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(54) **SYRINGE FOR CRYOPRESERVATION OF CELLS, NUCLEIC ACIDS, OR PROTEINS**

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(57) **ABSTRACT**

According to the present invention, a syringe for cryopreservation of cells, a nucleic acid, or a protein comprising: a barrel having a spout and an opening and for containing cells, a nucleic acid, or a protein; a gasket slidably accommodated in the barrel; a plunger connected to the gasket; and a cap that seals the spout of the barrel, wherein the barrel is formed from a fluororesin, and the gasket is formed from a fluororesin or a polyethylene resin. The syringe is excellent in usability, durability, and safety.

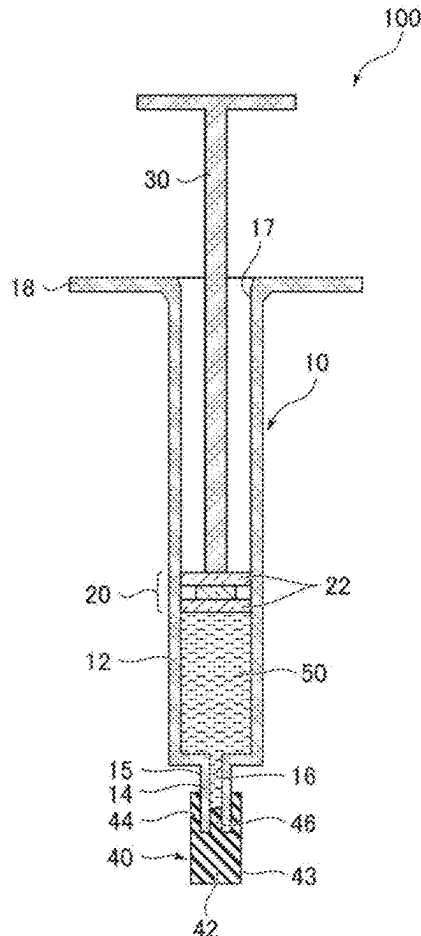
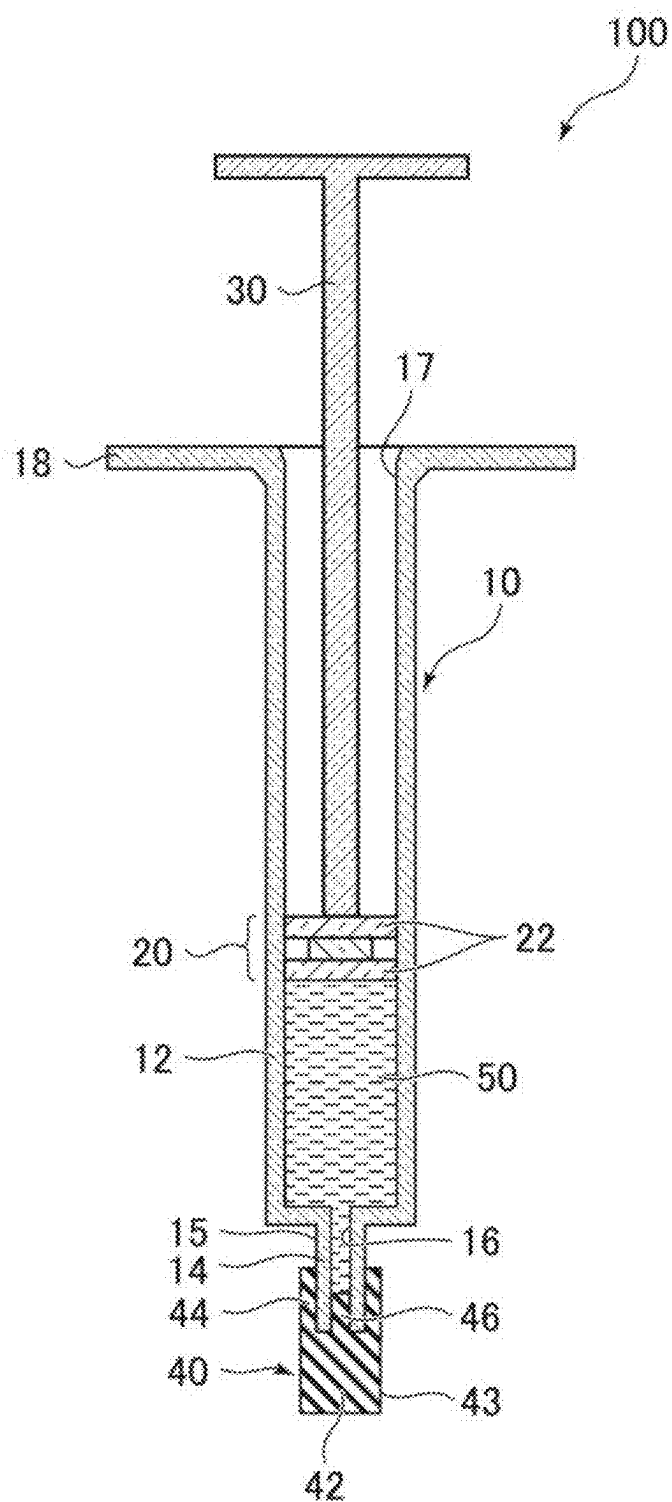


Fig. 1



SYRINGE FOR CRYOPRESERVATION OF CELLS, NUCLEIC ACIDS, OR PROTEINS

TECHNICAL FIELD

[0001] The present invention relates to a syringe for cryopreservation of cells, a nucleic acid, or a protein, a prefilled syringe, and a method for cryopreserving cells, a nucleic acid, or a protein.

BACKGROUND ART

[0002] In recent years, opportunities for using various cells are increasing in the medical field, the field of pharmaceutical manufacturing technology, and the field of testing and research of medicine, pharmacy, agriculture, and biology or the like. In particular, due to the development of stem cell research including iPS cells (induced pluripotent stem cells), there are increasing expectations for practical application of cell transplantation therapy in the field of regenerative medicine.

[0003] Therapeutic cells such as stem cells used in regenerative medicine are often administered to patients through culture processes such as selection, concentration, and proliferation depending on the purpose. Therefore, therapeutic cells often have opportunities to be transported between medical facilities and cell processing and culturing facilities or preserved at those facilities, before being administered to patients. In order to maintain the viability of the therapeutic cells and maintain the therapeutic ability of the cells during transportation and preservation, therapeutic cells generally need to be frozen at a very low temperature such as -80°C . or less. Accordingly, containers storing therapeutic cells are also required to withstand extremely low temperatures during transportation and preservation of therapeutic cells.

[0004] Further, therapeutic cells will eventually be injected into patients, and syringes are generally used for injecting the therapeutic cells to patients. Replacing the therapeutic cells from containers for transportation and preservation into syringes is laborious, and there are risks of the mix-up of therapeutic cells in the process, microbial contamination, and foreign matter contamination. Therefore, in the field of regenerative medicine, there is a need for prefilled syringes in which therapeutic cells are stored in advance, in view of the work efficiency and the sanitary conditions. When the contents of prefilled syringes are cells, freezing at very low temperatures of -80°C . or less is required during transportation and preservation, as described above, and therefore the syringes of course need to withstand such very low temperatures of -80°C . or less.

[0005] Commercially available syringes for medical care generally have a barrel formed from polypropylene-based materials and a gasket formed from rubber-based materials. However, since polypropylene-based materials and rubber-based materials generally easily embrittle at a low temperature, syringes by combining these materials cannot withstand a very low temperature such as -80°C . or less. Accordingly, when preservation at a very low temperature such as -80°C . or less is assumed, use of a special syringe is required.

[0006] Patent Document 1 discloses a syringe for freezing and thawing therapeutic cell mixtures, comprising a barrel made of a cyclic olefin polymer or the like for reasons such as transparency. Further, Patent Document 2 discloses a syringe for storing and dispensing cryopreserved cells,

which is made of a cycloolefin polymer or cycloolefin copolymer as a biocompatible material. However, it is disclosed that the syringe of Patent Document 1 requires a special structure such as a cap or a seal near the opening of a barrel, since the gasket alone is not sufficient to protect the sterility in the barrel during preservation at a temperature lower than -80°C . Further, the syringe of Patent Document 2 also requires a cap at the opening of a barrel as an essential configuration although it includes a gasket in the barrel. This is probably because, similarly to the syringe of Patent Document 1, it is impossible to prevent contamination into the barrel with microorganisms or foreign matter without the cap at the opening of the barrel in addition to the gasket in the barrel under very low temperature conditions during cell freezing.

[0007] The 17th edition of the Japanese Pharmacopoeia states that it is necessary to confirm that plastic pharmaceutical containers used for injections are “tight containers free from microorganisms”. Here, the term “tight containers” refer to “containers that are impervious to solid or liquid foreign matter and can prevent the loss, efflorescence, deliquescence, or evaporation of the pharmaceuticals stored under normal handling, transportation, or preservation”. Accordingly, it is essential for plastic syringes, especially for medical use, to have at least the ability to prevent the intrusion of microorganisms, solid foreign matter, or liquid foreign matter (that is, integrity).

[0008] Known cell syringes for cryopreservation require a special structure for maintaining the integrity during freezing and thawing such as a cap and a seal at the opening of the barrel, which is not found in general syringes, as mentioned above. However, since such a special structure needs to be removed during use of syringes, and the user must also perform the operation to connect the gasket and the plunger, it takes time and effort to use syringes, thereby increasing the work load. Thus, there is a risk of hindering prompt medical practice especially in the medical field. In addition, during these operations, a strong force may be partially applied to the barrel, the gasket, or the plunger of syringes, which may cause distortion or cracking, leading to contamination of syringes with microorganisms or foreign matter. Further, there is also a problem that the need for a special structure such as a cap or a seal, which is not found in general syringes, leads to an increase in manufacturing costs of syringes.

[0009] As a method for improving the integrity of a syringe, there is a technique of forming the gasket from a rubber-based material that has excellent adhesion to the inner wall of the barrel, or laminating the gasket with a rubber-based material. However, rubber-based materials generally vitrify to embrittle at a very low temperature such as -80°C . or less, as mentioned above, and there is a concern that the durability may decrease. As another method, there is also a technique of coating the inside of the barrel or the gasket with a silicone compound. Since the use of a silicone compound may also contribute to the improvement of slidability, they are often used for coating syringes. However, when the contents of syringes contain proteins, the silicone compound may cause aggregation of proteins, which may lead to serious safety problems especially in syringes intended for medical use.

PRIOR ART DOCUMENTS

Patent Documents

- [0010] [Patent Document 1] Japanese unexamined Patent Application Publication (Translation of PCT Application) No. 2020-510477
- [0011] [Patent Document 2] Japanese unexamined Patent Application Publication (Translation of PCT Application) No. 2008-507536

SUMMARY OF THE INVENTION

Object to be Solved by the Invention

[0012] It is an object of the present invention is to provide a syringe for cryopreservation of cells, a nucleic acid, or a protein that can maintain its integrity during preservation not only at room temperature but also at a very low temperature and heating and is excellent in usability, durability, and safety, without requiring a special structure such as a cap or a seal at the opening of the barrel or using a silicone compound or a rubber-based material.

Means to Solve the Object

[0013] As a result of dedicated studies in order to solve the aforementioned problems, the inventor has found that a syringe comprising: a barrel having a spout and an opening, and for storing a cell, a nucleic acid, or a protein; a gasket slidably accommodated in the barrel; a plunger connected to the gasket; and a cap that seals the spout of the barrel, wherein the barrel is formed from a fluororesin, and the gasket is formed from a fluororesin or a polyethylene resin, is excellent in usability, durability, and safety, and can be used for cryopreservation of cells, a nucleic acid, or a protein, and thereby accomplishing the present invention.

[0014] That is, the present invention is as specified by the following items.

[0015] (1) A syringe for cryopreservation of a cell, a nucleic acid, or a protein, comprising:

[0016] a barrel having a spout and an opening, and for storing a cell, a nucleic acid, or a protein;

[0017] a gasket slidably accommodated in the barrel;

[0018] a plunger connected to the gasket; and

[0019] a cap that seals the spout of the barrel,

[0020] wherein the barrel is formed from a fluororesin, and

[0021] the gasket is formed from a fluororesin or a polyethylene resin.

[0022] (2) The syringe according to (1) above, wherein

[0023] the barrel is formed from a fluororesin selected from the group consisting of a tetrafluoroethylene-hexafluoropropylene-based copolymer and a tetrafluoroethylene-perfluoroalkyl vinyl ether-based copolymer, and

[0024] the gasket is formed from a fluororesin selected from the group consisting of a tetrafluoroethylene-hexafluoropropylene-based copolymer, a tetrafluoroethylene-perfluoroalkyl vinyl ether-based copolymer, and polytetrafluoroethylene, or a polyethylene resin selected from the group consisting of low density polyethylene, medium density polyethylene, high density polyethylene, and ultrahigh molecular weight polyethylene.

[0025] (3) The syringe according to (1) or (2) above, wherein the fluororesin is a fluororesin having a total number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends in the fluororesin of 70 or less per 1×10^6 of carbon atoms.

[0026] (4) The syringe according to any one of (1) to (3) above, wherein the fluororesin is a fluororesin containing one or more $-\text{CF}_3$ end groups.

[0027] (5) The syringe according to any one of (1) to (4) above, wherein the cap is formed from a fluororesin or a polyethylene resin.

[0028] (6) The syringe according to any one of (1) to (5) above, wherein the plunger is formed from a fluororesin or a polyethylene resin.

[0029] (7) The syringe according to any one of (1) to (6) above, wherein the syringe is further stored in an outer bag.

[0030] (8) The syringe according to (7) above, wherein the outer bag is formed from a fluororesin or a polyethylene resin.

[0031] (9) The syringe according to any one of (1) to (8) above, wherein the syringe is free of silicone compounds.

[0032] (10) The syringe according to any one of (1) to (9) above, wherein the syringe is free of elastomers, natural rubber, or synthetic rubber.

[0033] (11) The syringe according to any one of (1) to (10) above, wherein the cell is a mammalian cell.

[0034] (12) The syringe according to any one of (1) to (11) above, wherein the syringe is for use in regenerative medicine.

