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Leonard et al.

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(54) **ENGINEERED RED BLOOD CELL-BASED
BIOSENSORS**

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U.S.C. 154(b) by 1332 days.

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Feb. 28, 2018, now Pat. No. 10,753,930.

(60) Provisional application No. 62/464,754, filed on Feb.
28, 2017.

(51) **Int. Cl.**

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C12N 5/078 (2010.01)

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G01N 33/80 (2006.01)

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CPC **G01N 33/555** (2013.01); **C12N 5/0641**
(2013.01); **G01N 21/6428** (2013.01); **G01N**
33/566 (2013.01); **C12N 2510/00** (2013.01);
G01N 33/80 (2013.01)

(58) **Field of Classification Search**

None

See application file for complete search history.

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(57)

ABSTRACT

Disclosed are systems and methods for detecting extracel-
lular ligands. The disclosed systems and method for detect-
ing extracellular ligands typically comprise or utilize engi-
neered red blood cells (eRBCs) that comprises modular
extracellular sensors. The eRBCs may comprise: (i) a first
exogenous extracellular sensor; the first extracellular sensor
comprising: a) a ligand binding domain, b) a transmembrane
domain, and c) a first fragment of a functional protein, and
(ii) a second exogenous extracellular sensor; the second
extracellular sensor comprising: a) a ligand binding domain,
b) a transmembrane domain, and c) a second fragment of the
functional protein. In the eRBCs, the ligand binding domain
of the first exogenous sensor and the ligand binding domain
of the second exogenous sensor bind to the same ligand to
form a tertiary complex, and the first fragment of the
functional protein and the second fragment of the functional
protein interact in the tertiary complex to reconstitute func-
tional activity of the functional protein. Suitable functional
proteins for the disclosed eRBCs may include fluorescent
proteins that emit fluorescence when the ligand binding
domain of the first exogenous sensor and the ligand binding
domain of the second exogenous sensor bind to the same
ligand, and enzymatic proteins that exhibit enzymatic activ-
ity when ligand binding domain of the first exogenous
sensor and the ligand binding domain of the second exog-
enous sensor bind to the same ligand.

19 Claims, 13 Drawing Sheets

Specification includes a Sequence Listing.

