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### (54) APPARATUS AND METHOD FOR PRODUCTION OF FORMATE FROM CARBON DIOXIDE

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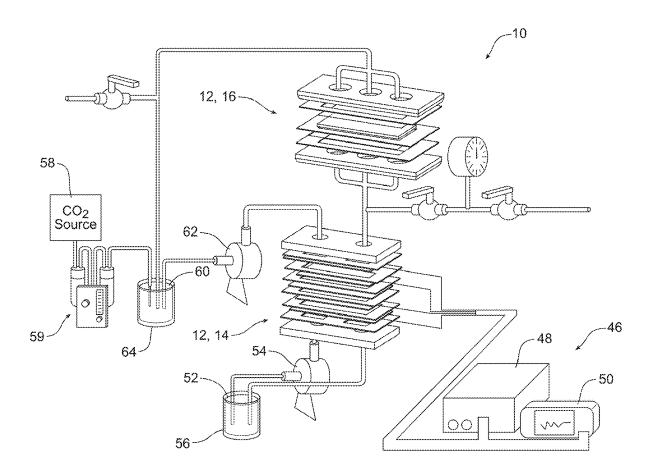
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#### (57)**ABSTRACT**

An apparatus is adapted for the production of formate from carbon dioxide. The apparatus includes a reactor including a reduction cell and a production cell and a formate dehydrogenase catalyst immobilized in the production cell. A cathode solution circulates through the reduction cell and the production cell. The cathode solution includes a charge carrier. An anode solution circulates through the anode compartment. The charge carrier is reduced in the reduction cell and reoxidized in the production cell. The carbon dioxide is converted to formate in the production cell.



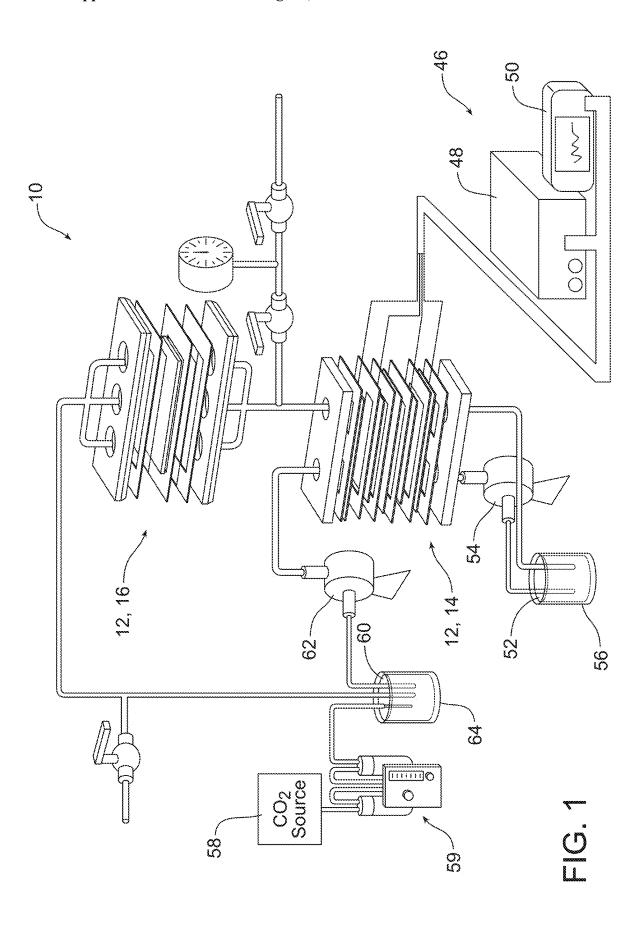


FIG. 2

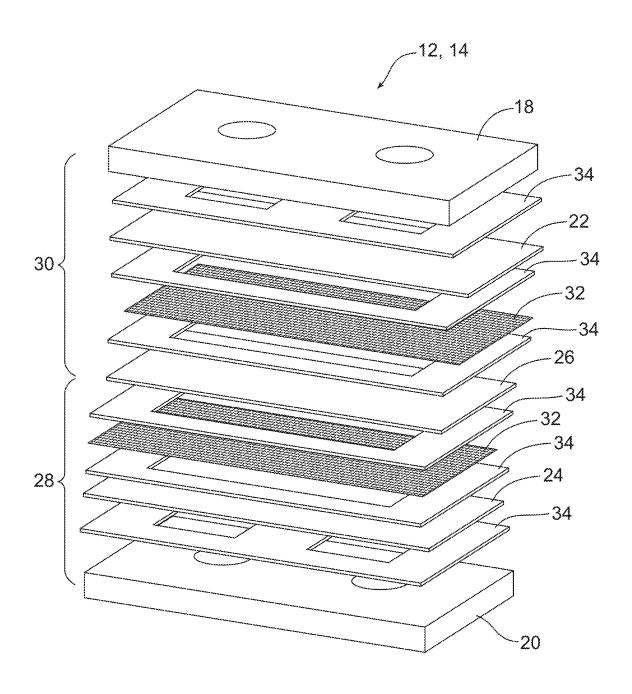
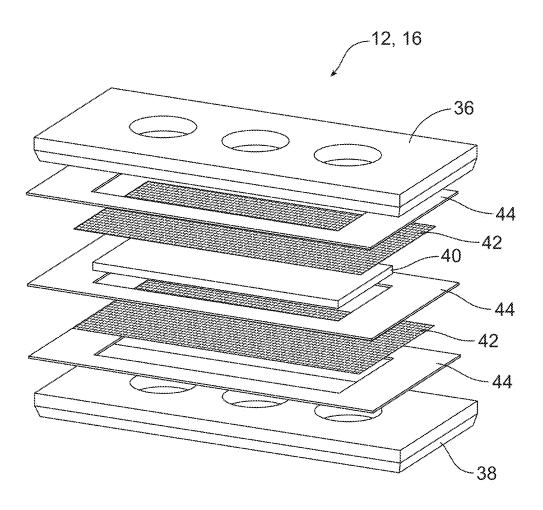