[0035] (13) The syringe according to any one of (1) to (12) above, wherein the syringe is used at -80°C . or less for cryopreservation.

[0036] (14) The syringe according to any one of (1) to (12) above, wherein the syringe is used at -150°C . or less for cryopreservation.

[0037] (15) A prefilled syringe for cryopreserving a cell, a nucleic acid, or a protein, wherein the cell, the nucleic acid, or the protein is stored in the syringe according to any one of (1) to (14) above.

[0038] (16) The prefilled syringe according to (15) above, wherein the prefilled syringe is used at -80°C . or less for cryopreservation.

[0039] (16') The prefilled syringe according to (15) above, wherein the cell, the nucleic acid, or the protein is cryopreserved in the syringe at -80°C . or less.

[0040] (17) The prefilled syringe according to (15) above, wherein the prefilled syringe is used at -150°C . or less for cryopreservation.

[0041] (17') The prefilled syringe according to (15) above, wherein the cell, the nucleic acid, or the protein is cryopreserved in the syringe at -150°C . or less.

[0042] (18) A method for cryopreserving a cell, a nucleic acid, or a protein, comprising cryopreserving the cell, the nucleic acid, or the protein in the syringe according to any one of (1) to (14) above.

[0043] (19) The method according to (18) above, wherein the cryopreservation is a cryopreservation at -80°C . or less.

[0044] (20) The method according to (18) above, wherein the cryopreservation is a cryopreservation at -150°C . or less.

[0045] As another embodiment of the present invention, the following can be mentioned, for example.

[0046] (21) A method for cryopreserving a cell, a nucleic acid, or a protein, the method comprising the steps of:

[0047] providing a cell, a nucleic acid, or a protein to be cryopreserved;

[0048] providing a syringe comprising: a barrel having a spout and an opening and for storing the cell, a nucleic acid, or a protein; a gasket slidably accommodated in the barrel, a plunger connected to the gasket; and a cap that seals the spout of the barrel, wherein the barrel is formed from a fluororesin, and the gasket is formed from a fluororesin or a polyethylene resin;

[0049] storing the cell, a nucleic acid, or a protein in the barrel of the syringe and sealing the barrel with the gasket and the cap; and

[0050] putting the syringe storing the cell, the nucleic acid, or the protein under a desired low temperature to cryopreserve the cell, the nucleic acid, or the protein.

[0051] (22) A method for administering a cell, a nucleic acid, or a protein, the method comprising the steps of:

[0052] providing cells, a nucleic acid, or a protein to be administered;

[0053] providing a syringe comprising: a barrel having a spout and an opening and for storing the cell, nucleic acid, or protein; a gasket slidably accommodated in the barrel, a plunger connected to the gasket; and a cap that seals the spout of the barrel, wherein the barrel is formed from a fluororesin, and the gasket is formed from a fluororesin or a polyethylene resin;

[0054] storing the cells, the nucleic acid, or the protein in the barrel of the syringe and sealing the barrel with the gasket and the cap;

[0055] cryopreserving the cell, the nucleic acid, or the protein by putting the syringe storing the cell, the nucleic acid, or the protein under a desired low temperature;

[0056] thawing the cell, the nucleic acid, or the protein by putting the syringe under a desired high temperature

[0057] administering the thawed cell, the thawed nucleic acid, or the thawed protein to a subject in need of administration of the cell, nucleic acid, or protein from the syringe.

[0058] (23) The method according to (22) above, wherein the subject is a mammal (for example, a human).

[0059] (24) The method according to (22) or (23) above, wherein the desired high temperature is room temperature, a temperature of 30° C. or more, or 37° C.

[0060] (25) The method according to any one of (21) to (24) above, wherein the desired low temperature is a temperature of -80° C. or less.

[0061] (26) The method according to any one of (21) to (24) above, wherein the desired low temperature is a temperature of -150° C. or less.

[0062] (27) Use of the syringe according to any one of (1) to (14) above, for cryopreserving a cell, a nucleic acid, or a protein.

[0063] (28) The use according to (27) above, wherein the cryopreservation is a cryopreservation at -80° C. or less.

[0064] (29) The use according to (27) above, wherein the cryopreservation is a cryopreservation at -150° C. or less.

[0065] (30) A kit comprising the syringe according to any one of (1) to (12) above, and an instruction for use describing the method according to any one of (18) to (26) above (that is, for performing the method according to any one of (18) to (26) above).

[0066] The syringe of the present invention is a syringe which use is limited as “for use in the cryopreservation of a cell, a nucleic acid, or a protein”. Therefore, it can be said that the present invention is a use limited invention.

Effect of the Invention

[0067] According to the present invention, a syringe for cryopreservation of cells, a nucleic acid, or a protein that has sufficient impact resistance and integrity during preservation not only at room temperature but also at a very low temperature and good slidability, and is excellent in usability, durability, and safety can be provided without requiring use of a special structure for imparting integrity or using a silicone compound or a rubber-based material, and in particular, it can contribute to the field of regenerative medicine.

BRIEF DESCRIPTION OF DRAWINGS

[0068] FIG. 1 is a schematic view showing a configuration of the syringe of this embodiment during preservation.

MODE OF CARRYING OUT THE INVENTION

[0069] Hereinafter, an embodiment for carrying out the present invention (which will be hereinafter referred to simply as “this embodiment”) will be described in detail. The following embodiment is shown as an example for describing the present invention, and the present invention is not limited to the embodiment. The present invention can be appropriately modified and implemented within the scope of the gist thereof.

<Configuration of Syringe>

[0070] The configuration of the syringe of this embodiment will be described. The following configuration is an aspect of the syringe of this embodiment, and the syringe of the present invention is not limited by the following configuration.

[0071] FIG. 1 is a schematic view showing a configuration of the syringe of this embodiment during preservation. As shown in FIG. 1, the syringe 100 comprises a barrel 10, a gasket 20, a plunger 30, and a cap 40. The syringe 100 is a so-called prefilled syringe that stores cells 50 in an enclosed state. The syringe 100 is stored in an outer bag 60 (not shown). In the present invention, the term “enclosed state” means that the exchange of liquids and solids inside and outside the barrel is blocked.

[0072] As shown in FIG. 1, the barrel 10 has a cylindrical body 12, and the cells 50 are stored therein. Further, the cylindrical body 12 has a spout 14 for spouting (discharging) the cells 50 on its tip and has an opening 17 into which the gasket 20 is inserted on the other end. Further, the opening 17 has a finger grip 18.

[0073] As shown in FIG. 1, the cap 40 is attached on the spout 14 of the cylindrical body 12 during preservation of the syringe 100. The cap 40 is detached when the syringe 100 is used, and an injection needle (not shown) is attached on the spout 14. The spout 14 consists of a portion having a smaller diameter than that of the cylindrical body 12.

[0074] The gasket 20 is connected to one end of the plunger 30 and slidably installed within the barrel 10. The gasket 20 is accommodated in the barrel 10 through the opening 17 of the barrel 10, adheres closely the inner surface of the barrel 10, and is pushed to move in a direction from the opening 17 toward the spout 14 when the syringe 100 is used (that is, the gasket 20 slides within the barrel 10). When the gasket 20 is thus pushed to move, the cells 50 in the barrel 10 are subjected to a pressure in the same direction. As described above, an injection needle (not shown) is attached on the spout 14 when the syringe 100 is used, and the cells 50 subjected to the pressure by the gasket 20 are spouted through the injection needle (not shown) via the spout 14. The gasket 20 is composed of two gasket lips 22.

[0075] The cap 40 comprises a substrate 42, a mantle tube 44 fitted from a circumferential edge 43 of the substrate 42 to an outer circumferential surface 15 of the spout 14 of the barrel 10, and a projection 46 protruding from an inner center of the substrate 42 and fitted into an inner circumferential surface 16 of the spout 14 of the barrel 10. The mantle tube 44 is fitted and adheres closely to the outer circumferential surface 15 of the spout 14 of the barrel 10, and the projection 46 is fitted and adheres closely into the inner circumferential surface 16 of the spout 14 of the barrel 10, thereby sealing the spout 14 of the barrel 10.

[0076] Hereinafter, each member will be described.

<Barrel>

[0077] In the present invention, the barrel is formed from a fluororesin. Specifically, examples of the fluororesin in the present invention can include polytetrafluoroethylene (PTFE), tetrafluoroethylene (TFE)-ethylene copolymer (ETFE), tetrafluoroethylene (TFE)-hexafluoropropylene (HFP)-based copolymer (FEP), and TFE-perfluoroalkyl vinyl ether (PAVE)-based copolymer (PFA).

[0078] The “TFE-HFP-based copolymer” means a copolymer containing at least TFE and HFP. That is, the “TFE-HFP-based copolymer” includes ternary copolymers such as a copolymer of TFE, HFP, and vinyl fluoride (VF) (TFE/HFP/VF copolymer), a copolymer of TFE, HFP, and vinylidene fluoride (VDF) (TFE/HFP/VDF copolymer), and a copolymer of TFE, HFP, and perfluoro(alkyl vinyl ether) (PAVE) (TFE/HFP/PAVE copolymer), quaternary copolymers such as a copolymer of TFE, HFP, VF, and VDF (TFE/HFP/VF/VDF copolymer), a copolymer of TFE, HFP, VF, and PAVE (TFE/HFP/VF/PAVE copolymer), and a copolymer of TFE, HFP, VDF, and PAVE (TFE/HFP/VDF/PAVE copolymer), and quinary copolymers such as a copolymer of TFE, HFP, VF, VDF, and PAVE (TFE/HFP/VF/VDF/PAVE copolymer), in addition to binary copolymers such as a copolymer of TFE and HFP (TFE/HFP copolymer; FEP).

[0079] The TFE-HFP-based copolymer is preferably a TFE/HFP copolymer and a TFE/HFP/PAVE copolymer. The mass ratio of TFE and HFP in such a TFE/HFP copolymer is preferably 80 to 97/3 to 20, more preferably 84 to 92/8 to 16. When the mass ratio of HFP in the TFE/HFP copolymer is lower than the aforementioned numerical range, the

formability is poor, and when it is higher than the aforementioned numerical range, the mechanical strength and the hardness are low, and the dimensional stability is poor. Further, the mass ratio of TFE, HFP, and PAVE in the TFE/HFP/PAVE copolymer is preferably 70 to 97/3 to 20/0.1 to 10, more preferably 81 to 92/5 to 16/0.3 to 5. When the mass ratio of HFP and PAVE in the TFE/HFP/PAVE copolymer is lower than the aforementioned numerical range, the formability is poor, and when it is higher than the aforementioned numerical range, the mechanical strength and the hardness are low, and the dimensional stability is poor.

[0080] The “TFE-PAVE-based copolymer” means a copolymer containing at least TFE and PAVE. That is, the “TFE-PAVE-based copolymer” includes ternary copolymers such as a copolymer of TFE, PAVE, and hexafluoropropylene (HFP) (TFE/PAVE/HFP copolymer), a copolymer of TFE, PAVE, and vinylidene fluoride (VDF) (TFE/PAVE/VDF copolymer), and a copolymer of TFE, PAVE, and chlorotrifluoroethylene (CTFE) (TFE/PAVE/CTFE copolymer), quaternary copolymers such as a copolymer of TFE, PAVE, HFP, and VDF (TFE/PAVE/HFP/VDF copolymer), a copolymer of TFE, PAVE, HFP, and CTFE (TFE/PAVE/HFP/CTFE copolymer), and a copolymer of TFE, PAVE, VDF, and CTFE (TFE/PAVE/VDF/CTFE copolymer), and quinary copolymers such as a copolymer of TFE, PAVE, HFP, VDF, and CTFE (TFE/PAVE/HFP/VDF/CTFE copolymer), in addition to binary copolymers such as a copolymer of TFE and PAVE (TFE/PAVE copolymer; PFA).

[0081] The PAVE constituting the PAVE unit is not specifically limited, and examples thereof can include perfluoro(methyl vinyl ether) [PMVE], perfluoro(ethyl vinyl ether) [PEVE], perfluoro(propyl vinyl ether) [PPVE], perfluoro(butyl vinyl ether), perfluoro(pentyl vinyl ether), perfluoro(hexyl vinyl ether), and perfluoro(heptyl vinyl ether).

[0082] The mass ratio of TFE and PAVE in the TFE-PAVE-based copolymer is preferably 90 to 98/2 to 10, more preferably 92 to 97/3 to 8. When the mass ratio of PAVE in the TFE-PAVE-based copolymer is lower than the aforementioned numerical range, the formability is poor, and when it is higher than the aforementioned numerical range, the mechanical strength and the hardness are low, and the dimensional stability is poor.

[0083] Further, the fluororesin in the present invention preferably has a total number of non-fluorinated group ends (for example, functional groups such as —COF, —COOH, —COOH associated with water, —CH₂OH, —CONH₂, and —COOCH₃) and —CF₂H group ends in the fluororesin of 70 or less per 1×10⁶ of carbon atoms, preferably 50 or less per 1×10⁶ of carbon atoms, more preferably 35 or less per 1×10⁶ of carbon atoms, for suppressing the adsorption of cells, a nucleic acid, or a protein stored in the barrel onto the inner surface of the barrel and/or for suppressing the reduction in cell viability. Further, it is more preferably 20 or less per 1×10⁶ of carbon atoms, particularly preferably 10 or less per 1×10⁶ of carbon atoms. Further, it is possible to not contain —CF₂ end group, and in case of not containing —CF₂ end group, the number of non-fluorinated group ends in the fluororesin is preferably 70 or less per 1×10⁶ of carbon atoms, and more preferably 35 or less. Further, it is more preferably 20 or less per 1×10⁶ of carbon atoms, particularly preferably 10 or less per 1×10⁶ of carbon atoms. The numbers of —COF, —COOH, —COOH associated with

water, $-\text{CH}_2\text{OH}$, $-\text{CONH}_2$, $-\text{COOCH}_3$, and $-\text{CF}_2\text{H}$ per 1×10^6 of carbon atoms can be calculated by FT-IR.

