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(54) COMPOSITIONS AND METHODS FOR EPIGENOME EDITING

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(72) Inventors: Charles A. Gersbach, Chapel Hill, NC (US); Isaac Hilton, Houston, TX (US)

(21) Appl. No.: 18/952,981

(22) Filed: Nov. 19, 2024

Related U.S. Application Data

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- (60) Provisional application No. 62/113,569, filed on Feb. 9, 2015.

Publication Classification

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C12N 9/10 (2006.01)
C12N 15/11 (2006.01)
C12N 15/63 (2006.01)
C12N 15/85 (2006.01)

(52) U.S. Cl.

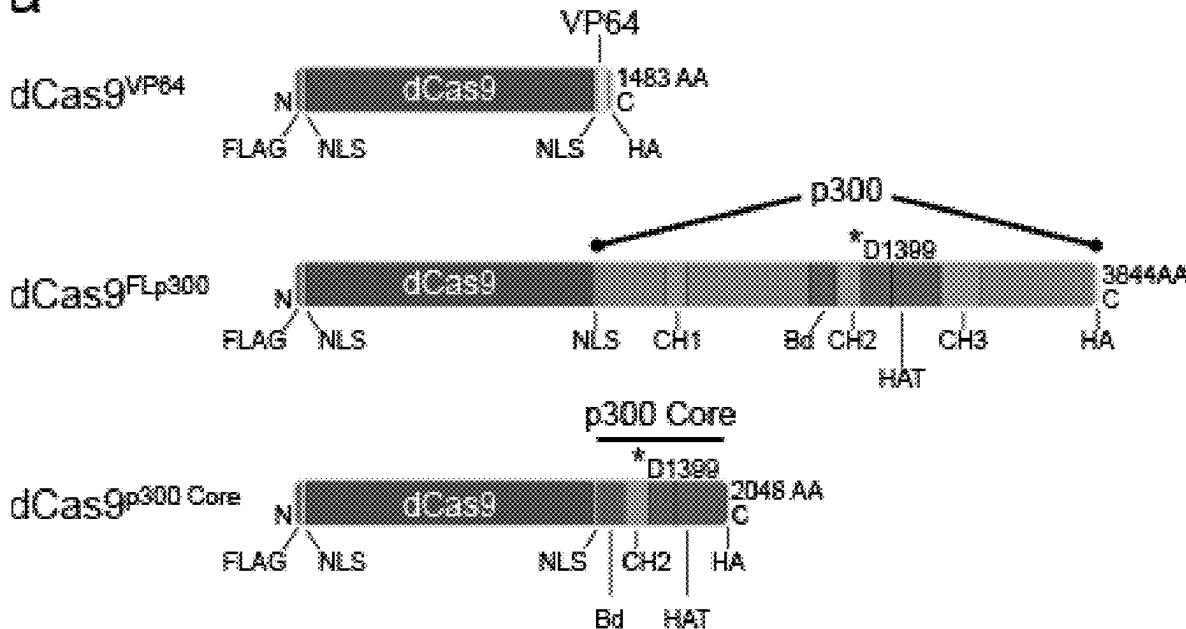
CPC C12N 9/22 (2013.01); C12N 9/1029 (2013.01); C12N 15/11 (2013.01); C12N 15/63 (2013.01); C12N 15/85 (2013.01); C12Y 203/01048 (2013.01); C07K 2319/80 (2013.01); C12N 2310/20 (2017.05)

(57) ABSTRACT

Disclosed herein are CRISPR/Cas9-based gene activation systems that include a fusion protein of a Cas9 protein and a protein having histone acetyltransferase activity, and methods of using said systems.

Specification includes a Sequence Listing.

a



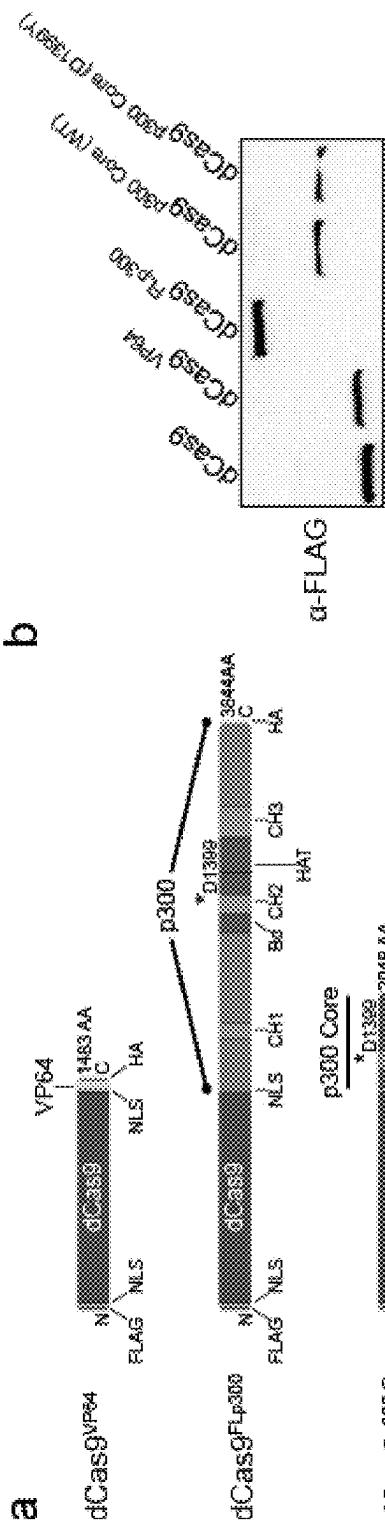
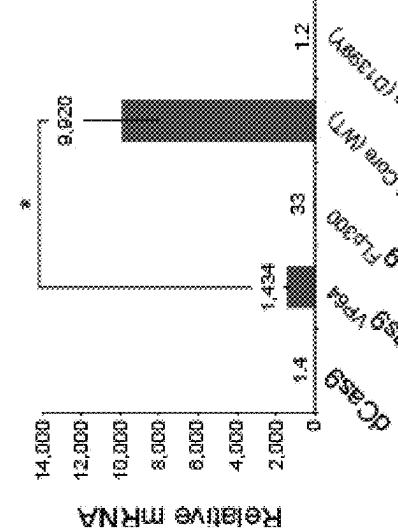


FIG. 1A

L1RN Promoter



MYOD Promoter

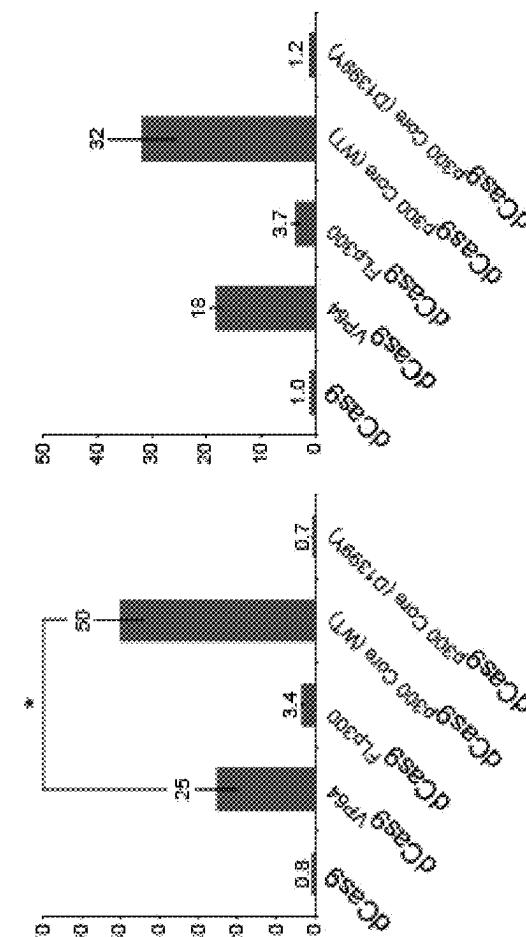


FIG. 1B

OCT4 Promoter

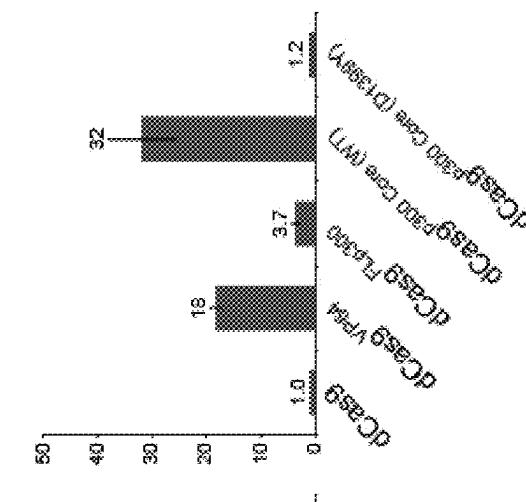
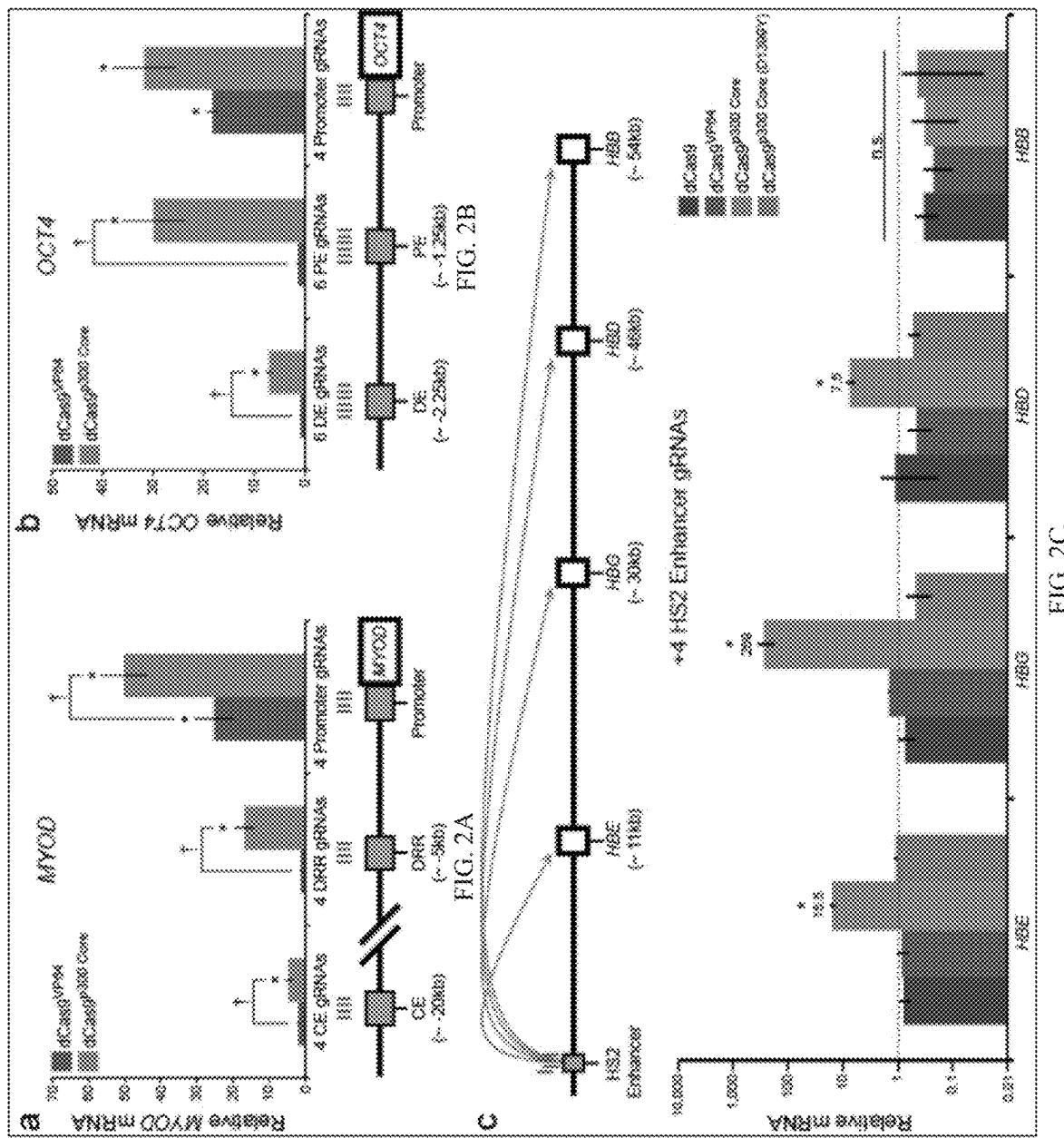


FIG. 1C



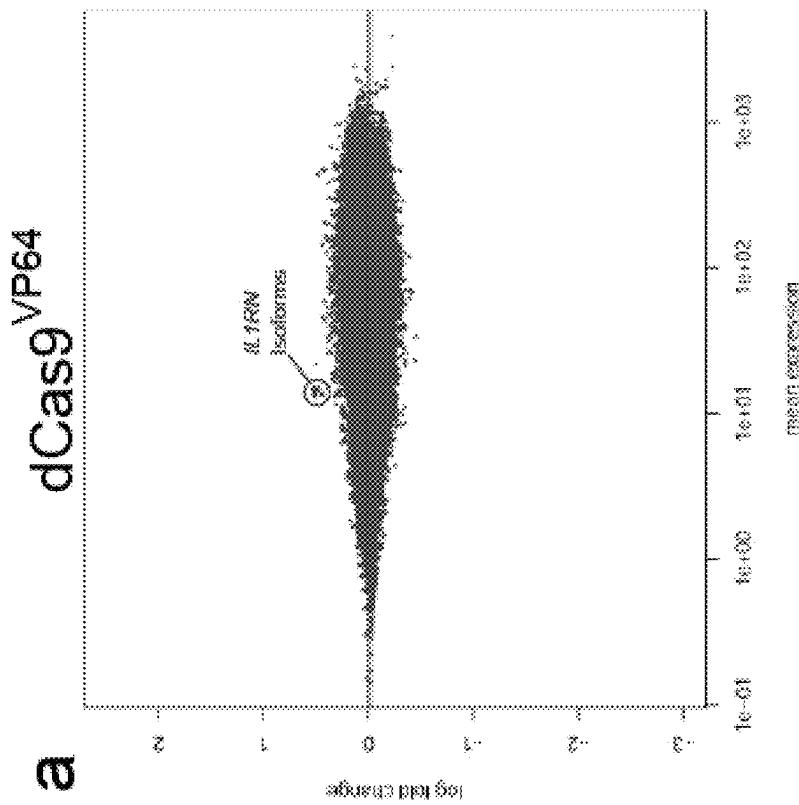


FIG. 3A

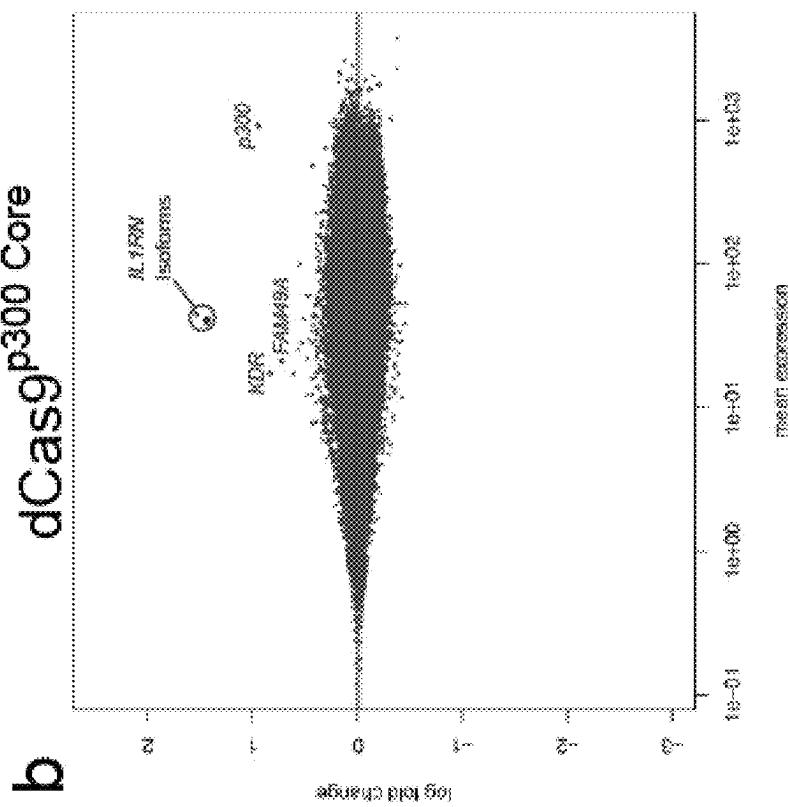


FIG. 3B

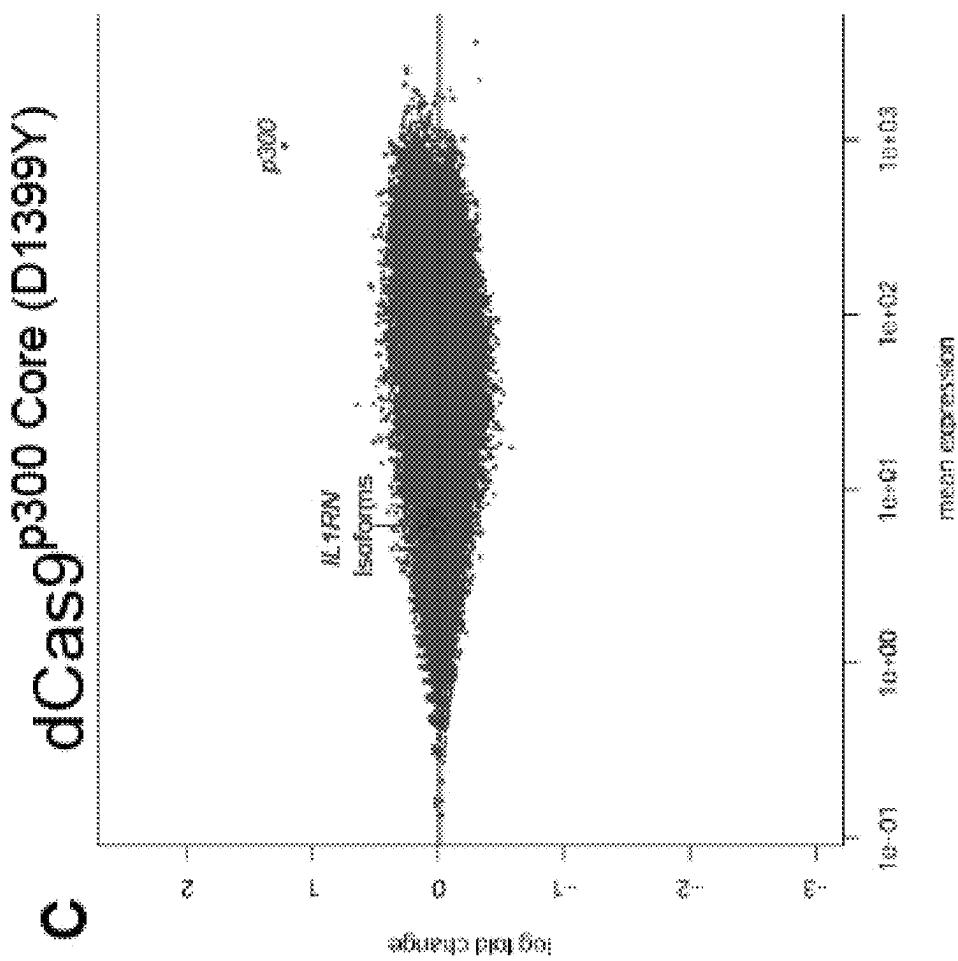


FIG. 3C

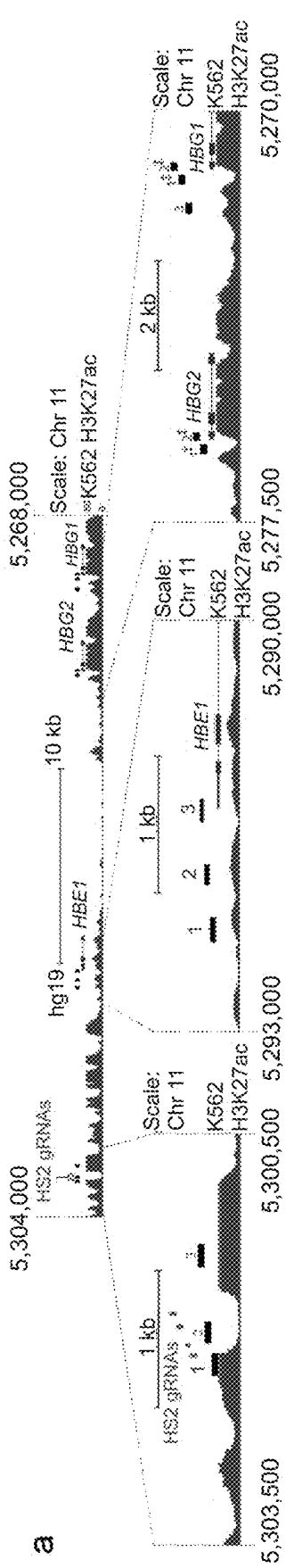


FIG. 4A

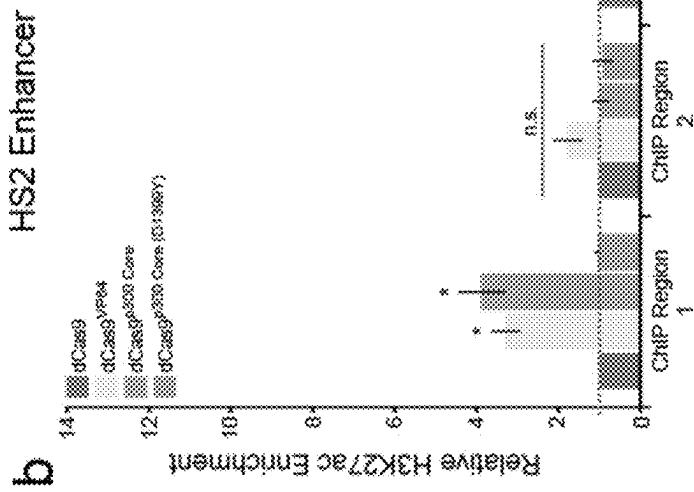
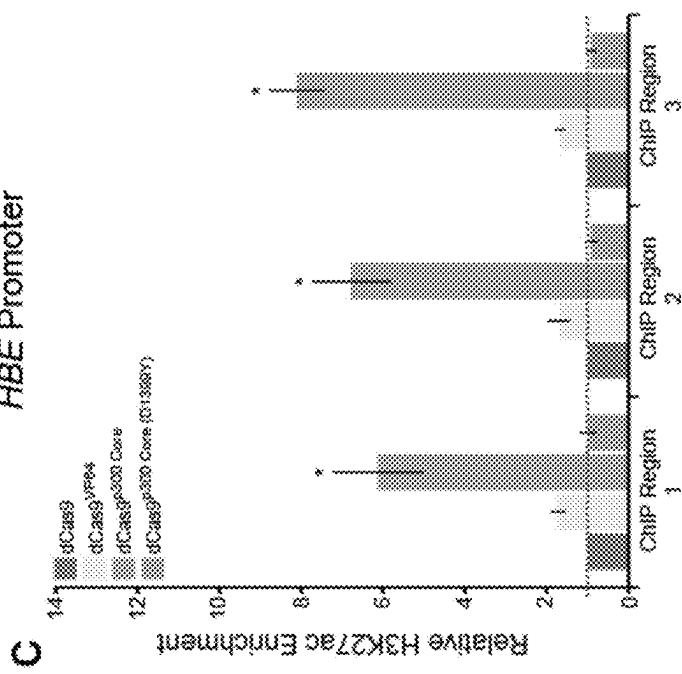


FIG. 4



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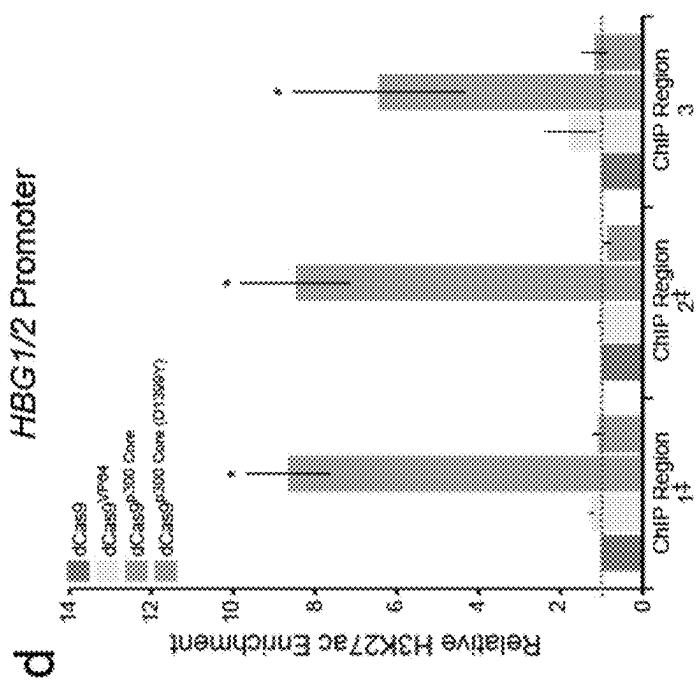


FIG. 4D

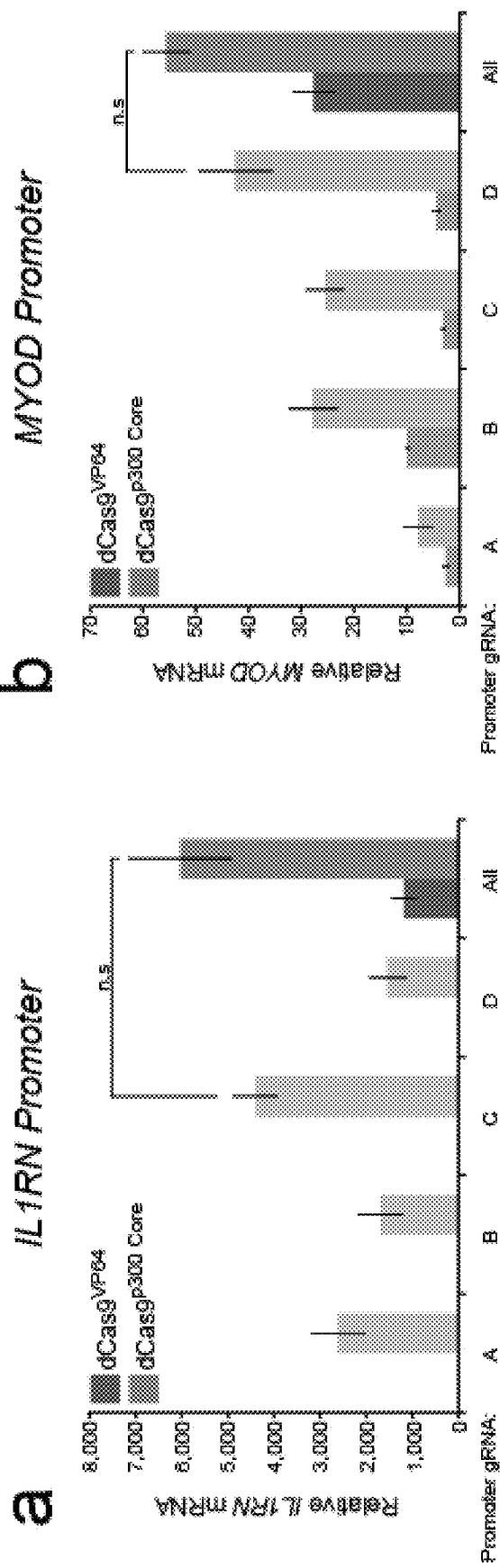


FIG. 5A

FIG. 5B

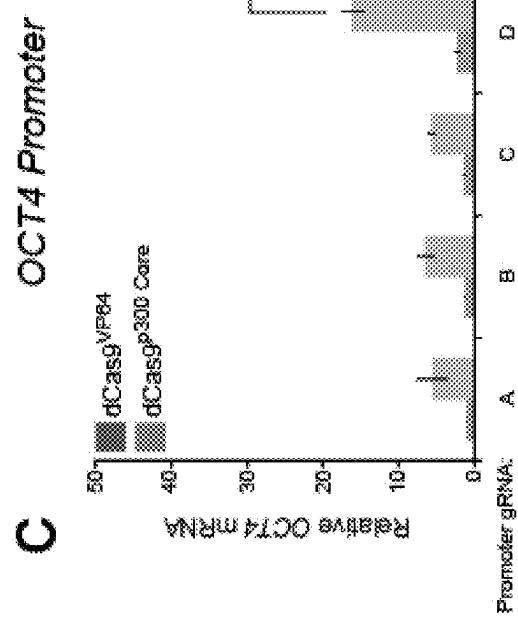


FIG. 5C

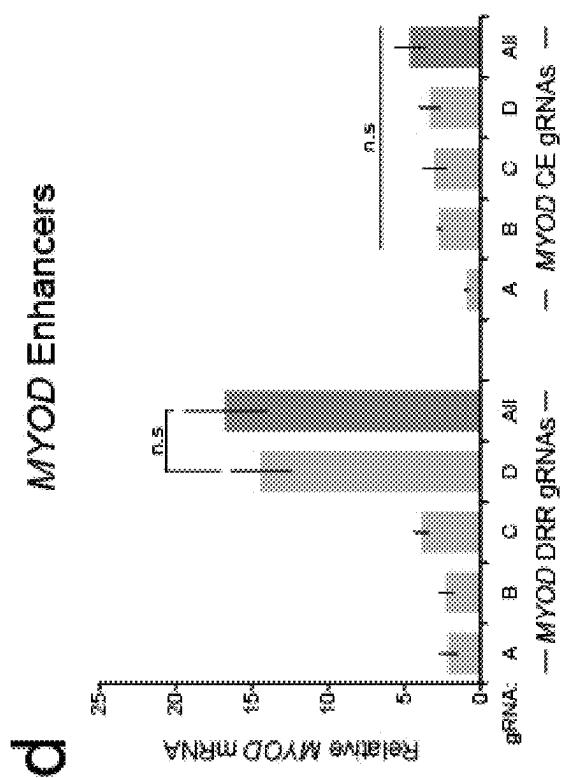
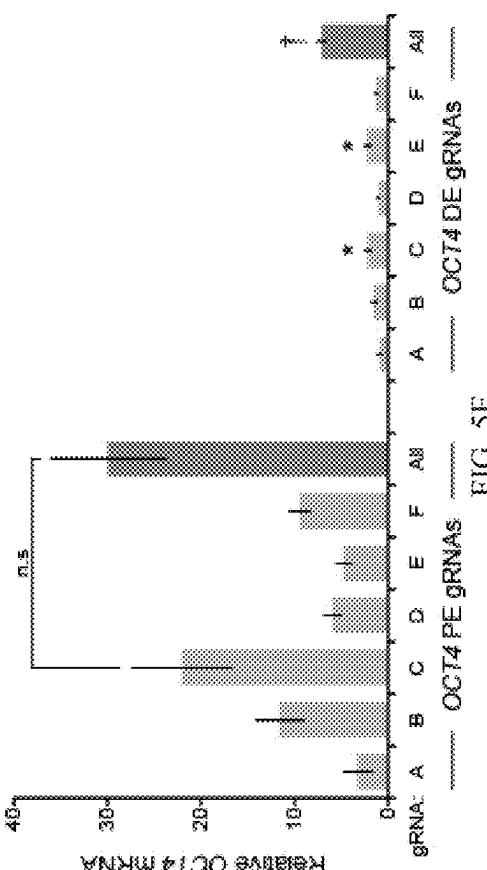
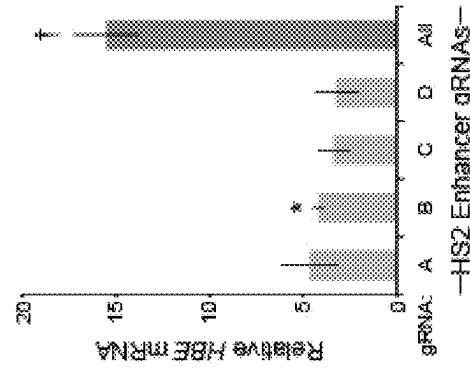


FIG. 5D

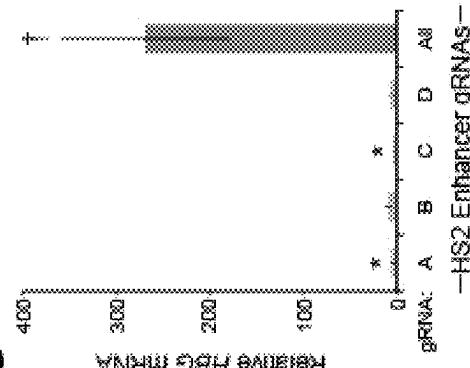
e OCT4 Enhancers



f HBE



g HBG



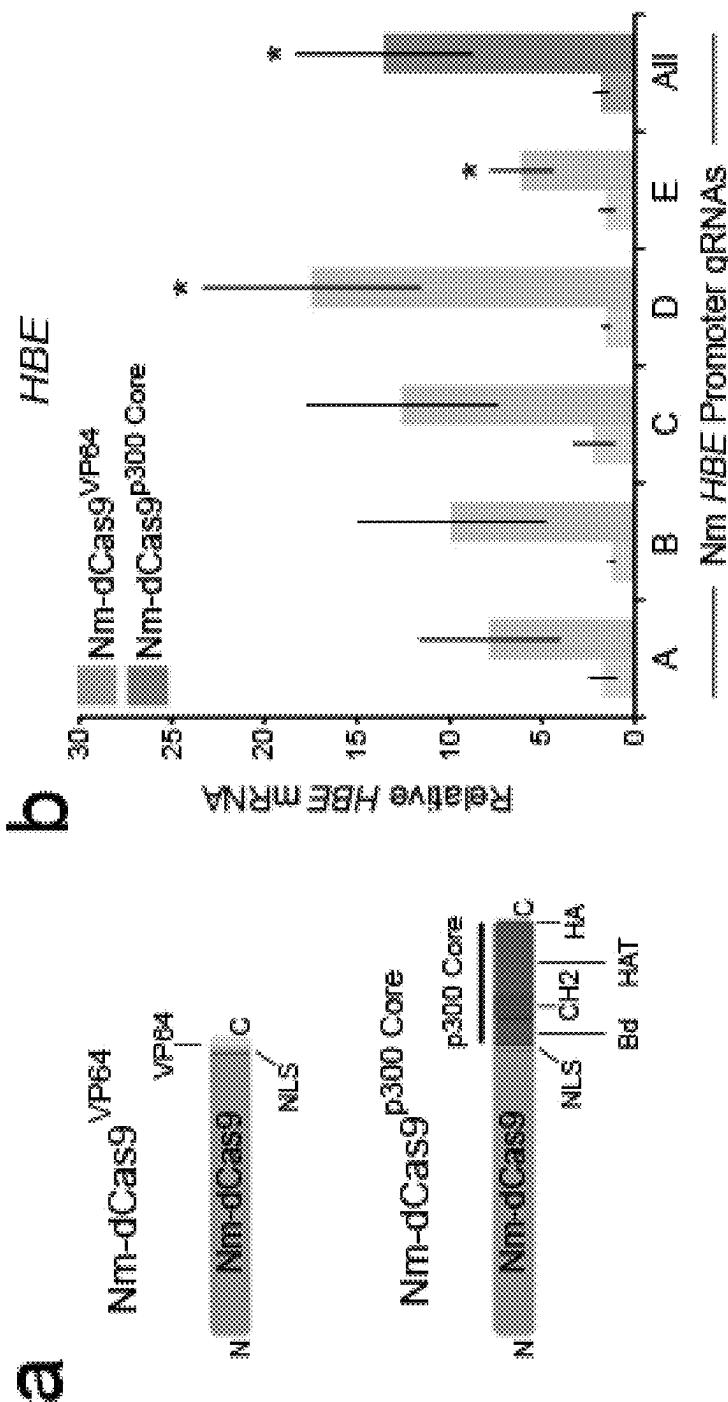


FIG. 6A

FIG. 6B

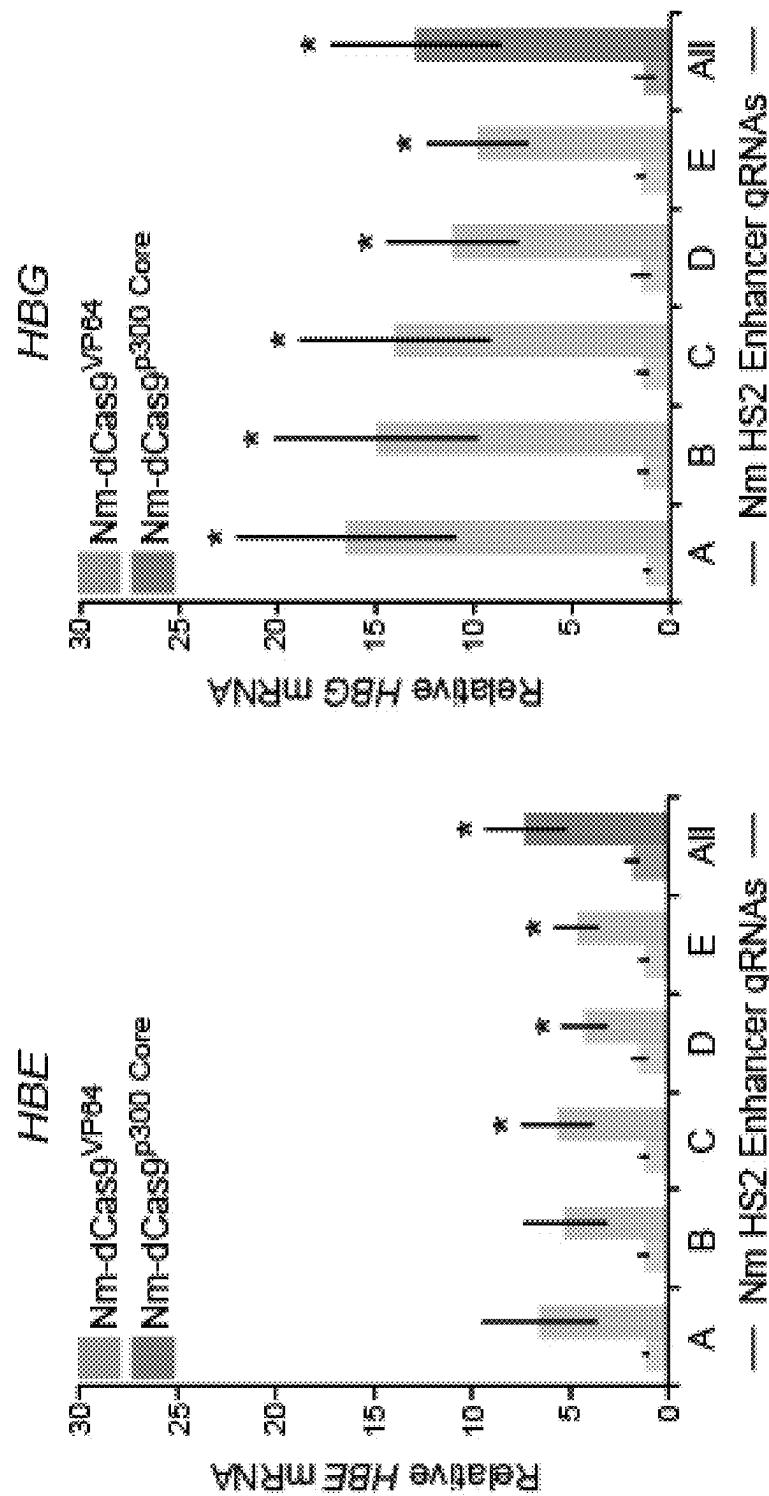


FIG. 6D

FIG. 6C

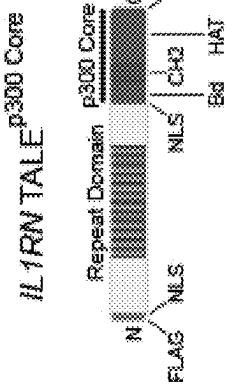
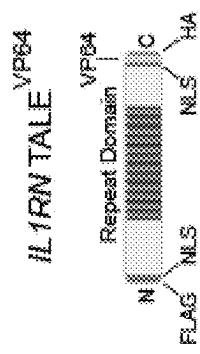


