and the chymotrypsin proteases derived from *Cellumonas* described in WO 2005/052161 and WO 2005/052146.

[0288] A further preferred protease is the alkaline protease from *Bacillus lentus* DSM 5483, as described for example in WO 95/23221, and variants thereof which are described in WO 92/21760, WO 95/23221, EP 1921147 and EP 1921148.

[0289] Examples of metalloproteases are the neutral metalloproteases as described in WO 2007/044993 (Genencor Int.) such as those derived from *Bacillus amyloliquefaciens*.

[0290] Examples of useful proteases are the variants described in: WO 92/19729, WO 96/034946, WO 98/20115, WO 98/20116, WO 99/11768, WO 01/44452, WO 03/006602, WO 2004/03186, WO 2004/041979, WO 2007/006305, WO 2011/036263, WO 2011/036264, especially protease variants comprising a substitution in one or more of the following positions: 3, 4, 9, 15, 24, 27, 42, 55, 59, 60, 66, 74, 85, 96, 97, 98, 99, 100, 101, 102, 104, 116, 118, 121, 126, 127, 128, 154, 156, 157, 158, 161, 164, 176, 179, 182, 185, 188, 189, 193, 198, 199, 200, 203, 206, 211, 212, 216, 218, 226, 229, 230, 239, 246, 255, 256, 268 and 269, wherein the positions correspond to the positions of the *Bacillus lentus* protease shown in SEQ ID NO: 79. More

preferred the protease variants may comprise one or more of the mutations selected from the group consisting of: S3T, V41, S9R, S9E, A15T, S24G, S24R, K27R, N42R, S55P, G59E, G59D, N60D, N60E, V66A, N74D, S85R, A96S, S97G, S97D, S97A, S97SD, S99E, S99D, S99G, S99M, S99N, S99R, S99H, S101A, V102I, V102Y, V102N, S104A, G116V, G116R, H118D, H118N, A120S, S126L, P127Q, S128A, S154D, A156E, G157D, G157P, S158E, Y161A, R164S, Q176E, N179E, S182E, Q185N, A188P, G189E, V193M, N198D, V199I, Y203W, S206G, L211Q, L211D, N212D, N212S, M216S, A226V, K229L, Q230H, Q239R, N246K, N255W, N255D, N255E, L256E, L256D T268A and R269H. The protease variants are preferably variants of the *Bacillus lenthus* protease (Savinase®) shown in SEQ ID NO: 79 or the *Bacillus amyloliquefaciens* protease (BPN') shown in SEQ ID NO: 80. The protease variants preferably have at least 80% sequence identity to SEQ ID NO: 79 or SEQ ID NO: 80.

[0291] A protease variant comprising a substitution at one or more positions corresponding to positions 171, 173, 175, 179, or 180 of SEQ ID NO: 81, wherein said protease variant has a sequence identity of at least 75% but less than 100% to SEQ ID NO: 81. Suitable commercially available protease enzymes include those sold under the trade names Alcalase®, Duralase™, Durazym™, Relase®, Relase® Ultra, Savinase®, Savinase® Ultra, Primase®, Polarzyme®, Kannase®, Liquanase®, Liquanase® Ultra, Ovozyme®, Coronase®, Coronase® Ultra, Blaze®, Blaze Euity® 100T, Blaze Euity® 125T, Blaze Euity® 150T, Neutraser®, Ever-lase® and Esperase® (Novozymes A/S), those sold under the tradename Maxataze®, Maxacal®, Maxapem®, Purafect Ox®, Purafect OxP®, Puramax®, FN2®, FN3®, FN4®, Excellase®, Excellenz P1000™, Excellenz P1250™, Eraser®, Preferenz P100™, Purafect Prime®, Preferenz P110™, Effectenz P1000™, Purafect®™, Effectenz P1050™, Purafect Ox®™, Effectenz P2000™, Purafast®, Properase®, Opticlean® and Optimase® (Danisco/DuPont), Axapem™ (Gist-Brocades N.V.), BLAP (sequence shown in FIG. 29 of U.S. Pat. No. 5,352,604) and variants hereof (Henkel AG) and KAP (*Bacillus alkalophilus* subtilisin) from Kao.

Peroxidases/Oxidases

[0292] Suitable peroxidases/oxidases include those of plant, bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Examples of useful peroxidases include peroxidases from *Coprinus*, e.g., from *C. cinereus*, and variants thereof as those described in WO 93/24618, WO 95/10602, and WO 98/15257. Commercially available peroxidases include Guardzyme™ (Novozymes A/S).

Lipases and Cutinases:

[0293] Suitable lipases and cutinases include those of bacterial or fungal origin. Chemically modified or protein engineered mutant enzymes are included. Examples include lipase from *Thermomyces*, e.g., from *T. lanuginosus* (previously named *Humicola lanuginosa*) as described in EP 258068 and EP 305216, cutinase from *Humicola*, e.g., *H. insolens* (WO 96/13580), lipase from strains of *Pseudomonas* (some of these now renamed to *Burkholderia*), e.g., *P. alcaligenes* or *P. pseudoalcaligenes* (EP 218272), *P. cepacia* (EP 331376), *P. sp.* strain SD705 (WO 95/06720 & WO

96/27002), *P. wisconsinensis* (WO 96/12012), GDSL-type *Streptomyces* lipases (WO 2010/065455), cutinase from *Magnaporthe grisea* (WO 2010/107560), cutinase from *Pseudomonas mendocina* (U.S. Pat. No. 5,389,536), lipase from *Thermobifida fusca* (WO 2011/084412), *Geobacillus stearothermophilus* lipase (WO 2011/084417), lipase from *Bacillus subtilis* (WO 2011/084599), and lipase from *Streptomyces griseus* (WO 2011/150157) and *S. pristinaespiralis* (WO 2012/137147).

[0294] Other examples are lipase variants such as those described in EP 407225, WO 92/05249, WO 94/01541, WO 94/25578, WO 95/14783, WO 95/30744, WO 95/35381, WO 95/22615, WO 96/00292, WO 97/04079, WO 97/07202, WO 00/34450, WO 00/60063, WO 01/92502, WO 2007/87508 and WO 2009/109500.

[0295] Preferred commercial lipase products include Lipolase™, Lipex™; Lipolex™ and Lipoclean™ (Novozymes A/S), Lumafast (originally from Genencor) and Lipomax (originally from Gist-Brocades).

[0296] Still other examples are lipases sometimes referred to as acyltransferases or perhydrolases, e.g., acyltransferases with homology to *Candida antarctica* lipase A (WO 2010/111143), acyltransferase from *Mycobacterium smegmatis* (WO 2005/056782), perhydrolases from the CE 7 family (WO 2009/067279), and variants of the *M. smegmatis* perhydrolase in particular the S54V variant used in the commercial product Gentle Power Bleach from Huntsman Textile Effects Pte Ltd (WO 2010/100028).

Peroxidases/Oxidases

[0297] A peroxidase according to the invention is a peroxidase enzyme comprised by the enzyme classification EC 1.11.1.7, as set out by the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology (IUBMB), or any fragment derived therefrom, exhibiting peroxidase activity.

[0298] Suitable peroxidases include those of plant, bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Examples of useful peroxidases include peroxidases from *Coprinopsis*, e.g., from *C. cinerea* (EP 179486), and variants thereof as those described in WO 93/24618, WO 95/10602, and WO 98/15257.

[0299] A suitable peroxidase includes a haloperoxidase enzyme, such as chloroperoxidase, bromoperoxidase and compounds exhibiting chloroperoxidase or bromoperoxidase activity. Haloperoxidases are classified according to their specificity for halide ions. Chloroperoxidases (E.C. 1.11.1.10) catalyze formation of hypochlorite from chloride ions. Preferably, the haloperoxidase is a vanadium haloperoxidase, i.e., a vanadate-containing haloperoxidase. Haloperoxidases have been isolated from many different fungi, in particular from the fungus group dematiaceous hyphomycetes, such as *Caldariomyces*, e.g., *C. fumago*, *Alternaria*, *Curvularia*, e.g., *C. verruculosa* and *C. inaequalis*, *Drechslera*, *Ulocladium* and *Botrytis*.

[0300] Haloperoxidases have also been isolated from bacteria such as *Pseudomonas*, e.g., *P. pyrrocinia* and *Streptomyces*, e.g., *S. aureofaciens*.

[0301] A suitable oxidase includes, in particular, any laccase enzyme comprised by the enzyme classification EC 1.10.3.2, or any fragment derived therefrom exhibiting laccase activity, or a compound exhibiting a similar activity, such as a catechol oxidase (EC 1.10.3.1), an o-aminophenol oxidase (EC 1.10.3.4), or a bilirubin oxidase (EC 1.3.3.5).

Preferred laccase enzymes are enzymes of microbial origin. The enzymes may be derived from plants, bacteria or fungi (including filamentous fungi and yeasts). Suitable examples from fungi include a laccase derivable from a strain of *Aspergillus*, *Neurospora*, e.g., *N. crassa*, *Podospora*, *Botryotis*, *Collybia*, *Fomes*, *Lentinus*, *Pleurotus*, *Trametes*, e.g., *T. villosa* and *T. versicolor*, *Rhizoctonia*, e.g., *R. solani*, *Coprinopsis*, e.g., *C. cinerea*, *C. comatus*, *C. friesii*, and *C. plicatilis*, *Psathyrella*, e.g., *P. condelleana*, *Panaeolus*, e.g., *P. papilionaceus*, *Myceliophthora*, e.g., *M. thermophila*, *Schytalidium*, e.g., *S. thermophilum*, *Polyporus*, e.g., *P. pinsitus*, *Phlebia*, e.g., *P. radiata* (WO 92/01046), or *Coriolus*, e.g., *C. hirsutus* (JP 2238885). Suitable examples from bacteria include a laccase derivable from a strain of *Bacillus*. A laccase derived from *Coprinopsis* or *Myceliophthora* is preferred; in particular, a laccase derived from *Coprinopsis cinerea*, as disclosed in WO 97/08325; or from *Myceliophthora thermophila*, as disclosed in WO 95/33836.

Dispersants

[0302] The cleaning compositions of the present invention can also contain dispersants. In particular, powdered detergents may comprise dispersants. Suitable water-soluble organic materials include the homo- or co-polymeric acids or their salts, in which the polycarboxylic acid comprises at least two carboxyl radicals separated from each other by not more than two carbon atoms. Suitable dispersants are for example described in Powdered Detergents, Surfactant science series volume 71, Marcel Dekker, Inc.

Dye Transfer Inhibiting Agents

[0303] The cleaning compositions of the present invention may also include one or more dye transfer inhibiting agents. Suitable polymeric dye transfer inhibiting agents include, but are not limited to, polyvinylpyrrolidone polymers, polyamine N-oxide polymers, copolymers of N-vinylpyrrolidone and N-vinylimidazole, polyvinyloxazolidones and polyvinylimidazoles or mixtures thereof. When present in a subject composition, the dye transfer inhibiting agents may be present at levels from about 0.0001% to about 10%, from about 0.01% to about 5% or even from about 0.1% to about 3% by weight of the composition.

Fluorescent Whitening Agent

[0304] The cleaning compositions of the present invention will preferably also contain additional components that may tint articles being cleaned, such as fluorescent whitening agent or optical brighteners. Where present the brightener is preferably at a level of about 0.01% to about 0.5%. Any fluorescent whitening agent suitable for use in a laundry detergent composition may be used in the composition of the present invention. The most commonly used fluorescent whitening agents are those belonging to the classes of diaminostilbene-sulfonic acid derivatives, diarylpyrazoline derivatives and bisphenyl-distyryl derivatives. Examples of the diaminostilbene-sulfonic acid derivative type of fluorescent whitening agents include the sodium salts of: 4,4'-bis-(2-diethanolamino-4-anilino-s-triazin-6-ylamino) stilbene-2,2'-disulfonate, 4,4'-bis-(2,4-dianilino-s-triazin-6-ylamino) stilbene-2,2'-disulfonate, 4,4'-bis-(2-anilino-4-(N-methyl-N-2-hydroxy-ethylamino)-s-triazin-6-ylamino) stilbene-2,2'-disulfonate, 4,4'-bis-(4-phenyl-1,2,3-triazol-2-yl)stilbene-2,2'-disulfonate and sodium 5-(2H-naphtho[1,2-d][1,2,3]

triazol-2-yl)-2-[(E)-2-phenylvinyl]benzenesulfonate. Preferred fluorescent whitening agents are Tinopal DMS and Tinopal CBS available from Ciba-Geigy AG, Basel, Switzerland. Tinopal DMS is the disodium salt of 4,4'-bis-(2-morpholino-4-anilino-s-triazin-6-ylamino) stilbene-2,2'-disulfonate. Tinopal CBS is the disodium salt of 2,2'-bis(phenyl-styryl)-disulfonate. Also preferred are fluorescent whitening agents is the commercially available Parawhite KX, supplied by Paramount Minerals and Chemicals, Mumbai, India. Other fluorescers suitable for use in the invention include the 1,3-diaryl pyrazolines and the 7-alkylaminocoumarins. Suitable fluorescent brightener levels include lower levels of from about 0.01, from 0.05, from about 0.1 or even from about 0.2 wt. % to upper levels of 0.5 or even 0.75 wt. %.

Soil Release Polymers

[0305] The cleaning compositions of the present invention may also include one or more soil release polymers which aid the removal of soils from fabrics such as cotton and polyester based fabrics, in particular, the removal of hydrophobic soils from polyester based fabrics. The soil release polymers may for example be nonionic or anionic terephthalate based polymers, polyvinyl caprolactam and related copolymers, vinyl graft copolymers, polyester polyamides see for example Chapter 7 in Powdered Detergents, Surfactant science series volume 71, Marcel Dekker, Inc. Another type of soil release polymers is amphiphilic alkoxyLATED grease cleaning polymers comprising a core structure and a plurality of alkoxylate groups attached to that core structure. The core structure may comprise a polyalkyleneimine structure or a polyalkanolamine structure as described in detail in WO 2009/087523 (hereby incorporated by reference). Furthermore, random graft co-polymers are suitable soil release polymers. Suitable graft co-polymers are described in more detail in WO 2007/138054, WO 2006/108856 and WO 2006/113314 (hereby incorporated by reference). Suitable polyethylene glycol polymers include random graft copolymers comprising: (i) hydrophilic backbone comprising polyethylene glycol; and (ii) side chain(s) selected from the group consisting of: C4-C25 alkyl group, polypropylene, polybutylene, vinyl ester of a saturated C1-C6 mono-carboxylic acid, C1-C6 alkyl ester of acrylic or methacrylic acid, and mixtures thereof. Suitable polyethylene glycol polymers have a polyethylene glycol backbone with random grafted polyvinyl acetate side chains. The average molecular weight of the polyethylene glycol backbone can be in the range of from 2,000 Da to 20,000 Da, or from 4,000 Da to 8,000 Da. The molecular weight ratio of the polyethylene glycol backbone to the polyvinyl acetate side chains can be in the range of from 1:1 to 1:5, or from 1:1.2 to 1:2. The average number of graft sites per ethylene oxide units can be less than 1, or less than 0.8, the average number of graft sites per ethylene oxide units can be in the range of from 0.5 to 0.9, or the average number of graft sites per ethylene oxide units can be in the range of from 0.1 to 0.5, or from 0.2 to 0.4. A suitable polyethylene glycol polymer is Sokalan HP22. Other soil release polymers are substituted polysaccharide structures especially substituted cellulosic structures such as modified cellulose derivatives such as those described in EP 1867808 or WO 2003/040279 (both are hereby incorporated by reference). Suitable cellulosic polymers include cellulose, cellulose ethers, cellulose esters, cellulose amides and mixtures thereof. Suitable cellulosic

polymers include anionically modified cellulose, nonionically modified cellulose, cationically modified cellulose, zwitterionically modified cellulose, and mixtures thereof. Suitable cellulosic polymers include methyl cellulose, carboxy methyl cellulose, ethyl cellulose, hydroxyl ethyl cellulose, hydroxyl propyl methyl cellulose, ester carboxy methyl cellulose, and mixtures thereof.