[0084] The fluororesin in the present invention is further preferably a fluororesin containing one or more $-\text{CF}_3$ end groups, to have a more stabilized end structure. The fluororesin containing one or more $-\text{CF}_3$ end groups effectively suppresses the adsorption of cells, a nucleic acid, or a protein onto the inner surface of the barrel and/or the reduction in cell viability. The $-\text{CF}_3$ end groups can be analyzed by ^{19}F NMR measurement at high temperature.

[0085] Such a fluororesin can be fabricated by fluorination using a known method such as a method of performing stabilization by bringing the end group of a fluororesin synthesized according to a conventional method such as suspension polymerization or emulsion polymerization into contact with a fluorine-containing compound (for example, fluorine radical source) before melt extrusion of the fluororesin or a method of performing fluorination by bringing pellets of the fluororesin obtained into contact with a fluorine-containing compound after melt extrusion of the fluororesin. Further, a fluororesin can be obtained also by using a chain transfer agent or a polymerization catalyst that can control the end group together with fluorine monomers during production (polymerization reaction). Further, a commercially available product can be used as a fluororesin in the present invention. Further, fluorination can be performed by bringing a syringe that is a formed product formed by melting a fluororesin into contact with a fluorine-containing compound. Further, these treatment methods can be combined.

[0086] That is, the total of nonfluorinated group ends and $-\text{CF}_2\text{H}$ group ends is not necessarily 70 or less per 1×10^6 of carbon atoms at the stage of a fluororesin, pellets, and the like as a raw material and may be 70 or less per 1×10^6 of carbon atoms on the surface in contact with the cells, nucleic acids, or proteins in the barrel of a syringe as a final product. Further, a fluororesin containing one or more $-\text{CF}_3$ end groups does not need to contain one or more $-\text{CF}_3$ end groups at the stage of a fluororesin, pellets, and the like as a raw material and the fluororesin may contain one or more $-\text{CF}_3$ end groups on the surface in contact with the cells, nucleic acids, or proteins in the barrel of a syringe as a final product.

[0087] The fluorine radical source is not specifically limited, but examples thereof can include halogen fluorides such as IF_5 and ClF_3 , F_2 gas, CoF_3 , AgF_2 , UF_6 , OF_2 , N_2F_2 , and CF_3OF . Such F_2 gas may have a concentration of 100% but is mixed with an inert gas to 5 to 50 mass %, preferably diluted to 15 to 30 mass % for use, in view of the safety. Examples of the inert gas can include nitrogen gas, helium gas, and argon gas, and nitrogen gas is preferable in view of the cost-effectiveness.

[0088] The fluorination is preferably performed at a temperature of 20 to 220° C., more preferably 100 to 200° C. The fluorination is preferably performed for 5 to 30 hours, more preferably 10 to 20 hours.

[0089] In the present invention, the barrel may have a laminated structure, or may not have a laminated structure, as long as the effects of the present invention are not impaired. In the case of forming a laminated structure using a plurality of layers made of different materials, the combination of materials is not particularly limited, as long as the shrinkage rates when cooled to a desired temperature are the same or within a close range, and the layers do not

separate during cooling. The same applies to the gasket, the plunger, the cap, the finger grip, and the outer bag, which will be described below.

[0090] In the present invention, the volume of the barrel (volume of the syringe) is not particularly limited, and examples thereof include 0.5 mL or more, 1 mL or more, 2.5 mL or more, 100 mL or less, 70 mL or less, 50 mL or less, 30 mL or less, 20 mL or less, 15 mL or less, 10 mL or less, 5 mL or less, and 2.5 mL or less, that is 0.5 mL to 50 mL, 1 mL to 30 mL, 1 mL to 20 mL, 1 mL to 10 mL, 1 mL to 5 mL, 1 mL to 2.5 mL can be exemplified.

[0091] In the present invention, the inner diameter (A) of the barrel is not particularly limited, and examples thereof include 2 mm or more, 3 mm or more, 4 mm or more, 5 mm or more, 6 mm or more, 7 mm or more, 8 mm or more, 9 mm or more, 10 mm or more, 40 mm or less, 35 mm or less, 30 mm or less, 25 mm or less, 20 mm or less, 18 mm or less, 16 mm or less, 15 mm or less, 14 mm or less, 13 mm or less, 12 mm or less, 11 mm or less, and 10 mm or less, that is 2 mm to 40 mm, 3 mm to 30 mm, 4 mm to 20 mm, 4 mm to 18 mm, 5 mm to 18 mm, 5 mm to 15 mm, 5 mm to 12 mm, 6 mm to 10 mm can be exemplified.

[0092] In the present invention, the thickness of the barrel is not particularly limited, and examples thereof include 0.2 mm or more, 0.3 mm or more, 0.4 mm or more, 0.5 mm or more, 0.6 mm or more, 0.7 mm or more, 0.8 mm or more, 0.9 mm or more, 1.0 mm or more, 1.1 mm or more, 1.2 mm or more, 1.3 mm or more, 1.4 mm or more, 1.5 mm or more, 2.5 mm or less, 2.2 mm or less, 2.0 mm or less, 1.9 mm or less, 1.8 mm or less, 1.7 mm or less, 1.6 mm or less, 1.5 mm or less, 1.4 mm or less, 1.3 mm or less, 1.2 mm or less, 1.1 mm or less, 1.0 mm or less, 0.9 mm or less, 0.8 mm or less, 0.7 mm or less, 0.6 mm or less, and 0.5 mm or less, that is 0.2 mm to 2.5 mm, 0.3 mm to 2.2 mm, 0.4 mm to 2.0 mm, 0.3 mm to 1.9 mm, 0.4 mm to 1.8 mm, 0.5 mm to 1.7 mm, 0.6 mm to 1.6 mm, 0.7 mm to 1.5 mm, 0.4 mm to 1.2 mm, 0.5 mm to 1.1 mm, 0.6 mm to 1.0 mm, 0.7 mm to 0.9 mm can be exemplified.

[0093] The resin forming the barrel preferably has a transparency to the extent that the contents such as cells stored in the barrel are easily visually recognized from the outside of the barrel, even in the case where the resin has a certain thickness. From this viewpoint, FEP or PFA is preferable as the fluororesin. Here, examples of the transparency include a transmittance at a wavelength of 450 nm of 15% or more, 25% or more, 35% or more, 45% or more, and 55% or more.

[0094] The barrel that is a member particularly susceptible to the impact from the outside among the constituents of the syringe is required to have a particularly high impact resistance. In particular, the syringe of the present invention that is assumed to be used for cryopreservation of cells or the like is required to have a sufficient impact resistance during preservation not only at room temperature but also at a very low temperature that is suitable for cryopreservation of cells or the like. Examples of the absorption energy as the impact resistance of the resin forming the barrel in the present invention in a tensile impact test under liquid nitrogen cooling conditions include 0.23 J or more, 0.24 J or more, 0.25 J or more, 0.26 J or more, 0.27 J or more, 0.28 J or more, 0.29 J or more, 0.30 J or more, 0.31 J or more, 0.32 J or more, 0.33 J or more, 0.34 J or more, or 0.35 J or more, preferably 0.25 J or more, more preferably 0.30 J or more. From this viewpoint, the fluororesin is preferably PTFE,

FEP, or PFA. Here, a larger absorbed energy (J) means a higher impact resistance. Examples of the method for conducting the tensile impact test under liquid nitrogen cooling conditions include a method of sufficiently cooling a test piece, crosshead, and fixing jig with a liquid nitrogen (-196°C .) and measuring the absorbed energy using a commercially available tensile impact test.

[0095] In the present invention, the method for producing the barrel is not particularly limited, and it can be produced, for example, by general injection molding. The same applies to the gasket, the plunger, the cap, and the finger grip, which will be described below.

<Gasket>

[0096] In the present invention, the gasket is formed from a fluororesin or a polyethylene (PE) resin. Examples of the fluororesin include polytetrafluoroethylene (PTFE), tetrafluoroethylene (TFE)-ethylene copolymer (ETFE), tetrafluoroethylene (TFE)-hexafluoropropylene (HFP)-based copolymer (FEP), and TFE-perfluoroalkyl vinyl ether (PAVE)-based copolymer (PFA), like the fluororesin forming the barrel. Further, since the gasket is in contact with the contents such as cells, examples of the fluororesin forming the gasket suitably include fluororesins having a total number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends in the fluororesin of 70 or less per 1×10^6 of carbon atoms or fluororesins containing one or more $-\text{CF}_3$ end groups, for suppressing the adsorption of cells, a nucleic acid, or a protein stored in the barrel onto the inner surface of the barrel or for suppressing the reduction in cell viability, like the fluororesin forming the barrel.

[0097] Examples of the polyethylene resin forming the gasket include low density polyethylene (LDPE) resin, medium density polyethylene (MDPE) resin, high density polyethylene (HDPE) resin, ultrahigh density polyethylene resin, and ultrahigh molecular weight polyethylene (UHPE) resin.

[0098] In the present invention, PTFE, FEP, PFA, LDPE, MDPE, HDPE, or UHPE is preferable as the fluororesin or polyethylene resin forming the gasket, for a high impact resistance at a very low temperature. Further, the resin forming the gasket may have a transparency so that the contents are easily visually recognized even when the resin has a certain thickness, like the resin forming the barrel. From this viewpoint, FEP, PFA, LDPE, MDPE, or HDPE is more preferable, and FEP or PFA is particularly preferable.

[0099] In the present invention, since the shrinkage rate (X) when the resin forming the barrel is cooled from room temperature to a certain low temperature and the shrinkage rate (Y) when the resin forming the gasket is cooled from room temperature to the low temperature are the same or in a close range by combining the gasket formed from a fluororesin or polyethylene resin with the barrel formed from a fluororesin, the integrity of the syringe can be maintained during preservation not only at room temperature but also at a very low temperature. When (Y) is larger than (X), a gap occurs between the barrel and the gasket when cooled to the low temperature, and thus the integrity of the syringe cannot be maintained theoretically, unless $1 \leq (X/Y)$. Further, when (X) is excessively large with respect to (Y), there is a fear that the gasket or barrel may be broken because they cannot withstand the shrinkage pressure of the barrel when cooled to the low temperature, or the gasket may be disengaged from the barrel. Therefore, in particular

when the ratio (X/Y) of (X) to (Y) is focused on, examples thereof include $1 \leq (X/Y)$, $1 \leq (X/Y) < 1.5$, $1 \leq (X/Y) < 1.45$, $1 \leq (X/Y) < 1.4$, $1 \leq (X/Y) < 1.35$, $1 \leq (X/Y) < 1.3$, $1 \leq (X/Y) < 1.25$, $1 \leq (X/Y) < 1.2$, $1 \leq (X/Y) < 1.15$, $1 \leq (X/Y) < 1.1$, and $1 \leq (X/Y) < 1.05$, most preferably $(X/Y) = 1$.

[0100] Further, the combination of a gasket formed from a fluororesin or a polyethylene resin with a barrel formed from a fluororesin in the present invention allows a syringe with good slidability of the gasket in the barrel in which the integrity of the syringe can be maintained at room temperature and even at a very low temperature to be produced. In such a view, in addition to the combination of the materials of the barrel and the gasket, the ratio (B/A) of the diameter (B) of the gasket to the inner diameter (A) of the barrel can be further focused on. When the ratio (B/A) is excessively large, the resistance of the gasket when slid in the barrel increases, and the slidability deteriorates. Meanwhile, when the ratio (B/A) is excessively small, a gap occurs between the barrel and the gasket, and in particular, the integrity of the syringe cannot be maintained theoretically, unless $1 \leq (B/A)$. Therefore, examples of the ratio (B/A) of (B) to (A) in the syringe of the present invention include $1 \leq (B/A)$, $1 \leq (B/A)$, $1.002 \leq (B/A)$, $1.005 \leq (B/A)$, $1.006 \leq (B/A)$, $1.010 \leq (B/A)$, $1.015 \leq (B/A)$, and $1.020 \leq (B/A)$, for example, $(B/A) < 1.100$, $(B/A) < 1.090$, $(B/A) < 1.080$, $(B/A) < 1.070$, $(B/A) < 1.060$, $(B/A) < 1.050$, $(B/A) < 1.040$, $(B/A) < 1.030$, and $(B/A) < 1.025$, that is, $1 \leq (B/A) < 1.100$, $1 \leq (B/A) < 1.090$, $1 \leq (B/A) < 1.080$, $1 \leq (B/A) < 1.070$, $1 \leq (B/A) < 1.060$, $1 \leq (B/A) < 1.050$, $1 \leq (B/A) < 1.040$, $1 \leq (B/A) < 1.030$, $1 \leq (B/A) < 1.025$, $1.002 \leq (B/A) < 1.050$, $1.005 \leq (B/A) < 1.030$, $1.006 \leq (B/A) < 1.050$, $1.006 \leq (B/A) < 1.030$, preferably $1 \leq (B/A) < 1.050$, and more preferably $1 \leq (B/A) < 1.030$.

[0101] In the present invention, the slidability of the gasket when slid in the barrel can be evaluated by a known method. For example, it can be evaluated by measuring the maximum load (N) at a moving distance of 40 mm at a moving speed of 100 mm/minute using a tensile tester. The slidability that can be evaluated in this way is preferably a maximum load of 10 N or less, more preferably 8 N or less, further preferably 7 N or less, particularly preferably 6 N or less, in which case, the slidability can be determined to be good. That is, a syringe with the maximum load (N) when the gasket is slid in the barrel falling within such a range can be provided as a syringe having good slidability and thus excellent usability.