FIG. 6E

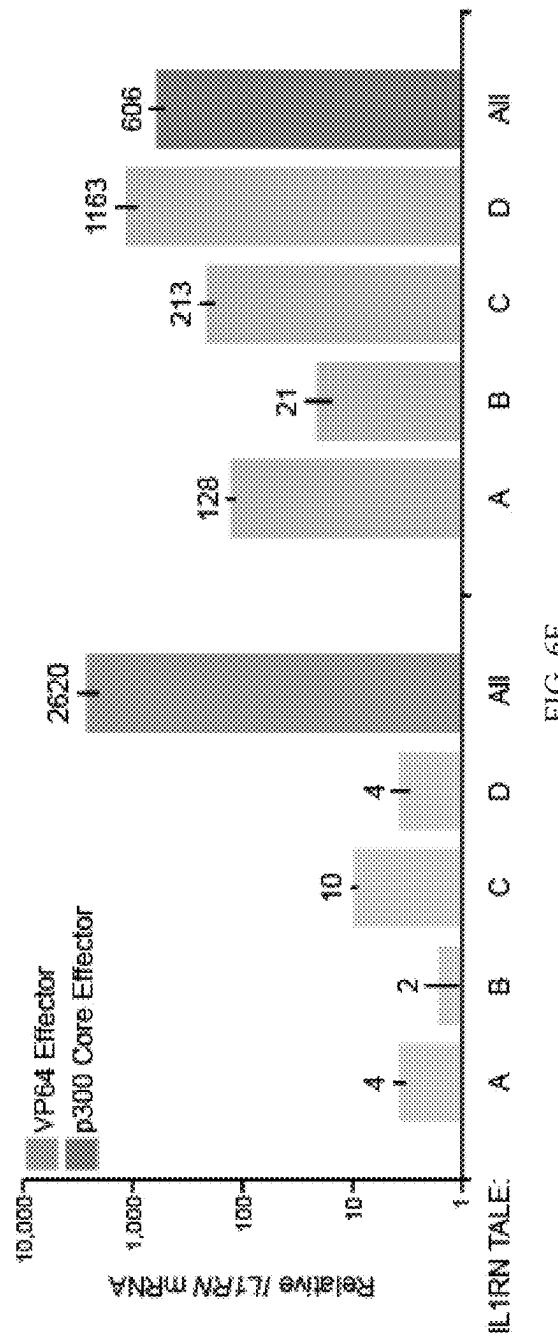


FIG. 6F

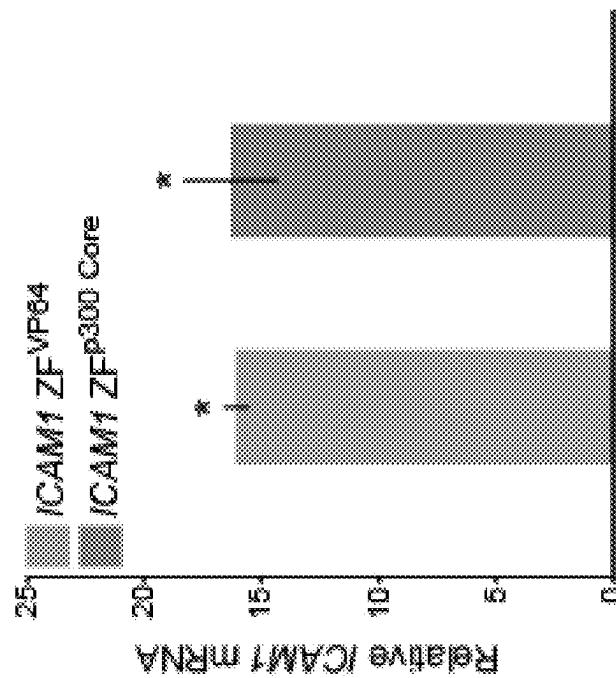


FIG. 6H

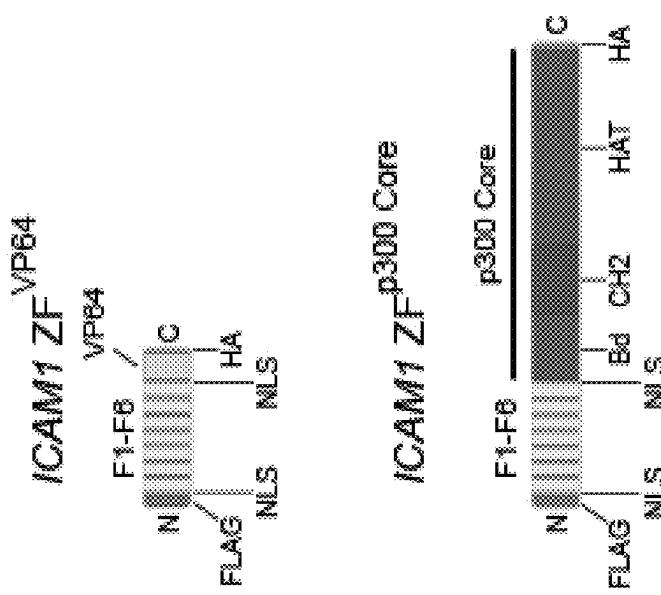


FIG. 6G

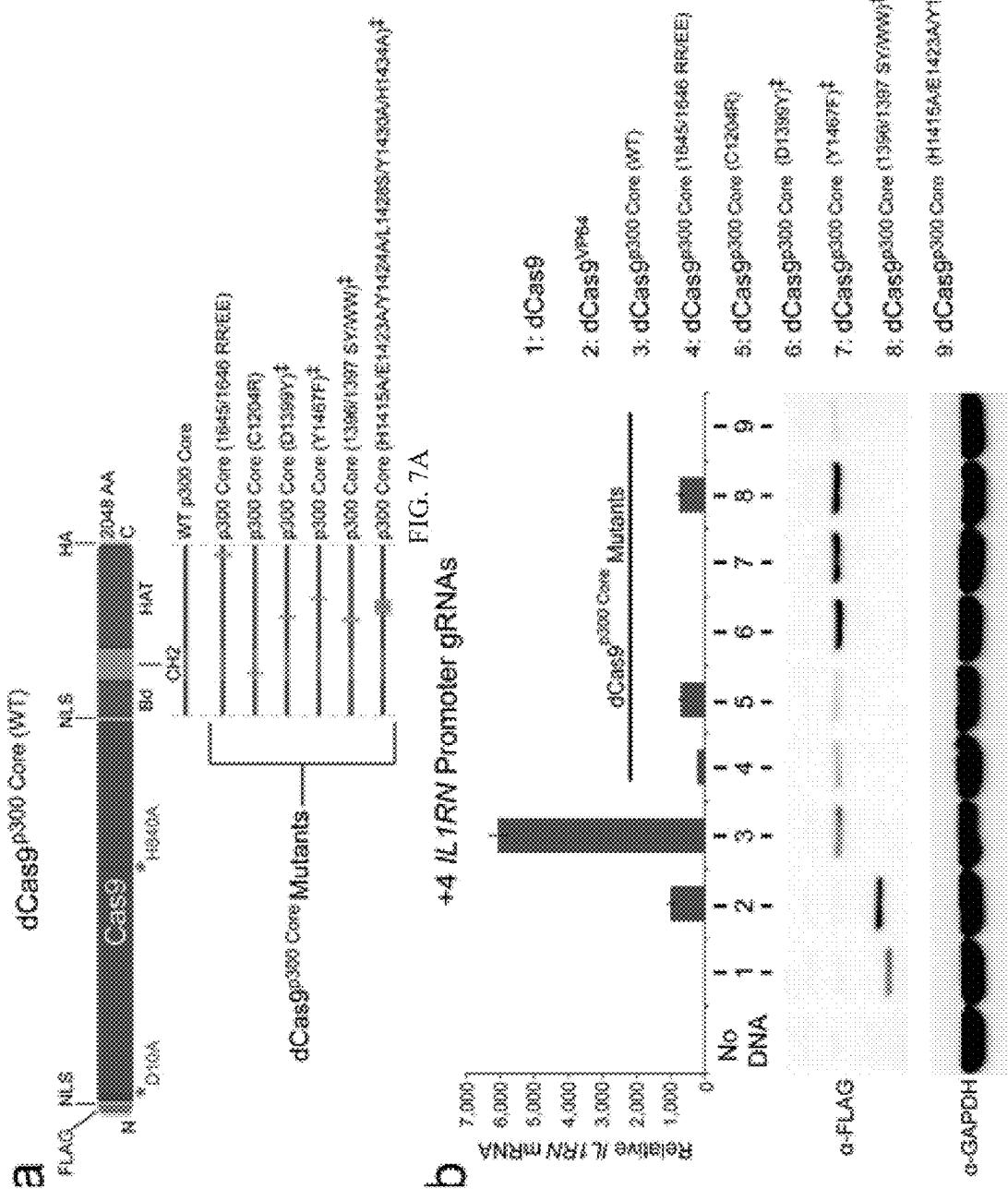


FIG. 7B

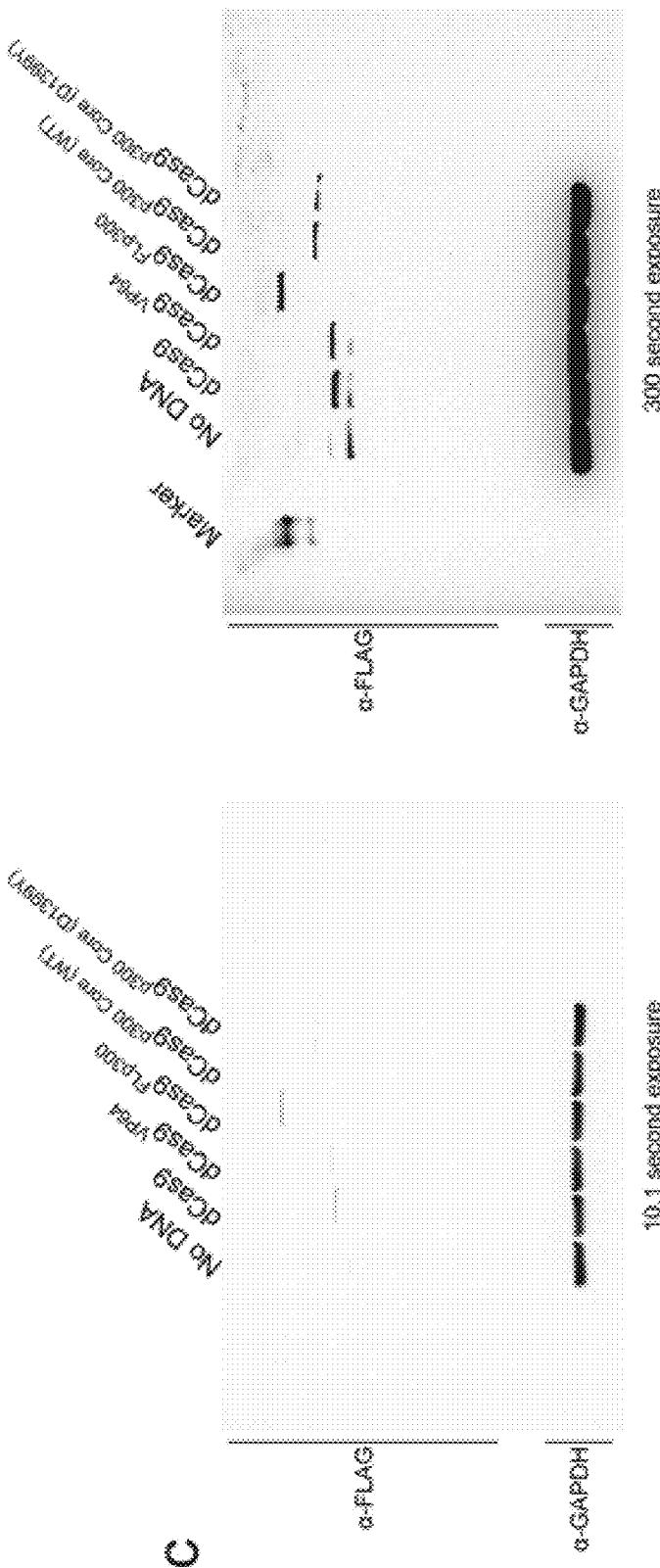


FIG. 7C

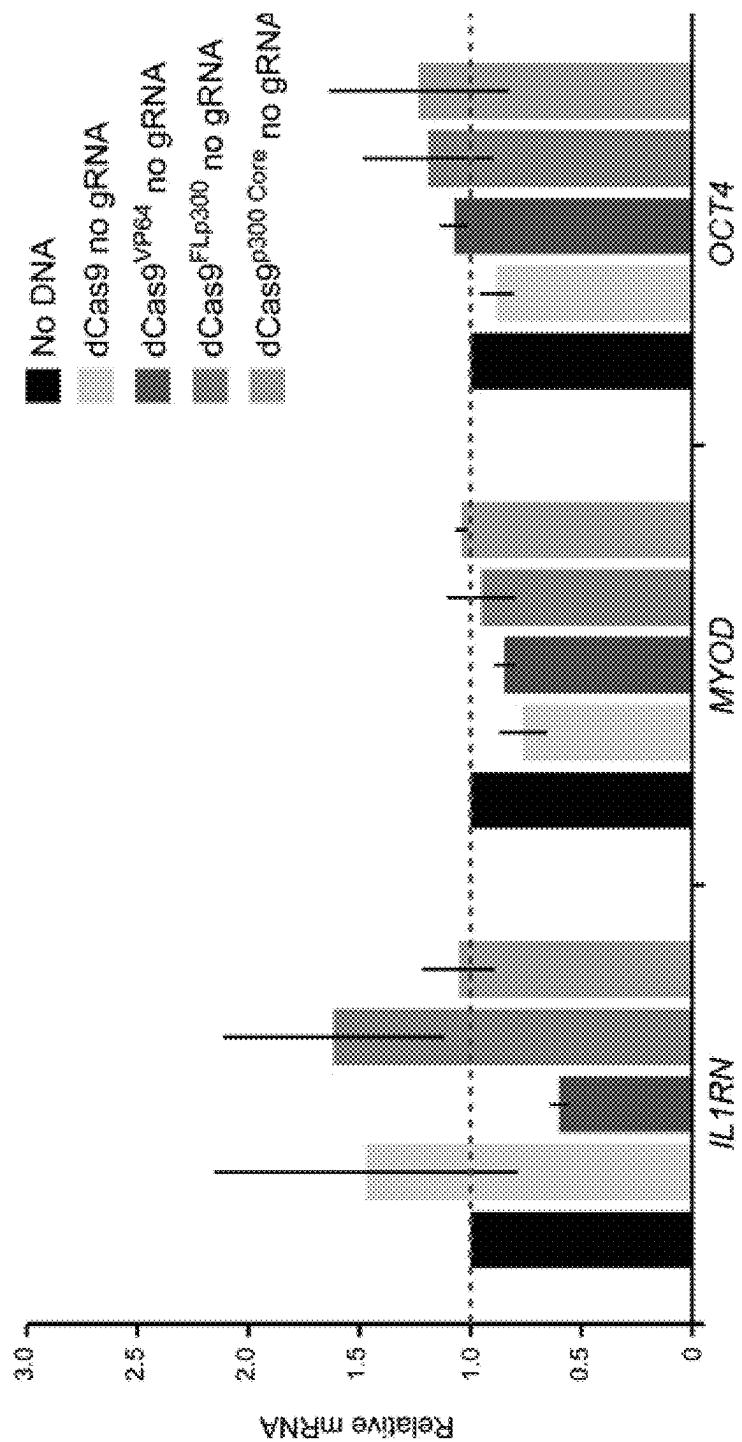


FIG. 8

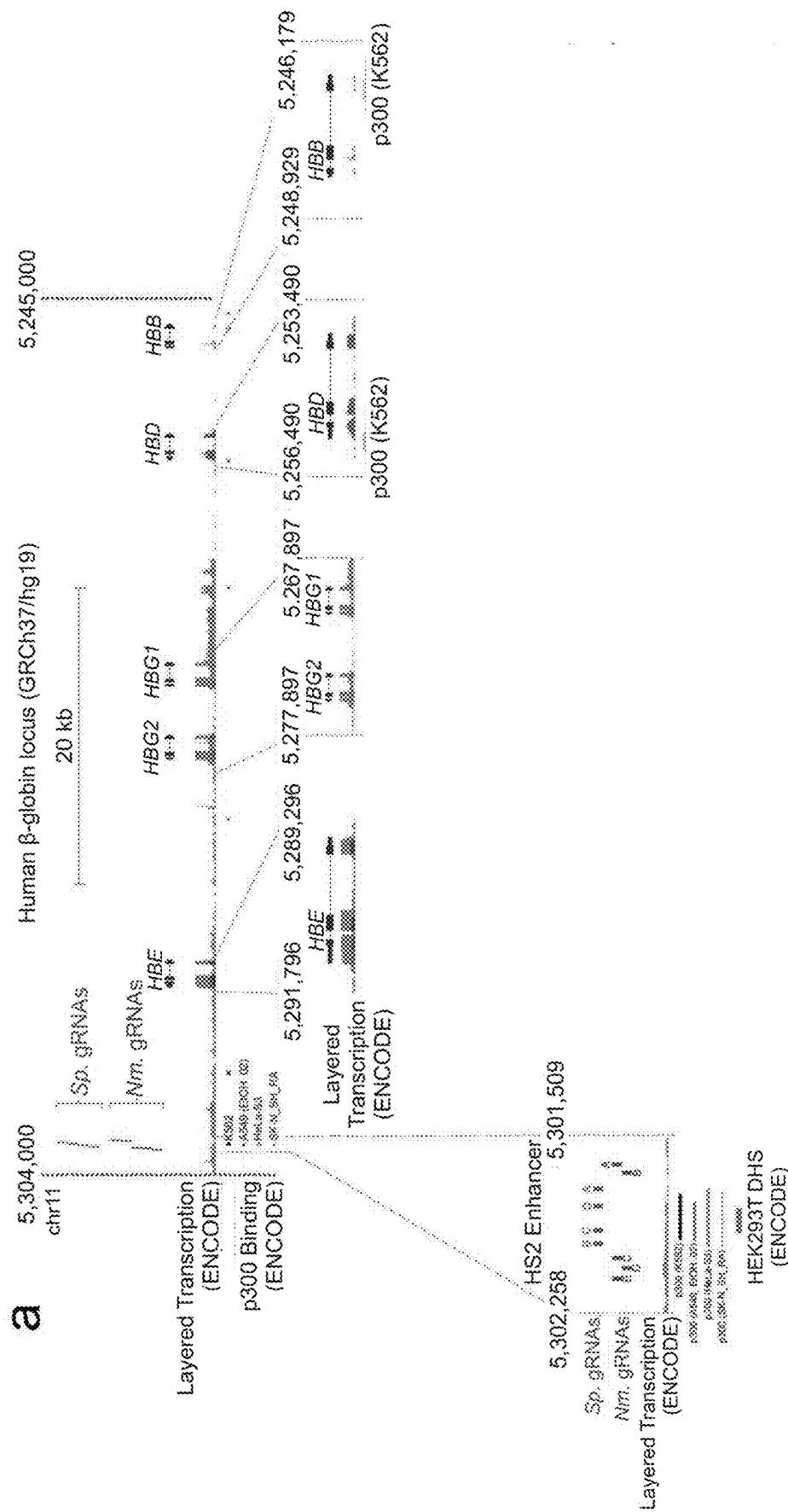


FIG. 9A

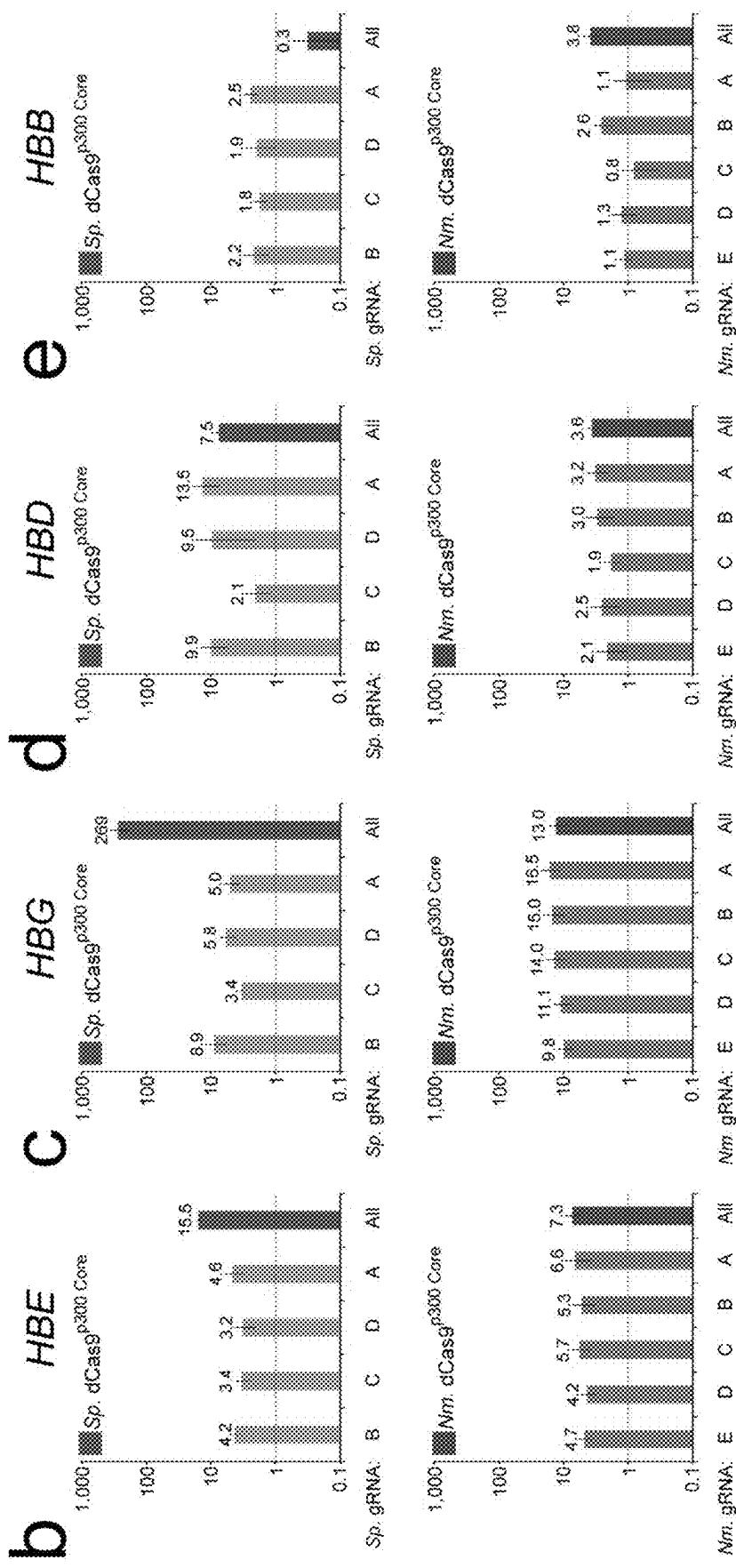
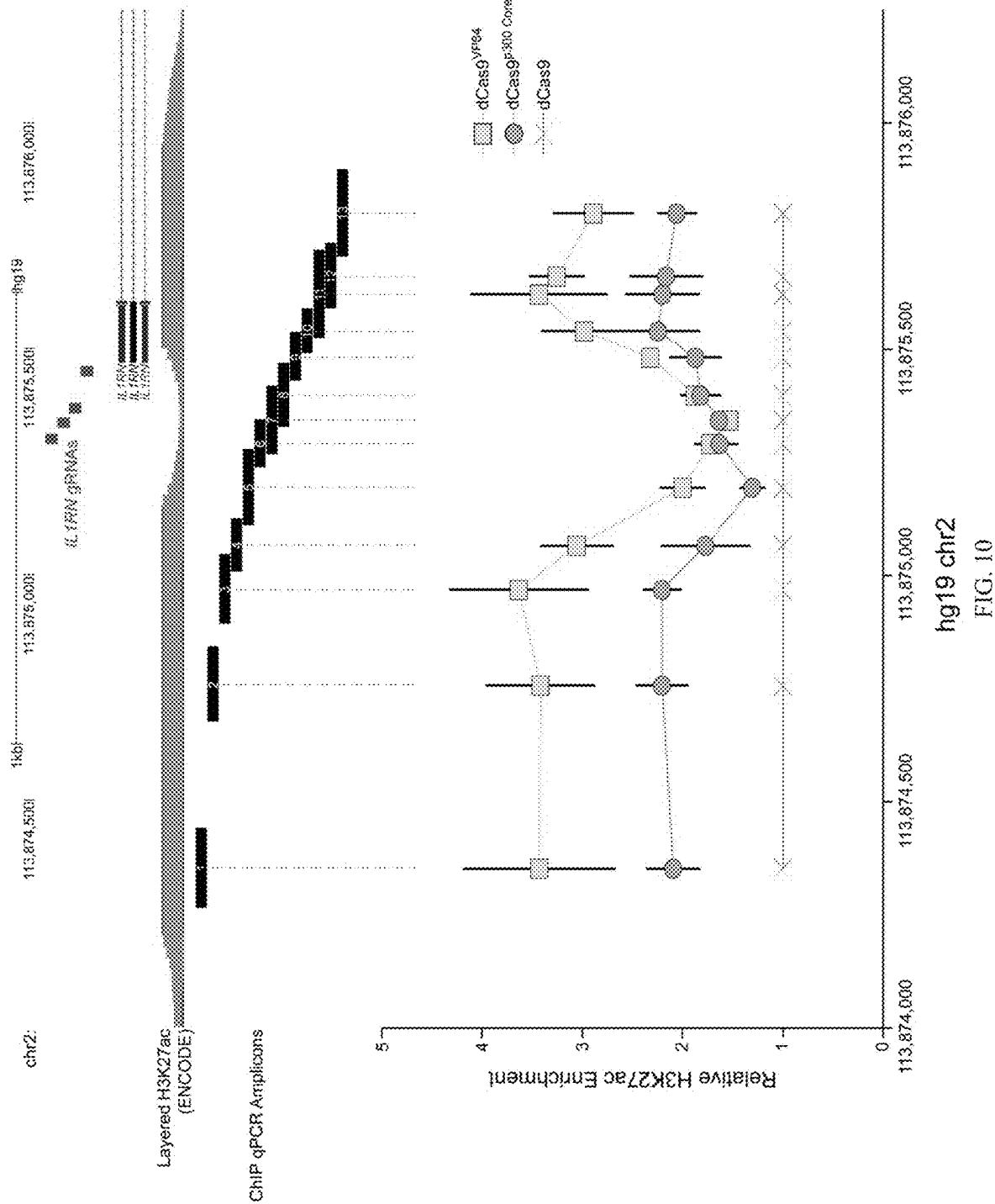


FIG. 9B

FIG. 9C

FIG. 9D

FIG. 9E



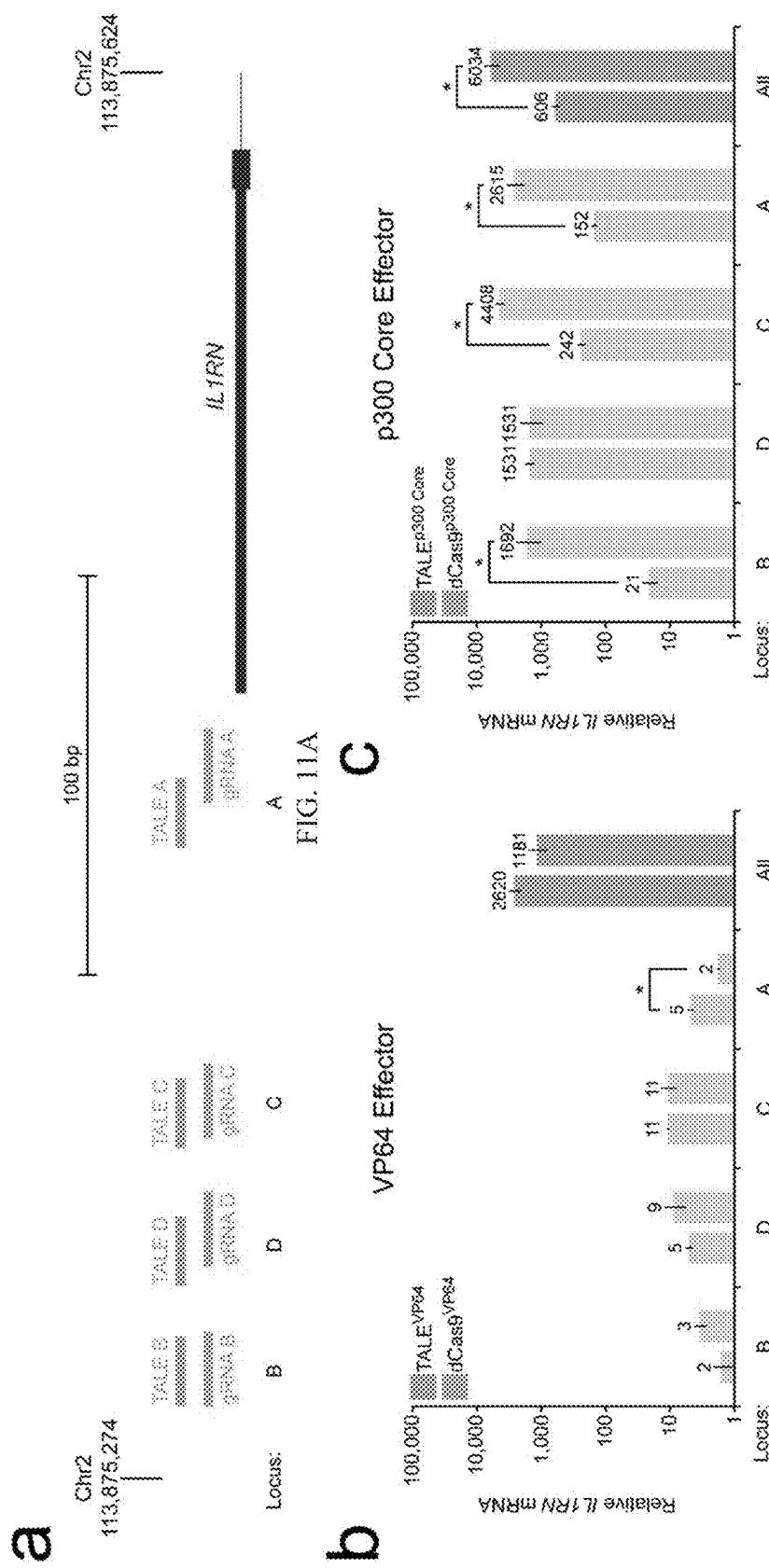


FIG. 11C

FIG. 11B

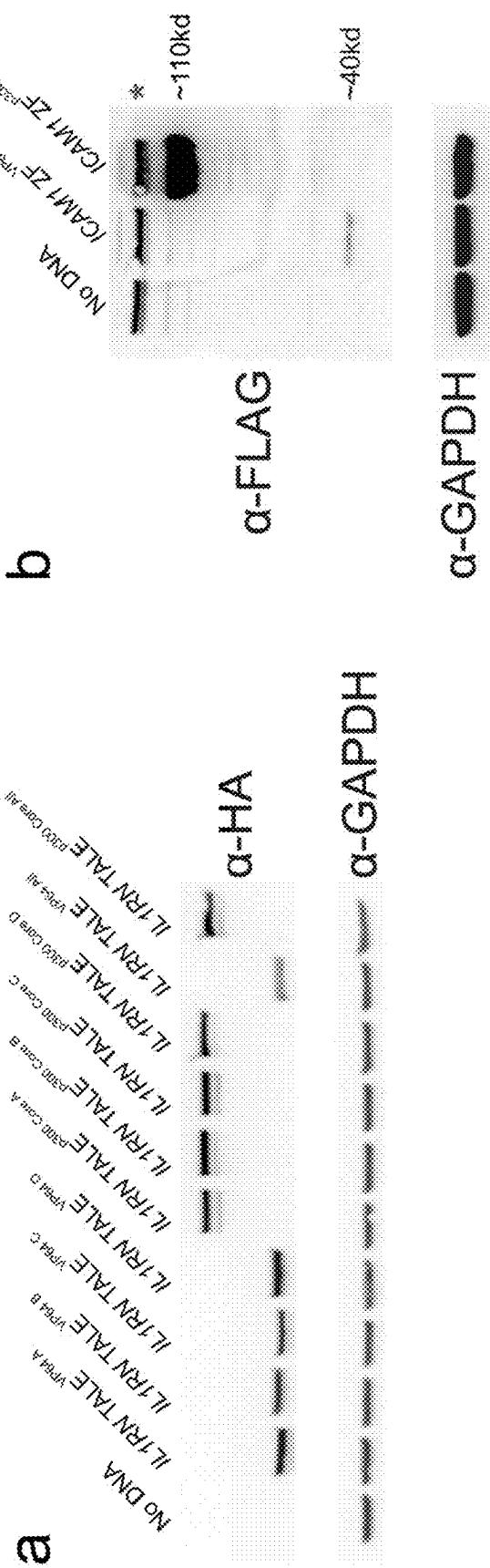
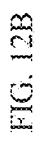


FIG. 12A



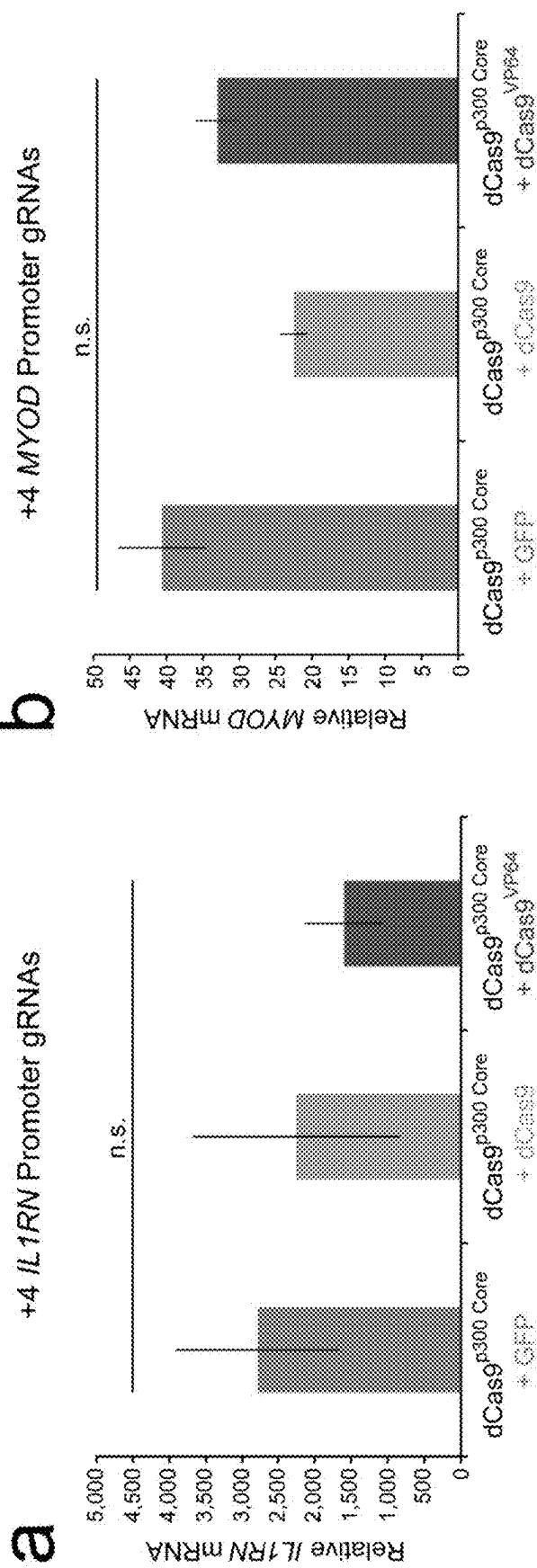


FIG. 13A

FIG. 13B

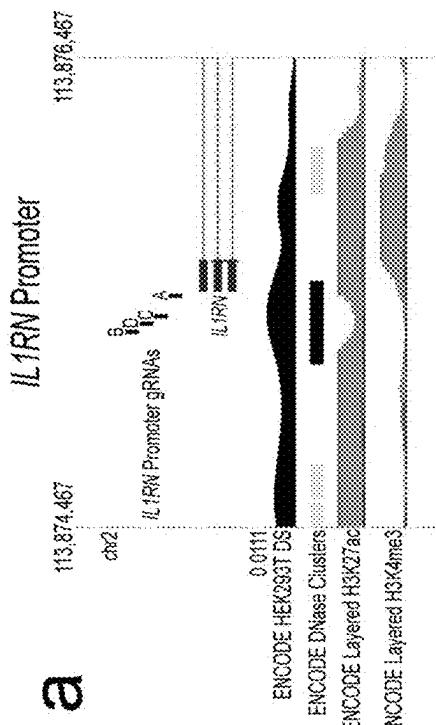


FIG. 14A

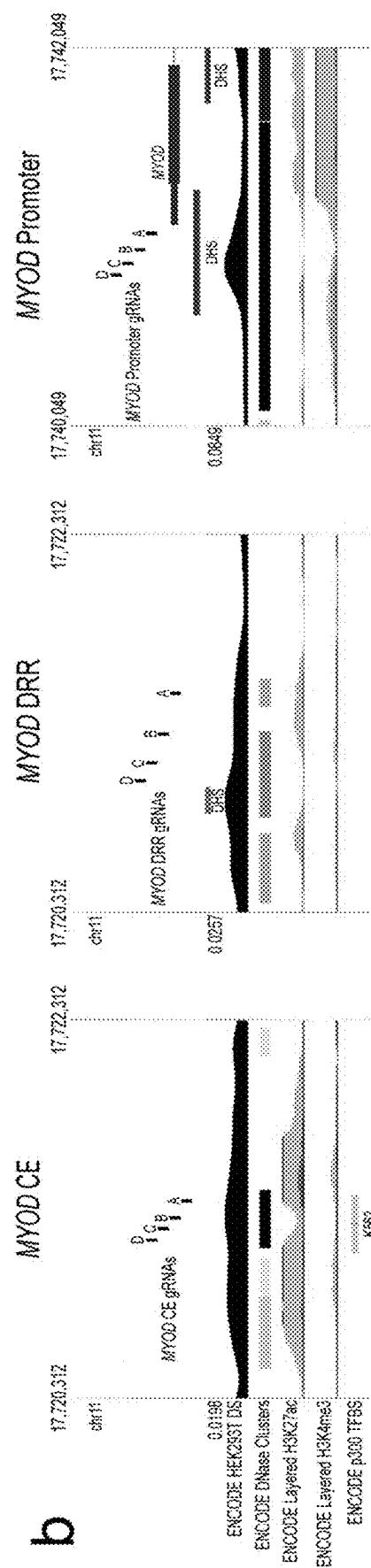


FIG. 14B

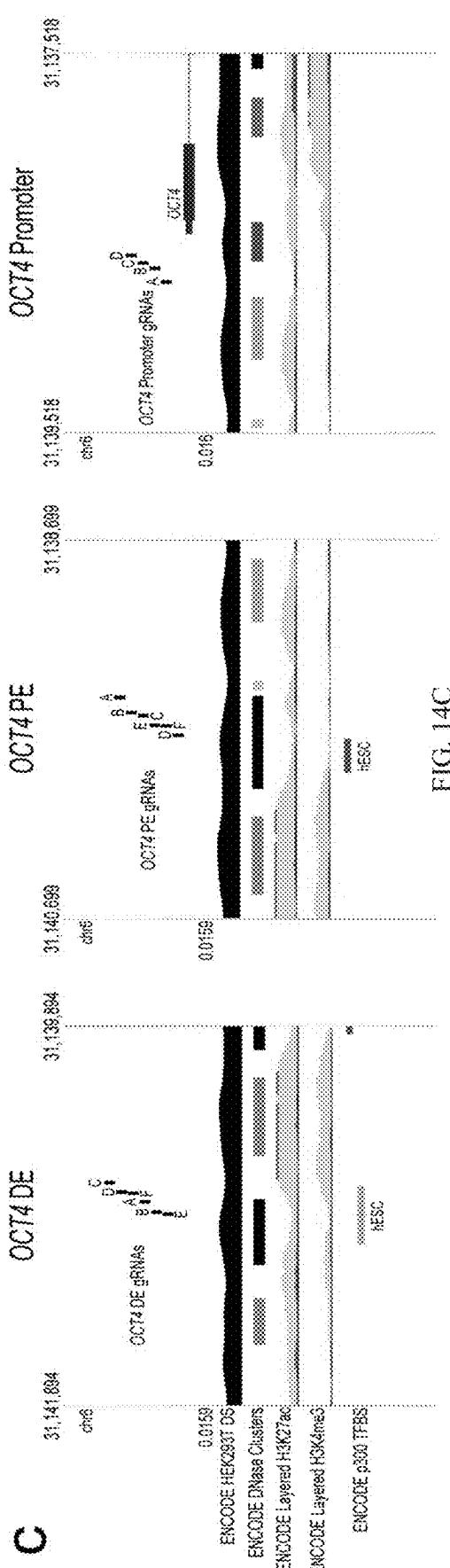


FIG. 14C

HS2 Enhancer

d

FIG. 14D

e

gRNA-Targeted Locus	Overlap DHS in HEK293T	Overlap DHS in Other ENCODE Lines	Multiple gRNAs Required for Maximal dCas9-gRNA Target Activation	Overlap Endogenous p300 in ENCORE Lines
IL17AN Promoter	N	Y	N	N
MYC2 CE	N	Y	N	Y
MYC2 DRN	Marginal	Y	N	N
MYC2 Promoter	N	Y	N	N
OCT4 CE	N	Y	Y	Y
OCT4 DR	N	Y	Y	N
OCT4 Promoter	N	Y	Y	Marginal
HE2 Enhancer	Y	Y	Y	Y

FIG. 14E

3dCas9 HA: (Addgene plasmid 67355) amino acid sequence: **HA**" Edition (SEQ ID NO: 138) Nuclear Localization Sequence, Streptavidin recognition Class (D10A, K44Q),

Nuclear Localization Sequence, *Streptococcus pyogenes* Cass (D10A).

MAPKKKRKVGRGDDKXKYSQSLACTIVSGWAVIDEXYKVSSKKFKULGNTDKHNSKMLALPDSQETAEATRKRTAER
YTRRKRNCYLGFSNEAKVQDVSFRRLSFLVTEENKKHEBHPGNVDEVAYHEKPYTHLBRKLVNSTUKANRMLAAMKFRGHFLLGCHNPTNSDV
DOKLICRVCOTYNOLFEENPHASGVCAKALSAEKSRSRBLMELAGLPCEKHGLFNLIALSLGALTPKFSKNDLAEDAKLQLSKOTYQDDBLNLAOQDQYADL
FLIAKNUSDAHLISDHRYNTTCAPIASABMKYDKEKBDLTILKALVQODLPERKVKEDPDKSKNGYADYINGASOEEFYKFBPHXEKDCTEILVNLNEOU
SKOTYDGGSPHQBLGELALBRCEFFPKDNREKHNUTPRPVYVGLARONGSRFAMNTRKSEETTPWNFEEVNDKASADSFERMTNPURKPHKVL
PSKSLLYEFTYVTELTKVVKVTYKOLDEKTKRKTCKANDLIFTRKVTYKOLDEKTKRKTCKANDLIFTRKVTYKOLDEKTKRKTCKANDLIFTRKVTYKOLDE
EVVYLILFEDREMEERLKTYAHLEDKVVKOLKFRYTOWGRSLSKLNGDHDGCKTIDLSKSGFAHNNKOLKEDSLTRKEDOKACNSOGCDLHENDA
AGSPANKGQCATVKVADLVVKVAGBHEPENWEMARENGTIGKGSMSERKMRKREEGKELGSQKKEHEVNTGONEXKLVYLYLNGKHMVYUQCLHUNR
SOYODAYVQCSFLKODSINKVATRDKRCKSDNVESEFTVKKKSVROLLNAKLITORNPHNLTRKRGSLSELOKAGEWRLVETRQTKHNCQCLCSEB
NTKYDMDKLIKEV/KVTLKSKLVSEEDDFOYKVNENKVKHADAYLHAVVGTAIKCKYPKLESEVYQCYKXKDVVKMKAESOENGSKAYAKYTFYSHMNFCT
STLAKSENKORPLKETNGTGYEVWDKGDFATVVKVLSPQVNNWKTTEVGTOGFSKESLPKRNSDULKARKEWDOPPKYGFOSPTVAYSMLVVAEKGSKX
KXKSVWLLGTCIERSSEFKAPUWLEAKQYKVKDULKLRVSEELNGGKSMWASACELQGNELAQPSKXVWELVASHYXKLUKOSSETEKONQOLFVEQAK
KXKRVGRA
INPVDPDAS

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dCas9^{FL} psico: Addgene Plasmid 61356) amino acid sequence; **“HA”** epitope (SEQ ID NO: 140).

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Nuclear Localization Sequence, *Streptococcus pyogenes* Cas9 (S9A).

MAPKKRKVGRGMOKKVSGLAIGTNSVGWAVITBEYKPSKPKVAGLNTDRHSSKRMUIGALIPOSGETAEATLKRYTARRY
TRKARRSITYLOEFSNEAKVDSFFHRLLESPLVEEDKHERWIFGNYDVEVAYHEKPYTTHLRRKLVOSTDKADLRLLTALLAHWKFRSHYJEGELNPDKNSOVD
KLPHOLVCTYWHOLFENPMNASCYDAKAI.SARLSKSRRLENLJACSLPGEKKGNGLEGNLALSGLTPNFKSNFDLAEDAKLOLSCKOTYDDLDNLLAGDQGYADFL
AAKNA.SDAA.LSDA.RVNTETKAPLASSMKRVDYDANQJTLKA.VRQOL.REKYKHEFDQSINGVAGYDGSQEEFYKEKPFLKMDCTEELLWKLNGEDLURK
GRTFGNGSIPHCHLSELHAR.KRGEOFYFLKDNREKIEKIL.TRPYFYVGLPARCHSEFABWTRSKSEETHTPVNFEVVTKGASACGSPIERMTNDKMLPNEKVLPK
HSLLYEFYTIVNELTKVKYTCOMRKPAFLSCEOKWAVOLLFIKTNRKVUTVKOLKEDYFVKIECEGSVESOWEDRFNASLGTYHDLRUKDOKDFLMEEDLIEDVL
TYLFEDUREMEEERLKTYAHLUOKVVKMQLKRRYTGWLRSKELINGRDKOSOKYULDLSUDGFAMBNENOLHDDSLTEKDOKKAGQVSQGOSLMEHANLAC
SPANKGLOTYKVOELVKMNGBHNKPNENVNEMASENCOTOKGOKNREMRMCKREEGRMGSOKKEHNPVENTCLOKEMVLYYLCKACRDMVYDQGDLDNRI.SOVD
VOAIVPCSFKXODSIIKVLTKLVSDFBKDFCYKVRNNYHANCHAYNAVYGTIAJMKRKPKESEFVYGDYKVKYDVWKRMIAKSECHOKATAKYFFYSHNNNNFFKTHILANG
NOKLJERBYKVTLSKLVSDFBKDFCYKVRNNYHANCHAYNAVYGTIAJMKRKPKESEFVYGDYKVKYDVWKRMIAKSECHOKATAKYFFYSHNNNNFFKTHILANG
EIRKPLFLETINGETGEVNWOKRDFAUTVKRVLSPMPQVNUKCTEVOTCGFSKESEPKRNSKJLAJKRWDHPKKGFSKUJKSVKEL.
LGHIMMERSSEFKEPKPFLLEANGYKEVKKCOLLKLPKSLSFELENCRRKRMJASAGELOKGMELALPSKVNFLYASHYECKKESSPEDENEOKGOLFVEQHCKYLDENEON
SEFSKRNVAJADKLYDVALSAYWKHRKPIREDAEKHLFTLNLCSAPAKYFOTMOKRATVISTKENDATMHSOTOLYETRDLSCUSGPKASPKKKRKVG

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dCas9-gRNA Core (D1398Y); (Addgene Plasmid 61358) amino acid sequence: D1398Y, KEGG ID: Q10A, "HA" Epitope (SEQ ID NO: 142) Nuclear Localization Sequence, Sphingomyelins Biosynthesis Class 9

MAPKKKKVGRGMWKKYKXSLAGTNGVWAVITDEKKYKPKKVKYI. CNTDRKSHKKNLGALI. FDSCTAYAATLKRTARRY
TRKMKRIVOLQEFNEMANCOJOSSEFHRLVEESFLVIEEDKXKEHESKPKKVKYI. CNTDRKSHKKNLGALI. FDSCTAYAATLKRTARRY
KLFQLOLVOYVQHLEEMPKASGVUAKAHLASARLSSCRRLMIAOLPGEKWNCHFONLMSGLTPAKKSNBLAOUAKLQLSKOTYDUDLWLLAQGQOYQYLFLR
AAKNI SDALLSHD RWTNLTSAKSMKRYD-HPDLTLKALVRCQOLPEKXKEEFDCKSKWGYACYDQGASQEELYKPKPKLKEKOGTEBLRK
GRTYINGSPHQHAGELHAI. ERGDNFYPFLKONSEMEKLTRIPYVCPPLRGKSRAMWTTKSEETTPPKNNEFVQKGSASGSCFEEFTNFCKOLPKVLPK
HSLLVYCFYNELTKVYNTCEGKAVNOLLKTKHCKVTKOKEDYFKKECDFSVESGVEDRPNASLCTYHDLKIKOKDOLNEENEDEEDVL
LTLDEKREMEHEHUKVYAH. FUDKVKHOKLKRRTYTGWRSLKLNCHDKCOKSKTKDPLKSGDFARNPQCGHNDLTFKEDOKAONVSQGDOSLKHMHMHAAS
SPAKKQGCOLTVANVDELVKVKGKPERKETTAKERQQTGKGCNSTERMKRKEESEKPKKVKYI. CNTDRKSHKKNLGALI. FDSCTAYAATLKRTARRY
VOMAVPSFLKINDSINWKLTRSDKWRKSKINVPSEEVKRMKNNYVROLN. KAROKTORKFINTKAERQSLSELRAKSFNKWLVETRQTKHAGLDSRMTKYD
ENDKREYVITLKSFLVSDFRKDFGTYKUREVNPHANDAYLNAVVGFLUKKPKLESEFVGDYKVKYDVKRMKNNYVROLN. KAROKTORKFINTKAERQSLSELRAKSFNKWLVETRQTKHAGLDSRMTKYD
GERKPKPLTETNGTGEVWQKDFATVRKVLSEHQVNVKKTTEVGTCSKESILPKRNNSKMLAUSKQWPKYQGSDPTVASYVLVAKVECKSKLKSVK
ELL GTHMERSSFERKPDFLAEKGKWEVKMDLULKPKYSLFELNOKRMLASAGELQSNELALSKYVNLFLASHYKLGKSPDNEQGKOLFGVQHMYLDNE
QSEFSKRYHADMLWVA. SAVKHWOKPREGAHHM. PTHQAPKAKHOTTKRKYTSKWEVLA. LKNGTCS. YETRKO. SON COPIAGSKASPKKKVKY

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dCas9-g30 Cons (C120R); (Addgene Plasmid 61361) amino acid sequence; C10A, H34A, C110A, "HA" Eptope (SEQ ID NO: 144)

Nuclear Localization Sequence, Streptococcus pyogenes Class I

M**MAPKKKKRKVGRGDDKKYKSYCLAGTNSYQAVITYDEYKVPSSKKFVKLGHTHSHSKKLIGALLFSGETAYAFLKRTA>**
YTRKKKRCYLOEFSNEMAKVDOSFFHLLEESLVEETKKHEERPHPSNEVDEVAVHENKYPTIVHRRKVLVDTSTKADLRLVIA
DKLFLCHVQTYNCGLFEERPHASQVDAKALSAABLSSKSLERLALPGEKKKGLVNLALPGLCITPNFKSNFDLAEADAKLCLS
PLAARNLSDAHLSDHHRVNTETYAPLSASWKRVDENHEDLTLKALVRQDLPKREKEKFLFPPFLKDWREKEKFLFPPFLKDWREKE
PKQRTTFCNGSSPHQHNLGELMAUERQDFVPPFLKDWREKEKFLFPPFLKDWREKEKFLFPPFLKDWREKEKFLFPPFLKDWREKE
PKSLLYEFVTVNELLWVQVVTGKVKVTPGKVKVTPGKVKVTPGKVKVTPGKVKVTPGKVKVTPGKVKVTPGKVKVTPGKVK
DVLTLTLEDFRMEERFLKTYAHLDOKVWKOLKPRRTYTGWGRLLRKLINGRDKGSGKTLDFLXSGFANAMWQLNDLSLT
WLAGSPANKKGLQTVWVNLNKVWGRMHNARENQTYCKGOKNSBERMKIEGKELGQILKEMPVENATQLONEWLLYLQNGCOWVDOELDNR
LSDYDVA>PQSELKEDQCDMMLNACLTGCRGKQNLNAKLTGCRGKQNLNAKLTGCRGKQNLNAKLTGCRGKQNLNAKLTGCR
NRYDENMLKREVKVWTLKSKLVWVKDFQYVWENKAWHADAYNANVGTALKKPKLESEEVYGDVKVYDVKVYDVKVYDVKV
ETLANGEFRNRFQIETNGETGEGTGEVGTGEGESKESHPKNSKQIARKNWSDEKKFSEGDFTVYDVKVYDVKVYDVKVYDVKV
KLSVSKELGQTMRSSEFEKPNPDLFLEAKQVKEVKQVKEVKQVKEVKQVKEVKQVKEVKQVKEVKQVKEVKQVKEVKQVKEVK
HYDDEEQQSEFSKRVHADANLDRVLSAYNHKTKPBDQEAENHHLFTLNGAPAAFKYFOTIDKRYTSIKEVLDATINCSTISIYETRDI
PKKKRKVGR**A**

FIG. 15F

dCas9aa Core (Y145F): (Addgene Plasmid 61362) amino acid sequence: **[REDACTED]**, Nuclear Localization Sequence, **[REDACTED]**, Streptococcus pyogenes Cas9

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dCas9p300 Core (1386-1397 SVWVW): Addgene Plasmid 61363 amino acid sequence: **[REDACTED]** Nuclear Localization Sequence, **[REDACTED]** Sypapoxocysnycggees

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Nuclear Localization Sequence, “NLS” Epitope (SEQ ID NO: 147) H1136A E1423A Y1428S Y1430A H1134A, “HA” Epitope (SEQ ID NO: 147) H1136A E1423A Y1428S Y1430A H1134A

VEDA VEDAS

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Nm-dCas9^{rec}, amino acid sequence (Addgene Plasmid #48676), *Neisseria meningitidis* Cass 1016A, 0387A, 0388A, 0611A, Nuclear Localization
Sequence: [REDACTED] (SEQ ID NO: 148)

MAAYKPNPINYLGLAGIASVGVWANVEDEDENPCLIDGVWYFPERAEVYVKTGOSLAMABBLARSVERLTYRAHRLURABBLIKREGVYQAADEFNENGLKS
 LPNTPWOLRRAAALDKLTPLEVSAILHLKHKGVLISQRKNEGETADKELGALLKGVADNAHALOTGDEFTPAELALNKKEKESGHINNORGDOYSHFSRKD
 LOAEILLLFEKOKFEGFMPHNSGLKEGETLLMTPORPLALSGDAVOKMLGHCIFTEPAEPPKAAKNTYTAERFIWLTNLNRLIEOGSERPLTTERATLMDEPYR
 KSKLTYAQSKRLLGLEDIAFFKGLRKGKNAEASTLWEMKAYVAKRSHALEKEGSKLXKUKSPNLSFLPELQDEGIAFSLFKTDUEHTGRUKUNQPEEALLKHSR
 DKPVQGSLSKALARRVPLMEQGKRYDEACAEYGDHYGKNTEEKVYKPPAEMERNPQVVRULSQUARKVINGVVRYYGSPARHETAREVSKSFKDRKEKRCGE
 ENRKDREKAAKFREYFPNPFYGEPKSKDLKLRLYEQOHCKLGLYSKEMNLGRNEKSVEAALPFFSRTRWIOSPNNKVLVLSGEAQNKGQCPYEFVNSKD
 NSREWQEFKARVETSRSPSKKORILLQKFEDDSKFERALNOTRYVNRFLLCQFVADMWLTGKGKCRVFAANGQITLRLRGWLGWLXVRAENDRHALDAVV
 VACSTVAKQGKITHVYKREKHNADFOKTIDKETGEWUHQKTHFYQPKDGFEEAOIPEKLRLLAELKLSRPEAVHEYVTPLFVSRSA
 PWRKMKSSQGHMETVSKAKRDLGEVSVLVPLTQLKAKRLEAHKDOPAKAPAKAEPKYDKAGNRTOCQKVAVREQVQTKSV
 WSYKHNHGQADNAATWVYDVFYKQKDYKLYVPSWQVAKGIPDRAVVGQKQKDYKQYDKEEDWQKIDOSPFNSLJPNOLVETVTKARMFGYFASCHRTGHNHRQD
 OHMGKNGGLEGIGVYKTALESFQYQDELGKEFRPCRLKKRPPVRSRADPKKKRKRVEASRA
 SR

Nm-dCas9^{rec}, (Addgene Plasmid 61365) amino acid sequence; *Neisseria meningitidis* Cass 1016A, 0387A, 0388A, 0611A, Nuclear Localization
Sequence: [REDACTED] (SEQ ID NO: 149)
"HA" Epitope: [REDACTED] (SEQ ID NO: 149)
 MAAYKPNPINYLGLAGIASVGVWANVEDEDENPCLIDGVWYFPERAEVYVKTGOSLAMABBLARSVERLTYRAHRLURABBLIKREGVYQAADEFNENGLKS
 PNTPWOLRRAAALDKLTPLEVSAILHLKHKGVLISQRKNEGETADKELGALLKGVADNAHALOTGDEFTPAELALNKKEKESGHINNORGDOYSHFSRKD
 AEILLLFEKOKFEGFMPHNSGLKEGETLLMTPORPLALSGDAVOKMLGHCIFTEPAEPPKAAKNTYTAERFIWLTNLNRLIEOGSERPLTTERATLMDEPYR
 LLYAQASRLLGLEDIAFFKGLRKGKNAEASTLWEMKAYHAKRSHALEKEGSKLXKUKSPNLSFLPELQDEGIAFSLFKTDUEHTGRUKUNQPEEALLKHSR
 QSIKALARRVPLMEQGKRYDEACAEYGDHYGKNTEEKVYKPPAEMERNPQVVRULSQUARKVINGVVRYYGSPARHETAREVSKSFKDRKEKRCGE
 DREKAAAKFREYFPNPFYGEPKSKDLKLRLYEQOHCKLGLYSKEMNLGRNEKSVEAALPFFSRTRWIOSPNNKVLVLSGEAQNKGQCPYEFVNSKD
 WOEFKARVETSRSPSKKORILLQKFEDDSKFERALNOTRYVNRFLLCQFVADMWLTGKGKCRVFAANGQITLRLRGWLGWLXVRAENDRHALDAVV
 TVAKQGKITHVYKREKHNADFOKTIDKETGEWUHQKTHFYQPKDGFEEAOIPEKLRLLAELKLSRPEAVHEYVTPLFVSRSA
 WSYQGHMETVSKAKRDLGEVSVLVPLTQLKAKRLEAHKDOPAKAEPKYDKAGNRTOCQKVAVREQVQTKSV
 NGIADNAATWVYDVFYKQKDYKLYVPSWQVAKGIPDRAVVGQKQKDYKQYDKEEDWQKIDOSPFNSLJPNOLVETVTKARMFGYFASCHRTGHNHRQD
 GLEGIVYKTALESFQYQDELGKEFRPCRLKKRPPVRSRADPKKKRKRVEASRA
 YPYDVPDYAS

FIG. 15J

ICAM1 Zfp389 Core amino acid sequence; Nuclear Localization Sequence, Zinc Finger Motif, "HA" Epitope (SEQ ID NO: 151)

WYDŁUDZAS

४५

Myocd gRNAs Screening

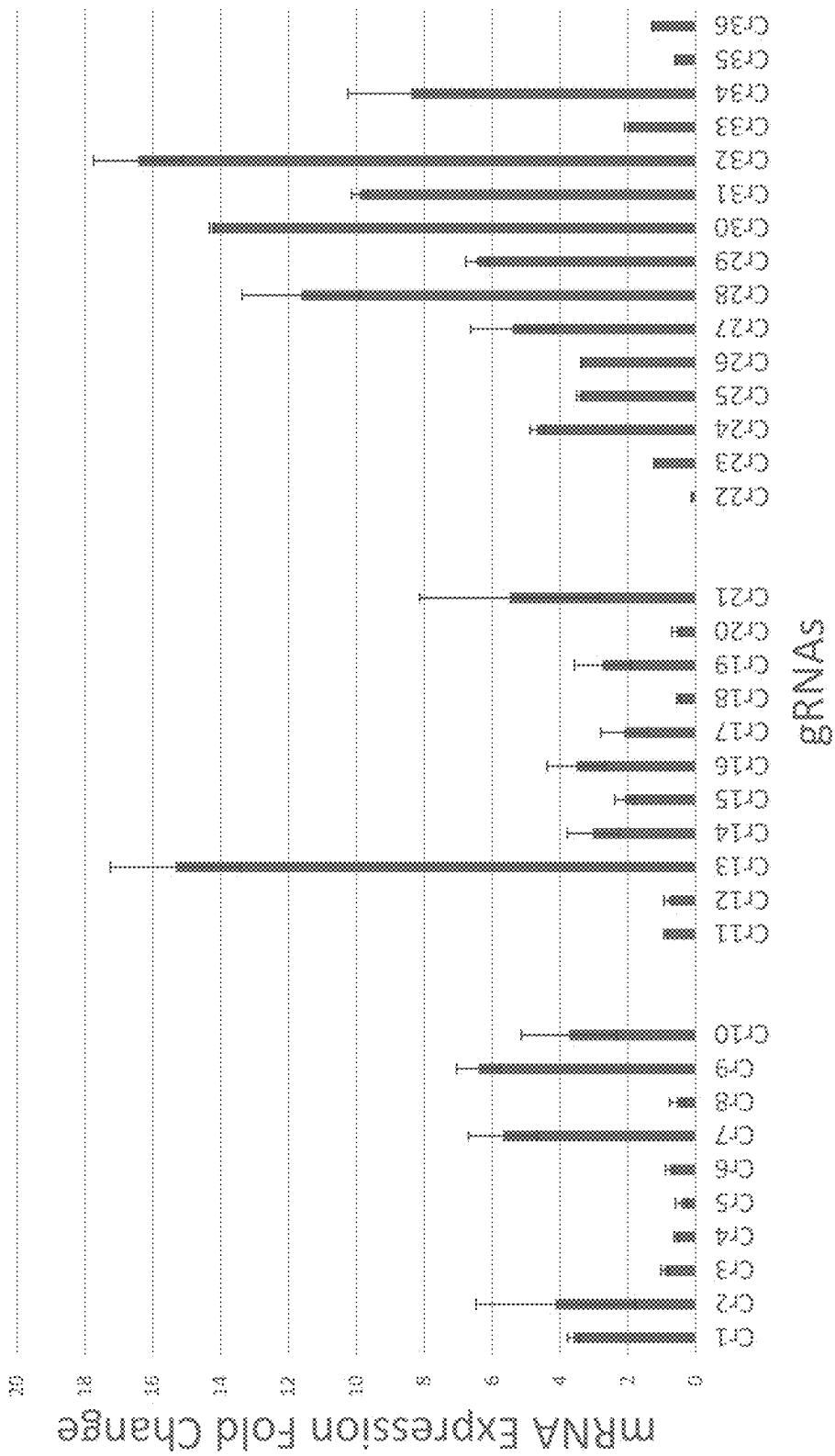


FIG. 17

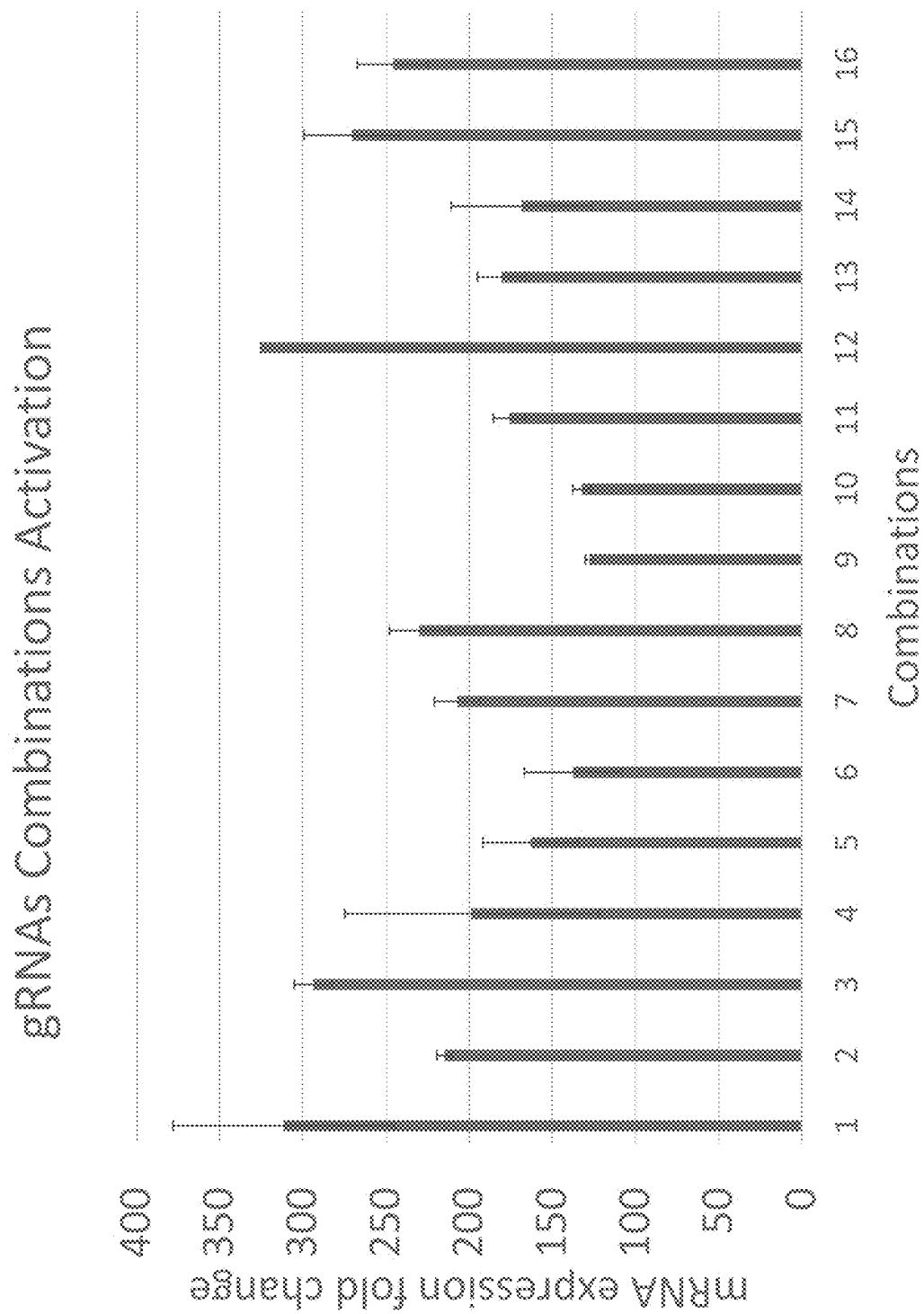
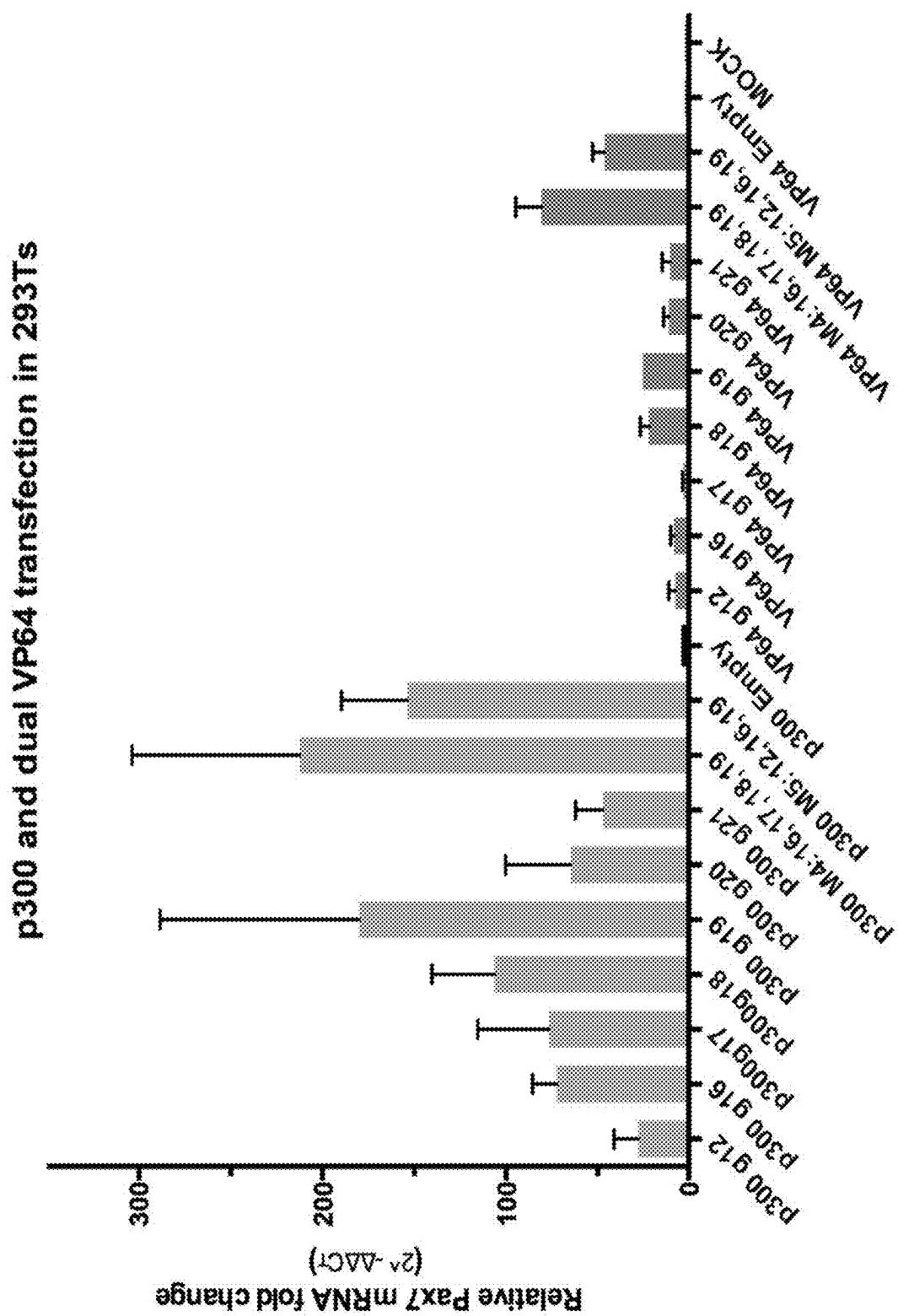