FIG. 3



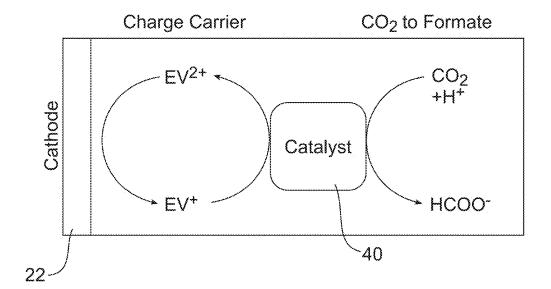


FIG. 4

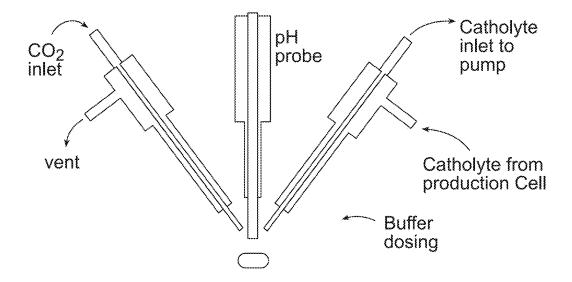


FIG. 5A

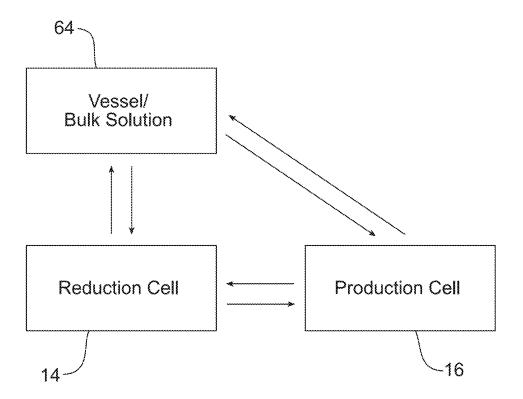


FIG. 5B

#### APPARATUS AND METHOD FOR PRODUCTION OF FORMATE FROM CARBON DIOXIDE

#### RELATED APPLICATIONS

[0001] This document claims the benefit of U.S. Provisional Patent Application Ser. No. 63/553,116, filed on Feb. 13, 2024, and U.S. Provisional Patent Application Ser. No. 63/756,510, filed Feb. 10, 2025, the full disclosure of each are incorporated herein by reference.

#### TECHNICAL FIELD

[0002] This document relates to an apparatus and method for converting carbon dioxide to formate using an enzymatic catalyst.

#### BACKGROUND

[0003] While the shift to renewable energy continues, fossil fuel-based resources will nonetheless continue to be used for the foreseeable future. One way to mitigate emissions from such resources is by converting the emitted carbon dioxide ( $\mathrm{CO}_2$ ) into other usable products, such as formic acid. Formic acid has a number of useful applications, including in the fuel cell and liquid hydrogen storage fields. Furthermore, formic acid requires a minimum activation energy relative to other products and minimal water for  $\mathrm{CO}_2$  conversion. Formic acid, formaldehyde, and oxalic acid, are ranked highest among a myriad of other fuel products due to economic feasibility and high capacity to decrease  $\mathrm{CO}_2$  emissions.

[0004] The state-of-the-art process for formic acid production is the Kemira process, which first reforms methane into methanol, both of which are already valuable commodities, then reacts methanol with carbon monoxide, including heterogenous catalysis under elevated pressure and mild to high temperatures, followed by hydrolysis to form formic acid. Subsequently, distillation separates the formic acid from the methanol and any water. This overall process is energy intensive and can contribute to further carbon emissions. Alternatively, formic acid production can be done electrochemically, first converting CO<sub>2</sub> to formate, and then to formic acid in post-processing using techniques such as ion exchange. The electrochemical route for formate (referred to as such henceforth) production can circumvent the need for elevated temperatures/pressures while providing the capability for negative emissions.

[0005] A significant challenge arising from electrochemical processes is product selectivity from competing electrochemical reactions. Typically, this is done using metal catalysts, and the most popular catalysts for electrochemical reduction are lead, tin, and bismuth. The metal catalyst used will affect the products formed, such as carbon monoxide, hydrogen, and higher-order carbon products. However, the electrochemical conversion of CO<sub>2</sub> can be difficult to control and often generates unwanted side reactions. Some studies have reported efficiencies approaching 100% for different types of catalyst, but these are usually short-term. Long-term, stability remains a challenge, and protonation on the catalyst can expose a vulnerability to unwanted reactions such as hydrogen evolution, and thus form additional products.

[0006] This document relates to an apparatus and method for converting carbon dioxide to formate using an enzymatic

catalyst, formate dehydrogenase 1 from *Methylobacterium extorquens* AM1 (MeFDH1), that provides a number of benefits and advantages over the state of the art devices and processes described above. The enzymatic catalyst, MeFDH1, produces high yields of the desired formate product, and operates reliably at near ambient temperature and pressure conditions at higher current densities that are used to accelerate formate production.

#### SUMMARY

[0007] In accordance with the purposes and benefits set forth herein, a new and improved apparatus is provided for the production of formate from carbon dioxide. The apparatus comprises, consists of or consists essentially of:

[0008] a reactor including a reduction cell and a production cell;

[0009] a proton exchange membrane partitioning the reduction cell into an anode compartment and a cathode compartment;

[0010] an anode in the anode compartment;

[0011] a cathode in the cathode compartment;

[0012] a voltage source connected across the anode and the cathode;

[0013] a formate dehydrogenase catalyst immobilized in the production cell;

[0014] a cathode solution circulating through the cathode compartment and the production cell, the cathode solution including a charge carrier; and

[0015] an anode solution circulating through the anode compartment whereby (a) the charge carrier is reduced in the cathode compartment and reoxidized in the production cell and (b) carbon dioxide is converted to formate in the production cell.

