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### System and method for calculating focus variation for a digital microscope

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#### Abstract

Apparatus and methods are described for use with a digital microscope unit that includes a digital microscope. A biological cell sample that is disposed within a sample carrier is received into the digital microscope unit. It is determined that there is a variation in the focal depth of the biological sample with respect to the microscope due to curvature in the sample carrier and/or due to tolerance in setup of the microscope. In response to determining that there is the variation in the focal depth of the biological sample with respect to the microscope, the variation in the focal depth of the biological sample with respect to the microscope is accounted for. Other applications are also described.

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## References Cited

### U.S. PATENT DOCUMENTS

Patent No.	Issued Date	Patentee Name	U.S. Cl.	CPC
3603156	12/1970	Konkol	N/A	N/A
3676076	12/1971	Grady	N/A	N/A
3786184	12/1973	Pieters	250/204	G02B 27/40
3967056	12/1975	Yata et al.	N/A	N/A
4076419	12/1977	Kleker	N/A	N/A
4209548	12/1979	Bacus	N/A	N/A
4350884	12/1981	Dieter	N/A	N/A
4454235	12/1983	Johnson	N/A	N/A
4494479	12/1984	Drury et al.	N/A	N/A
4580895	12/1985	Patel	N/A	N/A
4700298	12/1986	Palcic et al.	N/A	N/A
4761381	12/1987	Blatt et al.	N/A	N/A
4774192	12/1987	Terminiello et al.	N/A	N/A
4803352	12/1988	Bierleutgeb	N/A	N/A
4849340	12/1988	Oberhardt	N/A	N/A
4902101	12/1989	Fujihara et al.	N/A	N/A
5001067	12/1990	Coleman et al.	N/A	N/A
5064282	12/1990	Curtis	N/A	N/A
5229265	12/1992	Tometsko	N/A	N/A
5300779	12/1993	Hillman et al.	N/A	N/A
5331958	12/1993	Oppenheimer	N/A	N/A
5430542	12/1994	Shepherd et al.	N/A	N/A
5499097	12/1995	Ortyn	N/A	N/A
5672861	12/1996	Fairley et al.	N/A	N/A
5674457	12/1996	Williamsson et al.	N/A	N/A

5745804	12/1997	Iwane	N/A	N/A
5782770	12/1997	Mooradian et al.	N/A	N/A
5827190	12/1997	Palcic et al.	N/A	N/A
5834217	12/1997	Levine et al.	N/A	N/A
5932872	12/1998	Price	N/A	N/A
5948686	12/1998	Wardlaw	N/A	N/A
5985595	12/1998	Krider et al.	N/A	N/A
6027695	12/1999	Oldenburg et al.	N/A	N/A
6064474	12/1999	Lee et al.	N/A	N/A
6074879	12/1999	Zelmnanovic et al.	N/A	N/A
6101404	12/1999	Yoon et al.	N/A	N/A
6262423	12/2000	Hell et al.	N/A	N/A
6262798	12/2000	Shepherd et al.	N/A	N/A
6320979	12/2000	Melen	N/A	N/A
6339472	12/2001	Hafeman et al.	N/A	N/A
6350613	12/2001	Wardlaw et al.	N/A	N/A
6448024	12/2001	Bruegger	N/A	N/A
6519355	12/2002	Nelson	N/A	N/A
6538249	12/2002	Takane	850/10	G06T 5/50
6554788	12/2002	Hunley et al.	N/A	N/A
6582964	12/2002	Samsoondar et al.	N/A	N/A
6611777	12/2002	Samsoondar	N/A	N/A
6632681	12/2002	Chu	N/A	N/A
6658143	12/2002	Hansen et al.	N/A	N/A
6664528	12/2002	Cartlidge et al.	N/A	N/A
6711516	12/2003	Samsoondar	N/A	N/A
6819408	12/2003	Scrivens et al.	N/A	N/A
6831733	12/2003	Pettersson et al.	N/A	N/A
6834237	12/2003	Noergaard et al.	N/A	N/A
6836559	12/2003	Abdel-Fattah et al.	N/A	N/A
6842233	12/2004	Narisada et al.	N/A	N/A
6866823	12/2004	Wardlaw	N/A	N/A
6872930	12/2004	Cartlidge et al.	N/A	N/A
6898451	12/2004	Wuori	N/A	N/A
6903323	12/2004	Cartlidge et al.	N/A	N/A
6929953	12/2004	Wardlaw	N/A	N/A
6949384	12/2004	Samsoondar	N/A	N/A
6955872	12/2004	Maples et al.	N/A	N/A
6956650	12/2004	Boas et al.	N/A	N/A
6989891	12/2005	Braig et al.	N/A	N/A
7027628	12/2005	Gagnon	382/233	G06T 5/50
7030351	12/2005	Wasserman	250/201.3	H04N 23/56
7034883	12/2005	Rosenqvist	N/A	N/A
7105795	12/2005	Cartlidge et al.	N/A	N/A
7132636	12/2005	Cartlidge et al.	N/A	N/A
7133547	12/2005	Marcelpoil et al.	N/A	N/A
7151246	12/2005	Fein et al.	N/A	N/A
7155049	12/2005	Wetzel	378/43	G02B 21/0016
7214936	12/2006	Sato	250/310	H01J 37/222

7248716	12/2006	Fein et al.	N/A	N/A
7274810	12/2006	Reeves et al.	N/A	N/A
7283217	12/2006	Ikeuchi et al.	N/A	N/A
7288751	12/2006	Cartlidge et al.	N/A	N/A
7305109	12/2006	Gagnon	382/233	G01N 15/1433
7324694	12/2007	Chapoulaud et al.	N/A	N/A
7329537	12/2007	Qiu	N/A	N/A
7338168	12/2007	Cartlidge et al.	N/A	N/A
7344890	12/2007	Perez et al.	N/A	N/A
7346205	12/2007	Walker, Jr.	N/A	N/A
7369696	12/2007	Arini et al.	N/A	N/A
7385168	12/2007	Cartlidge et al.	N/A	N/A
7411680	12/2007	Chang et al.	N/A	N/A
7417213	12/2007	Krief	250/201.3	G02B 21/244
7439478	12/2007	Cartlidge et al.	N/A	N/A
7450223	12/2007	Ikeuchi et al.	N/A	N/A
7450762	12/2007	Morell	N/A	N/A
7460222	12/2007	Kalveram et al.	N/A	N/A
7490085	12/2008	Walker et al.	N/A	N/A
7493219	12/2008	Qi et al.	N/A	N/A
7580120	12/2008	Hamada et al.	N/A	N/A
7599893	12/2008	Sapir et al.	N/A	N/A
7601938	12/2008	Cartlidge et al.	N/A	N/A
7602954	12/2008	Marcelpoil et al.	N/A	N/A
7605356	12/2008	Krief	250/201.3	G06T 5/50
7609369	12/2008	Simon-Lopez	N/A	N/A
7630063	12/2008	Padmanabhan et al.	N/A	N/A
7633604	12/2008	Ikeuchi et al.	N/A	N/A
7638748	12/2008	Krief et al.	N/A	N/A
7663738	12/2009	Johansson	N/A	N/A
7668362	12/2009	Olson	382/128	G02B 21/245
7692131	12/2009	Fein et al.	N/A	N/A
7697764	12/2009	Kataoka	N/A	N/A
7702181	12/2009	Gouch	N/A	N/A
7706862	12/2009	Alfano et al.	N/A	N/A
7713474	12/2009	Schulman et al.	N/A	N/A
7747153	12/2009	Ibaraki	N/A	N/A
7765069	12/2009	Ostoich et al.	N/A	N/A
7777869	12/2009	Nerin et al.	N/A	N/A
7787109	12/2009	Dosmann et al.	N/A	N/A
7796797	12/2009	Nakaya et al.	N/A	N/A
7863552	12/2010	Cartlidge et al.	N/A	N/A
7869009	12/2010	Dosmann et al.	N/A	N/A
7894047	12/2010	Hamada et al.	N/A	N/A
7911617	12/2010	Padmanabhan	N/A	N/A
7925070	12/2010	Sumida et al.	N/A	N/A

7929121	12/2010	Wardlaw et al.	N/A	N/A
7933435	12/2010	Hunter et al.	N/A	N/A
7936913	12/2010	Nordell et al.	N/A	N/A
7951599	12/2010	Levine et al.	N/A	N/A
7995200	12/2010	Matsumoto	N/A	N/A
7998435	12/2010	Reed	N/A	N/A
8000511	12/2010	Perz	250/201.3	G02B 21/244
8044974	12/2010	Sumida et al.	N/A	N/A
8045782	12/2010	Li et al.	N/A	N/A
8055471	12/2010	Qi et al.	N/A	N/A
8064680	12/2010	Ramoser et al.	N/A	N/A
8077296	12/2010	Wardlaw et al.	N/A	N/A
8081303	12/2010	Levine et al.	N/A	N/A
8105554	12/2011	Kanigan et al.	N/A	N/A
8125643	12/2011	Hansen et al.	N/A	N/A
D655421	12/2011	Lee et al.	N/A	N/A
8131035	12/2011	Grady et al.	N/A	N/A
8131052	12/2011	Alexandrov	N/A	N/A
8150114	12/2011	Svanberg et al.	N/A	N/A
8154713	12/2011	Simon-Lopez	N/A	N/A
8165385	12/2011	Reeves et al.	N/A	N/A
8175353	12/2011	Westphal et al.	N/A	N/A
8184273	12/2011	Dosmann et al.	N/A	N/A
8216832	12/2011	Battrell et al.	N/A	N/A
8224058	12/2011	Lindberg et al.	N/A	N/A
8269954	12/2011	Levine et al.	N/A	N/A
8280134	12/2011	Hoyt	N/A	N/A
8310659	12/2011	Wardlaw et al.	N/A	N/A
8314837	12/2011	Kenny	348/79	G02B 21/367
8320655	12/2011	Sarachan et al.	N/A	N/A
8331627	12/2011	Xiong	382/109	G06T 3/4053
8331642	12/2011	Zerfass et al.	N/A	N/A
8339586	12/2011	Zahniser et al.	N/A	N/A
8345227	12/2012	Zahniser et al.	N/A	N/A
8351676	12/2012	Dai et al.	N/A	N/A
8363221	12/2012	Hansen et al.	N/A	N/A
8379944	12/2012	Grady et al.	N/A	N/A
8428331	12/2012	DiMarzio et al.	N/A	N/A
8432392	12/2012	Kim et al.	N/A	N/A
8477294	12/2012	Zahniser et al.	N/A	N/A
8481303	12/2012	Faris et al.	N/A	N/A
8488111	12/2012	Zahniser et al.	N/A	N/A
8491499	12/2012	Choi et al.	N/A	N/A
8526704	12/2012	Dobbe	N/A	N/A
8565480	12/2012	Eichhorn	382/103	G02B 30/52
8570496	12/2012	Chen	N/A	N/A
8582924	12/2012	De La Torre- Bueno et al.	N/A	N/A

8638427	12/2013	Wardlaw et al.	N/A	N/A
8712142	12/2013	Rajpoot et al.	N/A	N/A
8736824	12/2013	Matsui et al.	N/A	N/A
8743195	12/2013	Najmabadi	382/128	G06T 7/30
8744165	12/2013	Liu et al.	N/A	N/A
8778687	12/2013	Levine et al.	N/A	N/A
8792693	12/2013	Satish et al.	N/A	N/A
8837803	12/2013	Wang et al.	N/A	N/A
8849024	12/2013	Shinoda et al.	N/A	N/A
8873827	12/2013	McCulloch et al.	N/A	N/A
8877458	12/2013	Maurer	N/A	N/A
8878923	12/2013	Henderson	382/199	G02B 21/244
8885154	12/2013	Wardlaw et al.	N/A	N/A
8885912	12/2013	Sui	N/A	N/A
8891851	12/2013	Spaulding	N/A	N/A
8922761	12/2013	Zahniser et al.	N/A	N/A
8942458	12/2014	Takahashi et al.	N/A	N/A
8964171	12/2014	Zahniser et al.	N/A	N/A
8994930	12/2014	Levine et al.	N/A	N/A
9012868	12/2014	Courtney et al.	N/A	N/A
9041791	12/2014	Zahniser	348/79	G02B 21/244
9041792	12/2014	Van Leeuwen et al.	N/A	N/A
9046473	12/2014	Levine et al.	N/A	N/A
9050595	12/2014	Miller et al.	N/A	N/A
9064301	12/2014	Zie et al.	N/A	N/A
9088729	12/2014	Takayama	N/A	G02B 21/367
9176121	12/2014	Winkelman et al.	N/A	N/A
9186843	12/2014	Chan et al.	N/A	N/A
9235041	12/2015	Crandall	N/A	G02B 21/367
9240043	12/2015	Christiansen et al.	N/A	N/A
9322767	12/2015	Ehrenkranz	N/A	N/A
9329129	12/2015	Pollak et al.	N/A	N/A
9332190	12/2015	Murakami	N/A	G02B 21/365
9342734	12/2015	Lin et al.	N/A	N/A
9349208	12/2015	Eichhorn	N/A	G06T 15/08
9404737	12/2015	Segale	N/A	G02B 26/10
9404852	12/2015	Braig et al.	N/A	N/A
9470609	12/2015	Wimberger-Friedl et al.	N/A	N/A
9477875	12/2015	Ohya et al.	N/A	N/A
9522396	12/2015	Bachelet et al.	N/A	N/A
9588033	12/2016	Zahniser et al.	N/A	N/A
9690974	12/2016	Obrien	N/A	G01N 15/1433

9767343	12/2016	Jones et al.	N/A	N/A
9930241	12/2017	Oshima	N/A	G02B 21/244
9934571	12/2017	Ozaki et al.	N/A	N/A
10024858	12/2017	Smith et al.	N/A	N/A
10061972	12/2017	Champlin et al.	N/A	N/A
10093957	12/2017	Pollak et al.	N/A	N/A
10132738	12/2017	Obrien	N/A	G02B 21/365
10139613	12/2017	Hing	N/A	G02B 21/367
10169861	12/2018	Ozaki et al.	N/A	N/A
10176565	12/2018	Greenfield et al.	N/A	N/A
10210376	12/2018	Hedlund	N/A	G03B 13/32
10269094	12/2018	Fuchs	N/A	G06T 5/50
10281386	12/2018	Hsu et al.	N/A	N/A
10330910	12/2018	Iwase	N/A	G02B 21/367
10353190	12/2018	Hulsken	N/A	G02B 21/367
10488644	12/2018	Eshel et al.	N/A	N/A
11207836	12/2020	Ramos	N/A	B33Y 10/00
2002/0009711	12/2001	Wada et al.	N/A	N/A
2002/0028158	12/2001	Wardlaw	N/A	N/A
2002/0028471	12/2001	Oberhardt	N/A	N/A
2003/0017085	12/2002	Kercso et al.	N/A	N/A
2003/0161514	12/2002	Curry	N/A	N/A
2003/0197925	12/2002	Hamborg	N/A	N/A
2003/0224522	12/2002	de Jong et al.	N/A	N/A
2003/0227612	12/2002	Fein et al.	N/A	N/A
2003/0227673	12/2002	Nakagawa	359/368	G02B 21/365
2003/0228038	12/2002	Douglass	382/128	G06T 7/60
2003/0231791	12/2002	Torre-Bueno et al.	N/A	N/A
2004/0004614	12/2003	Bacus	345/419	G02B 21/367
2004/0132171	12/2003	Rule et al.	N/A	N/A
2004/0170312	12/2003	Soenksen	N/A	N/A
2004/0185447	12/2003	Maples et al.	N/A	N/A
2004/0218804	12/2003	Affleck et al.	N/A	N/A
2004/0240050	12/2003	Ogihara	N/A	N/A
2004/0241677	12/2003	Lin et al.	N/A	N/A
2004/0256538	12/2003	Olson	250/201.3	G02B 21/002
2005/0089208	12/2004	Dong	382/133	G06T 3/4038
2005/0109959	12/2004	Wasserman et al.	N/A	N/A
2005/0270611	12/2004	Oshiro	713/186	G02B 21/244
2005/0282292	12/2004	Torre-Bueno	436/180	G01N 1/31
2005/0286800	12/2004	Gouch	N/A	N/A

