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### Compositions and methods for epigenetic regulation of HBV gene expression

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#### Abstract

This invention relates to compositions, methods, strategies, and treatment modalities related to the epigenetic modification of hepatitis B virus (HBV) genes.

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## **Background/Summary**

CROSS-REFERENCE (1) This application is a continuation of International Application No. PCT/US2024/029529, filed on May 15, 2024, which claims the benefit of U.S. Provisional Application No. 63/502,325, filed May 15, 2023, U.S. Provisional Application No. 63/516,096, filed Jul. 27, 2023, and U.S. Provisional Application No. 63/581,236, filed Sep. 7, 2023, each of which is incorporated herein by reference in its entirety.

### **SEQUENCE LISTING**

(1) The instant application contains a Sequence Listing which has been submitted electronically in XML file format and is hereby incorporated by reference in its entirety. Said XML copy, created on Jul. 11, 2024, is named 59073-720.602\_SL.xml and is 1,435,558 bytes in size.

### **BACKGROUND OF THE INVENTION**

(2) Despite available treatments, chronic hepatitis B (CHB) remains a high unmet medical need, with more than 250 million carriers of hepatitis B virus (HBV) worldwide and approximately 800,000 annual deaths due to HBV-related liver disease. Current approved CHB therapies elicit a functional cure rate (defined as durable HBsAg loss and undetectable serum HBV after completing a course of treatment) of

less than 20%. Accordingly, there is a need for improved clinical modalities targeting HBV.

## SUMMARY OF THE INVENTION

(3) Some aspects of the present disclosure provide systems, compositions, strategies, and methods for the epigenetic modification of HBV, including HBV in host cells and organisms.

(4) Some aspects of this disclosure provide methods of modifying an epigenetic state of a hepatitis B virus (HBV) gene or genome, comprising contacting the HBV gene or genome with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, optionally, wherein the first DNA binding domain binds a first target region of the HBV gene or genome, and wherein the contacting results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, and/or expression of a protein product encoded by the HBV gene or genome, wherein said reduction is at least about 20% compared to contacting the HBV gene or genome with a suitable control or without contacting the HBV gene or genome with the epigenetic editing system, and/or wherein said reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least 20%, at least 60%, at least 70%, at least 80%, at least 90% (i.e., at least a 1-log reduction), at least 95%, at least 99% (i.e., at least a 2-log reduction), or at least 99.9% (i.e., at least a 3-log reduction), compared to the number, replication, and/or expression in the subject before the contacting.

(5) Some aspects of this disclosure provide methods of treating an HBV infection in a subject comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, optionally, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the administering results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, and/or expression of a protein product encoded by the HBV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control or without administering the epigenetic editing system, and/or wherein said reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least 20%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, compared to the number, replication, and/or expression in the subject before administering.

(6) Some aspects of this disclosure provide methods of modulating expression of an HBV gene or genome comprising contacting the HBV gene or genome with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of the HBV gene or genome, and wherein the contacting results in a reduction of expression of a gene product encoded by the HBV gene or genome, optionally, wherein the gene product is a nucleic acid or a protein, wherein said reduction is at least about 20% compared to contacting the HBV genome with a suitable control or without contacting the HBV gene or genome with the epigenetic editing system, and/or wherein said reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least 20%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, compared to the number, replication, and/or expression in the subject before the contacting.

(7) Some aspects of this disclosure provide methods of inhibiting viral replication in a cell infected with an HBV comprising contacting the cell with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, optionally, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the epigenetic editing system targets a target region of the HBV gene or genome, and wherein the contacting results in a reduction of number of HBV viral episomes or replication of the HBV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control or without contacting the HBV gene or genome with the epigenetic editing system, and/or wherein said reduction of the number

of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least 20%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, compared to the number, replication, and/or expression in the subject before the contacting.

(8) Some aspects of this disclosure provide methods comprising administering an epigenetic editing system to a subject in need thereof, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the contacting results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by the HBV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control, and/or wherein said reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 20% compared to the number, replication, and/or expression in the subject before administering.

(9) Some aspects of this disclosure provide methods of inhibiting viral replication in a subject infected with an HBV comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the epigenetic editing system targets a target region of the HBV gene or genome, and wherein the administering results in a reduction of number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by an HBV gene or genome, wherein the reduction is at least about 20% compared to administering a suitable control or without administering the epigenetic editing system. In some embodiments, the reduction is at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9% compared to administering a suitable control or compared to the respective number or level in the subject before the administering. In some embodiments, the reduction is maintained for at least 6 days, for at least 19 days, for at least 27 days, for at least 42 days, or for at least 168 days.

(10) In some embodiments, the contacting further results in a reduction of a protein product. In some embodiments, the protein product comprises an HBV antigen, for example an HBe antigen (HBeAg). In some embodiments, the protein product comprises an HBs antigen (HBsAg).

(11) In some embodiments, the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA. In some embodiments, the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H. In some embodiments, the HBV genome comprises a sequence with at least 80% identity to an HBV genome sequence provided herein. In some embodiments, the first target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome provided herein. In some embodiments, the first target region of the HBV genome is located in a CpG island. In some embodiments, the first target region of the HBV genome is located in a promotor. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a preCore mRNA, a preS mRNA, a S mRNA, and a X mRNA. In some embodiments, the first DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region. In some embodiments, the gRNA comprises a sequence selected from a gRNA provided herein, e.g., in Table 12 or 13. In some embodiments, the first DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1. In some embodiments, the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein. In some embodiments, the transcriptional repressor domain comprises ZIM3. In some embodiments, the first DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the first DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the epigenetic editing system further comprises a second DNMT domain or a

nucleic acid encoding thereof. In some embodiments, the second DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the second DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the epigenetic editing system comprises a fusion protein or a nucleic acid encoding thereof, and wherein the fusion protein comprises the first DNA binding domain, the first DNMT domain, the repressor domain and the second DNMT domain. In some embodiments, the fusion protein further comprises a nuclear localization sequence (NLS). In some embodiments, the fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the epigenetic editing system further comprises a second DNA binding domain or a nucleic acid encoding thereof, wherein the second DNA binding domain binds a second target region of the HBV genome. In some embodiments, the second target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182. In some embodiments, the second target region of the HBV genome is located in a CpG island. In some embodiments, the second target region of the HBV genome is located in a promotor. In some embodiments, the second target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a preCore mRNA, a preS mRNA, a S mRNA, and a X mRNA. In some embodiments, the second DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a second gRNA that comprises a region complementary to a strand of the second target region. In some embodiments, the gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., a sequence provided in Table 12 or 13. In some embodiments, the second DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif sequence provided herein, e.g., a zinc finger motif provided in Table 1. In some embodiments, the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1. In some embodiments, the epigenetic editing system comprises a first fusion protein or a first nucleic acid encoding thereof and a second fusion protein or a second nucleic acid encoding thereof, wherein the first fusion protein comprises the first DNA binding domain and the first DNMT domain, and wherein the second fusion protein comprises the second DNA binding domain and the transcriptional repressor domain. In some embodiments, the first fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the second fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the epigenetic editing system further comprises a third DNA binding domain or a nucleic acid encoding thereof, wherein the third DNA binding domain binds to a third target region of the HBV genome. In some embodiments, the third target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182. In some embodiments, the third target region of the HBV genome is located in a CpG island. In some embodiments, the third target region of the HBV genome is located in a promotor. In some embodiments, the third target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a preCore mRNA, a preS mRNA, a S mRNA, and a X mRNA. In some embodiments, the third DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a third gRNA that comprises a region complementary to a strand of the third target region. In some embodiments, the third gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., of a gRNA sequence provided in Table 12 or 13. In some embodiments, the third DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein. In some embodiments, the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1. In some embodiments, the epigenetic editing system further comprises a second DNMT domain or a nucleic acid encoding thereof. In some embodiments, the second DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the epigenetic editing system comprises a third fusion protein or a nucleic acid encoding thereof, wherein the third fusion protein comprises the third DNA binding domain and the second DNMT domain. In some embodiments, the third fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the epigenetic editing system comprises a nucleic acid sequence provided in Table 18. In some embodiments, the reduction of the number of HBV viral episomes, of replication of the

HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 20% compared to the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome measured or observed before contacting the HBV genome with the epigenetic editing system, or before administering the epigenetic editing system to the subject. In some embodiments, the reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 25%, at least about 50%, at least about 75%, at least about 80%, at least about 90%, at least about 95%, at least about 99%, at least about 99.5%, at least about 99.8%, at least about 99.9%, at least about 99.95%, at least about 99.99%, or more than 99.99%, compared to the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome measured or observed before contacting the HBV genome with the epigenetic editing system, or before administering the epigenetic editing system to the subject.

(12) Some aspects of this disclosure provide epigenetic editing systems comprising: a fusion protein or a nucleic acid encoding the fusion protein, wherein the fusion protein comprises: (a) a DNA-binding domain that binds a target region of a HBV gene or genome, (b) a first DNA methyltransferase (DNMT) domain, and (c) a transcriptional repressor domain. In some embodiments, the epigenetic editing system is capable of reducing a number of the HBV viral episome, replication of the HBV, or expression of a gene product encoded by the HBV gene or genome, wherein said reduction is at least about 20% compared to contacting the HBV gene or genome with a suitable control. In some embodiments, the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA. In some embodiments, the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H. In some embodiments, the HBV genome comprises a sequence with at least 80% identity to an HBV genome sequence provided herein. In some embodiments, the target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome sequence provided herein. In some embodiments, the target region of the HBV genome is located in a CpG island. In some embodiments, the target region of the HBV genome is located in a promotor. In some embodiments, the target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a preCore mRNA, a preS mRNA, a S mRNA, and a X mRNA. In some embodiments, the DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a gRNA that comprises a region complementary to a strand of the target region. In some embodiments, the gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., in Table 12 or 13. In some embodiments, the DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein. In some embodiments, the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1. In some embodiments, the transcriptional repressor domain comprises a sequence of a transcriptional repressor provided herein. In some embodiments, the first DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the fusion protein further comprises a second DNMT domain. In some embodiments, the second DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the fusion protein further comprises a nuclear localization sequence (NLS). In some embodiments, the fusion protein comprises a sequence of a fusion protein provided herein.

(13) Some aspects of the present disclosure provide epigenetic editing systems comprising: a first fusion protein or a nucleic acid encoding the first fusion protein, wherein the first fusion protein comprises a first DNA binding domain and a first DNMT domain, wherein the first DNA binding domain binds a first target region of a HBV genome, and a second fusion protein or a nucleic acid encoding the second fusion protein, wherein the second fusion protein comprises a second DNA binding domain and a transcriptional repressor domain, wherein the second DNA binding domain binds a second target region of the HBV genome. In some embodiments, the epigenetic editing system is capable of reducing a

number of the HBV viral episomes, of replication of the HBV, or expression of a gene product encoded by the HBV genome, wherein said reduction is at least about 20% compared to contacting the HBV genome with a suitable control. In some embodiments, the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA. In some embodiments, the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H. In some embodiments, the HBV genome comprises a sequence with at least 80% identity to an HBV genome provided herein. In some embodiments, the epigenetic editing system further comprises a third fusion protein or a nucleic acid encoding the third fusion protein, wherein the third fusion protein comprises a third DNA binding domain and a second DNMT domain, wherein the third DNA binding domain binds a third target region of the HBV genome. In some embodiments, the first target region, the second target region or the third target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome provided herein. In some embodiments, the first target region, the second target region or the third target region of the HBV genome is located in a CpG island. In some embodiments, the first target region, the second target region or the third target region of the HBV genome is located in a promotor. In some embodiments, the first target region, the second target region or the third target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a preCore mRNA, a preS mRNA, a S mRNA, and a X mRNA. In some embodiments, the first DNA binding domain, the second DNA binding domain or the third DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a first gRNA that comprises a region complementary to a strand of the first target region, a second gRNA that comprises a region complementary to a strand of the second target region or a third RNA that comprises a region complementary to a strand of the third target region. In some embodiments, the first gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 or 13, the second gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 or 13, and/or the third gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 or 13. In some embodiments, the first DNA binding domain, the second DNA binding domain or the third DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein. In some embodiments, the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1. In some embodiments, the transcriptional repressor domain comprises ZIM3. In some embodiments, the first DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the first DNMT domain comprises a sequence of a DNMT provided herein. In some embodiments, the second DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the second DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the first fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the second fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the third fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the epigenetic editing system comprises a nucleic acid sequence provided in Table 18. In some embodiments, the reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 20% compared to the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome measured or observed before contacting the HBV genome with the epigenetic editing system, or before administering the epigenetic editing system to the subject. In some embodiments, the reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 25%, at least about 50%, at least about 75%, at least about 80%, at least about 90%, at least about 95%, at least about 99%, at least about 99.5%, at least about 99.8%, at least about 99.9%, at least about 99.95%, at least about 99.99%, or more than 99.99%, compared to the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome measured or observed before

contacting the HBV genome with the epigenetic editing system, or before administering the epigenetic editing system to the subject.

(14) Some aspects of the present disclosure provide a method of treating an HDV infection in a subject comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the contacting results in a reduction of: number of HDV viral episomes, replication of the HDV gene or genome, or expression of a protein product encoded by the HDV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control. Some aspects of the present disclosure provide a method of inhibiting viral replication in a cell infected with an HDV comprising administering an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the epigenetic editing system targets a target region of the HBV gene or genome, and wherein the contacting results in a reduction of number of HDV viral episomes or replication of the HDV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control. In some embodiments, the first DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region. In some embodiments, the gRNA comprises a sequence selected from a gRNA provided herein, e.g., in Table 12 and/or 13. In some embodiments, the first DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1 or Table 18. In some embodiments, the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein. In some embodiments, the transcriptional repressor domain comprises ZIM3. In some embodiments, the first DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the first DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the epigenetic editing system further comprises a second DNMT domain or a nucleic acid encoding thereof. In some embodiments, the second DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the second DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the epigenetic editing system comprises a fusion protein or a nucleic acid encoding thereof, and wherein the fusion protein comprises the first DNA binding domain, the first DNMT domain, the repressor domain and the second DNMT domain. In some embodiments, the fusion protein further comprises a nuclear localization sequence (NLS). In some embodiments, the fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the first DNA binding domain binds a target region of an HBV gene or genome encoding or controlling expression of an S-antigen. In some embodiments, the epigenetic editing system comprises a nucleic acid sequence provided in Table 18. In some embodiments, the reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 20% compared to the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome measured or observed before contacting the HBV genome with the epigenetic editing system, or before administering the epigenetic editing system to the subject. In some embodiments, the reduction of the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome is at least about 25%, at least about 50%, at least about 75%, at least about 80%, at least about 90%, at least about 95%, at least about 99%, at least about 99.5%, at least about 99.8%, at least about 99.9%, at least about 99.95%, at least about 99.99%, or more than 99.99%, compared to the number of HBV viral episomes, of replication of the HBV gene or genome, or of expression of a protein product encoded by the HBV gene or genome measured or observed before contacting the HBV genome with the epigenetic editing system, or before administering the epigenetic editing system to the subject.

(15) Some aspects of this disclosure provide methods comprising administering an epigenetic editing system to a subject characterized by the presence of detectable levels of HBV DNA, HBsAg, and/or HBeAg in the plasma of the subject, for example, a subject having a chronic HBV infection. In some such embodiments, the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding the same, wherein the first DNA binding domain binds a first target region of an HBV gene or genome, and the administering results in a reduction of the level of HBV DNA, the level of HBsAg, and/or the level of HBeAg in the plasma of the subject, and the reduction of the level of HBV DNA, of the level of HBsAg, and/or of the level of HBeAg in the plasma of the subject, is at least 90% (a 1-log reduction) compared to the respective level observed or observable in the plasma of the subject prior to the administering, and the 1-log reduction is maintained for at least 14 days after the administering. In some embodiments, the reduction of the level of HBV DNA in the plasma of the subject is at least 90% (a 1-log reduction). In some embodiments, the reduction of the level of HBV DNA in the plasma of the subject is at least 99% (a 2-log reduction). In some embodiments, the reduction of the level of HBsAg in the plasma of the subject is at least 90% (a 1-log reduction). In some embodiments, the reduction of the level of HBsAg in the plasma of the subject is at least 99% (a 2-log reduction). In some embodiments, the reduction of the level of HBeAg in the plasma of the subject is at least 90% (a 1-log reduction). In some embodiments, the reduction of the level of HBeAg in the plasma of the subject is at least 99% (a 2-log reduction). In some embodiments, the reduction is maintained for at least 21 days. In some embodiments, the reduction is maintained for at least 28 days. In some embodiments, the reduction is maintained for at least 35 days. In some embodiments, the reduction is maintained for at least 42 days. In some embodiments, the reduction is maintained for at least 56 days. In some embodiments, the reduction is maintained for at least 70 days. In some embodiments, the reduction is maintained for at least 84 days. In some embodiments, the reduction is maintained for at least 112 days. In some embodiments, the reduction is maintained for at least 140 days. In some embodiments, the reduction is maintained for at least 168 days. In some embodiments, the reduction is maintained for at least 6 months. In some embodiments, the reduction is maintained for at least 9 months. In some embodiments, the reduction is maintained for at least 12 months. In some embodiments, the reduction is maintained for at least 24 months. In some embodiments, the HBV genome comprises HBV genotype A. In some embodiments, the HBV genome comprises HBV genotype B. In some embodiments, the HBV genome comprises HBV genotype C. In some embodiments, the HBV genome comprises HBV genotype D. In some embodiments, the HBV genome comprises HBV genotype E. In some embodiments, the HBV genome comprises HBV genotype F. In some embodiments, the HBV genome comprises HBV genotype G. In some embodiments, the HBV genome comprises HBV genotype H. In some embodiments, the HBV genome comprises a sequence with at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99% sequence identity to an HBV genome sequence provided herein. In some embodiments, the first target region is located in a region of the HBV genome within nucleotides 0-303 of an HBV genome provided herein. In some embodiments, the first target region is located within nucleotides 0-303 of SEQ ID NO: 1082. In some embodiments, the first target region is located within nucleotides 0-303 of SEQ ID NO: 1083. In some embodiments, the first target region is located in a region of the HBV genome within nucleotides 1000-2448 of an HBV genome provided herein. In some embodiments, the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1082. In some embodiments, the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1083. In some embodiments, the first target region is located in a region of the HBV genome within nucleotides 2802-3182 of an HBV genome provided herein. In some embodiments, the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1082. In some embodiments, the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1083. In some embodiments, the first target region of the HBV genome is located in an HBV CpG island (CGI). In some embodiments, the CGI is an HBV canonical CGI. In some embodiments, the CGI is canonical CGI-I. In some embodiments, CGI is canonical CGI-I of HBV genotype D. In some embodiments, CGI-I spans nucleotides 186-288 of SEQ ID NO: 1082. In some embodiments, CGI-I spans nucleotides 186-288 of SEQ ID NO: 1083. In some embodiments, the CGI is canonical CGI-II. In some embodiments, the CGI



is canonical CGI-II HBV genotype D. In some embodiments, the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1082. In some embodiments, the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1083. In some embodiments, the CGI is canonical CGI-III. In some embodiments, the CGI is canonical CGI-III HBV genotype D. In some embodiments, the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1082. In some embodiments, the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1083. In some embodiments, the first target region of the HBV genome is located in a promoter. In some embodiments, the first target region of the HBV genome is located in the sp1 promoter. In some embodiments, the first target region of the HBV genome is located in sp2 promoter. In some embodiments, the first target region of the HBV genome is located in cp promoter. In some embodiments, the first target region of the HBV genome is located in xp promoter. In some embodiments, the first target region of the HBV genome is located in an enhancer region. In some embodiments, the first target region of the HBV genome is located in Enh I. In some embodiments, the first target region of the HBV genome is located in Enh II. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a pgRNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a preCore RNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a preS RNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes an S RNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes an HBx RNA transcript. In some embodiments, the first target region of the HBV genome is within 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) of an HBV transcription start site (TSS). In some embodiments, the TSS is a pg RNA TSS. In some embodiments, the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the pg RNA TSS. In some embodiments, the pg RNA TSS is located at nucleotide 1820 of SEQ ID NO: 1082 or at nucleotide 1820 of SEQ ID NO: 1083. In some embodiments, the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 100 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 100 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the TSS is a preC RNA TSS. In some embodiments, the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the preC RNA TSS. In some embodiments, the preC RNA TSS is located at nucleotide 1791 of SEQ ID NO: 1082 or at nucleotide 1791 of SEQ ID NO: 1083. In some embodiments, the first target region is within 600 base pairs of nucleotide 1791 in SEQ ID NO: 1082. In some embodiments, the first target region is within 600 base pairs of nucleotide 1791 in SEQ ID NO: 1083. In some embodiments, the first target region is within 500 base pairs of nucleotide 1791 in SEQ ID NO: 1082. In some embodiments, the first target region is within 500 base pairs of nucleotide 1791 in SEQ ID NO: 1083. In some embodiments, the first target region is within 400 base pairs of nucleotide 1791 in SEQ ID NO: 1082. In some embodiments, the first target region is within 400 base pairs of nucleotide 1791 in SEQ ID NO: 1083. In some embodiments, the first target region is within 300 base pairs of nucleotide 1791 in SEQ ID NO: 1082. In some embodiments, the first target region is within 300 base pairs of nucleotide 1791 in SEQ ID NO: 1083. In some embodiments, the first target region is within 200 base pairs of nucleotide 1791 in SEQ

[illegible]

some embodiments, the reduction is a reduction in a level of expression of a protein product encoded by the HBV genome. In some embodiments, the reduction is a reduction in a level of HBsAg. In some embodiments, the reduction is a reduction in a level of HBeAg. In some embodiments, the reduction is a reduction of total HBV DNA of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and the reduction is maintained for at least 14 days after the contacting or the administering. In some embodiments, the reduction is a reduction of HBeAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and the reduction is maintained for at least 14 days after the contacting or the administering. In some embodiments, the reduction is a reduction of HBsAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and the reduction is maintained at or below that level for at least 14 days after the contacting or the administering. In some embodiments, the reduction is a reduction of at least 90%. In some embodiments, the reduction is a reduction of at least 95%. In some embodiments, the reduction is a reduction of at least 99%. In some embodiments, the reduction is a reduction of at least 99.9%. In some embodiments, the reduction is maintained for at least 14 days after the contacting or the administering. In some embodiments, the reduction is maintained for at least 21 days. In some embodiments, the reduction is maintained for at least 28 days. In some embodiments, the reduction is maintained for at least 35 days. In some embodiments, the reduction is maintained for at least 42 days. In some embodiments, the reduction is maintained for at least 56 days. In some embodiments, the reduction is maintained for at least 70 days. In some embodiments, the reduction is maintained for at least 84 days. In some embodiments, the reduction is maintained for at least 112 days. In some embodiments, the reduction is maintained for at least 140 days. In some embodiments, the reduction is maintained for at least 168 days. In some embodiments, the reduction is maintained for at least 6 months. In some embodiments, the reduction is maintained for at least 7 months. In some embodiments, the reduction is maintained for at least 8 months. In some embodiments, the reduction is maintained for at least 9 months. In some embodiments, the reduction is maintained for at least 12 months. In some embodiments, the reduction is maintained for at least 18 months. In some embodiments, the reduction is maintained for at least 24 months. In some embodiments, the epigenetic editing system is administered as a monotherapy. Accordingly, in some embodiments, the method does not comprise administering a nucleoside or nucleotide analog (NUC) to the subject. In some embodiments, the method further comprises administering a NUC to the subject. In some embodiments, the first DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region. In some embodiments, the gRNA comprises a sequence selected from a gRNA provided herein, and preferably the gRNA comprises a sequence provided in Table 12 or 13. In some embodiments, the first DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1 or Table 18. In some embodiments, the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein. In some embodiments, the transcriptional repressor domain comprises ZIM3. In some embodiments, the first DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the first DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the epigenetic editing system comprises the fusion protein provided in SEQ ID NO: 1248 or the fusion protein provided in SEQ ID NO: 1252 and at least one guide RNA provided as gRNA #003, gRNA #007, gRNA #008, gRNA #009, gRNA #011, or gRNA #015 herein. Some aspects of this disclosure provide epigenetic editing systems for use in the methods described herein. In some embodiments, the epigenetic editing system comprises a fusion protein or a nucleic acid encoding the fusion protein, and the fusion protein comprises: (a) a DNA-binding domain that binds a target region of a HBV gene or genome, (b) a first DNA methyltransferase (DNMT) domain, and (c) a transcriptional repressor domain. In some embodiments, the fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the DNA-binding domain is a CRISPR-Cas DNA binding domain, and the epigenetic editing system comprises at least gRNA provided herein. In some embodiments, the epigenetic editing system comprises the fusion protein provided in SEQ ID NO: 1248 or the fusion

protein provided in SEQ ID NO: 1252 and at least one guide RNA provided as gRNA #003, gRNA #007, gRNA #008, gRNA #009, gRNA #011, or gRNA #015 herein.

(16) Other features, objectives, and advantages of the invention are apparent in the detailed description that follows. It should be understood, however, that the detailed description, while indicating embodiments and embodiments of the invention, is given by way of illustration only, not limitation. Various changes and modifications within the scope of the invention will become apparent to those skilled in the art from the detailed description.

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## Description

### BRIEF DESCRIPTION OF THE DRAWINGS

(1) FIG. 1 is a diagram illustrating an exemplary structure of a circular HBV genome. HBV genes and CpG islands are indicated. Exemplary target sites for CRISPR-based epigenetic repressors (red arrows) as well as for zinc-finger-based epigenetic repressors (green arrows) are identified.

(2) FIG. 2 is a heat map showing conservation of guide RNA target domains across different HBV genotypes.

(3) FIG. 3 is a bar graph illustrating the geographical distribution of different HBV genotypes.

(4) FIG. 4A is a diagram describing the experimental timeline for testing different CRISPR-based epigenetic repressors in HepAD38 cells, which express HPV in a doxycycline-inducible manner. FIG. 4B is a diagram showing the repression of HBV by various CRISPR-based epigenetic repressors (#1.1-3.2). Controls: UT: untransfected control; GFP: transfection control without repressor; HBV-KO: CRISPR nuclease mediated knockout; sgRNA scramble: CRISPR-based repressor with sgRNA not targeting HBV; B2M: CRISPR-based repressor with sgRNA targeting B2M.

(5) FIG. 5A is a diagram describing the experimental timeline for testing different CRISPR-based epigenetic repressors in a HepG2-NTCP infection model (see, e.g., Methods Mol Biol. 2017; 1540:1-14). FIG. 5B is a diagram showing the expression of HBe antigen (via ELISA) at different times after treatment of HBV-infected Hep2G-NTCT cells with different doses of CRISPR-based epigenetic repressors (ETRs), or with different doses of Cas9 nuclease targeting HBV (Cas9), plotted normalized to the expression value of HBe antigen measured for a negative control (empty).

(6) FIG. 6 is a diagram describing the experimental timeline for a guide RNA screen testing different CRISPR-based epigenetic repressor systems in a HepG2-NTCP infection model with ELISA readout for HBe and HBs antigens at day 6.

(7) FIG. 7 is a diagram showing QC results from different LNP batches used in the guide screen.

(8) FIG. 8 is a bar graph showing the expression of HBe and HBs for an exemplary CRISPR-based epigenetic repressor (#3.2), calculated as the percentage of the expression of the respective antigen measured for a non-targeting control.

(9) FIG. 9 is a diagram showing HBe expression values measured in the guide RNA screen for different guides (calculated as a percentage of the expression of HBe measured for a non-targeting control). Each guide/repressor combination is represented by a dot. A 50% repression cutoff is shown as a horizontal line. The position of the respective guide RNA within the HBV genome (shown at the bottom of the graph) is mapped on the X-axis. The position and the measured modulation of HBe expression for exemplary guide RNA #3.2 is indicated by red lines.

(10) FIG. 10 is a diagram showing HBs expression values measured in the guide RNA screen for different guides (calculated as a percentage of the expression of HBs measured for a non-targeting control). Each guide/repressor combination is represented by a dot. A 50% repression cutoff is shown as a horizontal line. The position of the respective guide RNA within the HBV genome (shown at the bottom of the graph) is mapped on the X-axis. The position and the measured modulation of HBs expression for exemplary guide RNA #3.2 is indicated by red lines.

(11) FIG. 11 is a diagram showing a correlation between HBs and HBe expression for the guides tested. The graph on the right shows HBe and HBs repression efficiencies for 25 exemplary guides.

(12) FIG. 12A is a diagram describing the experimental timeline for a guide RNA assay testing CRISPR-off single construct epigenetic editor in combination with individual exemplary gRNAs in a HepG2-

NTCP infection model with ELISA readout for HBe and HBs antigens at day 6; and FIG. 12B is a graph summarizing the percentage reduction in HBV antigens at day 6 relative to non-targeting control.

(13) FIG. 13A is a diagram describing the experimental timeline for a guide RNA assay testing CRISPR-off single construct epigenetic editor in combination with individual exemplary gRNAs in a PLC/PRF/5 cell model with ELISA readout for HBs antigen at day 4; and FIG. 13B is a graph summarizing the percentage reduction in HBs antigen at day 4 relative to non-targeting control.

(14) FIG. 14A is a diagram describing the experimental timeline for a guide RNA assay testing CRISPR-off single construct epigenetic editor in combination with individual exemplary gRNAs in a PXB cell model with ELISA readout for HBe and HBs antigens at day 6; and FIG. 14B is a graph summarizing the percentage reduction in HBV antigens at day 6 relative to non-targeting control. FIG. 14C is a diagram describing the experimental timeline for a guide RNA assay testing CRISPR-off single construct epigenetic editor in combination with individual exemplary gRNAs in a PXB cell model with ELISA readout for HBe and HBs antigens at day 12. FIG. 14D is a graph summarizing the percentage reduction in HBV antigens at day 12 relative to non-targeting control. Bars represent mean $\pm$ SEM; N=5. EE1=PLA002 and gRNA #007, EE2=PLA002 and gRNA #008, EE3=PLA002 and gRNA #009, EE4=PLA002 and gRNA #015, and EE5=PLA002 and gRNA #011.

(15) FIG. 15 is a diagram describing the design for in vivo experiments testing CRISPR-off single construct epigenetic editor in combination with individual exemplary gRNAs in AAV-HBV mouse HBV genotype D persistent infection model, and transgenic HBV genotype A mouse persistent infection model, respectively.

(16) FIG. 16 shows time course graphs summarizing the level of serum HBV DNA, HBs and HBe antigens in transgenic mouse HBV model before and after single administration of an epigenetic editor (CRISPR-off with gRNA or ETR with gRNA), Cas9 with gRNA, or control vehicle at day 0.

(17) FIG. 17 shows time course graphs summarizing the level of serum HBV DNA, HBs and HBe antigens in AAV-HBV mouse model before and after single administration of an epigenetic editor (CRISPR-off with gRNA or ETR with gRNA), Cas9 with gRNA, or control vehicle at day 0.

(18) FIG. 18A shows time course graphs summarizing the level of serum HBV DNA, HBs and HBe antigens in transgenic mouse HBV model, and a schematic of the timeline for the experiment. All mice received a single administration of an epigenetic editor (CRISPR-off with gRNA or ETR with gRNA), Cas9 with gRNA, or control vehicle at day 0, and some mice received a designated redosing at day 35. FIG. 18B shows results for the single-administration (no redosing) groups and controls to 168 days duration for HBV DNA and HBsAg. The lefthand panels shows the group data at each timepoint, whereas the righthand panels show the readouts for individual animals at two timepoints. EE=epigenetic editor (CRISPR-off with gRNA #011).

(19) FIG. 19 shows time course graphs summarizing the level of serum HBV DNA, HBs and HBe antigens in AAV-HBV mouse model, and a schematic of the timeline for the experiment. All mice received a single administration of an epigenetic editor (CRISPR-off with gRNA or ETR with gRNA), Cas9 with gRNA, or control vehicle at day 0, and some mice received a designated redosing at day 35.

(20) FIG. 20A is a diagram describing the experimental timeline for a zinc finger assay testing ZF-off single construct epigenetic editor that contains individual exemplary zinc finger motif in a HepG2-NTCP infection model with ELISA readout for HBe and HBs antigens at day 6; and FIG. 20B is a graph summarizing the percentage reduction in HBV antigens at day 6 relative to non-targeting control. “N” denotes non-targeting control, “P” denotes the positive control, and the individual numbers on the x-axis denote exemplary constructs tested in the experiment, for instance, “1” represents “mRNA0001” construct, and “20” represents “mRNA0020” construct.

(21) FIG. 21A is a graph summarizing the results of top ten ZF-off constructs from FIG. 20B. FIG. 21B is a diagram showing HBsAg (top) and HBeAg (middle) expression values measured in the ZF-off screen (calculated as a percentage of the expression of HBsAg or HBeAg—top and middle, respectively—measured for a non-targeting control). Each ZF-off construct is represented by a dot. 50% and 60% repression cutoffs are shown as horizontal lines. The position of the respective guide RNA within the HBV genome (bottom) is mapped on the X-axis.

(22) FIG. 22 is an experimental timeline for testing dose response (top) and two graphs showing dose

response of % HbsAg (bottom left) and % HbeAg (bottom right) in HepG2-NTCP cells upon administration of ZF fusion proteins. The mRNA corresponding to the ZF motif for each fusion protein is indicated.

(23) FIGS. 23A-23C show an experimental timeline for testing durable silencing of HBsAg (FIG. 23A), a graph showing the durability of HBsAg silencing by ZF fusion proteins (FIG. 23B), and a graph showing the durability of HBsAg silencing by CRISPR-off fusion proteins with guide RNAs (FIG. 23C) in an integrated cell line. The mRNA corresponding to the ZF motif for each fusion protein is indicated. Error bars represent mean  $\pm$  SEM; in FIG. 23C, N=3, EE1=PLA002 and gRNA #007, EE2=PLA002 and gRNA #008, EE3=PLA002 and gRNA #009, EE4=PLA002 and gRNA #015, and EE5=PLA002 and gRNA #011).

(24) FIG. 24 is an experimental timeline for testing HBsAg silencing in a PLC/PRF/5 in vitro model (top) and a graph showing % HBsAg relative to control on Day 14 after administration of ZF fusion proteins. The mRNA corresponding to the ZF motif for each fusion protein is indicated. Information about the % match to target for each construct is also indicated.