[0016] In at least some of the many possible embodiments of the apparatus, the formate dehydrogenase catalyst is formate dehydrogenase 1 from *Methylobacterium extorquens* AM1 (MeFDH1). Preferably, the MeFDH1 is immobilized on agarose beads.

[0017] The charge carrier may be selected from a group consisting of ethyl viologen, methyl viologen dibromide, 1,1'-diheptyl-4,4'-bipyridinium dibromide, 1,1'-di-n-octyl-4, 4'-bipyridinium dibromide, 1,1'-dibenzyl-4,4'-bipyridinium dichloride, 1,1'-diphenyl-4,4'-bipyridinium dichloride, 1,1'-diphenyl-4,4'-bipyridinium dichloride, propyl viologen, butyl viologen, benzyl viologen, ethanol viologen, propanol viologen, methyl acetate viologen, acetate viologen, ethylamine viologen, propylamine viologen, and methyl ethylamine viologen, chloride salts thereof, bromide salts thereof and combinations thereof.

[0018] In at least some embodiments of the apparatus, the apparatus further comprises an oxygen scavenger in the cathode solution. The oxygen scavenger may be selected from a group consisting of sodium thiosulfate, ascorbic acid, ferrous carbonate, sodium sulphite, sodium bisulphite, sodium metabisulphite, tannin, sodium erythorbate, hydrazine/carbohydrazine, N, N-diethylhydroxylamine and combinations thereof.

[0019] In at least some embodiments of the apparatus, the apparatus further comprises a pH buffer in the cathode solution. The pH buffer may be selected from a group consisting of Tris buffer, Tris-Bis buffer, BES, buffer, MOPS buffer, TES buffer, HEPES buffer, MOPSO buffer, Imidazole (glyoxaline)-HCl buffer, potassium hydroxide, sodium hydroxide, potassium carbonate, potassium phosphate and

mixtures thereof, adapted to maintain a pH of the cathode solution between pH 6 and pH 8.

[0020] The anode may be selected from a group consisting of a platinum wire mesh electrode, a platinum oxide coated carbon electrode, an iridium oxide coated carbon electrode, a cobalt oxide coated carbon electrode, a lead oxide/titanium alloy iridium-platinum coated carbon electrode, an iridium-ruthenium coated carbon electrode, and an iridium-tantalum coated carbon electrode. The cathode may be selected from a group consisting of a copper electrode, a tin-coated copper electrode, a bismuth-coated copper electrode, a lead-coated copper electrode, and bismuth-tin-lead alloys coated copper electrode, and bismuth-tin-lead alloys coated on porous carbon, carbon felt, carbon cloth, woven carbon, or carbon nanotube electrodes.

[0021] In accordance with yet another aspect, an apparatus adapted for production of formate from carbon dioxide, comprises, consists of or consists essentially of:

[0022] a reactor including a reduction cell and a production cell;

[0023] a formate dehydrogenase catalyst immobilized in the production cell;

[0024] a cathode solution circulating through the reduction cell and the production cell, the cathode solution including a charge carrier; and

[0025] an anode solution circulating through the anode compartment whereby (a) the charge carrier is reduced in the reduction cell and reoxidized in the production cell and (b) carbon dioxide is converted to formate in the production cell.

[0026] The formate dehydrogenase catalyst may be MeFDH1. The MeFDH1 may be immobilized on agarose beads. The apparatus may further comprise an oxygen scavenger and a pH buffer in the cathode solution.

[0027] In accordance with yet another aspect, a new and improved method of producing formate from carbon dioxide in a dual cell reactor, comprises, consists of or consists essentially of:

[0028] circulating a charge carrier solution between a reduction cell and a production cell;

[0029] reducing a charge carrier in the charge carrier solution in the reduction cell:

[0030] reoxidizing the charge carrier in the production cell: and

[0031] converting carbon dioxide and protons to formate in presence of a formate dehydrogenase catalyst in the production cell.

[0032] In at least some embodiments, the method further comprises one or more of the following steps: (a) partitioning the reduction cell into an anode compartment and a cathode compartment with a proton exchange membrane, (b) locating an anode in the anode compartment, (c) locating a cathode in the cathode compartment, and (d) applying a voltage across the anode and the cathode.

[0033] The method may further comprise circulating an oxygen scavenger with the charge carrier in the charge carrier solution. The method may further comprise circulating a pH buffer with the charge carrier in the charge carrier solution. The method may further comprise maintaining the charge carrier solution at a temperature between  $20^{\circ}$  C. and  $40^{\circ}$  C. and/or a pH of between pH 6 and pH 8. The method may further comprise maintaining the applied voltage between -0.75 V to -0.95 V vs. Ag/AgCl.

[0034] In accordance with still another aspect, a method of producing formate from carbon dioxide in a dual cell reactor, including a reduction cell and a production cell, comprises, consists of or consists essentially of:

[0035] partitioning the reduction cell into an anode compartment and a cathode compartment with a proton exchange membrane;

[0036] locating an anode in the anode compartment;

[0037] locating a cathode in the cathode compartment;

[0038] applying a voltage across the anode and the cathode:

[0039] immobilizing a formate dehydrogenase catalyst in the production cell;

[0040] circulating a cathode solution, including a charge carrier, through the cathode compartment and the production cell; and

[0041] circulating an anode solution through the anode compartment whereby (a) the charge carrier is reduced in the cathode compartment and reoxidized in the production cell and (b) carbon dioxide is converted to formate in the production cell.

[0042] The method may further comprise circulating an oxygen scavenger with the charge carrier in the charge carrier solution. The method may further comprise circulating a pH buffer with the charge carrier in the charge carrier solution. The method may further comprise maintaining the charge carrier solution at a temperature between  $20^{\circ}$  C. and  $40^{\circ}$  C. and/or a pH of between pH 6 and pH 8. The method may further comprise maintaining the applied voltage between -0.75 V to -0.95 V vs. Ag/AgCl.