[0102] As mentioned above, the present invention can maintain the integrity of the syringe during at room temperature and even at a very low temperature by combining a gasket formed from a fluororesin or a polyethylene resin with a barrel formed from a fluororesin. Therefore, the syringe of the present invention can be an aspect free from special members for sealing the opening of the barrel such as caps or seals other than the gasket in the barrel. That is, the syringe of the present invention can be an aspect in which the opening of the barrel is sealed only with the gasket. That is, the syringe of the present invention can be provided as a syringe with high usability that does not require the work to remove such a special cap or seal or the work to connect the gasket to the plunger in use. The term "seal" means to block the exchange of liquids and solids inside and outside the barrel to prevent the intrusion of microorganisms, solid foreign matter, or liquid foreign matter. Further, the syringe of the present invention can be an

aspect of being free or substantially free of (not using or substantially not using) a silicone compound for securing good slidability or good integrity. That is, the syringe of the present invention can be provided as a syringe with high safety in which the contents are not adversely affected by a silicone compound or contaminated by foreign matter. Further, the syringe of the present invention can be an aspect of being free from or substantially free of (not using or substantially not using) rubber-based materials for securing good slidability or good integrity, e.g., elastomers such as thermoplastic elastomers, natural rubber, or synthetic rubber. That is, the syringe of the present invention can be provided as a syringe with high durability that does not become brittle even at a very low temperature. Here, the silicone compound is not particularly limited, as long as it is a compound having a siloxane bond, and may be in any state of a liquid such as silicone oil, a solid such as silicone resin, or a semi-solid mixture thereof. Further, the natural rubber or synthetic rubber is not particularly limited, and examples thereof include isoprene rubber, polyisoprene/styrene butadiene rubber, butadiene rubber, butyl rubber, and silicone rubber. Further, “substantially free of” or “substantially not using” means that it is allowed to comprise or use if it is an embodiment that does not negatively affect the contents of the syringe (for example, embodiment that does not contact the contents of the syringe), or a minute amount that does not negatively affect (for example, a minute amount in a level that does not cause aggregation, degeneration, or inactivation of the contents). The syringe of the present invention is a syringe in which at least the exchange of liquids and solids inside and outside the barrel is blocked to prevent the intrusion of microorganisms, solid foreign matter, or liquid foreign matter into the barrel but can be an aspect in which the exchange of gases is also completely or almost completely blocked.

[0103] In the present invention, the gasket may be composed of one gasket lip or may be composed of two or more gasket lips. Providing two or more gasket lips can further enhance the integrity of the syringe. The thickness of the gasket in the portion in contact with the inner surface of the barrel (size in the sliding contact direction of the sliding contact portion of the gasket and the inner surface of the barrel) is not particularly limited but there is a concern that as the contact area of the gasket with the inner surface of the barrel increases, the friction increases, and the slidability deteriorates.

[0104] Examples of the thickness (C) of the gasket include 0.1 mm or more, 0.2 mm or more, 0.3 mm or more, 0.4 mm or more, 0.5 mm or more, 0.6 mm or more, 0.7 mm or more, 0.8 mm or more, and 0.9 mm or more and 1.3 mm or less, 1.2 mm or less, 1.1 mm or less, 1.0 mm or less, less than 1.0 mm, 0.9 mm or less, 0.8 mm or less, 0.7 mm or less, 0.6 mm or less, 0.5 mm or less, and 0.4 mm or less, that is, 0.1 mm to 0.9 mm, 0.1 mm to 0.7 mm, 0.1 mm to 0.6 mm, 0.1 mm to 0.5 mm, 0.2 mm to 0.8 mm, 0.2 mm to 0.7 mm, 0.2 mm to 0.6 mm, 0.2 mm to 0.5 mm, 0.3 mm to 1.2 mm, 0.4 mm to 1.1 mm, 0.5 mm to 1.0 mm, 0.3 mm to 0.7 mm, 0.2 mm to 0.6 mm, 0.3 mm to 0.5 mm, and 0.4 mm to 0.6 mm, and it is preferably 0.7 mm or less, and more preferably 0.5 mm or less.

[0105] Further, examples of the ratio (C/B) of a thickness (C) of the gasket to a diameter (B) of the gasket include 0.01 or more, 0.02 or more, 0.03 or more, 0.04 or more, 0.05 or more, 0.06 or more, and 0.07 or more and 0.15 or less, 0.14

or less, 0.13 or less, 0.12 or less, 0.11 or less, 0.10 or less, 0.09 or less, and 0.08 or less, that is, $0.01 \leq (C/B) \leq 0.15$, $0.01 \leq (C/B) \leq 0.10$, $0.01 \leq (C/B) \leq 0.09$, $0.01 \leq (C/B) \leq 0.08$, $0.02 \leq (C/B) \leq 0.14$, $0.03 \leq (C/B) \leq 0.14$, $0.03 \leq (C/B) \leq 0.13$, $0.02 \leq (C/B) \leq 0.12$, $0.02 \leq (C/B) \leq 0.11$, $0.02 \leq (C/B) \leq 0.10$, $0.02 \leq (C/B) \leq 0.09$, $0.02 \leq (C/B) \leq 0.08$, $0.03 \leq (C/B) \leq 0.10$, $0.03 \leq (C/B) \leq 0.09$, and $0.03 \leq (C/B) \leq 0.08$, and it is preferably 0.10 or less, and more preferably 0.08 or less.

[0106] When the gasket is composed of one gasket lip, the thickness of the gasket lip is preferably within such a range, and when the gasket is composed of two or more gasket lips, the total thickness of the two or more gasket lips is preferably within such a range.

[0107] Further, in the present invention, the thickness of the gasket refers to the thickness of the portion in contact with the inner surface of the barrel of the gasket (that is the sliding contact portion) or of the portion that can be substantially in contact with the inner surface of the barrel (that is the portion that can be substantially in sliding contact). For example, in case the shape of the sliding contact portion is a flat surface or approximate flat surface as shown in FIG. 1, it refers to the thickness of the flat surface or approximate flat surface, and in case the shape of the sliding contact portion is a curved surface with an arc-shape or approximate arc-shape, or the like, it refers to the chord length or less of the arc or approximate arc. Here, the chord length of the arc or approximate arc refers to the distance between the end points of the arc or approximate arc. Further, in the present invention, the shape of sliding contact portion, or portions other than the sliding contact portion is not limited as long as it does not impair the effect of the present invention.

[0108] In the present invention, the gasket is required to have a sufficient impact resistance not only at room temperature but also at a very low temperature that is suitable for cryopreservation of cells or the like, similarly to the barrel, in order to secure the integrity of the syringe. In the present invention, examples of the absorption energy as the impact resistance of the resin forming the gasket in a tensile impact test under liquid nitrogen cooling conditions include 0.23 J or more, 0.24 J or more, 0.25 J or more, 0.26 J or more, 0.27 J or more, 0.28 J or more, 0.29 J or more, 0.30 J or more, 0.31 J or more, 0.32 J or more, 0.33 J or more, 0.34 J or more, and 0.35 J or more, preferably 0.25 J or more, more preferably 0.30 J or more.

[0109] In the syringe of the present invention, the gasket is slidably accommodated in the barrel configured to store a cell, a nucleic acid, or a protein, but the position of the gasket in the barrel is not particularly limited and can be appropriately adjusted depending on the presence or absence or the amount of the contents stored in the barrel. For example, when nothing is stored in the barrel, the gasket may be accommodated adhering closely to the spout side of the barrel, when the amount of the contents in the barrel is small, the gasket may be accommodated on the spout side of the barrel, or when the amount of the contents in the barrel is large, the gasket may be accommodated on the opening side of the barrel.

<Plunger>

[0110] The plunger is connected to the tip of the gasket, the user of the syringe pushes the plunger in the direction from the opening of the barrel toward the spout to move, and the gasket is configured to slide against the inner surface of the barrel. Therefore, the gasket and the plunger may be

integrally formed, or the gasket and the plunger may be integrated by having a groove or a threaded structure for connecting them. The aspect of the plunger is not particularly limited, as long as the function of sliding the gasket is not impaired, as mentioned above but when the gasket is slidably accommodated in the barrel, the plunger is preferably connected to the gasket with a gap from the barrel. At this time, it is more preferable that the plunger is connected to the gasket perpendicularly or substantially perpendicularly. The gap provided between the plunger and the barrel can eliminate or mitigate the resistance when the gasket slides in the barrel due to the contact between the plunger and the barrel, so that the gasket can slide better. The gap may be provided so that the plunger and the barrel are not in contact at all, or may be provided so that they are partially in contact, but it is preferably provided so that they are not in contact at all in view of the slidability, as mentioned above. The plunger is also generally referred to as a push rod.

[0111] In the present invention, the plunger is not in contact with the contents such as cells or involved in the integrity or slidability of the syringe, but in the syringe of the present invention assumed to be used for cryopreservation of cells or the like, the plunger is preferably formed from a material that can withstand preservation at a very low temperature. Further, as mentioned above, the plunger in the present invention can be molded integrally with the gasket. From these points of view, the plunger in the present invention is preferably formed from the same fluororesin or polyethylene resin as the fluororesin or polyethylene resin mentioned above as the examples of the fluororesin or polyethylene resin forming the gasket, particularly, the resin forming the gasket.

<Cap>

[0112] The cap in the present invention is mounted on the spout of the barrel and seals the spout of the barrel. The term "seal" means to block the exchange of liquids and solids inside and outside the barrel to prevent the intrusion of microorganisms, solid foreign matter, or liquid foreign matter. In the present invention, the cap is in contact with the contents such as cells and involved in securing the integrity of the syringe. Therefore, suitable examples of the resin forming the cap include the same fluororesin or polyethylene resin mentioned above as the examples of the fluororesin or polyethylene resin forming the barrel or gasket that is similarly in contact with the contents and involved in securing the integrity of the syringe. Further, in consideration that it is necessary to secure the integrity of the syringe also during cryopreservation of cells or the like at a very low temperature by closely adhering to the barrel, the shrinkage rate (X) when the resin forming the barrel is cooled from room temperature to a certain low temperature and the shrinkage rate (Z) when the resin forming the cap is cooled from room temperature to the low temperature are preferably the same or in a close range. When (X) is larger than (Z), a gap occurs between the barrel and the cap when cooled to the low temperature, and thus the integrity of the syringe cannot be maintained theoretically, unless $1 \leq (Z/X)$. Further, when (Z) is excessively large with respect to (X), there is a fear that the cap or barrel may be broken because they cannot withstand the shrinkage pressure of the barrel when cooled to the low temperature, or the cap may be disengaged from the barrel. Therefore, in particular when the ratio (Z/X)

of (Z) to (X) is focused on, examples thereof include $1 \leq (Z/X)$, $1 \leq (Z/X) < 1.5$, $1 \leq (Z/X) < 1.45$, $1 \leq (Z/X) < 1.4$, $1 \leq (Z/X) < 1.35$, $1 \leq (Z/X) < 1.3$, $1 \leq (Z/X) < 1.25$, $1 \leq (Z/X) < 1.2$, $1 \leq (Z/X) < 1.15$, $1 \leq (Z/X) < 1.1$, and $1 \leq (Z/X) < 1.05$, most preferably, $(Z/X) = 1$.

[0113] The shape of the cap is not particularly limited, as long as it can adhere closely to the spout of the barrel to secure the integrity of the syringe but examples thereof may include an aspect of comprising a substrate and a mantle tube fitted from the circumferential edge of the substrate into the outer circumferential surface of the spout of the barrel, or an aspect of further comprising a projection projecting from the inner center of the substrate and fitted into the inner circumferential surface of the spout.

<Finger Grip>

[0114] The syringe of the present invention may further comprise a finger grip. The stability when using the syringe is enhanced to increase the usability by comprising the finger grip, since the plunger can be pressed with the thumb while fixing the syringe with the index finger and the middle finger on the finger grip when using the syringe. In the present invention, the finger grip is not in contact with the contents such as cells or involved in the integrity or slidability of the syringe, but in the syringe of the present invention assumed to be used for cryopreservation of cells or the like, the finger grip is also preferably formed from a material that can withstand preservation at a very low temperature. From this viewpoint, the finger grip in the present invention is preferably formed from the same fluororesin or polyethylene resin mentioned above as the examples of the fluororesin or polyethylene resin forming the gasket, plunger, or cap. Here, the finger grip may be integrally formed with the barrel to be connected thereto, and in this case, it is particularly preferable that it is formed from the same resin as the resin from which the barrel is formed, or may be detachable.

[0115] In the case where the finger grip is detachable, it is preferable that the finger grip is not attached when the syringe is stored in the later-described outer bag, and the finger grip is attached when the syringe is taken out from the outer bag for use. The finger grip is not attached when the syringe is stored in the outer bag, thereby allowing the outer bag to easily adhere more closely to the syringe, so that vacuum packaging that does not allow a gas phase to exist between the outer bag and the syringe can be enabled. In the state of vacuum packaging that does not allow a gas phase to exist between the outer bag and the syringe, heat is directly conducted to the syringe without passing through the gas phase when the syringe stored in the outer bag is cooled and heated, and thus cooling and heating efficiency are improved, so that the time required for freezing and thawing the contents can be shortened. Therefore, particularly when cells or the like that are sensitive to the temperature change are stored in the syringe, quality deterioration and death of cells or the like due to a decrease in thawing speed can be suppressed.

[0116] Further, in the case where the finger grip is not detachable, it is preferable in the same manner when the outer circumferential surface of the finger grip and the outer circumferential surface of the barrel are partially flush or substantially flush with each other, since the outer bag easily adheres more closely to the syringe. Here, to be partially flush means to have a portion with no step between the outer

circumferential surface of the finger grip and the outer circumferential surface of the barrel. To be substantially flush includes a state in which there is a step that does not hinder the adhesion of the outer bag to the syringe, other than the state of being flush. As an example of the syringe structure in which the outer circumferential surface of the finger grip and the outer circumferential surface of the barrel are partially flush or substantially flush with each other, the following aspect can be mentioned. That is, in the case where the cross section of the barrel is circular, and the finger grip is substantially elliptical, an aspect in which the diameter of the circle and the minor axis of the ellipse are the same or substantially the same can be mentioned as an example. Further, in the case where the cross section of the barrel is circular, and the finger grip is substantially rectangular, an aspect in which the diameter of the circle and the length of the short side of the rectangle is the same or substantially the same can be mentioned as an example.

<Outer Bag>

[0117] The syringe of the present invention may be an aspect of being stored in an outer bag, for improving the sanitary conditions during transportation, preservation, freezing, thawing or the like and facilitating the handling. In the present invention, the outer bag is not in contact with the contents such as cells or involved in the integrity or slidability of the syringe, but in the syringe of the present invention assumed to be used for cryopreservation of cells or the like, the outer bag is also preferably formed from a material that can withstand preservation at a very low temperature. From this viewpoint, the outer bag in the present invention is preferably formed from the same fluororesin or polyethylene resin mentioned above as the examples of the fluororesin or polyethylene resin forming the gasket, plunger, or cap.