(25) FIG. 25A is a volcano plot showing differentially expressed (DE) genes for an exemplary ZF specificity assay. DE genes are shown with dots. FIG. 25B is a volcano plot showing DE for CRISPR-off and gRNA epigenetic editors. Points represent genes with their change in expression (x-axis) and statistical significance of that change (y-axis). EE1=PLA002 and gRNA #007, EE2=PLA002 and gRNA #008, EE3=PLA002 and gRNA #009, EE4=PLA002 and gRNA #015, and EE5=PLA002 and gRNA #011. Also shown are results for low specificity and host target gene controls. FIGS. 25C-25D are scatter plots showing methylation levels between treatment (y-axis) and control (x-axis) for 935,000 CpG sites in the human genome. Lines represent thresholds for changes in methylation considered significant (absolute [methylation difference] $\geq$ 0.2). DMRs are noted on each figure. Results for a host target (PCSK9, next-to-final panel) as well as a low specificity control (final panel) are also shown. FIG. 25C shows the results versus effector only; FIG. 25D shows the results versus no treatment.

EE1=PLA002 and gRNA #007, EE2=PLA002 and gRNA #008, EE3=PLA002 and gRNA #009, EE4=PLA002 and gRNA #015, EE5=PLA002 and gRNA #011, EE6=PLA002 and gRNA #003, and EE7=PLA002 and gRNA #016.

(26) FIG. 26 is a schematic of an in vivo experiment testing ZF-off constructs.

(27) FIG. 27 shows graphs showing log fold change, relative to baseline, for HBV DNA (left), HBsAg (middle), and HBeAg (right) in plasma of mice treated with the plasmids indicated in the experiment shown in FIG. 26.

(28) FIG. 28 is an experimental schematic for an in vivo study of multiplexing ZF fusion protein effectors.

(29) FIG. 29 is a schematic for a dose response experiment using CRISPR-Off in an AAV-HBV in vivo model.

(30) FIG. 30 is a line graph of plasma HBsAg levels for a dose response experiment using CRISPR-Off in an AAV-HBV in vivo model.

(31) FIG. 31 is a schematic for a dose response experiment using CRISPR-Off in a Tg-HBV in vivo model.

(32) FIG. 32 shows line graphs of plasma HBV DNA, HBsAg, and HBeAg levels for a dose response experiment using CRISPR-Off in a Tg-HBV in vivo model.

(33) FIG. 33 is a dot plot of HBsAg levels of individual mice at the 207 day time point of a dose response experiment using CRISPR-Off in a Tg-HBV in vivo model.

(34) FIG. 34 shows line graphs of HBV-DNA and HBsAg in plasma in AAV mice treated with CRISPR-Off mRNA with various single guide RNAs. n=5 for each guide RNA treatment group; n=4 for vehicle-only control.

(35) FIG. 35A shows line graphs of HBV-DNA and HBsAg in plasma in AAV mice treated with a single dose of ZF-Off mRNA.

(36) FIG. 35B shows line graphs of HBV-DNA and HBsAg in plasma in AAV mice treated with multiple doses of ZF-Off mRNA.

(37) FIG. 36 shows line graphs of HBV-DNA, HBsAg, and HBeAg in plasma in AAV mice treated with

single versus multiple doses of 1 mg/kg CRISPR-Off mRNA with guide RNA.

(38) FIG. 37 shows line graphs of HBV-DNA and HBsAg in plasma in AAV mice treated with a single bolus dose of 3 mg/kg versus three doses of 1 mg/kg CRISPR-Off mRNA with guide RNA.

(39) FIG. 38 shows line graphs of HBsAg in plasma in response to treatment with two different CRISPR-Off effectors (left, SEQ ID NO: 1248; right, SEQ ID NO: 1252) delivered via mRNA in combination with the same guide RNA.

(40) FIGS. 39A-39G show methylation of the HBV genome upon treatment with CRISPR-Off with various single guide RNAs versus wild type Cas9, CRISPRi, and non-targeting controls. The box in FIG. 39A represents the region 500 bp both upstream and downstream of the target site. The arrows indicate the position of the target sequence for the guide RNA used in the depicted experiment.

(41) FIG. 40 shows volcano plots of RNA-Seq (top) and methylation (bottom) experiments at Day 14 after treatment in HepG2.2.15 cells treated with ZF-Off (left, SEQ ID NO: 36; center, SEQ ID NO: 73) and CRISPR-Off (right, SEQ ID NO: 1248) constructs (delivered as mRNA) targeting HBV. DE, differentially expressed. DMR: differentially methylated region.

(42) FIG. 41 shows HBsAg levels over 14 days for the cells treated for the RNA-Seq and methylation plots in FIG. 40.

(43) FIG. 42 shows a schematic (top) and dose curves (bottom) for CRISPR-Off dose curve experiments in HepG2.2.15 cells using various single guide RNAs and measuring HBsAg and HBeAg.

(44) FIG. 43 shows dose curves for a CRISPR-Off variant, delivered with guide RNA, in HepG2.2.15 cells measuring HBsAg and HBeAg.

#### DETAILED DESCRIPTION OF THE INVENTION

(45) The present disclosure provides epigenetic editors, and strategies and methods of using such epigenetic editors, for regulating expression of HBV. By altering expression of HBV, and in particular, by repressing expression of HBV, e.g., of a gene comprised in the HBV genome or a gene product encoded by the HBV genome, the compositions and methods described herein are useful to suppress viral function in infected cells, e.g., in the context of treating an HBV infection in a human subject, or in the context of treating CHB.

(46) The structure and biology of HBV as well as HBV-associated diseases have been reported (see, for example, Yuen, M F., Chen, D S., Dusheiko, G. et al. Hepatitis B virus infection. *Nat Rev Dis Primers* 4, 18035 (2018), incorporated herein by reference in its entirety).

(47) Exemplary HBV sequences can be found at various NCBI database entries, e.g., representative sequences can be found under accession numbers NC\_00397 and U95551, which are incorporated herein by reference in their entirety, and the sequences of which are provided elsewhere herein.

(48) A number of treatment options for HBV has been reported, but there remains a need for effective treatment of HBV infections. Genetic editing approaches targeting HBV genomes for cutting of genomic DNA are associated with a risk of off-target cutting and genomic translocations. The present epigenetic editors and related methods of use have several advantages compared to other genome engineering methods, including increased efficiency, decreased risk of translocation, and durable silencing of HBV.

(49) The present disclosure also provides methods for treating Hepatitis D virus (HDV). HDV is the smallest pathogen known to infect humans. HDV infection is only found in patients infected with HBV, as HDV relies on HBV functions for most of its functions, including viral packaging, infectivity, transmission, and inhibition of host immunity. About 5% of patients with HBV infection also have an HDV infection. HDV uses HBV S-antigen (HBsAg) as a capsid protein, and HDV infection is therefore dependent on HBV S-antigen production. Decreasing HBV S-antigen expression also reduces HDV infectivity. The structure and biology of HDV has been reported (see, for example, Asselah and Rizzetto, *Hepatitis D Virus Infection*, *The New England Journal of Medicine* (389; 1; Jul. 6, 2023), incorporated herein by reference in its entirety). In some embodiments of the present disclosure, HDV infection is addressed through methods targeting an HBV gene or genome that reduce the level of HBsAg.

(50) In some embodiments, an epigenetic editor as described herein may comprise one or more fusion proteins, wherein each fusion protein comprises a DNA-binding domain linked to one or more effector domains for epigenetic modification. In certain embodiments, where the DNA-binding domain is a polynucleotide guided DNA-binding domain, the epigenetic editor may further comprise one or more

guide polynucleotides. DNA-binding domains, effector domains, and guide polynucleotides of an epigenetic editor as described herein may be selected, e.g., from those described below, in any functional combination.

(51) The epigenetic editors described herein may be expressed in a host cell transiently, or may be integrated in a genome of the host cell; such cells and their progeny are also contemplated by the present disclosure. Both transiently expressed and integrated epigenetic editors or components thereof can effect stable epigenetic modifications. For example, after introducing to a host cell an epigenetic editor described herein, the target gene in the host cell may be stably or permanently repressed or silenced. For example, in some embodiments provided herein, a transiently expressed epigenetic editor comprising a DNMT3A domain, a DNMT3L domain, and a KRAB domain effects stable epigenetic modifications. For example, in some embodiments provided herein, a constitutively expressed epigenetic editor comprising DNMT3A and a DNMT3L domain effects stable epigenetic modifications. In some embodiments, expression of the target gene is reduced or silenced for at least 1 week, at least 2 weeks, at least 3 weeks, at least 4 weeks, at least 5 weeks, at least 6 weeks, at least 7 weeks, at least 2 months, at least 3 months, at least 4 months, at least 5 months, at least 6 months, at least 1 year, at least 2 years, or for the entire lifetime of the cell or the subject carrying the cell, as compared to the level of expression in the absence of the epigenetic editor. The epigenetic modification may be inherited by the progeny of the host cells into which the epigenetic editor was introduced. In some embodiments, the host cell is a liver cell characterized by the presence of an HBV genome in the cell.

(52) The present epigenetic editors may be introduced to a patient in need thereof (e.g., a human patient), e.g., into the patient's hepatocytes, biliary epithelial cells (cholangiocytes), stellate cells, Kupffer cells, and liver sinusoidal endothelial cells.

#### I. DNA-Binding Domains

(53) An epigenetic editor described herein may comprise one or more DNA-binding domains that direct the effector domain(s) of the epigenetic editor to target sequences within an HBV genome. A DNA-binding domain as described herein may be, e.g., a polynucleotide guided DNA-binding domain, a zinc finger protein (ZFP) domain, a transcription activator like effector (TALE) domain, a meganuclease DNA-binding domain, and the like. Examples of DNA-binding domains can be found in U.S. Pat. No. 11,162,114, which is incorporated by reference herein in its entirety.

(54) In some embodiments, a DNA-binding domain described herein is encoded by its native coding sequence. In other embodiments, the DNA-binding domain is encoded by a nucleotide sequence that has been codon-optimized for optimal expression in human cells.

##### (55) A. Polynucleotide Guided DNA-Binding Domains

(56) In some embodiments, a DNA-binding domain herein may be a protein domain directed by a guide nucleic acid sequence (e.g., a guide RNA sequence) to a target site in an HBV genome. In certain embodiments, the protein domain may be derived from a CRISPR-associated nuclease, such as a Class I or II CRISPR-associated nuclease. In some embodiments, the protein domain may be derived from a Cas nuclease such as a Type II, Type IIA, Type IIB, Type IIC, Type V, or Type VI Cas nuclease. In certain embodiments, the protein domain may be derived from a Class II Cas nuclease selected from Cas1, Cas1B, Cas2, Cas3, Cas4, Cas5, Cas6, Cas7, Cas8, Cas9, Cas10, Cas14a, Cas14b, Cas14c, CasX, CasY, CasPhi, C2c4, C2c8, C2c9, C2c10, Csy1, Csy2, Csy3, Cse1, Cse2, Csc1, Csc2, Csa5, Csn2, Csm2, Csm3, Csm4, Csm5, Csm6, Cmr1, Cmr3, Cmr4, Cmr5, Cmr6, Csb1, Csb2, Csb3, Csx17, Csx14, Csx10, Csx16, CsaX, Csx3, Csx1, Csx1S, Csf1, Csf2, CsO, Csf4, and homologues and modified versions thereof. "Derived from" is used to mean that the protein domain comprises the full polypeptide sequence of the parent protein, or comprises a variant thereof (e.g., with amino acid residue deletions, insertions, and/or substitutions). The variant retains the desired function of the parent protein (e.g., the ability to form a complex with the guide nucleic acid sequence and the target DNA).

(57) In some embodiments, the CRISPR-associated protein domain may be a Cas9 domain described herein. Cas9 may, for example, refer to a polypeptide with at least about 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% sequence identity and/or sequence similarity to a wildtype Cas9 polypeptide described herein. In some embodiments, said wildtype polypeptide is Cas9 from *Streptococcus pyogenes* (NCBI Ref. No. NC\_002737.2 (SEQ ID NO: 1)) and/or UniProt Ref. No.



Q99ZW2 (SEQ ID NO: 2). In some embodiments, said wildtype polypeptide is Cas9 from *Staphylococcus aureus* (SEQ ID NO: 3). In some embodiments, the CRISPR-associated protein domain is a Cpf1 domain or protein, or a polypeptide with at least about 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% sequence identity and/or sequence similarity to a wildtype Cpf1 polypeptide described herein (e.g., Cpf1 from *Francisella novicida* (UniProt Ref. No. U2UMQ6 or SEQ ID NO: 4). In certain embodiments, the CRISPR-associated protein domain may be a modified form of the wildtype protein comprising one or more amino acid residue changes such as a deletion, an insertion, or a substitution; a fusion or chimera; or any combination thereof.

(58) Cas9 sequences and structures of variant Cas9 orthologs have been described for various organisms. Exemplary organisms from which a Cas9 domain herein can be derived include, but are not limited to, *Streptococcus pyogenes*, *Streptococcus thermophilus*, *Streptococcus* sp., *Staphylococcus aureus*, *Listeria innocua*, *Lactobacillus gasseri*, *Francisella novicida*, *Wolinella succinogenes*, *Sutterella wadsworthensis*, *Gamma proteobacterium*, *Neisseria meningitidis*, *Campylobacter jejuni*, *Pasteurella multocida*, *Fibrobacter succinogene*, *Rhodospirillum rubrum*, *Nocardiopsis dassonvillei*, *Streptomyces pristinaespiralis*, *Streptomyces viridochromogenes*, *Streptomyces viridochromogenes*, *Streptosporangium roseum*, *Alicyclobacillus acidocaldarius*, *Bacillus pseudomycolides*, *Bacillus selenitireducens*, *Exiguobacterium sibiricum*, *Lactobacillus delbrueckii*, *Lactobacillus salivarius*, *Lactobacillus buchneri*, *Treponema denticola*, *Microscilla marina*, *Burkholderiales bacterium*, *Polaromonas naphthalenivorans*, *Polaromonas* sp., *Crocospaera watsonii*, *Cyanotheca* sp., *Microcystis aeruginosa*, *Synechococcus* sp., *Acetohalobium arabaticum*, *Ammonifex degensii*, *Caldicelulosiruptor beccii*, *Candidatus Desulforudis*, *Clostridium botulinum*, *Clostridium difficile*, *Fingoldia magna*, *Natronaerobius thermophilus*, *Pelotomaculum thermopropionium*, *Acidithiobacillus caldus*, *Acidithiobacillus ferrooxidans*, *Allochrochromatium vinosum*, *Marinobacter* sp., *Nitrosococcus halophilus*, *Nitrosococcus watsoni*, *Pseudoalteromonas haloplanktis*, *Ktedonobacter racemifer*, *Methanohalobium evestigatum*, *Anabaena variabilis*, *Nodularia spumigena*, *Nostoc* sp., *Arthrospira maxima*, *Arthrospira platensis*, *Arthrospira* sp., *Lyngbya* sp., *Microcoleus chthonoplastes*, *Oscillatoria* sp., *Petrogloa mobilis*, *Thermosiphon africanus*, *Streptococcus pasteurianus*, *Neisseria cinerea*, *Campylobacter lari*, *Parvibaculum lavamentivorans*, *Corynebacterium diphtheriae*, and *Acaryochloris marina*. Cas9 sequences also include those from the organisms and loci disclosed in Chylinski et al., RNA Biol. (2013) 10(5):726-37.

(59) In some embodiments, the Cas9 domain is from *Streptococcus pyogenes*. In some embodiments, the Cas9 domain is from *Staphylococcus aureus*.

(60) Other Cas domains are also contemplated for use in the epigenetic editors herein. These include, for example, those from CasX (Cas12E) (e.g., SEQ ID NO: 5), CasY (Cas12d) (e.g., SEQ ID NO: 6), CasΦ (CasPhi) (e.g., SEQ ID NO: 7), Cas12f1 (Cas14a) (e.g., SEQ ID NO: 8), Cas12f2 (Cas14b) (e.g., SEQ ID NO: 9), Cas12f3 (Cas14c) (e.g., SEQ ID NO: 10), and C2c8 (e.g., SEQ ID NO: 11).

(61) For epigenetic editing, the nuclease-derived protein domain (e.g., a Cas9 or Cpf1 domain) may have reduced or no nuclease activity through mutations such that the protein domain does not cleave DNA or has reduced DNA-cleaving activity while retaining the ability to complex with the guide nucleic acid sequence (e.g., guide RNA) and the target DNA. For example, the nuclease activity may be reduced by at least 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or 99% compared to the wildtype domain. In some embodiments, a CRISPR-associated protein domain described herein is catalytically inactive ("dead"). Examples of such domains include, for example, dCas9 ("dead" Cas9), dCpf1, ddCpf1, dCasPhi, ddCas12a, dLbCpf1, and dFnCpf1. A dCas9 protein domain, for example, may comprise one, two, or more mutations as compared to wildtype Cas9 that abrogate its nuclease activity. The DNA cleavage domain of Cas9 is known to include two subdomains: the HNH nuclease subdomain and the RuvC1 subdomain. The HNH subdomain cleaves the strand complementary to the gRNA, whereas the RuvC1 subdomain cleaves the non-complementary strand. Mutations within these subdomains can silence the nuclease activity of Cas9. For example, the mutations D10A (in RuvC1) and H840A (in HNH) completely inactivate the nuclease activity of SpCas9. SaCas9, similarly, may be inactivated by the mutations D10A and N580A. In some embodiments, the dCas9 comprises at least one mutation in the HNH subdomain and/or the RuvC1

subdomains that reduces or abrogates nuclease activity. In some embodiments, the dCas9 only comprises a RuvC1 subdomain, or only comprises an HNH subdomain. It is to be understood that any mutation that inactivates the RuvC1 and/or the HNH domain may be included in a dCas9 herein, e.g., insertion, deletion, or single or multiple amino acid substitution in the RuvC1 domain and/or the HNH domain.

(62) In some embodiments, a dCas9 protein herein comprises a mutation at position(s) corresponding to position D10 (e.g., D10A), H840 (e.g., H840A), or both, of a wildtype SpCas9 sequence as numbered in the sequence provided at UniProt Accession No. Q99ZW2 (SEQ ID NO: 2). In particular embodiments, the dCas9 comprises the amino acid sequence of dSpCas9 (D10A and H840A) (SEQ ID NO: 12).

(63) In some embodiments, a dCas9 protein as described herein comprises a mutation at position(s) corresponding to position D10 (e.g., D10A), N580 (e.g., N580A), or both, of a wildtype SaCas9 sequence (e.g., SEQ ID NO: 9). In particular embodiments, the dCas9 comprises the amino acid sequence of dSaCas9 (D10A and N580A) (SEQ ID NO: 13).

(64) Additional suitable mutations that inactivate Cas9 will be apparent to those of skill in the art based on this disclosure and knowledge in the field and are within the scope of this disclosure. Such mutations may include, but are not limited to, D839A, N863A, and/or K603R in SpCas9. The present disclosure contemplates any mutations that reduce or abrogate the nuclease activity of any Cas9 described herein (e.g., mutations corresponding to any of the Cas9 mutations described herein).

(65) A dCpf1 protein domain may comprise one, two, or more mutations as compared to wildtype Cpf1 that reduce or abrogate its nuclease activity. The Cpf1 protein has a RuvC-like endonuclease domain that is similar to the RuvC domain of Cas9, but does not have an HNH endonuclease domain, and the N-terminal of Cpf1 does not have the alpha-helical recognition lobe of Cas9. In some embodiments, the dCpf1 comprises one or more mutations corresponding to position D917A, E1006A, or D1255A as numbered in the sequence of the *Francisella novicida* Cpf1 protein (FnCpf1; SEQ ID NO: 4). In certain embodiments, the dCpf1 protein comprises mutations corresponding to D917A, E1006A, D1255A, D917A/E1006A, D917A/D1255A, E1006A/D1255A, or D917A/E1006A/D1255A, or corresponding mutation(s) in any of the Cpf1 amino acid sequences described herein. In some embodiments, the dCpf1 comprises a D917A mutation. In particular embodiments, the dCpf1 comprises the amino acid sequence of dFnCpf1 (SEQ ID NO: 14).

(66) Further nuclease inactive CRISPR-associated protein domains contemplated herein include those from, for example, dNmeCas9 (e.g., SEQ ID NO: 15), dCjCas9 (e.g., SEQ ID NO: 16), dSt1Cas9 (e.g., SEQ ID NO: 17), dSt3Cas9 (e.g., SEQ ID NO: 18), dLbCpf1 (e.g., SEQ ID NO: 19), dAsCpf1 (e.g., SEQ ID NO: 20), denAsCpf1 (e.g., SEQ ID NO: 21), dHFAsCpf1 (e.g., SEQ ID NO: 22), dRVRAcCpf1 (e.g., SEQ ID NO: 23), dRRAsCpf1 (e.g., SEQ ID NO: 24), dCasX (e.g., SEQ ID NO: 25), and dCasPhi (e.g., SEQ ID NO: 26).

(67) In some embodiments, a Cas9 domain described herein may be a high fidelity Cas9 domain, e.g., comprising one or more mutations that decrease electrostatic interactions between the Cas9 domain and the sugar-phosphate backbone of DNA to confer increased target binding specificity. In certain embodiments, the high fidelity Cas9 domain may be nuclease inactive as described herein.

(68) A CRISPR-associated protein domain described herein may recognize a protospacer adjacent motif (PAM) sequence in a target gene. A “PAM” sequence is typically a 2 to 6 bp DNA sequence immediately following the sequence targeted by the CRISPR-associated protein domain. The PAM sequence is required for CRISPR protein binding and cleavage but is not part of the target sequence. The CRISPR-associated protein domain may either recognize a naturally occurring or canonical PAM sequence or may have altered PAM specificity. CRISPR-associated protein domains that bind to non-canonical PAM sequences have been described in the art. For example, Cas9 domains that bind non-canonical PAM sequences have been described in Kleinstiver et al., Nature (2015) 523(7561):481-5 and Kleinstiver et al., Nat Biotechnol. (2015) 33:1293-8. Such Cas9 domains may include, for example, those from “VRER (SEQ ID NO: 1261)” SpCas9, “EQR” SpCas9, “VQR” SpCas9, “SpG Cas9,” “SpRYCas9,” and “KKH” SaCas9. Nuclease inactive versions of these Cas9 domains are also contemplated, such as nuclease inactive VRER (SEQ ID NO: 1261) SpCas9 (e.g., SEQ ID NO: 27), nuclease inactive EQR SpCas9 (e.g., SEQ ID NO: 28), nuclease inactive VQR SpCas9 (e.g., SEQ ID NO: 29), nuclease inactive SpG Cas9 (e.g., SEQ ID NO: 30), nuclease inactive SpRY Cas9 (e.g., SEQ ID NO: 31), and nuclease

inactive KKH SaCas9 (e.g., SEQ ID NO: 32). Another example is the Cas9 of *Francisella novicida* engineered to recognize 5'-YG-3' (where "Y" is a pyrimidine).

(69) Additional suitable CRISPR-associated proteins, orthologs, and variants, including nuclease inactive variants and sequences, will be apparent to those of skill in the art based on this disclosure.

(70) Guide RNAs that can be used in conjunction with the CRISPR-associated protein domains herein are further described in Section II below.

(71) B. Zinc Finger Protein Domains

(72) In some embodiments, the DNA-binding domain of an epigenetic editor described herein comprises a zinc finger protein (ZFP) domain (or "ZF domain" as used herein). ZFPs are proteins having at least one zinc finger, and bind to DNA in a sequence-specific manner. A "zinc finger" (ZF) or "zinc finger motif" (ZF motif) refers to a polypeptide domain comprising a beta-beta-alpha ( $\beta\beta\alpha$ )-protein fold stabilized by a zinc ion. A ZF binds from two to four base pairs of nucleotides, typically three or four base pairs (contiguous or noncontiguous). Each ZF typically comprises approximately 30 amino acids. ZFP domains may contain multiple ZFs that make tandem contacts with their target nucleic acid sequence. A tandem array of ZFs may be engineered to generate artificial ZFPs that bind desired nucleic acid targets. ZFPs may be rationally designed by using databases comprising triplet (or quadruplet) nucleotide sequences and individual ZF amino acid sequences, in which each triplet or quadruplet nucleotide sequence is associated with one or more amino acid sequences of ZFs that bind the particular triplet or quadruplet sequence. See, e.g., U.S. Pat. Nos. 6,453,242, 6,534,261, and 8,772,453.

(73) ZFPs are widespread in eukaryotic cells, and may belong to, e.g., C2H2 class, CCHC class, PHD class, or RING class. An exemplary motif characterizing one class of these proteins (C2H2 class) is -Cys-(X).sub.2-4-Cys-(X).sub.12-His-(X).sub.3-5-His-(SEQ ID NO:1091), where X is any independently chosen amino acid. In some embodiments, a ZFP domain herein may comprise a ZF array comprising sequential C2H2-ZFs each contacting three or more sequential nucleotides. Additional architectures, e.g. as described in Paschon et al., Nat. Commun. 10, 1133 (2019), are also possible.

(74) A ZFP domain of an epigenetic editor described herein may include 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, or more ZFs. The ZFP domain may include an array of two-finger or three-finger units, e.g., 3, 4, 5, 6, 7, 8, 9 or 10 or more units, wherein each unit binds a subsite in the target sequence. In some embodiments, a ZFP domain comprising at least three ZFs recognizes a target DNA sequence of 9 or 10 nucleotides. In some embodiments, a ZFP domain comprising at least four ZFs recognizes a target DNA sequence of 12 to 14 nucleotides. In some embodiments, a ZFP domain comprising at least six ZFs recognizes a target DNA sequence of 18 to 21 nucleotides.

(75) In some embodiments, ZFs in a ZFP domain described herein are connected via peptide linkers. The peptide linkers may be, e.g., 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more amino acids in length. In some embodiments, a linker comprises 5 or more amino acids. In some embodiments, a linker comprises 7-17 amino acids. The linker may be flexible or rigid.

(76) In some embodiments a zinc finger array may have the sequence:

(77) TABLE-US-00001 SRPGERPFQCRICMRNFSXXXXXXXXHXXTHTGEKPFQCRICMRNF  
XXXXXXXXHXXTH[linker]FQCRICMRNFSXXXXXXXXHXXTHT  
GEKPFQCRICMRNFSXXXXXXXXHXXTH[linker]PFQCRICMRN

FSXXXXXXXXHXXTHTGEKPFQCRICMRNFSXXXXXXXXHXXTHLRG S (SEQ ID NOs:  
1084 and 1258-1259, respectively, in order of appearance),

or a sequence at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical thereto,

where "XXXXXXXX" represents the amino acids of the ZF recognition helix, which confers DNA-binding specificity upon the zinc finger; each X may be independently chosen. In the above sequence, "XX" in italics may be TR, LR or LK, and "[linker]" represents a linker sequence. In some

embodiments, the linker sequence is TGSQKP (SEQ ID NO: 1085); this linker may be used when sub-sites targeted by the ZFs are adjacent. In some embodiments, the linker sequence is TGGGGSQKP (SEQ ID NO: 1086); this linker may be used when there is a base between the sub-sites targeted by the zinc fingers. The two indicated linkers may be the same or different.

(78) ZFP domains herein may contain arrays of two or more adjacent ZFs that are directly adjacent to one another (e.g., separated by a short (canonical) linker sequence), or are separated by longer, flexible

or structured polypeptide sequences. In some embodiments, directly adjacent fingers bind to contiguous nucleic acid sequences, i.e., to adjacent trinucleotides/triplets. In some embodiments, adjacent fingers cross-bind between each other's respective target triplets, which may help to strengthen or enhance the recognition of the target sequence, and leads to the binding of overlapping sequences. In some embodiments, distant ZFs within the ZFP domain may recognize (or bind to) non-contiguous nucleotide sequences.

(79) The amino acid sequences of the ZF DNA-recognition helices of exemplary ZFP domains herein, and their HBV target sequences, are shown below in Table 1. Table 1. Zinc finger transcriptional repressors for silencing HBV. ZF sequences of exemplary ZFP domains are presented. SEQ ID Nos for target sequences and ZF can be found in Table 18 sequence listing.

(80) TABLE-US-00002 TABLE 18 sequence listing. SEQ Target ZFP ID Sequence Start End Strd F1 F2 F3 F4 F5 F6 ZFP894 33 GATGAGGCAT 415 432 – KKEN RQDN RSHN QSTT RNTN IKHN AGCAGCAG LLQ LNS LKL LKR LTR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 102) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 125) 156) 189) 222) 257) 297) ZFP895 34 GATGAGGCAT 415 432 – KKEN RKDY RSHN QSTT RQDN VVNN AGCAGCAG LLQ LIS LKI LKR LGR LNR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 102) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 125) 157) 189) 222) 258) 298) ZFP896 35 GATGAGGCAT 415 432 – KKEN RKDY RSHN QSTT RQDN VVNN AGCAGCAG LLO LIS LRL LKR LGR LNR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 102) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 125) 157) 190) 222) 258) 298) ZFP899 36 GATGATTAGG 1828 1845 – RRHI RQDN QSTT RRDG VHHN ISHN CAGAGGTG LDR LGR LKR LAG LVR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 103) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 126) 158) 191) 223) 259) 299) ZFP900 37 GATGATTAGG 1828 1845 – RREV RRDN QSTT RRDG VHHN ISHN CAGAGGTG LEN LNR LKR LAG LVR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 103) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 127) 159) 191) 223) 259) 299) ZFP901 38 GATGATTAGG 1828 1845 – RRAV RQDN QSTT RRDG VHHN ISHN CAGAGGTG LDR LGR LKR LAG LVR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 103 ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 128) 158) 191) 223) 259) 299) ZFP902 39 GGATTCAGCG 1433 1450 – RQEH EGGN SDRR SFQS RPNH QSPH CCGACGGG LVR LMR DLD YLE LAI LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 104) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 129) 160) 192) 224) 260) 300) ZFP903 40 GGATTCAGCG 1433 1450 – RREH DPSN SDRR SFQS RPNH QSPH CCGACGGG LVR LOR DLD YLE LAI LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 104) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 130) 161) 192) 224) 260) 300) ZFP904 41 GGATTCAGCG 1433 1450 – RREH DMGN SDRR SFQS RPNH QSPH CCGACGGG LVR LGR DLD YLE LAI LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 104) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 130) 162) 192) 224) 260) 300) ZFP907 42 GGCAGTAGTC 90 108 – KKDH QKEI QSAH ETGS QSHS ESGH GGAACAGGG LHR LTR LKR LRR LKS LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 105) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 131) 163) 193) 225) 261) 301) ZFP908 43 GGCAGTAGTC 90 108 – KKDH QKEI QSAH DRTP QSHS ESGH GGAACAGGG LHR LTR LKR LNR LKS LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 105) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 131) 163) 193) 226) 261) 301) ZFP909 44 GGCAGTAGTC 90 108 – KTDH QKEI QSAH ETGS QKHH ENSK GGAACAGGG LAR LTR LKR LRR LVT LRR (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 132) 163) 193) 225) 262) 302) ZFP912 45 GTAAACTGAG 664 682 – QAGN QNSH DLST QNEH GGTA QRSS CCAGGAGAA LVR LRR LRR LKV LRM LVR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 106) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 133) 164) 194) 227) 263) 303) ZFP913 46 GTAAACTGAG 664 682 – QRGN QTTH DGST QKTH GGTA QRSS CCAGGAGAA LQR LSR LRR LAV LRM LVR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 106) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 134) 165) 195) 228) 263) 303) ZFP914 47 GTAAACTGAG 664 682 – QRGN QTTH DLST QNEH GGSA QRSS CCAGGAGAA LQR LSR LRR LKV LSM LVR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 106) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 134) 165) 194) 227) 264) 303) ZFP930 48 ACGGTGGTCT 1605 1623 – DRGN QARS EKAS DHSS RRFI RNDS

