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COMPOSITIONS, SYSTEMS, AND METHODS FOR EXTRACTION OF METALS FROM MINERALS

Abstract

Provided herein are methods, systems, and compositions for degrading minerals. The methods, systems, and compositions provided herein involve the use of enzymes having silicase activity and an increased ability to degrade minerals such as silicate materials. The methods, systems, and compositions provided herein may be used to release metal from the amorphous silica. The methods, systems, and compositions provided herein may further involve collecting, extracting, and/or purifying the metal released from the minerals.

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Background/Summary

CROSS-REFERENCE [0001] This application is a continuation of International Application No. PCT/US2024/046780, filed Sep. 13, 2024, which claims the benefit of U.S. Provisional Application No. 63/583,201, filed Sep. 15, 2023, each of which is incorporated herein by reference in its entirety.

SEQUENCE LISTING

[0002] The instant application contains a Sequence Listing which has been submitted electronically in XML format and is hereby incorporated by reference in its entirety. Said XML copy, created on Oct. 14, 2024, is named 66122_702_301.xml and is 614,155 bytes in size.

BACKGROUND

[0003] Metals such as lithium, aluminum, iron, nickel, cobalt, strontium, and rare earth elements have vast industrial applications and are in high demand across various industries. For example, lithium is widely used for energy storage, rechargeable batteries, electronic motors, electric vehicles, air mobility, clean energy, energy storage from solar panels, and other applications. Lithium has pharmaceutical applications, such as in lithium-based bipolar disorder treatments. Currently available sources and technologies for obtaining metals for use in industrial applications and products are limited, inefficient, costly, energy-intensive, and harmful to the environment. SUMMARY

[0004] There is a significant unmet need for compositions, methods, and systems that facilitate access to existing sources of metals, such as to extract and/or collect the metals from their sources, separate them, process them, and make them available for use in various industrial applications and/or products, in a manner that is industrially scalable, efficient, and inexpensive. This is at least in part to meet the demand for such metals in the industrial applications and products in need thereof. Many of the currently available methods and techniques for doing so are limited with respect to efficiency, scalability, and high cost. In many cases, such existing technologies may require performing processes and reactions at high temperatures and pressures that are energyintensive, costly, and harmful to the environment. The compositions, methods, and systems of the present disclosure address the aforementioned needs and shortcomings, in some aspects, by providing compositions, methods, and systems for extracting, separating, and/or collecting metals from mineral sources, such as mineral materials including solid mineral materials, natural mineral materials, man-made mineral materials, rocks, ores, deposits, and/or other sources in an efficient, inexpensive, and scalable manner. In some cases, the disclosure further provides methods and systems for processing the extracted metals and/or using them in a product (e.g., rechargeable batteries). The extracted metal may be processed and turned into an industrial grade metal, battery grade metal, pharmaceutical grade metal, or other useful forms of metals.

[0005] In an aspect, provided herein is a method of extracting a metal from a mineral, the method comprising: (a) contacting the mineral material with an enzyme having silicase activity under reaction conditions such that the metal contained within the mineral material is solubilized and released; and (b) collecting the released metal, thereby extracting the metal from the mineral material. In some embodiments, the mineral material comprises an ore, a rock, a natural mineral material, a man-made mineral material, or any combination thereof. In some embodiments, the mineral material comprises a silicate. In some embodiments, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, a tectosilicate, or any combination thereof. In

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some embodiments, the amorphous silicate is selected from the group consisting of: obsidian, coal
fly ash, pumice, glass, and any combination thereof. In some embodiments, the tectosilicate
comprises quarts, sand, or both. In some embodiments, the enzyme having silicase activity has a
sequence identity of at least about at least about 30%, at least about 40%, at least about 50%, at
least about 60%, at least about 70%, at least about 80%, at least about 90%, or more identity with a
carbonic anhydrase. In some embodiments, the enzyme having silicase activity has a sequence
identity of at least about 30%, at least about 400%, at least about 50%, at least about 60%, at least
about 70%, at least about 80%, at least about 90%, or more identity with an alpha carbonic
anhydrase. In some embodiments, the enzyme having silicase activity has a sequence identity at
least at least about 30%, at least about 40%, about 50%, at least about 60%, at least about 70%, at
least about 80%, at least about 90%, or more identity with a gamma carbonic anhydrase. In some
embodiments, the enzyme having silicase activity is derived from an organism selected from the
group consisting of: of: Methanosarcina thermophila, Bacillus licheniformis CG-B52, Pelobacter
carbinolicus, Syntrophus aciditrophicus, Methanosarcina barkeri, Methanosarcina mazei, Bacillus
halodurans, Alkalihalobacillus clausii (strain KSM-K16) (Bacillus clausii), Methanosarcina
acetivorans, Kofleriaceae bacterium SLC26A/SulP, Thermodesulfitimonas autotrophica,
Fischerella thermalis/Mastigocladus laminosus, Thermosynechococcus vestitus BP-
1/(Thermosynechococcus elongatus BP-1) carboxysome assembly protein CcmM, Methanothrix
thermoacetophila, Thermosyntropha lipolytica, isoleucine patch superfamily, Desulfofundulus
thermobenzoicus, Archaeoglobus veneficus, Suberites domuncula, and any combination thereof. In
some embodiments, the enzyme having silicase activity has a sequence identity of at least about
30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about
80%, at least about 90% or more, with a Methanosarcina thermophila gamma carbonic anhydrase,
Bacillus licheniformis CG-B52 gamma carbonic anhydrase, Pelobacter carbinolicus gamma
carbonic anhydrase, Syntrophus aciditrophicus gamma carbonic anhydrase, Methanosarcina
barkeri gamma carbonic anhydrase, Methanosarcina mazei carbonic anhydrase, Bacillus
halodurans alpha carbonic anhydrase, Alkalihalobacillus clausii (strain KSM-K16) (Bacillus
clausii) alpha carbonic anhydrase, Methanosarcina acetivorans carbonate dehydratase,
Kofleriaceae bacterium SLC26A/SulP transporter domain-containing protein,
Thermodesulfitimonas autotrophica carbonic anhydrase/acetyltransferase-like protein (Isoleucine
patch superfamily), Fischerella thermalis/Mastigocladus laminosus JSC-11 carboxysome assembly
protein CcmM, Thermosynechococcus vestitus BP-1/(Thermosynechococcus elongatus BP-1)
carboxysome assembly protein CcmM, Methanothrix thermoacetophila carbonate dehydratase,
Thermosyntropha lipolytica carbonic anhydrase or acetyltransferase, isoleucine patch superfamily,
Desulfofundulus thermobenzoicus transferase, Archaeoglobus veneficus carbonate dehydratase,
Suberites domuncula carbonic anhydrase. In some embodiments, the enzyme having silicase
activity has an amino acid sequence having at least 50%, at least 55%, at least 60%, at least 65%, at
least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least
93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least
99.5%, or at least 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ
ID NOS: 1-18. In some embodiments, the enzyme having silicase activity is an engineered enzyme,
and wherein the engineered enzyme has the sequence of any one of SEQ ID NOS: 19-402, or an
amino acid sequence having at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at
least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least
94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least
99.9%, or more sequence identity to any one of SEQ ID NOS: 19-402. In some embodiments, the
enzyme having silicase activity is an enzyme having at least one amino acid variation as compared
to a wild-type enzyme. In some embodiments, the enzyme having silicase activity has a pKd of at
least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at
least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat
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value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 100 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 800 mol/s, at least about 900 mol/s, at least about 1000 mol/s, or higher. In some embodiments, the reaction conditions comprise a temperature from about 23 to about 85 degrees Celsius (C). In some embodiments, the reaction conditions comprise a temperature from about 45 to about 50 degrees Celsius (C). In some embodiments, the reaction conditions comprise a temperature of about 50 degrees Celsius (C). In some embodiments, the reaction conditions comprise a pH from about 4 to about 11. In some embodiments, the reaction conditions comprise a pH of about 5. In some embodiments, the reaction conditions comprise a pH of about 10. In some embodiments, the reaction conditions comprise contacting the enzyme having silicase activity with a co-factor. In some embodiments, the co-factor is selected from the group consisting of: iron, zinc, copper, nickel, and cobalt. In some embodiments, the co-factor is iron. In some embodiments, the enzyme having silicase activity depolymerizes silicate mineral in the mineral material. In some embodiments, the enzyme having silicase activity cleaves one or more Si—O bonds in the mineral material to generate silicic acid (Si(OH)4). In some embodiments, the metal is lithium, aluminum, iron, nickel, cobalt, strontium, or a rare earth element. In some embodiments, the metal is lithium. In some embodiments, the metal is iron. In some embodiments, the metal is aluminum. In some embodiments, the metal is strontium. In some embodiments, the metal is released into a solution. In some embodiments, the method further comprises extracting the metal from the solution. In some embodiments, the method further comprises purifying the metal from the solution, thereby generating a purified metal. In some embodiments, the purified metal has a purity of at least about 80%. In some embodiments, the purified metal has a purity of at least about 90%. In some embodiments, the purified metal has a purity of at least about 95%. In some embodiments, the purified metal has a purity of at least about 99%. In some embodiments, the purified metal has a purity of at least about 99.99%. In some embodiments, the purified metal has a purity of at least about 99.999%. In some embodiments, the purified metal is purified lithium. In some embodiments, the purified lithium is industrial grade, battery grade, or pharmaceutical grade. In some embodiments, the method is performed in situ or ex situ. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the enzyme having silicase activity is recombinantly produced in a host cell or in a cell-free production system. In some embodiments, the host cell is a bacterial cell or yeast cell. In some embodiments, the bacterial cell is *Escherichia coli* or the yeast cell is *Pichia pastoris* or Saccharomyces cerevisiae. In some embodiments, the reaction conditions comprise a rock to liquid ratio from about 1-40% (w/v). In some embodiments, the reaction conditions comprise a rock to liquid ratio of about 30% (w/v). In some embodiments, the reaction conditions comprise a buffer. In some embodiments, the buffer is TRIS, PBS, citrate, monosodium glutamate, or a combination thereof.

[0006] In an aspect, provided herein is a method of extracting a metal from a mineral. The method comprises: extracting a metal from a mineral material, the method comprising: contacting the mineral material with an enzyme having silicase activity under reaction conditions such that the metal contained within the mineral material is solubilized and released, wherein the reaction conditions comprise a temperature from about 23-85 degrees Celsius (C), a pH from about 4-11, a co-factor, and a rock to liquid ratio from about 1-40% (w/v), and further comprises collecting the released metal, thereby extracting the metal from the mineral material. In some embodiments, the

reaction conditions proceed for about 1-48 hours. In some embodiments, the reaction conditions proceed for about 48 hours. In some embodiments, the reaction conditions comprise a temperature of about 50 degrees Celsius (C). In some embodiments, the reaction condition comprises a pH of about 10. In some embodiments, the co-factor is zinc, iron, copper, cobalt, or any combination thereof. In some embodiments, the co-factor is iron. In some embodiments, the rock to liquid ratio is about 30% (w/v).

[0007] In an aspect, provided herein is a non-naturally occurring enzyme having silicase activity, the enzyme comprising at least one amino acid variation relative to a wild-type enzyme and having increased ability to release metals from mineral materials as compared to the wild-type enzyme. In some embodiments, the wild-type enzyme is selected from the group consisting of: Methanosarcina thermophila gamma carbonic anhydrase, Bacillus licheniformis CG-B52 gamma carbonic anhydrase, Pelobacter carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma carbonic anhydrase, *Methanosarcina barkeri* gamma carbonic anhydrase, *Methanosarcina* mazei carbonic anhydrase, Bacillus halodurans alpha carbonic anhydrase, Alkalihalobacillus clausii (strain KSM-K16) (Bacillus clausii) alpha carbonic anhydrase, Methanosarcina acetivorans carbonate dehydratase, Kofleriaceae bacterium SLC26A/SulP transporter domain-containing protein, Thermodesulfitimonas *autotrophica* carbonic anhydrase/acetyltransferase-like protein (Isoleucine patch superfamily), *Fischerella thermalis/Mastigocladus laminosus* JSC-11 carboxysome assembly protein CcmM, *Thermosynechococcus vestitus* BP-1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix* thermoacetophila carbonate dehydratase, Thermosyntropha lipolytica carbonic anhydrase or acetyltransferase, isoleucine patch superfamily, Desulfofundulus thermobenzoicus transferase, Archaeoglobus veneficus carbonate dehydratase, Suberites domuncula carbonic anhydrase, and any combination thereof. In some embodiments, the non-naturally occurring enzyme comprises an amino acid sequence having at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-18. In some embodiments, the wild-type enzyme is a carbonic anhydrase. In some embodiments, the carbonic anhydrase is a gamma carbonic anhydrase or alpha carbonic anhydrase. In some embodiments, the mineral materials comprise rock, ore, natural mineral, man-made mineral, or any combination thereof. In some embodiments, the mineral materials comprise a silicate. In some embodiments, the mineral materials comprise inosilicates, phyllosilicates, amorphous silicates, tectosilicates, or any combination thereof. In some embodiments, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some embodiments, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some embodiments, the amorphous silicate is selected from the group consisting of obsidian, coal fly ash, pumice, and any combination thereof. In some embodiments, the tectosilicate comprises sand, glass, quartz, or any combination thereof. In some embodiments, the non-naturally occurring enzyme has increased ability to depolymerize silicate mineral in the mineral material as compared to the wild-type enzyme, increased selectivity or specificity toward a mineral structure in the mineral material, or both. In some embodiments, the non-naturally occurring enzyme has increased ability to cleave one or more Si—O bonds in the mineral material to generate silicic acid (Si(OH)4) as compared to the wild-type enzyme. In some embodiments, the metal comprises lithium, aluminum, iron, strontium, or any combinations thereof. In some embodiments, the metal comprises lithium. In some embodiments, the metal comprises iron. In some embodiments, the metal comprises aluminum. In some embodiments, the metal comprises strontium. In some embodiments, the non-naturally occurring enzyme is recombinantly produced in a host cell or in a cell-free production system. In

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some embodiments, the host cell is a bacterial cell or yeast cell. In some embodiments, the bacterial
cell is Escherichia coli or the yeast cell is Pichia pastoris or Saccharomyces cerevisiae.
[0008] In an aspect, provided herein is a reaction mixture comprising a mineral material and a non-
naturally occurring enzyme having silicase activity, wherein the non-naturally occurring enzyme
comprises at least one amino acid variation relative to a wild-type enzyme and has increased ability
to release metals from the mineral material as compared to the wild-type enzyme. In some
embodiments, the mineral material comprises an ore, a rock, a natural mineral material, a man-
made mineral material, or any combination thereof. In some embodiments, the reaction mixture has
a pH from about 4 to about 11. In some embodiments, the reaction mixture has a pH of about 5. In
some embodiments, the reaction mixture has a pH of about 10. In some embodiments, the reaction
mixture has a temperature from about 23 to about 85 degrees Celsius (C). In some embodiments,
the reaction mixture has a temperature from about 45 to about 50 degrees Celsius (C). In some
embodiments, the reaction mixture has a temperature of about 50 degrees Celsius (C). In some
embodiments, the reaction mixture further comprises a co-factor of the non-naturally occurring
enzyme. In some embodiments, the co-factor is selected from the group consisting of: iron, zinc,
copper, nickel, and cobalt. In some embodiments, the co-factor is copper. In some embodiments,
the co-factor is iron. In some embodiments, the reaction mixture further comprises a buffered saline
solution. In some embodiments, the reaction mixture further comprises an activator co-factor of the
non-naturally occurring enzyme. In some embodiments, the activator co-factor is glycine. In some
embodiments, the wild-type enzyme is selected from the group consisting of: Methanosarcina
thermophila gamma carbonic anhydrase, Bacillus licheniformis CG-B52 gamma carbonic
anhydrase, Pelobacter carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma
carbonic anhydrase, Methanosarcina barkeri gamma carbonic anhydrase, Methanosarcina mazei
carbonic anhydrase, Bacillus halodurans alpha carbonic anhydrase, Alkalihalobacillus clausii
(strain KSM-K16) (Bacillus clausii) alpha carbonic anhydrase, Methanosarcina acetivorans
carbonate dehydratase, Kofleriaceae bacterium SLC26A/SulP transporter domain-containing
protein, Thermodesulfitimonas autotrophica carbonic anhydrase/acetyltransferase-like protein
(Isoleucine patch superfamily), Fischerella thermalis/Mastigocladus laminosus JSC-11
carboxysome assembly protein CcmM, Thermosynechococcus vestitus BP-
1/(Thermosynechococcus elongatus BP-1) carboxysome assembly protein CcmM, Methanothrix
thermoacetophila carbonate dehydratase, Thermosyntropha lipolytica carbonic anhydrase or
acetyltransferase, isoleucine patch superfamily, Desulfofundulus thermobenzoicus transferase,
Archaeoglobus veneficus carbonate dehydratase, Suberites domuncula carbonic anhydrase, and any
combination thereof. In some embodiments, the non-naturally occurring enzyme comprises an
amino acid sequence having at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at
least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least
94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least
99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-18. In
some embodiments, the wild-type enzyme is a carbonic anhydrase. In some embodiments, the
carbonic anhydrase is a gamma carbonic anhydrase or alpha carbonic anhydrase. In some
embodiments, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate,
a tectosilicate or any combination thereof. In some embodiments, the inosilicate is selected from
the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite,
shattuckite, augite, tremolite, and any combination thereof. In some embodiments, the
phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite,
muscovite, montmorillonite, and any combination thereof. In some embodiments, the amorphous
silicate is selected from the group consisting of: obsidian, coal fly ash, pumice, glass, and any
combination thereof. In some embodiments, the non-naturally occurring enzyme has increased
ability to depolymerize silicate in the mineral material as compared to the wild-type enzyme. In
some embodiments, the non-naturally occurring enzyme has increased ability to cleave one or more
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Si—O bonds in the mineral material to generate silicic acid (Si(OH)4) as compared to the wild-type enzyme. In some embodiments, the metal comprises lithium. In some embodiments, the metal comprises aluminum. In some embodiments, the metal comprises iron. In some embodiments, the metal comprises strontium. In some embodiments, the non-naturally occurring enzyme is recombinantly produced in a host cell. In some embodiments, the host cell is a bacterial cell or a yeast cell. In some embodiments, the bacterial cell is *Escherichia coli* or the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*. In some embodiments, the reaction mixture has a rock to liquid ratio from about 1-40% (w/v). In some embodiments, the reaction mixture has a buffer. In some embodiments, the buffer is TRIS, PBS, citrate, monosodium glutamate, or a combination thereof. In some embodiments, the reaction mixture proceeds for about 1-48 hours. In some embodiments, the reaction mixture proceeds for about 48 hours.

[0009] In an aspect, provided herein is a polynucleotide comprising a nucleotide sequence encoding the non-naturally occurring enzyme disclosed herein.

[0010] In an aspect, provided herein is a vector comprising the polynucleotide disclosed herein. [0011] In an aspect, provided herein is a method of increasing silicase activity of an enzyme, the method comprising contacting the enzyme with a non-natural co-factor, wherein the non-natural co-factor increases silicase activity of the enzyme as compared to the enzyme in the presence of a natural co-factor. In some embodiments, the non-natural co-factor is copper. In some embodiments, the natural co-factor is zinc. In some embodiments, the natural co-factor is iron. In some embodiments, the method is performed in the absence of the natural co-factor. In some embodiments, the non-natural co-factor does not act as a co-factor for the enzyme having silicase activity in nature. In some embodiments, the method further comprises contacting the enzyme and the non-natural co-factor with a mineral material under reaction conditions such that a metal contained within the mineral material is solubilized and released from the mineral material. In some embodiments, the amount of metal solubilized and released from the mineral material when the enzyme is contacted with the natural co-factor.

[0012] In some embodiments, the mineral material comprises a rock, an ore, a natural mineral material, a man-made mineral material, or any combination thereof. In some embodiments, the mineral material comprises a silicate. In some embodiments, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, a tectosilicate, or any combination thereof. In some embodiments, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some embodiments, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some embodiments, the amorphous silicate is selected from the group consisting of: obsidian, coal fly ash, pumice, glass, and any combination thereof. In some embodiments, the enzyme having silicase activity is a carbonic anhydrase. In some embodiments, the carbonic anhydrase is a gamma carbonic anhydrase or alpha carbonic anhydrase. In some embodiments, the enzyme having silicase activity is derived from an organism selected from the group consisting of: *Methanosarcina thermophila, Bacillus licheniformis* CG-B52, Pelobacter carbinolicus, Syntrophus aciditrophicus, Methanosarcina barkeri, Methanosarcina mazei, Bacillus halodurans, Alkalihalobacillus clausii (strain KSM-K16) (Bacillus clausii), Methanosarcina acetivorans, Kofleriaceae bacterium, Thermodesulfitimonas autotrophica, Fischerella thermalis/Mastigocladus laminosus JSC-11 carboxysome assembly protein CcmM, *Thermosynechococcus vestitus* BP-1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix* thermoacetophila, Thermosyntropha lipolytica, Desulfofundulus thermobenzoicus, Archaeoglobus *veneficus*, Suberites domuncula, and any combination thereof. In some embodiments, the enzyme having silicase activity comprises an amino acid sequence having at least 50%, at least 55%, at

least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-18. In some embodiments, the enzyme having silicase activity is an engineered enzyme, optionally wherein the engineered enzyme has the sequence of any one of SEQ ID NOS: 19-402. In some embodiments, the enzyme having silicase activity is an enzyme having at least one amino acid variation as compared to a wild-type enzyme. In some embodiments, the enzyme having silicase activity has a pKd of at least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 100 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 800 mol/s, at least about 900 mol/s, at least about 1000 mol/s, or higher. In some embodiments, the method is performed under reaction conditions. In some embodiments, the reaction conditions comprise a temperature from about 23 to about 85 degrees Celsius (C). In some embodiments, the reaction conditions comprise a temperature from about 45 to about 50 degrees Celsius (C). In some embodiments, the reaction conditions comprise a temperature of about 50 degrees Celsius (C). In some embodiments, the reaction conditions comprise a pH from about 4 to about 11. In some embodiments, the reaction conditions comprise a pH of 5. In some embodiments, the reaction conditions comprise a pH of 10. In some embodiments, the enzyme having silicase activity depolymerizes silicate mineral in the mineral material. In some embodiments, the enzyme having silicase activity cleaves one or more Si—O bonds in the mineral material to generate silicic acid (Si(OH)4). In some embodiments, the metal is selected from the group consisting of: lithium, aluminum, iron, nickel, cobalt, strontium, and rare earth metals. In some embodiments, the metal is lithium. In some embodiments, the metal is iron. In some embodiments, the metal is aluminum. In some embodiments, the metal is strontium. In some embodiments, the metal is released into a solution. In some embodiments, the method further comprises extracting the metal from the solution. In some embodiments, the method further comprises purifying the metal from the solution, thereby generating a purified metal, a solid metal complex, a metal precipitate, or any combination thereof. In some embodiments, the purified metal has a purity of at least about 80%, at least about 90%, at least about 95%, at least about 99%, at least about 99.99%, at least about 99.9999% or greater. In some embodiments, the method is performed in situ or ex-situ. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the enzyme having silicase activity is recombinantly produced in a host cell or in a cell-free production system. In some embodiments, the host cell is a bacterial cell or a yeast cell. In some embodiments, the bacterial cell is *Escherichia coli* or the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*. In some embodiments, the reaction conditions comprise a rock to liquid ratio from about 1-40% (w/v). In some embodiments, the reaction conditions comprise a rock to liquid ratio of about 30% (w/v). In some embodiments, the reaction conditions comprise a buffer. In some embodiments, the buffer is TRIS, PBS, citrate, monosodium glutamate, or a combination thereof. [0013] In an aspect, provided herein is a reaction mixture comprising an enzyme having silicase activity, and a non-natural co-factor.