[0043] The above and other aspects and features are described and exemplified by the following figures and detailed description.

# BRIEF DESCRIPTION OF THE DRAWING FIGURES

[0044] The following descriptions should not be considered limiting in any way.

[0045] FIG. 1 is a schematic illustration of the apparatus adapted for production of formate from carbon dioxide using a reactor including a reduction cell and a production cell and a formate dehydrogenase catalyst.

[0046] FIG. 2 is a detailed schematic illustration of the reduction cell of the apparatus illustrated in FIG. 1.

[0047] FIG. 3 is a detailed schematic illustration of the production cell of the apparatus illustrated in FIG. 1.

[0048] FIG. 4 is an illustration of the catalyst activity that converts carbon dioxide and protons to formate.

[0049] FIG. 5A is a schematic illustration a vessel for the cathode solution in order to accommodate dosing with a pH buffer

[0050] FIG. 5B is a schematic illustration of the cathode solution flow system.

#### DETAILED DESCRIPTION

[0051] Reference is now made to FIG. 1 which schematically illustrates one possible embodiment of the new and improved apparatus 10 adapted for converting carbon dioxide (CO<sub>2</sub>) to formate (HCOO<sup>-</sup>). In the illustrated embodiment, the apparatus 10 includes a dual cell reactor 12, comprising a reduction cell 14 and a production cell 16.

[0052] As shown in FIG. 2, the reduction cell 14 includes two opposed end plates 18, 20. A cathode 22 is provided

adjacent the end plate 18 and an anode 24 is provided adjacent the end plate 20. A proton exchange membrane 26, of a type known in the art, partitions the reduction cell 14 into an anode compartment 28, between the membrane 26 and the end plate 20, and a cathode compartment 30, between the membrane 26 and the end plate 18. Mesh sheets 32, on each side of the membrane 26, help support the membrane in position. Gaskets 34 seal the anode and cathode compartments 28, 30 provided between the components.

[0053] As shown in FIG. 3, the production cell 16 includes two opposed end plates 36, 38. An immobilized formate dehydrogenase catalyst 40 is supported between the end plates 36, 38 and sandwiched between mesh sheets 42. Gaskets 44 seal the production cell 16. Preferably, the formate dehydrogenase catalyst 40 is formate dehydrogenase 1 from Methylobacterium extorquens AM1 (MeFDH1) immobilized on agarose beads or other appropriate support material known to those skilled in the art. In at least some embodiments, the production cell 16 may comprise a packed bed reactor and for purposes of this document, the terminology "production cell" includes such a packed bed reactor. [0054] The cathode 22 may be made from various materials. Porous, conductive, inert materials appear to be the most viable choices. Useful materials include, but not necessarily limited to, a copper electrode, a tin-coated copper electrode, a bismuth-coated copper electrode, a lead-coated copper electrode, a bismuth-tin-lead alloys coated copper electrode, and bismuth-tin-lead alloys coated on porous carbon, carbon felt, carbon cloth, woven carbon, or carbon nanotube electrodes. The anode 24 may comprise a platinum wire mesh electrode, a platinum oxide coated carbon electrode, an iridium oxide coated carbon electrode, a cobalt oxide coated carbon electrode, a lead oxide/titanium alloy iridium-platinum coated carbon electrode, an iridium-ruthenium coated carbon electrode, and an iridium-tantalum coated carbon electrode.

[0055] A voltage source, generally designated by reference number 46, is connected across the cathode 22 and the anode 24. In the illustrated embodiment, the voltage source 46 includes a potentiostat 48 and a data logger 50.

[0056] An anode solution 52 is circulated by a pump 54 from the anode solution reservoir 56 through the anode compartment 28, across the anode, and then back to the anode solution reservoir. The anode solution 52 may comprise a sulfuric acid solution to serve as a proton source for converting carbon dioxide into formate. Other anode solutions, serving as proton donors, include, but are not necessarily limited to, hydrochloric acid, acetic acid, formic acid, carbonic acid, citric acid, ascorbic acid, and nitric acid.

[0057] A cathode solution or charge carrier solution 60 is circulated by the pump 62 from the cathode solution reservoir 64 through the cathode compartment 30, across the cathode 22, and then to the production cell 16 and, finally, back to the cathode solution reservoir. In the most preferred embodiment, the cathode solution includes a charge carrier, an oxygen scavenger and a pH buffer.

[0058] In one possible embodiment, the charge carrier is ethyl viologen (EV), the oxygen scavenger is sodium thiosulfate, and the pH buffer is potassium carbonate. The charge carrier is adapted to facilitate the conversion of carbon dioxide by first being electrochemically reduced in the cathode compartment 30 of the reduction cell 14 at a sufficient voltage (~-0.7 V vs. Ag/AgCl) to an EV<sup>+</sup> species,

then donating its charge to the catalyst enzyme 40 in the production cell 16 to produce formate, while reoxidizing back to dissolved  $\mathrm{EV}^{2+}$  in the process. The oxygen scavenger functions to eliminate competition with dissolved oxygen (O2) present in the cathode solution: that is, deceases the use of applied current for undesired chemical reactions. The pH buffer maintains the pH of the cathode solution within a desired range of pH 6 to pH 8; that is, the pH range most conducive to the catalytic activity and service life of the MeFDH1 catalyst 40. The concentration of the charge carrier in the cathode solution may range from about 1-100 mM, or 1-50 mM, or 1-25 mM, or 1-20 mM or 10 mM.