Anti-Redeposition Agents

[0306] The cleaning compositions of the present invention may also include one or more anti-redeposition agents such as carboxymethylcellulose (CMC), polyvinyl alcohol (PVA), polyvinylpyrrolidone (PVP), polyoxyethylene and/or polyethyleneglycol (PEG), homopolymers of acrylic acid, copolymers of acrylic acid and maleic acid, and ethoxylated polyethyleneimines. The cellulose based polymers described under soil release polymers above may also function as anti-redeposition agents.

Rheology Modifiers

[0307] The cleaning compositions of the present invention may also include one or more rheology modifiers, structurants or thickeners, as distinct from viscosity reducing agents. The rheology modifiers are selected from the group consisting of non-polymeric crystalline, hydroxy-functional materials, polymeric rheology modifiers which impart shear thinning characteristics to the aqueous liquid matrix of a liquid detergent composition. The rheology and viscosity of the detergent can be modified and adjusted by methods known in the art, for example as shown in EP 2169040.

[0308] Other suitable cleaning composition components include, but are not limited to, anti-shrink agents, anti-wrinkling agents, bactericides, binders, carriers, dyes, enzyme stabilizers, fabric softeners, fillers, foam regulators, hydrotropes, perfumes, pigments, sod suppressors, solvents, and structurants for liquid detergents and/or structure elasticizing agents.

Formulation of Detergent Products

[0309] The cleaning composition of the invention may be in any convenient form, e.g., a bar, a homogenous tablet, a tablet having two or more layers, a pouch having one or more compartments, a regular or compact powder, a granule, a paste, a gel, or a regular, compact or concentrated liquid.

[0310] Pouches can be configured as single or multicompartments. It can be of any form, shape and material which is suitable for hold the composition, e.g., without allowing the release of the composition to release of the composition from the pouch prior to water contact. The pouch is made from water soluble film which encloses an inner volume. Said inner volume can be divided into compartments of the pouch. Preferred films are polymeric materials preferably polymers which are formed into a film or sheet. Preferred polymers, copolymers or derivates thereof are selected polyacrylates, and water soluble acrylate copolymers, methyl cellulose, carboxy methyl cellulose, sodium dextrin, ethyl cellulose, hydroxyethyl cellulose, hydroxypropyl methyl cellulose, malto dextrin, poly methacrylates, most preferably polyvinyl alcohol copolymers and, hydroxypropyl methyl cellulose (HPMC). Preferably the level of polymer in the film for example PVA is at least about 60%. Preferred average molecular weight will typically be about 20,000 to about 150,000. Films can also be of blended compositions

comprising hydrolytically degradable and water soluble polymer blends such as polylactide and polyvinyl alcohol (known under the Trade reference M8630 as sold by Mono-Sol LLC, Indiana, USA) plus plasticisers like glycerol, ethylene glycerol, propylene glycol, sorbitol and mixtures thereof. The pouches can comprise a solid laundry cleaning composition or part components and/or a liquid cleaning composition or part components separated by the water-soluble film. The compartment for liquid components can be different in composition than compartments containing solids: US 2009/0011970.

[0311] Detergent ingredients can be separated physically from each other by compartments in water dissolvable pouches or in different layers of tablets. Thereby negative storage interaction between components can be avoided. Different dissolution profiles of each of the compartments can also give rise to delayed dissolution of selected components in the wash solution.

[0312] A liquid or gel detergent, which is not unit dosed, may be aqueous, typically containing at least 20% by weight and up to 95% water, such as up to about 70% water, up to about 65% water, up to about 55% water, up to about 45% water, up to about 35% water. Other types of liquids, including without limitation, alkanols, amines, diols, ethers and polyols may be included in an aqueous liquid or gel. An aqueous liquid or gel detergent may contain from 0-30% organic solvent. A liquid or gel detergent may be non-aqueous.

Granular Detergent Formulations

[0313] Non-dusting granulates may be produced, e.g., as disclosed in U.S. Pat. Nos. 4,106,991 and 4,661,452 and may optionally be coated by methods known in the art. Examples of waxy coating materials are poly(ethylene oxide) products (polyethyleneglycol, PEG) with mean molar weights of 1000 to 20000; ethoxylated nonylphenols having from 16 to 50 ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols; fatty acids; and mono- and di- and triglycerides of fatty acids. Examples of film-forming coating materials suitable for application by fluid bed techniques are given in GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. Protected enzymes may be prepared according to the method disclosed in EP 238216.

[0314] The DNase may be formulated as a granule for example as a co-granule that combines one or more enzymes. Each enzyme will then be present in more granules securing a more uniform distribution of enzymes in the detergent. This also reduces the physical segregation of different enzymes due to different particle sizes. Methods for producing multi-enzyme co-granulate for the detergent industry is disclosed in the IP.com disclosure IPCOM000200739D.

[0315] Another example of formulation of enzymes by the use of co-granulates are disclosed in WO 2013/188331, which relates to a detergent composition comprising (a) a multi-enzyme co-granule; (b) less than 10 wt. % zeolite (anhydrous basis); and (c) less than 10 wt. % phosphate salt (anhydrous basis), wherein said enzyme co-granule comprises from 10 to 98 wt. % moisture sink component and the

composition additionally comprises from 20 to 80 wt. % detergent moisture sink component. WO 2013/188331 also relates to a method of treating and/or cleaning a surface, preferably a fabric surface comprising the steps of (i) contacting said surface with the detergent composition as claimed and described herein in aqueous wash liquor, (ii) rinsing and/or drying the surface.

[0316] An embodiment of the invention relates to an enzyme granule/particle comprising the DNase and at least one carbohydراse and a cleaning component, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase. The granule is composed of a core, and optionally one or more coatings (outer layers) surrounding the core. Typically, the granule/particle size, measured as equivalent spherical diameter (volume based average particle size), of the granule is 20-2000 µm, particularly 50-1500 µm, 100-1500 µm or 250-1200 µm. The core may include additional materials such as fillers, fiber materials (cellulose or synthetic fibers), stabilizing agents, solubilising agents, suspension agents, viscosity regulating agents, light spheres, plasticizers, salts, lubricants and fragrances. The core may include binders, such as synthetic polymer, wax, fat, or carbohydrate. The core may comprise a salt of a multivalent cation, a reducing agent, an antioxidant, a peroxide decomposing catalyst and/or an acidic buffer component, typically as a homogenous blend. The core may consist of an inert particle with the enzyme absorbed into it, or applied onto the surface, e.g., by fluid bed coating. The core may have a diameter of 20-2000 µm, particularly 50-1500 µm, 100-1500 µm or 250-1200 µm. The core can be prepared by granulating a blend of the ingredients, e.g., by a method comprising granulation techniques such as crystallization, precipitation, pan-coating, fluid bed coating, fluid bed agglomeration, rotary atomization, extrusion, prilling, spheronization, size reduction methods, drum granulation, and/or high shear granulation.

[0317] Methods for preparing the core can be found in Handbook of Powder Technology; Particle size enlargement by C. E. Capes; Volume 1; 1980; Elsevier.

[0318] The core of the enzyme granule/particle may be surrounded by at least one coating, e.g., to improve the storage stability, to reduce dust formation during handling, or for coloring the granule. The optional coating(s) may include a salt coating, or other suitable coating materials, such as polyethylene glycol (PEG), methyl hydroxy-propyl cellulose (MHPC) and polyvinyl alcohol (PVA). Examples of enzyme granules with multiple coatings are shown in WO 93/07263 and WO 97/23606. The coating may be applied in an amount of at least 0.1% by weight of the core, e.g., at least 0.5%, 1% or 5%. The amount may be at most 100%, 70%, 50%, 40% or 30%. The coating is preferably at least 0.1 µm thick, particularly at least 0.5 µm, at least 1 µm or at least 5 µm. In a one embodiment, the thickness of the coating is below 100 µm. In another embodiment, the thickness of the coating is below 60 µm. In an even more particular embodiment the total thickness of the coating is below 40 µm. The coating should encapsulate the core unit by forming a substantially continuous layer. A substantially continuous layer is to be understood as a coating having few or no holes, so that the core unit it is encapsulating/enclosing has few or none uncoated areas. The layer or coating should be homogeneous in thickness. The coating can further contain other materials as known in the art, e.g., fillers, antisticking agents, pigments, dyes, plasticizers and/or binders, such as

titanium dioxide, kaolin, calcium carbonate or talc. A salt coating may comprise at least 60% by weight w/w of a salt, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95% or at least 99% by weight w/w. The salt may be added from a salt solution where the salt is completely dissolved or from a salt suspension wherein the fine particles is less than 50 µm, such as less than 10 µm or less than 5 µm. The salt coating may comprise a single salt or a mixture of two or more salts. The salt may be water soluble, and may have a solubility of at least 0.1 grams in 100 g of water at 20° C., preferably at least 0.5 g per 100 g water, e.g., at least 1 g per 100 g water, e.g., at least 5 g per 100 g water. The salt may be an inorganic salt, e.g., salts of sulfate, sulfite, phosphate, phosphonate, nitrate, chloride or carbonate or salts of simple organic acids (less than 10 carbon atoms, e.g., 6 or less carbon atoms) such as citrate, malonate or acetate. Examples of cations in these salts are alkali or earth alkali metal ions, the ammonium ion or metal ions of the first transition series, such as sodium, potassium, magnesium, calcium, zinc or aluminium. Examples of anions include chloride, bromide, iodide, sulfate, sulfite, bisulfite, thiosulfate, phosphate, monobasic phosphate, dibasic phosphate, hypophosphate, dihydrogen pyrophosphate, tetraborate, borate, carbonate, bicarbonate, metasilicate, citrate, malate, maleate, malonate, succinate, lactate, formate, acetate, butyrate, propionate, benzoate, tartrate, ascorbate or gluconate. In particular, alkali- or earth alkali metal salts of sulfate, sulfite, phosphate, phosphonate, nitrate, chloride or carbonate or salts of simple organic acids such as citrate, malonate or acetate may be used. The salt in the coating may have a constant humidity at 20° C. above 60%, particularly above 70%, above 80% or above 85%, or it may be another hydrate form of such a salt (e.g., anhydrate). The salt coating may be as described in WO 00/01793 or WO 2006/034710. Specific examples of suitable salts are NaCl ($\text{CH}_{20^\circ \text{ C.}}=76\%$), Na_2CO_3 ($\text{CH}_{20^\circ \text{ C.}}=92\%$), NaNO_3 ($\text{CH}_{20^\circ \text{ C.}}=73\%$), Na_2HPO_4 ($\text{CH}_{20^\circ \text{ C.}}=95\%$), Na_3PO_4 ($\text{CH}_{25^\circ \text{ C.}}=92\%$), NH_4Cl ($\text{CH}_{20^\circ \text{ C.}}=79.5\%$), $(\text{NH}_4)_2\text{HPO}_4$ ($\text{CH}_{20^\circ \text{ C.}}=93.0\%$), $\text{NH}_4\text{H}_2\text{PO}_4$ ($\text{CH}_{20^\circ \text{ C.}}=93.1\%$), $(\text{NH}_4)_2\text{SO}_4$ ($\text{CH}_{20^\circ \text{ C.}}=81.1\%$), KCl ($\text{CH}_{20^\circ \text{ C.}}=85\%$), K_2HPO_4 ($\text{CH}_{20^\circ \text{ C.}}=92\%$), KH_2PO_4 ($\text{CH}_{20^\circ \text{ C.}}=96.5\%$), KNO_3 ($\text{CH}_{20^\circ \text{ C.}}=93.5\%$), Na_2SO_4 ($\text{CH}_{20^\circ \text{ C.}}=93\%$), K_2SO_4 ($\text{CH}_{20^\circ \text{ C.}}=98\%$), KHSO_4 ($\text{CH}_{20^\circ \text{ C.}}=86\%$), MgSO_4 ($\text{CH}_{20^\circ \text{ C.}}=90\%$), ZnSO_4 ($\text{CH}_{20^\circ \text{ C.}}=90\%$) and sodium citrate ($\text{CH}_{25^\circ \text{ C.}}=86\%$). Other examples include NaH_2PO_4 , $(\text{NH}_4)\text{H}_2\text{PO}_4$, CuSO_4 , $\text{Mg}(\text{NO}_3)_2$ and magnesium acetate. The salt may be in anhydrous form, or it may be a hydrated salt, i.e., a crystalline salt hydrate with bound water(s) of crystallization, such as described in WO 99/32595. Specific examples include anhydrous sodium sulfate (Na_2SO_4), anhydrous magnesium sulfate (MgSO_4), magnesium sulfate heptahydrate ($\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$), zinc sulfate heptahydrate ($\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$), sodium phosphate dibasic heptahydrate ($\text{Na}_2\text{HPO}_4 \cdot 7\text{H}_2\text{O}$), magnesium nitrate hexahydrate ($\text{Mg}(\text{NO}_3)_2(6\text{H}_2\text{O})$), sodium citrate dihydrate and magnesium acetate tetrahydrate. Preferably, the salt is applied as a solution of the salt, e.g., using a fluid bed.

[0319] One embodiment of the present invention provides a granule, which comprises:

[0320] (a) a core comprising a DNase and a carbohydراse, wherein the carbohydراse is a cellulase, an amylase, a mannanase or a xylanase, and

[0321] (b) optionally a coating consisting of one or more layer(s) surrounding the core.

[0378] (b) optionally a coating consisting of one or more layer(s) surrounding the core.

[0379] One embodiment of the invention relates to a granule, which comprises:

[0380] (a) a core comprising a DNase and a xylanase, wherein the xylanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68, and

[0381] (b) optionally a coating consisting of one or more layer(s) surrounding the core.

Uses

[0382] The present invention is also directed to methods for using the compositions thereof. Laundry/textile/fabric (House hold laundry washing, Industrial laundry washing). Hard surface cleaning (ADW, car wash, Industrial surface). The compositions of the invention comprise a blend of DNase and carbohydrase, selected from a cellulase, an amylase, a mannanase or a xylanase and effectively reduce or remove organic components, such as protein and DNA from surfaces such as textiles and hard surfaces, e.g., dishes.

[0383] The compositions of the invention comprise a blend of DNase and carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase, and effectively reduce or remove organic components, such as mannan, starch, cellulose, xyloglucan and DNA from surfaces such as textiles and hard surfaces, e.g., dishes. One embodiment of the invention relates to the use of a cleaning composition comprising a DNase, a carbohydrase, selected from a cellulase, an amylase, a mannanase or a xylanase and at least one cleaning component for reduction or removal of components of biofilm, such as DNA and at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase, of an item, wherein the item is a textile or a hard surface.

[0384] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase for deep cleaning of an item, wherein the item is a textile or a surface.

[0385] One embodiment of the invention relates to the use of a composition comprising a DNase and a carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase for reduction or removal of biofilm and/or compounds such as mannan, starch, cellulose, xyloglucan and DNA of an item. One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and a carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase for reduction or removal of biofilm and/or compounds such as mannan, starch, cellulose, xyloglucan and DNA of an item such as textile. One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and a carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a

xylanase for deep cleaning when the cleaning composition is applied in, e.g., laundry process.

[0386] One embodiment of the invention relates to the use of a composition comprising a DNase and carbohydrase, selected from a cellulase, an amylase, a mannanase or a xylanase for reduction of redeposition or reduction of malodor. One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and carbohydrase, selected from a cellulase, an amylase, a mannanase or a xylanase for reduction of redeposition or reduction of malodor.

[0387] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and carbohydrase, selected from a cellulase, an amylase, a mannanase or a xylanase for reduction of redeposition or reduction of malodor when the cleaning composition is applied in, e.g., laundry process. One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and carbohydrase, selected from a cellulase, an amylase, a mannanase or a xylanase for reduction of redeposition or reduction of malodor on an item, e.g., textile. In one embodiment, the composition is an anti-redeposition composition.

[0388] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and a cellulase for deep cleaning of an item or reduction of redeposition or malodor, wherein the cellulase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 86.