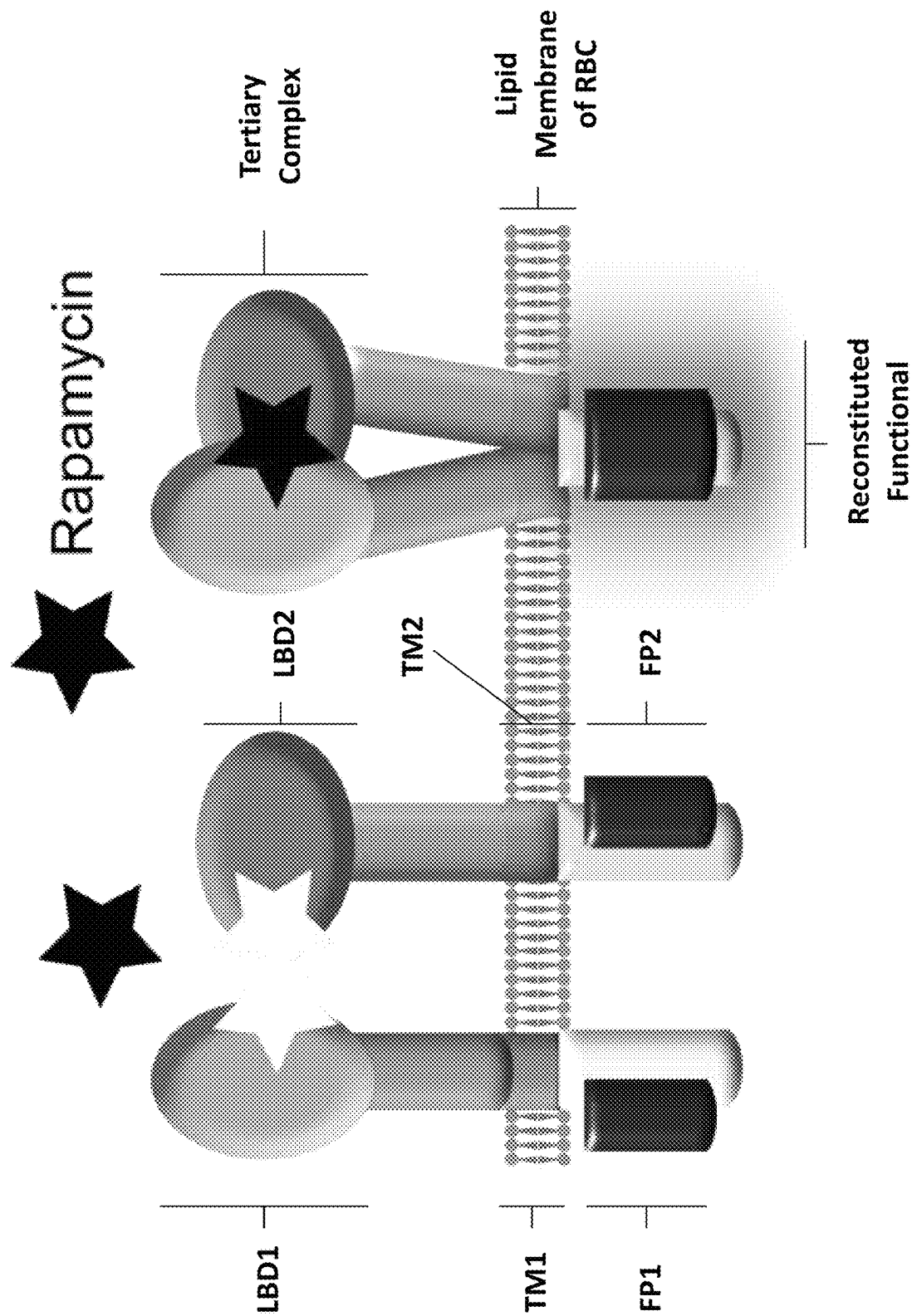


Figure 1

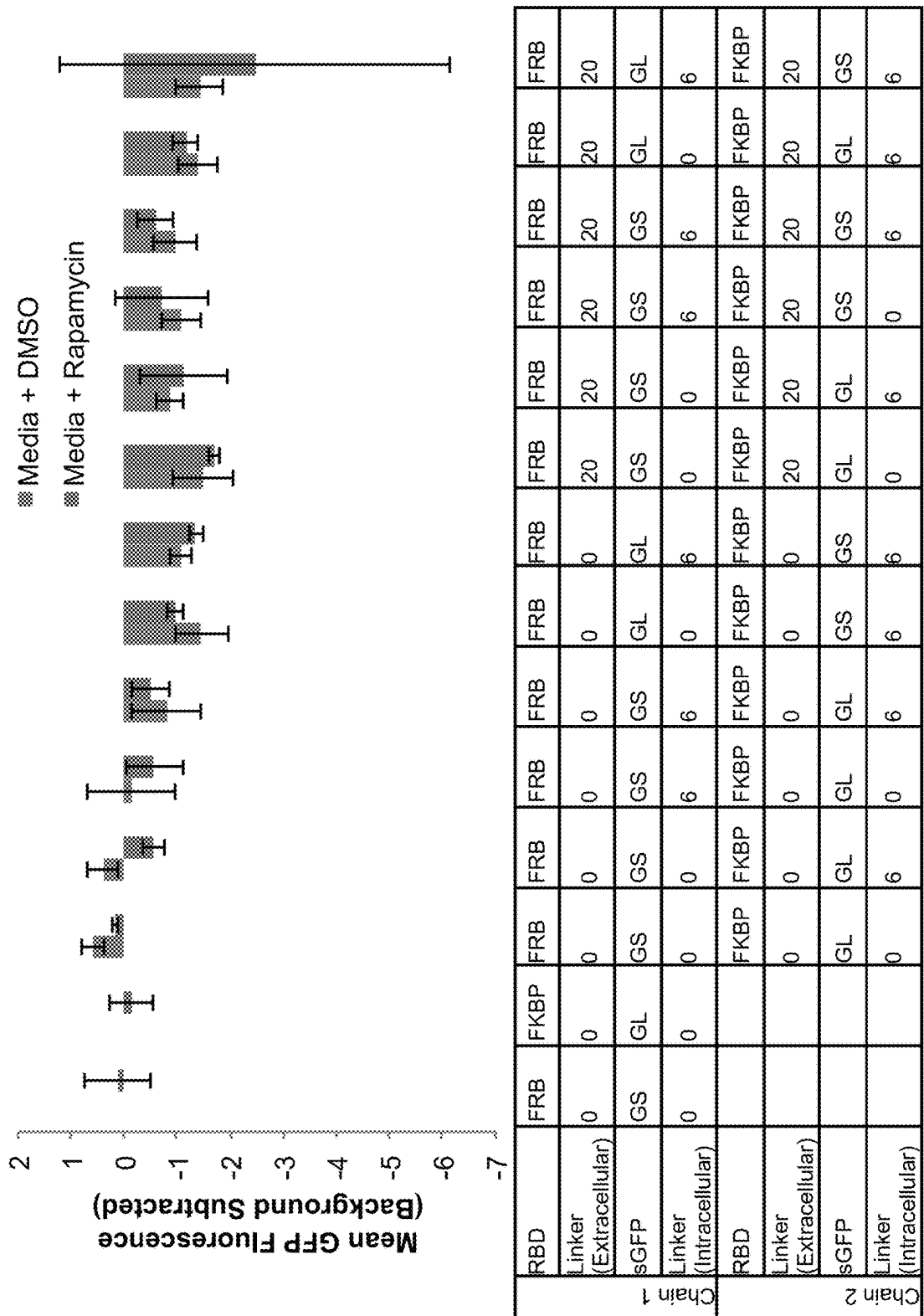


Figure 2

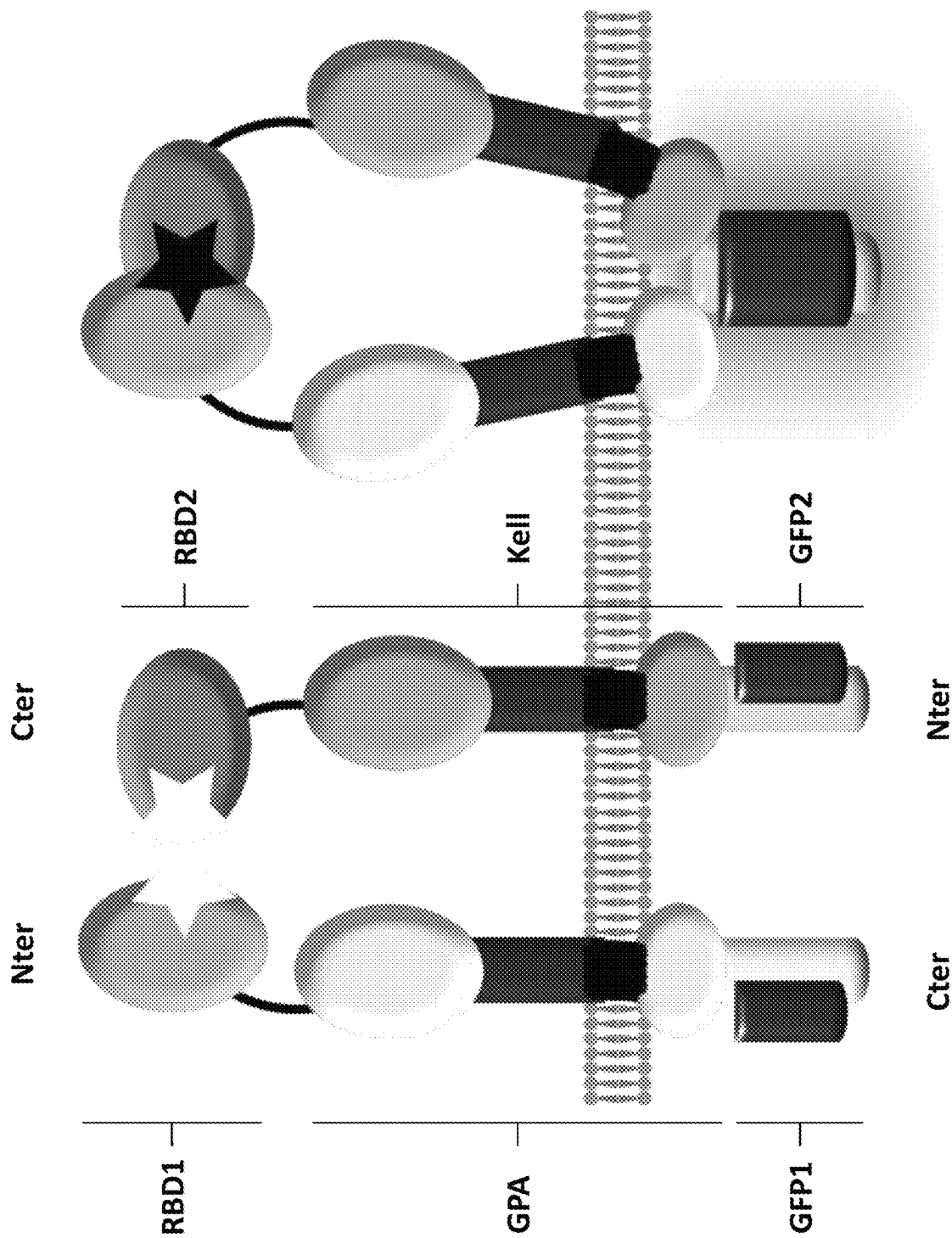


Figure 3

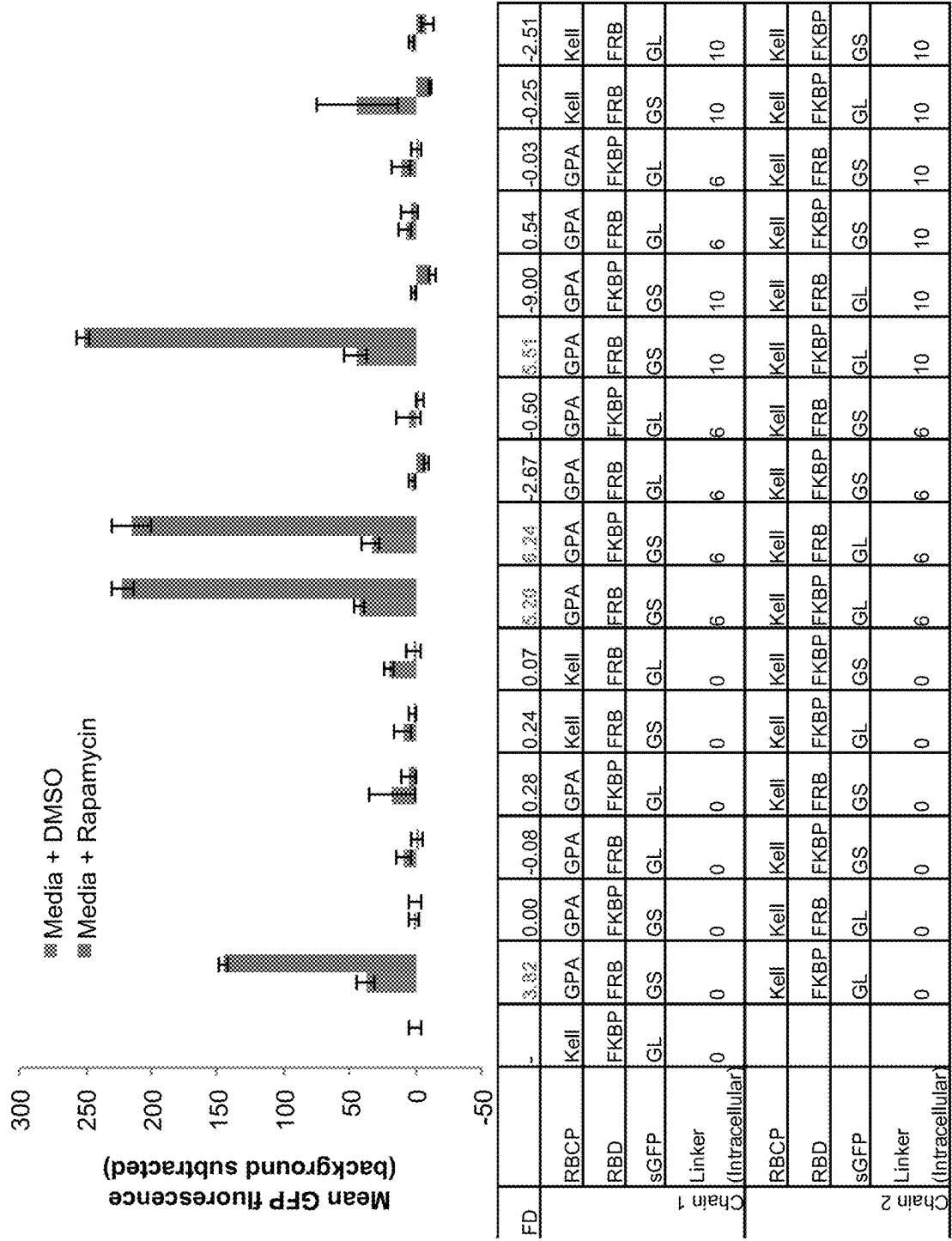


Figure 4

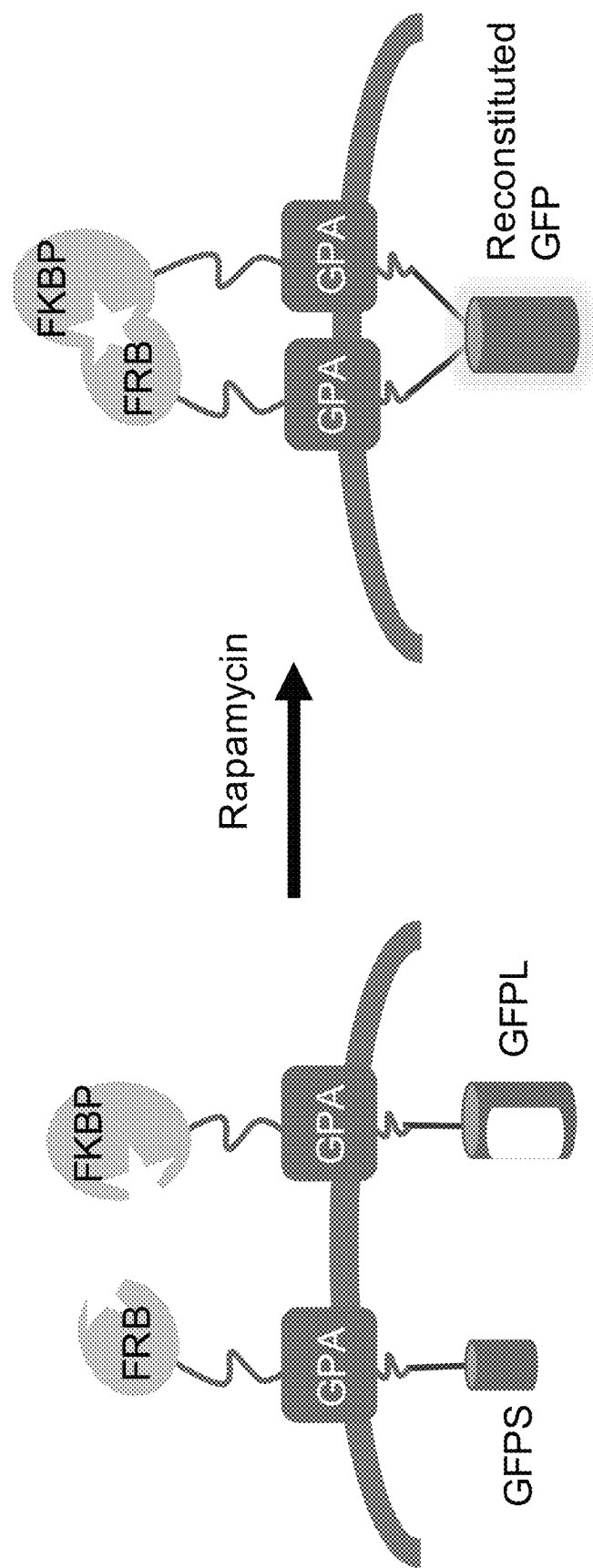


Figure 5

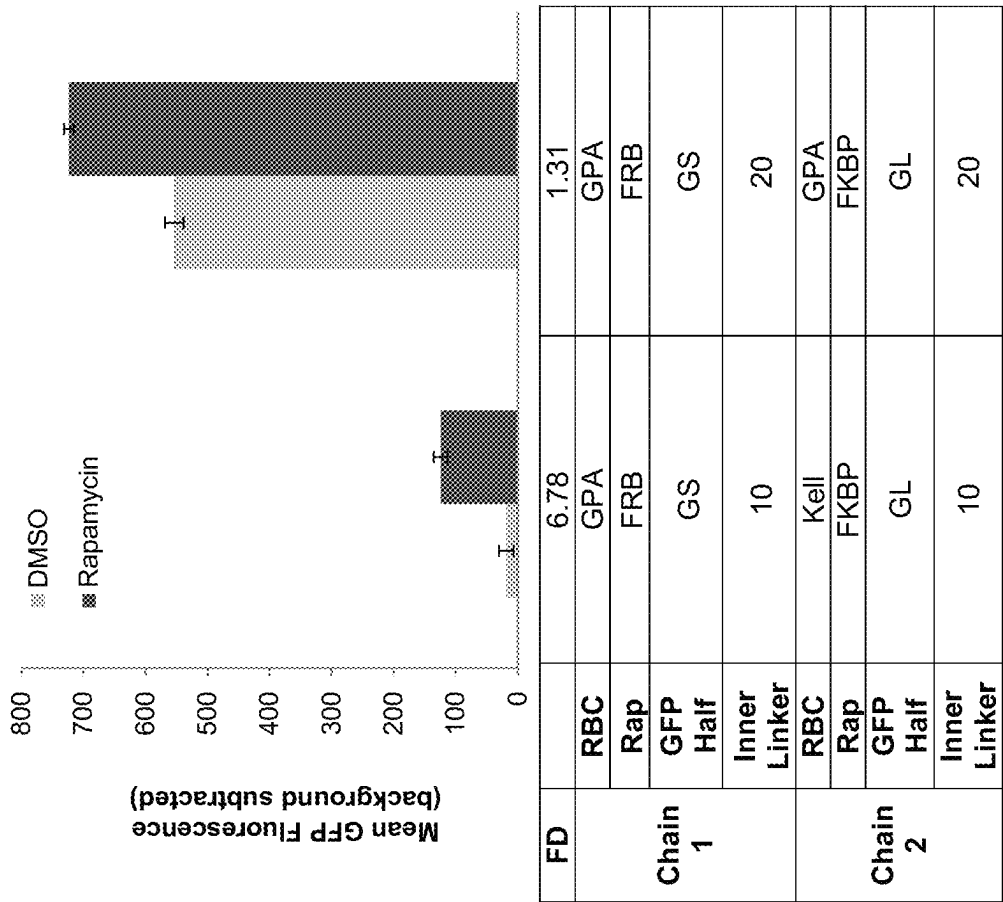


Figure 6

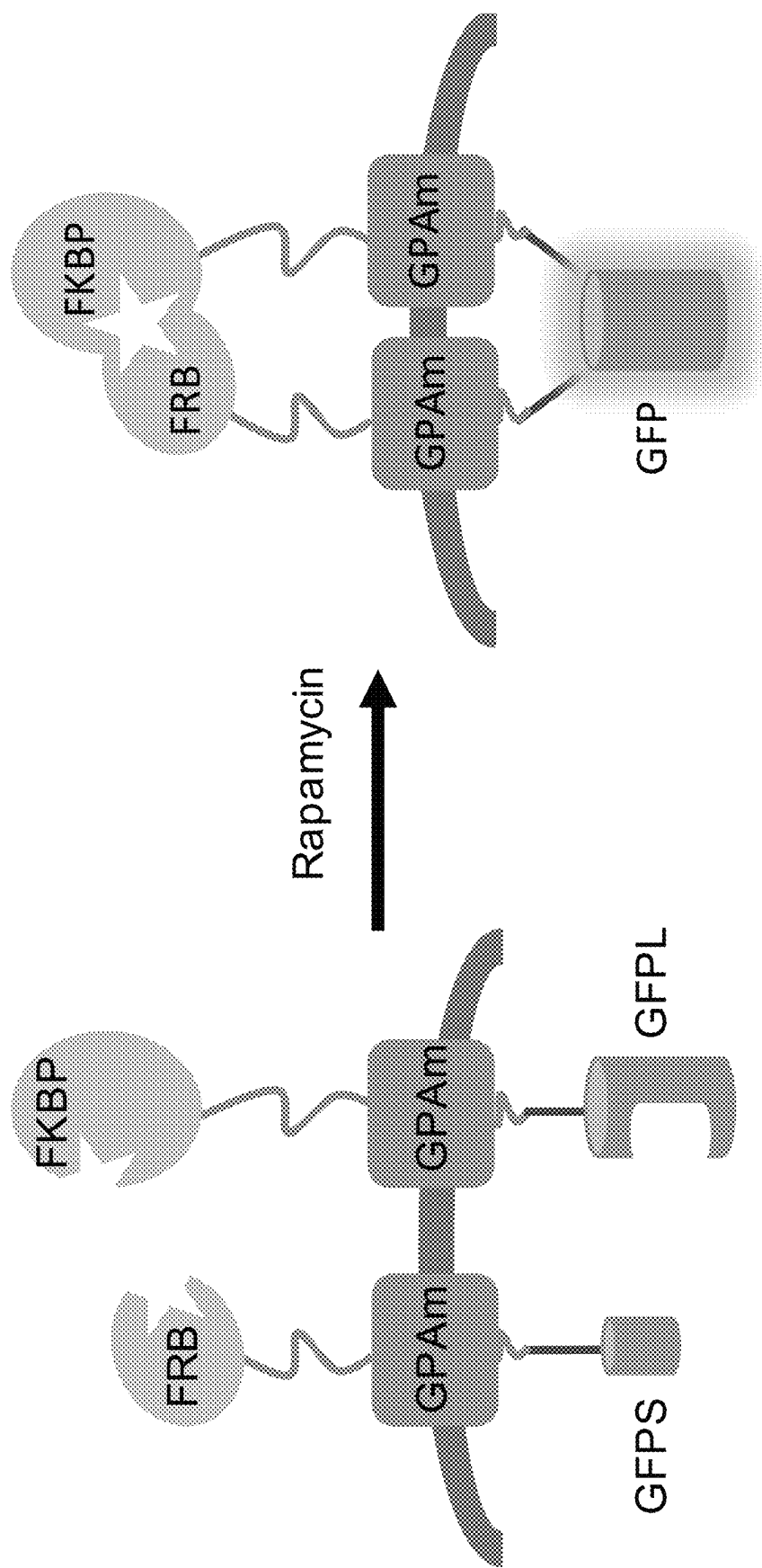


Figure 7

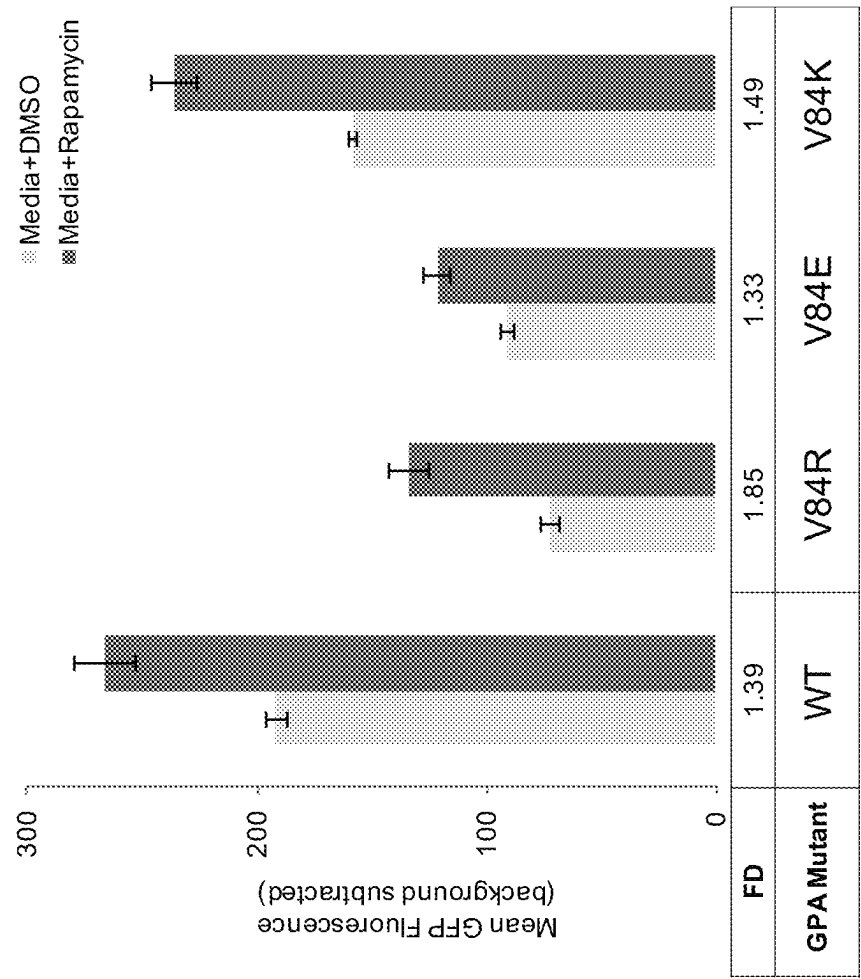
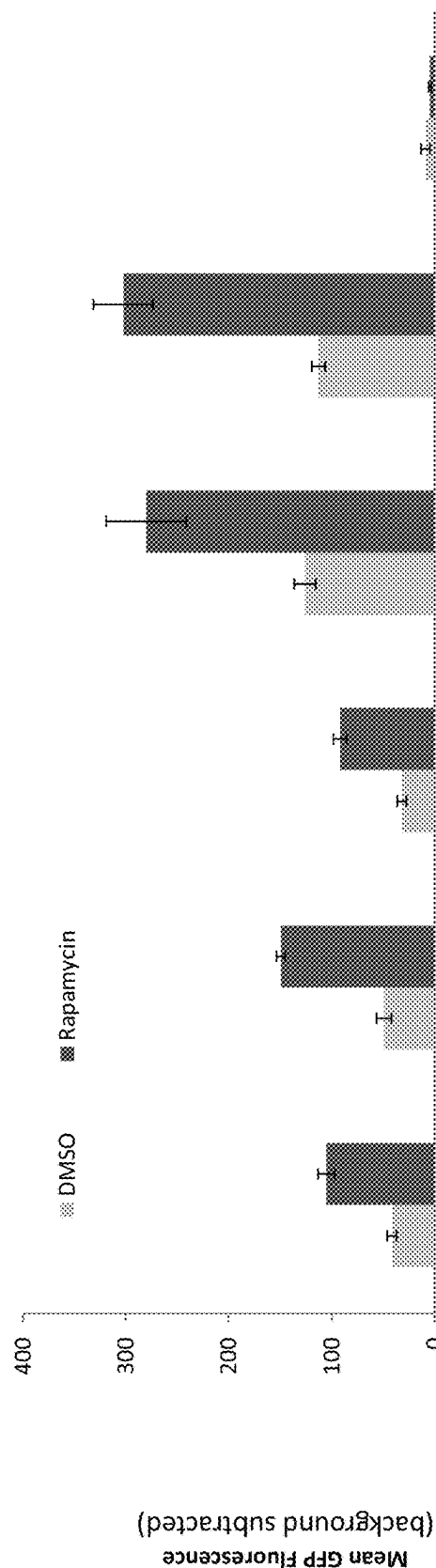