2006/0001955	12/2005	Kinney	359/391	G02B 21/244
2006/0007345	12/2005	Olson	348/345	G02B 21/245
2006/0045505	12/2005	Zeineh	396/89	G02B 21/367
2006/0063185	12/2005	Vannier	N/A	N/A
2006/0098861	12/2005	See	382/145	G01B 11/08
2006/0187442	12/2005	Chang et al.	N/A	N/A
2006/0223052	12/2005	MacDonald et al.	N/A	N/A
2006/0223165	12/2005	Chang et al.	N/A	N/A
2006/0226362	12/2005	Kitsuki et al.	N/A	N/A
2007/0054350	12/2006	Walker	N/A	N/A
2007/0243117	12/2006	Wardlaw	N/A	N/A
2007/0250301	12/2006	Vaisberg et al.	N/A	N/A
2007/0252984	12/2006	Van Beek et al.	N/A	N/A
2008/0012978	12/2007	Wang	N/A	N/A
2008/0020128	12/2007	van Ryper	359/368	G02B 21/362
2008/0059135	12/2007	Murugkar et al.	N/A	N/A
2008/0118399	12/2007	Fleming	N/A	N/A
2008/0187466	12/2007	Wardlaw	N/A	N/A
2008/0212069	12/2007	Goldberg et al.	N/A	N/A
2008/0260369	12/2007	Ibaraki	N/A	N/A
2008/0273776	12/2007	Krief et al.	N/A	N/A
2008/0305514	12/2007	Alford et al.	N/A	N/A
2009/0046909	12/2008	Rutenberg	382/128	G06T 5/50
2009/0066934	12/2008	Gao et al.	N/A	N/A
2009/0075324	12/2008	Pettersson	N/A	N/A
2009/0128618	12/2008	Fahn et al.	N/A	N/A
2009/0185734	12/2008	Lindberg et al.	N/A	N/A
2009/0191098	12/2008	Beard et al.	N/A	N/A
2009/0195688	12/2008	Henderson	348/E5.024	G02B 21/245
2009/0213214	12/2008	Yamada	348/80	G06T 7/0012
2009/0238437	12/2008	Levine et al.	N/A	N/A
2009/0258347	12/2008	Scott	N/A	N/A
2009/0269799	12/2008	Winkelman et al.	N/A	N/A
2009/0291854	12/2008	Wiesinger-Mayr et al.	N/A	N/A
2010/0104169	12/2009	Yamada	N/A	N/A
2010/0112631	12/2009	Hur et al.	N/A	N/A
2010/0120129	12/2009	Amshey et al.	N/A	N/A
2010/0152054	12/2009	Love	N/A	N/A
2010/0157086	12/2009	Segale et al.	N/A	N/A
2010/0172020	12/2009	Price	359/383	G02B 21/244
2010/0234703	12/2009	Sterling et al.	N/A	N/A
2010/0254596	12/2009	Xiong	382/159	G06T 3/4053
2010/0256918	12/2009	Chen et al.	N/A	N/A



2010/0265323	12/2009	Perz	N/A	N/A
2010/0300563	12/2009	Ramunas et al.	N/A	N/A
2011/0007178	12/2010	Kahlman	N/A	N/A
2011/0009163	12/2010	Fletcher et al.	N/A	N/A
2011/0030458	12/2010	Paark et al.	N/A	N/A
2011/0102571	12/2010	Yoneyama	359/368	G02B 21/367
2011/0123398	12/2010	Carrilho et al.	N/A	N/A
2011/0144480	12/2010	Lu et al.	N/A	N/A
2011/0149097	12/2010	Danuser et al.	N/A	N/A
2011/0151502	12/2010	Kendall et al.	N/A	N/A
2011/0178716	12/2010	Krockenberger et al.	N/A	N/A
2011/0212486	12/2010	Yamada	435/40.5	G02B 21/365
2011/0249910	12/2010	Henderson	382/278	G06V 10/993
2011/0260055	12/2010	Wang	356/612	G01N 23/20058
2011/0275111	12/2010	Pettigrew et al.	N/A	N/A
2011/0301012	12/2010	Dolecek et al.	N/A	N/A
2012/0002195	12/2011	Wu et al.	N/A	N/A
2012/0007977	12/2011	Yamamoto	348/79	G02B 27/0075
2012/0021951	12/2011	Hess et al.	N/A	N/A
2012/0030618	12/2011	Leong et al.	N/A	N/A
2012/0044342	12/2011	Hing	348/79	G02B 21/365
2012/0058504	12/2011	Li et al.	N/A	N/A
2012/0092477	12/2011	Kawano	348/79	G02B 21/365
2012/0120221	12/2011	Dong et al.	N/A	N/A
2012/0162495	12/2011	Ogawa	N/A	N/A
2012/0169863	12/2011	Bachelet et al.	N/A	N/A
2012/0225446	12/2011	Wimberger-Friedl et al.	N/A	N/A
2012/0249770	12/2011	Hamm et al.	N/A	N/A
2012/0288157	12/2011	Kishima	382/106	G06T 7/571
2012/0312957	12/2011	Loney	250/201.3	G02B 21/245
2012/0320045	12/2011	Yao et al.	N/A	N/A
2013/0016885	12/2012	Tsujimoto	382/128	G06T 5/50
2013/0023007	12/2012	Zahniser et al.	N/A	N/A
2013/0078668	12/2012	Levine et al.	N/A	N/A
2013/0100272	12/2012	Price	348/E5.045	G02B 21/365
2013/0130262	12/2012	Battrell et al.	N/A	N/A
2013/0170730	12/2012	Yu et al.	N/A	N/A
2013/0176551	12/2012	Wardlaw et al.	N/A	N/A
2013/0258058	12/2012	Watanabe	348/46	G06T 5/50

2013/0273968	12/2012	Rhoads	455/566	H04W 4/50
2013/0284924	12/2012	Mizuochi	250/310	H01J 37/21
2013/0290225	12/2012	Kamath et al.	N/A	N/A
2013/0342902	12/2012	Krueger et al.	N/A	N/A
2014/0139625	12/2013	Mathuis et al.	N/A	N/A
2014/0139630	12/2013	Kowalevich	N/A	N/A
2014/0186859	12/2013	Calderwood	348/79	G01N 33/54366
2014/0205176	12/2013	Obrien et al.	N/A	N/A
2014/0347459	12/2013	Greenfield	382/133	G02B 21/361
2015/0006105	12/2014	Zhou	702/150	G02B 21/26
2015/0037806	12/2014	Pollack et al.	N/A	N/A
2015/0124082	12/2014	Kato	348/135	G06V 20/693
2015/0168705	12/2014	O'Neill	348/80	G02B 21/244
2015/0185459	12/2014	Clark et al.	N/A	N/A
2015/0190063	12/2014	Zakharov et al.	N/A	N/A
2015/0278575	12/2014	Allano et al.	N/A	N/A
2015/0302237	12/2014	Ohya et al.	N/A	N/A
2015/0316477	12/2014	Pollak et al.	N/A	N/A
2016/0202460	12/2015	Zheng	N/A	N/A
2016/0208306	12/2015	Pollak et al.	N/A	N/A
2016/0246046	12/2015	Yorav Raphael et al.	N/A	N/A
2016/0279633	12/2015	Bachelet et al.	N/A	N/A
2017/0052110	12/2016	Malissek et al.	N/A	N/A
2017/0160185	12/2016	Minemura et al.	N/A	N/A
2017/0218425	12/2016	Chen et al.	N/A	N/A
2017/0307496	12/2016	Zahniser et al.	N/A	N/A
2017/0328924	12/2016	Jones et al.	N/A	N/A
2018/0246313	12/2017	Eshel et al.	N/A	N/A
2018/0296102	12/2017	Satish et al.	N/A	N/A
2019/0002950	12/2018	Pollak et al.	N/A	N/A
2019/0087953	12/2018	Yorav-Raphael et al.	N/A	N/A
2019/0347467	12/2018	Ohsaka et al.	N/A	N/A
2020/0209604	12/2019	Chen et al.	N/A	N/A

#### FOREIGN PATENT DOCUMENTS

Patent No.	Application Date	Country	CPC
2655024	12/2013	CA	N/A
101403650	12/2009	CN	N/A
0073551	12/1982	EP	N/A
0479231	12/1991	EP	N/A
1 381 229	12/2003	EP	N/A
1698883	12/2005	EP	N/A
2145684	12/2009	EP	N/A

3001174	12/2015	EP	N/A
3 123 927	12/2016	EP	N/A
2211165	12/2016	EP	N/A
3482189	12/2018	EP	N/A
1 873 232	12/2019	EP	N/A
61198204	12/1985	JP	N/A
H11-73903	12/1998	JP	N/A
2000-199845	12/1999	JP	N/A
2003-315684	12/2002	JP	N/A
2004-257768	12/2003	JP	N/A
2006-294301	12/2005	JP	N/A
2006-301270	12/2005	JP	N/A
2007-040814	12/2006	JP	N/A
2007-93470	12/2006	JP	N/A
2009223164	12/2008	JP	G02B 21/244
2017-209530	12/2016	JP	N/A
1985/005446	12/1984	WO	N/A
1996/001438	12/1995	WO	N/A
1996/012981	12/1995	WO	N/A
1996/013615	12/1995	WO	N/A
2000/006765	12/1999	WO	N/A
2000/052195	12/1999	WO	N/A
2000/055572	12/1999	WO	N/A
2002/033400	12/2001	WO	N/A
02/39059	12/2001	WO	N/A
03/056327	12/2002	WO	N/A
2003/073365	12/2002	WO	N/A
2003/081525	12/2002	WO	N/A
2004/111610	12/2003	WO	N/A
2005/121863	12/2004	WO	N/A
2006/121266	12/2005	WO	N/A
2008/063135	12/2007	WO	N/A
2010/056740	12/2009	WO	N/A
2010/116341	12/2009	WO	N/A
2010/126903	12/2009	WO	N/A
2011/076413	12/2010	WO	N/A
2011/123070	12/2010	WO	N/A
2011/143075	12/2010	WO	N/A
2012/000102	12/2011	WO	N/A
2012/030313	12/2011	WO	N/A
2012/107468	12/2011	WO	N/A
2012/090198	12/2011	WO	N/A
2012/154333	12/2011	WO	N/A
2013/041951	12/2012	WO	N/A
2013/098821	12/2012	WO	N/A
2014/159620	12/2013	WO	N/A
2014/188405	12/2013	WO	N/A
2015/001553	12/2014	WO	N/A
2015/029032	12/2014	WO	N/A
2016/030897	12/2015	WO	N/A

2017/046799	12/2016	WO	N/A
2017/168411	12/2016	WO	N/A
2017/195205	12/2016	WO	N/A
2017/195208	12/2016	WO	N/A
2018/009920	12/2017	WO	N/A
2019/035084	12/2018	WO	N/A
2019/097387	12/2018	WO	N/A
2019/102277	12/2018	WO	N/A
2019/198094	12/2018	WO	N/A
2021/079305	12/2020	WO	N/A
2021/079306	12/2020	WO	N/A

## OTHER PUBLICATIONS

Guy, Rebecca, Paul Liu, Peter Pennefather and Ian Crandall, "The use of fluorescence enhancement to improve the microscopic diagnosis of falciparum malaria", Malaria Journal 2007 6:89, <https://malariajournal.biomedcentral.com/articles/10.1186/1475-2875-6-89>, (Jul. 9, 2007). cited by applicant

Rappaz, Benjamin, et al. "Comparative study of human erythrocytes by digital holographic microscopy, confocal microscopy, and impedance volume analyzer." Cytometry Part A 73.10 (2008): 895-903. cited by applicant

An Office Action dated May 16, 2024 which issued during the prosecution of U.S. Appl. No. 17/063,320. cited by applicant

An Office Action dated Feb. 17, 2023 which issued during the prosecution of U.S. Appl. No. 17/576,023. cited by applicant

An Office Action dated Sep. 14, 2023 which issued during the prosecution of U.S. Appl. No. 17/063,320. cited by applicant

Notice of Allowance dated Jun. 22, 2023 which issued during the prosecution of U.S. Appl. No. 17/576,023. cited by applicant

Communication dated Aug. 19, 2024, issued in U.S. Appl. No. 18/376,680. cited by applicant

Rodrigo et al., "Fast control of temporal and spatial coherence properties of microscope illumination using DLP projector", Proc. of SPIE, 2015, vol. 9336, pp. 93360F-1 to 93360F-6 (7 pages total). cited by applicant

Pan et al., "High-resolution and large field-of-view Fourier ptychographic microscopy and its applications in biomedicine", Reports on Progress in Physics, 2020, vol. 83, 1 page (abstract only). cited by applicant

Communication dated Mar. 11, 2025, issued in U.S. Appl. No. 18/376,680. cited by applicant

"Blood specimens: Microscopic Examination", Centers for Disease Control and Prevention CDC, Diagnostic Procedures, 2009, [http://www.dpd.cdc.gov/dpdx/HTML/Frames/DiagnosticProcedures/body\\_dp\\_bloodexamin.htm](http://www.dpd.cdc.gov/dpdx/HTML/Frames/DiagnosticProcedures/body_dp_bloodexamin.htm). cited by applicant

A European Examination Report dated Dec. 10, 2021 which issued during the prosecution of Applicant's European App No. 15778059.4. cited by applicant

A European Search Report dated Dec. 14, 2016, which issued during the prosecution of Applicant's European App No. 14800352.8. cited by applicant

A European Search Report dated Mar. 23, 2017, which issued during the prosecution of Applicant's European App No. 14839661.7. cited by applicant