CCATGCGAC LTR LRA LKR LSK (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ NO: 107) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 135) 166) 196) 229) 265) 304) ZFP931 49  
ACGGTGGTCT 1605 1623 – DRGN QARS DKSS DHSS RNFI RNDT CCATGCGAC LTR LRA LRK  
LKR LQR LII (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 107) ID ID ID ID ID ID NO: NO:  
NO: NO: NO: NO: 135) 166) 197) 229) 266) 305) ZFP932 50 ACGGTGGTCT 1605 1623 – DRGN  
QARS CNGS DHSS RNFI RNDT CCATGCGAC LTR LRA LKK LKR LQR LII (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 107) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 135) 166)  
198) 229) 266) 305) ZFP933 51 GCTGGATGTG 372 393 + RTDT RTDS DHSS QPHG QSAH VGNS  
TCTGCGGCG LAR LPR LKR LAH LKR LSR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
108) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 136) 167) 199) 230) 267) 306) ZFP934 52  
GCTGGATGTG 372 393 + RTDT RTDS DHSS QPHG QSAH VGNS TCTGCGGCG LAR LPR LKR  
LRH LKR LSR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 108) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: 136) 167) 199) 231) 267) 306) ZFP935 53 GCTGGATGTG 372 393 + RTDT  
RLDM DHSS QPHG QQAQ VHES TCTGCGGCG LAR LAR LKR LST LVR LKR (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 108) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 136) 168)  
199) 232) 268) 307) ZFP938 54 GTCTGCGAGG 2381 2398 – RADN RNTH RGDG RRDN RARN  
DPSS CGAGGGAG LGR LSY LRR LNR LTL LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
109) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 137) 169) 200) 233) 269) 308) ZFP939 55  
GTCTGCGAGG 2381 2398 – RADN RNTH RKLK RQDN RARN DPSS CGAGGGAG LGR LSY  
LLR LGR LTL LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 109) ID ID ID ID ID ID  
NO: NO: NO: NO: NO: NO: 137) 169) 201) 234) 269) 308) ZFP940 56 GTCTGCGAGG 2381 2398 –  
RADN RNTH RKLK RODN RRRN DHSS CGAGGGAG LGR LSY LLR LGR LQL LKR (SEQ ID  
(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 109) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 137)  
169) 201) 234) 270) 309) ZFP943 57 GTTGCCGGGC 1146 1164 – QQSS RREH GLTA ERAK AKRD  
VNSS AACGGGGTA LLR LVR LRT LIR LDR LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
110) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 138) 170) 202) 235) 271) 310) ZFP944 58  
GTTGCCGGGC 1146 1164 – QQSS RREH GLTA ERAK LRKD VRHS AACGGGGTA LLR LVR  
LRT LIR LVR LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 110) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: 138) 170) 202) 235) 272) 311) ZFP945 59 GTTGCCGGGC 1146 1164 –  
QASA RREH GLTA ERAK AKRD VNSS AACGGGGTA LSR LVR LRT LIR LDR LTR (SEQ ID  
(SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 110) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 139)  
170) 202) 235) 271) 310) ZFP951 60 CGAGAAAGTG 1085 1103 – RGRN DSSV QNAN QKHH  
QRSN QKVH AAAGCCTGC LEM LRR LKR LAV LAR LEA (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ  
(SEQ (SEQ NO: 111) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 140) 171) 203) 236) 273) 312)  
ZFP952 61 CGAGAAAGTG 1085 1103 – RRRN DSSV QNAN QKHH QRSN QKVH AAAGCCTGC  
LDV LRR LKR LAV LAR LEA (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 111) ID ID ID  
ID ID ID NO: NO: NO: NO: NO: NO: 141) 171) 203) 236) 273) 312) ZFP953 62 CGAGAAAGTG  
1085 1103 – RGRN DSSV LKSN LKQH LKTN QKCH AAAGCCTGC LAI LRR LHR LVV LAR  
LKA (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 111) ID ID ID ID ID ID NO: NO: NO: NO:  
NO: NO: 142) 171) 204) 237) 274) 313) ZFP956 63 GAGGCTTGAA 1856 1874 – DGSN RIDN QRRY  
QQTN QRSD RGDN CAGTAGGAC LRR LDG LVE LAR LTR LNR (SEQ ID (SEQ (SEQ (SEQ  
(SEQ (SEQ (SEQ NO: 112) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 143) 172) 205) 238)  
275) 314) ZFP957 64 GAGGCTTGAA 1856 1874 – DPSN RRDN TTFN QTQN HKET REDN  
CAGTAGGAC LOR LPK LRV LTR LNR LGR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
112) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 144) 173) 206) 239) 276) 315) ZFP958 65  
GAGGCTTGAA 1856 1874 – DPSN RRDN QRRY QQTN QRSD RGDN CAGTAGGAC LOR LPK  
LVE LAR LTR LNR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 112) ID ID ID ID ID ID  
NO: NO: NO: NO: NO: NO: 144) 173) 205) 238) 275) 314) ZFP961 66 GAGGTTGGGG 312 329 –  
QQTN ANRT EEAN RGEH TNSS RIDN ACTGCGAA LTR LVH LRR LTR LTR LIR (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 113) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 145) 174)  
207) 240) 277) 316) ZFP962 67 GAGGTTGGGG 312 329 – QQTN ANRT EEAN RREH MTSS  
RQDN ACTGCGAA LTR LVH LRR LVR LRR LGR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ

ID ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 145) 174) 207) 241) 278) 317) ZFP963 68  
GAGGTTGGGG 312 329 – QQTN ANRT EEAN RGEH MTSS RQDN ACTGCGAA LTR LVH LRR  
LTR LRR LGR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 113) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: 145) 174) 207) 240) 278) 317) ZFP964 69 GATGATGTGG 742 762 + RATH  
RADV QRSS RKDA VHHN ISHN TATTGGGG LTR LKG LVR LHV LVR LAR (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 114) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 146) 175)  
208) 242) 259) 299) ZFP965 70 GATGATGTGG 742 762 + RATH RADV QSSS RKER VRHN ISHN  
TATTGGGG LTR LKG LVR LAT LTR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
114) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 146) 175) 209) 243) 279) 299) ZFP966 71  
GATGATGTGG 742 762 + KKDH RKES QSSS RKER VHHN ISHN TATTGGGG LHR LTV LVR  
LAT LVR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 114) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: NO: 131) 176) 209) 243) 259) 299) ZFP969 72 GATGATGTGG 742 763 + RVDH  
RREH QSSS RKER VAHN ISHN TATTGGGGG LHR LSG LVR LAT LTR LAR (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 115) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 147) 177)  
209) 243) 280) 299) ZFP970 73 GATGATGTGG 742 763 + RKHH RREH QSSS RKER VAHN ISHN  
TATTGGGGG LGR LTI LVR LAT LTR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
115) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 148) 178) 209) 243) 280) 299) ZFP971 74  
GATGATGTGG 742 763 + RVDH RSDH QSSS RKER VAHN ISHN TATTGGGGG LHR LSL LVR  
LAT LTR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 115) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: NO: 147) 179) 209) 243) 280) 299) ZFP984 75 GCAGTAGTCG 90 107 – KTDH  
QKEI QSAH ETGS QSSS QTNT GAACAGGG LAR LTR LKR LRR LVR LGR (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 116) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 132) 163)  
193) 225) 281) 318) ZFP985 76 GCAGTAGTCG 90 107 – KKDH QKEI QSAH ETGS QSSS QGGT  
GAACAGGG LHR LTR LKR LRR LVR LRR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
116) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 131) 163) 193) 225) 281) 319) ZFP986 77  
GCAGTAGTCG 90 107 – KKDH QKEI QSAH DPTS QSSS QTNT GAACAGGG LHR LTR LKR  
LNR LVR LGR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 116) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: NO: 131) 163) 193) 244) 281) 318) ZFP989 78 GCATAGCAGC 409 426 – QQTN  
VGGN KRYN RQDN RSHN QSTT AGGATGAA LTR LAR LYQ LNT LKL LKR (SEQ ID (SEQ  
(SEQ (SEQ (SEQ (SEQ (SEQ NO: 117) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 145) 180)  
210) 245) 283) 320) ZFP990 79 GCATAGCAGC 409 426 – QQTN VGGN KRYN RQDN RSHN  
QSTT AGGATGAA LTR LSR LYQ LNT LRL LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
117) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 145) 181) 210) 245) 283) 320) ZFP991 80  
GCATAGCAGC 409 426 – QQTN VGGN KKEN RRDN RSHN QSTT AGGATGAA LTR LSR LLQ  
LKS LKI LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 117) ID ID ID ID ID ID NO: NO:  
NO: NO: NO: NO: NO: NO: NO: 145) 181) 211) 246) 282) 320) ZFP994 81 GGCGTTCACG 1612 1630 – DKSS  
DHSS RNFI RNDT TSTL LKEH GTGGTCTCC LRK LKR LOR LII LKR LTR (SEQ ID (SEQ (SEQ  
(SEQ (SEQ (SEQ (SEQ NO: 118) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 149) 182) 212)  
247) 284) 321) ZFP995 82 GGCGTTCACG 1612 1630 – CNGS DHSS RNFI RQDI HKSS ESGH  
GTGGTCTCC LKK LKR LAR LVV LTR LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO:  
118) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 150) 182) 213) 248) 285) 301) ZFP996 83  
GGCGTTCACG 1612 1630 – CNGS DHSS RNFI RQDI TSTL LKEH GTGGTCTCC LKK LKR LAR  
LVV LKR LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 118) ID ID ID ID ID ID NO:  
NO: NO: NO: NO: NO: NO: NO: 150) 182) 213) 248) 284) 321) ZFP999 84 GTTGGTGAGT 327 344 – TNNN  
RTDS QREH RRDN RRQK HKSS GATTGGAG LAR LTL LTT LNR LTI LTR (SEQ ID (SEQ (SEQ  
(SEQ (SEQ (SEQ (SEQ NO: 119) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 151) 183) 214)  
233) 286) 322) ZFP1000 85 GTTGGTGAGT 327 344 – TNNN RTDS QREH RGDN RRQK HKSS  
GATTGGAG LAR LTL LTT LKR LTI LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 119)  
ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: NO: 151) 183) 214) 249) 286) 322) ZFP1001 86  
GTTGGTGAGT 327 344 – TNNN RTDS QREH RGDN RRQK HKSS GATTGGAG LAR LTL LNG  
LAR LTI LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 119) ID ID ID ID ID ID NO: NO:  
NO: NO: NO: NO: NO: NO: NO: 151) 183) 215) 250) 286) 322) ZFP1005 87 GGAGGTTGGG 312 330 – QQTN

ANRT DPAN RQEH QNSH GACTGCGAA LTR LVH LRR LRR LRR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ NO: 120) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 145) 174) 216) 251) 287) 323) ZFP1006 88 GGAGGTTGGG 312 330 – QQTN ANRT EEAN RREH MKHH QNSH GACTGCGAA LTR LVH LRR LVR LGR LRR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 120) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 145) 174) 207) 241) 287) 323) ZFP1007 89 GGAGGTTGGG 312 330 – QQTN ANRT DPAN RQEH LKQH QGGH GACTGCGAA LTR LVH LRR LVR LVR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 120) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 145) 174) 216) 251) 288) 324) ZFP1008 90 GGATGATGTG 741 762 + RNTH RADV QRSS RKDA QNEH QNSH GTATTGGGG LAR LKG LVR LHV LKV LRR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 121) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 152) 175) 208) 242) 289) 323) ZFP1009 91 GGATGATGTG 741 762 + RNTH RADV QSSS RKER QKTH QGGH GTATTGGGG LAR LKG LVR LAT LAV LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 121) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 152) 175) 209) 243) 290) 325) ZFP1010 92 GGATGATGTG 741 762 + RNTH RADV QSSS RKER QKTH QNSH GTATTGGGG LAR LKG LVR LAT LAV LRR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 121) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 152) 175) 209) 243) 290) 323) ZFP1013 93 GGATGTGTCT 375 395 + HKSS ESGH RRRN DRSS QPHS QKPH GCGGCGTT LTR LKR LTL LKR LAV LSR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 122) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 153) 184) 217) 252) 291) 326) ZFP1014 94 GGATGTGTCT 375 395 + HKSS EGGH RRRN DHSS RRQH QSAH GCGGCGTT LTR LKR LQL LKR LQY LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 122) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 153) 185) 218) 229) 292) 327) ZFP1015 95 GGATGTGTCT 375 395 + HKSS EGGH RRRN DRSS RRQH QSAH GCGGCGTT LTR LKR LTL LKR LQY LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 122) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 153) 185) 217) 252) 292) 327) ZFP1018 96 GGGGGTTGCG 1184 1202 – GHTA QSGT DHSS AMRS RRSR RGEH TCAGCAAAC LRN LHR LKR LMG LVR LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 123) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 154) 186) 199) 253) 293) 328) ZFP1019 97 GGGGGTTGCG 1184 1202 – GHTA QSTT DHSS QQRS EAAH RTEH TCAGCAAAC LRN LKR LKR LVG LSR LAR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 123) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 154) 187) 199) 254) 294) 329) ZFP1020 98 GGGGGTTGCG 1184 1202 – GHTA QSTT DHSS AMRS RQSR RREH TCAGCAAAC LRN LKR LKR LMG LQR LVR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 123) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 154) 187) 199) 253) 295) 330) ZFP1023 99 GTTGTTAGAC 2342 2363 + QGET RADN DKAN DOGN HRHV TNSS GACGAGGCA LKR LRR LTR LIR LIN LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 124) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 155) 188) 219) 255) 296) 331) ZFP1024 100 GTTGTTAGAC 2342 2363 + QGET RADN DSSN DQGN HKSS IRTS GACGAGGCA LKR LRR LRR LIR LTR LKR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 124) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 155) 188) 220) 255) 285) 332) ZFP1025 101 GTTGTTAGAC 2342 2363 + QGET RADN EQGN DGN HRHV TNSS GACGAGGCA LKR LRR LLR LGR LIN LTR (SEQ ID (SEQ (SEQ (SEQ (SEQ (SEQ (SEQ NO: 124) ID ID ID ID ID ID NO: NO: NO: NO: NO: NO: 155) 188) 221) 256) 296) 331)

(81) In some embodiments, the ZFP domain of the present epigenetic editor binds to a target sequence provided herein. In further embodiments, the ZFP domain comprises, in order, the F1-F6 amino acid sequences of any one of the zinc finger proteins as shown in Table 1 and Table 18. The F1-F6 amino acid sequences may be placed within the ZF framework sequence of SEQ ID NOs: 1084 and 1258-1259, or within any other ZF framework known in the art.

(82) C. TALEs

(83) In some embodiments, the DNA-binding domain of an epigenetic editor described herein comprises a transcription activator-like effector (TALE) domain. The DNA-binding domain of a TALE comprises a highly conserved sequence of about 33-34 amino acids, with a repeat variable di-residue (RVD) at positions 12 and 13 that is central to the recognition of specific nucleotides. TALEs can be engineered to bind practically any desired DNA sequence. Methods for programming TALEs are known in the art. For example, such methods are described in Carroll et al., *Genet Soc Amer.* (2011) 188(4):773-82; Miller et

al., *Nat Biotechnol.* (2007) 25(7):778-85; Christian et al., *Genetics* (2008) 186(2):757-61; Li et al., *Nucl Acids Res.* (2010) 39(1):359-72; and Moscou et al., *Science* (2009) 326(5959):1501.

(84) D. Other DNA-Binding Domains

(85) Other DNA-binding domains are contemplated for the epigenetic editors described herein. In some embodiments, the DNA-binding domain comprises an argonaute protein domain, e.g., from *Natronobacterium gregoryi* (NgAgo). NgAgo is a ssDNA-guided endonuclease that is guided to its target site by 5' phosphorylated ssDNA (gDNA), where it produces double-strand breaks. In contrast to Cas9, the NgAgo-gDNA system does not require a protospacer-adjacent motif (PAM). Thus, using a nuclease inactive NgAgo (dNgAgo) can greatly expand the bases that may be targeted. The characterization and use of NgAgo have been described, e.g., in Gao et al., *Nat Biotechnol.* (2016) 34(7):768-73; Swarts et al., *Nature* (2014) 507(7491):258-61; and Swarts et al., *Nucl Acids Res.* (2015) 43(10):5120-9.

(86) In some embodiments, the DNA-binding domain comprises an inactivated nuclease, for example, an inactivated meganuclease. Additional non-limiting examples of DNA-binding domains include tetracycline-controlled repressor (tetR) DNA-binding domains, leucine zippers, helix-loop-helix (HLH) domains, helix-turn-helix domains,  $\beta$ -sheet motifs, steroid receptor motifs, bZIP domains homeodomains, and AT-hooks.

## II. Guide Polynucleotides

(87) Epigenetic editors described herein that comprise a polynucleotide guided DNA-binding domain may also include a guide polynucleotide that is capable of forming a complex with the DNA-binding domain. The guide polynucleotide may comprise RNA, DNA, or a mixture of both. For example, where the polynucleotide guided DNA-binding domain is a CRISPR-associated protein domain, the guide polynucleotide may be a guide RNA (gRNA). A "guide RNA" or "gRNA" refers to a nucleic acid that is able to hybridize to a target sequence and direct binding of the CRISPR-Cas complex to the target sequence. Methods of using guide polynucleotide sequences with programmable DNA-binding proteins (e.g., CRISPR-associated protein domains) for site-specific DNA targeting (e.g., to modify a genome) are known in the art.

(88) A guide polynucleotide sequence (e.g., a gRNA sequence) may comprises two parts: 1) a nucleotide sequence comprising a "targeting sequence" that is complementary to a target nucleic acid sequence ("target sequence"), e.g., to a nucleic acid sequence comprised in a genomic target site; and 2) a nucleotide sequence that binds a polynucleotide guided DNA-binding domain (e.g., a CRISPR-Cas protein domain). The nucleotide sequence in 1) may comprise a targeting sequence that is 100% complementary to a genomic nucleic acid sequence, e.g., a nucleic acid sequence comprised in a genomic target site, and thus may hybridize to the target nucleic acid sequence. The nucleotide sequence in 1) may be referred to as, e.g., a crispr RNA, or crRNA. The nucleotide sequence in 2) may be referred to as a scaffold sequence of a guide nucleic acid, e.g., a tracrRNA, or an activating region of a guide nucleic acid, and may comprise a stem-loop structure. Parts 1) and 2) as described above may be fused to form one single guide (e.g., a single guide RNA, or sgRNA), or may be on two separate nucleic acid molecules. In some embodiments, a guide polynucleotide comprises parts 1) and 2) connected by a linker. In some embodiments, a guide polynucleotide comprises parts 1) and 2) connected by a non-nucleic acid linker, for example, a peptide linker or a chemical linker.

(89) Part 2 (the scaffold sequence) of a guide polynucleotide as described herein may be, for example, as described in Jinek et al., *Science* (2012) 337:816-21; U.S. Patent Publication 2016/0208288; or U.S. Patent Publication 2016/0200779. Variants of part 2) are also contemplated by the present disclosure. For example, the tetraloop and stem loop of a gRNA scaffold (tracrRNA) sequence may be modified to include RNA aptamers, which can be bound by specific protein domains. In some embodiments, such modified gRNAs can be used to facilitate the recruitment of repressive or activating domains fused to the protein-interacting RNA aptamers.

(90) A gRNA as provided herein typically comprises a targeting domain and a binding domain. The targeting domain (also termed "targeting sequence") may comprise a nucleic acid sequence that binds to a target site, e.g., to a genomic nucleic acid molecule within a cell. The target site may be a double-stranded DNA sequence comprising a PAM sequence as well as the target sequence, which is located on





embodiments, the targeting domain is 19 nucleotides in length. In some embodiments, the targeting domain is 20 nucleotides in length. In some embodiments, the targeting domain is 21 nucleotides in length. In some embodiments, the targeting domain is 22 nucleotides in length. In some embodiments, the targeting domain is 23 nucleotides in length. In some embodiments, the targeting domain is 24 nucleotides in length. In some embodiments, the targeting domain is 25 nucleotides in length. In certain embodiments, the targeting domain fully corresponds, without mismatch, to a target sequence provided herein, or a part thereof. In some embodiments, the targeting domain of a gRNA provided herein comprises 1 mismatch relative to a target sequence provided herein. In some embodiments, the targeting domain comprises 2 mismatches relative to the target sequence. In some embodiments, the target domain comprises 3 mismatches relative to the target sequence.

(97) Methods for designing, selecting, and validating gRNAs are described herein and known in the art. Software tools can be used to optimize the gRNAs corresponding to a target DNA sequence, e.g., to minimize total off-target activity across the genome. For example, DNA sequence searching algorithms can be used to identify a target sequence in crRNAs of a gRNA for use with Cas9. Exemplary gRNA design tools include the ones described in Bae et al., *Bioinformatics* (2014) 30:1473-5.

(98) Guide polynucleotides (e.g., gRNAs) described herein may be of various lengths. In some embodiments, the length of the spacer or targeting sequence depends on the CRISPR-associated protein component of the epigenetic editor system used. For example, Cas proteins from different bacterial species have varying optimal targeting sequence lengths. Accordingly, the spacer sequence may comprise, e.g., 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, or more than 50 nucleotides in length. In some embodiments, the spacer comprises 10-24, 11-20, 11-16, 18-24, 19-21, or 20 nucleotides in length. In some embodiments, a guide polynucleotide (e.g., gRNA) is from 15-100 (e.g., 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50) nucleotides in length and comprises a spacer sequence of at least 10 (e.g., 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50) contiguous nucleotides complementary to the target sequence. In some embodiments, a guide polynucleotide described herein may be truncated, e.g., by 1, 2, 3, 4, 5, 10, 15, 20, 25, 30, 40, 50 or more nucleotides.

(99) In certain embodiments, the 3' end of the HBV target sequence is immediately adjacent to a PAM sequence (e.g., a canonical PAM sequence such as NGG for SpCas9). The degree of complementarity between the targeting sequence of the guide polynucleotide (e.g., the spacer sequence of a gRNA) and the target sequence may be at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%. In particular embodiments, the targeting and the target sequence may be 100% complementary. In other embodiments, the targeting sequence and the target sequence may contain, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 mismatches.

(100) A guide polynucleotide (e.g., gRNA) may be modified with, for example, chemical alterations and synthetic modifications. A modified gRNA, for instance, can include an alteration or replacement of one or both of the non-linking phosphate oxygens and/or of one or more of the linking phosphate oxygens in the phosphodiester backbone linkage, an alteration of the ribose sugar (e.g., of the 2' hydroxyl on the ribose sugar), an alteration of the phosphate moiety, modification or replacement of a naturally occurring nucleobase, modification or replacement of the ribose-phosphate backbone, modification of the 3' end and/or 5' end of the oligonucleotide, replacement of a terminal phosphate group or conjugation of a moiety, cap, or linker, or any combination thereof.

(101) In some embodiments, one or more ribose groups of the gRNA may be modified. Examples of chemical modifications to the ribose group include, but are not limited to, 2'-O-methyl (2'-OMe), 2'-fluoro (2'-F), 2'-deoxy, 2'-O-(2-methoxyethyl) (2'-MOE), 2'-NH<sub>2</sub>, 2'-O-allyl, 2'-O-ethylamine, 2'-O-cyanoethyl, 2'-O-acetalester, or a bicyclic nucleotide such as locked nucleic acid (LNA), 2'-(5'-constrained ethyl (S-cEt)), constrained MOE, or 2'-0,4'-C-aminomethylene bridged nucleic acid (2',4'-BNANC). 2'-O-methyl modification and/or 2'-fluoro modification may increase binding affinity and/or nuclease stability of the gRNA oligonucleotides.

(102) In some embodiments, one or more phosphate groups of the gRNA may be chemically modified.

Examples of chemical modifications to a phosphate group include, but are not limited to, a phosphorothioate (PS), phosphonoacetate (PACE), thiophosphonoacetate (thioPACE), amide, triazole, phosphonate, and phosphotriester modification. In some embodiments, a guide polynucleotide described herein may comprise one, two, three, or more PS linkages at or near the 5' end and/or the 3' end; the PS linkages may be contiguous or noncontiguous.

(103) In some embodiments, the gRNA herein comprises a mixture of ribonucleotides and deoxyribonucleotides and/or one or more PS linkages.

(104) In some embodiments, one or more nucleobases of the gRNA may be chemically modified. Examples of chemically modified nucleobases include, but are not limited to, 2-thiouridine, 4-thiouridine, N6-methyladenosine, pseudouridine, 2,6-diaminopurine, inosine, thymidine, 5-methylcytosine, 5-substituted pyrimidine, isoguanine, isocytosine, and nucleobases with halogenated aromatic groups. Chemical modifications can be made in the spacer region, the tracr RNA region, the stem loop, or any combination thereof.

(105) Table 2 below lists exemplary target sequences for epigenetic modification of HBV, as well as the coordinates of the start and end positions of the targeted site on the HBV genome.

(106) TABLE-US-00005 TABLE 2 Targeting Domain Sequences of Exemplary gRNAs Targeting HBV. The following target sites were identified as suitable for targeting with an epigenetic repressor: SEQ Target domain IDs sequence Start End Strand

333	CCTGCTGGTGGCTCCAGTTC	57	77	+	334	CTGAACTGGAGCCACCAGCA	59	79	-	335
	CCTGAACTGGAGCCACCAGC	60	80	-	336	CCTCGAGAAGATTGACGATA	115	135	-	337
	TCGTCAATCTTCTCGAGGAT	117	137	+	338	CGTCAATCTTCTCGAGGATT	118	138	+	339
	GTCAATCTTCTCGAGGATTG	119	139	+	340	AACATGGAGAACATCACATC	153	173	+	341
	AACATCACATCAGGATTCCT	162	182	+	342	CTAGACTCTGCGGTATTGTG	233	253	-	343
	TACCGCAGAGTCTAGACTCG	238	258	+	344	CGCAGAGTCTAGACTCGTGG	241	261	+	345
	CACCACGAGTCTAGACTCTG	243	263	-	346	TGGACTTCTCTCAATTTTCT	261	281	+	347
	GGACTTCTCTCAATTTTCTA	262	282	+	348	GACTTCTCTCAATTTTCTAG	263	283	+	349
	ACTTCTCTCAATTTTCTAGG	264	284	+	350	CGAATTTTGGCCAAGACACA	295	315	-	351
	AGGTTGGGGACTGCGAATTT	309	328	-	352	GGCATAGCAGCAGGATGAAG	408	427	-	353
	AGAAGATGAGGCATAGCAGC	417	436	-	354	GCTATGCCTCATCTTCTTGT	420	439	+	355
	GAAGAACCAACAAGAAGATG	429	448	-	356	CATCTTCTTGTGTTGGTTCTTC	429	448	+	357
	CCCGTTTGTCTCTAATTCC	469	488	+	358	CCTGGAATTAGAGGACAAAC	472	491	-	359
	TCCTGGAATTAGAGGACAAA	473	492	-	360	TACTAGTGCCATTTGTTTCAG	680	699	+	361
	CCATTTGTTTCAGTGGTTCGT	688	707	+	362	CATTTGTTTCAGTGGTTCGTA	689	708	+	363
	CCTACGAACCACTGAACAAA	691	710	-	364	TTTCAGTTATATGGATGATG	731	750	+	365
	CAAAAGAAAATTGGTAACAG	799	818	-	366	TACCAATTTTCTTTTGTCTT	803	822	+	367
	ACCAATTTTCTTTTGTCTTT	804	823	+	368	ACCCAAAGACAAAAGAAAAT	808	827	-	369
	TGACATACTTTCCAATCAAT	975	994	-	370	CACTTTCTCGCCAACTTACA	1093	1113	+	371
	CACAGAAAGGCCTTGTAAGT	1106	1126	-	372	TGAACCTTTACCCCGTTGCC	1137	1157	+	373
	GGGCAACGGGGTAAAGGTTT	1138	1158	-	374	TTTACCCCGTTGCCCGGCAA	1143	1163	+	375
	GTTGCCGGGCAACGGGGTAA	1144	1164	-	376	CCCGTTGCCCGGCAACGGCC	1148	1168	+	377
	CTGGCCGTTGCCGGGCAACG	1150	1170	-	378	CCTGGCCGTTGCCGGGCAAC	1151	1171	-	379
	ACCTGGCCGTTGCCGGGCAA	1152	1172	-	380	GCACAGACCTGGCCGTTGCC	1158	1178	-	381
	GGCACAGACCTGGCCGTTGC	1159	1179	-	382	GCAAACACTTGGCACAGACC	1169	1189	-	383
	GGGTTGCGTCAGCAAACACT	1180	1200	-	384	TTTGCTGACGCAACCCCCAC	1184	1204	+	385
	CTGACGCAACCCCCACTGGC	1188	1208	+	386	TGACGCAACCCCCACTGGCT	1189	1209	+	387
	GACGCAACCCCCACTGGCTG	1190	1210	+	388	AACCCCCACTGGCTGGGGCT	1195	1215	+	389
	TCCTCTGCCGATCCATACTG	1255	1275	+	390	TCCGCAGTATGGATCGGCAG	1259	1279	-	391
	AGGAGTTCCGCAGTATGGAT	1265	1285	-	392	CGGCTAGGAGTTCCGCAGTA	1270	1290	-	393
	TGCGAGCAAAACAAGCGGCT	1285	1305	-	394	CCGCTTGTTTTGCTCGCAGC	1287	1307	+	395
	CCTGCTGCGAGCAAAACAAG	1290	1310	-	396	TGTTTTGCTCGCAGCAGGTC	1292	1312	+	397
	GCAGCACAGCCTAGCAGCCA	1376	1396	-	398	TGCTAGGCTGTGCTGCCAAC	1380	1400	+	399
	GCTGCCAACTGGATCCTGCG	1391	1411	+	400	CTGCCAACTGGATCCTGCGC	1392	1412	+	401

CGTCCGCGACGATCCAGCT 1398 1418 – 402 AAACAAAGACGTCACGCGC 1408 1428 – 403  
 GTCCTTTGTTTACGTCCCGT 1417 1437 + 404 CGCCGACGGGACGTAAACAA 1422 1442 – 405  
 TGCCGTTCCGACCGACCACG 1504 1523 + 406 AGGTGCGCCCCGTGGTCCGT 1513 1533 – 407  
 AGAGAGGTGCGCCCCCGTGGT 1517 1537 – 408 GTAAAGAGAGGTGCGCCCCG 1521 1541 –  
 409 GGGGCGCACCTCTCTTTACG 1522 1542 + 410 CGGGGAGTCCGCGTAAAGAG 1533 1553 –  
 411 CAGATGAGAAGGCACAGACG 1551 1571 – 412 GTCTGTGCCTTCTCATCTGC 1552 1572 +  
 413 GGCAGATGAGAAGGCACAGA 1553 1573 – 414 GCAGATGAGAAGGCACAGAC 1553 1572  
 – 415 ACACGGTCCGGCAGATGAGA 1562 1582 – 416 GAAGCGAAGTGCACACGGTC 1574  
 1594 – 417 GAGGTGAAGCGAAGTGCACA 1579 1599 – 418 CTTACCTCTGCACGTCGCA 1590  
 1610 + 419 GGTCTCCATGCGACGTGCAG 1598 1618 – 420 TGCCCAAGGTCTTACATAAG 1640  
 1660 + 421 GTCCTCTTATGTAAGACCTT 1645 1665 – 422 AGTCCTCTTATGTAAGACCT 1646  
 1666 – 423 GTCTTACATAAGAGGACTCT 1648 1668 + 424 AATGTCAACGACCGACCTTG 1680  
 1700 + 425 TTTGAAGTATGCCTCAAGGT 1694 1714 – 426 AGTCTTTGAAGTATGCCTCA 1698  
 1718 – 427 AAGACTGTTTGTTTAAAGAC 1712 1732 + 428 AGACTGTTTGTTTAAAGACT 1713  
 1733 + 429 CTGTTTGTTTAAAGACTGGG 1716 1736 + 430 GTTTAAAGACTGGGAGGAGT 1722  
 1742 + 431 TCTTTGTACTAGGAGGCTGT 1766 1786 + 432 AGGAGGCTGTAGGCATAAAT 1776  
 1796 + 433 GTGAAAAAGTTGCATGGTGC 1810 1830 – 434 GCAGAGGTGAAAAAGTTGCA 1816  
 1836 – 435 AACAAGAGATGATTAGGCAG 1832 1852 – 436 GACATGAACAAGAGATGATT 1838  
 1858 – 437 AGCTTGGAGGCTTGAACAGT 1860 1880 – 438 CAAGCCTCCAAGCTGTGCCT 1866  
 1886 + 439 AAGCCTCCAAGCTGTGCCTT 1867 1887 + 440 CCTCCAAGCTGTGCCTTGGG 1871  
 1890 + 441 CCACCCAAGGCACAGCTTGG 1873 1893 – 442 AGCTGTGCCTTGGGTGGCTT 1876  
 1896 + 443 AAGCCACCCAAGGCACAGCT 1876 1896 – 444 GCTGTGCCTTGGGTGGCTTT 1877  
 1897 + 445 CTGTGCCTTGGGTGGCTTTG 1878 1898 + 446 TAGCTCCAAATTCTTTATAA 1916  
 1936 – 447 GTAGCTCCAAATTCTTTATA 1917 1937 – 448 TAAAGAATTTGGAGCTACTG 1919  
 1939 + 449 ATGACTCTAGCTACCTGGGT 2097 2117 + 450 CACATTTCTTGTCTCACTTT 2211  
 2231 + 451 TAGTTTCCGGAAGTGTTGAT 2321 2341 – 452 CGTCTAACAACAGTAGTTTC 2334  
 2354 – 453 ACTACTGTTGTTAGACGACG 2337 2357 + 454 CTGTTGTTAGACGACGAGGC 2341  
 2361 + 455 CGAGGGAGTTCTTCTTCTAG 2368 2388 – 456 GCGAGGGAGTTCTTCTTCTA 2369  
 2389 – 457 GGCGAGGGAGTTCTTCTTCT 2370 2390 – 458 CTCCCTCGCCTCGCAGACGA 2380  
 2400 + 459 GACCTTCGTCTGCGAGGCGA 2385 2405 – 460 AGACCTTCGTCTGCGAGGCG 2386  
 2406 – 461 GATTGAGACCTTCGTCTGCG 2391 2411 – 462 GATTGAGATCTTCTGCGACG 2415  
 2435 – 463 GTCGCAGAAGATCTCAATCT 2416 2436 + 464 TCGCAGAAGATCTCAATCTC 2417  
 2437 + 465 ATATGGTGACCCACAAAATG 2807 2827 – 466 TTTGTGGGTCACCATATTCT 2810  
 2830 + 467 TTGTGGGTCACCATATTCTT 2811 2831 + 468 GCTGGATCCAACCTGGTGGTC 2894  
 2914 – 469 CACCCCAAAGGCCTCCGTG 3026 3046 – 470 CCTTTTGGGGTGGAGCCCTC 3034  
 3054 + 471 CCGAGGGCTCCACCCCAA 3037 3057 – 472 GGGGTGGAGCCCTCAGGCTC 3040  
 3060 + 473 GGGTGGAGCCCTCAGGCTCA 3041 3061 + 474 CGATTGGTGGAGGCAGGAGG 3092  
 3112 – 475 CTCATCCTCAGGCCATGCAG 3159 3179 + 102 GATGAGGCATAGCAGCAG 415 432  
 – 103 GATGATTAGGCAGAGGTG 1828 1845 – 104 GGATTACGCGCCGACGGG 1433 1450 – 105  
 GGCAGTAGTCGGAACAGGG 90 108 – 106 GTAAACTGAGCCAGGAGAA 664 682 – 107  
 ACGGTGGTCTCCATGCGAC 1605 1623 – 108 GCTGGATGTGTCTGCGGCG 372 393 + 109  
 GTCTGCGAGGCGAGGGAG 2381 2398 – 110 GTTGCCGGGCAACGGGGTA 1146 1164 – 111  
 CGAGAAAGTGAAAGCCTGC 1085 1103 – 112 GAGGCTTGAACAGTAGGAC 1856 1874 – 113  
 GAGGTTGGGGACTGCGAA 312 329 – 114 GATGATGTGGTATTGGGG 742 762 + 115  
 GATGATGTGGTATTGGGGG 742 763 + 116 GCAGTAGTCGGAACAGGG 90 107 – 117  
 GCATAGCAGCAGGATGAA 409 426 – 118 GGCGTTCACGGTGGTCTCC 1612 1630 – 119  
 GTTGGTGAGTGATTGGAG 327 344 – 120 GGAGGTTGGGGACTGCGAA 312 330 – 121  
 GGATGATGTGGTATTGGGG 741 762 + 122 GGATGTGTCTGCGGCGTT 375 395 + 123  
 GGGGGTTGCGTCAGCAAAC 1184 1202 – 124 GTTGTTAGACGACGAGGCA 2342 2363 +  
 (107) Target domains identified above that are adjacent to a PAM sequence, e.g., an *S. pyogenes* Cas9  
 PAM sequence, can be targeted by a CRISPR-based epigenetic repressor, e.g., an epigenetic repressor  
 comprising a dCas9 DNA-binding domain. For example, target sites 1-143 are suitable for dCas9-based

epigenetic repressor targeting.