Figure 8



FD		2.53	3.03	2.88	2.22	2.68	0.52
Chain 1	RBC	GPA	GPA	GPA	GPA	GPA	GPA
	Mutate	X	X	V84R	V84R	V84R	V84R
	Rap	FRB	FRB	FRB	FRB	FRB	FRB
	GFP Half	GS	GS	GS	GS	GS	GS
	Inner Linker	6	10	10	10	20	20
Chain 2	RBC	Kell	Kell	Kell	GPA	GPA	Kell
	Mutate	X	X	X	V84R	V84R	X
	Rap	FKBP	FKBP	FKBP	FKBP	FKBP	FKBP
	GFP Half	GL	GL	GL	GL	GL	GL
	Inner Linker	6	10	10	10	20	10

Figure 9

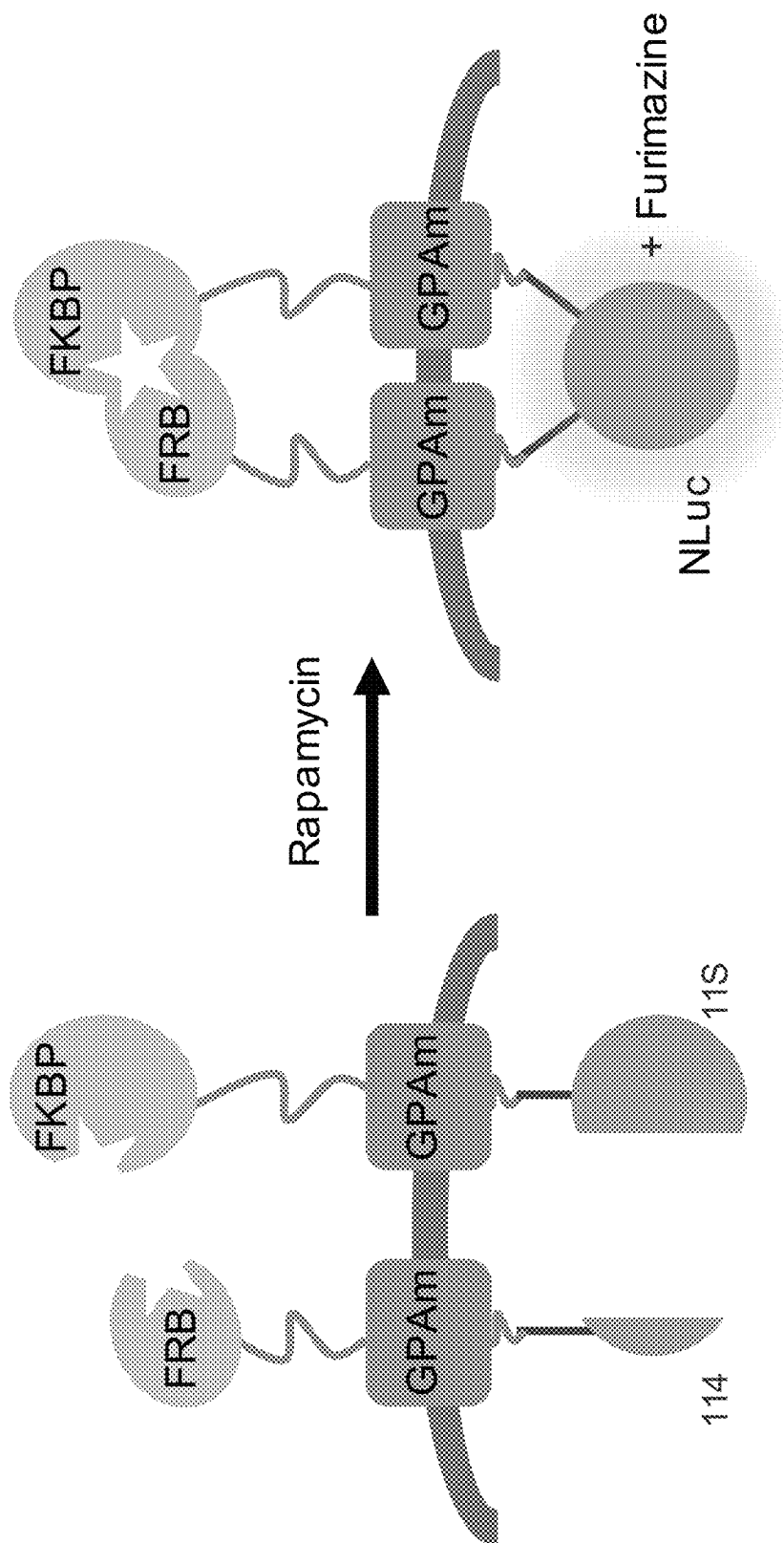
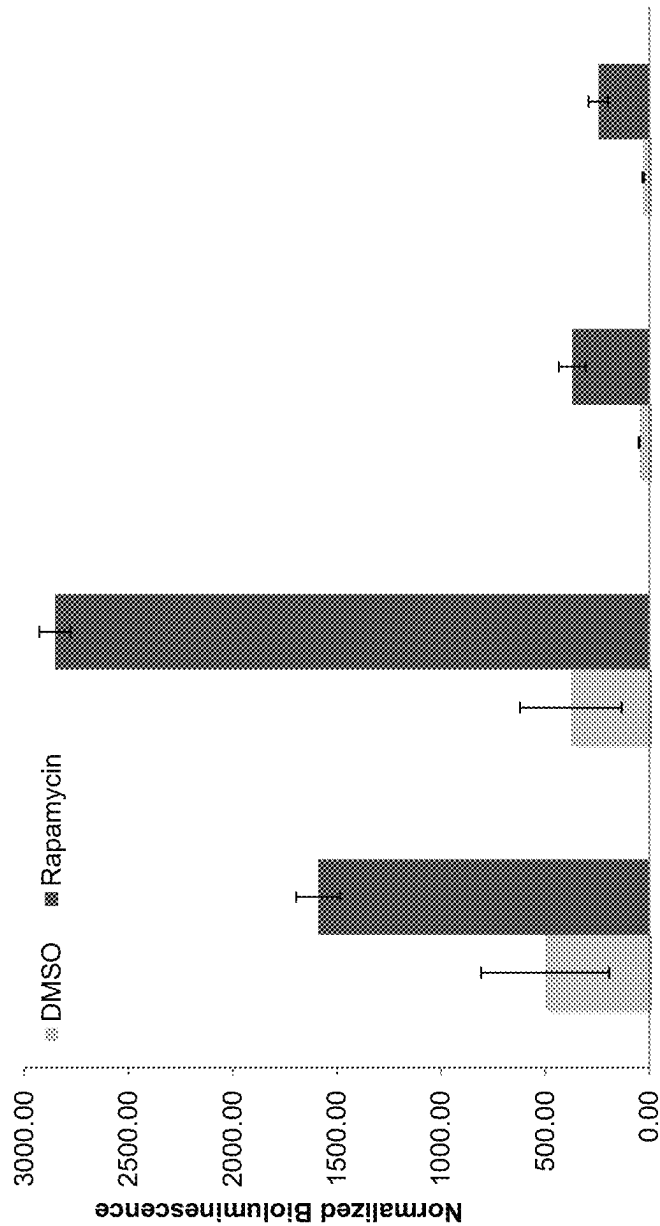
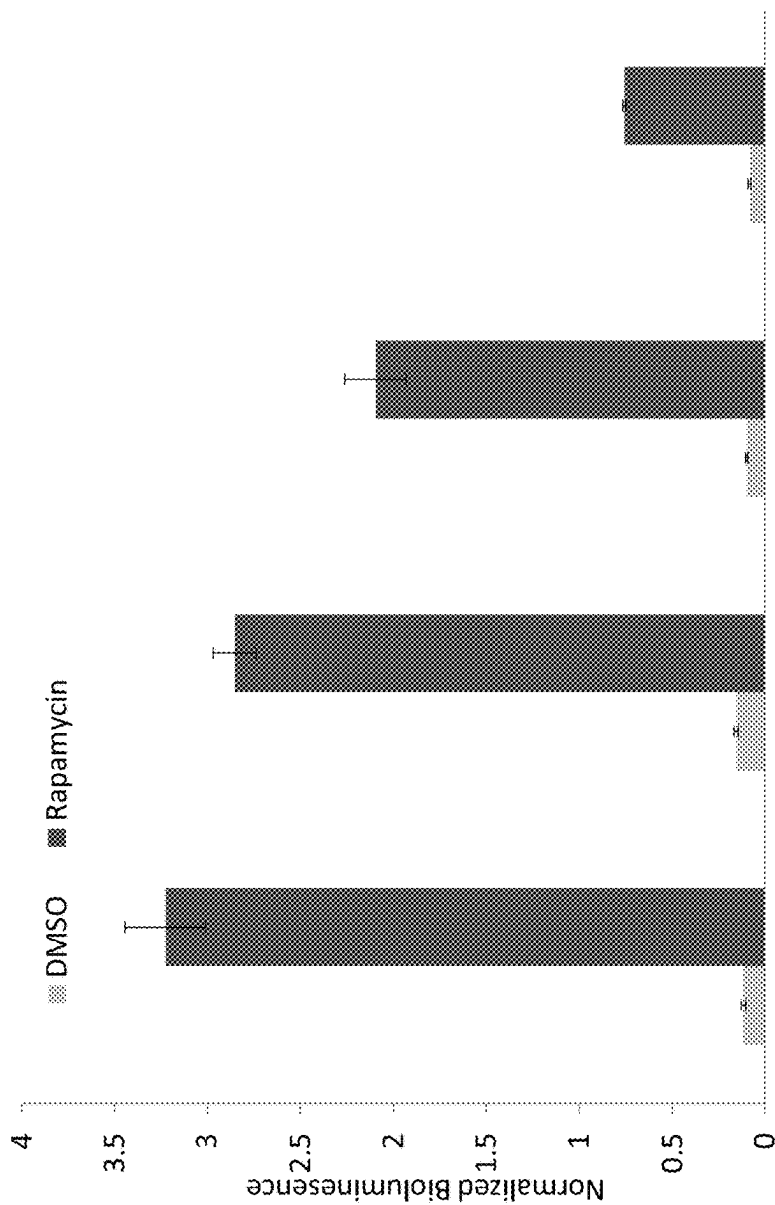


Figure 10



FD	3.18	7.57	7.46	8.33
GPA Mutant	X	X	V84R	V84R
Chain 1	11S	114	11S	114
Chain 2	114	11S	114	11S

Figure 11



FD	27.30	18.14	21.08	9.24
Condition	Ethanol	β -estradiol	Ethanol	β -estradiol
Time Point	24 hours	24 hours	48 hours	48 hours

Figure 12

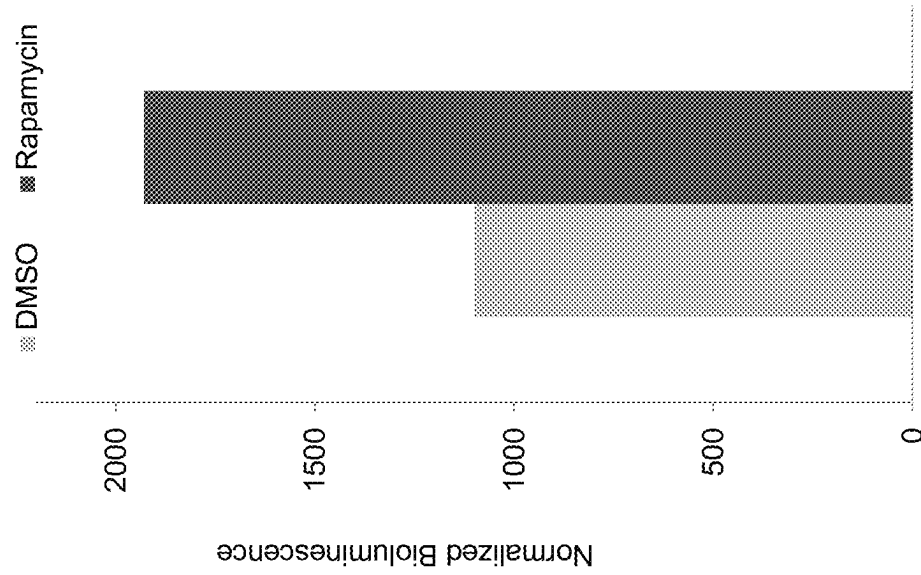


Figure 13

1

**ENGINEERED RED BLOOD CELL-BASED
BIOSENSORS****CROSS-REFERENCE TO RELATED PATENT
APPLICATIONS**

The present application is a Continuation of U.S. application Ser. No. 15/908,077, filed Feb. 28, 2018, which claims the benefit of priority under 35 U.S.C. § 119(e) to U.S. Provisional Application No. 62/464,754, filed on Feb. 28, 2017, the content of which is incorporated herein by reference in its entirety.

**STATEMENT REGARDING FEDERALLY
SPONSORED RESEARCH OR DEVELOPMENT**

This invention was made with government support under 11-23-CCM-DT-FP-008 awarded by The Defense Advanced Research Projects Agency (DARPA). The government has certain rights in the invention

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted in TXT format and is hereby incorporated by reference in its entirety. Said TXT copy, created on Feb. 20, 2025, is named 121384-0147 SL.txt and is 10,151 bytes.

BACKGROUND

The present invention provides modular extracellular sensors, nucleic acids encoding such sensors, cells expressing such sensors, systems and methods of employing such sensors and cells for detecting extracellular ligands. In particular, the invention relates to red blood cells that have been engineered to express modular extracellular sensors for detecting extracellular ligands.

SUMMARY

Disclosed are systems and methods for detecting extracellular ligands. The disclosed systems and method for detecting extracellular ligands typically comprise or utilize engineered red blood cells (eRBCs) that comprises modular extracellular sensors. The eRBCs may comprise: (i) a first exogenous extracellular sensor; the first extracellular sensor comprising: a) an extracellular ligand binding domain or a portion thereof, b) a transmembrane domain, and c) a first fragment of a functional protein, and (ii) a second exogenous extracellular sensor; the second extracellular sensor comprising: a) an extracellular ligand binding domain or a portion thereof, b) a transmembrane domain, and c) a second fragment of the functional protein. In the eRBCs, the extracellular ligand binding domain of the first exogenous sensor and the ligand binding domain of the second exogenous sensor bind to a ligand to form a ternary complex (which may be the same ligand), and the first fragment of the functional protein and the second fragment of the functional protein interact in the ternary complex to reconstitute functional activity of the functional protein.