[0014] In some embodiments, the non-natural co-factor is bound to the enzyme having silicase activity. In some embodiments, the non-natural co-factor increases a function of the enzyme having

silicase activity as compared to a reaction mixture comprising the enzyme having silicase activity and a natural co-factor. In some embodiments, the non-natural co-factor is copper. In some embodiments, the natural co-factor is zinc. In some embodiments, the natural co-factor is iron. In some embodiments, the reaction mixture does not contain the natural co-factor. In some embodiments, the non-natural co-factor does not act as a co-factor for the enzyme having silicase activity in nature. In some embodiments, the reaction mixture further comprises a mineral material and reaction conditions such that a metal contained within the mineral material is solubilized and released from the mineral material. In some embodiments, the enzyme having silicase activity has increased ability to release metals from the mineral material in the presence of the non-natural cofactor as compared to the enzyme having silicase activity in the presence of the natural co-factor. In some embodiments, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, or any combination thereof. In some embodiments, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some embodiments, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some embodiments, the amorphous silicate is selected from the group consisting of: obsidian, coal, pumice, glass, and any combination thereof. In some embodiments, the enzyme having silicase activity has a sequence identity of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more identity with a carbonic anhydrase. In some embodiments, the enzyme having silicase activity has a sequence identity of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more identity with an alpha carbonic anhydrase or a gamma carbonic anhydrase. In some embodiments, the enzyme having silicase activity is derived from an organism selected from the group consisting of: *Methanosarcina thermophila*, *Bacillus licheniformis* CG-B52, Pelobacter carbinolicus, Syntrophus aciditrophicus, Methanosarcina barkeri, Methanosarcina mazei, Bacillus halodurans, Alkalihalobacillus clausii (strain KSM-K16) (Bacillus clausii), *Methanosarcina acetivorans*, Kofleriaceae bacterium SLC26A/SulP, Thermodesulfitimonas autotrophica, Fischerella thermalis/Mastigocladus laminosus, Thermosynechococcus vestitus BP-1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix* thermoacetophila, Thermosyntropha lipolytica, isoleucine patch superfamily, Desulfofundulus thermobenzoicus, Archaeoglobus veneficus, Suberites domuncula, and any combination thereof. In some embodiments, the enzyme having silicase activity comprises an amino acid sequence having at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-18. In some embodiments, the enzyme having silicase activity is an engineered enzyme, optionally wherein the enzyme has the sequence of any one of SEQ ID NOS: 19-402. In some embodiments, the enzyme having silicase activity is an enzyme having at least one amino acid variation as compared to a wild-type enzyme. In some embodiments, the enzyme having silicase activity has a pKd of at least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 100 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 800 mol/s, at least about 900 mol/s, at least about 1000 mol/s, or higher. In some embodiments, the reaction mixture has a temperature from about 23 to about 85 degrees Celsius (C). In some embodiments, the reaction mixture has a temperature from about 45 to about 50 degrees Celsius (C). In some embodiments, the reaction mixture has a temperature of about 50 degrees Celsius (C). In some embodiments, the reaction

mixture has a pH from about 4 to about 11. In some embodiments, the reaction mixture has a pH of 5. In some embodiments, the reaction mixture has a pH of 10. In some embodiments, the reaction mixture further comprises a buffered saline solution. In some embodiments, the reaction mixture further comprises an activator co-factor of the non-naturally occurring enzyme. In some embodiments, the activator co-factor is glycine or an iron ion. In some embodiments, the metal is lithium, aluminum, iron, nickel, cobalt, strontium, or a rare earth element. In some embodiments, the metal is lithium. In some embodiments, the metal is iron. In some embodiments, the metal is aluminum. In some embodiments, the metal is strontium. In some embodiments, the enzyme having silicase activity is recombinantly produced in a host cell. In some embodiments, the host cell is a bacterial cell or yeast cell. In some embodiments, the bacterial cell is *Escherichia coli* or the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*. In some embodiments, the reaction conditions comprise a rock to liquid ratio from about 1-40% (w/v). In some embodiments, the reaction conditions comprise a rock to liquid ratio of about 30% (w/v). In some embodiments, the reaction conditions comprise a buffer. In some embodiments, the buffer is TRIS, PBS, citrate, monosodium glutamate, or a combination thereof. In some embodiments, the reaction conditions proceed for about 1-48 hours. In some embodiments, the reaction conditions proceed for about 48 hours.

INCORPORATION BY REFERENCE

[0015] All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference.

Description

BRIEF DESCRIPTION OF THE DRAWINGS

[0016] The novel features of the invention are set forth with particularity in the appended claims. A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings of which:

[0017] FIG. **1** shows an example workflow according to the methods of the present disclosure; [0018] FIG. **2** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals(Alpha Spodumene and Beta Spodumene) using an enzyme having silicase activity according to the embodiments of the present disclosure;

[0019] FIG. **3** presents the rate of production of Si(OH)4 corresponding to reaction rates of degrading silicate minerals (iron ore, platinum group metal (PGM) tailing, and Bauxite) using an enzyme having silicase activity according to the embodiments of the present disclosure; [0020] FIG. **4** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (Rhyolite and Olivine) using an enzyme having silicase activity according to the embodiments of the present disclosure;

[0021] FIG. **5** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (Hectorite mix, Clay, a silicate named Maverick source, and a Lepidolite) using an enzyme having silicase activity according to the embodiments of the present disclosure;

[0022] FIG. **6** presents the rate of production of Si(OH)4 corresponding to reaction rates of degrading silicate minerals (crushed glass and Perlite) using an enzyme having silicase activity according to the embodiments of the present disclosure; and

[0023] FIG. 7 presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (Oil Shale and Fly Ash) using an enzyme having silicase activity according to the embodiments of the present disclosure.

DETAILED DESCRIPTION

[0024] Metals such as lithium, aluminum, iron, nickel, cobalt, strontium, and rare earth elements have vast applications across various industries and in different products. The demand for such metals continues to increase, and there is an unmet need for efficient technologies to facilitate access to metal sources, and to extract and collect the metals for use in products and industries in need thereof and to meet demand. As an example, lithium is highly in demand for rechargeable batteries which can be used in a variety of products such as electronics, electric motors and electric vehicles, clean energy industry, solar panels, and beyond. As these industries advance and become more prominent in global markets, so does the demand for lithium. Lithium can be found in a number of sources, including in brine, for example, in brine deposits generated as a result of accumulations of saline groundwater enriched in dissolved lithium. However, brine sources of lithium are limited in abundance and can only be found in limited geographical locations, mostly located in South America.

[0025] Another prominent and abundant source of metals are mineral materials, such as natural minerals (e.g., rock, ore, clay) and man-made minerals that are commonly available across the world in a diverse range of geographical areas, constituting a major primary source of metals. The currently available technologies for extracting metals from solid mineral materials/sources, rocks, and ores are limited, inefficient, costly, and environmentally harmful. An example of such process is acid leaching or acid roasting which involves contacting a mineral with a strong acid (e.g., sulfuric acid) to extract a metal, such as lithium, from the mineral. Acid leaching/roasting usually requires reaction conditions involving highly acidic pH, high temperatures (e.g., 200 degrees Celsius (C) and above) and significant energy consumption (e.g., over 6000 megajoules (MJ) per ton of Li.sub.2O extracted). This process is expensive, energy-inefficient, and harmful to the environment. Therefore, there is an unmet need for improved compositions, methods, and systems to address these shortcomings.

[0026] Provided herein are compositions, methods, and systems that can efficiently extract metals from minerals (e.g., natural minerals (e.g., rock, ore, clay), man-made minerals) in an efficient, industrially scalable, inexpensive, and environmentally friendly fashion. For example, in some cases, the methods may avoid reaction conditions requiring substances (e.g., highly acidic solvents), high temperatures and pressures, and the like, which may cause harm to the environment and/or increase the cost, energy demands, and/or environmental footprint of the process. In some cases, this is accomplished by performing an enzymatic reaction on a mineral material, to extract and separate a metal (e.g., metal ion/atom) therefrom. The enzymatic reaction may have improved features such as higher reaction rate, specificity toward degrading a mineral material, acting on a certain substrate in the mineral material with high/improved substrate specificity, such that performing the reaction can be industrially scaled and implemented for releasing and collecting the metal. For example, in some cases, the reaction may not require temperatures that are significantly higher than room temperature, pressures significantly higher than atmospheric pressure, highly acidic or highly basic pH conditions, and other conditions that are environmentally harmful and costly. Instead, in many cases, the reactions of the present disclosure may be efficiently performed in near-ambient temperature, near-atmospheric pressure, and/or near-neutral pH conditions, reducing their cost, energy demand, and environmental footprint. The details of such reaction conditions are further elaborated on herein.

[0027] In some cases, the enzymatic reaction may comprise using one or more enzymes and/or co-factors. The enzymatic reaction may comprise an enzymatic degradation/digestion reaction performed with the aid of one or more enzymes and/or co-factors. The enzymes may catalyze the reaction and facilitate the extraction of the metal from the mineral material encasing it. For example, enzymes can be used to degrade, dissolve, and/or depolymerize silicates, and liberate metals (e.g., metal ions) therefrom at near-ambient temperatures without the need for an energy-intensive, high temperature acid separation process. This process significantly decreases the

environmental impact of refining lithium and other metals deposited in mineral materials. Provided herein are also enzymes and co-factors with enhanced features and capabilities for performing such reactions. Such enzymes may be semi-synthetic and/or engineered enzymes with features, capabilities, sequences, methods of generation, and methods of use that are presented and further elaborated on in the present disclosure.

[0028] The term "sequence identity" as used herein generally refers to an exact nucleotide-tonucleotide or amino acid-to-amino acid correspondence of two polynucleotides or polypeptide sequences, respectively. Typically, techniques for determining sequence identity include determining the nucleotide sequence of a polynucleotide and/or determining the amino acid sequence encoded thereby, and comparing these sequences to a second nucleotide or amino acid sequence. Two or more sequences (polynucleotide or amino acid) can be compared by determining their percentage (%) of "sequence identity". The % of sequence identity of two sequences, whether nucleic acid or amino acid sequences, is the number of exact matches between two aligned sequences divided by the length of the longer sequence and multiplied by 100. Percent identity may also be determined, for example, by comparing sequence information using the advanced BLAST computer program, including version 2.2.9, available from the National Institutes of Health. The BLAST program is based on the alignment method of Karlin and Altschul, Proc. Natl. Acad. Sci. USA, 87:2264-2268 (1990) and as discussed in Altschul, et al., J. Mol. Biol., 215:403-410 (1990); Karlin And Altschul, Proc. Natl. Acad. Sci. USA, 90:5873-5877 (1993); and Altschul et al., Nucleic Acids Res., 25:3389-3402 (1997). The program may be used to determine percent identity over the entire length of the proteins being compared. Default parameters are provided to optimize searches with short query sequences in, for example, with the blastp program. The program also allows use of an SEG filter to mask-off segments of the query sequences as determined by the SEG program of Wootton and Federhen, Computers and Chemistry 17:149-163 (1993). Ranges of desired degrees of sequence identity are approximately 50% to 100% and integer values therebetween. In general, this disclosure encompasses sequences with at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least 99.9% sequence identity with any sequence provided herein. [0029] The terms "variant enzyme", "enzyme variant", "modified enzyme", "synthetic enzyme",

[0029] The terms "variant enzyme", "enzyme variant", "modified enzyme", "synthetic enzyme", "truncated enzyme", and "engineered enzyme" are used interchangeably throughout to generally refer to non-naturally occurring polypeptides. The non-naturally occurring polypeptides have been designed and sequences included herein.

[0030] The term "about" or "approximately" generally means within an acceptable error range for the particular value as determined by one of ordinary skill in the art, which will depend in part on how the value is measured or determined, i.e., the limitations of the measurement system. For example, "about" can mean within 1 or more than 1 standard deviation, per the practice in the art. Alternatively, "about" can mean a range of up to 20%, up to 10%, up to 5%, or up to 1% of a given value. Alternatively, particularly with respect to biological systems or processes, the term can mean within an order of magnitude, preferably within 5-fold, and more preferably within 2-fold, of a value. Where particular values are described in the application and claims, unless otherwise stated, the term "about" meaning within an acceptable error range for the particular value should be assumed.

[0031] In an aspect, provided herein is a method of extracting a metal from a mineral material (e.g., natural mineral material, man-made mineral material, rock, ore, clay and/or other kinds of mineral material). The method may comprise contacting the mineral material with an enzyme having silicase activity under reaction conditions such that the metal contained within the mineral material is solubilized and released. The method may further comprise collecting the released metal, thereby extracting the metal from the mineral material. In some cases, the mineral material comprises or is a natural mineral material or a man-made mineral material. In some cases, the natural mineral

material comprises or is a rock, an ore, or a clay. In some cases, the metal is a metal ion or metal atom. FIG. 1 shows an example workflow according to the embodiments of the present disclosure. [0032] In some cases, the enzyme comprises silicase activity such as the capability to digest or degrade a silicate. In some cases, the mineral material (e.g., rock/ore/clay) comprises a silicate. Silicate may comprise any kind of silicate. In some cases, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, or any combination thereof. In some cases, the mineral material comprises similar or near-similar unit cell geometries. In some cases, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some cases, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some cases, silicate may be an amorphous silicate. In some cases, the amorphous silicate is selected from the group consisting of a tectosilicate, obsidian, coal fly ash, pumice, glass, and any combination thereof.

[0033] In some cases, the enzyme having silicase activity disclosed herein is an engineered enzyme with improved characteristics compared to a wild-type enzyme, such that the modified enzyme exhibits improved ability (e.g., increased reaction rate, increased efficiency, increased ability to degrade silicates, increased specificity for a silicate type) to release metals from mineral materials. The enzyme used in the methods of the present disclosure are generally capable of degrading silicates. In some cases, the enzyme having silicase activity comprises or is a carbonic anhydrase, such as a gamma carbonic anhydrase, or an alpha carbonic anhydrase. In some cases, a wild-type enzyme may be used to perform the methods of the present disclosure. Alternatively or in addition, a modified, mutated, and/or engineered enzyme may be used to perform the methods of the present disclosure. In some cases, an enzyme used in a reaction/process of the present disclosure is a synthetic or semi-synthetic engineered enzyme. In some cases, an enzyme of the present disclosure may be a variant of a carbonic anhydrase, such as a gamma carbonic anhydrase, or alpha carbonic anhydrase. In some cases, an enzyme of the present disclosure may share some similarities (e.g., sequence identity, enzymatic activity) with a carbonic anhydrase. Any combination of wild-type and engineered/modified enzymes may be used.

[0034] In some cases, the enzyme having silicase activity is an engineered enzyme. In some cases, the enzyme having silicase activity is an enzyme having at least one amino acid variation as compared to a wild-type enzyme. In some cases, the enzyme having silicase activity may comprise an amino acid sequence having at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about 99.5%, or greater, sequence identity to an amino acid sequence of a wild-type silicase enzyme. In some cases, the enzyme having silicase activity comprises an amino acid sequence having at most about 95%, at most about 90%, at most about 80%, at most about 70%, at most about 60%, at most about 50%, at most about 40%, at most about 30%, at most about 20%, or at most about 10% sequence identity to an amino acid sequence of a wild-type silicase enzyme. In some cases, the enzyme having silicase activity comprises an amino acid sequence having about 95%, about 90%, about 85%, about 80%, about 75%, about 70%, about 65%, about 60%, about 55%, about 50%, about 45%, about 40%, about 35, about 30%, about 25%, about 20%, about 15%, or about 10% sequence identity to an amino acid sequence of a wild-type silicase enzyme.

[0035] In some cases, the enzyme having silicase activity is derived from an organism selected from the group consisting of: *Methanosarcina thermophila*, *Bacillus* lichenmformis CG-B52, Pelobacter carbinolicus, Syntrophus aciditrophicus, *Methanosarcina barkeri*, *Methanosarcina*

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mazei, Bacillus halodurans, Alkalihalobacillus clausii (strain KSM-K16) (Bacillus clausii),
Methanosarcina acetivorans, Kofleriaceae bacterium SLC26A/SulP, Thermodesulfitimonas
autotrophica, Fischerella thermalis/Mastigocladus laminosus, Thermosynechococcus vestitus BP-
1/(Thermosynechococcus elongatus BP-1) carboxysome assembly protein CcmM, Methanothrix
thermoacetophila, Thermosyntropha lipolytica, isoleucine patch superfamily, Desulfofundulus
thermobenzoicus, Archaeoglobus veneficus, Suberites domuncula, and any combination thereof.
[0036] In some cases, the enzyme having silicase activity has a sequence identity of at least about
10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about
60%, at least about 70%, at least about 80%, at least about 90% or more with a Methanosarcina
thermophila gamma carbonic anhydrase, Bacillus lichenmformis CG-B52 gamma carbonic
anhydrase, Pelobacter carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma
carbonic anhydrase, Methanosarcina barkeri gamma carbonic anhydrase, Methanosarcina mazei
carbonic anhydrase, Bacillus halodurans alpha carbonic anhydrase, Alkalihalobacillus clausii
(strain KSM-K16) (Bacillus clausii) alpha carbonic anhydrase, Methanosarcina acetivorans
carbonate dehydratase, Kofleriaceae bacterium SLC26A/SulP transporter domain-containing
protein, Thermodesulfitimonas autotrophica carbonic anhydrase/acetyltransferase-like protein
(Isoleucine patch superfamily), Fischerella thermalis/Mastigocladus laminosus JSC-11
carboxysome assembly protein CcmM, Thermosynechococcus vestitus BP-
1/(Thermosynechococcus elongatus BP-1) carboxysome assembly protein CcmM, Methanothrix
thermoacetophila carbonate dehydratase, Thermosyntropha lipolytica carbonic anhydrase or
acetyltransferase, isoleucine patch superfamily, Desulfofundulus thermobenzoicus transferase,
Archaeoglobus veneficus carbonate dehydratase, Suberites domuncula carbonic anhydrase.
[0037] In some cases, the enzyme having silicase activity has a sequence identity of at most about
80%, at most about 70%, at most about 60%, at most about 50%, at most about 40%, at most about
30%, at most about 20%, at most about 10% or less with a Methanosarcina thermophila gamma
carbonic anhydrase, Bacillus lichenmformis CG-B52 gamma carbonic anhydrase, Pelobacter
carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma carbonic anhydrase,
Methanosarcina barkeri gamma carbonic anhydrase, Methanosarcina mazei carbonic anhydrase,
Bacillus halodurans alpha carbonic anhydrase, Alkalihalobacillus clausii (strain KSM-K16)
(Bacillus clausii) alpha carbonic anhydrase, Methanosarcina acetivorans carbonate dehydratase,
Kofleriaceae bacterium SLC26A/SulP transporter domain-containing protein,
Thermodesulfitimonas autotrophica carbonic anhydrase/acetyltransferase-like protein (Isoleucine
patch superfamily), Fischerella thermalis/Mastigocladus laminosus JSC-11 carboxysome assembly
protein CcmM, Thermosynechococcus vestitus BP-1/(Thermosynechococcus elongatus BP-1)
carboxysome assembly protein CcmM, Methanothrix thermoacetophila carbonate dehydratase,
Thermosyntropha lipolytica carbonic anhydrase or acetyltransferase, isoleucine patch superfamily,
Desulfofundulus thermobenzoicus transferase, Archaeoglobus veneficus carbonate dehydratase,
Suberites domuncula carbonic anhydrase.
[0038] In some embodiments, the enzyme having silicase activity comprises an amino acid
sequence having at least about 50%, at least about 60%, at least about 70%, at least about 80%, at
least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at
least about 84%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at
least about 99%, at least about 99.5%, at least about 99.9%, or more sequence identity sequence
identity to an amino acid sequence of any one of SEQ ID NOS: 1-18 provided in Table 1.
TABLE-US-00001 TABLE
                            1 Library of Carbonic Anhydrase Enzymes Uniprot SEQ ID
                                 number SEQUENCE
Accession NO: Name Enzyme
                                                        1 y Carbonic
                                                                        Anhydrase P40881
MMFNKQIFTILILSLSLALAGSGCI SEGAEDNVAQEITVDEFSNIRENP
VTPWNPEPSAPVIDPTAYIDPQAS VIGEVTIGANVMVSPMASIRSDE
GMPIFVGDRSNVQDGVVLHALE TINEEGEPIEDNIVEVDGKEYAVY
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IGNNVSLAHQSQVHGPAAVGDD TFIGMQAFVFKSKVGNNCVLEPR