[0059] Charge carriers useful in the apparatus 10 include, but are not necessarily limited to, ethyl viologen, methyl viologen dibromide, 1,1'-diheptyl-4,4'-bipyridinium dibromide, 1,1'-di-n-octyl-4,4'-bipyridinium dibromide, 1,1'-dibenzyl-4,4'-bipyridinium dichloride, 1,1'-bis(2,4-dinitrophenyl)-4,4'-bipyridinium dichloride, 1,1'-diphenyl-4,4'-bipyridinium dichloride, propyl viologen, butyl viologen, benzyl viologen, ethanol viologen, propanol viologen, methyl acetate viologen, acetate viologen, ethylamine viologen, propylamine viologen, and methyl ethylamine viologen, chloride salts thereof, bromide salts thereof and combinations thereof.

[0060] Oxygen scavengers useful in the apparatus 10 include, but are not necessarily limited to, sodium thiosulfate, ascorbic acid, ferrous carbonate, sodium sulphite, sodium bisulphite, sodium metabisulphite, tannin, sodium erythorbate, hydrazine/carbohydrazine, N, N-diethylhydroxylamine and combinations thereof.

[0061] The pH buffers useful in the apparatus 10 include, but are not necessarily limited to, Tris buffer, Tris-Bis buffer, BES, buffer, MOPS buffer, TES buffer, HEPES buffer, MOPSO buffer, Imidazole (glyoxaline)-HCl buffer, potassium hydroxide, sodium hydroxide, potassium carbonate, potassium phosphate and mixtures thereof, adapted to maintain a pH of the cathode solution between pH 6 and pH 8. The concentration of the oxygen scavenger in the cathode solution may range from about 1-100 mM, or 5-95 mM, or 10-50 mM, or 20-40 mM. In some embodiments, the cathode solution reservoir 64 may include structure to accommodate dosing the cathode solution 60 with potassium hydroxide (KOH). See FIG. 5A. FIG. 5B illustrates the flow system components in the order through which the cathode solution flows. In such an embodiment, the cathode solution may first be pumped into the production cell 16, then into the reduction cell 14 so that the cathode solution reservoir 64 is capable of measuring pH immediately before entering the production cell. Such a configuration is particularly useful when the production cell 16 is a packed bed reactor.

[0062] The apparatus 10 described herein is useful in a method of producing formate from carbon dioxide in a dual cell reactor 12 including a reduction cell 14 and a production cell 16. The method may include steps of: (a) partitioning the reduction cell 14 into an anode compartment 28 and a cathode compartment 30 with a proton exchange membrane 26, (b) locating an anode 24 in the anode compartment, (c) locating a cathode 22 in the cathode compartment, (d) applying a voltage across the anode and the cathode, (e) immobilizing a formate dehydrogenase catalyst 40 in the production cell, (f) circulating a cathode solution 60, including a charge carrier, through the cathode compartment and the production cell, and (g) circulating an anode solution 52 through the anode compartment whereby (i) the charge

carrier is reduced in the cathode compartment and reoxidized in the production cell and (ii) carbon dioxide is converted to formate in the production cell.

[0063] The method may further comprise one or more of the following steps: (a) circulating an oxygen scavenger with the charge carrier in the charge carrier solution, (b) circulating a pH buffer with the charge carrier in the charge carrier solution, (c) maintaining the charge carrier solution at a temperature between 20° C. and 40° C., or 25° C. and 35° C. or at 30° C., (d) maintaining the cathode solution at a pH of between pH 6 and pH 8 or pH 6.5 and pH 7.5, or pH 6.3 and pH 6.8, and (e) maintaining the applied voltage between -0.75 V to -0.95 V vs. Ag/AgCl or -0.8 to -0.9 V vs. Ag/AgCl, or at -0.85 V vs. Ag/AgCl.

[0064] The apparatus 10 and method advantageously decouple the electrochemical charge carrier reduction and subsequent catalyst interaction into two separate processes, each in their own unit cell. If done in a flow cell configuration, the system would have two primary benefits. One would be that the product catalyst in the production cell 16 would be protected from high overpotentials in the electrode. The configuration would also greatly reduce the chance of fouling deposits from the enzyme accumulating in the reduction cell 14 and subsequently weakening its performance.

#### **Experimental Section**

[0065] The flow system initially considered in the study involved two cells 14, 16 consisting of stainless steel end plates 18, 20 and 36, 38 hand-tightened with sufficiently shaped, compressed gaskets 34, 44 (See FIG. 1). The contact area which the solution could interact with either the electrodes (reduction cell) 22, 24 or enzyme (production cell) 40 was approximately 33 cm<sup>2</sup> in both cells. In the reduction cell 14, a Nafion 115 membrane 26 separated the two streams in the anode and cathode compartments 28, 30. The anode side or compartment 28 used a platinum-iridium coated electrode 24 (average loading 0.7 mg/cm<sup>2</sup> on carbon) and circulated a 100 mM aqueous H<sub>2</sub>SO<sub>4</sub> solution 52. The dissociated protons from the sulfuric acid replenished by the oxygen evolution reaction serve as a proton source through the Nafion membrane 26, and travel to the production cell 16 to react with CO2 and EV+ in the enzyme catalyst 40 to produce formate. In the absence of an applied current between both cells, charge neutrality is conserved by the presence of other species, including  $HCO_3^-$ ,  $CO_3^{2-}$ ,  $OH^-$ ,  $CI^-$ ,  $HCOO^-$ ,  $K^+$ ,  $H^+$ ,  $EV^{2+}$ ,  $EV^+$ ,  $EV^{22+}$ , and  $EVH^+$ . Therefore, the protons can remain in the solution into the production cell 16, although this will alter the concentration of some other ionic species.

[0066] The cathode side or compartment 30 used a carbon felt electrode 22, of a type commonly used in batteries, and a 200 mM potassium carbonate (KHCO<sub>3</sub>) buffer. Using the carbon felt electrode 22 increased current density on average by over 10× for the same applied voltage, which may result in larger formate production rates over traditional copper electrodes that typically produce a wide array of other products. Both cathode and anode solutions 60, 52 were initially filled with a 100 mL volume. The cathode volume doubled to 200 mL when a pH buffer was employed to balance the proton flux from the anode 24, which diluted the charge mediator/carrier concentration. The charge mediator used in this study is 10 mM ethyl viologen (EV). The pH

buffer was only employed during the third set of testing when the packed bed reactor was present.