A Final Office Action dated Jul. 29, 2019 from the United States Patent and Trademark Office in U.S. Appl. No. 14/914,329. cited by applicant

A Notice of Allowance issued on Jan. 27, 2021 for U.S. Appl. No. 16/592,335. cited by applicant

A Notice of Allowance issued on May 13, 2021 for U.S. Appl. No. 16/592,335. cited by applicant

Agero, U., Mesquita, L.G., Neves, B.R.A., Gazzinelli, R.T. and Mesquita, O.N., 2004. Defocusing microscopy. *Microscopy research and technique*, 65(3), pp. 159-165. cited by applicant

Ahirwar, Neetu, Sapnojit Pattnaik, and Bibhudendra Acharya. "Advanced image analysis based system for automatic detection and classification of malarial parasite in blood images." *International Journal of Information Technology and Knowledge Management* 5.1 (2012): 59-64. cited by applicant

An Advisory Action dated Dec. 27, 2019 which issued during the prosecution of U.S. Appl. No. 14/914,329. cited by applicant

An Examination Report for Indian Application No. 201727008982 on Jan. 29, 2021. cited by applicant

An Extended European Search Report issued for European Patent Application No. 21154596.7 on Jul. 23, 2021. cited by applicant

An Extended European Search Report issued for European Patent Application No. 21194040.8 on Jan. 18, 2022. cited by applicant

An Indian Examination Report dated Apr. 22, 2020. which issued during the prosecution of Indian App No. 201617009399. cited by applicant

An International Preliminary Report on Patentability dated Feb. 28, 2017, which issued during the prosecution of Applicant's PCT/IL2015/050864. cited by applicant

An International Search Report and a Written Opinion both dated Feb. 12, 2015, which issued during the prosecution of Applicant's PCT/IL2014/050770. cited by applicant

An International Search Report and a Written Opinion both dated Jan. 15, 2016, which issued during the prosecution of Applicant's PCT/IL2015/050864. cited by applicant

An International Search Report and a Written Opinion both dated Sep. 29, 2014, which issued during the prosecution of Applicant's PCT/IL2014/050423. cited by applicant

An International Search Report and Written Opinion for Application No. PCT/IB2020/061731 issued on Feb. 22, 2021. cited by applicant

An International Search Report and Written Opinion for PCT Application No. PCT/IB2020/061732 mailed on May 7, 2021. cited by applicant

An Office Action dated Apr. 4, 2019, which issued during the prosecution of U.S. Appl. No. 14/914,329. cited by applicant

An Office Action dated Jun. 13, 2017, which issued during the prosecution of U.S. Appl. No. 14/285,672. cited by applicant

An Office Action dated Mar. 26, 2018, which issued during the prosecution of U.S. Appl. No. 14/285,672. cited by applicant

An Office Action dated Mar. 3, 2022, which issued during the prosecution of U.S. Appl. No. 17/063,320. cited by applicant

An Office Action dated Mar. 9, 2020 which issued during the prosecution of U.S. Appl. No. 16/232,124. cited by applicant

An Office Action dated Oct. 5, 2016, which issued during the prosecution of U.S. Appl. No. 14/285,672. cited by applicant

Anand, A., et al. "Automatic identification of malaria-infected RBC with digital holographic microscopy using correlation algorithms." *Photonics Journal, IEEE* 4.5 (2012): 1456-1464. cited by applicant

Bacus, J.W., 1985. Cytometric approaches to red blood cells. *Pure and Applied Chemistry*, 57(4), pp. 593-598. cited by applicant

Ben-Suliman-2018-Computerized Counting-Based System for Acute Lymphoblastic Leukemia Detection in Microscopic Blood Images: 27th International Conference on Artificial Neural Networks, Rhodes, Greece, Oct. 4-7, 2018, Proceedings, Part II. cited by applicant

Bovik, Alan C., ed. "The essential guide to image processing", chapter 27, "Computer assisted Microscopy", pp. 777-831, Academic Press, 2009. cited by applicant

Bravo-Zanoguera, M.E., Laris, C.A., Nguyen, L.K., Oliva, M. and Price, J.H., 2007. "Dynamic autofocus for continuous-scanning time-delay-and-integration image acquisition in automated microscopy", *Journal of biomedical optics*, 12(3), pp. 034011-1-034011-16. cited by applicant

C. Briggs, et al., "Continuing developments with the automated platelet count", Blackwell Publishing Ltd, *International Journal of Laboratory Hematology*, Jan. 18, 2007, pp. 77-91, vol. 29 (15 pages total). cited by applicant

Centers for Disease Control and Prevention. "DPDx—Laboratory Identification of Parasitic Diseases of Public Health Concern", <<http://www.cdc.gov/dpdx/diagnosticProcedures/blood/microexam.html>>, Nov. 29, 2013. cited by applicant

Emma Eriksson et al: "Automated focusing of nuclei for time lapse experiments on single cells using holographic optical tweezers", *Optics Express* vol. 17, No. 7, Mar. 24, 2009 pp. 5585-5594. cited by applicant

Frean, John. "Microscopic determination of malaria parasite load: role of image analysis." *Microscopy: Science, technology, Applications, and Education* (2010) : 862-866. cited by applicant

Groen F C A et al: :A Comparison of Different Focus Functions for Use in Autofocus Algorithms, *Cytometry*, Alan Liss, New York, US, vol. 6, No. 2, Mar. 1, 1985 (Mar. 1, 1985), pp. 81-91. cited by applicant

Guy, Rebecca, Paul Liu, Peter Pennefather and Ian Crandall, "The use of fluorescence enhancement to improve the microscopic diagnosis of falciparum malaria", *Malaria Journal* 2007 6:89, <https://malariajournal.biomedcentral.com/articles/10.1186/1475-2875-6-89>, (Jul. 9, 2007). cited by applicant

Hiremath, P.S., et al., "Automated Identification and Classification of White Blood Cells (Leukocytes) in Digital Microscopic Images", *IJCA Special Issue on "Recent Trends in Image Processing and Pattern Recognition"* RTIPPR, 2010. cited by applicant

Houri-Yafin, A., et al. "An enhanced computer vision platform for clinical diagnosis of malaria." *Malar Control Elimin* 5.138.10 (2016): 4172. cited by applicant

International Search Report and Written Opinion for PCT Application No. PCT/IB2020/061728 mailed on May 7, 2021. cited by applicant

International Search Report and Written Opinion for PCT Application No. PCT/IB2020/061736 mailed on May 3, 2021. cited by applicant

International Search Report issued for PCT Application No. PCT/IB2020/059924 on Mar. 22, 2021. cited by applicant

International Search Report issued for PCT Application No. PCT/IB2020/059925 on Mar. 26, 2021. cited by applicant

International Search Report issued for PCT Application No. PCT/IB2020/061724 on Mar. 18, 2021. cited by applicant

Invitation to pay fees and Partial Search Report issued for PCT Application No. PCT/IB2020/059924 on Jan. 28, 2021. cited by applicant

Invitation to pay fees and Partial Search Report issued for PCT Application No. PCT/IB2020/059925 on Feb. 4, 2021. cited by applicant

Invitation to pay fees and Partial Search Report issued for PCT Application No. PCT/IB2020/061728 on Mar. 15, 2021. cited by applicant

Invitation to pay fees and Partial Search Report issued for PCT Application No. PCT/IB2020/061732 on Mar. 10, 2021. cited by applicant

Invitation to pay fees and Partial Search Report issued for PCT Application No. PCT/IB2020/061736 on Mar. 12, 2021. cited by applicant

Brenner, et al., "An Automated Microscope for Cytologic Research a Preliminary Evaluation", *The Journal of Histochemistry and Cytochemistry*, 1976, pp. 100-111, vol. 24, No. 1 (12 pages total). cited by applicant

Jahnmehr, S A H et al., “Simple Technique for Fluorescence Staining of Blood Cells with Acridine Orange”, *Journal of Clinical Pathology*, Feb. 12, 1987, pp. 926-929 (4 pages total). cited by applicant

Kawamoto, F. and P. F. Billingsley. “Rapid diagnosis of malaria by fluorescence microscopy.” *Parasitology today* 8.2 (1992): 69-71. cited by applicant

Kawamoto, Fumihiko, “Rapid diagnosis of malaria by fluorescence microscopy with light microscope and interference filter”. *The Lancet*, vol. 337, pp. 200-202, Jan. 26, 1991. cited by applicant

Knesel, “Roche Image Analysis Systems, Inc.”, *Acta Cytologica*, vol. 40, pp. 60-66, (1996). cited by applicant

Le, et al., “A novel semi-automatic image processing approach to determine Plasmodium falciparum parasitemia in Giemsa-stained thin blood smears”, *BMC Cell Biology*, Mar. 28, 2008 (pp. 1-12). cited by applicant

Leif, “Methods for Preparing Sorted Cells as Monolayer Specimens”, *Springer Lab Manuals*, Section 7—Chapter 5, pp. 592-619, (2000). cited by applicant

Life Technologies Corporation, “Counting blood cells with Countless Automated Cell Counter” found at <http://www.lifetechnologies.com/content/dam/LifeTech/migration/files/cell-tissue-analysis/pdfs.par.83996.file.dat/w-082149-countless-application-blood-cells.pdf>, four pages, (2009). cited by applicant

An Office Action dated Sep. 2, 2022 which issued during the prosecution of U.S. Appl. No. 17/063,320. cited by applicant

Matcher, S. J., M. Cope, and D. T. Delpy. “Use of the water absorption spectrum to quantify tissue chromophore concentration changes in near-infrared spectroscopy.” *Physics in medicine and biology* 39.1 (1994): 177. cited by applicant

Moody, “Rapid Diagnostic Tests for Malaria Parasites”, *Clinical Microbiology Reviews*, vol. 15, No. 1, pp. 66-78, (2002). cited by applicant

Moon S, Lee S, Kim H, Freitas-Junior LH, Kang M, Ayong L, et al. (2013) “An Image Analysis Algorithm for Malaria Parasite Stage Classification and Viability Quantification.”, *PLoS ONE* 8(4): e61812. <https://doi.org/10.1371/journal.pone.0061812>. cited by applicant

Non-Final Office Action dated Oct. 6, 2021, which issued during the prosecution of U.S. Appl. No. 17/063,320. cited by applicant

Notice of Allowance dated Jun. 24, 2021 which issued during the prosecution of U.S. Appl. No. 16/232,124. cited by applicant

Notice of Allowance dated Mar. 20, 2019, which issued during the prosecution of U.S. Appl. No. 15/506,997. cited by applicant

Omucheni et al. “Application of principal component analysis to multispectral-multimodal optical image analysis for malaria diagnostics”, *Malaria Journal* 2014, 13:485 <http://www.malariajournal.com/content/13/1/485>. cited by applicant

Ortyn, William E., et al. “Extended depth of field imaging for high speed cell analysis.” *Cytometry Part A* 71.4 (2007): 215-231. cited by applicant

Osibote, O. A., et al. “Automated focusing in bright-field microscopy for tuberculosis detection.” *Journal of microscopy* 240.2 (2010): 155-163. cited by applicant

Panel Decision dated Mar. 2, 2020 which issued during the prosecution of U.S. Appl. No. 14/914,329. cited by applicant

Piruska, et al., “The autofluorescence of plastic materials and chips measured under laser irradiation”, *The Royal Society of Chemistry, Lab Chip*, vol. 5, 2005 (pp. 1348-1354). cited by applicant

Price, Jeffrey H., and David A. Gough. “Comparison of phase—contrast and fluorescence digital autofocus for scanning microscopy.” *Cytometry* 16.4 (1994): 283-297. cited by applicant

Purwar, Yashasvi, et al. “Automated and unsupervised detection of malarial parasites in

microscopic images.” *Malaria journal* 10.1 (2011): 364. cited by applicant

Putzu, et al., “Leucocyte classification for leukaemia detection using image processing techniques.”, *Artificial Intelligence in Medicine*, vol. 63, No. 3, Nov. 1, 2014. cited by applicant

Ran, Qiong et al. “Spatial-spectral blood cell classification with microscopic hyperspectral imagery”, *Proc. SPIE 10461, AOPC 2017: Optical Spectroscopy and Imaging*, 1046102 (Oct. 24, 2017). cited by applicant

Rappaz, Benjamin, et al. “Comparative study of human erythrocytes by digital holographic microscopy, confocal microscopy, and impedance vol. analyzer.” *Cytometry Part A* 73.10 (2008): 895-903. cited by applicant

Roma, P. M. S., et al. “Total three-dimensional imaging of phase objects using defocusing microscopy: Application to red blood cells.” *Applied Physics Letters* 104.25 (2014): 251107. cited by applicant

An Office Action dated Nov. 10, 2022 which issued during the prosecution of U.S. Appl. No. 17/362,009. cited by applicant

Ross, Nicholas E., et al. “Automated image processing method for the diagnosis and classification of malaria on thin blood smears.” *Medical and Biological Engineering and Computing* 44.5 (2006): 427-436. cited by applicant

Saraswat, et al. “Automated microscopic image analysis for leukocytes identification: A survey”, *ABV-Indian Institute of Information Technology and Management, Gwalior, India, Micron*, 2014, vol. 65, pp. 20-33. cited by applicant

Sheikh, et al., “Blood Cell Identification Using Neural Networks”, *IEEE*, 1996 (p. 119-120). cited by applicant

Shen, Feimo, Louis Hodgson, and Klaus Hahn. “Digital autofocus methods for automated microscopy.” *Methods in enzymology* 414 (2006): 620-632. cited by applicant

Shute, G. T., and T. M. Sodeman. “Identification of malaria parasites by fluorescence microscopy and acridine orange staining.” *Bulletin of the World Health Organization*, 48.5 (1973): 591. cited by applicant

Sun, Yu, S. Duthaler and B.J. Nelson, “Autofocusing algorithm selection in computer microscopy”, 2005 *IEEE/RSJ International Conference on Intelligent Robots and Systems*. cited by applicant

Tek, F. Boray, Andrew G. Dempster, and Izzet Kale. “Computer vision for microscopy diagnosis of malaria.” *Malaria Journal* 8.1 (2009): 153. cited by applicant

Thung, Ferdian, and Iping Supriana Suwardi. “Blood parasite identification using feature based recognition.” *Electrical Engineering and Informatics (ICEEI)*, 2011 *International Conference on*. *IEEE*, 2011. cited by applicant

U.S. Appl. No. 61/870,106, filed Aug. 26, 2013. cited by applicant

U.S. Appl. No. 62/042,388, filed Aug. 27, 2014. cited by applicant

UNITAID Malaria Diagnostic Technology and Market Landscape, 2nd Edition (2014). cited by applicant

Varga, et al., “An automated scoring procedure for the micronucleus test by image analysis,”, *Mutagenesis* vol. 19 No. 5 pp. 391-397, 2004. cited by applicant