<Contents>

[0118] Since the syringe of the present invention can withstand preservation at a very low temperature, as mentioned above, it is a syringe that can be used for cryopreservation of cells, a nucleic acid, or a protein, that is, specified for use in “cryopreservation of cells, a nucleic acid, or a protein”. Here, “cryopreservation of cells, a nucleic acid, or a protein” means to preserve the cells, nucleic acid, and/or protein as the contents of the syringe in a frozen state for a certain period of time. Here, “preservation” is a state where the contents are not used, and for example, it can be a state where it is allowed to stand in a deep freezer, etc. or a state during transportation, etc. The preservation period is not particularly limited but may be, for example, several minutes to several years. Cells, a nucleic acid, or a protein can be preserved stably over a long period of time by being frozen. Cells, a nucleic acid, or a protein may be stored in a syringe alone or may be stored in a syringe in any state of liquid, gas, solid, or mixture thereof containing them.

[0119] In addition, examples of the temperature for cryopreservation of cells, a nucleic acid, or a protein include -20°C . or less, -30°C . or less, -40°C . or less, -50°C . or less, -60°C . or less, -70°C . or less, -80°C . or less, -90°C . or less, -100°C . or less, -110°C . or less, -120°C . or less, -130°C . or less, -140°C . or less, -150°C . or less, -160°C . or less, -170°C . or less, -180°C . or less, -190°C . or less, and -196°C . or less, and -200°C . or more, -196°C .

or more, -190°C . or more, -180°C . or more, -170°C . or more, -160°C . or more, -150°C . or more, -140°C . or more, -130°C . or more, -120°C . or more, -110°C . or more, -100°C . or more, -90°C . or more, -80°C . or more, -70°C . or more, -60°C . or more, -50°C . or more, -40°C . or more, and -30°C . or more, preferably -80°C . or less, for example, -80°C . to -196°C ., -80°C . to -180°C ., -80°C . to -150°C ., -80°C . to -130°C ., -80°C . to -100°C ., -150°C . to -196°C ., and -150°C . to -180°C .

[0120] The syringe of the present invention can be a syringe that is used at the above-mentioned temperature for cryopreservation, and examples include to be a syringe that is used at -80°C . or less for cryopreservation, a syringe that is used at -150°C . or less for cryopreservation. That is, the syringe of the present invention can be a syringe that is for use in the cryopreservation at the above-mentioned temperature, and examples include to be a syringe that is for use in the cryopreservation at -80°C . or less, a syringe that is for use in the cryopreservation at -150°C . or less. It is the same for the prefilled syringe described later.

[0121] In general, it is said that the lower the cryopreservation temperature, the better the long-term preservation stability, and when it is -196°C . or less, semi-permanent preservation is possible. Cells in particular may be preserved at about -20°C . for a short period of time and then preserved at -80°C . or less for a long period of time. Although the method of freezing cells, nucleic acids, or proteins is not particularly limited, examples thereof include a freezing method using a deep freezer with a cooling capacity of -80°C . or less, and a freezing method using the gas phase (-130°C . or less) or liquid phase (-196°C .) of liquid nitrogen. Further, the syringe of the present invention can be used by cooling to the above temperature to freeze the contents and then heating to thaw the contents. The thawing temperature is not particularly limited but examples thereof include -20°C . or more, -10°C . or more, 0°C . or more, 10°C . or more, 20°C . or more, 30°C . or more, 37°C . or more, 40°C . or less, and room temperature, for example, 0°C . to 40°C ., 20°C . to 40°C ., and 30°C . to 40°C . The thawing method is not particularly limited but examples thereof include a thawing method gently on ice, or a thawing method by setting the desired temperature using a heating device such as a hot water bath.

[0122] In the present invention, cells, a nucleic acid, or a protein stored in the syringe can be directly administered to an administration target without being transferred to another container after cryopreservation and thawing. That is, the syringe of the present invention can be provided as a prefilled syringe storing cells, a nucleic acid, or a protein. Examples of the administration target can include mammals, amphibians, reptiles, birds, and fish, particularly suitably mammals. Examples of the mammals can include humans, pigs, cows, horses, goats, sheep, wild boars, rabbits, mice, rats, hamsters, guinea pigs, monkeys, rhesus monkeys, cynomolgus monkey, marmosets, orangutans, chimpanzees, dogs, and cats, suitably humans.

[0123] In the present invention, examples of the cells include mammalian cells, and the mammalian cells may be buoyant cells or adherent cells. Examples of the buoyant cells can include erythrocytes, (peripheral blood-derived) leukocytes (neutrophils, mononuclear cells [monocytes, lymphocytes], macrophages). Examples of adherent cells include stem cells including pluripotent stem cells such as embryonic stem cells (ES cells), embryonic germ cells (EG

cells), germline stem cells (GS cells), and induced pluripotent stem cells (iPS cells), multipotent stem cells such as mesenchymal stem cells, hematopoietic stem cells, and nervous system stem cells, unipotent stem cells (progenitor cells) such as cardiac progenitor cells, vascular endothelial progenitor cells, neural progenitor cells, adipose progenitor cells, dermal fibroblasts, skeletal muscle myoblasts, osteoblasts, and odontoblasts, mature cells including cardiomyocytes, vascular endothelial cells, nerve cells, adipocytes, dermal fiber cells, skeletal muscle cells, osteocytes, hepatocyte (liver) cells, umbilical vein endothelial cell, dermal microlymphatic endothelial cells, epidermal keratinocytes, bronchial epithelial cells, melanocytes, smooth muscle cells, and dentin cells. Examples of the application of cells include medical uses and testing and research uses, and cells such as stem cells (therapeutic cells) for use in medical treatments such as regenerative medicine are particularly suitable. The cells may be suspended in a cell culture medium, cell preservative solution, or the like.

[0124] In the present invention, the nucleic acids are not particularly limited, as long as they are natural, modified, or synthetic nucleotides, and may be DNA, RNA, or chimeras thereof. Generally, a nucleic acid is a gene or an analogue thereof, more specifically the gene is, for example, a tumor suppressor gene, and more specifically a polynucleotide or an oligonucleotide as a gene analogue. Here, nucleotides include oligonucleotides modified to make them less susceptible to degradation in vivo such as an oligonucleotide having a thiophosphodiester bond (S-oligo) in which the oxygen atom of the phosphodiester bond is replaced with a sulfur atom, or an oligonucleotide in which the phosphodiester bond is replaced with an uncharged methylphosphate group. Further specifically, examples of the oligonucleotide include decoys (decoy molecules), antisense, ribozymes, aptamers, or siRNA or mRNA. Examples of the application of nucleic acids include medical uses and testing and research uses, and these nucleic acids as pharmaceuticals are particularly preferable. Here, pharmaceuticals include RNA vaccines. Nucleic acids may be in a lyophilized (freeze-dried) state, or may be in a state of being suspended or dissolved in a nucleic acid preservation solution or the like.

[0125] In the present invention, proteins refer to single or multiple polymer compounds formed by linking (polymerizing) a large number of L-amino acids in a chain via amide bonds (also called peptide bonds) and are not limited by the number of constituent amino acids. Accordingly, the proteins of the present invention also include so-called peptides. Further, the proteins of the present invention also include glycoproteins in which sugars and proteins are bound and lipoproteins in which lipids and proteins are bound. Examples of the proteins include cell growth factors such as albumin, fibrinogen, globulin (α 1-globulin, α 2-globulin, β -globulin, and γ -globulin), erythropoietin, collagen, elastin, keratin, lactoferrin, avidin, cadherin, proteoglycan, mucin, LDL (Low Density Lipoprotein), HDL (High Density Lipoprotein), VLDL (Very Low Density Lipoprotein), insulin, and transferrin, cytokines and growth factors such as activin A, Bone Morphogenetic Proteins 4 (BMP-4), Epidermal Growth Factor (EGF), Stem Cell Factor (SCF), interleukin. Examples of the application of proteins include medical uses and testing and research uses, and these proteins as pharmaceuticals are particularly preferable. The proteins may be

in a lyophilized (freeze-dried) state, or may be in a state of being suspended or dissolved in a protein preservation solution or the like.

[0126] The purpose of use of the syringe of the present invention is not particularly limited, but the syringe of the present invention can be a syringe that is used for medical procedure (regenerative medicine, dosing, vaccination, etc.), manufacture of medicine, research, or the like. Examples of regenerative medicine include, administering therapeutic cells such as stem cells to a patient, going through processes such as collecting, selecting, concentrating, and/or proliferation, according to the purpose. The syringe of the present invention can be cryopreserved one or more times when using, according to the purpose.

[0127] The specific method of use of the syringe of the present invention is not particularly limited, but the syringe of the present invention can be used for example in a method for cryopreserving a cell, a nucleic acid, or a protein, as in the following.

[0128] (1) providing a cell, a nucleic acid, or a protein to be cryopreserved (that should be cryopreserved);

[0129] (2) providing a syringe comprising: a barrel having a spout and an opening and for storing the cell, the nucleic acid, or the protein; a gasket slidably accommodated in the barrel, a plunger connected to the gasket; and a cap that seals the spout of the barrel, wherein the barrel is formed from a fluororesin, and the gasket is formed from a fluororesin or a polyethylene resin;

[0130] (3) pulling to move the gasket by an operation of the plunger to the direction from the spout to the opening of the barrel to store the cell, the nucleic acid, or the protein in the barrel;

[0131] (4) attaching the cap to the spout of the barrel to seal the barrel;

[0132] (5) storing the syringe in the outer bag if necessary, and further close or seal the outer bag if necessary;

[0133] (6) putting the syringe under a desired low temperature (for example, at -80°C . or less, or -150°C . or less) to freeze the cell, the nucleic acid or the protein in the syringe to preserve for a desired period;

[0134] Further, the syringe of the present invention can be used for example in a method for administering a cell, a nucleic acid, or a protein, as in the following.

[0135] (1) providing a cell, a nucleic acid, or a protein to be administered (that should be administered);

[0136] (2) providing a syringe comprising: a barrel having a spout and an opening and for storing the cell, the nucleic acid, or the protein; a gasket slidably accommodated in the barrel, a plunger connected to the gasket; and a cap that seals the spout of the barrel, wherein the barrel is formed from a fluororesin, and the gasket is formed from a fluororesin or a polyethylene resin;

[0137] (3) pulling to move the gasket by an operation of the plunger to the direction from the spout to the opening of the barrel to store the cell, the nucleic acid, or the protein in the barrel;

[0138] (4) attaching the cap to the spout of the barrel to seal the barrel;

[0139] (5) storing the syringe in the outer bag if necessary, and to further close or seal the outer bag if necessary;

- [0140] (6) putting the syringe under a desired low temperature (for example, at -80°C . or less, or -150°C . or less) to freeze the cell, the nucleic acid or the protein in the syringe to preserve for a desired period;
- [0141] (7) putting the syringe under a desired high temperature (for example, at room temperature, 30°C . or more, or 37°C .) to thaw the cell, the nucleic acid, or the protein for a desired period;
- [0142] (8) taking out the syringe from the outer bag in case the syringe is stored in the outer bag;
- [0143] (9) detaching the cap from the spout of the barrel of the syringe, and attaching the injection needle to the spout;
- [0144] (10) administering the cell, the nucleic acid, or the protein from the syringe to a subject in need of administration of the cell, the nucleic acid, or the protein.

<Prefilled Syringe>

[0145] One aspect of the prefilled syringe includes a syringe comprising the barrel, the gasket, the plunger, and, the cap, described above and storing cells, a nucleic acid, or a protein, wherein the cap is configured to be detached from the syringe, and the injection needle is configured to be attached in use.

[0146] The prefilled syringe of the present invention can be a prefilled syringe that is used at each temperature described above for cryopreservation, and examples include to be a prefilled syringe that is used at -80°C . or less for cryopreservation, a prefilled syringe that is used at -150°C . or less for cryopreservation. That is, the prefilled syringe of the present invention can be a prefilled syringe that is for use in the cryopreservation at -80°C . or less, a prefilled syringe that is for use in the cryopreservation at -150°C . or less.

<Kit>

[0147] The syringe or prefilled syringe of the present invention may be provided by being contained in a kit. As kits, examples include a kit comprising the syringe or prefilled syringe of the present invention, and an instruction for use describing the method for cryopreserving or administering cells, a nucleic acid, and a protein using the syringe or prefilled syringe of the present invention (i.e. for performing the method). The kit of the present invention can be a kit that is used for cryopreserving and/or administering cells, a nucleic acid, or a protein. The kit of the present invention may further contain other constituents. Examples of other constituents include a cryopreservation solution for cryopreserving cells, a nucleic acid, or a protein, a dissolution buffer, a carrier, a pH buffer, a stabilizer, an instruction manual, other attached documents, an injection needle, a tube, etc.

Examples

[0148] Hereinafter, the present invention will be described further specifically by way of examples, but the technical scope of the present invention is not limited to these examples.

1. Analytical Evaluation of Fluororesin

[0149] It is known that a fluororesin in which a total number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends in the fluororesin is 70 or less per 1×10^6 of carbon

atoms, and/or containing one or more $-\text{CF}_3$ end groups can suppress adsorption of cells (i.e. can suppress decrease in cell recovery rate), can suppress decrease in cell viability, and further can suppress adsorption of protein (see WO2017/010100 and WO2018/135228). Therefore, resin Q and resin R shown in Table 1 were used for producing syringes. For both resin Q and resin R, in the sample before molding syringes (pellet), the number of non-fluorinated group ends and the number of $-\text{CF}_2\text{H}$ group ends were both 0 per 1×10^6 of carbon atoms. Further, in the sample after molding syringes of resin R, the number of non-fluorinated group ends was 48 to 66 per 1×10^6 of carbon atoms, and the number of $-\text{CF}_2\text{H}$ group ends is 0 per 1×10^6 of carbon atoms as shown in Table 1, that is the total number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends was 70 or less per 1×10^6 of carbon atoms. Here, since similar syringe molding processing as for resin R is performed for resin Q, it is estimated that the change in the number of non-fluorinated group ends and the number of $-\text{CF}_2\text{H}$ group ends after syringe molding has the same tendency. As such, it is inferred that in the sample after molding syringes of resin Q, the total number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends is 70 or less per 1×10^6 of carbon atoms, and the number of $-\text{CF}_2\text{H}$ group ends is 0 per 1×10^6 of carbon atoms. The number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends was measured according to the method described in WO2017/010100 and WO2018/135228.