The first exogenous sensor and/or the second exogenous sensor may be derived, prepared, and/or engineered from native membrane proteins of RBCs or portions or fragments thereof. For example, the first exogenous sensor and/or the second exogenous sensor may be derived, prepared, and/or engineered from a portion of a native membrane protein of

2

a RBC comprising at least the transmembrane portion to which is fused a ligand binding domain at the extracellular terminus and a functional protein (or a portion of a functional protein) at the intracellular terminus. The transmembrane portion further may be mutated in order to prevent homodimerization of the transmembrane portion with another transmembrane portion, for example, where the native membrane protein form a homodimer in native conditions.

Suitable functional proteins for the disclosed eRBCs may include fluorescent proteins that emit fluorescence when the ligand binding domain of the first exogenous sensor and the ligand binding domain of the second exogenous sensor bind to the same ligand, and enzymatic proteins that exhibit enzymatic activity when ligand binding domain of the first exogenous sensor and the ligand binding domain of the second exogenous sensor bind to the same ligand. Substrates for the enzymatic proteins may include substrates that are luminescent after they are metabolized by the enzymatic protein.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1: Proposed mechanism of protein-based biosensors.

FIG. 2: Evaluation of first-generation biosensor proteins containing a simple transmembrane domain. All samples were analyzed in biological triplicate, the mean was subtracted from the mean of negative control cells (expressing only one receptor chain), and error bars represent one standard deviation. Abbreviations: RBD: Rapamycin Binding Domain—FRB or FKBP; sGFP: split GFP—GS (small fragment) or GL (large fragment)

FIG. 3: Proposed mechanism of Kell-based biosensor protein and GPA-based biosensor protein. Kell and GPA, two native RBC proteins, were modified to contain extracellular rapamycin-binding domains and intracellular split GFP halves.

FIG. 4: Evaluation of Kell-based biosensor protein and GPA-based biosensor protein. All samples were analyzed in biological triplicate, the mean was subtracted from the mean of cells transfected with only one receptor half, and error bars represent one standard deviation. Abbreviations: RBD: Rapamycin Binding Domain—FRB or FKBP; sGFP: split GFP—GS (small fragment) or GL (large fragment); RBC: Red Blood Cell Protein.

FIG. 5: Proposed mechanism of GPA only-based biosensor proteins.

FIG. 6: Evaluation of GPA only-based biosensor proteins compared to GPA-based and Kell-based biosensor proteins. All samples were analyzed in biological triplicate, the background auto-fluorescence of the cells with only the color control was subtracted off, and error bars represent one standard deviation. Abbreviations: Rap: Rapamycin Binding Domain—FRB or FKBP; GFP Half: split GFP—GS (small fragment) or GL (large fragment); RBC: Red Blood Cell Protein.

FIG. 7: Second generation proposed mechanism of mutated GPA (GPAm)-based biosensor proteins. Abbreviations: Rap: Rapamycin Binding Domain—FRB or FKBP; GFP Half: split GFP—GS (small fragment) or GL (large fragment); RBC: Red Blood Cell Protein.

FIG. 8: Mutation-driven improvement of eRBC biosensors. All samples were analyzed in biological triplicate, the mean was subtracted from the mean of negative control cells, and error bars represent one standard deviation. Abbreviations: FD: fold-difference; GPA Mutant: WT: Wild Type,

V84R: Valine 84 to Arginine, V84E: Valine 84 to Glutamic Acid, V84K: Valine 84 to Lysine.

FIG. 9: Evaluation of mutated GPA-based biosensor proteins compared to GPA-based and Kell-based biosensor proteins. All samples were analyzed in biological triplicate, error bars represent one standard deviation. Abbreviations: Rap: Rapamycin Binding Domain—FRB or FKBP; GFP Half: split GFP—GS (small fragment) or GL (large fragment); RBC: Red Blood Cell Protein; Mutate: X—wild type or V84R: Valine 84 to Arginine mutation.

FIG. 10: Third generation proposed mechanism of mutated GPA-based biosensor proteins.

FIG. 11: Evaluation of GPA-based biosensor proteins compared to mutated GPA-based biosensor proteins with NANOLUCIFERASE™ output. All samples were analyzed in biological triplicate, error bars represent one standard deviation. Abbreviations: GPA mutant—X: wild type GPA, V84R: Valine 84 to Arginine; NANOLUCIFERASE™ Half—11S (larger half) or 114 (smaller half); FD—fold difference.

FIG. 12: Evaluation of GPA-based biosensor proteins with NANOLUCIFERASE™ output in Gler cells, which approximate red blood cells. Abbreviations: FD—fold difference.

FIG. 13: Evaluation of GPA-based biosensor proteins with NANOLUCIFERASE™ output in human CD34+ cells.

DETAILED DESCRIPTION

The present invention is described herein using several definitions, as set forth below and throughout the application.

Unless otherwise specified or indicated by context, the terms “a”, “an”, and “the” mean “one or more.” For example, “a receptor,” “ligand,” and “complex” should be interpreted to mean “one or more receptors,” “ligands,” and “complexes,” respectively.

As used herein, “about,” “approximately,” “substantially,” and “significantly” will be understood by persons of ordinary skill in the art and will vary to some extent on the context in which they are used. If there are uses of these terms which are not clear to persons of ordinary skill in the art given the context in which they are used, “about” and “approximately” will mean plus or minus ?10% of the particular term and “substantially” and “significantly” will mean plus or minus >10% of the particular term.

As used herein, the terms “include” and “including” have the same meaning as the terms “comprise” and “comprising” in that these latter terms are “open” transitional terms that do not limit claims only to the recited elements succeeding these transitional terms. The term “consisting of,” while encompassed by the term “comprising,” should be interpreted as a “closed” transitional term that limits claims only to the recited elements succeeding this transitional term. The term “consisting essentially of,” while encompassed by the term “comprising,” should be interpreted as a “partially closed” transitional term which permits additional elements succeeding this transitional term, but only if those additional elements do not materially affect the basic and novel characteristics of the claim.

The disclosed technology relates to “extracellular sensors.” As disclosed herein, an “extracellular sensor” is a molecule or a system of molecules that can be used to bind to a ligand and provide a detectable response based on binding the ligand. Extracellular sensors are disclosed in the art. (See, e.g., Daringer et al., “Modular Extracellular Sensor Architecture for Engineering Mammalian Cell-based

Devices,” Nichole M. Daringer, Rachel M. Dudek, Kelly A. Schwarz, and Josh N. Leonard, ACS Synth. Biol. 2014, 3, 892-902, published Feb. 25, 2014; WO 2013/022739, published on Feb. 14, 2013; and U.S. Publication No. 2014-0234851; the contents of which are incorporated herein by reference in their entirety).

The disclosed extracellular sensors typically include a ligand-binding domain of a ligand-binding protein. As contemplated herein, a “ligand-binding protein” is a macromolecule, typically a protein, which binds to a ligand. For example, a ligand-binding protein may include a receptor for a ligand or a portion of a receptor for a ligand, for example, where the receptor is a membrane protein and the ligand-binding protein comprises the extracellular portion of the receptor that binds an extracellular ligand. A suitable ligand for the ligand-binding domains of the disclosed extracellular sensors may include more than one binding site for a ligand-binding protein. As such, a suitable ligand can bind more than one ligand-binding domain of one or more extracellular sensors as contemplated herein, and as such, a suitable ligand may form a ternary or high order complex with two or more extracellular sensors.

The disclosed exogenous sensors may be utilized for sensing an extracellular ligand and providing a molecular signal when the ligand is sensed. Suitable molecular signals may be generated via so-called “bimolecular complementation” that occurs between two or more exogenous sensors (or between portions of two or more exogenous sensors) when the ligand is sensed. “Bimolecular complementation” is known in the art, for example, bimolecular fluorescence complementation and split enzyme complementation or protein-fragment complementation are known in the art. (See Kodama et al., “Bimolecular fluorescence complementation (BiFC): A 5-year update and future perspectives, BioTechniques, Vol. 53, No. 5, November 2012, pp. 285-298. See also Remy et al., “Application of protein-fragment complementation assays in cell biology,” BioTechniques, Vol. 42, No. 2, 2007, pages 137-145; and Azad et al., “Split-luciferase complementary assay: Applications, recent developments, and future perspectives,” Anal. and Bioanal. Chem 406(23) 2014, pages 5541-5560; the contents of which are incorporated herein by reference in their entirety).

Reference is made herein to nucleic acid and nucleic acid sequences. The terms “nucleic acid” and “nucleic acid sequence” refer to a nucleotide, oligonucleotide, polynucleotide (which terms may be used interchangeably), or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin (which may be single-stranded or double-stranded and may represent the sense or the antisense strand).

Reference also is made herein to peptides, polypeptides, proteins and compositions comprising peptides, polypeptides, and proteins. As used herein, a polypeptide and/or protein is defined as a polymer of amino acids, typically of length >100 amino acids (Garrett & Grisham, Biochemistry, 2nd edition, 1999, Brooks/Cole, 110). A peptide is defined as a short polymer of amino acids, of a length typically of 20 or less amino acids, and more typically of a length of 12 or less amino acids (Garrett & Grisham, Biochemistry, 2nd edition, 1999, Brooks/Cole, 110).

As disclosed herein, exemplary peptides, polypeptides, proteins may comprise, consist essentially of, or consist of any reference amino acid sequence disclosed herein, or variants of the peptides, polypeptides, and proteins may comprise, consist essentially of, or consist of an amino acid sequence having at least about 80%, 90%, 95%, 96%, 97%, 98%, or 99% sequence identity to any amino acid sequence

5

disclosed or contemplated herein. Variant peptides, polypeptides, and proteins may include peptides, polypeptides, and proteins having one or more amino acid substitutions, deletions, additions and/or amino acid insertions relative to a reference peptide, polypeptide, or protein. Also disclosed are nucleic acid molecules that encode the disclosed peptides, polypeptides, and proteins (e.g., polynucleotides that encode any of the peptides, polypeptides, and proteins disclosed herein and variants thereof).

The term "amino acid," includes but is not limited to amino acids contained in the group consisting of alanine (Ala or A), cysteine (Cys or C), aspartic acid (Asp or D), glutamic acid (Glu or E), phenylalanine (Phe or F), glycine (Gly or G), histidine (His or H), isoleucine (Ile or I), lysine (Lys or K), leucine (Leu or L), methionine (Met or M), asparagine (Asn or N), proline (Pro or P), glutamine (Gln or Q), arginine (Arg or R), serine (Ser or S), threonine (Thr or T), valine (Val or V), tryptophan (Trp or W), and tyrosine (Tyr or Y) residues. The term "amino acid residue" also may include amino acid residues contained in the group consisting of homocysteine, 2-Aminoadipic acid, N-Ethylasparagine, 3-Aminoadipic acid, Hydroxylysine, ?-alanine, ?-Amino-propionic acid, allo-Hydroxylysine acid, 2-Aminobutyric acid, 3-Hydroxyproline, 4-Aminobutyric acid, 4-Hydroxyproline, piperidinic acid, 6-Aminocaproic acid, Isodesmosine, 2-Aminoheptanoic acid, allo-Isoleucine, 2-Aminoisobutyric acid, N-Methylglycine, sarcosine, 3-Aminoisobutyric acid, N-Methylisoleucine, 2-Aminopimelic acid, 6-N-Methyllysine, 2,4-Diaminobutyric acid, N-Methylvaline, Desmosine, Norvaline, 2,2'-Diaminopimelic acid, Norleucine, 2,3-Diaminopropionic acid, Ornithine, and N-Ethylglycine. Typically, the amide linkages of the peptides are formed from an amino group of the backbone of one amino acid and a carboxyl group of the backbone of another amino acid.

The amino acid sequences contemplated herein may include conservative amino acid substitutions relative to a reference amino acid sequence. For example, a variant peptides, polypeptides, and proteins as contemplated herein may include conservative amino acid substitutions relative to an amino acid sequence of a reference peptide, polypeptide, or protein. "Conservative amino acid substitutions" are those substitutions that are predicted to interfere least with the properties of the reference peptide, polypeptide, or protein. In other words, conservative amino acid substitutions substantially conserve the structure and the function of the reference peptide, polypeptide, or protein. The following table provides a list of exemplary conservative amino acid substitutions.

Table of Conservative Amino Acid Substitutions	
Original Residue	Conservative Substitution
Ala	Gly, Ser
Arg	His, Lys
Asn	Asp, Gln, His
Asp	Asn, Glu
Cys	Ala, Ser
Gln	Asn, Glu, His
Glu	Asp, Gln, His
Gly	Ala
His	Asn, Arg, Gln, Glu
Ile	Leu, Val
Leu	Ile, Val
Lys	Arg, Gln, Glu
Met	Leu, Ile
Phe	His, Met, Leu, Trp, Tyr

6

-continued

Table of Conservative Amino Acid Substitutions	
Original Residue	Conservative Substitution
Ser	Cys, Thr
Thr	Ser, Val
Trp	Phe, Tyr
Tyr	His, Phe, Trp
Val	Ile, Leu, Thr

"Non-conservative amino acid substitutions" are those substitutions that are predicted to interfere most with the properties of the reference peptide, polypeptide, or protein. For example, a non-conservative amino acid substitution might replace a basic amino acid at physiological pH such as Arg, His, or Lys, with a non-basic or acidic amino acid at physiological pH such as Asp or Glu. A non-conservative amino acid substitution might replace a non-polar amino acid at physiological pH such as Ala, Gly, Ile, Leu, Phe, or Val, with a polar amino acid at physiological pH such as Arg, Asp, Glu, His, or Lys.

Variants comprising deletions relative to a reference amino acid sequence or nucleotide sequence are contemplated herein. A "deletion" refers to a change in a reference amino acid sequence that results in the absence of one or more amino acid residues. A deletion removes at least 1, 2, 3, 4, 5, 10, 20, 50, 100, or 200 amino acids residues or a range of amino acid residues bounded by any of these values (e.g., a deletion of 5-10 amino acids). A deletion may include an internal deletion or a terminal deletion (e.g., an N-terminal truncation or a C-terminal truncation of a reference polypeptide). A "variant" of a reference polypeptide sequence may include a deletion relative to the reference polypeptide sequence (e.g., relative to any of SEQ ID NOs:1-3).

The words "insertion" and "addition" refer to changes in an amino acid sequence resulting in the addition of one or more amino acid residues. An insertion or addition may refer to 1, 2, 3, 4, 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 150, or 200 amino acid residues or a range of amino acid residues bounded by any of these values (e.g., an insertion or addition of 5-10 amino acids). A "variant" of a reference polypeptide sequence may include an insertion or addition relative to the reference polypeptide sequence (e.g., relative to any of SEQ ID NOs:1-3).

A "fusion polypeptide" refers to a polypeptide comprising at the N-terminus, the C-terminus, or at both termini of its amino acid sequence a heterologous amino acid sequence, for example, a heterologous amino acid sequence that extends the half-life of the fusion polypeptide in serum. A "variant" of a reference polypeptide sequence may include a fusion polypeptide comprising the reference polypeptide.