(108) A suitable gRNA for targeting any of the target domain sequences would, in some embodiments, comprise a target domain sequence that is the RNA-equivalent sequence of the provided DNA sequence of the targeting domain sequence (i.e., an RNA nucleotide of that sequence instead of the provided DNA nucleotide, with uracil instead of thymine), and a suitable tracr RNA sequence.

(109) Any tracr sequence known in the art is contemplated for a gRNA described herein. In some embodiments, a gRNA described herein has a tracr sequence shown in Table 3 below, or a tracr sequence at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical to the tracr sequence shown below (SEQ: SEQ ID NO).

(110) TABLE-US-00006 TABLE 3 Exemplary TRACR Sequences SEQ Sequence (5' to 3')

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1087 GUUUAAGAGCUAUGCUGGAAACAGCAUAGCAAGUUUAAAUAAG
GCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGC UUUUUUU 1088
GUUUUAGAGCUAGAAAUAGCAAGUUAAAAUAAGGCUAGUCCGU
UAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUU 1089
GUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAUAAG
GCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGC UUUUUUU 1090
GUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAUAAG
GCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGC UUUUUUU
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(111) In some embodiments, the gRNA herein is provided to the cell directly (e.g., through an RNP complex together with the CRISPR-associated protein domain). In some embodiments, the gRNA is provided to the cell through an expression vector (e.g., a plasmid vector or a viral vector) introduced into the cell, where the cell then expresses the gRNA from the expression vector. Methods of introducing gRNAs and expression vectors into cells are well known in the art.

### III. Effector Domains

(112) Epigenetic editors described herein include one or more effector protein domains (also “epigenetic effector domains,” or “effector domains,” as used herein) that effect epigenetic modification of a target gene. An epigenetic editor with one or more effector domains may modulate expression of a target gene without altering its nucleobase sequence. In some embodiments, an effector domain described herein may provide repression or silencing of expression of HBV or an HBV gene, e.g., by repressing transcription or by modifying or remodeling HBV chromatin. Such effector domains are also referred to herein as “repression domains,” “repressor domains,” “epigenetic repressor domains,” or “epigenetic repression domains.” Non-limiting examples of chemical modifications that may be mediated by effector domains include methylation, demethylation, acetylation, deacetylation, phosphorylation, SUMOylation and/or ubiquitination of DNA or histone residues.

(113) In some embodiments, an effector domain of an epigenetic editor described herein may make histone tail modifications, e.g., by adding or removing active marks on histone tails.

(114) In some embodiments, an effector domain of an epigenetic editor described herein may comprise or recruit a transcription-related protein, e.g., a transcription repressor. The transcription-related protein may be endogenous or exogenous.

(115) In some embodiments, an effector domain of an epigenetic editor described herein may, for example, comprise a protein that directly or indirectly blocks access of a transcription factor to the gene of interest harboring the target sequence.

(116) An effector domain may be a full-length protein or a fragment thereof that retains the epigenetic effector function (a “functional domain”). Functional domains that are capable of modulating (e.g., repressing) gene expression can be derived from a larger protein. For example, functional domains that can reduce target gene expression may be identified based on sequences of repressor proteins. Amino acid sequences of gene expression-modulating proteins may be obtained from available genome browsers, such as the UCSD genome browser or Ensembl genome browser. Protein annotation databases such as UniProt or Pfam can be used to identify functional domains within the full protein sequence. As a starting point, the largest sequence, encompassing all regions identified by different databases, may be tested for gene expression modulation activity. Various truncations then may be tested to identify the minimal functional unit.

(117) Variants of effector domains described herein are also contemplated by the present disclosure. A variant may, for example, refer to a polypeptide with at least about 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% sequence identity and/or sequence similarity to a wildtype effector domain described herein. In particular embodiments, the variant retains at least about 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% of the epigenetic effector function of the wildtype effector domain.

(118) In some embodiments, an epigenetic editor described herein may comprise 1 effector domain, 2 effector domains, 3 effector domains, 4 effector domains, 5 effector domains, 6 effector domains, 7 effector domains, 8 effector domains, 9 effector domains, 10 effector domains, or more. In certain embodiments, the epigenetic editor comprises one or more fusion proteins (e.g., one, two, or three fusion proteins), each with one or more effector domains (e.g., one, two, or three effector domains) linked to a DNA-binding domain. In some embodiments, the effector domains may induce a combination of epigenetic modifications, e.g., transcription repression and DNA methylation, DNA methylation and histone deacetylation, DNA methylation and histone demethylation, DNA methylation and histone methylation, DNA methylation and histone phosphorylation, DNA methylation and histone ubiquitylation, DNA methylation, and histone SUMOylation.

(119) In certain embodiments, an effector domain described herein (e.g., DNMT3A and/or DNMT3L) is encoded by a nucleotide sequence as found in the native genome (e.g., human or murine) for that effector domain. In other embodiments, an effector domain described herein is encoded by a nucleotide sequence that has been codon-optimized for optimal expression in human cells.

(120) Effector domains described herein may include, for example, transcriptional repressors, DNA methyltransferases, and/or histone modifiers, as further detailed below.

(121) A. Transcriptional Repressors

(122) In some embodiments, an epigenetic effector domain described herein mediates repression of a target gene's expression (e.g., transcription). The effector domain may comprise, e.g., a Krüppel-associated box (KRAB) repression domain, a Repressor Element Silencing Transcription Factor (REST) repression domain, a KRAB-associated protein 1 (KAP1) domain, a MAD domain, a FKHR (forkhead in rhabdosarcoma gene) repressor domain, an EGR-1 (early growth response gene product-1) repressor domain, an ets2 repressor factor repressor domain (ERD), a MAD smSIN3 interaction domain (SID), a WRPW motif (SEQ ID NO: 1257) of the hairy-related basic helix-loop-helix (bHLH) repressor proteins, an HP1 alpha chromo-shadow repression domain, an HP1 beta repression domain, or any combination thereof. The effector domain may recruit one or more protein domains that repress expression of the target gene, e.g., through a scaffold protein. In some embodiments, the effector domain may recruit or interact with a scaffold protein domain that recruits a PRMT protein, a HDAC protein, a SETDB1 protein, or a NuRD protein domain.

(123) In some embodiments, the effector domain comprises a functional domain derived from a zinc finger repressor protein, such as a KRAB domain. KRAB domains are found in approximately 400 human ZFP-based transcription factors. Descriptions of KRAB domains may be found, for example, in Ecco et al., *Development* (2017) 144(15):2719-29 and Lambert et al., *Cell* (2018) 172:650-65.

(124) In certain embodiments, the effector domain comprises a repression domain (e.g., KRAB) derived from KOX1/ZNF10, KOX8/ZNF708, ZNF43, ZNF184, ZNF91, HPF4, HTF10, or HTF34. In some embodiments, the effector domain comprises a repression domain (e.g., KRAB) derived from ZIM3, ZNF436, ZNF257, ZNF675, ZNF490, ZNF320, ZNF331, ZNF816, ZNF680, ZNF41, ZNF189, ZNF528, ZNF543, ZNF554, ZNF140, ZNF610, ZNF264, ZNF350, ZNF8, ZNF582, ZNF30, ZNF324, ZNF98, ZNF669, ZNF677, ZNF596, ZNF214, ZNF37, ZNF34, ZNF250, ZNF547, ZNF273, ZNF354, ZFP82, ZNF224, ZNF33, ZNF45, ZNF175, ZNF595, ZNF184, ZNF419, ZFP28-1, ZFP28-2, ZNF18, ZNF213, ZNF394, ZFP1, ZFP14, ZNF416, ZNF557, ZNF566, ZNF729, ZIM2, ZNF254, ZNF764, ZNF785, or any combination thereof. For example, the repression domain may be a KRAB domain derived from KOX1, ZIM3, ZFP28, or ZNF627. In particular embodiments, the repression domain is a ZIM3 KRAB domain. In further embodiments, the effector domain is derived from a human protein, e.g., a human ZIM3, a human KOX1, a human ZFP28, or a human ZNF627.

(125) Exemplary effector domains that may reduce or silence target gene expression are provided in

Table 4 below (SEQ: SEQ ID NO, see Table 18 for sequences of exemplary effector domains). Further examples of repressors and transcriptional repressor domains can be found, e.g., in PCT Patent Publication WO 2021/226077 and Tycko et al., *Cell* (2020) 183(7):2020-35, each of which is incorporated herein by reference in its entirety.

(126) TABLE-US-00007 TABLE 4 Exemplary Effector Domains Suitable for Silencing Gene Expression Protein SEQ ZIM3 495 ZNF436 496 ZNF257 497 ZNF675 498 ZNF490 499 ZNF320 500 ZNF331 501 ZNF816 502 ZNF680 503 ZNF41 504 ZNF189 505 ZNF528 506 ZNF543 507 ZNF554 508 ZNF140 509 ZNF610 510 ZNF264 511 ZNF350 512 ZNF8 513 ZNF582 514 ZNF30 515 ZNF324 516 ZNF98 517 ZNF669 518 ZNF677 519 ZNF596 520 ZNF214 521 ZNF37A 522 ZNF34 523 ZNF250 524 ZNF547 525 ZNF273 526 ZNF354A 527 ZFP82 528 ZNF224 529 ZNF33A 530 ZNF45 531 ZNF175 532 ZNF595 533 ZNF184 534 ZNF419 535 ZFP28-1 536 ZFP28-2 537 ZNF18 538 ZNF213 539 ZNF394 540 ZFP1 541 ZFP14 542 ZNF416 543 ZNF557 544 ZNF566 545 ZNF729 546 ZIM2 547 ZNF254 548 ZNF764 549 ZNF785 550 ZNF10 (KOX1) 551 CBX5 (chromoshadow domain) 552 RYBP (YAF2\_RYBP 553 component of PRC1) YAF2 (YAF2\_RYBP 554 component of PRC1) MGA (component of PRC1.6) 555 CBX1 (chromoshadow) 556 SCMHI (SAM\_1/SPM) 557 MPP8 (Chromodomain) 558 SUMO3 (Rad60-SLD) 559 HERC2 (Cyt-b5) 560 BIN1 (SH3\_9) 561 PCGF2 (RING finger protein 562 domain) TOX (HMG box) 563 FOXA1 (HNF3A C-terminal 564 domain) FOXA2 (HNF3B C-terminal 565 domain) IRF2BP1 (IRF-2BP1\_2 N- 566 terminal domain) IRF2BP2 (IRF-2BP1\_2 N- 567 terminal domain) IRF2BPL IRF-2BP1\_2 N- 568 terminal domain HOXA13 (homeodomain) 569 HOXB13 (homeodomain) 570 HOXC13 (homeodomain) 571 HOXA11 (homeodomain) 572 HOXC11 (homeodomain) 573 HOXC10 (homeodomain) 574 HOXA10 (homeodomain) 575 HOXB9 (homeodomain) 576 HOXA9 (homeodomain) 577 ZFP28\_HUMAN 578 ZN334\_HUMAN 579 ZN568\_HUMAN 580 ZN37A\_HUMAN 581 ZN181\_HUMAN 582 ZN510\_HUMAN 583 ZN862\_HUMAN 584 ZN140\_HUMAN 585 ZN208\_HUMAN 586 ZN248\_HUMAN 587 ZN571\_HUMAN 588 ZN699\_HUMAN 589 ZN726\_HUMAN 590 ZIK1\_HUMAN 591 ZNF2\_HUMAN 592 Z705F\_HUMAN 593 ZNF14\_HUMAN 594 ZN471\_HUMAN 595 ZN624\_HUMAN 596 ZNF84\_HUMAN 597 ZNF7\_HUMAN 598 ZN891\_HUMAN 599 ZN337\_HUMAN 600 Z705G\_HUMAN 601 ZN529\_HUMAN 602 ZN729\_HUMAN 603 ZN419\_HUMAN 604 Z705A\_HUMAN 605 ZNF45\_HUMAN 606 ZN302\_HUMAN 607 ZN486\_HUMAN 608 ZN621\_HUMAN 609 ZN688\_HUMAN 610 ZN33A\_HUMAN 611 ZN554\_HUMAN 612 ZN878\_HUMAN 613 ZN772\_HUMAN 614 ZN224\_HUMAN 615 ZN184\_HUMAN 616 ZN544\_HUMAN 617 ZNF57\_HUMAN 618 ZN283\_HUMAN 619 ZN549\_HUMAN 620 ZN211\_HUMAN 621 ZN615\_HUMAN 622 ZN253\_HUMAN 623 ZN226\_HUMAN 624 ZN730\_HUMAN 625 Z585A\_HUMAN 626 ZN732\_HUMAN 627 ZN681\_HUMAN 628 ZN667\_HUMAN 629 ZN649\_HUMAN 630 ZN470\_HUMAN 631 ZN484\_HUMAN 632 ZN431\_HUMAN 633 ZN382\_HUMAN 634 ZN254\_HUMAN 635 ZN124\_HUMAN 636 ZN607\_HUMAN 637 ZN317\_HUMAN 638 ZN620\_HUMAN 639 ZN141\_HUMAN 640 ZN584\_HUMAN 641 ZN540\_HUMAN 642 ZN75D\_HUMAN 643 ZN555\_HUMAN 644 ZN658\_HUMAN 645 ZN684\_HUMAN 646 RBAK\_HUMAN 647 ZN829\_HUMAN 648 ZN582\_HUMAN 649 ZN112\_HUMAN 650 ZN716\_HUMAN 651 HKR1\_HUMAN 652 ZN350\_HUMAN 653 ZN480\_HUMAN 654 ZN416\_HUMAN 655 ZNF92\_HUMAN 656 ZN100\_HUMAN 657 ZN736\_HUMAN 658 ZNF74\_HUMAN 659 CBX1\_HUMAN 660 ZN443\_HUMAN 661 ZN195\_HUMAN 662 ZN530\_HUMAN 663 ZN782\_HUMAN 664 ZN791\_HUMAN 665 ZN331\_HUMAN 666 Z354C\_HUMAN 667 ZN157\_HUMAN ZN727\_HUMAN 669 ZN550\_HUMAN 670 ZN793\_HUMAN 671 ZN235\_HUMAN 672 ZNF8\_HUMAN 673 ZN724\_HUMAN 674 ZN573\_HUMAN 675 ZN577\_HUMAN 676 ZN789\_HUMAN 677 ZN718\_HUMAN 678 ZN300\_HUMAN 679 ZN383\_HUMAN 680 ZN429\_HUMAN 681 ZN677\_HUMAN 682 ZN850\_HUMAN 683 ZN454\_HUMAN 684 ZN257\_HUMAN 685 ZN264\_HUMAN 686 ZFP82\_HUMAN 687 ZFP14\_HUMAN 688 ZN485\_HUMAN 689 ZN737\_HUMAN 690 ZNF44\_HUMAN 691 ZN596\_HUMAN 692 ZN565\_HUMAN 693 ZN543\_HUMAN 694 ZFP69\_HUMAN 695 SUMO1\_HUMAN 696 ZNF12\_HUMAN 697 ZN169\_HUMAN 698 ZN433\_HUMAN 699

SUMO3\_HUMAN 700 ZNF98\_HUMAN 701 ZNF98\_HUMAN 702 ZN347\_HUMAN 703  
ZNF25\_HUMAN 704 ZN519\_HUMAN 705 Z585B\_HUMAN 706 ZIM3\_HUMAN 707  
ZN517\_HUMAN 708 ZN846\_HUMAN 709 ZN230\_HUMAN 710 ZNF66\_HUMAN 711  
ZFP1\_HUMAN 712 ZN713\_HUMAN 713 ZN816\_HUMAN 714 ZN426\_HUMAN 715  
ZN674\_HUMAN 716 ZN627\_HUMAN 717 ZNF20\_HUMAN 718 Z587B\_HUMAN 719  
ZN316\_HUMAN 720 ZN233\_HUMAN 721 ZN611\_HUMAN 722 ZN556\_HUMAN 723  
ZN234\_HUMAN 724 ZN560\_HUMAN 725 ZNF77\_HUMAN 726 ZN682\_HUMAN 727  
ZN614\_HUMAN 728 ZN785\_HUMAN 729 ZN445\_HUMAN 730 ZFP30\_HUMAN 731  
ZN225\_HUMAN 732 ZN551\_HUMAN 733 ZN610\_HUMAN 734 ZN528\_HUMAN 735  
ZN284\_HUMAN 736 ZN418\_HUMAN 737 MPP8\_HUMAN 738 ZN490\_HUMAN 739  
ZN805\_HUMAN 740 Z780B\_HUMAN 741 ZN763\_HUMAN 742 ZN285\_HUMAN 743  
ZNF85\_HUMAN 744 ZN223\_HUMAN 745 ZNF90\_HUMAN 746 ZN557\_HUMAN 747  
ZN425\_HUMAN 748 ZN229\_HUMAN 749 ZN606\_HUMAN 750 ZN155\_HUMAN 751  
ZN222\_HUMAN 752 ZN442\_HUMAN 753 ZNF91\_HUMAN 754 ZN135\_HUMAN 755  
ZN778\_HUMAN 756 RYBP\_HUMAN 757 ZN534\_HUMAN 758 ZN586\_HUMAN 759  
ZN567\_HUMAN 760 ZN440\_HUMAN 761 ZN583\_HUMAN 762 ZN441\_HUMAN 763  
ZNF43\_HUMAN 764 CBX5\_HUMAN 765 ZN589\_HUMAN 766 ZNF10\_HUMAN 767  
ZN563\_HUMAN 768 ZN561\_HUMAN 769 ZN136\_HUMAN 770 ZN630\_HUMAN 771  
ZN527\_HUMAN 772 ZN333\_HUMAN 773 Z324B\_HUMAN 774 ZN786\_HUMAN 775  
ZN709\_HUMAN 776 ZN792\_HUMAN 777 ZN599\_HUMAN 778 ZN613\_HUMAN 779  
ZF69B\_HUMAN 780 ZN799\_HUMAN 781 ZN569\_HUMAN 782 ZN564\_HUMAN 783  
ZN546\_HUMAN 784 ZFP92\_HUMAN 785 YAF2\_HUMAN 786 ZN723\_HUMAN 787  
ZNF34\_HUMAN 788 ZN439\_HUMAN 789 ZFP57\_HUMAN 790 ZNF19\_HUMAN 791  
ZN404\_HUMAN 792 ZN274\_HUMAN 793 CBX3\_HUMAN 794 ZNF30\_HUMAN 795  
ZN250\_HUMAN 796 ZN570\_HUMAN 797 ZN675\_HUMAN 798 ZN695\_HUMAN 799  
ZN548\_HUMAN 800 ZN132\_HUMAN 801 ZN738\_HUMAN 802 ZN420\_HUMAN 803  
ZN626\_HUMAN 804 ZN559\_HUMAN 805 ZN460\_HUMAN 806 ZN268\_HUMAN 807  
ZN304\_HUMAN 808 ZIM2\_HUMAN 809 ZN605\_HUMAN 810 ZN844\_HUMAN 811  
SUMO5\_HUMAN 812 ZN101\_HUMAN 813 ZN783\_HUMAN 814 ZN417\_HUMAN 815  
ZN182\_HUMAN 816 ZN823\_HUMAN 817 ZN177\_HUMAN 818 ZN197\_HUMAN 819  
ZN717\_HUMAN 820 ZN669\_HUMAN 821 ZN256\_HUMAN 822 ZN251\_HUMAN 823  
CBX4\_HUMAN 824 PCGF2\_HUMAN 825 CDY2\_HUMAN 826 CDYL2\_HUMAN 827  
HERC2\_HUMAN 828 ZN562\_HUMAN 829 ZN461\_HUMAN 830 Z324A\_HUMAN 831  
ZN766\_HUMAN 832 ID2\_HUMAN 833 TOX\_HUMAN 834 ZN274\_HUMAN 835 SCMH1\_HUMAN  
836 ZN214\_HUMAN 837 CBX7\_HUMAN 838 ID1\_HUMAN 839 CREM\_HUMAN 840  
SCX\_HUMAN 841 ASCL1\_HUMAN 842 ZN764\_HUMAN 843 SCML2\_HUMAN 844  
TWST1\_HUMAN 845 CREB1\_HUMAN 846 TERF1\_HUMAN 847 ID3\_HUMAN 848  
CBX8\_HUMAN 849 CBX4\_HUMAN 850 GSX1\_HUMAN 851 NKX22\_HUMAN 852  
ATF1\_HUMAN 853 TWST2\_HUMAN 854 ZNF17\_HUMAN 855 TOX3\_HUMAN 856  
TOX4\_HUMAN 857 ZMYM3\_HUMAN 858 I2BP1\_HUMAN 859 RHXF1\_HUMAN 860  
SSX2\_HUMAN 861 I2BPL\_HUMAN 862 ZN680\_HUMAN 863 CBX1\_HUMAN 864  
TRI68\_HUMAN 865 HXA13\_HUMAN 866 PHC3\_HUMAN 867 TCF24\_HUMAN 868  
CBX3\_HUMAN 869 HXB13\_HUMAN 870 HEY1\_HUMAN 871 PHC2\_HUMAN 872  
ZNF81\_HUMAN 873 FIGLA\_HUMAN 874 SAM11\_HUMAN 875 KMT2B\_HUMAN 876  
HEY2\_HUMAN 877 JDP2\_HUMAN 878 HXC13\_HUMAN 879 ASCL4\_HUMAN 880  
HHEX\_HUMAN 881 HERC2\_HUMAN 882 GSX2\_HUMAN 883 BIN1\_HUMAN 884  
ETV7\_HUMAN 885 ASCL3\_HUMAN 886 PHC1\_HUMAN 887 OTP\_HUMAN 888 I2BP2\_HUMAN  
889 VGLL2\_HUMAN 890 HXA11\_HUMAN 891 PDLI4\_HUMAN 892 ASCL2\_HUMAN 893  
CDX4\_HUMAN 894 ZN860\_HUMAN 895 LMBL4\_HUMAN 896 PDIP3\_HUMAN 897  
NKX25\_HUMAN 898 CEBPB\_HUMAN 899 ISL1\_HUMAN 900 CDX2\_HUMAN 901  
PROP1\_HUMAN 902 SIN3B\_HUMAN 903 SMBTI\_HUMAN 904 HXC11\_HUMAN 905  
HXC10\_HUMAN 906 PRS6A\_HUMAN 907 VSX1\_HUMAN 908 NKX23\_HUMAN 909



MTG16\_HUMAN 910 HMX3\_HUMAN 911 HMX2\_HUMAN 912 KIF22\_HUMAN 913  
 CSTF2\_HUMAN 914 CEBPE\_HUMAN 915 DLX2\_HUMAN 916 ZMYM3\_HUMAN 917  
 PPARG\_HUMAN 918 PRICI\_HUMAN 919 UNC4\_HUMAN 920 BARX2\_HUMAN 921  
 ALX3\_HUMAN 922 TCF15\_HUMAN 923 TERA\_HUMAN 924 VSX2\_HUMAN 925  
 HXD12\_HUMAN 926 CDX1\_HUMAN 927 TCF23\_HUMAN 928 ALX1\_HUMAN 929  
 HXA10\_HUMAN 930 RX\_HUMAN 931 CXXC5\_HUMAN 932 SCML1\_HUMAN 933  
 NFIL3\_HUMAN 934 DLX6\_HUMAN 935 MTG8\_HUMAN 936 CBX8\_HUMAN 937  
 CEBPD\_HUMAN 938 SEC13\_HUMAN 939 FIP1\_HUMAN 940 ALX4\_HUMAN 941  
 LHX3\_HUMAN 942 PRIC2\_HUMAN 943 MAGI3\_HUMAN 944 NELLI\_HUMAN 945  
 PRRX1\_HUMAN 946 MTG8R\_HUMAN 947 RAX2\_HUMAN 948 DLX3\_HUMAN 949  
 DLX1\_HUMAN 950 NKX26\_HUMAN 951 NABI\_HUMAN 952 SAMD7\_HUMAN 953  
 PITX3\_HUMAN 954 WDR5\_HUMAN 955 MEOX2\_HUMAN 956 NAB2\_HUMAN 957  
 DHX8\_HUMAN 958 FOXA2\_HUMAN 959 CBX6\_HUMAN 960 EMX2\_HUMAN 961  
 CPSF6\_HUMAN 962 HXC12\_HUMAN 963 KDM4B\_HUMAN 964 LMBL3\_HUMAN 965  
 PHX2A\_HUMAN 966 EMX1\_HUMAN 967 NC2B\_HUMAN 968 DLX4\_HUMAN 969  
 SRY\_HUMAN 970 ZN777\_HUMAN 971 NELLI\_HUMAN 972 ZN398\_HUMAN 973  
 GATA3\_HUMAN 974 BSH\_HUMAN 975 SF3B4\_HUMAN 976 TEADI\_HUMAN 977  
 TEAD3\_HUMAN 978 RGAP1\_HUMAN 979 PHF1\_HUMAN 980 FOXA1\_HUMAN 981  
 GATA2\_HUMAN 982 FOXO3\_HUMAN 983 ZN212\_HUMAN 984 IRX4\_HUMAN 985  
 ZBED6\_HUMAN 986 LHX4\_HUMAN 987 SIN3A\_HUMAN 988 RBBP7\_HUMAN 989  
 NKX61\_HUMAN 990 TRI68\_HUMAN 991 R51A1\_HUMAN 992 MB3L1\_HUMAN 993  
 DLX5\_HUMAN 994 NOTCI\_HUMAN 995 TERF2\_HUMAN 996 ZN282\_HUMAN 997  
 RGS12\_HUMAN 998 ZN840\_HUMAN 999 SPI2B\_HUMAN 1000 PAX7\_HUMAN 1001  
 NKX62\_HUMAN 1002 ASXL2\_HUMAN 1003 FOXO1\_HUMAN 1004 GATA3\_HUMAN 1005  
 GATAI\_HUMAN 1006 ZMYM5\_HUMAN 1007 ZN783\_HUMAN 1008 SPI2B\_HUMAN 1009  
 LRP1\_HUMAN 1010 MIXLI\_HUMAN 1011 SGT1\_HUMAN 1012 LMCDI\_HUMAN 1013  
 CEBPA\_HUMAN 1014 GATA2\_HUMAN 1015 SOX14\_HUMAN 1016 WTIP\_HUMAN 1017  
 PRP19\_HUMAN 1018 CBX6\_HUMAN 1019 NKX11\_HUMAN 1020 RBBP4\_HUMAN 1021  
 DMRT2\_HUMAN 1022 SMCA2\_HUMAN 1023 ZNF10\_HUMAN 1024 EED\_HUMAN 1025  
 RCOR1\_HUMAN 1026

(127) A functional analog of any one of the above-listed proteins, i.e., a molecule having the same or substantially the same biological function (e.g., retaining 70% or more, 80% or more, 90% or more, 95% or more, or 98% or more) of the protein's transcription factor function) is encompassed by the present disclosure. For example, the functional analog may be an isoform or a variant of the above-listed protein, e.g., containing a portion of the above protein with or without additional amino acid residues and/or containing mutations relative to the above protein. In some embodiments, the functional analog has a sequence identity that is at least 75, 80, 85, 90, 95, 98, or 99% to one of the sequences listed in Table 4. Homologs, orthologs, and mutants of the above-listed proteins are also contemplated.

(128) In certain embodiments, an epigenetic editor described herein comprises a KRAB domain derived from KOX1, ZIM3, ZFP28, or ZN627, and/or an effector domain derived from KAP1, MECP2, HP1a, HP1b, CBX8, CDYL2, TOX, TOX3, TOX4, EED, EZH2, RBBP4, RCOR1, or SCML2, optionally wherein the parental protein is a human protein. In particular embodiments, an epigenetic editor described herein comprises a domain derived from KOX1, ZIM3, ZFP28, and/or ZN627, optionally wherein the parental protein is a human protein. In certain embodiments, the epigenetic editor may comprise a KRAB domain derived from KOX1 (ZNF10), e.g., a human KOX1. In certain embodiments, the epigenetic editor may comprise a KRAB domain derived from ZIM3 (ZNF657 or ZNF264), e.g., a human ZIM3. In certain embodiments, the epigenetic editor may comprise a KRAB domain derived from ZFP28, e.g., a human ZFP28. In certain embodiments, the epigenetic editor may comprise a KRAB domain derived from ZN627, e.g., a human ZN627. In certain embodiments, an epigenetic editor described herein may comprise a CDYL2, e.g., a human CDYL2, and/or a TOX domain (e.g., a human TOX domain) in combination with a KOX1 KRAB domain (e.g., a human KOX1 KRAB domain).

(129) In certain embodiments, an epigenetic effector described herein comprises a repression domain

derived from ZNF10 (SEQ ID NO: 1024). For example, the repression domain may comprise the sequence of SEQ ID NO: 1024, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1024.

### (130) B. DNA Methyltransferases

(131) In some embodiments, an effector domain of an epigenetic editor described herein alters target gene expression through DNA modification, such as methylation. Highly methylated areas of DNA tend to be less transcriptionally active than less methylated areas. DNA methylation occurs primarily at CpG sites (shorthand for “C-phosphate-G-” or “cytosine-phosphate-guanine” sites). Many mammalian genes have promoter regions near or including CpG islands (nucleic acid regions with a high frequency of CpG dinucleotides).

(132) An effector domain described herein may be, e.g., a DNA methyltransferase (DNMT) or a catalytic domain thereof, or may be capable of recruiting a DNA methyltransferase. DNMTs encompass enzymes that catalyze the transfer of a methyl group to a DNA nucleotide, such as canonical cytosine-5 DNMTs that catalyze the addition of methyl groups to genomic DNA (e.g., DNMT1, DNMT3A, DNMT3B, and DNMT3C). This term also encompasses non-canonical family members that do not catalyze methylation themselves but that recruit (including activate) catalytically active DNMTs; a non-limiting example of such a DNMT is DNMT3L. See, e.g., Lyko, *Nat Review* (2018) 19:81-92. Unless otherwise indicated, a DNMT domain may refer to a polypeptide domain derived from a catalytically active DNMT (e.g., DNMT1, DNMT3A, and DNMT3B) or from a catalytically inactive DNMT (e.g., DNMT3L). A DNMT may repress expression of the target gene through the recruitment of repressive regulatory proteins. In some embodiments, the methylation is at a CG (or CpG) dinucleotide sequence. In some embodiments, the methylation is at a CHG or CHH sequence, where H is any one of A, T, or C. In some embodiments, DNMTs in the epigenetic editors may include, e.g., DNMT1, DNMT3A, DNMT3B, and/or DNMT3C. In some embodiments, the DNMT is a mammalian (e.g., human or murine) DNMT. In particular embodiments, the DNMT is DNMT3A (e.g., human DNMT3A). In certain embodiments, an epigenetic editor described herein comprises a DNMT3A domain comprising SEQ ID NO: 1028, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1028. In certain embodiments, an epigenetic editor described herein comprises a DNMT3A domain comprising SEQ ID NO: 1029, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1029. In some embodiments, the DNMT3A domain may have, e.g., a mutation at position H739 (such as H739A or H739E), R771 (such as R771L) and/or R836 (such as R836A or R836Q), or any combination thereof (numbering according to SEQ ID NO: 1028).

(133) In some embodiments, an effector domain described herein may be a DNMT-like domain. As used herein a “DNMT-like domain” is a regulatory factor of DNA methyltransferase that may activate or recruit other DNMT domains, but does not itself possess methylation activity. In some embodiments, the DNMT-like domain is a mammalian (e.g., human or mouse) DNMT-like domain. In certain embodiments, the DNMT-like domain is DNMT3L, which may be, for example, human DNMT3L or mouse DNMT3L. In certain embodiments, an epigenetic editor described herein comprises a DNMT3L domain comprising SEQ ID NO: 1032, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1032. In certain embodiments, an epigenetic editor herein comprises a DNMT3L domain comprising SEQ ID NO: 1033, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1033. In certain embodiments, an epigenetic editor described herein comprises a DNMT3L domain comprising SEQ ID NO: 1034, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1034. In certain embodiments, an epigenetic editor described herein comprises a DNMT3L domain comprising SEQ ID NO: 1035, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to SEQ ID NO: 1035. In some embodiments, the DNMT3L domain may have, e.g., a mutation corresponding to that at position D226 (such as D226V), Q268 (such as Q268K), or both (numbering according to SEQ ID NO: 1032).

(134) In certain embodiments, an epigenetic editor herein may comprise comprising both DNMT and

DNMT-like effector domains. For example, the epigenetic editor may comprise a DNMT3A-3L domain, wherein DNMT3A and DNMT3L may be covalently linked. In other embodiments, an epigenetic editor described herein may comprise an effector domain that comprises only a DNMT3A domain (e.g., human DNMT3A), or only a DNMT-like domain (e.g., DNMT3L, which may be human or mouse DNMT3L). (135) Table 5 below provides exemplary methyltransferases from which an effector domain of an epigenetic editor described herein may be derived. See Table 18 for sequences of these exemplary methyltransferases.