TABLE 1

Analytical evaluation of fluororesin				
Sample after molding syringe				
Resin name	Material of resin	number of non-fluorinated group ends (per 1×10^6 of carbon atoms)	number of $-\text{CF}_2\text{H}$ group ends (per 1×10^6 of carbon atoms)	Presence or absence of $-\text{CF}_3$ end groups
Resin Q	FEP	(no data)	(no data)	Present
Resin R	PFA	1.0 mL volume syringe: 48 2.5 mL volume syringe: 66	1.0 mL volume syringe: 0 2.5 mL volume syringe: 0	1.0 mL volume syringe: present 2.5 mL volume syringe: present

Non-fluorinated group ends: $-\text{COF}$, $-\text{COOH}$, $-\text{COOH}$ associated with water, $-\text{CH}_2\text{OH}$, $-\text{CONH}_2$, and $-\text{COOCH}_3$

2. Evaluation of Durability During Cooling (1): Tensile Impact Test

[0150] In order to evaluate the impact resistance during cooling of each material, a tensile impact test was conducted.

2-1 Method

2-1-1 Test Sample

[0151] Each resin shown in Table 2 was subjected to the test.

2-1-2 Test Operation

[0152] [1] A test piece obtained by molding each resin into the shape of type 2 of JIS K 7160, a crosshead, and a fixing jig were cooled over about 1 minute in liquid nitrogen.

[0153] [2] The test piece, the crosshead, and the fixing jig cooled in [1] above were attached to a tensile impact tester (fully automatic Charpy impact tester, No. 190, manufactured by YASUDA SEIKI SEISAKUSHO, LTD.).

[0154] [3] Liquid nitrogen was spun around the test piece attached to the tester to cool it again.

[0155] [7] The test was conducted immediately after [3] above. This test was performed three times for the same resin, and the average and standard deviation were obtained. The weights of the crosshead hammer and the hammer were appropriately adjusted according to the test piece and measurement conditions.

2-2 Results

[0156] Table 2 shows the test results. The results indicate that the higher the absorbed energy (J), the higher the impact resistance. It was shown that fluororesin and PE resin have high impact resistance under liquid nitrogen cooling conditions, and among them, UHPE has extremely high impact resistance under liquid nitrogen cooling conditions. It was also shown that among fluororesins, PTFE in particular has high impact resistance under liquid nitrogen cooling conditions. Meanwhile, PP resin was shown to have low impact resistance under liquid nitrogen cooling conditions as compared to the above. As can be seen from the above, it was shown that fluororesin or PE resin are suitable materials for syringes that are assumed to be used under liquid nitrogen cooling conditions.

TABLE 2

Absorption energy		
Resin sample No.	Material of resin	Absorption energy (J)
1	FEP	0.34 ± 0.08
2	PFA	0.32 ± 0.06
3	PTFE	0.78 ± 0.13
4	UHPE	2.99 ± 0.14
5	HDPE	0.40 ± 0.02
6	LDPE	0.37 ± 0.13
7	PP	0.22 ± 0.04

Abbreviations in the table indicate the following resins.

FEP: Resin Q in Table 1

PFA: Resin R in Table 1

PTFE: Manufactured by Flonchemical, Co., Ltd.

UHPE: Newlight, manufactured by Shimonoseki Packing, Co., Ltd.

HDPE: Manufactured by Flonchemical, Co., Ltd.

LDPE: Manufactured by Flonchemical, Co., Ltd.

PP: Manufactured by Flonchemical, Co., Ltd.

3. Evaluation of Durability During Cooling (2) Drop Test

[0157] In order to evaluate the embrittlement resistance of a syringe formed from each material during cooling, a drop test was conducted.

3-1 Method

3-1-1 Test Sample

[0158] The 1.0 mL volume syringes of syringe sample Nos. 7-1 to 7-3 each consisting of a barrel, a gasket, a plunger, a finger grip, and a cap (see Table 3) were subjected to the test. The cap was common to all syringes, and a

combination cap (product number: 4495209 made of PE, manufactured by B. Braun Melsungen AG) was used.

3-1-2 Test Operation

[0159] [1] Four syringes were prepared for each, and each was filled with 1.0 mL of water and capped.

[0160] [2] The syringes of [1] above were put into a deep freezer (−80° C.) and preserved for one day or more.

[0161] [3] The syringes were taken out from the deep freezer and dropped onto an iron plate from a height of 120 cm.

[0162] [4] The syringes dropped in [3] above were observed with the cap detached, to confirm the presence or absence of damage.

3-2 Results

[0163] Table 3 shows the test results. One of the four syringes 7-2 subjected to the test was damaged by cracking at the spout of the barrel. Syringe 7-2 was a syringe with a barrel formed from PP resin. PP resin generally has a glass transition temperature of about 0° C. to −10° C., that is, it is known to be embrittled at a low temperature below it, and this test also shows that PP resin is unsuitable as a material for syringes that are assumed to be cryopreserved at −80° C. or less.

[0164] Meanwhile, syringe 7-1 having a barrel and a gasket formed from PFA resin was not damaged at all even when dropped onto an iron plate from a height of 120 cm and was not damaged when cooled in the gas phase of liquid nitrogen (−150° C. or less) and thereafter hit against the iron plate. This result showed that PFA resin is highly suitable as a material for forming a syringe that is assumed to be cryopreserved at −80° C. or less, further at −150° C. or less.

TABLE 3

The number of damaged syringes			
Syringe sample No.	Barrel material	Gasket material	The number of damaged syringes
7-1	PFA	PFA	0
7-2	PP	Thermoplastic elastomer	1
7-3	PP	PE	0

n = 4. The terms in the table indicate the following resins.

PFA: Resin R in Table 1

7-1: Prototype manufactured by injection molding, Volume: 1.0 mL

7-2: Manufactured by JMS Co., Ltd. (Model: JS-SM01C), Volume: 1.0 mL

7-3: Manufactured by Henke-Sass, Wolf GmbH (Code: S4010-LT), Volume: 1.0 mL

4. Evaluation of Shrinkage Rate During Cooling

[0165] In order to evaluate the shrinkage rate of each material during cooling, the following test was conducted.

4-1 Method

4-1-1 Test Sample

[0166] The resins (or elastomers) shown in Table 4 were subjected to the test.

4-1-2 Test Operation

[0167] [1] A disc-shaped resin piece having a diameter of 5 mm and a thickness of 1 mm was fabricated for each resin.

[0168] [2] The resin piece was left standing at room temperature and then the diameter (a) was measured.

[0169] [3] The resin piece was cooled using a microscope cooling and heating stage at 30° C./minute, followed by standing at -130° C. for 20 minutes, and the diameter (b) was measured.

[0170] [4] The shrinkage rate of the resin during cooling (-130° C.) was determined by the following calculation formula. Calculation formula: Shrinkage rate (%) = $\{(a-b)/a\} \times 100$

4-2 Results

[0171] Table 4 shows the test results. All of the resins were shown to shrink more when cooled at -130° C. than at room temperature, but the magnitude of the shrinkage rate significantly differed among the resins as shown in Table 4.

TABLE 4

Shrinkage rate during cooling at low temperature (-130° C.)		
Resin sample No.	Material of resin	Shrinkage rate (%)
8-1	PP	1.2
8-2	PE	1.6
8-3	Silicone	3.8
8-4	SEBS	0.8
8-5	FEP	2.0
8-6	PFA	1.6

n = 1. The terms in the table indicate the following resins.

SEBS: SEBE (styrene ethylene butylene styrene; thermoplastic elastomer) cut from the gasket of Terumo Syringe, manufactured by Terumo Corporation

FEP: Resin Q in Table 1

PFA: Resin R in Table 1

[0172] On the assumption that the barrel and the gasket of a syringe are formed by combining these resins, Table 5 shows a value calculated as a ratio (X/Y) of a shrinkage rate (X) of a resin forming the barrel during cooling at a low temperature (-130° C.) to a shrinkage rate (Y) of a resin forming the gasket during cooling at a low temperature (-130° C.) in each case. The value of Table 4 was used for calculation.

[0173] Theoretically, a gap occurs between the barrel and the gasket when (Y) is larger than (X), and therefore the integrity of the syringe cannot be maintained. Therefore, in order to form a syringe that is assumed to be cryopreserved, $1 \leq (X/Y)$ is considered necessary to be satisfied. Further, it is considered that there is a fear that the gasket or the barrel may be damaged when the ratio (X/Y) is excessively large, since it cannot withstand the shrinkage pressure of the barrel during cooling at low temperature, or the gasket may be disengaged from the barrel. However, as shown in Table 5, the ratio (X/Y) was expected to range very widely from 0.32 to 2.50 depending on the combination of resins. As can be seen from the above, it was suggested that it was important to consider that the ratio of the shrinkage rate of each resin during cooling at low temperature satisfies $1 \leq (X/Y)$, and the ratio (X/Y) falls within the range comparatively close to 1 in the combination of the materials as resins forming the barrel and the gasket of a syringe that is assumed to be preserved at very low temperatures.

TABLE 5

Ratio (X/Y) of shrinkage rate (X) of resin forming barrel during cooling at low temperature (-130° C.) to shrinkage rate (Y) of resin forming gasket during cooling at low temperature (-130° C.)							
		Material of resin forming gasket					
		PP	PE	Silicone	SEBS	FEP	PFA
Material of resin forming barrel	PP	1.00	0.75	0.32	1.50	0.60	0.75
	PE	1.33	1.00	0.42	2.00	0.80	1.00
	FEP	1.67	1.25	0.53	2.50	1.00	1.25
	PFA	1.33	1.00	0.42	2.00	0.80	1.00

*n = 1. Each value was determined from value shown in Table 4. The terms in the table indicate the following resins.

SEBS: SEBE cut from the gasket of Terumo Syringe, manufactured by Terumo Corporation

FEP: Resin Q in Table 1

PFA: Resin R in Table 1

5. Evaluation of Slidability of Syringe, and Evaluation of Integrity During Cooling and Heating (1)

[0174] In order to determine whether or not a syringe formed from each material can withstand a practical use for cryopreservation at -80° C. by evaluating the slidability of the syringe, and the integrity during cooling and heating, the following tests for slidability and integrity were conducted.

5-1 Method

5-1-1 Test Sample

[0175] The syringes of sample Nos. 9-1-1 to 9-6-4 shown in Table 6 were produced by injection molding and subjected to the tests. These were 1.0 mL volume syringes each consisting of a barrel, a gasket composed of one gasket lip, a plunger, and a finger grip. The gasket had a discotic shape with uniform thickness, and the shape of the sliding contact portion with the inner surface of the barrel was a flat surface extending in a sliding contact direction. Further, as comparative controls, commercially available syringes (syringe sample Nos. 9-7 to 9-8) were also subjected to the tests (see Table 8). All these syringes subjected to the tests had an integrity at room temperature. Further, the cap used in the integrity test was common to all syringes, and a combination cap (product number: 4495209 made of polyethylene, manufactured by B. Braun Melsungen AG) was used. The cap had a shape including a substrate and a mantle tube fitted from a circumferential edge of the substrate to an outer circumferential surface of the spout of the barrel.

5-1-2 Test operation: Slidability test

[0176] [1] Using a tensile tester (TENSILON, RTG-1225, manufactured by A&D Company, Limited), the maximum load (N) at a moving distance of 40 mm at a moving speed of 100 mm/minute was measured. The measurement was performed 3 times with respect to the same syringe, and the average was obtained.

[0177] [2] The slidability of the syringe was determined to be good (Good) when the maximum load was 8 N or less and poor (Poor) when it was greater than 8 N.

5-1-3 Test Operation: Integrity Test

[0178] [1] A yellow pigment was dissolved in 99.5% ethanol to fabricate a colored ethanol (freezing point: about -120° C.). The colored ethanol was filled into a PE bag.

- [0179] [2] The gasket was pulled to the display volume of the syringe, the cap was attached without anything stored in the barrel, the syringe was immersed in the colored ethanol in the PE bag, and the PE bag was closed.
- [0180] [3] The PE bag was put into a deep freezer ($-80^{\circ}\text{C}.$) and preserved for 3 to 7 days.
- [0181] [4] The PE bag was taken out from the deep freezer and heated in a hot water bath at $37^{\circ}\text{C}.$ for 5 minutes or more.
- [0182] [5] The syringe was taken out from the PE bag and lightly washed with water, and then the water was wiped off.
- [0183] [6] The presence or absence of the intrusion of the colored ethanol into the barrel of the syringe was visually observed. For each syringe, when the intrusion of the colored ethanol was confirmed, it was determined to have no integrity (Poor), and when the intrusion of the colored ethanol was not confirmed, it was determined to have integrity (Good).

5-2 Results

[0184] Table 6 shows the test results. In the combination of the barrel made of PFA material and the gasket made of PFA, PTFE, UHPE, HDPE, or LDPE material, a syringe with good slidability in which the integrity can be maintained not only at room temperature, but also during any of cooling from room temperature to a very low temperature such as $-80^{\circ}\text{C}.$, preservation at the temperature, and heating from the temperature to $37^{\circ}\text{C}.$ could be obtained.

[0185] Further, the average of the maximum load (N) of each gasket material was calculated from the data shown in Table 6 (see Table 7). As a result, the average of the maximum load (N) was smaller when a gasket made of a PTFE, UHPE, HDPE, or LDPE material was combined with a barrel made of a PFA material than when a gasket made of

a PFA material was combined. This fact suggested that it is more preferable to combine a gasket made of PTFE, UHPE, HDPE, or LDPE material that is different from a barrel made of PFA material than a combination of a gasket made of the same PFA material with the barrel, particularly in view of slidability.