Vink, J. P., et al. “An automatic vision based malaria diagnosis system. ” *Journal of microscopy* 250.3(2013): 166-178. cited by applicant

Wissing, et al., “Illumination of the Malaria Parasite *Plasmodium falciparum* Alters Intracellular pH”, *The Journal of Biological Chemistry*, vol. 277, No. 40, Oct. 4, 2002 (pp. 37747-37755). cited by applicant

Witt, et al. “Establishing traceability of photometric absorbance values for accurate measurements of the haemoglobin concentration in blood.”, *Metrologia* 50 (2013) 539-548. cited by applicant

Wu, Qiang, Fatima Merchant, and Kenneth Castleman. “Microscope image processing”, Chapter 16, “Autofocusing”, pp. 441-467, *Academic press*, 2010. cited by applicant

Xu, Lili, Asok Chaudhuri, “*Plasmodium yoelii*: A differential fluorescent technique using Acridine



Orange to identify infected erythrocytes and reticulocytes in Duffy knockout mouse”, *Experimental Parasitology*, vol. 110, Issue 1, May 2005, pp. 80-87,  
<https://www.sciencedirect.com/science/article/pii/S001448940500038X>, (May 31, 2005). cited by applicant

Notice of Allowance issued for U.S. Appl. No. 17/362,009 on Mar. 21, 2023. cited by applicant

Yang, Ming, and Li Luo. “A rapid auto-focus method in automatic microscope.” *Signal Processing*, 2008, ICSP 2008. 9th International Conference on. IEEE, 2008. cited by applicant

Yazdanfar, S., Kenny, K.B., Tasimi, K., Corwin, A.D., Dixon, E.L. and Filkins, R.J., 2008. Simple and robust image-based autofocusing for digital microscopy. *Optics express*, 16(12), pp. 8670-8677. cited by applicant

Zahniser et al., Automated Slide Preparation System for the Clinical Laboratory, *Cytometry*, vol. 26, No. 10, pp. 30-64, (1996). cited by applicant

An International Search Report and a Written Opinion dated Sep. 17, 2021, for International Application No. PCT/IB2021/056073. cited by applicant

Cervantes, Serena , Jacques Prudhomme, David Carter, Krishna G Gopi, Qian Li, Young-Tae Chang, and Karine G Le Roch, High-content live cell imaging with RNA probes: advancements in high-throughput antimalarial drug discovery, *BMC Cell Biology* 2009, 10:45,  
<https://bmcmolcellbiol.biomedcentral.com/track/pdf/10.1186/1471-2121-10-45> (Jun. 10, 2009). cited by applicant

Chong, Shau Poh, Shilpa Pant, and Nanguang Chen. “Line-scan Focal Modulation Microscopy for Rapid Imaging of Thick Biological Specimens.” *SPIE/OSA/IEEE Asia Communications and Photonics. International Society for Optics and Photonics*, 2011. cited by applicant

Hoover Karl “SimpleAutofocus.cpp—Micro-Manager Device Adapter Code”, Dec. 31, 2009. cited by applicant

Keiser, J. et al., “Acridine Orange for malaria diagnosis: its diagnostic performance, its promotion and implementation in Tanzania, and the implications for malaria control”, *Annals of Tropical Medicine and parasitology*, 96.7 (2002): 643-654. cited by applicant

MetaXpress® 6 Software Guide, Configuring Image-Based Autofocus During Acquisition-2015. cited by applicant

MicroManager Documentation: SimpleAutofocus-2009. cited by applicant

Non-Final Office Action dated Jun. 25, 2021 which issued during the prosecution of U.S. Appl. No. 16/706,159. cited by applicant

A Notice of Allowance issued on May 14, 2021 for U.S. Appl. No. 16/232,124. cited by applicant

D5-Micro-Manager-User's-Guide-Micro-Manager-2016. cited by applicant

Gordon, Andrew et al. “Single-cell quantification of molecules” *Nature Methods* 4, Jan. 21, 2007, pp. 175-181. cited by applicant

Gordon, Andrew et al. Supplementary Note to Gordon et al.: “Single-cell quantification of molecules” *Nature Methods*, Jan. 21, 2007, pp. 1-35. cited by applicant

Notice of Allowance dated Jul. 10, 2019 in U.S. Appl. No. 15/506,997. cited by applicant

Poon et al., “Automated Image Detection and Segmentation in Blood Smears”, [*Cytometry* 1992 13:766-774]. cited by applicant

Wu, Caicai et al., “Feasibility study of the spectroscopic measurement of oxyhemoglobin using whole blood without pre-treatment”, *The Analyst*, Mar. 1998, pp. 477-481, vol. 123 (5 pages total). cited by applicant

An Office Action dated Jan. 6, 2023 which issued during the prosecution of U.S. Appl. No. 17/063,320. cited by applicant

Parasight LTD, WO2012/090198 A3, WIPO International Publication, “An Apparatus and Method for Automatic Detection of Pathogens,” published Nov. 15, 2012, including International Search Report mailed Jul. 27, 2012. cited by applicant

A Chinese Office Action dated Dec. 19, 2019 for Chinese Application No. 201580058510.7. cited

by applicant

A European Examination Report dated Dec. 11, 2019 which issued during the prosecution of Applicant's European App No. 14800352.8. cited by applicant

A European Examination Report dated Jan. 7, 2020 which issued during the prosecution of Applicant's European App No. 15778059.4. cited by applicant

An International Search Report and a Written Opinion both dated Sep. 29 (24), 2014 for International Application No. PCT/IL2012/050423. cited by applicant

An Office Action dated Jan. 19, 2021 for U.S. Appl. No. 16/706,159. cited by applicant

An Office Action dated Nov. 16, 2018 which issued during the prosecution of U.S. Appl. No. 14/914,329. cited by applicant

Fohlen-Walter, Anne PhD, et al., "Laboratory Identification of Cryoglobulinemia From Automated Blood Cell Counts, Fresh Blood Samples, and Blood Films", American Society for Clinical Pathology, Am J Clin Pathol, 2002, pp. 606-614, vol. 117 (9 pages total). cited by applicant

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## **Background/Summary**

CROSS-REFERENCE TO RELATED APPLICATIONS (1) This application is a continuation of U.S. patent application Ser. No. 17/362,009 to Yorav-Raphael (published as US 2021/0327064), filed Oct. 21, 2021, which is a continuation of U.S. patent application Ser. No. 16/592,335 to Yorav-Raphael (issued as U.S. Pat. No. 11,100,637), filed Oct. 3, 2019, which is a continuation of U.S. patent application Ser. No. 15/506,997 to Yorav-Raphael (issued as U.S. Pat. No. 10,482,595), which is a US national phase application of PCT Application No. PCT/IL/2015/050864 to Yorav-Raphael (published as WO 16/030897), filed Aug. 27, 2015, which claims the benefit of U.S. Provisional Application No. 62/042,388, filed on Aug. 27, 2014.

### **TECHNICAL FIELD**

(1) The presently disclosed subject matter relates to the field of microscopy and more specifically to calculating focus variation for a digital microscope.

### **BACKGROUND**

(2) References considered to be relevant as background to the presently disclosed subject matter are listed below. Acknowledgement of the references herein is not to be inferred as meaning that these are in any way relevant to the patentability of the presently disclosed subject matter.

(3) U.S. Pat. No. 4,700,298 (Palcic et al.) published on Oct. 13, 1987; U.S. Pat. No. 4,803,352 (Bierleutgeb) published on Feb. 7, 1989; U.S. Pat. No. 5,932,872 (Price), published on Aug. 3, 1999; and Vink et al. Journal of Microscopy, 2013 1-13, "An automatic vision-based malaria diagnosis system.

### **General Description**

(4) In accordance with certain aspects of the presently disclosed subject matter, there is provided a digital microscopy method, comprising: obtaining, in a memory comprised in or operatively coupled to a digital microscope, data representative of at least one image of at least a portion of a cell sample, the at least one image acquired by the digital microscope, wherein the portion encompasses a mapping field, and the mapping field comprises a plurality of focus analysis regions; and calculating, by processing resources operatively connected to the digital microscope, using the obtained data, a focus configuration for each of at least two of the focus analysis regions within the mapping field, giving rise to data indicative of focus variation within the mapping field.

- (5) In accordance with certain other aspects of the presently disclosed subject matter, there is provided a digital microscope comprising: a memory configured to obtain data representative of at least one image of at least a portion of a cell sample, wherein the portion encompasses a mapping field, and the mapping field comprises a plurality of focus analysis regions; and one or more processing resources configured to calculate, using the obtained data, a focus configuration for each of at least two of the focus analysis regions within the mapping field, giving rise to data indicative of focus variations within the mapping field.
- (6) In accordance with certain other aspects of the presently disclosed subject matter, there is provided a program storage device readable by a machine, tangibly embodying a program of instructions executable by the machine to perform method steps of: obtaining, in a memory comprised in or operatively coupled to a digital microscope, data representative of at least one image of at least a portion of a cell sample, the at least one image acquired by the digital microscope, wherein the portion encompasses a mapping field, and the mapping field comprises a plurality of focus analysis regions; and calculating, using the obtained data, a focus configuration for each of at least two of the focus analysis regions within the mapping field, giving rise to data indicative of focus variation within the mapping field.
- (7) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, the obtained data can be representative of a series of images, the images captured by performing a depth scan using the digital microscope, the series of images being associated with a respective series of depth levels of the cell sample.
- (8) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, the calculating can include operating a statistical function on the obtained data.
- (9) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, the calculating can include detecting at least one depth level corresponding to a drop in image contrast, wherein the detected depth level is such that image contrast at the detected depth level is lower than image contrast associated with a depth level immediately preceding the detected depth level in the series of depth levels and lower than image contrast associated with a depth level immediately following the detected depth level in the series of depth levels. The image contrast can be calculated from any of the following contrast functions: variance, standard deviation, sum of absolute-value of derivatives.
- (10) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, the mapping field and at least one diagnostic field within the sample can be of substantially or precisely same size and shape. At least one of the diagnostic field and the mapping field can be the largest field that can be captured by the microscope at a magnification selected for analysis of the sample.
- (11) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, at least one focus field can cover an area of the cell sample that is smaller than the area covered by at least one diagnostic field and the area covered by the mapping field.
- (12) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, at least one image of at least one diagnostic field within the cell sample can be analyzed using the data indicative of focus variation.
- (13) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, a confidence score can be calculated for each of the at least two focus analysis regions of the mapping field, the confidence score being indicative of an accuracy of a diagnostic analysis performed on the at least one image at a corresponding region. The at least one image of at least one diagnostic field can be acquired at a specific focus configuration, and a plurality of regions within the at least one image can be correlated with at least two of the focus analysis regions of the mapping field, and the confidence scores can be utilized for the analyzing.

- (14) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, the data indicative of focus variations can be utilized for defining at least one of a size or a shape of one or more diagnostic fields within the cell sample.
- (15) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, there is further provided: capturing at least one image of a given diagnostic field at a specific focus configuration; analyzing the at least one image to detect a pathogen candidate; correlating a plurality of regions within the at least one image with at least two of the focus analysis regions of the mapping field; and capturing at least one additional image of the pathogen candidate at an imaging focus configuration calculated using the focus configuration of at least one of the focus analysis region correlated with the position of the pathogen candidate in the diagnostic field. The capturing of at least one additional image can be performed only if the pathogen candidate is detected in a region of the diagnostic field that is correlated with a given focus analysis region of the mapping field associated with a focus configuration that differs from the specific focus configuration by at least a threshold value. The imaging focus configuration can be the focus configuration of the at least one of the focus analysis region.
- (16) In accordance with further aspects and, optionally, in combination with other aspects of the presently disclosed subject matter, there is further provided: capturing at least one image of a given diagnostic field at a specific focus configuration; correlating a plurality of regions within the at least one image with at least two of the focus analysis regions of the mapping field; and selecting a portion of the diagnostic field for analysis such that the focus variation between the specific focus configuration and the focus configuration of the focus analysis regions within the portion is below a specific value.
- (17) The present disclosure further provides a method of determining a reference depth level within a cell sample. The method comprises obtaining data representative of a series of images captured by performing a depth scan of the cell sample using a digital microscope, the series of images being associated with a series of depth levels of the cell sample; processing the data for detecting at least one depth level corresponding to a drop in image contrast; and identifying the detected depth level as the reference depth level.
- (18) In some embodiments, the detected depth level is such that image contrast at the detected depth level is lower than image contrast associated with a depth level immediately preceding the detected depth level in the series of depth levels and lower than image contrast associated with a depth level immediately following the detected depth level in the series of depth levels.
- (19) In some embodiments, the method further comprises calculating image contrast using a contrast function increasing with the image contrast and wherein detecting at least one depth level corresponding to a drop in image contrast comprises detecting a well in a contrast curve representing image contrast as a function of the depth level.
- (20) In some embodiments, the detected depth level corresponds to a bottom of the well in the contrast curve.
- (21) In some embodiments, the method further comprises calculating image contrast using a contrast function decreasing with the image contrast and wherein detecting at least one depth level corresponding to a drop in image contrast comprises detecting a roof of a contrast curve representing image contrast as a function of the depth level.
- (22) In some embodiments, the detected depth level corresponds to a top of the roof of the contrast curve.
- (23) In some embodiments, a span of the series of depth levels is of 5 to 1000 micrometers.
- (24) In some embodiments, a span of the series of depth levels is less than 50 micrometers.
- (25) In some embodiments, focusing the microscope at an investigation level further comprises shifting a focus plane of the digital microscope from the reference depth level by a specific value.
- (26) In some embodiments, the image contrast of an image is calculated from any of the following contrast functions: variance, standard deviation, sum of absolute-value of derivatives.

(27) Further, it is understood that the term “well” is used to refer to a point or region of a curve where the curve passes from decreasing (curving down) to increasing (curving up). It is understood that the term well refers to a drop of contrast. In the following, it is generally considered that contrast function is such that a drop of contrast between two images generates a drop in the contrast function values (i.e. the contrast function is an increasing function of the image contrast). However, it is noted that if the contrast function is a function decreasing with the contrast, a drop of contrast would generate an increase in the contrast function thereby turning a “well” into a “roof”. It will be appreciated that a decreasing function can be transformed into an increasing function of the contrast by multiplying the function by  $-1$ . Therefore, the present disclosure also applies to decreasing function of the contrast. It will also be appreciated that another way of applying the teaching of the present invention to a decreasing function of the contrast would be to determine a “roof” instead of a well for a decreasing contrast function of the contrast (i.e. contrast functions so that a drop of contrast generates an increase in the contrast function values), wherein the roof refers to a point or region of the curve where the curve passes from increasing (curving up) to decreasing (curving down). However, since in the art most contrast functions are increasing with the contrast, the present disclosure refers generally, without any limitation, to a well.

(28) Furthermore, it is noted that the term “bottom” of the well should be understood as the minimum point within a well and that the term “top of a roof” should be understood as a maximum point within a roof. It is also understood that the term “series” refers to an ordered set of values. In particular, the series of depth levels may be arranged in increasing or decreasing order.