[0389] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and an amylase for deep cleaning of an item or reduction of redeposition or malodor, wherein the amylase selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0390] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and a mannanase for deep cleaning of an item or reduction of rede-

[0403] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and mannanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 65.

[0404] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and mannanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 66.

[0405] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and mannanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 67.

[0406] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and mannanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0407] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and xylanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the xylanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 13.

[0408] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and xylanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the xylanase is selected from a polypep-

tide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 65.

[0409] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and xylanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the xylanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 66.

[0410] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and xylanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the xylanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 67.

[0411] One embodiment of the invention relates to the use of a cleaning composition comprising a DNase and xylanase for deep cleaning of an item or reduction of redeposition or malodor, wherein the xylanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the is DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

[0412] The invention further relates to a method of deep cleaning of an item, wherein the item may be textile or hard surface preferably is a textile.

[0413] One embodiment of the invention relates to a method of deep cleaning on an item, comprising the steps of:

[0414] a) contacting the item with a cleaning composition according to the invention; and

[0415] b) optionally rinsing the item, wherein the item is preferably a textile.

[0416] One embodiment of the invention relates to a method of deep cleaning on an item, comprising the steps of:

[0417] a) contacting the item with a solution comprising an enzyme mixture comprising a DNase, a carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase; and a cleaning component, wherein the cleaning component is selected from 0.1 to 50 wt. % of at least one a surfactant; 0.5 to 30 wt. % of at least one builder; and 0.01 to 20 wt. % of at least one bleach component; and

or 100% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 91 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 13, SEQ ID NO: 65, SEQ ID NO: 66, SEQ ID NO: 67 or SEQ ID NO: 68.

[0437] One embodiment of the invention relates to a method of deep cleaning on an item, comprising the steps of:

[0438] a) contacting the item with a solution comprising an enzyme mixture comprising a DNase, mannanase; and a cleaning component, wherein the cleaning component is selected from 0.1 to 50 wt. % of at least one a surfactant; 0.5 to 30 wt. % of at least one builder; and 0.01 to 20 wt. % of at least one bleach component; and

[0439] b) optionally rinsing the item, wherein the item is preferably a textile;

wherein the mannanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 82 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 13, SEQ ID NO: 65, SEQ ID NO: 66, SEQ ID NO: 67 or SEQ ID NO: 68.

[0440] One embodiment of the invention relates to a method of deep cleaning on an item, comprising the steps of:

[0441] a) contacting the item with a solution comprising an enzyme mixture comprising a DNase, xylanase; and a cleaning component, wherein the cleaning component is selected from 0.1 to 50 wt. % of at least one a surfactant; 0.5 to 30 wt. % of at least one builder; and 0.01 to 20 wt. % of at least one bleach component; and

[0442] b) optionally rinsing the item, wherein the item is preferably a textile;

wherein the xylanase is selected from a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the polypeptide shown in SEQ ID NO: 87 and wherein the DNase has at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 98% or 100% sequence identity to the amino acid sequence shown in SEQ ID NO: 13, SEQ ID NO: 65, SEQ ID NO: 66, SEQ ID NO: 67 or SEQ ID NO: 68.

[0443] The invention is further described in the following paragraphs

[0444] Paragraph 1. A cleaning composition comprising at least 0.001 ppm DNase and at least 0.001 ppm carbohydrase and a cleaning component, wherein the cleaning component is selected from

[0445] a. 0.1 to 15 wt. % of at least one a surfactant;
 [0446] b. 0.5 to 20 wt. % of at least one builder; and
 [0447] c. 0.01 to 10 wt. % of at least one bleach component.

[0448] Paragraph 2. The cleaning composition according to paragraph 1, wherein the DNase comprises one or both of the motif(s) [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73), ASXNRSKG (SEQ ID NO: 74) and the carbohydrase is a cellulase.

[0449] Paragraph 3. The cleaning composition according to paragraph 1, wherein the DNase comprises one or both of the motif(s) [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73), ASXNRSKG (SEQ ID NO: 74) and the carbohydrase is a mannanase.

[0450] Paragraph 4. The cleaning composition according to paragraph 1, wherein the DNase comprises one or both of the motif(s) [D/M/L][S/T]GYSR[D/N](SEQ ID NO: 73), ASXNRSKG (SEQ ID NO: 74) and the carbohydrase is a amylase.

[0451] Paragraph 5. The cleaning composition according to any of paragraphs 1 to 4, wherein the DNase is selected from the group of polypeptides:

[0452] a) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 1,

[0453] b) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 2,

[0454] c) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 3,

[0455] d) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 4,

[0456] e) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 5,

[0457] f) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 6,

[0458] g) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 7,

[0459] h) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 8,

[0460] i) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 9,

[0461] j) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 10,

[0462] k) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 11,

[0463] l) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 12,

[0464] m) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 13,

[0465] n) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 14,

[0466] o) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 15,

[0467] p) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 16,

[0468] q) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 17,

[0469] r) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 18,

[0470] s) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 19,

[0471] t) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 20,

[0472] u) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 21,

- [0473] v) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 22,
- [0474] w) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 23,
- [0475] x) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 24, and
- [0476] y) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 25, and

wherein the carbohydrase is selected from the group consisting of:

- [0477] i. a cellulase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 86;
- [0478] ii. a xylanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 87;
- [0479] iii. a mannanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 82; and
- [0480] iv. an amylase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0481] Paragraph 6. The cleaning composition according to paragraph 1, wherein the DNase comprises one or both of the motifs [V/I]PL[S/A]NAWK (SEQ ID NO: 75) or NPQL (SEQ ID NO: 76) and the carbohydrase is a cellulase.

[0482] Paragraph 7. The cleaning composition according to paragraph 1, wherein the DNase comprises one or both of the motifs [V/I]PL[S/A]NAWK (SEQ ID NO: 75) or NPQL (SEQ ID NO: 76) and the carbohydrase is a mannanase.

[0483] Paragraph 8. The cleaning composition according to paragraph 1, wherein the DNase comprise one or both of the motifs [V/I]PL[S/A]NAWK (SEQ ID NO: 75) or NPQL (SEQ ID NO: 76) and the carbohydrase is a amylase.

[0484] Paragraph 9. The cleaning composition according to any of paragraphs 1 and 6 to 8, wherein the DNase is selected from the group of polypeptides:

- [0485] a) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 26,
- [0486] b) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 27,
- [0487] c) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 28,
- [0488] d) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 29,
- [0489] e) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 30,
- [0490] f) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 31,

[0491] g) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 32,

[0492] h) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 33,

[0493] i) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 34,

[0494] j) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 35,

[0495] k) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 36,

[0496] l) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 37,

[0497] m) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 38, and

wherein the carbohydrase is selected from the group consisting of:

[0498] i. a cellulase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 86;

[0499] ii. a xylanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 87;

[0500] iii. a mannanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 82; and

[0501] iv. an amylase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0502] Paragraph 10. The cleaning composition according to paragraph 1, wherein the DNase comprises one or both of the motifs P[Q/E]L[W/Y] or [K/H/E]NAW and the carbohydrase is a cellulase.

[0503] Paragraph 11. The cleaning composition according to paragraph 1, wherein the DNase comprise one or both of the motifs P[Q/E]L[W/Y] or [K/H/E]NAW and the carbohydrase is a mannanase.

[0504] Paragraph 12. The cleaning composition according to paragraph 1, wherein the DNase comprise one or both of the motifs P[Q/E]L[W/Y] or [K/H/E]NAW and the carbohydrase is an amylase.

[0505] Paragraph 13. The cleaning composition according to paragraph 1 or 10 to 12, wherein the DNase is selected from the group of polypeptides:

[0506] a) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 39,

[0507] b) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 40,

[0508] c) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 41,

[0509] d) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 42,

- [0510] e) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 43
- [0511] f) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 44,
- [0512] g) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 45,
- [0513] h) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 46,
- [0514] i) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 47,
- [0515] j) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 48,
- [0516] k) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 49,
- [0517] l) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 50,
- [0518] m) a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 51, and

wherein the carbohydrase is selected from the group consisting of;

- [0519] i. a cellulase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 86;
- [0520] ii. a xylanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 87;
- [0521] iii. a mannanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 82; and
- [0522] iv. an amylase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0523] Paragraph 14. The cleaning composition according to paragraph 1, wherein the DNase is selected from the group consisting of:

- [0524] a) polypeptide obtainable from *Bacillus licheniformis* having a sequence identity to the polypeptide shown in SEQ ID NO: 65 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity,
- [0525] b) polypeptide obtainable from *Bacillus subtilis* having a sequence identity to the polypeptide shown in SEQ ID NO: 66 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity,
- [0526] c) polypeptide obtainable from *Aspergillus oryzae* having a sequence identity to the polypeptide

shown in SEQ ID NO: 67 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity,

[0527] d) polypeptide obtainable from *Trichoderma harzianum* having a sequence identity to the polypeptide shown in SEQ ID NO: 68 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% and which have DNase activity, and

wherein the carbohydrase is selected from the group consisting of;

[0528] i. a cellulase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 83, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 84, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 85, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 86;

[0529] ii. a xylanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 87;

[0530] iii. a mannanase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 82; and

[0531] iv. an amylase selected from a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 88, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 89, a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 90, and a polypeptide having at least 80% sequence identity to the polypeptide shown in SEQ ID NO: 91.

[0532] Paragraph 15. The use of a composition according to any of the previous paragraphs for deep cleaning of an item, wherein the item is a textile or a surface.

[0533] Paragraph 16. A method of formulating a cleaning composition comprising adding a DNase, a carbohydrase and at least one cleaning component.

[0534] Paragraph 17. A kit intended for deep cleaning, wherein the kit comprises a solution of an enzyme mixture comprising a DNase, carbohydrase and optionally a protease.

[0535] Paragraph 18. A method of deep cleaning on an item, comprising the steps of:

[0536] a) contacting the item with a solution comprising an enzyme mixture comprising a DNase and a carbohydrase and optionally a protease; and a cleaning component, wherein the cleaning component is selected from 0.1 to 15 wt. % of at least one a surfactant; 0.5 to 20 wt. % of at least one builder; and 0.01 to 10 wt. % of at least one bleach component; and

[0537] b) optionally rinsing the item, wherein the item is preferably a textile.

Definitions

[0538] Biofilm is produced by any group of microorganisms in which cells stick to each other or stick to a surface, such as a textile, dishware or hard surface or another kind of

surface. These adherent cells are frequently embedded within a self-produced matrix of extracellular polymeric substance (EPS). Biofilm EPS is a polymeric conglomeration generally composed of extracellular DNA, proteins, and polysaccharides. Biofilms may form on living or non-living surfaces. The microbial cells growing in a biofilm are physiologically distinct from planktonic cells of the same organism, which, by contrast, are single-cells that may float or swim in a liquid medium. Bacteria living in a biofilm usually have significantly different properties from planktonic bacteria of the same species, as the dense and protected environment of the film allows them to cooperate and interact in various ways. One benefit of this environment for the microorganisms is increased resistance to detergents and antibiotics, as the dense extracellular matrix and the outer layer of cells protect the interior of the community.

[0539] On laundry biofilm producing bacteria can be found among the following species: *Acinetobacter* sp., *Aeromicrobium* sp., *Brevundimonas* sp., *Microbacterium* sp., *Micrococcus luteus*, *Pseudomonas* sp., *Staphylococcus epidermidis*, and *Stenotrophomonas* sp. On hard surfaces biofilm producing bacteria can be found among the following species: *Acinetobacter* sp., *Aeromicrobium* sp., *Brevundimonas* sp., *Microbacterium* sp., *Micrococcus luteus*, *Pseudomonas* sp., *Staphylococcus epidermidis*, *Staphylococcus aureus* and *Stenotrophomonas* sp. In one aspect, the biofilm producing strain is *Brevundimonas* sp. In one aspect, the biofilm producing strain is *Pseudomonas alcaliphila* or *Pseudomonas fluorescens*. In one aspect, the biofilm producing strain is *Staphylococcus aureus*.

[0540] By the term "deep cleaning" is meant reduction, disruption or removal of components of organic matter, e.g., biofilm, such as polysaccharides, proteins, DNA, soil or other components present in the organic matter.

[0541] Cleaning component: The cleaning component, e.g., the detergent adjunct ingredient is different to the DNase and carbohydrase. The precise nature of these additional cleaning components, e.g., adjunct components, and levels of incorporation thereof, will depend on the physical form of the composition and the nature of the operation for which it is to be used. Suitable cleaning components, e.g., adjunct materials include, but are not limited to the components described below such as surfactants, builders, flocculating aid, chelating agents, dye transfer inhibitors, enzymes, enzyme stabilizers, enzyme inhibitors, catalytic materials, bleach activators, hydrogen peroxide, sources of hydrogen peroxide, preformed peracids, polymeric agents, clay soil removal/anti-redeposition agents, brighteners, suds suppressors, dyes, perfumes, structure elasticizing agents, fabric softeners, carriers, hydrotropes, builders and co-builders, fabric huing agents, anti-foaming agents, dispersants, processing aids, and/or pigments.

[0542] Cleaning Composition: The term "cleaning composition" refers to compositions that find use in the removal of undesired compounds from items to be cleaned, such as textiles. The detergent composition may be used to, e.g., clean textiles for both household cleaning and industrial cleaning. The terms encompass any materials/compounds selected for the particular type of cleaning composition desired and the form of the product (e.g., liquid, gel, powder, granulate, paste, or spray compositions) and includes, but is not limited to, detergent compositions (e.g., liquid and/or solid laundry detergents and fine fabric detergents; fabric fresheners; fabric softeners; and textile and laundry pre-

spotters/pretreatment). In addition to containing the enzyme blend of the invention, the detergent formulation may contain one or more additional enzymes (such as proteases, lipases, cutinases, endoglucanases, xyloglucanases, pectinases, pectin lyases, xanthanases, peroxidases, haloperoxigenases and catalases or any mixture thereof), and/or detergent adjunct ingredients such as surfactants, builders, chelators or chelating agents, bleach system or bleach components, polymers, fabric conditioners, foam boosters, suds suppressors, dyes, perfume, tannish inhibitors, optical brighteners, bactericides, fungicides, soil suspending agents, anti-corrosion agents, enzyme inhibitors or stabilizers, enzyme activators, transferase(s), hydrolytic enzymes, oxido reductases, bluing agents and fluorescent dyes, anti-oxidants, and solubilizers.

[0543] The term "enzyme detergency benefit" is defined herein as the advantageous effect an enzyme may add to a detergent compared to the same detergent without the enzyme. Important detergency benefits which can be provided by enzymes are stain removal with no or very little visible soils after washing and/or cleaning, prevention or reduction of redeposition of soils released in the washing process (an effect that also is termed anti-redeposition), restoring fully or partly the whiteness of textiles which originally were white but after repeated use and wash have obtained a greyish or yellowish appearance (an effect that also is termed whitening). Textile care benefits, which are not directly related to catalytic stain removal or prevention of redeposition of soils, are also important for enzyme detergency benefits. Examples of such textile care benefits are prevention or reduction of dye transfer from one fabric to another fabric or another part of the same fabric (an effect that is also termed dye transfer inhibition or anti-backstaining), removal of protruding or broken fibers from a fabric surface to decrease pilling tendencies or remove already existing pills or fuzz (an effect that also is termed anti-pilling), improvement of the fabric-softness, colour clarification of the fabric and removal of particulate soils which are trapped in the fibers of the fabric or garment. Enzymatic bleaching is a further enzyme detergency benefit where the catalytic activity generally is used to catalyze the formation of bleaching components such as hydrogen peroxide or other peroxides. Textile care benefits, which are not directly related to catalytic stain removal or prevention of redeposition of soils, are also important for enzyme detergency benefits. Examples of such textile care benefits are prevention or reduction of dye transfer from one textile to another textile or another part of the same textile (an effect that is also termed dye transfer inhibition or anti-backstaining), removal of protruding or broken fibers from a textile surface to decrease pilling tendencies or remove already existing pills or fuzz (an effect that also is termed anti-pilling), improvement of the textile-softness, colour clarification of the textile and removal of particulate soils which are trapped in the fibers of the textile. Enzymatic bleaching is a further enzyme detergency benefit where the catalytic activity generally is used to catalyze the formation of bleaching component such as hydrogen peroxide or other peroxides or other bleaching species."