[0186] In the preliminary consideration, when inserting a gasket made of HDPE with a diameter (B) of about 7 mm to a barrel made of PFA with an inner diameter (A) of 6.37 mm ($B/A \approx 1.100$), it was felt that the slidability was not good. Therefore, in the present test, a gasket having a smaller diameter (B) than the above was produced, and subjected to the test.

[0187] Further, in the syringe shown in Table 6, the thickness of the barrel of the currently designed syringe was in the range of 0.72 to 0.94 mm, and the slidability was not significantly affected by the thickness of the barrel in such a range.

[0188] Meanwhile, it was shown that commercially available syringes shown in Table 8 could not be used for cryopreservation since the integrity is lost during any of cooling from room temperature to $-80^{\circ}\text{C}.$, preservation at the temperature, or heating from the temperature to $37^{\circ}\text{C}.$

[0189] It was shown from the above that a syringe that can be used for cryopreservation with good slidability in which the integrity can be maintained not only at room temperature but also during cold preservation at a very low temperature of such as $-80^{\circ}\text{C}.$ and heating, and the usability and the safety are excellent can be provided by forming by a barrel from a fluororesin typified by PFA and forming a gasket from a fluororesin or PE resin typified by PFA, PTFE, UHPE, HDPE, or LDPE in a syringe formed by a barrel, a gasket, a plunger, and a cap without providing a special structure such as a cap or a seal at the barrel opening or without using a silicone compound or a rubber-based material.

TABLE 6

Evaluation of slidability and evaluation of integrity during cooling and heating										
Syringe sample	Barrel		Gasket			Slidability				
	Material	Inner diameter (A) (mm)	Material	Diameter (B) (mm)	Thickness (C) (mm)	Ratio		Maximum load (N)	Evaluation	Integrity Evaluation
						B/A	C/B			
9-1-1	PFA	6.37	PFA	6.40	1.0	1.005	0.156	18.4	Poor	Good
9-1-2	PFA	6.37	PFA	6.37	1.0	1.000	0.157	18.2	Poor	Good
9-1-3	PFA	6.37	PFA	6.38	1.0	1.002	0.157	18.5	Poor	Good
9-1-4	PFA	6.37	PFA	6.38	1.0	1.002	0.157	20.9	Poor	Good
9-2-1	PFA	6.38	PFA	6.42	0.5	1.006	0.078	6.7	Good	Good
9-2-2	PFA	6.37	PFA	6.43	0.5	1.009	0.078	7.6	Good	Good
9-3-1	PFA	6.37	PTFE	6.43	0.5	1.009	0.078	6.1	Good	Good
9-3-2	PFA	6.37	PTFE	6.42	0.5	1.008	0.078	6.2	Good	Good
9-3-3	PFA	6.37	PTFE	6.42	0.5	1.008	0.078	5.7	Good	Good
9-3-4	PFA	6.37	PTFE	6.42	0.5	1.008	0.078	4.6	Good	Good
9-4-1	PFA	6.37	UHPE	6.45	0.5	1.013	0.078	5.3	Good	Good
9-4-2	PFA	6.37	UHPE	6.44	0.5	1.011	0.078	6.1	Good	Good
9-4-3	PFA	6.37	UHPE	6.52	0.5	1.024	0.077	5.9	Good	Good
9-4-4	PFA	6.37	UHPE	6.47	0.5	1.016	0.077	4.2	Good	Good
9-4-5	PFA	6.37	UHPE	6.44	0.5	1.011	0.078	5.7	Good	Good
9-5-1	PFA	6.37	HDPE	6.49	0.5	1.019	0.077	6.0	Good	Good
9-5-2	PFA	6.37	HDPE	6.51	0.5	1.022	0.077	6.9	Good	Good
9-5-3	PFA	6.37	HDPE	6.50	0.5	1.020	0.077	6.7	Good	Good
9-5-4	PFA	6.37	HDPE	6.50	0.5	1.020	0.077	5.9	Good	Good
9-5-5	PFA	6.37	HDPE	6.49	0.5	1.019	0.077	6.4	Good	Good
9-6-1	PFA	6.37	LDPE	6.40	0.5	1.005	0.078	3.4	Good	Good

TABLE 6-continued

Evaluation of slidability and evaluation of integrity during cooling and heating										
Syringe sample	Barrel		Gasket		Slidability			Integrity		
	Material	diameter (A) (mm)	Material	Diameter (B) (mm)	Thickness (C) (mm)	Ratio B/A	C/B	Maximum load (N)	Evaluation	Evaluation
9-6-2	PFA	6.37	LDPE	6.38	0.5	1.002	0.078	3.7	Good	Good
9-6-3	PFA	6.37	LDPE	6.42	0.5	1.008	0.078	3.9	Good	Good
9-6-4	PFA	6.37	LDPE	6.47	0.5	1.016	0.077	4.7	Good	Good

Preservation period in deep freezer was 3 days in the evaluation of integrity.

The terms in the table indicate the following resins.

PFA: Resin R in Table 1

UHPE: Newlight, manufactured by Shimomoseki Packing, Co., Ltd.

HDPE: Manufactured by Flonchemical, Co., Ltd.

LDPE: Manufactured by Flonchemical, Co., Ltd.

TABLE 7

Average of maximum load (N) for each gasket material when evaluating slidability			
Syringe sample No.	Barrel material	Gasket material	Average of maximum load (N)
9-2-1 to 2(n = 2)	PFA	PFA	7.2
9-3-1 to 4(n = 4)	PFA	PTFE	5.7
9-4-1 to 5(n = 5)	PFA	UHPE	5.4
9-5-1 to 5(n = 5)	PFA	HDPE	6.4
9-6-1 to 4(n = 4)	PFA	LDPE	3.9

It was calculated based on data of syringe having 0.5 mm thick gasket shown in Table 6.

TABLE 8

Evaluation of integrity during cooling and heating of commercially available syringe			
Syringe Sample No.	Barrel material	Gasket material	Determination of integrity
9-7	PP	Thermoplastic elastomer	Poor
9-8	PP	Thermoplastic elastomer	Poor

n = 4. Storage period in deep freezer was 7 days. The numbers in the table indicate the following syringes.

9-7: Manufactured by Terumo Corporation (Code No.: SS-02LZ), Volume: 2.5 mL, Thermoplastic elastomer was SEBS, containing silicone oil as lubricant.

9-8: Manufactured by JMS Co., Ltd. (Model: JS-SM01C), Volume: 1.0 mL

6. Studies on Shape of Finger Grip

[0190] In order to evaluate how the shape of the finger grip of the syringe affects the syringe when preserved while being stored in the outer bag, the following test was conducted.

6-1 Method

6-1-1 Test Sample

[0191] A commercially available syringe (manufactured by NIPRO CORPORATION, volume: 2.5 mL) was used. The syringe had a general shape consisting of a barrel, a gasket, a plunger, and a finger grip, wherein the finger grip had a substantially elliptical shape, but there was no portion

without steps between the outer circumferential surface of the finger grip and the outer circumferential surface of the barrel (that is, there was no portion flush with each other). Syringes in which finger grips having such a shape were processed into a substantially elliptical shape so that there were two portions without steps between the outer circumferential surface of the finger grip and the outer circumferential surface of the barrel (that is, there were two portions flush with each other) were referred to as “processed group”. Meanwhile, syringes that were not subjected to such processing were referred to as “unprocessed group”.

6-1-2 Test Operation

[0192] [1] Three syringes from each of the processed group and the unprocessed group (syringe sample Nos. 10-1-1 to 10-2-3) were each filled with 2.5 mL of an aqueous solution (aqueous solution containing 10% DMSO and 0.81% NaCl) imitating a cell cryopreservation solution and capped (combination cap, product number: 4495209, manufactured by B. Braun Melsungen AG).

[0193] [2] Each syringe was packaged one by one with an outer bag made of PE, closed with the air removed from the outer bag, and stored in a deep freezer (−80° C.), to completely freeze the aqueous solution in the syringe.

[0194] [3] Each syringe was taken out from the deep freezer and immersed in a hot water bath at 37° C. without opening the outer bag, to start thawing. 2 minutes, 4 minutes, and 6 minutes after thawing was started, the syringe was visually observed, and it was determined to be in any state in which the aqueous solution in the syringe was completely thawed (Good) or not completely thawed (Poor). At the time when the aqueous solution in the syringe completely thawed the observation of the syringe was ended.

6-2 Results

[0195] Table 9 shows the test results. As compared with that in the syringes of the unprocessed group, the aqueous solution in the syringes of the processed group could be completely thawed in a comparatively short time. This is thought to be because the outer circumferential surface of the finger grip and the outer circumferential surface of the

barrel were partially flush with each other in the syringes of the processed group, and thus the outer bag easily adheres closely to the syringe, that is, the gas phase existing between the outer bag and the syringe could be reduced than in the unprocessed group, thereby allowing the heat to be conducted directly to the syringe without passing through the gas phase during thawing and improving the heating efficiency. In this case, it is considered that not only the heating efficiency but also the cooling efficiency would be improved. Therefore, the aforementioned results show that, in syringes that are assumed to be cryopreserved and thawed, the outer circumferential surface of the finger grip and the outer circumferential surface of the barrel are preferably partially flush with each other.

TABLE 9

Evaluation of thawing time required				
Syringe		Elapsed time after starting thawing		
	sample No.	2 minutes	4 minutes	6 minutes
Processed group	10-1-1	Poor	Good	—
	10-1-2	Poor	Poor	Good
	10-1-3	Poor	Poor	Good
Unprocessed group	10-2-1	Poor	Good	—
	10-2-2	Poor	Poor	Poor
	10-2-3	Poor	Poor	Poor

Good: Aqueous solution in syringe was completely thawed.

Poor: Aqueous solution in syringe was not completely thawed.

7. Evaluation of Slidability of Syringe, and Evaluation of Integrity During Cooling and Heating (2)

[0196] In order to determine whether or not the syringe of the present invention can withstand a practical use for cryopreservation at -150°C . by evaluating the slidability of the syringe, and the integrity during cooling and heating, the following tests for slidability and integrity were conducted.

7-1-1 Test Sample

[0197] The barrels and gaskets shown in Table 10 were produced by injection molding and subjected to the tests. These were 2.5 mL volume syringes each consisting of a barrel, a gasket composed of one gasket lip, a plunger, and a finger grip. The gasket had a discotic shape with uniform thickness of 0.31 mm, and the shape of the sliding contact portion with the inner surface of the barrel was a curved surface with an arc-shape with a chord length of 0.31 mm. All these syringes subjected to the tests had an integrity at

room temperature. Further, the cap used in the integrity test was common to all syringes, and a combination cap (made of polyethylene, manufactured by B. Braun Melsungen AG, product number: 4495209) was used. The cap had a shape including a substrate and a mantle tube fitted from a circumferential edge of the substrate to an outer circumferential surface of the spout of the barrel.

7-1-2 Test Operation: Slidability Test

[0198] [1] Using a tensile tester (manufactured by A&D Company, Limited TENSILON, RTG-1225), the maximum load (N) at a moving distance of 40 mm at a moving speed of 100 mm/minute was measured. The measurement was performed once with respect to the same syringe.

[0199] [2] The slidability of the syringe was determined to be good (Good) when the maximum load was 8 N or less and poor (Poor) when it was greater than 8 N.

7-1-3 Test Operation: Integrity Test

[0200] [1] Isopentane (freezing point: about -160°C .) was filled into a PE bag.

[0201] [2] The gasket was pulled to the display volume of the syringe, the cap was attached without anything stored in the barrel, the syringe was immersed in the isopentane in the PE bag, and the PE bag was closed.

[0202] [3] The PE bag was put into a deep freezer (-150°C .) and preserved for 3 or more days.

[0203] [4] The PE bag was taken out from the deep freezer and immersed in water at room temperature for 5 minutes or more.

[0204] [5] The syringe was taken out from the PE bag and lightly washed with water, and then the water was wiped off.

[0205] [6] The presence or absence of the intrusion of the isopentane into the barrel of the syringe was visually observed. For each syringe, when the intrusion of the isopentane was confirmed, it was determined to have no integrity (Poor), and when the intrusion of the isopentane was not confirmed, it was determined to have integrity (Good).

[0206] Table 10 shows the test results. In the combination of the barrel made of PFA material and the gasket made of HDPE, a syringe with good slidability in which the integrity can be maintained not only at room temperature, but also during any of cooling from room temperature to a very low temperature such as -150°C ., preservation at the temperature, and heating from the temperature to room temperature could be obtained.

TABLE 10

Evaluation of slidability and evaluation of integrity during cooling and heating (2)										
Syringe		Barrel		Gasket			Slidability			
sample		Inner diameter		Diameter	Thickness	Ratio	Maximum load (N)	Evaluation	Integrity	
No.	Material	(A) (mm)	Material	(B) (mm)	(C) (mm)	B/A	C/B	load (N)	Evaluation	Evaluation
11-1-1	PFA	9.10	HDPE	9.15	0.31	1.005	0.034	4.9	Good	Good
11-1-2	PFA	9.10	HDPE	9.15	0.31	1.000	0.034	5.0	Good	Good

TABLE 10-continued

Evaluation of slidability and evaluation of integrity during cooling and heating (2)										
Syringe sample	Barrel		Gasket			Slidability			Integrity	
	Inner diameter		Diameter		Thickness	Ratio		Maximum load (N)	Evaluation	Evaluation
	No.	Material	(A) (mm)	Material	(B) (mm)	(C) (mm)	B/A	C/B		
11-1-3	PFA	9.10	HDPE	9.15	0.31	1.002	0.034	6.1	Good	Good
11-1-4	PFA	9.10	HDPE	9.15	0.31	1.002	0.034	7.1	Good	Good

Outer diameter of the barrel: 10.6 mm for all syringes

The terms in the table indicate the following resins.

PFA: Resin R in Table 1

HDPE: Manufactured by Flonchemical, Co., Ltd.

8. Evaluation of Integrity During Cooling and Heating (3)

[0207] In order to determine whether or not the syringe of the present invention can withstand a practical use for cryopreservation at -80°C . by evaluating the integrity during cooling and heating of the syringe when storing liquid in the syringe, the following tests for integrity were conducted.