FIG. 18



١٣

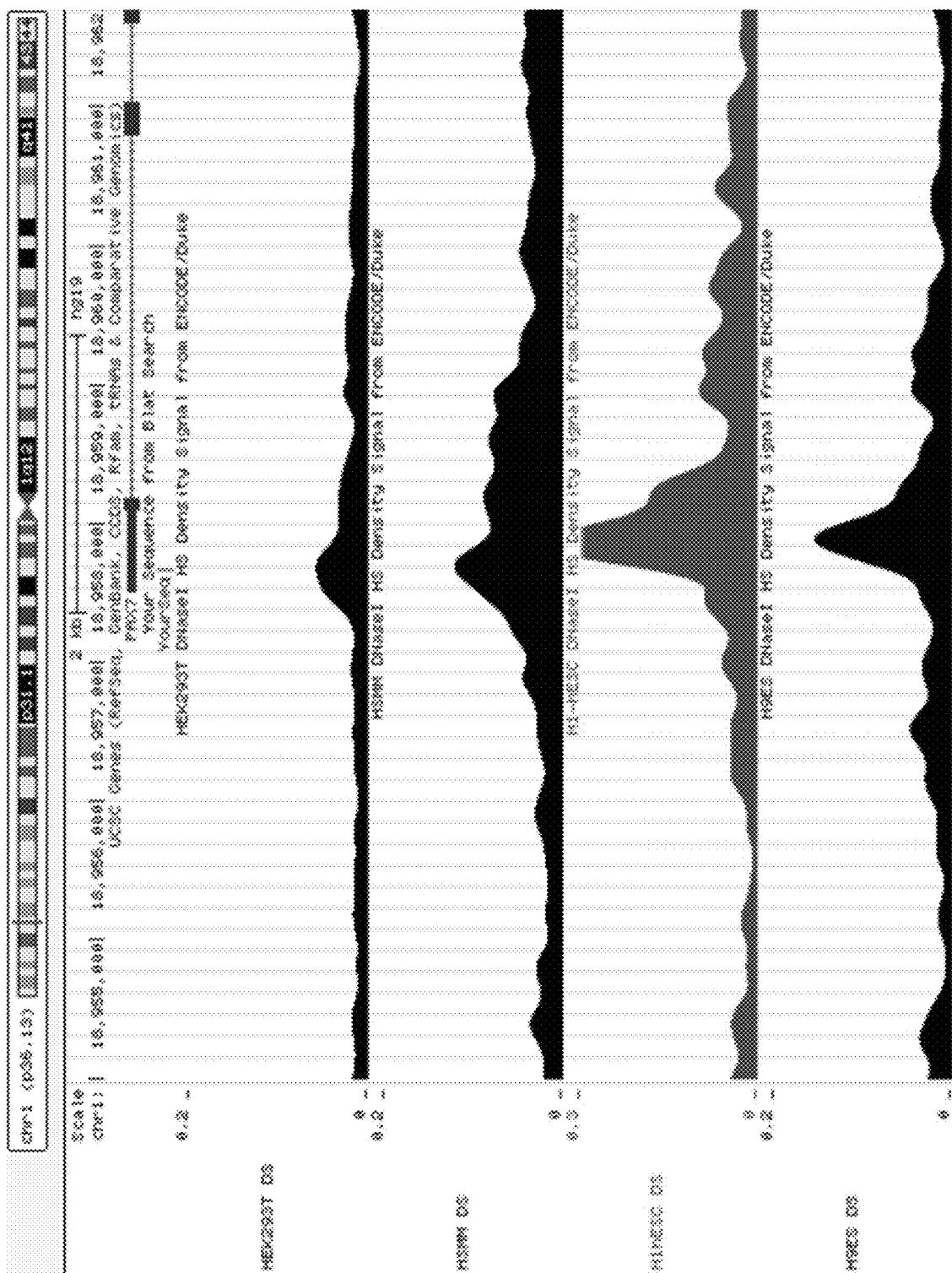


FIG. 20

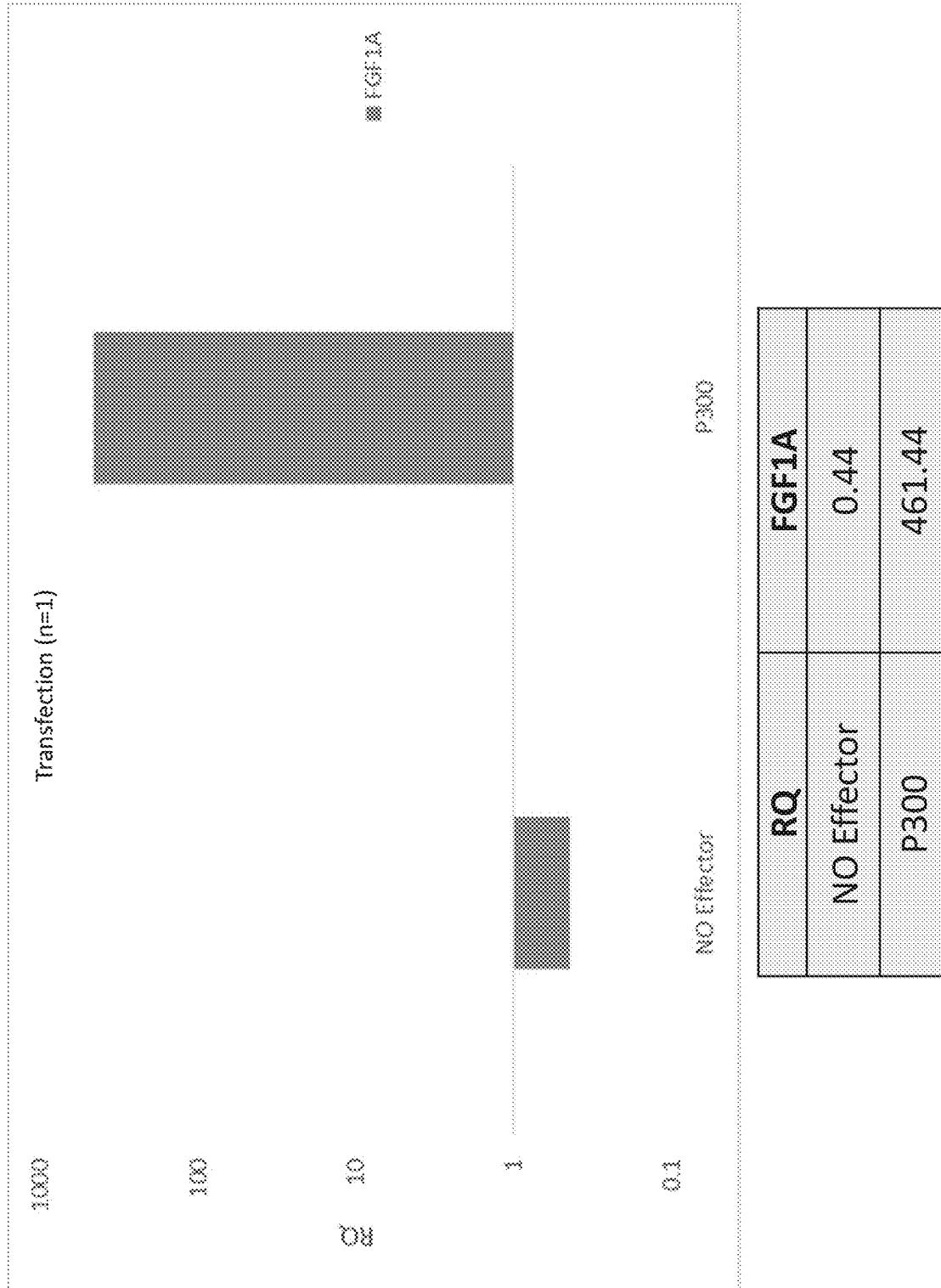
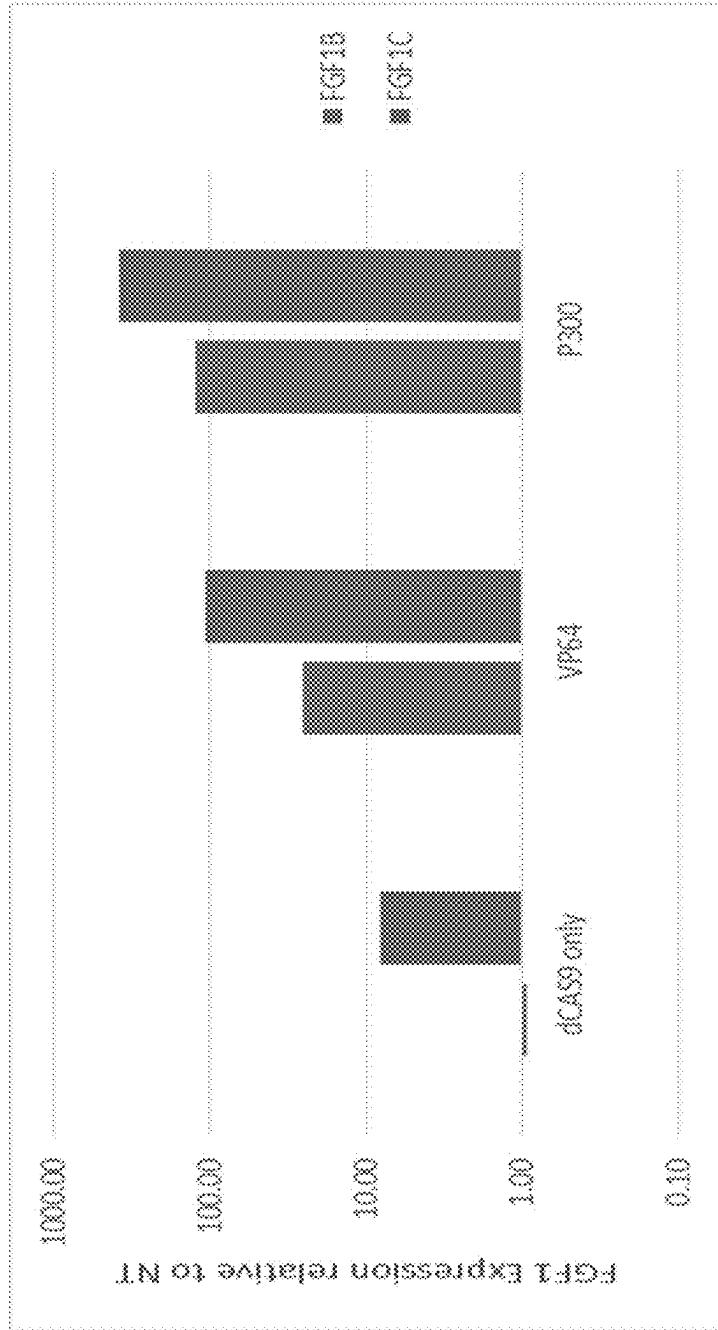


FIG. 21



RQ	FGFR1B	FGFR1C
dCASS9 only	0.94	8.11
VP64	25.50	107.54
P300	125.29	388.36

FIG. 22

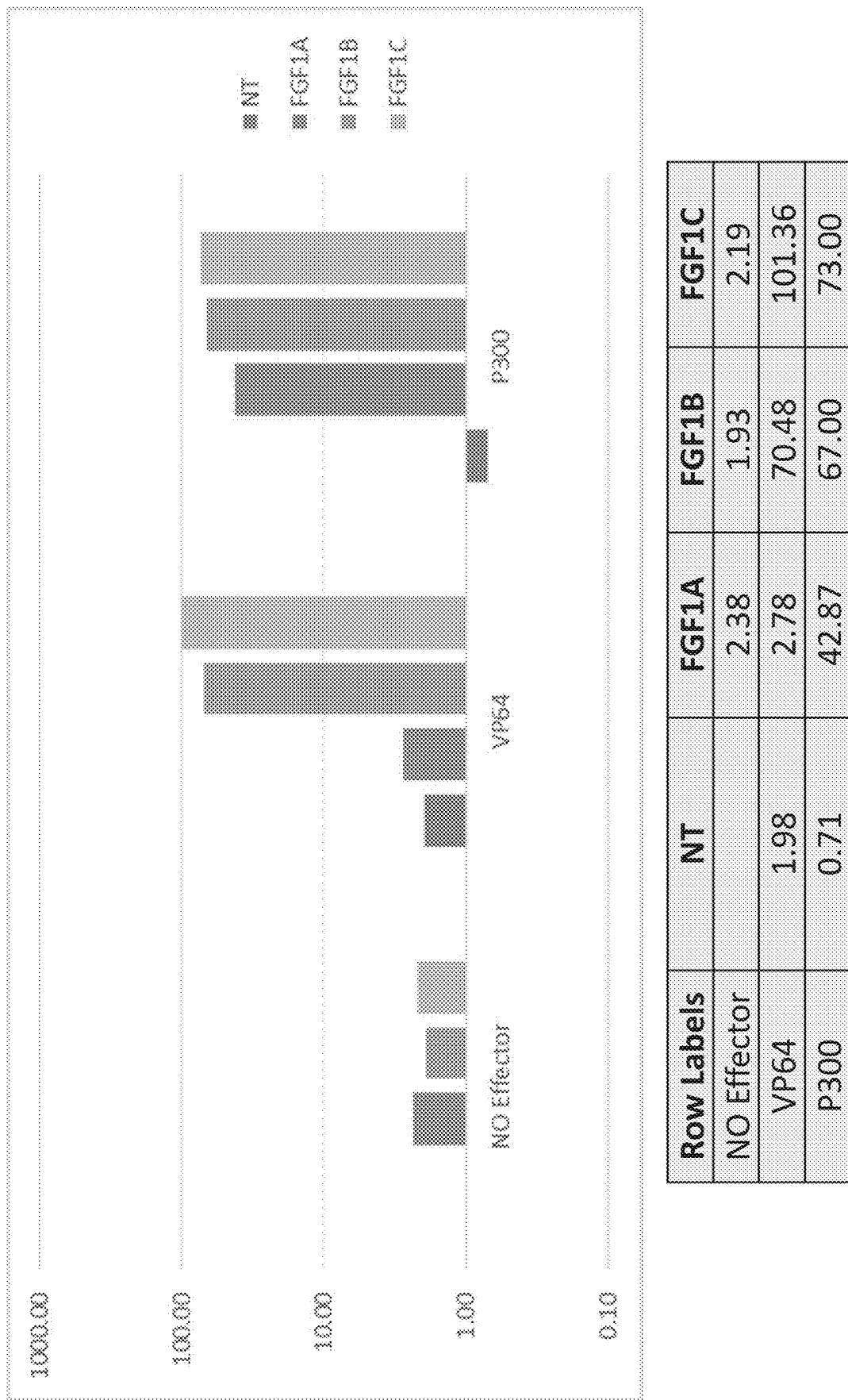


FIG. 23

COMPOSITIONS AND METHODS FOR EPIGENOME EDITING

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This patent application is a continuation of Ser. No. 17/471,935, filed Sep. 10, 2021, which is a divisional of Ser. No. 16/865,151, filed May 1, 2020, which is a divisional of U.S. patent application Ser. No. 15/549,842, filed Aug. 9, 2017, which is the U.S. national stage entry, under 35 U.S.C. § 371, of International Application Number PCT/US2016/017221, filed Feb. 9, 2016, which claims priority to U.S. Provisional Application No. 62/113,569, filed Feb. 9, 2015, the entire contents of each of which are incorporated herein by reference.

STATEMENT OF GOVERNMENT INTEREST

[0002] This invention was made with Government support under Federal Grant No. 1R01DA036865 awarded by the National Institutes of Health. The Government has certain rights to this invention.

SEQUENCE LISTING

[0003] The sequence listing is filed with the application in electronic format only and is incorporated by reference herein. The sequence listing XML file named “028193-9190-US04_Sequence Listing” was created on Apr. 4, 2025, and is 407,760 bytes in size.

TECHNICAL FIELD

[0004] The present disclosure is directed to CRISPR/Cas9-based gene activation systems and methods of using said systems.

BACKGROUND

[0005] The Human Genome Project was funded and pursued based on the premise that the sequencing of the human genome would reveal the genetic basis for complex diseases that have a strong inheritable component, including cardiovascular disease, neurodegenerative conditions, and metabolic diseases such as diabetes. It was believed that this information would lead to new drug targets for these widespread diseases. However, thousands of genome-wide association studies (GWAS) have shown that the genetic variation associated with these complex diseases does not occur within genes, but rather in intergenic regulatory regions that control the levels of particular genes. Similarly, approximately 20% of Mendelian disorders do not have a detectable coding mutation, suggesting that the causal mutation is in a gene regulatory element. Importantly, it is very difficult to assign functional roles to these regulatory elements as they often are located in distant locations from their target genes. Moreover, many genes and regulatory elements fall into each positive hit of each GWAS study. In fact, follow-up projects to the Human Genome Project, such as the NIH-funded Encyclopedia of DNA Elements (ENCODE) and the Roadmap Epigenomics Project, have identified millions of putative regulatory elements across the human genome for many human cell types and tissues.

[0006] A primary challenge of functional genomics is to develop technologies that directly and precisely manipulate genome function at individual loci. Projects such as

ENCODE and the Roadmap Epigenomics Project have identified millions of epigenetic marks across the human genome for many human cell types and tissues. Studying the function of those marks, however, has been largely limited to statistical associations with gene expression. Technologies for targeted direct manipulation of these epigenetic properties are necessary to transform such association-based findings into mechanistic principles of gene regulation. Such advances have the potential to benefit human health, as they could lead to gene therapies that modify the epigenetic code at targeted regions of the genome, strategies for regenerative medicine and disease modeling based on the epigenetic reprogramming of cell lineage specification, and the engineering of epigenome-specific drug screening platforms.

[0007] Manipulation of the epigenome is possible by treating cells with small molecule drugs, such as inhibitors of histone deacetylases or DNA methyltransferases, or differentiating cells into specific lineages. However, small molecule-based methods globally alter the epigenome and transcriptome, and are not suitable for targeting individual loci. Epigenome editing technologies, including the fusion of epigenome-modifying enzymes to programmable DNA-binding proteins such as zinc finger proteins and transcription activator-like effectors (TALEs), have been effective at achieving targeted DNA methylation, DNA hydroxymethylation, and histone demethylation, methylation, and deacetylation.

[0008] Fused to activation domains, such as oligomers of the herpes simplex viral protein 16 (VP16), dCas9 can function as a synthetic transcriptional regulator. However, limitations in the use of dCas9 activators remain, including the need for multiple activation domains or combinations of gRNAs to achieve high levels of gene induction by synergistic effects between activation domains. The conventional activator domains used in these engineered transcriptional factors, such as the VP16 tetramer VP64, function as a scaffold for recruiting multiple components of the preinitiation complex and do not have direct enzymatic function to specifically modulate the chromatin state. This indirect method of epigenetic remodeling does not allow for testing the role of specific epigenetic marks and may not be as potent as the direct programming of epigenetic states. There remains a need for the ability to target direct manipulation of epigenetic properties.

SUMMARY

[0009] The present invention is directed to a fusion protein comprising two heterologous polypeptide domains, wherein the first polypeptide domain comprises a Clustered Regularly Interspaced Short Palindromic Repeats associated (Cas) protein and the second polypeptide domain

[0010] The present invention is directed to a DNA targeting system comprising the fusion protein, described above, and at least one guide RNA (gRNA).

[0011] The present invention is directed to a method of activating gene expression of a target gene in a cell, the method comprising contacting the cell with a polynucleotide encoding a DNA targeting system, wherein the DNA targeting system comprises the fusion protein, described above, and at least one guide RNA (gRNA).

BRIEF DESCRIPTION OF THE DRAWINGS

[0012] FIGS. 1A-1C show that dCas9^{p300 Core} fusion protein activates transcription of endogenous genes from proxi-

mal promoter regions. FIG. 1A shows a schematic of dCas9 fusion proteins dCas9^{VP64} dCas9^{FL p300}, and dCas9^{p300 Core}. *Streptococcus pyogenes* dCas9 contains nuclease inactivating mutations D10A and H840A. The D1399 catalytic residue in the p300 HAT domain is indicated. FIG. 1B shows Western blot showing expression levels of dCas9 fusion proteins and GAPDH in co-transfected cells (full blot shown in FIG. 7C). FIG. 1C shows relative mRNA expression of IL1RN, MYOD, and OCT4, determined by qRT-PCR, by the indicated dCas9 fusion protein co-transfected with four gRNAs targeted to each promoter region (Tukey-test, *P-value <0.05, n=3 independent experiments each, error bars: s.e.m.). Numbers above bars indicate mean expression. FLAG, epitope tag; NLS, nuclear localization signal; HA, hemagglutinin epitope tag; CH, cysteine-histidine-rich region; Bd, bromodomain; HAT, histone acetyltransferase domain.

[0013] FIGS. 2A-2C show that dCas9^{p300 Core} fusion protein activates transcription of endogenous genes from distal enhancer regions. FIG. 2A shows relative MYOD mRNA production in cells co-transfected with a pool of gRNAs targeted to either the proximal or distal regulatory regions and dCas9^{VP64} or dCas9^{p300 Core}; promoter data from FIG. 1C (Tukey-test, *P-value <0.05 compared to mock-transfected cells, Tukey test †P-value <0.05 between dCas9^{p300 Core} and dCas9^{VP64}, n=3 independent experiments, error bars: s.e.m.). The human MYOD locus is schematically depicted with corresponding gRNA locations in red. CE, MyoD core enhancer; DRR, MyoD distal regulatory region. FIG. 2B shows relative OCT4mRNA production in cells co-transfected with a pool of gRNAs targeted to the proximal and distal regulatory regions and dCas9^{VP64} or dCas9^{p300 Core}; promoter data from FIG. 1C (Tukey-test, *P-value <0.05 compared to mock-transfected cells, Tukey test †P-value <0.05 between dCas9^{p300 Core} and dCas9^{VP64}, n=3 independent experiments, error bars: s.e.m.). The human OCT4 locus is schematically depicted with corresponding gRNA locations in red. DE, Oct4 distal enhancer; PE, Oct4 proximal enhancer. FIG. 2C shows the human β-globin locus is schematically depicted with approximate locations of the hypersensitive site 2 (HS2) enhancer region and downstream genes (HBE, HBG, HBD, and HBB). Corresponding HS2 gRNA locations are shown in red. Relative mRNA production from distal genes in cells co-transfected with four gRNAs targeted to the HS2 enhancer and the indicated dCas9 proteins. Note logarithmic y-axis and dashed red line indicating background expression (Tukey test among conditions for each β-globin gene, †P-value <0.05, n=3 independent experiments, error bars: s.e.m.). n.s., not significant.

[0014] FIGS. 3A-3C show that dCas9^{p300 Core} targeted transcriptional activation is specific and robust. FIGS. 3A-3C show MA plots generated from DEseq2 analysis of genome-wide RNA-seq data from HEK293T cells transiently co-transfected with dCas9^{VP64} (FIG. 3A) dCas9^{p300 Core}(FIG. 3B) or dCas9^{p300 Core} (D1399Y) (FIG. 3C) and four IL1RN promoter-targeting gRNAs compared to HEK293T cells transiently co-transfected with dCas9 and four IL1RN promoter-targeting gRNAs. mRNAs corresponding to IL1RN isoforms are shown in blue and circled in each of FIGS. 3A-3C. Red labeled points in FIGS. 3B and 3C correspond to off-target transcripts significantly enriched after multiple hypothesis testing (KDR, (FDR=1.4×10⁻³);

FAM49A, (FDR=0.04); p300, (FDR=1.7×10⁻⁴) in FIG. 3B; and p300, (FDR=4.4×10⁻¹⁰) in FIG. 3C.

[0015] FIGS. 4A-4D show that dCas9^{p300 Core} fusion protein acetylates chromatin at a targeted enhancer and corresponding downstream genes. FIG. 4A shows the region encompassing the human β-globin locus on chromosome 11 (5,304,000-5,268,000; GRCh37/hg19 assembly) is shown. HS2 gRNA target locations are indicated in red and ChIP-qPCR amplicon regions are depicted in black with corresponding green numbers. ENCODE/Broad Institute H3K27ac enrichment signal in K562 cells is shown for comparison. Magnified insets for the HS2 enhancer, HBE, and HBG1/2promoter regions are displayed below. FIGS. 4B-4D show H3K27ac ChIP-qPCR enrichment (relative to dCas9; red dotted line) at the HS2 enhancer, HBE promoter, and HBG1/2 promoters in cells co-transfected with four gRNAs targeted to the HS2 enhancer and the indicated dCas9 fusion protein. HBG ChIP amplicons 1 and 2 amplify redundant sequences at the HBG1 and HBG2 promoters (denoted by ‡). Tukey test among conditions for each ChIP-qPCR region, *P-value <0.05 (n=3 independent experiments, error bars: s.e.m.).

[0016] FIGS. 5A-5G show that dCas9^{p300 Core} fusion protein activates transcription of endogenous genes from regulatory regions with a single gRNA. Relative IL1RN (FIG. 5A), MYOD (FIG. 5B) or OCT4 (FIG. 5C) mRNA produced from cells co-transfected with dCas9^{p300 Core} or dCas9^{VP64} and gRNAs targeting respective promoters (n=3 independent experiments, error bars: s.e.m.). Relative MYOD (FIG. 5D) or OCT4 (FIG. 5E) mRNA produced from cells co-transfected with dCas9^{p300 Core} and indicated gRNAs targeting the indicated MYOD or OCT4 enhancers (n=3 independent experiments, error bars: s.e.m.). DRR, MYOD distal regulatory region; CE, MYOD core enhancer; PE, OCT4 proximal enhancer; DE, OCT4 distal enhancer. (Tukey test between dCas9^{p300 Core} and single OCT4 DE gRNAs compared to mock-transfected cells, *P-value <0.05, Tukey test among dCas9^{p300 Core} and OCT4 DE gRNAs compared to All, †P<0.05, n=3 independent experiments, error bars: s.e.m.). Relative HBE (FIG. 5F) or HBG (FIG. 5G) mRNA production in cells co-transfected with dCas9^{p300 Core} and the indicated gRNAs targeted to the HS2 enhancer (Tukey test between dCas9^{p300 Core} and single HS2 gRNAs compared to mock-transfected cells, *P-value <0.05, Tukey test among dCas9^{p300 Core} and HS2 single gRNAs compared to All, †P<0.05, n=3 independent experiments, error bars: s.e.m.). HS2, β-globin locus control region hypersensitive site 2; n.s., not significant using Tukey test.

[0017] FIGS. 6A-6H show that the p300 Core can be targeted to genomic loci by diverse programmable DNA-binding proteins. FIG. 6A shows schematic of the *Neisseria meningitidis* (Nm) dCas9 fusion proteins Nm-dCas9^{VP64} and Nm-dCas9^{p300 Core} *Neisseria meningitidis* dCas9 contains nuclease-inactivating mutations D16A, D587A, H588A, and N611A. FIG. 6B shows relative HBE mRNA in cells co-transfected with five individual or pooled (A-E) Nm gRNAs targeted to the HBE promoter and Nm-dCas9^{VP64} or Nm-dCas9^{p300 Core} FIGS. 6C-6D Relative HBE (FIG. 6C) or HBG (FIG. 6D) mRNA in cells co-transfected with five individual or pooled (A-E) Nm gRNAs targeted to the HS2 enhancer and Nm-dCas9^{VP64} or Nm-dCas9^{p300 Core} FIG. 6E shows schematic of TALEs with domains containing IL1RN-targeted repeat variable diresidues (Repeat Domain). FIG. 6F shows relative IL1RN mRNA in cells transfected

with individual or pooled (A-D) IL1RN TALE^{VP64} or IL1RN TALE^{p300 Core} encoding plasmids. FIG. 6G shows schematic of ZF fusion proteins with zinc finger helices 1-6 (F1-F6) targeting the ICAM1 promoter. FIG. 6H shows relative ICAM1 mRNA in cells transfected with ICAM1 ZF^{VP64} or ICAM1 ZF^{p300 Core} Tukey-test, *P-value <0.05 compared to mock-transfected control, n=3 independent experiments each, error bars: s.e.m. NLS, nuclear localization signal; HA, hemagglutinin tag; Bd, bromodomain; CH, cysteine-histidine-rich region; HAT, histone acetyltransferase domain.

[0018] FIGS. 7A-7C show dCas9^{p300 Core} mutant fusion protein activities. FIG. 7A shows schematic depiction of the WT dCas9^{p300 Core} fusion protein and p300 Core mutant derivatives. Relative locations of mutated amino acids are displayed as yellow bars within the p300 Core effector domain. FIG. 7B shows dCas9^{p300 Core} variants were transiently co-transfected with four IL1RN promoter gRNAs and were screened for hyperactivity¹ (amino acid 1645/1646 RR/EE and C1204R mutations) or hypoactivity (denoted by \$) via mRNA production from the IL1RN locus (top panel, n=2 independent experiments, error bars: s.e.m.). Experiments were performed in duplicate with one well used for RNA isolation and the other for western blotting to validate expression (bottom panels). The nitrocellulose membrane was cut and incubated with α -FLAG primary antibody (top, Sigma-Aldrich cat. #F7425) or α -GAPDH (bottom, Cell Signaling Technology cat. #14C10) then α -Rabbit HRP secondary antibody (Sigma-Aldrich cat. #A6154). FIG. 7C shows full membranes from western blot shown in main text (FIG. 1). The nitrocellulose membrane was cut and incubated with α -FLAG primary antibody (top, Sigma-Aldrich cat. #F7425) or α -GAPDH (bottom, Cell Signaling Technology cat. #14C10) then α -Rabbit HRP secondary antibody (Sigma-Aldrich cat. #A6154). Membrane was imaged for the indicated durations after careful re-alignment of trimmed pieces.

[0019] FIG. 8 shows target gene activation is unaffected by overexpression of synthetic dCas9 fusion proteins.

[0020] FIGS. 9A-9E show a comparison of Sp. dCas9 and Nm. dCas9 gene induction from the HS2 enhancer with individual and pooled gRNAs. FIG. 9A shows schematic display of the human β -globin locus including *Streptococcus pyogenes* dCas9 (Sp. dCas9) and *Neisseria meningitidis* dCas9 (Nm. dCas9) gRNA locations at the HS2 enhancer. Layered transcription profiles scaled to a vertical viewing range of 8 from nine ENCODE cell lines (GM12878, H1-hESC, HeLa-S3, HepG2, HSMM, HUVEC, K562, NHEK, and NHLF) is shown in addition to ENCODE p300 binding peaks in K562, A549 (EtOH 0.02), HeLa-S3, and SKN_SH_RA cell lines. An ENCODE HEK293T DNase hypersensitive site (HEK293T DHS) is shown in the HS2 Enhancer inset. FIGS. 9B-9E shows relative transcriptional induction of HBE, HBG, HBD, and HBD transcripts from single and pooled Sp. dCas9 gRNAs (A-D) or single and pooled Nm. dCas9 gRNAs (A-E) in response to co-transfection with Sp. dCas9^{p300 Core} or Nm. dCas9^{p300 Core} respectively. gRNAs are tiled for each dCas9 ortholog corresponding to their location in GRCh37/hg19. Gray dashed line indicates background expression level in transiently co-transfected HEK293T cells. Note shared logarithmic scale among FIGS. 9B-9E. Numbers above bars in FIGS. 9B-9E indicate mean expression (n=at least 3 independent experiments, error bars. s.e.m.).

[0021] FIG. 10 shows that dCas9^{VP64} and dCas9^{p300 Core} induce H3K27ac enrichment at IL1RN gRNA-targeted chromatin.

[0022] FIGS. 11A-11C show a direct comparison of VP64 and p300 Core effector domains between TALE and dCas9 programmable DNA binding proteins. FIG. 11A shows the GRCh37/hg19 region encompassing the IL1RN transcription start site is shown schematically along with IL1RN TALE binding sites and dCas9 IL1RN gRNA target sites. FIG. 11B shows direct comparison of IL1RN activation in HEK293T cells when transfected with individual or pooled (A-D) IL1RN TALE^{VP64} fusion proteins or when co-transfected with dCas9^{VP64} and individual or pooled (A-D) IL1RN-targeting gRNAs. FIG. 11C shows direct comparison of IL1RN activation in HEK293T cells when transfected with individual or pooled (A-D) IL1RN TALE^{p300 Core} fusion proteins or when co-transfected with dCas9^{p300 Core} and individual or pooled (A-D) IL1RN-targeting gRNAs. Note shared logarithmic scale between FIG. 11B and FIG. 11C. Numbers above bars in FIGS. 11B and 11C indicate mean values. Tukey test, *P-value <0.05, n=at least 3 independent experiments, error bars: s.e.m.

[0023] FIGS. 12A-12B show TALE and ZF fusion protein expression. FIG. 12A shows Western blotting was carried out on cells transiently transfected with individual or pooled IL1RN TALE proteins. Nitrocellulose membranes were cut and probed with α -HA primary antibody (1:1000 dilution in TBST+5% Milk, top, Covance cat. #MMS-101P) or α -GAPDH (bottom, Cell Signaling Technology cat. #14C10) then α -Mouse HRP (Santa Cruz, sc-2005) or α -Rabbit HRP (Sigma-Aldrich cat. #A6154) secondary antibody, respectively. FIG. 12B shows Western blotting was carried out on cells transiently transfected with ICAM1 ZF-effector proteins and nitrocellulose membranes were cut and probed with α -FLAG primary antibody (top, Sigma-Aldrich cat. #F7425) or α -GAPDH (bottom, Cell Signaling Technology cat. #14C10) then α -Rabbit HRP secondary antibody (Sigma-Aldrich cat. #A6154). Red asterisk indicates non-specific band.

[0024] FIGS. 13A-13B show that dCas9^{p300 Core} and dCas9^{VP64} do not display synergy in transactivation. FIG. 13A shows dCas9^{p300 Core} was co-transfected at a 1:1 mass ratio to PL-SIN-EF1 α -EGFP³ (GFP), dCas9, or dCas9^{VP64} with four IL1RN promoter gRNAs as indicated (n=2 independent experiments, error bars: s.e.m.). FIG. 13B shows dCas9^{p300 Core} was co-transfected at a 1:1 mass ratio to GFP, dCas9, or dCas9^{VP64} with four MYOD promoter gRNAs as indicated (n=2 independent experiments, error bars: s.e.m.). No significant differences were observed using Tukey's test (n.s.).

[0025] FIGS. 14A-14D show the underlying chromatin context of dCas9^{p300 Core} target loci. FIGS. 14A-14D show indicated loci along with associated *Streptococcus pyogenes* gRNAs used in this study at corresponding genomic locations in GRCh37/hg19. ENCODE HEK293T DNase hypersensitivity enrichment is shown (note changes in scale) along with regions of significant DNase hypersensitivity in HEK293T cells ("DHS"). In addition ENCODE master DNase clusters across 125 cell types are shown. Layered ENCODE H3K27ac and H3K4me3 enrichment across seven cell lines (GM12878, H1-hESC, HSMM, HUVEC, K562, NHEK, and NHLF) is also displayed and scaled to a vertical

viewing range of 50 and 150 respectively. Endogenous p300 binding profiles are also indicated for each locus and respective cell line.

[0026] FIG. 14E shows an overview of the information provided in FIGS. 14A-14D.

[0027] FIGS. 15A-15J show the amino acid sequences of dCas9 constructs.

[0028] FIG. 16 shows the amino acid sequences of ICAM1 Zinc Finger¹⁰ effectors.

[0029] FIG. 17 shows gRNA design and screening.

[0030] FIG. 18 shows gRNA combination activation.

[0031] FIG. 19 shows Pax7 guide screening in 293 Ts.

[0032] FIG. 20 shows that gRNA19 localizes to a DHS.

[0033] FIG. 21 shows the relative quantity of FGF1A mRNA in 293 Ts with or without dCas9^{p300 Core}.

[0034] FIG. 22 shows expression levels of FGF1B and FGF1C in 293 Ts with dCas9^{p300 Core} dCas9^{VP64} or dCas9 alone.

[0035] FIG. 23 shows expression levels of FGF1A, FGF1B, and FGF1C in 293 Ts with dCas9^{p300 Core} dCas9^{VP64} or dCas9 alone.

DETAILED DESCRIPTION

[0036] Disclosed herein are CRISPR/Cas9-based gene activation systems and methods of using said systems. The systems provide an easily programmable approach to facilitate robust control of the epigenome and downstream gene expression. The CRISPR/Cas9-based gene activation system includes a CRISPR/Cas9-based acetyltransferase, which is a fusion protein of a Cas9 protein and a protein having histone acetyltransferase activity, such as the catalytic histone acetyltransferase (HAT) core domain of the human E1A-associated protein p300. The Cas9 protein may not have nuclelease activity. An example of a Cas9 protein where the nuclelease activity has been abolished is dCas9. Recruitment of the acetyltransferase function by dCas9 and a gRNA to the genomic target site allow direct modulation of epigenetic structure, and thus provide an effective means of gene activation.

[0037] The disclosed CRISPR/Cas9-based acetyltransferase catalyzes acetylation of histone H3 lysine 27 at its target sites, leading to robust transcriptional activation of target genes from promoters and both proximal and distal enhancers. As disclosed herein, gene activation by these targeted acetyltransferases is highly specific across the genome. The CRISPR/Cas9-based acetyltransferase, which can be targeted to any site in the genome, is uniquely capable of activating distal regulatory elements. In contrast to conventional dCas9-based activators, the CRISPR/Cas9-based acetyltransferase effectively activates genes from enhancer regions and with individual or single guide RNAs.

1. Definitions

[0038] The terms “comprise(s),” “include(s),” “having,” “has,” “can,” “contain(s),” and variants thereof, as used herein, are intended to be open-ended transitional phrases, terms, or words that do not preclude the possibility of additional acts or structures. The singular forms “a,” “and” and “the” include plural references unless the context clearly dictates otherwise. The present disclosure also contemplates other embodiments “comprising,” “consisting of” and “consisting essentially of,” the embodiments or elements presented herein, whether explicitly set forth or not.

[0039] For the recitation of numeric ranges herein, each intervening number there between with the same degree of precision is explicitly contemplated. For example, for the range of 6-9, the numbers 7 and 8 are contemplated in addition to 6 and 9, and for the range 6.0-7.0, the number 6.0, 6.1, 6.2, 6.3, 6.4, 6.5, 6.6, 6.7, 6.8, 6.9, and 7.0 are explicitly contemplated.

[0040] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art. In case of conflict, the present document, including definitions, will control. Preferred methods and materials are described below, although methods and materials similar or equivalent to those described herein can be used in practice or testing of the present invention. All publications, patent applications, patents and other references mentioned herein are incorporated by reference in their entirety. The materials, methods, and examples disclosed herein are illustrative only and not intended to be limiting.

[0041] “Adeno-associated virus” or “AAV” as used interchangeably herein refers to a small virus belonging to the genus *Dependovirus* of the Parvoviridae family that infects humans and some other primate species. AAV is not currently known to cause disease and consequently the virus causes a very mild immune response.

[0042] “Chromatin” as used herein refers to an organized complex of chromosomal DNA associated with histones.

[0043] “Cis-regulatory elements” or “CREs” as used interchangeably herein refers to regions of non-coding DNA which regulate the transcription of nearby genes. CREs are found in the vicinity of the gene, or genes, they regulate. CREs typically regulate gene transcription by functioning as binding sites for transcription factors. Examples of CREs include promoters and enhancers.

[0044] “Clustered Regularly Interspaced Short Palindromic Repeats” and “CRISPRs”, as used interchangeably herein refers to loci containing multiple short direct repeats that are found in the genomes of approximately 40% of sequenced bacteria and 90% of sequenced archaea.

[0045] “Coding sequence” or “encoding nucleic acid” as used herein means the nucleic acids (RNA or DNA molecule) that comprise a nucleotide sequence which encodes a protein. The coding sequence can further include initiation and termination signals operably linked to regulatory elements including a promoter and polyadenylation signal capable of directing expression in the cells of an individual or mammal to which the nucleic acid is administered. The coding sequence may be codon optimize.

[0046] “Complement” or “complementary” as used herein means a nucleic acid can mean Watson-Crick (e.g., A-T/U and C-G) or Hoogsteen base pairing between nucleotides or nucleotide analogs of nucleic acid molecules. “Complementarity” refers to a property shared between two nucleic acid sequences, such that when they are aligned antiparallel to each other, the nucleotide bases at each position will be complementary.

[0047] “Endogenous gene” as used herein refers to a gene that originates from within an organism, tissue, or cell. An endogenous gene is native to a cell, which is in its normal genomic and chromatin context, and which is not heterologous to the cell. Such cellular genes include, e.g., animal genes, plant genes, bacterial genes, protozoal genes, fungal genes, mitochondrial genes, and chloroplastic genes.

[0048] “Enhancer” as used herein refers to non-coding DNA sequences containing multiple activator and repressor binding sites. Enhancers range from 200 bp to 1 kb in length and may be either proximal, 5' upstream to the promoter or within the first intron of the regulated gene, or distal, in introns of neighboring genes or intergenic regions far away from the locus. Through DNA looping, active enhancers contact the promoter independently of the core DNA binding motif promoter specificity. 4 to 5 enhancers may interact with a promoter. Similarly, enhancers may regulate more than one gene without linkage restriction and may “skip” neighboring genes to regulate more distant ones. Transcriptional regulation may involve elements located in a chromosome different to one where the promoter resides. Proximal enhancers or promoters of neighboring genes may serve as platforms to recruit more distal elements.

[0049] “Fusion protein” as used herein refers to a chimeric protein created through the joining of two or more genes that originally coded for separate proteins. The translation of the fusion gene results in a single polypeptide with functional properties derived from each of the original proteins.

[0050] “Genetic construct” as used herein refers to the DNA or RNA molecules that comprise a nucleotide sequence that encodes a protein. The coding sequence includes initiation and termination signals operably linked to regulatory elements including a promoter and polyadenylation signal capable of directing expression in the cells of the individual to whom the nucleic acid molecule is administered. As used herein, the term “expressible form” refers to gene constructs that contain the necessary regulatory elements operable linked to a coding sequence that encodes a protein such that when present in the cell of the individual, the coding sequence will be expressed.

[0051] “Histone acetyltransferases” or “HATs” are used interchangeably herein refers to enzymes that acetylate conserved lysine amino acids on histone proteins by transferring an acetyl group from acetyl CoA to form ε-N-acetyllysine. DNA is wrapped around histones, and, by transferring an acetyl group to the histones, genes can be turned on and off. In general, histone acetylation increases gene expression as it is linked to transcriptional activation and associated with euchromatin. Histone acetyltransferases can also acetylate non-histone proteins, such as nuclear receptors and other transcription factors to facilitate gene expression.

[0052] “Identical” or “identity” as used herein in the context of two or more nucleic acids or polypeptide sequences means that the sequences have a specified percentage of residues that are the same over a specified region. The percentage may be calculated by optimally aligning the two sequences, comparing the two sequences over the specified region, determining the number of positions at which the identical residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the specified region, and multiplying the result by 100 to yield the percentage of sequence identity. In cases where the two sequences are of different lengths or the alignment produces one or more staggered ends and the specified region of comparison includes only a single sequence, the residues of single sequence are included in the denominator but not the numerator of the calculation. When comparing DNA and RNA, thymine (T) and uracil (U) may be considered equiva-

lent. Identity may be performed manually or by using a computer sequence algorithm such as BLAST or BLAST 2.0.

[0053] “Nucleic acid” or “oligonucleotide” or “polynucleotide” as used herein means at least two nucleotides covalently linked together. The depiction of a single strand also defines the sequence of the complementary strand. Thus, a nucleic acid also encompasses the complementary strand of a depicted single strand. Many variants of a nucleic acid may be used for the same purpose as a given nucleic acid. Thus, a nucleic acid also encompasses substantially identical nucleic acids and complements thereof. A single strand provides a probe that may hybridize to a target sequence under stringent hybridization conditions. Thus, a nucleic acid also encompasses a probe that hybridizes under stringent hybridization conditions.

[0054] Nucleic acids may be single stranded or double stranded, or may contain portions of both double stranded and single stranded sequence. The nucleic acid may be DNA, both genomic and cDNA, RNA, or a hybrid, where the nucleic acid may contain combinations of deoxyribo- and ribo-nucleotides, and combinations of bases including uracil, adenine, thymine, cytosine, guanine, inosine, xanthine hypoxanthine, isocytosine and isoguanine. Nucleic acids may be obtained by chemical synthesis methods or by recombinant methods.

[0055] “Operably linked” as used herein means that expression of a gene is under the control of a promoter with which it is spatially connected. A promoter may be positioned 5' (upstream) or 3' (downstream) of a gene under its control. The distance between the promoter and a gene may be approximately the same as the distance between that promoter and the gene it controls in the gene from which the promoter is derived. As is known in the art, variation in this distance may be accommodated without loss of promoter function. [0056] “p300 protein,” “EP300,” or “E1A binding protein p300” as used interchangeably herein refers to the adenovirus E1A-associated cellular p300 transcriptional co-activator protein encoded by the EP300 gene. p300 is a highly conserved acetyltransferase involved in a wide range of cellular processes. p300 functions as a histone acetyltransferase that regulates transcription via chromatin remodeling and is involved with the processes of cell proliferation and differentiation.

[0056] “Promoter” as used herein means a synthetic or naturally-derived molecule which is capable of conferring, activating or enhancing expression of a nucleic acid in a cell. A promoter may comprise one or more specific transcriptional regulatory sequences to further enhance expression and/or to alter the spatial expression and/or temporal expression of same. A promoter may also comprise distal enhancer or repressor elements, which may be located as much as several thousand base pairs from the start site of transcription. A promoter may be derived from sources including viral, bacterial, fungal, plants, insects, and animals. A promoter may regulate the expression of a gene component constitutively, or differentially with respect to cell, the tissue or organ in which expression occurs or, with respect to the developmental stage at which expression occurs, or in response to external stimuli such as physiological stresses, pathogens, metal ions, or inducing agents. Representative examples of promoters include the bacteriophage T7 promoter, bacteriophage T3 promoter, SP6 promoter, lac operator-promoter, tac promoter, SV40 late promoter, SV40 early

promoter, RSV-LTR promoter, CMV IE promoter, SV40 early promoter or SV40 late promoter and the CMV IE promoter.

[0057] “Target enhancer” as used herein refers to enhancer that is targeted by a gRNA and CRISPR/Cas9-based gene activation system. The target enhancer may be within the target region.

[0058] “Target gene” as used herein refers to any nucleotide sequence encoding a known or putative gene product. The target gene includes the regulatory regions, such as the promoter and enhancer regions, the transcribed regions, which include the coding regions, and other function sequence regions.

[0059] “Target region” as used herein refers to a cis-regulatory region or a trans-regulatory region of a target gene to which the guide RNA is designed to recruit the CRISPR/Cas9-based gene activation system to modulate the epigenetic structure and allow the activation of gene expression of the target gene.

[0060] “Target regulatory element” as used herein refers to a regulatory element that is targeted by a gRNA and CRISPR/Cas9-based gene activation system. The target regulatory element may be within the target region.

[0061] “Transcribed region” as used herein refers to the region of DNA that is transcribed into single-stranded RNA molecule, known as messenger RNA, resulting in the transfer of genetic information from the DNA molecule to the messenger RNA. During transcription, RNA polymerase reads the template strand in the 3' to 5' direction and synthesizes the RNA from 5' to 3'. The mRNA sequence is complementary to the DNA strand.

[0062] “Transcriptional Start Site” or “TSS” as used interchangeably herein refers to the first nucleotide of a transcribed DNA sequence where RNA polymerase begins synthesizing the RNA transcript.

[0063] “Transgene” as used herein refers to a gene or genetic material containing a gene sequence that has been isolated from one organism and is introduced into a different organism. This non-native segment of DNA may retain the ability to produce RNA or protein in the transgenic organism, or it may alter the normal function of the transgenic organism’s genetic code. The introduction of a transgene has the potential to change the phenotype of an organism.