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SAAIGVTIPDGRYIPAGMVVTSQ AEADKLPEVTDDYAYSHTNEAV VYVNVHLAEGYKETS
 2 y Carbonic Anhydrase T5H8M4 MKLSSKLILGLTVSSLAGKFLEKL
LIQDNVSPNITASFNQEADIPDIDA SSYIHHFASVIGSVVIGRNVFIGPF
SSIRGDVGLKIFISHDCNIQDGVV LHGLKNYEYNSPVTEHSVFKDRE
SYSIYIGEKVSLAPQCQIYGPVRID KNVFVGMQSLVFDAYIQEDTVIE
PGAKIIGVTIPPKRFVSAGRVISNQ EDANRLPEITDSYPYHDLNSKMT
SVNLELAKGYKKEERQWKL 3 y Carbonic Anhydrase Q3A3H4
MIEKNVVTDFCSEASEPVIDASTY VHPLAAVIGNVILGKNIMVSPTA
VVRGDEGQPLHVGDDSNIQDGV VIHALETEMNGKPVAKNLYQVD
GRSYGAYVGCRVSLAHQVQIHG PAVVLDDTFVGMKSLVFKSFVG
KGCVIEPGSIVMGVTVADGRYVP AGSVIRTQEDADALPEIGADYPFR
AMNPGVVHVNTALAKGYMVKQ GN 4 y Carbonic Anhydrase
MIGKNVLTDFSARASEPVIGSFTF VHPLAAVIGNVILGDNIMVSPGA
SIRGDEGQPLYVGSDSNVQDGVV IHALETELDGKPVEKNLVEVDGK
KYAVYVGNRVSLAHQVQVHGP AVIRDDTFVGMKSLVFKSYVGSN
CVIEPGVLLMGVTVADGRYVPA GSVVKTQEQADALPVITDDYPM
KEMNKGVLHVNKALARGYLAA GS 5 y Carbonic Anhydrase Q2LUP7
MRFNKQTFTILILSLSLALLGSGCI SEGEGAEGNVTQGITESEFSNIRE
NPVTPWNPVPVAPVIDPTAFIDPQ ASVIGNVTIGASVMVSPMASIRSD
EGMPIFVGDRSNVQDGVVLHAL ETIDEEGEPVENNIVEVGGKKYA
VYIGENVSLAHQAQVHGPASVG NDTFIGMQAFVFKSKIGNNCVLE
PTSAAIGVTVPDGRYIPAGMVVT SQAEADNLSEITDDYAYKHTNEA
VVYVNVHLAEGYNKA 6 Carbonic Anhydrase Q467M8
MALLLSLAITLAGSGCVSQGEGA EEGENIEAEEVEANVEESNIRANP
VTPWNPEPTEPVIDPTAYIHPQAS VIGDVTIGASVMVSPMASVRSDE
GMPIFVGDECNIQDGVILHALET VNEEGEPVEENQVEVDGKKYAV
YIGERVSLAHQAQVHGPSLVGND TFIGMQTFVFKAKIGNNCVLEPTS
AAIGVTVPDGRYIPAGTVVTSQD EADKLPEVTDDYAYKHTNEAVV YVNTNLAEGYNA
7 α Carbonic Q8PSJ1 MKKYLWGKTCLVVSLSVMVTA Anhydrase
CSSAPSTEPVDEPSETHEETSGGA HEVHWSYTGDTGPEHWAELDSE
YGACAQGEEQSPINLDKTEAIDT DTEIHVHYEPSSFTIKNNGHTIQA
ETTSDKNTIEIDGKEYTLVQFHFH IPSEHEMEGKNLDMELHFVHKNE
NDELAVLGVLMKAGEENEELAQ LWSKLPAEETEENISLDESIDLNV
LLPESKEGFHYNGSLTTPPCSEGV KWTVLSEPITVSQEQIDAFAEIFP
DNHRPVQPWNDRDVYDVITE 8 α Carbonic A0A0N9WRG3
MKRSHLFTSITLASVVTLATAPA Anhydrase ASAASFLSPLQALKASWSYEGET
GPEFWGDLDEAFAACSNGKEQSP INLFYDREQTSKWNWAFSYSEAA
FSVENNGHTIQANVENEDAGGLE INGEAYQLIQFHFHTPSEHTIEETS
FPMELHLVHANHAGDLAVLGVL MEMGNDHEGIEAVWEVMPEEEG
TAAYSISLDPNLFLPESVTAYQYD GSLTTPPCSEGVKWTVLNDTISIS
ETQLDAFRDIYPQNYRPVQELGD REIGFHYH 9 Carbonate Q5WD44
MKINRIFLALLFSLALTLAGSGCV dehydratase SQGEGAEDGESADTEVESEVSNI
RANPVTPWNPEPTEPVIDSTAYIH PQAAVIGDVTIGASVMVSPMASV
RSDEGTPIFVGDETNIQDGVVLH ALETVNEEGEPVESNLVEVDGEK
YAVYVGERVSLAHQSQIHGPAY VGNDTFIGMQALVFKANVGDNC
VLEPKSGAIGVTIPDGRYIPAGTV VTSQAEADELPEVTDDYGYKHT
NEAVVYVNVNLAAGYNA 10 Kofleriaceae Q8TMW3 MRTNRVRTAGASKWSGVSDIRT
bacterium Carbonic TLRERWSEIAAQGLSYHDVLAGL Anhydrase
TVATVAIPLNVALAISAGLPPSAG LLAGAVGGLFAAAFGGSNFQVS
GPAAALNVMVFGVVAKFGLGGA AAAALVCGIVGIALGVSGLGKYS
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NLMPKLVLAGFTTGVGLKLLDQ QIPILLGSDLALWHMLSNFWAME
WLREVEWFSVVCGLLVAWITVG LAHLKSFPSALLGIVLATLIAYEL
DWNVARVGEVDLSDLALALPSIA DGTSWFALIAVALPLAVLSSVES
LISAKAVDAMANGKSGYSANTE LFGQGVGSIASALVGGMPLAGV
VVRSSVNQQSGARTRLAAMCHA VFLGIVAYFFGGLLGVIPVAALA
GLLVVIATRLMKLSYFFSALREN KLHALAFLAAAIGTLLGYLISGLA
LGCALVYIAHKLAHRPVKDAPVL RPSPTIRAVISQAGERAQDHTPSI
DEQAKWSRHVRTRPKIHPTAYV HPTASVIGWVELGREVNIAADTS
VRADEGAPFYVGDRSNVQDGVV IHALKDKWVMVDGRRWAVWIG
SDVSLAHQALVHGPSMIGSRSFIG FKAIVHDSVVGEGCFIGLGAVVV
GVEIPAGKRVPNGWIVDSPEKVR ELPDVEHAHAHFNEDVVQVNRG
LVVAYSRHVPTEELPQRTPSDSPL FHLKPL 11 Thermodesulfitimonas A0A7Y6PMB4
MRLPKMLTVVAVGATLCFTAGC autotrophica ASTQTTATKEPAKPANIRPNVVT Carbonic
Anhydrase TFNPTTETPVIAKDAYIDPLASVI GNVEIGSKVYVAPFASVRGDEGQ
PIYVGEGSNVQDGVVLHALETED NGKPVEKNLVEYGGKKYAVYIG
KHVSLAHQAQVHGPALVDDGTF VGMQALVFKAQVGKNCVIEPGA
KLLNGVKVPDGRYVPAGTVVTT QAQADKLPVITDAYPLKNLNKG
VLHVNEQLAEGYLKAQEGATGE TKSH 12 Fischerella A0A3N5BJ34
MAVRSIAEAAPPTPWSRNLAEPTI thermalis/ HPSAFLHSFSNIIGDVRIGANVIIA
Mastigocladus PGTSVRADEGTPFYIGENTNLQD laminosus JSC-11
GVVVHGLEKGRVIGDDRQEYSV Carbonic Anhydrase WIGKNNCITHMALIHGPCYIGDD
CFIGFRSTVFNARVGAGCIVMMH ALIQDVEIPPGKYVPSGAIITNQQ
QADRLPDVQADDKEFAHHVVGI NQALRAGYLCAADSKCIRAIRDE
LNNSYTSIEVDVLERSDEVSSNSL GAETVEQVRYLLQQGYHIGTEH
VDQRRFRTGSWTSCKPIEARSLG EAIAALEACLRDHSGEYVRLFGI
DPKGKRRVLENIIQRPDGVVQAS SSLKAPAYSSNNGSYNGNGSSRL
SSETIDQIRQLLAGGYKIGTEHVD ERRFRTGSWQSCKPIESSSPGDVV
AALEDCMDNHQGEYVRLIGIDPK AKRRVLESIIQRPNGPVSTPSSKST
ATTTSYAASGTTATATSSKLSSEA IEQLQQLLAGGFKISAEHVDGRR
FRTGSWASCGQIQANSIREAIAAL EGYMNEYQGEYVRLIGIDPKVKR RVLELIVQRP 13
Thermosynechococcus G6FUV4 MLRKNPRTSWNSQESMPSVATT vestitus BP-
AYVDETAVVIGDVRIGERVYVGP 1/(Thermosynechococcus
CASIRADEATPIVISEECNVQDGA elongatus BP-1 IFHGLKGSSIKLGKKVSVAHGAV
Carbonic Anhydrase VHGPMTIGDESFIGFNAVVHAST VGERCFIGHRALVMGVKLKDGS
FVPHGSVIDTQDKADALGPVPDS LKGFNAEVVEVNCEFAKGYRSL R 14 Methanothrix
Q8DKB5 MSENLRLNPQGDKPVIDPSSYVD thermoacetophila
PTAVIIGPVTIGKNCYIGPHTVIRA Carbonic Anhydrase DEVDEKTGKVAPVIIGDNVNLQD
GVIIHALAGTSVEVGSNTSLAHG CVVHGPCKIEAGCFIGFRAVVFK
TVIGSGSMVKHGAIVEGVNIPSG KLVPTGEIITSEDHLVKLKEVGQA
EKEFMQEVVHVNMELAHGYKK 15 Thermosyntropha A0B700
MSENLRLNPQGDKPVIDPSSYVD lipolytica Carbonic PTAVIIGPVTIGKNCYIGPHTVIRA
Anhydrase DEVDEKTGKVAPVIIGDNVNLQD GVIIHALAGTSVEVGSNTSLAHG
CVVHGPCKIEAGCFIGFRAVVFK TVIGSGSMVKHGAIVEGVNIPSG
KLVPTGEIITSEDHLVKLKEVGQA EKEFMQEVVHVNMELAHGYKK 16 Desulfofundulus
A0A1M5PQH8 MSENLRLNPQGDKPVIDPSSYVD thermobenzoicus
PTAVIIGPVTIGKNCYIGPHTVIRA Carbonic Anhydrase DEVDEKTGKVAPVIIGDNVNLQD
GVIIHALAGTSVEVGSNTSLAHG CVVHGPCKIEAGCFIGFRAVVFK
TVIGSGSMVKHGAIVEGVNIPSG KLVPTGEIITSEDHLVKLKEVGQA
EKEFMQEVVHVNMELAHGYKK 17 Archaeoglobus A0A6N7IXF4
MLQKSPAVSWKPAGYPRISSLAF veneficus Carbonic VHPTAVLIGEVVIHDGAIIFPLAII
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VIGDETFVGFRAMVINSRIGRGCF IDHGALIEGVEIPDGKYIPGLTRV
SSQEQVSRLAGITEQQKDFAAEV LAVNGELKEAMQVIITSRDDAYP GQ 18 y Carbonic
Anhydrase F2KNT3 MRWAIILTTVLFAALLLGCAAEK GAIEPLETPEEKASNIHANPITEW
NDEQTMPDIDPTAFVHPYATVIG DVHIGKYVCISPHASVRGDEGMP
IYVGDYSNIQDCVVIHALETRDA EGNPIEKNLVVGDDGKKYAVYIA
DHVSLAHQSQVHGPAYVGSGTFI GMQALVFKAKVGKNCVIEPGAK
VIGVTIPDGRYVPAGMAVTNQSV ADNLPEITEDYPFKHTNEAVVHV
NIELAKGYNAMFGGESTEGTEGE GGH
[0039] In some embodiments, an enzyme of the present disclosure is an engineered enzyme. In
some cases, the engineered enzyme may have the sequence of any one of SEQ ID NOS: 19-402
provided in Table 2, or an amino acid sequence having at least about 50%, at least about 60%, at
least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at
least about 92%, at least about 93%, at least about 84%, at least about 95%, at least about 96%, at
least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about 99.9%,
or more sequence identity to any one of SEQ ID NOS: 19-402.
[0040] In some embodiments, an enzyme of the present disclosure is an engineered enzyme.
[0041] In some cases, the engineered enzyme may have the sequence of any one of SEQ ID NOS:
403-464 provided in Table 2, or an amino acid sequence having at least about 50%, at least about
60%, at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about
91%, at least about 92%, at least about 93%, at least about 84%, at least about 95%, at least about
96%, at least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about
99.9%, or more sequence identity to any one of SEQ ID NOS: 403-464.
TABLE-US-00002 TABLE 2 Library of modified or engineered
                                                         enzymes SEQ.ID.
NO Sequence 19 MQEITVTRFENIRPSPVTPWNPEPRYPEIHPTAYIDPAAVVQGD
VKIGANVLVMANAVIRADEGYPIYIGDNSSVQDNVVLHALETR
DADGRDLEENIVRVGDERYAVYVGDNVVLAHNAQVHGPAAV
GDNTFVGMNALVFRSRVGADCVLAPLAAAIGVTVPDGRYVPA
GTVVTTQAAAAALPAVTPDHPFAGLNARVVAVNVALAKGYL ALS 20
MSKIYLAFVCGPEQWHRDFPTANGLRQSPIDIIPSKAVYDPKLR
PLELKYDPSTCLHILNNGHSFQVEFDDSQDKSVLKGGPLDGIY
RLIQFHFHWGSVDGQGSEHTVDKKKYAAELHLVHWNTKYGD
FGKAVQQPDGLAVLGIFLKVGRHKPELQKLVDALSSIKHKDTL
VDFGNFDPSCLMPTCPDYWTYSGSLTTPPLSESVTWIIKKQPVE
VDHDQLEQFRSLLFTSEGEKEKRMVDNFRPLQPLMNRTVRSSF R 21
MKRSLVATIFGYCPEWNDHQSEWGYGETNGPKTWGKHFPEA
NGLLQSPIDIKTEETQHDPNLRPLTLKYDPSTAKEILNNGHSFQ
VTFVDDTDSSTLTDGPITGTYRLKQFHFHWGSSDDKGSEHTVD
GAKYPAELHLVHWNTKYASFGEAASKPDGLAVVGVFLKIGKE
HPGLKKLTDALYMVRFKGTKAQFTNFNPKCLLPTSLDYWTYP
GSLTTPPLSECVTWIVLKEPISVSSAQMEKFRNLLFTSEGEKAC CMVDNYRPPQPLKG 22
MQEITVTNYNNIRPSPVTSWNPTPKLPKIHPTAYIDPAAVVQGD
VTIGENVMVSANASIRSDEGYPIYIGDNSNVQDNVVLHALETV
DADGNVLEENVVTVGDKKYAVYIGKNVSLAHQAQVHGPAAV
GDNTFIGMQAFVFNSVVGKDCVLMPLAAAIGVTIPDGKYIPAG
TVVTTQEEADKLPEVTPDHPFANTNKAVVAVNVELAKGYLAL A 23
MRFFECSCSPFPSQLSSFLTHLLILYTLSSSVEASSRNNYQWSYD
SDVFGGPDFWGLVEKDWWMCRKGRLQSPIDIQPDRLLFDASV
KPVRLDKLPVLSEFVNTGQMVRIRIGYSTKKPSVNITNGPLYG
YRYRVQRIDFHMGRGKENGSEHTINGRRFPMEVQLVAFNTDL
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Anhydrase RADEGFPIIVGENTNIQDGVIIHCL KGGRVEIGRRVSLAHGAVIHGPC

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YPNFTAASKSPHGIAILSVLVDFGAQTNQELTKLTIATASISYKD
QRVQMADFEPWRLLPFTRDIITYEGSLTSPGCHETVTWIILNQPI
FITREHFEEWSHLYHTMEGAEKVPVAPNYRKIQETNNRLVRTN IQHKV 24
MQEITVLEFSNVTKNEVTPWNPKPVTPVIDPTAYIDPTATVIGD
VTIGANCYIAASAVIRADEGKPIVIGDRSNVQDGVVLHALESV
DDGGKVREDNVVIHGDNWYAVYIGENVSLAHQSQVHGPAYV
GDDSFVGMKSLVFKSIVGSNCVIEPEAAAIGVTIPDGKYIPAGT
VVTTQAEADKLPEVTPDYAFYTQVAAVVTVNVNLCRAYRNL S 25
MQEITVAEYSNITKNEVTPWNPKPSTPVIDPTSYVDPNATVIGD
VTIGKNCYIAASAVIRADEGKPIVIGDRSNVQDGVVLHALESV
DDGGMIIGDNVVVEGDKYYAVYIGNNVKLAHQSQVHGPAMV
GDDSFVGMQSFVFNSIVGSNCVIEPEAAAIGVTVPDNKYIPAGT
VVTTQAEADKLPEVTPDDAAFTKNAAVVNVNVGLAKAYREK A 26
MQEITVTNYNNIRPSPVTSWNPEPKLPEIHPTAYIDPAAVVQGD
VTIGENVLVMANAVIRADEGYPIVIGDNSAVQDNVVLHALETV
DENGNRIEENIVKVGDEEYAVYIGKNVVLAHNAQVHGPAIVG
DNTFVGMNALVFRSRVGKNCVLEHNAAAIGVTVPDGKYIPAG
TVVTTQEEADKLPEVTPDHPHYKLNERVVKVNVELAKGYLAL K 27
MQEITVTRYENIQPSPVTPWNPTPKRPQIHPTAYVHPLAYVQG
DVTIGANVMISPNASIRSDEGYPIKIGDNSNVQDNVVLHALETV
DADGKRIEENIVKVGDEEYAVYIGDNVSLAHQAQVHGPAAVG
DNTFIGMQAFVFRSIVGKNCVLEPLAAAIGVTIPDGTYIPAGTV
VTTQEEADKLPKVTPDHPFAKTNAAVVAVNVALAKGYLALA 28
MLRKNPSGHIPQVAETAFIDPTAIICGKVIIEDYVFIGPYAVIRA
DEVNEQGDMEAIVIKRDTNIQDGVVIHSKAGAAVTIGERSSIAH
RSIIHGPCWVGDDVFIGFNSVVFNAKIGKGCVIRHNSVVDGLD
LPENFHVPPMTNIGPGFDLESISKVPPEYSAFSESVVSANHELV QGYRRIANEL 29
MGRSCLTLSRYQAKVSANFLKNRVMASWGYKTDNGPSQWHI
GYPVAKTGTRQSPVNIVPSTVTRDDLLKALKYEYTPSMIKMIN
TGSSWRMDFSPEGSNLSGGPLGDDYKVLQMHAHWGDKAGR
GSEHTMDGKMFDAELHIVHYNSKYGEPAIALDKPDGLAVLGM
FIKTGWRSHPEFDKLCDNLKLIEMKGESLQLQEYLNPANCLPN
NKTFVTYPGSLTTPPLFESVTWIVFLEPIEMSSKQLDSMRALKI
GDTADCGCMVNNYRPPCALGNRKIRVKV 30
MSLVPIERETARRGRPPVAPRALGALLALASAVAATPAIAWQS
GIAVPDPNAMPQWRYTGERGPEHWSELDPSYGACAHTDTQSP
VALTESMAVAVACEPLRFRYRSGPLYVINDGRALRLGYDRGS
HLLVEGLSYELVELRFHAPAEHVINGSRADAELQLIHANNRGD
IAVVAVALMPGPRANSMLQRLLKHAPRLSGESFYGRNVGVNP
LFLLPGRKDYFAYRGSVTRPPCTEGVRWYVLRTPLEVADADL
QRLVGFMEPNARPLQPLGGRRVTKACGP 31
MQEITVTRYENIRPSPVTPWNPEPKLPKIHPTAYIDPAAVVQGD
VTIGANVMVSANASIRSDEGYPIYIGDNSNVQDNVVLHALETV
DENGNVIEENVVTVGDKKYAVAIGDNVSLAHQAQVHGPAIVG
DNTFIGMQAFVFRSRVGKNCVLAPLAAAIGVTVPDGTYIPAGK
VVTTQEEAAKLPKVTPDHPFANTNAAVVKVNVALAKGYLAL A 32
MQEITVHHYSNVTKNEVTPTNPKPVTPVIDPTSYVDPNATVIG
DVTIGKNVLIAANAVIRADEGAPIVIGDRSTVQDGVVLHALES
VDDAGKVREDNVVIEGDEEYAVYIGKDVSLAHQSQVHGPAR
VGDHSFVGMKSLVFKSIVGSNCVLEPEAAAIGVTVPDGKYIPA
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GTVVTTQEEAAKLPEITPDYPLYHANQVVVNVNVLLCQAYKA LS 33
MAKTSFFPVVLSFIFILSYTMCINANATGKHEVDDEEPFSYLLG
TAEGPYKWGTLKPDWEICNTGLFQSPINFRNKTVKVTKHIPHF
TPNYKIASATIMNRGHDIKLQWEGDAGSITLNGTVYKLIQCHW
HTPSEHKVDGQSLAMEAHLIHQSVNGKLIAVIGILFNIGPPDPFL
NELIHHAKKVDHKGKKVGLVDPNKLGVKAEPFYRYIGSLTIPP
CTEGIVWNVLHQPRTVSMDQMMALRNAVNDGFQANARPAQ GLRRRPVYLVM 34
MQEITVTTYNNIRPSPVTSWNPEPRLPKIHPTAYIDPAAVVTGD
VTIGANVLVMANAVIRADEGYPIVIGDNSSVQDNVVLHALETV
DADGNVLEENVVLVGDERYAVYVGDNVVIAHNAQVHGPAIV
GDNTFVGMNALVFRSRVGANCVLAPLAAAIGVTIPDGTYIPAG
KVVTTQEEAAKLPRVTPDHPFADLVARVVKVNVELAKGYLAL S 35
MQEITVTDYSNITKNEVTSTNPKPTTPVIDPTSYVDPNATVTGD
VTIGKNVMISDSASIRSDEGRPIVIGDRSNVQDGVVLHALESVD
DDGEILEDNVVEVGDENYAVYVGKNVSLAHQSQVHGPAAVG
DDSFIGMQAFVFKSKVGSNCVIEPDAAAIGVTVPDGKYIPAGT
VVTTQEEAAKLPEITPDYEYSDTVEAVVEVNVALREAYKEKS 36
MLAFVALVSLIFLGVQAQHGADWTYSEGMLDETHWPEEYPD
CGGQRQSPIDLQRRKVRFNPDLQPLELTGYGDSQGSPFLMTNN
GHTVQITLPPTMQLTAPDGAVYKATQMHYHWGGASYELSGS
EHTIDGIRRVIEMHLVHYNAKYESYDVAKDKPDGLAVMAAFV
EIEEYAENTHYSSLISHLANIRYPGQTTYLTDFDILDMLPGDMY
HYYTYNGSLTTPPCTQNVRWFVMSDSVKISKAQVIKLENSVM
NHQNQTLHNGYRKTQPLHSRVVEANFPYFPNTMPGEGSGLRA
KDPAREFGSRRHCYAWRGWQPAAAAALEGHGEPRRRWRPLE EASTPPP 37
MQEITVTRYNNIRPSPVTPWNPEPKLPEIHPTAYIDPAAVVQGD
VTIGANVMVSANASIRSDEGYPIYIGDNSNVQDNVVLHALETV
DADGKRIEENVVKVGDKDYAVYVGDNVSLAHQAQVHGPAA
VGDNTFIGMQSFVFRSIVGKNCVLEPLAAAIGVTVPDNTYIPAG
KVVTTQEEADKLPKVTPDHPFANTNAAVVKVNVALAKGYLA LA 38
MKNRRIKPEIMKTKFLFAILTLFFFSGCQFFDKNKSTEIESKPSS
HEKWSYTGESGPEHWAELEDQAVCDGQHQSPVNISDIDIKPGK
LIQESLDLSYQEVTTIKSITNNGHTIQYNFDANSNLVSLHDKQY
KLKQFHFHSPSEHTINGTHSPLEIHLVHHSEATNSYIVIAILVQQ
GEPDDAFDFLEKYLPINVGETKEINSKYYFGSTFPEMYGKDTL
NIYTYEGSLTTPPCTESVLWVVIKDPAYASSSQIVMLQKLMPK
DNYREVQSLNGRLIYNEIIEDDISVLNH 39
MTKLSFAVIGPENWHRYCDQAQGDQQSPINIQTRDVKHDPTL
RPLTLRYDPSTAREIVNNGHSFNVEFEDSTDRSVLRGGPLTDRY
RLTQFHFHWGSSDDHGSEHTVDGVKYAAELHLVHWNTKYGD
FGEAASKPDGLAVVGVFLKVGRHNPRLQKILDALHAIKTKGK
RASFTNFDPSVLLPGCLDYWTYSGSLTTPPLSESVTWIVLREPIS VSPSQMAKFRSLLFTS 40
MQEITVTTYNNIQPSPVTPWNPEPKLPKIHPTAYIHPKAVVQGD
VTIGKNVMVSANASIRSDEGYPIVIGDNSNVQDNVVLHALETV
DENGNEIEENIVTVGDKKYAVYIGKNVSLAHQAQVHGPAIVG
DNTFIGMQAFVFNSNVGSNCYLAPLAAAIGVTVPDGTYIPAGK
VVTTQEEAAKLPKITPDHPFYNTNAAVVKVNVALAKGYLALS 41
MQEITVDEFSNITKNEVTPFNPKPTIPVIDPTAYVDPNATVIGDV
TIGKNCYIAPFASIRADEGKPIVIGDNSNVQDGVVLHALESIDD
GGKLIEENVVVEGEKRYAVYIGKNVSLAHQSQVHGPARVGDD
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SFVGMNSLVFNSKVGSNCVIEPFAAAIGVTVPDGKYVPAGTVV
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MQEITVLEYSNVTKNEVTSQNPKPVTPVIDPTSYVDPNATVVG
DVTIGENCLVWPTAVIRADEGRPIVIGNESSVQDGVVLHALES
VDDGGELVEDNVVVVGDKNYAVYVGKNVSLAHQAQVHGPA
RVGDDSFVGMKSLVFKSDVGSNCVIEPFAAAIGVTVPDGKYIP
AGTVVTTQEEAAQLPEVTPDHAEYTTQATVVTVNVELNEAYR NQR 43
MTEKLWGYDSHNGPARWFQICVPAQGKRQSPIDIQPDKAVLD
STLKPLELKYDPSTARRIVNVGHSFHVEFEDSTDKSVLQGGPLT
GSYRLRQFHFHWGKKDDVGSEHVLDGVKYSAELHVVHWNA
DKYSSFVEAAHEPDGLVVLGVFLQIGDQHPGLQRLTDALYAV
RFKGTKAQFACFNPKCLLPTSRHYWTYPGSLTTPPLSESVTWI
VLREPISVSERQMEKFRSLLFTSEDDERIHMVNNFRPLQPLMNR TVRSSF 44
MYHNALFLTPITVFYVAAHKFGYDAEDGPSTWRGVCQTGKR
QSPVDIRAFEIEIAPLDPLQFLNYDLTGHIHLANNGHTVVGSGF
ERWGEKRPYISGGGLNGTYQLSQFHFHWSQQNDTGSEHTIASL
HYPGELHLVHIKKEPSPDEVNTIAVVAAFIKLDDHAGSLHNLK
PYVHNIRMPNTELVVPGFSVSSLLPEHRENFYRYEGSLTTPGCD
EVVVWTLMADPIAVTPSQMGAFHQVHFASGKTGHNWRPTQP
LNGRKILFRPSITLRTFKSGGAMLKPVFQPFISIWLYGIYHIISVF 45
MQEITVTKYNNIRPSPVTPWNPEPKLPEIHPTAYIDPAAVVRGD
VKIGENVLVMANAVIRADEGYPIYIGNNSSVQDNVVLHALETV
DENGNRIEENIVLVGDKEYAVYIGDNVVIAHNAQVHGPAAVG
DNTFIGMNSLVFRSRVGSNCVLAPLAAAIGVTVPDGTYIPAGK
VVTTQEEAAKLPKITPDHPFANLNDRVVKVNVALAKGYLAQA 46
MKMFPLDCLILPCCYFFFISTPHFANADVHIADWDHDHHHTHP
DNWEGMCKEGQRQSPIDIITNETTKEKWGQPFIFHGYERKLSM
NVKNNRHSMVVEFDNDKKYEDIWIRGGGLGESKFRFAQLHFH
WGSTNDQGSEHTIDGKASPMEMHIVHWNLDVGKDVKEATEK
DAYNSLEVLGVLFKLGKFNKDYDAIFNAARKVEKENTNATLE
KDVRLRDLLPEDTNAFYRYVGSLTTPPCNQIVMWTIFKDPIEIS
QEQLDIMRKGSYRLEGENDVRYIANNYRSTQTLYERDVLDIDT
HIVHLACNSKGSTRYHFEEGSEGFVHNTGNSLNSPIVTCMLFY LSIFVISMRLLH 47
MQEITVTRYENIRPSPVTPWNPEPRRPVIHPTAYVDPLAYVQG
DVTIGANVMVSANASIRSDEGYPIVIGDNSNVQDNVVLHALET
RDADGRVLEENVVVVGDERYAVYVGDNVSLAHQAQVHGPA
AVGDNTFIGMQAFVFRSRVGKNCVLEPLAAAIGVTVPDGTYV
PAGKVVTTQEEAAKLPKVTPDHPFATTNAAVVAVNVALAKG YLALA 48
MQEITVLEFSNITKNEVTSFNPEPVTPVIDPTAYIDPNATVIGDV
TIGANVLIWPTAVIRADEGKPIVIGDRSNVQDGVVLHALESVD
DGGKVREDNVVIEGDEEYAVYIGKNVTLAHQSQVHGPARVG
DDSFVGMKSLVFNSDVGENCVIEPFAAAIGVTVPDGKYIPAGT
VVTTQAEAATLPEVTPDYAFYTQVAAVVSVNVGLCQAYKNE A 49
MQEITVDEFSNVTKNEVTPWNPKPTTPVIDPTSYIDPEATVIGD
VTIGKNCYIAPFAVIRADEGSPIVIGDDSTIQDGVVLHALESVD
DGGKLIEDNVVLEGDQYYAVYIGRNVVLAHQSQVHGPAWVG
DDSFVGMKSLVFKSTVGSNCVLEPNAAAIGVTVPDGKYIPAGQ
VVTTQAEADNLPEVTADDAYYTKVAAVVKVNVALCEAYREQ S 50
MQEITVTKYNNIRPSPVTPWNPEPKLPEIHPTAYIDPAAVVQGD
VTIGANVMVSANASIRSDEGYPIKIGDNSNVQDNVVLHALETV
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DADGKELTENVVTVGDEKYAVYVGDNVSLAHQAQVHGPAA
VGDNTFIGMQAFVFRSTVGKNCVLAPLAAAIGVTVPDGRYIPA
GLVVTTQEEADKLPKVTPDHPFYNTNAAVVAVNVALAKGYL AQA 51
MSRPVALTIFGYEDKNQWHCCYPSAQGNRQSPINIDIKKTVYD
PKLKPLELSYDPATAKGILNNGHSFNVEFEDSQDKSVLKGGPL
TGTYRLIQFHFHWGATDDKGSEHTVDGVKYPSELHLVHWNA
VKYSSFAEAASKPDGLAVLGVFLKVGDHNAALQKLTDALYM
VRFKGTKAQFTGFNPKCLLPASLDYWTYSGSLTTPPLLESVTW
IVLKEPISVSSEQMAKFRSLLFTSEGEAECCMVDNYRPPQPLKG R 52
MQEITVTTYTNIRKSPVTSWNPTPKYPKIHPTAYIDPAAVVQGD
VTIGENVMVSANASIRSDEGYPIYIGDNSNVQDNVVLHALETV
DANGNVIEENVVTVGDKKYAVYVGNNVSLAHQAQVHGPAA
VGDNTFIGMQAFVFNSRVGKNCVLEPLAAAIGVTVPDGTYIPA
GEVVTTQEAADKLPKVTPDHPFANTNAAVVKVNIELAKGYLA QA 53
MQEITVTVYTNIQPSPVTSWNPTPKLPKIDETAYVHPQAVVQG
DVTIGKNVMISANASIRSDEGYPIVIGDNSNVQDNVVLHALET
VDADGKEIEENVVTVGDKKYAVYIGDNVSLAHQAQVHGPAA
VGDNTFIGMQAFVFRSNVGKDCVLEPLAAAIGVTVPDGTYIPA
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LTPDYAMYHTNETVVAVNLALAAAAKAAATAYIAPGAEVIGE VEIGANVMVSPMASIRSDEGMPIYLGDNTNVQDGVTLHGLEV **EDEEGEEDESVYVEVNGKKYRVYIGNNVSLAHQAQVHGPAY** VGDDTFIGMGATVFKSRIGNGCVLEPGATVIGVTIPDGRYVPA GKTVTTQAEADALPVLTPDYAMYHTNETVVAVNLALAAAAK AAA 417 TAYIAPTAEVIGDVIIGDNVMISPNASIRSDEGMPIYIGENVNVQ DGVTITADRTKDEAGNDIPENWVTVNGKKYAVYLGKNVVLA HNATVNGRTVLGENVLVQENATLTASTLGENVIVQENATLTG VTVAEGKVVEAGKTITTQAEADKLKDLTKDHPLYNKNKEVV AKNLAILEEKKKLETAYIAPTAEVIGDVIIGDNVMISPNASIRSD EGMPIYIGENVNVQDGVTITADRTKDEAGNDIPENWVTVNGK KYAVYLGKNVVLAHNATVNGRTVLGENVLVQENATLTASTL GENVIVQENATLTGVTVAEGKVVEAGKTITTQAEADKLKDLT KDHPLYNKNKEVVAKNLAILEEKKKLETAYIAPTAEVIGDVIIG DNVMISPNASIRSDEGMPIYIGENVNVQDGVTITADRTKDEAG NDIPENWVTVNGKKYAVYLGKNVVLAHNATVNGRTVLGENV LVQENATLTASTLGENVIVQENATLTGVTVAEGKVVEAGKTIT TQAEADKLKDLTKDHPLYNKNKEVVAKNLAILEEKKKLE 418 TAYIAPTATVIGDVEIADNVMISPNASIRSDEGMPIYIGENANLQ DNVVLHALETKDEEGNDIEENWVEVDGKKYAVYIGRRVSLGH QAQIHGPALVGDDTFIGMNAKVFKSRIGNRCVLEPNAQVIGVT IPDGRYVPAGKVVTTQEEADKLPLLTPDYAMYHTNERVNAVN LALAAEARALATAYIAPTATVIGDVEIADNVMISPNASIRSDEG MPIYIGENANLQDNVVLHALETKDEEGNDIEENWVEVDGKKY AVYIGRRVSLGHQAQIHGPALVGDDTFIGMNAKVFKSRIGNRC VLEPNAQVIGVTIPDGRYVPAGKVVTTQEEADKLPLLTPDYAM YHTNERVNAVNLALAAEARALATAYIAPTATVIGDVEIADNV MISPNASIRSDEGMPIYIGENANLQDNVVLHALETKDEEGNDIE ENWVEVDGKKYAVYIGRRVSLGHQAQIHGPALVGDDTFIGMN AKVFKSRIGNRCVLEPNAQVIGVTIPDGRYVPAGKVVTTQEEA DKLPLLTPDYAMYHTNERVNAVNLALAAEARALA 419 TAYIHPTAEVIGDVEIGDNVMISPNASIRADEGMPIVIEENVNV QDGVEITALRSDLPEEEVEKLDLQEVDGKKVRAYFGKGAVLA HGAKILVASTRLKVEPVPGVTVLKQDNAVLRNVLLTEMHGLIL EVNAETGSIVIRESSDPALESKAKTWKVTPEDKAKIAAVIAANA AARQEALAAATAYIHPTAEVIGDVEIGDNVMISPNASIRADEG MPIVIEENVNVQDGVEITALRSDLPEEEVEKLDLQEVDGKKVR AYFGKGAVLAHGAKILVASTRLKVEPVPGVTVLKQDNAVLRN VLLTEMHGLILEVNAETGSIVIRESSDPALESKAKTWKVTPEDK AKIAAVIAANAAARQEALAAATAYIHPTAEVIGDVEIGDNVMI SPNASIRADEGMPIVIEENVNVQDGVEITALRSDLPEEEVEKLD LQEVDGKKVRAYFGKGAVLAHGAKILVASTRLKVEPVPGVTV LKQDNAVLRNVLLTEMHGLILEVNAETGSIVIRESSDPALESKA KTWKVTPEDKAKIAAVIAANAAARQEALAAA 420 TAYIEPNAEVIGDVKIGENVMISPNASIRSDEGMPIVIKENVNV QDGVVINAKLKKNENGEVDESQLNTINGEKVQIYLEKNVQLA HNVTIEDSVVLKENVLLQENVVLKNSTLGEGVVLAENVVIENV TLPENTVVEAGTVIKNQEEVKTLKQLTADSPAIVQLQAVLAKN AALWEELKAAETAYIEPNAEVIGDVKIGENVMISPNASIRSDEG MPIVIKENVNVQDGVVINAKLKKNENGEVDESQLNTINGEKV