[0544] The term "hard surface cleaning" is defined herein as cleaning of hard surfaces wherein hard surfaces may include floors, tables, walls, roofs etc. as well as surfaces of hard objects such as cars (car wash) and dishes (dish wash). Dish washing includes but are not limited to cleaning of

plates, cups, glasses, bowls, cutlery such as spoons, knives, forks, serving utensils, ceramics, plastics, metals, china, glass and acrylics.

[0545] The term “wash performance” is used as an enzyme’s ability to remove stains present on the object to be cleaned during, e.g., wash or hard surface cleaning.

[0546] The term “whiteness” is defined herein as a greying, yellowing of a textile. Loss of whiteness may be due to removal of optical brighteners/hueing agents. Greying and yellowing can be due to soil redeposition, body soils, coloring from, e.g., iron and copper ions or dye transfer. Whiteness might include one or several issues from the list below: colourant or dye effects; incomplete stain removal (e.g., body soils, sebum etc.); redeposition (greying, yellowing or other discolourations of the object) (removed soils reassociate with other parts of textile, soiled or unsoiled); chemical changes in textile during application; and clarification or brightening of colors.

[0547] The term “laundering” relates to both household laundering and industrial laundering and means the process of treating textiles with a solution containing a cleaning or detergent composition of the present invention. The laundering process can for example be carried out using, e.g., a household or an industrial washing machine or can be carried out by hand.

[0548] The term “malodor” means an odor which is not desired on clean items. The cleaned item should smell fresh and clean without malodors adhered to the item. One example of malodor is compounds with an unpleasant smell, which may be produced by microorganisms. Another example is unpleasant smells can be sweat or body odor adhered to an item which has been in contact with human or animal. Another example of malodor can be the odor from spices, which sticks to items for example curry or other exotic spices which smells strongly.

[0549] The term “mature polypeptide” means a polypeptide in its final form following translation and any post-translational modifications, such as N-terminal processing, C-terminal truncation, glycosylation, phosphorylation, etc.

[0550] The term “textile” means any textile material including yarns, yarn intermediates, fibers, non-woven materials, natural materials, synthetic materials, and any other textile material, fabrics made of these materials and products made from fabrics (e.g., garments and other articles). The textile or fabric may be in the form of knits, wovens, denims, non-wovens, felts, yarns, and towelling. The textile may be cellulose based such as natural cellulosics, including cotton, flax/linen, jute, ramie, sisal or coir or manmade cellulosics (e.g., originating from wood pulp) including viscose/rayon, cellulose acetate fibers (tricell), lyocell or blends thereof. The textile or fabric may also be non-cellulose based such as natural polyamides including wool, camel, cashmere, mohair, rabbit and silk or synthetic polymers such as nylon, aramid, polyester, acrylic, polypropylene and spandex/elastane, or blends thereof as well as blends of cellulose based and non-cellulose based fibers. Examples of blends are blends of cotton and/or rayon/viscose with one or more companion material such as wool, synthetic fiber (e.g., polyamide fiber, acrylic fiber, polyester fiber, polyvinyl chloride fiber, polyurethane fiber, polyurea fiber, aramid fiber), and/or cellulose-containing fiber (e.g., rayon/viscose, ramie, flax/linen, jute, cellulose acetate fiber, lyocell). Fabric may be conventional washable laundry, for

example stained household laundry. When the term fabric or garment is used it is intended to include the broader term textiles as well.

[0551] The term “variant” means a polypeptide having the activity of the parent or precursor polypeptide and comprising an alteration, i.e., a substitution, insertion, and/or deletion, at one or more (e.g., several) positions compared to the precursor or parent polypeptide. A substitution means replacement of the amino acid occupying a position with a different amino acid; a deletion means removal of the amino acid occupying a position; and an insertion means adding an amino acid adjacent to and immediately following the amino acid occupying a position.

[0552] Sequence identity: The relatedness between two amino acid sequences or between two nucleotide sequences is described by the parameter “sequence identity”. For purposes of the present invention, the sequence identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, J. Mol. Biol. 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, Trends Genet. 16: 276-277), preferably version 6.6.0 or later. The parameters used are a gap open penalty of 10, a gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled “longest identity” (obtained using the -nobrief option) is used as the percent identity and is calculated as follows:

$$\frac{(\text{Identical Residues} \times 100)}{(\text{Length of Alignment} - \text{Total Number of Gaps in Alignment})}$$

EXAMPLES

Assays

Assay I: Testing of DNase Activity

[0553] DNase activity was determined on DNase Test Agar with Methyl Green (BD, Franklin Lakes, NJ, USA), which was prepared according to the manual from supplier. Briefly, 21 g of agar was dissolved in 500 ml water and then autoclaved for 15 min at 121° C. Autoclaved agar was tempered to 48° C. in water bath, and 20 ml of agar was poured into petri dishes with and allowed to solidify by incubation o/n at room temperature. On solidified agar plates, 5 µl of enzyme solutions are added and DNase activity is observed as colorless zones around the spotted enzyme solutions.

Assay II: Testing of Mannanase Activity

[0554] Mannanase activity may be tested according to standard test procedures known in the art, such as by applying a solution to be tested to 4 mm diameter holes punched out in agar plates containing 0.2% AZCL galactomannan (carob), i.e., substrate for the assay of endo-1,4-beta-D-mannanase available as CatNo.I-AZGMA from the company Megazyme (Megazyme’s Internet address: megazyme.com/Purchase/index.html).

Assay III: Testing of Xyloglucanase Activity

[0555] The reaction involves endo hydrolysis of 1,4-beta-D-glucosidic linkages in xyloglucan. For purposes of the present invention, xyloglucanase activity is determined

using AZCL-xyloglucan (from Megazyme) as the reaction substrate. The assay can be performed in several ways, e.g., as described in Example 2 of the present application or as described in WO 01/62903. One unit of xyloglucanase activity (XyloU) is defined by reference to the assay method described in WO 01/62903, page 60, lines 3-17.

Assay IV: Testing of Cellulase Activity

[0556] The term “cellulase activity” is defined herein as an enzyme catalyzed hydrolysis of 1,4-beta-D-glucosidic linkages in beta-1,4-glucan (cellulose). For purposes of the present invention, cellulase activity is determined using AZCL-HE-cellulose (from Megazyme) as the reaction substrate.

Example 1

Isolating Laundry Specific Bacterial Strains

[0557] One strain of *Brevundimonas* sp. isolated from laundry was used in the present example. The *Brevundimonas* sp. was isolated during a study, where the bacterial diversity in laundry after washing at 15, 40 and 60° C., respectively, was investigated. The study was conducted on laundry collected from Danish households. For each wash, 20 g of laundry items (tea towel, towel, dish cloth, bib, T-shirt armpit, T-shirt collar, socks) in the range 4:3:2:2:1:1:1 was used. Washing was performed in a Laundr-O-Meter (LOM) at 15, 40 or 60° C. For washing at 15 and 40° C., Ariel Sensitive White & Color was used, whereas WFK IEC-A* model detergent was used for washing at 60° C. Ariel Sensitive White & Color was prepared by weighing out 5.1 g and adding tap water up to 1000 ml followed by stirring for 5 minutes. WFK IEC-A model detergent (which is available from WFK Testgewebe GmbH) was prepared by weighing out 5 g and adding tap water up to 1300 ml followed by stirring for 15 min. Washing was performed for 1 hour at 15, 40 and 60° C., respectively, followed by 2 times rinsing with tap water for 20 min at 15° C. Laundry was sampled immediately after washing at 15, 40 and 60° C., respectively. Twenty grams of laundry was added 0.9% (w/v) NaCl (1.06404; Merck, Darmstadt, Germany) with 0.5% (w/w) tween 80 to yield a 1:10 dilution in stomacher bag. The mixture was homogenized using a Stomacher for 2 minutes at medium speed. After homogenization, ten-fold dilutions were prepared in 0.9% (w/v) NaCl. Bacteria were enumerated on Tryptone Soya Agar (TSA) (CM0129, Oxoid, Basingstoke, Hampshire, UK) incubated aerobically at 30° C. for 5-7 days. To suppress growth of yeast and moulds, 0.2% sorbic acid (359769, Sigma) and 0.1% cycloheximide (18079; Sigma) were added. Bacterial colonies were selected from countable plates and purified by restreaking twice on TSA. For long time storage, purified isolates were stored at -80° C. in TSB containing 20% (w/v) glycerol (49779; Sigma).

Preparation of Swatches with Biofilm

[0558] Swatches with biofilm of *Brevundimonas* sp. was included in the present study. Bacteria was pre-grown on Tryptone Soya Agar (TSA) (pH 7.3) (CM0131; Oxoid Ltd, Basingstoke, UK) for 2-5 days at 30° C. From a single colony, a loop-full was transferred to 10 mL of TSB and incubated for 1 day at 30° C. with shaking (240 rpm). After propagation, cells were pelleted by centrifugation (Sigma Laboratory Centrifuge 6K15) (3000 g at 21° C. in 7 min) and

resuspended in 10 mL of TSB diluted twice with water. Optical density (OD) at 600 nm was measured using a spectrophotometer (POLARstar Omega (BMG Labtech, Ortenberg, Germany). Fresh TSB diluted twice with water was inoculated to an OD₆₀₀ nm of 0.03, and 50 mL was added into a petri dish (diameter 125 mm), in which a swatch (80 mmx120 mm) of sterile cotton (WFK10A). After incubation (48 hours at 15° C. with shaking (100 rpm), swatches were rinsed twice with 0.9% (w/v) NaCl and dried in LAF bench for 60 min. Swatches were stored at 4° C. prior to wash.

Example 2

Wash Experiment

[0559] Wash experiment was performed using the Automatic Mechanical Stress Assay (AMSA). With AMSA, the wash performance of many small volume enzyme-detergent solutions can be examined at the same time. The AMSA plate has many slots for test solutions, and a lid that firmly squeezes the textile to be washed against the slot openings. During the wash, the plate, test solutions, textile and lid are vigorously shaken to bring the test solution in contact with the textile and apply mechanical stress in a regular, periodic, oscillating manner.

[0560] The wash experiment was conducted under the experimental conditions specified below:

Detergent dosage	3.3 g/L (liquid detergent)
Test solution volume	160 micro L
pH	pH 8
Wash time	20 minutes
Temperature	30° C.
Water hardness	15° dH
Soil	Wfk09V 0.7 g/L

Model Detergents and Test Materials were as Follows:

Laundry liquid model detergent	Model detergent A
Test material	<i>Brevundimonas</i> sp. 2-day biofilm grown on WFK10 (cotton) or WFK30A (polyester)

[0561] For wash experiments, Model detergent A (containing 12% LAS, 11% AEO Biosoft N25-7 (NI), 7% AEOS (SLES), 6% MPG, 3% ethanol, 3% TEA, 2.75% cocoa soap, 2.75% soya soap, 2% glycerol, 2% sodium hydroxide, 2% sodium citrate, 1% sodium formate, 0.2% DTMPA and 0.2% PCA (all percentages are w/w)) (3.3 g/L) dissolved in water hardness 15°dH (Ca:Mg:NaHCO₃=4:1:1.5) was used. Soil was subsequently added to reach a concentration of 0.7 g soil/L (WFK09V pigment soil) to reveal biofilm. After washing, textiles were flushed in tap water and dried over night before scanning. Wash experiments were done twice.

[0562] Wash performance was measured as the brightness of the WFK09V pigment soiled, washed textile. Brightness can also be expressed as the intensity of the light reflected from the sample when illuminated with white light. When the sample is soiled, the intensity of the reflected light is lower, than that of a clean sample. Therefore, the intensity of the reflected light can be used to measure wash performance. Intensity measurements were made with a professional flatbed scanner (Kodak iQsmart, Kodak, Midtager 29,

DK-2605 Brøndby, Denmark), which was used to capture an image of the washed and dried textile. To extract a value for the light intensity from the scanned images, 24-bit pixel values from the image were converted into values for red, green and blue (RGB). The intensity value (Int) was calculated by adding the RGB values together as vectors and then taking the length of the resulting vector:

$$Int = \sqrt{r^2 + g^2 + b^2}$$

Example 3: Wash Synergy Between DNase and Cellulase

[0563] To assess wash synergy between DNase (SEQ ID NO: 13) and cellulase (SEQ ID NO: 85) biofilm-harboring textile was AMSA washed a) in the absence of enzyme (blank), b) in the presence of DNase alone, c) in the presence of cellulase alone and d) with a mixture of DNase and cellulase. The resulting textile intensities and corresponding wash performances (WPs) are listed in Tables 1 and 2. Wash performances attributable to DNase (WP_{DNase}), cellulase (WP_{Cellu}) and the mixture of the two ($WP_{DNase+Cellu}$) were quantified as the difference in intensity between textile washed with and without enzyme: $WP_{DNase} = I_{DNase} - I_{Blank}$, $WP_{Cellu} = I_{Cellu} - I_{Blank}$, $WP_{DNase+Cellu} = I_{DNase+Cellu} - I_{Blank}$. The synergistic component of wash performance WP_{syn} was quantified as the extent to which wash performance of mixed DNase and cellulase ($WP_{DNase+Cellu}$) exceeded the sum of the individual wash performances of DNase alone and cellulase alone: $WP_{syn} = WP_{DNase+Cellu} - (WP_{DNase} + WP_{Cellu})$.

TABLE 1

synergistic wash effect of DNase and cellulase (wash experiment 1)				
	Cellulase (SEQ ID NO: 85)	I	WP	WP_{syn}
Blank	No enzyme	285.460	—	—
DNase	0.00002 ppm DNase	298.681	13.22	—
Cellulase	0.015% Cellulase	289.347	3.89	—
	0.15% Cellulase	290.955	5.49	—
DNase +	0.00002 ppm DNase +	304.770	19.31	2.20
Cellulase	0.015% Cellulase			
	0.00002 ppm DNase +	310.821	25.36	6.65
	0.15% Cellulase			

TABLE 2

synergistic wash effect of DNase and cellulase (wash experiment 2)				
	Cellulose (SEQ ID NO: 91)	I	WP	WP_{syn}
Blank	No enzyme	286.616	—	—
DNase	0.00002 ppm DNase	322.453	35.84	—
Cellulase	0.015% Cellulase	287.859	1.24	—
	0.15% Cellulase	291.329	4.71	—
DNase +	0.00002 ppm DNase +	326.890	40.27	3.19
Cellulase	0.015% Cellulase			
	0.00002 ppm DNase +	332.886	46.27	5.72
	0.15% Cellulase			

SEQUENCE LISTING

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Sequence total quantity: 91
SEQ ID NO: 1      moltype = AA  length = 182
FEATURE           Location/Qualifiers
source            1..182
mol_type = protein
organism = Bacillus sp.

SEQUENCE: 1
LPPDLPSKST TQAQLNSLNV KNEESMSGYS REKFPHWISQ GDGCDTRQVI LKRDADNYSG 60
NCPVTSGKWKY SYYDGITFND PSQLIDHVV PLAEAWRSGA SSWSTAKRED FANDLNGPQL 120
IAVSASSNRN KGDQDPSTWQ PPRAGANCAY AKMWINTKYN WGLHLQSSEK TALQGMLNSC 180
SY                                         182

SEQ ID NO: 2      moltype = AA  length = 182
FEATURE           Location/Qualifiers
source            1..182
mol_type = protein
organism = Bacillus horikoshii

SEQUENCE: 2
LPPGTPTKSE AQNQLNSLTV KSEGSMTGYS RDLFPHWSGQ GNGCDTRQIV LQRDADYYTG 60
TCPTTSGKWKY SYFDGVIVYS PSEIDIDHIV PLAEAWRSGA SSWTTEQRRA FANDLNGPQL 120
IAVTASVNRN KGDQDPSTWQ PPRAGARCAY AKWWINTKHR WNLHLQSSEK SSLQTMNLNGC 180
AY                                         182

SEQ ID NO: 3      moltype = AA  length = 182
FEATURE           Location/Qualifiers
source            1..182
mol_type = protein
organism = Bacillus sp.