[0064] “Trans-regulatory elements” as used herein refers to regions of non-coding DNA which regulate the transcription of genes distant from the gene from which they were transcribed. Trans-regulatory elements may be on the same or different chromosome from the target gene.

[0065] “Variant” used herein with respect to a nucleic acid means (i) a portion or fragment of a referenced nucleotide sequence; (ii) the complement of a referenced nucleotide sequence or portion thereof; (iii) a nucleic acid that is substantially identical to a referenced nucleic acid or the complement thereof; or (iv) a nucleic acid that hybridizes under stringent conditions to the referenced nucleic acid, complement thereof, or a sequences substantially identical thereto.

[0066] “Variant” with respect to a peptide or polypeptide that differs in amino acid sequence by the insertion, deletion, or conservative substitution of amino acids, but retain at least one biological activity. Variant may also mean a protein with an amino acid sequence that is substantially identical to a referenced protein with an amino acid sequence that retains at least one biological activity. A conservative sub-

stitution of an amino acid, i.e., replacing an amino acid with a different amino acid of similar properties (e.g., hydrophilicity, degree and distribution of charged regions) is recognized in the art as typically involving a minor change. These minor changes may be identified, in part, by considering the hydrophilicity index of amino acids, as understood in the art. Kyte et al., *J. Mol. Biol.* 157:105-132 (1982). The hydrophilicity index of an amino acid is based on a consideration of its hydrophobicity and charge. It is known in the art that amino acids of similar hydrophilicity indexes may be substituted and still retain protein function. In one aspect, amino acids having hydrophilicity indexes of 2 are substituted. The hydrophilicity of amino acids may also be used to reveal substitutions that would result in proteins retaining biological function. A consideration of the hydrophilicity of amino acids in the context of a peptide permits calculation of the greatest local average hydrophilicity of that peptide. Substitutions may be performed with amino acids having hydrophilicity values within ±2 of each other. Both the hydrophobicity index and the hydrophilicity value of amino acids are influenced by the particular side chain of that amino acid. Consistent with that observation, amino acid substitutions that are compatible with biological function are understood to depend on the relative similarity of the amino acids, and particularly the side chains of those amino acids, as revealed by the hydrophobicity, hydrophilicity, charge, size, and other properties.

[0067] “Vector” as used herein means a nucleic acid sequence containing an origin of replication. A vector may be a viral vector, bacteriophage, bacterial artificial chromosome or yeast artificial chromosome. A vector may be a DNA or RNA vector. A vector may be a self-replicating extrachromosomal vector, and preferably, is a DNA plasmid. For example, the vector may encode a CRISPR/Cas9-based acetyltransferase having an amino acid sequence of SEQ ID NO: 140, 141, or 149 and/or at least one gRNA nucleotide sequence of any one of SEQ ID NOs: 23-73, 188-223, or 224-254.

2. CRISPR/Cas9-Based Gene Activation System

[0068] Provided herein are CRISPR/Cas9-based gene activation systems for use in activating gene expression of a target gene. The CRISPR/Cas9-based gene activation system includes a fusion protein of a Cas9 protein that does not have nuclease activity, such as dCas9, and a histone acetyltransferase or histone acetyltransferase effector domain. Histone acetylation, carried out by histone acetyltransferases (HATs), plays a fundamental role in regulating chromatin dynamics and transcriptional regulation. The histone acetyltransferase protein releases DNA from its heterochromatin state and allows for continued and robust gene expression by the endogenous cellular machinery. The recruitment of an acetyltransferase by dCas9 to a genomic target site may directly modulate epigenetic structure.

[0069] The CRISPR/Cas9-based gene activation system may catalyze acetylation of histone H3 lysine 27 at its target sites, leading to robust transcriptional activation of target genes from promoters and proximal and distal enhancers. The CRISPR/Cas9-based gene activation system is highly specific and may be guided to the target gene using as few as one guide RNA. The CRISPR/Cas9-based gene activation system may activate the expression of one gene or a family of genes by targeting enhancers at distant locations in the genome.

a) CRISPR System

[0070] The CRISPR system is a microbial nuclease system involved in defense against invading phages and plasmids that provides a form of acquired immunity. The CRISPR loci in microbial hosts contain a combination of CRISPR-associated (Cas) genes as well as non-coding RNA elements capable of programming the specificity of the CRISPR-mediated nucleic acid cleavage. Short segments of foreign DNA, called spacers, are incorporated into the genome between CRISPR repeats, and serve as a ‘memory’ of past exposures. Cas9 forms a complex with the 3' end of the single guide RNA (“sgRNA”), and the protein-RNA pair recognizes its genomic target by complementary base pairing between the 5' end of the sgRNA sequence and a predefined 20 bp DNA sequence, known as the protospacer. This complex is directed to homologous loci of pathogen DNA via regions encoded within the CRISPR RNA (“crRNA”), i.e., the protospacers, and protospacer-adjacent motifs (PAMs) within the pathogen genome. The non-coding CRISPR array is transcribed and cleaved within direct repeats into short crRNAs containing individual spacer sequences, which direct Cas nucleases to the target site (protospacer). By simply exchanging the 20 bp recognition sequence of the expressed chimeric sgRNA, the Cas9 nuclease can be directed to new genomic targets. CRISPR spacers are used to recognize and silence exogenous genetic elements in a manner analogous to RNAi in eukaryotic organisms.

[0071] Three classes of CRISPR systems (Types I, II and III effector systems) are known. The Type II effector system carries out targeted DNA double-strand break in four sequential steps, using a single effector enzyme, Cas9, to cleave dsDNA. Compared to the Type I and Type III effector systems, which require multiple distinct effectors acting as a complex, the Type II effector system may function in alternative contexts such as eukaryotic cells. The Type II effector system consists of a long pre-crRNA, which is transcribed from the spacer-containing CRISPR locus, the Cas9 protein, and a tracrRNA, which is involved in pre-crRNA processing. The tracrRNAs hybridize to the repeat regions separating the spacers of the pre-crRNA, thus initiating dsRNA cleavage by endogenous RNase III. This cleavage is followed by a second cleavage event within each spacer by Cas9, producing mature crRNAs that remain associated with the tracrRNA and Cas9, forming a Cas9:crRNA-tracrRNA complex.

[0072] An engineered form of the Type II effector system of *Streptococcus pyogenes* was shown to function in human cells for genome engineering. In this system, the Cas9 protein was directed to genomic target sites by a synthetically reconstituted “guide RNA” (“gRNA”, also used interchangeably herein as a chimeric sgRNA, which is a crRNA-tracrRNA fusion that obviates the need for RNase III and crRNA processing in general).

[0073] The Cas9:crRNA-tracrRNA complex unwinds the DNA duplex and searches for sequences matching the crRNA to cleave. Target recognition occurs upon detection of complementarity between a “protospacer” sequence in the target DNA and the remaining spacer sequence in the crRNA. Cas9 mediates cleavage of target DNA if a correct protospacer-adjacent motif (PAM) is also present at the 3' end of the protospacer. For protospacer targeting, the sequence must be immediately followed by the protospacer-adjacent motif (PAM), a short sequence recognized by the

Cas9 nuclease that is required for DNA cleavage. Different Type II systems have differing PAM requirements. The *S. pyogenes* CRISPR system may have the PAM sequence for this Cas9 (SpCas9) as 5'-NRG-3', where R is either A or G, and characterized the specificity of this system in human cells. A unique capability of the CRISPR/Cas9 system is the straightforward ability to simultaneously target multiple distinct genomic loci by co-expressing a single Cas9 protein with two or more sgRNAs. For example, the *Streptococcus pyogenes* Type II system naturally prefers to use an “NGG” sequence, where “N” can be any nucleotide, but also accepts other PAM sequences, such as “NAG” in engineered systems (Hsu et al., *Nature Biotechnology* (2013) doi:10.1038/nbt.2647). Similarly, the Cas9 derived from *Neisseria meningitidis* (NmCas9) normally has a native PAM of NNNNGATT, but has activity across a variety of PAMs, including a highly degenerate NNNNGNNN PAM (Esvelt et al. *Nature Methods* (2013) doi:10.1038/nmeth.2681).

b) Cas9

[0074] The CRISPR/Cas9-based gene activation system may include a Cas9 protein or a Cas9 fusion protein. Cas9 protein is an endonuclease that cleaves nucleic acid and is encoded by the CRISPR loci and is involved in the Type II CRISPR system. The Cas9 protein may be from any bacterial or archaea species, such as *Streptococcus pyogenes*, *Streptococcus thermophiles*, or *Neisseria meningitidis*. The Cas9 protein may be mutated so that the nuclease activity is inactivated. In some embodiments, an inactivated Cas9 protein from *Streptococcus pyogenes* (iCas9, also referred to as “dCas9”; SEQ ID NO: 1) may be used. As used herein, “iCas9” and “dCas9” both refer to a Cas9 protein that has the amino acid substitutions D10A and H840A and has its nuclease activity inactivated. In some embodiments, an inactivated Cas9 protein from *Neisseria meningitidis*, such as NmCas9 having an amino acid sequence of SEQ ID NO: 10, may be used.

c) Histone Acetyltransferase (HAT) Protein

[0075] The CRISPR/Cas9-based gene activation system may include a histone acetyltransferase protein, such as a p300 protein, CREB binding protein (CBP; an analog of p300), GCN5, or PCAF, or fragment thereof. The p300 protein regulates the activity of many genes in tissues throughout the body. The p300 protein plays a role in regulating cell growth and division, prompting cells to mature and assume specialized functions (differentiate) and preventing the growth of cancerous tumors. The p300 protein may activate transcription by connecting transcription factors with a complex of proteins that carry out transcription in the cell’s nucleus. The p300 protein also functions as a histone acetyltransferase that regulates transcription via chromatin remodeling.

[0076] The histone acetyltransferase protein may include a human p300 protein or a fragment thereof. The histone acetyltransferase protein may include a wild-type human p300 protein or a mutant human p300 protein, or fragments thereof. The histone acetyltransferase protein may include the core lysine-acetyltransferase domain of the human p300 protein, i.e., the p300 HAT Core (also known as “p300 Core”). In some embodiments, the histone acetyltransferase protein includes an amino acid sequence of SEQ ID NO: 2 or 3.

i) dCas9^{p300 Core}

[0077] The CRISPR/Cas9-based gene activation system may include a histone acetylation effector domain. The histone acetylation effector domain may be the catalytic histone acetyltransferase (HAT) core domain of the human E1A-associated protein p300 (also referred to herein as “p300 Core”). In some embodiments, the p300 Core includes amino acids 1048-1664 of SEQ ID NO: 2 (i.e., SEQ ID NO: 3). In some embodiments, the CRISPR/Cas9-based gene activation system includes a dCas9^{p300 Core} fusion protein of SEQ ID NO: 141 or an Nm-dCas9^{p300 Core} fusion protein of SEQ ID NO: 149. The p300 Core acetylates lysine 27 on histone H3 (H3K27ac) and may provide H3K27ac enrichment.

[0078] The dCas9^{p300 Core} fusion protein is a potent and easily programmable tool to synthetically manipulate acetylation at targeted endogenous loci, leading to regulation of proximal and distal enhancer-regulated genes. The fusion of the catalytic core domain of p300 to dCas9 may result in substantially higher transactivation of downstream genes than the direct fusion of full-length p300 protein despite robust protein expression. The dCas9^{p300 Core} fusion protein may also exhibit an increased transactivation capacity relative to dCas9^{VPG4}, including in the context of the Nm-dCas9 scaffold, especially at distal enhancer regions, at which dCas9^{VPG4} displayed little, if any, measurable downstream transcriptional activity. Additionally, the dCas9^{p300 Core} displays precise and robust genome-wide transcriptional specificity. dCas9^{p300 Core} may be capable of potent transcriptional activation and co-enrichment of acetylation at promoters targeted by the genetically modified enhancer.

[0079] The dCas9^{p300 Core} may activate gene expression through a single gRNA that target and bind a promoters and/or a characterized enhancer. This technology also affords the ability to synthetically transactivate distal genes from putative and known regulatory regions and simplifies transactivation via the application of a single programmable effector and single target site. These capabilities allow multiplexing to target several promoters and/or enhancers simultaneously. The mammalian origin of p300 may provide advantages over virally-derived effector domains for in vivo applications by minimizing potential immunogenicity.

d) gRNA

[0080] The CRISPR/Cas9-based gene activation system may include at least one gRNA that targets a nucleic acid sequence. The gRNA provides the targeting of the CRISPR/Cas9-based gene activation system. The gRNA is a fusion of two noncoding RNAs: a crRNA and a tracrRNA. The sgRNA may target any desired DNA sequence by exchanging the sequence encoding a 20 bp protospacer which confers targeting specificity through complementary base pairing with the desired DNA target. gRNA mimics the naturally occurring crRNA:tracrRNA duplex involved in the Type II Effector system. This duplex, which may include, for example, a 42-nucleotide crRNA and a 75-nucleotide tracrRNA, acts as a guide for the Cas9.

[0081] The gRNA may target and bind a target region of a target gene. The target region may be a cis-regulatory region or trans-regulatory region of a target gene. In some embodiments, the target region is a distal or proximal cis-regulatory region of the target gene. The gRNA may target and bind a cis-regulatory region or trans-regulatory region of a target gene. In some embodiments, the gRNA may target and bind an enhancer region, a promoter region,

or a transcribed region of a target gene. For example, the gRNA may target and bind the target region is at least one of HS2 enhancer of the human β-globin locus, distal regulatory region (DRR) of the MYOD gene, core enhancer (CE) of the MYOD gene, proximal (PE) enhancer region of the OCT4 gene, or distal (DE) enhancer region of the OCT4 gene. In some embodiments, the target region may be a viral promoter, such as an HIV promoter.

[0082] The target region may include a target enhancer or a target regulatory element. In some embodiments, the target enhancer or target regulatory element controls the gene expression of several target genes. In some embodiments, the target enhancer or target regulatory element controls a cell phenotype that involves the gene expression of one or more target genes. In some embodiments, the identity of one or more of the target genes is known. In some embodiments, the identity of one or more of the target genes is unknown. The CRISPR/Cas9-based gene activation system allows the determination of the identity of these unknown genes that are involved in a cell phenotype. Examples of cell phenotypes include, but not limited to, T-cell phenotype, cell differentiation, such as hematopoietic cell differentiation, oncogenesis, immunomodulation, cell response to stimuli, cell death, cell growth, drug resistance, or drug sensitivity.

[0083] In some embodiments, at least one gRNA may target and bind a target enhancer or target regulatory element, whereby the expression of one or more genes is activated. For example, between 1 gene and 20 genes, between 1 gene and 15 genes, between 1 gene and 10 genes, between 1 gene and 5 genes, between 2 genes and 20 genes, between 2 genes and 15 genes, between 2 genes and 10 genes, between 2 genes and 5 genes, between 5 genes and 20 genes, between 5 genes and 15 genes, or between 5 genes and 10 genes are activated by at least one gRNA. In some embodiments, at least 1 gene, at least 2 genes, at least 3 genes, at least 4 genes, at least 5 gene, at least 6 genes, at least 7 genes, at least 8 genes, at least 9 gene, at least 10 genes, at least 11 genes, at least 12 genes, at least 13 gene, at least 14 genes, at least 15 genes, or at least 20 genes are activated by at least one gRNA.

[0084] The CRISPR/Cas9-based gene activation system may activate genes at both proximal and distal locations relative the transcriptional start site (TSS). The CRISPR/Cas9-based gene activation system may target a region that is at least about 1 base pair to about 100,000 base pairs, at least about 100 base pairs to about 100,000 base pairs, at least about 250 base pairs to about 100,000 base pairs, at least about 500 base pairs to about 100,000 base pairs, at least about 1,000 base pairs to about 100,000 base pairs, at least about 2,000 base pairs to about 100,000 base pairs, at least about 5,000 base pairs to about 100,000 base pairs, at least about 10,000 base pairs to about 100,000 base pairs, at least about 20,000 base pairs to about 100,000 base pairs, at least about 50,000 base pairs to about 100,000 base pairs, at least about 75,000 base pairs to about 100,000 base pairs, at least about 1 base pair to about 75,000 base pairs, at least about 100 base pairs to about 75,000 base pairs, at least about 250 base pairs to about 75,000 base pairs, at least about 500 base pairs to about 75,000 base pairs, at least about 1,000 base pairs to about 75,000 base pairs, at least about 2,000 base pairs to about 75,000 base pairs, at least about 5,000 base pairs to about 75,000 base pairs, at least about 10,000 base pairs to about 75,000 base pairs, at least about 20,000 base pairs to about 75,000 base pairs, at least

about 50,000 base pairs to about 75,000 base pairs, at least about 1 base pair to about 50,000 base pairs, at least about 100 base pairs to about 50,000 base pairs, at least about 250 base pairs to about 50,000 base pairs, at least about 500 base pairs to about 50,000 base pairs, at least about 1,000 base pairs to about 50,000 base pairs, at least about 2,000 base pairs to about 50,000 base pairs, at least about 5,000 base pairs to about 50,000 base pairs, at least about 10,000 base pairs to about 50,000 base pairs, at least about 20,000 base pairs to about 50,000 base pairs, at least about 1 base pair to about 25,000 base pairs, at least about 100 base pairs to about 25,000 base pairs, at least about 250 base pairs to about 25,000 base pairs, at least about 500 base pairs to about 25,000 base pairs, at least about 1,000 base pairs to about 25,000 base pairs, at least about 2,000 base pairs to about 25,000 base pairs, at least about 5,000 base pairs to about 25,000 base pairs, at least about 10,000 base pairs to about 25,000 base pairs, at least about 20,000 base pairs to about 25,000 base pairs, at least about 1 base pair to about 10,000 base pairs, at least about 100 base pairs to about 10,000 base pairs, at least about 250 base pairs to about 10,000 base pairs, at least about 500 base pairs to about 10,000 base pairs, at least about 1,000 base pairs to about 10,000 base pairs, at least about 2,000 base pairs to about 10,000 base pairs, at least about 5,000 base pairs to about 10,000 base pairs, at least about 10,000 base pairs to about 25,000 base pairs, at least about 1 base pair to about 5,000 base pairs, at least about 100 base pairs to about 5,000 base pairs, at least about 250 base pairs to about 5,000 base pairs, at least about 500 base pairs to about 5,000 base pairs, at least about 1,000 base pairs to about 5,000 base pairs, at least about 2,000 base pairs to about 5,000 base pairs, at least about 5,000 base pairs to about 5,000 base pairs, at least about 10,000 base pairs to about 5,000 base pairs, at least about 20,000 base pairs to about 5,000 base pairs, at least about 1 base pair to about 11,000 base pairs, at least about 20,000 base pairs, at least about 30,000 base pairs, at least about 46,000 base pairs, at least about 50,000 base pairs, at least about 54,000 base pairs, at least about 75,000 base pairs, or at least about 100,000 base pairs upstream from the TSS.

[0085] The CRISPR/Cas9-based gene activation system may target a region that is at least about 1 base pair to at least about 500 base pairs, at least about 1 base pair to at least about 250 base pairs, at least about 1 base pair to at least about 200 base pairs, at least about 1 base pair to at least about 100 base pairs, at least about 50 base pairs to at least about 500 base pairs, at least about 50 base pairs to at least about 250 base pairs at least about 50 base pairs to at least about 200 base pairs, at least about 50 base pairs to at least about 100 base pairs, at least about 100 base pairs to at least about 500 base pairs, at least about 100 base pairs to at least about 250 base pairs, or at least about 100 base pairs to at least about 200 base pairs downstream from the TSS. The CRISPR/Cas9-based gene activation system may target a region that is at least about 1 base pair, at least about 2 base pairs, at least about 3 base pairs, at least about 4 base pairs, at least about 5 base pairs, at least about 10 base pairs, at least about 15 base pairs, at least about 20 base pairs, at least about 25 base pairs, at least about 30 base pairs, at least about 40 base pairs, at least about 50 base pairs, at least about 60 base pairs, at least about 70 base pairs, at least

about 80 base pairs, at least about 90 base pairs, at least about 100 base pairs, at least about 110 base pairs, at least about 120, at least about 130, at least about 140 base pairs, at least about 150 base pairs, at least about 160 base pairs, at least about 170 base pairs, at least about 180 base pairs, at least about 190 base pairs, at least about 200 base pairs, at least about 210 base pairs, at least about 220, at least about 230, at least about 240 base pairs, or at least about 250 base pairs downstream from the TSS.

[0086] In some embodiments, the CRISPR/Cas9-based gene activation system may target and bind a target region that is on the same chromosome as the target gene but more than 100,000 base pairs upstream or more than 250 base pairs downstream from the TSS. In some embodiments, the CRISPR/Cas9-based gene activation system may target and bind a target region that is on a different chromosome from the target gene.

[0087] The CRISPR/Cas9-based gene activation system may use gRNA of varying sequences and lengths. The gRNA may comprise a complementary polynucleotide sequence of the target DNA sequence followed by NGG. The gRNA may comprise a “G” at the 5' end of the complementary polynucleotide sequence. The gRNA may comprise at least a 10 base pair, at least a 11 base pair, at least a 12 base pair, at least a 13 base pair, at least a 14 base pair, at least a 15 base pair, at least a 16 base pair, at least a 17 base pair, at least a 18 base pair, at least a 19 base pair, at least a 20 base pair, at least a 21 base pair, at least a 22 base pair, at least a 23 base pair, at least a 24 base pair, at least a 25 base pair, at least a 30 base pair, or at least a 35 base pair complementary polynucleotide sequence of the target DNA sequence followed by NGG. The gRNA may target at least one of the promoter region, the enhancer region or the transcribed region of the target gene. The gRNA may include a nucleic acid sequence of at least one of SEQ ID NOS: 23-73, 188-223, or 224-254.

[0088] The CRISPR/Cas9-based gene activation system may include at least 1 gRNA, at least 2 different gRNAs, at least 3 different gRNAs at least 4 different gRNAs, at least 5 different gRNAs, at least 6 different gRNAs, at least 7 different gRNAs, at least 8 different gRNAs, at least 9 different gRNAs, or at least 10 different gRNAs. The CRISPR/Cas9-based gene activation system may include between at least 1 gRNA to at least 10 different gRNAs, at least 1 gRNA to at least 8 different gRNAs, at least 1 gRNA to at least 4 different gRNAs, at least 2 gRNA to at least 10 different gRNAs, at least 2 different gRNAs to at least 4 different gRNAs, at least 4 gRNA to at least 10 different gRNAs, or at least 4 different gRNAs to at least 8 different gRNAs.

3. Target Genes

[0089] The CRISPR/Cas9-based gene activation system may be designed to target and activate the expression of any target gene. The target gene may be an endogenous gene, a transgene, or a viral gene in a cell line. In some embodiments, the target region is located on a different chromosome as the target gene. In some embodiments, the CRISPR/Cas9-based gene activation system may include more than 1 gRNA. In some embodiments, the CRISPR/Cas9-based gene activation system may include more than 1 different gRNAs. In some embodiments, the different gRNAs bind to different target regions. For example, the different gRNAs

may bind to target regions of different target genes and the expression of two or more target genes are activated.

[0090] In some embodiments, the CRISPR/Cas9-based gene activation system may activate between about one target gene to about ten target genes, about one target genes to about five target genes, about one target genes to about four target genes, about one target genes to about three target genes, about one target genes to about two target genes, about two target gene to about ten target genes, about two target genes to about five target genes, about two target genes to about four target genes, about two target genes to about three target genes, about three target genes to about ten target genes, about three target genes to about five target genes, or about three target genes to about four target genes. In some embodiments, the CRISPR/Cas9-based gene activation system may activate at least one target gene, at least two target genes, at least three target genes, at least four target genes, at least five target genes, or at least ten target genes. For example, the may target the hypersensitive site 2 (HS2) enhancer region of the human β -globin locus and activate downstream genes (HBE, HBG, HBD and HBB).

[0091] In some embodiments, the CRISPR/Cas9-based gene activation system induces the gene expression of a target gene by at least about 1 fold, at least about 2 fold, at least about 3 fold, at least about 4 fold, at least about 5 fold, at least about 6 fold, at least about 7 fold, at least about 8 fold, at least about 9 fold, at least about 10 fold, at least 15 fold, at least 20 fold, at least 30 fold, at least 40 fold, at least 50 fold, at least 60 fold, at least 70 fold, at least 80 fold, at least 90 fold, at least 100 fold, at least about 110 fold, at least 120 fold, at least 130 fold, at least 140 fold, at least 150 fold, at least 160 fold, at least 170 fold, at least 180 fold, at least 190 fold, at least 200 fold, at least about 300 fold, at least 400 fold, at least 500 fold, at least 600 fold, at least 700 fold, at least 800 fold, at least 900 fold, or at least 1000 fold compared to a control level of gene expression. A control level of gene expression of the target gene may be the level of gene expression of the target gene in a cell that is not treated with any CRISPR/Cas9-based gene activation system.

[0092] The target gene may be a mammalian gene. For example, the CRISPR/Cas9-based gene activation system may target a mammalian gene, such as IL1RN, MYOD1, OCT4, HBE, HBG, HBD, HBB, MYOCD (Myocardin), PAX7 (Paired box protein Pax-7), FGF (fibroblast growth factor-1) genes, such as FGF1A, FGF1B, and FGF1C. Other target genes include, but not limited to, Atf3, Axud1, Btg2, c-Fos, c-Jun, Cxcl1, Cxcl2, Edn1, Ereg, Fos, Gadd45b, Ier2, Ier3, Ifrd1, I11b, Il6, Irf1, Junb, Lif, Nfkbia, Nfkbiz, Ptgs2, S1c25a25, Sqstm1, Tieg, Tnf, Tnfaip3, Zfp36, Birc2, Ccl2, Ccl20, Ccl7, Cebpd, Ch25h, CSF1, Cx3cl1, Cxcl10, Cxcl5, Gch, Icam1, Ifi47, Ifngr2, Mmp10, Nfkbie, Npal1, p21, Relb, Ripk2, Rnd1, Slpr3, Stx11, Tgtp, Tlr2, Timem140, Tnfaip2, Tnfrsf6, Vcam1, 1110004C05Rik (GenBank accession number BC010291), Abca1, AI561871 (GenBank accession number B1143915), AI882074 (GenBank accession number BB730912), Arts1, AW049765 (GenBank accession number BC026642.1), C3, Casp4, Cel5, Ccl9, Cdsn, Enpp2, Gbp2, H2-D1, H2-K, H2-L, Ifit1, Ii, Il13ra1, Il1rl1, Lcn2, Lhfpl2, LOC677168 (GenBank accession number AK019325), Mmp13, Mmp3, Mt2, Nafl, Ppicap, Prnd, Psmb10, Saa3, Serpina3g, Serpinfl, Sod3, Stat1, Tapbp, U90926 (GenBank accession number NM_020562), Ubd, A2AR (Adenosine A2A receptor), B7-H3 (also called

CD276), B7-H4 (also called VTCN1), BTLA (B and T Lymphocyte Attenuator; also called CD272), CTLA-4 (Cytotoxic T-Lymphocyte-Associated protein 4; also called CD152), IDO (Indoleamine 2,3-dioxygenase) KIR (Killer-cell Immunoglobulin-like Receptor), LAG3 (Lymphocyte Activation Gene-3), PD-1 (Programmed Death 1 (PD-1) receptor), TIM-3 (T-cell Immunoglobulin domain and Mucin domain 3), and VISTA (V-domain Ig suppressor of T cell activation).

4. Compositions for Gene Activation

[0093] The present invention is directed to a composition for activating gene expression of a target gene, target enhancer, or target regulatory element in a cell or subject. The composition may include the CRISPR/Cas9-based gene activation system, as disclosed above. The composition may also include a viral delivery system. For example, the viral delivery system may include an adeno-associated virus vector or a modified lentiviral vector.

[0094] Methods of introducing a nucleic acid into a host cell are known in the art, and any known method can be used to introduce a nucleic acid (e.g., an expression construct) into a cell. Suitable methods include, include e.g., viral or bacteriophage infection, transfection, conjugation, protoplast fusion, lipofection, electroporation, calcium phosphate precipitation, polyethyleneimine (PEI)-mediated transfection, DEAE-dextran mediated transfection, liposome-mediated transfection, particle gun technology, calcium phosphate precipitation, direct micro injection, nanoparticle-mediated nucleic acid delivery, and the like. In some embodiments, the composition may be delivered by mRNA delivery and ribonucleoprotein (RNP) complex delivery.

a) Constructs and Plasmids

[0095] The compositions, as described above, may comprise genetic constructs that encodes the CRISPR/Cas9-based gene activation system, as disclosed herein. The genetic construct, such as a plasmid or expression vector, may comprise a nucleic acid that encodes the CRISPR/Cas9-based gene activation system, such as the CRISPR/Cas9-based acetyltransferase and/or at least one of the gRNAs. The compositions, as described above, may comprise genetic constructs that encodes the modified AAV vector and a nucleic acid sequence that encodes the CRISPR/Cas9-based gene activation system, as disclosed herein. The genetic construct, such as a plasmid, may comprise a nucleic acid that encodes the CRISPR/Cas9-based gene activation system. The compositions, as described above, may comprise genetic constructs that encodes a modified lentiviral vector. The genetic construct, such as a plasmid, may comprise a nucleic acid that encodes the CRISPR/Cas9-based acetyltransferase and at least one sgRNA. The genetic construct may be present in the cell as a functioning extrachromosomal molecule. The genetic construct may be a linear minichromosome including centromere, telomeres or plasmids or cosmids.

[0096] The genetic construct may also be part of a genome of a recombinant viral vector, including recombinant lentivirus, recombinant adenovirus, and recombinant adenovirus associated virus. The genetic construct may be part of the genetic material in attenuated live microorganisms or recombinant microbial vectors which live in cells. The genetic constructs may comprise regulatory elements for

gene expression of the coding sequences of the nucleic acid. The regulatory elements may be a promoter, an enhancer, an initiation codon, a stop codon, or a polyadenylation signal. [0097] The nucleic acid sequences may make up a genetic construct that may be a vector. The vector may be capable of expressing the fusion protein, such as the CRISPR/Cas9-based gene activation system, in the cell of a mammal. The vector may be recombinant. The vector may comprise heterologous nucleic acid encoding the fusion protein, such as the CRISPR/Cas9-based gene activation system. The vector may be a plasmid. The vector may be useful for transfecting cells with nucleic acid encoding the CRISPR/Cas9-based gene activation system, which the transformed host cell is cultured and maintained under conditions wherein expression of the CRISPR/Cas9-based gene activation system takes place.

[0098] Coding sequences may be optimized for stability and high levels of expression. In some instances, codons are selected to reduce secondary structure formation of the RNA such as that formed due to intramolecular bonding.

[0099] The vector may comprise heterologous nucleic acid encoding the CRISPR/Cas9-based gene activation system and may further comprise an initiation codon, which may be upstream of the CRISPR/Cas9-based gene activation system coding sequence, and a stop codon, which may be downstream of the CRISPR/Cas9-based gene activation system coding sequence. The initiation and termination codon may be in frame with the CRISPR/Cas9-based gene activation system coding sequence. The vector may also comprise a promoter that is operably linked to the CRISPR/Cas9-based gene activation system coding sequence. The CRISPR/Cas9-based gene activation system may be under the light-inducible or chemically inducible control to enable the dynamic control of gene activation in space and time. The promoter operably linked to the CRISPR/Cas9-based gene activation system coding sequence may be a promoter from simian virus 40 (SV40), a mouse mammary tumor virus (MMTV) promoter, a human immunodeficiency virus (HIV) promoter such as the bovine immunodeficiency virus (BIV) long terminal repeat (LTR) promoter, a Moloney virus promoter, an avian leukosis virus (ALV) promoter, a cytomegalovirus (CMV) promoter such as the CMV immediate early promoter, Epstein Barr virus (EBV) promoter, or a Rous sarcoma virus (RSV) promoter. The promoter may also be a promoter from a human gene such as human ubiquitin C (hUbC), human actin, human myosin, human hemoglobin, human muscle creatine, or human metallothionein. The promoter may also be a tissue specific promoter, such as a muscle or skin specific promoter, natural or synthetic. Examples of such promoters are described in US Patent Application Publication No. US20040175727, the contents of which are incorporated herein in its entirety.

[0100] The vector may also comprise a polyadenylation signal, which may be downstream of the CRISPR/Cas9-based gene activation system. The polyadenylation signal may be a SV40 polyadenylation signal, LTR polyadenylation signal, bovine growth hormone (bGH) polyadenylation signal, human growth hormone (hGH) polyadenylation signal, or human (3-globin polyadenylation signal. The SV40 polyadenylation signal may be a polyadenylation signal from a pCEP4 vector (Invitrogen, San Diego, CA).

[0101] The vector may also comprise an enhancer upstream of the CRISPR/Cas9-based gene activation system, i.e., the CRISPR/Cas9-based acetyltransferase coding

sequence or sgRNAs. The enhancer may be necessary for DNA expression. The enhancer may be human actin, human myosin, human hemoglobin, human muscle creatine or a viral enhancer such as one from CMV, HA, RSV or EBV. Polynucleotide function enhancers are described in U.S. Pat. Nos. 5,593,972, 5,962,428, and WO94/016737, the contents of each are fully incorporated by reference. The vector may also comprise a mammalian origin of replication in order to maintain the vector extrachromosomally and produce multiple copies of the vector in a cell. The vector may also comprise a regulatory sequence, which may be well suited for gene expression in a mammalian or human cell into which the vector is administered. The vector may also comprise a reporter gene, such as green fluorescent protein ("GFP") and/or a selectable marker, such as hygromycin ("Hygro").

[0102] The vector may be expression vectors or systems to produce protein by routine techniques and readily available starting materials including Sambrook et al., Molecular Cloning and Laboratory Manual, Second Ed., Cold Spring Harbor (1989), which is incorporated fully by reference. In some embodiments the vector may comprise the nucleic acid sequence encoding the CRISPR/Cas9-based gene activation system, including the nucleic acid sequence encoding the CRISPR/Cas9-based acetyltransferase and the nucleic acid sequence encoding the at least one gRNA comprising the nucleic acid sequence of at least one of SEQ ID NOs: 23-73, 188-223, or 224-254.

b) Combinations

[0103] The CRISPR/Cas9-based gene activation system composition may be combined with orthogonal dCas9s, TALEs, and zinc finger proteins to facilitate studies of independent targeting of particular effector functions to distinct loci. In some embodiments, the CRISPR/Cas9-based gene activation system composition may be multiplexed with various activators, repressors, and epigenetic modifiers to precisely control cell phenotype or decipher complex networks of gene regulation.

5. Methods of Use

[0104] Potential applications of the CRISPR/Cas9-based gene activation system are diverse across many areas of science and biotechnology. The CRISPR/Cas9-based gene activation system may be used to activate gene expression of a target gene or target a target enhancer or target regulatory element. The CRISPR/Cas9-based gene activation system may be used to transdifferentiate a cell and/or activate genes related to cell and gene therapy, genetic reprogramming, and regenerative medicine. The CRISPR/Cas9-based gene activation system may be used to reprogram cell lineage specification. Activation of endogenous genes encoding the key regulators of cell fate, rather than forced overexpression of these factors, may potentially lead to more rapid, efficient, stable, or specific methods for genetic reprogramming and transdifferentiation. The CRISPR/Cas9-based gene activation system could provide a greater diversity of transcriptional activators to complement other tools for modulating mammalian gene expression. The CRISPR/Cas9-based gene activation system may be used to compensate for genetic defects, suppress angiogenesis, inactivate oncogenes, activate silenced tumor suppressors, regenerate tissue or reprogram genes.

6. Methods of Activating Gene Expression

[0105] The present disclosure provides a mechanism for activating the expression of target genes based on targeting a histone acetyltransferase to a target region via a CRISPR/Cas9-based gene activation system, as described above. The CRISPR/Cas9-based gene activation system may activate silenced genes. The CRISPR/Cas9-based gene activation system target regions upstream of the TSS of the target gene and substantially induced gene expression of the target gene. The polynucleotide encoding the CRISPR/Cas9-based gene activation system can also be transfected directly to cells.

[0106] The method may include administering to a cell or subject a CRISPR/Cas9-based gene activation system, compositions of CRISPR/Cas9-based gene activation system, or one or more polynucleotides or vectors encoding said CRISPR/Cas9-based gene activation system, as described above. The method may include administering a CRISPR/Cas9-based gene activation system, compositions of CRISPR/Cas9-based gene activation system, or one or more polynucleotides or vectors encoding said CRISPR/Cas9-based gene activation system, as described above, to a mammalian cell or subject.

7. Pharmaceutical Compositions

[0107] The CRISPR/Cas9-based gene activation system may be in a pharmaceutical composition. The pharmaceutical composition may comprise about 1 ng to about 10 mg of DNA encoding the CRISPR/Cas9-based gene activation system. The pharmaceutical compositions according to the present invention are formulated according to the mode of administration to be used. In cases where pharmaceutical compositions are injectable pharmaceutical compositions, they are sterile, pyrogen free and particulate free. An isotonic formulation is preferably used. Generally, additives for isotonicity may include sodium chloride, dextrose, mannitol, sorbitol and lactose. In some cases, isotonic solutions such as phosphate buffered saline are preferred. Stabilizers include gelatin and albumin. In some embodiments, a vasoconstriction agent is added to the formulation.

[0108] The pharmaceutical composition containing the CRISPR/Cas9-based gene activation system may further comprise a pharmaceutically acceptable excipient. The pharmaceutically acceptable excipient may be functional molecules as vehicles, adjuvants, carriers, or diluents. The pharmaceutically acceptable excipient may be a transfection facilitating agent, which may include surface active agents, such as immune-stimulating complexes (ISCOMS), Freunds incomplete adjuvant, LPS analog including monophosphoryl lipid A, muramyl peptides, quinone analogs, vesicles such as squalene and squalene, hyaluronic acid, lipids, liposomes, calcium ions, viral proteins, polyanions, polycations, or nanoparticles, or other known transfection facilitating agents.

[0109] The transfection facilitating agent is a polyanion, polycation, including poly-L-glutamate (LGS), or lipid. The transfection facilitating agent is poly-L-glutamate, and more preferably, the poly-L-glutamate is present in the pharmaceutical composition containing the CRISPR/Cas9-based gene activation system at a concentration less than 6 mg/ml. The transfection facilitating agent may also include surface active agents such as immune-stimulating complexes (ISCOMS), Freunds incomplete adjuvant, LPS analog including monophosphoryl lipid A, muramyl peptides, qui-

none analogs and vesicles such as squalene and squalene, and hyaluronic acid may also be used administered in conjunction with the genetic construct. In some embodiments, the DNA vector encoding the CRISPR/Cas9-based gene activation system may also include a transfection facilitating agent such as lipids, liposomes, including lecithin liposomes or other liposomes known in the art, as a DNA-liposome mixture (see for example WO9324640), calcium ions, viral proteins, polyanions, polycations, or nanoparticles, or other known transfection facilitating agents. Preferably, the transfection facilitating agent is a polyanion, polycation, including poly-L-glutamate (LGS), or lipid.

8. Methods of Delivery

[0110] Provided herein is a method for delivering the pharmaceutical formulations of the CRISPR/Cas9-based gene activation system for providing genetic constructs and/or proteins of the CRISPR/Cas9-based gene activation system. The delivery of the CRISPR/Cas9-based gene activation system may be the transfection or electroporation of the CRISPR/Cas9-based gene activation system as one or more nucleic acid molecules that is expressed in the cell and delivered to the surface of the cell. The CRISPR/Cas9-based gene activation system protein may be delivered to the cell. The nucleic acid molecules may be electroporated using BioRad Gene Pulser Xcell or Amaxa Nucleofector IIb devices or other electroporation device. Several different buffers may be used, including BioRad electroporation solution, Sigma phosphate-buffered saline product #D8537 (PBS), Invitrogen OptiMEM I (OM), or Amaxa Nucleofector solution V (N.V.). Transfections may include a transfection reagent, such as Lipofectamine 2000.

[0111] The vector encoding a CRISPR/Cas9-based gene activation system protein may be delivered to the mammal by DNA injection (also referred to as DNA vaccination) with and without in vivo electroporation, liposome mediated, nanoparticle facilitated, and/or recombinant vectors. The recombinant vector may be delivered by any viral mode. The viral mode may be recombinant lentivirus, recombinant adenovirus, and/or recombinant adeno-associated virus.

[0112] The nucleotide encoding a CRISPR/Cas9-based gene activation system protein may be introduced into a cell to induce gene expression of the target gene. For example, one or more nucleotide sequences encoding the CRISPR/Cas9-based gene activation system directed towards a target gene may be introduced into a mammalian cell. Upon delivery of the CRISPR/Cas9-based gene activation system to the cell, and thereupon the vector into the cells of the mammal, the transfected cells will express the CRISPR/Cas9-based gene activation system. The CRISPR/Cas9-based gene activation system may be administered to a mammal to induce or modulate gene expression of the target gene in a mammal. The mammal may be human, non-human primate, cow, pig, sheep, goat, antelope, bison, water buffalo, bovids, deer, hedgehogs, elephants, llama, alpaca, mice, rats, or chicken, and preferably human, cow, pig, or chicken.

9. Routes of Administration

[0113] The CRISPR/Cas9-based gene activation system and compositions thereof may be administered to a subject by different routes including orally, parenterally, sublin-

gually, transdermally, rectally, transmucosally, topically, via inhalation, via buccal administration, intrapleurally, intravenous, intraarterial, intraperitoneal, subcutaneous, intramuscular, intranasal intrathecal, and intraarticular or combinations thereof. For veterinary use, the composition may be administered as a suitably acceptable formulation in accordance with normal veterinary practice. The veterinarian may readily determine the dosing regimen and route of administration that is most appropriate for a particular animal. The CRISPR/Cas9-based gene activation system and compositions thereof may be administered by traditional syringes, needless injection devices, "microparticle bombardment gone guns", or other physical methods such as electroporation ("EP"), "hydrodynamic method", or ultrasound. The composition may be delivered to the mammal by several technologies including DNA injection (also referred to as DNA vaccination) with and without in vivo electroporation, liposome mediated, nanoparticle facilitated, recombinant vectors such as recombinant lentivirus, recombinant adenovirus, and recombinant adenovirus associated virus.

10. Cell Types

[0114] The CRISPR/Cas9-based gene activation system may be used with any type of cell. In some embodiments, the cell is a bacterial cell, a fungal cell, an archaea cell, a plant cell or an animal cell. In some embodiments, the cell may be an ENCODE cell line, including but not limited to, GM12878, K562, H1 human embryonic stem cells, HeLa-S3, HepG2, HUVEC, SK-N-SH, IMR90, A549, MCF7, HMEC or LHCN, CD14+, CD20+, primary heart or liver cells, differentiated H1 cells, 8988T, Adult_CD4_naive, Adult_CD4_Th0, Adult_CD4_Th1, AG04449, AG04450, AG09309, AG09319, AG10803, AoAF, AoSMC, BC_Adipose_UHN00001, BC_Adrenal_Gland_H12803N, BC_Bladder_01-11002, BC_Brain_H11058N, BC_Breast_02-03015, BC_Colon_01-11002, BC_Colon_H12817N, BC_Esophagus_01-11002, BC_Esophagus_H12817N, BC_Jejunum_H12817N, BC_Kidney_01-11002, BC_Kidney_H12817N, BC_Left_Ventricle_N41, BC_Leukocyte_UHN00204, BC_Liver_01-11002, BC_Lung_01-11002, BC_Lung_H12817N, BC_Pancreas_H12817N, BC_Penis_H12817N, BC_Pericardium_H12529N, BC_Placenta_UHN00189, BC_Prostate_Gland_H12817N, BC_Rectum_N29, BC_Skeletal_Muscle_01-11002, BC_Skeletal_Muscle_H12817N, BC_Skin_01-11002, BC_Small_Intestine_01-11002, BC_Spleen_H12817N, BC_Stomach_01-11002, BC_Stomach_H12817N, BC_Testis_N30, BC_Uterus_BN0765, BE2_C, BG02ES, BG02ES-EBD, BJ, bone_marrow_HS27a, bone_marrow_HS5, bone_marrow_MSC, Breast_OC, Caco-2, CD20+_RO01778, CD20+_RO01794, CD34+_Mobilized, CD4+_Naive_Wb11970640, CD4+_Naive_Wb78495824, Cerebellum_OC, Cerebrum_frontal_OC, Chorion, CLL, CMK, Colo829, Colon_BC, Colon_OC, Cord_CD4_naive, Cord_CD4_Th0, Cord_CD4_Th1, Decidua, Dnd41, ECC-1, Endometrium_OC, Esophagus_BC, Fibrobl, Fibrobl_GM03348, FibroP, FibroP_AG08395, FibroP_AG08396, FibroP_AG20443, Frontal_cortex_OC, GC_B_cell, Gliobla, GM04503, GM04504, GM06990, GM08714, GM10248, GM10266, GM10847, GM12801, GM12812, GM12813, GM12864, GM12865, GM12866, GM12867, GM12868, GM12869, GM12870, GM12871, GM12872, GM12873, GM12874, GM12875, GM12878-XiMat, GM12891, GM12892, GM13976, GM13977, GM15510, GM18505, GM18507,

GM18526, GM18951, GM19099, GM19193, GM19238, GM19239, GM19240, GM20000, H0287, H1-neurons, H7-hESC, H9ES, H9ES-AFP-, H9ES-AFP+, H9ES-CM, H9ES-E, H9ES-EB, H9ES-EBD, HAc, HAEpiC, HA-h, HAL, HAoAF, HAoAF_6090101.11, HAoAF_6111301.9, HAoEC, HAoEC_7071706.1, HAoEC_8061102.1, HA-sp, HBMEC, HBVP, HBVSMC, HCF, HCFAa, HCH, HCH_0011308.2P, HCH_8100808.2, HCM, HConF, HCPepiC, HCT-116, Heart_OC, Heart_STL003, HEPEpiC, HEK293, HEK293T, HEK293-T-REx, Hepatocytes, HFDPC, HFDPC_0100503.2, HFDPC_0102703.3, HFF, HFF-Myc, HFL11W, HFL24W, HGF, HHSEC, HEPEpiC, HL-60, HMEpC, HMEpC_6022801.3, HMF, hMNC-CB, hMNC-CB_8072802.6, hMNC-CB_9111701.6, hMNC-PB, hMNC-PB_0022330.9, hMNC-PB_0082430.9, hMSC-AT, hMSC-AT_0102604.12, hMSC-AT_9061601.12, hMSC-BM, hMSC-BM_0050602.11, hMSC-BM_0051105.11, hMSC-UC, hMSC-UC_0052501.7, hMSC-UC_0081101.7, HMVEC-dAd, HMVEC-dBI-Ad, HMVEC-dBI-Neo, HMVEC-dLy-Ad, HMVEC-dLy-Neo, HMVEC-dNeo, HMVEC-LBI, HMVEC-LLy, HNPCEpiC, HOB, HOB_0090202.1, HOB_0091301, HPAEC, HPAEpiC, HPAF, HPC-PL, HPC-PL_0032601.13, HPC-PL_0101504.13, HPDE6-E6E7, HPdLF, HPF, HPIEpC, HPIEpC_9012801.2, HPIEpC_9041503.2, HRCEpiC, HRE, HRGEC, HRPEpiC, HSaVEC, HSaVEC_0022202.16, HSaVEC_9100101.15, HSMM, HSMM_emb, HSMM_FSHD, HSMMtube, HSMMtube_emb, HSMMtube_FSHD, HT-1080, HTR8svn, Huh-7, Huh-7.5, HVMF, HVMF_6091203.3, HVMF_6100401.3, HWP, HWP_0092205, HWP_8120201.5, iPS, iPS_CWRU1, iPS_hFib2_iPS4, iPS_hFib2_iPS5, iPS_NI-Hil 1, iPS_NIH7, Ishikawa, Jurkat, Kidney BC, Kidney_OC, LHCN-M2, LHSR, Liver_OC, Liver_STL004, Liver-STLO11, LNCap, Loucy, Lung_BC, Lung_OC, Lymphoblastoid_cell_line, M059J, MCF10A-Er-Src, MCF-7, MDA-MB-231, Medullo, Medullo_D341, Mel 2183, Melano, Monocytes-CD14+, Monocytes-CD14+_RO01746, Monocytes-CD14+_RO01826, MRT_A204, MRT_G401, MRT_TTC549, Myometr, Naive_B_cell, NB4, NH-A, NHBE, NHBE_RA, NHDF, NHDF_0060801.3, NHDF_7071701.2, NHDF-Ad, NHDF-neo, NHEK, NHEM.f_M2, NHEM.f_M2_5071302.2, NHEM.f_M2_6022001, NHEM_M2, NHEM_M2_7011001.2, NHEM_M2_7012303, NHLF, NT2-D1, Olf_neurosphere, Osteobl, ovarc-3, PANC-1, Pancreas_OC, PanIsletD, PanIslets, PBDE, PBDEFetal, PBMC, PFSK-1, pHTe, Pons_OC, PrEC, ProgFib, Prostate, Prostate_OC, Psoas_muscle_OC, Raji, RCC_7860, RPMI-7951, RPTEC, RWPE1, SAEC, SH-SY5Y, Skeletal_Muscle_BC, SkMC, SkMC, SkMC_8121902.17, SkMC_9011302, SK-N-MC, SK-N-SH_RA, Small_intestine_OC, Spleen_OC, Stellate, Stomach_BC, T_cells_CD4+, T-47D, T98G, TBEC, Th1, Th1_Wb33676984, Th1_Wb54553204, Th17, Th2, Th2_Wb33676984, Th2_Wb54553204, Treg_Wb78495824, Treg_Wb83319432, U2OS, U87, UCH-1, Urothelia, WERI-Rb-1, and WI-38.

11. Kits

[0115] Provided herein is a kit, which may be used to activate gene expression of a target gene. The kit comprises a composition for activating gene expression, as described above, and instructions for using said composition. Instructions included in kits may be affixed to packaging material or may be included as a package insert. While the instruc-

tions are typically written or printed materials they are not limited to such. Any medium capable of storing such instructions and communicating them to an end user is contemplated by this disclosure. Such media include, but are not limited to, electronic storage media (e.g., magnetic discs, tapes, cartridges, chips), optical media (e.g., CD ROM), and the like. As used herein, the term "instructions" may include the address of an internet site that provides the instructions. [0116] The composition for activating gene expression may include a modified AAV vector and a nucleotide sequence encoding a CRISPR/Cas9-based gene activation system, as described above. The CRISPR/Cas9-based gene activation system may include CRISPR/Cas9-based acetyltransferase, as described above, that specifically binds and targets a cis-regulatory region or trans-regulatory region of a target gene. The CRISPR/Cas9-based acetyltransferase, as described above, may be included in the kit to specifically bind and target a particular regulatory region of the target gene.

12. Examples

[0117] The foregoing may be better understood by reference to the following examples, which are presented for purposes of illustration and are not intended to limit the scope of the invention.

Example 1

Methods and Materials—Activator

[0118] Cell lines and transfection. HEK293T cells were procured from the American Tissue Collection Center (ATCC, Manassas VA) through the Duke University Cell Culture Facility. Cells were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% FBS and 1% penicillin/streptomycin and maintained at 37° C. and 5% CO₂. Transfections were performed in 24-well plates using 375 ng of respective dCas9 expression vector and 125 ng of equimolar pooled or individual gRNA expression vectors mixed with Lipofectamine 2000 (Life Technologies, cat. #11668019) as per manufacturer's instruction. For ChIP-qPCR experiments, HEK293T cells were transfected in 15 cm dishes with Lipofectamine 2000 and 30 µg of respective dCas9 expression vector and 10 µg of equimolar pooled gRNA expression vectors as per manufacturer's instruction. [0119] Plasmid constructs. pcDNA-dCas9^{VP64} (dCas9^{VP64}; Addgene, plasmid #47107) was used (Perez-Pinera, P. et al, *Nature methods* 10:973-976 (2013)). An HA epitope tag was added to dCas9 (no effector) by removing

the VP64 effector domain from dCas9^{VP64} via Ascl/PacI restriction sites and using isothermal assembly (Gibson et al. *Nat. Methods* 6:343-345 (2009)) to include an annealed set of oligos containing the appropriate sequence as per manufacturers instruction (NEB cat. #2611). pcDNA-dCas9^{FLp300} (dCas9^{FLp300}) was created by amplifying full-length p300 from pcDNA3.1-p300 (Addgene, plasmid #23252) (Chen et al. *EMBO J.* 21:6539-6548 (2002)) in two separate fragments and cloning these fragments into the dCas9^{VP64} backbone via isothermal assembly. A substitution in the full-length p300 protein (L553M), located outside of the HAT Core region, was identified in dCas9^{FLp300} and in the precursor pcDNA3.1-p300 during sequence validation. pcDNA-dCas9^{p300 Core} (dCas9^{p300 Core}) was generated by first amplifying amino acids 1048-1664 of human p300 from cDNA and then subcloning the resulting amplicon into pCR-Blunt (pCR-Blunt^{p300 Core}) (Life Technologies cat. #K2700). An Ascl site, HA-epitope tag, and a Pmel site were added by PCR amplification of the p300 Core from pCR-Blunt^{p300 Core} and subsequently this amplicon was cloned into pCR-Blunt (pCR-Blunt^{p300 Core+HA}) (Life Technologies cat. #K2700). The HA-tagged p300 Core was cloned from pCR-Blunt^{p300 Core+HA} into the dCas9^{VP64} backbone via shared Ascl/Pmel restriction sites. pcDNA-dCas9^{p300 Core} (D13991) (dCas9^{p300 Core} (D13991)) was generated by amplification of the p300 Core from dCas9^{p300 Core} in overlapping fragments with primer sets including the specified nucleic acid mutations, with a subsequent round of linkage PCR and cloning into the dCas9^{p300 Core} backbone using shared Ascl/Pmel restriction sites. All PCR amplifications were carried out using Q5 high-fidelity DNA polymerase (NEB cat. #M0491). Protein sequences of all dCas9 constructs are shown in FIGS. 15A-15J.