[0067] In the reduction cell 14 (FIG. 2), the anode solution 52 was circulated on the bottom side and the cathode 22 at the top, to minimize cathode transit time between the two cells. Unless otherwise stated, the default flow rate used during most experiments was 10 mL/min. The production cell 16 (FIG. 3) was essentially a bioelectrochemical cell, with no external wiring connections and served to house the catalyst 40. The catalyst 40 would receive reduced EV<sup>+</sup> from the reduction cell 14, along with CO<sub>2</sub> and H<sup>+</sup>, to produce HCOO<sup>-</sup>, converting the reduced EV<sup>+</sup> back to EV<sup>2+</sup> in the process (See FIG. 4). The production cell 16 was placed at a higher elevation than the reduction cell 14 and operated in a flow-through mode. From the production cell 16, one pathway would be to separate the formate from the solution and perform an additional protonation step using available technologies such as ion exchange (IEX) to produce a purified formic acid stream.

[0068] The pump pressure was used to oppose gravitational effects, which in this configuration, would ensure that the catholyte 60 would maximize its contact with the enzyme 40 in the production cell 16. Initially, microfiltration membranes were used, but they resulted in high pressure increases in the system even at small flow rates. Darcy's law was used to relate flow rate and pressure:

$$\Delta P = \frac{\mu LQ}{k} \tag{1}$$

where  $\mu$  represents the fluid viscosity, L is the length through which a pressure increase is expected, and Q is the flow rate. The permeability constant, k, was determined. All values obtained were on the order of  $10^{-19}~\text{m}^2$ , in line with impervious surfaces such as granite. Such results verified the difficulty of using high filtration membranes, even for water with viscosity of approximately 1 mPa·s. The size of the immobilized agarose beads attached to the catalyst 40 enabled it to be used in a much larger mesh, which kept the overall system pressure at more manageable levels. Ultimately, it was determined that the catalyst 40 was large enough to be contained in a  $121{\times}121$  size mesh membrane (McMaster-Carr).

[0069] The  $\rm CO_2$  reduction catalyst 40 in the production cell 16 was a formate dehydrogenase 1 from *Methylobacterium extorquens* AM1 (MeFDH1). An affinity tag was used to immobilize the enzyme by binding it onto agarose beads. Kinetic studies determined the enzyme's most favorable operating conditions, suggesting that temperatures above ambient would decrease activity. An econo-column (Bio-Rad) enabled that the enzyme detached from the beads, where UV-vis could be used to confirm the enzyme's optimal pH range of 6.3<pH<6.8. The enzyme samples were preserved in a MOPS (3-(N-morpholino) propanesulfonic acid) buffer and stored at 4° C. The approximate enzyme activity used in the experiment was 6.7  $\mu$ M, or approximately 200 U.

[0070] The cathode solution 60 initially consisted of 200 mM potassium bicarbonate (KHCO<sub>3</sub>) buffer with 10 mM ethyl viologen (EV). To minimize the contribution of dissolved atmospheric molecular oxygen present in the cathode solution, a 5 mM oxygen scavenger (sodium thiosulfate) was also employed. The scavenger served to decrease the

contribution of the current for unwanted reactions and therefore improves the  $\mathrm{CO}_2$  to formate conversion efficiency. The EV charge carrier was reduced in the reduction cell 14 and then reoxidized in the production cell 16 upon interaction with the catalyst 40 to produce formate. This formate was released by the catalyst back into the bulk solution where liquid samples could be drawn to determine the quantity produced. Samples were analyzed with ion chromatography (IC), mixed with a 6 M sulfuric acid solution to prevent the produced formate from reoxidizing. Samples in the flow system were collected at intervals sufficient to evaluate the formate production changes over time.

[0071] Voltage was applied using a Gamry REF-600 potentiostat 48. The system pressure was measured using an Omega PX-409 transducer. System voltages, along with pH, were also recorded using an external MIDI GL220 data logger 50. A pH buffer has also been implemented to offset pH drops in the system which could denature the enzymes 40, either due to increased presence of protons in the cathode side or production of formate, due to its low pKa number.

[0072] The pH controller 70 and pump 72 used to maintain the pH in the cathode compartment 30 by gradual dosing of potassium hydroxide (KOH)

[0073] The second set of experiments sought to enable better contact with EV in the production cell 16 to produce formate. Here, the production cell in FIG. 1 was replaced with a packed bed column. This column served a similar function as the production cell 16, with the catalyst 40 secured through the same 121×121 mesh membrane. The catalyst was embedded in between volumes of sand to better distribute contact with the packed bed. To determine the dependence of pressure with sand packing on hydrodynamic conditions, the Ergun equation was used:

$$\Delta P = \frac{150\mu L}{D_p^2} \frac{(1-\varepsilon)^2}{\varepsilon^3} v_s + \frac{1.75L\rho}{D_p} \frac{(1-\varepsilon)}{\varepsilon^3} v_s^2$$
 (2)

where  $\mu$  again represents fluid viscosity, L is the length of the bed,  $D_n$  is the packing spherical diameter,  $\varepsilon$  is the bed porosity, v<sub>s</sub> is the velocity of the fluid entering the bed, and  $\rho$  is the fluid density.  $D_p$  was obtained as 0.3 mm using known properties of the packing sand used, while porosity was obtained using the void/pore volume fractions. When the enzyme/catalyst 40 was included in the system, this porosity value was estimated at 0.1, but other values are included for reference. It should be noted that the variation in pressure with flow rate appears entirely linear; we attribute this to the extremely small velocities obtained in the system through the packed bed (reaching no more than 1 mm/s, corresponding to entirely laminar flow). The Ergun equation predicted pressure increases to over 200 kPa. This result is coupled with the initial equation based on the membrane selected, and suggests that, coupled with a microfiltration membrane, porosity should be kept high enough to prevent over-pressurizing the system. Thus, if system pressure is minimal, pumping energy for the system will be low, and thus not as major a concern when considering energy costs for dual-cell configuration long-term. This confirms that pressure and corresponding energy requirements for the system are decreased compared with microfiltration.