(136) TABLE-US-00008

Protein Name	Species	Target Sequence	DNMT1	Human	5mC	SEQ ID NO:	Protein
DNMT3A	Human	5mC	1027	1027	1027	DNMT3A	
Human 5mC	Human	5mC	1028	1028	1028	DNMT3A	
Human 5mC	Human	5mC	1029	1029	1029	DNMT3A	
Human 5mC	Human	5mC	1030	1030	1030	DNMT3B	
Human 5mC	Human	5mC	1031	1031	1031	DNMT3C	
Human 5mC	Human	5mC	1032	1032	1032	DNMT3L	
Human 5mC	Human	5mC	1033	1033	1033	DNMT3L	
Human 5mC	Human	5mC	1034	1034	1034	DNMT3L	
Human 5mC	Human	5mC	1035	1035	1035	DNMT3L	
Human 5mC	Human	5mC	1036	1036	1036	DNMT2	
Human 5mC	Human	5mC	1037	1037	1037	DNMT2	
Human 5mC	Human	5mC	1038	1038	1038	DNMT2	
Human 5mC	Human	5mC	1039	1039	1039	DNMT2	
Human 5mC	Human	5mC	1040	1040	1040	DNMT2	
Human 5mC	Human	5mC	1041	1041	1041	DNMT2	
Human 5mC	Human	5mC	1042	1042	1042	DNMT2	
Human 5mC	Human	5mC	1043	1043	1043	DNMT2	
Human 5mC	Human	5mC	1044	1044	1044	DNMT2	
Human 5mC	Human	5mC	1045	1045	1045	DNMT2	
Human 5mC	Human	5mC	1046	1046	1046	DNMT2	
Human 5mC	Human	5mC	1047	1047	1047	DNMT2	
Human 5mC	Human	5mC	1048	1048	1048	DNMT2	
Human 5mC	Human	5mC	1049	1049	1049	DNMT2	
Human 5mC	Human	5mC	1050	1050	1050	DNMT2	
Human 5mC	Human	5mC	1051	1051	1051	DNMT2	
Human 5mC	Human	5mC	1052	1052	1052	DNMT2	
Human 5mC	Human	5mC	1053	1053	1053	DNMT2	
Human 5mC	Human	5mC	1054	1054	1054	DNMT2	
Human 5mC	Human	5mC	1055	1055	1055	DNMT2	
Human 5mC	Human	5mC	1056	1056	1056	DNMT2	
Human 5mC	Human	5mC	1057	1057	1057	DNMT2	
Human 5mC	Human	5mC	1058	1058	1058	DNMT2	
Human 5mC	Human	5mC	1059	1059	1059	DNMT2	
Human 5mC	Human	5mC	1060	1060	1060	DNMT2	
Human 5mC	Human	5mC	1061	1061	1061	DNMT2	

(137) A functional analog of any one of the above-listed proteins, i.e., a molecule having the same or substantially the same biological function (e.g., retaining 70% or more, 80% or more, 90% or more, 95% or more, or 98% or more) of the protein's DNA methylation function or recruiting function) is encompassed by the present disclosure. For example, the functional analog may be an isoform or a variant of the above-listed protein, e.g., containing a portion of the above protein with or without additional amino acid residues and/or containing mutations relative to the above protein. In some embodiments, the functional analog has a sequence identity that is at least 75, 80, 85, 90, 95, 98, or 99% to one of the sequences listed in Table 5. In some embodiments, the effector domain herein comprises only the functional domain (or functional analog thereof), e.g., the catalytical domain or recruiting domain, of the above-listed proteins.

(138) As used herein, a DNMT domain (e.g., a DNMT3A domain or a DNMT3L domain) refers to a protein domain that is identical to the parental protein (e.g., a human or murine DNMT3A or DNMT3L) or a functional analog thereof (e.g., having a functional fragment, such as a catalytic fragment or recruiting fragment, of the parental protein; and/or having mutations that improve the activity of the DNMT protein).

(139) An epigenetic editor herein may effect methylation at, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 300, 400, 500, 600, 700, 800, 900, or 1000 or more CpG dinucleotide sequences in the target gene or chromosome. The CpG dinucleotide sequences may be located within or near the target gene in CpG islands, or may be located in a region that is not a CpG island. A CpG island generally refers to a nucleic acid sequence or chromosome region that comprises a high frequency of CpG dinucleotides. For example, a CpG island may comprise at least 50% GC content. The CpG island may have a high observed-to-expected CpG ratio, for example, an observed-to-expected CpG ratio of at least 60%. As used herein, an observed-to-expected CpG ratio is

determined by  $\text{Number of CpG}^*/(\text{sequence length})/(\text{Number of C}^*\text{Number of G})$ . In some embodiments, the CpG island has an observed-to-expected CpG ratio of at least 60%, 70%, 80%, 90% or more. A CpG island may be a sequence or region of, e.g., at least 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, or 800 nucleotides. In some embodiments, only 1, or less than 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, or 50 CpG dinucleotides are methylated by the epigenetic editor.

(140) In some embodiments, an epigenetic editor herein effects methylation at a hypomethylated nucleic acid sequence, i.e., a sequence that may lack methyl groups on the 5-methyl cytosine nucleotides (e.g., in CpG) as compared to a standard control. Hypomethylation may occur, for example, in aging cells or in cancer (e.g., early stages of neoplasia) relative to a younger cell or non-cancer cell, respectively.

(141) In some embodiments, an epigenetic editor described herein induces methylation at a hypermethylated nucleic acid sequence.

(142) In some embodiments, methylation may be introduced by the epigenetic editor at a site other than a CpG dinucleotide. For example, the target gene sequence may be methylated at the C nucleotide of CpA, CpT, or CpC sequences. In some embodiments, an epigenetic editor comprises a DNMT3A domain and effects methylation at CpG, CpA, CpT, CpC sequences, or any combination thereof. In some embodiments, an epigenetic editor comprises a DNMT3A domain that lacks a regulatory subdomain and only maintains a catalytic domain. In some embodiments, the epigenetic editor comprising a DNMT3A catalytic domain effects methylation exclusively at CpG sequences. In some embodiments, an epigenetic editor comprising a DNMT3A domain that comprises a mutation, e.g. a R836A or R836Q mutation (numbering according to SEQ ID NO: 1028), has higher methylation activity at CpA, CpC, and/or CpT sequences as compared to an epigenetic editor comprising a wildtype DNMT3A domain.

(143) C. Histone Modifiers

(144) In some embodiments, an effector domain of an epigenetic editor herein mediates histone modification. Histone modifications play a structural and biochemical role in gene transcription, such as by formation or disruption of the nucleosome structure that binds to the histone and prevents gene transcription. Histone modifications may include, for example, acetylation, deacetylation, methylation, phosphorylation, ubiquitination, SUMOylation and the like, e.g., at their N-terminal ends ("histone tails"). These modifications maintain or specifically convert chromatin structure, thereby controlling responses such as gene expression, DNA replication, DNA repair, and the like, which occur on chromosomal DNA. Post-translational modification of histones is an epigenetic regulatory mechanism and is considered essential for the genetic regulation of eukaryotic cells. Recent studies have revealed that chromatin remodeling factors such as SWI/SNF, RSC, NURF, NRD, and the like, which facilitate transcription factor access to DNA by modifying the nucleosome structure; histone acetyltransferases (HATs) that regulate the acetylation state of histones; and histone deacetylases (HDACs), act as important regulators.

(145) In particular, the unstructured N-termini of histones may be modified by acetylation, deacetylation, methylation, ubiquitylation, phosphorylation, SUMOylation, ribosylation, citrullination O-GlcNAcylation, crotonylation, or any combination thereof. For example, histone acetyltransferases (HATs) utilize acetyl-CoA as a cofactor and catalyze the transfer of an acetyl group to the epsilon amino group of the lysine side chains. This neutralizes the lysine's positive charge and weakens the interactions between histones and DNA, thus opening the chromosomes for transcription factors to bind and initiate transcription. Acetylation of K14 and K9 lysines of histone H3 by histone acetyltransferase enzymes may be linked to transcriptional competence in humans. Lysine acetylation may directly or indirectly create binding sites for chromatin-modifying enzymes that regulate transcriptional activation. On the other hand, histone methylation of lysine 9 of histone H3 may be associated with heterochromatin, or transcriptionally silent chromatin.

(146) In certain embodiments, an effector domain of an epigenetic editor described herein comprises a histone methyltransferase domain. The effector domain may comprise, for example, a DOT1L domain, a SET domain, a SUV39H1 domain, a G9a/EHMT2 protein domain, an EZH1 domain, an EZH2 domain, a SETDB1 domain, or any combination thereof. In particular embodiments, the effector domain comprises a histone-lysine-N-methyltransferase SETDB1 domain.

(147) In some embodiments, the effector domain comprises a histone deacetylase protein domain. In certain embodiments, the effector domain comprises a HDAC family protein domain, for example, a HDAC1, HDAC3, HDAC5, HDAC7, or HDAC9 protein domain. In particular embodiments, the effector domain comprises a nucleosome remodeling and deacetylase complex (NURD), which removes acetyl groups from histones.

#### (148) D. Other Effector Domains

(149) In some embodiments, the effector domain comprises a tripartite motif containing protein (TRIM28, TIF1-beta, or KAP1). In certain embodiments, the effector domain comprises one or more KAP1 proteins. A KAP1 protein in an epigenetic editor herein may form a complex with one or more other effector domains of the epigenetic editor or one or more proteins involved in modulation of gene expression in a cellular environment. For example, KAP1 may be recruited by a KRAB domain of a transcriptional repressor. A KAP1 protein domain may interact with or recruit one or more protein complexes that reduces or silences gene expression. In some embodiments, KAP1 interacts with or recruits a histone deacetylase protein, a histone-lysine methyltransferase protein, a chromatin remodeling protein, and/or a heterochromatin protein. For example, a KAP1 protein domain may interact with or recruit a heterochromatin protein 1 (HP1) protein, a SETDB1 protein, an HDAC protein, and/or a NuRD protein complex component. In some embodiments, a KAP1 protein domain interacts with or recruits a ZFP90 protein (e.g., isoform 2 of ZFP90), and/or a FOXP3 protein. An exemplary KAP1 amino acid sequence is shown in SEQ ID NO: 1062.

(150) In some embodiments, the effector domain comprises a protein domain that interacts with or is recruited by one or more DNA epigenetic marks. For example, the effector domain may comprise a methyl CpG binding protein 2 (MECP2) protein that interacts with methylated DNA nucleotides in the target gene (which may or may not be at a CpG island of the target gene). An MECP2 protein domain in an epigenetic editor described herein may induce condensed chromatin structure, thereby reducing or silencing expression of the target gene. In some embodiments, an MECP2 protein domain in an epigenetic editor described herein may interact with a histone deacetylase (e.g. HDAC), thereby repressing or silencing expression of the target gene. In some embodiments, an MECP2 protein domain in an epigenetic editor described herein may block access of a transcription factor or transcriptional activator to the target sequence, thereby repressing or silencing expression of the target gene. An exemplary MECP2 amino acid sequence is shown in SEQ ID NO: 1063.

(151) Also contemplated as effector domains for the epigenetic editors described herein are, e.g., a chromoshadow domain, a ubiquitin-2 like Rad60 SUMO-like (Rad60-SLD/SUMO) domain, a chromatin organization modifier domain (Chromo) domain, a Yaf2/RYPB C-terminal binding motif domain (YAF2\_RYPB), a CBX family C-terminal motif domain (CBX7\_C), a zinc finger C3HC4 type (RING finger) domain (ZF-C3HC4\_2), a cytochrome b5 domain (Cyt-b5), a helix-loop-helix domain (HLH), a helix-hairpin-helix motif domain (e.g., HHH\_3), a high mobility group box domain (HMG-box), a basic leucine zipper domain (e.g., bZIP\_1 or bZIP\_2), a Myb\_DNA-binding domain, a homeodomain, a MYM-type Zinc finger with FCS sequence domain (ZF-FCS), an interferon regulatory factor 2-binding protein zinc finger domain (IRF-2BP1\_2), an SSX repression domain (SSXRD), a B-box-type zinc finger domain (ZF-B\_box), a CXXC zinc finger domain (ZF-CXXC), a regulator of chromosome condensation 1 domain (RCC1), an SRC homology 3 domain (SH3\_9), a sterile alpha motif domain (SAM\_1), a sterile alpha motif domain (SAM 2), a sterile alpha motif/Pointed domain (SAM\_PNT), a Vestigial/Tondu family domain (Vg\_Tdu), a LIM domain, an RNA recognition motif domain (RRM\_1), a paired amphipathic helix domain (PAH), a proteasomal ATPase OB C-terminal domain (Prot\_ATP\_ID\_OB), a nervy homology 2 domain (NHR2), a hinge domain of cleavage stimulation factor subunit 2 (CSTF2\_hinge), a PPAR gamma N-terminal region domain (PPARgamma\_N), a CDC48 N-terminal domain (CDC48\_2), a WD40 repeat domain (WD40), a Fip1 motif domain (Fip1), a PDZ domain (PDZ\_6), a Von Willebrand factor type C domain (VWC), a NAB conserved region 1 domain (NCD1), an S1 RNA-binding domain (S1), an HNF3C-terminal domain (HNF\_C), a Tudor domain (Tudor\_2), a histone-like transcription factor (CBF/NF-Y) and archaeal histone domain (CBFD\_NFYB\_HMF), a zinc finger protein domain (DUF3669), an EGF-like domain (cEGF), a GATA zinc finger domain (GATA), a TEA/ATTS domain (TEA), a phorbol

esters/diacylglycerol binding domain (C1-1), polycomb-like MTF2 factor 2 domain (Mtf2<sub>2</sub>C), a transactivation domain of FOXO protein family (FOXO-TAD), a homeobox KN domain (Homeobox\_KN), a BED zinc finger domain (ZF-BED), a zinc finger of C3HC4-type RING domain (ZF-C3HC4\_4), a RAD51 interacting motif domain (RAD51\_interact), a p55-binding region of a methyl-CpG-binding domain protein MBD (MBDa), a Notch domain, a Raf-like Ras-binding domain (RBD), a Spin/Ssty family domain (Spin-Ssty), a PHD finger domain (PHD\_3), a Low-density lipoprotein receptor domain class A (Ldl\_recept\_a), a CS domain, a DM DNA-binding domain, and a QLQ domain.

(152) In some embodiments, the effector domain is a protein domain comprising a YAF2\_RYBP domain or homeodomain or any combination thereof. In certain embodiments, the homeodomain of the YAF2\_RYBP domain is a PRD domain, an NKL domain, a HOXL domain, or a LIM domain. In particular embodiments, the YAF2\_RYBP domain may comprise a 32 amino acid Yaf2/RYPB C-terminal binding motif domain (32 aa RYBP).

(153) In some embodiments, the effector domain comprises a protein domain selected from a group consisting of SUMO3 domain, Chromo domain from M phase phosphoprotein 8 (MPP8), chromoshadow domain from Chromobox 1 (CBX1), and SAM\_1/SPM domain from Scm Polycomb Group Protein Homolog 1 (SCMH1).

(154) In some embodiments, the effector domain comprises an HNF3 C-terminal domain (HNF\_C). The HNF\_C domain may be from FOXA1 or FOXA2. In certain embodiments, the HNF\_C domain comprises an EH1 (engrailed homology 1) motif.

(155) In some embodiments, the effector domain may comprise an interferon regulatory factor 2-binding protein zinc finger domain (IRF-2BP1\_2), a Cyt-b5 domain from DNA repair factor HERC2 E3 ligase, a variant SH3 domain (SH3\_9) from Bridging Integrator 1 (BIN1), an HMG-box domain from transcription factor TOX or ZF-C3HC4-2 RING finger domain from the polycomb component PCGF2, a Chromodomain-helicase-DNA binding protein 3 (CHD3) domain, or a ZNF783 domain.

#### IV. Epigenetic Editors

(156) Provided herein are epigenetic editors, also referred to herein as epigenetic editing systems, that direct epigenetic modification(s) to a target sequence in a gene of interest, e.g., using one or more DNA-binding domains as described herein and one or more effector domains (e.g., epigenetic repression domains) as described herein, in any combination. The DNA-binding domain (in concert with a guide polynucleotide such as one described herein, where the DNA-binding domain is a polynucleotide guided DNA-binding domain) directs the effector domain to epigenetically modify the target sequence, resulting in gene repression or silencing that may be durable and inheritable across cell generations. In some aspects, the epigenetic editors described herein can repress or silence genes reversibly or irreversibly in cells.

(157) In particular embodiments, an epigenetic editor described herein comprises one or more fusion proteins, each comprising (1) DNA-binding domain(s) and (2) effector domain(s). The effector domains may be on one or more fusion proteins comprised by the epigenetic editor. For example, a single fusion protein may comprise all of the effector domains with a DNA-binding domain. Alternatively, the effector domains or subsets thereof may be on separate fusion proteins, each with a DNA-binding domain (which may be the same or different). A fusion protein described herein may further comprise one or more linkers (e.g., peptide linkers), detectable tags, nuclear localization signals (NLSs), or any combination thereof. As used herein, a “fusion protein” refers to a chimeric protein in which two or more coding sequences (e.g., for DNA-binding domain(s) and/or effector domain(s)) are covalently or non-covalently joined, directly or indirectly.

(158) In some embodiments, an epigenetic editor described herein comprises 2, 3, 4, 5, 6, 7, 8, 9, 10, or more effector (e.g., repression) domains, which may be identical or different. In certain embodiments, two or more of said effector domains function synergistically. Combinations of effector domains may comprise DNA methylation domains, histone deacetylation domains, histone methylation domains, and/or scaffold domains that recruit any of the above. For example, an epigenetic editor described herein may comprise one or more transcriptional repressor domains (e.g., a KRAB domain such as KOX1, ZIM3, ZFP28, or ZN627 KRAB) in combination with one or more DNA methylation domains (e.g., a

DNMT domain) and/or recruiter domain (e.g., a DNMT3L domain). Such an epigenetic editor may comprise, for instance, a KRAB domain, a DNMT3A domain, and a DNMT3L domain. An epigenetic editor can comprise a DNMT3A domain and a DNMT3L domain and preferably further comprise a KRAB domain. In some embodiments, the epigenetic editor further comprises an additional effector domain (e.g., a KAP1, MECP2, HP1b, CBX8, CDYL2, TOX, TOX3, TOX4, EED, RBBP4, RCOR1, or SCML2 domain). In some embodiments, the additional effector domain is a CDYL2, TOX, TOX3, TOX4, or HP1a domain. For example, an epigenetic editor described herein may comprise a CDYL2 and/or a TOX domain in combination with a KRAB domain (e.g., a KOX1 KRAB domain).

(159) A. Linkers

(160) A fusion protein as described herein may comprise one or more linkers that connect components of the epigenetic editor. A linker may be a peptide or non-peptide linker.

(161) In some embodiments, one or more linkers utilized in an epigenetic editor provided herein is a peptide linker, i.e., a linker comprising a peptide moiety. A peptide linker can be any length applicable to the epigenetic editor fusion proteins described herein. In some embodiments, the linker can comprise a peptide between 1 and 200 (e.g., between 1 and 80) amino acids. In some embodiments, the linker comprises from 1 to 5, 1 to 10, 1 to 20, 1 to 30, 1 to 40, 1 to 50, 1 to 60, 1 to 80, 1 to 100, 1 to 150, 1 to 200, 5 to 10, 5 to 20, 5 to 30, 5 to 40, 5 to 60, 5 to 80, 5 to 100, 5 to 150, 5 to 200, 10 to 20, 10 to 30, 10 to 40, 10 to 50, 10 to 60, 10 to 80, 10 to 100, 10 to 150, 10 to 200, 20 to 30, 20 to 40, 20 to 50, 20 to 60, 20 to 80, 20 to 100, 20 to 150, 20 to 200, 30 to 40, 30 to 50, 30 to 60, 30 to 80, 30 to 100, 30 to 150, 30 to 200, 40 to 50, 40 to 60, 40 to 80, 40 to 100, 40 to 150, 40 to 200, 50 to 60, 50 to 80, 50 to 100, 50 to 150, 50 to 200, 60 to 80, 60 to 100, 60 to 150, 60 to 200, 80 to 100, 80 to 150, 80 to 200, 100 to 150, 100 to 200, or 150 to 200 amino acids in length. Longer or shorter linkers are also contemplated. In some embodiments, the peptide linker is 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 25, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, or 100 amino acids in length. For example, the peptide linker may be 4, 5, 16, 20, 24, 27, 32, 40, 64, 92, or 104 amino acids in length. The peptide linker may be a flexible or rigid linker. In particular embodiments, the peptide linker comprises the amino acid sequence of any one of SEQ ID NOs: 1064-1068 or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical thereto.

(162) In certain embodiments, the peptide linker is an XTEN linker. Such a linker may comprise part of the XTEN sequence (Schellenberger et al., *Nat Biotechnol* (2009) 27(1):1186-90), an unstructured hydrophilic polypeptide consisting only of residues G, S, P, T, E, and A. The term “XTEN” as used herein refers to a recombinant peptide or polypeptide lacking hydrophobic amino acid residues. XTEN linkers typically are unstructured and comprise a limited set of natural amino acids. Fusion of XTEN to proteins alters its hydrodynamic properties and reduces the rate of clearance and degradation of the fusion protein. These XTEN fusion proteins are produced using recombinant technology, without the need for chemical modifications, and degraded by natural pathways. The XTEN linker may be, for example, 5, 10, 16, 20, 26, or 80 amino acids in length. In some embodiments, the XTEN linker is 16 amino acids in length. In some embodiments, the XTEN linker is 80 amino acids in length. In certain embodiments, the XTEN linker may be XTEN10, XTEN16, XTEN20, or XTEN80. In certain embodiments, the XTEN linker may comprise the amino acid sequence of any one of SEQ ID NOs: 1069-1073 and 1092 or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical thereto. In some embodiments, the XTEN linker may be XTEN10, XTEN16, XTEN20, or XTEN80.

(163) In some embodiments, one or more linkers utilized in an epigenetic editor provided herein is a non-peptide linker. For example, the linker may be a carbon bond, a disulfide bond, or carbon-heteroatom bond. In certain embodiments, the linker is a carbon-nitrogen bond of an amide linkage. In certain embodiments, the linker is a cyclic or acyclic, substituted or unsubstituted, or branched or unbranched aliphatic or heteroaliphatic linker.

(164) In some embodiments, one or more linkers utilized in an epigenetic editor provided herein is polymeric (e.g., polyethylene, polyethylene glycol, polyamide, polyester, etc.). The linker may comprise, for example, a monomer, dimer, or polymer of aminoalkanoic acid; an aminoalkanoic acid (e.g., glycine, ethanoic acid, alanine, beta-alanine, 3-aminopropanoic acid, 4-aminobutanoic acid, 5-

pentanoic acid, etc.); a monomer, dimer, or polymer of aminohexanoic acid (Ahx); or a polyethylene glycol moiety (PEG); or an aryl or heteroaryl moiety. In certain embodiments, the linker may be based on a carbocyclic moiety (e.g., cyclopentane or cyclohexane) or a phenyl ring. The linker may include functionalized moieties to facilitate attachment of a nucleophile (e.g., thiol, amino) from the peptide to the linker. Any electrophile may be used as part of the linker. Exemplary electrophiles include, but are not limited to, activated esters, activated amides, alkyl halides, aryl halides, acyl halides, and isothiocyanates.

(165) Various linker lengths and flexibilities can be employed between any two components of an epigenetic editor (e.g., between an effector domain (e.g., a repressor domain) and a DNA-binding domain (e.g., a Cas9 domain), between a first effector domain and a second effector domain, etc.). The linkers may range from very flexible linkers, such as glycine/serine-rich linkers, to more rigid linkers, in order to achieve the optimal length for effector domain activity for the specific application. In some embodiments, the more flexible linkers are glycine/serine-rich linkers (GS-rich linkers), where more than 45% (e.g., more than 48, 50, 55, 60, 70, 80, or 90%) of the residues are glycine or serine residues. Non-limiting examples of the GS-rich linkers are (GGGS)<sub>n</sub> (SEQ ID NO: 485), (G)<sub>n</sub> (SEQ ID NO: 1260), and W linker. In some embodiments, the more rigid linkers are in the form of the form (EAAAK)<sub>n</sub> (SEQ ID NO: 487), (SGGS)<sub>n</sub> (SEQ ID NO: 488), and (XP)<sub>n</sub> (SEQ ID NO: 489). In the aforementioned formulae of flexible and rigid linkers, n may be any integer between 1 and 30. In some embodiments, n is 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, or 15. In some embodiments, the linker comprises a (GGS)<sub>n</sub> motif, wherein n is 1, 3, or 7 (SEQ ID NO: 490). In some embodiments, the linker comprises a (GGGS)<sub>n</sub> motif, wherein n is 4 (SEQ ID NO: 491).

(166) In some embodiments, a linker in an epigenetic editor described herein comprises a nuclear localization signal, for example, with the amino acid sequence of any one of SEQ ID NOs: 1074-1079. In some embodiments, a linker in an epigenetic editor described herein comprises an expression tag, e.g., a detectable tag such as a green fluorescence protein.

#### (167) B. Nuclear Localization Signals

(168) A fusion protein described herein may comprise one or more nuclear localization signals, and in certain embodiments, may comprise two or more nuclear localization signals. For example, the fusion protein may comprise 1, 2, 3, 4, or 5 nuclear localization signals. As used herein, a “nuclear localization signal” (NLS) is an amino acid sequence that directs proteins to the nucleus. In certain embodiments, the NLS may be an SV40 NLS. The fusion protein may comprise an NLS at its N-terminus, C-terminus, or both, and/or an NLS may be embedded in the middle of the fusion protein (e.g., at the N- or C-terminus of a DNA-binding domain or an effector domain). In certain embodiments, an NLS comprises the amino acid sequence of any one of SEQ ID NOs: 1074-1079, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to the selected sequence. Additional NLSs are known in the art.

#### (169) C. Tags

(170) Epigenetic editors provided herein may comprise one or more additional sequences (“tags”) for tracking, detection, and localization of the editors. In some embodiments, the epigenetic editor comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more detectable tags. Each of the detectable tags may be the same or different.

(171) For example, an epigenetic editor fusion protein may comprise cytoplasmic localization sequences, export sequences, such as nuclear export sequences, or other localization sequences, as well as sequence tags that are useful for solubilization, purification, or detection of the fusion proteins. Suitable protein tags provided herein include, but are not limited to, biotin carboxylase carrier protein (BCCP) tags, myc-tags, calmodulin-tags, FLAG-tags, hemagglutinin (HA)-tags, poly-histidine tags (also referred to as histidine tags or His-tags), maltose binding protein (MBP)-tags, nus-tags, glutathione-S-transferase (GST)-tags, green fluorescent protein (GFP)-tags, thioredoxin-tags, S-tags, Softags (e.g., Softag 1 or Softag 3), strep-tags, biotin ligase tags, FlAsH tags, V5 tags, and SBP-tags. Additional suitable sequences will be apparent to those of skill in the art. Sequences disclosed herein that are presented with tag sequences included are also contemplated without the presented tag sequences; similarly, sequences disclosed herein without tag sequences are also contemplated to include the



addition of suitable tag sequences apparent to those of skill in the art.

(172) D. Fusion Protein Configurations

(173) A fusion protein of an epigenetic editor described herein may have its components structured in different configurations. For example, the DNA-binding domain may be at the C-terminus, the N-terminus, or in between two or more epigenetic effector domains or additional domains. In some embodiments, the DNA-binding domain is at the C-terminus of the epigenetic editor. In some embodiments, the DNA-binding domain is at the N-terminus of the epigenetic editor. In some embodiments, the DNA-binding domain is linked to one or more nuclear localization signals. In some embodiments, the DNA-binding domain is flanked by an epigenetic effector domain and/or an additional domain on both sides. In some embodiments, where “DBD” indicates DNA-binding domain and “ED” indicates effector domain, the epigenetic editor comprises the configuration of: N’-[ED1]-[DBD]-[ED2]-[C’ N’]-[ED1]-[DBD]-[ED2]-[ED3]-[C’ N’]-[ED1]-[ED2]-[DBD]-[ED3]-[C’ or N’]-[ED1]-[ED2]-DBD-[ED3]-[ED4]-[C’.

(174) In some embodiments, an epigenetic editor comprises a DNA-binding domain (DBD), a DNA methyltransferase (DNMT) domain, and a transcriptional repressor (“repressor”) domain that represses or silences expression of a target gene. The DBD, DNMT, and transcriptional repressor domains may be any as described herein, in any combination. For example, an epigenetic editor can comprise a DBD, a DNMT3A domain, and a DNMT3L domain. An epigenetic editor can comprise a DBD, a DNMT3A domain, a DNMT3L domain, and preferably further comprise a KRAB domain. In some embodiments, the epigenetic editor comprises a fusion protein with the configuration of N’-[DNA methyltransferase domain]-[DBD]-[repressor domain]-[C’ N’]-[repressor domain]-[DBD]-[DNA methyltransferase domain]-[C’ N’]-[DNA methyltransferase domain]-[repressor domain]-[DBD]-[C’ or N’]-[repressor domain]-[DNA methyltransferase domain]-[DBD]-[C’.

(175) In some embodiments, a connecting structure “]-[” in any one of the epigenetic editor structures is a linker, e.g., a peptide linker; a detectable tag; a peptide bond; a nuclear localization signal; and/or a promoter or regulatory sequence. In an epigenetic editor structure, the multiple connecting structures “]-[” may be the same or may each be a different linker, tag, NLS, or peptide bond. In particular embodiments, the DNA methyltransferase domain comprises DNMT3A, DNMT3L, or both. In particular embodiments, the DBD is a catalytically inactive polynucleotide guided DNA-binding domain (e.g., a dCas9) or a ZFP domain. In particular embodiments, the repressor domain is a KRAB domain.

(176) In some embodiments, the epigenetic editor comprises a configuration selected from N’-[DNMT3A-DNMT3L]-[DBD]-[KRAB]-[C’ N’]-[KRAB]-[DBD]-[DNMT3A-DNMT3L]-[C’ N’]-[KRAB]-[DBD]-[DNMT3A]-[C’ N’]-[DNMT3A]-[DBD]-[KRAB]-[C’ N’]-[KRAB]-[DBD]-[DNMT3A]-[DNMT3L]-[C’ N’]-[DNMT3A]-[DNMT3L]-[DBD]-[KRAB]-[C’ N’]-[DNMT3A]-[DBD]-[C’ N’]-[DBD]-[DNMT3A]-[C’ N’]-[DNMT3L]-[DBD]-[C’ N’]-[DBD]-[DNMT3L]-[C’ wherein [DNMT3A-DNMT3L] indicates that the DNMT3A and DNMT3L domains are directly fused via a peptide bond, and wherein the connecting structure]-[is any one of the linkers as described herein, a detectable tag, an affinity domain, a peptide bond, a nuclear localization signal, a promoter, and/or a regulatory sequence. The DBD, KRAB, DNMT3A, and DNMT3L domains may be any as described herein, in any combination. In particular embodiments, the DBD is a CRISPR-associated protein domain (e.g., dCas9) or a ZFP domain; the KRAB domain is derived from KOX1, ZIM3, ZFP28, or ZN627; the DNMT3A domain is a human DNMT3A domain; and the DNMT3L domain is a human or mouse DNMT3L domain; any combination of these components is also contemplated by the present disclosure.

(177) In some embodiments, the epigenetic editor comprises a configuration selected from N’-[DNMT3A]-[DBD]-[SETDB1]-[C’ N’]-[DNMT3A]-[DNMT3L]-[DBD]-[SETDB1]-[C’ N’]-[DNMT3A-DNMT3L]-[DBD]-[SETDB1]-[C’ N’]-[SETDB1]-[DBD]-[DNMT3A]-[DNMT3L]-[C’ N’]-[SETDB1]-[DBD]-[DNMT3A]-[C’

wherein [DNMT3A-DNMT3L] indicates that the DNMT3A and DNMT3L domains are directly fused via a peptide bond, and wherein the connecting structure]-[is any one of the linkers as described herein, a detectable tag, an affinity domain, a peptide bond, a nuclear localization signal, a promoter, and/or a regulatory sequence. The DBD, SETDB1, DNMT3A, and DNMT3L domains may be any as described herein, in any combination. In particular embodiments, the DBD is a CRISPR-associated protein domain

(e.g., dCas9) or a ZFP domain; the SETDB1 domain is derived from human SETDB1, ZIM3, ZFP28, or ZN627; the DNMT3A domain is a human DNMT3A domain; and the DNMT3L domain is a human or mouse DNMT3L domain; any combination of these components is also contemplated by the present disclosure.

(178) Particular constructs contemplated herein include: DNMT3A-DNMT3L-XTEN80-NLS-dCas9-NLS-XTEN16-KOX1 KRAB (Configuration 1), and DNMT3A-DNMT3L-XTEN80-NLS-ZFP domain-NLS-XTEN16-KOX1 KRAB (Configuration 2).

(179) In particular embodiments, the DNMT3L and DNMT3A are both derived from human parental proteins. In particular embodiments, the DNMT3L and DNMT3A are derived from human and mouse parental proteins, respectively. In particular embodiments, the DNMT3L and DNMT3A are derived from mouse and human parental proteins, respectively. In particular embodiments, the DNMT3L and DNMT3A are both derived from mouse parental proteins. In some embodiments, the dCas9 is dSpCas9. In some embodiments, the KOX1 is human KOX1.