QIYLEKNVQLAHNVTIEDSVVLKENVLLQENVVLKNSTLGEGV VLAENVVIENVTLPENTVVEAGTVIKNQEEVKTLKQLTADSPA IVQLQAVLAKNAALWEELKAAETAYIEPNAEVIGDVKIGENV MISPNASIRSDEGMPIVIKENVNVQDGVVINAKLKKNENGEVD ESQLNTINGEKVQIYLEKNVQLAHNVTIEDSVVLKENVLLQEN VVLKNSTLGEGVVLAENVVIENVTLPENTVVEAGTVIKNQEEV KTLKQLTADSPAIVQLQAVLAKNAALWEELKAAE 421 TAYIHPSAEVIGDVTIGDNVMISPNASIRADEGMPIYLGDNANV QDGVTLHGLETKDEEGNIIEENLVEVNGKKYAVYVGDNVSLA HQAQIHGPAIVGDDTFIGMGATVRRSILGDGVLLGEGVQIENA TLPAGLCLGPGRVIRTPEELVDDCTEEQRAELKKKHAEVVAKN LALHEELKAAATAYIHPSAEVIGDVTIGDNVMISPNASIRADEG MPIYLGDNANVQDGVTLHGLETKDEEGNIIEENLVEVNGKKY AVYVGDNVSLAHQAQIHGPAIVGDDTFIGMGATVRRSILGDG VLLGEGVQIENATLPAGLCLGPGRVIRTPEELVDDCTEEQRAEL KKKHAEVVAKNLALHEELKAAATAYIHPSAEVIGDVTIGDNV MISPNASIRADEGMPIYLGDNANVQDGVTLHGLETKDEEGNIIE ENLVEVNGKKYAVYVGDNVSLAHQAQIHGPAIVGDDTFIGMG ATVRRSILGDGVLLGEGVQIENATLPAGLCLGPGRVIRTPEELV DDCTEEQRAELKKKHAEVVAKNLALHEELKAAA 422 TAYIAPNAQVIGEVTIGENVMISPNASIRSDEGMPIYIGENANLQ DNVVLHGLEVYTEEGELIEENLVEVDGKKYVVYIGKNVSLGH QAQIHGPAKVGDDTFIGMNAKVFKSVIGNRCVLEPNATVIGVT IPDGRYVPAGKVVTTQAEADALPVLTPDYALYHTNELVNEVN LALAAEGRAAATAYIAPNAQVIGEVTIGENVMISPNASIRSDEG MPIYIGENANLQDNVVLHGLEVYTEEGELIEENLVEVDGKKYV VYIGKNVSLGHQAQIHGPAKVGDDTFIGMNAKVFKSVIGNRC VLEPNATVIGVTIPDGRYVPAGKVVTTQAEADALPVLTPDYAL YHTNELVNEVNLALAAEGRAAATAYIAPNAQVIGEVTIGENV MISPNASIRSDEGMPIYIGENANLQDNVVLHGLEVYTEEGELIE ENLVEVDGKKYVVYIGKNVSLGHQAQIHGPAKVGDDTFIGMN AKVFKSVIGNRCVLEPNATVIGVTIPDGRYVPAGKVVTTQAEA DALPVLTPDYALYHTNELVNEVNLALAAEGRAAA 423 TAYIHPSAEVIGDVEIADNVMISPNASIRADEGMPIYLGENTNV QDGVSLHALENSSEEGEEDESNWVEVNGKKYRVYIGNNVSLG HQAQVHGPAIVGDDTFIGMGAKVFKSTIGNGCVLEPGVTVIGV TIPDGRYLEAGTVLRSQADIEKAKPIKEDMPTYKKVKEHKEKL KEEREKLKKERTAYIHPSAEVIGDVEIADNVMISPNASIRADEG MPIYLGENTNVQDGVSLHALENSSEEGEEDESNWVEVNGKKY RVYIGNNVSLGHQAQVHGPAIVGDDTFIGMGAKVFKSTIGNG CVLEPGVTVIGVTIPDGRYLEAGTVLRSQADIEKAKPIKEDMPT YKKVKEHKEKLKEEREKLKKERTAYIHPSAEVIGDVEIADNVM ISPNASIRADEGMPIYLGENTNVQDGVSLHALENSSEEGEEDES NWVEVNGKKYRVYIGNNVSLGHQAQVHGPAIVGDDTFIGMG AKVFKSTIGNGCVLEPGVTVIGVTIPDGRYLEAGTVLRSQADIE KAKPIKEDMPTYKKVKEHKEKLKEEREKLKKER 424 TAIIAPGATVIGEVHIADGVMISPNASIRADEGMPIYLGEYTNLQ DNVVLHALETYDEEGNLIEENLVEVNGKKYAVYVGKNVSLG HQAQLHGPTIVGDDTFIGMNAKVIRSTLGEGVVLEENVVVEGQ TLEKGTYLEKGMKLLTPEDLKKAKKIKEEDPVKKKLEAHIKEQ

KAQAKAAQAAATAIIAPGATVIGEVHIADGVMISPNASIRADE **GMPIYLGEYTNLQDNVVLHALETYDEEGNLIEENLVEVNGKK** YAVYVGKNVSLGHQAQLHGPTIVGDDTFIGMNAKVIRSTLGE GVVLEENVVVEGQTLEKGTYLEKGMKLLTPEDLKKAKKIKEE DPVKKKLEAHIKEQKAQAKAAQAAATAIIAPGATVIGEVHIAD GVMISPNASIRADEGMPIYLGEYTNLQDNVVLHALETYDEEGN LIEENLVEVNGKKYAVYVGKNVSLGHQAQLHGPTIVGDDTFIG MNAKVIRSTLGEGVVLEENVVVEGQTLEKGTYLEKGMKLLTP EDLKKAKKIKEEDPVKKKLEAHIKEQKAQAKAAQAAA 425 TAYIHPSAEVIGEVEIGANVMVSPMASIRSDEGMPIKIGDNVNV **QDGVVLHGLETKNEEGEEIEENLVEVDGEKYVVYLGKNVSLA** HQAQVHGPSIVGDDTFIGMGAKVEGSTLGDGVFLGEGATVTG LTIPAGAVVSPGTVLTTPAQLASLKPLTADDPLLKKKKDVVEN NLATAAALKALETAYIHPSAEVIGEVEIGANVMVSPMASIRSD EGMPIKIGDNVNVQDGVVLHGLETKNEEGEEIEENLVEVDGEK YVVYLGKNVSLAHQAQVHGPSIVGDDTFIGMGAKVEGSTLGD GVFLGEGATVTGLTIPAGAVVSPGTVLTTPAQLASLKPLTADD PLLKKKKDVVENNLATAAALKALETAYIHPSAEVIGEVEIGAN VMVSPMASIRSDEGMPIKIGDNVNVQDGVVLHGLETKNEEGE EIEENLVEVDGEKYVVYLGKNVSLAHQAQVHGPSIVGDDTFIG MGAKVEGSTLGDGVFLGEGATVTGLTIPAGAVVSPGTVLTTP AQLASLKPLTADDPLLKKKKDVVENNLATAAALKALE 426 TAYIHPSASVIGDVEIADNVMISPNASIRADEGMPIKIGPNANV QDGVTLHGLETYDEEGNLIEENYVEVNGERYVVYIGDNVSLG HQAQIHGPAKVGDDTFIGMKATVFKSVIGNNCVLEPGATVIGV TIPDGRYVPAGKVVTTQAEADALPVLTPDYALYHTNERVNAV NLKLAEKARLEATAYIHPSASVIGDVEIADNVMISPNASIRADE GMPIKIGPNANVQDGVTLHGLETYDEEGNLIEENYVEVNGERY VVYIGDNVSLGHQAQIHGPAKVGDDTFIGMKATVFKSVIGNN CVLEPGATVIGVTIPDGRYVPAGKVVTTQAEADALPVLTPDYA LYHTNERVNAVNLKLAEKARLEATAYIHPSASVIGDVEIADNV MISPNASIRADEGMPIKIGPNANVQDGVTLHGLETYDEEGNLIE ENYVEVNGERYVVYIGDNVSLGHQAQIHGPAKVGDDTFIGMK ATVFKSVIGNNCVLEPGATVIGVTIPDGRYVPAGKVVTTQAEA DALPVLTPDYALYHTNERVNAVNLKLAEKARLEA 427 TAYIAPSAEVIGDVEIGANVMVSPMASIRADEGMPIYIGDNAN VQDGVVLHALETYDEEGNLIEEAYVEVDGKKYAVYVGDNVS LAHQAQIHGPAKVGEDTFIGMGAKVVGSTLGKGVFLAEGVVV ENATLPEGTILEKGTVVTPSDKELPKAPEELRAKLAAEHKAVV AANIAAAAAKAAATAYIAPSAEVIGDVEIGANVMVSPMASIR ADEGMPIYIGDNANVQDGVVLHALETYDEEGNLIEEAYVEVD GKKYAVYVGDNVSLAHQAQIHGPAKVGEDTFIGMGAKVVGS TLGKGVFLAEGVVVENATLPEGTILEKGTVVTPSDKELPKAPE ELRAKLAAEHKAVVAANIAAAAAAKAAATAYIAPSAEVIGDV EIGANVMVSPMASIRADEGMPIYIGDNANVQDGVVLHALETY DEEGNLIEEAYVEVDGKKYAVYVGDNVSLAHQAQIHGPAKV GEDTFIGMGAKVVGSTLGKGVFLAEGVVVENATLPEGTILEKG TVVTPSDKELPKAPEELRAKLAAEHKAVVAANIAAAAAAKAA A 428 TAYIAPGAEVIGDVEIGANVMVSPMASIRADEGMPIYVGDNAN VQDGVVLHGLETLDEEGNLIEENWVEVDGKKYVVYLGKNVS

LAHQAQIHGPAKVGEDTFIGMQALVFKSTIGNGCVLEPGAAVI GVTVPDGRYVPAGAVVTSQAEADALPKMTPDYAYAHTNETV VAVNNALAAGYKAAATAYIAPGAEVIGDVEIGANVMVSPMA SIRADEGMPIYVGDNANVQDGVVLHGLETLDEEGNLIEENWV EVDGKKYVVYLGKNVSLAHQAQIHGPAKVGEDTFIGMQALV FKSTIGNGCVLEPGAAVIGVTVPDGRYVPAGAVVTSQAEADA LPKMTPDYAYAHTNETVVAVNNALAAGYKAAATAYIAPGAE VIGDVEIGANVMVSPMASIRADEGMPIYVGDNANVQDGVVLH GLETLDEEGNLIEENWVEVDGKKYVVYLGKNVSLAHQAQIHG PAKVGEDTFIGMQALVFKSTIGNGCVLEPGAAVIGVTVPDGRY VPAGAVVTSQAEADALPKMTPDYAYAHTNETVVAVNNALAA GYKAAA 429 TAYIHPSATVIGQVNIGANVMVSPMASIRADEGMPITLEDNVN VQDGVLIQNESLKNESGEIDYSKVHPKNKRIESIVLKKNVSLAH QATVYSNTELSEGVFLQEGVVVKNSVIEGRVVLQRGVTVENV YIGEEVVIAEGTVLKGDEDLKKTTLAPLTPEQVAQIQAVIAQNL AAAAAAKAAATAYIHPSATVIGQVNIGANVMVSPMASIRADE **GMPITLEDNVNVQDGVLIQNESLKNESGEIDYSKVHPKNKRIES** IVLKKNVSLAHQATVYSNTELSEGVFLQEGVVVKNSVIEGRVV LQRGVTVENVYIGEEVVIAEGTVLKGDEDLKKTTLAPLTPEQV AQIQAVIAQNLAAAAAAKAAATAYIHPSATVIGQVNIGANVM VSPMASIRADEGMPITLEDNVNVQDGVLIQNESLKNESGEIDYS KVHPKNKRIESIVLKKNVSLAHQATVYSNTELSEGVFLQEGVV VKNSVIEGRVVLQRGVTVENVYIGEEVVIAEGTVLKGDEDLKK TTLAPLTPEQVAQIQAVIAQNLAAAAAAKAAA 430 TAYIAPGAQVIGDVEIADNVMISPNASIRADEGMPIYIGENANL QDNVQLHGLEVYTEEGELIEENFVEVDGKKYVVYIGRRVSLA HQAQVHGPAKVGDDTFIGMNAKVFKSIVGNRCVLEPNATVIG VTIPDGRYVPAGKTVTTQAEADALPVLTPDYALYHTNELVNA VNLALAAEARAAATAYIAPGAQVIGDVEIADNVMISPNASIRA DEGMPIYIGENANLQDNVQLHGLEVYTEEGELIEENFVEVDGK KYVVYIGRRVSLAHQAQVHGPAKVGDDTFIGMNAKVFKSIVG NRCVLEPNATVIGVTIPDGRYVPAGKTVTTQAEADALPVLTPD YALYHTNELVNAVNLALAAEARAAATAYIAPGAQVIGDVEIA DNVMISPNASIRADEGMPIYIGENANLQDNVQLHGLEVYTEEG ELIEENFVEVDGKKYVVYIGRRVSLAHQAQVHGPAKVGDDTFI GMNAKVFKSIVGNRCVLEPNATVIGVTIPDGRYVPAGKTVTTQ AEADALPVLTPDYALYHTNELVNAVNLALAAEARAAA 431 TAYIHPTARVIGEVTIAAGVMISPGASIRADEGMPIVIGENANV **QDGVSLHGLEVYDEEGNLIEENLVEVNGEKYVVYIGENVSLG** HQAQIHGPALVGDDTFIGMGAKVTRSILGEGVILEEGAQLTNVI VPDGAYVKSGQVFVSTGEPVVLSELKQTPEQKEKLAAQLAAE RAARAAQAAATAYIHPTARVIGEVTIAAGVMISPGASIRADE GMPIVIGENANVQDGVSLHGLEVYDEEGNLIEENLVEVNGEKY VVYIGENVSLGHQAQIHGPALVGDDTFIGMGAKVTRSILGEGV ILEEGAQLTNVIVPDGAYVKSGQVFVSTGEPVVLSELKQTPEQ KEKLAAQLAAERAARAAAQAAATAYIHPTARVIGEVTIAAGV MISPGASIRADEGMPIVIGENANVQDGVSLHGLEVYDEEGNLIE ENLVEVNGEKYVVYIGENVSLGHQAQIHGPALVGDDTFIGMG AKVTRSILGEGVILEEGAQLTNVIVPDGAYVKSGQVFVSTGEP VVLSELKQTPEQKEKLAAQLAAERAARAAAQAAA 432

TAYIHPTAEVIGNVKIGENVMISPNASIRSDEGMPIVIKENANV QDGVVIRADPTKDENGNDIEENWVTVNGEKYAVYLEKNVVL AHNAVVEGRTVLKEGVLVQENAVVRRSTLGEGVILQENAVLE GVTVADGKIVPAGATIRTQAEADTLATLTPDHPLYNLNKVVN AKNLALLKENLAAKTAYIHPTAEVIGNVKIGENVMISPNASIRS DEGMPIVIKENANVQDGVVIRADPTKDENGNDIEENWVTVNG EKYAVYLEKNVVLAHNAVVEGRTVLKEGVLVQENAVVRRST LGEGVILQENAVLEGVTVADGKIVPAGATIRTQAEADTLATLT PDHPLYNLNKVVNAKNLALLKENLAAKTAYIHPTAEVIGNVKI GENVMISPNASIRSDEGMPIVIKENANVQDGVVIRADPTKDEN GNDIEENWYTVNGEKYAVYLEKNVVLAHNAVVEGRTVLKEG VLVQENAVVRRSTLGEGVILQENAVLEGVTVADGKIVPAGATI RTOAEADTLATLTPDHPLYNLNKVVNAKNLALLKENLAAK 433 TAIIAPGATVIGEVEIGDNVMISPNASIRSDEGMPIVLGEGANLQ DNVELHALEVYDEEGNLIEENYVEVNGKKYAVYIGNNVSLGH QAQIHGPAIVGDDTFIGMNAEVFKSIIGNGCVLEPNARVIGVTIP DGRYVKAGTTITDQAEIPSLKQLKDSDPIKAKVEAHKAALKAE RERLLAERTAIIAPGATVIGEVEIGDNVMISPNASIRSDEGMPIV LGEGANLQDNVELHALEVYDEEGNLIEENYVEVNGKKYAVYI GNNVSLGHQAQIHGPAIVGDDTFIGMNAEVFKSIIGNGCVLEP NARVIGVTIPDGRYVKAGTTITDQAEIPSLKQLKDSDPIKAKVE AHKAALKAERERLLAERTAIIAPGATVIGEVEIGDNVMISPNASI RSDEGMPIVLGEGANLQDNVELHALEVYDEEGNLIEENYVEV NGKKYAVYIGNNVSLGHQAQIHGPAIVGDDTFIGMNAEVFKSI IGNGCVLEPNARVIGVTIPDGRYVKAGTTITDQAEIPSLKQLKD SDPIKAKVEAHKAALKAERERLLAER 434 QEITVDEFSNIRENPVTPWNPEPSAPVIDPTAYIDPQASVIGEVTI GANVMVSPMASIRSDEGMPIFVGDRSNVQDGVVLHALETINEE GEPIEDNIVEVDGKEYAVYIGNNVSLAHQSQVHGPAAVGDDT FIGMQAFVFKSKVGNNCVLEPRSAAIGVTIPDGRYIPAGMVVT SQAEADKLPEVTDDYAYSHTNEAVVYVNVHLAEGYKETS 435 GKVYVKKPVFIPARHIPSDKTIIEPEIDEEAVIEEGAIITGGVIIKG RVYIASGATIRSDEGVPIVIEENSSIQDGALVHADETVDEDGNPI EENIVEVNGKPYAVYIGENVVLEHNATVHGPAAVGKNSLIGEG ALVRNSVIGENCVLEEGASAENVTIPAGRYVPAGVTVTTQAAA AALPAVTPDHPLYKRNEELVKENIEKAEKLLAEA 436 GSVLVEPSDIQCSPPNKYHKEPRCPTIAKGAYIEKGALIEGDVII EENVYIESGAIIRSDEGTPIYIGKNSVIQDGALVHADETVDEDG NPIEENIVEVNGKPYAVYIGENVVLEHNAEVHGPAAVGKNSLI GEGALVRNSIIGENCVLEEGASAENVTIPAGRYVPAGKTVTTQ AEAAALPKMTPDHPLYKRNEELVKENLEKVKKANAAA 437 GVVLVEEEGIRPSPATPRYPEPRAPIIHPSAYVADGALITGEVIIE DNVLIAEGAVIRSDEGRPIYIGKNSSVQDGAVIHADETVDAEGK EIEENIVEVNGKKYAVYIGENVVIEHGATVHGPAKIGENSLIGR GALVENSVIGKNCVLEEGASAIGVTIPEGRYIPAGVTVTTQEEA DALPEVTPDHPDYNRVAELVAKNIALAKELNAAR 438 PAPVRGHEAVFEDSLHPVTGKKLVTTIAETAYIEEGATISGAVI LADNVYVESGATIRSDEGIPIYVGENSAIQDGAVLHADETVDA DGNPIPENIVEVNGEPYAVYIGENVVLEHGATVHGPAAVGKNS LIGKNAVVRNSVVGENCVLEEGASAENVTIPAGRYVPAGKKV