(29) It is also appreciated that the present disclosure also applies to functions which are usually classified in the art as representative of sharpness and that the expression “contrast function” should be understood as referring generally to contrast and/or sharpness function i.e. functions for assessing a contrast and/or sharpness of an image.

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## Description

### BRIEF DESCRIPTION OF THE DRAWINGS

(1) In order to understand the presently disclosed subject matter and to see how it may be carried out in practice, the subject matter will now be described, by way of non-limiting examples only, with reference to the accompanying drawings, in which:

(2) FIG. 1A is a top view illustration of a sample carrier that may be used in connection with some embodiments of the presently disclosed subject matter;

(3) FIG. 1B is an illustration of some cross section lateral views of sample carriers as having a top view as shown in FIG. 1A;

(4) FIG. 2A is a top view illustration of a sample chamber comprising diagnostic fields, focus fields and mapping fields, in accordance with some embodiments of the presently disclosed subject matter;

(5) FIG. 2B is an illustration of some alternative shapes and distributions of focus analysis regions within a mapping field, in accordance with some embodiments of the presently disclosed subject matter;

(6) FIG. 3A is an illustration of a three-dimensional view of raw data indicative of focus configurations calculated for focus analysis regions of a mapping field, in accordance with some embodiments of the presently disclosed subject matter;

(7) FIG. 3B is an illustration of three-dimensional view of a flat plane matched to the raw data shown in FIG. 3A, in accordance with some embodiments of the presently disclosed subject matter;

(8) FIG. 4 is a block diagram schematically illustrating one example of a system for calculating focus configurations for a digital microscope, in accordance with some embodiments of the presently disclosed subject matter;

- (9) FIG. 5 is a flowchart illustrating one example of a sequence of operations carried out for imaging and analyzing a biological cell sample, in accordance with some embodiments of the presently disclosed subject matter;
- (10) FIG. 6 is an illustration of a mapping field including a quality confidence score, in accordance with some embodiments of the presently disclosed subject matter;
- (11) FIG. 7 is a flowchart illustrating steps of a method of imaging a cell sample, in accordance with some embodiments of the presently disclosed subject matter;
- (12) FIG. 8 illustrates an image variance calculation, in accordance with some embodiments of the presently disclosed subject matter;
- (13) FIG. 9 illustrates an example for a focus curve obtained by depth scanning of a cell sample and computing of a focus function according to some embodiments of the present disclosure;
- (14) FIG. 10A is an illustration of an image having a slant on the X axis, and an example of a division thereof to diagnostic fields;
- (15) FIG. 10B is an illustration of an image having a slant on both the X and Y axis, and an example of a division thereof to diagnostic fields;
- (16) FIG. 11 illustrates a non-limiting example of a mapping field having a plurality of focus analysis regions;
- (17) FIG. 12 illustrates a non-limiting example of a correlation function; and
- (18) FIG. 13 illustrates a non-limiting example of variation in the appearance of objects in two or more focus analysis regions of a mapping field.

#### DETAILED DESCRIPTION

(19) In the following detailed description, numerous specific details are set forth in order to provide a thorough understanding of the invention. However, it will be understood by those skilled in the art that the present invention may be practiced without these specific details. In other instances, well-known methods, procedures, and components have not been described in detail so as not to obscure the present invention.

(20) In the drawings and descriptions set forth, identical reference numerals indicate those components that are common to different embodiments or configurations.

(21) In many cases, a biological cell sample being analyzed through a microscope is larger than the optical field of the microscope at the desired magnification. Therefore, in some cases, a plurality of images are acquired, each having a different position along an XY plane parallel to the focal plane of the microscope. Each of these images presents a so called “diagnostic field” within the biological sample. The images or parts thereof can be analyzed in order to detect the presence of a pathogen suspected to be included in the cell sample and/or for characterizing the composition of a cell sample (for example in a complete blood cell count).

(22) In order for a captured image to be clear, the microscope needs to be focused on the cell sample. The focusing can be performed by overlaying a focal plane of the microscope with an investigation plane of the sample being imaged. For example, one may focus the microscope by changing a distance between the sample carrier holding the biological cell sample and the microscope's lens and/or by adjusting properties of the lens or any other way known in the art.

(23) As used herein, a focus configuration means a configuration of the microscope characterized by the relative positions of the microscope focal plane and an investigation plane in the cell sample. Optionally, the focus configuration can be defined by the distance between these relative positions. Optionally, the focus configuration for any given focus analysis region can be a value related to a distance between the investigative plane of the given focus analysis region and the investigative plane of another focus analysis region, such as a prior analyzed focus analysis region. A focus configuration of a microscope may be manipulated for example by changing one or more of (a) a distance between the microscope objective lens and the cell sample; and (b) one or more properties of the microscope lens. It is to be noted that the focus configuration can be manipulated in other ways, including ways that are known in the art, to control the position of a microscope's focal plane

relative to a cell sample investigation plane.

(24) A focus configuration may be selected by analyzing data representative of a series of overlapping images of a cell sample or a portion thereof, captured by performing a depth scan using a digital microscope, the series of images being associated with a series of depth levels of the cell sample. Analyzing may include by applying a focus function to the data to select a focus configuration, as further detailed herein. The selected focus configuration may be indicative of an imaging focus configuration, namely a focus configuration at which to capture at least one image of the sample for analysis.

(25) Selecting an imaging focus configuration may be considered as focusing. Selecting an imaging focus configuration may include selecting a reference focus configuration and calculating an imaging focus configuration therefrom (e.g. by adding or subtracting a fixed value or a percentage), as further detailed herein.

(26) Due to various reasons such as tolerance in the microscope setup and fabrication and/or curvature of some sample carriers, the investigation plane in the sample carrier is not necessarily perfectly parallel to the microscope's focal plane. In addition, the focal plane is not necessarily perfectly flat at a given magnification. Accordingly, the focus configuration applicable for one location (e.g. a Cartesian (X,Y) position) in a sample may differ from the focus configuration applicable for another location in the same sample.

(27) FIG. 1A shows a top view illustration of a sample carrier, that may be used in connection with some embodiments of the presently disclosed subject matter, and FIG. 1B shows an illustration of some cross section lateral views of sample carriers having a top view as shown in FIG. 1A. The lateral cross sections may be for example taken along a line A-A as shown in FIG. 1A.

(28) Sample carrier **100** can comprise one or more chambers, each configured to accommodate a cell sample. In the illustrated example ten chambers are depicted, namely chambers **101-110**, however this is merely a non-limiting example and any other number and configuration of chambers is also contemplated.

(29) As used herein, a "sample carrier" can be any carrier known in the art for holding a biological cell sample. Examples include carrier slides, laboratory receptacles, dishes, plates, multi-well plates, test tubes (e.g. with a flat bottom), microfluidic cells and cartridges and the like, provided that it has at least one transparent side allowing imaging.

(30) It can be appreciated that in the illustrated examples, in the cross section view of sample carrier **200** the chambers **201-205** are all flat, which enable positioning thereof parallel to a surface on which the sample carrier **200** is to be placed and/or parallel to the microscope's focal plane. However, looking at sample carriers **220** and **240**, it can be appreciated that they are bent. Accordingly, the chambers in the bent sample carriers **220** and **240** (chambers **221-225** and **241-245** respectively) have different angles (i.e. they are not parallel) with respect to a surface holding the sample holder **220** or **240** and/or with respect to the microscope's focal plane. While chamber **243** may be essentially parallel to such surface and/or to such focal plane, chamber **244** would have an angle with respect to the surface and/or to the focal plane and chamber **245** would have an even sharper angle with respect to the surface and/or to the focal plane. It is to be noted that the bends in the illustration are drawn very prominently for illustrative purposes. In reality, the angles of all or part of the chambers with respect to the surface holding the corresponding sample carrier and/or with respect to the microscope's focal plane, can sometimes be difficult or even impossible to detect by the naked eye but might still have a significant effect on focus under microscope magnification. It is to be further noted that although the bends shown in the illustration are on the longitudinal axis, this is of course not limiting, and the bends can additionally or alternatively be on the latitudinal axis. Moreover, the internal bottom surface of the carrier may have a different contour than that of the external bottom surface, such that even if the carrier is positioned in parallel to the surface holding the corresponding sample carrier and/or with respect to the microscope's focal plane, the cell sample within it may lie on a relatively bent surface, thereby

having a bent investigation plane.

(31) In some cases, irrespective of any bending of the sample carrier, tolerance in the microscope's setup and/or fabrication can also result in (or contribute to) the investigation plane in the sample carrier not being parallel to the microscope's focal plane. In addition, the microscope's focal plane itself might not be flat at a given magnification. Any one or any combination of two or more of the scenarios illustrated above, and/or any other reason that results in the investigation plane in the sample carrier not being parallel to the microscope's focal plane, optionally results in a difference between a focus configuration applicable for one location (e.g. a Cartesian (X,Y) position) in a cell sample and a focus configuration applicable for another location in the same cell sample or the same chamber or the same portion of a chamber or for a plurality of chambers, as further detailed herein.

(32) Attention is now drawn to FIG. 2A, showing a top view illustration of a sample chamber comprising diagnostic fields, focus fields and mapping fields, in accordance with some embodiments of the presently disclosed subject matter. Optionally, a given image covers precisely a given diagnostic field, focus field or mapping field. In some embodiments, a given diagnostic field, focus fields or mapping field is defined as one or more portions of a specific image. Optionally, several images or portions thereof are assembled to define a diagnostic field, focus field or mapping field.

(33) Sample chamber **300** comprises a plurality of diagnostic fields (e.g. diagnostic fields **301**, **302**, etc.). A diagnostic field is an area covering a portion of the sample chamber **300** that comprises a cell sample to be imaged, and is imaged as a single image under at least one lighting configuration of the microscope, to be analyzed for various purposes, including diagnostics, detection of pathogens, blood count, and the like. Optionally, a plurality of images of a single diagnostic field are taken for analysis, each image encompassing the entire diagnostic field. It is to be noted that although the sample chamber **300** shown in the illustration comprises a plurality of diagnostic fields (namely thirty six (36) diagnostic fields), in some cases the sample chamber can comprise a single diagnostic field, or any other number of diagnostic fields, in any configuration. Likewise, while the sample chamber, the diagnostic fields, the focus fields and the mapping fields in FIG. 2A are depicted as rectangular shapes, any other shapes may be used, covering the entire surface of the chamber or only one or more portions thereof.

(34) For each diagnostic field, a corresponding focus field is shown in the example, as a gray rectangle (e.g. focus fields **311**, **312** in diagnostic fields **301**, **302**, respectively). A focus field is an area covering a portion of the sample chamber **300** used for calculating a focus configuration to be used for acquiring images of at least one diagnostic field, as further detailed herein. It is to be noted that although the focus fields (e.g. focus fields **311**, **312**) are shown as a portion of the diagnostic fields, in some cases the focus field can be identical to the diagnostic field. In addition, although the focus fields (e.g. focus fields **311**, **312**) are shown as fully overlapping the diagnostic field, in some cases the focus field can overlap the diagnostic field only partially, and in other cases it is possible that no such overlap will exist. Still further, although in the illustrated example each diagnostic field has a corresponding focus field therein (focus field **311** for diagnostic field **301**, focus field **312** for diagnostic field **302**), in some cases fewer or more focus fields can exist. Moreover, a single focus field can overlap, or partially overlap, more than one diagnostic field. In addition, as indicated herein, a given focus field can be used to calculate a focus configuration to be used for acquiring images of more than one diagnostic field, including for one or more diagnostic fields that do not overlap the given focus field at all.

(35) Also shown in the illustration are examples for mapping fields **350** and **355**. A mapping field is an area covering a portion of the sample chamber **300** used for obtaining data indicative of focus variation within at least a portion of the sample holder (e.g. within the portion covered by the mapping field), as further detailed herein. Such data can further be used for extrapolating additional data indicative of focus variations in portions external to the mapping field, as further detailed



herein.

(36) In the illustration, mapping field **355** is shown as identical or essentially identical to the diagnostic field shown immediately below diagnostic field **302**, whereas mapping field **350** is shown to partially overlap four (4) diagnostic fields. It is to be noted however that the mapping fields are not necessarily of the same shape and/or size of the diagnostic field and that they do not necessarily overlap any given diagnostic field. However, in some cases they can be of the same shape and/or size as of the diagnostic field, or substantially of the same shape and/or size of the diagnostic field. Optionally, at least one diagnostic field is used also as a mapping field. It is to be noted that in some cases, at least one diagnostic field and at least one mapping field are the largest fields (i.e. imaged areas) acquired by the microscope at a magnification selected for analysis of the cell sample.

(37) Each mapping field comprises a plurality of focus analysis regions. For illustration purposes, looking at the right hand side of the FIG. 2A, there is shown an enlarged view of mapping field **350**, comprising a plurality of focus analysis regions (e.g. focus analysis region **351**, **352**, **353**, etc.). A focus analysis region is an area covering a portion of the mapping field for which a corresponding focus configuration is calculated, as further detailed herein.

(38) It is to be noted that in some cases at least one focus field covers an area of the cell sample that is smaller than the area covered by at least one diagnostic field and at least one mapping field.

(39) FIG. 2B shows an illustration of some alternative shapes and distributions of focus analysis regions within a mapping field, in accordance with some embodiments of the presently disclosed subject matter. In the illustration, mapping fields **360**, **370** and **380** are depicted as rectangles. Mapping field **360** is split to a three by three (3×3) grid or matrix of rectangle shaped focus analysis regions a-i. However, the focus analysis regions may have any shape, and any number of focus analysis regions may be included in any mapping field. In some cases a mapping field can comprise a single focus analysis region, and in some cases (not limiting) such single focus analysis region can overlap the mapping field. While the focus analysis regions need not cover the entire mapping field (as shown, for example, in mapping fields **370** and **380**) they may do so (as shown for example in mapping field **360**). Optionally, some or all of the focus analysis regions may partially overlap one or more other focus analysis regions. It is to be noted that in some cases there is a direct relation between the number and/or size of focus analysis regions and the accuracy of the data indicative of focus variations. However, there may also be a direct relation between the number of focus analysis regions and the background noise, and the time needed for analysis of focus variation of a mapping field may increase in proportion to the number of focus analysis regions. In some non-limiting cases, the number of focus analysis ranges within a given mapping field may range between 2 to 1000 focus analysis regions. In more specific cases, the number of focus analysis ranges within a given mapping field may range between 9 to 120 focus analysis regions.

(40) FIG. 3A is an illustration of a three-dimensional view of raw data indicative of focus configurations calculated for focus analysis regions of a mapping field, in accordance with some embodiments of the presently disclosed subject matter.

(41) In this example, data representative of a series of images of a mapping field spanning 500  $\mu\text{m}$  by 700  $\mu\text{m}$  of the surface of a sample carrier was analyzed. The data represented images that were captured by performing a depth scan (as detailed inter alia with respect to FIG. 5) of the mapping field using a digital microscope. In the illustration, the mapping field was divided into 35 identical rectangular focus analysis regions.