8-1-1 Test Sample

[0208] The barrels and gaskets shown in Table 11 were produced by injection molding and subjected to the tests. These were 2.5 mL volume syringes each consisting of a barrel, a gasket composed of one gasket lip, a plunger, and a finger grip. The gasket had a discotic shape with uniform thickness of 0.31 mm, and the shape of the sliding contact portion with the inner surface of the barrel was a curved surface with an arc-shape with a chord length of 0.31 mm. All these syringes subjected to the tests had an integrity at room temperature. Further, the cap was common to all syringes, and a cap produced by injection molding (made of PE) was used. The cap had a shape including a substrate and a mantle tube fitted from a circumferential edge of the substrate to an outer circumferential surface of the spout of the barrel.

8-1-2 Test Operation: Integrity Test

[0209] [1] A yellow pigment (Yellow No. 4) was dissolved in 99.5% ethanol to fabricate a colored ethanol (freezing point: about -120°C .). The colored ethanol was filled into a PE bag.

[0210] [2] 2.5 mL of water was filled in the syringe, and adjusted so that the head space becomes within 0.1 to

0.15 mL. The cap was attached, the syringe was immersed in the colored ethanol in the PE bag, and the PE bag was closed.

[0211] [3] The PE bag was put into a deep freezer (-80°C .) and preserved for 3 or more days.

[0212] [4] The PE bag was taken out from the deep freezer and heated in a hot water bath at 37°C . for 5 minutes.

[0213] [5] The syringe was taken out from the PE bag and lightly washed with water, and then the water was wiped off.

[0214] [6] The colored ethanol was diluted using water, and an aqueous solution with a concentration of 1 v/v %, 0.75 v/v %, 0.5 v/v %, 0.25 v/v %, 0.1 v/v % was prepared.

[0215] [7] The wavelength of the spectrophotometer was set to 428 nm, and the absorbance of each solution prepared in [6] was measured to prepare a standard curve. Water was used as control.

[0216] [8] Similarly, the wavelength of the spectrophotometer was set to 428 nm, and the absorbance of the filled water of each syringe used in the test was measured, and the concentration of the colored ethanol was calculated using the standard curve.

[0217] Table 12 shows the test results. The filled water of any of the syringes subjected to the tests showed an equal absorbance as water used as control. From these results, it has been revealed that there was no intrusion of colored ethanol in the syringe. Therefore, it was shown that all syringes had maintained integrity not only at room temperature, but also during any of cooling from room temperature to a very low temperature such as -80°C ., preservation at the temperature, and heating from the temperature to 37°C .

TABLE 11

Evaluation of integrity during cooling and heating of the syringe (3) sample							
Syringe sample No.	Barrel		Gasket			Ratio	
	Inner diameter		Diameter		Thickness	Ratio	
	Material	(A) (mm)	Material	(B) (mm)	(C) (mm)	B/A	C/B
12-1-1	PFA	9.10	HDPE	9.15	0.31	1.005	0.034
12-1-2	PFA	9.10	HDPE	9.15	0.31	1.000	0.034

TABLE 11-continued

Evaluation of integrity during cooling and heating of the syringe (3) sample						
Syringe sample No.	Barrel		Gasket			Ratio
	Material	Inner diameter (A) (mm)	Material	Diameter (B) (mm)	Thickness (C) (mm)	
12-1-3	PFA	9.10	HDPE	9.15	0.31	1.002 0.034
12-1-4	PFA	9.10	HDPE	9.15	0.31	1.002 0.034

Outer diameter of the barrel: 10.6 mm for all syringes

The terms in the table indicate the following resins.

PFA: Resin R in Table 1

HDPE: Manufactured by Flochemical, Co., Ltd.

TABLE 12

Evaluation of integrity during cooling and heating of the syringe (3) results	
Sample name	Absorbance (Abs)
Water (control) 1	0.000
Colored ethanol 1 v/v % aqueous solution	0.067
Colored ethanol 0.75 v/v % aqueous solution	0.051
Colored ethanol 0.5 v/v % aqueous solution	0.034
Colored ethanol 0.25 v/v % aqueous solution	0.018
Colored ethanol 0.1 v/v % aqueous solution	0.006
Filled water of syringe sample No. 12-1-1	-0.003
Filled water of syringe sample No. 12-1-2	0.000
Filled water of syringe sample No. 12-1-3	0.000
Filled water of syringe sample No. 12-1-4	0.000
Water (control) 2	0.000

9. Cell Preservation Test

[0218] In order to determine whether or not the syringe of the present invention can withstand a practical use for cryopreservation of cells at -80°C ., the following tests were conducted.

9-1-1 Test Sample

[0219] The barrels and gaskets shown in Table 13 were produced by injection molding and subjected to the tests. These were 2.5 mL volume syringes each consisting of a barrel, a gasket composed of one gasket lip, a plunger, and a finger grip. The gasket had a discotic shape with uniform thickness of 0.31 mm, and the shape of the sliding contact portion with the inner surface of the barrel was a curved surface with an arc-shape with a chord length of 0.31 mm. All these syringes subjected to the tests had an integrity at room temperature. Further, the cap was common to all syringes, and a cap produced by injection molding (made of PE) was used. The cap had a shape including a substrate and a mantle tube fitted from a circumferential edge of the substrate to an outer circumferential surface of the spout of the barrel.

9-1-2 Test Operation: Cell Culture

[0220] Human adipose derived stem cells (hAD-MSC; manufactured by Lonza Walkersville; Lot: 19TL200176) were seeded and subcultured according to a conventional method. Specifically, by using a hADSC culture kit (ADSC-BulletKit, ADSC-Adipose-Derived Stem Cells Growth Medium BulletKit, manufactured by Lonza Walkersville),

15 mL of hADSC culture was put into a 75 cm² flask, incubated at 37° C. with a 5% CO₂ incubator, and subcultured at 80 to 100% confluence.

9-1-3 Test Operation: Cell Preservation Test

[0221] [1] Human adipose derived stem cells after subculture was suspended in cell preservative solution (STEM-CELLBANKER®GMP grade) to prepare a cell suspension solution (5×10^5 cells/mL).

[0222] [2] 2 mL of the cell suspension solution was filled in the syringe (1×10^6 cells/syringe), and adjusted so that the head space becomes within 0.1 to 0.15 mL. The cap was attached.

[0223] [3] Each syringe was packaged one by one with an PE outer bag, closed with the air removed from the outer bag.

[0224] [4] The PE outer bag was put into a deep freezer (-80°C .) and preserved for 10 or more days.

[0225] [5] The PE outer bag was taken out from the deep freezer and heated in a hot water bath at 37° C. for 1 to 2 minutes.

[0226] [6] The syringe is taken out from the PE outer bag, the cap was detached, and the cell suspension solution was poured from the spout of the barrel by pushing the plunger.

[0227] [7] 20 μL of the poured cell suspension solution was mixed with 20 μL of 0.4% Trypan blue (Trypan Blue Solution, 0.4%; manufactured by Life Technologies).

[0228] [8] The total number of cells, the number of dead cells, and the total cell concentration of the cell suspension solution were measured with a cell counting chamber under a microscope, and the cell viability, the cell recovery rate, and the living cell recovery rate were calculated as follows, respectively.

Cell viability (%): (Total number of cells – number of dead cells)/

total number of cells $\times 100$

Cell recovery rate (%): (total cell concentration after cryopreservation)/

(total cell concentration before cryopreservation) $\times 100$

Living cell recovery rate (%):

Cell viability (%) \times Cell recovery rate (%) / 100

[0229] Table 14 shows the test results. When cryopreserving cells in any of the syringes subjected to the tests, the cell viability, the cell recovery rate and the living cell recovery rate showed a very high level of approximately 100%. It is thought that the error when measuring the cell number caused the cell recovery rate to be over 100% in some of the tests, and it was determined that a number of cells equal to before cryopreservation could be recovered. Further, by visually observing after the test operation [5] of the above 9-1-3, no abnormality such as leakage of cell suspension solution was observed in any of the syringes. Further, when spouting the cell suspension solution from the syringe in the test operation [6] in the above 9-1-3, when pushing the plunger, the gasket smoothly slid, and it was thought that the usability was good. By observing the spouted cell suspension solution with a microscope, there was no abnormality in the cells, and no foreign matter contamination was observed. Therefore, a syringe that while being possible to suppress absorption of the preserved cells to the syringe and death of cells as much as possible (that is, being possible to recover living cells at an approximately maximum rate), in which the integrity can be maintained not only at room temperature, but also during any of cooling from room temperature to a very low temperature such as -80°C ., preservation at the temperature, and heating from the temperature to 37°C ., and with good slidability could be obtained. From these results, it was shown that the syringe of the present invention could be particularly preferably used for cryopreservation of not only cells but also biological materials such as nucleic acid or protein which similarly require freezing at a very low temperature. Further, since the slidability at actual use was good, it can be said that when attaching an injection needle to the syringe of the present invention to directly administer the contents from the syringe to the subject of administration such as human, it is possible to administer without imposing strain to the body of the subject of administration, and the spout amount could be readily adjusted. Therefore, it was revealed that the syringe of the present invention can be preferably used particularly for medical use such as regenerative medicine (for example, administer therapeutic cells to human).

TABLE 13

Cell preservation test sample							
Syringe	Barrel		Gasket			Ratio	
	Inner diameter		Diameter	Thickness			
sample No.	Material	(A) (mm)	Material	(B) (mm)	(C) (mm)	B/A	C/B
13-1-1	PFA	9.10	HDPE	9.15	0.31	1.005	0.034
13-1-2	PFA	9.10	HDPE	9.15	0.31	1.000	0.034
13-1-3	PFA	9.10	HDPE	9.15	0.31	1.002	0.034

Outer diameter of the barrel: 10.6 mm for all syringes

The terms in the table indicate the following resins.

PFA: Resin R in Table 1

HDPE: Manufactured by Flonchemical, Co., Ltd.

TABLE 14

Cell preservation test results				
	Before cryopreservation	After cryopreservation		
		Syringe sample No. 13-1-1	Syringe sample No. 13-1-2	Syringe sample No. 13-1-3
Total cell concentration (cell/mL)	4.97×10^5	4.97×10^5	5.07×10^5	5.05×10^5
Cell viability (%)	98.3	97.3	98.7	98.0
Cell recovery rate (%)	—	100.0	102.0	101.7
Living cell recovery rate (%)	—	97.3	100.7	99.7

Values in the table all represent mean values of n=3.

EXPLANATION OF LETTERS OR NUMERALS

- [0230] 10: Barrel
 [0231] 12: Cylindrical body
 [0232] 14: Spout
 [0233] 15: Outer circumferential surface of spout
 [0234] 16: Inner circumferential surface of spout
 [0235] 17: Opening
 [0236] 18: Finger grip
 [0237] 20: Gasket
 [0238] 22: Gasket lip
 [0239] 30: Plunger
 [0240] 40: Cap
 [0241] 42: Substrate
 [0242] 43: Circumferential edge of substrate
 [0243] 44: Mantle tube
 [0244] 46: Projection
 [0245] 50: Cells
 [0246] 60: Outer bag
 [0247] 100: Syringe

1. A syringe for cryopreservation of a cell, a nucleic acid, or a protein, comprising:
 - a barrel having a spout and an opening, and for storing a cell, a nucleic acid, or a protein;
 - a gasket slidably accommodated in the barrel;
 - a plunger connected to the gasket; and
 - a cap that seals the spout of the barrel,wherein the barrel is formed from a fluoro-resin, and the gasket is formed from a fluoro-resin or a polyethylene resin.
2. The syringe according to claim 1, wherein the barrel is formed from a fluoro-resin selected from the group consisting of a tetrafluoroethylene-hexafluoropropylene-based copolymer and a tetrafluoroethylene-perfluoroalkyl vinyl ether-based copolymer, and the gasket is formed from a fluoro-resin selected from the group consisting of a tetrafluoroethylene-hexafluoropropylene-based copolymer, a tetrafluoroethylene-perfluoroalkyl vinyl ether-based copolymer, and polytetrafluoroethylene, or a polyethylene resin selected from the group consisting of low density polyethylene, medium density polyethylene, high density polyethylene, and ultrahigh molecular weight polyethylene.
3. The syringe according to claim 1, wherein the fluoro-resin is a fluoro-resin having a total number of non-fluorinated group ends and $-\text{CF}_2\text{H}$ group ends in the fluoro-resin of 70 or less per 1×10^6 of carbon atoms.
4. The syringe according to claim 1, wherein the fluoro-resin is a fluoro-resin containing one or more $-\text{CF}_3$ end groups.
5. The syringe according to claim 1, wherein the cap is formed from a fluoro-resin or a polyethylene resin.
6. The syringe according to claim 1, wherein the plunger is formed from a fluoro-resin or a polyethylene resin.

7. The syringe according to claim 1, wherein the syringe is further stored in an outer bag.
8. The syringe according to claim 7, wherein the outer bag is formed from a fluoro-resin or a polyethylene resin.
9. The syringe according to claim 1, wherein the syringe is free of silicone compounds.
10. The syringe according to claim 1, wherein the syringe is free of elastomers, natural rubber, or synthetic rubber.
11. The syringe according to claim 1, wherein the cell is a mammalian cell.
12. The syringe according to claim 1, wherein the syringe is used for use in regenerative medicine.
13. The syringe according to claim 1, wherein the syringe is used at -80°C . or less for cryopreservation.
14. The syringe according to claim 1, wherein the syringe is used at -150°C . or less for cryopreservation.
15. A prefilled syringe for cryopreserving a cell, a nucleic acid, or a protein, wherein the cell, the nucleic acid, or the protein is stored in the syringe according to claim 1.
16. The prefilled syringe according to claim 15, wherein the prefilled syringe is used at -80°C . or less for cryopreservation.
17. The prefilled syringe according to claim 15, wherein the cell, wherein the prefilled syringe is used at -150°C . or less for cryopreservation.
18. A method for cryopreserving a cell, a nucleic acid, or a protein, comprising cryopreserving the cell, the nucleic acid, or the protein in the syringe according to claim 1.
19. The method according to claim 18, wherein the cryopreservation is a cryopreservation at -80°C . or less.
20. The method according to claim 18, wherein the cryopreservation is a cryopreservation at -150°C . or less.

* * * * *