[0074] During optimal operation, the following reactions are expected to take place within the system:

Anode:

Oxygen Evolution/Proton Formation:

[0075]

$$2H_2O \rightarrow 4H^+ + 4e^- + O_2$$
 (1.032 V vs. Ag/AgCl, pH=0) (3)

Cathode, Reduction Cell:

First Reduction of EV:

[0076]

$$EV^{2+}+e^{-}\rightarrow EV^{+}(-0.646 \text{ V vs. Ag/AgCl, pH=6.3})$$
 (4)

Cathode, Production Cell:

Re-Oxidation of EV:

[0077]

$$EV^{+} \rightarrow EV^{2+} + e^{-}(-0.646 \text{ V vs. Ag/AgCl, pH=6.3})$$
 (5)

CO<sub>2</sub> to Formate: 
$$2\text{CO}_2 + 2\text{H}^+ + 4e^- \rightarrow 2\text{HCOO}^-$$
 (6)

[0078] While the initial buffer (KHCO<sub>3</sub>) is alkaline and purging the system with  $CO_2$  decreases the pH, the pH remains >6 and the product is formate. Additional, unwanted side reactions may also take place on the cathode side of the reduction cell:

Second Reduction of EV:

[0079]

$$EV^{+}+e^{-}\rightarrow EV^{0} (-1.09 \text{ V vs. Ag/AgCl, pH=6.3})$$
 (7)

Dimerization of EV:

[0800]

$$2EV^{+} \rightarrow [EV^{+}]_{2} (-0.9 \text{ V vs. Ag/AgCl, pH=6.3})$$
 (8)

Hydrogen Evolution:

[0081]

$$2H^++2e^- \rightarrow H_2 (-0.57 \text{ V vs. Ag/AgCl, pH=6.3})$$
 (9)

**[0082]** Dimerization of EV, however, has been shown to be minimal, particularly when compared with other charge carriers such as methyl viologen. Hydrogen, while also a desired product in electrochemical fuel production, is neither analyzed nor captured in this study.

[0083] The Coulombic efficiency (CE) of the system is calculated by relating the molar formate production in the system to the total electronic input. As a percentage, this value is expressed as:

$$CE = \frac{\text{(mol HCOO}^-)(\text{\# electrons required)(Faraday constant)}}{\text{(Charge Input)}} *100\%$$
 (10)

where the number of moles of formate produced is found by taking its concentration and multiplying by the solution volume in the system. The electrochemical conversion of  $CO_2$  into formate requires 2 electrons (Eq. 6). Charge input is given in Coulombs:

Charge Input = 
$$\int I(t)dt$$
 (11)

where I represents the input current (in A). For Eq. 10, an absolute value of this quantity is taken since the applied current is negative, typically consistent with system potential.

[0084] In summary, we report on the unique design and construction of a dual-cell flow system which electrochemically converts  $\mathrm{CO}_2$  to formate first using an electrochemical reduction of a charge carrier, then having said reduced charge carrier reoxidize with an enzymatic catalyst to produce HCOO $^-$ . The two processes are carried out in separate cells within the reactor. This is believed to be a first of its kind configuration that demonstrates successful conversion to formate and can be implemented into a range of different systems.

[0085] Each of the following terms written in singular grammatical form: "a", "an", and "the", as used herein, means "at least one", or "one or more". Use of the phrase "One or more" herein does not alter this intended meaning of "a", "an", or "the". Accordingly, the terms "a", "an", and "the", as used herein, may also refer to, and encompass, a plurality of the stated entity or object, unless otherwise specifically defined or stated herein, or, unless the context clearly dictates otherwise. For example, the phrase: "a charge carrier", as used herein, may also refer to, and encompass, a plurality of charge carriers.

[0086] Each of the following terms: "includes", "including", "has", "having", "comprises", and "comprising", and, their linguistic/grammatical variants, derivatives, or/and conjugates, as used herein, means "including, but not limited to", and is to be taken as specifying the stated component(s), feature(s), characteristic(s), parameter(s), integer(s), or step (s), and does not preclude addition of one or more additional component(s), feature(s), characteristic(s), parameter(s), integer(s), step(s), or groups thereof.

[0087] The phrase "consisting of", as used herein, is closed-ended and excludes any element, step, or ingredient not specifically mentioned. The phrase "consisting essentially of", as used herein, is a semi-closed term indicating that an item is limited to the components specified and those that do not materially affect the basic and novel characteristic(s) of what is specified.

[0088] Terms of approximation, such as the terms about, substantially, approximately, etc., as used herein, refers to  $\pm 10\%$  of the stated numerical value.

[0089] Although the apparatus and method of this disclosure have been illustratively described and presented by way of specific exemplary embodiments, and examples thereof, it is evident that many alternatives, modifications, or/and variations, thereof, will be apparent to those skilled in the art. Accordingly, it is intended that all such alternatives, modifications, or/and variations, fall within the spirit of, and are encompassed by, the broad scope of the appended claims.