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TTQEEADALPEVTPDHPDYKRNEKLVAENNAKVKAYNAAR 439
GVVLTPVSDIRPSAPTPRYKESKAPTIHPSAYIAPGATIVGDVTI
AANVYVEAGATIRSDEGVPIYVGANSAVQDGAVLHADETVDE
NGNPIEENIVEVNGKKYAVYVGENVVLEHGATVHGPAAIGAN
SLVGEGALVANSIVGANCVLEPGASAINVTIPAGRYVPAGVTV
TTQAAADALPAVTPDHPLANRNAELVAKNVAKAKAAVAAR 440
GTVVVATSPIRPSEPTPWRKESRAPTLAPGAYVHPDATVEGAV
ILEEGALVQGGATIRSDEGVPIYVGRNSVIQDGATLHADETVDE
EGNPIPENIVEVDGKPYAVYVGENVVIQHGATIHGPAAVGENS
LIGENALVENSVVGKNCVIQEGGAARNVTIPEGRYIPAGKTVT
TQAEADALPKVTPDHPYYNKNAALVAENLARRAELLAAR 441
MVVLVEEAGLRPSPPTPRHREPRAPTLAEGAWVAPGATIEGEV
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DGKVIEENVVTVNGEPYAVYIGENVVIEHNATIHGPAAVGANS
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MLLVEEEPLIRPSEPTPWRGTRREPTIAEGAYIEPGAVITGDTIIE
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NLIEENVVEVNGKKYAVYVGANVVIEHNAVIHGPAAVGANSL
IGEGAVVRNSIVGANCVLEPGASVENVTVPAGRYVPAGVRVT
TQAAADALPAVTPDHPLANRNAELVAKNVAKVKAANAAK 443
GLKKRKLTPAELRKPDPRYTGPRVSTIGETCLFAPGAVISPGVT
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LIGEGATVRNSTVGSNCVLEPGASAIGVTIPAGRYVPAGKTVTT
QAEADALPAVTPDHPYANRVAELVAKNLAKVKAANAAR 444
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DNVLIEKGATIRSDEGVPIYIGENSSIQDGATLHADETVDEAGN
PIEENIVTVNGKPYAVYIGENVVIEHGATVHGPAAIGRNSLIGE
GATVRNSIVGENCVLEEGASAIGVTIPAGRYIPAGKVVTTQEEA
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TAYIHPSATVIGDVIIGKNAMISPNASIRSDEGMPIVLEENAIVQ
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TAYIHPSATVIGDVEIGENAMISPNASIRSDEGMPIVLEKNVVV
QDNATIEANPVLDENGELIEENVREVNGKKYAVYLEEGVVLQ
KNATIKGGTVVKKNVLVQENATLTNSTLGENVIVQENATLTG
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VTVAEGKVVPEGATITTQAEAEKLAPLTPDHPLYNLGAEIRAK ALALRELKLALE 450
TAYIHPSAKVEGEVEIGANVMVSPMATINSTEGEPIFLGDRVNV
ODGVTITCEPVYNADGELDESKLVEIDGKKYCVYVAENVSLA
HQSTLTSGTVLKKNVFVQEGAVLKNSTLGENVVVRENAVIEG
VTIEEGTVAEEGTVVLTEEDLAKLRPLTPEDPLLEIHKKVVEEN IAKAKALKAAA 451
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QDGVSLHALETLDEEGNLIEENVVEVDGKKYAVYVGDNVSLA
HQAQVHGPAIVGEDTFIGMGAVVRRSVLGKGVILREGATVEG
VTIEEGTVVEENTVLTKQEEVKKLKKLTPEHPYYNLNKKVVE QNIKKVQAARAAA 452
TAVIHPSATVIGDVTIADNVMISPNASIRADEGMPIVLGENVNL
QDNVSLHALETLDEEGNLIEENVVEVNGKKYAVYLGEGVSLA
HQAQVHGPAIVGKDTFIGMNAKVSGSILGEGVILQDNATVEGA
TIAEGKVVPEGAVITTQEEADKLAPLTPDHPYYELVKRTREEN LRLRDLLLELE 453
TAYIHPSAKVIGDVEIGENVMISPNASIRADEGMPIKLEKNVIV
QDNAVIEADRIYDENGELIESAVVTVNGKKYAVYLGENVILQK
NATVRGGTVLGKNVLVQENAVLTNSTLGENVIVQENATVTGA
TLKEGTIVPEGATITTQEEADKLEKLTPDHPLYNLHAALLAAGL ALRDLLLALE 454
TAYIHPSAEVIGDVTIGANVMVSPMASIRADEGMPIVLGNNVN
VQDGVVLHGLEVLNEEGELIEENVVEVDGKKYVVYVGEGVSL
AHQATVVGATVLGKGVFVGEGVVLERSILGEGVIVGENAVIK
GVTIAEGKVVKEGTTVTTQEEADKLEKLTPDHPYYELNARVV AENIAKARLLKLLE 455
TAYIAPGASVIGEVEIGDNVMISPNASLRSDEGMPIKLGDNVNV
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GVTIKEGMYAKEGTVIRTQEDVKSLEMIKDLAKHKAKVQAVI
DANLRIHQEALAAATAYIAPGASVIGEVEIGDNVMISPNASLRS
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GKYVYVEKGVVIEGVTIKEGMYAKEGTVIRTQEDVKSLEMIK
DLAKHKAKVQAVIDANLRIHQEALAAATAYIAPGASVIGEVEI
GDNVMISPNASLRSDEGMPIKLGDNVNVQDGVTLHGLETKDE
EGNLIEENVVEVNGEKYVVYVGKNVVLAHNVTLTSRTVVEDN
VYLEENVTLTRSTLGKYVYVEKGVVIEGVTIKEGMYAKEGTVI
RTQEDVKSLEMIKDLAKHKAKVQAVIDANLRIHQEALAAA 456
TAYIHPTATVIGSVTIADGVMISPYASIRADEGMPIYIGEGANV
QDGVQLHGLETRDEEGNLIEENVVEVNGKKYVVYIGKGVSLG
HQAQVHGPAIVGDDTFIGMGAQVTGAILPEGLVLAEGVRVES
VDLSGYRYLPPGTVIRTQADKERVREDESLAAEVEARRAALA
AERAAAEAARAAATAYIHPTATVIGSVTIADGVMISPYASIRAD
EGMPIYIGEGANVQDGVQLHGLETRDEEGNLIEENVVEVNGK
KYVVYIGKGVSLGHQAQVHGPAIVGDDTFIGMGAQVTGAILP
EGLVLAEGVRVESVDLSGYRYLPPGTVIRTQADKERVREDESL
AAEVEARRAALAAERAAAEAARAAATAYIHPTATVIGSVTIAD
GVMISPYASIRADEGMPIYIGEGANVQDGVQLHGLETRDEEGN
LIEENVVEVNGKKYVVYIGKGVSLGHQAQVHGPAIVGDDTFIG
MGAQVTGAILPEGLVLAEGVRVESVDLSGYRYLPPGTVIRTQA
DKERVREDESLAAEVEARRAALAAERAAAEAARAAA 457
TAFIHPSATVIGDVTIGANVMISPMASIRSDEGMPIYVGANANL
QDQVTLHALEVFDEEGNLIEENWVEVNGEKYAVYLGDNVSLA
HQAQVHGPAIVGEDTFIGMQASVFKSTIGNGCVLEPGAAVIGV
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TIPDGRYVPAGKVVTSQAEADALPKMTPDYAYYHTNEKVVA VNRALAAGYRAAQTAFIHPSATVIGDVTIGANVMISPMASIRS DEGMPIYVGANANLQDQVTLHALEVFDEEGNLIEENWVEVNG EKYAVYLGDNVSLAHQAQVHGPAIVGEDTFIGMQASVFKSTI GNGCVLEPGAAVIGVTIPDGRYVPAGKVVTSQAEADALPKMT PDYAYYHTNEKVVAVNRALAAGYRAAQTAFIHPSATVIGDVT IGANVMISPMASIRSDEGMPIYVGANANLQDQVTLHALEVFDE EGNLIEENWVEVNGEKYAVYLGDNVSLAHQAQVHGPAIVGE DTFIGMQASVFKSTIGNGCVLEPGAAVIGVTIPDGRYVPAGKV VTSQAEADALPKMTPDYAYYHTNEKVVAVNRALAAGYRAA Q 458 TAYIHPSATVIGDVTIGANVMVSPMASIRSDEGMPIYLGDNVN VQDGVSLHGLETVDEEGNVIEENLVEVDGKKYVVYLGDNVSL AHQAVVEGGTVLGENVFLQEGVRVRRSTLGEGVILREGATVE GVTIAPGKVVAAGQTVTTQAAADALPTLTASDPLYSEHATVV AANIAAAAAKAAATAYIHPSATVIGDVTIGANVMVSPMASIR SDEGMPIYLGDNVNVQDGVSLHGLETVDEEGNVIEENLVEVD GKKYVVYLGDNVSLAHQAVVEGGTVLGENVFLQEGVRVRRS TLGEGVILREGATVEGVTIAPGKVVAAGQTVTTQAAADALPTL TASDPLYSEHATVVAANIAAAAAAKAAATAYIHPSATVIGDVT IGANVMVSPMASIRSDEGMPIYLGDNVNVQDGVSLHGLETVD EEGNVIEENLVEVDGKKYVVYLGDNVSLAHQAVVEGGTVLG ENVFLQEGVRVRRSTLGEGVILREGATVEGVTIAPGKVVAAGQ TVTTQAAADALPTLTASDPLYSEHATVVAANIAAAAAAKAAA 459 TAYIAPSATVIGDVTIGANVMISPMASIRADEGMPIKVGDNAN VQDQVTLHALETKDEEGNDIEENWVEVNGEKYAVYVGANVS LAHQAQLHGPCIVGDDTFIGMQARVFKSSIGNGCVLEPQAAVI GVTIPDGRYVPAGAVVTSQAEADALPKLTDDYAYAHTNEKVV AVNLALAKGYLTTPTAYIAPSATVIGDVTIGANVMISPMASIRA DEGMPIKVGDNANVQDQVTLHALETKDEEGNDIEENWVEVN GEKYAVYVGANVSLAHQAQLHGPCIVGDDTFIGMQARVFKSS IGNGCVLEPQAAVIGVTIPDGRYVPAGAVVTSQAEADALPKLT DDYAYAHTNEKVVAVNLALAKGYLTTPTAYIAPSATVIGDVTI GANVMISPMASIRADEGMPIKVGDNANVQDQVTLHALETKDE EGNDIEENWVEVNGEKYAVYVGANVSLAHQAQLHGPCIVGD DTFIGMQARVFKSSIGNGCVLEPQAAVIGVTIPDGRYVPAGAV VTSQAEADALPKLTDDYAYAHTNEKVVAVNLALAKGYLTTP 460 TAYIHPSAVVIGQVEIGANVMVSPMASIRSDEGMPIKIEANANV QDGVLIQSLVSKENDKELLEKLKKLNNGEYYNIYLEEGVSLAH QATILNSCYLSSGCFLAEGVVLENSVLNDAVFLGRGVTVTNAE VLEPHVFEAGDVITEEKVEPVEIPEELRAAIAAQRAAVIAANLA AAAAAKAAATAYIHPSAVVIGQVEIGANVMVSPMASIRSDEG MPIKIEANANVQDGVLIQSLVSKENDKELLEKLKKLNNGEYYN IYLEEGVSLAHQATILNSCYLSSGCFLAEGVVLENSVLNDAVFL GRGVTVTNAEVLEPHVFEAGDVITEEKVEPVEIPEELRAAIAAQ RAAVIAANLAAAAAKAAATAYIHPSAVVIGQVEIGANVMVS PMASIRSDEGMPIKIEANANVQDGVLIQSLVSKENDKELLEKL KKLNNGEYYNIYLEEGVSLAHQATILNSCYLSSGCFLAEGVVL ENSVLNDAVFLGRGVTVTNAEVLEPHVFEAGDVITEEKVEPVE IPEELRAAIAAQRAAVIAANLAAAAAAKAAA 461 TAYIHPQANVIGDVEIGANVMVSPMASIRSDEGMPIFVGENAN

VQDQVTLHALETYDEEGNPIEENIVEVDGKKYAVYLGKNVSL AHQAQIHGPSIVGDDTFIGMQALVFKSVLGNNCVLEPQAAAIG VTIPDGRYIPAGKVVTSQAEADALPEVTPDYAYYHTNEQVVY VNTQLAEGYRAAATAYIHPQANVIGDVEIGANVMVSPMASIRS DEGMPIFVGENANVQDQVTLHALETYDEEGNPIEENIVEVDGK KYAVYLGKNVSLAHQAQIHGPSIVGDDTFIGMQALVFKSVLG NNCVLEPQAAAIGVTIPDGRYIPAGKVVTSQAEADALPEVTPD YAYYHTNEQVVYVNTQLAEGYRAAATAYIHPQANVIGDVEIG ANVMVSPMASIRSDEGMPIFVGENANVQDQVTLHALETYDEE GNPIEENIVEVDGKKYAVYLGKNVSLAHQAQIHGPSIVGDDTFI GMQALVFKSVLGNNCVLEPQAAAIGVTIPDGRYIPAGKVVTSQ AEADALPEVTPDYAYYHTNEQVVYVNTQLAEGYRAAA 462 TAYIHPSAEVIGSVEIGENVMISPNASIRSDEGMPIVIGDNANVQ DGVTLHGLETKTEEGELIEENYVEVDGKKYVVYIGENVSLAH QAQVHGPAKVGEDTFIGMGATVTQSILGEGVLLREGAQITGVT LAPGTVVDRGTVLTTQADVASLRKLEPSDPLLKENEEVRKKN LALWEELKKAETAYIHPSAEVIGSVEIGENVMISPNASIRSDEG MPIVIGDNANVQDGVTLHGLETKTEEGELIEENYVEVDGKKY VVYIGENVSLAHQAQVHGPAKVGEDTFIGMGATVTQSILGEG VLLREGAQITGVTLAPGTVVDRGTVLTTQADVASLRKLEPSDP LLKENEEVRKKNLALWEELKKAETAYIHPSAEVIGSVEIGENV MISPNASIRSDEGMPIVIGDNANVQDGVTLHGLETKTEEGELIE ENYVEVDGKKYVVYIGENVSLAHQAQVHGPAKVGEDTFIGM GATVTQSILGEGVLLREGAQITGVTLAPGTVVDRGTVLTTQAD VASLRKLEPSDPLLKENEEVRKKNLALWEELKKAE 463 TAVIAPNAQVIGEVHIGDNVMISPNASIRSDEGMPIYIGENANL QDNVQLHGLEVLDEEGNVIEEALVEVDGKKYVVYIGKNVSLG HQAQIHGPALVGDDTFIGMNAKVFKSRIGNGCVLEPNAQVIGV TIPDGRYVPAGKVVTTQAEADALPVLTPDYAMAHTNERVVAV NLALAAAARAAATAVIAPNAQVIGEVHIGDNVMISPNASIRSD EGMPIYIGENANLQDNVQLHGLEVLDEEGNVIEEALVEVDGK KYVVYIGKNVSLGHQAQIHGPALVGDDTFIGMNAKVFKSRIG NGCVLEPNAQVIGVTIPDGRYVPAGKVVTTQAEADALPVLTPD YAMAHTNERVVAVNLALAAAARAAATAVIAPNAQVIGEVHIG DNVMISPNASIRSDEGMPIYIGENANLQDNVQLHGLEVLDEEG NVIEEALVEVDGKKYVVYIGKNVSLGHQAQIHGPALVGDDTFI GMNAKVFKSRIGNGCVLEPNAQVIGVTIPDGRYVPAGKVVTT QAEADALPVLTPDYAMAHTNERVVAVNLALAAAARAAA 464 TAYIHPQATVIGDVTIGANVMVSPMASIRSDEGMPIFVGDNAN VQDGVTLHALETYDEEGNPIEENWVEVDGKKYAVYLGDNVS LAHQAQVHGPAAVGEDTFIGMQATVFKSKLGNNCVLEPGAA AIGVTIPDGRYIPAGKVVTSQAEADALPEVTPDYAYYHTNEDV VYVNIALAEGYKKLSTAYIHPQATVIGDVTIGANVMVSPMASI RSDEGMPIFVGDNANVQDGVTLHALETYDEEGNPIEENWVEV DGKKYAVYLGDNVSLAHQAQVHGPAAVGEDTFIGMQATVFK SKLGNNCVLEPGAAAIGVTIPDGRYIPAGKVVTSQAEADALPE VTPDYAYYHTNEDVVYVNIALAEGYKKLSTAYIHPQATVIGD VTIGANVMVSPMASIRSDEGMPIFVGDNANVQDGVTLHALET YDEEGNPIEENWVEVDGKKYAVYLGDNVSLAHQAQVHGPAA VGEDTFIGMQATVFKSKLGNNCVLEPGAAAIGVTIPDGRYIPA

GKVVTSQAEADALPEVTPDYAYYHTNEDVVYVNIALAEGYK KLS

[0042] The enzyme comprising silicase activity may be specific or substantially specific to a substrate in the silicate such that the enzyme acts specifically on that substrate and facilitates extracting the metal from the mineral material. In some cases, the enzyme having silicase activity may degrade, digest, and/or disintegrate the silicate. As a result, metals, such as in the form of metal ions, metal atoms, or metal precipitates, may be released from the mineral material (e.g., into a solution). In some cases, the method may comprise collecting the metal or the solution containing the metal. In some cases, the method may comprise separating the metal from the solution. In some cases, the metal may be water soluble. Alternatively or in addition, the metal may precipitate in the solution. The solution may be an aqueous solution comprising water and/or a buffer described anywhere herein.

[0043] In some embodiments, the enzyme having silicase activity has a pKd of at least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 800 mol/s, at least about 900 mol/s, at least about 1000 mol/s, or higher.

[0044] In some cases, the specificity of the enzyme having silicate activity to the substrate may be higher than a wild-type version of the enzyme. For example, a wild-type enzyme may be engineered to improve its specificity for a substrate. In some cases, the enzyme provided herein may be engineered by directed evolution. In some cases, machine learning and artificial intelligence may be used for performing such enzyme engineering and directed evolution methods to design the enzymes of the present disclosure. The engineered enzyme may be synthesized or semi-synthesized. In some embodiments, the enzyme (e.g., the engineered enzyme) has the sequence of any one of SEQ ID NOS: 19-402, or an amino acid sequence having at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or more sequence identity to any one of SEQ ID NOS: 19-402. In some embodiments, the enzyme (e.g., the engineered enzyme) has the sequence of any one of SEQ ID NOS: 403-464, or an amino acid sequence having at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or more sequence identity to any one of SEQ ID NOS: 403-464. [0045] In some embodiments, the enzyme having silicase activity comprises a purification tag. The purification tag can comprise commonly used tags known in the art to aide in the purification of the enzyme having silicase activity from a host cell. The purification tag can comprise, for example, a GST tag, a His tag, a NEXT tag, a FLAG tag, or any other tags known in the art. In some cases, the purification tag is conjugated to the N-terminus of the enzyme having silicase activity. In some cases, the purification tag is conjugated to the C-terminus of the enzyme having silicase activity. In some cases, the purification tag is cleaved from the enzyme having silicase activity after purification. In some cases, the purification tag is not cleaved from the enzyme having silicase activity after purification. In some cases, the purification tag aides in enzyme stability. In some cases, the purification tag does not affect enzymatic efficacy. In some cases, the purification tag does not affect the enzyme having silicase activity polymerization.

[0046] In some cases, the enzyme having silicase activity may have a catalytic activity that is superior to the wild-type version of the same enzyme. For example, the enzymes provided herein may have a higher catalytic rate. This may decrease the time required to extract a given amount of

metal from the mineral material, compared to when using a wild-type enzyme. In some cases, the enzymes provided herein may have reduced energy requirements, as compared to a wild-type enzyme. In some cases, catalytic activity/rate/efficiency may be characterized by using one or more metrics.

[0047] In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 98% at least about 99%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher.

[0048] In some embodiments, the enzyme having silicase activity has a pKd of at least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 100 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 900 mol/s, at least about 1000 mol/s, or higher.

[0049] The methods of the present disclosure, such as the enzymatic reactions provided herein, may be performed under a set of reaction conditions. In some cases, the reaction conditions may comprise a reaction temperature (e.g., a temperature under which the enzymatic reaction, or a portion thereof, is performed). In some cases, the reaction temperature is from about 20 to about 90 degrees Celsius (C). In some cases, the reaction temperature is from about 23 to about 90 degrees Celsius (C). In some cases, the reaction temperature is from about 23 to about 85 degrees Celsius (C). In some cases, the reaction temperature is from about 30 to about 90 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 30 to about 80 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 30 to about 70 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 30 to about 60 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 30 to about 50 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 45 to about 55 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 45 to about 50 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 20 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 23 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 25 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 30 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 35 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 40 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 45 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 50 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 55 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 60 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 70 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 75 degrees Celsius (C). In some cases, the reaction conditions comprises a temperature about 80 degrees Celsius (C). In some cases, the reaction

conditions comprises a temperature about 85 degrees Celsius (C). In some cases, the methods of the present disclosure may be performed in near-ambient temperatures and/or pressures. In some cases, the temperature ranges of the methods of the present disclosure may be significantly lower than temperatures required in acid roasting. (e.g., 200 degrees Celsius (C) and above). This may reduce the energy requirements of the process performed using the methods of the present disclosure

[0050] In some cases, the enzymatic reaction may be performed at a pH from about 4 to about 11. In some cases, the enzymatic reaction may be performed at a pH from about 4 to about 10. In some cases, the enzymatic reaction may be performed at a pH from about 4 to about 9. In some cases, the enzymatic reaction may be performed at a pH from about 4 to about 8. In some cases, the enzymatic reaction may be performed at a pH from about 4 to about 7. In some cases, the enzymatic reaction may be performed at a pH from about 4 to about 6. In some cases, the enzymatic reaction may be performed at a pH from about 5 to about 11. In some cases, the enzymatic reaction may be performed at a pH from about 5 to about 10. In some cases, the enzymatic reaction may be performed at a pH from about 5 to about 9. In some cases, the enzymatic reaction may be performed at a pH from about 5 to about 8. In some cases, the enzymatic reaction may be performed at a pH from about 5 to about 7. In some cases, the enzymatic reaction may be performed at a pH from about 5 to about 6. In some cases, the enzymatic reaction may be performed at a pH from about 6 to about 11. In some cases, the enzymatic reaction may be performed at a pH from about 6 to about 10. In some cases, the enzymatic reaction may be performed at a pH from about 7 to about 11. In some cases, the enzymatic reaction may be performed at a pH from about 7 to about 10. In some cases, the enzymatic reaction may be performed at a pH from about 8 to about 11. In some cases, the enzymatic reaction may be performed at a pH from about 8 to about 10. In some cases, the enzymatic reaction may be performed at a pH from about 9 to about 11. In some cases, the enzymatic reaction may be performed at a pH from about 9 to about 10. In some cases, the enzymatic reaction may be performed at a pH of 4. In some cases, the enzymatic reaction may be performed at a pH of 5. In some cases, the enzymatic reaction may be performed at a pH of 6. In some cases, the enzymatic reaction may be performed at a pH of 7. In some cases, the enzymatic reaction may be performed at a pH of 8. In some cases, the enzymatic reaction may be performed at a pH of 9. In some cases, the enzymatic reaction may be performed at a pH of 10. In some cases, the enzymatic reaction may be performed at a pH of 11. In some cases, the methods of the present disclosure may be performed in pH ranges that are substantially neutral, or in other words, not highly/strongly acidic or highly/strongly basic. In some cases, the method/reaction of the present disclosure may not comprise using a strong acid, such as is used in acid leaching. Instead, the enzymes presented throughout the disclosure may perform enzymatic degradation of the mineral material in neutral pH conditions.

[0051] In some cases, the methods provided herein comprise crushing or grinding the rocks or ores to achieve a particulate size. In some cases, the ground ores comprise a size of about 50 μm to 1 mm. In some cases, the ground ores comprise a size from about 50 μm to 750 μm . In some cases, the ground ores comprise a size from about 50 μm to 250 μm . In some cases, the ground ores comprise a size from about 50 μm to 150 μm . In some cases, the ground ores comprise a size from about 50 μm to 100 μm . In some cases, the ground ores comprise a size of about 50 μm . In some cases, the ground ores comprise a size of about 100 μm . In some cases, the ground ores comprise a size of about 150 μm . In some cases, the ground ores comprise a size of about 500 μm . In some cases, the ground ores comprise a size of about 500 μm . In some cases, the ground ores comprise a size of about 500 μm . In some cases, the ground ores comprise a size of about 750 μm . In some cases, the ground ores comprise a size of about 750 μm . In some cases, the ground ores comprise a size of about 750 μm .