A "fragment" is a portion of an amino acid sequence which is identical in sequence to but shorter in length than a reference sequence (e.g., a fragment of any of SEQ ID NOs:1-3). A fragment may comprise up to the entire length of the reference sequence, minus at least one amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous amino acid residues of a reference polypeptide. In some embodiments, a fragment may comprise at least 5, 10, 15, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, 150, 250, or 500 contiguous amino acid residues of a reference polypeptide; or a fragment may comprise no more than 5, 10, 15, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, 150, 250, or 500 contiguous amino acid residues of a reference polypeptide; or a fragment may comprise a range of contiguous

amino acid residues of a reference polypeptide bounded by any of these values (e.g., 40-80 contiguous amino acid residues). Fragments may be preferentially selected from certain regions of a molecule. The term "at least a fragment" encompasses the full length polypeptide. A "variant" of a reference polypeptide sequence may include a fragment of the reference polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polypeptide sequences. Homology, sequence similarity, and percentage sequence identity may be determined using methods in the art and described herein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide. Percent identity for amino acid sequences may be determined as understood in the art. (See, e.g., U.S. Pat. No. 7,396,664, which is incorporated herein by reference in its entirety). A suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S. F. et al. (1990) *J. Mol. Biol.* 215:403-410), which is available from several sources, including the NCBI, Bethesda, Md., at its website. The BLAST software suite includes various sequence analysis programs including "blastp," that is used to align a known amino acid sequence with other amino acid sequences from a variety of databases.

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 100, at least 150, at least 200, at least 250, at least 300, at least 350, at least 400, at least 450, at least 500, at least 550, at least 600, at least 650, or at least 700 contiguous amino acid residues; or a fragment of no more than 15, 20, 30, 40, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, or 700 amino acid residues; or over a range bounded by any of these values (e.g., a range of 500-600 amino acid residues). Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

In some embodiments, a "variant" of a particular polypeptide sequence may be defined as a polypeptide sequence having at least 20% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool available at the National Center for Biotechnology Information's website. (See Tatiana A. Tatusova, Thomas L. Madden (1999), "Blast 2 sequences—a new tool for comparing protein and nucleotide sequences", *FEMS Microbiol. Lett.* 174:247-250). Such a pair of polypeptides may show, for example, at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%,

or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides, or range of percentage identity bounded by any of these values (e.g., range of percentage identity of 80-99%).

The disclosed fusion polypeptides may comprise a amino acid sequence fused directly to a heterologous amino acid sequence or fused indirectly via a linker sequence. Suitable linker sequences may include amino acid sequences of 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, or 15 amino acids or more, or a range bounded by any of these values (e.g., a linker of 5-25 amino acids). In some embodiments, the linker sequence comprises only glycine and serine residues.

Fusion polypeptide disclosed herein may include an amino acid tag sequence, for example, which may be utilized for purifying and/or identifying the fusion polypeptide. Suitable amino acid tag sequences may include, but are not limited to, histidine tag sequences comprising 5-10 histidine residues.

A variant polypeptide may have substantially the same functional activity as a reference polypeptide. For example, a variant polypeptide may exhibit or more biological activities associated with binding a ligand, exhibiting fluorescence, and/or enzymatic activity.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences. Percent identity for a nucleic acid sequence may be determined as understood in the art. (See, e.g., U.S. Pat. No. 7,396,664, which is incorporated herein by reference in its entirety). A suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S. F. et al. (1990) *J. Mol. Biol.* 215:403-410), which is available from several sources, including the NCBI, Bethesda, Md., at its website. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at the NCBI website. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed above).

Percent identity may be measured over the length of an entire defined polynucleotide sequence or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length may be used to describe a length over which percentage identity may be measured.

A "full length" polynucleotide sequence of a gene is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

A "variant," "mutant," or "derivative" of a particular nucleic acid sequence may be defined as a nucleic acid sequence having at least 50% sequence identity to the

particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the “BLAST 2 Sequences” tool available at the National Center for Biotechnology Information’s website. (See Tatiana A. Tatusova, Thomas L. Madden (1999), “Blast 2 sequences—a new tool for comparing protein and nucleotide sequences”, FEMS Microbiol Lett. 174:247-250). In some embodiments a variant polynucleotide may show, for example, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length relative to a reference polynucleotide.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

A “recombinant nucleic acid” is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, J. et al. (1989) *Molecular Cloning: A Laboratory Manual*, 2nd ed., vol. 1 3, Cold Spring Harbor Press, Plainview N.Y. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

“Transfection” and “transformation” describe a process by which exogenous DNA is introduced into a recipient cell. Transfection and transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transfection or transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The terms “transfected cells” and “transformed cells” include stably transfected cells or transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transfected cells or transformed cells which express the inserted DNA or RNA for limited periods of time.

The polynucleotide sequences contemplated herein may be present in expression cassettes and/or expression vectors (e.g., an expression vector comprising an expression cassette). For example, the vectors may comprise a polynucleotide encoding an ORF of a recombinant protein (e.g., an exogenous sensor as disclosed herein). The polynucleotide present in the vector may be operably linked to a promoter (e.g., a eukaryotic promoter or prokaryotic promoter). “Operably linked” refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close

proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame. Vectors contemplated herein may comprise a heterologous promoter (e.g., a eukaryotic or prokaryotic promoter) operably linked to a polynucleotide that encodes a protein. A “heterologous promoter” refers to a promoter that is not the native or endogenous promoter for the protein or RNA that is being expressed. For example, a heterologous promoter for a LAMP may include a eukaryotic promoter or a prokaryotic promoter that is not the native, endogenous promoter for the LAMP.

As used herein, “expression” refers to the process by which a polynucleotide is transcribed from a DNA template (such as into and mRNA or other RNA transcript) and/or the process by which a transcribed mRNA is subsequently translated into peptides, polypeptides, or proteins. Transcripts and encoded polypeptides may be collectively referred to as “gene product.” If the polynucleotide is derived from genomic DNA, expression may include splicing of the mRNA in a eukaryotic cell.

As used herein, an “expression cassette” minimally refers to a recombinant polynucleotide comprising a promoter operably linked to a recombinant coding sequence. An expression cassette may be present in a vector (e.g., an episomal vector which is transfected into a cell and remains episomal and/or which recombines into the genome of the cell). A vector may include one or more expression cassettes which express one or more coding sequences (e.g., one or more coding sequences for sensors as disclosed herein).

The term “vector” refers to some means by which nucleic acid (e.g., DNA) can be introduced into a host organism or host tissue. There are various types of vectors including plasmid vector, bacteriophage vectors, cosmid vectors, bacterial vectors, and viral vectors. As used herein, a “vector” may refer to a recombinant nucleic acid that has been engineered to express a heterologous polypeptide (e.g., the fusion proteins disclosed herein). The recombinant nucleic acid typically includes cis-acting elements for expression of the heterologous polypeptide.

Any of the conventional vectors used for expression in eukaryotic cells may be used for directly introducing DNA into a subject. Expression vectors containing regulatory elements from eukaryotic viruses may be used in eukaryotic expression vectors (e.g., vectors containing SV40, CMV, or retroviral promoters or enhancers). Exemplary vectors include those that express proteins under the direction of such promoters as the SV40 early promoter, SV40 later promoter, metallothionein promoter, human cytomegalovirus promoter, murine mammary tumor virus promoter, and Rous sarcoma virus promoter. Expression vectors as contemplated herein may include eukaryotic or prokaryotic control sequences that modulate expression of a heterologous protein (e.g. the fusion protein disclosed herein). Prokaryotic expression control sequences may include constitutive or inducible promoters (e.g., T3, T7, Lac, trp, or phoA), ribosome binding sites, or transcription terminators.

The vectors contemplated herein may be introduced and propagated in a prokaryote, which may be used to amplify copies of a vector to be introduced into a eukaryotic cell or as an intermediate vector in the production of a vector to be introduced into a eukaryotic cell (e.g. amplifying a plasmid as part of a viral vector packaging system). A prokaryote may be used to amplify copies of a vector.

The presently disclosed methods may include delivering one or more polynucleotides, such as or one or more vectors as described herein, one or more transcripts thereof, and/or one or more proteins transcribed therefrom, to a host cell. Further

contemplated are host cells produced by such methods, and organisms (such as animals, plants, or fungi) comprising or produced from such cells. The disclosed extracellular vesicles may be prepared by introducing vectors that express mRNA encoding a fusion protein as contemplated herein. Conventional viral and non-viral based gene transfer methods can be used to introduce nucleic acids in mammalian cells or target tissues. Non-viral vector delivery systems include DNA plasmids, RNA (e.g. a transcript of a vector described herein), naked nucleic acid, and nucleic acid complexed with a delivery vehicle, such as a liposome. Viral vector delivery systems include DNA and RNA viruses, which have either episomal or integrated genomes after delivery to the cell.

In the methods contemplated herein, a host cell may be transiently or non-transiently transfected (i.e., stably transfected) with one or more vectors described herein. In some embodiments, a cell is transfected as it naturally occurs in a subject (i.e., in situ). In some embodiments, a cell that is transfected is taken from a subject (i.e., explanted). In some embodiments, the cell is derived from cells taken from a subject, such as a cell line. Suitable cells may include stem cells (e.g., embryonic stem cells and pluripotent stem cells). A cell transfected with one or more vectors described herein may be used to establish a new cell line comprising one or more vector-derived sequences. In the methods contemplated herein, a cell may be transiently transfected with the components of a system as described herein (such as by transient transfection of one or more vectors, or transfection with RNA), and modified through the activity of a complex, in order to establish a new cell line comprising cells containing the modification but lacking any other exogenous sequence.

A “composition comprising a given polypeptide” and a “composition comprising a given polynucleotide” refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. The compositions may be stored in any suitable form including, but not limited to, freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. The compositions may be aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components.

“Substantially isolated or purified” nucleic acid or amino acid sequences are contemplated herein. The term “substantially isolated or purified” refers to nucleic acid or amino acid sequences that are removed from their natural environment, and are at least 60% free, preferably at least 75% free, and more preferably at least 90% free, even more preferably at least 95% free from other components with which they are naturally associated.

The disclosed sensors may include a protease cleavage sequence and/or a cognate protease that recognizes and cleaves the protease cleavage sequence. The sensors are not limited to any particular protease or corresponding protease cleavage site. In some embodiments, the protease and cleavage site are from a virus. For example, in certain embodiments, the protease and protease cleavage site are from a virus selected from: tobacco etch virus (TEV), a chymotrypsin-like serine protease and corresponding cleavage sites, alphavirus proteases and cleavage sites, Hepatitis C virus proteases (e.g., NS3 proteases) and corresponding cleavage sites, chymotrypsin-like cysteine proteases and corresponding cleavage sites, papain-like cysteine proteases and cleavage sites, picornavirus leader proteases and cleavage sites, HIV proteases and cleavage sites, Herpesvirus

proteases and cleavage sites, and adenovirus proteases and cleavage sites (see, Tong, Chem. Rev. 2002, 102, 4609-4626, herein incorporated by reference in its entirety). In particular embodiments, the proteases and cleavage sites are bacterial in origin, such as, for example, from *Streptomyces griseus* protease A (SGPA), SGPB, and alpha-lytic protease and corresponding cleavage sites. In some embodiments, the proteases and cleavage sites are mammalian. For example, the proteases could be one of the five major classes of proteases known in mammals which include serine proteases, cysteine proteases, metallo proteases, aspartic proteases, and threonine proteases (see, e.g., Turk et al., The EMBO Journal, 2012, 31, 1630-1643; Lopez-Otin and Overall, 2002, Nat. Rev. Mol. Cell Biol., 2:509-519; Overall and Blobel, 2007, Nat. Rev. Mol. Cell Biol., 8: 245-257; and Lopez-Otin and Bond, 2008, J. Biol. Chem., 283:30422-30437, all of which are herein incorporated in their entirety by references).

The disclosed subject matter relates to engineered red blood cells (eRBCs) which interchangeably may be referred to as red blood corpuscles, haematids, erythroid cells, or erythrocytes. The disclosed eRBCs typically include recombinant membrane proteins for detecting an extracellular ligand and generating a signal if the extracellular ligand is detected (i.e., recombinant exogenous sensors). The disclosed exogenous sensors may be prepared from native and/or non-native proteins of RBCs.

The disclosed eRBCs are anucleatic (i.e., lack a nucleus) and may be prepared from precursor RBCs (e.g., by inducing precursor RBCs to differentiate to RBCs). Precursor RBCs may include, but are not limited to, hemocytoblasts, multipotent hematopoietic stem cells, common myeloid progenitors, multipotent stem cells, unipotent stem cells, pronormoblasts, proerythroblasts, rubriblasts, normoblasts, erythroblasts, polychromatophilic normoblasts, intermediate normoblasts, orthochromatic normoblasts, and/or late normoblasts. The disclosed eRBCs may be prepared by transfecting precursor RBCs with expression cassettes and/or expression vectors for expressing biosensors as disclosed herein, and subsequently inducing the transfected precursor RBCs to differentiate to RBCs, thereby obtaining the disclosed eRBCs. The transfected precursor RBCs may be induced to differentiate by methods known in the art including contacting the transfected precursor RBCs and/or culturing the transfected precursor RBCs with differentiation factors (e.g., hormones and/or growth factors).

In some embodiments, the recombinant exogenous sensors may be derived, prepared, or engineered from native membrane proteins of RBCs or portions or fragments of membrane proteins of RBCs. Membrane proteins of RBCs suitable for deriving, preparing, and/or engineering the disclosed sensors may include, but are not limited to: glycoporphins (e.g., glycophorin A (GPA or GYPA), glycophorin B (GPB or GYPB), glycophorin C (GPC or GYPC), and/or glycophorin E (GPE or GYPE)); Band 3 (major anion transporter); Aquaporin 1 (water transporter); Glut1 (glucose and L-dehydroascorbic acid transporter); Kidd antigen protein (urea transporter); RhAG (gas transporter); Na⁺/K⁺-ATPase; Ca²⁺-ATPase; Na⁺-Cl⁻-cotransporter; Na⁺ K⁺2Cl⁻-cotransporter; Na-H exchanger; K-Cl cotransporter; Gardos channel; ICAM-4; BCAM; Kell antigen (XK); RHD/RhCE; Duffy protein; Adducin; and/or Dematin. In some embodiments, the native membrane protein from which the disclosed recombinant extracellular sensors are derived, prepared, and/or engineered is a type I membrane protein (i.e., single pass membrane protein with an extracellular N-terminus and an intracellular C-terminus).

such as GPA) and/or a type II membrane protein (i.e., single pass membrane protein with an extracellular C-terminus and an intracellular N-terminus such as Kell).

The disclosed exogenous sensors include one or more recombinant membrane proteins. In some embodiments, the disclosed exogenous sensors may include two recombinant membrane proteins which are fusion proteins comprising a membrane protein of a RBC or a portion or fragment thereof. In some embodiments, the fusion proteins may comprise a membrane protein of a RBC or a portion or fragment thereof to which a ligand binding domain has been fused at the N-terminus (i.e., N_{ter}-(ligand binding domain)-(RBC membrane protein)-C_{ter}) or at the C-terminus (i.e., N_{ter}-(RBC membrane protein)-(ligand binding domain)-C_{ter}). Further, the fusion proteins may comprise a membrane protein of a RBC or a portion or fragment thereof to which a functional protein has been fused at the N-terminus (i.e., N_{ter}-(ligand binding domain)-(RBC membrane protein)-(functional protein)-C_{ter}) or at the C-terminus (i.e., N_{ter}-(functional protein)-(RBC membrane protein)-(ligand binding domain)-C_{ter}). A first fusion protein may comprise a non-functional portion of a functional protein, and a second fusion protein may comprise a second non-functional portion of a functional protein such that when the first fusion protein and the second fusion protein bind the ligand for the ligand binding domain, the first non-functional portion and the second non-functional portion interact to constitute the functional protein.