SEQUENCE: 3
LPPGTPSKSE AQSQNLNALTV KPEDPMTGYS RDHFPHWISQ GNGCNTRQIV LQRDADYYSG 60
ACPVTTGKWKY SYFDGVIVYS PSEIDIDHIV PLAEAWRSGA SSWTTEKRRS FANDLNGPQL 120
IAVTASVNRN KGDQDPSTWQ PPRAGARCAY AKWWINTKHR WGLHLQSSEK SSLQSMNLNGC 180
AY                                         182

SEQ ID NO: 4      moltype = AA  length = 182

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FEATURE                               Location/Qualifiers
source                                1..182
                                         mol_type = protein
                                         organism = Bacillus sp.

SEQUENCE: 4
LPPGTPSKSE AQSQLNALT V KPEDPMTGYS RDHFPHWISQ GNGCNTRQIV LQRDADYYSG 60
ACPVTGKWK Y SYFDGVIVYS PSEIDIDHIV PLAEAWRSGA SSWTTEQRSS FANDLNGPQL 120
IAVTASVNR S KGDQDPSTWQ PPRAGARCAY AKWWINTKHR WGLHLQSSEK SSLQSMMLNGC 180
AY                                         182

SEQ ID NO: 5                         moltype = AA length = 182
FEATURE                               Location/Qualifiers
source                                1..182
                                         mol_type = protein
                                         organism = Bacillus horikoshii

SEQUENCE: 5
LPPGTPSKSE AQSQLNLSLT V KSEDPMGYS RDHFPHWSQ GNGCDTRQIV LQRDADYYSG 60
NCPTSGKWK Y SYFDGVIVYS PSEIDIDHV PLAEAWRSGA SSWTTEQRSS FANDLNGPQL 120
IAVTASVNR S KGDQDPSTWQ PPRAGARCAY AKWWINTKHR WNLHLQSSEK SALQTMLNGC 180
VY                                         182

SEQ ID NO: 6                         moltype = AA length = 182
FEATURE                               Location/Qualifiers
source                                1..182
                                         mol_type = protein
                                         organism = Bacillus horikoshii

SEQUENCE: 6
LPPGTPSKSE AQSQLNLSLT V KTEDPMGYS RDLFPHWSQ GSGCDTRQIV LQRDADYYTG 60
TCPTTSGKWK Y SYFDGVIVYS PSEIDVDHV PLAEAWRSGA SSWTTEQRRA FANDLNGPQL 120
IAVTASVNR S KGDQDPSTWQ PPRAGARCAY AKWWINTKHR WNLHLQSSEK SSLQTMMLNGC 180
AY                                         182

SEQ ID NO: 7                         moltype = AA length = 182
FEATURE                               Location/Qualifiers
source                                1..182
                                         mol_type = protein
                                         organism = Bacillus sp.

SEQUENCE: 7
LPPGTPSKSE AQSQLNALT V KAEDPMGYS RNLFPHWNSQ GNGCNTRQLV LQRDADYYSG 60
NCPTSGRW Y SYFDGVVVT S PSEIDIDHIV PLAEAWRSGA SSWTTEKRKE FANDLNGPQL 120
IAVTASVNR S KGDQDPSTWQ PPRAAARCGY AKWWINTKYR WDLSLQSSEK SSLQTMMLNTC 180
SY                                         182

SEQ ID NO: 8                         moltype = AA length = 182
FEATURE                               Location/Qualifiers
source                                1..182
                                         mol_type = protein
                                         organism = Bacillus sp.

SEQUENCE: 8
LPPGTPSKS Q AQSQLNALT V KAEDPMGYS RNLFPHWSSQ GNGCNTRQLV LQRDADYYSG 60
NCPTSGRW Y SYFDGVVVT S PSEIDIDHIV PLAEAWRSGA SSWTTEKRRE FANDLNGPQL 120
IAVTASVNR S KGDQDPSTWQ PPRVAARCGY AKWWINTKYR WDLSLQSSEK SSLQTMMLNTC 180
SY                                         182

SEQ ID NO: 9                         moltype = AA length = 182
FEATURE                               Location/Qualifiers
source                                1..182
                                         mol_type = protein
                                         organism = Bacillus sp.

SEQUENCE: 9
LPPGTPSKSE AQSQLTSLT V KPEDPMGYS RDHFPHWISQ GNGCNTRQIV LQRDADYYSG 60
NCPTTGKWK Y SYFDGVIVYS PSEIDIDHIV PLAEAWRSGA SSWTAEQRNN FANDLNGPQL 120
IAVTASVNR S KGDQDPSTWQ PPRTGARCAY AKWWINTKYR WGLHLQSSEK SSLQSMMLNGC 180
AY                                         182

SEQ ID NO: 10                        moltype = AA length = 183
FEATURE                               Location/Qualifiers
source                                1..183
                                         mol_type = protein
                                         organism = Bacillus sp.

SEQUENCE: 10
AFPPGTPSKS TAQSQLNLSLT VKSEGSMTGY SRDKFPHWIS QGDGCDTRQL VLKRDGDYYS 60
GNCPVTSGKW Y SYYDGIAVY SPSEIDIDHI VPPLAEAWRSGA ASGWTTEKRQ NFANDLNGPQ 120
IAVTASVNR SKGDQDPSTW QPPRSGSHCA YAKMWVNTKY RWGLHLQSSEK KSALQSMMLNA 180
CSY                                         183

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SEQ ID NO: 11      moltype = AA length = 185
FEATURE          Location/Qualifiers
source           1..185
mol_type = protein
organism = Bacillus horneckiae

SEQUENCE: 11
ASAFPPGTPS KSTAQSQLNS LTVKSEGSMT GYSRDKFPHW ISQGDGCDTR QLVLKRDGYD 60
YSGNCVPVTSQ KWYSYYDGIT VYSPSEIDID HIVPLAEAWR SGASGWTTEK RQSFDNLNG 120
PQLIAVTASV NRSKGDQDPS TWQPPRSGSH CAYAKMWNT KYRWGLHVQS AEKSALQSM 180
NACSY                                         185

SEQ ID NO: 12      moltype = AA length = 182
FEATURE          Location/Qualifiers
source           1..182
mol_type = protein
organism = Bacillus sp.

SEQUENCE: 12
FPPEIPIPSKST AQSQLNLSLT V KSEADAMTGY S RDKFPHWISQ GDGCDTRQMV LKRDADYYSG 60
SCPVTSGKWY SYYDGITVYS PSEIDIDHIV PLAEAWRS GA SSWTTEKRNN FANDLNGPQL 120
IAVTASVNR S KGDDQDPSTWQ PPRSGARCAY AKMWVNTKYR WGLHLQS A EK SGLESMLNTC 180
SY                                         182

SEQ ID NO: 13      moltype = AA length = 182
FEATURE          Location/Qualifiers
source           1..182
mol_type = protein
organism = Bacillus cibi

SEQUENCE: 13
TPPGTPSKA AQSQLNALT V KTEGMSGYS RDLFPHWISQ GSGCDTRQVV LKRDADSYSG 60
NCPVTSGSWY SYYDGVTFTN PSDLDIDHIV PLAEAWRS GA SSWTTSKRQD FANDLNGPQL 120
IAVSASTNR S KGDDQDPSTWQ PPRSGAACGY SKWWISTKYK WGLSLQSSEK TALQGMLNSC 180
SY                                         182

SEQ ID NO: 14      moltype = AA length = 182
FEATURE          Location/Qualifiers
source           1..182
mol_type = protein
organism = Bacillus sp.

SEQUENCE: 14
FPPGTPSKST AQSQLNLSLT V KSEGSMGTGYS RDKFPHWIGQ GSGCDTRQLV LQRDADYYSG 60
SCPVTSGKWY SYYDGVTFYD PSDLDIDHVV PLAEAWRS GA SSWSTQKRKD FANDLNGPQL 120
IAVSASSNR S KGDDQDPSTWQ PTRSGAACGY SKWWISTKHK WGLSLQSSEK NALQGMLNSC 180
VY                                         182

SEQ ID NO: 15      moltype = AA length = 182
FEATURE          Location/Qualifiers
source           1..182
mol_type = protein
organism = Bacillus idriensis

SEQUENCE: 15
LPPGTPSKST AQSQLNALT V QTEGSMGTGYS RDKFPHWISQ GNGCDTRQVV LQRDADYYSG 60
TCPVTSGKWY SYYDGVTLYN PSDLDIDHVV ALAEAWRS GA SSWTTDKRED FANDLNGTQL 120
IAVSASTNR S KGDDQDPSTWQ PPRSGAACGY AKWWISTKYK WNLNLSQSEK TALQGMLNSC 180
SY                                         182

SEQ ID NO: 16      moltype = AA length = 182
FEATURE          Location/Qualifiers
source           1..182
mol_type = protein
organism = Bacillus algicola

SEQUENCE: 16
FPPGTPSKSE AQSQLNLSLT V QSEGSMGTGYS RDKFPHWIGQ GNGCDTRQLV LQRDADYYSG 60
DCPVTSGKWY SYFDGVTVD PSDLDIDHMV PMAEAWRS GA SSWSTQKRKD FANDLNGPHL 120
IAVTASSNR S KGDDQDPSTWK PTRYGAHCGY AKWWINTKYV YDLTLQSSEK TELQGMLNTC 180
SY                                         182

SEQ ID NO: 17      moltype = AA length = 182
FEATURE          Location/Qualifiers
source           1..182
mol_type = protein
note = Environmental sample J
organism = unidentified

SEQUENCE: 17
LPPNIPSKAD ALTKLNALT V QTEGPMTGYS RDLPFHWSSQ GNGCNTRHVV LKRDADSVVD 60
TCPVTTGRWY SYYDGLVFTS ASDIDIDHVV PLAEAWRS GA SSWTSTKRQS FANDLNGPQL 120
IAVSATSNR S KGDDQDPSTWQ PPRAGARCAY AKMWVETKSR WGLTLQSSEK AALQTA INAC 180

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-continued

SY	182
SEQ ID NO: 18	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Bacillus vietnamensis</i>
SEQUENCE: 18	
FPPGTPSKST AQSQLNALT V KSESSMTGYS RDKFPHWIGQ RNGCDTRQLV LQRDADSYSG	60
SCPVTSGSWY SYYDGVTFTD PSDLDIDHVV PLAEAWRSGA SSWTTAKRED FANDLSGPQL	120
IAVSASSNRS KGQDPSTWQ PPRSGAACGY SKWWISTKYK WGLSLQSSEK TALQGMLNSC	180
IY	182
SEQ ID NO: 19	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Bacillus hwajinpoensis</i>
SEQUENCE: 19	
IPPGTPSKSA AQSQLDSLAV QSEGMSGYS RDKFPHWIGQ GNGCDTRQLV LQRDADYSG	60
DCPVTSGKWY SYFDGVQVYD PSYLDIDHMV PLAEAWRSGA SSWTQKRED FANDLDPHQL	120
IAVTASSNRS KGQDPSTWK PTRYSAHCGY AKWWINTKYV YDLNLQSSEK SALQSMILNTC	180
SY	182
SEQ ID NO: 20	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Paenibacillus mucilaginosus</i>
SEQUENCE: 20	
LPPGTPSKST AQSQLNLT V KSESSMTGYS RDGFPHWTSQ GGGCDTRQVV LKRDADYSG	60
SCPVTSGKWY SYYDGITVYS PSEIDIDHIV PLAEAWRSGA SSWTTEKRQN FANDLGGPQL	120
IAVTASSNRA KGQDPSTWK PTRSGAHCA Y AKWWINTKYR WGLHLQSSEK TALQSMILNTC	180
SY	182
SEQ ID NO: 21	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Bacillus indicus</i>
SEQUENCE: 21	
TPPGTPSKST AQTQLNALT V KTEGAMTGY S RDGFPHWISQ GSGCDTRQVV LKRDADYSG	60
SCPVTSGKWY SYYDGVTFYD PSDLDIDHIV PLAEAWRSGA SSWTTSKRQD FANDLSGPQL	120
IAVSASTNRS KGQDPSTWQ PPRAGAACGY SKWWISTKYK WGLSLQSSEK TALQGMLNSC	180
SY	182
SEQ ID NO: 22	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Bacillus marisflavi</i>
SEQUENCE: 22	
TPPVTPSKAT SQSQLNGLTV KTEGAMTGY S RDGFPHWSSQ GGGCDTRQVV LKRDADSYSG	60
NCPVTSGSWY SYYDGVKFTN PSDLDIDHIV PLAEAWRSGA SSWTTAQREA FANDLSGSQ	120
IAVSASSNRS KGQDPSTWQ PPRAGAKCGY AKWWISTKS V WNLSLQSSEK TALQGMLNSC	180
VY	182
SEQ ID NO: 23	moltype = AA length = 184
FEATURE	Location/Qualifiers
source	1..184
	mol_type = protein
	organism = <i>Bacillus luciferensis</i>
SEQUENCE: 23	
ASLPPGIPSL STAQLNLSL TVKSEGSLTG YSRDVFPHWI SQGSGCDTRQ VVLKRDADYY	60
SGNCPVTSGK WYSYYDGVTW YSPSEIDIDH VVPLAEAWRS GASSWTTEKR QNFANDLNGP	120
QIAVATASSN RSKGDQDPST WQPTRTGARC AYAKMWINTK YRWGLHLQSS EKSALQSMLN	180
TCSY	184
SEQ ID NO: 24	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Bacillus marisflavi</i>
SEQUENCE: 24	
TPPVTPSKET SQSQLNGLTV KTEGAMTGY S RDGFPHWSSQ GGGCDTRQVV LKRDADSYSG	60
NCPVTSGSWY SYYDGVKFTH PSDLDIDHIV PLAEAWRSGA SSWTTAQREA FANDLSGSQ	120