(180) In particular embodiments, a fusion construct described herein may have Configuration 1 and comprise SEQ ID NO: 1080, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical thereto. In SEQ ID NO: 1080 below, the XTEN linkers are underlined, the NLS sequences are bolded, the DNMT3A sequence is italicized, the DNMT3L sequence is underlined and italicized, the dCas9 domain is bolded and italicized, and the KOX1 KRAB domain is underlined and bolded:

(181) TABLE-US-00009 (SEQ ID NO: 1080)

MNHDQEFDPKVPVPAEKRPKIRVLSLEDGIATGLLVLKDLGIQVDRY  
IASEVCEDSITVGMVRHQGKIMYVGDVRSVTQKHIQEWGPFDLVIGGSPC  
NDLSIVNPARKGLYEGTGRLFFEFYRLLHDARPKEGDDRPFFWLFENVVA  
MGVSDKRDISRFLSNPVMIDAKEVSAHRARYFWGNLPGMNRPLASTVN  
DKLELQECLEHGRIAKESKVRTITTRSNSIKQGKDQHFPVFMNEKEDILW  
CTEMERVFGFPVHYTDVSNMSRLARQRLGRSWSVPVIRHLFAPLKEYFA  
CVSSGNSNANSRGPSFSSGLVPLSLRGSHMGPMIYKTVSAWKRPVRVL  
SLERNIDKVLKSLGFLESGSGSGGGTLKYVEDVTNVVRRDVEKWGPEDLV  
YGSTQPLGSSCDRCPGWYMEQFHRILQYALPRQESQRPFFWIFMDNLLLT  
EDDQETTTREFLQTEAVTLQDVRGRDYQNAMRVWSNIPGLKSKHAPLTPKE  
EEYLQAQVRSRSLDAPKVDLLVKNCLLPLREYFKYFSQNSLPLGGPSSG  
APPPSGGSPAGSPTSTEEGTSESATPESGPGTSTEPSEGSAPGSPAGSPT  
STEEGTSTEPSEGSAPGTSTEPSEPKKKRKVYMDKKYSIGLAIGTNSVGW  
AVITDEYKVPSSKFKVLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTA  
RRRYTRRKNRICYLQEIFSNEMAKVDDSFHRLVESFLVEEDKKHERHPI  
FGNIVDEVAYHEKYPTIYHLRKKLVDSTDKADLRILIYALAHMIKERGHF  
LIEGDLNPDNSDVKLFIQLVQTYNQLFEENPINASGVDAKAILSARLSK  
SRRLLENLIAQLPGEKKNGLGNLIALSLGLTPNEKSNEDLAEDAKLQLSK  
DTYDDDLNLLAQIGDQYADLFLAAKNLSDAILLSDILRVNTEITKAPLS  
ASMIKRYDEHHQDLTLLKALVRQQLPKEYKEIFFDQSKNGYAGYIDGGAS  
QEEFYKFIKPILEKMDGTEELLVKNLREDLLRKQRTFDNGSIPHQIHLGE  
LHAILRRQEDFYFPLKDNREKIEKILTFRIPIYYVGPLARGNSRFAWMTRK  
SEETITPWNFEVVDKGASAQSFIERMTNEDKNLPNEKVLPKHSLLYEYF  
TVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLEKTNRKVTVKQLKEDY  
FKKIECFDSVEISGVEDRENASLGTYHDLLKIIKDKDELDNEENEDILED  
IVLTTLTFEDREMIEERLKYAHLEDDKVMKQLKRRRYTGWGRLSRKLIN  
GIRDKQSGKTILDELKSDGEANRNEMQLIHDDSLTFKEDIQKAQVSGQGD  
SLHEHIANLAGSPAIIKKGILQTVKVVDDELVKVMGRHKPENIVIEMARENQ  
TTQKGQKNSRERMKRIEELGSGILKEHPVENTQLQNEKLYLYYLQN  
GRDMYVDQELDINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSD  
NVPSEEVVKKMKNYWRQLLNAKLITQRKEDNLTKAERGGLSELDKAGFIK  
RQLVETRQITKHVAQILDSRMNTKYDENDKLIREVKVITLKSCLVSDERK

DFQFYKYNREIYKYNHAAHNAVVGTAIIKKYPKLESEFVYGDYKVVYDV  
 RKMIKSEQEIGKATAKYFFYSNIMNFFKTEITLANGEIRKRPLIETNGE  
 TGEIVWDKGRDFATVRKVLSPQVNVKKTEVQTGGFSKESILPKRNSDK  
 LIARKKDWDPKKYGGFDSPTVAYSVLVVAKEKGKSKKLKSVKELLGITI  
 MERSSFENPIDFLEAKGYKEVKKDLIIKLPKYSLFELENGRKRMLASAG  
 ELQKGNELALPSKYVNFYLYASHYEKLKGSPEDEQKQLFVEQHKHYLDE  
 IIEQISEFSKRVLADANLDKVL SAYNKH RD KPIREQAENIIHLFTLTNL  
 GAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ SITGLYETRIDLSQLGGD  
 PKKKRKVSGSETPGTSESATPESTGRTLVTFKDVFVDFTREEWKLLDTAQ  
 QIVYRNVMLENYKNLVSLGYQLTKPDVILRLEKGEEP

(182) In particular embodiments, a fusion construct described herein may have Configuration 2 and comprise SEQ ID NO: 1081, or a sequence at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% identical thereto. In SEQ ID NO: 1081 below, the XTEN linkers are underlined, the NLS sequences are bolded and underlined, the DNMT3A sequence is italicized, the DNMT3L sequence is underlined and italicized, the ZFP domain is bolded, and the KOX1 KRAB domain is underlined and bolded. Variable amino acids represented by Xs are the amino acids of the DNA-recognition helix of the zinc finger and XX in italics may be either TR, LR or LK.

(183) TABLE-US-00010 MNHDQEFDPKVVPPVPAEKRKPIRVLSLEDGIATGLLVKDLGIQVDRY  
 IASEVCEDSITVGMVRHQGKIMYVG D VRSVTQKH IQEWGPEDLVIGGSPC  
 NDLSIVNPARKGLYEGTGRLFFEFYRLLHDARPKEGDDRPFFWLFENVVA  
 MGVSDKRDISRFLSNPVMIDAKEVSAAHRARYFWGNLPGMNRPLASTVN  
 DKLELQECLEHGRIAKFSKVRTITTRSNSIKQGKDQHFPVFMNEKEDILW  
 CTEMERVEGFVHYTDVSNMSRLARQRLGRSWSPVIRHLFAPLKEYFA  
 CVSSGNSNANSRGPSESSGLVPLSLRGSHMGPM E IYKTVSAWKRPVRVL  
 SLERNIDKVLKSLGFLESGSGSGGGTLKYVEDVTNVVRRDVEKWGPEDLV  
 YGSTQPLGSSCDRCPGWYMFQFHRILQYALPRQESQRPFFWIEMDNLLLT  
 EDDQETTRELQTEAVTLQDVRGRDYQ NAMRVWSNIPGLKSKHAPLTPKE  
 EEYLQAQVRSRSLDAPKVDLLVKNCLLPLREYFKYFSQNSLPLGGPSSG  
 APPPSGGSPAGSPTSTEEGTSESATPESGPGTSTEPSEGSAPGSPAGSPT  
 STEEGTSTEPSEGSAPGTSTEPSEPKKKRKVYSRPGERPFQCRICMRNFS  
 XXXXXXXHXXTHTGEKPFQCRICMRNFSXXXXXXXXHXXTH[linker]PF  
 QCRICMRNFSXXXXXXXXHXXTHTGEKPFQCRICMRNFSXXXXXXXXHXXTH  
 [linker]PFQCRICMRNFSXXXXXXXXHXXTHTGEKPFQCRICMRNFSXX  
 XXXXXHXXTHLRGSPKKRKVSGSETPGTSESATPESTGRTLVTFKDVFV  
 DFTREEWKLLDTAQQIVYRNVMLENYKNLVSLGYQLTKPDVILRLEKGEE P (SEQ

ID NOs: 1081, 1262 and 1263, respectively, in order of appearance)

(184) In certain embodiments, the six “XXXXXXX” regions in SEQ ID NO: 1081, 1262 or 1263 comprise, in order, the F1-F6 amino acid sequences shown in Table 1. [linker] represents a linker sequence. In some embodiments, one or both linker sequences may be TGSQKP (SEQ ID NO: 1085). In some embodiments, one or both linker sequences may be TGGGGSQKP (SEQ ID NO: 1086). In some embodiments, one linker sequence may have the amino acid sequence of SEQ ID NO: 1085 and the other linker sequence may have the amino acid sequence of SEQ ID NO: 1086.

(185) Multiple epigenetic editors may be used to effect activation or repression of a target gene or multiple target genes. For example, an epigenetic editor fusion protein comprising a DNA-binding domain (e.g., a dCas9 domain) and an effector domain may be co-delivered with two or more guide polynucleotides (e.g., gRNAs), each targeting a different target DNA sequence. The target sites for two of the DNA-binding domains may be the same or in the vicinity of each other, or separated by, for example, about 100 base pairs, about 200 base pairs, about 300 base pairs, about 400 base pairs, about 500 base pairs, or about 600 or more base pairs. In addition, when targeting double-strand DNA, such as an endogenous gene locus, the guide polynucleotides may target the same or different strands (one or more to the positive strand and/or one or more to the negative strand).

## V. Target Sequences

(186) An epigenetic editor herein may be directed to an HBV target sequence to effect epigenetic modification of HBV or an HBV gene. As used herein, a “target sequence,” a “target site,” or a “target region” is a nucleic acid sequence present in a genome or gene of interest, e.g., in an HBV genome or an HBV gene; in some instances, the target sequence may be outside but in the vicinity of the gene of interest wherein methylation or binding by a repressor of the target sequence represses expression of the gene. In some embodiments, the target sequence may be a hypomethylated or hypermethylated nucleic acid sequence.

(187) The structure and biology of HBV as well as HBV-associated diseases have been reported (see, for example, Yuen, MF., Chen, DS., Dusheiko, G. et al. Hepatitis B virus infection. *Nat Rev Dis Primers* 4, 18035 (2018); R. Koshy and W. H. Caselman (Eds.), *Hepatitis B Virus: Molecular Mechanism in Disease and Novel Strategies for Antiviral Therapy*, Imperial College Press, London (1998), ISBN 1783262737; the entire contents of each of which are incorporated herein by reference). HBV genotypes and sub-types, as well as their genomic, transcript, and protein sequences have been described and are known to the skilled artisan. Some exemplary HBV sequences, e.g., those under accession numbers NC\_00397 and U95551 are provided elsewhere herein, and the entire content of each such database entry is incorporated herein by reference.

(188) Without wishing to be bound by any particular theory, it has been reported that HBV persists as a covalently closed circular DNA (cccDNA) of approximately 3.2 kb, as well as in an integrated form. The HBV genome has been extensively characterized. The HBV genome has been shown to comprise four genes (the S gene, the P gene, the C gene, and the X gene), regulated by four promoter elements (sp1, sp2, cp and xp) and two enhancer elements (Enh I and Enh II) that control the expression of four defined (and overlapping) protein-encoding open reading frames (S, C, X, and P). See FIG. 1. The HBV genome has been described to express six major viral RNA transcripts encoding the viral proteins: (1) the preCore (preC) RNA, which encodes the C protein (also referred to as Core protein, HBe Antigen, or HBeAg); (2), the pre-genomic (pg)RNA, which encodes the two viral proteins C (core) and P (polymerase), and also serves as the template for the synthesis of viral DNA, which is mediated by the reverse transcriptase activity of the viral P protein once pg RNA and the P protein are encapsidated into the nucleocapsids formed by the C protein; (3) the large surface protein (preS1) RNA, which encodes the Large S Antigen (also referred to as L-HBsAg); (4) the middle surface protein (preS2) RNA, which encodes the Middle S Antigen (also referred to as M-HBsAg); (5) the small surface protein (S) RNA, which encodes the Small S Antigen (also referred to as S-HBsAg); and (6) the X protein (HBx) RNA, which encodes the X protein. Transcription start sites (TSSs) as well as the termination site of the HBV transcripts have been mapped in various HBV genotypes and sub-types. Notably, HBV transcripts have been described to terminate at a single termination/polyadenylation signal located downstream of the Hbx CDS and comprising a canonical ATAAA motif. It has further been reported that HBV DNA may be methylated by infected cells and such methylation has been postulated to correlate with inhibition of viral gene expression. However, naturally occurring cell-mediated methylation of viral DNA is typically insufficient to silence viral expression to a level that would result in control of HBV infection. DNA methylation typically occurs at CpG dinucleotides. Several CpG-rich genomic regions, also referred to as CpG islands or CGIs, have been identified in the HBV genome. CGIs are typically identified in HBV genomic sequences as sequences of a specific minimal length (e.g., at least 100 bp) that comprise a minimum percentage of G and C nucleotides (e.g., at least 50% or at least 60% GC content) and a ratio of observed vs. expected CpG dinucleotides of at least 0.6. CGIs satisfying these criteria have been identified in all HBV genotypes, and it has been demonstrated that HBV genomes typically contain three CpG islands (CGI-I, CGI-II, and CGI-III, respectively), which are also sometimes referred to as ‘conventional’ HBV CpG islands. Some HBV genotypes or sub-types have been reported to comprise additional, ‘non-conventional’ CGIs. FIG. 1 is a diagram illustrating an exemplary structure of a circular HBV genome (the underlying sequence of which is provided herein as SEQ ID NO: 1082), identifying the coding regions of HBV genes and CpG islands CGI-I-III. See, for example, M. J. Kosovsky, et al., *The regulation of hepatitis B virus gene expression: an overview of the cis- and trans-acting components* in R. Koshy and W. H. Caselman (Eds.), *Hepatitis B Virus: Molecular Mechanism in Disease and Novel Strategies for Antiviral Therapy*, Imperial College Press, London (1998), ISBN 1783262737; Miller et al

Compact organization of the hepatitis B virus genome. *Hepatology*. 1989 February; 9(2):322-7; Stadelmayer et al., Full-length 5'RACE identifies all major HBV transcripts in HBV-infected hepatocytes and patient serum. *J Hepatol*. 2020 July; 73(1):40-51; Meier-Stephenson et al., Comprehensive Analysis of Hepatitis B Virus Promoter Region Mutations. *Viruses*. 2018 Nov. 1; 10(11):603; Vivekanandan et al., Hepatitis B viral DNA is methylated in liver tissues. *J Viral Hepat*. 2008, 15(2):103-7; Chen et al., Detection of hepatitis B virus DNA in hepatocellular carcinoma: methylation of integrated viral DNA. *J Virol Methods*. 1988, 19(3-4):257-63; Zhang et al., Comparative Analysis of CpG Islands among HBV Genotypes. *PLOS ONE* 2013, 8(2):e56711; Jain et al., Comprehensive DNA methylation analysis of hepatitis B virus genome in infected liver tissues. *Sci Rep* 5, 10478 (2015); Low et al., Hepatitis B virus DNA methylation and its potential role in chronic hepatitis B. *Expert Reviews in Molecular Medicine*. 2023; 25:ell; Hou et al., CpG islands of hepatitis B virus genome isolated from Chinese patients. *Gene* (2015) 561:261-267; Mouzannar et al., The Post-Transcriptional Regulatory Element of Hepatitis B Virus: From Discovery to Therapy. *Viruses*. 2024 Mar. 29; 16(4):528; Peng et al., Nonproductive Hepatitis B Virus Covalently Closed Circular DNA Generates HBx-Related Transcripts from the HBx/Enhancer I Region and Acquires Reactivation by Superinfection in Single Cells. *J Virol*. 2023 Jan. 31; 97(1):e0171722; Altinel et al., Single-Nucleotide Resolution Mapping of Hepatitis B Virus Promoters in Infected Human Livers and Hepatocellular Carcinoma. *J Virol*. 2016 Nov. 14; 90(23):10811-10822; the entire contents of each of which, and, where applicable, including any supplemental information, are incorporated herein by reference.

(189) The target sequence (also referred to herein as target site or target region) of an epigenetic editor provided herein may be any suitable HBV sequence.

(190) The target sequence may be in any part of a target gene. In some embodiments, the target sequence is part of or near a noncoding sequence of the gene. In some embodiments, the target sequence is part of an exon of the gene. In some embodiments, the target sequence is part of or near a transcriptional regulatory sequence of the gene, such as a promoter or an enhancer. In some embodiments, the target sequence is adjacent to, overlaps with, or encompasses a CpG island, e.g., a CpG island identified within the HBV genome. In some embodiments, the target sequence is outside of a CpG island. In certain embodiments, the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS. In certain embodiments, the target sequence is within 500 bp flanking the HBV TSS. In certain embodiments, the target sequence is within 1000 bp flanking the HBV TSS.

(191) Some exemplary embodiments in which the target sequence is part of a target gene are provided herein and additional embodiments will be apparent to the skilled artisan based on the present disclosure and the knowledge of the genomic structure of HBV in the art. For example, in some embodiments, the target sequence is part of the HBV S gene, the HBV P gene, the HBV C gene, or the HBV X gene. In some embodiments, the target sequence is part of the HBV S gene. In some embodiments, the target sequence is part of the HBV P gene. In some embodiments, the target sequence is part of the HBV C gene. In some embodiments, the target sequence is part of the HBV X gene. Some exemplary embodiments in which the target sequence is part of a noncoding sequence of a target gene are provided herein and additional embodiments will be apparent to the skilled artisan based on the present disclosure and the knowledge of the genomic structure of HBV in the art. For example, in some embodiments the target sequence is part of a noncoding sequence of the HBV S gene, of the HBV P gene, of the HBV C gene, or of the HBV X gene. For example, in some embodiments, the target sequence is part of a noncoding sequence of the HBV S gene. In some embodiments, the target sequence is part of a noncoding sequence of the HBV P gene. In some embodiments, the target sequence is part of a noncoding sequence of the HBV C gene. In some embodiments, the target sequence is part of a noncoding sequence of the HBV X gene. Noncoding sequences of the various HBV genes are known in the art and include, for example, the promoter and enhancer sequences of the HBV genome. Accordingly, in some embodiments, the target sequence is part of an HBV promoter sequence (e.g., of a promoter sequence within the HBV genome driving the transcription of one of the HBV transcripts described elsewhere herein, including, for example, of a sequence of the sp1, the sp2, the cp, and the xp

promoter elements). In some embodiments, the target sequence is part of an HBV enhancer sequence (e.g., of the Enh I or of the Enh II sequence).

(192) Some exemplary embodiments, in which the target sequence is adjacent to, overlaps with, or encompasses a CpG island, e.g., a CpG island identified within the HBV genome include embodiments in which the target sequence is adjacent to, overlaps with, or encompasses a conventional CGI of HBV, e.g., CGI I, CGI II, or CGI III. CGIs of HBV have been identified and described in numerous publications and are thus known to the skilled artisan. Bioinformatics tools for the identification of CGIs in any specific HBV sequence, e.g., in a sequence of a specific HBV genotype or sub-type, or in an HBV sequence isolated from a patient, are known in the art, including, for example, EMBOSS CpG plot (EMBL-EBI) and Methprimer (Li LC and Dahiya R. MethPrimer: designing primers for methylation PCRs. Bioinformatics. 2002 November; 18(11):1427-31). Conventional CGIs of HBV include CGI I, which overlaps the S and the P gene ORFs; CGI-II, which overlaps the P gene and X gene ORFs; and CGI III, which overlaps the C gene and P gene ORFs (see FIG. 1). In some embodiments, an HBV CGI is identified as a sequence within the HBV genome that is (1) at least 100 nucleotides long; (2) is characterized by a GC content of at least 50%; and (3) is characterized by an observed-to-expected CpG dinucleotide ratio of at least 0.6. According to these criteria, in the exemplary HBV genome referenced in FIG. 1, i.e., NC\_003977 (provided herein as SEQ ID NO: 1082), CGI I spans nucleotides 186-288, CGI II spans nucleotides 1,217-1,670, and CGI III spans nucleotides 2,282-2,448 (see FIG. 1). CGIs of HBV fulfilling these criteria, including conventional HBV CGIs I-III, of other HBV sequences, including other genotypes, sub-types, or specific HBV sequences, will be apparent to the skilled artisan. In some embodiments, the target sequence overlaps with HBV CGI I. In some embodiments, the target sequence overlaps with HBV CGI II. In some embodiments, the target sequence overlaps with CGI III.

(193) Exemplary embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS (transcription start site) include embodiments, in which the target sequence is within the respective number of base pairs of the TSS of any of the six major viral RNA transcripts, i.e., the TSS of the preCore (pre-C) RNA, the TSS of the pre-genomic (pg)RNA, the TSS of the large surface protein (preS1) RNA, the TSS of the middle surface protein (preS2) RNA, the TSS of the small surface protein (S) RNA, and the TSS of the X protein (HBx) RNA. The positions of the transcription start sites of the various HBV transcripts have been identified in various HBV genotypes and sub-types and are thus known to the skilled artisan. For example, for HBV of genotype D, as exemplified by NCBI database entries NC\_003977 and U95551.1 (provided as SEQ ID NOs 1082 and 1083 herein), the TSS of the pg RNA transcript has been identified as nucleotide 1820, the TSS of the pre-C RNA as nucleotide 1791, and the TSS of the pre-S2 RNA as nucleotide 3159. The initiation of HBx RNA transcripts encoded by HBV genomes has been reported to not be limited to a single nucleotide, but to be spread over a short sequence. For example, TSSs for canonical HBx transcripts have been reported to initiate closely upstream of the first ATG in the sequence encoding the X protein, with HBx transcript TSS positions having been mapped to nucleotides 1243-1338 of HBV of genotype D, as exemplified by NCBI database entries NC\_003977 and U95551.1 (provided as SEQ ID NOs 1082 and 1083 herein). TSSs for additional transcripts have also been identified and TSSs have been mapped to various HBV genotypes and sub-types.

(194) In some embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS, the HBV TSS is an HBV pg RNA TSS. For example, in some embodiments provided herein, the target sequence of an epigenetic editor is within 100 bp flanking an HBV pg RNA TSS, e.g., within 100 bp of nucleotide 1820 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 200 bp flanking an HBV pg RNA TSS, e.g., within 200 bp of nucleotide 1820 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 300 bp flanking an HBV pg RNA TSS, e.g., within 300 bp of nucleotide 1820 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 400 bp flanking an HBV pg RNA TSS, e.g., within 400 bp of nucleotide 1820

of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 500 bp flanking an HBV pg RNA TSS, e.g., within 500 bp of nucleotide 1820 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 600 bp flanking an HBV pg RNA TSS, e.g., within 600 bp of nucleotide 1820 of SEQ ID NO: 1082 or 1083.

(195) In some embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS, the HBV TSS is an HBV preCore (preC) RNA TSS. For example, in some embodiments provided herein, the target sequence of an epigenetic editor is within 100 bp flanking an HBV preC RNA TSS, e.g., within 100 bp of nucleotide 1791 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 200 bp flanking an HBV preC RNA TSS, e.g., within 200 bp of nucleotide 1791 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 300 bp flanking an HBV preC RNA TSS, e.g., within 300 bp of nucleotide 1791 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 400 bp flanking an HBV preC RNA TSS, e.g., within 400 bp of nucleotide 1791 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 500 bp flanking an HBV preC RNA TSS, e.g., within 500 bp of nucleotide 1791 of SEQ ID NO: 1082 or 1083.

(196) In some embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS, the HBV TSS is an HBV preS2 RNA TSS. For example, in some embodiments provided herein, the target sequence of an epigenetic editor is within 100 bp flanking an HBV preS2 RNA TSS, e.g., within 100 bp of nucleotide 3159 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 200 bp flanking an HBV preS2 RNA TSS, e.g., within 200 bp of nucleotide 3159 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 300 bp flanking an HBV preS2 RNA TSS, e.g., within 300 bp of nucleotide 3159 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 400 bp flanking an HBV preS2 RNA TSS, e.g., within 400 bp of nucleotide 3159 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 500 bp flanking an HBV preS2 RNA TSS, e.g., within 500 bp of nucleotide 3159 of SEQ ID NO: 1082 or 1083.

(197) In some embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS, the HBV TSS is an HBV HBx RNA TSS. For example, in some embodiments provided herein, the target sequence of an epigenetic editor is within 100 bp flanking an HBV HBx RNA TSS, e.g., within 100 bp of nucleotide 1243 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 200 bp flanking an HBV HBx RNA TSS, e.g., within 200 bp of nucleotide 1243 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 300 bp flanking an HBV HBx RNA TSS, e.g., within 300 bp of nucleotide 1243 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 400 bp flanking an HBV HBx RNA TSS, e.g., within 400 bp of nucleotide 1243 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 500 bp flanking an HBV HBx RNA TSS, e.g., within 500 bp of nucleotide 1243 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 600 bp flanking an HBV HBx RNA TSS, e.g., within 600 bp of nucleotide 1243 of SEQ ID NO: 1082 or 1083.

(198) In some embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS, the HBV TSS is an

HBV HBx RNA TSS. For example, in some embodiments provided herein, the target sequence of an epigenetic editor is within 100 bp flanking an HBV HBx RNA TSS, e.g., within 100 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 200 bp flanking an HBV HBx RNA TSS, e.g., within 200 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 300 bp flanking an HBV HBx RNA TSS, e.g., within 300 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 400 bp flanking an HBV HBx RNA TSS, e.g., within 400 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 500 bp flanking an HBV HBx RNA TSS, e.g., within 500 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 600 bp flanking an HBV HBx RNA TSS, e.g., within 600 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083.

(199) In some embodiments in which the target sequence is within about 3000, 2900, 2800, 2700, 2600, 2500, 2400, 2300, 2200, 2100, 2000, 1900, 1800, 1700, 1600, 1500, 1400, 1300, 1200, 1100, 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) flanking an HBV TSS, the HBV TSS is an HBV HBx RNA TSS. For example, in some embodiments provided herein, the target sequence of an epigenetic editor is within 100 bp flanking an HBV HBx RNA TSS, e.g., within 100 bp of nucleotide 1243 and within 100 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 200 bp flanking an HBV HBx RNA TSS, e.g., within 200 bp of nucleotide 1243 and within 200 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 300 bp flanking an HBV HBx RNA TSS, e.g., within 300 bp of nucleotide 1243 and within 300 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 400 bp flanking an HBV HBx RNA TSS, e.g., within 400 bp of nucleotide 1243 and within 400 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 500 bp flanking an HBV HBx RNA TSS, e.g., within 500 bp of nucleotide 1243 and within 500 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083. In some embodiments provided herein, the target sequence of an epigenetic editor is within 600 bp flanking an HBV HBx RNA TSS, e.g., within 600 bp of nucleotide 1243 and within 600 bp of nucleotide 1338 of SEQ ID NO: 1082 or 1083.

(200) In some embodiments, the target sequence may hybridize to a guide polynucleotide sequence (e.g., gRNA) complexed with a fusion protein comprising a polynucleotide guided DNA-binding domain (e.g., a CRISPR protein such as dCas9) and effector domain(s). The guide polynucleotide sequence may be designed to have complementarity to the target sequence, or identity to the opposing strand of the target sequence. In some embodiments, the guide polynucleotide comprises a spacer sequence that is about 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% identical to a protospacer sequence in the target sequence. In particular embodiments, the guide polynucleotide comprises a spacer sequence that is 100% identical to a protospacer sequence in the target sequence.

(201) In some embodiments, where the DNA-binding domain of an epigenetic editor described herein is a zinc finger array, the target sequence may be recognized by said zinc finger array.

(202) In some embodiments, where the DNA-binding domain of an epigenetic editor described herein is a TALE, the target sequence may be recognized by said TALE.

(203) A target sequence described herein may be specific to one genotype of HBV, to one copy of an HBV target gene, or may be specific to one allele of an HBV target gene. In some embodiments, however, the target sequence may be conserved across two or more HBV genotypes, across two or more copies of an HBV gene, and across alleles of an HBV gene. Accordingly, the epigenetic modification and modulation of expression thereof may be specific to one copy or one allele of the target gene, or, in other embodiments, may be universal to different HBV genotypes, or HBV gene copies or alleles.

(204) In some embodiments, the target sequence is comprised in the following sequence:

(205) TABLE-US-00011 >NC\_003977.2 Hepatitis B virus (strain ayw) genome (SEQ ID No. 1082) AATTCCACAACCTTCCACCAAACCTCTGCAAGATCCCAGAGTGAGAGGCCT



GTATTTCCTTCCCTGCTGGTGGGCTCCAGTTTCAGGAACAGTAACACCCTGTTCTGA  
CTACTGCCTCTCCCTTATCGTCAATCTTCTCGAGGATTGGGGACCCTGCG  
CTGAACATGGAGAACATCACATCAGGATTCCTAGGACCCCTTCTCGTGTT  
ACAGGCGGGGTTTTTCTTGTTGACAAGAATCCTCACAATACCGCAGAGTC  
TAGACTCGTGGTGGACTTCTCTCAATTTTCTAGGGGGAACCTACCGTGTGT  
CTTGGCCAAAATTCGCAGTCCCCAACCTCCAATCACTCACCAACCTCTTG  
TCCTCCAACCTTGTCCTGGTTATCGCTGGATGTGTCTGCGGCGTTTTATCA  
TCTTCCTCTTCATCCTGCTGCTATGCCTCATCTTCTTGTTGGTTCTTCTG  
GACTATCAAGGTATGTTGCCCGTTTGTCTCTAATTCAGGATCCTCAAC  
AACCAGCACGGGACCATGCCGGACCTGCATGACTACTGCTCAAGGAACCT  
CTATGTATCCCTCCTGTTGCTGTACCAAACCTTCGGACGGAAATTGCACC  
TGTATTCCCATCCCATCATCCTGGGCTTTCGGAAAATTCCTATGGGAGTG  
GGCCTCAGCCCGTTTCTCCTGGCTCAGTTTACTAGTGCCATTTGTTCACT  
GGTTCGTAGGGCTTTCCTCCCACTGTTTGGCTTTCAGTTATATGGATGATG  
TGGTATTGGGGGCCAAGTCTGTACAGCATCTTGAGTCCCTTTTTACCGCT  
GTTACCAATTTTCTTTTGTCTTTGGGTATACATTTAAACCCTAACAAAAC  
AAAGAGATGGGGTTACTCTCTAAATTTTATGGGTATGTCATTGGATGTT  
ATGGGTCCTTGCCACAAGAACACATCATACAAAAAATCAAAGAATGTTTT  
AGAAAACCTTCCTATTAACAGGCCTATTGATTGGAAAGTATGTCAACGAAT  
TGTGGGTCTTTTGGGTTTTGCTGCCCTTTTACACAATGTGGTTATCCTG  
CGTTGATGCCTTTGTATGCATGTATTCAATCTAAGCAGGCTTTCACCTTC  
TCGCCAACTTACAAGGCCTTCTGTGTAAACAATACCTGAACCTTTACCC  
CGTTGCCCGGCAACGGCCAGGTCTGTGCCAAGTGTTTGCTGACGCAACCC  
CCACTGGCTGGGGCTTGGTCATGGGCCATCAGCGCATGCGTGGAACCTTT  
TCGGCTCCTCTGCCGATCCATACTGCGGAACCTCCTAGCCGCTTGTTTTGC  
TCGCAGCAGGTCTGGAGCAAACATTATCGGGACTGATAACTCTGTTGTCC  
TATCCCGCAAATATACATCGTTTCCATGGCTGCTAGGCTGTGCTGCCAAC  
TGGATCCTGCGCGGGACGTCCTTTGTTTACGTCCCGTCGGCGCTGAATCC  
TGC GGACGACCCCTTCTCGGGGTGCTTGGGACTCTCTCGTCCCTTCTCC  
GTCTGCCGTTCCGACCGACACGGGGCGCACCTCTCTTTACGCGGACTCC  
CCGTCTGTGCCTTCTCATCTGCCGGACCGTGTGCACTTCGCTTCACCTCT  
GCACGTCGCATGGAGACCACCGTGAACGCCCAACCAATATTGCCCAAGGT  
CTTACATAAGAGGACTCTTGGACTCTCAGCAATGTCAACGACCGACCTTG  
AGGCATACTTCAAAGACTGTTTGTTTAAAGACTGGGAGGAGTTGGGGGAG  
GAGATTAGGTAAAGGTCTTTGTACTAGGAGGCTGTAGGCATAAATTGGT  
CTGCGCACCAGCACCATGCAACTTTTTACCTCTGCCTAATCATCTCTTG  
TTCATGTCCTACTGTTCAAGCCTCCAAGCTGTGCCTTGGGTGGCTTTGGG  
GCATGGACATCGACCCTTATAAAGAATTTGGAGCTACTGTGGAGTTACTC  
TCGTTTTTGCCTTCTGACTTCTTTCCTTCAGTACGAGATCTTCTAGATAC  
CGCTCAGCTCTGTATCGGGAAGCCTTAGAGTCTCCTGAGCATTGTTTAC  
CTCACCTACTGCACTCAGGCAAGCAATTCTTTGCTGGGGGGAACCTAATG  
ACTCTAGCTACCTGGGTGGGTGTTAATTTGGAAGATCCAGCGTCTAGAGA  
CCTAGTAGTCAGTTATGTCAACACTAATATGGGCCTAAAGTTCAGGCAAC  
TCTTGTTGGTTTACATTTCTTGTTCTCACTTTTGGAAAGAGAAACAGTTATA  
GAGTATTTGGTGTCTTTCGGAGTGTGGATTGCGCACTCCTCCAGCTTATAG  
ACCACCAAATGCCCTATCCTATCAACACTTCCGGAGACTACTGTTGTTA  
GACGACGAGGCAGGTCCCCTAGAAGAAGAACTCCCTCGCTCGCAGACGA  
AGGTCTCAATCGCCGCGTCGCAGAAGATCTCAATCTCGGGAATCTCAATG  
TTAGTATTCCTTGGACTCATAAGGTGGGGAACTTTACTGGGCTTTATTCT  
TCTACTGTACCTGTCTTTAATCCTCATTGGAAAACACCATCTTTTCCTAA  
TATACATTTACACCAAGACATTATCAAAAAATGTGAACAGTTTGTAGGCC  
CACTCACAGTTAATGAGAAAAGAAGATTGCAATTGATTATGCCTGCCAGG

TTTATTCAACGATTAATTAACCAATTTGATTAAGGTTATTAAACC  
TTATTATCCAGAACATCTAGTTAATCATTACTTCCAAACTAGACACTATT  
TACACACTCTATGGAAGGCGGGTATATTATATAAGAGAGAAACAACACAT  
AGCGCCTCATTTTGTGGGTCACCATATTCTTGGGAACAAGATCTACAGCA  
TGGGGCAGAATCTTTCCACCAGCAATCCTCTGGGATTCTTTCCCGACCAC  
CAGTTGGATCCAGCCTTCAGAGCAAACACCGCAAATCCAGATTGGGACTT  
CAATCCCAACAAGGACACCTGGCCAGACGCCAACAAGGTAGGAGCTGGAG  
CATTCGGGCTGGGTTTCACCCCCACCGCACGGAGGCCTTTTGGGGTGGAGC  
CCTCAGGCTCAGGGCATACTACAAACTTTGCCAGCAAATCCGCCTCCTGC  
CTCCACCAATCGCCAGTCAGGAAGGCAGCCTACCCCGCTGTCTCCACCTT  
TGAGAAACACTCATCCTCAGGCCATGCAGTGG

FIG. 1 provides a diagram illustrating the structure of a circular HBV genome comprising SEQ ID NO: 1082. The coding regions of the HBV genes and CpG islands CGI-I-III are identified. Nucleotides 2309-1625 of SEQ ID NO: 1082 encode the P protein (NCBI reference number YP\_009173866.1).