[0120] IL1RN, MYOD, and OCT4 promoter gRNA protospacers have been described previously (Perez-Pinera, P. et al, *Nature methods* 10:973-976 (2013); Hu, J. et al., *Nucleic Acids Res* 42:4375-4390 (2014)). *Neisseria meningitidis* dCas9^{VP64} (Nm-dCas9^{VP64}) was obtained from Addgene (plasmid #48676). Nm-dCas9^{p300 Core} was created by amplifying the HA-tagged p300 Core from dCas9^{p300 Core} with primers to facilitate subcloning into the Ael/Agel-digested Nm-dCas9^{VP64} backbone using isothermal assembly (NEB cat. #2611). IL1RN TALE^{p300 Core} TALEs were generated by subcloning the HA-tagged p300 Core domain from dCas9^{p300 Core} into previously published (Perez-Pinera, P. et al, *Nature methods* 10:973-976 (2013)) IL1RN TALE^{VP64} constructs via shared Ascl/Pmel restriction sites. IL1RN TALE target sites are shown in Table 1.

TABLE 1

IL1RN TAL effector information.				
Name	Target Site	SEQ ID NO	Location (GRCh37/hg19 assembly)	
IL1RN TALE ^{VP64 A}	GGGCTCCTCCTTGTACT	15	chr2: 113875431-113875447	
IL1RN TALE ^{VP64 B}	ACGCAGATAAGAACCGAT	16	chr2: 113875291-113875308	
IL1RN TALE ^{VP64 C}	GGCATCAAGTCAGCCAT	17	chr2: 113875356-113875372	
IL1RN TALE ^{VP64 D}	AGCCTGAGTCACCCTCCT	18	chr2: 113875321-113875338	
IL1RN TALE ^{p300 Core A}	GGGCTCCTCCTTGTACT	19	chr2: 113875431-113875447	

TABLE 1-continued

IL1RN TAL effector information.			
Name	Target Site	SEQ ID NO	Location (GRCh37/hg19 assembly)
IL1RN TALE ^{p300 Core B}	ACCGAGATAAGAACAGT	20	chr2: 113875291-113875308
IL1RN TALE ^{p300 Core C}	GGCATCAAGTCAGCCAT	21	chr2: 113875356-113875372
IL1RN TALE ^{p300 Core D}	AGCCTGAGTCACCCCTCCT	22	chr2: 113875321-113875338

[0121] ICAM1 ZF^{VP64} and ICAM1 ZF^{p300 Core} were constructed by subcloning the ICAM1 ZF from pMX-CD54-31Opt-VP64⁵⁴ into dCas9^{VP64} and dCas9^{p300 Core} backbones, respectively, using isothermal assembly (NEB cat. #2611). Protein sequences of ICAM1 ZF constructs are shown in FIG. 16. Transfection efficiency was routinely above 90% as assayed by co-transfection of PL-SIN-EF1 α -EGFP (Addgene plasmid #21320) and gRNA empty vector in all experiments. All *Streptococcus pyogenes* gRNAs were

annealed and cloned into pZdonor-pSPgRNA (Addgene plasmid #47108) for expression (Cong, L. et al., *Science* 339:819-823 (2013)) with slight modifications using NEB BbsI and T4 ligase (Cat. #s R0539 and M0202). Nm-dCas9 gRNA oligos were rationally designed using published PAM requirements (Esvelt, K. M. et al., *Nature Methods* 10:1116-1121 (2013)), and then cloned into pZDonor-Nm-Cas9-gRNA-hU6 (Addgene, plasmid #61366) via BbsI sites. Plasmids are available through Addgene (Table 2).

TABLE 2

Referenced plasmids in this study available at Addgene.	
Plasmid Name	Addgene Plasmid #
pcDNA-dCas9 ^{VP64} (SEQ ID NO: 139)	47107
pcDNA-dCas9-HA (SEQ ID NO: 138)	61355
pcDNA3.1-p300	23252
pcDNA-dCas9 ^{FLp300} (SEQ ID NO: 140)	61356
pcDNA-dCas9 ^{p300 Core} (SEQ ID NO: 141)	61357
pcDNA-dCas9 ^{p300 Core (D1399F)} (SEQ ID NO: 142)	61358
pcDNA-dCas9 ^{p300 Core (1645/1646 RR/EE)} (SEQ ID NO: 143)	61359
pcDNA-dCas9 ^{p300 Core (C1204R)} (SEQ ID NO: 144)	61361
pcDNA-dCas9 ^{p300 Core (Y1467F)} (SEQ ID NO: 145)	61362
pcDNA-dCas9 ^{p300 Core (1396/1397 SY/WW)} (SEQ ID NO: 146)	61363
pcDNA-dCas9 ^{p300 Core (H1415A/E1423A/Y1424A/L1428S/Y1430A/H1434A)} (SEQ ID NO: 147)	61364
pZdonor-pSPgRNA	47108
pcDNA3.1-300(HAT-)	23254
pcDNA3.3-Nm-dCas9 ^{VP64} (SEQ ID NO: 148)	48676
pcDNA3.3-Nm-dCas9 ^{p300 Core} (SEQ ID NO: 149)	61365
pZDonor-NmCas9-gRNA-hU6	61366
PL-SIN-EF1 α -EGFP	21320

[0122] All gRNA protospacer targets are listed in Tables 3 and 4.

TABLE 3

gRNA information.			
Target Location	Protospacer Sequence (5'-3')	SEQ ID NO	Genomic Location (GRCh37/hg19 Assembly)
IL1RN Promoter A	TGTACTCTCTGAGGTGCTC	23	chr2: 113875442-113875460
IL1RN Promoter B	ACGCAGATAAGAACAGTT	24	chr2: 113875291-113875309
IL1RN Promoter C	CATCAAGTCAGCCATCAGC	25	chr2: 113875358-113875376
IL1RN Promoter D	GAGTCACCCCTCCTGGAAAC	26	chr2: 113875326-113875344

TABLE 3-continued

gRNA information.					
Target Location	Protospacer Sequence (5'-3')	SEQ	ID NO	Genomic Location (GRCh37/hg19 Assembly)	
MYOD Promoter A	CCTGGGCTCCGGGGCGTTT	27	chr11: 17741056-17741074		
MYOD Promoter B	GGCCCCCTGCGCCACCCCG	28	chr11: 17740969-17740987		
MYOD Promoter C	CTCCCTCCCTGCCCGGTAG	29	chr11: 17740897-17740915		
MYOD Promoter D	AGGTTTGAAAGGGCGTGC	30	chr11: 17740837-17740855		
OCT4 Promoter A	ACTCCACTGCACTCCAGTCT	31	chr6: 31138711-31138730		
OCT4 Promoter B	TCTGTGGGGACCTGCACTG	32	chr6: 31138643-31138662		
OCT4 Promoter C	GGGGCGCCAGTTGTCTCC	33	chr6: 31138613-31138632		
OCT4 Promoter D	ACACCATTGCCACCACCATT	34	chr6: 31138574-31138593		
MYOD DRR A	TGTTTCAGCTTCCAAACT	35	chr11: 17736528-17736546		
MYOD DRR B	CATGAAGACAGCAGAACCC	36	chr11: 17736311-17736329		
MYOD DRR C	GGCCCACATTCCCTTCCAG	37	chr11: 17736158-17736176		
MYOD DRR D	GGCTGGATTGGTTCCAG	38	chr11: 17736065-17736083		
MYOD CE A	CAAATGAGTCCTGAGGTTT	39	chr11: 17721347-17721365		
MYOD CE B	CTCACAGCACAGCCAGTGT	40	chr11: 17721257-17721275		
MYOD CE C	CAGCAGCTGGTCACAAAGC	41	chr11: 17721200-17721218		
MYOD CE D	CTTCCTATAAACTTCTGAG	42	chr11: 17721139-17721157		
OCT4 PE A	AGTGATAAGAACACCCGCTTT	43	chr6: 31139524-31139543		
OCT4 PE B	CAGACATCTAATACCACGGT	44	chr6: 31139604-31139623		
OCT4 PE C	AGGGAGAACGGGCCTACCG	45	chr6: 31139620-31139639		
OCT4 PE D	ACTTCAGGTCACAAAGGCC	46	chr6: 31139725-31139744		
OCT4 PE E	TTTCCCCACCCAGGGCCTA	47	chr6: 31139671-31139690		
OCT4 PE F	CCCTGGGTGGGGAAAACAG	48	chr6: 31139675-31139694		
OCT4 DE A	GGAGGAACATGCTCGAAC	49	chr6: 31140809-31140828		
OCT4 DE B	GTGCCGTGATGGTCTGTCC	50	chr6: 31140864-31140883		
OCT4 DE C	GGTCTGCCGGAAGGTCTACA	51	chr6: 31140707-31140726		
OCT4 DE D	TCGGCCTTTAACTGCCAAA	52	chr6: 31140757-31140776		
OCT4 DE E	GCATGACAAAGGTGCCGTGA	53	chr6: 31140875-31140894		
OCT4 DE F	CCTGCCCTTGGGCAGTTAA	54	chr6: 31140764-31140783		
HS2 A	AATATGTCACATTCTGTCTC	55	chr11: 5301800-5301819		
HS2 B	GGACTATGGGAGGTCACTAA	56	chr11: 5302108-5302127		
HS2 C	GAAGGTTACACAGAACAGA	57	chr11: 5302033-5302052		

TABLE 3 -continued

qRNA information.				
Target Location	Protospacer Sequence (5'-3')	SEQ ID NO	Genomic Location (GRCh37/hg19 Assembly)	
HS2 D	GCCCTGTAAGCATCCTGCTG	58	chr11: 5301898-5301917	

TABLE 4

Target Location	Protospacer Sequence (5'-3')	SEQ ID NO	Genomic Location (GRCh37/hg19 Assembly)
HBG Promoter A	CCACTGCTAACTGAAAGAGA	59	chr11: 5271570-5271589
HBG Promoter B	AGCCACAGTTTCAGCGCAGT	60	chr11: 5271692-5271711
HBG Promoter C	CTGTTTCATCTTAGAAAAAT	61	chr11: 5271793-5271812
HBG Promoter D	GAATGTTCTTGCGAGGTAC	62	chr11: 5271942-5271961
HBG Promoter E	CGCACATCTTATGTCTTAGA	63	chr11: 5272021-5272040
HBE Promoter A	CTTAAGAGAGCTAGAACTGG	64	chr11: 5291618-5291637
HBE Promoter B	TCCCCAAAGTACAGTACCTTG	65	chr11: 5291758-5291777
HBE Promoter C	TCCCTAGAGAGGGACAGACAG	66	chr11: 5291785-5291804
HBE Promoter D	TCATAGAGAAATGAAAAGAG	67	chr11: 5291840-5291859
HBE Promoter E	ATAATATACCCTGACTCCTA	68	chr11: 5292038-5292057
HS2 A	AGGCCACCTGCAAGATAAAAT	69	chr11: 5301662-5301681
HS2 B	TGTTGTTATCAATTGCCATA	70	chr11: 5301708-5301727
HS2 C	ATCCCCTCCAGCATCCTCAT	71	chr11: 5302187-5302206
HS2 D	GTGCTTCAAAACCATTGCT	72	chr11: 5302245-5302264
HS2 E	GATACATGTTTATTCTTAT	73	chr11: 5302306-5302325

[0123] Western Blotting. 20 µg of protein was loaded for SDS PAGE and transferred onto a nitrocellulose membrane for western blots. Primary antibodies (as-FLAG; Sigma-Aldrich cat. #F7425 and α-GAPDH; Cell Signaling Technology cat. #14C10) were used at a 1:1000 dilution in TBST+500 Milk. Secondary α-Rabbit HRP (Sigma-Aldrich cat. #A6154) was used at a 1:5000 dilution in TBST+500 Milk. Membranes were exposed after addition of ECL (Bio-Rad cat. #170-5060).

[0124] Quantitative reverse-transcription PCR. RNA was isolated from transfected cells using the RNeasy Plus mini kit (Qiagen cat. #74136) and 500 ng of purified RNA was used as template for cDNA synthesis (Life Technologies,

cat. #11754). Real-time PCR was performed using PerfeCTa SYBR Green FastMix (Quanta Biosciences, cat. #95072) and a CFX96 Real-Time PCR Detection System with a C1000 Thermal Cycler (Bio-Rad). Baselines were subtracted using the baseline subtraction curve fit analysis mode and thresholds were automatically calculated using the Bio-Rad CFX Manager software version 2.1. Results are expressed as fold change above control mock transfected cells (No DNA) after normalization to GAPDH expression using the ΔΔCt method (Schmittgen et al., *Nat. Protoc.* 3:1101-1108 (2008)). All qPCR primers and conditions are listed in Table 5.

TABLE 5

Quantitative reverse transcription PCR and ChIP-qPCR primers and conditions.						
Target	Primer (5'-3')	SEQ ID NO	Reverse Primer (5'-3')	SEQ ID NO	Cycling Parameters	
GAPDH	CAATGACCCCTTCATT GACC	74	TTGATTTGGAGGGA TCTCG	75	95° C. 30 sec 45X 95° C. 5 sec 53° C. 20 sec	

TABLE 5-continued

Quantitative reverse transcription PCR and ChIP-qPCR primers and conditions.							
Target	Primer (5'-3')	SEQ ID NO	Reverse Primer (5'-3')	SEQ ID NO	Cycling Parameters		
IL1RN	GGAATCCATGGAGGG AAGAT	76	TGTTCTCGCTCAGGTC AGTG	77	95° C. 30 sec	45X	
					95° C. 5 sec		
					58° C. 20 sec		
MYOD	TCCCTCTTCACGGTC TCAC	78	AACACCCGACTGCTG TATCC	79	95° C. 30 sec	45X	
					95° C. 5 sec		
					53° C. 20 sec		
OCT4	CGAAAGAGAAAGCGA ACCAGTATCGAGAAC	80	CGTTGTGCATAGTCG CTGCTTGATCGC	81	95° C. 30 sec	45X	
					95° C. 5 sec		
					53° C. 20 sec		
HBB	GCACGTGGATCCTGAG AACT	82	ATTGGACAGCAAGAA AGCGAG	83	95° C. 30 sec	45X	
					95° C. 5 sec		
					58° C. 20 sec		
HBD	GCACGTGGATCCTGAG AACT	84	CAGGAAACAGTCCAG GATCTCA	85	95° C. 30 sec	45X	
					95° C. 5 sec		
					58° C. 20 sec		
HBG	GCTGAGTGAAGTCAC TGTGA	86	GAATTCTTGCAGAA ATGGA	87	95° C. 30 sec	45X	
					95° C. 5 sec		
					58° C. 20 sec		
HBE	TCACTAGCAAGCTCTC AGGC	88	AAACAACGAGGAGTCT GCC	89	95° C. 30 sec	45X	
					95° C. 5 sec		
					62° C. 20 sec		
ICAM1	GCAGACAGTGACCATC TACAGCTT	90	CAATCCCTCTCGTCC AGTCG	91	95° C. 30 sec	45X	
					95° C. 5 sec		
					58° C. 20 sec		
HS2 ChIP Region 1	TGCTTGGACTATGGGA GTC	92	GCAGGTGCTTCAAAA CCATT	93	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HS2 ChIP Region 2	TCAGGTGGTCAGCTTC TCCT	94	AAGCAAACCTCTGG CTCAA	95	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HS2 ChIP Region 3	CCACACAGGTGAACCC TTTT	96	GGACACATGCTCACA TACGG	97	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HBE ChIP Region 1	ATTCGATCCATGTGCC TGA	98	CAATGCTGGAATTG TGGAA	99	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HBE ChIP Region 2	GGGGTGATTCCCTAGA GAGG	100	AAGCAGGACAGACA GGCAAG	101	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HBE ChIP Region 3	GAGGGTCAGCAGTGA TGGAT	102	TGGAAAAGGAGAATG GGAGA	103	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HBG1/2 ChIP Region 1	TGGTCAAAGTTGCCTT GTCA	104	GGAATGACTGAATCG GAACAA	105	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HBG1/2 ChIP Region 2	CCTCCAGCATCTTCCA CATT	106	GAAGCACCCCTTCAGC AGTTC	107	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
HBG1/2 ChIP Region 3	CCACAGTTCAAGCGCA GTAATA	108	ATCAGCCAGCACACA CACTT	109	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		

TABLE 5-continued

Quantitative reverse transcription PCR and ChIP-qPCR primers and conditions.							
Target	Primer (5'-3')	SEQ ID NO	Reverse Primer (5'-3')	SEQ ID NO	Cycling Parameters		
IL1RN ChIP Region 1	CCCTGTCAGGAGGGAC AGAT	110	GGCTCACCGGAAGCA TGAAT	111	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 2	AAGCTACAAGCAGGTT CGCT	112	AATAAACAGGGTCCAT CCCGC	113	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 3	TGTTCCCTCCACCTGG AATA	114	GGGAAAATCCAAAGC AGGAT	115	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 4	TCCTAGGTCCCTCAA AGCA	116	GTCCCCAACGCTCTA ACAAA	117	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 5	GTTAGAGCGTTGGGA CCTT	118	CACATGCAGAGAACT GAGCTG	119	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 6	GTTGGGTAAGCACG AAGG	120	TTTCCAGGAGGGTGA CTCAG	121	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 7	TTCTCTGCATGTGACC TCCC	122	ACACACTCACAGAGG GTTGG	123	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 8	TGAGTCACCCCTCTGG AAAC	124	CTCCTTCCAGAGCAC CTCAG	125	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 9	GCTGGGCTCCTCCTTG TACT	126	GCTGCTGCCCATAAA GTAGC	127	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 10	GGACTGTGGCCAGGT ACT	128	GGCCTCATAGGACAG GAGGT	129	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 11	TTATGGCAGCAGCTC AGTT	130	GACATTTCCTGGAC GCTTG	131	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 12	CCCTCCCCATGGCTTT AGGT	132	AGCTCCATCGCTTG ACATT	133	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
IL1RN ChIP Region 13	ACCGTCCAGGAAAT GTCAA	134	ATGACCCCTCACACTC CAAGG	135	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		
Upstream β-actin ChIP NEG CTRL	GTTGGGTGCTCCAGCT TTTA	136	CCTAAAAACTCCTGG ACTCG	137	95° C. 30 sec	45X	
					95° C. 5 sec		
					60° C. 20 sec		

[0125] RNA-seq. RNA-seq was performed using three replicates per experimental condition. RNA was isolated from transfected cells using the RNeasy Plus mini kit (Qiagen cat. #74136) and 1 µg of purified mRNA was used as template for cDNA synthesis and library construction using the PrepX RNA-Seq Library Kit (Wafergen Biosys-

tems, cat. #400039). Libraries were prepared using the Apollo 324 liquid handling platform, as per manufacturer's instruction. Indexed libraries were validated for quality and size distribution using the Tapestation 2200 (Agilent) and quantified by qPCR using the KAPA Library Quantification Kit (KAPA Biosystems; KK4835) prior to multiplex pooling

and sequencing at the Duke University Genome Sequencing Shared Resource facility. Libraries were pooled and then 50 bp single-end reads were sequenced on a Hiseq 2500 (Illumina), de-multiplexed and then aligned to the HG19 transcriptome using Bowtie 2 (Langmead et al. *Nat. Methods* 9:357-359 (2012)). Transcript abundance was calculated using the SAMtools (Li et al. *Bioinformatics* 25:2078-2079 (2009)) suite and differential expression was determined in R using the DESeq2 analysis package. Multiple hypothesis correction was performed using the method of Benjamini and Hochberg with a FDR of <5%. RNA-seq data is deposited in the NCBI's Gene Expression Omnibus and is accessible through GEO Series accession number GSE66742.

[0126] ChIP-qPCR. HEK293T cells were co-transfected with four HS2 enhancer gRNA constructs and indicated dCas9 fusion expression vectors in 15 cm plates in biological triplicate for each condition tested. Cells were cross-linked with 1% Formaldehyde (final concentration; Sigma F8775-25ML) for 10 min at RT and then the reaction was stopped by the addition of glycine to a final concentration of 125 mM. From each plate ~2.5e7 cells were used for H3K27ac ChIP-enrichment. Chromatin was sheared to a median fragment size of 250 bp using a Bioruptor XL (Diagenode). H3K27ac enrichment was performed by incubation with 5 µg of Abcam ab4729 and 200 µl of sheep anti-rabbit IgG magnetic beads (Life Technologies 11203D) for 16 hrs at 4° C. Cross-links were reversed via overnight incubation at 65° C. with sodium dodecyl sulfate, and DNA was purified using MinElute DNA purification columns (Qiagen). 10 ng of DNA was used for subsequent qPCR reactions using a CFX96 Real-Time PCR Detection System with a C1000 Thermal Cycler (Bio-Rad). Baselines were subtracted using the baseline subtraction curve fit analysis mode and thresholds were automatically calculated using the Bio-Rad CFX Manager software version 2.1. Results are expressed as fold change above cells co-transfected with dCas9 and four HS2 gRNAs after normalization to β-actin enrichment using the ΔΔCt method (Schmittgen et al., *Nat. Protoc.* 3:1101-1108 (2008)). All ChIP-qPCR primers and conditions are listed in Table 5.

Example 2

A dCas9 fusion to the p300 HAT domain activates target genes

[0127] The full-length p300 protein was fused to dCas9 (dCas9^{FLp300}; FIGS. 1A-1*i*) and assayed for its capacity for transactivation by transient co-transfection of human HEK293T cells with four gRNAs targeting the endogenous promoters of IL1RN, MYOD1 (MYOD), and POU5F1/OCT4 (OCT4) (FIG. 1C). A combination of four gRNAs targeting each promoter was used. dCas9^{FLp300} was well expressed and induced modest activation above background compared to the canonical dCas9 activator fused to the VP64 acidic activation domain (dCas9^{VP64}) (FIGS. 1A-1C). The full-length p300 protein is a promiscuous acetyltransferase which interacts with a multitude of endogenous proteins, largely via its termini. In order to mitigate these interactions the contiguous region of full-length p300 (2414 aa) solely required for inherent HAT activity (amino acids 1048-1664), known as the p300 HAT core domain (p300 Core) was isolated. When fused to the C-terminus of dCas9 (dCas9^{p300}_{core}, FIGS. 1A-1) the p300 Core domain induced high levels

of transcription from endogenous gRNA-targeted promoters (FIG. 1C). When targeted to the IL1RN and MYOD promoters, the dCas9^{p300}_{core} fusion displayed significantly higher levels of transactivation than dCas9^{VP64} (P-value 0.01924 and 0.0324 respectively; FIGS. 1A-1C). These dCas9-effector fusion proteins were expressed at similar levels (FIG. 1B, FIGS. 7A-7C) indicating that the observed differences were due to differences in transactivation capacity. Additionally, no changes to target gene expression were observed when the effector fusions were transfected without gRNAs (FIG. 8). For FIG. 8, dCas9 fusion proteins were transiently co-transfected with an empty gRNA vector backbone and mRNA expression of IL1RN, MYOD, and OCT4 was assayed as in the main text. Red dashed line indicates background expression level from No DNA-transfected cells. n=2 independent experiments, error bars: s.e.m., no significant activation was observed for any target gene assayed.

[0128] To ensure that the p300 Core acetyltransferase activity was responsible for gene transactivation using the dCas9^{p300}_{core} fusion, a panel of dCas9^{p300}_{core} HAT-domain mutant fusion proteins was screened (FIGS. 7A-7C). A single inactivating amino acid substitution within the HAT core domain (WT residue D1399 of full-length p300) of dCas9^{p300}_{core} (dCas9^{p300}_{core}(D1399Y)) (FIG. 1A) abolished the transactivation capacity of the fusion protein (FIG. 1C), demonstrating that intact p300 Core acetyltransferase activity was required for dCas9^{p300}_{core}-mediated transactivation.

Example 3

dCas9^{p300} Core Activates Genes from Proximal and Distal Enhancers

[0129] As p300 plays a role and is localized at endogenous enhancers, the dCas9^{p300}_{core} may effectively induce transcription from distal regulatory regions with appropriately targeted gRNAs. The distal regulatory region (DRR) and core enhancer (CE) of the human MYOD locus was targeted through co-transfection of four gRNAs targeted to each region and either dCas9^{VP64} or dCas9^{p300}_{core} (FIG. 2A). Compared to a mock-transfected control, dCas9^{VP64} did not show any induction when targeted to the MYOD DRR or CE region. In contrast, dCas9^{p300}_{core} induced significant transcription when targeted to either MYOD regulatory element with corresponding gRNAs (P-value 0.0115 and 0.0009 for the CE and DRR regions respectively). The upstream proximal (PE) and distal (DE) enhancer regions of the human OCT4 gene were also targeted by co-transfection of six gRNAs and either dCas9^{VP64} or dCas9^{p300}_{core} (FIG. 2B). dCas9^{p300}_{core} induced significant transcription from these regions (P-value <0.0001 and P-value <0.003 for the DE and PE, respectively), whereas dCas9^{VP64} was unable to activate OCT4 above background levels when targeted to either the PE or DE regions.

[0130] The well-characterized mammalian β-globin locus control region (LCR) orchestrates transcription of the downstream hemoglobin genes; hemoglobin epsilon 1 (HBE, from ~11 kb), hemoglobin gamma 1 and 2 (HBG, from ~30 kb), hemoglobin delta (HBD, from ~46 kb), and hemoglobin beta (HBB, from ~54 kb) (FIG. 2C). DNase hypersensitive sites within the β-globin LCR serve as docking sites for transcriptional and chromatin modifiers, including p300, which coordinate distal target gene expression. Four gRNAs targeting the DNase hypersensitive site 2 within the LCR

enhancer region (HS2 enhancer) were generated. These four HS2-targeted gRNAs were co-transfected with dCas9, dCas9^{VP64}, dCas9^{p300 Core}, or dCas9^{p300 Core (D1399Y)} and the resulting mRNA production from HBE, HBG, HBD, and HBB was assayed (FIG. 2C). dCas9, dCas9^{VP64} and dCas9^{p300 Core (D1399Y)} were unable to transactivate any downstream genes when targeted to the HS2 enhancer. In contrast, targeting of dCas9^{p300 Core} to the HS2 enhancer led to significant expression of the downstream HBE, HBG, and HBD genes (P-value <0.0001, 0.0056, and 0.0003 between dCas9^{p300 Core} and mock-transfected cells for HBE, HBG, and HBD respectively). Overall, HBD and HBE appeared relatively less responsive to synthetic p300 Core-mediated activation from the HS2 enhancer; a finding consistent with lower rates of general transcription from these two genes across several cell lines (FIGS. 9A-9E).

[0131] Nevertheless, with the exception of the most distal HBB gene, dCas9^{p300 Core} exhibited a capacity to activate transcription from downstream genes when targeted to all characterized enhancer regions assayed, a capability not observed for dCas9^{VP64}. Together, these results demonstrate that dCas9^{p300 Core} is a potent programmable transcription

factor that can be used to regulate gene expression from a variety of promoter-proximal and promoter-distal locations.

Example 4

Gene Activation by dCas9^{p300 Core} is Highly Specific

[0132] Recent reports indicate that dCas9 may have widespread off-target binding events in mammalian cells in combination with some gRNAs, which could potentially lead to off-target changes in gene expression. In order to assess the transcriptional specificity of the dCas9^{p300 Core} fusion protein, transcriptome was performed profiling by RNA-seq in cells co-transfected with four IL1RN-targeted gRNAs and either dCas9, dCas9^{VP64}, dCas9^{p300 Core}, or dCas9^{p300 Core (D1399Y)}. Genome-wide transcriptional changes were compared between dCas9 with no fused effector domain and either dCas9^{VP64}, dCas9^{p300 Core} or dCas9^{p300 Core (D1399Y)} (FIGS. 3A-3C). While both dCas9^{VP64} and dCas9^{p300 Core} upregulated all four IL1RN isoforms, only the effects of dCas9^{p300 Core} reached genome-wide significance (FIGS. 3A-3B, Table 6; P-value 1.0×10⁻³-5.3×10⁻⁴ for dCas9^{VP64}; P-value 1.8×10⁻¹⁷-1.5×10⁻¹⁹ for dCas9^{p300 Core})

TABLE 6

Ten most enriched mRNAs for dCas9 IL1RN-targeted RNA-seq experiments							
dCas9 ^{VP64} + 4 IL1RN gRNAs compared to dCas9 + 4 IL1RN gRNAs							
Refseq ID	Gene	Base Mean	log2 Fold Change	IfcSE	stat	pvalue	padj
1 NM_173842	IL1RN (transcript variant 1)	14.764	0.529	0.152	3.48	0.000494857	0.99992134
2 NM_173843	IL1RN (transcript variant 4)	13.606	0.517	0.149	3.47	0.000530109	0.99992134
3 NR_073102	ZNF551	21.505	0.505	0.159	3.17	0.00152863	0.99992134
4 NM_000577	IL1RN (transcript variant 3)	14.890	0.497	0.152	3.28	0.001039353	0.99992134
5 NM_001077441	BCLAF1 (transcript variant 3)	437.814	0.482	0.153	3.14	0.001665925	0.99992134
6 NM_173841	IL1RN (transcript variant 2)	13.711	0.448	0.15	3.00	0.002716294	0.99992134
7 NM_001268	RCBTB2	46.265	0.440	0.167	2.64	0.008335513	0.99992134
8 NM_000922	PDE3B	143.947	0.439	0.167	2.63	0.008471891	0.99992134
9 NM_001077440	BCLAF1 (transcript variant 2)	463.743	0.439	0.156	2.82	0.004790762	0.99992134
10 NM_014739	BCLAF1 (transcript variant 1)	474.598	0.432	0.158	2.74	0.006232218	0.99992134

dCas9 ^{p300 Core} + 4 IL1RN gRNAs compared to dCas9 + 4 IL1RN gRNAs							
Refseq ID	Gene	Base Mean	log2 Fold Change	IfcSE	stat	pvalue	padj
1 NM_173843	IL1RN (transcript variant 4)	45.517	1.548	0.171	9.04	1.52E-19	5.24E-15
2 NM_173841	IL1RN (transcript variant 2)	40.690	1.457	0.171	8.50	1.83E-17	3.16E-13
3 NM_173842	IL1RN (transcript variant 1)	39.568	1.448	0.171	8.45	2.88E-17	3.30E-13
4 NM_000577	IL1RN (transcript variant 3)	41.821	1.437	0.171	8.39	4.88E-17	4.20E-13
5 NM_001429	p300	928.435	0.955	0.171	5.57	2.50E-08	0.000171838
6 NM_002253	KDR	17.477	0.842	0.163	5.17	2.36E-07	0.00135472
7 NM_030797	FAM49A	21.286	0.736	0.166	4.44	8.91E-06	0.043823927
8 NM_012074	DPF3	17.111	0.609	0.164	3.72	0.000202676	0.871938986
9 NM_031476	CRISPLD2	25.148	0.569	0.167	3.41	0.000653132	0.999954424
10 NM_007365	PADI2	99.012	0.554	0.162	3.41	0.000641145	0.999954424

dCas9 ^{p300 Core (D1399Y)} + 4 IL1RN gRNAs compared to dCas9 + 4 IL1RN gRNAs							
Refseq ID	Gene	Base Mean	log2 Fold Change	IfcSE	stat	pvalue	padj
1 NM_001429	p300	935.659	1.234	0.198	6.24	4.36E-10	1.49E-05
2 NM_001270493	SREK1 (transcript variant 4)	30.118	0.651	0.203	3.20	0.001388089	0.999938051
3 NM_001079802	FKTN (transcript variant 1)	148.558	0.546	0.203	2.69	0.007212168	0.999938051
4 NM_000922	PDE3B	140.122	0.535	0.201	2.66	0.007805491	0.999938051
5 NM_206937	LIG4 (transcript variant 2)	30.589	0.521	0.203	2.56	0.010513626	0.999938051
6 NM_001136116	ZNF879	18.421	0.520	0.201	2.59	0.009600802	0.999938051
7 NM_018374	TMEM106B (transcript variant 1)	280.758	0.516	0.196	2.64	0.008329592	0.999938051

TABLE 6-continued

Ten most enriched mRNAs for dCas9 IL1RN-targeted RNA-seq experiments							
8 NM_019863	F8 (transcript variant 2)	8.048	0.515	0.178	2.89	0.003827553	0.999938051
9 NM_001193349	MEF2C (transcript variant 5)	18.934	0.510	0.202	2.53	0.011492452	0.999938051
10 NM_183245	INVS (transcript variant 2)	38.545	0.497	0.203	2.45	0.014125973	0.999938051

[0133] In contrast, dCas9^{p300 Core (D1399Y)} did not significantly induce any IL1RN expression (FIG. 3C; P-value >0.5 for all 4 IL1RN Isoforms). Comparative analysis to dCas9 revealed limited dCas9^{p300 Core} off-target gene induction, with only two transcripts induced significantly above background at a false discovery rate (FDR)<5%: KDR (FDR=1.4×10⁻³); and FAM49A (FDR=0.04) (FIG. 3B, Table 6). Increased expression of p300 mRNA was observed in cells transfected with dCas9^{p300 Core} and dCas9^{p300 Core (D1399Y)}. This finding is most likely explained by RNA-seq reads mapping to mRNA from the transiently transfected p300 core fusion domains. Thus the dCas9^{p300 Core} fusion displayed high genome-wide targeted transcriptional specificity and robust gene induction of all four targeted IL1RN isoforms.

Example 5

dCas9^{p300 Core} Acetylates H3K27 at Enhancers and Promoters

[0134] Activity of regulatory elements correlates with covalent histone modifications such as acetylation and methylation. Of those histone modifications, acetylation of lysine 27 on histone H3 (H3K27ac) is one of the most widely documented indicators of enhancer activity. Acetylation of H3K27 is catalyzed by p300 and is also correlated with endogenous p300 binding profiles. Therefore H3K27ac enrichment was used as a measurement of relative dCas9^{p300 Core}-mediated acetylation at the genomic target site. To quantify targeted H3K27 acetylation by dCas9^{p300 Core} chromatin immuno-precipitation was performed with an anti-H3K27ac antibody followed by quantitative PCR (ChIP-qPCR) in HEK293T cells co-transfected with four HS2 enhancer-targeted gRNAs and either dCas9, dCas9^{VP64}, dCas9^{p300 Core} or dCas9^{p300 Core (D1399Y)} (FIGS. 4A-4D). Three amplicons were analyzed at or around the target site in the HS2 enhancer or within the promoter regions of the HBE and HBG genes (FIG. 4A). Notably, H3K27ac is enriched in each of these regions in the human K562 erythroid cell line that has a high level of globin gene expression (FIG. 4A). Significant H3K27ac enrichment was observed at the HS2 enhancer target locus compared to treatment with dCas9 in both the dCas9^{VP64} (P-value 0.0056 for ChIP Region land P-value 0.0029 for ChIP Region 3) and dCas9^{p300 Core} (P-value 0.0013 for ChIP Region land P-value 0.0069 for ChIP Region 3) co-transfected samples (FIG. 4B).

[0135] A similar trend of H3K27ac enrichment was also observed when targeting the IL1RN promoter with dCas9^{VP64} or dCas9^{p300 Core} (FIG. 10). FIG. 10 shows the IL1RN locus on GRCh37/hg19 along with IL1RN gRNA target sites. In addition, layered ENCODE H3K27ac enrichment from seven cell lines (GM12878, H1-hESC, HSMM, HUVEC, K562, NHEK, and NHLF) is indicated with the vertical range setting set to 50. Tiled IL1RN ChIP qPCR amplicons (1-13) are also shown in corresponding locations

on GRCh37/hg19. H3K27ac enrichment for dCas9^{VP64} and dCas9^{p300 Core} co-transfected with four IL1RN-targeted gRNAs and normalized to dCas9 co-transfected with four IL1RN gRNAs is indicated for each ChIP qPCR locus assayed. 5ng of ChIP-prepared DNA was used for each reaction (n=3 independent experiments, error bars: s.e.m.).

[0136] In contrast to these increases in H3K27ac at the target sites by both dCas9^{VP64} or dCas9^{p300 Core} robust enrichment in H3K27ac at the HS2-regulated HBE and HBG promoters was observed only with dCas9^{p300 Core} treatment (FIGS. 4C-4D). Together these results demonstrate that dCas9^{p300 Core} uniquely catalyzes H3K27ac enrichment at gRNA-targeted loci and at enhancer-targeted distal promoters. Therefore the acetylation established by dCas9^{p300 Core} at HS2 may catalyze enhancer activity in a manner distinct from direct recruitment of preinitiation complex components by dCas9^{VP64}, as indicated by the distal activation of the HBE, HBG, and HBD genes from the HS2 enhancer by dCas9^{p300 Core} but not by dCas9^{VP64} (FIG. 2C, FIGS. 9A-9E).

Example 6

dCas9^{p300 Core} Activates Genes with a Single gRNA

[0137] Robust transactivation using dCas9-effector fusion proteins currently relies upon the application of multiple gRNAs, multiple effector domains, or both. Transcriptional activation could be simplified with the use of single gRNAs in tandem with a single dCas9-effector fusion. This would also facilitate multiplexing distinct target genes and the incorporation of additional functionalities into the system. The transactivation potential of dCas9^{p300 Core} with single gRNAs was compared to that of dCas9^{p300 Core} with four pooled gRNAs targeting the IL1RN, MYOD and OCT4 promoters (FIGS. 5A-5B). Substantial activation was observed upon co-transfection of the dCas9^{p300 Core} and a single gRNA at each promoter tested. For the IL1RN and MYOD promoters, there was no significant difference between the pooled gRNAs and the best individual gRNA (FIGS. 5A-5B; IL1RN gRNA “C”, P-value 0.78; MYOD gRNA “D”, P-value 0.26). Although activation of the OCT4 promoter produced additive effects when four gRNAs were pooled with dCas9^{p300 Core} the most potent single gRNA (gRNA “D”) induced a statistically comparable amount of gene expression to that observed upon co-transfection of dCas9^{VP64} with an equimolar pool of all four promoter gRNAs (P-value 0.73; FIG. 5C). Compared to dCas9^{p300 Core} levels of gene activation with dCas9^{VP64} and single gRNAs were substantially lower. Also, in contrast to dCas9^{p300 Core}, dCas9^{VP64} demonstrated synergistic effects with combinations of gRNAs in every case (FIGS. 5A-5C).

[0138] Based on the transactivation ability of dCas9^{p300 Core} at enhancer regions and with single gRNAs at promoter regions, it was hypothesized that dCas9^{p300 Core} might also be able to transactivate enhancers via a single targeted

gRNA. The MYOD (DRR and CE), OCT4 (PE and DE), and HS2 enhancer regions were tested with equimolar concentrations of pools or single gRNAs (FIGS. 5D-5G). For both MYOD enhancer regions, co-transfection of dCas9^{p300 Core} and a single enhancer-targeting gRNA was sufficient to activate gene expression to levels similar to cells co-transfected with dCas9^{p300 Core} and the four pooled enhancer gRNAs (FIG. 5D). Similarly, OCT4 gene expression was activated from the PE via dCas9^{p300 Core} localization with a single gRNA to similar levels as dCas9^{p300 Core} localized with a pool of six PE-targeted gRNAs (FIG. 5E). dCas9^{p300 Core}-mediated induction of OCT4 from the DE (FIG. 5E) and HBE and HBG genes from the HS2 enhancer (FIGS. 5F-5G) showed increased expression with the pooled gRNAs relative to single gRNAs. Nevertheless, there was activation of target gene expression above control for several single gRNAs at these enhancers (FIGS. 5E-5G).

Example 7

The p300 HAT Domain is Portable to Other DNA-Binding Proteins

[0139] The dCas9/gRNA system from *Streptococcus pyogenes* has been widely adopted due to its robust, versatile, and easily programmable properties. However, several other programmable DNA-binding proteins are also under development for various applications and may be preferable for particular applications, including orthogonal dCas9 systems from other species, TALEs, and zinc finger proteins. To determine if the p300 Core HAT domain was portable to these other systems, fusions were created to dCas9 from *Neisseria meningitidis* (Nm-dCas9), four different TALEs targeting the IL1RN promoter, and a zinc finger protein targeting ICAM1 (FIGS. 6A-6H). Co-transfection of Nm-dCas9^{p300 Core} and five Nm-gRNAs targeted to the HBE or the HBG promoters led to significant gene induction compared to mock-transfected controls (P-value 0.038 and 0.0141 for HBE and HBG respectively) (FIG. 6B). When co-transfected with five Nm-gRNAs targeted to the HS2 enhancer, Nm-dCas9^{p300 Core} also significantly activated the distal HBE and HBG, globin genes compared to mock-transfected controls (p=0.0192 and p=0.0393, respectively)

(FIGS. 6C-6D). Similar to dCas9^{p300 Core}, Nm-dCas9^{p300 Core} activated gene expression from promoters and the HS2 enhancer via a single gRNA. Nm-dCas9^{VP64} displayed negligible capacity to transactivate HBE or HBG regardless of localization to promoter regions or to the HS2 enhancer either with single or multiple gRNAs (FIGS. 6B-6D). Transfection of the expression plasmids for a combination of four TALE^{p300 Core} fusion proteins targeted to the IL1RN promoter (IL1RN TALE^{p300 Core}) also activated downstream gene expression, although to a lesser extent than four corresponding TALE^{VP64} fusions (IL1RN TALE^{VP64}) (FIGS. 6E-6F). However, single p300 Core effectors were much more potent than single VP64 domains when fused to IL1RN TALEs. Interestingly, dCas9^{p300 Core} directed to any single binding site generated comparable IL1RN expression relative to single or pooled IL1RN TALE effectors and direct comparisons suggest that dCas9 may be a more robust activator than TALEs when fused to the larger p300 Core fusion domain (FIGS. 11A-11C). The p300 Core effector domain did not display synergy with either additional gRNAs or TALEs (see FIGS. 5A-5G, 6A-6H, 9A-9E, and 11A-11C) or in combination with VP64 (see FIGS. 13A-13B). The underlying chromatin context of the dCas9^{p300 Core} target loci is shown in FIGS. 14A-14E.

[0140] The ZF^{p300 Core} fusion targeted to the ICAM1 promoter (ICAM1 ZF^{p300 Core}) also activated its target gene relative to control and at a similar level as ZF^{VP64} (ICAM1 ZF^{VP64}) (FIGS. 6G-6H). The versatility of the p300 Core fusion with multiple targeting domains is evidence that this is a robust approach for targeted acetylation and gene regulation. The various p300 core fusion proteins were expressed well, as determined by western blot (FIGS. 12A-12B), but differences in p300 Core activity between different fusion proteins could be attributable to binding affinity or protein folding.

Example 8

Myocardin

[0141] 36 gRNAs were designed to span -2000 bp to +250 bp (coordinates relative to TSS) region of the MYOCD gene (Table 7).

TABLE 7

MyoCD gRNAs Information									
Target Name	gRNA Name	Protospacer (N20)	SEQ ID NO	SEQ PAM	SEQ ID NO	+/−	Length	Coordinates Relative to TSS	
MyoCD	Cr1	cctgggtttcaatgagaaga	152	NGG	188	−	20	-1991	-1971
MyoCD	Cr2	gatttaggacatgaacatggg	153	NGG	189	−	20	-1897	-1877
MyoCD	Cr3	cctcttctacattaacctta	154	NGG	190	−	20	-1771	-1751
MyoCD	Cr4	tttttgaaggccagcaatcgt	155	NGG	191	−	20	-1693	-1673
MyoCD	Cr5	cgttagtttctggaggctct	156	NGG	192	−	20	-1597	-1577
MyoCD	Cr6	acaaattaccacgaatgtag	157	NGG	193	−	20	-1480	-1460
MyoCD	Cr7	tggcctggcgccctgttat	158	NGG	194	−	20	-1395	-1375
MyoCD	Cr8	attttgtaaataaggtttc	159	NGG	195	−	20	-1297	-1277
MyoCD	Cr9	agcaacaggggatgggcag	160	NGG	196	+	20	-1221	-1201

TABLE 7-continued

Myo cd gRNAs Information									
Target Name	gRNA Name	Protospacer (N20)	SEQ ID NO	SEQ ID NO	+/-	Length	Coordinates Relative to TSS		
				PAM					
Myo cd	Cr10	aggactcgtagtatgcaggc	161	NGG	197	+	20	-1120	-1100
Myo cd	Cr11	ctgagccaccaactatTTAA	162	NGG	198	+	20	-1005	-985
Myo cd	Cr12	ctgagccaccaactatTTAA	163	NGG	199	+	20	-945	-925
Myo cd	Cr13	actctgggtcggttacggaa	164	NGG	200	+	20	-907	-887
Myo cd	Cr14	gggctgggcttagcttggga	165	NGG	201	-	20	-837	-817
Myo cd	Cr15	atagggaggggctctggagc	166	NGG	202	-	20	-798	-778
Myo cd	Cr16	atgggaaaagataacctgagt	167	NGG	203	-	20	-751	-731
Myo cd	Cr17	tgggagcgttgtgcgcagc	168	NGG	204	+	20	-713	-693
Myo cd	Cr18	tggaaaaggcttcatttct	169	NGG	205	-	20	-642	-622
Myo cd	Cr19	gtatctcgagctccaatac	170	NGG	206	-	20	-594	-574
Myo cd	Cr20	acgcattcccctcggttga	171	NGG	207	-	20	-544	-524
Myo cd	Cr21	tccggaaagctttcttcag	172	NGG	208	+	20	-511	-491
Myo cd	Cr22	cggaaaggcggtgcgcgcccc	173	NGG	209	-	20	-449	-429
Myo cd	Cr23	cggcgcgaaaggaaagcgccc	174	NGG	210	-	20	-396	-376
Myo cd	Cr24	ggctgcgcacgcggcatcccc	175	NGG	211	+	20	-352	-332
Myo cd	Cr25	ggggcttgcagggtggttgc	176	NGG	212	-	20	-297	-277
Myo cd	Cr26	cgagctaaagagcggtgcc	177	NGG	213	-	20	-246	-226
Myo cd	Cr27	agagggcgggagcaggGCC	178	NGG	214	-	20	-200	-180
Myo cd	Cr28	aaccggctttaacttttg	179	NGG	215	-	20	-153	-133
Myo cd	Cr29	caggagcgccgagcggggtc	180	NGG	216	-	20	-101	-81
Myo cd	Cr30	gggtatcagatggcaaagt	181	NGG	217	+	20	-54	-34
Myo cd	Cr31	tcataggctgcggcgattg	182	NGG	218	-	20	0	20
Myo cd	Cr32	gagggtggccaggagcagcg	183	NGG	219	-	20	47	67
Myo cd	Cr33	aattagccccgcacggcgag	184	NGG	220	+	20	100	120
Myo cd	Cr34	tccccctgggttaggatcacag	185	NGG	221	-	20	157	177
Myo cd	Cr35	ggttgttagctgcggcagc	186	NGG	222	+	20	203	223
Myo cd	Cr36	ggtgaggaaacaggGGGCGCC	187	NGG	223	+	20	246	266

[0142] The gRNAs were cloned into a spCas9 gRNA expression vector containing hU6 promoter and BbsI restriction site. The gRNAs were transiently co-transfected with dCas9^{p300 Core} into BTEK293T cells. The resulting mRNA production for myocardin was assayed in samples harvested three days post-transfection (FIG. 17). Combinations of Cr32, Cr13, Cr30, Cr28, Cr31, and Cr34 were analyzed with dCas9^{p300 Core} (Table 8; FIG. 18).

TABLE 8

Condition	Cr32	Cr13	Cr30	Cr28	Cr31	Cr34
1	X	X	X	X	X	X
2	X	X	X	X		
3	X	X	X		X	
4	X	X	X			X
5	X	X		X	X	
6	X	X			X	X
7	X	X		X		X
8	X		X	X	X	
9		X	X	X		X
10		X		X	X	X
11		X	X	X	X	X
12	X		X	X	X	X
13	X	X		X	X	X
14	X	X	X		X	X
15	X	X	X	X		X
16	X	X	X	X	X	

Example 9

Pax7

[0143] gRNAs were designed to span the region surrounding the PAX7 gene (Table 9). The gRNAs were cloned into a spCas9 gRNA expression vector containing hU6 promoter and BbsI restriction site. The gRNAs were transiently co-transfected with dCas9^{p300 Core} or dCas9^{VP64} into HEK293T cells. The resulting mRNA production for Pax7 was assayed in samples harvested three days post-transfection (FIG. 19). The gRNA19 (“g19”) was used in further experiments and shown to localize to a DNase hypersensitive site (DHS) (FIG. 20).

TABLE 9

TSS	Target	Target	SEQ ID	
position	Strand	name	Oligo in sense strand	NO
138	AS	JK12	GGGGCGCGAGTGATCAGCT	224
27	S	JK16	CCCGGGTCTCCTAGGGGACG	225

TABLE 9-continued

Pax7 qRNAs				
TSS position	Target Strand	Target name	SEQ ID Oligo in sense strand	ID NO
+95	S	JK17	TGGTCCGGAGAAAGAAGGCG	226
+187	S	JK18	GTCCTCGGGCTCGAAACTT	227
+223	S	JK19	AGGCCAGAGCGCGAGAGCG	228
+273	S	JK20	CGATTCCGGCCGCCTCCCC	229
+335	AS	JK21	GTTGTGCGGGCTGATGCGCC	230

Example 10

FGF1

[0144] gRNAs were designed for the FGF1A, FGF1B, and FGF1C genes (Tables 10 and 11). The 25 nM of gRNAs were transiently co-transfected with dCas9^{p300 Core} or dCas9^{VP64} into HEK293T cells. The resulting mRNA production for FGF1 expression was determined (FIGS. 21-23). In FIG. 23, the number of stable cell-lines transfected with the lentivirus vector was 2, except for FGF1A where n=1.

TABLE 10

gRNA	Gene	Type	Name
1	FGF1A	F_7sk	1FGF1AF_7sk
2	FGF1A	F_h1	2FGF1AF_h1
3	FGF1A	F_hU6	3FGF1AF_hU6
4	FGF1A	F_mU6	4FGF1AF_mU6
1	FGF1A	R_7sk	1FGF1AR_7sk
2	FGF1A	R_h1	2FGF1AR_h1
3	FGF1A	R_hU6	3FGF1AR_hU6
4	FGF1A	R_mU6	4FGF1AR_mU6
1	FGF1B	F_7sk	1FGF1BF_7sk
2	FGF1B	F_h1	2FGF1BF_h1
3	FGF1B	F_hU6	3FGF1BF_hU6
4	FGF1B	F_mU6	4FGF1BF_mU6
1	FGF1B	R_7sk	1FGF1BR_7sk
2	FGF1B	R_h1	2FGF1BR_h1
3	FGF1B	R_hU6	3FGF1BR_hU6
4	FGF1B	R_mU6	4FGF1BR_mU6
1	FGF1C	F_7sk	1FGF1CF_7sk
2	FGF1C	F_h1	2FGF1CF_h1
3	FGF1C	F_hU6	3FGF1CF_hU6
4	FGF1C	F_mU6	4FGF1CF_mU6
1	FGF1C	R_7sk	1FGF1CR_7sk
2	FGF1C	R_h1	2FGF1CR_h1
3	FGF1C	R_hU6	3FGF1CR_hU6
4	FGF1C	R_mU6	4FGF1CR_mU6

TABLE 11

FGF1 qRNAs Information				
gRNA	Final Sequence	SEQ ID 1st NO addition	Sequence	SEQ ID 2nd NO addition
1	CCTCGTGTGTTCTGGGC CTGCTGC	231 CCTCG	TGTGTTCTGGGCCTG CTGC	255
2	TCCCATAAACAGGATT TGCTCAGA	232 TCCCA	TAAACAGGATTCTGCT CAGA	256

TABLE 11-continued

FGF1 gRNAs Information						
	gRNA Final Sequence	SEQ ID NO	1st addition	Sequence	SEQ ID NO	2nd addition
3	CACCGGCCAGATGACAGAACAGAAA	233	CACCG	GCCAGATGACAGAACAGAAA	257	
4	TTGTTTGAAATGCCATTGTAGGGCT	234	TTGTTTG	AAAAATGCCATTGTAGGGCT	258	
1	AAACGCAGCAGGCCAGGAACACAC	235	AAAC	GCAGCAGGCCAGGAACACAC	259	C
2	AAACTCTGAGCAGAACCTGTTTAT	236	AAAC	TCTGAGCAGAACCTGTTTAT	260	T
3	AAACTTCTGTTCTGTCACTGGCC	237	AAAC	TTTCTGTTCTGTCACTGGCC	261	C
4	AAACAGCCCTACAAATGCGATTTCAA	238	AAAC	AGCCCTACAAATGGCATTTT	262	CAA
1	CCTCGtctgcttcgtccgaacctca	239	CCTCG	tctgcttcgtccgaacctca	263	
2	TCCCCacctaagagctttaggcg	240	TCCCA	cctaaagagctttaggcg	264	
3	CACCGagagctggtaaccgtccct	241	CACCG	agagctggtaaccgtccct	265	
4	TTGTTTGcggtccttgttatcagtag	242	TTGTTTG	cggtccttgttatcagtag	266	
1	AAACtgaggttcggcagaaggcaga	243	AAAC	tgagggttcggcagaaggcaga	267	C
2	AAACccgcctacaagcttttaggT	244	AAAC	cggcctacaagcttttagg	268	T
3	AAACaggacgggtagccagctct	245	AAAC	aggacgggtagccagctct	269	C
4	AAACctactgataaacaaaaggaccgC	246	AAAC	ctactgataaacaaaaggaccg	270	CAA
1	CCTCGGAGCTGGCTACCCTTA	247	CCTCG	GAGCTGGCTACCCGTCCCTA	271	
2	TCCCCACTTGGCTGGGTTAAACCA	248	TCCCA	CTTTGGCTGGGTTAAACCA	272	
3	CACCGGTCAAGTCAGGGTTTGTTGTA	249	CACCG	GTCAGCTCAGGGTTTTGGTA	273	
4	TTGTTTGAGTTAGCTCCCGACCCAG	250	TTGTTTG	GAGTTAGCTCCCGACCCAG	274	
1	AAACTAGGGACGGGTAGCCAGCTCC	251	AAAC	TAGGGACGGGTAGCCAGCTCC	275	C
2	AAACTGGTTAACCCAGCCGCCAAAGT	252	AAAC	TGGTTAACCCAGCCGCCAAAGT	276	T
3	AAACTACCAAAACCTGAGCTGACC	253	AAAC	TACCAAAACCTGAGCTGACC	277	C
4	AAACCTGGGTGGGAGCTAACTCCAA	254	AAAC	CTGGGTGGGAGCTAACTCCAA	278	CAA

[0145] It is understood that the foregoing detailed description and accompanying examples are merely illustrative and are not to be taken as limitations upon the scope of the invention, which is defined solely by the appended claims and their equivalents.