[0052] In some cases, the methods provided herein comprise creating a slurry of crushed rock and liquid. In some cases, the rock to liquid ratio is from about 1-40% (w/v). In some cases, the rock to

liquid ratio is from about 1-35% (w/v). In some cases, the rock to liquid ratio is from about 1-30% (w/v). In some cases, the rock to liquid ratio is from about 1-25% (w/v). In some cases, the rock to liquid ratio is from about 1-20% (w/v). In some cases, the rock to liquid ratio is from about 1-15% (w/v). In some cases, the rock to liquid ratio is from about 1-10% (w/v). In some cases, the rock to liquid ratio is from about 1-5% (w/v). In some cases, the rock to liquid ratio is from about 10-40% (w/v). In some cases, the rock to liquid ratio is from about 10-35% (w/v). In some cases, the rock to liquid ratio is from about 10-30% (w/v). In some cases, the rock to liquid ratio is from about 10-25% (w/v). In some cases, the rock to liquid ratio is from about 15-35% (w/v). In some cases, the rock to liquid ratio is from about 15-30% (w/v). In some cases, the rock to liquid ratio is from about 20-35% (w/v). In some cases, the rock to liquid ratio is from about 20-30% (w/v). In some cases, the rock to liquid ratio is from about 25-35% (w/v). In some cases, the rock to liquid ratio is from about 25-30% (w/v). In some cases, the rock to liquid ratio is about 1% (w/v). In some cases, the rock to liquid ratio is about 5% (w/v). In some cases, the rock to liquid ratio is about 10% (w/v). In some cases, the rock to liquid ratio is about 15% (w/v). In some cases, the rock to liquid ratio is about 20% (w/v). In some cases, the rock to liquid ratio is about 25% (w/v). In some cases, the rock to liquid ratio is about 30% (w/v). In some cases, the rock to liquid ratio is about 35% (w/v). In some cases, the rock to liquid ratio is about 40% (w/v).

[0053] In some cases, the methods provided herein comprise an enzymatic reaction that proceeds for a set period of time. In some cases, the method comprises the enzymatic reaction proceeding for about 1-72 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1-60 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1-48 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1-36 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1-24 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1-12 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1-6 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 12-72 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 12-60 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 12-48 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 12-36 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 12-24 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 24-72 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 24-60 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 24-48 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 24-36 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 36-72 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 36-60 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 36-48 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 48-72 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 48-60 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 1 hour. In some cases, the method comprises the enzymatic reaction proceeding for about 6 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 12 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 24 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 36 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 48 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 60 hours. In some cases, the method comprises the enzymatic reaction proceeding for about 72 hours.

[0054] In some cases, the methods provided herein comprise contacting the enzyme having silicase activity with a co-factor. The co-factor may help further increase the reaction rate when used in combination with the enzyme having silicase activity. In some cases, the co-factor is selected from

the group consisting of: iron, zinc, copper, nickel, cobalt, and any combination thereof. Such cofactor may be used with any enzyme disclosed anywhere in the present disclosure. In some cases, the co-factor may be a non-natural co-factor, such as a co-factor that is typically not used by an enzyme as a co-factor in nature.

[0055] In some cases, the enzyme having silicase activity depolymerizes silicate mineral in the mineral material (e.g., ore/rock). In some cases, the enzyme having silicase activity cleaves one or more Si—O bonds in the mineral material to generate silicic acid (Si(OH)4). In some cases, the metal is lithium, aluminum, iron, nickel, cobalt, strontium, and/or a rare earth element. In some cases, the metal is lithium. In some cases, the metal is aluminum. In some cases, the metal is iron. In some cases, the metal is strontium. As a result of enzymatic degradation and/or disintegration of the mineral material, the metal may be released and extracted from the mineral, in some cases, in a solution, in some cases in the form of a metal ion or a metal atom. The solution may be an aqueous solution.

[0056] In some cases, the method comprises extracting the metal from the solution. In some cases, the method comprises purifying the metal from the solution, thereby generating a purified metal, a metal ion, a metal atom, a solid metal complex, a metal precipitate, or any combination thereof. In some cases, the purified metal has a purity of at least about 60%, at least about 70%, at least about 90%, at least about 99.99, at least about 99.99, at least about 99.99, at least about 99.99%, at least about 99.999%, or higher purity. In some cases, the purified metal is lithium, aluminum, iron, nickel, cobalt, strontium, a rare earth element, and/or uranium. In some cases, the purified metal is industry-grade, battery-grade, and/or pharmaceutical grade. In some cases, the purified metal is industry-grade lithium, battery-grade lithium, and/or pharmaceutical-grade lithium.

[0057] In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 98% at least about 99%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher.

[0058] In some aspects, the enzyme having silicase activity is recombinantly produced in a host cell or in a cell-free production system. In some cases, the host cell is a bacterial cell or yeast cell. In some cases, the bacterial cell is *Escherichia coli*. In some cases, the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*.

[0059] In some embodiments, the methods of the present disclosure comprise performing a reaction involving an enzyme and a mineral material, as described throughout the disclosure. The reaction may be performed inside any suitable container. In some cases, the reaction may be performed on a rock or ore. In some cases, the mineral material may be placed inside a container and added to a solution comprising the enzyme, water, a buffer, and/or potential other reagents for performing the reaction. In some cases, the reaction may be performed in one or more of a container, a dish, a beaker, a device, a tank, a reactor, and/or any combination thereof. The containers (e.g., one or more reactors and/or tanks) may be connected to one another to perform one or more reactions according to the embodiments of the present disclosure. The containers may also be reaction units and/or process units each of which may serve a function as part of the method and/or in combination with the method. For example, one or more tanks, reactors, and/or processing units,

may be connected to each other with any configuration, such as in series, in parallel, or any combination thereof to perform the method steps such as contact the enzyme with the mineral, extracting the metal, separating the metal from the solution, purifying the metal, processing the metal, and converting the metal into an industry-grade, battery-grade, or pharmaceutical-grade metal. In some cases, the method further comprises grinding the mineral material (e.g., rock/ore) prior to performing the reaction (e.g., the enzymatic degradation). In some cases, the method further comprises using a filtration/chelating system, a precipitation system, a recycle system, or any combination thereof.

[0060] In an aspect, provided herein is a non-naturally occurring enzyme having silicase activity, the enzyme comprising at least one amino acid variation relative to a wild-type enzyme, and having increased ability to release metals from a mineral material (e.g., rock/ore) as compared to the wildtype enzyme. In some cases, the enzyme having silicase activity is used in a reaction/process of the present disclosure for metal extraction, as described anywhere in the present disclosure. In some cases, a wild-type carbonic anhydrase, gamma carbonic anhydrase, or alpha carbonic anhydrase may be used as a starting point for the enzyme engineering and performing the modifications to design and synthesize the non-naturally occurring enzyme having silicase activity. The nonnaturally occurring enzyme having silicase activity may in some cases be a modified, engineered, and semi-synthetic enzyme (e.g., the enzyme having silicase activity for performing the methods and reactions of the present disclosure). In some cases, a wild-type carbonic anhydrase, gamma carbonic anhydrase, or alpha carbonic anhydrase may be used as a starting point for the enzyme engineering and performing the modifications to design and synthesize the non-naturally occurring enzyme (e.g., the enzyme having silicase activity for performing the methods and reactions of the present disclosure). As such, the resulting modified enzyme may have some similarities, for example with respect to characteristics and sequence, to a wild-type Carbonic Anhydrase, gamma Carbonic Anhydrase, or alpha Carbonic Anhydrase, and some differences, for example with respect to characteristics and sequence, to a wild-type carbonic anhydrase, gamma carbonic anhydrase, or alpha carbonic anhydrase.

[0061] In some cases the wild-type enzyme is selected from the group consisting of: *Methanosarcina thermophila* gamma carbonic anhydrase, *Bacillus* lichenmformis CG-B52 gamma carbonic anhydrase, Pelobacter carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma carbonic anhydrase, *Methanosarcina barkeri* gamma carbonic anhydrase, *Methanosarcina mazei* carbonic anhydrase, *Bacillus halodurans* alpha carbonic anhydrase, Alkalihalobacillus *clausii* (strain KSM-K16) (*Bacillus clausii*) alpha carbonic anhydrase, *Methanosarcina acetivorans* carbonate dehydratase, Kofleriaceae bacterium SLC26A/SulP transporter domain-containing protein, Thermodesulfitimonas *autotrophica* carbonic anhydrase/acetyltransferase-like protein (Isoleucine patch superfamily), *Fischerella thermalis/Mastigocladus laminosus* JSC-11 carboxysome assembly protein CcmM, *Thermosynechococcus vestitus* BP-

1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix thermoacetophila* carbonate dehydratase, *Thermosyntropha lipolytica* carbonic anhydrase or acetyltransferase, isoleucine patch superfamily, Desulfofundulus thermobenzoicus transferase, *Archaeoglobus veneficus* carbonate dehydratase, Suberites domuncula carbonic anhydrase, and any combination thereof.

[0062] In some cases, the non-naturally occurring enzyme is derived from an organism selected from the group consisting of: *Methanosarcina thermophila*, *Bacillus* lichenmformis CG-B52, Pelobacter carbinolicus, Syntrophus aciditrophicus, *Methanosarcina* barkeri, *Methanosarcina mazei*, *Bacillus halodurans*, *Alkalihalobacillus clausii* (strain KSM-K16) (*Bacillus clausii*), *Methanosarcina acetivorans*, Kofleriaceae bacterium, Thermodesulfitimonas *autotrophica*, *Fischerella thermalis/Mastigocladus laminosus* JSC-11 carboxysome assembly protein CcmM, *Thermosynechococcus vestitus* BP-1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix thermoacetophila*, *Thermosyntropha lipolytica*,

Desulfofundulus thermobenzoicus, Archaeoglobus veneficus, Suberites domuncula, and any combination thereof.

[0063] In some cases, the non-naturally occurring enzyme comprises an amino acid sequence having at least about 10%, at least about 20%, at least about 30%, at least about 40%, having at least about 50%, at least about 60%, at least about 70%, at least about 93%, at least about 94%, at least about 95%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-402. In some cases, the non-naturally occurring enzyme comprises an amino acid sequence having at least about 10%, at least about 20%, at least about 30%, at least about 40%, having at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 403-464. In some cases, the wild-type enzyme is a carbonic anhydrase. In some cases, the carbonic anhydrase is a gamma carbonic anhydrase or alpha carbonic anhydrase.

[0064] In some cases, the methods of the present disclosure are performed on a mineral material. In some cases, the mineral material comprises or is a rock, an ore, a deposit, a clay, a natural mineral material, a man-made mineral material, or any combination thereof. The method may be according to the embodiments described anywhere herein. According to the embodiments described anywhere in the present disclosure, in some cases, an enzyme having silicase activity acts on a mineral material. The enzyme and the method of the present disclosure may degrade, digest, or disintegrate the mineral material and extract a metal therefrom. The mineral material such as a rock, ore, natural mineral materials, and/or man-made mineral materials may by abundant sources of valuable metals such as lithium, aluminum, iron, nickel, cobalt, copper, strontium, rare earth elements, uranium, and other metals with vast industrial use and applications. The methods and enzymes of the present disclosure facilitate access to such sources. In some cases, the mineral material comprises a silicate. In some cases, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, a tectosilicate or any combination thereof. In some cases, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some cases, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some cases, amorphous silicate is selected from the group consisting of obsidian, coal fly ash, pumice, glass, and any combination thereof. In some cases, the tectosilicate comprises quartz, sand, or glass.

[0065] In some cases, the non-naturally occurring enzyme has an increased ability to depolymerize silicate mineral in the mineral material, rock, ore, or other kind of mineral material as compared to the wild-type enzyme, increased selectivity or specificity toward a mineral structure in the mineral material, or both. In some cases, the non-naturally occurring enzyme has increased ability to cleave one or more Si—O bonds in the mineral material to generate silicic acid (Si(OH)4) as compared to the wild-type enzyme. In some cases, the metal comprises lithium, aluminum, iron, nickel, cobalt, or a rare earth element. In some cases, the metal comprises lithium. In some cases, the metal comprises strontium. In some cases, the metal comprises iron. In some cases, the metal comprises strontium. In some cases, the non-naturally occurring enzyme is recombinantly produced in a host cell or in a cell-free production system. In some cases, the host cell is a bacterial cell or yeast cell. In some cases, the bacterial cell is *Escherichia coli*. In some cases, the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*.

[0066] In an aspect, provided herein is a reaction mixture comprising a mineral material and a non-naturally occurring enzyme having silicase activity, wherein the non-naturally occurring enzyme comprises at least one amino acid variation relative to a wild-type enzyme, and has increased

ability to release metal from the mineral material as compared to the wild-type enzyme. The mineral material may be according to any embodiment described anywhere herein. It may comprise silicates and/or Si—O bonds. In some cases, the mineral material comprises silicates. In some cases, the mineral material may comprise an inosilicate, a phyllosilicate, an amorphous silicate, or any combination thereof. In some cases, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some cases, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some cases, the amorphous silicate is selected from the group consisting of obsidian, coal fly ash, pumice, glass, and any combination thereof. The mineral material may be a source of a metal.

[0067] The metal may be extracted from the mineral material (in some cases a rock or ore) by using the methods and enzymes described anywhere in the present disclosure. The enzyme having silicase activity may be any enzyme disclosed anywhere in the present disclosure. In some cases, the reaction mixture has a pH from about 4 to about 11. In some cases, the reaction mixture has a pH from about 4 to about 10. In some cases, the reaction mixture has a pH from about 4 to about 9. In some cases, the reaction mixture has a pH from about 4 to about 8. In some cases, the reaction mixture has a pH from about 4 to about 7. In some cases, the reaction mixture has a pH from about 4 to about 6. In some cases, the reaction mixture has a pH from about 5 to about 11. In some cases, the reaction mixture has a pH from about 5 to about 10. In some cases, the reaction mixture has a pH from about 5 to about 9. In some cases, the reaction mixture has a pH from about 5 to about 8. In some cases, the reaction mixture has a pH from about 5 to about 7. In some cases, the reaction mixture has a pH from about 5 to about 6. In some cases, the reaction mixture has a pH from about 6 to about 11. In some cases, the reaction mixture has a pH from about 6 to about 10. In some cases, the reaction mixture has a pH from about 7 to about 11. In some cases, the reaction mixture has a pH from about 7 to about 10. In some cases, the reaction mixture has a pH from about 8 to about 11. In some cases, the reaction mixture has a pH from about 8 to about 10. In some cases, the reaction mixture has a pH from about 9 to about 11. In some cases, the reaction mixture has a pH from about 9 to about 10. In some cases, the reaction mixture has a pH of 4. In some cases, the reaction mixture has a pH of 5. In some cases, the reaction mixture has a pH of 6. In some cases, the reaction mixture has a pH of 7. In some cases, the reaction mixture has a pH of 8. In some cases, the reaction mixture has a pH of 9. In some cases, the reaction mixture has a pH of 10. In some cases, the reaction mixture has a pH of 11. In some cases, the reaction mixture has a temperature from about 20 to about 90 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 23 to about 90 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 23 to about 85 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 30 to about 90, from about 30 to about 80, from about 30 to about 70, from about 30 to about 60, or from about 30 to about 50 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 45 to about 55 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 45 to about 50 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 20 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 23 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 25 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 30 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 35 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 40 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 45 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 50 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 55 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 60 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 70 degrees Celsius (C). In some cases, the reaction mixture has a

temperature about 75 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 80 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 85 degrees Celsius (C). In some cases, the reaction mixture further comprises a co-factor of the non-naturally occurring enzyme. In some cases, the co-factor is selected from the group consisting of iron, zinc, copper, nickel, and cobalt. In some cases, the co-factor is copper. In some cases, the co-factor is iron. In some cases, the reaction mixture further comprises a buffered saline solution. In some cases, the buffered solution comprises saline, glycine, iron ions, or any combination thereof. In some cases the buffered solution comprises TRIS, PBS, citrate, monosodium glutamate, or any combination thereof. In some cases, the reaction mixture further comprises an activator co-factor of the non-naturally occurring enzyme. In some cases, the activator co-factor is glycine. The reaction mixture may be used to perform any method described anywhere herein using any enzyme, any co-factor, and/or any reaction condition disclosed anywhere herein.

[0068] In some cases, the reaction mixture provided herein comprise crushing or grinding the rocks or ores to achieve a particulate size. In some cases, the ground ores comprise a size of about 50 μ m to 1 mm. In some cases, the ground ores comprise a size from about 50 μ m to 750 μ m. In some cases, the ground ores comprise a size from about 50 μ m to 250 μ m. In some cases, the ground ores comprise a size from about 50 μ m to 150 μ m. In some cases, the ground ores comprise a size from about 50 μ m. In some cases, the ground ores comprise a size of about 50 μ m. In some cases, the ground ores comprise a size of about 150 μ m. In some cases, the ground ores comprise a size of about 150 μ m. In some cases, the ground ores comprise a size of about 500 μ m. In some cases, the ground ores comprise a size of about 500 μ m. In some cases, the ground ores comprise a size of about 500 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m.

[0069] In some cases, the reaction mixture provided herein comprise creating a slurry of crushed rock and liquid. In some cases, the rock to liquid ratio is from about 1-40% (w/v). In some cases, the rock to liquid ratio is from about 1-35% (w/v). In some cases, the rock to liquid ratio is from about 1-30% (w/v). In some cases, the rock to liquid ratio is from about 1-25% (w/v). In some cases, the rock to liquid ratio is from about 1-20% (w/v). In some cases, the rock to liquid ratio is from about 1-15% (w/v). In some cases, the rock to liquid ratio is from about 1-10% (w/v). In some cases, the rock to liquid ratio is from about 1-5% (w/v). In some cases, the rock to liquid ratio is from about 10-40% (w/v). In some cases, the rock to liquid ratio is from about 10-35% (w/v). In some cases, the rock to liquid ratio is from about 10-30% (w/v). In some cases, the rock to liquid ratio is from about 10-25% (w/v). In some cases, the rock to liquid ratio is from about 15-35% (w/v). In some cases, the rock to liquid ratio is from about 15-30% (w/v). In some cases, the rock to liquid ratio is from about 20-35% (w/v). In some cases, the rock to liquid ratio is from about 20-30% (w/v). In some cases, the rock to liquid ratio is from about 25-35% (w/v). In some cases, the rock to liquid ratio is from about 25-30% (w/v). In some cases, the rock to liquid ratio is about 1% (w/v). In some cases, the rock to liquid ratio is about 5% (w/v). In some cases, the rock to liquid ratio is about 10% (w/v). In some cases, the rock to liquid ratio is about 15% (w/v). In some cases, the rock to liquid ratio is about 20% (w/v). In some cases, the rock to liquid ratio is about 25% (w/v). In some cases, the rock to liquid ratio is about 30% (w/v). In some cases, the rock to liquid ratio is about 35% (w/v). In some cases, the rock to liquid ratio is about 40% (w/v). [0070] In some cases, the reaction mixture provided herein comprise an enzymatic reaction that proceeds for a set period of time. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-24 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-12 hours. In some cases,

the reaction mixture comprises the enzymatic reaction proceeding for about 1-6 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-24 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 48-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 48-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1 hour. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 6 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 72 hours. [0071] The reaction mixture comprises a non-naturally occurring enzyme described anywhere herein. As described throughout the present disclosure, in some cases, the non-naturally occurring enzyme may be an engineered and/or semi-synthetic enzyme having a modification compared to a wild-type enzyme. In some cases, the wild-type enzyme is selected from the group consisting of:

herein. As described throughout the present disclosure, in some cases, the non-naturally occurring enzyme may be an engineered and/or semi-synthetic enzyme having a modification compared to a wild-type enzyme. In some cases, the wild-type enzyme is selected from the group consisting of: *Methanosarcina thermophila* gamma carbonic anhydrase, *Bacillus* lichenmformis CG-B52 gamma carbonic anhydrase, Pelobacter carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma carbonic anhydrase, *Methanosarcina barkeri* gamma carbonic anhydrase, *Methanosarcina mazei* carbonic anhydrase, *Bacillus halodurans* alpha carbonic anhydrase, Alkalihalobacillus *clausii* (strain KSM-K16) (*Bacillus clausii*) alpha carbonic anhydrase, *Methanosarcina acetivorans* carbonate dehydratase, Kofleriaceae bacterium SLC26A/SulP transporter domain-containing protein, Thermodesulfitimonas *autotrophica* carbonic anhydrase/acetyltransferase-like protein (Isoleucine patch superfamily), *Fischerella thermalis/Mastigocladus laminosus* JSC-11 carboxysome assembly protein CcmM, *Thermosynechococcus vestitus* BP-

1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix thermoacetophila* carbonate dehydratase, *Thermosyntropha lipolytica* carbonic anhydrase or acetyltransferase, isoleucine patch superfamily, Desulfofundulus thermobenzoicus transferase, *Archaeoglobus veneficus* carbonate dehydratase, Suberites domuncula carbonic anhydrase, and any combination thereof.

[0072] In some cases, the non-naturally occurring enzyme comprises an amino acid sequence having at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80% at least about 95%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-402. In some cases, the non-naturally occurring enzyme comprises an amino acid sequence having at least

about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80% at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 403-464. In some cases, the wild-type enzyme is a carbonic anhydrase. In some cases, the carbonic anhydrase is a gamma carbonic anhydrase or alpha carbonic anhydrase.

[0073] In some cases, the non-naturally occurring enzyme has increased ability to depolymerize silicate mineral in the mineral material as compared to the wild-type enzyme. In some cases, the non-naturally occurring enzyme has increased ability to cleave one or more Si—O bonds in the mineral material (e.g., rock/ore) to generate silicic acid (Si(OH)4) as compared to the wild-type enzyme. In some cases, the metal comprises lithium. In some cases, the metal comprises aluminum. In some cases, the metal comprises iron. In some cases, the metal comprises strontium. In some cases, the metal may comprise lithium, aluminum, nickel, iron, cobalt, copper, a rare earth element, uranium, strontium, another metal, or any combination thereof. In some cases, the non-naturally occurring enzyme is recombinantly produced in a host cell. In some cases, the host cell is a bacterial cell or yeast cell. In some cases, the bacterial cell is *Escherichia coli*. In some cases, the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*.

[0074] In an aspect, provided herein is a polynucleotide comprising a nucleotide sequence encoding the non-naturally occurring enzyme of any one of the preceding embodiments. In an aspect, provided herein is a vector comprising the polynucleotide comprising the nucleotide sequence encoding the non-naturally occurring enzyme. The non-naturally occurring enzyme may be according to any embodiment mentioned anywhere in the present disclosure. The polynucleotide and vector may be designed and engineered. The polynucleotide and the vector may get synthesized. The polynucleotide and/or vector may be used to generate and/or produce the non-naturally occurring enzyme of the present disclosure.

[0075] In an aspect, provided herein is a method of increasing silicase activity of an enzyme, the method comprising contacting or combining the enzyme with a non-natural co-factor. In some cases, the enzyme and the co-factor may be added to a solution. In some cases, the co-factor may be brought in proximity of the enzyme. In some cases, the enzyme and the co-factor may be part of the same system, the same reaction mixture, or the same kit for performing the methods of the present disclosure. In some cases, the non-natural co-factor increases silicase activity of the enzyme as compared to the enzyme in the presence of a natural co-factor. In some cases, the nonnatural co-factor may be copper. In some cases, natural co-factor is zinc. In some cases, natural cofactor is iron. In some cases, the method is performed in the absence of the natural co-factor. In some cases, the non-natural co-factor does not act as a co-factor for the enzyme having silicase activity in nature. In some cases, the method further comprises contacting the enzyme and the nonnatural co-factor with the mineral material under reaction conditions such that a metal contained within the mineral material is solubilized and released from the mineral material such as rock/ore. [0076] As an example, a method of the present disclosure comprises using an enzyme having silicase activity (wild-type or engineered/modified/semi-synthetic) to degrade a mineral material such as a rock/ore comprising metal-bearing silicates so as to extract the metal from the mineral material. Zinc may act as a natural co-factor for the enzyme comprising silicase activity in nature. For example, a natural silicate rock may get degraded by a wild-type silicase enzyme such as a gamma carbonic anhydrase in nature, zinc may act as a co-factor for the wild-type enzyme, catalyze the digestion/degradation reaction, and speed it up and/or increase its efficiency. Alternatively, in some cases, the method of the present disclosure may comprise bringing an enzyme having silicase activity to a mineral material such as a rock/ore comprising metal-bearing silicates, and further provide a co-factor other than zinc (the natural co-factor) to increase the catalytic effects of the enzyme having silicase activity on the mineral material. The co-factor other

than zinc may be a non-natural co-factor. The non-natural co-factor is in some cases, copper, iron, nickel, cobalt, or glycine. In some cases, the non-natural co-factor may work better than the natural co-factor. For example, copper, iron, nickel, cobalt, or glycine may work better than zinc in increasing the catalytic efficiency of the enzyme and increasing the reaction rate. The enzyme used in the method of the present disclosure may comprise using a wild-type enzyme or a modified enzyme described anywhere in the present disclosure.