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IAVSASSNRS KGDQDPSTWQ PPRAGAKCGY AKWWISTKSK WNLSLQSSEK TALQGMLNSC	180
VY	182
SEQ ID NO: 25	moltype = AA length = 182
FEATURE	Location/Qualifiers
source	1..182
	mol_type = protein
	organism = <i>Bacillus</i> sp.
SEQUENCE: 25	
LPSGIPSKST AQSQLNSLTIV KSEGSMTGYS RDKFPHWISQ GGGCDTRQVV LKRDADYYSG	60
NCPVTSGKWY SYYDGISVYS PSEIDIDHVV PLAEAWRSGA SSWTTTKRQN FANDLNGPQL	120
IAVTASVNRS KGDQDPSTWQ PPRYGARCAY AKMWINTKYR WDLNLQSSEK SSLQSMLDTC	180
SY	182
SEQ ID NO: 26	moltype = AA length = 191
FEATURE	Location/Qualifiers
source	1..191
	mol_type = protein
	organism = <i>Pyrenopochaetopsis</i> sp.
SEQUENCE: 26	
LPSPLLIARS PPNIPSATTA KTQLAGLTV PQGPQTGYSR DLFPHWITQS GTCNTREVVL	60
KRDGTNVVTN SACASTSGSW LSPYDGKWTD SASDIQIDHL VPLSNAWKSG AAAWTTAQRO	120
FANDLTHPQ LVAVTGSVNE SKGDDGPEDW KPPLASYYCT YASMWAVKS NYKLTITSAE	180
KSALTSMLAT C	191
SEQ ID NO: 27	moltype = AA length = 190
FEATURE	Location/Qualifiers
source	1..190
	mol_type = protein
	organism = <i>Vibrissea flavovirens</i>
SEQUENCE: 27	
TPLPIIARTP PNIPTTATAK SQLAALTVAAGPQTGYSRDLFPHWITQSGTCNTRETVL	60
RDGTNVVDS ACVATSGSWY SPYDGATWTASDVVIDHMVPLSNAWKSGAAASWTAAQRQ	120
FANDLTPQ LVAVTDNVNQAKGDGPEDWPSLTSYCTYAKMWVKVTVYDLTITSAE	180
TALTMLNTC	190
SEQ ID NO: 28	moltype = AA length = 192
FEATURE	Location/Qualifiers
source	1..192
	mol_type = protein
	organism = <i>Setosphaeria rostrata</i>
SEQUENCE: 28	
APTSSPLVAR APPNVPSKAE ATSPLLAGLTV APQGPQTGYS RDLFPHWITQSGTCNTRETV	60
LKRDGTNVVTN NSACASTSGS WFSPYDGATW TAASDVVIDHMVPLSNAWKSGAASWTARR	120
QAFANDLTPQ QLLAVTDNVNQAKGDGPEDWPKPLTSYCTYAKMWVKVSVWGLTITS	180
EKSALTSMTC	192
SEQ ID NO: 29	moltype = AA length = 192
FEATURE	Location/Qualifiers
source	1..192
	mol_type = protein
	organism = <i>Endophragmiella valdina</i>
SEQUENCE: 29	
APVPGHLMR APPNVPTTAA AKTALAGLTV QAQGSQTGYS RDLFPHWITQSGTCNTREVV	60
LKRDGTNVVT DSACAAATSGT WVSPYDGATW TAASDVVIDHMVPLSNAWKSGAAASWTAAQR	120
QAFANDLTPQ QLLAVTDNVNQSKGDGPEDWPKPLTSYCTYAKMWVKVSVYSLTITS	180
EKTALTSMTC	192
SEQ ID NO: 30	moltype = AA length = 190
FEATURE	Location/Qualifiers
source	1..190
	mol_type = protein
	organism = <i>Corynespora cassiicola</i>
SEQUENCE: 30	
LPAPLVPRAP PGIPPTSAAR SQLAGLTVAA QGPQTGYSRDLFPHWITQSGSCNTREVLA	60
RDGTGVVQDS SCAATSGTWR SPFDGATWTASDVVIDHMVPLSNAWKSGAASWTTSRQA	120
FANDLTPQ LVAVTDNVNQSKGDGPEDWPPLTSYCTYAKMWVKVSYSLTITSAE	180
SALTSMMLTC	190
SEQ ID NO: 31	moltype = AA length = 192
FEATURE	Location/Qualifiers
source	1..192
	mol_type = protein
	organism = <i>Paraphoma</i> sp.
SEQUENCE: 31	
APAPVHLVAR APPNVPTAAQ AQTQLAGLTV AAQGPQTGYS RDLFPHWITQSGACNTRETV	60

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LKRDGTGVQ	DSACAATSGT	WKSPYDGATW	TAASDVIDH	MVPLSNAWKS	GAASWTTARR	120
QAFANDLTNP	QLLAVTDNVN	QAKGDKGPED	WKPPLTSYYC	IYARMWIKVK	SVYSLTITSA	180
EKSALTSMLG	TC					192
 SEQ ID NO: 32		moltype = AA	length = 186			
FEATURE		Location/Qualifiers				
source		1..186				
		mol_type = protein				
		organism = Monilinia fructicola				
 SEQUENCE: 32						
TPVPAPTGIP	STSVANTQLA	ALTVAAGSQ	DGYSRDLPPH	WITISGACNT	RETVLKRDGT	60
NVVVNSACAA	TSGTWSPYD	GATWTAASDV	DIDHLVPLSN	AWKAGASSWT	TAQRQAFAND	120
LVNPQLLAVT	DSVNQGKSDS	GPEAWKPSLK	SYWCTYAKMW	IKVKYVYDLT	ITSAEKSALV	180
TMMDT						186
 SEQ ID NO: 33		moltype = AA	length = 190			
FEATURE		Location/Qualifiers				
source		1..190				
		mol_type = protein				
		organism = Curvularia lunata				
 SEQUENCE: 33						
APAPLALARAP	PNIPSKADAT	SQLAGLTVA	QGPQTGYSRD	LFPHWITQSG	TCNTRETVLK	60
RDGTONVVTSS	SCAATSGTW	SPYDGATWTA	ASDVIDHVV	VPLSNAWKS	ASWTTARRQA	120
FANDLTNPQL	IAVTDSVNQA	KGDKGPEDWK	PPLSSYYCTY	SKMWIKVKSV	YGLTVTSAEK	180
SALSSMLATC						190
 SEQ ID NO: 34		moltype = AA	length = 191			
FEATURE		Location/Qualifiers				
source		1..191				
		mol_type = protein				
		organism = Penicillium reticulisperorum				
 SEQUENCE: 34						
LPAPEALPAP	PGVPSASTAQ	SELAALTVA	QGSQDGYSRS	KFPHWITQSG	SCDTRDVVLK	60
RDGTONVVSQA	SGCTITSGKW	VSPYDGATWT	ASSDVIDHVV	VPLSNAWKS	ASGWTTAARQ	120
AFANDLTPQ	LLVVTDNVNE	SKGDKGPEEW	KPPLTSYYCT	YAEMWVKVKS	VYKLITITSAE	180
KSALTSMLST	C					191
 SEQ ID NO: 35		moltype = AA	length = 191			
FEATURE		Location/Qualifiers				
source		1..191				
		mol_type = protein				
		organism = Penicillium quercetorum				
 SEQUENCE: 35						
LPAPEPAPSP	PGIPSASTAR	SELASLTVP	QGSQDGYSRA	KFPHWIKQSG	SCDTRDVLE	60
RDGTONVQSS	TGCTITGGTW	VSPYDGATWT	ASSDVIDHVL	VPLSNAWKS	ASAWTTAQRO	120
AFANDLTPQ	LVAVIDNVNE	AKGDKGPEEW	KPPLTSYYCT	YAEMWVKVKS	VYKLITITSAE	180
KSALSSMLNT	C					191
 SEQ ID NO: 36		moltype = AA	length = 192			
FEATURE		Location/Qualifiers				
source		1..192				
		mol_type = protein				
		organism = Setophaeosphaeria sp.				
 SEQUENCE: 36						
LPAVPTLEAR	APPNIPTAS	ANTLLLAGLT	AAQGSQQTGYS	RDLFPHWITQ	SGTCNTRETV	60
LKRDGTGVVT	DSACASTSGS	WYSVYDGATW	TAASDVIDH	VVPLSNAWKS	GAASWTTARR	120
QSFANDLTNP	QLIAVTDNVN	QAKGDKGPED	WKPPLTSYYC	TYAKMWVKV	SVYSLTITSA	180
EKTALTSMLN	TC					192
 SEQ ID NO: 37		moltype = AA	length = 192			
FEATURE		Location/Qualifiers				
source		1..192				
		mol_type = protein				
		organism = Alternaria sp. XZ2545				
 SEQUENCE: 37						
LPAVPTLEAR	APPNIPTAA	AKTQLAGLT	AAQGPQTGYS	RDLFPHWITQ	SGTCNTRETV	60
LKRDGTGVVT	DSACASTSGS	WFSVYDGATW	TAASDVIDH	VVPLSNAWKS	GAASWTTARR	120
QSFANDLTNP	QLIAVTDNVN	QAKGDKGPED	WKPPLTSYYC	TYAKMWVKV	SVYALTITSA	180
EKTALTSMLN	TC					192
 SEQ ID NO: 38		moltype = AA	length = 192			
FEATURE		Location/Qualifiers				
source		1..192				
		mol_type = protein				
		organism = Alternaria sp.				
 SEQUENCE: 38						

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LPAPVTLEAR APPNIPTAA AKTQLAGLTV AAQGPQTGYS RDLFPHWITQ SGSCNTREVV LQRDGTGVVT DSACAAATSGS WYSVYDGATW TAASDVDDIH MVPLSNAWKS GAASWTARR QAFANDLTNP QLLAVTDNVN QAKGDKGPED WKPPPLTSYYC TYAKMWVKV SVYALTITSA EKTALTSMILN TC	60 120 180 192
 SEQ ID NO: 39 FEATURE source SEQUENCE: 39 APLPAPPPIP SEDTARTQLA GLTVAVVGSG TGYSRDLFP TWAISGN CNA REYVLKRDGE GVQVNNA CEA QSGSWISP YDGETTNSAS DLDIDHMVPL KNAWISGAAT WTAAQRTSFA VSRRPQLWAVS ASSNRSKGDR SPDQWKPP LTTFYCTYAK SWITVKYN YGLSITSAEKSA SMLDTC	 moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Trichoderma reesei moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Chaetomium thermophilum moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Scyphomyces thermophilum moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Metapochonia suchlasporia moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Daldinia fissa moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Acremonium sp. XZ2007 moltype = AA length = 186 Location/Qualifiers 1..186 mol_type = protein organism = Acremonium dichromosporum
SEQ ID NO: 40 FEATURE source SEQUENCE: 40 APAPQPTPPG IPSRSTAQS Y LNSLTVAASY DDGNYNRDLF PHWNTVSGTC NTREYVLKD GSNVVTNSAC QATSGTWYSP YDGETTNSAS DLDIDHMVPL KNAWISGAAT WTAAQRTSFA NDINSPQLWA VTDSVNQSKG DKSPDKWPK LTTFYCTYAK SWITVKYN YGLSITSAEKSA LQNMINTC	60 120 180 186
SEQ ID NO: 41 FEATURE source SEQUENCE: 41 LPAPAPMPTP PGIPSKSTAQ SQLNALTVKA SYDDGKYKRD LFPHWNTVSG TCNTREYVLK RDGVNVNTNSAC ACAATSGTWY SPFDGATWTAS ASDVDIDHMVPL KNAWISGA NNWTSTKRTQ FANDINLQLW WAVTDDVNQA KGDKSPDKWPK PPLTSFYCTYAK SWITVKYN YGLSITSAEK SALTSMINTC	60 120 180 190
SEQ ID NO: 42 FEATURE source SEQUENCE: 42 VPVPAPPPIP STSTAKTLLA GLKVAVPLSG DGYSREKFPL WETIQGTCNA REFVLKRDGT DVKTNNACVA ESGNWSPYD GVKFTAARDL DIDHMVPL KNAWISGAAT WTAAQRTSFA ITRPQLWAVS AHANRGKSDD SPDEWKPPPLK TFWCTYAK SWITVKYN YGLSITSAEK GMLDSC	60 120 180 186
SEQ ID NO: 43 FEATURE source SEQUENCE: 43 APAPIPV AEP APMPMPTPPG IPSASSAKSQ LASLTVAAY DDGGYQRDLF PTWDTITGTC NTREYVLKD GANVQVGS DC YPTSGTWTSP YDGGKWTSPS DV DIDHMVPL KNAWISGAAT WTAAQRTSFA NDVDRPQLWA VTDSVNSSKG DKSPDTWPK L TSFYCTYAS AYVAVKSYWG LTTSAEKSA LSDMLGTC	60 120 180 198
SEQ ID NO: 44 FEATURE source SEQUENCE: 44 LPLQSRDPG IPSTATAKSL LNGLTVKAWS NEGTYDRDLF PHWQTIETGTC NAREYVLKD GQN VVVSAC TAQSGTWKSV YDGETTNSAS DLDIDHMVPL KNAWISGAAT WTAAQRTSFA NDI SS PQLWA VTAGVNRSKS DRSPDTWVPP LASFHCTYKG AWVQVKSKWA LSITSAEKSA LTG LLNK C	60 120 180 188
SEQ ID NO: 45 FEATURE source SEQUENCE: 45 moltype = AA length = 182 Location/Qualifiers 1..182 mol_type = protein organism = Acremonium dichromosporum	182

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SEQUENCE: 45
IPPGIPEAT ARSLLSSLTV APTVDDGTYD RDLFPHWSSV EGNCNAREFV LRRDGDGCSV 60
GNDCYPTAGT WTCPYDGKRH SVPSDVSIDH MVPLHNNAWMT GASEWTTAER EAFANDIDGP 120
QLWAVTSTTN SQKGSDAPDE WQPQTSIHC KYAAAWIQVK STYDLTVSSA EQAALEEMLG 180
RC 182

SEQ ID NO: 46      moltype = AA length = 188
FEATURE
source
1..188
mol_type = protein
organism = Sarocladium sp. XZ2014

SEQUENCE: 46
VPIPLPDPPG IPSSSTANTL LAGLTVRASS NEDTYNRDLF PHWVAISGNC NAREYVLRRD 60
GTNVVVNTAC VPQSGTWRSP YDGESTTNAS DLDIDHMVPLK KNAWISGAAS WTTAKRQDF 120
NDVSGPQLWA VTAGVNRSKG DKSPDSWVPP LASFHCTYAR SWIQUVKSSWA LSVTSAEKA 180
LTDLLSTC 188

SEQ ID NO: 47      moltype = AA length = 186
FEATURE
source
1..186
mol_type = protein
organism = Metarhizium sp. HNA15-2

SEQUENCE: 47
VPVPAPPPIP TASTARTLLA GLKVATPLSG DGYSRTLFPET WETIEGTCNA REFVLKRDGT 60
DVQTNTACVA QSGNWVSPYD GVAFTAASDL DIDHMVPLKN AWISGASQWT TDKRKGLAND 120
ITRPQLWAVS AHANRAKGDS SPDEWKPPPLK TFWCTYARSW VQVKSYALT ITDAEKGALS 180
GMLDSC 186

SEQ ID NO: 48      moltype = AA length = 188
FEATURE
source
1..188
mol_type = protein
organism = Acremonium sp. XZ2414

SEQUENCE: 48
APIAVRDPPG IPSASTANTL LAGLTVRASS NEDSYDRNLF PHWSAISGNC NAREFVLERD 60
GTNVVVNNAC VAQSGTWRSP YDGETTGNA S DLDIDHMVPLK KNAWISGASS WSTTRQEFA 120
NDVSGPQLWA VTAGVNRSKG DRSPDSWVPP LASFHCTYAK SWVQVKSSWS LSVTSAEKA 180
LSDLGGTC 188

SEQ ID NO: 49      moltype = AA length = 186
FEATURE
source
1..186
mol_type = protein
organism = Isaria tenuipes

SEQUENCE: 49
APVPEPPPIP STSTAQSDLN SLQVAASGSG DGYSRAEPPH WVSVEGSCDS REYVLKRDGQ 60
DVQADSSCKI TSGTNVSPYD ATTWNNSKV DIDHLVPLKN AWISGASSWT KAQRQDFAND 120
IKRPQLYAVS ENANRSKGDR SPDWKPPPLK SFYCTYAKSW VAVKSYYKLT ITSAEKSALG 180
DMLDTC 186

SEQ ID NO: 50      moltype = AA length = 184
FEATURE
source
1..184
mol_type = protein
organism = Scytalidium circinatum

SEQUENCE: 50
APPGIPEAST ASSLLGELAV AEPVDDGSYD RDLFPHWEPI PGETACSCARE YVLRDGTV 60
ETGSDCYPTS GTWSSPYDGG SWTAPSVDI DHMVPLKN AWISGASQWT EREAFANDID 120
GPOLWAVTDE VNQSKSDQSP DEWKPPLSF YCTYACAWIQ VKSTYLSIS SAEQAALDM 180
LGSC 184

SEQ ID NO: 51      moltype = AA length = 186
FEATURE
source
1..186
mol_type = protein
organism = Metarhizium lepidiotae

SEQUENCE: 51
VPVPAPPPIP TASTARTLLA GLKVATPLSG DGYSRTLFPET WETIEGTCNA REFVLKRDGT 60
DVQTNTACVA ESGNWVSPYD GVSFTAASDL DIDHMVPLKN AWISGASQWT TDKRKDLAND 120
ITRPQLWAVS AHANRSKGDS SPDEWKPPPLQ TFWCTYSKSW IQVKSHYSLT ITDAEKGALS 180
GMLDSC 186

SEQ ID NO: 52      moltype = AA length = 226
FEATURE
source
1..226
mol_type = protein