Illustrative Embodiments

The following Embodiments are illustrative and should not be interpreted to limit the scope of the claimed subject matter.

Embodiment 1. An engineered red blood cell comprising: (a) a first exogenous extracellular sensor; the first extracellular sensor comprising: (i) a first extracellular ligand binding domain (LBD1), (ii) a transmembrane domain (TM1), and (iii) an intracellular first fragment of a functional protein (FP1), and (b) a second exogenous extracellular sensor; the second extracellular sensor comprising: (i) a second extracellular ligand binding domain (LBD2), (ii) a transmembrane domain (TM2), and (iii) an intracellular second fragment of the functional protein (FP2); wherein: the first extracellular ligand binding domain of the first exogenous sensor and the second extracellular ligand binding domain of the second exogenous sensor bind to the same ligand to form a ternary complex, and the first fragment of the functional protein and the second fragment of the functional protein interact in the ternary complex to reconstitute functional activity of the functional protein (i.e., the ligand comprises binding sites for LBD1 and LBD2). Optionally, TM1 and TM2 may be the same or different and or may comprise a native amino acid sequence of a RBC membrane protein and/or a mutated amino acid sequence of a RBC membrane protein (e.g., where the native amino acid sequence forms multimers and the mutated amino acid sequence does not form multimers). Optionally, where FP1 comprises a N-terminal portion of a full-length functional protein and FP2 comprises a C-terminal portion of a full-length functional protein or where FP1 comprises a C-terminal portion of a full-length functional protein and FP2 comprises a N-terminal portion of a full-length functional protein.

Embodiment 2. The engineered red blood cell of embodiment 1, wherein the functional protein is a fluorescent protein (e.g., a split fluorescent protein) and the fluorescent protein emits fluorescence when the ternary complex is

formed and the first fragment of the fluorescent protein and the second fragment of the fluorescent protein interact to reconstitute the fluorescent protein.

Embodiment 3. The engineered red blood cell of embodiment 2, wherein the fluorescent protein is selected from the group consisting of green fluorescent protein (GFP), EGFP (enhanced green fluorescent protein), Emerald, Superfolder GFP, Azami Green, mWasabi, TagGFP, TurboGFP, AcGFP, ZsGreen, T-Sapphire, GFP-S65T, frGFP, sfGFP, EBFP, EBFP2, Azurite, mTagBFP, ECFP, mECFP, Cerulean, mTurquoise, CyPet, AmCyan1, Midori-Ishi Cyan, TagCFP, mTFP1 (Teal), Dronpa, EYFP, Topaz, Venus, mCitrine, YPet, TagYFP, PhiYFP, ZsYellow1, mBanana, Kusabira Orange, Kusabira Orange2, mOrange, mOrange2, dTomato, dTomato-Tandem, TagRFP, TagRFP-T, DsRed, DsRed2, DsRed-Express (T1), DsRed-Monomer, mTangerine, mRuby, mApple, mStrawberry, AsRed2, mRFP1, JRed, mCherry, mKate, HcRed1, mRaspberry, dKeima-Tandem, HcRed-Tandem, mPlum, and AQ143.

Embodiment 4. The engineered red blood cell of any of the foregoing embodiments, wherein the first extracellular ligand binding domain (LBD1) and the transmembrane domain of the first exogenous extracellular sensor (TM1) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., LBD1-GGGSGGGGS-TM1; SEQ ID NO: 4); and/or wherein the transmembrane domain (TM1) and the intracellular first fragment of the functional protein of the first exogenous extracellular sensor (FP1) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., TM1-GGGSGGGGS-FP1; SEQ ID NO: 4).

Embodiment 5. The engineered red blood cell of any of the foregoing embodiments, wherein the second extracellular ligand binding domain (LBD2) and the transmembrane domain of the second exogenous extracellular sensor (TM2) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., LBD2-GGGSGGGGS-TM2; SEQ ID NO: 4); and/or wherein the transmembrane domain TM2 and the intracellular first fragment of the functional protein of the second exogenous extracellular sensor (FP2) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., TM2-GGGSGGGGS-FP2; SEQ ID NO: 4).

Embodiment 6. The engineered red blood cell of any of the foregoing embodiments, wherein the functional protein is an enzyme and the enzyme exhibits enzymatic activity emits when the ternary complex is formed and the first fragment of the fluorescent protein and the second fragment of the enzyme interact to reconstitute the enzyme.

Embodiment 7. The engineered red blood cell of embodiment 6, wherein the enzyme is a luciferase.

Embodiment 8. A combination of expression cassettes for preparing an engineered red blood cell, the combination comprising: (a) a first cassette expressing a first exogenous extracellular sensor; the first extracellular sensor comprising: (i) a first extracellular ligand binding domain (LBD1), (ii) a transmembrane domain (TM1), and (iii) an intracellular first fragment of a functional protein (FP1), and (b) a second cassette expressing a second exogenous extracellular sensor; the second extracellular sensor comprising: (i) a second extracellular ligand binding domain (LBD2), (ii) a transmembrane domain (TM2), and (iii) an intracellular second fragment of the functional protein (FP2); wherein: the first extracellular ligand binding domain of the first exogenous sensor and the second extracellular ligand bind-

15

ing domain of the second exogenous sensor bind to the same ligand to form a ternary complex, and the first fragment of the functional protein and the second fragment of the functional protein interact in the ternary complex to reconstitute functional activity of the functional protein (i.e., the ligand comprises binding sites for LBD1 and LBD2). Optionally, TM1 and TM2 may be the same or different and or may comprise a native amino acid sequence of a RBC membrane protein and/or a mutated amino acid sequence of a RBC membrane protein (e.g., where the native amino acid sequence forms multimers and the mutated amino acid sequence does not form multimers). Optionally, FP1 comprises an N-terminal portion of a full-length functional protein and FP2 comprises a C-terminal portion of a full-length functional protein or where FP1 comprises a C-terminal portion of a full-length functional protein and FP2 comprises an N-terminal portion of a full-length functional protein.

Embodiment 9. The combination of embodiment 8, wherein the expression cassettes are present on separate vectors.

Embodiment 10. The combination of embodiment 8, wherein the expression cassettes are present on the same vector.

Embodiment 11. The combination of any of the foregoing embodiments, wherein the functional protein is a fluorescent protein (e.g., a split fluorescent protein) and the fluorescent protein emits fluorescence when the ternary complex is formed and the first fragment of the fluorescent protein and the second fragment of the fluorescent protein interact to reconstitute the fluorescent protein.

Embodiment 12. The combination of embodiment 11, wherein the fluorescent protein is selected from the group consisting of green fluorescent protein (GFP), EGFP, Emerald, Superfolder GFP, Azami Green, mWasabi, TagGFP, TurboGFP, AcGFP, ZsGreen, T-Sapphire, GFP-S65T, frGFP, sfGFP, EBFP, EBFP2, Azurite, mTagBFP, ECFP, mECFP, Cerulean, mTurquoise, CyPet, AmCyan1, Midori-Ishi Cyan, TagCFP, mTFP1 (Teal), Dronpa, EYFP, Topaz, Venus, mCitrine, YFP, TagYFP, PhiYFP, ZsYellow1, mBanana, Kusabira Orange, Kusabira Orange2, mOrange, mOrange2, dTomato, dTomato-Tandem, TagRFP, TagRFP-T, DsRed, DsRed2, DsRed-Express (T1), DsRed-Monomer, mTangerine, mRuby, mApple, mStrawberry, AsRed2, mRFP1, JRed, mCherry, mKate, HcRed1, mRaspberry, dKeima-Tandem, HcRed-Tandem, mPlum, and AQ143.

Embodiment 13. The combination of any of the foregoing embodiments, wherein the first extracellular ligand binding domain (LBD1) and the transmembrane domain of the first exogenous extracellular sensor (TM1) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., LBD1-GGGSGGGG-TM1; SEQ ID NO: 4); and/or wherein the transmembrane domain (TM1) and the intracellular first fragment of the functional protein of the first exogenous extracellular sensor (FP1) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., TM1-GGGSGGGG-FP1; SEQ ID NO: 4).

Embodiment 14. The combination of any of the foregoing embodiments, wherein the second extracellular ligand binding domain (LBD2) and the transmembrane domain of the second exogenous extracellular sensor (TM2) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., LBD2-GGGSGGGG-TM2; SEQ ID NO: 4); and/or wherein the transmembrane domain TM2 and the intracellular first frag-

16

ment of the functional protein of the second exogenous extracellular sensor (FP2) are linked by a 5-25 amino acid linking sequence of amino acids selected from glycine, serine, and combination thereof (e.g., TM2-GGGSGGGG-FP2; SEQ ID NO: 4).

Embodiment 15. The combination of any of the foregoing embodiments, wherein the functional protein is an enzyme and the enzyme exhibits enzymatic activity emits when the ternary complex is formed and the first fragment of the fluorescent protein and the second fragment of the enzyme interact to reconstitute the enzyme.

Embodiment 16. The combination of embodiment 15, wherein the enzyme is a luciferase.

Embodiment 17. A method for detecting a ligand (or a metabolite), the method comprising contacting the engineered red blood cell of any of embodiments 1-7 with the ligand (or metabolite), and detecting functional activity of the functional protein.

Embodiment 18. The method of embodiment 17, wherein the functional protein is a fluorescent protein and detecting functional activity comprises detecting fluorescence.

Embodiment 19. The method of embodiment 17, wherein the functional protein is a luciferase protein and detecting functional activity comprises contacting the engineered red blood cell with a substrate for the luciferase protein and detecting light emitted from the engineered red blood cell.

Embodiment 20. A method for preparing the engineered red blood cells of any of embodiments 1-7, the method comprising: (I) transfecting a precursor red blood cell with a combination of expression cassettes of any of embodiments 8-16; and (II) inducing the transfected precursor red blood cell precursor cell to differentiate into a red blood cell, thereby preparing the engineered red blood cell.

Embodiment 21. A precursor red blood cell transfected with the cassettes of any of embodiments 8-16.

Embodiment 22. The engineered red blood cells of any of embodiments 1-7, wherein the first exogenous extracellular sensor and/or the second exogenous extracellular sensor comprise an amino acid sequence of a native membrane protein of red blood cells or a portion or fragment thereof (e.g., at least a portion of the transmembrane amino acid sequence of a native membrane protein of red blood cells, optionally at least a portion of the extracellular amino acid sequence of a native membrane protein of red blood cells, and optionally at least a portion of the intracellular amino acid sequence of a native membrane protein of red blood cells).

Embodiment 23. The combination of expression cassettes of any of claims 8-16, wherein the first exogenous extracellular sensor and/or the second exogenous extracellular sensor comprise an amino acid sequence of a native membrane protein of red blood cells or a portion or fragment thereof (e.g., at least a portion of the transmembrane amino acid sequence of a native membrane protein of red blood cells, optionally at least a portion of the extracellular amino acid sequence of a native membrane protein of red blood cells, and optionally at least a portion of the intracellular amino acid sequence of a native membrane protein of red blood cells).

Embodiment 24. The method of any of embodiments 17-20, wherein the first exogenous extracellular sensor and/or the second exogenous extracellular sensor comprise an amino acid sequence of a native membrane protein of red blood cells or a portion or fragment thereof (e.g., at least a portion of the transmembrane amino acid sequence of a native membrane protein of red blood cells, optionally at least a portion of the extracellular amino acid sequence of a

native membrane protein of red blood cells, and optionally at least a portion of the intracellular amino acid sequence of a native membrane protein of red blood cells).

Embodiment 25. The precursor red blood cell of embodiment 21, wherein the first exogenous extracellular sensor and/or the second exogenous extracellular sensor comprise an amino acid sequence of a native membrane protein of red blood cells or a portion or fragment thereof (e.g., at least a portion of the transmembrane amino acid sequence of a native membrane protein of red blood cells, optionally at least a portion of the extracellular amino acid sequence of a native membrane protein of red blood cells, and optionally at least a portion of the intracellular amino acid sequence of a native membrane protein of red blood cells).

EXAMPLES

The following Examples are illustrative and should not be interpreted to limit the scope of the claimed subject matter.

Example 1

Abstract

Cell-based therapies comprise a blossoming, multi-billion dollar industry addressing markets ranging from cancer to autoimmune disease. An emerging frontier is the use of red blood cells (RBC), which have exceptionally long circulation times (up to 120 days—far longer than synthetic vehicles), lack DNA (and thus are safe), and can be loaded with drugs, proteins, or other cargo. Technologies that enable one to engineer RBCs to perform specific functions in vivo could serve unmet diagnostic and therapeutic needs. This invention comprises a technology for generating engineered RBC (eRBC) to serve as long-lived biosensors. Each eRBC would emit a fluorescent readout when bound to the analyte of interest, such that eRBC state could be monitored non-invasively using established technologies for fluorescent imaging (e.g., fluorescent imaging of the retina).

Applications

Applications for the disclosed technology may include, but are not limited to: (a) Diagnostics: using simple imaging technology, a patient could perform regular self-analysis and enable real-time, high frequency monitoring outside clinical settings, none of which is possible with existing technologies requiring specialized equipment, trained personnel, and/or sample collection; (b) Exposure monitoring: by a similar approach, military personal and first responders could determine whether or not they have been exposed to a potential infectious agent via monitoring output from a tailored eRBC; and (c) Basic science: eRBC could provide a facile approach to non-invasively monitoring soluble species in the serum of experimental animals.

Advantages

Advantages of the disclosed technology may include, but are not limited to: (a) eRBC are longer-lived than any injectable particle or small molecule; a single injection could enable monitoring for weeks or months; (b) eRBC biosensor output is amenable to noninvasive monitoring, without requiring sample collection, complicated analysis, or trained personnel; (c) the disclosed protein biosensors are readily customized to detect novel analytes—all that is needed is replacing the ligand binding domain with binding proteins

(e.g., single chain antibodies) that bind the analyte of interest; and (c) RBC have been engineered for other applications but this represents the first technology enabling the engineering of eRBC to serve as biosensors for a range of analytes found in the blood.

Brief Summary of the Technology

This technology comprises a protein biosensor platform for engineering RBCs such that sensing of an extracellular analyte results in the reconstitution of a split functional protein in the RBC cytosol. The biosensor comprises two protein chains, each of which includes an extracellular ligand binding domain, a membrane-spanning domain, and an intracellular domain comprising half of a split functional protein (FIG. 1). In the proposed mechanism, upon binding of the ligand to the ligand binding domains, the two chains dimerize, bringing the two split protein halves into proximity causing the split functional protein to reconstitute and therefore exhibit functionality. Suitable functional proteins may include, but are not limited to, fluorescent proteins and enzymatic proteins.

In FIG. 1, each biosensor is composed of two protein chains each with an extracellular ligand binding domain (LBD1 and LBD2, respectively) and a single transmembrane pass domain (TM1 and TM2, respectively). The intracellular portion of the receptors contains one half of a split fluorescent protein (FP1 and FP2, respectively). Upon ligand binding, the two chains dimerize, allowing to two split protein halves to come into contact and reconstitute the fluorescent protein and then emit fluorescence.