(42) In the illustration, the XY plane is a plane parallel to the focal plane of the microscope. In an ideal case, an unbent diagnostic field should have a focus variation map that is perfectly parallel to the XY plane. In the example, the focus configuration is depicted as a function of the distance between the microscope objective and the investigation plane in the sample. The focus configuration at the field having the most extreme focus configuration was set at 0 and all other

fields are shown relative thereto. In this example, focus configurations calculated for the focus analysis regions in the shown mapping field varied along the z axis by up to 2.5  $\mu\text{m}$ .

(43) It is to be noted that the three dimensional view is provided for illustrative purposes only and it is based on the actual raw data that is the focus configuration calculated for each corresponding focus analysis region. It is to be noted that the focus configuration can be actually represented by a single calculated number, e.g. a number indicative of a relative position of the microscope focal plane and an investigation plane in the cell sample.

(44) When looking at a single mapping field within a sample chamber, it is expected that it will be essentially flat and linear, but possibly at an angle with respect to the focal plane of the microscope. It can be appreciated that in the illustrated example, there is a clearly a deviation from this expectation. Such deviation can be explained mainly by optical distortion (a property of the lens) and some background noise. Therefore, in some cases, a flat plane can be matched to the raw data. This may be performed for example using least squares regression, which finds the equation of a plane  $z=ax+by+c$  that minimizes the sum of the square of vertical distance between the plane and the raw data. FIG. 3B shows an illustration of a three-dimensional view of a flat plane matched to the raw data shown in FIG. 3A, in accordance with some embodiments of the presently disclosed subject matter.

(45) By matching a plane to the raw data, the noise may be reduced, but some information regarding optic distortion can be lost. In some cases, e.g. where focus finding is performed using brightfield images whereas florescent images are used for diagnosis (alone or together with brightfield images), the optic distortion of brightfield may be ignored.

(46) FIG. 4 is a block diagram schematically illustrating one example of a system for calculating focus configurations for a digital microscope, in accordance with some embodiments of the presently disclosed subject matter.

(47) According to some examples of the presently disclosed subject matter, all or parts of the system **400** can be comprised within a digital microscope, or be otherwise operatively connected to a digital microscope.

(48) System **400** can comprise one or more processing resources **410**. The one or more processing resources **410** can be a processing unit, a microprocessor, a microcontroller or any other computing device or module, including multiple and/or parallel and/or distributed processing units, which are adapted to independently or cooperatively process data for controlling relevant system **400** resources and for enabling operations related to system **400** resources.

(49) According to some examples of the presently disclosed subject matter, system **400** can comprise (or be otherwise associated with) a data repository **420**, configured to store data, including inter alia, data relating to focus configurations, images acquired by the microscope, etc. Data repository **420** can be any volatile and/or non-volatile memory, including, for example, magnetic media such as a hard disk drive (HDD), Solid-State Drives (SSD), FLASH memory or FLASH drives, Electrically Erasable Programmable Read-Only Memory (EEPROM), battery backed DRAM or SRAM, Random-access memory (RAM), etc.

(50) According to some examples of the presently disclosed subject matter, the processing resources **410** can include (or be otherwise associated with) one or more of the following modules: image acquisition module **430**, focus calculation module **440** and analysis module **450**.

(51) In some cases image acquisition module **430** can be configured to control the digital microscope for acquiring images of the cell sample and/or parts thereof, as further detailed herein.

(52) In some cases focus calculation module **440** can be configured to calculate focus configurations to be used for acquiring images of the cell sample or parts thereof by the digital microscope, as further detailed herein.

(53) In some cases analysis module **450** can be configured to analyze the images acquired by the digital microscope, as further detailed herein.

(54) Attention is drawn to FIG. 5, showing a flowchart illustrating one example of a sequence of

operations carried out for imaging and analyzing a biological cell sample, in accordance with some embodiments of the presently disclosed subject matter.

(55) According to some examples of the presently disclosed subject matter, system **400** can be configured to obtain data representative of a series of images (also referred to as a set of images) of a focus field within a sample chamber of a sample holder placed on a digital microscope stage, the images captured by focusing the digital microscope at a corresponding series of depths levels along the optical axis (e.g. the Z axis) of the microscope (block **510**). As indicated herein, a focus field is an area covering a portion of the sample chamber **300** used for calculating a focus configuration to be used for acquiring images of at least one diagnostic field. The process of capturing the images by focusing a digital microscope at a corresponding series of depths levels along the optical axis of the microscope is also referred to as in-depth scanning.

(56) The in-depth scanning can be performed, for example, by varying a distance between a focus plane of the microscope and a sample holder intended to accommodate a cell sample by methods well known in the art. In some cases, obtaining the set of images can be carried out using an image sensor unit of the microscope connected with the processing resources **410** of the system **400**. In some cases, in-depth scanning may be performed with brightfield illumination.

(57) It is to be noted that the term “obtain data representative of a series of images” encompasses one or both of actual capturing of images by imaging of a portion of a cell sample to acquire the set of images (in-depth scanning) and respective data (e.g. utilizing the image acquisition module **430**), and also loading/downloading from a computer storage media (e.g. data repository **420**) a set of images of the field preliminarily acquired by a digital microscope and/or the data relating to the set. In some cases the portion of the cell sample which is imaged is a field within the sample (e.g. a focus field, mapping field, diagnostic field, etc.). Optionally, the imaged field can be captured in whole or in parts. Optionally, one or more images covering one focus analysis region within a mapping field may be captured separately from one or more images covering another focus analysis region within the same mapping field, for example by performing a plurality of depth scans, each for a different portion of the sample.

(58) Further, the term depth level may be understood as a coordinate value along the optical axis of the microscope corresponding to a position that is optionally inside the cell sample or the sample holder/chamber. The actual direction of the axis and the origin of the axis to quantify depth may be arbitrarily chosen. The images may be obtained in any order, and may have an equal distance along the axis between pairs of consequent images or at a varying distance i.e. the in-depth scanning may be performed with a fixed step or with a variable step. For example, the origin of the axis may be positioned at an outer surface of the cell sample facing an objective of the microscope and the direction of the coordinate axis may be chosen so that the coordinates increase when progressing toward the cell sample. It is also understood that since the series of depth levels may be understood as an ordered set along the axis, it is possible to define endpoint depth levels (hereinafter referred to as endpoints) of the series of depth levels. As used herein, the term scanning depth interval refers to a series of depth levels between two endpoints' depth levels. One endpoint level of a series of depth levels, for example a minimum depth level of the set, may be referred to as a first scanning depth level and the other endpoint of the set, for example the maximum depth level, may be referred to as a second scanning depth level. In such case, the scanning depth interval refers to the depth levels comprised between the first and second scanning depth levels.

(59) In some cases, an estimated reference depth level may be preliminarily provided. For example, the cell sample may comprise a plurality of diagnostic fields to be investigated and reference depth levels determined for one or more previous diagnostic fields may be used to estimate the estimated reference level for a subsequent diagnostic field. In these cases, the scanning depth interval may be selected so as to cover the estimated depth reference level i.e. distances between the estimated depth reference level and the first and second scanning depth levels may be above a specific threshold. In some cases, a span of the depth scanning interval may be of around 5 micrometers to

1000 micrometers. In some cases, the span of the depth scanning interval may be between 150 and 250 micrometers, or less than 50 micrometers or even between 10 and 30 micrometers. Optionally, the estimated depth level is approximately in the midpoint of the span of the depth scanning interval.

(60) In some cases, the series of images and associated depth levels (or the data representative thereof) are utilized for calculating a focus configuration to be used for imaging one or more diagnostic fields (block 520). The focus configuration can be calculated by operating any desired focus function on the series of images and their respective associated depth levels, e.g. utilizing the focus calculation module 440. Several non-limiting examples of such focus functions are disclosed in: Groen, Frans CA, Ian T. Young, and Guido Ligthart. "A comparison of different focus functions for use in autofocus algorithms." *Cytometry* 6.2 (1985): 81-91. An additional example is provided herein, with reference to FIGS. 7-9 and to US Provisional Application No. U.S. 61/826,718, filed May 23, 2013, which is incorporated herein by reference. In some cases, the focus function can be a statistical function.

(61) It is to be noted that in some cases, the smaller the focus field is, the faster a focus configuration can be calculated thereto. Therefore, in some cases the focus field can be smaller than the diagnostic fields. On the other hand, the smaller the focus field is, the accuracy of the calculated focus configuration may be reduced (which can have a negative effect on the accuracy of the diagnostic field analysis). It is to be noted that in some non-limiting cases the size of the focus field can range between 1 to 50 percent of the size of the diagnostic field. In more specific cases, the size of the focus field can range between 1 to 25 percent of the size of the diagnostic field. In even more specific cases, the size of the focus field can range between 1 to 10 percent of the size of the diagnostic field. In even more specific cases, the size of the focus field can range between 1 to 5 percent of the size of the diagnostic field. In even more specific cases, the size of the focus field can range between 1 to 3 percent of the size of the diagnostic field.

(62) In some cases, the focus field can at least partially overlap one or more diagnostic fields that are to be acquired using the configuration calculated using the focus field. It is to be noted that using a focus configuration that is calculated using a focus field that at least partially overlaps a given diagnostic field in order to image the given diagnostic field, can yield better analysis accuracy compared to using a focus configuration that is calculated using a focus field that does not overlap the given diagnostic field.

(63) In some cases, a focus configuration is calculated to one or more diagnostic fields using a focus field that is comprised within a corresponding diagnostic field. It is to be noted that in some cases, the focus field can be identical to one of the diagnostic fields (i.e. have the same size and shape and cover the same area of the cell sample).

(64) According to some examples of the presently disclosed subject matter, system 400 can be configured to capture at least one image, of one or more diagnostic fields, at the focus configuration calculated for the corresponding focus fields or at an imaging focus configuration that is calculated therefrom (e.g. by adding or subtracting a fixed value or a percentage from the focus configuration calculated for the corresponding focus fields) (block 530), e.g. utilizing the image acquisition module 430. Optionally, system 400 can be further configured to analyze the images of the diagnostic fields (block 540), e.g. utilizing the analysis module 450.

(65) At any time before image analysis is performed, data indicative of focus variations within one or more mapping fields can be obtained. For this purpose, system 400 (e.g. utilizing the image acquisition module 430) can be configured to obtain data representative of a series of images (also referred to as a set of images) of at least a portion of a cell sample, the portion encompassing one or more mapping fields within a sample chamber of a sample holder placed on a digital microscope stage (block 550). The images are captured by focusing the digital microscope at a corresponding series of depths levels along the optical axis (e.g. the Z axis) of the microscope. It is to be noted that the process of obtaining the data representative of a series of images is similar to the process

described above with respect to block **510**, with the exception of the imaged portion described with respect to block **510** being a focus field whereas the imaged portion described with respect to block **550** being a mapping field. Optionally, the mapping field is not imaged as a whole but rather different portions thereof are imaged separately, each at a plurality of different depth levels (for example one or more focus analysis regions may be imaged separately from one or more other focus analysis regions). This may provide the depth scan spanning a mapping field, even if one scan comprises images taken at different depth levels than another scan. For example, a plurality of depth scans may be performed for different sub-portions of the sample within a single mapping field, for example, each depth scan may relate to one or more focus analysis regions. As indicated herein, a mapping field is an area covering a portion of the sample chamber **300** used for obtaining data indicative of focus variations within at least a portion of the sample chamber (e.g. within the portion covered by the mapping field). For this purpose, each mapping field comprises a plurality of focus analysis regions. As indicated herein, a focus analysis region is an area covering a portion of the mapping field for which a corresponding focus configuration is calculated.

(66) In some cases, the series of images and associated depth levels (or the data representative thereof) are utilized for calculating a focus configuration for at least two focus analysis regions within the mapping fields, giving rise to data indicative of focus variations within the mapping fields (block **560**). It is to be noted that any distribution of the focus analysis regions within the mapping fields as disclosed with respect to FIG. **2B** is contemplated. The focus configuration can be calculated by operating any desired focus function on the series of images and their respective associated depth levels, e.g. utilizing the focus calculation module **440**. The non-limiting examples provided with respect to block **520** are applicable also for this purpose.

(67) In some cases, a single image of a mapping field, or a portion thereof encompassing at least two focus analysis regions, can be used to calculate the focus configuration. For example, the system can obtain data representative of an image of a portion of the cell sample comprising a mapping field having a plurality of focus analysis regions, and can compare the appearance of objects depicted in at least two focus analysis regions in the mapping field. Certain variations in the appearance of objects of a given type in different focus analysis regions can sometimes be attributed to differences in the respective objects' positions along the z-axis relative to the focal plane of the microscope. For example, cells (e.g. red blood cells) depicted in the image which lie at different heights relative to the focal plane of microscope can exhibit variances in appearance such as width, sharpness of contour lines, color, intensity, etc. These variations in appearance can be analyzed and used to calculate focus configurations for the different focus analysis regions of the mapping field, thereby giving rise to data indicative of focus variation within the mapping field, as will be detailed below with reference to FIG. **11**. As used herein, "obtain data representative of an image" encompasses one or both of actual capturing of the image by imaging of a portion of a cell sample to acquire the image and respective data (e.g. utilizing the image acquisition module **430**), and also loading/downloading the image from a computer storage media (e.g. data repository **420**), such as an image preliminarily acquired by a digital microscope, and/or the data relating thereto.

(68) By way of non-limiting example, the system can use variations in intensity or in intensity related data between images spanning at different focus analysis regions to calculate, estimate or otherwise obtain information relating to a focus configuration by:

(69) 1. Obtaining a correlation function which provides known intensity related data, e.g. intensity statistics (e.g. variance) at each of several focus configurations. A correlation function can be obtained in any manner known in the art (e.g. in the form of a graph similar to FIG. **9** or a mathematical equation representing the same correlation). Optionally, the correlation function can be obtained in advance and stored in a memory, e.g. data repository **420**. Optionally, the correlation function can be generated on the fly by capturing or otherwise obtaining a series of depth scanned images of at least a portion of the mapping field, and calculating an intensity statistic for each image in the series of depth scanned images, essentially as detailed below with reference to FIG. **8**.

Since certain intensity statistics (e.g. variance) may be affected by the number of cells appearing therein, the obtained correlation function is optionally normalized or otherwise corrected to a number or concentration of cells (for example the number of red blood cells). Optionally, the intensity statistic for each focus analysis region may be adjusted according to number or concentration of cells in that region and according to the cell count or concentration of cells in images based upon which the correlation function was produced. Additionally, certain intensity statistics (e.g. variance) may be affected by the surface area of the sample carrier captured in an image being analyzed. Thus, the obtained correlation function is optionally normalized or otherwise corrected to the surface area. Optionally, the intensity statistic for each focus analysis region may be adjusted according to the surface area spanned by the region and according to the surface area of the sample carrier covered in images based upon which the correlation function was produced.