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SEQUENCE: 52          organism = Thermobispora bispora
LDIADGRPAG GKAAEAATGT SPLANPDGTR PGLAAITSAD ERAEARALIE RLRTKGRGPK 60
TGYEREKFQY AWADSDVGIP FGRNGCDTRN DVLKRDGORL QFRSGSDCVV ISMLTFDPYT 120
GKTIEWTKQN AAEVQIDHVV PLSYSWQMGA SRWSDEKRRQ LANDPLNLMP VDGATNSRKG 180
DSGPASWLPP RREIRCAYV RFAQVALKYD LPVTTADKET MLQQCS                226

SEQ ID NO: 53          moltype = AA length = 191
FEATURE               Location/Qualifiers
source                1..191
mol_type = protein
organism = Sporomia fimetaria

SEQUENCE: 53          moltype = AA length = 191
LPAPAVLEKRT PPNIPSTSTA QSLLSGLTVVA PQGSQTGYSR DLFPHWITVS GTCNTRETVL 60
KRDGSNVTD SACASVSGSW YSTYDGATWT AASDVDDIDHV VPLSNAWKSG AASWTTARRQ 120
AFANDLTNPQ LIAVTDNVNQ AKGDQGPESW KPPLTSYYCT YAKMWVKVKS VYSLTVTSAE 180
KSALSSMLGT C          191

SEQ ID NO: 54          moltype = AA length = 193
FEATURE               Location/Qualifiers
source                1..193
mol_type = protein
organism = Pycnidiphora cf. dispersa

SEQUENCE: 54          moltype = AA length = 193
LPAPAPVLVA REPPNIPSTS SAQSMSLGLT VKAQGPQDGY SRDLFPHWIT ISGTCNTRET 60
VLRDGTNTVV TNSACASTSG SWYSPYDGAT WTAASDVDDID HIVPLSNAWK SGAASWTTSR 120
RQQFANDLTN PQLIAVTDSV NQAKGDKGPE DWKPSRTSYH CTYAKMWIKV KSVYSLTVTS 180
AEKSALTMM NTC          193

SEQ ID NO: 55          moltype = AA length = 199
FEATURE               Location/Qualifiers
source                1..199
mol_type = protein
note = Environmental sample D
organism = unidentified

SEQUENCE: 55          moltype = AA length = 199
DTDPPEPVAGS ALEALAGLEV KGPGPDTGYE RALFGPPWAD VDGNGCDTRN DILARDLTDL 60
TFSTRGDVCE VRTGTFDDPY TGETIDFRRG NATSAAVQID HVPVLLDAWR KGARAWDDET 120
RQQFANDPLN LLASDG PANQ SKGARDASAW LPPNHAFRCP YVARQIAVKA AYELSVTPSE 180
SEAMARVLAD CPAEPLPAG                199

SEQ ID NO: 56          moltype = AA length = 199
FEATURE               Location/Qualifiers
source                1..199
mol_type = protein
note = Environmental sample O
organism = unidentified

SEQUENCE: 56          moltype = AA length = 199
DDEPEPARGS ALEALARLEV VGPGPDTGYE RELFGPAWAD VDGNGCDTRN DILARDLTDL 60
TFSTRGEVCE VRTGTFQDPY TGETIDFRRG NATSMAVQID HVPVLLDAWR KGARAWDDET 120
RQQFANDPLN LLASDG PANQ SKGARDASAW LPPNHAFRCP YVARQIAVKT AYELSVTPSE 180
SEAMARVLED CPAEPVPAG                199

SEQ ID NO: 57          moltype = AA length = 186
FEATURE               Location/Qualifiers
source                1..186
mol_type = protein
organism = Clavicipitaceae sp-70249

SEQUENCE: 57          moltype = AA length = 186
VPVPAPPGIP STSTAKTLLA GLKVATPLSG DGYSRDKFPT WETIQGTCNA REFVIKRDGT 60
DVKITNSACVA ESGNNVSPYD GVKFTAARDL DIDHMVPLKN AWISGASQWT TEQRKALAND 120
ITRPQLWAHS AHANRGKSDD SPDEWKPLK TFWCTYAKSW VQVKSFYKLT ITDTEKGALA 180
GMLDTC                         186

SEQ ID NO: 58          moltype = AA length = 187
FEATURE               Location/Qualifiers
source                1..187
mol_type = protein
organism = Westerdykella sp. AS85-2

SEQUENCE: 58          moltype = AA length = 187
FPAPASVLEA RAPPNIPSAS TAQSLLVGLT VQPQGPQDGY SRDLFPHWIT ISGTCNTRET 60
VLKRDGSNVV TNSACAATSG TWYSPYDGAT WTSASDVDDID HLVPLSNAWK SGAASWTTAK 120
RQQFANDLTN PQLLAUTDRV NQAKGDKGPE AWKPSLASYH CTYAKMWVKV KSKDVRLTGN 180
WTKDDGW                         187

SEQ ID NO: 59          moltype = AA length = 194

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FEATURE	Location/Qualifiers
source	1..194
	mol_type = protein
	organism = <i>Humiclopsis cephalosporioides</i>
SEQUENCE: 59	
APTPAPVELE RRTPPNIPTT ASAKSLLAGL TVAAQGPQTG YSRDLFPHWI TISGSCNTRE 60	
TVLKRDGTGV TVDSACASTA GSWYSPYDGA TWTAASDVDI DHMVPLSNAW KSGAAQWTTA 120	
RQRODFFANDLT NPQLFAVTDN VNQEKGDKGP EDWKPSLTSY YCTYAKAWVK VKSVWALTIT 180	
SAEKSALTTM LNTC	194
SEQ ID NO: 60	moltype = AA length = 190
FEATURE	Location/Qualifiers
source	1..190
	mol_type = protein
	organism = <i>Neosartorya massa</i>
SEQUENCE: 60	
IPAPVALPTP PGIPSAATAE SELAALTVA QGSSSGYSRD LFPHWISQGG SCNTREVVLA 60	
RDGSGVVVKDS NCYPTSGSWY SPYDGATWT ASDVDIDHV PLANAWRSGA SKWTTSQRQA 120	
FANDLTNPQL MAVTDVNQKA KGDDGPEAWK PPLTSYYCTY AKMWVVRKVYV YDLTITSAEK 180	
SALVSMMLDTC	190
SEQ ID NO: 61	moltype = AA length = 191
FEATURE	Location/Qualifiers
source	1..191
	mol_type = protein
	organism = <i>Roussoella intermedia</i>
SEQUENCE: 61	
APTPALLPRA PPNIPISTATA KSQLAALTVA AQGPQDGYSR DLFPHWITQS GSCNTREVVL 60	
KRDGTNVVQD SSCAATSGTW VSPFDGATWT AASDVDIDHL VPLSNAWKSG AASWTTARRQ 120	
SFANDLTNPQ LLAVTDEVNQ AKGDKGPESW KPPLTSYHCT YAKMWVVKVS TYSLTITSAE 180	
KSALTTMLNT C	191
SEQ ID NO: 62	moltype = AA length = 191
FEATURE	Location/Qualifiers
source	1..191
	mol_type = protein
	organism = <i>Pleosporales sp.</i>
SEQUENCE: 62	
LPTPSLVKRP PPNIPSTSTA KSLLAGLTV AQGPQDGYSR DLFPHWITIS GTCNTRETVL 60	
KRDGTNVVTD SACASTSGW YSTYDGATWT AASDVDIDHV VPLSNAWKSG AASWTTARRQ 120	
SFANDLTNPQ LIAVTDVNQ SKGDKGPESW KPPLTSYHCT YAKMWVVKVD VYSLTVTSAE 180	
KSALTTMLNT C	191
SEQ ID NO: 63	moltype = AA length = 192
FEATURE	Location/Qualifiers
source	1..192
	mol_type = protein
	organism = <i>Phaeosphaeria sp.</i>
SEQUENCE: 63	
LPAPIHLTAR APPNIPSASE ARTQLAGLT AAQGPQDGYS RDLPFWITQ SGTCNTRETV 60	
LKDGTNVNT NSACASTSGS WFSPYDGATW TAASDVDIDH MVPLSNAWKS GAASWTTARR 120	
QAFANDLTNP QLLAVTDVNQ QAKGDKGPED WKPLTSYYC TYAKMWVVKV SVYALTVTSA 180	
EKSALTSMLG TC	192
SEQ ID NO: 64	moltype = AA length = 189
FEATURE	Location/Qualifiers
source	1..189
	mol_type = protein
	organism = <i>Didymosphaeria futilis</i>
SEQUENCE: 64	
LPTPNLTLEAR APPNIPSTSA AQSQLSALTVA AAQGPQTGYS RDLPFWITQ SGTCNTRETV 60	
LKDGTNVLT DSACASTSGS WKSPYDGATW TAASDVDIDH VVPLSNAWKS GAASWTTARR 120	
QSFANDLTNP QLIAVTDVNQ QAKGDKGPED WKPLTSYYC TYAKMWVVKV SVYSLTITS 180	
EKSALTMLA	189
SEQ ID NO: 65	moltype = AA length = 109
FEATURE	Location/Qualifiers
source	1..109
	mol_type = protein
	organism = <i>Bacillus licheniformis</i>
SEQUENCE: 65	
ARYDDILYFP ASRYPETGAH ISDAIKAGHS DVCTIERSGA DKRRQESLKG IPTKPGFDRD 60	
EWPMAMCEEG GKGASVRYVS SSDNRGAGSW VGNRLSGFAD GTRILFIVQ	109
SEQ ID NO: 66	moltype = AA length = 110
FEATURE	Location/Qualifiers

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source          1..110
               mol_type = protein
               organism = Bacillus subtilis
SEQUENCE: 66
ASSYDKVLYF PLSRYPETGS HIRDAIAEGH PDICTIDRDG ADKRREESLK GIPTKPGYDR 60
DEWPMAVCEE GGAGADVRYV TPSDNRGAGS WVGNQMSSYP DGTRVLFIVQ           110

SEQ ID NO: 67      moltype = AA length = 221
FEATURE          Location/Qualifiers
source           1..221
               mol_type = protein
               organism = Aspergillus oryzae
SEQUENCE: 67
VPVNPEPDAT SVENVALKTG SGDSQSDPIK ADLEVKGQSA LPFDVDCWAI LCKGAPNVLQ 60
RVNEKTKNSN RDRSGANKGP FKDPQKWGIK ALPPKNPSWS AQDFKSPEY AFASSLQQGT 120
NAILAPVNLA SQNSQGGVLN GFYSANKVAQ FDPSKPQQTG GTWFQITKFT GAAGPYCKAL 180
GSNDKSVCDK NKNIAGDWGF DPAKWAYQYD EKNNKFNYVG K                   221

SEQ ID NO: 68      moltype = AA length = 188
FEATURE          Location/Qualifiers
source           1..188
               mol_type = protein
               organism = Trichoderma harzianum
SEQUENCE: 68
APAPMPTPPG IPTESSARTQ LAGLTVAVAG SGTGYSRDLF PTWDAISGNC NAREYVLKRD 60
GEGVQVNNAAC ESQSGTWISP YDNASFTNAS SLIDDHMVPL KNAWISGASS WTTAQREALA 120
NDVSRPQLWA VSASANRSKG DRSPDQWKPP LTSFYCTYAK SWIDVKSFYK LTITSAEKTA 180
LSSMLDTC

SEQ ID NO: 69      moltype = length =
SEQUENCE: 69
000

SEQ ID NO: 70      moltype = length =
SEQUENCE: 70
000

SEQ ID NO: 71      moltype = length =
SEQUENCE: 71
000

SEQ ID NO: 72      moltype = length =
SEQUENCE: 72
000

SEQ ID NO: 73      moltype = AA length = 7
FEATURE          Location/Qualifiers
source           1..7
               mol_type = protein
               organism = synthetic construct
VARIANT          1
note = X=D, M or L
VARIANT          2
note = X=S or T
VARIANT          7
note = X=D or N
SEQUENCE: 73
XXGYSRX

SEQ ID NO: 74      moltype = AA length = 8
FEATURE          Location/Qualifiers
source           1..8
               mol_type = protein
               organism = synthetic construct
VARIANT          3
note = X=any amino acid
SEQUENCE: 74
ASXNRSKG

SEQ ID NO: 75      moltype = AA length = 8
FEATURE          Location/Qualifiers
source           1..8
               mol_type = protein
               organism = synthetic construct
VARIANT          1
note = X=V or I

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VARIANT	4		
	note = X=S or A		
SEQUENCE: 75			
XPLXNAWK		8	
SEQ ID NO: 76	moltype = AA length = 4		
FEATURE	Location/Qualifiers		
REGION	1..4		
source	note = Motif		
	1..4		
	mol_type = protein		
SEQUENCE: 76	organism = synthetic construct		
NPQL		4	
SEQ ID NO: 77	moltype = length =		
SEQUENCE: 77			
000			
SEQ ID NO: 78	moltype = length =		
SEQUENCE: 78			
000			
SEQ ID NO: 79	moltype = AA length = 269		
FEATURE	Location/Qualifiers		
source	1..269		
	mol_type = protein		
SEQUENCE: 79	organism = Bacillus lentus		
AQSVWPWGISR VQAPAAHNRG	LGTSGVKVAV LDTGISTHPP LNIRGGASFV PGEPSTQDGN	60	
GHGTHVAGTI AALNNNSIGVL	GVAPSAELYA VKVLGASGSG SVSSIAQGLE WAGNNNGMHVA	120	
NLSLGSPSPS ATLEQAVNSA	TSRGVLVVA SGNSGAGSIS YPARYANAMA VGATDQNNNR	180	
ASFSQYQAGL DIVAPGVNVQ	STYPGSTYAS LNGTSMATPH VAGAAALVKQ KNPSWSNVQI	240	
RNHLKNTATS LGSTNLGYSG	LVNAEAAATR	269	
SEQ ID NO: 80	moltype = AA length = 275		
FEATURE	Location/Qualifiers		
source	1..275		
	mol_type = protein		
SEQUENCE: 80	organism = Bacillus amyloliquefaciens		
AQSVPYVGVSQ IKAPALHSQG	YTGSNVKVAV IDSGIDSSH DLKVAGGASM VPSETNPQD	60	
NNSHGTHVAG TVAALNNNSIGV	YAVKVLGADG SGQYSWIING IEWAIANNNMD	120	
VINMSLGGPS GSAALKAAVD	KAVASGVVVV AAAGNEGTS SSSTVGYPGK YPSVIAVGAV	180	
DSSNQRASF S VGPPELDVMA	PGVSIQSTLP GNKYGAYNGT SMASPHVAGA AALILSKHPN	240	
WTNTQVRSSL ENT TTKLGDS	FYYGKGLINV QAAAQ	275	
SEQ ID NO: 81	moltype = AA length = 311		
FEATURE	Location/Qualifiers		
source	1..311		
	mol_type = protein		
SEQUENCE: 81	organism = Bacillus subtilis		
AVPSTQTTPWG IKSINYDQSI	TKTTGGSGIK VAVALDTGVYT SHDLLAGSSE QCKDFTQSNP	60	
LVDGSCTDRQ GHGTHVAGTV	LAHGGNSNGQG VYGVAPQAKL WAYKVLGDN SGYSDIAAA	120	
IRHVVADEASR TGSKVVINMS	LGSSAKDSL ASAVDYAYGK GVLIVAAAGN SGSGSNTIGF	180	
PGLLVNAVAV AALEVNQQNG	TYRVADEFSSR GNPATAGDYI IQERDIEVSA PGASVESTWY	240	
TGGYNTISGT SMATPHVAGL	AAKIWSANTS LSHSQLRTTEL QNRAKVDIK GGIGAGTGDD	300	
YASGFQYPRV K		311	
SEQ ID NO: 82	moltype = AA length = 298		
FEATURE	Location/Qualifiers		
source	1..298		
	mol_type = protein		
SEQUENCE: 82	organism = Bacillus bogoriensis		
ANSGFYVSGT TLYDANGNPF	VMRGINHGH A WKDQATTAI E GIANTGANT VRIVLSDGGQ	60	
WTKDDIHTVR NLISLAEDNH	LVALEVHDA TGYDSIASLN RAVDYWIEMR SALIGKEDTV	120	
IINIANEWFG SWEGDAWADG	YKQAPIPRLRN AGLNHTLMVD AAGWGQFPQS IHDYGREVFN	180	
ADPQRNTMFS IHMYEYAGGN	ASQVRTNIDR VLNQDLALVII GEFGHRHTNG DVDEATIMSY	240	
SEQRGVGWLA WSWKGNGPEW	EYLDLSNDWA GNNLTAWGNT IVNGPYGLRE TSRLSTVF	298	
SEQ ID NO: 83	moltype = AA length = 278		
FEATURE	Location/Qualifiers		
source	1..278		
	mol_type = protein		