[0146] Various changes and modifications to the disclosed embodiments will be apparent to those skilled in the art. Such changes and modifications, including without limitation those relating to the chemical structures, substituents, derivatives, intermediates, syntheses, compositions, formulations, or methods of use of the invention, may be made without departing from the spirit and scope thereof.

[0147] For reasons of completeness, various aspects of the invention are set out in the following numbered clauses:

[0148] Clause 1. A fusion protein comprising two heterologous polypeptide domains, wherein the first polypeptide domain comprises a Clustered Regularly Interspaced Short Palindromic Repeats associated (Cas) protein and the second polypeptide domain comprises a peptide having histone acetyltransferase activity.

[0149] Clause 2. The fusion protein of clause 1, wherein the fusion protein activates transcription of a target gene.

[0150] Clause 3. The fusion protein of clause 1 or 2, wherein the Cas protein comprises Cas9.

[0151] Clause 4. The fusion protein of clause 3, wherein the Cas9 comprises at least one amino acid mutation which knocks out nuclease activity of Cas9.

[0152] Clause 5. The fusion protein of clause 4, wherein the Cas protein comprises SEQ ID NO: 1 or SEQ ID NO: 10.

[0153] Clause 6. The fusion protein of any one of clauses 1-5, wherein the second polypeptide domain comprises a histone acetyltransferase effector domain.

[0154] Clause 7. The fusion protein of clause 6, wherein the histone acetyltransferase effector domain is a p300 histone acetyltransferase effector domain.

[0155] Clause 8. The fusion protein of any one of clauses 1-7, wherein the second polypeptide domain comprises SEQ ID NO: 2 or SEQ ID NO: 3.

[0156] Clause 9. The fusion protein of any one of clauses 1-8, wherein the first polypeptide domain comprises SEQ ID NO: 1 or SEQ ID NO: 10 and the second polypeptide domain comprises SEQ ID NO: 2 or SEQ ID NO: 3.

[0157] Clause 10. The fusion protein of any one of clauses 1-9, wherein the first polypeptide domain comprises SEQ ID NO: 1 and the second polypeptide domain comprises SEQ ID NO: 3, or the first polypeptide domain comprises SEQ ID NO: 10 and the second polypeptide domain comprises SEQ ID NO: 3.

[0158] Clause 11. The fusion protein of any one of clauses 1-10, further comprising a linker connecting the first polypeptide domain to the second polypeptide domain.

[0159] Clause 12. The fusion protein of any one of clauses 1-11, wherein the fusion protein comprises an amino acid sequence of SEQ ID NO: 140, 141, or 149.

[0160] Clause 13. A DNA targeting system comprising the fusion protein of any one of clauses 1-12 and at least one guide RNA (gRNA).

[0161] Clause 14. The DNA targeting system of clause 13, wherein the at least one gRNA comprises a 12-22 base pair complementary polynucleotide sequence of the target DNA sequence followed by a protospacer-adjacent motif.

[0162] Clause 15. The DNA targeting system of clause 13 or 14, wherein the at least one gRNA targets a target region, the target region comprises a target enhancer, target regulatory element, a cis-regulatory region of a target gene, or a trans-regulatory region of a target gene.

[0163] Clause 16. The DNA targeting system of clause 15, wherein the target region is a distal or proximal cis-regulatory region of the target gene.

[0164] Clause 17. The DNA targeting system of clause 15 or 16, wherein the target region is an enhancer region or a promoter region of the target gene.

[0165] Clause 18. The DNA targeting system of any one of clauses 15-17, wherein the target gene is an endogenous gene or a transgene.

[0166] Clause 19. The DNA targeting system of clause 15, wherein the target region comprises a target enhancer or a target regulatory element.

[0167] Clause 20. The DNA targeting system of clause 19, wherein the target enhancer or target regulatory element control the gene expression of more than one target gene.

[0168] Clause 21. The DNA targeting system of any one of clauses 15-20, wherein the DNA targeting system comprises between one and ten different gRNAs.

[0169] Clause 22. The DNA targeting system of any one of clauses 15-21, wherein the DNA targeting system comprises one gRNA.

[0170] Clause 23. The DNA targeting system of any one of clauses 15-22, wherein the target region is located on the same chromosome as the target gene.

[0171] Clause 24. The DNA targeting system of clause 23, wherein the target region is located about 1 base pair to about 100,000 base pairs upstream of a transcription start site of the target gene.

[0172] Clause 25. The DNA targeting system of clause 24, wherein the target region is located about 1000 base pairs to about 50,000 base pairs upstream of the transcription start site of the target gene.

[0173] Clause 26. The DNA targeting system of any one of clauses 15-22, wherein the target region is located on a different chromosome as the target gene.

[0174] Clause 27. The DNA targeting system of any one of clauses 15-28, wherein the different gRNAs bind to different target regions.

[0175] Clause 28. The DNA targeting system of clause 27, wherein the different gRNAs bind to target regions of different target genes.

[0176] Clause 29. The DNA targeting system of clause 27, wherein the expression of two or more target genes are activated.

[0177] Clause 30. The DNA targeting system of any one of clauses 15-29, wherein the target gene is selected from the group consisting of IL1RN, MYOD1, OCT4, HBE, HBG, HBD, HBB, MYOCD, PAX7, FGF1A, FGF1B, and FGF1C.

[0178] Clause 31. The DNA targeting system of clause 30, wherein the target region is at least one of HS2 enhancer of the human β-globin locus, distal regulatory region (DRR) of the MYOD gene, core enhancer (CE) of the MYOD gene, proximal (PE) enhancer region of the OCT4 gene, or distal (DE) enhancer region of the OCT4 gene.

[0179] Clause 32. The DNA targeting system of any one of clauses 13-31, wherein the gRNA comprises at least one of SEQ ID NOS: 23-73, 188-223, or 224-254.

[0180] Clause 33. An isolated polynucleotide encoding the fusion protein of any one of clauses 1-12 or the DNA targeting system of any one of clauses 13-32.

[0181] Clause 34. A vector comprising the isolated polynucleotide of clause 33.

- [0182] Clause 35. A cell comprising the isolated polynucleotide of clause 33 or the vector of clause 34.
- [0183] Clause 36. A kit comprising the fusion protein of any one of clauses 1-12, the DNA targeting system of clauses 13-32, the isolated polynucleotide of clause 33, the vector of clause 34, or the cell of clause 35.
- [0184] Clause 37. A method of activating gene expression of a target gene in a cell, the method comprising contacting the cell with the fusion protein of any one of clauses 1-12, the DNA targeting system of clauses 13-32, the isolated polynucleotide of clause 33, or the vector of clause 34.
- [0185] Clause 38. A method of activating gene expression of a target gene in a cell, the method comprising contacting the cell with a polynucleotide encoding a DNA targeting system, wherein the DNA targeting system comprises the fusion protein of any one of clauses 1-12 and at least one guide RNA (gRNA).
- [0186] Clause 39. The method of clause 38, wherein the at least one gRNA comprises a 12-22 base pair complementary polynucleotide sequence of the target DNA sequence followed by a protospacer-adjacent motif.
- [0187] Clause 40. The method of clause 38 or 39, wherein the at least one gRNA targets a target region, the target region is a cis-regulatory region or a trans-regulatory region of a target gene.
- [0188] Clause 41. The method of clause 40, wherein the target region is a distal or proximal cis-regulatory region of the target gene.
- [0189] Clause 42. The method of clause 40 or 41, wherein the target region is an enhancer region or a promoter region of the target gene.
- [0190] Clause 43. The method of clause 40-42, wherein the target gene is an endogenous gene or a transgene.
- [0191] Clause 44. The method of clause 43, wherein the DNA targeting system comprises between one and ten different gRNAs.
- [0192] Clause 45. The method of clause 43, wherein the DNA targeting system comprises one gRNA.

- [0193] Clause 46. The method of clause 40-45, wherein the target region is located on the same chromosome as the target gene.
- [0194] Clause 47. The method of clause 46, wherein the target region is located about 1 base pair to about 100,000 base pairs upstream of a transcription start site of the target gene.
- [0195] Clause 48. The method of clause 46, wherein the target region is located about 1000 base pairs to about 50,000 base pairs upstream of the transcription start site of the target gene.
- [0196] Clause 49. The method of clause 40-45, wherein the target region is located on a different chromosome as the target gene.
- [0197] Clause 50. The method of clause 40-45, wherein the different gRNAs bind to different target regions.
- [0198] Clause 51. The method of clause 50, wherein the different gRNAs bind to target regions of different target genes.
- [0199] Clause 52. The method of clause 51, wherein the expression of two or more target genes are activated.
- [0200] Clause 53. The method of clause 40-52, wherein the target gene is selected from the group consisting of IL1RN, MYOD1, OCT4, HBE, HBG, HBD, HBB, MYOCD, PAX7, FGF1A, FGF1B, and FGF1C.
- [0201] Clause 54. The method of clause 53, wherein the target region is at least one of HS2 enhancer of the human β -globin locus, distal regulatory region (DRR) of the MYOD gene, core enhancer (CE) of the MYOD gene, proximal (PE) enhancer region of the OCT4 gene, or distal (DE) enhancer region of the OCT4 gene.
- [0202] Clause 55. The method of clause 37-54, wherein the gRNA comprises at least one of SEQ ID NOs: 23-73, 188-223, or 224-254.
- [0203] Clause 56. The method of any one of clauses 37-55, wherein the DNA targeting system is delivered to the cell virally or non-virally.
- [0204] Clause 57. The method of any one of clauses 37-56, wherein the cell is a mammalian cell.

Appendix - Sequences

Streptococcus pyogenes Cas 9 (with D10A, H849A) (SEQ ID NO: 1)
MDKKYSIGLAIGTNSGVWAVITDEYKVPSSKKFVLGNTDRHSIKKNLIGALLFDSGETA
EATRLKRTARRRYTRRKNRICYLQEIIFSNEMAKVDDSFHRLLEESFLVEEDKKHERHPIF
GNIVDEVAYHEKPYTIYHLRKKLVDSTDKADLRILTYLALAHMIFKFGHFIEGDLNPDNS
DVKDLFIQLVQTYNQLFEENPINASGVDAKAIISARLSKSRRLENLIAQLPGEEKNGLFG
NLIALSLGLTPNPKNSNPDLAEDAKLQLSKDTYDDLDNLLAQIGDQYADLFLAAKNLSD
ATLLSDILRNVTEITKAPLSASIMIKYREDEHHQDLTLLKALVRQOLPEKYKEIFFDQSKNGY
AGYIDGGASQEFPYKPIKPILEKMDGTEELLVVLNREDLLFKQRTFDNGSIPHQIHGEL
HAILRRQEDFYPFLKDNRREKIEKILTFRIPYVGPLARGNSRFAMTRKSEETITPWNFEE
VVVKKGASAQSFTERMTNFDKNLPNEKVLPKHSSLLYEYFTVNEYLTVKVYVTEGMRKPA
FLSGEQKKAIVDLLFKTNRKVTQQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLL
KIIKDKDFLDNEENEDEDIVLTLLTFEDREMIEERLKTYAHLFDDKVMQKLKRRYTG
WGRLSRKLINGIRDKQSGKTIIDFLFLKSDGFANRFMQLIHDDSLTFKEDIQKAQVSGQG
DSLHEHIANLAGSPAIIKGQILQTVKVVDELVKVMGRHKPENIVIEMARENQTTQKGQKN
SRERMKRIEEGIGELGSQILKEHPVENTQQLNEKLYLYLQNGRDMYVQELDINRLSD
YDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKNYWRQLNAAKLT
QRKFDNLTKAERGGLSELDKAQFIKRQLVETRQITKXVAQILDSRMNTKYDENDKLIRE
VKVITLKS KLVSFRKDFQFYKVR EIN NYHHAD YLN AV GTALIK KPKL ESEF VYG
DYK VYD VRK MIA KSE QE I G KATA KY FF Y S N IM NFF K T E IT L A N G E I R K R P L I E T N G E T G E I
VWD KG R D F A T V R K V L S M P Q V N I V K K T E V Q T G G F S K E I I P K R N S D K L I A R K K D W D P K K
YGG F DS P T V A Y S V L V V A K V E K G K S K K L K S V K E L L G I T I M E R S S F E K N P I D F L E A K G Y K E
VKK D L I I K L P K Y S L F E L E N G R K R M L A S A G E L Q K G N E L A L P S K Y V N F L Y L A S H Y E K L K G S
P E D N E Q K Q L F V E Q H K H Y L D E I I E Q I S E F S K R V I L A D A N L D K V L S A Y N K H R D K P I R E Q A E N I
I H L F T L T N L G A P A A F K Y F D T T I D R K R Y T S T K E V I L D A T L I H Q S I T G L Y E T R I D L S Q L G G D

-continued

Appendix-Sequences

Human p300 (with L553M mutation) (SEQ ID NO: 2)

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MAENVVEPGPPSAKRPKLSSPALSASASDGTDFGSLFDLEHLDPELINSTELGLTNGGDI
NQLQTSLGMDAASKHQQLSBLRSGSSPNLNMGVGGPGVMASQAQQSSPGLGLIN
SMVKSPMTQAGLTSNPMGMGTSPNQGPQTSTGMNNSPVNQPMGNTGMNAGMN
PGMLAAGNGQGIMPNQVMNGS1GAGGRQRQNQCYPNCAGMSAGNLLTEPLQQGSPOM
GGQTGLRPQPLKMGMNNPNPYGSPYTQNPQQIGASGLGLQIQTKTVLNSNLSPFA
MDKKAAPGGMPQGPQVQGPGLVTPTVQGMGSGAHTADPEKRKLITQQQLVL
LIHAHKCQRREQANGEVRQCNLPHCRTMKVNLNMMTHCQSGKSCQVAHCASSRQIISH
WKNCTRHDPCPVCLPLKNAGDKRNQQPILTGAPEVGLGNPSSLGVGQOSAPNLSTVSQIDP
SSTERAYAALGLPYQVNQMPQTQPVQAKNQNNQGPQSPQGMRPMNSMSAPMVNG
GVGVQTPSLLSDSMLHSAINSQNPMMSENASVPSMGPMPAAQPSTTGIRKQWHEDITQ
DLRNHHLVHLVQLQAIFFTPDPALKDRRMENLVAYARKVEGDMYESANNRAEYYHLLA
EKYKIKIKELEKRRTRLQKQMLPNAAGMVPVSMNPQPMGQCPQPGMTSNCPLPDPS
MIRGSVPVNQMMPRITPQSGLNQFGQMSMAQPPIVPRQTPPLQOHGOLAQPGALNPPMG
YGPRMQQPSNQGQFLPQTQFSPSQGMNVNTNIP LAPSSGQAPVSQAQMSSSCPVNSPIMP
GSQGSHIHCPLQPALHQNSPSPVSPSRTPHTPPS1GAQQPPATTIPAPVPTPPAMPPG
PQSQALHPPPRQPTPTPPTQLQPVQPSLPAAPSADQPQQPRSQOSTAAASVPTPTAPLLP
PQPATPLSQAQPSAVSVPVNQNPSTSSTEVNSQIAEKQPSQEVKMEAKMEVDQPEPADTQ
PEDIISSESKVEDCKMESTETEERSTELEKTEIKEEDQPSATQSSPAPGQSKKLFKPEELR
QALMPTLEALYRQDPESLPFRQPVDPQQLGIPDYFDIVKSPMDLSTIKRKLDTGQYQEPW
QYVDDIWLNFNNAWLYNRKTSRVYKCSKLSEVFEQEIDPVMQSLGYCCGRKLEFSPQ
TLCYCQKQLCTIPRDATYYSYQNRYHFCCEKCFNEIQGESVSLGDDPSQPQTTINKEQFSK
RKNDTLDPELFVTECTCRKMHQICVLHHEIIWPAFVCDGCLKKSTARTRKENKFSAKR
LPSTRLGTFLENRVNDFLRRQNHPESEGEVTVRVVHASDKTVEVKPGMKARFVDSEMA
ESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSVHFFPKCLR
TAVYHEILIGYLEVKKLGYTTGHIACPPSEGDDYIFHCHPPDQKIPKPRLQEWYKK
MIDKAVSERIDFKQATEDRLTSAKELYFEGDFWPVNLEESIKELEQEEERKR
EENTSNESTDVTKGDSNAKKNNKKTSKNKSLSRGNKKPGMPNVSNLSQLIYAT
MEKHKEVFFVIRLIAQPAANSLPIVDPDPLIPCDLMGDRDAFTLARDKHLLEFSSLRRA
QWSTMCMVLVELHTQSQDRFVYTCNECKHHVETRWHCTVCEYDLCITCYNTKHNHDHK
MEKLGGLDDDESNNQQAATQSPGDSSRLSIQRCIQSLVHACQCRNANCSLPSQCKMK
RVVQHTGCKCRKTNGQCPICKQLIALCCYAHKCOCENKCPVFPCLNIKQKLRLQQQLQH
RLQQAQMLRRLPMSMQRPTVQVQGQQLPSPPTPATPTPQTPQPTSQPQPTP
PNSMPPYLPRTAQAGQVTPPTPQTAQPPPLPGPPPAAVEMAMQIQRRAET
QRQMAHVQIFQRPQIHQHMPPTMAMPMGMNPPMTRGPSPGHLEPGMGPTGMQQQPPW
SQGGLPQPQQLQSGMPRPARMSVAQHGQPLNMAPQPLGQVGVISPLKPGTVSQQALQ
NLLRTLRSPPSLQQQQSLIHLANPOLLAFIKQRAAKYANSNPOPPIPQOPGMQPGQPG
LQPPTMPGQQGVHSNPAMQNMNPQAGVQRAGLPLQQQQLQPPMGGMSPQAQQ
MNMMNHNTMPMSQFRDILRRQMMQQQQGAGPGIGPGMANHNQFQQPQGVGYPQQ
QQRMQHMMQMQGQGNGMCQIGQLPQALGAEAGASLQAYQQRLLQQQMGSPVQPNPM
SPQQHMLPNQQAQSPHLQGQQIPNSLSNQVRSPQPVSPRPSQSPHSSPSPRMQPQSPH
HVSPTQSSPBPGLVAAQANPMBQGHFASPDQNMSLSQLASNGMANLHGASATDGLS
TDNSDLNSNLSQLSTLDIH

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p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2) (SEQ ID NO: 3)

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IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPQQLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNAWLYNRKTSRVYKCSKLSEVFEQEIDPVMQSLGYCCG
RKLEFSPQTLCCYKQLCTIPRDATYYSYQNRYHFCCEKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRNKDLDPELFVTECTCRKMHQICVLHHEIIWPAFVCDGCLKKSTARTRK
ENKFSAKRLPSTRLGTFLENRVNDFLRRQNHPESEGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSV
HFFRPKCLR TAVYHEILIGYLEVKKLGYTTGHIACPPSEGDDYIFHCHPPDQKIPKP
RLEQEWYKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELYFEGDFWPVNLEESIKE
EQEEERKREENTSNESTDVTKGDSNAKKNNKKT SKNKSLSRGNKKPGMPNVSN
DLSQLKLYATMEKHKEVFFVIRLIAQPAANSLPIVDPDPLIPCDLMGDRDAFTLARDK
LEFSSLRRAQWSTMCMVLVELHTQSQD

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p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2 with D1399Y mutation) (SEQ ID NO: 4)

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IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPQQLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNAWLYNRKTSRVYKCSKLSEVFEQEIDPVMQSLGYCCG
RKLEFSPQTLCCYKQLCTIPRDATYYSYQNRYHFCCEKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRNKDLDPELFVTECTCRKMHQICVLHHEIIWPAFVCDGCLKKSTARTRK
ENKFSAKRLPSTRLGTFLENRVNDFLRRQNHPESEGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSV
HFFRPKCLR TAVYHEILIGYLEVKKLGYTTGHIACPPSEGDDYIFHCHPPDQKIPKP
RLEQEWYKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELYFEGDFWPVNLEESIKE
EQEEERKREENTSNESTDVTKGDSNAKKNNKKT SKNKSLSRGNKKPGMPNVSN
DLSQLKLYATMEKHKEVFFVIRLIAQPAANSLPIVDPDPLIPCDLMGDRDAFTLARDK
LEFSSLRRAQWSTMCMVLVELHTQSQD

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-continued

Appendix-Sequences

p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2 with 1645/1646 RR/EE mutations) (SEQ ID NO: 5)

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IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPOQLLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNNAWLWNRKTSRVYKCSKLSEVFQEIDPVMQSLGYCCG
RKEFSPOTLCCYGKQLCTIPRDATYSSYQNRYHFCFKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRKNLTDLDELFVECTECGRKMHQICVLHHEIIWPAFVCDGCLKKSARTRK
ENKFSAKRLPSTRNGTFLENRVDLFLRRQNHPESGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSV
HFFRPKCLRTAVYHEILIGLEYVKKLGYTGHIACPPSEGDDYIFHCHPPDQKIPKPK
RLQEWYKKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELPYFEGDFWPNVLEESIKEL
EQEEERKREENTSNESTDVTKGDSNAKKNNKTKSKNSSLRGNGKKPGMPNVSN
DLSQKLYATMEKHKEVFFVIRLIAGPAANSLPPIVDPDPLIPCDLMGRDAFLTLARDKH
LEFSSLREAQWSTMCMVLVELHTQSQD
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p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2 with C1204R mutation) (SEQ ID NO: 6)

```
IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPOQLLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNNAWLWNRKTSRVYKCSKLSEVFQEIDPVMQSLGYCCG
RKEFSPOTLCCYGKQLCTIPRDATYSSYQNRYHFCFKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRKNLTDLDELFVECTECGRKMHQICVLHHEIIWPAFVCDGCLKKSARTRK
ENKFSAKRLPSTRNGTFLENRVDLFLRRQNHPESGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSV
HFFRPKCLRTAVYHEILIGLEYVKKLGYTGHIACPPSEGDDYIFHCHPPDQKIPKPK
RLQEWYKKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELPYFEGDFWPNVLEESIKEL
EQEEERKREENTSNESTDVTKGDSNAKKNNKTKSKNSSLRGNGKKPGMPNVSN
DLSQKLYATMEKHKEVFFVIRLIAGPAANSLPPIVDPDPLIPCDLMGRDAFLTLARDKH
LEFSSLRRAQWSTMCMVLVELHTQSQD
```

p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2 with Y1467F mutation) (SEQ ID NO: 7)

```
IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPOQLLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNNAWLWNRKTSRVYKCSKLSEVFQEIDPVMQSLGYCCG
RKEFSPOTLCCYGKQLCTIPRDATYSSYQNRYHFCFKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRKNLTDLDELFVECTECGRKMHQICVLHHEIIWPAFVCDGCLKKSARTRK
ENKFSAKRLPSTRNGTFLENRVDLFLRRQNHPESGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSV
HFFRPKCLRTAVYHEILIGLEYVKKLGYTGHIACPPSEGDDYIFHCHPPDQKIPKPK
RLQEWYKKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELPYFEGDFWPNVLEESIKEL
EQEEERKREENTSNESTDVTKGDSNAKKNNKTKSKNSSLRGNGKKPGMPNVSN
DLSQKLYATMEKHKEVFFVIRLIAGPAANSLPPIVDPDPLIPCDLMGRDAFLTLARDKH
LEFSSLRRAQWSTMCMVLVELHTQSQD
```

p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2 with 1396/1397 SY/WW mutations) (SEQ ID NO: 8)

```
IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPOQLLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNNAWLWNRKTSRVYKCSKLSEVFQEIDPVMQSLGYCCG
RKEFSPOTLCCYGKQLCTIPRDATYSSYQNRYHFCFKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRKNLTDLDELFVECTECGRKMHQICVLHHEIIWPAFVCDGCLKKSARTRK
ENKFSAKRLPSTRNGTFLENRVDLFLRRQNHPESGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYIWLDLS
VHFFRPKCLRTAVYHEILIGLEYVKKLGYTGHIACPPSEGDDYIFHCHPPDQKIPKPK
RLQEWYKKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELPYFEGDFWPNVLEESIKE
LEQEEERKREENTSNESTDVTKGDSNAKKNNKTKSKNSSLRGNGKKPGMPNVSN
NDLSQKLYATMEKHKEVFFVIRLIAGPAANSLPPIVDPDPLIPCDLMGRDAFLTLARDKH
LEFSSLRRAQWSTMCMVLVELHTQSQD
```

p300 Core Effector (aa 1048-1664 of SEQ ID NO: 2 with H1415A, E1423A, Y1424A, L14285, Y1430A, and H1434A mutations) (SEQ ID NO: 9)

```
IFKPEELRQALMPTLEALYRQDPESLPFRQPVDPOQLLGIPDYFDIVKSPMDLSTIKRKLDT
GQYQEPWQYVDDIWLMPNNNAWLWNRKTSRVYKCSKLSEVFQEIDPVMQSLGYCCG
RKEFSPOTLCCYGKQLCTIPRDATYSSYQNRYHFCFKCFNEIQGESVSLGDDPSQPQTT
INKEQFSKRKNLTDLDELFVECTECGRKMHQICVLHHEIIWPAFVCDGCLKKSARTRK
ENKFSAKRLPSTRNGTFLENRVDLFLRRQNHPESGEVTVRVVHASDKTVEVKPGMKAR
FVDSGEMAESFPYRTKALFAFEEDGVDLCCFGMHVQEYGSDCPPNQRRVYISYLDSV
HFFRPKCLRTAVYAEILIGLYLAALKSGATTGAIWACPPSEGDDYIFHCHPPDQKIPKPK
RLQEWYKKMLDKAVSERIVHDYKDI FKQATEDRLTSAKELPYFEGDFWPNVLEESIKEL
EQEEERKREENTSNESTDVTKGDSNAKKNNKTKSKNSSLRGNGKKPGMPNVSN
DLSQKLYATMEKHKEVFFVIRLIAGPAANSLPPIVDPDPLIPCDLMGRDAFLTLARDKH
LEFSSLRRAQWSTMCMVLVELHTQSQD
```

-continued

Appendix-Sequences

Neisseria meningitidis Cas9 (with D16A, D587A, H588A, and N611A mutations) (SEQ ID NO: 10)

MAAFKPNPINYLGLAIGIASVGWAMVEIDEDENPCLIDLGVRVFERAEVPKTGDSLAM
ARRLARSVRRRLTRRRHRRLLRREGVLQAADFDENLIKSLPNTPWQLRAAAL
DRKLTPLEWSAVLLHLIKHRYGLSQRKNEGETADKELGALLKGVAADNAHALQTDGFRT
PAELALNKFEKESEGHIRNQRGDYSHTFSRKDLQAEILLFEQKQEGFNGPHVSGGLKEGIE
TLLMTQRPALSGDAVQKMLGHCTFEPAAPEPKAAKNTYTAERFIWLTKLNNLRILEQGSER
PLTDTERATLMDPEYRKSKLTYAQARKLGLLEDTAFKGLRYGKDNEAESTLMEAKA
YHAISRALEKEGLKDCKSPNLNSPELQDEIGTAFSLFKTEDITGRLKDRIQPELEALLKH
ISFDKFVQISLKLRRVPLMEQGKRYDEACAEIYGDHYGKNTKEEYLPPIPADEFIRNP
VVLRALSQARKINGVRRYSPAHIIETAREVGKSFDRKEIEKRQEENRKDREKAA
AKFREYFPNFVGEPKSKDILKLRLYEQQHKGKCLYSGKEINLGLNEKGYVEIAAALPFSR
TWDDSFNKKVLVLSGEAQNKGNQTPYEYFNNGKDNRSREWQEFKARVETSRFPRSKQRI
LLQKFDEDGFKERNLNDTRYVNFRLCQFVADRMRLTGKGKRVFASNGQITNLLRGFW
GLRKVRAENDRRHALDAAAACSCTVAMQQKITRFVRYKEMNAFDGKTIDKETGEVLH
QKTHFPWPWEFFAQEVMRVFPKPDGKPEFEAADTPEKRLTLLAEKLSSRPEAVHEYVTP
LFVSRAPRNKMSGQGHMETVKSAKRLDEGVSVLRVPLTQLKLKDLEKMVNREREPKL
YEALKARLEAHKDDPAKAFAYFYKDGAGNRTOQVKAVRVEQVQKTGVVWRNHNG
IADNATMVRVDPEKGDKEYLPIYSWQVAKGILPDRAVVQGKDEEDWQLIDDSFNFK
FSLHPNDLVEVITKKARMFGYFASCHRG TGNIINIRIHLDHKIGNGILEGIGVKTALSFPQ
KYQIDEGLKEIRPCRLKKRPPVR

3X "Flag" Epitope (SEQ ID NO: 11)
DYKDHDGDYKDHDIDYKDDDK

Nuclear Localization Sequence (SEQ ID NO: 12)
PKKKRKVG

HA Epitope (SEQ ID NO: 13)
YPYDVPDYAS

VP64 Effector (SEQ ID NO: 14)
DALDDDFDLDMLGSDALDDFDLDMGSDALDDFDLDM

SEQUENCE LISTING

Sequence total quantity: 278

SEQ ID NO: 1 moltype = AA length = 1368
FEATURE Location/Qualifiers
source 1..1368
mol_type = protein
organism = Streptococcus pyogenes

SEQUENCE: 1

MDKYSIGLA IGTNSVGWAV ITDEYKVPSK KFKVLGNNTDR HSIKKNLIGA LLFDSGETA
ATRLKRTARR RYTRRKNRIC YLQEIFSNEA AKVDDSFHRR LEESFLVEED KKHERHPIFG
NIVDEVAYHE KYPTIYHLRK KLVSTDKA LRLIYLALAH MIKFRGHFLI EGDLNPDNSD
VDKLFQIQLVQ TYNQFLFEENP INASGVDAKA ILSARLSKSR RLENLIAQLP GEKKNGLFGN
LIALSLGLTP NFKSNFLDAE DAQKLQLSQTDT YDDDDLNLLA QIGDQYADLF LAAKNLSDAI
LLSDILRVNT EITKAPLSAS MIKRYDEHHQ DLTLKLALVR QOLPEKYKEI FFDQSKNGYA
GYIDGGASQE EFYKFKIPIL EKMDGTEELL VKLNREDLLR KQRTFDNGSI PHQIHLGEHL
AIZRRQDFY PFLKDNRKEK EKILTRFRPY YVGPLARGNS RFAWMTRKSE ETITPWNFEE
VVVKGASAQS FIERMTNFDPK NLPNPKVLPK HSLLYEEYFTV YNELTVKVYV TEGMRKP AFL
SGBQKKAIVD LLFKTMNRKVY VQQLKEDYFP KIECFDSEVI SGVEDRFNAS LGTYHDLKI
IKDKDFLDNE ENEDILEDIV LTTLTFEDRE MIEERLKTYA HLFDDKVMKQ LKRRRTGNG
RLSRRKNGI RDKQSGKTTL RNMFMQLIHDD SLTFKEDIQK AQVSQGDSDL
HEHIANLAGS KAKGKILQTV KVVKVDELKV MGRHKPENIV IEMARENQTT QKGQKNSRER
MKRIEEGIKE LGSQILKEHP VENTQLQNEK LYLYYLQNGR DMVVDQELDI NRSLSDYDVDA
IVPQSFLLKDD SIDNKVLTRS DKNRGKSDNV PSEEVVKMMK NYWRQLLNAK LITQRKFDSL
TKAERGLSE LDKAGFIKRQ LVTTRQITKH VAOILDLSRMN TKYDENDKLL REVKVITLKS
KLVSDFRKDF QFYKVREINN YHHAHDAYLN AVVGTALIKK YPKLESEFVY GDYKVYDVRK
MIAKSEQBIG KATAKYFFFYS NIMNFFKTEI TLANGEIRKR PLIETNGETG EIVWDKGDRF
ATVRKVLSMP QVNIVKKTEV QTGGFSKESI LPKRNSDKLI ARKKDWDPKK YGGFDSPV
YSVLVVAKVE KGKSKKLKSV KELLGITIME RSSFEKNPID FLEAKGYKEV KKDLI1KLPK
YSLFELENGR KRMLASAGEL KQGNEALAPS KYVNFLYLAS HYEKLKGSP E DNEQKQLFVE
QHKHYLDEII EQISEFSKRV ILADANLKV LSAYNKHDK PIREQAEINII HLFTLTNLGA
PAAFKYFDTT IDRKRYTSTK EVLDATLHQ SITGLYETRI DLSQLGGD

SEQ ID NO: 2 moltype = AA length = 2414
FEATURE Location/Qualifiers

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source 1..2414
 mol_type = protein
 organism = Homo sapiens

SEQUENCE: 2

MAENVVEPGP PSAKRPKLSS PALSASASDG TDFGSLFDLE HDLPDELINS TELGLTNGGD 60
 INQLQTSLGM VQDAASKHKQ LSELLRSGSS PNLNMGVGGP QVVMASQAQQ SSPGLGLINS 120
 MVKSPMTQAG LTSPNMGMT SGPNQGPTQS TGMMNSPVNQ PAMGMNTGMN AGMNPGLMLAA 180
 GNGQGIMPMNQ VMNGS1GAGE GRQNMQYPNP GMGSAGNLIT EPLQQGSPQM GGQTGLRGPO 240
 PLKMGMMNNP NPYGSPYTQN PGQQIGASGL GLQIQTKTVEL SNNLSPFAMD KKAVPGGGMP 300
 NMQOQPAPQV QOPGLTPVVA QGMGSGAHTA DPEKRKLIIQO QLVLLLHAH CQRREQANGE 360
 VRQCNLPHCR TMKVNVLNHTM HCQSQSGSCV AHCASSRQII SHWKNCNTRHD CPVCLPLKNA 420
 GDKRNQQPIL TGAPVGLGNP SSLGVGQQSA PNLSLTVSQID PSSIERAYAA LGLPYQVNQM 480
 PTQPQVQAKN QQNQQPQGSP QGMRPMNSMS ASPMGVNGV GVQTPSSLSD SMLHSAINSQ 540
 NPMMSENASV PSMGPMPMIAA QPSTTGTGK WHEDITQDLR NHLVHKLVQA IFPTPDPAAL 600
 KDRRMENLVA YARKVEGDMY ESANNEAEYY HLLAEKIYK1 QKEEKKRT RLQKQNMPLN 660
 AAGMVPVSMN PGPNMGQOPQ GMTSNGPLPD PSMIRGSVPN QMMPRTTPQS GLNQFGQMSM 720
 AQPPIVPRQT PPLQHHGQLA QPGALNPPMG YGPRMQQPSN QGQFLPQTQF PSQGMNVNTNI 780
 PIAPSSQAOQ VSQAOQMSSSS CPVNSPIMP GSQGSHIHCP QLPQPAHLQH SPSPVPSRTP 840
 TPHHTPPSISI AQQPPATTIP APVPTPQPM PGPQSQALHP PPRQTPTPPT TQLPQQVQPS 900
 LPAPAPSADQP QQQPRSQST AASVPTPTAP LLPPQPATPL SQAPAVSIEGQ VSNPSTST 960
 EVNSQIAEAK QPSQEVKMEA KMEVDQPEPA DTQPEDISES KVEDCKMEST ETEERSTELK 1020
 TEIKEEEDQP STSATQSSPA PGQSKKKPK PEELRQALMP TLEALYRQDP ESLPFRQPVD 1080
 PQLLGIPDYF PDKVSPMDLS TIKRKLDTGQ YQEPWQYVDD IWL MFNNNAWL YNRKTSRVYK 1140
 YCSKLSEVFE QEIDPVMQSL GYCCGRKLEF SPQTLCCYKGK QLCLTIPRDAY YYSYQNRYHF 1200
 CEKCFNEIQG ESVSLGDDPS QPQTTINKEQ FSKRKNLTD LPELFVCTEC GRKMHQICCVL 1260
 HHEIIIWPAFG VCDGCLKKSA RTRKENKFSA KRLPSTRRLGT FLENRVMDFL RRQNHPESGE 1320
 VTVRVVHASD KTVEVPGKGM ARFVDSGEMA ESFPPYRTKAL FAFFEEIDGVD LCFFGMHVQE 1380
 YGSDCPPPNQ RRVYIYLDs VHFFRPKCLR TAVYHEILIG YLEYVKKLG Y TTGHIVACPP 1440
 SEGDYYIFHC HPPDQKIPKP KRLQEWYKKM LDKAVSERIV HDYKDFIKQA TEDRLTSAKE 1500
 LPYFEGDFWP NVLEESIKEL EQUEEERERKRE ENTSNESTDV TKGDSKNAKK KNNKKTTSKN 1560
 SSLSRGNKKK PGMPNVSNDL SQKLYATMKEK HKEVFFVIRL IAGPAANSLP PIVDPDPLIP 1620
 CDLMGDGRDAF LTLLARDKHL E FSSLRRAQWS TMCMVLVELHT QSQDRFVYTC NECKHHVETR 1680
 WHCTVCEDYD LCITCYNTKN HDHKMEKGLG GLDDESNQQ AAATQSPGDS RRLS1QRCIQ 1740
 SLVHACQCRN ANCSLSPSCN MKRVRVQHKTG CKRKTNNGCP ICKQLIALCC YHAKHCQENK 1800
 CPVPFCNLKQ QKLRLQQOLQH RLQQAQMLRR RMASMQRTGVB GVGQQGLPSP TPATPTTPTG 1860
 QQPPTPQPTPQ PTSQPOPTPP NSMPMPLPRT QAAQPVSQKQ AAGQVTPTP PQTAQPLPLPG 1920
 PPPAAVEMAM QIQRAAETQR QMAHVQIFQR P1QHQMPMPT PMAPMGMNPP PMTRGPSGHL 1980
 EPGMGPTGMO QQPWSQGGL PQQPQLQSGM PRPAMMSVAQ HGQPLNMAPQ PGLGQVGISP 2040
 LKPGTVSSQQA LQNLLRTLRS PSSPLQVQ LSI LHNAPL LAAFIKQRAA KYANSNPQPI 2100
 PGQPGMPGQG PGLQPPPTMP QGGVHSNPAM QNNNPMQAGV QRAGLPQQQP QQQLQPPMGG 2160
 MSPQAQQMNM NHINTMPSQFR DILRRQQMMQ QQQQQGAGPG IGPGMANHNQ FQQQPGVGYP 2220
 PQQQORMQHH MQQMQQGNMG QIGQLPQALG AEAGASLQAY QRQLLQQQMG SPVQPNPMSP 2280
 QQQMLPNQAOQ SPHLQGQIP NSLSNQVRSP QPVPSPRQOS QPPHSSPSPR MQQPSPHHV 2340
 SPQTSSPHPG LVAAQANPME QGHFASPDQN SMLSQLASNP GMANLHGASA TDLGLSTDNS 2400
 DLNSNLSQST LDIH 2414

SEQ ID NO: 3 moltype = AA length = 617
 FEATURE Location/Qualifiers
 source 1..617
 mol_type = protein
 organism = Homo sapiens

SEQUENCE: 3

IFKPEELRQA LMPTLEALYR QDPESLPFRQ PVDPQLLGIP DYFDIVKSPM DLSTIKRKL 60
 TGQYQEPWQY VDDIWL MFNN AWLYN RKT SR VYKYCSKLSE VFEQEIDPVM QSLGYCCGRK 120
 LEFSPQTLCC YGKQLCTIPR DATYSSYQNR YHFC EKCFNE IQGESVSLGD DPSQPTTIN 180
 KEQFSKRKND TLDPELFVEC TECGRKMHQI CVLHHEIIW P AGFVCDGCLK KSARTRKENK 240
 FSAKRLPSTR LGTFLENRVN DFLRRQNHPE SGEVTVRVVH ASDKTVEVKP GMKARFVDSL 300
 EMAESFPYRT KALFAFEEID GVDLCFFGMH VQEYGSDCPP PNQRRVYISY LDSVHFFRPK 360
 CLRTAVYHEI LIGYLEYVKK LGYTTGHIWA CPPSEGDDYI FHCHPPDQKI PKPKRKLQEWY 420
 KMKLDKAVSE RIVHDYKDIQ KQATEDRLTS AKELPYFEGD FWPVNLEESI KELEQEEER 480
 KREBNTNES TDVTKGDSK AKKKNNKKT KNKSSLRGN KKKPGMPNVS NDLSQKLYAT 540
 MEKHKEVFFF IRLIAGPAAN SLPIVDPDP LIPCDLMDGR DAFTLARDK HLEFSSLRRA 600
 QWSTMCMVLE LHTQSQD 617

SEQ ID NO: 4 moltype = AA length = 617
 FEATURE Location/Qualifiers
 source 1..617
 mol_type = protein
 organism = Homo sapiens

SEQUENCE: 4

IFKPEELRQA LMPTLEALYR QDPESLPFRQ PVDPQLLGIP DYFDIVKSPM DLSTIKRKL 60
 TGQYQEPWQY VDDIWL MFNN AWLYN RKT SR VYKYCSKLSE VFEQEIDPVM QSLGYCCGRK 120
 LEFSPQTLCC YGKQLCTIPR DATYSSYQNR YHFC EKCFNE IQGESVSLGD DPSQPTTIN 180
 KEQFSKRKND TLDPELFVEC TECGRKMHQI CVLHHEIIW P AGFVCDGCLK KSARTRKENK 240
 FSAKRLPSTR LGTFLENRVN DFLRRQNHPE SGEVTVRVVH ASDKTVEVKP GMKARFVDSL 300
 EMAESFPYRT KALFAFEEID GVDLCFFGMH VQEYGSDCPP PNQRRVYISY LYSVHFFRPK 360

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CLRTAVYHEI	LIGYLEYVKK	LGYYTGHIA	CPPSEGDDYI	FHCHPPDQKI	PKPKRLQEWY	420
KMMLDKAVSE	RIVHDYKDIF	KQATEDRLTS	AKELPYFEGD	FWPNVLEESI	KELEQEEER	480
KREENTSNES	TDVTKGDSKN	AKKKNNKCTS	KNKSSLRGN	KKPGMPNVS	NDLSQKLYAT	540
MEKHKEVFFV	IRLIAGPAAN	SLPPIVDPDP	LIPCDLMDGR	DAFLTLARDK	HLEFSSLRRA	600
QWSTMCMMLVE	LHTQSQD					617

SEQ ID NO: 5	moltype = AA	length = 617
FEATURE	Location/Qualifiers	
source	1..617	
	mol_type = protein	
	organism = Homo sapiens	

SEQUENCE: 5						
IFKPEELRQA	LMPTLEALYR	QDPESLPFRQ	PVDPQLLGIP	DYFDIVKSPM	DLSTIKRKLD	60
TGQYQEPWQY	VDDIILMFNN	AWLYNRKTSR	VYKYCSKLSE	VFEQEIDPVM	QSLGYCCGRK	120
LEFSPQTLCC	YGKQLCTIPR	DATYSSYQNR	YHPCFKCFNE	IQGESVSLGD	DPSQPQTIN	180
KEQFSKRKND	TLDPELFVEC	TECGRKMHQI	CVLHHEIIW	AGFVCDGCLK	KSARTRKENK	240
FSAKRLPSTR	LGTFLNRVN	DFLRRQNHP	SGEVTVRVHH	ASDKTVEVKP	GMKARFVDSL	300
EMAESFPYRT	KALFAFEIID	GVDLCFFGMH	VQBYGSDCP	PNQRRVYISY	LDSVHFPRPK	360
CLRTAVYHEI	LIGYLEYVKK	LGYYTGHIA	CPPSEGDDYI	FHCHPPDQKI	PKPKRLQEWY	420
KMMLDKAVSE	RIVHDYKDIF	KQATEDRLTS	AKELPYFEGD	FWPNVLEESI	KELEQEEER	480
KREENTSNES	TDVTKGDSKN	AKKKNNKCTS	KNKSSLRGN	KKPGMPNVS	NDLSQKLYAT	540
MEKHKEVFFV	IRLIAGPAAN	SLPPIVDPDP	LIPCDLMDGR	DAFLTLARDK	HLEFSSLRRA	600
QWSTMCMMLVE	LHTQSQD					617

SEQ ID NO: 6	moltype = AA	length = 617
FEATURE	Location/Qualifiers	
source	1..617	
	mol_type = protein	
	organism = Homo sapiens	

SEQUENCE: 6						
IFKPEELRQA	LMPTLEALYR	QDPESLPFRQ	PVDPQLLGIP	DYFDIVKSPM	DLSTIKRKLD	60
TGQYQEPWQY	VDDIILMFNN	AWLYNRKTSR	VYKYCSKLSE	VFEQEIDPVM	QSLGYCCGRK	120
LEFSPQTLCC	YGKQLCTIPR	DATYSSYQNR	YHPCFKCFNE	IQGESVSLGD	DPSQPQTIN	180
KEQFSKRKND	TLDPELFVEC	TECGRKMHQI	CVLHHEIIW	AGFVCDGCLK	KSARTRKENK	240
FSAKRLPSTR	LGTFLNRVN	DFLRRQNHP	SGEVTVRVHH	ASDKTVEVKP	GMKARFVDSL	300
EMAESFPYRT	KALFAFEIID	GVDLCFFGMH	VQBYGSDCP	PNQRRVYISY	LDSVHFPRPK	360
CLRTAVYHEI	LIGYLEYVKK	LGYYTGHIA	CPPSEGDDYI	FHCHPPDQKI	PKPKRLQEWY	420
KMMLDKAVSE	RIVHDYKDIF	KQATEDRLTS	AKELPYFEGD	FWPNVLEESI	KELEQEEER	480
KREENTSNES	TDVTKGDSKN	AKKKNNKCTS	KNKSSLRGN	KKPGMPNVS	NDLSQKLYAT	540
MEKHKEVFFV	IRLIAGPAAN	SLPPIVDPDP	LIPCDLMDGR	DAFLTLARDK	HLEFSSLRRA	600
QWSTMCMMLVE	LHTQSQD					617

SEQ ID NO: 7	moltype = AA	length = 617
FEATURE	Location/Qualifiers	
source	1..617	
	mol_type = protein	
	organism = Homo sapiens	

SEQUENCE: 7						
IFKPEELRQA	LMPTLEALYR	QDPESLPFRQ	PVDPQLLGIP	DYFDIVKSPM	DLSTIKRKLD	60
TGQYQEPWQY	VDDIILMFNN	AWLYNRKTSR	VYKYCSKLSE	VFEQEIDPVM	QSLGYCCGRK	120
LEFSPQTLCC	YGKQLCTIPR	DATYSSYQNR	YHPCFKCFNE	IQGESVSLGD	DPSQPQTIN	180
KEQFSKRKND	TLDPELFVEC	TECGRKMHQI	CVLHHEIIW	AGFVCDGCLK	KSARTRKENK	240
FSAKRLPSTR	LGTFLNRVN	DFLRRQNHP	SGEVTVRVHH	ASDKTVEVKP	GMKARFVDSL	300
EMAESFPYRT	KALFAFEIID	GVDLCFFGMH	VQBYGSDCP	PNQRRVYISY	LDSVHFPRPK	360
CLRTAVYHEI	LIGYLEYVKK	LGYYTGHIA	CPPSEGDDYI	FHCHPPDQKI	PKPKRLQEWY	420
KMMLDKAVSE	RIVHDYKDIF	KQATEDRLTS	AKELPYFEGD	FWPNVLEESI	KELEQEEER	480
KREENTSNES	TDVTKGDSKN	AKKKNNKCTS	KNKSSLRGN	KKPGMPNVS	NDLSQKLYAT	540
MEKHKEVFFV	IRLIAGPAAN	SLPPIVDPDP	LIPCDLMDGR	DAFLTLARDK	HLEFSSLRRA	600
QWSTMCMMLVE	LHTQSQD					617

SEQ ID NO: 8	moltype = AA	length = 617
FEATURE	Location/Qualifiers	
source	1..617	
	mol_type = protein	
	organism = Homo sapiens	

SEQUENCE: 8						
IFKPEELRQA	LMPTLEALYR	QDPESLPFRQ	PVDPQLLGIP	DYFDIVKSPM	DLSTIKRKLD	60
TGQYQEPWQY	VDDIILMFNN	AWLYNRKTSR	VYKYCSKLSE	VFEQEIDPVM	QSLGYCCGRK	120
LEFSPQTLCC	YGKQLCTIPR	DATYSSYQNR	YHPCFKCFNE	IQGESVSLGD	DPSQPQTIN	180
KEQFSKRKND	TLDPELFVEC	TECGRKMHQI	CVLHHEIIW	AGFVCDGCLK	KSARTRKENK	240
FSAKRLPSTR	LGTFLNRVN	DFLRRQNHP	SGEVTVRVHH	ASDKTVEVKP	GMKARFVDSL	300
EMAESFPYRT	KALFAFEIID	GVDLCFFGMH	VQBYGSDCP	PNQRRVYISY	LDSVHFPRPK	360
CLRTAVYHEI	LIGYLEYVKK	LGYYTGHIA	CPPSEGDDYI	FHCHPPDQKI	PKPKRLQEWY	420
KMMLDKAVSE	RIVHDYKDIF	KQATEDRLTS	AKELPYFEGD	FWPNVLEESI	KELEQEEER	480
KREENTSNES	TDVTKGDSKN	AKKKNNKCTS	KNKSSLRGN	KKPGMPNVS	NDLSQKLYAT	540
MEKHKEVFFV	IRLIAGPAAN	SLPPIVDPDP	LIPCDLMDGR	DAFLTLARDK	HLEFSSLRRA	600