[0077] In some cases, the amount of metal extracted, solubilized/precipitated in the extraction solution, and/or released from the mineral material (e.g., ore/rock) is greater than an amount of metal extracted from the mineral material when the enzyme is contacted with the natural co-factor (e.g., in unit time when other conditions are equal). In some cases, the amount of metal extracted from the mineral material is greater than an amount of metal extracted from the mineral material when the enzyme is contacted with the natural co-factor. In some cases, the mineral material comprises silicate. In some cases, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, a tectosilicate, or any combination thereof. In some cases, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some cases, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some cases, the amorphous silicate is selected from the group consisting of obsidian, coal fly ash, pumice, glass, and any combination thereof. In some cases, the enzyme having silicase activity is a carbonic anhydrase. In some cases, the carbonic anhydrase is a gamma carbonic anhydrase or an alpha carbonic anhydrase. In some cases, the enzyme is a wild-type enzyme. In some cases, the enzyme is a modified or engineered enzyme.

[0078] In some cases, the enzyme having silicase activity is derived from an organism selected from the group consisting of: *Methanosarcina thermophila*, *Bacillus* lichenmformis CG-B52, Pelobacter carbinolicus, Syntrophus aciditrophicus, *Methanosarcina* barkeri, *Methanosarcina mazei*, *Bacillus halodurans*, *Alkalihalobacillus clausii* (strain KSM-K16) (*Bacillus clausii*), *Methanosarcina acetivorans*, Kofleriaceae bacterium, Thermodesulfitimonas *autotrophica*, *Fischerella thermalis/Mastigocladus laminosus* JSC-11 carboxysome assembly protein CcmM, *Thermosynechococcus vestitus* BP-1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix thermoacetophila*, *Thermosyntropha lipolytica*, *Desulfofundulus thermobenzoicus*, *Archaeoglobus veneficus*, Suberites domuncula, and any combination thereof.

[0079] In some cases, the enzyme having silicase activity comprises an amino acid sequence having at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, having at least about 50%, at least about 60%, at least about 70%, tale at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-402. In some cases, the enzyme having silicase activity comprises an amino acid sequence having at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, having at least about 50%, at least about 60%, at least about 70%, tale at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 403-464. In some cases, the enzyme having silicase activity is an engineered enzyme. In some cases, the enzyme having silicase activity is an enzyme having at least one amino acid variation as compared to a wild-type enzyme.

[0080] In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at

least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher. In some embodiments, the enzyme having silicase activity has a pKd of at least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 100 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 800 mol/s, at least about 900 mol/s, at least about 1000 mol/s, or higher.

[0081] In some cases, the reaction conditions comprise a temperature from about 20 to about 90 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 23 to about 90 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 23 to about 85 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 30 to about 90, from about 30 to about 80, from about 30 to about 70, from about 30 to about 60, or from about 30 to about 50 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature from about 45 to about 55 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 45 to about 50 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 20 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 23 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 25 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 30 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 35 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 40 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 45 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 50 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 55 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 60 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 70 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 75 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 80 degrees Celsius (C). In some cases, the reaction conditions comprise a temperature about 85 degrees Celsius (C). In some cases, the reaction conditions comprise a pH from about 4 to about 11. In some cases, the reaction conditions comprise a pH from about 4 to about 10. In some cases, the reaction conditions comprise a pH from about 4 to about 9. In some cases, the reaction conditions comprise a pH from about 4 to about 8. In some cases, the reaction conditions comprise a pH from about 4 to about 7. In some cases, the reaction conditions comprise a pH from about 4 to about 6. In some cases, the reaction conditions comprise a pH from about 5 to about 11. In some cases, the reaction conditions comprise a pH from about 5 to about 10. In some cases, the reaction conditions comprise a pH from about 5 to about 9. In some cases, the reaction conditions comprise a pH from about 5 to about 8. In some cases, the reaction conditions comprise a pH from about 5 to about 7. In some cases, the reaction conditions comprise a pH from about 5 to about 6. In some cases, the reaction conditions comprise a pH from about 6 to about 11. In some cases, the reaction conditions comprise a pH from about 6 to about 10.

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In some cases, the reaction conditions comprise a pH from about 7 to about 11. In some cases, the
reaction conditions comprise a pH from about 7 to about 10. In some cases, the reaction conditions
comprise a pH from about 8 to about 11. In some cases, the reaction conditions comprise a pH from
about 8 to about 10. In some cases, the reaction conditions comprise a pH from about 9 to about 11.
In some cases, the reaction conditions comprise a pH from about 9 to about 10. In some cases, the
reaction conditions comprise a pH of 4. In some cases, the reaction conditions comprise a pH of 5.
In some cases, the reaction conditions comprise a pH of 6. In some cases, the reaction conditions
comprise a pH of 7. In some cases, the reaction conditions comprise a pH of 8. In some cases, the
reaction conditions comprise a pH of 9. In some cases, the reaction conditions comprise a pH of 10.
In some cases, the reaction conditions comprise a pH of 11. In some cases, the reaction conditions
comprise contacting the enzyme having silicase activity with a co-factor. In some cases, the co-
factor is selected from the group consisting of: Iron, zinc, copper, nickel, and cobalt.
[0082] In some cases, the reaction conditions provided herein comprise crushing or grinding the
rocks or ores to achieve a particulate size. In some cases, the ground ores comprise a size of about
50 \mu m to 1 mm. In some cases, the ground ores comprise a size from about 50 \mu m to 750 \mu m. In
some cases, the ground ores comprise a size from about 50 µm to 500 µm. In some cases, the
ground ores comprise a size from about 50 µm to 250 µm. In some cases, the ground ores comprise
a size from about 50 μm to 150 μm. In some cases, the ground ores comprise a size from about 50
μm to 100 μm. In some cases, the ground ores comprise a size of about 50 μm. In some cases, the
ground ores comprise a size of about 100 µm. In some cases, the ground ores comprise a size of
about 150 μm. In some cases, the ground ores comprise a size of about 250 μm. In some cases, the
ground ores comprise a size of about 500 µm. In some cases, the ground ores comprise a size of
about 750 μm. In some cases, the ground ores comprise a size of about 1 mm.
[0083] In some cases, the reaction conditions provided herein comprise creating a slurry of crushed
rock and liquid. In some cases, the rock to liquid ratio is from about 1-40% (w/v). In some cases,
the rock to liquid ratio is from about 1-35% (w/v). In some cases, the rock to liquid ratio is from
about 1-30% (w/v). In some cases, the rock to liquid ratio is from about 1-25% (w/v). In some
cases, the rock to liquid ratio is from about 1-20% (w/v). In some cases, the rock to liquid ratio is
from about 1-15% (w/v). In some cases, the rock to liquid ratio is from about 1-10% (w/v). In some
cases, the rock to liquid ratio is from about 1-5% (w/v). In some cases, the rock to liquid ratio is
from about 10-40% (w/v). In some cases, the rock to liquid ratio is from about 10-35% (w/v). In
some cases, the rock to liquid ratio is from about 10-30% (w/v). In some cases, the rock to liquid
ratio is from about 10-25% (w/v). In some cases, the rock to liquid ratio is from about 15-35%
(w/v). In some cases, the rock to liquid ratio is from about 15-30% (w/v). In some cases, the rock to
liquid ratio is from about 20-35% (w/v). In some cases, the rock to liquid ratio is from about 20-
30% (w/v). In some cases, the rock to liquid ratio is from about 25-35% (w/v). In some cases, the
rock to liquid ratio is from about 25-30% (w/v). In some cases, the rock to liquid ratio is about 1%
(w/v). In some cases, the rock to liquid ratio is about 5% (w/v). In some cases, the rock to liquid
ratio is about 10% (w/v). In some cases, the rock to liquid ratio is about 15% (w/v). In some cases,
the rock to liquid ratio is about 20% (w/v). In some cases, the rock to liquid ratio is about 25%
(w/v). In some cases, the rock to liquid ratio is about 30% (w/v). In some cases, the rock to liquid
ratio is about 35% (w/v). In some cases, the rock to liquid ratio is about 40% (w/v).
[0084] In some cases, the reaction conditions provided herein comprise an enzymatic reaction that
proceeds for a set period of time. In some cases, the reaction conditions comprise the enzymatic
reaction proceeding for about 1-72 hours. In some cases, the reaction conditions comprise the
enzymatic reaction proceeding for about 1-60 hours. In some cases, the reaction conditions
comprise the enzymatic reaction proceeding for about 1-48 hours. In some cases, the reaction
conditions comprise the enzymatic reaction proceeding for about 1-36 hours. In some cases, the
reaction conditions comprise the enzymatic reaction proceeding for about 1-24 hours. In some
cases, the reaction conditions comprise the enzymatic reaction proceeding for about 1-12 hours. In
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some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 1-6 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 12-72 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 12-60 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 12-48 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 12-36 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 12-24 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 24-72 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 24-60 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 24-48 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 24-36 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 36-72 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 36-60 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 36-48 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 48-72 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 48-60 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 1 hour. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 6 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 12 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 24 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 36 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 48 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 60 hours. In some cases, the reaction conditions comprise the enzymatic reaction proceeding for about 72 hours. [0085] In some cases, the enzyme having silicase activity depolymerizes silicate mineral in the mineral material (e.g., ore/rock). In some cases, the enzyme having silicase activity cleaves one or more Si—O bonds in the mineral material to generate silicic acid (Si(OH)4). In some cases, the metal (e.g., metal ion) extracted from the mineral material is lithium, aluminum, iron, nickel, cobalt, uranium, strontium, a rare earth element, or any combination thereof. In some cases, the metal is lithium. In some cases, the metal ion is aluminum. In some cases, the metal ion is iron. In some cases, the metal ion is strontium.

[0086] In some cases, the metal is released into a solution. In some cases, the metal is extracted and released in form of metal ion. In some cases, the metal is extracted and released in form of a metal atom. In some cases, the metal is solubilized in a solution comprising water and/or buffer. In some cases the buffer comprises TRIS, PBS, citrate, monosodium glutamate, or any combination thereof. In some cases, the metal precipitates in the solution. In some cases, the metal is released and/or extracted in form of a metal complex. In some cases, the method further comprises extracting, and/or separating the metal from the solution. Any proper separation technique may be used. In some cases, an electromagnetic force may be used to separate metal ions from the solution. In some cases, a solid-liquid separation technique may be used to separate a metal precipitate from the solution. Any combination of separation and processing methods may be used. The method may comprise collecting the metal from the mineral material (e.g., source rock/ore) and from the system or solution used to perform the extraction according to the embodiments of the present disclosure and provide the metal for use in its intended application. In some cases, the metal may be processed as an industry-grade metal, battery-grade metal, or pharmaceutical-grade metal. An example of this may comprise industry-grade metal, battery-grade metal, or pharmaceutical-grade lithium. [0087] In some cases, the method further comprises comprising purifying the metal from the solution, thereby generating a purified metal, a metal ion, a metal atom, a solid metal complex, a metal precipitate, or any combination thereof. In some cases, the purified metal ion a purity of at

least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, at least about 95%, at least about 99.99%, at least about 99.99%, at least about 99.999% or greater.

[0088] In some cases, the method is performed in situ or ex-situ. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 95%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher.

[0089] In some cases, the enzyme having silicase activity is recombinantly produced in a host cell or in a cell-free production system. In some cases, the host cell is a bacterial cell or a yeast cell. In some cases, the bacterial cell is *Escherichia coli* or the yeast cell is *Pichia pastoris* or *Saccharomyces cerevisiae*.

[0090] In an aspect, provided herein is a reaction mixture comprising an enzyme having silicase activity, and a non-natural co-factor. In some cases, the non-natural co-factor is bound to the enzyme having silicase activity. In some cases, the non-natural co-factor increases a function of the enzyme having silicase activity as compared to a reaction mixture comprising the enzyme having silicase activity and a natural co-factor. In some cases, the non-natural co-factor is copper, iron, nickel, cobalt, or glycine. In some cases, the natural co-factor is zinc. In some cases, the natural cofactor is iron. In some cases, the reaction mixture does not contain the natural co-factor. In some cases, the non-natural co-factor does not act as a co-factor for the enzyme having silicase activity in nature. In some cases, the reaction mixture further comprises the mineral material comprising silicate. In some cases, the reaction mixture has reaction conditions such that a metal contained within the mineral material is extracted, released, or solubilized, or precipitated into a solution from the mineral material. For example, a solution may be provided in proximity of a mineral material comprising a metal-bearing silicate. An enzyme having silicase activity and a non-natural co-factor according to the embodiments disclosed anywhere herein may be present in the solution. The enzyme may facilitate breaking Si—O bonds in the mineral material, thereby digesting and/or degrading the mineral material (e.g., rock/ore) and releasing the metal encased in the mineral material in form of metal ion, metal atom, metal solubilized in solution, metal precipitated in solution, or any combination thereof. The non-natural co-factor combined with or bound to the enzyme may further enhance the catalytic efficiency of the enzyme in releasing the metal from the mineral material in any of the mentioned forms.

[0091] In some cases, the enzyme having silicase activity has increased ability to release metal in any form mentioned anywhere herein, from mineral materials (e.g., ore/rock) in the presence of the non-natural co-factor as compared to the enzyme having silicase activity in the presence of the natural co-factor. In some cases, the mineral material may comprise silicates. In some cases, the mineral material may comprise a metal bearing silicate. In some cases, the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, or any combination thereof. In some cases, the inosilicate is selected from the group consisting of: spodumene, wollastonite, balangeroite, eveslogite, holmquistite, jadeite, shattuckite, augite, tremolite, and any combination thereof. In some cases, the phyllosilicate is selected from the group consisting of: lepidolite, hectorite, kaolinite, vermiculite, muscovite, montmorillonite, and any combination thereof. In some cases, the amorphous silicate is selected from the group consisting of obsidian, coal fly ash,

pumice, glass, and any combination thereof.

[0092] In some cases, the enzyme having silicase activity has a sequence identity of at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50% or more identity with a carbonic anhydrase. In some cases, the enzyme having silicase activity has a sequence identity of at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50% or more identity with an alpha carbonic anhydrase or a gamma carbonic anhydrase. In some cases, the enzyme having silicase activity is derived from an organism selected from the group consisting of: *Methanosarcina thermophila*, *Bacillus* lichenmformis CG-B52, Pelobacter carbinolicus, Syntrophus aciditrophicus, *Methanosarcina* barkeri, *Methanosarcina mazei*, *Bacillus halodurans*, *Alkalihalobacillus clausii* (strain KSM-K16) (*Bacillus clausii*), *Methanosarcina acetivorans*, Kofleriaceae bacterium SLC26A/SulP, Thermodesulfitimonas *autotrophica*, *Fischerella thermalis/Mastigocladus laminosus*, *Thermosynechococcus vestitus* BP-1/(*Thermosynechococcus elongatus* BP-1) carboxysome assembly protein CcmM, *Methanothrix thermoacetophila*, *Thermosyntropha lipolytica*, isoleucine patch superfamily, Desulfofundulus thermobenzoicus, *Archaeoglobus veneficus*, Suberites domuncula, and any combination thereof.

[0093] In some cases, the non-naturally occurring enzyme comprises an amino acid sequence having at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-402. In some cases, the non-naturally occurring enzyme comprises an amino acid sequence having at least about 5%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, at least about 99.5%, at least about 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 403-464. In some cases, the enzyme having silicase activity is an engineered enzyme. In some cases, the enzyme having silicase activity is an enzyme having at least one amino acid variation as compared to a wild-type enzyme.

[0094] In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 99.5% or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 98% at least about 99%, at least about 98.5% or higher. In some embodiments, the enzyme having silicase activity has a catalytic efficiency of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher. In some embodiments, the method has a maximum metal extraction rate of at least about 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 2.5, 2.75, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 8, 9, 10 times or higher.

[0095] In some embodiments, the enzyme having silicase activity has a pKd of at least about 5, at least about 6, at least about 7, at least about 8, at least about 9, at least about 10, at least about 11, or higher. In some embodiments, the enzyme having silicase activity has a Kcat value of at least about 2 mol per second (mol/s), at least about 10 mol/s, at least about 50 mol/s, at least about 200 mol/s, at least about 300 mol/s, at least about 400 mol/s, at least about 500 mol/s, at least about 600 mol/s, at least about 700 mol/s, at least about 800 mol/s, at least about 900

mol/s, at least about 1000 mol/s, or higher.

[0096] In some cases, the reaction mixture has a pH from about 4 to about 11. In some cases, the reaction mixture has a pH from about 4 to about 10. In some cases, the reaction mixture has a pH from about 4 to about 9. In some cases, the reaction mixture has a pH from about 4 to about 8. In some cases, the reaction mixture has a pH from about 4 to about 7. In some cases, the reaction mixture has a pH from about 4 to about 6. In some cases, the reaction mixture has a pH from about 5 to about 11. In some cases, the reaction mixture has a pH from about 5 to about 10. In some cases, the reaction mixture has a pH from about 5 to about 9. In some cases, the reaction mixture has a pH from about 5 to about 8. In some cases, the reaction mixture has a pH from about 5 to about 7. In some cases, the reaction mixture has a pH from about 5 to about 6. In some cases, the reaction mixture has a pH from about 6 to about 11. In some cases, the reaction mixture has a pH from about 6 to about 10. In some cases, the reaction mixture has a pH from about 7 to about 11. In some cases, the reaction mixture has a pH from about 7 to about 10. In some cases, the reaction mixture has a pH from about 8 to about 11. In some cases, the reaction mixture has a pH from about 8 to about 10. In some cases, the reaction mixture has a pH from about 9 to about 11. In some cases, the reaction mixture has a pH from about 9 to about 10. In some cases, the reaction mixture has a pH of 4. In some cases, the reaction mixture has a pH of 5. In some cases, the reaction mixture has a pH of 6. In some cases, the reaction mixture has a pH of 7. In some cases, the reaction mixture has a pH of 8. In some cases, the reaction mixture has a pH of 9. In some cases, the reaction mixture has a pH of 10. In some cases, the reaction mixture has a pH of 11. In some cases, the reaction mixture has a temperature from about 20 to about 90 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 23 to about 90 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 23 to about 85 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 30 to about 90, from about 30 to about 80, from about 30 to about 70, from about 30 to about 60, or from about 30 to about 50 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 45 to about 55 degrees Celsius (C). In some cases, the reaction mixture has a temperature from about 45 to about 50 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 20 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 23 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 25 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 30 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 35 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 40 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 45 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 50 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 55 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 60 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 70 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 75 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 80 degrees Celsius (C). In some cases, the reaction mixture has a temperature about 85 degrees Celsius (C). In some cases, the buffered solution comprises saline, glycine, iron ions, or any combination thereof. In some cases the buffered solution comprises TRIS, PBS, citrate, monosodium glutamate, or any combination thereof. In some cases, the reaction mixture further comprises an activator co-factor of the non-naturally occurring enzyme. In some cases, the activator co-factor comprises glycine, iron ion, or both. In some cases, the metal is lithium, aluminum, iron, nickel, cobalt, strontium, or a rare earth element. In some cases, the metal is lithium. In some cases, the metal is aluminum. In some cases, the metal is iron. In some cases, the metal is strontium. In some cases, the enzyme having silicase activity is recombinantly produced in a host cell or in a cell-free production system. In some cases, the host cell is a bacterial cell or yeast cell. In some cases, the bacterial cell is *Escherichia coli*, or the yeast cell is Pichia pastoris or Saccharomyces cerevisiae.

[0097] In some cases, the reaction mixture provided herein comprise crushing or grinding the rocks or ores to achieve a particulate size. In some cases, the ground ores comprise a size of about 50 μ m to 1 mm. In some cases, the ground ores comprise a size from about 50 μ m to 750 μ m. In some cases, the ground ores comprise a size from about 50 μ m to 500 μ m. In some cases, the ground ores comprise a size from about 50 μ m to 150 μ m. In some cases, the ground ores comprise a size from about 50 μ m. In some cases, the ground ores comprise a size of about 50 μ m. In some cases, the ground ores comprise a size of about 100 μ m. In some cases, the ground ores comprise a size of about 150 μ m. In some cases, the ground ores comprise a size of about 500 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m. In some cases, the ground ores comprise a size of about 750 μ m.

[0098] In some cases, the reaction mixture provided herein comprise creating a slurry of crushed rock and liquid. In some cases, the rock to liquid ratio is from about 1-40% (w/v). In some cases, the rock to liquid ratio is from about 1-35% (w/v). In some cases, the rock to liquid ratio is from about 1-30% (w/v). In some cases, the rock to liquid ratio is from about 1-25% (w/v). In some cases, the rock to liquid ratio is from about 1-20% (w/v). In some cases, the rock to liquid ratio is from about 1-15% (w/v). In some cases, the rock to liquid ratio is from about 1-10% (w/v). In some cases, the rock to liquid ratio is from about 1-5% (w/v). In some cases, the rock to liquid ratio is from about 10-40% (w/v). In some cases, the rock to liquid ratio is from about 10-35% (w/v). In some cases, the rock to liquid ratio is from about 10-30% (w/v). In some cases, the rock to liquid ratio is from about 10-25% (w/v). In some cases, the rock to liquid ratio is from about 15-35% (w/v). In some cases, the rock to liquid ratio is from about 15-30% (w/v). In some cases, the rock to liquid ratio is from about 20-35% (w/v). In some cases, the rock to liquid ratio is from about 20-30% (w/v). In some cases, the rock to liquid ratio is from about 25-35% (w/v). In some cases, the rock to liquid ratio is from about 25-30% (w/v). In some cases, the rock to liquid ratio is about 1% (w/v). In some cases, the rock to liquid ratio is about 5% (w/v). In some cases, the rock to liquid ratio is about 10% (w/v). In some cases, the rock to liquid ratio is about 15% (w/v). In some cases, the rock to liquid ratio is about 20% (w/v). In some cases, the rock to liquid ratio is about 25% (w/v). In some cases, the rock to liquid ratio is about 30% (w/v). In some cases, the rock to liquid ratio is about 35% (w/v). In some cases, the rock to liquid ratio is about 40% (w/v). [0099] In some cases, the reaction mixture provided herein comprise an enzymatic reaction that proceeds for a set period of time. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-24 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-12 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1-6 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12-24 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24-36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36-72 hours. In some

cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 48-72 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 48-60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 1 hour. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 6 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 12 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 24 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 36 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 48 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 60 hours. In some cases, the reaction mixture comprises the enzymatic reaction proceeding for about 72 hours.

[0100] In some cases, provided herein are engineered enzymes. In some cases, the method of enzyme engineering comprises performing directed molecular evolution on an enzyme sequence and generating an evolved enzyme sequence, wherein the evolved enzyme sequence has a higher specificity to a substrate in a mineral material (e.g., a silicate rock), a higher catalytic rate for acting on the mineral material, or both, compared to the first enzyme sequence. The first enzyme sequence may be a wild-type enzyme. In some cases, the wild-type enzyme is a carbonic anhydrase, a gamma carbonic anhydrase, or an alpha carbonic anhydrase.

[0101] The evolved enzyme, the wild-type enzyme, or both may cleave Si—O bonds in the substrate and extract a metal from the mineral material, in some cases a silicate rock. In some examples, directed molecular evolution is performed using a Machine Learning (ML) or Artificial Intelligence (AI) Algorithm. In some cases, performing directed molecular evolution comprises deoxyribonucleic acid (DNA) shuffling. In some cases, the ML or AI algorithm comprises one or more of structural sequence generation, sequence ranking, and sequence fine-tuning. In some cases, the ML or AI algorithm comprises a transformer model system. In some cases, the ML or AI algorithm comprises using natural language processing (NLP). In some cases, the evolved enzyme sequence is the sequence of the synthetic enzyme used in the methods of any of the embodiments of the present disclosure.

EXAMPLES

Example 1. Methods for Enzymatic Degradation Assay

Materials

[0102] Buffer recipe: 50 micro-Molar (μ M) amino acid activator, 250 micro-Molar (μ M) co-factor, and 0.15 Molar (M) Sodium Chloride (NaCl) diluted in distilled water [0103] Sample comprising mineral material and enzyme in buffer (positive sample): 0.175 grams (g) of mineral material (e.g., ore or rock or other mineral), 7 milliliters (mL) of buffer, 350 microliters (μ L) of purified enzyme (0.51 mg/mL enzyme). The enzyme having silicase activity according to the embodiments and sequences described anywhere in the present disclosure. [0104] Negative control sample: 0.175 g of mineral material, 7350 μ L of buffer.

Procedure

[0105] Mineral samples were crushed and sifted to a grain size of 150 μ M. they were subsequently washed with distilled water and ethanol. Samples were then centrifuged, and the pellets dried for 12 hours at 100° C. 2.5% (weight/volume (w/v)) solutions were made for both the positive sample and the negative control. Samples were resuspended in 96-well flat bottom plates and placed in an OT-2 heater shaker set to 300 rounds per minute (rpm) and a temperature from about 50 to about 60° C., overnight with periodic samples taken at regular time intervals. Samples were degraded and assayed in triplicate. The molybdenum blue photometry method in solution was used to determine the concentration of colloidal silica and soluble silica in the samples as well as for the standard curve. The assays were conducted with an automated procedure on an OT-2 Opentrons and

absorbance taken on a Byuonoy Absorbance 96.

[0106] Positive reaction rates minus the negative baseline rates were calculated and normalized based on an established silica concentration reference curve.