Nucleotides 2850-837 of SEQ ID NO: 1082 encode the long surface protein (L-HBsAg or LHBS; NCBI reference number YP\_009173869.1). Nucleotides 3174-837 of SEQ ID NO: 1082 encode the middle surface protein (M-HBsAg or MHBS; NCBI reference number YP\_009173870.1). Nucleotides 157-837 of SEQ ID NO: 1082 encode the small surface protein (S-HBsAg or SHBs; NCBI reference number YP\_009173871.1). Nucleotides 1816-2454 of SEQ ID NO: 1082 encode the C Protein (core protein, NCBI reference number AAB59971.1). Nucleotides 1376-1840 of SEQ ID NO: 1082 encode the X protein (HBx, NCBI reference number YP\_009173867.1). CGI I spans nucleotides 186-288, CGI II spans nucleotides 1,217-1,670, and CGI III spans nucleotides 2,282-2,448. See, NCBI database entry NC 003977.2. TSSs of various transcripts have been mapped: pg RNA TSS: 1820; pre-C RNA TSS: 1791; pre-S2 RNA TSS: 3159; HBx RNA TSSs: 1243-1338. The ATAAA motif of the transcription termination/polyadenylation site is located at nucleotide 1919. See references cited elsewhere herein. See also, e.g., Abraham, T. M. and Loeb, D. D., The topology of hepatitis B virus pregenomic RNA promotes its replication, *J. Virol.* 81 (21), 11577-11584 (2007); Chen, A., Kao, Y. F. and Brown, C. M., Translation of the first upstream ORF in the hepatitis B virus pregenomic RNA modulates translation at the core and polymerase initiation codons, *Nucleic Acids Res.* 33 (4), 1169-1181 (2005); Borisova, G. P., Pumpen, P. P., Bychko, V. V., Pushko, P. M., Kalis, Y. V., Dishler, A. V., Gren, E. Y., Tsibinogin, V. V. and Kukain, R. A., Structure and expression of the gene of the core antigen of human hepatitis B virus (HBV) in *Escherichia coli* cells, *Dokl. Biochem.* 279, 386-390 (1985); Galibert, F., Mandart, E., Fitoussi, F., Tiollais, P. and Chamay, P., Nucleotide sequence of the hepatitis B virus genome (subtype ayw) cloned in *E. coli*, *Nature* 281 (5733), 646-650 (1979), the entire contents of each of which are incorporated herein by reference.

(206) In some embodiments, the target sequence is comprised in the following sequence:

(207) TABLE-US-00012 >U95551.1 Hepatitis B virus subtype ayw, complete genome (SEQ ID No. 1083)

AATTCCACAACCTTTACCAAACCTCTGCAAGATCCCAGAGTGAGAGGCCT  
GTATTTCCCTGCTGGTGGCTCCAGTTCAGGAGCAGTAAACCCTGTTCCGA  
CTACTGCCTCTCCCTTATCGTCAATCTTCTCGAGGATTGGGGACCCTGCG  
CTGAACATGGAGAACATCACATCAGGATTCCTAGGACCCCTTCTCGTGTT  
ACAGGCGGGGTTTTTCTTGTTGACAAGAATCCTCACAAATACCGCAGAGTC  
TAGACTCGTGGTGGACTTCTCTCAATTTTCTAGGGGGAACCTACCGTGTGT  
CTTGGCCAAAATTCGCAGTCCCCAACCTCCAATCACTACCAACCTCCTG  
TCCTCCAACCTTGTCTGCTGCTATGCCTCATCTTCTTGTTGGTTCTTCTG  
GACTATCAAGGTATGTTGCCCCGTTTGTCTCTAATTCCAGGATCCTCAAC  
CACCAGCACGGGACCATGCCGAACCTGCATGACTACTGCTCAAGGAACCT  
CTATGTATCCCTCCTGTTGCTGTACCAAACCTTCGGACGGAAATTGCACC  
TGTATTCCCATCCCATCATCCTGGGCTTTCGGAAAATTCCTATGGGAGTG  
GGCCTCAGCCCGTTTCTCCTGGCTCAGTTTACTAGTGCCATTTGTTCACT

GGTTCGCTGATGTTCCCTCCCTGTTTCCAGTTATATGGATGAT  
TGGTATTGGGGGCCAAGTCTGTACAGCATCTTGAGTCCCTTTTTACCGCT  
GTTACCAATTTTCTTTTGTCTTTGGGTATACATTTAAACCCTAACAAAAC  
AAAGAGATGGGGTTACTCTCTGAATTTTATGGGTTATGTCATTGGAAGTT  
ATGGGTCCTTGCCACAAGAACACATCATACAAAAAATCAAAGAATGTTTT  
AGAAAACCTTCCTATTAACAGGCCTATTGATTGGAAAGTATGTCAACGAAT  
TGTGGGTCTTTTGGGTTTTGCTGCCCCATTTACACAATGTGGTTATCCTG  
CGTTAATGCCCTTGTATGCATGTATTCAATCTAAGCAGGCTTTCACTTTC  
TCGCCAACTTACAAGGCCTTTCTGTGTAAACAATACTGAACCTTTACCC  
CGTTGCCCCGGCAACGGCCAGGTCTGTGCCAAGTGTTTGCTGACGCAACCC  
CCACTGGCTGGGGCTTGGTCATGGGCCATCAGCGCGTGCGTGGAACCTTT  
TCGGCTCCTCTGCCGATCCATACTGCGGAACCTCCTAGCCGCTTGTTTTGC  
TCGCAGCAGGTCTGGAGCAAACATTATCGGGACTGATAACTCTGTTGTCC  
TCTCCCGCAAATATACATCGTATCCATGGCTGCTAGGCTGTGCTGCCAAC  
TGGATCCTGCGCGGGACGTCCTTTGTTTACGTCCCGTCGGCGCTGAATCC  
TGCGGACGACCCTTCTCGGGGTGCTTGGGACTCTCTCGTCCCCTTCTCC  
GTCTGCCGTTCCGACCGACCACGGGGCGCACCTCTCTTTACGCGGACTCC  
CCGTCTGTGCCTTCTCATCTGCCGGACCGTGTGCACTTCGCTTCACCTCT  
GCACGTCGCATGGAGACCACCGTGAACGCCACCGAATGTTGCCCAAGGT  
CTTACATAAGAGGACTCTTGGACTCTCTGCAATGTCAACGACCGACCTTG  
AGGCATACTTCAAAGACTGTTTGTAAAGACTGGGAGGAGTTGGGGGAG  
GAGATTAGATTAAAGGTCTTTGTACTAGGAGGCTGTAGGCATAAATTGGT  
CTGCGCACCAGCACCATGCAACTTTTTACCTCTGCCTAATCATCTCTTG  
TTCATGTCTACTGTTCAAGCCTCCAAGCTGTGCCTTGGGTGGCTTTGGG  
GCATGGACATCGACCCTTATAAAGAATTTGGAGCTACTGTGGAGTTACTC  
TCGTTTTTGCCTTCTGACTTCTTTCCTTCAGTACGAGATCTTCTAGATAC  
CGCCTCAGCTCTGTATCGGGAAGCCTTAGAGTCTCCTGAGCATTGTTTAC  
CTCACCATACTGCACTCAGGCAAGCAATTCTTTGCTGGGGGGAACATAATG  
ACTCTAGCTACCTGGGTGGGTGTTAATTTGGAAGATCCAGCATCTAGAGA  
CCTAGTAGTCAGTTATGTCAACACTAATATGGGCCTAAAGTTCAGGCAAC  
TCTTGTGGTTTCACATTTCTTGTCTCACTTTTGGAAGAGAAACCGTTATA  
GAGTATTTGGTGTCTTTCGGAGTGTGGATTTCGCACTCCTCCAGCTTATAG  
ACCACCAAATGCCCTATCCTATCAACACTTCCGGAAACTACTGTTGTTA  
GACGACGAGGCAGGTCCCCTAGAAGAAGAACTCCCTCGCCTCGCAGACGA  
AGGTCTCAATCGCCGCGTCGCAGAAGATCTCAATCTCGGGAACCTCAATG  
TTAGTATTCCTTGGACTCATAAGGTGGGGAACTTTACTGGTCTTTATTCT  
TCTACTGTACCTGTCTTTAATCCTCATTGGAACACCATCTTTTCCTAA  
TATACATTTACACCAAGACATTATCAAAAAATGTGAACAGTTTGTAGGCC  
CACTTACAGTTAATGAGAAAAGAAGATTGCAATTGATTATGCCTGCTAGG  
TTTTATCCAAAGGTTACCAAATATTTACCATTGGATAAGGGTATTAAACC  
TTATTATCCAGAACATCTAGTTAATCATTACTTCCAAACTAGACACTATT  
TACACACTCTATGGAAGGCGGGTATATTATATAAGAGAGAAACAACACAT  
AGCGCCTCATTTTGTGGGTCACCATATTCTTGGGAACAAGATCTACAGCA  
TGGGGCAGAATCTTTCACCAGCAATCCTCTGGGATTCTTTCCCGACCAC  
CAGTTGGATCCAGCCTTCAGAGCAAACACAGCAAATCCAGATTGGGACTT  
CAATCCCAACAAGGACACCTGGCCAGACGCCAACAAGGTAGGAGCTGGAG  
CATTCGGGCTGGGTTTCACCCACCGCACGGAGGCCTTTTGGGGTGGAGC  
CCTCAGGCTCAGGGCATACTACAAACTTTGCCAGCAAATCCGCCTCCTGC  
CTCCACCAATCGCCAGACAGGAAGGCAGCCTACCCCGCTGTCTCCACCTT  
TGAGAAACACTCATCCTCAGGCCATGCAGTGG.

(208) Annotation of SEQ ID NO: 1083: P protein CDS: 2309-1625; L-HBsAG CDS: 2850-837; M-HBsAg CDS: 3174-837; S-HBsAg CDS: 157-837; C Protein CDS: 1816-2454; X protein CDS: 1376-

186; CGI II: 1,217-1,670; CGI III: 2,282-2,448; pg RNA TSS: 1820; pre-C RNA TSS: 1791; pre-S2 RNA TSS: 3159; HBx RNA TSSs: 1243-1338; termination/polyA site: 1919. See references cited elsewhere herein.

## VI. Epigenetic Modifications

(209) An epigenetic editor described herein may perform sequence-specific epigenetic modification(s) (e.g., alteration of chemical modification(s)) of a target gene that harbors the target sequence. Such epigenetic modulation may be safer and more easily reversible than modulation due to gene editing, e.g., with generation of DNA double-strand breaks. In some embodiments, the epigenetic modulation may reduce or silence the target gene. In some embodiments, the modification is at a specific site of the target sequence. In some embodiments, the modification is at a specific allele of the target gene. Accordingly, the epigenetic modification may result in modulated (e.g., reduced) expression of one copy of a target gene harboring a specific allele, and not the other copy of the target gene. In some embodiments, the specific allele is associated with a disease, condition, or disorder.

(210) In some embodiments, the epigenetic modification reduces or abolishes transcription of the target gene harboring the target sequence. In some embodiments, the epigenetic modification reduces or abolishes transcription of a copy of the target gene harboring a specific allele recognized by the epigenetic editor. In some embodiments, the epigenetic editor reduces the level of or eliminates expression of a protein encoded by the target gene. In some embodiments, the epigenetic editor reduces the level of or eliminates expression of a protein encoded by a copy of the target gene harboring a specific allele recognized by the epigenetic editor. The target HBV gene may be epigenetically modified *in vitro*, *ex vivo*, or *in vivo*.

(211) The effector domain of an epigenetic editor described herein may alter (e.g., deposit or remove) a chemical modification at a nucleotide of the target gene or at a histone associated with the target gene. The chemical modification may be altered at a single nucleotide or a single histone, or may be altered at 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1500, 2000, 2500, 3000 or more nucleotides.

(212) In some embodiments, an effector domain of an epigenetic editor described herein may alter a CpG dinucleotide within the target gene. In some embodiments, all CpG dinucleotides within 2000, 1500, 1000, 500, or 200 bps flanking a target sequence (e.g., in an alteration site as described herein) are altered according to a modification type described herein, as compared to the original state of the gene or the gene in a comparable cell not contacted with the epigenetic editor. In some embodiments, at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700 or more of the CpG dinucleotides are altered as compared to the original state of the gene or the gene in a comparable cell not contacted with the epigenetic editor. In some embodiments, at least 5%, 6%, 7%, 8%, 9%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% of the CpG dinucleotides are altered as compared to the original state of the gene or the gene in a comparable cell not contacted with the epigenetic editor. In some embodiments, one single CpG dinucleotide is altered, as compared to the original state of the gene or the gene in a comparable cell not contacted with the epigenetic editor.

(213) An effector domain of an epigenetic editor described herein may alter a histone modification state of a histone associated with or bound to the target gene. For example, an effector domain may deposit a modification on one or more lysine residues of histone tails of histones associated with the target gene. In some embodiments, the effector domain may result in deacetylation of one or more histone tails of histones associated with the target gene, thereby reducing or silencing expression of the target gene. In some embodiments, the histone modification state is a methylation state. For example, the effector domain may result in a H3K9, H3K27 or H4K20 methylation (e.g. one or more of a H3K9me2, H3K9me3, H3K27me2, H3K27me3, and H4K20me3 methylation) at one or more histone tails associated with the target gene, thereby reducing or silencing expression of the target gene.

(214) In some embodiments, all histone tails of histones bound to DNA nucleotides within 2000, 1500, 1000, 500, or 200 bps flanking the target sequence are altered according to a modification type as described herein, as compared to the original state of the chromosome or the chromosome in a

comparable cell not contacted with the epigenetic editor. In some embodiments, at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120 or more histone tails of the bound histones are altered as compared to the original state of the chromosome or the chromosome in a comparable cell not contacted with the epigenetic editor. In some embodiments, at least 5%, 6%, 7%, 8%, 9%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% of histone tails of the bound histones are altered as compared to the original state of the chromosome or the chromosome in a comparable cell not contacted with the epigenetic editor. For example, one single histone tail of the bound histones may be altered as compared to the original state of the chromosome or the chromosome in a comparable cell not contacted with the epigenetic editor. As another example, one single bound histone octamer may be altered as compared to the original state of the chromosome or the chromosome in a comparable cell not contacted with the epigenetic editor.

(215) The chemical modification deposited at target gene DNA nucleotides or histone residues may be at or in close proximity to a target sequence in the target gene. In some embodiments, an effector domain of an epigenetic editor described herein alters a chemical modification state of a nucleotide or histone tail bound to a nucleotide 100-200, 200-300, 300-400, 400-55, 500-600, 600-700, or 700-800 nucleotides 5' or 3' to the target sequence in the target gene. In some embodiments, an effector domain alters a chemical modification state of a nucleotide or histone tail bound to a nucleotide within 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, or 2000 nucleotides flanking the target sequence. As used herein, "flanking" refers to nucleotide positions 5' to the 5' end of and 3' to the 3' end of a particular sequence, e.g. a target sequence.

(216) In some embodiments, an effector domain mediates or induces a chemical modification change of a nucleotide or a histone tail bound to a nucleotide distant from a target sequence. Such modification may be initiated near the target sequence, and may subsequently spread to one or more nucleotides in the target gene distant from the target sequence. For example, an effector domain may initiate alteration of a chemical modification state of one or more nucleotides or one or more histone residues bound to one or more nucleotides within 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 300, 400, 500 nucleotides flanking the target sequence, and the chemical modification state alteration may spread to one or more nucleotides at least 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500, 1600, 1700, 1800, 1900, 2000, 2500, 3000, or more nucleotides from the target sequence in the target gene, either upstream or downstream of the target sequence. In certain embodiments, the chemical modification may be initiated at less than 2, 3, 5, 10, 20, 30, 40, 50, or 100 nucleotides in the target gene and spread to at least 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 2000, or more nucleotides in the target gene. In some embodiments, the chemical modification spreads to nucleotides in the entire target gene. Additional proteins or transcription factors, for example, transcription repressors, methyltransferases, or transcription regulation scaffold proteins, may be involved in the spreading of the chemical modification. Alternatively, the epigenetic editor alone may be involved.

(217) In some embodiments, an epigenetic editor described herein reduces expression of a target gene by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, at least about 95%, at least about 99%, or more, as measured by transcription of the target gene in a cell, a tissue, or a subject as compared to a control cell, control tissue, or a control subject (e.g., in the absence of the epigenetic editor). In some embodiments, the epigenetic editors described herein reduces expression of a copy of target gene by at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, at least about 95%, at least about 99%, at least about 99.9%, or more, as measured by transcription of the copy of the target gene in a cell, a tissue, or a subject as compared to a control cell, control tissue, or a control subject. For example, in some embodiments, an epigenetic editor described herein reduces expression of an HBV target gene in vitro or in vivo (e.g., as measured as the level of an HBV biomarker in a subject), by at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, at least 99.9%, or more, as measured for example, by transcription of the target gene, or by

assessing an HBV biomarker (e.g., plasma HBV DNA, plasma HBV sAg, or plasma HBV eAg) in a cell, a tissue, or a subject contacted or administered with the epigenetic editor as compared to a control cell, control tissue, or a control subject (e.g., in the absence of the epigenetic editor). In certain embodiments, the copy of the target gene harbors a specific sequence or allele recognized by the epigenetic editor. In particular embodiments, the epigenetically modified copy encodes a functional protein, and accordingly an epigenetic editor disclosed herein may reduce or abolish expression and/or function of the protein. For example, an epigenetic editor described herein may reduce expression and/or function of a protein encoded by the target gene by at least 3-fold, at least 4-fold, at least 5-fold, at least 6-fold, at least 7-fold, at least 8-fold, at least 9-fold, at least 10-fold, at least 11-fold, at least 12-fold, at least 13-fold, at least 14-fold, at least 15-fold, at least 20-fold, at least 25-fold, at least 30-fold, at least 35-fold, at least 40-fold, at least 45-fold, at least 50-fold, at least 60-fold, at least 70-fold, at least 80-fold, at least 90-fold, or at least 100 fold in a cell, a tissue, or a subject as compared to a control cell, control tissue, or a control subject.

(218) Modulation of target gene expression can be assayed by determining any parameter that is indirectly or directly affected by the expression of the target gene. Such parameters include, e.g., changes in RNA or protein levels; changes in protein activity; changes in product levels; changes in downstream gene expression; changes in transcription or activity of reporter genes such as, for example, luciferase, CAT, beta-galactosidase, or GFP; changes in signal transduction; changes in phosphorylation and dephosphorylation; changes in receptor-ligand interactions; changes in concentrations of second messengers such as, for example, cGMP, cAMP, IP3, and Ca<sup>2+</sup>; changes in cell growth; changes in neovascularization; and/or changes in any functional effect of gene expression. Measurements can be made in vitro, in vivo, and/or ex vivo, and can be made by conventional methods, e.g., measurement of RNA or protein levels, measurement of RNA stability, and/or identification of downstream or reporter gene expression. Readout can be by way of, for example, chemiluminescence, fluorescence, colorimetric reactions, antibody binding, inducible markers, ligand binding assays, changes in intracellular second messengers such as cGMP and inositol triphosphate (IP3), changes in intracellular calcium levels; cytokine release, and the like.

(219) Methods for determining the expression level of a gene, for example the target of an epigenetic editor, may include, e.g., determining the transcript level of a gene by reverse transcription PCR, quantitative RT-PCR, droplet digital PCR (ddPCR), Northern blot, RNA sequencing, DNA sequencing (e.g., sequencing of complementary deoxyribonucleic acid (cDNA) obtained from RNA); next generation (Next-Gen) sequencing, nanopore sequencing, pyrosequencing, or Nanostring sequencing. Levels of protein expressed from a gene may be determined, e.g., by Western blotting, enzyme linked immuno-absorbance assays, mass-spectrometry, immunohistochemistry, or flow cytometry analysis. Gene expression product levels may be normalized to an internal standard such as total messenger ribonucleic acid (mRNA) or the expression level of a particular gene, e.g., a housekeeping gene.

(220) In some embodiments, the effect of an epigenetic editor in modulating target gene expression may be examined using a reporter system. For example, an epigenetic editor may be designed to target a reporter gene encoding a reporter protein, such as a fluorescent protein. Expression of the reporter gene in such a model system may be monitored by, e.g., flow cytometry, fluorescence-activated cell sorting (FACS), or fluorescence microscopy. In some embodiments, a population of cells may be transfected with a vector that harbors a reporter gene. The vector may be constructed such that the reporter gene is expressed when the vector transfects a cell. Suitable reporter genes include genes encoding fluorescent proteins, for example green, yellow, cherry, cyan or orange fluorescent proteins. The population of cells carrying the reporter system may be transfected with DNA, mRNA, or vectors encoding the epigenetic editor targeting the reporter gene.

## VII. Pharmaceutical Compositions

(221) Another aspect of the present disclosure is a pharmaceutical composition comprising as an active ingredient (or as the sole active ingredient) one or more epigenetic editors described herein or component(s) (e.g., fusion proteins and/or guide polynucleotides) thereof, or nucleic acid molecule(s) encoding said epigenetic editors or component(s) thereof. For example, a pharmaceutical composition may comprise nucleic acid molecule(s) encoding the fusion protein(s) (and guide polynucleotides, where

applicable) of an epigenetic editor described herein. In some embodiments, separate pharmaceutical compositions comprise the fusion protein(s) and the guide polynucleotide(s). In some embodiments, multiple pharmaceutical compositions, each comprising one epigenetic editor, are administered simultaneously. A pharmaceutical composition may also comprise cells that have undergone epigenetic modification(s) mediated or induced by an epigenetic editor provided herein.

(222) Generally, the epigenetic editors described herein or component(s) thereof, or nucleic acid molecule(s) encoding said epigenetic editors or component(s) thereof, of the present disclosure are suitable to be administered as a formulation in association with one or more pharmaceutically acceptable excipient(s), e.g., as described below.

(223) The term “excipient” is used herein to describe any ingredient other than the compound(s) of the present disclosure. The choice of excipient(s) will to a large extent depend on factors such as the particular mode of administration, the effect of the excipient on solubility and stability, and the nature of the dosage form. As used herein, “pharmaceutically acceptable excipient” includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Some examples of pharmaceutically acceptable excipients are water, saline, phosphate buffered saline, dextrose, glycerol, ethanol and the like, as well as combinations thereof. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, or sodium chloride in the composition. Additional examples of pharmaceutically acceptable substances are wetting agents or minor amounts of auxiliary substances such as wetting or emulsifying agents, preservatives, or buffers, which enhance the shelf life or effectiveness of the antibody.

(224) Formulations of a pharmaceutical composition suitable for parenteral administration typically comprise the active ingredient combined with a pharmaceutically acceptable carrier, such as sterile water or sterile isotonic saline. Such formulations may be prepared, packaged, or sold in a form suitable for bolus administration or for continuous administration. In some embodiments, the epigenetic editor or its component(s) are introduced to target cells in the form of nucleic acid molecule(s) encoding the epigenetic editor or its component(s); accordingly, the pharmaceutical compositions herein comprise the nucleic acid molecule(s). Such nucleic acid molecule(s) may be, for example, DNA, RNA or mRNA, and/or modified nucleic acid sequence(s) (e.g., with chemical modifications, a 5' cap, or one or more 3' modifications). In some embodiments, the nucleic acid molecule(s) may be delivered as naked DNA or RNA, for instance by means of transfection or electroporation, or can be conjugated to molecules (e.g., N-acetylgalactosamine) promoting uptake by target cells. In some embodiments, the nucleic acid molecule(s) may be in nucleic acid expression vector(s), which may include expression control sequences such as promoters, enhancers, transcription signal sequences, transcription termination sequences, introns, polyadenylation signals, Kozak consensus sequences, internal ribosome entry sites (IRES), etc. Such expression control sequences are well known in the art. A vector may also comprise a sequence encoding a signal peptide (e.g., for nuclear localization, nucleolar localization, or mitochondrial localization), associated with (e.g., inserted into or fused to) a sequence coding for a protein.

(225) Examples of vectors include, but are not limited to, plasmid vectors; viral vectors based on vaccinia virus, poliovirus, adenovirus, adeno-associated virus, SV40, herpes simplex virus, human immunodeficiency virus, retrovirus (e.g., Murine Leukemia Virus, or spleen necrosis virus, vectors derived from retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, a lentivirus, human immunodeficiency virus, myeloproliferative sarcoma virus, and mammary tumor virus); and other recombinant vectors. In certain embodiments, the vector is a plasmid or a viral vector. Viral particles may also be used to deliver nucleic acid molecule(s) encoding epigenetic editors or component(s) thereof as described herein. For example, “empty” viral particles can be assembled to contain any suitable cargo. Viral vectors and viral particles may also be engineered to incorporate targeting ligands to alter target tissue specificity.

(226) In certain embodiments, an epigenetic editor as described herein or component(s) thereof are encoded by nucleic acid sequence(s) present in one or more viral vectors, or a suitable capsid protein of any viral vector. Examples of viral vectors include adeno-associated viral vectors (e.g., derived from

AAV3, AAV3b, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAVrh8, AAV10, and/or variants thereof); retroviral vectors (e.g., Maloney murine leukemia virus, MML-V), adenoviral vectors (e.g., AD100), lentiviral vectors (e.g., HIV and FIV-based vectors), and herpesvirus vectors (e.g., HSV-2).

(227) In some embodiments, delivery involves an adeno-associated virus (AAV) vector. AAV vector delivery may be particularly useful where the DNA-binding domain of an epigenetic editor fusion protein is a zinc finger array. Without wishing to be bound by any theory, the smaller size of zinc finger arrays compared to larger DNA-binding domains such as Cas protein domains may allow such a fusion protein to be conveniently packed in viral vectors such as an AAV vector.

(228) Any AAV serotype, e.g., human AAV serotype, can be used for an AAV vector as described herein, including, but not limited to, AAV serotype 1 (AAV1), AAV serotype 2 (AAV2), AAV serotype 3 (AAV3), AAV serotype 4 (AAV4), AAV serotype 5 (AAV5), AAV serotype 6 (AAV6), AAV serotype 7 (AAV7), AAV serotype 8 (AAV8), AAV serotype 9 (AAV9), AAV serotype 10 (AAV10), and AAV serotype 11 (AAV11), as well as variants thereof. In some embodiments, an AAV variant has at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% amino acid sequence identity to a wildtype AAV. In certain embodiments, the AAV variant may be engineered such that its capsid proteins have reduced immunogenicity or enhanced transduction ability in humans. In some instances, one or more regions of at least two different AAV serotype viruses are shuffled and reassembled to generate a chimeric variant. For example, a chimeric AAV may comprise inverted terminal repeats (ITRs) that are of a heterologous serotype compared to the serotype of the capsid. The resulting chimeric AAV can have a different antigenic reactivity or recognition compared to its parental serotypes. In some embodiments, a chimeric variant of an AAV includes amino acid sequences from 2, 3, 4, 5, or more different AAV serotypes.

(229) Non-viral systems are also contemplated for delivery as described herein. Non-viral systems include, but are not limited to, nucleic acid transfection methods including electroporation, sonoporation, calcium phosphate transfection, microinjection, DNA biolistics, lipid-mediated transfection, transfection through heat shock, compacted DNA-mediated transfection, lipofection, cationic agent-mediated transfection, and transfection with liposomes, immunoliposomes, or cationic facial amphiphiles (CFAs). In certain embodiments, one or more mRNAs encoding epigenetic editor fusion proteins as described herein may be co-electroporated with one or more guide polynucleotides (e.g., gRNAs) as described herein. One important category of non-viral nucleic acid vectors is nanoparticles, which can be organic (e.g., lipid) or inorganic (e.g., gold). For instance, organic (e.g. lipid and/or polymer) nanoparticles can be suitable for use as delivery vehicles in certain embodiments of this disclosure.

(230) In some embodiments, delivery is accomplished using a lipid nanoparticle (LNP). LNP compositions are typically sized on the order of micrometers or smaller and may include a lipid bilayer. In some embodiments, a LNP refers to any particle that has a diameter of less than 1000 nm, 500 nm, 250 nm, 200 nm, 150 nm, 100 nm, 75 nm, 50 nm, or 25 nm. Nanoparticle compositions encompass lipid nanoparticles (LNPs), liposomes (e.g., lipid vesicles), and lipoplexes.

(231) An LNP as described herein may be made from cationic, anionic, or neutral lipids. In some embodiments, an LNP may comprise neutral lipids, such as the fusogenic phospholipid 1,2-Dioleoyl-sn-glycero-3-phosphoethanolamine (DOPE) or the membrane component cholesterol, as helper lipids to enhance transfection activity and nanoparticle stability. In some embodiments, an LNP may comprise hydrophobic lipids, hydrophilic lipids, or both hydrophobic and hydrophilic lipids. Any lipid or combination of lipids that are known in the art can be used to produce an LNP. The lipids may be combined in any molar ratios to produce the LNP. In some embodiments, the LNP is a liver-targeting (e.g., preferentially or specifically targeting the liver) LNP.

(232) LNP formulations and methods of LNP delivery that can be used will be apparent to those skilled in the art based on the present disclosure and the state of the art. Non-limiting exemplary compositions and methods can be found in Shah, R., Eldridge, D., Palombo, E., and Harding, I., *Lipid Nanoparticles: Production, Characterization and Stability*, Springer, 2015, ISBN-13 978-3319107103; Ziegler, S., *Lipid Nanoparticles: Advances in Research and Applications*, Nova Science Pub., Inc, ISBN-13 978-1536186536; Mitchell, M. J., Billingsley, M. M., Haley, R. M. et al. *Engineering precision*



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(234) Other methods of delivery to target cells will be known to those skilled in the art and can be used with the compositions of the present disclosure.

(235) Any type of cell may be targeted for delivery of an epigenetic editor or component(s) thereof as described herein. For example, the cells may be eukaryotic or prokaryotic. In some embodiments, the cells are mammalian (e.g., human) cells. Human cells may include, for example, hepatocytes, biliary epithelial cells (cholangiocytes), stellate cells, Kupffer cells, and liver sinusoidal endothelial cells.

(236) In some embodiments, an epigenetic editor described herein, or component(s) thereof, are delivered to a host cell for transient expression, e.g., via a transient expression vector. Transient expression of the epigenetic editor or its component(s) may result in prolonged or permanent epigenetic modification of the target gene. For example, the epigenetic modification may be stable for at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, or 12 weeks or more; or 3, 4, 5, 6, 7, 8, 9, 10, 11, or 12 months or more, after introduction of the epigenetic editor into the host cell. The epigenetic modification may be maintained after one or more mitotic and/or meiotic events of the host cell. In particular embodiments, the epigenetic modification is maintained across generations in offspring generated or derived from the host cell.

## VIII. Therapeutic Uses of Epigenetic Editors

(237) The present disclosure also provides methods for treating or preventing a condition in a subject, comprising administering to the subject an epigenetic editor or pharmaceutical composition as described herein. The epigenetic editor may effectuate an epigenetic modification of a target polynucleotide sequence in a target gene associated with a disease, condition, or disorder in the subject, thereby modulating expression of the target gene to treat or prevent the disease, condition, or disorder. In some embodiments, the epigenetic editor reduces the expression of the target gene to an extent sufficient to

achieve a desired effect, e.g., a therapeutically relevant effect such as the prevention or treatment of the disease, condition, or disorder.

(238) In some embodiments, a subject is administered a system for modulating (e.g., repressing) expression of HBV or of an HBV gene, wherein the system comprises (1) the fusion protein(s) and, where relevant, guide polynucleotide(s) of an epigenetic editor as described herein, or (2) nucleic acid molecules encoding said fusion protein(s) and, where relevant, guide polynucleotide(s).

(239) “Treat,” “treating” and “treatment” refer to a method of alleviating or abrogating a biological disorder and/or at least one of its attendant symptoms. As used herein, to “alleviate” a disease, disorder or condition means reducing the severity and/or occurrence frequency of the symptoms of the disease, disorder, or condition. Further, references herein to “treatment” include references to curative, palliative and prophylactic treatment. In some embodiments, as compared with an equivalent untreated control, alleviating a symptom may involve reduction of the symptom by at least 3%, 5%, 10%, 20%, 40%, 50%, 60%, 80%, 90%, 95%, 98%, 99%, 99.5%, 99.9%, or 100% as measured by any standard technique.

(240) In some embodiments, the subject may be a mammal, e.g., a human. In some embodiments, the subject is selected from a non-human primate such as chimpanzee, cynomolgus monkey, or macaque, and other apes and monkey species.