Technical Description

As an initial proof of concept, we developed a receptor with extracellular ligand binding domains that heterodimerize upon binding the small molecule rapamycin (FRB and FKBP form a heterodimer in the presence of rapamycin) (FIG. 1) (for a review of the FKBP-Rapamycin-FRB ternary complex, see Banaszynski, et al., J. Am. Chem. Soc. (JACS) Articles, Mar. 9, 2005, 127, 4715-4721, the content of which is incorporate herein by reference in its entirety). In this preliminary work, we used split GFP (sGFP) as the split fluorescent protein [Ghosh 2000], although the eventual application of this technology in vivo would probably utilize reconstitution of a split infrared fluorescent protein [Filonov 2013] or split luciferase [Dixon 2016]. Our initial constructs utilized a simple single-pass transmembrane domain, and none of these receptors exhibited ligand-inducible reconstitution of GFP (FIG. 2).

The results in FIG. 2 represent experiments in which a library of receptors encoding a rapamycin binding domain, transmembrane domain, and split GFP half was created and transfected in pair-wise combinations into HEK293FT cells. The background signaling was very low, with most cells actually showing GFP values at or below that of the control cells (transfected with only one receptor chain). Furthermore, no combination of receptors, regardless of linker length, showed any induction of GFP upon addition of rapamycin. All transfections were performed using the CaCl₂-HEPES buffered saline methodology. Sixteen (16) hours post transfection, rapamycin was added with media change, and the cells were allowed to incubate for an additional 24 hours. Cells were harvested and analyzed by flow cytometry.

We next investigated whether biosensors could be constructed by fusing ligand-binding and sGFP domains to the

RBC-resident proteins Kell and Glycophorin A (GPA) (FIG. 3). Kell and GPA are expressed in RBC even when expressed as genetic fusion constructs [Shi 2014]. Moreover, since GPA is a Type I membrane protein and Kell is a Type II membrane protein (Kell and GPA are expressed in opposite orientation relative to the plasma membrane), we hypothesized that fusion of sGFP fragments to the intracellular terminus of each protein may bring the sGFP fragments together in the antiparallel orientation required for reconstitution of GFP. As illustrated in FIG. 3, Kell and GPA were modified to contain extracellular rapamycin-binding domains (RBD1 and RBD2 at their extracellular N-terminus and extracellular C-terminus, respectively) and intracellular split GFP halves (GFP1 and GFP2 at their intracellular C-terminus and intracellular C-terminus, respectively). Upon the addition of rapamycin, these two protein chains can dimerize, allowing for the reconstitution of GFP. These two proteins are naturally antiparallel to one another: Kell possesses an extracellular C-terminus, and GPA an extracellular N-terminus.

Several of these second-generation biosensors exhibited ligand-inducible reconstitution of fluorescence (FIG. 4). Because only some combinations of receptor designs conferred ligand-inducible reconstitution, we hypothesized that geometric constraints may limit reconstitution in these receptors. Thus, we introduced longer linkers into the intracellular portions of these functional receptors (i.e., glycine-serine linkers between the transmembrane domain portion and functional protein portion of the sensors), and we observed that ligand-induced reconstitution was indeed dramatically improved (FIG. 4).

The results in FIG. 4 represent experiments in which pairs of receptors were expressed in HEK 293FT cells. These data indicate that split protein reconstitution on a membrane is feasible, given that the receptors achieve the necessary orientation for protein reconstitution. All transfections were performed using the CaCl₂-HEPES buffered saline methodology. Sixteen (16) hours post-transfection, rapamycin was added along with media change, and cells were incubated for an additional 24 hours before being harvested and analyzed by flow cytometry.

These initial experiments demonstrated that RBC-resident proteins may be engineered into ligand-inducible biosensor proteins, and that such proteins may be iteratively improved using design-driven approaches. These data also support the feasibility of adapting this modular receptor design to incorporate novel ligand binding domains and novel fluorophore outputs

Because GPA is known to be expressed at higher levels than is Kell, we also investigated GPA-only biosensors (FIG. 5). As illustrated in FIG. 5, GPA, a native RBC protein, was modified to create two sensors (i.e., two fusion proteins) having extracellular rapamycin-binding domains (FRB or FKBP) and intracellular split GFP halves (GFPS and GFPL). Upon the addition of rapamycin, these two protein chains can dimerize, allowing for the reconstitution of GFP. These biosensors also conferred ligand-inducible reconstitution of sGFP (FIG. 6).

The results in FIG. 6 represent experiments in which pairs of receptors were expressed in HEK 293FT cells. All transfections were performed using the CaCl₂-HEPES buffered saline methodology. Sixteen (16) hours post-transfection, rapamycin was added along with media change, and cells were incubated for an additional 24 hours before being harvested and analyzed by flow cytometry.

We hypothesized that biosensors based upon wild-type GPA exhibited high background signaling due to homodi-

merization in the absence of ligand, and therefore we investigated biosensors in which one or both GPA domains required for homodimerization were mutated to reduce homodimerization in the absence of ligand (FIG. 7). As illustrated in FIGS. 7-9, biosensor proteins described in FIG. 6 were modified to mutate the transmembrane domain of GPA to decrease native background dimerization. As illustrated in FIG. 7 and FIG. 8, GPA was mutated in the transmembrane domain (GPAm) by converting the neutral valine at position 84 to a charged amino acid (V84R: Valine 84 to Arginine, V84E: Valine 84 to Glutamic Acid, V84K: Valine 84 to Lysine), and GPA was modified to contain N-terminal extracellular rapamycin-binding domains (FRB or FKBP) and C-terminal intracellular split GFP halves (GFPS or GFPL). Upon the addition of rapamycin, these two protein chains can dimerize, allowing for the reconstitution of GFP.

The mutated proteins then were expressed in HEK293FT cells to test dimerization in the presence of rapamycin. (See FIG. 8 and FIG. 9). The results in FIG. 8 and FIG. 9 represent experiments in which pairs of receptors were expressed in HEK 293FT cells. All transfections were performed using the CaCl₂-HEPES buffered saline methodology. Sixteen (16) hours post-transfection, rapamycin was added along with media change, and cells were incubated for an additional 24 hours before being harvested and analyzed by flow cytometry. As observed, the mutated biosensors exhibited sGFP reconstitution that was lower than that exhibited by wild-type GPA based receptors but higher than that exhibited by Kell-based biosensors (FIG. 8 and FIG. 9).

Because sGFP reconstitution efficiency may be limited by parameters not directly related to the eRBC biosensor mechanism, we also investigated whether we could generate a biosensor based upon reconstitution of a different signaling domain—split NANOLUCIFERASE™ [Dixon 2016](FIG. 10). As illustrated in FIG. 10, GPA was mutated in the transmembrane domain (GPAm, see FIGS. 7-9) and was modified to contain N-terminal extracellular rapamycin-binding domains (FRB or FKBP) and C-terminal intracellular split NANOLUCIFERASE™ halves (114 and 11S). Upon the addition of rapamycin, these two protein chains can dimerize, allowing for the reconstitution of NANOLUCIFERASE™, and luminescence upon addition of the substrate, furimazine.

These NANOLUCIFERASE™-based biosensors exhibited robust ligand-inducible reconstitution of luciferase activity, when expressed in HEK293FT cells (FIG. 11). The results in FIG. 11 represent experiments in which pairs of receptors were expressed in HEK 293FT cells. All transfections were performed using the CaCl₂-HEPES buffered saline methodology. Sixteen (16) hours post-transfection, rapamycin was added along with media change, and cells were incubated for an additional 24 hours before being harvested and analyzed by plate reader for luciferase luminescence.

To test whether such eRBC biosensors would be expressed in functional in red blood cells (erythrocytes), we next expressed our biosensors in Gler cells. Gler cells can be induced to differentiate into erythrocytes by exposure to beta-estradiol. We observed that our biosensors were highly functional in Gler cells, without or without differentiation stimuli, and when differentiated for various lengths of time (FIG. 12). As illustrated in FIG. 12, Gler cells were spinoculated with retrovirus containing pairs of receptors and GFP expressed in one plasmid. Cells were sorted for GFP expression. GFP+ cells received either ethanol or -estradiol (to drive differentiation into an erythrocyte phenotype) and

either DMSO or rapamycin. Cell lysates were collected in passive lysis buffer 24 hours and 48 hours after receiving treatment. Technical replicates of cell lysates were analyzed for GFP fluorescence on a plate reader. Then a luciferase assay was performed on the cell lysates with the addition of substrate. Abbreviations: FD—fold difference. These results indicate that our eRBC biosensors are expressed, retained, and functional in erythrocytes, validating the core concept described in this disclosure.

We also tested the biosensors in human CD34+ cells. The results in FIG. 13 represent experiments in which human CD34+ cells were spinoculated with a lentiviral vector expressing a pair of receptors and Puromycin resistance. Cells were selected with Puromycin and then differentiated for 14 days. Differentiated cells received either DMSO or rapamycin. One hour after treatment, cell lysates were collected in passive lysis buffer. A luciferase assay was performed on cell lysates.

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Example 2

Reference is made to the Abstract entitled “Engineering Red Blood Cell-Based Biosensors for Physiological Monitoring,” Authors: Dolberg, T. B., Schwarz, K. A., Leonard, J. M. which was presented at the 2016 Synthetic Biology: Engineering, Evolution & Design (SEED) conference on Jul. 19, 2016, and which Abstract is incorporated by reference herein in its entirety.

Cell-based therapies have a wide range of applications ranging from cancer immunotherapy to regenerative medicine. A promising emerging frontier of this field is the development of engineered red blood cells (eRBCs) for therapeutic and diagnostic applications. RBCs have exceptionally long circulation times (around 40 days—far longer than synthetic vehicles), lack DNA (and thus are safe), and can be loaded with drugs, proteins, or other cargo. Technologies that enable one to engineer RBCs to perform specific functions in vivo could serve unmet diagnostic and

therapeutic needs. In particular, new technologies are required for non-invasive, routine monitoring for pathogen exposure (e.g., in the context of first responders) and for “actionable” analytes (e.g., markers of inflammation post-surgery).

In this project, we sought to develop eRBC biosensors that detect highly toxic agents, with the long-term goal of enabling one to detect exposure to these agents prior to the onset of physical symptoms. As a first step towards this goal, we designed and evaluated a novel biosensor strategy that is suitable to achieving biosensing in eRBCs, which lack DNA and thus require a readout other than gene expression. Towards this end, we engineered a novel cell-surface receptor protein in which ligand binding induces receptor dimerization, which then facilitates reconstitution of an intracellular split fluorescent protein. Ultimately, eRBC fluorescence may be monitored noninvasively using established technologies for fluorescent imaging of the retina. Importantly, our strategy involves modification of RBC-resident proteins, since retention of membrane proteins during RBC maturation is a tightly regulated and an incompletely understood process. In this study, we comparatively evaluated a range of biosensor architectures that implement the proposed mechanism, identified design biosensor features that successfully conferred significant ligand-induced generation of fluorescent output, and investigated strategies for improving biosensor performance (e.g., minimization of background fluorescence and enhancing fold-induction upon exposure to ligand). This crucial proof-of-principle demonstration establishes a foundation for developing eRBC biosensors that could ultimately address an unmet need for noninvasive monitoring of physiological signals for a range of diagnostic applications.

Example 3

Reference is made to the Abstract entitled “Engineering Red Blood Cell-Based Biosensors for Physiological Monitoring,” Authors: Dolberg, T. B., Schwarz, K. A., Leonard, J. M., which was presented at the 2016 annual conference of the American Institute of Chemical Engineers (AIChE) on Nov. 14, 2016 and which Abstract is incorporated by reference herein in its entirety.

Cell-based therapies have a wide range of applications ranging from cancer immunotherapy to regenerative medicine. A promising emerging frontier of this field is the development of engineered red blood cells (eRBCs) for therapeutic and diagnostic applications. RBCs have exceptionally long circulation times (around 120 days—far longer than synthetic vehicles), lack DNA (and thus are safe), and can be loaded with drugs, proteins, or other cargo. Technologies that enable one to engineer RBCs to act as biosensors, performing specific functions in vivo, could serve unmet diagnostic and therapeutic needs. In particular, new technologies are required for non-invasive, routine monitoring for pathogen exposure (e.g., in the context of first responders) and for other “actionable” analytes (e.g., markers of inflammation post-surgery).

In this project, we are developing eRBC biosensors that generate a fluorescent output upon detection of the analyte of interest. Ultimately, eRBC biosensor fluorescent output may be monitored non-invasively using established technologies for fluorescent imaging of the retina. Using this simple imaging technology, a patient could perform regular self-analysis and enable real time, high frequency monitoring outside clinical settings, none of which is possible with existing technologies requiring specialized equipment,

trained personnel, and/or sample collection. Thus, such biosensors that enable the detection of actionable analytes could benefit exposed personnel by accelerating the initiation of treatment (perhaps before obvious symptoms present) and reduction of further exposure risks when possible.

As a first step towards the goal of building eRBC biosensors, we designed and evaluated a novel biosensor strategy that is suitable for achieving biosensing in eRBCs, which lack DNA and thus require a readout other than gene expression. Towards this end, we engineered a novel cell-surface receptor protein in which ligand binding induces receptor dimerization, which then facilitates reconstitution of an intracellular split fluorescent protein. Importantly, our strategy involves modification of RBC-resident proteins, since retention of membrane proteins during RBC maturation is a tightly regulated and an incompletely understood process. We comparatively evaluated a range of biosensor architectures that implement the proposed mechanism, enabling us to identify biosensor designs and design features that successfully conferred significant ligand-induced generation of fluorescent output. We also evaluated and implemented strategies for improving biosensor performance, including minimization of background fluorescence and enhancing fold-induction upon exposure to ligand. This crucial proof-of-principle demonstration establishes a foundation for developing eRBC biosensors that could ultimately address an unmet need for non-invasive monitoring of physiological signals for a range of diagnostic applications.

Example 4

Reference is made to the Abstract entitled "Engineering Red Blood Cell-Based Biosensors for Physiological Monitoring," Authors: Dolberg, T. B., Schwarz, K. A., Leonard, J. M.," which is to be presented at the 2017 Midwest Regional Conference (MRC) of the American Institute of Chemical Engineers (AIChE) on Mar. 1, 2017 and which Abstract is incorporated by reference herein in its entirety.

Cell-based therapies have a wide range of applications ranging from cancer immunotherapy to regenerative medicine. A promising emerging frontier of this field is the development of engineered red blood cells (eRBCs) for therapeutic and diagnostic applications. RBCs are an attractive platform for diagnostics because they have exceptionally long circulation times (around 120 days—far longer than synthetic vehicles), lack DNA (and thus are safe), and can be loaded with drugs, proteins, or other cargo. Recent technological advances have enabled the large-scale production of RBCs from precursor cells, which may potentially be harnessed to generate off-the-shelf eRBC-based products to meet medical needs, including both diagnostic and therapeutic applications.

The specific goal of this project is to generate eRBC-based technologies enabling noninvasive monitoring for pathogen exposure (e.g., in the context of first responders) and for other "actionable" analytes (e.g., markers of inflammation post-surgery). Towards this goal, we are developing eRBC biosensors that generate a fluorescent output upon detection of the analyte of interest, and this output may be monitored non-invasively using established technologies for fluorescent imaging of the retina. These biosensors would enable the detection of actionable analytes thus benefitting exposed personnel by accelerating the initiation of treatment (perhaps before obvious symptoms present) and reducing of further exposure risks when possible.