Results

- [0107] The method was performed on a variety of mineral materials, and the results are present in FIGS. **2-7**.
- [0108] FIG. **2** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals(Alpha Spodumene and Beta Spodumene) using an enzyme having silicase activity according to the embodiments of the present disclosure.
- [0109] FIG. **3** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (iron ore, platinum group metal (PGM) tailing, and Bauxite) using an enzyme having silicase activity according to the embodiments of the present disclosure.
- [0110] FIG. **4** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (Rhyolite and Olivine) using an enzyme having silicase activity according to the embodiments of the present disclosure.
- [0111] FIG. **5** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (Hectorite mix, Clay, a silicate named Maverick source, and a Lepidolite) using an enzyme having silicase activity according to the embodiments of the present disclosure.
- [0112] FIG. **6** presents the rate of production of Si(OH).sub.4 corresponding to reaction rates of degrading silicate minerals (crushed glass and Perlite) using an enzyme having silicase activity according to the embodiments of the present disclosure.
- [0113] FIG. 7 presents the rate of production of Si(OH)4 corresponding to reaction rates of degrading silicate minerals (Oil Shale and Fly Ash) using an enzyme having silicase activity according to the embodiments of the present disclosure.
- Example 2. Inosilicate Degradation with Enzymes Having Silicase Activity
- [0114] The enzyme Gamma Carbonic Anhydrase from *M. thermophila* degrades silicate mineral material which allows for the extraction of metals such as lithium and aluminum, for example. Enzymes having silicase activity were designed from gamma carbonic anhydrase to improve the efficiency of the degradation reaction of silicate mineral material, specifically inosilicates, such as alpha spodumene, augite, and tremolite.
- [0115] The 17 amino acid residue signal sequence as well as part of the disordered region (about residues 18 to 64, starting from the N-terminus) of wildtype Gamma Carbonic Anhydrase from *M. thermophila* were truncated. The wildtype truncation is as shown in SEQ ID NO: 403. From there, further mutations were generated to optimize for enzymatic efficacy, as shown in SEQ ID NOs: 404-433. Table 3 shows the percentage identity, as calculated by comparing sequence information using the advanced BLAST computer program, of the enzymes having silicase activity compared to wildtype gamma carbonic anhydrase from *M. thermophila*.
- TABLE-US-00003 TABLE 3 Percentage Identity of Enzymes Having Silicase Activity to Wildtype Gamma Carbonic Anhydrase % Identity to Gamma Carbonic Anhydrase from *M. thermophila* (SEQ ID SEQ ID NO: NO: 1) 403 100 404 65.0 405 67.8 406 54.7 407 61.1 408 65.4 409 73.6 410 71.5 411 67.6 412 66.5 413 64.3 414 58.0 415 69.5 416 72.8 417 46.1 418 70.5 419 51.7 420 38.3 421 60.1 422 71.6 423 67.1 424 50.0 425 58.9 426 68.9 427 58.2 428 74.3 429 42.9 430 69.3 431 58.7 432 46.3 433 67.5
- [0116] Enzymes having silicase activity of SEQ ID NOs: 403-433 were generated comprising a GST tag and expressed in *E. coli* BL21 using a pGEX-6P1 GST codon-optimized vector. *E. coli* cultures were grown at 37° C. with shaking and induced. After induction, the cells were harvested by centrifugation and lysed. The lysate was supplemented with ferrous gluconate to provide a source of Fe. The lysate was heated to 55° C. to aide in protein folding and stability. Enzymes were generated comprising an N-terminal GST tag. Other common purification tags, such as NEXT or 6-

HIS, or no tags may be utilized in purification and experimental use of the enzymes. The enzymes were purified using a GST affinity column. The GST tag was found to not affect enzymic activity under reaction conditions.

[0117] The degradation reactions were tested for the various enzymes having silicase activity with alpha spodumene, augite, or tremolite. Degradation reactions were conducted in plastic containers to avoid silicate dissolution from glass. A range of reaction conditions were tested. The reaction was operational in a buffer comprising TRIS, PBS, 0.1 M citrate, or 0.9% monosodium glutamate in a pH range of 4-11, the rock to liquid rock ratio was tested between 1-40% (w/v), the minerals were crushed with a small grinder to ground ore sizes between 50 µm to 1 mm, the reaction was shaken with a range of 1-220 RPM, at a temperature between 23-85° C., the enzymes having silicase activity of SEQ ID NOs: 403-433 with GST, NEXT, 6-His or no N-terminal tag were tested, Zn, Fe, Cu, or Co metal cofactors were included, and the reaction ran for a time range of 1-48 hours. The range of reaction conditions tested resulted in degradation of silicate material. Data not shown.

[0118] Optimal reaction conditions were developed and used to compare the enzymes having silicase activity of SEQ ID NOs: 403-433. The enzymes were tested with alpha spodumene, augite, and tremolite. Optimized reaction conditions were found to be the following: the minerals were crushed with a small grinder and sifted to achieve a particulate size between 50 and 150 μ m. The reaction proceeded in a 0.1 M TRIS buffer at pH 10 in a rock to liquid rock ration of 30% (w/v) at a temperature of 51° C. and shaking at 220 RPM for 48 hours. Reactions were completed in triplicate. After the reaction proceeded for 48 hours, the suspensions were centrifuged at 14,000 RPM for 15 minutes. The supernatant was filtered using a 45 μ m, 13 mm diameter syringe filter. [0119] The elemental composition of the supernatant was determined using a Laser Induced Breakdown Spectroscopy (LIBS) lithium brine analyzer and X-ray fluorescence analyzer. The results of the extracted metals were measured as PPM in solution as shown in Table 4. Degradation results were normalized to compared to a truncated wildtype Gamma Carbonic Anhydrase from *M. thermophila*, SEQ ID NO: 403.

[0120] The hydrolytic activity of the enzymes having silicase activity on the crystalline structure was also assessed using a silica degradation assay. The assay measured the amount of free silica (Si(OH).sub.4, orthosilicic acid) using a molybdenum blue assay. The reaction proceeded as described above. 100 μL of the supernatant was deposited into a 96-well plate. 10 μL 1:1 sulfuric acid solution was added to the sample and mixed with the sample. 20 μL of 5% ammonium molybdate solution was added to the acidified sample and allowed to rest. Next, 20 μL of 0.5% ascorbic acid reducing reagent was added to the mixture and allowed the color to develop for 10 minutes. The molybdenum blue assay reaction mixture was transferred to a microcuvette and the absorbance was measured at 810 nm. The amount of free orthosilicic acid was calculated against a calibrated standard curve The results of the degradation of enzymes having silicase activity of SEQ ID NO: 403 as shown in Table 5. The results show superior extraction of Fe, Li and Al as well as enzyme degradation activity in the enzymes having silicase activity of SEQ ID NOs: 404-433 as compared to the designed truncated wildtype enzyme having silicase activity of SEQ ID NO: 403.

TABLE-US-00004 TABLE 4 Metal Extraction Reaction Results Degradation activity compared to truncated SEQ ID NO: wildtype enzyme Degradation of alpha spodumene 403 1 404 2.3 405 4.6 406 1.6 407 3.5 408 1.7 409 1.5 410 2.1 411 2 412 1.5 413 1.1 Degradation of augite 403 1 414 3.9 415 1.4 416 5 417 1.5 418 3.4 419 1.3 420 6.9 421 2.1 422 1.5 423 3.2 Degradation of tremolite 403 1 424 2.1 425 1.2 426 1.2 427 1.8 428 1.4 429 4.2 430 1.2 431 3.2 432 2.5 433 2 TABLE-US-00005 TABLE 5 Degradation Activity of Enzymes Having Silicase Activity SEQ ID NO: Fe (PPM) Li (PPM) Al (PPM) Metal extraction of alpha spodumene 403 29177 2500 29744 404 33654 3128 35912 405 58434 4970 71002 406 31598 2774 33592 407 51460 4350 62887 408

32134 2802 34201 409 31025 2659 32378 410 33380 3020 35312 411 33014 2982 34789 412

30896 2640 32157 413 29756 2536 30298 Metal extraction of augite 403 21072 2877 31841 414 41234 5604 62197 415 23918 3265 36029 416 51043 6820 77102 417 24368 3321 36901 418 39692 5382 59748 419 22996 3104 34219 420 68129 9072 102432 421 30112 4098 45412 422 24560 3350 37129 423 37850 5137 57092 Metal extraction of tremolite 403 27594 29727 31841 424 31267 33564 39753 425 28345 30218 32053 426 28409 30301 32198 427 30567 32674 31252 428 29123 31089 34506 429 35981 38374 45098 430 28390 30296 39587 431 34012 36102 39584 432 32878 34906 38128 433 31856 33870 37656

Example 3. Inosilicate Degradation with Host Cells Expressing Enzymes Having Silicase Activity [0121] The enzyme Gamma Carbonic Anhydrase from *M. thermophila* degrades silicate mineral material which allows for the extraction of metals such as lithium and aluminum, for example. Enzymes having silicase activity are designed to improve the efficiency of the degradation reaction of silicate mineral material, specifically inosilicates, such as alpha spodumene, augite, and tremolite.

[0122] Enzymes having silicase activity are generated comprising a tag and expressed in a bacteria host cell, such as *E. coli*, using a host cell appropriate vector. Host cell cultures are grown and induced. After induction, the cells are harvested.

[0123] The degradation reactions are tested using the host cells expressing the enzymes having silicase activity with inosilicate materials, such as alpha spodumene, augite, and tremolite. Degradation reactions are conducted in plastic containers to avoid silicate dissolution from glass. A range of reaction conditions are tested. The reaction is tested in a buffer comprising TRIS, PBS, 0.1 M citrate, or 0.9% monosodium glutamate in a pH range of 4-11, the rock to liquid rock ratio is tested between 1-40% (w/v), the minerals are crushed with a small grinder to ground ore sizes between 50 μ m to 1 mm, the reaction is shaken with a range of 1-220 RPM, at a temperature between 23-85° C., Zn, Fe, Cu, or Co metal cofactors are included, and the reaction is run for a time range of 1-48 hours.

[0124] The elemental composition of the supernatant is determined using a Laser Induced Breakdown Spectroscopy (LIBS) lithium brine analyzer and X-ray fluorescence analyzer. The results of the extracted metals are measured as PPM in solution.

[0125] The hydrolytic activity of the enzymes on the crystalline structure is also assessed using a silica degradation assay. The assay measures the amount of free silica (Si(OH)4, orthosilicic acid) using a molybdenum blue assay. Supernatant is deposited into a 96-well plate. 1:1 sulfuric acid solution is added to the sample and mixed with the sample. 5% ammonium molybdate solution is added to the acidified sample and allowed to rest. 0.5% ascorbic acid reducing reagent is added to the mixture and allowed the color to develop for 10 minutes. The molybdenum blue assay reaction mixture is transferred to a microcuvette and the absorbance is measured at 810 nm. The amount of free orthosilicic acid is calculated against a calibrated standard curve.

Example 4. Phyllosilicate Degradation with Enzymes Having Silicase Activity [0126] The enzyme gamma carbonic anhydrase from *M. thermophila* degrades silicate mineral material which allows for the extraction of metals such as lithium and aluminum, for example. Enzymes having silicase activity were designed from Gamma Carbonic Anhydrase to improve the efficiency of the degradation reaction of silicate mineral material, specifically phyllosilicates, such as lepidolite, montmorillonite, and muscovite.

[0127] The 17 amino acid residue signal sequence or the 17 amino acid residue signal sequence and part of the disordered region (about residues 18 to 64, starting from the N-terminus) of wildtype Gamma Carbonic Anhydrase from *M. thermophila* were truncated (SEQ ID NO: 403 and SEQ ID NO: 434 respectively). From there, further mutations were generated to optimize for enzymatic efficacy, as shown in SEQ ID NOs: 435-464. Table 6 shows the percentage identity, as calculated by comparing sequence information using the advanced BLAST computer program, of the enzymes having silicase activity compared to wildtype gamma carbonic anhydrase from *M. thermophila*.

TABLE-US-00006 TABLE 6 Percentage Identity of Enzymes Having Silicase Activity to Wildtype Gamma Carbonic Anhydrase % Identity to Gamma Carbonic Anhydrase from *M. thermophila* (SEQ ID SEQ ID NO: NO: 1) 403 100 434 100 435 52.2 436 51.3 437 54.2 438 52.6 439 55.1 440 54.4 441 48.8 442 49.3 443 53.3 444 54.2 445 42.8 446 50.6 447 43.3 448 43.6 449 42.8 450 43.9 451 59.0 452 59.0 453 41.1 454 53.9 455 45.5 456 56.5 457 73.3 458 55.3 459 74.9 460 41.1 461 81.7 462 51.2 463 66.7 464 84.2

[0128] Enzymes having silicase activity of SEQ ID NOs: 403 and 434-464 were generated comprising a GST tag and expressed in *E. coli* BL21 using a pGEX-6P1 GST codon-optimized vector. *E. coli* cultures were grown at 37° C. with shaking and induced. After induction, the cells were harvested by centrifugation and lysed. The lysate was supplemented with ferrous gluconate to provide a source of Fe. The lysate was heated to 55° C. to aide in protein folding and stability. Enzymes were generated comprising an N-terminal GST tag. Other common purification tags, such as NEXT or 6-HIS, or no tags may be utilized in purification and experimental use of the enzymes. The enzymes were purified using a GST affinity column. The GST tag was found to not affect enzymic activity under reaction conditions.

[0129] The degradation reactions were tested for the various enzymes having silicase activity with lepidolite, montmorillonite, and muscovite. Degradation reactions were conducted in plastic containers to avoid silicate dissolution from glass. A range of reaction conditions were tested. The reaction was operational in a buffer comprising TRIS, PBS, 0.1 M citrate, or 0.9% monosodium glutamate in a pH range of 4-11, the rock to liquid rock ratio was tested between 1-40% (w/v), the minerals were crushed with a small grinder to ground ore sizes between 50 μ m to 1 mm, the reaction was shaken with a range of 1-220 RPM, at a temperature between 23-85° C., the enzymes having silicase activity of SEQ ID NOs: 403 and 434-464 with GST, NEXT, 6-His or no N-terminal tag were tested, Zn, Fe, Cu, or Co metal cofactors were included, and the reaction ran for a time range of 1-48 hours. The range of reaction conditions tested resulted in degradation of silicate material. Data not shown.

[0130] Optimal reaction conditions were developed and used to compare the enzymes having silicase activity of SEQ ID NOs: 403 and 434-464. The enzymes were tested with lepidolite, montmorillonite, and muscovite. Optimized reaction conditions were found to be the following: the minerals were crushed with a small grinder and sifted to achieve a particulate size between 50 and 150 μ m. The reaction proceeded in a 0.1 M TRIS buffer at pH 10 in a rock to liquid rock ration of 30% (w/v) at a temperature of 51° C. and shaking at 220 RPM for 48 hours. Reactions were completed in triplicate. After the reaction proceeded for 48 hours, the suspensions were centrifuged at 14,000 RPM for 15 minutes. The supernatant was filtered using a 45 μ m, 13 mm diameter syringe filter.

[0131] The elemental composition of the supernatant was determined using a Laser Induced Breakdown Spectroscopy (LIBS) lithium brine analyzer and X-ray fluorescence analyzer. The results of the extracted metals were measured as PPM in solution as shown in Table 7. Degradation results were normalized to compared to a truncated wildtype Gamma Carbonic Anhydrase from *M. thermophila* (either of SEQ ID NOs: 403 or 434).

[0132] The hydrolytic activity of the enzymes having silicase activity on the crystalline structure was also assessed using a silica degradation assay. The assay measured the amount of free silica (Si(OH).sub.4, orthosilicic acid) using a molybdenum blue assay. The reaction proceeded as described above. 100 μL of the supernatant was deposited into a 96-well plate. 10 μL 1:1 sulfuric acid solution was added to the sample and mixed with the sample. 20 μL of 5% ammonium molybdate solution was added to the acidified sample and allowed to rest. Next, 20 μL of 0.5% ascorbic acid reducing reagent was added to the mixture and allowed the color to develop for 10 minutes. The molybdenum blue assay reaction mixture was transferred to a microcuvette and the absorbance was measured at 810 nm. The amount of free orthosilicic acid was calculated against a calibrated standard curve. The results of the degradation of enzymes were normalized to activity of

a truncated wildtype Gamma Carbonic Anhydrase from *M. thermophila* as shown in Table 8. The results of the degradation using enzymes having silicase activity of SEQ ID NOs: 435-464 were normalized to activity of SEQ ID NOs: 403 or 434 as shown in Table 5. The results show superior extraction of Fe, Li and Al as well as enzyme degradation activity in the enzymes having silicase activity of SEQ ID NOs: 435-464 as compared to the designed truncated wildtype enzyme having silicase activity of SEQ ID NO: 403 and SEQ ID NO: 434.

TABLE-US-00007 TABLE 7 Metal Extraction Reaction Results Degradation activity compared to

SEQ ID NO: SEQ ID NO: 403 (truncated wildtype) Degradation of lepidolite 434 1 435 2 436 2.2 437 1.9 438 1.5 439 1.7 440 1.5 441 1.6 442 1.7 443 1.7 444 1.6 Degradation of montmorillonite 434 1 445 3.6 446 2.4 447 1.2 448 1.1 449 2 450 1.5 451 2.2 452 2.4 453 1.3 454 1.2 Degradation activity compared to SEQ ID NO: SEQ ID NO: 424 (truncated wildtype) Degradation of muscovite 403 1 455 1.7 456 2.8 457 2 458 1.5 459 1 460 3 461 1.7 462 1.4 463 4 464 2.4 TABLE-US-00008 TABLE 8 Degradation Activity of Enzymes Having Silicase Activity SEQ ID NO: Fe (PPM) Li (PPM) Al (PPM) Sr (PPM) Metal extraction of lepidolite 434 21287 3514 24903 N.D. 435 24021 3598 25968 N.D. 436 24341 3678 26245 N.D. 437 23123 3627 25873 N.D. 438 21786 3552 25514 N.D. 439 22259 3642 26031 N.D. 440 21892 3557 25487 N.D. 441 22018 3589 25716 N.D. 442 22230 3638 25955 N.D. 443 22242 3643 26028 N.D. 444 22011 3594 25732 N.D. Metal extraction of montmorillonite 434 21061 450 25105 2194 445 41584 789 55613 4216 446 32235 654 46147 3798 447 23018 478 26695 2389 448 23240 461 26821 2453 449 34012 652 37334 3728 450 24287 634 27568 3560 451 34763 658 32849 3851 452 35047 662 38312 3917 453 23391 592 26743 2329 454 23674 594 27058 2290 Metal extraction of muscovite 403 20364 29624 24959 N.D. 455 21547 31789 28125 N.D. 456 25432 34812 38912 N.D. 457 23489 32950 34968 N.D. 458 22015 30970 27605 N.D. 459 20456 29730 24964 N.D. 460 26710 35890 40025 N.D. 461 21570 31810 28060 N.D. 462 21030 30200 25302 N.D. 463 28020 37250 46390 N.D. 464 24080 33420 32705 N.D. N.D. = no data

Example 5. Phyllosilicate Degradation with Host Cells Expressing Enzymes Having Silicase Activity

[0133] The enzyme Gamma Carbonic Anhydrase from *M. thermophila* degrades mineral material which allows for the extraction of metals such as lithium and aluminum, for example. Enzymes having silicase activity are designed to improve the efficiency of the degradation reaction of silicate mineral material, specifically phyllosilicates, such as lepidolite, montmorillonite, and muscovite. [0134] Enzymes having silicase activity are generated comprising a tag and expressed in a bacteria host cell, such as *E. coli*, using a host cell appropriate vector. Host cell cultures are grown and induced. After induction, the cells are harvested.

[0135] The degradation reactions are tested using the host cells expressing the enzymes with phyllosilicate materials, such as lepidolite, montmorillonite, and muscovite. Degradation reactions are conducted in plastic containers to avoid silicate dissolution from glass. A range of reaction conditions are tested. The reaction is tested in a buffer comprising TRIS, PBS, 0.1 M citrate, or 0.9% monosodium glutamate in a pH range of 4-11, the rock to liquid rock ratio is tested between 1-40% (w/v), the minerals are crushed with a small grinder to ground ore sizes between 50 µm to 1 mm, the reaction is shaken with a range of 1-220 RPM, at a temperature between 23-85° C., Zn, Fe, Cu, or Co metal cofactors are included, and the reaction is run for a time range of 1-48 hours. [0136] The elemental composition of the supernatant is determined using a Laser Induced Breakdown Spectroscopy (LIBS) lithium brine analyzer and X-ray fluorescence analyzer. [0137] The results of the extracted metals are measured as PPM in solution. [0138] The hydrolytic activity of the enzymes on the crystalline structure is also assessed using a

silica degradation assay. The assay measures the amount of free silica (Si(OH).sub.4, orthosilicic acid) using a molybdenum blue assay. Supernatant is deposited into a 96-well plate. 1:1 sulfuric acid solution is added to the sample and mixed with the sample. 5% ammonium molybdate solution is added to the acidified sample and allowed to rest. 0.5% ascorbic acid reducing reagent is added

to the mixture and allowed the color to develop for 10 minutes. The molybdenum blue assay reaction mixture is transferred to a microcuvette and the absorbance is measured at 810 nm. The amount of free orthosilicic acid is calculated against a calibrated standard curve. [0139] While preferred embodiments of the present invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only. Numerous variations, changes, and substitutions will now occur to those skilled in the art without departing from the invention. It should be understood that various alternatives to the embodiments of the invention described herein may be employed in practicing the invention. It is intended that the following claims define the scope of the invention and that methods and structures within the scope of these claims and their equivalents be covered thereby.

Claims

- **1**. A method of extracting a metal from a mineral material, the method comprising: (a) contacting the mineral material with an enzyme having silicase activity under reaction conditions such that the metal contained within the mineral material is solubilized and released; and (b) collecting the released metal, thereby extracting the metal from the mineral material.
- **2**. The method of claim 1, wherein the mineral material comprises an ore, a rock, a natural mineral material, a man-made mineral material, or any combination thereof.
- **3.** The method of claim 1, wherein the mineral material comprises a silicate.
- **4.** The method of claim 1, wherein the mineral material comprises an inosilicate, a phyllosilicate, an amorphous silicate, a tectosilicate, or any combination thereof.
- **5.** The method of claim 1, wherein the enzyme having silicase activity has a sequence identity of at least about 30%, at least about 40%, about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or more with an amino acid sequence of a gamma carbonic anhydrase.
- **6**. The method of claim 1, wherein the enzyme having silicase activity has a sequence identity of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90% or more, with an amino acid sequence of an enzyme selected from the group consisting of: Methanosarcina thermophila gamma carbonic anhydrase, Bacillus licheniformis CG-B52 gamma carbonic anhydrase, Pelobacter carbinolicus gamma carbonic anhydrase, Syntrophus aciditrophicus gamma carbonic anhydrase, *Methanosarcina barkeri* gamma carbonic anhydrase, *Methanosarcina mazei* carbonic anhydrase, *Bacillus halodurans* alpha carbonic anhydrase, Alkalihalobacillus *clausii* (strain KSM-K16) (*Bacillus clausii*) alpha carbonic anhydrase, *Methanosarcina acetivorans* carbonate dehydratase, Kofleriaceae bacterium SLC26A/SulP transporter domain-containing protein, Thermodesulfitimonas *autotrophica* carbonic anhydrase/acetyltransferase-like protein (Isoleucine patch superfamily), *Fischerella* thermalis/Mastigocladus laminosus JSC-11 carboxysome assembly protein CcmM, Thermosynechococcus vestitus BP-1/(Thermosynechococcus elongatus BP-1) carboxysome assembly protein CcmM, *Methanothrix thermoacetophila* carbonate dehydratase, *Thermosyntropha lipolytica* carbonic anhydrase or acetyltransferase, isoleucine patch superfamily, Desulfofundulus thermobenzoicus transferase, Archaeoglobus veneficus carbonate dehydratase, Suberites domuncula carbonic anhydrase.
- 7. The method of claim 1, wherein the enzyme having silicase activity has an amino acid sequence having at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 95%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5%, or at least 99.9%, or more sequence identity to an amino acid sequence of any one of SEQ ID NOS: 1-402.
- **8.** The method of claim 1, wherein the reaction conditions comprise a temperature from about 23 to about 85 degrees Celsius (C).

- **9**. The method of claim 1, wherein the reaction conditions comprise a pH from about 4 to about 11.
- **10**. The method of claim 1, wherein the reaction conditions comprise contacting the enzyme having silicase activity with a co-factor.
- **11**. The method of claim 10, wherein the co-factor is selected from the group consisting of: iron, zinc, copper, nickel, and cobalt.
- **12**. The method of claim 1, wherein the metal is selected from the group consisting of: lithium, aluminum, iron, nickel, cobalt, strontium, and a rare earth element.
- **13**. The method of claim 1, wherein the metal is released into a solution.
- **14.** The method of claim 13, further comprising extracting the metal from the solution.
- **15**. The method of claim 13, further comprising purifying the metal from the solution, thereby generating a purified metal.
- **16**. The method of claim 15, wherein the purified metal has a purity of at least about 80%.
- **17**. The method of claim 1, wherein the method is performed in situ or ex situ.
- **18**. The method of claim 1, wherein the reaction conditions comprise a rock to liquid ratio from about 1-40% (w/v).
- **19**. The method of claim 1, wherein the reaction conditions comprise a buffer.
- **20**. The method of claim 19, wherein the buffer is selected from the group consisting of: TRIS, PBS, citrate, monosodium glutamate, and any combination thereof.