-continued

QWSTMCMVLE LHTQSQD	617
SEQ ID NO: 9	moltype = AA length = 617
FEATURE	Location/Qualifiers
source	1..617
	mol_type = protein
	organism = Homo sapiens
SEQUENCE: 9	
IFKPPELROA LMPTLEALYR QDPESLPFRQ PVDPQLLGIP DYFDIVKSPM DLSTIKRKLD	60
TGQYQEPWQY VDDIWLMLFNN AWLYNRKTSR VYKYCSKLSE VFEQEIDPVM QSLGYCCGRK	120
LEFPSPQTLLC YGKQLCTIPR DATYYSYQNR YHFCFKCFNE IQGESVSLGD DPSQQTIN	180
KEQFSKRKNL TLDPELFVEC TECGRKMHQI CVLHHEIWP AGFVCDGCLL KSARTRKENK	240
FSAKRLPSTR LGTFLENRVN DFLRRQNHP ESGEVTVRVHH ASDKTVEVKP GMKARFVDSL	300
EMAESFPYRT VQKMLGHCTF GVDVLAFFGMH VQBYGSDCPP PNQRRVVIY LDSVHFPRPK	360
CLRATAVYAEI LIGYLAALKK SGATTGAIWA CPPSEGDDYI FHCHPPDQKI PKPKRLQEWY	420
KKMLDKAVSE RIVHDYKDIK QKATEDRLTS AKELPYFEGD FWPVNLEESI KELEQEEEER	480
KREENTSNES TDVTKGDSKN AKKKNNKTKS KNKSSLRGN KKPGMPNVS NDLSQKLYAT	540
MEKHKEVFV IRILIAGPAAN SLPPIVDPDP LIPCDLMDGR DAFLTLARDK HLEFSSLRRA	600
QWSTMCMVLE LHTQSQD	617
SEQ ID NO: 10	moltype = AA length = 1082
FEATURE	Location/Qualifiers
source	1..1082
	mol_type = protein
	organism = Neisseria meningitidis
SEQUENCE: 10	
MAAFKPNPIN YILGLAIGIA SVGWAMVEID EDENPICLID LGVRVFERAE VPKTGDSLAM	60
ARRLARSVRR LTRRRRAHRLL RARRLLKREG VLQAADFDEN GLIKSLPNTP WQLRAALDR	120
KLTPLEWSAV LLHLIKHRYG LSQRKNEGET ADKELGALLK GVADNAHALQ TGDFRTPAEL	180
ALNKFEKESG HIRNQRGDY HTFSPKDLQA ELILLFEEKQK EFGNPVHSGG LKEGIETLLM	240
TQRPALSGDA VQKMLGHCTF EPAEPKAAKN TYTAERFIWL TKLNLLRILE QGSERPLTDT	300
ERATLMDEPY RKS KLTYAQA RKLLGLEDTA FFKGRLRYGKD NAEASTLMEM KAYHAISRAL	360
EKEGLKDKKS PLNLSPELQD EIGTAFSLFK TDDEDITGRKLK DRIQPEILEA LLKHISFDKF	420
VQISLKLARR IVPMLEQGKR YDECAEYIG DHYGKKNTEE KIYLPPIPAD EIRNPVVLRA	480
LSQARKVING VVRRYGS PAR IHETAREVG KSF KDRKEIE KRQEENRKDR EKAAAKFREY	540
FPPNFVGEPKS KDILKLRLYE QQHGKCLYSG KEINLGRNLN KGYVEIAAL PFSRTWDASF	600
NNKVVLVGSE AQNKGNQTPY EYFNKGDNRSR EWQEKFARVE TSRFPERSKQ RILLQKFDED	660
GFKERNLNDT RYVNRLFCQF VADMRLT GKKR VFASNG QITNLRLRGFW GLRKVRAEND	720
RHHALDVAVV ACSTVAMQKQ ITRFVMM NAPDGKTI DK ETGEVLHQKT HFPQPWEFFA	780
QEV MIRVFGK PDGKPEFEEA DTPEKLRLL AEKLSSRPEA VHEYVTPLFV SRAPNRKMSG	840
QGHMETVKSA KRLDEGVSVL RVPLTQLKLK DLEKMVNRE R EPKLYE ALKA RLEAHKDDPA	900
KAF AEPFV KY DKAGNRTQOV KAVR VEQVQK TGVV VRHN HNG IADNATMVRV DVFEKGDKYY	960
LVP IYSWQVA KGILPDRAVV QGK DEEDWQL IDDSFNFKFS LHPNDLVEVI T KKARMFGYF	1020
ASCHRG TGNI NIRI HDL DHK IGKNGILEGI GVK TALS FQK YQIDELGKEI RPC RLKKRPP	1080
VR	1082
SEQ ID NO: 11	moltype = AA length = 22
FEATURE	Location/Qualifiers
REGION	1..22
source	note = Description of Artificial Sequence: Synthetic peptide 1..22
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 11	
DYKDHGDYK DH DIDYK DDD DK	22
SEQ ID NO: 12	moltype = AA length = 8
FEATURE	Location/Qualifiers
REGION	1..8
source	note = Description of Artificial Sequence: Synthetic peptide 1..8
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 12	
PKKKRKVG	8
SEQ ID NO: 13	moltype = AA length = 10
FEATURE	Location/Qualifiers
REGION	1..10
source	note = Description of Artificial Sequence: Synthetic peptide 1..10
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 13	
YPYDVPDYAS	10

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SEQ ID NO: 14	moltype = AA length = 50
FEATURE	Location/Qualifiers
REGION	1..50
	note = Description of Artificial Sequence: Synthetic
	polypeptide
source	1..50
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 14	DALDDDFDLD LGSDALDDFD LDMLGSDALD DFDLMLGSD ALDDDFDLDML
	50
SEQ ID NO: 15	moltype = DNA length = 17
FEATURE	Location/Qualifiers
misc_feature	1..17
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..17
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 15	gggctcctcc ttgtact
	17
SEQ ID NO: 16	moltype = DNA length = 18
FEATURE	Location/Qualifiers
misc_feature	1..18
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..18
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 16	acgcagataa gaaccagt
	18
SEQ ID NO: 17	moltype = DNA length = 17
FEATURE	Location/Qualifiers
misc_feature	1..17
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..17
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 17	ggcatcaagt cagccat
	17
SEQ ID NO: 18	moltype = DNA length = 18
FEATURE	Location/Qualifiers
misc_feature	1..18
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..18
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 18	agcctgagtc accctctt
	18
SEQ ID NO: 19	moltype = DNA length = 17
FEATURE	Location/Qualifiers
misc_feature	1..17
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..17
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 19	gggctcctcc ttgtact
	17
SEQ ID NO: 20	moltype = DNA length = 18
FEATURE	Location/Qualifiers
misc_feature	1..18
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..18
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 20	acgcagataa gaaccagt
	18

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SEQ ID NO: 21      moltype = DNA  length = 17
FEATURE
misc_feature
1..17
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..17
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 21
ggcatcaagt cagccat                                17

SEQ ID NO: 22      moltype = DNA  length = 18
FEATURE
misc_feature
1..18
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..18
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 22
agcctgagtc accctccat                                18

SEQ ID NO: 23      moltype = DNA  length = 19
FEATURE
misc_feature
1..19
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 23
tgtactctct gaggtgctc                                19

SEQ ID NO: 24      moltype = DNA  length = 19
FEATURE
misc_feature
1..19
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 24
acgcagataa gaaccaggat                                19

SEQ ID NO: 25      moltype = DNA  length = 19
FEATURE
misc_feature
1..19
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 25
catcaagtca gccatcagc                                19

SEQ ID NO: 26      moltype = DNA  length = 19
FEATURE
misc_feature
1..19
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 26
gagtcaccct cctggaaac                                19

SEQ ID NO: 27      moltype = DNA  length = 19
FEATURE
misc_feature
1..19
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 27

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cctgggctcc ggggcgtt	19
SEQ ID NO: 28	moltype = DNA length = 19
FEATURE	Location/Qualifiers
misc_feature	1..19
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..19
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 28	
ggccctcgcg gccaccccg	19
SEQ ID NO: 29	moltype = DNA length = 19
FEATURE	Location/Qualifiers
misc_feature	1..19
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..19
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 29	
ctccctccct gccccgttag	19
SEQ ID NO: 30	moltype = DNA length = 19
FEATURE	Location/Qualifiers
misc_feature	1..19
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..19
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 30	
agggtttggaa agggcgtgc	19
SEQ ID NO: 31	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 31	
actccactgc actccagtc	20
SEQ ID NO: 32	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 32	
tctgtggggg acctgcactg	20
SEQ ID NO: 33	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 33	
ggggcgccag ttgtgtctcc	20
SEQ ID NO: 34	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct

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SEQUENCE: 34 acaccattgc caccaccatt	20
SEQ ID NO: 35 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA organism = synthetic construct
SEQUENCE: 35 tgttttcagc ttccaaact	19
SEQ ID NO: 36 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA organism = synthetic construct
SEQUENCE: 36 catgaagaca gcagaagcc	19
SEQ ID NO: 37 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA organism = synthetic construct
SEQUENCE: 37 ggccccacatt ccttccag	19
SEQ ID NO: 38 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA organism = synthetic construct
SEQUENCE: 38 ggctggattg ggtttccag	19
SEQ ID NO: 39 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA organism = synthetic construct
SEQUENCE: 39 caactgagtc ctgaggttt	19
SEQ ID NO: 40 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA organism = synthetic construct
SEQUENCE: 40 ctcacagcac agccagtgt	19
SEQ ID NO: 41 FEATURE misc_feature source	moltype = DNA length = 19 Location/Qualifiers 1..19 note = Description of Artificial Sequence: Synthetic oligonucleotide 1..19 mol_type = other DNA

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SEQUENCE: 41	organism = synthetic construct	
cagcagctgg tcacaaaagc		19
SEQ ID NO: 42	moltype = DNA length = 19	
FEATURE	Location/Qualifiers	
misc_feature	1..19	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..19	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 42		
tttcctataaa acttctgag		19
SEQ ID NO: 43	moltype = DNA length = 20	
FEATURE	Location/Qualifiers	
misc_feature	1..20	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..20	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 43		
agtgtataaga caccgcgttt		20
SEQ ID NO: 44	moltype = DNA length = 20	
FEATURE	Location/Qualifiers	
misc_feature	1..20	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..20	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 44		
cagacatcta ataccacggt		20
SEQ ID NO: 45	moltype = DNA length = 20	
FEATURE	Location/Qualifiers	
misc_feature	1..20	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..20	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 45		
agggagaacg gggcctaccg		20
SEQ ID NO: 46	moltype = DNA length = 20	
FEATURE	Location/Qualifiers	
misc_feature	1..20	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..20	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 46		
acttcaggtt caaagaagcc		20
SEQ ID NO: 47	moltype = DNA length = 20	
FEATURE	Location/Qualifiers	
misc_feature	1..20	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..20	
	mol_type = other DNA	
	organism = synthetic construct	
SEQUENCE: 47		
ttttccccac ccagggccta		20
SEQ ID NO: 48	moltype = DNA length = 20	
FEATURE	Location/Qualifiers	
misc_feature	1..20	
	note = Description of Artificial Sequence: Synthetic	
	oligonucleotide	
source	1..20	

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SEQUENCE: 48 ccctgggtgg ggaaaaaccag	mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 49 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide source
SEQUENCE: 49 ggaggaacat gtttcggAAC	1..20 mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 50 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide source
SEQUENCE: 50 gtgccgtat ggTTCTGTCC	1..20 mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 51 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide source
SEQUENCE: 51 ggTCTGCGGG aaggTCTACa	1..20 mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 52 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide source
SEQUENCE: 52 tcggcctta actgccccaa	1..20 mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 53 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide source
SEQUENCE: 53 gcATGACAAA ggtGCCGTGA	1..20 mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 54 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide source
SEQUENCE: 54 cctgcctttt gggcagttaa	1..20 mol_type = other DNA organism = synthetic construct 20
SEQ ID NO: 55 FEATURE misc_feature	moltype = DNA length = 20 Location/Qualifiers 1..20 note = Description of Artificial Sequence: Synthetic oligonucleotide

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source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 55
aatatgtcac attctgtctc                                     20

SEQ ID NO: 56      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 56
ggactatggg aggtcaactaa                                     20

SEQ ID NO: 57      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 57
gaaggttaca cagaaccaga                                     20

SEQ ID NO: 58      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 58
gccctgtaa catccttgctg                                     20

SEQ ID NO: 59      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 59
ccactgctaa ctgaaagaga                                     20

SEQ ID NO: 60      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 60
agccacagtt tcagcgcagt                                     20

SEQ ID NO: 61      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
oligonucleotide
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 61
ctgtttcatc ttagaaaaat                                     20

SEQ ID NO: 62      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic

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source          oligonucleotide
 1..20
 mol_type = other DNA
 organism = synthetic construct
SEQUENCE: 62
gaatgttctt tggcaggtac                                20

SEQ ID NO: 63      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
source          1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 63
cgcacatctt atgtcttaga                                20

SEQ ID NO: 64      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
source          1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 64
cttaagagag ctagaactgg                                20

SEQ ID NO: 65      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
source          1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 65
tcccaaagta cagtaccttg                                20

SEQ ID NO: 66      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
source          1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 66
tcccttagaga ggacagacag                                20

SEQ ID NO: 67      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
source          1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 67
tcatacgagaa atgaaaagag                                20

SEQ ID NO: 68      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic
                 oligonucleotide
source          1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 68
ataatataacc ctgactccta                                20

SEQ ID NO: 69      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20

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note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 69
aggccacctg caagataaaat                                     20

SEQ ID NO: 70          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 70
tgttgttatac aattgccata                                     20

SEQ ID NO: 71          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 71
atcccttcca gcatacctcat                                     20

SEQ ID NO: 72          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 72
gtgcttcaaa accatggct                                     20

SEQ ID NO: 73          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 73
gatacatgtt ttattcttat                                     20

SEQ ID NO: 74          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic primer
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 74
caatgacccc ttcatttgacc                                     20

SEQ ID NO: 75          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic primer
source 1..20
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 75
ttgattttgg agggatctcg                                     20

SEQ ID NO: 76          moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic primer

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source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 76
ggaatccatg gagggaagat                                20

SEQ ID NO: 77      moltype = DNA  length = 20
FEATURE
misc_feature      Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 77
tgttctcgct caggtcagtg                                20

SEQ ID NO: 78      moltype = DNA  length = 20
FEATURE
misc_feature      Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 78
tcccttttc acggtctcac                                20

SEQ ID NO: 79      moltype = DNA  length = 20
FEATURE
misc_feature      Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 79
aacacccgac tgctgtatcc                                20

SEQ ID NO: 80      moltype = DNA  length = 30
FEATURE
misc_feature      Location/Qualifiers
1..30
note = Description of Artificial Sequence: Synthetic primer
source          1..30
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 80
cgaagagaa agcgaaccag tatcgagaac                                30

SEQ ID NO: 81      moltype = DNA  length = 27
FEATURE
misc_feature      Location/Qualifiers
1..27
note = Description of Artificial Sequence: Synthetic primer
source          1..27
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 81
cgtttgtcat agtcgctgct tgatcgc                                27

SEQ ID NO: 82      moltype = DNA  length = 20
FEATURE
misc_feature      Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 82
gcacgtggat cctgagaact                                20

SEQ ID NO: 83      moltype = DNA  length = 21
FEATURE
misc_feature      Location/Qualifiers
1..21
note = Description of Artificial Sequence: Synthetic primer
source          1..21
               mol_type = other DNA
               organism = synthetic construct
SEQUENCE: 83
attggacagc aagaaaagcga g                                21

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SEQ ID NO: 84      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 84      gcacgtggat cctgagaact
                           20

SEQ ID NO: 85      moltype = DNA length = 22
FEATURE           Location/Qualifiers
misc_feature      1..22
                  note = Description of Artificial Sequence: Synthetic primer
source            1..22
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 85      caggaaacag tccaggatct ca
                           22

SEQ ID NO: 86      moltype = DNA length = 21
FEATURE           Location/Qualifiers
misc_feature      1..21
                  note = Description of Artificial Sequence: Synthetic primer
source            1..21
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 86      gctgagtgaa ctgcactgtg a
                           21

SEQ ID NO: 87      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 87      gaattctttg ccgaaatgg
                           20

SEQ ID NO: 88      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic primer
source            1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 88      tcactagcaa gctctcaggc
                           20

SEQ ID NO: 89      moltype = DNA length = 19
FEATURE           Location/Qualifiers
misc_feature      1..19
                  note = Description of Artificial Sequence: Synthetic primer
source            1..19
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 89      aacaacgagg agtctgccc
                           19

SEQ ID NO: 90      moltype = DNA length = 24
FEATURE           Location/Qualifiers
misc_feature      1..24
                  note = Description of Artificial Sequence: Synthetic primer
source            1..24
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 90      gcagacagt accatctaca gctt
                           24

SEQ ID NO: 91      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic primer
source            1..20
                  mol_type = other DNA

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SEQUENCE: 91          organism = synthetic construct
caatccctct cgtccagtcg                                20

SEQ ID NO: 92          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 92          organism = synthetic construct
tgcttggact atgggaggtc                                20

SEQ ID NO: 93          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 93          organism = synthetic construct
gcaggtgctt caaaaaccatt                                20

SEQ ID NO: 94          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 94          organism = synthetic construct
tcaggtggtc agctttcct                                20

SEQ ID NO: 95          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 95          organism = synthetic construct
aagcaaacct tctggctcaa                                20

SEQ ID NO: 96          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 96          organism = synthetic construct
cacacacagt gaaccctttt                                20

SEQ ID NO: 97          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 97          organism = synthetic construct
ggacacatgc tcacatacgg                                20

SEQ ID NO: 98          moltype = DNA  length = 19
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..19
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 98          organism = synthetic construct
attcgatcca tgtgcctga                                19

SEQ ID NO: 99          moltype = DNA  length = 20
FEATURE

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misc_feature      1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 99
caatgctggaa                               20

SEQ ID NO: 100      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 100
gggggtgattc cctagagagg                         20

SEQ ID NO: 101      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 101
aagcaggaca gacaggcaag                           20

SEQ ID NO: 102      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 102
gagggcagtc agtgatggat                           20

SEQ ID NO: 103      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 103
tggaaaaggaa gaatgggaga                           20

SEQ ID NO: 104      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 104
tggtaaagg ttgccttgtca                           20

SEQ ID NO: 105      moltype = DNA length = 21
FEATURE          Location/Qualifiers
misc_feature     1..21
                  note = Description of Artificial Sequence: Synthetic primer
source           1..21
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 105
ggaatgactg aatcgaaaca a                           21

SEQ ID NO: 106      moltype = DNA length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic primer
source           1..20
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 106

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cctccagcat cttccacatt	20
SEQ ID NO: 107	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
source	note = Description of Artificial Sequence: Synthetic primer 1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 107	
gaaggcacct tcagcagttc	20
SEQ ID NO: 108	moltype = DNA length = 22
FEATURE	Location/Qualifiers
misc_feature	1..22
source	note = Description of Artificial Sequence: Synthetic primer 1..22
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 108	
ccacagttc agcgcgataa ta	22
SEQ ID NO: 109	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
source	note = Description of Artificial Sequence: Synthetic primer 1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 109	
atcagccagg acacacactt	20
SEQ ID NO: 110	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
source	note = Description of Artificial Sequence: Synthetic primer 1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 110	
ccctgtcagg agggacagat	20
SEQ ID NO: 111	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
source	note = Description of Artificial Sequence: Synthetic primer 1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 111	
ggctcacccg aagcatgaat	20
SEQ ID NO: 112	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
source	note = Description of Artificial Sequence: Synthetic primer 1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 112	
aagctacaag caggttcgct	20
SEQ ID NO: 113	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
source	note = Description of Artificial Sequence: Synthetic primer 1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 113	
aataacaggg tccatcccg	20
SEQ ID NO: 114	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic primer

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source          1..20
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 114
tgttccctcc acctggaata                                20

SEQ ID NO: 115      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 115
gggaaaatcc aaagcaggat                                20

SEQ ID NO: 116      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 116
tcctagggcc ctcaaaagca                                20

SEQ ID NO: 117      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 117
gtcccccaacg ctctaaacaaa                                20

SEQ ID NO: 118      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 118
gttagagcggt tggggacctt                                20

SEQ ID NO: 119      moltype = DNA length = 21
FEATURE
misc_feature       Location/Qualifiers
1..21
note = Description of Artificial Sequence: Synthetic primer
source          1..21
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 119
cacatgcaga gaactgagct g                                21

SEQ ID NO: 120      moltype = DNA length = 19
FEATURE
misc_feature       Location/Qualifiers
1..19
note = Description of Artificial Sequence: Synthetic primer
source          1..19
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 120
gttggggtaa gcacgaagg                                19

SEQ ID NO: 121      moltype = DNA length = 20
FEATURE
misc_feature       Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic primer
source          1..20
               mol_type = other DNA
               organism = synthetic construct

SEQUENCE: 121
tttccaggag ggtgactcag                                20

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SEQ ID NO: 122      moltype = DNA length = 20
FEATURE
misc_feature
source
SEQUENCE: 122
ttctctgcat gtgacccccc                                     20

SEQ ID NO: 123      moltype = DNA length = 20
FEATURE
misc_feature
source
SEQUENCE: 123
acacactcac agaggggttgg                                     20

SEQ ID NO: 124      moltype = DNA length = 20
FEATURE
misc_feature
source
SEQUENCE: 124
tgagtcaccc tcctggaaac                                     20

SEQ ID NO: 125      moltype = DNA length = 20
FEATURE
misc_feature
source
SEQUENCE: 125
ctccttcagg agcaccccttag                                     20

SEQ ID NO: 126      moltype = DNA length = 20
FEATURE
misc_feature
source
SEQUENCE: 126
gtggggctcc tccttgtact                                     20

SEQ ID NO: 127      moltype = DNA length = 20
FEATURE
misc_feature
source
SEQUENCE: 127
gctgctgcc ataaaagtgc                                     20

SEQ ID NO: 128      moltype = DNA length = 19
FEATURE
misc_feature
source
SEQUENCE: 128
ggactgttgc ccagggtact                                     19

SEQ ID NO: 129      moltype = DNA length = 20
FEATURE
misc_feature
source

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SEQUENCE: 129          organism = synthetic construct
ggcctcatag gacaggaggt                         20

SEQ ID NO: 130          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 130          organism = synthetic construct
ttatgggcag cagctcagtt                         20

SEQ ID NO: 131          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 131          organism = synthetic construct
gacattttcc tggacgcttg                         20

SEQ ID NO: 132          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 132          organism = synthetic construct
ccctccccat ggctttaggt                         20

SEQ ID NO: 133          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 133          organism = synthetic construct
agctccatgc gtttgacatt                         20

SEQ ID NO: 134          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 134          organism = synthetic construct
agcggtccagg aaaaatgtcaa                         20

SEQ ID NO: 135          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 135          organism = synthetic construct
atgaccctca cactccaagg                         20

SEQ ID NO: 136          moltype = DNA  length = 20
FEATURE
misc_feature
source
note = Description of Artificial Sequence: Synthetic primer
1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 136          organism = synthetic construct
gttgggtgct ccagctttta                         20

SEQ ID NO: 137          moltype = DNA  length = 20
FEATURE

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misc_feature	1..20	note = Description of Artificial Sequence: Synthetic primer
source	1..20	mol_type = other DNA organism = synthetic construct
SEQUENCE: 137		
cctcaaaact cctggactcg		20
SEQ ID NO: 138		moltype = AA length = 1434
FEATURE		Location/Qualifiers
REGION	1..1434	note = Description of Artificial Sequence: Synthetic polypeptide
source	1..1434	mol_type = protein organism = synthetic construct
SEQUENCE: 138		
MDYKDHGDY KDHDIDYKDD DDKMAPKKR KVGRGMDKKY SIGLAIGTNS VGWAVITDEY	60	
KVPSKKFKV GNTDRHSIKK NLIGALLFDs GETAEATRLK RTARRRYTRR KNRICYLQEI	120	
FSNEMAKVDD SFFHRLLEsF LVEEDKKHER HPIFGNIVDE VAYHEKYPTI YHLRKKLVDs	180	
TDKADLRLIY LALAHMIKFR GHFLIEGDLN PDNSDVKLF IQLVQTYNQL FEENPINASG	240	
VDAKAILSAR LSKSRRLENL IAQLPGEKKN GLFGNLTLAs LGLTPNFKSN FDLAEDAKLQ	300	
LSKDTYDDL DNLLAQIGDQ YADLFLAAKN LSDAIIILSDI LRVNTETTKA PLSASMIKRY	360	
DEHHQDLTLL KALVRQQLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG	420	
TEELLVKLNR EDLLRKQRTF DNGSIPHQIH LGELHAILRR QEDFYPLKD NREKIEKILT	480	
FRIPIYYVGPL ARGNSRFAWM TRKSEETITP WNFEVVVDKG ASAQSFIERM TNFDKNLPNE	540	
KVLPKHSSLY EFYFTVYNELT KVKVYVTEGMR KPAAFLSGEQK KAIVDLLFKT NRKVTVKQLK	600	
EDYFKKIECF DSVEISGVED RFNASLGTYH DLLKIIKDKD FLDNEEDEDI LEDIVLTTL	660	
FEDREMIEER LKTYAHLFDD KVMQKLKRR YTGWGRSLRK LINGIRDQs GKTILDFLKS	720	
DGFANRNFMQ LIHDDSLTFK EDIQAQVSG QGDSLHEHIA NLAGSPAIIK GILQTVKVVd	780	
ELVKVMGRHK PENIVIEMAR ENQTTQKGQK NSRERMKRIE EGIKELGSOI LKEHPVENTQ	840	
LQNEKLYL YY LQNGRDMYVD QELDINRLSD YDVAIVPOS FLKDDSIDNK VLTRSDKNRG	900	
KSDNPSEEV VKKMKNYWRQ LLNAKLITQR KFDNLTKAER GGLSELDKAG FIKRQLVETR	960	
QITKHVAQIL DSRMNtKYDE NDKLIREVKV ITLKSCLVSD FRKDFQFYKV REINNNYHHAH	1020	
DAYLNAVVTG ALIKKYPKLE SEFVYGDYKV YDVRKMTAKS EQEIGKATAK YFFYSNIMNF	1080	
FKTEITLANG EIRKRPLIET NGTGEIWD KGRDFATVRK VLSMPQVNIV KKTEVQTGGF	1140	
SKEsILPKNR SDKLIARKKD WDPKKYGGFD SPTVAYSVLU VAKVEKGSK KLKSVELLG	1200	
ITIMERSSFE KNPIDFLEAK GYKEVKKDLI IKLPKYSLF E LENGRKMLA SAGELOKGNE	1260	
LALPSKVYVF LYLAshYEKL KGSPEDNEQK QLVEQHKEY LDEIIIEQISE FSKRVILADA	1320	
NLDKVLSAYN KHRDKPIREQ AENIILHFLT TNLGAPAAFK YFDTTIDRKR YTSTKEVLDA	1380	
TLIHQsITGL YETRIDSQL GGDPIAGSKA SPKKRKVGR ADALDDFDLD MLGSDALDDF	1440	
DLDMLGSDAL DDFDLDMLGS DALDDFDLD LINPYDVDPD YAS	1483	

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SEQ ID NO: 140          moltype = AA  length = 3844
FEATURE           Location/Qualifiers
REGION            1..3844
                   note = Description of Artificial Sequence: Synthetic
                   polypeptide
source             1..3844
                   mol_type = protein
                   organism = synthetic construct
SEQUENCE: 140
MDYKDHDGDDY KDHDIDYKDD DDKMAPKKR KVGRGMDKKY SIGLAIGTNS VGWAVITDEY 60
KVPSSKKFKVL GNTDRHSIKK NLIGALLFDS GETAEATRLK RTARRRYTRR KNRICYLQEI 120
FSNEMAKVDD SFFHRLLEESF LVEEDKKHER HPIFGNIVDE VAYHEKYPTI YHLRKLVDS 180
TDKADLRLIY LALAHMIKFR GHFLIEGDLN PDNSDVKLKF IQLVQTYNQL FEENPINASG 240
VDAKAILSAR LSKSRRLENL IAQLPGEKKN GLFGNLIALS LGLTPNFKSN FDLAEDAKLQ 300
LSKDTYDSDL DNLLAQIGDQ YADLFLAAKN LSDAIIILSDI LRVNTETTKA PLSASMIKRY 360
DEHHQDLTLL KALVRQQLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG 420
TEELLVKLNR EDLLRKORTF DNGSIPHQIH LGELHAILRR QEDFYFPLKD NREKIEKILT 480
FRIPYYVGPL ARGNSRFAWM TRKSEETVWN WFNEEVVWDKG ASAQSFIERM TNFDKLPNE 540
KVLPKHSLLY EFYTVDYNELT KVXVTEGMR KPAFLSGEQK KAIVDFLLFKT NRKVTVKQLK 600
EDYFKKIECF DSVEISGVED RFNASLGTYH DLLKIIKDKD FLDNEEDEDI LEDIVLTTL 660
FEDREMIEER LKTYAHLF2 KVMQKLKRR YTCGWGRSLRK LINGIRDQKS GKTILDFLKS 720
DGFAKRNFMQ DGDLSLTFK EDIQAQVSG QGDSLHEHIA NLAGSPAIIKK GILQTVKVVD 780
ELVKVGMGRHK PENIVIEMAR ENQTQQGOK NSRERMKRIE EGIKELGSSI LKEHPVENTQ 840
LQNEKLYLYY LQNQGRDMYVD QELDINRLSD YDVAIVPQS FLKDDSIDNK VLTRSDKNRG 900
KSDNVPSEEV VKKMKNYWRQ LLNAKLITOR KFDNLTKAER GGLSELDKAG FIKRQLVETR 960
QITKHVAQIL DSRMNTKYD NDKLIREVKV ITLKSKLVSD FRKDFQFYKV REINNNYHHAH 1020
DAYLNAVVG7 ALIKKYPKLE SEFVYGYDKV YDVRKMTIAMS EOEIGKATAK YFFYSNIMNF 1080
FKTEITLANG EIRKRPLIET NGETGEIVWD KGDRDFATVRK VLSMPQVNIV KKTEVQTGGF 1140
SKESILPKRN SDKLIARKKD WDPKVVGGFD SPTVAYSVLV VAKVEKGKSK KLKSVKELLG 1200
ITIMERKSSFE KNPIDFPLEAK GYKEVKKD LIKLPKYSLFE LENGRKRMALA SAGELOKGNE 1260
LALPSKVNFM LYASHYKEK KGPSPDNEQK QLFVEQHKEY LDEIIEQISE FSKRVILADA 1320
NLTKVLSAYN KHRDKPIREQ AENIIHLFTL TNLGAPAAFK YFDTTIDRKR YTSTKEVLD 1380
TIIHQSTIGL YETRIDLSQ GGDPIAGSKA SPKKKRKVRG AAENVVEPGP PSAKRPKLSS 1440
PALSASASDG TDFGSLFDLE HDLPLDELINS TELGLTNGGD INQLQTSLSGM VQDAASKHKQ 1500
LSELLRSGSS PNLMNGVGPP QVMASQAAQ SSPGLGLINS MVKSPMTQAG LTSPNMGMGT 1560
SGPNQGPTQS TGMMNSPVNQ PAMGMNTGMN AGMNPGLMMA GNGQGIMPQ VMNGSIGAGR 1620
GRQNQYVNPN CMGSAGNLLT EPLQQGSPQM GGQTGLRGPQ PLKMGMMNPNP NPYGSPYTQN 1680
PGQQIGASGL GLQIQTKTBL SNNLSPFMD KKAVPGGGMP NMGGQQPAPQV QQPGLVTPVA 1740
QGMGSGAHTA DPEKRLKLIQQ QLVLLVHAHK CQRREQANGE VRQCNLPHCR TMKVNLNHMT 1800
HQCSGKSCQV AHCASSRQII SHWKNCTRHD CPVCLPLKNA GDKRNQOPIL TGAPVGLGNP 1860
SSLGVGQQSA PNLSVTSQID PSSIERAYAA LGLPYQVNQPM PTQPVQVAKN QQNQPGQSP 1920
QGQMRPMNSMS ASPMVGNGVQ GVQTPSLLSD SMLHSAINSQ NPMMSENASV PSMGPMPTAA 1980
QPSTTGIRKQ WHEDITQDLR NHLVHKVRA IFPTPDPAAL KDRRMENLVA YARKVEGDMY 2040
ESANNRIRKQY HLLAEKIYKI QKELEEKRT RLQKQNNMLPN AAGMVPVSMN PGPNMGQCPQ 2100
GMTSNGPLPD PSMIRGSPVN QMMPRITPQS GLNQFGQMSM AQPPIVPRQT PPLQHHGQLA 2160
QPGALNPPMG YGPRMCPQPSN QGQFLPQTOF PSQGMNVNTI PLAPSSGQAP VSQAOQMSSS 2220
CPVNSPIMPP GSQGSHIHC PQLQPALHQN SPSPVPSRTP TPHTTPPSIG AQQPPATTIP 2280
APVPTPPAMP PGPQSQALHP PPRQPTPTPT TQLPQQVQPS LPAAPSADQP QQQPRSQOST 2340
AASVPTPTAP LLPPQPATPL SQPAVSIEQG VSNNPSTSST EVNSQIAEK QPSQEVKMEA 2400
KMEVDQPEPA DTQPEDISES KVEDCKMEST ETBEERSTEKL TEIKEEEDQP STSATQSSPA 2460
PGQSKKKIFK PEELRQALMP TLEALYRQDP ESLPFRQPVQD PQLLGIPDYF DIVKSPMDLS 2520
TIKRKLDTGQ YQEPWQYVDD IWLMFPNNAWL YNRKTSRVYK YCSKLSSEVFE QEIDPVMQSL 2580
GYCCGRKLEF SPQTLCCYKG QLCCTIPRDAT YYSYQNRYHF CEKCFNEIQG ESVSLGDDPS 2640
QPOTTINKEQ FSKRKDNTL PELFVECTEC GRKMHQICVHL HEIIWIWAGB VCDGCLKKS 2700
RTRKENKFS A KRLPSTRGLT FLENRWNDFL RRQNHPESGE VTVRVVHASD KTVEVKPGMK 2760
ARFVDSGEMA ESFPYRTKAL FAEEFIDGVD LCFFGMHWQE YGSDCPPNQ RRVYISYLD 2820
VHFFRPKCLR TAVYHEILIG YLEYVKKLGY TTGHIWACPP SEGDDYIFHC HPPDQKIPKP 2880
KRLQEWYKHM LDKAVSERIV HDYKDIFKQ TEDRLTSAKE LPYFEGDFWP NVLEESIKEL 2940
EQEEEERKRE ESTNSNESTDV TKGDSKNAKK KNKKTSKNN SSSLRGNKKK PGMPNVSNLD 3000
SQKLYATMEK HKEVFFFVIRL IAGPAANSLP PIVDPDPLIP CDLMDGRDAF LTLARDKHL 3060
FSSLRRAQWS TMCMVLVELHT QSQDRFVYTC NECKHHVETR WHCTVCEDYD LCITCYNTKN 3120
HDHCKMEKLGL GLDDESNNQQ AAATQSPGDs RRLSIQRCIQ SLVHACQCRN ANCSLPSCK 3180
MKRVRVQHTKG CKRKTNNGCP ICKQQLIALCC YHAKHCQENK CPVPFCLNIK QKLRRQQQLQH 3240
RLQQAQMLRR RMASMQRTGV QRAGLPQQQP QQQLQPPMGG MSPQAQQMNM NHNTMPSQFR 3300
NSMPPYLPT QAAGPVSQGK AAGQVTPPTP POTAQPPPLPG PPPAAVEMAM QIQRRAETQR 3360
QMAHVQIFQR PIQHQMPPMT PMAPMGMNPP PMTRGPGSHL EPGMGPTGMQ QQPWSQGGL 3420
PQPQQLQSGM PRPAMMSVAQ HGQPLNMAPQ PGLGQVQGISP LKPGTVSQQA LQNLLRTLRS 3480
PSSPLQQQOV LSILHANPQL LAAPIKQRAA KYANSNPQPI PGQPGMPQGQ PGLQOPTMPG 3540
QQGVHSNPM QNMNPMPQAGV QRAGLPQQQP QQQLQPPMGG MSPQAQQMNM NHNTMPSQFR 3600
DILRRQQMMQ QQQQQGAGPG IGPGMANHQC FQQPQGVGYP PQQQQRMQQH MQQMCGQNNMG 3660
QIGQLPQALG AEAGASLQAY QQRLLQQQMG SPVQPNPMSP QQHMLPNQQAQ SPHLQGQQIP 3720
NSLSNQVRSP QPVPSPRQPS QPPHSPSPR MQPQPSPHV SPQTSSPHPG LVAAQANPME 3780
QGHFASPDQN SMLSQLASNP GMANLHGASA TDGLSTDNS DLNSNLSQST LDIHYPYDVP 3840
Dyas 3844

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SEQ ID NO: 141	moltype = AA length = 2048
FEATURE	Location/Qualifiers
REGION	1..2048
	note = Description of Artificial Sequence: Synthetic polypeptide
source	1..2048
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 141	
MDYKDHIDGYD KDHIDYKDD DDKMAPKKR KVGRGMDKKY SIGLAIGTNS VGWAVITDEY 60	
KVPSKKFKVL GNTDRHSIKK NLIGALFDs GETAEATRLK RTARRRYTRR KNRICYLQEI 120	
FSNEMAKVDD SFFHRLLESE LVVEDKKHER HP1FGNIVDE VAYHEKYPTI YHLRKKLVDs 180	
TDKADLRLIY LALAHMIKFR GHFLIEGDLN PDNSDVKLFL IQLVQTYNQL FEENPINASG 240	
VDAKAILSAR LSKSRRLENL IAQLPGEKKN GLPGNLITALS LGLTPNFKSN FDLAEDAKLQ 300	
LSKDTYDDDL DNLLAQIGDQ YADLFLAAKN LSDAIIILSDI LRVNTEITKA PLASAMIKRY 360	
DEHHQDLTLL KALVRQLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG 420	
TEELLVKLRN EDLLRKQRTF DNGSIPHQIH LGELHAILRR QEDFYPFLKD NREKIEKILT 480	
FRIPIYYVGPL ARGNSRFAWM TRKESEETITP WNFEVVVDKG ASAQSFIERM TNFDKKNLPNE 540	
KVLPKHSLLY EYFTVYNELT KVKVTEGMR KPAFLSGEQK KAIVDLLFKT NRKVTVKQLK 600	
EDYFKKIECF DSVEISGVED RFNAISLGTYH DLLKIIKDKD FLDNEENEDI LEDIVLTLT 660	
FEDREMIEER LKTYAHLFDD KVMQKLKRRR YTGWGRSLRK LINGIRDQs GKTILDFLKS 720	
DGFANRNFMQ LIHDDSLTFK EDIQAQVSG QGDSLHHEHIA NLAGSPAIIKK GILQTVKVV 780	
ELVKVMGRHK PENIVIEMAR ENQTTQKGOK NSRERMKRIE EGIKELGSQI LKEHPVENTQ 840	
LQNEKLYLYQ LQNGRDMYD QELDINRLSD YDVAIVPQS FLKDDSIDNK VLTRSDKNRG 900	
KSDNVPSEEV VKKMKNYWRQ LLNAKLTQR KFDNLTKAER GGLSELDKAG FIKRQLVETR 960	
QITKHVAQIL DSRMNTKYDE NDKLIREVKV ITLKSKLVSD FRKDFQFYKV REINNNYHHAH 1020	
DAYLNAVGT ALIKKPKLE SEFVYGDYKV YDVRKMIAKS QEIGKATAK YFFYSNIMNF 1080	
FKTEITLANG EIRKRPLIET NGETGEIVWD KGDRFATVRK VLSMPQVNIV KKTEVQTGGF 1140	
SKESILPKRN SDKLIARKKD WDPKKYGGFD SPTVAYSVLV VAKVEKGSK KLKSVKELLG 1200	
ITIMERSSFE KNPIDFLEAK GYKEVKKDLI IKLPKYSLFE LENGRKMLA SAGELOKGNE 1260	
LALPSKYVNF LYLAshyekl KGSPEDNEQK QLFVEQHKEY LDEIIEQISE FSKRVLADA 1320	
NLDKVL SAYN KHRDKPIREQ AENI IHLFTL TNLGAPAAFK YFDTTIDRKR YTSTKEVLD 1380	
TLIHQSIITGL YETRIDSQL GGDPIAGSKA SPKKKRKVGR AIFKPPELRQ ALMPTLEALY 1440	
RQDPESLFPF QPVDPOLLGI PDYDIVKSP MDLSTIKRKL DTGQYQEPWQ YVDDIWLMFN 1500	
NAWLBYNRKTS RYVYCSKSLK EVFQEIDPV MQLSGYCCGR KLEFSPQTL CYGKQLCTIP 1560	
RDATYSSYQVN RYHFCEKCFN EIQQESVSLG DDPSPQPTTI NKEQFSKRKN DTLDPELFVE 1620	
CTECGRKMHQ ICVLHHIIW PAGFVCDGCL KKSARTRKEN KFSAKRLPST RLGTFLNRV 1680	
NDPLRRQNHP ESGEVTVRVV HASDKTVEVK PGMKARFVDS GEMAESFPYR TKALFAFEEI 1740	
DGVDLCPFGM HVQEYGSDCP PPNVQYVIS YLDSVHFFRP KCLRTRAVYHE ILIGYLEYVK 1800	
KLGYTTGHIW ACPPSEGDDY IFCHCPPDQK IPKPKRQLKAVS ERIVHDYKDI 1860	
FIQATEDRLT SAKELPYFEG DFDPNVLEES IKBLEQEEE RKREENTSNE STDVTKGDSK 1920	
NAKKKNKKT SKNKSLSRG NKKKPGMPNV SNDLSQKLYA TMEKHKEFFF VIRLIAGPAA 1980	
NSLPPIVDPD PLIPCDLMGD RDAFLTLARD KHLEFSSLRR AOWSTMCMVL ELHTQSQDYP 2040	
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SEQ ID NO: 142	moltype = AA length = 2048
FEATURE	Location/Qualifiers
REGION	1..2048
	note = Description of Artificial Sequence: Synthetic polypeptide
source	1..2048
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 142	
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FSNEMAKVDD SFFHRLLESE LVVEDKKHER HP1FGNIVDE VAYHEKYPTI YHLRKKLVDs 180	
TDKADLRLIY LALAHMIKFR GHFLIEGDLN PDNSDVKLFL IQLVQTYNQL FEENPINASG 240	
VDAKAILSAR LSKSRRLENL IAQLPGEKKN GLPGNLITALS LGLTPNFKSN FDLAEDAKLQ 300	
LSKDTYDDDL DNLLAQIGDQ YADLFLAAKN LSDAIIILSDI LRVNTEITKA PLASAMIKRY 360	
DEHHQDLTLL KALVRQLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG 420	
TEELLVKLRN EDLLRKQRTF DNGSIPHQIH LGELHAILRR QEDFYPFLKD NREKIEKILT 480	
FRIPIYYVGPL ARGNSRFAWM TRKESEETITP WNFEVVVDKG ASAQSFIERM TNFDKKNLPNE 540	
KVLPKHSLLY EYFTVYNELT KVKVTEGMR KPAFLSGEQK KAIVDLLFKT NRKVTVKQLK 600	
EDYFKKIECF DSVEISGVED RFNAISLGTYH DLLKIIKDKD FLDNEENEDI LEDIVLTLT 660	
FEDREMIEER LKTYAHLFDD KVMQKLKRRR YTGWGRSLRK LINGIRDQs GKTILDFLKS 720	
DGFANRNFMQ LIHDDSLTFK EDIQAQVSG QGDSLHHEHIA NLAGSPAIIKK GILQTVKVV 780	
ELVKVMGRHK PENIVIEMAR ENQTTQKGOK NSRERMKRIE EGIKELGSQI LKEHPVENTQ 840	
LQNEKLYLYQ LQNGRDMYD QELDINRLSD YDVAIVPQS FLKDDSIDNK VLTRSDKNRG 900	
KSDNVPSEEV VKKMKNYWRQ LLNAKLTQR KFDNLTKAER GGLSELDKAG FIKRQLVETR 960	
QITKHVAQIL DSRMNTKYDE NDKLIREVKV ITLKSKLVSD FRKDFQFYKV REINNNYHHAH 1020	
DAYLNAVGT ALIKKPKLE SEFVYGDYKV YDVRKMIAKS QEIGKATAK YFFYSNIMNF 1080	
FKTEITLANG EIRKRPLIET NGETGEIVWD KGDRFATVRK VLSMPQVNIV KKTEVQTGGF 1140	
SKESILPKRN SDKLIARKKD WDPKKYGGFD SPTVAYSVLV VAKVEKGSK KLKSVKELLG 1200	
ITIMERSSFE KNPIDFLEAK GYKEVKKDLI IKLPKYSLFE LENGRKMLA SAGELOKGNE 1260	
LALPSKYVNF LYLAshyekl KGSPEDNEQK QLFVEQHKEY LDEIIEQISE FSKRVLADA 1320	

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NLDKVLSSAYN	KHRDKPIREQ	AENIIHLFTL	TNLGAPAAFK	YFDTTIDRKR	YTSTKEVILDA	1380
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RQDPESLPFR	QPVDPQQLGI	PDYFDIVKSP	MDLSTIKRKL	DTGQYQEPWQ	YVDDIWL MFN	1500
NAWLYNKRKS	RVYKYCSKLS	EVFQEIDPV	MQLGYC CGR	KLEFS PQTLC	CYGKQLCTIP	1560
RDATYYSYQN	RYHFCEKCFN	EIQGESVSLG	DDPSQPQTTI	NKEQFSKRKN	DTLDPELFVE	1620
CTECGRKMHQ	ICVLHHEIIW	PAGFVCDGCL	KKSARTRKEN	KFSAKRLPST	RLGTFLENRV	1680
NDFLRRQNHNP	ESGEVTVRVV	HASDKTVEVK	PGMKARFVDS	GEMAESFPYR	TKALFAEEI	1740
DGVDLCFFGM	HVQEYGSDCP	PPNQRVYIS	YLYSVHFFRP	KCLRATVYHE	ILIGLEYVK	1800
KLGYTTGHIW	ACPPSEGDDY	IFHCHPPDQK	IPKP KRLQEW	YKKMLDKAVS	ERIVHDYKDI	1860
FKQATEDRLT	SAKELYFEG	DWPVNVL EES	IKELEQEEEEE	RKREENTSNE	STDVTKGD SK	1920
NAKKNNNKKT	SKNKSSL SRG	NKKPGMPNV	SNDLSQKLYA	TMEKHKEVFF	VIRLIAGPAA	1980
NSLPPIVDPD	PLIPCDLM DG	RDAFLTLARD	KHLEFSSL R	AQWSTMCM LV	ELHTQS QDYP	2040
YDVDPDYAS						2048

SEQ ID NO: 143 moltype = AA length = 2048
 FEATURE Location/Qualifiers
 REGION 1..2048
 note = Description of Artificial Sequence: Synthetic
 polypeptide
 source 1..2048
 mol_type = protein
 organism = synthetic construct

SEQUENCE: 143
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 FSNEMAKVDD SFFHRLLEESF LVEEDKKHER HP1FGNIVDE VAYHEKPYTI YHLRKKL VDS 180
 TDKADLRLI Y LALAHMIKFR GHFLIEGDLN PDNSDVKDFL IQLVQTYNQL FEENPINASG 240
 VDAKAILS AR LSKSRLLENL IAQLPGEKKN GLFGNLIALS LGLTPNF KSN FDLAEDAKLQ 300
 LSKDTYDDD L DNLLAQIGDQ YADLFLAAKN LSDAILLSDI LRVNTEITKA PLSASMIKRY 360
 DEHHQDLT LL KALVRQQLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG 420
 TEELLVKL NR EDLLRKQRTF DNGSIPHQIH LGELHAILRR QEDFYFPLKD NREKIEKILT 480
 FRIPYYVG PL ARGNRSF AWM TRKSEETITP WNFEVVVDKG ASAQS FIERM TNFDKNLPNE 540
 KVLPHKSLL Y EYFTVYNE LT KVKVY TEGMR KPAFLS GEQK KAIVD LLLFKT NRKVTVKQLK 600
 EDYFKKIECF DSVEISGVED RFNASL GTYH DLLKIIKDKD FLDNEE NEDI LEDIVLTTL 660
 FEDREMIEER LKTYAHLFDD KVMKQLKRR Y TGWGRLSRK LINGIRDQKS GKTILDFLKS 720
 DGFANRNFMQ LIHDDSLTFK EDIQAQVSG QGDSLHEHIA NLAGSPAIIK GILQTVKVV 780
 ELVKVMGRH PENIVIEMAR ENQTTQKGQK NSRERMKRIE EGIKE LGSQI LKEHPVENTQ 840
 LQNEKLYL YY LQNGR DMYVD QELDINRLSD YDVA DIAV PQS FLKDD SIDL NK VLT RSDK NRG 900
 KSDNVPSE VVKMKN YWRQ LLNKLITK AER GGLN LDKAG FIKRQLVETR 960
 QITKHVAQ IL DSRMNTK YDE KDLK IREV KV ITL KS L VSD FRKDFQFYK V REIN NYH HAH 1020
 DAYLNAV VGT ALIKK YPKLE SEFVY GDY KV YD VRK MIAKS EOEIG KATAK Y FFYSNIMNF 1080
 FKTETITL ANG EIR KRPLI ET NGET CEI VWD KG RDFA T VRK VLS MPQVN IV KK TEV QT GGF 1140
 SKESILPKRN SDKL TARKKD WDPK YGG FD SPT VAYS VLV VAK VEG KSK KL KS V KELL G 1200
 ITIMERSS FE KN PID FLEAK GY KEV KKD L I KLP K YSL F E LEN GRK RML A SAGE LQ KG NE 1260
 LALPSKV VNF LY LASH YKE L KGS P DNE QK Q LF VEQ H KHY LDE II EQ ISE F SK RVI LADA 1320
 NLDKVLSSAYN KHRDKPIREQ AENIIHLFTL TNLGAPAAFK YFDTTIDRKR YTSTKEVILDA 1380
 TLIHQ SITGL Y ET RIDLSQLGI GGDPIAGSKA SPKKRKVGR AIFKPEELRQ ALMPTLEALY 1440
 RQDPESLPFR QPVDPQQLGI PDYFDIVKSP MDLSTIKRKL DTGQYQEPWQ YVDDIWL MFN 1500
 NAWLYNKRKS RVYKYCSKLS EVFQEIDPV MQLGYC CGR KLEFS PQTLC CYGKQLCTIP 1560
 RDATYYSYQN RYHFCEKCFN EIQGESVSLG DDPSQPQTTI NKEQFSKRKN DTLDPELFVE 1620
 CTECGRKMHQ ICVLHHEIIW PAGFVCDGCL KKSARTRKEN KFSAKRLPST RLGTFLENRV 1680
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 KLGYTTGHIW ACPPSEGDDY IFHCHPPDQK IPKP KRLQEW YKKMLDKAVS ERIVHDYKDI 1860
 FKQATEDRLT SAKELYFEG DWPNVLEES I K E LEQ EEE E R K REENT SNE STDVTKGD SK 1920
 NAKKKNNNKKT SKNKSSL SRG NKKPGMPNV SNDLSQKLYA TMEKHKEVFF VIRLIAGPAA 1980
 NSLPPIVDPD PLIPCDLM DG RDAFLTLARD KHLEFSSL E AQWSTMCM LV ELHTQS QDYP 2040
 YDVDPDYAS 2048

SEQ ID NO: 144 moltype = AA length = 2048
 FEATURE Location/Qualifiers
 REGION 1..2048
 note = Description of Artificial Sequence: Synthetic
 polypeptide
 source 1..2048
 mol_type = protein
 organism = synthetic construct

SEQUENCE: 144
 MDYKDHDG DY KDHDIDYKDD DDKMAPKKR KVGRGMDKKY SIGLAIGTNS VGWAVITDEY 60
 KVPSKKFKV L GNTDRHSIKK NLIGALLFDS GETAEATRLK RTARRRYTRR KNRICYLQEI 120
 FSNEMAKVDD SFFHRLLEESF LVEEDKKHER HP1FGNIVDE VAYHEKPYTI YHLRKKL VDS 180
 TDKADLRLI Y LALAHMIKFR GHFLIEGDLN PDNSDVKDFL IQLVQTYNQL FEENPINASG 240
 VDAKAILS AR LSKSRLLENL IAQLPGEKKN GLFGNLIALS LGLTPNF KSN FDLAEDAKLQ 300
 LSKDTYDDD L DNLLAQIGDQ YADLFLAAKN LSDAILLSDI LRVNTEITKA PLSASMIKRY 360
 DEHHQDLT LL KALVRQQLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG 420
 TEELLVKL NR EDLLRKQRTF DNGSIPHQIH LGELHAILRR QEDFYFPLKD NREKIEKILT 480