What is claimed:

1. An apparatus adapted for production of formate from carbon dioxide, comprising:

- a reactor including a reduction cell and a production cell;
- a proton exchange membrane partitioning the reduction cell into an anode compartment and a cathode compartment;
- an anode in the anode compartment;
- a cathode in the cathode compartment;
- a voltage source connected across the anode and the cathode:
- a formate dehydrogenase catalyst immobilized in the production cell;
- a cathode solution circulating through the cathode compartment and the production cell, the cathode solution including a charge carrier; and
- an anode solution circulating through the anode compartment whereby (a) the charge carrier is reduced in the cathode compartment and reoxidized in the production cell and (b) carbon dioxide is converted to formate in the production cell.
- **2**. The apparatus of claim **1**, wherein the formate dehydrogenase catalyst is formate dehydrogenase 1 from *Methylobacterium extorquens* AM1.
- **3**. The apparatus of claim **2**, wherein the formate dehydrogenase 1 from *Methylobacterium extorquens* AM1 is immobilized on agarose beads.
- 4. The apparatus of claim 1, wherein the charge carrier is selected from a group consisting of ethyl viologen, methyl viologen dibromide, 1,1'-diheptyl-4,4'-bipyridinium dibromide, 1,1'-di-n-octyl-4,4'-bipyridinium dibromide, 1,1'-dibenzyl-4,4'-bipyridinium dichloride, 1,1'-bis(2,4-dinitrophenyl)-4,4'-bipyridinium dichloride, 1,1'-diphenyl-4,4'-bipyridinium dichloride, propyl viologen, butyl viologen, benzyl viologen, ethanol viologen, propanol viologen, methyl acetate viologen, acetate viologen, ethylamine viologen, propylamine viologen, and methyl ethylamine viologen, chloride salts thereof, bromide salts thereof and combinations thereof.
- **5**. The apparatus of claim **1**, further including an oxygen scavenger in the cathode solution.
- **6**. The apparatus of claim **5**, wherein the oxygen scavenger is selected from a group consisting of sodium thiosulfate, ascorbic acid, ferrous carbonate, sodium sulphite, sodium bisulphite, sodium metabisulphite, tannin, sodium erythorbate, hydrazine/carbohydrazine, N,N-diethylhydroxylamine and combinations thereof.
- 7. The apparatus of claim 1, further including a pH buffer in the cathode solution.
- **8**. The apparatus of claim **7**, wherein the pH buffer is selected from a group consisting of Tris buffer, Tris-Bis buffer, BES, buffer, MOPS buffer, TES buffer, HEPES buffer, MOPSO buffer, Imidazole (glyoxaline)-HCl buffer, potassium hydroxide, sodium hydroxide, potassium carbonate, potassium phosphate and mixtures thereof, adapted to maintain a pH of the cathode solution between pH 6 and pH 8
- **9.** The apparatus of claim **1**, wherein the anode is selected from a group consisting of a platinum wire mesh electrode, a platinum oxide coated carbon electrode, an iridium oxide coated carbon electrode, a cobalt oxide coated carbon electrode, a lead oxide/titanium alloy iridium-platinum coated carbon electrode, an iridium-ruthenium coated carbon electrode, and an iridium-tantalum coated carbon electrode.
- **10**. The apparatus of claim **1**, wherein the cathode is selected from a group consisting of a copper electrode, a tin-coated copper electrode, a bismuth-coated copper electrode.

trode, a lead-coated copper electrode, a bismuth-tin-lead alloys coated copper electrode, and bismuth-tin-lead alloys coated on porous carbon, carbon felt, carbon cloth, woven carbon, or carbon nanotube electrodes.

- 11. An apparatus adapted for production of formate from carbon dioxide, comprising:
  - a reactor including a reduction cell and a production cell;
  - a formate dehydrogenase catalyst immobilized in the production cell;
  - a cathode solution circulating through the reduction cell and the production cell, the cathode solution including a charge carrier; and
  - an anode solution circulating through the anode compartment whereby (a) the charge carrier is reduced in the reduction cell and reoxidized in the production cell and (b) carbon dioxide is converted to formate in the production cell.
- **12**. The apparatus of claim **11**, wherein the formate dehydrogenase catalyst is formate dehydrogenase 1 from *Methylobacterium extorquens* AM1.
- **13**. The apparatus of claim **12**, wherein the formate dehydrogenase 1 from *Methylobacterium extorquens* AM1 is immobilized on agarose beads.
- 14. The apparatus of claim 13, further including an oxygen scavenger and a pH buffer in the cathode solution.
- **15**. A method of producing formate from carbon dioxide in a dual cell reactor, comprising:
  - circulating a charge carrier solution between a reduction cell and a production cell;
  - reducing a charge carrier in the charge carrier solution in the reduction cell;
  - reoxidizing the charge carrier in the production cell; and converting carbon dioxide and protons to formate in presence of a formate dehydrogenase catalyst in the production cell.

16. The method of claim 15, further comprising: partitioning the reduction cell into an anode compartment and a cathode compartment with a proton exchange membrane;

locating an anode in the anode compartment; locating a cathode in the cathode compartment; and applying a voltage across the anode and the cathode.

- 17. The method of claim 15, further comprising circulating an oxygen scavenger with the charge carrier in the charge carrier solution.
- 18. The method of claim 17, further comprising circulating a pH buffer with the charge carrier in the charge carrier solution.
- 19. The method of claim 18, further comprising maintaining the charge carrier solution at a temperature between  $20^{\circ}$  C. and  $40^{\circ}$  C. and a pH of between pH 6 and pH 8.
- 20. The method of claim 19, including maintaining the applied voltage between -0.75 V to -0.95 V vs. Ag/AgCl.
- 21. A method of producing formate from carbon dioxide in a dual cell reactor including a reduction cell and a production cell, the method comprising:
  - partitioning the reduction cell into an anode compartment and a cathode compartment with a proton exchange membrane:

locating an anode in the anode compartment;

locating a cathode in the cathode compartment;

applying a voltage across the anode and the cathode; immobilizing a formate dehydrogenase catalyst in the production cell;

- circulating a cathode solution, including a charge carrier, through the cathode compartment and the production cell: and
- circulating an anode solution through the anode compartment whereby (a) the charge carrier is reduced in the cathode compartment and reoxidized in the production cell and (b) carbon dioxide is converted to formate in the production cell.

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