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SEQUENCE: 83	organism = Thielavia terrestris
ASGSGQSTRY WDCKPSCAW PGKAAVSQPV YACDANFQLR SDFNVQSGCN GGSAYSCADQ 60	
TPWAVNDNLIA YGFAATSIAG GSESSWCCAC YALTFTSGPV AGKTMVQST STGGDLGSNQ 120	
FDIAMPGGV GIFNGCSSQF GGLPGAQYGG ISSRDQCDSF PAPLKPGCQW RFDWFQNADN 180	
PTTTFQQVQC PAEIVARSQG KRNDSSFPV FTPPSGGNGG TGTPTSTAPG SGQTSPGGG 240	
GCTSQKWAQC GGIGFSGCTT CVSGTTQKLN NDYFSQCL 278	
 SEQ ID NO: 84	moltype = AA length = 470
FEATURE Location/Qualifiers	
source 1..470	
mol_type = protein	
organism = Humicola insolens	
SEQUENCE: 84	
MRSSPLLRSA VVAALPVAL AADGRSTRYW DCKPSCGWKA KAPVNQPVF SCNANFQRI 60	
DFDAKSGCEP GGVAYSCADQ TPWAVNDFA LGFAATSIAG SNEAGWCCAC YELTFTSGPV 120	
AGKKMVVQST STGGDLGSNH FDNLNPQGGV GIFDGCTPQF GGLPGQRYGG ISSRNEDCRF 180	
PDALPKPGYVW RFDWFKNADNF PSFSPRQVQC PABELARTGC RRNDDGNFPA VQIPSSSTSS 240	
PVNQPTSTST TSTSTTSSPP VQPTTPSGCA DGRSTRYWDC CKPSCGWAKK APVNQPVFSC 300	
NANFQRIITDF DAKSGCEPVG VAYSCADQTP FAATSTAGSN EAGWCACAYE 360	
LTFSTGPVAG KKMVVQSTST GGDLGSNHF LNIPGGGVGI FDGCTPQFGG LPGQRYGGIS 420	
SRNECDRFPD ALKPGCYWRF DWFKNADNPS FSFRQVQCPA ELVARTGCC 470	
 SEQ ID NO: 85	moltype = AA length = 425
FEATURE Location/Qualifiers	
source 1..425	
mol_type = protein	
organism = Escherichia coli	
SEQUENCE: 85	
TALLLGLVNG QKPGETKEVH PQLTFRCKT RGCKPATNF IVLDSLSHPI HRAEGLGP 60	
CGDWGNPPPK DVCVDVESCA KNCIMEGIPD YSQYGVTTNG TSLRLQHILP DGRVPSPRVY 120	
LLDKTKRYY MLHHTGFEFT DVFDATKLPC GMNSALVLSM MHPTGAKSKY NPGGAYYGTG 180	
YCDACQCFVTP FINGLGNIEG KGSCCNEMDI WEANSRASHV APHTCNKGL YLCGEEECAF 240	
EGVCDKNGCG WNNYRVNVTD YYGRGEEFKV NTLKPFTVVT QFLANRGKL EKIHRFYVQD 300	
GKVIESTYTN KEGVPYTNMI DDEFCCEATGS RKYMELGATO GMGEALTRGM VLAMSIWWDQ 360	
GGNMEWLHDH EAGPCAKGEG APSNIVQVEP FPEVTVTNRL WGEIGSTYQE VQKPKPKPH 420	
GPRSD 425	
 SEQ ID NO: 86	moltype = AA length = 773
FEATURE Location/Qualifiers	
source 1..773	
mol_type = protein	
organism = Bacillus subtilis	
SEQUENCE: 86	
AEGNTREDNF KHLLGNDNVK RPSEAGALQL QEVDGQMTLV DQHGEKIQLR GMSTHGLQWF 60	
PEILNDNAYK ALANDWESNM IRLAMYVGEN GYASNPELIK SRVIKGIDLIA IENDMYVIVD 120	
WHVHAPGDPR DPVYAGAEDF FRDIAALYPN NPHIIYELAN EPSSNNNGGA GIPNNEEGWN 180	
AVKEYADPIV ALERDGSNAD DNIIIVGSPN WSQRPDLAAD NPINDHHTMY TVHFYTGSHA 240	
ASTESYPPEP PNSERGNVMS NTRYALENGV AVFATEWGTS QANGDGGPYF DEADWIEFL 300	
NENNISIWANW SLTNKNEVSG AFTPFLGKS NATNLDPGD HWAAPEELSL SGEYVRARIK 360	
GVNYEPIDRT KYTKVLWDFD DGTKQGFGVN SDSPNKEELIA VDNENNTLKV SGLDVSNDVS 420	
DGNFWANARL SADGWGKSVD ILGAEKLTMD VIVDEPTTVIA IAAIPQSSKS GWANPERAVR 480	
VNAEDFVQQT DGKYKAGLTI TGEDAPNLKN IAPHEEDNMM NNIILFVGTD AADVIYLDNI 540	
KVIGTEVEIP VVHDPKGEAV LPSVFEDGTR QGWDWAGESG VKTALTIEEA NGSNALSWEF 600	
GYPEVKPSDN WATAAPRLDFW KSDLVRGEND YVAFDFYLDP VRATEGAMNI NLVFQOPTNG 660	
YWWQAPKTYT INFDELEEEAN QVNGLYHYEV KINVRDITNI QDDTLLRNMM IIFADVESDF 720	
AGRVFVDNVR FEGAATTEPV EPEPVDPGEE TPPVDEKEAK CEQKEAEKEE KEE 773	
 SEQ ID NO: 87	moltype = AA length = 524
FEATURE Location/Qualifiers	
source 1..524	
mol_type = protein	
organism = Paenibacillus polymyxa	
SEQUENCE: 87	
VVHGQTAKTI TIKVDTFKDR KPISPYIYGT NQDLAGDENM AARRLGGNRM TGYNWENNMS 60	
NAGSDDWQHSS DNYLCNSNGGL TQAECEKPGA VVTSFHQSOL KLGTYSLVTI PMAGYVAADG 120	
NGSVQESEAA PSARWNQVNV AKNAPFQLQP DLNDNYVYVD EFVHFLVNKY GTASTKAGVK 180	
GYALDNEPAL WSHTHPRIHP EKVGAKELVD RSVSLSKAVK AIDAGAEVFG PVLYGFGAYK 240	
DLQTAQWDWS VKGNYSWFVD YYLDQMRLLSS QVEGKRLLDV FDVHWYPEAM GGGIRITNEV 300	
GNDETKARM QAPRTLWDPT YKEDSWIAQW FSEFLPILPR LKQSVDKYYP GTKLAMTEYS 360	
YGGENDISGG IAMTDLVGLIL GKNDVYMANW WLKDGVNMY VSAAYKLYRN YDGKNSTFGD 420	
TSVSAQTSDI VNSSVHASVT NASDKELHLV VMNKSMDSAF DAQFDLSGAK TYISGKVWGF 480	
DKNSSQIKEA APITQISGNR FTYTVPPPLTA YHIVLTTGND TSPV 524	
 SEQ ID NO: 88	moltype = AA length = 485
FEATURE Location/Qualifiers	

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source	1..485	
		mol_type = protein
		organism = <i>Bacillus</i> sp.
SEQUENCE: 88		
HHNGTNGTMM QYFEWYLPND GNHWNRLRSR ASNLDKKGIS AVWIPPAWKG ASQNDVGYGA	60	
YDLYDLGEFN QKGTVRTKYG TRNQLQAAVN ALKSNGIQVY GDFVVMNHKG ADATEMVR	120	A
EVNPNNRNQE VSGEYTIIEAW TKFDPPGRGN THSNFKWRWY HFDGVDWDQS RKLNNRIYKF	180	
RGDGKAWDWE VDTENGNYDV LMYADIDMDH PEVNVELRW GVWYTNLGL DGFRIDAVKH	240	
IKYSFTRDWI NHVRSATGKN MFAVAEFWKN DLGAIENYLN KTNWNHSVFD VPLHYNLYNA	300	
SKSGGNYDMR QIFNGTVVQR HPMHAVTFVD NHDSQPEEAL ESFVVEWFKP LAYALTLTRE	360	
QGYPSVVFYGD YYGIPHTHGV PAMSKIDPIL EARQKYAYGR QNDYLDHHNI IGWTREGNTA	420	
HPNSGLATIM SDGAGGNKWM FVGRNKAGQV WSDITGNRAG TVTINADGWG NFSVNGGSVS	480	
IWVNK	485	
SEQ ID NO: 89		moltype = AA length = 485
FEATURE		Location/Qualifiers
source	1..485	
		mol_type = protein
		organism = <i>Bacillus</i> sp. NCIB 12513
SEQUENCE: 89		
HHNGTNGTMM QYFEWHLPPND GNHWNRLRDD ASNLRNRGIT AIWIPPAWKG TSQNDVGYGA	60	
YDLYDLGEFN QKGTVRTKYG TRSOLQESAIH ALKNNGVQVY GDFVVMNHKG ADATENVLA	120	
EVNPNNRNQE VTEGTYIEAW TRFDPPGRGN THSSFKWRWY HFDGVDWDQS RQLNNRIYKF	180	
RGHGKAWDWE VDTENGNYDV LMYADIDMDH PEVNVELRW GVWYTNLGL DGFRIDAVKH	240	
IKYSFTRDWI THVRNATGKE MFAVAEFWKN DLGAIENYLN KTNWNHSVFD VPLHYNLYNA	300	
SNSGGNYDMA KLLNGTVVQK HPMHAVTFVD NHDSQPGESL ESFVQEWFKP LAYALTLTRE	360	
QGYPSVVFYGD YYGIPHTHGV PAMSKIDPIL EARQNFAYGT QHDYFDHHNI IGWTREGNTT	420	
HPNSGLATIM SDGPGGEKWM YVGQNKAGQV WHDITGNPKG TVTINADGWG NFSVNGGSVS	480	
IWVKR	485	
SEQ ID NO: 90		moltype = AA length = 485
FEATURE		Location/Qualifiers
source	1..485	
		mol_type = protein
		organism = <i>Bacillus</i> sp. no. 707
SEQUENCE: 90		
HHNGTNGTMM QYFEWYLPND GNHWNRLNSD ASNLSKGIT AVWIPPAWKG ASQNDVGYGA	60	
YDLYDLGEFN QKGTVRTKYG TRSOLQAAVT SLKNNGIQVY GDFVVMNHKG ADATEMVR	120	
EVNPNNRNQE VTGEYTIIEAW TRFDPPGRGN THSSFKWRWY HFDGVDWDQS RRLNNRIYKF	180	
RGHGKAWDWE VDTENGNYDV LMYADIDMDH PEVNVELRW GVWYTNLGL DGFRIDAVKH	240	
IKYSFTRDWI NHVRSATGKN MFAVAEFWKN DLGAIENYLQ KTNWNHSVFD VPLHYNLYNA	300	
SKSGGNYDMA NIFNGTVVQR HPSHAVTFVD NHDSQPEEAL ESFVQEWFKP LAYALTLTRE	360	
QGYPSVVFYGD YYGIPHTHGV PAMSKIDPIL EARQKYAYGR QNDYLDHHNI LGWTREGNTA	420	
HPNSGLATIM SDGAGGSKWM FVGRNKAGQV WSDITGNRTG TVTINADGWG NFSVNGGSVS	480	
IWVNK	485	
SEQ ID NO: 91		moltype = AA length = 485
FEATURE		Location/Qualifiers
REGION	1..485	
		note = fusion protein
source	1..485	
		mol_type = protein
		organism = synthetic construct
SEQUENCE: 91		
HHNGTNGTMM QYFEWNPND GQHWNRLHNN AQNLKNAGIT AIWIPPAWKG TSQNDVGYGA	60	
YDLYDLGEFN QKGTVRTKYG TKAELERAIR SLKANGIQVY GDFVVMNHKG ADFTERVQAV	120	
EVNPQNRRQE VSGTYIEAW TGFNFPPGRGN QHSSFKWRWY HFDGTDWDQS RQLANRIYKF	180	
RGDGKAWDWE VDTENGNYDV LMYADIDMDH PEVNVELRW GVWYANTLNL DGFRIDAVKH	240	
IKFSFTRDWL GHVRGQTGK LFAVAWEWN DLGAIENYLQ KTNWMTMSAFD VPLHYNLYQA	300	
SNSGGNYDMA NLLNGTLVQR HPSHAVTFVD NHDTQPGEAL ESFVQGWFKP LAYALTLTRE	360	
QGYPQVFYGD YYGIPSDGPV SYRQQIDPLL KARQQYAYGT QHDYLDHQDV IGWTREGDSA	420	
HAGSGLATVM SDGPGGSKTM YVGTAHAGQV FKDITGNRTD TTVTINSAGNG TFPNCNGGSVS	480	
IWVKQ	485	

1. A cleaning composition comprising a DNase, at least one carbohydrase and a cleaning component, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase.
2. A cleaning composition of claim 1, wherein the carbohydrase is a cellulase.
3. A cleaning composition of claim 1, wherein the carbohydrase is an amylase.
4. A cleaning composition of claim 1, wherein the carbohydrase is a mannanase.

5. A cleaning composition of claim 1, wherein the carbohydrase is a xylanase.
6. A cleaning composition of claim 1, wherein the DNase is obtained from bacteria or fungi.
7. A cleaning composition of claim 1, wherein the DNase is obtained from *Bacillus*.
8. A cleaning composition of claim 7, wherein the DNase comprises one or both of the motif(s) [D/M/L][S/T]GYSR [D/N](SEQ ID NO: 73) or ASXNRSKG (SEQ ID NO: 74).

9. A cleaning composition of claim **6**, wherein the DNase has at least 60% sequence identity to the amino acid sequence shown in SEQ ID NO 13.

10. A cleaning composition of claim **6**, wherein the DNase has at least 60% sequence identity to the amino acid sequence shown in SEQ ID NO: 65.

11. A cleaning composition of claim **6**, wherein the DNase has at least 60% sequence identity to the amino acid sequence shown in SEQ ID NO: 66.

12. A cleaning composition of claim **6**, wherein the DNase sequence identity to the amino acid sequence shown in SEQ ID NO: 67.

13. A cleaning composition of claim **6**, wherein the DNase is sequence identity to the amino acid sequence shown in SEQ ID NO: 68.

14. A cleaning composition of claim **1**, wherein the amount of DNase in the composition is from 0.01 to 1000 ppm and the amount of carbohydrase is from 0.01 to 1000 ppm.

15. A cleaning composition of claim **1**, wherein the cleaning component is selected from surfactants, builders and bleach components.

16. (canceled)

17. A method of formulating a cleaning composition of claim **1**, comprising adding a DNase, a carbohydrase and at least one cleaning component.

18. A kit intended for deep cleaning, wherein the kit comprises a solution of an enzyme mixture comprising a DNase and a carbohydrase, wherein the carbohydrase is a cellulase, an amylase, a mannanase or a xylanase.

19. A method of deep cleaning an item, comprising:

a) contacting the item with a cleaning composition of claim **1**; and

b) optionally rinsing the item.

20. The method of claim **19**, wherein the item is a textile.

* * * * *