(70) 2. Calculating, for each of the two or more focus analysis regions, intensity related data with respect to a reference intensity related data, e.g. as detailed below with reference to FIG. 8.

Optionally, this calculation provides a relative value between the different focus analysis regions, without providing an absolute value for some or any of the focus analysis regions; and

(71) 3. Estimating, for each of the two or more focus analysis regions, a focus configuration based on the correlation function and the calculated intensity related data of each respective focus analysis region, as detailed below with reference to FIG. 8. These values may be absolute (e.g. with respect to a given position and/or configuration of the imaging module and/or the sample carrier) and/or may be relative values relating to a degree of variation between the different focus analysis regions.

(72) FIG. 11 illustrates a non-limiting example of a mapping field **1100** having focus analysis regions **1101-1109**. FIG. 12 illustrates a non-limiting example of a correlation function, in which each of focus analysis regions **1101-1109** is mapped to a scanning depth level  $Z_n$  indicative of a focus configuration.

(73) In some embodiments (such as in the example of FIG. 12 where the correlation function provides a “well” correlation function), if the correlation function has a plurality of focus configurations that correspond to one calculated value, one or more of the following may apply: (a) The direction of the variation may be immaterial, and for example two symmetric results may thus be treated the same way. This is especially true when the variation value is high enough to reject the sample carrier, but also when the degree of variation (angle of the carrier internal bottom surface) is of more interest than a direction thereof; or (b) data from the image may be used in order to define a direction of the variation (e.g.  $Z_{sub.1}$  v.  $Z_{sub.5}$ ) and/or select between a plurality of values having the same direction (not shown in the example). By way of non-limiting example, since the slide is continuous, a degree of variation may be estimated in correlation to a distance from a defined point. In such cases, the shape of the curve may be followed (and/or extrapolated and/or interpolated) through a plurality of fields.

(74) By way of a further non-limiting example, FIG. 13 shows images **1102a**, **1105a** and **1108a** of red blood cells, the images captured within focus analysis regions **1102**, **1105** and **1108**, respectively. In image **1105a**, the cells appear to be almost transparent, indicative of the cells being located at a depth of or near  $Z_{sub.3}$ . In images **1102a** and **1108a**, the cells appear to be bordered by ring shapes. Furthermore, the cells in image **1102a** appear to have a dark circumferential border with a white outer ring, while cells in image **1108a** appear to have a white circumferential border with a dark outer ring. The appearance and colors of the ring shapes are indicative of a direction of the variation as well as the degree of variation, and can be used to define the focus configuration more precisely.

(75) It is to be noted that the data indicative of focus variations within at least a portion of the sample chamber pertains to a mapping of the investigation plane of the cell sample with respect to the focal plane of the microscope. This data may be depicted and/or represented in any form that

enables determination of differences between the focus configuration applicable to one focus analysis region with respect to the other focus analysis regions.

(76) As indicated herein, when looking at a single mapping field within a sample chamber, it is expected that it will be essentially flat and linear, but possibly at an angle with respect to the focal plane of the microscope. However, due to various reasons, including optical distortion and background noise, in some cases, this expectation is not fulfilled. Therefore, in some cases, as indicated with respect to FIGS. 3A and 3B, a flat plane can be matched to the raw data indicative of focus variations. This may be performed for example using least squares regression, which finds the equation of a plane  $z=ax+by+c$  that minimizes the sum of the square of vertical distance between the plane and the raw data indicative of focus variations.

(77) It is to be noted that any distribution of the mapping fields as disclosed with respect to FIG. 2A is contemplated. It is to be further noted, when assuming the size of the focus analysis regions is constant, that the larger the area covered by the mapping fields is, the accuracy of the calculated focus configurations may be enhanced (which can have a positive effect inter alia on the accuracy of the diagnostic field analysis and/or on the determination of the diagnostic fields distribution within the cell sample). It can also be appreciated that the larger the area covered by the mapping fields is, the more time is required for calculating the focus configuration for the focus analysis regions. It is to be noted that in some non-limiting cases the mapping fields cover an area of up to fifty (50) percent of the diagnostic fields. Furthermore, in some non-limiting cases the mapping fields cover an area of up to ten (10) percent of the diagnostic fields. Still further, in some non-limiting cases the mapping fields cover an area of up to one (1) percent of the diagnostic fields.

(78) In some cases, the data indicative of focus variations within a given mapping field can be utilized in order to extrapolate information of focus variations in areas not covered by the mapping field, using various known methods and/or techniques.

(79) Once the data indicative of focus variations within a given mapping field is calculated, it can be stored, e.g. in data repository 420. The data can be represented by absolute values (e.g. values indicative of a relative position of the microscope and the microscope's investigation plane) or relative values (e.g. a difference between the focus configuration at a given location and a given reference focus configuration). In some cases the data indicative of focus variations within a given mapping field can be represented by a mathematical formulae or any type of map that can enable determination of a focus configuration at a given location within the sample carrier.

(80) It is to be further noted that the data indicative of focus variations can be used to interpolate and/or extrapolate additional data indicative of focus variations, also in areas not covered by any mapping field.

(81) In some cases, the system 400 can be configured to provide an error notification upon detection of any calculated focus configuration exceeding a given threshold. Exceeding such given threshold can indicate, for example, a problem associated with either one (or a combination of more than one) of the following: the microscope, the sample carrier, the positioning of the sample carrier within the microscope stage, or the cell sample.

(82) As shown in FIG. 5, once the data indicative of focus variations within one or more mapping fields is calculated, it may be used, as further detailed herein, in one or more of: (a) controlling the capturing of the at least one image of one or more diagnostic fields detailed with respect to block 530; and (b) analyzing the images of the one or more diagnostic fields.

(83) In some cases, the system 400 can be configured to capture a plurality of images of each diagnostic field for analysis/diagnosis thereof. In some cases, the images can be captured using the focus configuration calculated using one or more mapping fields, as detailed herein.

(84) In some cases, the number of images captured for each diagnostic field (e.g. as detailed with respect to block 530) can be determined while taking into account the degree of focus variation within the diagnostic field. The degree of focus variation can be determined based on the focus configuration calculated for the focus analysis regions of one or more mapping fields (e.g. one or

more mapping fields that cover the area that is closest to, or overlap, the area covered by the diagnostic field). For example, if the degree of focus variation (e.g. the difference between a focus configuration of a first focus analysis region and the focus configuration of a second focus analysis region) does not exceed a given threshold, only a single image of the diagnostic field can be acquired, while in other cases, two or more images of the diagnostic field may be acquired using different focus configurations. It is to be noted that while this may increase the time needed for acquiring the images and analyzing them, it may improve the accuracy of the diagnosis and may thus be preferred for some applications.

(85) According to some examples of the presently disclosed subject matter, the system **400** can be configured to calculate a focus confidence score for at least two focus analysis regions (and in some cases for each focus analysis region) and/or a function that can provide such focus confidence score (e.g. given an indication of a certain focus analysis region). The focus confidence score can be indicative of a level of confidence in the focus configuration calculated for the corresponding focus analysis region. The focus confidence score can be represented by any scale (e.g. **0-1**, **0-100**, **1-10**, or any other scale).

(86) In some cases, the focus confidence score can be used as further detailed herein, in one or more of: (a) controlling the capturing the at least one image of one or more diagnostic fields detailed with respect to block **510**; and (b) analyzing the images of the one or more diagnostic fields, as further detailed herein.

(87) In some cases, the images can be analyzed using various computer vision and classification algorithms. In some cases, such algorithms can calculate a classification confidence score for the classifications calculated therewith. In some cases, such algorithms can calculate a classification confidence score, indicative of a level of confidence in the correctness of the calculated classification. In some cases, such a classification confidence score can be multiplied by the focus confidence score of the focus analysis region/s that is correlated with the corresponding areas of the diagnostic field (in some cases for that purpose the focus analysis regions are correlated with the corresponding areas of the diagnostic field using any method and/or technique). It is to be further noted that such multiplication can enable compensating for potential classification bias that may result from poor focus quality.

(88) For example, a detection confidence map (or function), may provide for each grid area (or focus analysis region) a confidence score. The confidence score may be indicative of the quality or sharpness of a portion of a diagnostic image of a diagnostic field. A diagnostic image is captured at an imaging focus configuration that may be selected according to a focusing step using a focus field as described herein. However, since the diagnostic plane may not be perfectly parallel to the focal plane of the microscope, some grid areas (or focus analysis regions) may be out of focus to some extent (or have a lower image quality). The degree of this focus variation is correlated to the difference between a focus configuration that would have been used for imaging a specific grid area (taken alone) and the actual imaging focus configuration that was selected for imaging the diagnostic area encompassing the grid area. Thus, a confidence score can be a function of a distance between the imaging focus configuration selected for imaging the diagnostic field (based on a focus field for example) and the focus configuration that would have been applied if only the given grid area would have been imaged. Such distance can have an effect on the image quality and subsequently on the accuracy of a diagnostic analysis performed on the diagnostic image at a corresponding grid area (or focus analysis region). The confidence score may define a potential expected computer vision and classification algorithm (hereon, the algorithm) quality as a function of that area focus quality. This confidence score may be used by/integrated into the algorithm in several ways.

(89) Optionally, the focus confidence score can be integrated into the algorithms in more complicated ways. One example is using the focus confidence score as a classifier of feature coordinates in the classification algorithms. In some cases, focus analysis regions having a focus



configuration that is too far (e.g. according to a given threshold, that in some cases can be defined by the depth of focus (DOF)) from the imaging focus configuration calculated for the focus field of a given diagnostic field that is being analyzed can optionally be ignored by the classification algorithms. For example, focus regions that are at least 1 DOF above or below imaging focus configuration for a given field, may be ignored.

(90) DOF is known as the distance between the nearest and farthest objects in a scene that appear acceptably sharp in an image. DOF is mostly a property of the microscope's objective lens and the magnification, the latter being determined by the resolution of interest. For example, for an object being about 1  $\mu\text{m}$  in dimensions (e.g. schizonts or some platelets), a resolution of at least 0.5  $\mu\text{m}$  would normally be required; similarly, for an object of about 2  $\mu\text{m}$  in dimension, a resolution of at least 1  $\mu\text{m}$  would normally be required. This also determines the magnification, and a magnification of at least 20 $\times$  would be used for a resolution of about 0.5  $\mu\text{m}$ , while a magnification of at least 10 $\times$  would be used for a resolution of about 1  $\mu\text{m}$ , in this connection, it is noted that a lens is chosen to provide a desired magnification. A lens is characterized by a numerical aperture (NA). For example, a lens for 20 $\times$  magnification may have a numerical aperture (NA) of about 0.4-0.5, while a lens for 10 $\times$  magnification may have a significantly smaller NA of about 0.2-0.25.

(91) According to the Shillaber equation, DOF relates to NA for a given wavelength of light ( $\lambda$ ) and medium refraction index ( $R_i$ ):

$$(92) \text{ DOF} = \frac{\sqrt{R_i - (\text{NA})^2}}{(\text{NA})^2}$$

(93) Below are provided non-limiting examples of DOF for several commercially available microscope objectives using 500 nm light and air as the medium ( $R_i=1.00$ ) between microscope objective and object:

(94) TABLE-US-00001 Numerical Depth of Aperture Field Magnification (NA) (DOF)    4 $\times$  0.10  
50    10 $\times$  0.25 7.7    20 $\times$  0.40 2.9    40 $\times$  0.65 0.9    60 $\times$  0.85 0.36    100 $\times$  0.95 0.17

(95) In some cases, when a suspect pathogen candidate is identified (e.g. when diagnosing a blood cell sample for the presence of a pathogen or when counting white blood cells in a cell sample) within an area of the diagnostic field that correlates with one or more focus analysis regions (in some cases for that purpose the focus analysis regions are correlated with the corresponding areas of the diagnostic field using any method and/or technique) having a focus confidence score lower than a given threshold or having a corresponding focus configuration that differs from a specific focus configuration used for acquiring the image or images of the diagnostic field by at least a threshold value, stricter scrutiny can be applied in analyzing the suspect candidate. For example, a new image or a set of images can be acquired at that location, at a calculated focus configuration having an improved focus confidence score (e.g. to confirm or negate the identification).

(96) In some cases the focus configuration data can be used for selecting a portion of the diagnostic field for analysis, such that the focus variation between the specific focus configuration in which the image or images of the diagnostic field were acquired, and the focus configuration of the focus analysis regions within the portion, is below a specific threshold, e.g. 2-5  $\mu\text{m}$  (non-limiting) or  $\pm 0.5-1$  DOF.

(97) It is to be noted that in some cases, the confidence score can be used for determining the size and/or shape of one or more diagnostic fields. In FIGS. 10A and 10B, an optical plane of a microscope is parallel to the shown XY plane, with the z axis being the optical axis of the microscope. As an example shown in FIG. 10A, the more slanted the mapping field 120 is along the X axis, the diagnostic fields can be narrower strips of the image. If the slant is in the Y axis, the strips will be perpendicular to those of the X axis. In cases where both the X and Y axis are slanted, the diagnostic fields can sometimes be defined as L shaped, as shown for diagnostic field 130 in FIG. 10B. It is to be noted that this is a mere example and a diagnostic field's size and/or shape can be determined in other manners. One example of such an alternative is to use the confidence score for determining a size of a rectangle or an ellipse or any other shape.

(98) It is to be noted that, with reference to FIG. 5 some of the blocks can be integrated into a consolidated block or can be broken down to a few blocks and/or other blocks may be added. It is to be further noted that some of the blocks are optional. It should be also noted that whilst the flow diagram is described also with reference to the system elements that realizes them, this is by no means binding, and the blocks can be performed by elements other than those described herein.

(99) FIG. 6 shows an illustration of a mapping field including a quality confidence score, in accordance with some embodiments of the presently disclosed subject matter.

(100) In the illustrated example, there is shown a rectangular diagnostic field **610**, split to twelve focus analysis regions (shown by a 3×4 table). Each focus analysis region has a corresponding focus configuration, where in the illustrated example the focus configuration is relative, however, as indicated above, this is by no means limiting. In addition to the focus configuration, each focus analysis region has a corresponding focus confidence score, having a value ranging from zero (e.g. for areas having a focus configuration that is less likely to be accurate) to one (e.g. for areas having a focus configuration that is most likely to be accurate). It is to be noted that the depicted illustration is by way of example only and is non-limiting.

(101) Attention is now drawn to FIG. 7, showing a flowchart illustrating steps of a method of imaging a cell sample, in accordance with some embodiments of the presently disclosed subject matter. The imaging method includes in a first stage, a method of determining a reference depth level within a cell sample and, in a second stage, focusing a digital microscope at an investigation level derived from the depth reference level. The method of determining a reference depth level may be carried out by system **400** (e.g. utilizing focus calculation module **440**). Advantageously, the system **400**, or parts thereof, may belong to an autofocus system of the microscope. The step of focusing the microscope may be performed automatically upon command by the autofocus system. The cell sample may comprise red blood cells and may optionally be a cell monolayer comprising red blood cells.