As a first step to enable RBCs to act as sensors, we designed and evaluated a novel biosensor strategy that is

suitable for achieving biosensing in eRBCs, which lack DNA and thus require a readout other than gene expression. Towards this end, we engineered a novel cell-surface receptor protein in which ligand binding induces receptor dimerization, which then facilitates reconstitution of an intracellular split fluorescent protein. Importantly, our strategy involves modification of RBC-resident proteins, since retention of membrane proteins during RBC maturation is a tightly regulated and an incompletely understood process. We comparatively evaluated a range of biosensor architectures that implement the proposed mechanism, enabling us to identify biosensor designs and design features that successfully conferred significant ligand-induced generation of fluorescent output. We also evaluated and implemented strategies for improving biosensor performance, including minimization of background fluorescence and enhancing fold-induction upon exposure to ligand. This crucial proof-of-principle demonstration establishes a foundation for developing eRBC biosensors that could ultimately address an unmet need for non-invasive monitoring of physiological signals for a range of diagnostic applications.

Example 5

Reference is made to U.S. Publication No. 2014-0234851, published on Aug. 21, 2014, which discloses modular extracellular sensor architecture (MESA) for cell-based biosensors, and which is incorporated herein by reference in its entirety. The modular exogenous extracellular sensor architecture disclosed in U.S. Publication No. 2014-0234851 may be adapted for use in the engineered red blood cell-based biosensors disclosed and contemplated herein.

Example 6

Background. This example describes how engineered mammalian biosensor technology can be applied to RBC-based technologies. The MESA family of engineered cell-surface biosensors (see, e.g., U.S. Publication No. 2014-0234851) enables the transduction of an extracellular ligand-binding event into a change in intracellular state. Of particular relevance to engineering RBCs, this intracellular state change can comprise reconstitution of a split protein, which may be an enzyme (as demonstrated using a split TEV protease) or potentially another functional domain (e.g., a split fluorescent protein). Briefly summarized below are several potential applications for engineering high value RBC using such technologies.

Pathogen exposure monitoring. RBCs could be engineered to monitor exposure to a pathogen and provide a readout that is easily monitored. A target pathogen could be dengue virus, for which serial exposure to multiple serotypes significantly increases risk for developing severe hemorrhagic symptoms. Thus, using engineered RBCs (eRBCs) to monitor patients or warfighters for secondary exposure could trigger rapid intervention to reduce morbidity or mortality. The biosensor readout could be reconstitution of a split fluorescent protein (e.g., an infrared fluorescent protein (IFP) that is spectrally distinguishable from hemoglobin in RBC), and eRBC fluorescence could be monitored using a technique similar to fluorescent angiography of the retina. Such eRBCs could also incorporate a second fluorophore to enable quantitative normalization of biosensor readout. Distinct biosensors could be engineered to provide pathogen or serotype-specific biosensing. Alternatively, biosensor readout could comprise reconstitution of a bioluminescent catalyst (e.g., luciferase or another enzyme, for

25

example that catalyzes production of a product that yields a color change in excreted urine) and/or eRBC biosensor state could be evaluated ex vivo using a blood sample.

Physiological monitoring. Using techniques similar to those described above, eRBC biosensors could be used to monitor blood concentrations of physiologically-important species over time using noninvasive techniques. Potential targets include (a) Low-density lipoprotein (LDL), certain forms of which promote immuno-inflammatory processes and drive atherosclerosis, which could be useful for evaluating or tracking cholesterol levels or (b) cytokines that predict the onset of acute transplant rejection, such as IL-6, which may predict acute kidney transplant rejection.

Incorporation of MESA-type biosensors into eRBCs. Because only a subset of proteins is retained in mature RBC, MESA-type biosensors may be redesigned to coopt localization of retained proteins such as glycophorin or Kell. Potential strategies include non-covalent tethering via the introduction of protein-protein interaction domains into both MESA proteins and glycophorin and/or Kell.

Induction of immune tolerance. Although expression of non-native protein domains on the surface of eRBC may enable immunological clearance of eRBCs, some evidence indicates that expression of proteins in or on RBC promotes active induction of immunological tolerance.

26

In the foregoing description, it will be readily apparent to one skilled in the art that varying substitutions and modifications may be made to the invention disclosed herein without departing from the scope and spirit of the invention. The invention illustratively described herein suitably may be practiced in the absence of any element or elements, limitation or limitations which is not specifically disclosed herein. The terms and expressions which have been employed are used as terms of description and not of limitation, and there is no intention that in the use of such terms and expressions of excluding any equivalents of the features shown and described or portions thereof, but it is recognized that various modifications are possible within the scope of the invention. Thus, it should be understood that although the present invention has been illustrated by specific embodiments and optional features, modification and/or variation of the concepts herein disclosed may be resorted to by those skilled in the art, and that such modifications and variations are considered to be within the scope of this invention.

Citations to a number of patent and non-patent references may be made herein. The cited references are incorporated by reference herein in their entireties. In the event that there is an inconsistency between a definition of a term in the specification as compared to a definition of the term in a cited reference, the term should be interpreted based on the definition in the specification.

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Ser	Ile	Met	Ala	His	Glu	Leu	Leu	His	Ile	Phe	Tyr	Gln	Leu	Leu	Leu
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Pro	Gly	Gly	Cys	Leu	Ala	Cys	Asp	Asn	His	Ala	Leu	Gln	Glu	Ala	His
		595					600					605			
Leu	Cys	Leu	Lys	Arg	His	Tyr	Ala	Ala	Phe	Pro	Leu	Pro	Ser	Arg	Thr
	610					615					620				
Ser	Phe	Asn	Asp	Ser	Leu	Thr	Phe	Leu	Glu	Asn	Ala	Ala	Asp	Val	Gly
625				630					635					640	
Gly	Leu	Ala	Ile	Ala	Leu	Gln	Ala	Tyr	Ser	Lys	Arg	Leu	Leu	Arg	His
		645						650						655	
His	Gly	Glu	Thr	Val	Leu	Pro	Ser	Leu	Asp	Leu	Ser	Pro	Gln	Gln	Ile
		660						665					670		
Phe	Phe	Arg	Ser	Tyr	Ala	Gln	Val	Met	Cys	Arg	Lys	Pro	Ser	Pro	Gln
		675					680					685			
Asp	Ser	His	Asp	Thr	His	Ser	Pro	Pro	His	Leu	Arg	Val	His	Gly	Pro
	690					695					700				
Leu	Ser	Ser	Thr	Pro	Ala	Phe	Ala	Arg	Tyr	Phe	Arg	Cys	Ala	Arg	Gly
705				710					715					720	
Ala	Leu	Leu	Asn	Pro	Ser	Ser	Arg	Cys	Gln	Leu	Trp				
			725					730							

<210> SEQ ID NO 3

<211> LENGTH: 238

<212> TYPE: PRT

<213> ORGANISM: Aequorea victoria

<400> SEQUENCE: 3

Met	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val
1				5					10					15	

Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu
		20						25					30		

Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys
		35						40				45			

-continued

Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Phe
50						55					60				
Ser	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln
65					70					75					80
His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg
				85					90					95	
Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val
				100				105						110	
Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile
				115				120					125		
Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn
				130			135					140			
Tyr	Asn	Ser	His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly
145					150					155					160
Ile	Lys	Val	Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val
				165					170						175
Gln	Leu	Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro
				180					185					190	
Val	Leu	Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu	Ser
				195				200					205		
Lys	Asp	Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val
				210			215					220			
Thr	Ala	Ala	Gly	Ile	Thr	His	Gly	Met	Asp	Glu	Leu	Tyr	Lys		
225					230					235					

<210> SEQ ID NO 4

<211> LENGTH: 8

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 4

Gly Gly Gly Ser Gly Gly Ser
1 5

We claim:

1. A biosensor expressed in a blood cell, comprising:

(a) a first protein comprising:

- (i) a first extracellular ligand binding domain,
- (ii) a first transmembrane domain comprising a portion of an amino acid sequence from a red blood cell native membrane protein,
- (iii) an intracellular first fragment of a functional protein, and
- (iv) an amino acid linker connecting the first transmembrane domain and the intracellular first fragment of the functional protein; and

(b) a second protein comprising:

- (i) a second extracellular ligand binding domain,
- (ii) a second transmembrane domain,
- (iii) an intracellular second fragment of the functional protein, and
- (iv) an amino acid linker connecting the second transmembrane domain and the intracellular second fragment of the functional protein;

wherein the first extracellular ligand binding domain and the second extracellular ligand binding domain bind to the same ligand, wherein the first fragment of the functional protein and the second fragment of the functional protein are capable of interacting to reconstitute activity of the func-

tional protein, and wherein the amino acid linker of the first protein or the amino acid linker of the second protein comprises 7-25 amino acids.

2. The biosensor of claim 1, wherein the amino acid linker of the first protein and the amino acid linker of the second protein each comprise 7-25 amino acids.

3. The biosensor of claim 1, wherein the amino acid linker of the first protein and the amino acid linker of the second protein each comprise a sequence of amino acids selected from glycine, serine, and combinations thereof.

4. The biosensor of claim 1, wherein the amino acid linker of the first protein and the amino acid linker of the second protein each comprise the sequence GGGSGGGS (SEQ ID NO: 4).

5. The biosensor of claim 1, wherein the functional protein is a fluorescent protein and the fluorescent protein emits fluorescence when the ternary complex is formed and the first fragment of the fluorescent protein and the second fragment of the fluorescent protein interact to reconstitute the fluorescent protein.

6. The biosensor of claim 5, wherein the fluorescent protein is selected from the group consisting of green fluorescent protein (GFP), EGFP, Emerald, Superfolder GFP, Azami Green, mWasabi, TagGFP, TurboGFP, AcGFP, ZsGreen, T-Sapphire, GFP-S65T, frGFP, sfGFP, EBFP,

33

EBFP2, Azurite, mTagBFP, ECFP, mECFP, Cerulean, mTurquoise, CyPet, AmCyan1, Midori-Ishi Cyan, TagCFP, mTFPI (Teal), Dronpa, EYFP, Topaz, Venus, mCitrine, YPet, TagYFP, PhiYFP, ZsYellow1, mBanana, Kusabira Orange, Kusabira Orange2, mOrange, mOrange2, dTomato, dTomato-Tandem, TagRFP, TagRFP-T, DsRed, DsRed2, DsRed-Express (TI), DsRed-Monomer, mTangerine, mRuby, mApple, mStrawberry, AsRed2, mRFPI, JRed, mCherry, mKate, HcRed1, mRaspberry, dKeima-Tandem, HcRed-Tandem, mPlum, and AQ143.

7. The biosensor of claim 1, wherein the first extracellular ligand binding domain and the first transmembrane domain of the first recombinant extracellular sensor are linked by a 5-25 amino acid sequence of amino acids selected from glycine, serine, and combinations thereof.

8. The biosensor of claim 1, wherein the second extracellular ligand binding domain and the second transmembrane domain of the first recombinant extracellular sensor are linked by a 5-25 amino acid sequence of amino acids selected from glycine, serine, and combinations thereof.

9. The biosensor of claim 1, wherein the first extracellular ligand binding domain and the first transmembrane domain of the first recombinant extracellular sensor are linked by a 5-25 amino acid sequence of amino acids selected from glycine, serine, and combinations thereof; and wherein the second extracellular ligand binding domain and the second transmembrane domain of the first recombinant extracellular sensor are linked by a 5-25 amino acid sequence of amino acids selected from glycine, serine, and combinations thereof.

10. The biosensor of claim 1, wherein the functional protein is an enzyme and the enzyme exhibits enzymatic activity when the ternary complex is formed and the first fragment of the enzyme and the second fragment of the enzyme interact to reconstitute the enzyme.

11. The biosensor of claim 10, wherein the enzyme is a luciferase.

12. A combination of expression cassettes for expressing the biosensor of claim 1 in a red blood cell, the combination comprising a first expression cassette and a second expression cassette, wherein the first expression cassette encodes the first protein and a second expression cassette encodes the second protein.

13. A biosensor expressed in a blood cell, comprising:

(a) a first protein comprising:

- (i) a first extracellular ligand binding domain,
- (ii) a first transmembrane domain comprising a portion of an amino acid sequence from a red blood cell native membrane protein,
- (iii) an extracellular linker comprising 7-25 amino acids and connecting the first extracellular ligand binding domain and the first transmembrane domain,
- (iv) an intracellular first fragment of a functional protein, and
- (v) an intracellular linker comprising 7-25 amino acids and connecting the first transmembrane domain and the intracellular first fragment of the functional protein; and

34

(b) a second protein comprising:

- (i) a second extracellular ligand binding domain,
- (ii) a second transmembrane domain,
- (iii) an extracellular linker comprising 7-25 amino acids and connecting the second extracellular ligand binding domain and the second transmembrane domain,
- (iv) an intracellular second fragment of a functional protein, and
- (v) an intracellular linker comprising 7-25 amino acids and connecting the second transmembrane domain and the intracellular second fragment of the functional protein;

wherein the first extracellular ligand binding domain and the second extracellular ligand binding domain bind to the same ligand, wherein the first fragment of the functional protein and the second fragment of the functional protein are capable of interacting to reconstitute activity of the functional protein, and wherein linkers comprise a sequence of amino acids selected from glycine, serine, and combinations thereof.

14. The biosensor of claim 13, wherein the intracellular linkers each comprise the sequence GGGSGGGS (SEQ ID NO: 4).

15. The biosensor of claim 13, wherein the functional protein is a fluorescent protein and the fluorescent protein emits fluorescence when the ternary complex is formed and the first fragment of the fluorescent protein and the second fragment of the fluorescent protein interact to reconstitute the fluorescent protein.

16. The biosensor of claim 15, wherein the fluorescent protein is selected from the group consisting of green fluorescent protein (GFP), EGFP, Emerald, Superfolder GFP, Azami Green, mWasabi, TagGFP, TurboGFP, AcGFP, ZsGreen, T-Sapphire, GFP-S65T, frGFP, sfGFP, EBFP, EBFP2, Azurite, mTagBFP, ECFP, mECFP, Cerulean, mTurquoise, CyPet, AmCyan1, Midori-Ishi Cyan, TagCFP, mTFPI (Teal), Dronpa, EYFP, Topaz, Venus, mCitrine, YPet, TagYFP, PhiYFP, ZsYellow1, mBanana, Kusabira Orange, Kusabira Orange2, mOrange, mOrange2, dTomato, dTomato-Tandem, TagRFP, TagRFP-T, DsRed, DsRed2, DsRed-Express (TI), DsRed-Monomer, mTangerine, mRuby, mApple, mStrawberry, AsRed2, mRFPI, JRed, mCherry, mKate, HcRed1, mRaspberry, dKeima-Tandem, HcRed-Tandem, mPlum, and AQ143.

17. The biosensor of claim 13, wherein the functional protein is an enzyme and the enzyme exhibits enzymatic activity when the ternary complex is formed and the first fragment of the enzyme and the second fragment of the enzyme interact to reconstitute the enzyme.

18. The biosensor of claim 17, wherein the enzyme is a luciferase.

19. A combination of expression cassettes for expressing the biosensor of claim 13 in a red blood cell, the combination comprising a first expression cassette and a second expression cassette, wherein the first expression cassette encodes the first protein and a second expression cassette encodes the second protein.

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