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FRIPIYYVGPL	ARGNSRFAWM	TRKSEETITP	WNFEEVVDKG	ASAQSFIERM	TNFDFKNLPNE	540
KVLPKHSLLY	EYFTVYNELT	KVKVTEGMR	KPAFLSGBQK	KAIVDILLFKT	NRKVTVKQLK	600
EDYFKKIECF	DSVEISGVED	RFNAISLGTYH	DLLKIIKDKD	FLDNEENEDI	LEDIVLTLTL	660
FEDREMEIER	LKTYAHLFDD	KVMQLKRRR	YTGWGRLSRK	LINGIRDQKS	GKTILDFLKS	720
DGFANRNFMQ	LIHDDSLTFK	EDIQKAQVSG	QGDSLHEHIA	NLAGSPAIIKK	GILQTVKVV	780
ELVKVMGRHK	PENIVIEMAP	ENQTTQKGQK	NSRERMKRIE	EGIKELGSQL	LKEHPVENTQ	840
LQNEKLYLYY	LQNGRDMYVD	QELDINRLSD	YDVDAIVPOS	FLKDDSIDNK	VLTRSDKNRG	900
KSDNVPSEEV	VKKMKNYWRQ	LLNAKLITOR	KFDNLTKAER	GGLSELDKAG	FIKRQLVETR	960
QITKHVAQIL	DSRMNTKYDE	NDKLIREVKV	ITLKSKLVD	FRKDFQFYKV	REINNNYHHAH	1020
DAYLNAVUGT	ALIKKYPKLE	SEFVYGDYKV	YDVRKMITAKS	EQEIGKATAK	YFFYSNIMNF	1080
FKTEITLANG	EIRKRPLIET	NGETGEIVWD	KGRDFPATVRK	VLSMPQVNIV	KKTEVQTGGF	1140
SKEISILPKRN	SDKLIAARKKD	WDPKYYGGFD	SPTVAYSVVL	VAKVEKGKSK	KLKSVKELLG	1200
ITIMERSSFE	KNPIDFLEAK	GYKEVKKDLI	IKLPKYSLF	LENGRKMLA	SAGELQKGNE	1260
LALPSKVVFN	LYLASHYEKL	KGSPEPDNEQK	QLFVEQHKHY	LDEIIIBQISE	FSKRVILADA	1320
NLDKVL SAYN	KHRDKPIREQ	AENIIHLFTL	TNLGAPAAFK	YFDTTIDRKR	YTSTKEVLD	1380
TLIHQSITGL	YETRIDSQL	GGDPPIAGSKA	SPKKKRKVGR	AIFKPEELRQ	ALMPTLEALY	1440
RQDPESLPFR	QPVDPQLLG	PDYFDIVKSP	MDLSTIKRKL	DTGQYQEPWQ	YVDDIWL MFN	1500
NAWLNRKTS	RVYKYCSKLS	EVFEQEI	MQSLGYCCGR	KLEFSPTOLC	CYGKQLCTIP	1560
RDATYYSYQN	RYHFCEKFRN	EIQGESVSLG	DDPSQPQTTI	NKEQFSKRKN	DTLDPELFVE	1620
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NDFLRRQNHP	ESGEVTVRVV	HASDKTVEVK	PGMKARFVDS	GEMAESFPYR	TKALFAEEI	1740
DGVDLFCFGM	HVQEYGSDCP	PPNQRVYIS	YLDHSVHFFRP	KCLRATAVYHE	ILIGYLEYVK	1800
KLGYTTGHIW	ACPPSEGDDY	IFHCHPPDQK	IPKPKRQLQEW	YKKMLDKAVS	ERIVHDYKDI	1860
FKQATEDRLT	SAKELPYFEG	DFWPNVLEES	IKELEQEEE	RKREENTSNE	STDVTKGDSK	1920
NAKKNNKKT	SKNKSSLSRG	NKKKPGMPNV	SNDLSQKLYA	TMEKHKEVFF	VIRLIAGPAA	1980
NSLPPIVDPD	PLIPCDLMDG	RDAFLTLARD	KHLEFSSLRR	AQWSTMCMVL	ELHTQSQDYP	2040
YDVPDYAS						2048

SEQ ID NO: 145	moltype = AA	length = 2048
FEATURE	Location/Qualifiers	
REGION	1..2048	
	note = Description of Artificial Sequence: Synthetic	
	polypeptide	
source	1..2048	
	mol_type = protein	
	organism = synthetic construct	

SEQUENCE: 145						
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KVPSKKPKV	GNTDRHSIKV	NLIGNLFD	GETAEATRLK	RTARRRYTRR	KNRICYLQE	120
FSNEMAKVDD	SFFHRLESF	VLEEDKKHER	HPIFGNIVDE	VAYHEKPTI	YHLRKKLVDS	180
TDKADLRLIY	LALAHMIKFR	GHFLIEGDLN	PDNSDVKL	IQLVQTYNQL	FEENPINASG	240
VDAKAILSAR	LSKSRRLENL	IAQLPGEKKN	GLFGNLIALS	LGLTPNFKSN	FDLABDAKLQ	300
LSKDTYDSDL	DNLLAQIGDQ	YADLFLAAKN	LSDAIIILSD	LRVNTEITKA	PLSASMIKRY	360
DEHHQDQLLL	KALVRQQLP	KYKEI	FFDQ	KNGYAGYIDG	GASQEEFYKF	420
TEELLVKLNR	EDLLRKQRTF	DNGSIPHQTH	LGELHAILRR	QEDFYFLKD	NREKIEKILT	480
FRIPIYYVGPL	ARGNSRFAWM	TRKSEETITP	WNFEEVVDKG	ASAQSFIERM	TNFDFKNLPNE	540
KVLPKHSLLY	EYFTVYNELT	KVKVTEGMR	KPAFLSGBQK	KAIVDILLFKT	NRKVTVKQLK	600
EDYFKKIECF	DSVEISGVED	RFNAISLGTYH	DLLKIIKDKD	FLDNEENEDI	LEDIVLTL	660
FEDREMEIER	LKTYAHLFDD	KVMQLKRRR	YTGWGRLSRK	LINGIRDQKS	GKTILDFLKS	720
DGFANRNFMQ	LIHDDSLTFK	EDIQKAQVSG	QGDSLHEHIA	NLAGSPAIIKK	GILQTVKVV	780
ELVKVMGRHK	PENIVIEMAP	ENQTTQKGQK	NSRERMKRIE	EGIKELGSQL	LKEHPVENTQ	840
LQNEKLYLYY	LQNGRDMYVD	QELDINRLSD	YDVDAIVPOS	FLKDDSIDNK	VLTRSDKNRG	900
KSDNVPSEEV	VKKMKNYWRQ	LLNAKLITOR	KFDNLTKAER	GGLSELDKAG	FIKRQLVETR	960
QITKHVAQIL	DSRMNTKYDE	NDKLIREVKV	ITLKSKLVD	FRKDFQFYKV	REINNNYHHAH	1020
DAYLNAVUGT	ALIKKYPKLE	SEFVYGDYKV	YDVRKMITAKS	EQEIGKATAK	YFFYSNIMNF	1080
FKTEITLANG	EIRKRPLIET	NGETGEIVWD	KGRDFPATVRK	VLSMPQVNIV	KKTEVQTGGF	1140
SKEISILPKRN	SDKLIAARKKD	WDPKYYGGFD	SPTVAYSVVL	VAKVEKGKSK	KLKSVKELLG	1200
ITIMERSSFE	KNPIDFLEAK	GYKEVKKDLI	IKLPKYSLF	LENGRKMLA	SAGELQKGNE	1260
LALPSKVVFN	LYLASHYEKL	KGSPEPDNEQK	QLFVEQHKHY	LDEIIIBQISE	FSKRVILADA	1320
NLDKVL SAYN	KHRDKPIREQ	AENIIHLFTL	TNLGAPAAFK	YFDTTIDRKR	YTSTKEVLD	1380
TLIHQSITGL	YETRIDSQL	GGDPPIAGSKA	SPKKKRKVGR	AIFKPEELRQ	ALMPTLEALY	1440
RQDPESLPFR	QPVDPQLLG	PDYFDIVKSP	MDLSTIKRKL	DTGQYQEPWQ	YVDDIWL MFN	1500
NAWLNRKTS	RVYKYCSKLS	EVFEQEI	MQSLGYCCGR	KLEFSPTOLC	CYGKQLCTIP	1560
RDATYYSYQN	RYHFCEKFRN	EIQGESVSLG	DDPSQPQTTI	NKEQFSKRKN	DTLDPELFVE	1620
CTECGRKMHQ	ICVLUHHEIIW	PAGFVCDGCL	KKSARTRKEN	KFSAKRLPST	RLGTFLENRV	1680
NDFLRRQNHP	ESGEVTVRVV	HASDKTVEVK	PGMKARFVDS	GEMAESFPYR	TKALFAEEI	1740
DGVDLFCFGM	HVQEYGSDCP	PPNQRVYIS	YLDHSVHFFRP	KCLRATAVYHE	ILIGYLEYVK	1800
KLGYTTGHIW	ACPPSEGDDY	IFHCHPPDQK	IPKPKRQLQEW	YKKMLDKAVS	ERIVHDYKDI	1860
FKQATEDRLT	SAKELPYFEG	DFWPNVLEES	IKELEQEEE	RKREENTSNE	STDVTKGDSK	1920
NAKKNNKKT	SKNKSSLSRG	NKKKPGMPNV	SNDLSQKLYA	TMEKHKEVFF	VIRLIAGPAA	1980
NSLPPIVDPD	PLIPCDLMDG	RDAFLTLARD	KHLEFSSLRR	AQWSTMCMVL	ELHTQSQDYP	2040
YDVPDYAS						2048

SEQ ID NO: 146	moltype = AA	length = 2048
FEATURE	Location/Qualifiers	
REGION	1..2048	

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note = Description of Artificial Sequence: Synthetic
      polypeptide
source   1..2048
mol_type = protein
organism = synthetic construct
SEQUENCE: 146
MDYKDHDGDDY KDHDIDYKDD DDKMAPKKR KVGRGMDKKY SIGLAIGTNS VGWAVITDEY 60
KVPSKKFKVL GNTDRHSIHK NLIGALLFDS GETAEATRLK RTARRYTRR KNRICYLQEI 120
FSNEMAKVDD SFFHRLEESF LVEEDKKHER HPIFGNIVDE VAYHEKYPTI YHLRKKLVDs 180
TDKAIDLRLIY LALAHMICKR GHFLIEGDLN PDNSDVKLDF IQLVQTYNQL FEENPINASG 240
VDAKAILSR LSKSRRLELN IAQLPGKEKNM GLFGNLMLLS LGLTPNFKSN FDLAEDAKLQ 300
LSKDTYDSDLN DNLLAQIGDQ YADLFLAKA LSDAIISSLSDI LRVNTEITKA PLASLMKRY 360
DEHHQDLTLL KALVRQOLPE KYKEIFFDQS KNGYAGYIDG GASQEEFYKF IKPILEKMDG 420
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VLPKLVHSLLY EFYFTVYNELT KVKGVTCEGM KPLAFLSGEQKG KAIVDVLFLKT NRKVTVPKQLK 600
EDYFKKIECF DSVEISGVED RFNASLGTYH DLLKIIKDKD FLDNEENEDI LEDIVLTLT 660
FEDREMIEER LKTYAHLFFF KVMKQLKRR YTGWGRSLRK LINGIIRDKQS GTKILDFLKS 720
DGFANRNFMQ LIHDDSLTFF EDIQKAQVSG QGDSLHEHIA NLAGSPAIFI GILQTVKVD 780
ELVKVGRMRHK PENIVIEMAR ENQTTQKGQK NSRERMKRVE EGIKELGSQL KLEHVENTQ 840
LQNEMKLYLYT LQNGRDMYVD QELDINRLSD YDVAIVPQS FLKDDDSIDNK VLTRSDKNRG 900
KSDNPVPSEEV VKKMKNYWRQ LLNAKLITQR KFDNLTKAER GGLSELDKAG FIKRQLVETR 960
QITKHVQAQIL DSRMNTKYDNDKLI REVKV ITLKSKLVSD FRKDFDQYKV REINNNYHHAA 1020
DAYLNAVVGTL ALIKKYPKLE SEFVYGDYKV D7VRKMIAKS EOEIGKATAK YFFYSNMNMF 1080
FKTEITLNG EIRKRPLIET NGETGEIWMV KGRDFATVVR VLSMPQVNIV KTEVQTVGGF 1140
SKESILPKRN SDKLIAKRD WDPKPYGGFD SPTVAYSVLU VAKVEKGKSK KLKSVKELLG 1200
ITIMERSSFF KNPIDFPLE GYKEVKKDLI IKLPKYSLFE LENGRKRMIA SAGELQKGNE 1260
LALPSKYVNF LYLASHYEKL KGSPEDNBQK QLFVEQHKHY DELIIIPQISE FSKEVILADA 1320
NDLKVLSAYN KHRDKPIREQ AEENIHLFLT TNLGPAPAFCP YFDTTIDRKR YTSTKEVLD 1380
TLIHOQSITGL YETRIDSQL GGDPIAGSKA SPKKKRVKGR AIFKPEELRQ ALMPTLEALY 1440
RQDPESLPPR QPVDPQQLGI PDYFDIVKSP MDLSTKRRKL DTGQYQEPWPQ YVDDIWLMPN 1500
NAWLYNRKTS RYVYCKSLSL EVFQEIDPV MQLSYGCCQR KLEFSPOTLC CYGKQLCITP 1560
RDRATVSYQIN RYHFECKCFN EIQQEVSGLG DDPSPQPTTI NKEQFQSKRKN DTLDLPELFVE 1620
CTECGRKRMHQ ICVLHHEIWI PAGFVCDGCL KKSARTRKEN KFSAKRLPST RLGTFLENRV 1680
NDFLRQRNHP ESGEFVTVVR HASDVKTEVK PGMKARFVDS GEMAESFPYR TKALFAFEEI 1740
DGVDLCCFFGM HVQEYGSDCP PNPNQRVYIW LLDVSHVFFCQK CLRTAVYHE ILIGLYEVK 1800
KLGYTTGHIW ACPPSEGEDY IFHCHPPDQK IPKPKRQLEW YKMMQLDKAVS ERIVHDYKDI 1860
FKQATEDRLT SAKELPYFEG DFWPNVLEES IKELEQEEE RKREENTSNE STDVTKGDSK 1920
NAKKNNKNTT SKNKSLSRG NKKKPGMPNV SNDLSQLYA TMEKHKEVFF VIRLIAQPA 1980
NSLPPIVDPD PLIPCDLMGD RDAFLTLARD KHLEFSSLRR AQWSTMCMVL ELHTQSQDYP 2040
YDVPDyas

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SEQUENCE : 147						
MDYKDHDDGKD	KDHIDYKDD	DDKMAPKKR	KVGRGMDKKY	SIGLAIGTNS	VGVAVITDEY	60
KVPSKKFKVL	GNTDRHSIKK	NLIGALLFD	GDTAEATRLK	TRARRYTRR	KNRICYLQEI	120
FSNEMAKVDD	SFFHRLLEESF	LVEEDKKHER	HPIFGNIVDE	VAYHEKYPTI	YHLRKKLVDS	180
TDKAIDLRLIY	LALAHMIKFR	GHFLIEGDLN	PDNSDVKLF	IQLVQTYNQL	FEENPINASG	240
VDAKAIALSR	LSKSRSRLENL	IAQLPGEKKN	GLFGNLIALS	LGLTTPNFKSN	FDLAEDAQLQ	300
LSKDTYDDDL	DNLLLAQIGDQ	YADLFLAAKN	LSDAIIILSDI	LRVNTEITKA	PLSAMIKRY	360
DEHHQDFTLTL	KALVRQOLPE	EDKTFDFDQS	KNGYAGYIDG	QASQEFPYKF	IKPILEKMDG	420
TEELLVKLNR	EDLLRKQRFT	DNGSIPHQIH	LGEHLAILRR	QEDFPYFLKD	NREKIEKILT	480
FRIPYYVGPF	ARGNSRFAWM	TRKSEETITP	WFNFNEVWDKG	ASAQSFIERM	TNFDFNLPNME	540
VKLPKHSLLY	EYFTVYNELT	KVVKYTEGMR	KPAFLSGEQK	KAIVDLLFKT	NRKVTVKQLK	600
EDYFKKIECF	DSVEISGVED	RFNANSALGTY	DLLKI1KDKD	FLDNEEDEDI	LEDIVLTTLT	660
FEDREMIEER	LKTYAHLFDD	KVMKQLKRRR	YTGWGRSLRK	LINGIRDQKS	GKTILDFLKS	720
DGFANRNFMQ	LIHDDSLLTFK	EDIQKAQVSG	QGDSLHEHIA	NLAGSPAIKK	GILQZTVKVV	780
ELVKVGMRKH	PENIVIEMAR	ENQTTQGQKQ	NSRERMKRIE	EGIKELGSIQ	LKEHPVENTQ	840
ZLQNKELYLWY	LQNQGRDMVY	QDLDINRLNS	YDVAIVPQSL	FLKDSDIDNK	VLTSDKRNQ	900
KSDNVPSEEV	VKKMKNYWRQ	LLNAKLITQR	KFDNLTKAER	GGLSELSDKAG	FIKQLVETR	960
QITKHVAQIL	DSRMNTKYDE	NDKLIREVKV	ITLKSCLVSD	FRKDFQFYKV	REINNYHHH	1020
DAYLNAVGGT	ALIKKPYKLE	SEFVGYDVKY	YDVRKMIAKS	EQEIQKATAK	YFFYSNIMNF	1080
FKTEITLANG	EIRKRPLIET	NGETGEIVWD	KGRDFATVRK	VLSMPQVNIV	KTVEQTGFF	1140
SKESILPLKRN	SDKLIAKRD	WDPKTYGGDF	SPTVAYSILV	VAKVEKGSK	KLKSVKELLG	1200
ITIMERSSFE	KNPIDFLEAK	GYKEVKDLDI	IKLPKYSLFE	LENGRKRLMA	SAGELQKGNE	1260
LALPSKVNF	LYLASHYEKL	KGSPEEDNEQK	QLFVEQHKHY	LDEIIEQISE	FSKRVILADA	1320
NLDKVLNSVAN	KHRDKPIREQ	AENIILHFLT	TNLGPAAFK	YFDFTIDRKR	YSTSTKEVLLA	1380
TLIHQSITGL	YETRIDLSQ	GGDPIAGSK	SPKKKRVKGR	AIFKPPEELRQ	ALMPTLEALY	1440
RDQPESLPLFR	QPVDPOLGI	PDYFDIVKSP	MDSLTIKRKL	DTGOYQEPWQ	VYDVIWLMPN	1500

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NAWLYNRKT	RVYKYCSKLS	EVFEQEIDPV	MQSLGYCCGR	KLEFSPQTLC	CYGKQLCTIP	1560
RDATYYSYQN	RYHFCEKCFN	EIQGESVSLG	DDPSQPQTTI	NKEQFSKRKN	DTLDPELFVE	1620
CTECGRKMHQ	ICVLUHHEIIW	PAGFVCDGCL	KKSARTRKEN	KFSAKRLPST	RLGTFLENRV	1680
NDFLRRQNHP	ESGEVTVRVV	HASDKTVEVK	PGMKARFVDS	GEMAESFPYR	TKALFAFEEI	1740
DGVDLCLFFGM	HVQEYGSDCP	PPNQRRTVYIS	YLDHSVHFFRP	KCLRTAVYAE	ILIGYLAALK	1800
KSGATTGAIW	ACPPSEGDDY	IFHCHPPDQK	IPKPKRLQEW	YKKMLDKAVS	ERIVHDYKDI	1860
FKQATEDRLT	SAKELPYFEG	DFWPNVLEES	IKELEQEEEEE	RKREENTSNE	STDVTKGDSK	1920
NAKKKNNKKT	SKNKSSLSRG	NKKKPGLMPNV	SNDLSQKLYA	TMEKHKEVFF	VIRLIAGPAA	1980
NSLPIVDPD	PLIPCDLMGD	RDAFLTLARD	KHLEFSSLRR	AQWSTMCMVL	ELHTQSQDYP	2040
YDVPDYAS						2048

SEQ ID NO: 148 moltype = AA length = 1155
 FEATURE Location/Qualifiers
 REGION 1..1155
 note = Description of Artificial Sequence: Synthetic
 polypeptide
 source 1..1155
 mol_type = protein
 organism = synthetic construct

SEQUENCE: 148
 MAAFKPNPIN YILGLAIGIA SVGWAMVEID EDENPICLID LGVRVFERAE VPKTGDSLAM 60
 ARRLARSVRR LTRRAHRRLL RARRLLKREG VLQAADFDEN GLIKSLPNTP WQLRAALDR 120
 KLTPLEWSAV LLHLIKHGRY LSQRKNEGET ADKELGALLK GVADNAHALQ TGDFRTPAEL 180
 ALNKFEKESG HIRNQRGDYS HTFSRKDLQA ELLLFEEKQK EFGNPVHSGG LKEGIETLLM 240
 TQRPALSGDA VQKMLGHCTF EPAEPKAAN TYTAERFIWL TKLNNRLILE QGSERPLTDT 300
 ERATLMDEPY RKSKLTYAQA RKLGLGEDTA FFKGLRYGKD NAEASTLMEM KAYHAISRAL 360
 EKEGLKDKKS PLNLSPELQD EIGTAFSLFK TDDEDITGRLK DRIQPEILEA LLKHISFDKF 420
 VQISLKLARR IVPLMEOGKR YDEACAEIYG DHYGKKNTEE KIYLPPPIPAD EIRNPVVLRA 480
 LSQARKVING VVRRYGPSPAR IHIETAREVG KSFKDRKEIE KRQEENRKDR EKAAAKFREY 540
 FPNFVGEPKS KDILKLRLYE QOHGKCLYSG KEINLGLRNLN KGYVEIAAL PFSRTWDSSF 600
 NNKVLVLGSE AQNKGNOTPY EYFNKGDNRSR EWQEFKARVE TSRFPRSKQ RILLQKFDED 660
 GFKERNLNDT RYVNRFQCF VADMRLTGT GKKRKFASNG QITNLLRGFW GLRKVRAEND 720
 RHHALDAVVV ACSTVAMQQK ITRFVRYKEM NAPDGKTIKD ETGEVLHQKT HFPQPWEFFA 780
 QEMVIRVGK PDGKPEFEEA DTPEKLRLL AEKLSSRPEA VHEYVTPLFV SRAPNRKMSG 840
 QGHMETVKSA KRLDEGVSVL RVPLTQLKLK DLEKVMVRER EPKLYEALKA RLEAHKDDPA 900
 KAPAEPFYKY DKAGNRTQQV KAVRVEQVK TGWVVRNHNG IADNATMVRV DVFEKGDKYY 960
 LVIPIYSWQVA KGILPDRAVV QGKDEEDWQL IDDSFNFKFS LHPNDLVEVI TKKARMFGYF 1020
 ASCHRGTTGNI NIRIHLDHK IKGNGILEGI GVKTALSFKQ YQIDELGKEI RPCRLKKRPP 1080
 VRSRADPKKK RKVEASGSGR ADALDDFDLDD MLGSDALDDF DLDMLGSDAL DDFDLDMLGS 1140
 DALDDFDLDM LINSR 1155

SEQ ID NO: 149 moltype = AA length = 1726
 FEATURE Location/Qualifiers
 REGION 1..1726
 note = Description of Artificial Sequence: Synthetic
 polypeptide
 source 1..1726
 mol_type = protein
 organism = synthetic construct

SEQUENCE: 149
 MAAFKPNPIN YILGLAIGIA SVGWAMVEID EDENPICLID LGVRVFERAE VPKTGDSLAM 60
 ARRLARSVRR LTRRAHRRLL RARRLLKREG VLQAADFDEN GLIKSLPNTP WQLRAALDR 120
 KLTPLEWSAV LLHLIKHGRY LSQRKNEGET ADKELGALLK GVADNAHALQ TGDFRTPAEL 180
 ALNKFEKESG HIRNQRGDYS HTFSRKDLQA ELLLFEEKQK EFGNPVHSGG LKEGIETLLM 240
 TQRPALSGDA VQKMLGHCTF EPAEPKAAN TYTAERFIWL TKLNNRLILE QGSERPLTDT 300
 ERATLMDEPY RKSKLTYAQA RKLGLGEDTA FFKGLRYGKD NAEASTLMEM KAYHAISRAL 360
 EKEGLKDKKS PLNLSPELQD EIGTAFSLFK TDDEDITGRLK DRIQPEILEA LLKHISFDKF 420
 VQISLKLARR IVPLMEOGKR YDEACAEIYG DHYGKKNTEE KIYLPPPIPAD EIRNPVVLRA 480
 LSQARKVING VVRRYGPSPAR IHIETAREVG KSFKDRKEIE KRQEENRKDR EKAAAKFREY 540
 FPNFVGEPKS KDILKLRLYE QOHGKCLYSG KEINLGLRNLN KGYVEIAAL PFSRTWDSSF 600
 NNKVLVLGSE AQNKGNOTPY EYFNKGDNRSR EWQEFKARVE TSRFPRSKQ RILLQKFDED 660
 GFKERNLNDT RYVNRFQCF VADMRLTGT GKKRKFASNG QITNLLRGFW GLRKVRAEND 720
 RHHALDAVVV ACSTVAMQQK ITRFVRYKEM NAPDGKTIKD ETGEVLHQKT HFPQPWEFFA 780
 QEMVIRVGK PDGKPEFEEA DTPEKLRLL AEKLSSRPEA VHEYVTPLFV SRAPNRKMSG 840
 QGHMETVKSA KRLDEGVSVL RVPLTQLKLK DLEKVMVRER EPKLYEALKA RLEAHKDDPA 900
 KAPAEPFYKY DKAGNRTQQV KAVRVEQVK TGWVVRNHNG IADNATMVRV DVFEKGDKYY 960
 LVIPIYSWQVA KGILPDRAVV QGKDEEDWQL IDDSFNFKFS LHPNDLVEVI TKKARMFGYF 1020
 ASCHRGTTGNI NIRIHLDHK IKGNGILEGI GVKTALSFKQ YQIDELGKEI RPCRLKKRPP 1080
 VRSRADPKKK RKVEASGRAF FKPEELRQAL MPTLEALYRQ DPESLPFRQP VDPQLLGIPD 1140
 YFDIVKSPMD LSTIKRKLDT GQYQEPWQYV DDIWLMFNNA WLYNRKTSRV YKYCSKLSEV 1200
 FFEQEIDPVMQ SLGYCCGRKL EFSPOTLCCY GKQLCTIPRD ATYYSYQNYR HFCEKCFNEI 1260
 QGESVSLGDD PSQPQTTINK EQFSKRKNDT LDPELFVECT ECGRKMHQIC VLHHEIIWPA 1320
 GFVCDGCLKK SARTRKENKF SAKRLPSTRL GTFLLENRVD FLRRQNHPES GEVTVRVVA 1380
 SDKTVEVKPG MKARFVDSGE MAESFPYRTK ALFAFEEIFD VGDCFVGMHV QEYGSDCPPP 1440
 NQRRVYISYL DSVHFFRPKC LRTAVYHEIL IGYLEYVKKL GYTTHIWIAC PPSEGDDYIF 1500
 HCHPPDQKIP KPKRQLQEWYK KMLDKAVSER IVHHDYKDFK QATEDRLTSA KELPYFEGDF 1560

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WPNVLEESIK ELEQEEERK REENTSNEST DVTKGDSKNA KKNNKKTSK NKSSLSRGNK	1620
KKPMPNVSN DLSQKLYATM EKHKEVFFVI RLIAGPAANS LPPIVDPDPL IPCDLMGIRD	1680
AFLTLARDKH LEFSSLRRAQ WSTMCMVEL HTQSQDYPYD VPDYAS	1726
SEQ ID NO: 150	moltype = AA length = 295
FEATURE	Location/Qualifiers
REGION	1..295
	note = Description of Artificial Sequence: Synthetic
	polypeptide
source	1..295
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 150	
MDYKDHGDY KDHDIDYKDD DDKMAPKKR KVGRGMAQAA LEPGEKPYAC PECGKSFSDC	60
RDLARHQRTH TGEKPYKCPE CGKSFRSRSD LVRHQRTHTG EKPYKCPCEG KSFQSQQNLV	120
RHQRHTGEK PYACPECGKS FSTSGELVRH QRHTGEKPY KCPECGKSFQ QRAHLERHQR	180
THTGEKPYKC PECGKSFQ QHLASHQRTH TGKKTSGQAG QASPKKKRKV GRADALDDFD	240
LIDMLGSDALD DFDLDMGLSD ALDDFDLDM GLSDALDDFDL DMLINYPYDV PDYAS	295
SEQ ID NO: 151	moltype = AA length = 860
FEATURE	Location/Qualifiers
REGION	1..860
	note = Description of Artificial Sequence: Synthetic
	polypeptide
source	1..860
	mol_type = protein
	organism = synthetic construct
SEQUENCE: 151	
MDYKDHGDY KDHDIDYKDD DDKMAPKKR KVGRGMAQAA LEPGEKPYAC PECGKSFSDC	60
RDLARHQRTH TGEKPYKCPE CGKSFRSRSD LVRHQRTHTG EKPYKCPCEG KSFQSQQNLV	120
RHQRHTGEK PYACPECGKS FSTSGELVRH QRHTGEKPY KCPECGKSFQ QRAHLERHQR	180
THTGEKPYKC PECGKSFQ QHLASHQRTH TGKKTSGQAG QASPKKKRKV GRAIFKPEEL	240
RQALMPTEA LYRQDPESLP FRQPVDPQLL GIPDVFIDVK SPMDLSTIKR KLDTGQYQEP	300
WQVVDDIWLM FNNAWLWNRK TSRVYKYCSK LSEVFEQEBID PVMQSLGYCC GRKLEFSPQT	360
LCCYGKQLCT IPRDATYSSY QNRYHFCEKC FNEIQGESVS LGDDPSQPQT TINKEQFSKR	420
KNDTLDPELF VECTECRKM HQICVLHHBI IWPGFVCDG CLKKSARTRK ENKFSAKRLP	480
STRLGTPLEN RVNDFLRRQN HPESGEVTVR VVHASDTVE VKPGMKAERV DSGEMAESFP	540
YRTKALFAFE EIDGVDSLCCF GMHVQEYGS CPPPQNRRVY ISYLDVHFFF RPKCLRTAVY	600
HEILIGLEY VKKLGYTTGH IWACPPSEGDI YIFHCHPPD QKIPKPKRLQ EYWKKMLDKA	660
VSERIVHDYK DIFKQATEDR LTSAKELPYF EGDFWPVNLE ESIKELEQEE EERKREENTS	720
NESTDVTKGD SKNAKKKNK KTSKNNKSSL RGNNKKPGMP NVSNLDSQKL YATMEKHKEV	780
FFVIRLIAGP AANSLPIVD PDPLIPCDLM DGRDAFLTLA RDKHLFSSL RRAQWSTMCM	840
LVELHTQSQD YPYDVPDYAS	860
SEQ ID NO: 152	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 152	
cctgggtttc aatgagaaga	20
SEQ ID NO: 153	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 153	
gattaggaca tgaacatggg	20
SEQ ID NO: 154	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 154	
cctcttctac attaacctta	20

-continued

```

SEQ ID NO: 155      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 155
ttttgaagc cagcaatcgta                                     20

SEQ ID NO: 156      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 156
cgtagtttc tggaggctct                                     20

SEQ ID NO: 157      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 157
acaattacc acgaaatgttag                                     20

SEQ ID NO: 158      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 158
tggcctgggc gcctgtctat                                     20

SEQ ID NO: 159      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 159
attttgtaaa taagggtttc                                     20

SEQ ID NO: 160      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 160
agcaacaggg gatggggcag                                     20

SEQ ID NO: 161      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 161

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-continued

aggactcgta gtatgcaggc	20
SEQ ID NO: 162	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 162	
ctgagccacc aactattttaa	20
SEQ ID NO: 163	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 163	
ctgagccacc aactattttaa	20
SEQ ID NO: 164	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 164	
actctgggtc ggttacggaa	20
SEQ ID NO: 165	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 165	
gggctgggct tagttggga	20
SEQ ID NO: 166	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 166	
atagggaggg gctctggagc	20
SEQ ID NO: 167	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 167	
atgggaaaag atacctgagt	20
SEQ ID NO: 168	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct

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SEQUENCE: 168
tgggagcgtt gtgtcgacgc                                     20

SEQ ID NO: 169      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA
organism         organism = synthetic construct

SEQUENCE: 169
tggaaaggct ttcattttct                                     20

SEQ ID NO: 170      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA
organism         organism = synthetic construct

SEQUENCE: 170
gtatctcgca gctccaataac                                     20

SEQ ID NO: 171      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA
organism         organism = synthetic construct

SEQUENCE: 171
acgcattccc ctcgggttga                                     20

SEQ ID NO: 172      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA
organism         organism = synthetic construct

SEQUENCE: 172
tcggaagctt ttcttctcag                                     20

SEQ ID NO: 173      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA
organism         organism = synthetic construct

SEQUENCE: 173
cggaaaggcg cg tgcgcgcggc                                     20

SEQ ID NO: 174      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA
organism         organism = synthetic construct

SEQUENCE: 174
ccggcgaag ggaagcggcc                                     20

SEQ ID NO: 175      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source           1..20
                  mol_type = other DNA

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SEQUENCE: 175          organism = synthetic construct
ggctgcgcac gccccatcccc                         20

SEQ ID NO: 176          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 176          organism = synthetic construct
ggggcttgca ggtgggttcgc                         20

SEQ ID NO: 177          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 177          organism = synthetic construct
cgagctaaag agcggatgcc                         20

SEQ ID NO: 178          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 178          organism = synthetic construct
agagggcggg agcaggggcca                         20

SEQ ID NO: 179          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 179          organism = synthetic construct
aaccggctct taactctttg                         20

SEQ ID NO: 180          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 180          organism = synthetic construct
caggagcggc gagcggggtc                         20

SEQ ID NO: 181          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 181          organism = synthetic construct
gggtatcaga tggcaaaggta                         20

SEQ ID NO: 182          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                 1..20

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mol_type = other DNA
organism = synthetic construct
SEQUENCE: 182
tcataggctg cccgcgattg                                         20

SEQ ID NO: 183      moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source            1..20
                   mol_type = other DNA
                   organism = synthetic construct
SEQUENCE: 183
gagggtggcc aggagcagcg                                         20

SEQ ID NO: 184      moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source            1..20
                   mol_type = other DNA
                   organism = synthetic construct
SEQUENCE: 184
aatttagcccc gcacggcgag                                         20

SEQ ID NO: 185      moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source            1..20
                   mol_type = other DNA
                   organism = synthetic construct
SEQUENCE: 185
tcccctgggt aggagtacag                                         20

SEQ ID NO: 186      moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source            1..20
                   mol_type = other DNA
                   organism = synthetic construct
SEQUENCE: 186
ggtttgttagc tgccggtcagc                                         20

SEQ ID NO: 187      moltype = DNA  length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source            1..20
                   mol_type = other DNA
                   organism = synthetic construct
SEQUENCE: 187
ggtgggagaac agggggcgcc                                         20

SEQ ID NO: 188      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                   note = Description of Artificial Sequence: Synthetic
                   oligonucleotide
source            1..23
                   mol_type = other DNA
                   organism = synthetic construct
variation          21
                   note = any nucleotide residue A, G, C, or T
SEQUENCE: 188
cctgggtcttc aatgagaaga ngg                                         23

SEQ ID NO: 189      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23

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note = Description of Artificial Sequence: Synthetic
      oligonucleotide
variation   21
note = any nucleotide residue A, G, C, or T
source     1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 189
gattaggaca tgaacatggg ngg                                23

SEQ ID NO: 190      moltype = DNA length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
variation   21
note = any nucleotide residue A, G, C, or T
source     1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 190
cctcttctac attaacctta ngg                                23

SEQ ID NO: 191      moltype = DNA length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
variation   21
note = any nucleotide residue A, G, C, or T
source     1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 191
tttttgaagc cagcaatcggt ngg                               23

SEQ ID NO: 192      moltype = DNA length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
variation   21
note = any nucleotide residue A, G, C, or T
source     1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 192
cgtagtttc tggaggctct ngg                                23

SEQ ID NO: 193      moltype = DNA length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
variation   21
note = any nucleotide residue A, G, C, or T
source     1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 193
acaaattacc acgaatgttag ngg                               23

SEQ ID NO: 194      moltype = DNA length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
variation   21
note = any nucleotide residue A, G, C, or T
source     1..23
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 194
tggcctgggc gcctgtctat ngg                               23

SEQ ID NO: 195      moltype = DNA length = 23

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FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
variation	21
	note = any nucleotide residue A, G, C, or T
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 195	
attttgtaaa taagggtcttc ngg	23
SEQ ID NO: 196	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
variation	21
	note = any nucleotide residue A, G, C, or T
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 196	
agcaaacaggg gatggggcag ngg	23
SEQ ID NO: 197	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
variation	21
	note = any nucleotide residue A, G, C, or T
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 197	
aggactcgta gtatgcaggc ngg	23
SEQ ID NO: 198	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
variation	21
	note = any nucleotide residue A, G, C, or T
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 198	
ctgagccacc aactatttaa ngg	23
SEQ ID NO: 199	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
variation	21
	note = any nucleotide residue A, G, C, or T
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 199	
ctgagccacc aactatttaa ngg	23
SEQ ID NO: 200	moltype = DNA length = 23
FEATURE	Location/Qualifiers
misc_feature	1..23
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
variation	21
	note = any nucleotide residue A, G, C, or T
source	1..23
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 200	
actctgggtc ggttacggaa ngg	23

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SEQ ID NO: 201      moltype = DNA  length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
variation           21
note = any nucleotide residue A, G, C, or T
source              1..23
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 201
gggctgggct tagcttggga ngg                                23

SEQ ID NO: 202      moltype = DNA  length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
variation           21
note = any nucleotide residue A, G, C, or T
source              1..23
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 202
atagggagggg gctctggagc ngg                                23

SEQ ID NO: 203      moltype = DNA  length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
variation           21
note = any nucleotide residue A, G, C, or T
source              1..23
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 203
atgggaaaag atacctgagt ngg                                23

SEQ ID NO: 204      moltype = DNA  length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
variation           21
note = any nucleotide residue A, G, C, or T
source              1..23
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 204
tgggagcggt gtgtcgcaagc ngg                               23

SEQ ID NO: 205      moltype = DNA  length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
variation           21
note = any nucleotide residue A, G, C, or T
source              1..23
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 205
tggaaaggct ttcattttct ngg                                23

SEQ ID NO: 206      moltype = DNA  length = 23
FEATURE
misc_feature        Location/Qualifiers
1..23
note = Description of Artificial Sequence: Synthetic
oligonucleotide
variation           21
note = any nucleotide residue A, G, C, or T
source              1..23
mol_type = other DNA
organism = synthetic construct

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SEQUENCE: 206
gtatctcgca gtcataatac ngg                                23

SEQ ID NO: 207      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation        21
                  note = any nucleotide residue A, G, C, or T
source          1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 207
acgcattccc ctcgggttga ngg                                23

SEQ ID NO: 208      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation        21
                  note = any nucleotide residue A, G, C, or T
source          1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 208
tcgaaagctt ttcttctcag ngg                                23

SEQ ID NO: 209      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation        21
                  note = any nucleotide residue A, G, C, or T
source          1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 209
cgaaggccg tgcgcgcggc ngg                                23

SEQ ID NO: 210      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation        21
                  note = any nucleotide residue A, G, C, or T
source          1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 210
ccggcgaag ggaagcggcc ngg                                23

SEQ ID NO: 211      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation        21
                  note = any nucleotide residue A, G, C, or T
source          1..23
                  mol_type = other DNA
                  organism = synthetic construct

SEQUENCE: 211
ggctgcgcac gccccatcccc ngg                                23

SEQ ID NO: 212      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation        21
                  note = any nucleotide residue A, G, C, or T
source          1..23

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mol_type = other DNA
organism = synthetic construct
SEQUENCE: 212
ggggcttgca ggtggttcgc ngg                                23

SEQ ID NO: 213      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation          21
                  note = any nucleotide residue A, G, C, or T
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 213
cgagctaaag agcggatgcc ngg                                23

SEQ ID NO: 214      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation          21
                  note = any nucleotide residue A, G, C, or T
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 214
agagggccgg agcagggccca ngg                                23

SEQ ID NO: 215      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation          21
                  note = any nucleotide residue A, G, C, or T
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 215
aaccggctct taactcttgc ngg                                23

SEQ ID NO: 216      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation          21
                  note = any nucleotide residue A, G, C, or T
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 216
caggagccgc gagccccggc ngg                                23

SEQ ID NO: 217      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation          21
                  note = any nucleotide residue A, G, C, or T
source            1..23
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 217
gggttatcaga tggcaaagtgc ngg                                23

SEQ ID NO: 218      moltype = DNA  length = 23
FEATURE           Location/Qualifiers
misc_feature      1..23
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
variation          21

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source          note = any nucleotide residue A, G, C, or T
                1..23
                mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 218
tcataggctg ccggcgattg ngg                                23

SEQ ID NO: 219      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
note = Description of Artificial Sequence: Synthetic
                oligonucleotide

variation        21
note = any nucleotide residue A, G, C, or T
1..23
source          mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 219
gagggttggcc aggagcagc ngg                                23

SEQ ID NO: 220      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
note = Description of Artificial Sequence: Synthetic
                oligonucleotide

variation        21
note = any nucleotide residue A, G, C, or T
1..23
source          mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 220
aattagcccc gcacggcgag ngg                                23

SEQ ID NO: 221      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
note = Description of Artificial Sequence: Synthetic
                oligonucleotide

variation        21
note = any nucleotide residue A, G, C, or T
1..23
source          mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 221
tcccctgggt aggagtacag ngg                                23

SEQ ID NO: 222      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
note = Description of Artificial Sequence: Synthetic
                oligonucleotide

variation        21
note = any nucleotide residue A, G, C, or T
1..23
source          mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 222
ggttgttagc tgccgtcagc ngg                                23

SEQ ID NO: 223      moltype = DNA  length = 23
FEATURE          Location/Qualifiers
misc_feature     1..23
note = Description of Artificial Sequence: Synthetic
                oligonucleotide

variation        21
note = any nucleotide residue A, G, C, or T
1..23
source          mol_type = other DNA
                organism = synthetic construct

SEQUENCE: 223
ggtgagaac agggggcgcc ngg                                23

SEQ ID NO: 224      moltype = DNA  length = 20
FEATURE          Location/Qualifiers
misc_feature     1..20
note = Description of Artificial Sequence: Synthetic

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source          oligonucleotide
 1..20
 mol_type = other DNA
 organism = synthetic construct

SEQUENCE: 224
gggggcgcga gtgatcagct                                20

SEQ ID NO: 225      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
note = Description of Artificial Sequence: Synthetic
               oligonucleotide

source          1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 225
cccggttctc ctaggggacg                                20

SEQ ID NO: 226      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
note = Description of Artificial Sequence: Synthetic
               oligonucleotide

source          1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 226
tggtccggag aaagaaggcg                                20

SEQ ID NO: 227      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
note = Description of Artificial Sequence: Synthetic
               oligonucleotide

source          1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 227
gtctccggc tcggaaactt                                20

SEQ ID NO: 228      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
note = Description of Artificial Sequence: Synthetic
               oligonucleotide

source          1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 228
agcgccagag cgcgagagcg                                20

SEQ ID NO: 229      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
note = Description of Artificial Sequence: Synthetic
               oligonucleotide

source          1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 229
cgattccggc cgcgttcccc                                20

SEQ ID NO: 230      moltype = DNA length = 20
FEATURE           Location/Qualifiers
misc_feature      1..20
note = Description of Artificial Sequence: Synthetic
               oligonucleotide

source          1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 230
gttgtgcggg ctgtatgcgc                                20

SEQ ID NO: 231      moltype = DNA length = 25
FEATURE           Location/Qualifiers
misc_feature      1..25

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note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..25
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 231
cctcggtgt tcctgggct gctgc                                25

SEQ ID NO: 232      moltype = DNA  length = 25
FEATURE          Location/Qualifiers
misc_feature     1..25
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..25
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 232
tcccataaac aggattctgc tcaga                                25

SEQ ID NO: 233      moltype = DNA  length = 25
FEATURE          Location/Qualifiers
misc_feature     1..25
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..25
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 233
caccggccag atgacagaac agaaa                                25

SEQ ID NO: 234      moltype = DNA  length = 27
FEATURE          Location/Qualifiers
misc_feature     1..27
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..27
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 234
ttgtttgaaa atgccatgg tagggct                                27

SEQ ID NO: 235      moltype = DNA  length = 25
FEATURE          Location/Qualifiers
misc_feature     1..25
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..25
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 235
aaacgcagca ggccccaggaa cacac                                25

SEQ ID NO: 236      moltype = DNA  length = 25
FEATURE          Location/Qualifiers
misc_feature     1..25
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..25
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 236
aaactctgag cagaatccctg tttat                                25

SEQ ID NO: 237      moltype = DNA  length = 25
FEATURE          Location/Qualifiers
misc_feature     1..25
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source 1..25
       mol_type = other DNA
       organism = synthetic construct
SEQUENCE: 237
aaactttctg ttctgtcatc tggcc                                25

SEQ ID NO: 238      moltype = DNA  length = 27
FEATURE          Location/Qualifiers

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misc_feature          1..27
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..27
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 238
aaacagccct acaaatggca ttttcaa                                         27

SEQ ID NO: 239          moltype = DNA  length = 25
FEATURE
misc_feature          1..25
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..25
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 239
cctcggtctgc ttctgcccga cctca                                         25

SEQ ID NO: 240          moltype = DNA  length = 25
FEATURE
misc_feature          1..25
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..25
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 240
tcccaccta agagcttgta ggccg                                         25

SEQ ID NO: 241          moltype = DNA  length = 25
FEATURE
misc_feature          1..25
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..25
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 241
caccgagagc tggctacccg tccct                                         25

SEQ ID NO: 242          moltype = DNA  length = 27
FEATURE
misc_feature          1..27
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..27
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 242
ttgtttcggg tccttgttta tcagtag                                         27

SEQ ID NO: 243          moltype = DNA  length = 25
FEATURE
misc_feature          1..25
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..25
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 243
aaactgaggc tcggcagaag cagac                                         25

SEQ ID NO: 244          moltype = DNA  length = 25
FEATURE
misc_feature          1..25
                      note = Description of Artificial Sequence: Synthetic
                      oligonucleotide
source               1..25
                      mol_type = other DNA
                      organism = synthetic construct
SEQUENCE: 244
aaaccggcct acaagcttt taggt                                         25

SEQ ID NO: 245          moltype = DNA  length = 25

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FEATURE          Location/Qualifiers
misc_feature    1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..25
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 245
aacacaggagc gggtagccag ctctc                                25

SEQ ID NO: 246      moltype = DNA length = 27
FEATURE          Location/Qualifiers
misc_feature    1..27
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..27
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 246
aacacctaactg ataaacaagg accgcaa                                27

SEQ ID NO: 247      moltype = DNA length = 25
FEATURE          Location/Qualifiers
misc_feature    1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..25
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 247
cctcggagct ggctaccgt cccta                                25

SEQ ID NO: 248      moltype = DNA length = 25
FEATURE          Location/Qualifiers
misc_feature    1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..25
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 248
tcccacttg gctgggtta aacca                                25

SEQ ID NO: 249      moltype = DNA length = 25
FEATURE          Location/Qualifiers
misc_feature    1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..25
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 249
caccggtag ctcagggttt tggta                                25

SEQ ID NO: 250      moltype = DNA length = 27
FEATURE          Location/Qualifiers
misc_feature    1..27
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..27
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 250
ttgtttggag ttagctcccc gaccag                                27

SEQ ID NO: 251      moltype = DNA length = 25
FEATURE          Location/Qualifiers
misc_feature    1..25
                  note = Description of Artificial Sequence: Synthetic
                  oligonucleotide
source          1..25
                  mol_type = other DNA
                  organism = synthetic construct
SEQUENCE: 251
aaacttaggga cgggtagccca gctcc                                25

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SEQ ID NO: 252	moltype = DNA length = 25
FEATURE	Location/Qualifiers
misc_feature	1..25
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..25
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 252	
aaactggttt aaaccaggcc aaagt	25
SEQ ID NO: 253	moltype = DNA length = 25
FEATURE	Location/Qualifiers
misc_feature	1..25
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..25
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 253	
aaactaccaa aaccctgagc tgacc	25
SEQ ID NO: 254	moltype = DNA length = 27
FEATURE	Location/Qualifiers
misc_feature	1..27
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..27
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 254	
aaacctgggtt cggggagcta actccaa	27
SEQ ID NO: 255	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 255	
tgtgttcctg ggcctgctgc	20
SEQ ID NO: 256	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 256	
taaacaggat tctgctcaga	20
SEQ ID NO: 257	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 257	
gccagatgac agaacagaaa	20
SEQ ID NO: 258	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 258	
aaaaatgccat ttgttagggct	20

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SEQ ID NO: 259      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 259
gcagcaggcc caggaacaca                                         20

SEQ ID NO: 260      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 260
tctgagcaga atcctgttta                                         20

SEQ ID NO: 261      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 261
tttctgttct gtcatctggc                                         20

SEQ ID NO: 262      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 262
agccctacaa atggcatttt                                         20

SEQ ID NO: 263      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 263
tctgcttctg ccgaacctca                                         20

SEQ ID NO: 264      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 264
cttaaagagc ttgttaggccg                                         20

SEQ ID NO: 265      moltype = DNA  length = 20
FEATURE
misc_feature
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source
1..20
mol_type = other DNA
organism = synthetic construct
SEQUENCE: 265

```

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agagctggct acccgccct	20
SEQ ID NO: 266	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 266	
cggtccttgt ttatcagtag	20
SEQ ID NO: 267	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 267	
tgagggttcgg cagaaggcaga	20
SEQ ID NO: 268	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 268	
cggcctacaa gctctttagg	20
SEQ ID NO: 269	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 269	
agggacgggt agccagctct	20
SEQ ID NO: 270	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 270	
ctactgataa acaaggaccg	20
SEQ ID NO: 271	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct
SEQUENCE: 271	
gagctggcta cccgtcccta	20
SEQ ID NO: 272	moltype = DNA length = 20
FEATURE	Location/Qualifiers
misc_feature	1..20
	note = Description of Artificial Sequence: Synthetic
	oligonucleotide
source	1..20
	mol_type = other DNA
	organism = synthetic construct

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```

SEQUENCE: 272
cttggctgg gtttaaacca                                20

SEQ ID NO: 273          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 273
gtcagctcg gttttggta                                20

SEQ ID NO: 274          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 274
gagttagtc cccgacccag                                20

SEQ ID NO: 275          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 275
tagggacggg tagccagctc                                20

SEQ ID NO: 276          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 276
tggtttaaac ccagccaaag                                20

SEQ ID NO: 277          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 277
tacccaaaacc ctgagctgac                                20

SEQ ID NO: 278          moltype = DNA  length = 20
FEATURE
misc_feature           Location/Qualifiers
1..20
note = Description of Artificial Sequence: Synthetic
      oligonucleotide
source                1..20
mol_type = other DNA
organism = synthetic construct

SEQUENCE: 278
ctgggtcggg gagctaactc                                20

```

1-57. (canceled)

58. A DNA targeting system comprising:

(1) a fusion protein comprising a first polypeptide domain and a second polypeptide domain, wherein the first polypeptide domain comprises a Clustered Regularly Interspaced Short Palindromic Repeats associated (Cas) protein, wherein the second polypeptide domain comprises a p300 histone acetyltransferase effector domain, and wherein the fusion protein activates transcription of a target gene; and

(2) at least one guide RNA (gRNA).

59. The DNA targeting system of claim **58**, wherein the first polypeptide domain comprises the sequence of SEQ ID NO: 1 or SEQ ID NO: 10.

60. The DNA targeting system of claim **58**, wherein the second polypeptide domain comprises the sequence of SEQ ID NO: 2 or SEQ ID NO: 3.

61. The DNA targeting system of claim **58**, wherein the first polypeptide domain comprises the polypeptide sequence of SEQ ID NO: 10 and the second polypeptide domain comprises the polypeptide sequence of SEQ ID NO: 3, or the first polypeptide domain comprises the polypeptide sequence of SEQ ID NO: 1 and the second polypeptide domain comprises the polypeptide sequence of SEQ ID NO: 2.

62. The DNA targeting system of claim **58**, wherein the fusion protein further comprises a linker connecting the first polypeptide domain to the second polypeptide domain.

63. The DNA targeting system of claim **58**, wherein the fusion protein comprises the polypeptide sequence of SEQ ID NO: 140, SEQ ID NO: 141, or SEQ ID NO: 149.

64. The DNA targeting system of claim **58**, wherein the at least one gRNA targets a target region of the target gene.

65. The DNA targeting system of claim **64**, wherein the target region comprises a target enhancer, target regulatory element, a cis-regulatory region of a target gene, or a trans-regulatory region of the target gene.

66. The DNA targeting system of claim **64**, wherein the target region is a distal or proximal cis-regulatory region of the target gene.

67. The DNA targeting system of claim **64**, wherein the target region is a promoter region of the target gene.

68. The DNA targeting system of claim **64**, wherein the target region is located on the same chromosome as the target gene.

69. The DNA targeting system of claim **64**, wherein the target region is located about 1 base pair to about 100,000 base pairs upstream of a transcription start site of the target gene.

70. The DNA targeting system of claim **64**, wherein the target region is located on a different chromosome as the target gene.

71. The DNA targeting system of claim **58**, wherein the target gene is selected from the group consisting of IL1RN, MYOD1, OCT4, HBE, HBG, HBD, HBB, MYOCD, PAX7, FGF1A, FGF1B, and FGF1C.

72. The DNA targeting system of claim **58**, wherein the target region is at least one of HS2 enhancer of the human β-globin locus, distal regulatory region (DRR) of the MYOD gene, core enhancer (CE) of the MYOD gene, proximal (PE) enhancer region of the OCT4 gene, or distal (DE) enhancer region of the OCT4 gene.

73. The DNA targeting system of claim **58**, wherein the gRNA comprises a polynucleotide encoded by a sequence selected from SEQ ID NO: 23-27.

74. A method of activating gene expression of a target gene in an isolated cell, the method comprising contacting the isolated cell with one or more polynucleotides encoding the DNA targeting system of claim **58**.

75. The method of claim **74**, wherein the DNA targeting system is delivered to the isolated cell virally.

76. The method of claim **74**, wherein the DNA targeting system is delivered to the isolated cell non-virally.

77. The method of claim **74**, wherein the isolated cell is a mammalian cell.

* * * * *