(102) In **700**, a series of images (also referred to as a set of images) representative of light captured by focusing a digital microscope at a corresponding series of depths levels within the cell sample is obtained. In some embodiments, obtaining the series of images comprises in-depth scanning of the cell sample with a digital microscope, for example by using an image sensor unit of the microscope connected with a computing module so as to provide the in-depth images (i.e. images captured during in-depth scanning) to the computing module. In some embodiments, in-depth scanning may be performed with brightfield illumination.

(103) The set (series) of images may be understood as a series of slices of the cell sample corresponding to different positions along the Z axis (optical axis of the microscope). Each image may be associated with a depth level. Optionally one or more images are associated with depth levels within the cell sample that are above or below the cells in cell sample. Optionally one or more images are associated with depth levels that are above or below the cell sample. The set of images may result from an in-depth scanning of the cell sample. Such in-depth scanning may for example be carried out by varying a distance between a focus plane of the microscope and a sample carrier intended to accommodate the cell sample by methods well known in the art.

(104) Further, the term depth level may be understood as a coordinate value along the optical axis of the microscope corresponding to a position that is optionally inside the cell sample. The actual direction of the axis and the origin of the axis to quantify depth may be arbitrarily chosen. The images may be obtained in any order, and may have an equal distance along the axis between pairs of consequent images or at a varying distance i.e. the in-depth scanning may be performed with a fixed step or with a variable step. For example, the origin of the axis may be positioned at an outer surface of the cell sample facing an objective of the microscope and the direction of the coordinate axis may be chosen so that the coordinates increase when progressing toward the cell sample. It is also understood that since the series of depth levels may be understood as an ordered set along the axis, it is possible to define endpoint depth levels (hereinafter referred to as endpoints) of the series

of depth levels. As used herein, the term scanning depth interval refers to a series of depth levels between two endpoints' depth levels. One endpoint level of a series of depth levels, for example a minimum depth level of the set, may be referred to as a first scanning depth levels and the other endpoint of the set, for example the maximum depth level, may be referred to as a second scanning depth level. In such cases, the scanning depth interval refers to the depth levels comprised between the first and second scanning depth levels.

(105) In some embodiments, an estimated reference depth level may be preliminarily provided. For example, the cell sample may comprise a plurality of fields to be investigated and reference depth levels determined for one or more previous fields may be used to estimate the estimated reference level for a subsequent field. In these embodiments, the scanning depth interval may be selected so as to cover the estimated depth reference level i.e. distances between the estimated depth reference level and the first and second scanning depth levels may be above a specific threshold. In some embodiments, a span of the depth scanning interval may be of around 5 micrometers to 1000 micrometers. In some embodiments, the span of the depth scanning interval may be between 150 and 250 micrometers, or less than 50 micrometers or even between 10 and 30 micrometers. Optionally, the estimated depth level is approximately in the midpoint of the span of the depth scanning interval.

(106) In **710**, the series of images and associated depth levels (or the data representative thereof) are processed for detecting at least one depth level corresponding to a drop in image contrast and the detected depth level is identified to be the reference depth level. The detected depth level may be such that an image contrast at the detected depth level is lower than the image contrast at the depth level immediately preceding and following the reference depth level (i.e. adjacent depth levels) in the series of depth levels. The drop in image contrast may be understood as a drop of image contrast over depth level (i.e. as a function of depth level). It is noted that when the contrast function used to calculate the image contrast is increasing with the contrast, **710** may be carried out by detecting a well of a contrast curve representing image contrast as a function of depth level. Image contrast of an image may be provided by applying a contrast function to the image. A well is considered to be formed on the contrast curve when a contrast function value is inferior at least to the previous and subsequent adjacent contrast function values. As used herein, some embodiments are described in which the image contrast is provided by the calculation of variance. It is understood that other functions can be contemplated to determine the contrast of an image. The set of images associated with the series of depth levels within the cell sample enables to analyze variations of an image parameter as a function of the depth level.

(107) In some embodiments, image variance may be computed for every image of the set of obtained images. FIG. 8 illustrates image variance calculation on an example image I comprising  $n \times p$  pixels ( $n, p$  integers) of pixel intensity  $I_{sub.ij}$  wherein  $1 \leq i \leq n$  and  $1 \leq j \leq p$ . The variance can be expressed as follows:

$$Var(I) = E[(I_{sub.ij} - E(I))^2]$$
, wherein  $E(I)$  is the mean value of the pixel intensity  $I_{sub.ij}$  over the example image.

(108) In some embodiments, a variance related value may be computed for each image of the set of images. It is understood that the variance related value encompasses transformations of the image variance enabling to derive the image variance i.e. transformations equivalent to image variance calculation, for example standard deviation.

(109) It is understood that implementation of the present disclosure does not require generation of the actual curve representing the variation of the contrast function over depth level but that a search for a well (or a roof if the contrast function is decreasing with the contrast) can be performed mathematically using the image representative data.

(110) In some embodiments, the cell sample may comprise predominantly red blood cells. In some embodiments, the cell sample may essentially be a monolayer of cells, wherein at least 80% of the cells or even at least 90% of the cells have direct contact with the surface on which the cell sample

is held. In the context of imaging blood samples, the Applicant found that the proposed method based on determining a reference depth level corresponding to a minimum of variance (contrast and/or sharpness) over the in-depth scanning, may be particularly advantageous.

(111) Indeed, the proposed method can be performed using brightfield illumination and can provide appropriate results for further investigation using fluorescent illumination. This may lead to reducing focus time because performing focus on fluorescent images is typically slower than when using brightfield, since obtaining fluorescent images typically requires longer exposure periods. Further, obtaining fluorescent images for focusing purpose may be problematic because fluorescent imaging is known to degrade the fluorescent response of the sample due to photo-bleaching.

(112) Applying contrast-based functions and/or sharpness based functions to brightfield microscopy of a blood sample containing mostly red blood cells (for example a blood sample) may yield graphs qualitatively similar to that shown in FIG. 9. FIG. 9 illustrates a curve representing variations of image variance (image contrast) over scanning depth level. The curve notably comprises a well 37 embraced between two maxima 38, 39 (in this example a saddle point which is a local minimum, but not an absolute one).

(113) The Applicant found that the depth level corresponding to the well 37 provides an efficient reference level providing robust and consistent results across different microscope hardware including different forms of brightfield illumination and different forms of sample preparation (dry thin smears, wet smears and microfluidic preparation). Moreover, a focus position based on the well 37 position provides a baseline for epifluorescent imaging.

(114) Therefore, imaging at the reference depth level or in its vicinity may provide efficient parasite detection. The Applicant further found that the consistency of the focus generated by the proposed method of determining a minimum of the contrast function may be explained as follows: the maxima 38, 39 surrounding the well 37 typically fall within 1 micrometer of each other. Consequently, the well 37 is steep thereby causing discernible shifts in the contrast function even for shifts of depth level of about a tenth of a micron. It is appreciated that having a consistent reference level within a cell sample enables to establish reliable automated diagnosis.

(115) Returning to FIG. 7, in 720 the digital microscope may be focused at an investigation level based on the determined reference level. In some embodiments, the investigation level may be equal to the reference level. In some embodiments, the investigation level may be shifted by a specific value with respect to the reference level. For example, this value may be in the range of 0.2-3 micrometers, or about 1-2 micrometers or about 1.5 micrometer. In some embodiments, switching to an investigation level that is different than the reference depth value enables to increase the contrast and/or sharpness of the image while preserving the consistency provided by the aforementioned method of determining a reference depth level. As explained above, focusing the microscope at the investigation level may enable to investigate the cell sample. In some embodiments, the investigation may be carried out with fluorescent illumination and/or with brightfield illumination. In some embodiments, the investigation level will provide a sharp image (or even the sharpest and/or highest contrast image).

(116) It is to be understood that the presently disclosed subject matter is not limited in its application to the details set forth in the description contained herein or illustrated in the drawings. The presently disclosed subject matter is capable of other embodiments and of being practiced and carried out in various ways. Hence, it is to be understood that the phraseology and terminology employed herein are for the purpose of description and should not be regarded as limiting. As such, those skilled in the art will appreciate that the conception upon which this disclosure is based may readily be utilized as a basis for designing other structures, methods, and systems for carrying out the several purposes of the present presently disclosed subject matter.

(117) It will also be understood that the system according to the presently disclosed subject matter can be implemented, at least partly, as a suitably programmed computer. Likewise, the presently disclosed subject matter contemplates a computer program being readable by a computer for

executing the disclosed method. The presently disclosed subject matter further contemplates a machine-readable memory tangibly embodying a program of instructions executable by the machine for executing the disclosed method.

(118) Unless specifically stated otherwise, as apparent from the above discussions, it is appreciated that throughout the specification discussions utilizing terms such as “obtaining”, “calculating”, “detecting”, “analyzing”, “correlating”, “capturing”, “selecting” or the like, include action and/or processes of a computer that manipulate and/or transform data into other data, said data represented as physical quantities, e.g. such as electronic quantities, and/or said data representing the physical objects. The terms “computer”, “processor”, and “controller” should be expansively construed to cover any kind of electronic device with data processing capabilities, including, by way of non-limiting example, a personal computer, a server, a computing system, a communication device, a processor (e.g. digital signal processor (DSP), a microcontroller, a field programmable gate array (FPGA), an application specific integrated circuit (ASIC), etc.), any other electronic computing device, and or any combination thereof.

(119) The operations in accordance with the teachings herein may be performed by a computer specially constructed for the desired purposes or by a general purpose computer specially configured for the desired purpose by a computer program stored in a non-transitory computer readable storage medium. The term “non-transitory” is used herein to exclude transitory, propagating signals, but to otherwise include any volatile or non-volatile computer memory technology suitable to the application.

(120) As used herein, the phrase “for example,” “such as”, “for instance” and variants thereof describe non-limiting embodiments of the presently disclosed subject matter. Reference in the specification to “one case”, “some cases”, “other cases” or variants thereof means that a particular feature, structure or characteristic described in connection with the embodiment(s) is included in at least one embodiment of the presently disclosed subject matter. Thus the appearance of the phrase “one case”, “some cases”, “other cases” or variants thereof does not necessarily refer to the same embodiment(s).

(121) It is appreciated that, unless specifically stated otherwise, certain features of the presently disclosed subject matter, which are, for clarity, described in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the presently disclosed subject matter, which are, for brevity, described in the context of a single embodiment, may also be provided separately or in any suitable sub-combination.

(122) In embodiments of the presently disclosed subject matter, fewer, more and/or different stages than those shown in FIGS. 5-7 may be executed. In embodiments of the presently disclosed subject matter one or more stages illustrated in FIGS. 5-7 may be executed in a different order and/or one or more groups of stages may be executed simultaneously. FIG. 4 illustrates a general schematic of the system architecture in accordance with an embodiment of the presently disclosed subject matter. Each module in FIG. 4 can be made up of any combination of software, hardware and/or firmware that performs the functions as defined and explained herein. The modules in FIG. 4 may be centralized in one location or dispersed over more than one location. In other embodiments of the presently disclosed subject matter, the system may comprise fewer, more, and/or different modules than those shown in FIG. 4.

## Claims

1. A method for use with a digital microscope unit that includes a digital microscope, comprising: receiving a biological cell sample that is disposed within a sample carrier, into the digital microscope unit; determining that there is a variation in the focal depth of the biological sample with respect to the microscope due to curvature in the sample carrier and/or due to tolerance in setup of the microscope, by determining a focus configuration at which the microscope should

image the biological cell sample, by using variations in intensity-related data between images acquired at a plurality of different focus configurations and deriving information relating to the focus configuration at which the microscope should image the biological cell sample based on the variations in intensity-related data between images acquired at a plurality of different focus configurations; capturing at least one image of the biological cell sample disposed within the sample carrier; determining a characteristic of the biological cell sample by analyzing the at least one captured image, the analyzing of the at least one captured image comprising, in response to determining that there is the variation in the focal depth of the biological sample with respect to the microscope, accounting for the variation in the focal depth of the biological sample with respect to the microscope, by using the calculated data that are indicative of focus variation as an input for calculating focus confidence scores indicative of levels of confidence in focus configurations calculated for respective focus analysis regions.

2. The method according to claim 1, wherein the biological cell sample includes a blood sample, and wherein determining a characteristic of the biological cell sample comprises determining a characteristic of the blood sample.

3. The method according to claim 2, wherein determining a characteristic of the blood sample comprises detecting a presence of a pathogen suspected to be included in the blood sample.

4. The method according to claim 2, wherein determining a characteristic of the blood sample comprises performing a blood count on the blood sample.

5. The method according to claim 1, the method further comprising identifying an entity within the sample by analyzing the at least one captured image, wherein identifying the entity comprises: identifying a candidate of the entity within the sample that is associated with a focus confidence score that is lower than a threshold; and in response thereto, acquiring one or more images of the candidate using different imaging parameters to those previously used to image the candidate.

6. Apparatus for use with a biological cell sample that is disposed within a sample carrier, the apparatus comprising: a microscopy unit, comprising: a digital microscope; a surface configured to receiving the biological cell sample that is disposed within the sample carrier; and a computer processor configured: to determine that there is a variation in the focal depth of the biological sample with respect to the microscope due to curvature in the sample carrier and/or due to tolerance in setup of the microscope; to determine a focus configuration at which the microscope should image the biological cell sample, by using variations in intensity-related data between images acquired at a plurality of different focus configurations and deriving information relating to the focus configuration at which the microscope should image the biological cell sample based on the variations in intensity-related data between images acquired at a plurality of different focus configurations; to capture at least one image of the biological cell sample that is disposed within the sample carrier; to determine a characteristic of the biological sample by analyzing the at least one captured image, the analyzing of the at least one captured image comprising, in response to determining that there is the variation in the focal depth of the biological sample with respect to the microscope, accounting for the variation in the focal depth of the biological sample with respect to the microscope, by using the calculated data that are indicative of focus variation as an input for calculating focus confidence scores indicative of levels of confidence in focus configurations calculated for respective focus analysis regions.

7. The apparatus according to claim 6, wherein the biological cell sample includes a blood sample, and the computer processor is configured to determine a characteristic of the blood sample.

8. The apparatus according to claim 7, wherein the computer processor is configured to detect a presence of a pathogen suspected to be included in the blood sample.

9. The apparatus according to claim 7, wherein the computer processor is configured to perform a blood count on the blood sample.

10. The apparatus according to claim 6, wherein the computer processor is further configured to identify an entity within the sample by analyzing the at least one captured image, by: identifying a

candidate of the entity within the sample that is associated with a focus confidence score that is lower than a threshold; and in response thereto, acquiring one or more images of the candidate using different imaging parameters to those previously used to image the candidate.

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