(241) Some aspects of this disclosure provide methods comprising administering an epigenetic editing system to a subject characterized by the presence of detectable levels of HBV DNA, HBsAg, and/or HBeAg in the plasma of the subject, for example, a subject having a chronic HBV infection. In some such embodiments, the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding the same, wherein the first DNA binding domain binds a first target region of an HBV gene or genome, and the administering results in a reduction of the level of HBV DNA, the level of HBsAg, and/or the level of HBeAg in the plasma of the subject, and the reduction of the level of HBV DNA, of the level of HBsAg, and/or of the level of HBeAg in the plasma of the subject, is at least 90% (a 1-log reduction) compared to the respective level observed or observable in the plasma of the subject prior to the administering, and the 1-log reduction is maintained for at least 14 days after the administering. In some embodiments, the reduction of the level of HBV DNA in the plasma of the subject is at least 90% (a 1-log reduction). In some embodiments, the reduction of the level of HBV DNA in the plasma of the subject is at least 99% (a 2-log reduction). In some embodiments, the reduction of the level of HBsAg in the plasma of the subject is at least 90% (a 1-log reduction). In some embodiments, the reduction of the level of HBsAg in the plasma of the subject is at least 99% (a 2-log reduction). In some embodiments, the reduction of the level of HBeAg in the plasma of the subject is at least 90% (a 1-log reduction). In some embodiments, the reduction of the level of HBeAg in the plasma of the subject is at least 99% (a 2-log reduction). In some embodiments, the reduction is maintained for at least 21 days. In some embodiments, the reduction is maintained for at least 28 days. In some embodiments, the reduction is maintained for at least 35 days. In some embodiments, the reduction is maintained for at least 42 days. In some embodiments, the reduction is maintained for at least 56 days. In some embodiments, the reduction is maintained for at least 70 days. In some embodiments, the reduction is maintained for at least 84 days. In some embodiments, the reduction is maintained for at least 112 days. In some embodiments, the reduction is maintained for at least 140 days. In some embodiments, the reduction is maintained for at least 168 days. In some embodiments, the reduction is maintained for at least 6 months. In some embodiments, the reduction is maintained for at least 9 months. In some embodiments, the reduction is maintained for at least 12 months. In some embodiments, the reduction is maintained for at least 24 months. In some embodiments, the HBV genome comprises HBV genotype A. In some embodiments, the HBV genome comprises HBV genotype B. In some embodiments, the HBV genome comprises HBV genotype C. In some embodiments, the HBV genome comprises HBV genotype D. In some embodiments, the HBV genome comprises HBV genotype E. In some embodiments, the HBV genome comprises HBV genotype F. In some embodiments, the HBV genome comprises HBV genotype G. In some embodiments, the HBV genome comprises HBV genotype H. In some embodiments, the HBV genome comprises a sequence with at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99% sequence identity to an HBV genome sequence provided herein. In some

embodiments, the first target region is located in a region of the HBV genome within nucleotides 0-303 of an HBV genome provided herein. In some embodiments, the first target region is located within nucleotides 0-303 of SEQ ID NO: 1082. In some embodiments, the first target region is located within nucleotides 0-303 of SEQ ID NO: 1083. In some embodiments, the first target region is located in a region of the HBV genome within nucleotides 1000-2448 of an HBV genome provided herein. In some embodiments, the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1082. In some embodiments, the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1083. In some embodiments, the first target region is located in a region of the HBV genome within nucleotides 2802-3182 of an HBV genome provided herein. In some embodiments, the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1082. In some embodiments, the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1083. In some embodiments, the first target region of the HBV genome is located in an HBV CpG island (CGI). In some embodiments, the CGI is an HBV canonical CGI. In some embodiments, the CGI is canonical CGI-I. In some embodiments, CGI is canonical CGI-I of HBV genotype D. In some embodiments, CGI-I spans nucleotides 186-288 of SEQ ID NO: 1082. In some embodiments, CGI-I spans nucleotides 186-288 of SEQ ID NO: 1083. In some embodiments, the CGI is canonical CGI-II. In some embodiments, the CGI is canonical CGI-II HBV genotype D. In some embodiments, the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1082. In some embodiments, the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1083. In some embodiments, the CGI is canonical CGI-III. In some embodiments, the CGI is canonical CGI-III HBV genotype D. In some embodiments, the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1082. In some embodiments, the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1083. In some embodiments, the first target region of the HBV genome is located in a promoter. In some embodiments, the first target region of the HBV genome is located in the sp1 promoter. In some embodiments, the first target region of the HBV genome is located in sp2 promoter. In some embodiments, the first target region of the HBV genome is located in cp promoter. In some embodiments, the first target region of the HBV genome is located in xp promoter. In some embodiments, the first target region of the HBV genome is located in an enhancer region. In some embodiments, the first target region of the HBV genome is located in Enh I. In some embodiments, the first target region of the HBV genome is located in Enh II. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a pgRNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a preCore RNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes a preS RNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes an S RNA transcript. In some embodiments, the first target region of the HBV genome is located in a section of the HBV genome that encodes an HBx RNA transcript. In some embodiments, the first target region of the HBV genome is within 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) of an HBV transcription start site (TSS). In some embodiments, the TSS is a pg RNA TSS. In some embodiments, the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the pg RNA TSS. In some embodiments, the pg RNA TSS is located at nucleotide 1820 of SEQ ID NO: 1082 or at nucleotide 1820 of SEQ ID NO: 1083. In some embodiments, the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1083. In some embodiments, the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1082. In some embodiments, the first

[illegible]

target region is within 100 base pairs of nucleotide 1243 in SEQ ID NO: 1083. In some embodiments, the first target region is within 600 base pairs of nucleotide 1338 in SEQ ID NO: 1083. In some embodiments, the first target region is within 500 base pairs of nucleotide 1338 in SEQ ID NO: 1082. In some embodiments, the first target region is within 500 base pairs of nucleotide 1338 in SEQ ID NO: 1083. In some embodiments, first target region is within 400 base pairs of nucleotide 1338 in SEQ ID NO: 1082. In some embodiments, the first target region is within 400 base pairs of nucleotide 1338 in SEQ ID NO: 1083. In some embodiments, the first target region is within 300 base pairs of nucleotide 1338 in SEQ ID NO: 1082. In some embodiments, the first target region is within 300 base pairs of nucleotide 1338 in SEQ ID NO: 1083. In some embodiments, the first target region is within 200 base pairs of nucleotide 1338 in SEQ ID NO: 1082. In some embodiments, the first target region is within 200 base pairs of nucleotide 1338 in SEQ ID NO: 1083. In some embodiments, the first target region is within 100 base pairs of nucleotide 1338 in SEQ ID NO: 1082. In some embodiments, the first target region is within 100 base pairs of nucleotide 1338 in SEQ ID NO: 1083. In some embodiments, the reduction is a reduction in the number of HBV viral episomes. In some embodiments, the reduction is a reduction in the number of cccDNA genomes. In some embodiments, the reduction is a reduction in total HBV DNA. In some embodiments, the reduction is a reduction in the replication of the HBV genome. In some embodiments, the reduction is a reduction in a level of expression of a protein product encoded by the HBV genome. In some embodiments, the reduction is a reduction in a level of HBsAg. In some embodiments, the reduction is a reduction in a level of HBeAg. In some embodiments, the reduction is a reduction of total HBV DNA of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and the reduction is maintained for at least 14 days after the contacting or the administering. In some embodiments, the reduction is a reduction of HBeAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and the reduction is maintained for at least 14 days after the contacting or the administering. In some embodiments, the reduction is a reduction of HBsAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and the reduction is maintained at or below that level for at least 14 days after the contacting or the administering. In some embodiments, the reduction is a reduction of at least 90%. In some embodiments, the reduction is a reduction of at least 95%. In some embodiments, the reduction is a reduction of at least 99%. In some embodiments, the reduction is a reduction of at least 99.9%. In some embodiments, the reduction is maintained for at least 14 days after the contacting or the administering. In some embodiments, the reduction is maintained for at least 21 days. In some embodiments, the reduction is maintained for at least 28 days. In some embodiments, the reduction is maintained for at least 35 days. In some embodiments, the reduction is maintained for at least 42 days. In some embodiments, the reduction is maintained for at least 56 days. In some embodiments, the reduction is maintained for at least 70 days. In some embodiments, the reduction is maintained for at least 84 days. In some embodiments, the reduction is maintained for at least 112 days. In some embodiments, the reduction is maintained for at least 140 days. In some embodiments, the reduction is maintained for at least 168 days. In some embodiments, the reduction is maintained for at least 6 months. In some embodiments, the reduction is maintained for at least 7 months. In some embodiments, the reduction is maintained for at least 8 months. In some embodiments, the reduction is maintained for at least 9 months. In some embodiments, the reduction is maintained for at least 12 months. In some embodiments, the reduction is maintained for at least 18 months. In some embodiments, the reduction is maintained for at least 24 months. In some embodiments, the epigenetic editing system is administered as a monotherapy. Accordingly, in some embodiments, the method does not comprise administering a nucleoside or nucleotide analog (NUC) to the subject. In some embodiments, the method further comprises administering a NUC to the subject. In some embodiments, the first DNA binding domain comprises a CRISPR-Cas protein. In some embodiments, the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region. In some embodiments, the gRNA comprises a sequence selected from a gRNA provided herein, and preferably the gRNA comprises a sequence provided in Table 12 or 13. In some embodiments, the first DNA binding domain comprises a zinc-finger protein. In some embodiments, the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc

finger motif provided herein, e.g., in Table 1 or Table 18. In some embodiments, the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein. In some embodiments, the transcriptional repressor domain comprises ZIM3. In some embodiments, the first DNMT domain is a DNMT3A domain or a DNMT3L domain. In some embodiments, the first DNMT domain comprises a sequence of a DNMT domain provided herein. In some embodiments, the epigenetic editing system comprises the fusion protein provided in SEQ ID NO: 1248 or the fusion protein provided in SEQ ID NO: 1252 and at least one guide RNA provided as gRNA #003, gRNA #007, gRNA #008, gRNA #009, gRNA #011, or gRNA #015 herein. Some aspects of this disclosure provide epigenetic editing systems for use in the methods described herein. In some embodiments, the epigenetic editing system comprises a fusion protein or a nucleic acid encoding the fusion protein, and the fusion protein comprises: (a) a DNA-binding domain that binds a target region of a HBV gene or genome, (b) a first DNA methyltransferase (DNMT) domain, and (c) a transcriptional repressor domain. In some embodiments, the fusion protein comprises a sequence of a fusion protein provided herein. In some embodiments, the DNA-binding domain is a CRISPR-Cas DNA binding domain, and the epigenetic editing system comprises at least gRNA provided herein. In some embodiments, the epigenetic editing system comprises the fusion protein provided in SEQ ID NO: 1248 or the fusion protein provided in SEQ ID NO: 1252 and at least one guide RNA provided as gRNA #003, gRNA #007, gRNA #008, gRNA #009, gRNA #011, or gRNA #015 herein.

(242) In some embodiments, the subject is a mammalian subject having, or having been diagnosed with, a Hepatitis B virus (HBV) infection. In some embodiments, the subject is a mammalian subject having, or having been diagnosed with, a Hepatitis D virus infection.

(243) In some embodiments, the subject is a mammalian subject, for example, a human subject, having, or having been diagnosed with, a Hepatitis B virus (HBV) infection. In some embodiments, the subject is a mammalian subject, for example, a human subject, having, or having been diagnosed with Hepatitis B. In some embodiments, the subject is a mammalian subject, for example, a human subject, having, or having been diagnosed with, a Hepatitis D virus infection. In some embodiments, a patient to be treated with an epigenetic editor of the present disclosure has received prior treatment for the condition to be treated (e.g., an HBV and/or HDV infection, or Hepatitis B). In other embodiments, the patient has not received such prior treatment. In some embodiments, the patient has failed on (or is refractory to) a prior treatment for the condition (e.g., a prior HBV treatment).

(244) In some embodiments, contacting the HBV gene or genome or a cell with an epigenetic editor as described herein results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by the HBV gene or genome. In some embodiments, the reduction is at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% compared to contacting the HBV gene or genome or the cell with a suitable control or without contacting the HBV gene or genome or the cell with the epigenetic editor described herein. In some embodiments, the reduction is maintained for at least 6 days, 19 days, 27 days, 42 days, or 168 days. In some embodiments, the protein product comprises a HBe antigen or a HBs antigen.

(245) In some embodiments, administering to the subject an epigenetic editor or pharmaceutical composition as described herein results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by the HBV gene or genome. In some embodiments, the reduction is at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% compared to administering a suitable control or without administering the epigenetic editor or pharmaceutical composition described herein. In some embodiments, the reduction is maintained for at least 6 days, 19 days, 27 days, 42 days, or 168 days. In some embodiments, the protein product comprises a HBe antigen or a HBs antigen.

(246) An epigenetic editor of the present disclosure may be administered in a therapeutically effective amount to a patient with a condition described herein. "Therapeutically effective amount," as used herein, refers to an amount of the therapeutic agent being administered that will relieve to some extent one or more of the symptoms of the disorder being treated, and/or result in clinical endpoint(s) desired

by healthcare professionals. An effective amount for therapy may be measured by its ability to stabilize disease progression and/or ameliorate symptoms in a patient, and preferably to reverse disease progression. The ability of an epigenetic editor of the present disclosure to reduce or silence HBV expression may be evaluated by in vitro assays, e.g., as described herein, as well as in suitable animal models that are predictive of the efficacy in humans. Suitable dosage regimens will be selected in order to provide an optimum therapeutic response in each particular situation, for example, administered as a single bolus or as a continuous infusion, and with possible adjustment of the dosage as indicated by the exigencies of each case.

(247) An epigenetic editor of the present disclosure may be administered without additional therapeutic treatments, i.e., as a stand-alone therapy (monotherapy). Alternatively, treatment with an epigenetic editor of the present disclosure may include at least one additional therapeutic treatment (combination therapy). In some embodiments, the additional therapeutic agent is any known in the art to treat an HBV infection. The current standard therapy for HBV employs nucleoside/nucleotide analogs (NUCs) and interferon (IFN). NUCs are viral polymerase and reverse transcriptase inhibitors that can efficiently suppress HBV viral replication, resulting in rapid HBV DNA reduction. NUCs do not directly target HBV cccDNA transcription, but NUC treatment of human HBV patients has been reported to reduce plasma HBV biomarkers such as HBeAg and HBsAg to some extent. Prolonged therapy with NUCs is frequently associated with the pathogen developing a resistance to the treatment, but some NUCs have been reported to be able to achieve long-term viral suppression and halt disease progression. IFN-based therapy has both direct antiviral and immunomodulatory effects, and has been reported to prevent the formation of replication-competent pregenomic RNA-containing HBV capsids, or otherwise accelerates their degradation, thereby inhibiting HBV replication. See, e.g., Su et al., Improving clinical outcomes of chronic hepatitis B virus infection. *Expert Rev Gastroenterol Hepatol*. 2015; 9:141-154; European Association for the Study of the Liver. EASL clinical practice guidelines: management of chronic hepatitis B virus infection. *J Hepatol*. 2012; 57:167-185; Wieland et al., Intrahepatic induction of alpha/beta interferon eliminates viral RNA-containing capsids in hepatitis B virus transgenic mice. *J Virol*. 2000; and Wieland et al., Interferon prevents formation of replication-competent hepatitis B virus RNA-containing nucleocapsids. *Proc Natl Acad Sci USA*. 2005; 102:9913-9917, the entire contents of each of which are incorporated herein by reference.

(248) In some embodiments, an epigenetic editor of the present disclosure is administered to a subject in need thereof, e.g., a subject having an HBV infection, without additional therapeutic treatment, e.g., without the co-administration of NUCs or IFN, or any other therapeutic treatment aimed at HBV, i.e., as a stand-alone therapy (monotherapy). In some such embodiments, a durable reduction of an HBV biomarker (e.g., as measured as the plasma level of HBV DNA, HBsAg, or HBeAg) by at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, at least 99.9%, or more, is achieved over a time period of at least 14 days, at least 21 days, at least 28 days, at least 35 days, at least 42 days, at least 56 days, at least 70 days, at least 84 days, at least 112 days, at least 140 days, at least 168 days, at least 6 months, at least 7 months, at least 8 months, at least 9 months, at least 10 months, at least 11 months, at least 12 months, at least 18 months, at least 24 months, or longer, after a single-dose administration of the epigenetic editor to the subject.

(249) In some embodiments, an epigenetic editor of the present disclosure is administered to a subject in need thereof, e.g., a subject having an HBV infection, in combination with (i.e., in temporal proximity) at least one additional HBV therapeutics, e.g., with NUCs and/or IFN therapeutics, or with any other therapeutic treatment aimed at HBV, i.e., as a combination therapy (monotherapy). In some such embodiments, a durable reduction of an HBV biomarker (e.g., as measured as the plasma level of HBV DNA, HBsAg, or HBeAg) by at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, at least 99.9%, or more, is achieved over a time period of at least 14 days, at least 21 days, at least 28 days, at least 35 days, at least 42 days, at least 56 days, at least 70 days, at least 84 days, at least 112 days, at least 140 days, at least 168 days, at least 6 months, at least 7 months, at least 8 months, at least 9 months, at least 10 months, at least 11 months, at least 12 months, at least 18 months, at least 24 months, or longer.

(250) An epigenetic editor of the present disclosure may be administered without additional therapeutic

treatments, i.e., as a stand-alone therapy (monotherapy). Alternatively, treatment with an epigenetic editor of the present disclosure may include at least one additional therapeutic treatment (combination therapy). In some embodiments, the additional therapeutic agent is any known in the art to HBV and/or HDV. In some embodiments, therapeutic agents include, but are not limited to, antivirals, such as entecavir, tenofovir, lamivudine, telvivudine, bictegravir, emtricitabine, or defovir, as well as immune modulators, such as pegylated interferon and interferon alpha.

(251) The epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure may be administered by any method accepted in the art (e.g., parenterally, intravenously, intradermally, or intramuscularly).

(252) The epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure may be administered to a subject once, twice, three times, or 4, 5, 6, 7, 8, 9, 10, or more times. In some embodiments, the one, two, three, or 4, 5, 6, 7, 8, 9, 10, or more administrations of epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) are in temporal proximity, e.g., within 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 1 week, 2 weeks, 4 weeks, 1 month or two months of each other. In some embodiments, a subject is re-dosed with the epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure for at least one more time after an initial dose. In some cases, a subject is administered with a subsequent dose of the epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure, which target a different DNA region of the HBV genome than the DNA region of the HBV genome that is targeted by the epigenetic editors or components thereof that the subject receives at the initial dose. In some cases, a subject is administered with multiple doses (e.g., 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) of the same epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure. In some cases, a subject is administered with a single dose of different epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure, at least two of which target different DNA regions of the HBV genome. In some cases, a subject is administered with multiple doses (e.g., 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) of different epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure, at least two of which target different DNA regions of the HBV genome. In some embodiments, redosing of the epigenetic editors or components thereof (or nucleic acid molecules encoding the epigenetic editors or components thereof) of the present disclosure has a better therapeutic efficacy than a single dose of the same, e.g., more potent suppression of HBV replication, or more profound reduction in HBV DNA and/or HBV antigens (e.g., HBsAg, HBeAg, and/or HBV core antigen (HBcAg)) present in the subject, e.g., in the circulation system and/or liver of the subject.

## XII. Definitions

(253) The term “nucleic acid” as used herein refers to any oligonucleotide or polynucleotide containing nucleotides (e.g., deoxyribonucleotides or ribonucleotides) in either single- or double-strand form, and includes DNA and RNA. “Nucleotides” contain a sugar deoxyribose (DNA) or ribose (RNA), a base, and a phosphate group, and are linked together through the phosphate groups. “Bases” include purines and pyrimidines, which include natural compounds such as adenine, thymine, guanine, cytosine, uracil, inosine, and natural analogs; as well as synthetic derivatives of purines and pyrimidines, which include, but are not limited to, modified versions which place new reactive groups such as amines, alcohols, thiols, carboxylates, alkylhalides, etc. Nucleic acids may contain known nucleotide analogs and/or modified backbone residues or linkages, which may be synthetic, naturally occurring, and non-naturally occurring. Such nucleotide analogs, modified residues, and modified linkages are well known in the art, and may provide a nucleic acid molecule with enhanced cellular uptake, reduced immunogenicity, and/or increased stability in the presence of nucleases.

(254) As used herein, an “isolated” or “purified” nucleic acid molecule is a nucleic acid molecule that exists apart from its native environment. For example, an “isolated” or “purified” nucleic acid molecule (1) has been separated away from the nucleic acids of the genomic DNA or cellular RNA of its source of



origin; and/or (2) does not occur in nature. In some embodiments, an “isolated” or “purified” nucleic acid molecule is a recombinant nucleic acid molecule.

(255) It will be understood that in addition to the specific proteins and nucleic acid molecules mentioned herein, the present disclosure also contemplates the use of variants, derivatives, homologs, and fragments thereof. A variant of any given sequence may have the specific sequence of residues (whether amino acid or nucleic acid residues) modified in such a manner that the polypeptide or polynucleotide in question substantially retains at least one of its endogenous functions. A variant sequence can be obtained by addition, deletion, substitution, modification, replacement and/or variation of at least one residue present in the naturally-occurring sequence (in some embodiments, no more than 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, or 20 residues). For specific proteins described herein (e.g., KRAB, dCas9, DNMT3A, and DNMT3L proteins described herein), the present disclosure also contemplates any of the protein's naturally occurring forms, or variants or homologs that retain at least one of its endogenous functions (e.g., at least 50%, 60%, 70%, 80%, 90%, 85%, 96%, 97%, 98%, or 99% of its function as compared to the specific protein described).

(256) As used herein, a homologue of any polypeptide or nucleic acid sequence contemplated herein includes sequences having a certain homology with the wildtype amino acid and nucleic sequence. A homologous sequence may include a sequence, e.g. an amino acid sequence which may be at least 50%, 55%, 65%, 75%, 85%, 90%, 91%, 92%<93%, 94%, 95%, 96%, 97%, 98%, or 99% identical to the subject sequence. The term “percent identical” in the context of amino acid or nucleotide sequences refers to the percent of residues in two sequences that are the same when aligned for maximum correspondence. In some embodiments, the length of a reference sequence aligned for comparison purposes is at least 30%, (e.g., at least 40, 50, 60, 70, 80, or 90%, or 100%) of the reference sequence. Sequence identity may be measured using sequence analysis software (for example, Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wis. 53705, BLAST, BESTFIT, GAP, or PILEUP/PRETTYBOX programs). Such software matches identical or similar sequences by assigning degrees of homology to various substitutions, deletions, and/or other modifications. In an exemplary approach to determining the degree of identity, a BLAST program may be used, with a probability score between  $e^{-3}$  and  $e^{-100}$  indicating a closely related sequence.

(257) The percent identity of two nucleotide or polypeptide sequences is determined by, e.g., BLAST® using default parameters (available at the U.S. National Library of Medicine's National Center for Biotechnology Information website). In some embodiments, the length of a reference sequence aligned for comparison purposes is at least 30%, (e.g., at least 40, 50, 60, 70, 80, or 90%) of the reference sequence.

(258) It will be understood that the numbering of the specific positions or residues in polypeptide sequences depends on the particular protein and numbering scheme used. Numbering might be different, e.g., in precursors of a mature protein and the mature protein itself, and differences in sequences from species to species may affect numbering. One of skill in the art will be able to identify the respective residue in any homologous protein and in the respective encoding nucleic acid by methods well known in the art, e.g., by sequence alignment and determination of homologous residues.

(259) The term “modulate” or “alter” refers to a change in the quantity, degree, or extent of a function. For example, an epigenetic editor as described herein may modulate the activity of a promoter sequence by binding to a motif within the promoter, thereby inducing, enhancing, or suppressing transcription of a gene operatively linked to the promoter sequence. As other examples, an epigenetic editor as described herein may block RNA polymerase from transcribing a gene, or may inhibit translation of an mRNA transcript. The terms “inhibit,” “repress,” “suppress,” “silence” and the like, when used in reference to an epigenetic editor or a component thereof as described herein, refers to decreasing or preventing the activity (e.g., transcription) of a nucleic acid sequence (e.g., a target gene) or protein relative to the activity of the nucleic acid sequence or protein in the absence of the epigenetic editor or component thereof. The term may include partially or totally blocking activity, or preventing or delaying activity. The inhibited activity may be, e.g., 10%, 20%, 30%, 40%, 50%, 60%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 99.9% less than that of a control, or may be, e.g., at

least 1.5-fold, 2-fold, 3-fold, 4-fold, 5-fold, or 10-fold less than that of a control. For example, in some embodiments, the inhibited activity (e.g., the transcription or expression of an HBV target gene, or the level of an HBV biomarker) may be at least 70% less than that of a control. In some embodiments, the inhibited activity may be at least 80% less than that of a control. In some embodiments, the inhibited activity may be at least 90% less than that of a control (1 log reduction). In some embodiments, the inhibited activity may be at least 91% less than that of a control. In some embodiments, the inhibited activity may be at least 92% less than that of a control. In some embodiments, the inhibited activity may be at least 93% less than that of a control. In some embodiments, the inhibited activity may be at least 94% less than that of a control. In some embodiments, the inhibited activity may be at least 95% less than that of a control. In some embodiments, the inhibited activity may be at least 96% less than that of a control. In some embodiments, the inhibited activity may be at least 97% less than that of a control. In some embodiments, the inhibited activity may be at least 98% less than that of a control. In some embodiments, the inhibited activity may be at least 99% less than that of a control (2 log reduction). In some embodiments, the inhibited activity may be at least 99.9% less than that of a control (3 log reduction).

(260) The term “about” or “approximately” means within an acceptable error range for the particular value as determined by one of ordinary skill in the art, which will depend in part on how the value is measured or determined, e.g., the limitations of the measurement system. For example, “about” can mean within one or more than one standard deviation, per the practice in the given value. Where particular values are described in the application and claims, unless otherwise stated, the term “about” should be assumed to mean an acceptable error range for the particular value.

(261) Ranges provided herein are understood to be shorthand for all of the values within the range. For example, a range of 1 to 50 is understood to include any number, combination of numbers, or sub-range from the group consisting of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50, as well as all intervening decimal values between the aforementioned integers such as, for example, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, and 1.9. With respect to sub-ranges, “nested sub-ranges” that extend from either end point of the range are specifically contemplated. For example, a nested sub-range of an exemplary range of 1 to 50 may comprise 1 to 10, 1 to 20, 1 to 30, and 1 to 40 in one direction, or 50 to 40, 50 to 30, 50 to 20, and 50 to 10 in the other direction.

(262) Unless otherwise defined herein, scientific and technical terms used in connection with the present disclosure shall have the meanings that are commonly understood by those of ordinary skill in the art. Exemplary methods and materials are described below, although methods and materials similar or equivalent to those described herein can also be used in the practice or testing of the present disclosure. In case of conflict, the present specification, including definitions, will control. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. Throughout this specification and embodiments, the words “have” and “comprise,” or variations such as “has,” “having,” “comprises,” or “comprising,” will be understood to imply the inclusion of a stated integer or group of integers but not the exclusion of any other integer or group of integers. The recitation of a listing of elements herein includes any of the elements singly or in any combination. The recitation of an embodiment herein includes that embodiment as a single embodiment, or in combination with any other embodiment(s) herein. All publications, patents, patent applications, and other references mentioned herein, including, where applicable, any supplementary information, are incorporated by reference in their entirety. To the extent that references incorporated by reference contradict the disclosure contained in the specification, the specification is intended to supersede and/or take precedence over any such contradictory material. Although a number of documents are cited herein, this citation does not constitute an admission that any of these documents forms part of the common general knowledge in the art.

#### Listings of Exemplary Embodiments

(263) In order that the present disclosure may be better understood, the following listings of exemplary embodiments is provided. This listing is for purposes of illustration of certain embodiments only. Additional embodiments will be apparent to the skilled artisan based on the present disclosure, and the

listing below is not to be construed as limiting the scope of the present disclosure.

(264) LISTING #1 of Exemplary Embodiments:

(265) 1. A method of modifying an epigenetic state of a hepatitis B virus (HBV) gene or genome, comprising contacting the HBV gene or genome with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding the same, wherein the first DNA binding domain binds a first target region of the HBV gene or genome, and wherein the contacting results in a reduction of number of HBV viral episomes, replication of the HBV gene or genome, and/or expression of a protein product encoded by the HBV gene or genome, wherein the reduction is at least about 50%, and preferably wherein the reduction is at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99%, compared to contacting the HBV gene or genome with a suitable control.

(266) 2. A method of treating an HBV infection in a subject comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the administering results in a reduction of number of HBV viral episomes, replication of the HBV gene or genome, and/or expression of a protein product encoded by the HBV gene or genome, wherein the reduction is at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99%, compared to administering a suitable control.

(267) 3. A method of modulating expression of an HBV gene or genome comprising contacting the HBV gene or genome with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of the HBV gene or genome, and wherein the contacting results in a reduction of expression of a gene product encoded by the HBV gene or genome, optionally, wherein the gene product is a nucleic acid or a protein, wherein the reduction is at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99%, compared to contacting the HBV genome with a suitable control.

(268) 4. A method of inhibiting viral replication in a cell infected with an HBV comprising administering an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the epigenetic editing system targets a target region of the HBV gene or genome, and wherein the administering results in a reduction of number of HBV viral episomes or replication of the HBV gene or genome, wherein the reduction is at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99%, compared to administering a suitable control.

(269) 5. The method of any one of embodiments 1-4, wherein the reduction is at least 70%.

(270) 6. The method of any one of embodiments 1-4, wherein the reduction is at least 80%.

(271) 7. The method of any one of embodiments 1-4, wherein the reduction is at least 90%.

(272) 8. The method of any one of embodiments 1-4, wherein the reduction is at least 95%.

(273) 9. The method of any one of embodiments 1-4, wherein the reduction is at least 99%.

(274) 10. The method of any one of embodiments 1-4, wherein the reduction is greater than 99%.

(275) 11. The method of any one of embodiments 1-10, wherein the HBV genome is a covalently closed circular DNA (cccDNA).

(276) 12. The method of any one of embodiments 1-10, wherein the HBV genome is an HBV integrated DNA.

(277) 13. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype A.

(278) 14. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype B.

(279) 15. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype C.

(280) 16. The method of any one of embodiments 1-12, wherein the HBV genome comprises, HBV genotype D.

(281) 17. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype E.

(282) 18. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype F.

(283) 19. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype G.

(284) 20. The method of any one of embodiments 1-12, wherein the HBV genome comprises HBV genotype H.

(285) 21. The method of any one of embodiments 1-12, wherein the HBV genome comprises a sequence with at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99% sequence identity to an HBV genome sequence provided herein.

(286) 22. The method of any one of embodiments 1-21, wherein the first target region is located in a region of the HBV genome within nucleotides 0-303 of an HBV genome provided herein.

(287) 23. The method of any one of embodiments 1-21, wherein the first target region is located within nucleotides 0-303 of SEQ ID NO: 1082.

(288) 24. The method of any one of embodiments 1-21, wherein the first target region is located within nucleotides 0-303 of SEQ ID NO: 1083.

(289) 25. The method of any one of embodiments 1-21, wherein the first target region is located in a region of the HBV genome within nucleotides 1000-2448 of an HBV genome provided herein.

(290) 26. The method of any one of embodiments 1-21, wherein the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1082.

(291) 27. The method of any one of embodiments 1-21, wherein the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1083.

(292) 28. The method of any one of embodiments 1-21, wherein the first target region is located in a region of the HBV genome within nucleotides 2802-3182 of an HBV genome provided herein.

(293) 29. The method of any one of embodiments 1-21, wherein the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1082.

(294) 30. The method of any one of embodiments 1-21, wherein the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1083.

(295) 31. The method of any one of embodiments 1-21, wherein the first target region of the HBV genome is located in an HBV CpG island (CGI).

(296) 32. The method of embodiment 31, wherein the CGI is an HBV canonical CGI.

(297) 33. The method of embodiment 31, wherein the CGI is canonical CGI-I.

(298) 34. The method of embodiment 31, wherein the CGI is canonical CGI-I of HBV genotype D.

(299) 35. The method of embodiment 33, wherein CGI-I spans nucleotides 186-288 of SEQ ID NO: 1082.

(300) 36. The method of embodiment 33, wherein CGI-I spans nucleotides 186-288 of SEQ ID NO: 1083.

(301) 37. The method of embodiment 31, wherein the CGI is canonical CGI-II.

(302) 38. The method of embodiment 31, wherein the CGI is canonical CGI-II HBV genotype D.

(303) 39. The method of embodiment 38, wherein the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1082.

(304) 40. The method of embodiment 38, wherein the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1083.

(305) 41. The method of embodiment 31, wherein the CGI is canonical CGI-III.

(306) 42. The method of embodiment 31, wherein the CGI is canonical CGI-III HBV genotype D.

(307) 43. The method of embodiment 42, wherein the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1082.

(308) 44. The method of embodiment 42, wherein the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1083.

(309) 45. The method of any one of embodiments 1-21, wherein the first target region of the HBV genome is located in a promotor.

(310) 46. The method of embodiment 45, wherein the first target region of the HBV genome is located in the sp1 promotor.

(311) 47. The method of embodiment 45, wherein the first target region of the HBV genome is located in sp2 promotor.

(312) 48. The method of embodiment 45, wherein the first target region of the HBV genome is located in cp promotor.

(313) 49. The method of embodiment 45, wherein the first target region of the HBV genome is located in xp promotor.

(314) 50. The method of any one of embodiments 1-21, wherein the first target region of the HBV genome is located in an enhancer region.

(315) 51. The method of embodiment 50, wherein the first target region of the HBV genome is located in Enh I.

(316) 52. The method of embodiment 50, wherein the first target region of the HBV genome is located in Enh II.

(317) 53. The method of any one of embodiments 1-21, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a transcript.

(318) 54. The method of embodiment 53, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a pgRNA transcript.

(319) 55. The method of embodiment 53, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a preCore RNA transcript.

(320) 56. The method of embodiment 53, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a preS RNA transcript.

(321) 57. The method of embodiment 53, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes an S RNA transcript.

(322) 58. The method of embodiment 53, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes an HBx RNA transcript.

(323) 59. The method of any one of embodiments 1-21, wherein the first target region of the HBV genome is within 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) of an HBV transcription start site (TSS).

(324) 60. The method of embodiment 59, wherein the TSS is a pg RNA TSS.

(325) 61. The method of embodiment 60, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the pg RNA TSS.

(326) 62. The method of embodiment 60, wherein the pg RNA TSS is located at nucleotide 1820 of SEQ ID NO: 1082 or at nucleotide 1820 of SEQ ID NO: 1083.

(327) 63. The method of embodiment 60, wherein the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(328) 64. The method of embodiment 60, wherein the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(329) 65. The method of embodiment 60, wherein the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(330) 66. The method of embodiment 60, wherein the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(331) 67. The method of embodiment 60, wherein the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(332) 68. The method of embodiment 60, wherein the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(333) 69. The method of embodiment 60, wherein the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(334) 70. The method of embodiment 60, wherein the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(335) 71. The method of embodiment 60, wherein the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(336) 72. The method of embodiment 60, wherein the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(337) 73. The method of embodiment 60, wherein the first target region is within 100 base pairs of nucleotide 1820 in SEQ ID NO: 1082 or wherein the first target region is within 100 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(338) 74. The method of embodiment 59, wherein the TSS is a preC RNA TSS.

(339) 75. The method of embodiment 74, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the preC RNA TSS.

(340) 76. The method of embodiment 74, wherein the preC RNA TSS is located at nucleotide 1791 of SEQ ID NO: 1082 or at nucleotide 1791 of SEQ ID NO: 1083.

(341) 77. The method of embodiment 74, wherein the first target region is within 600 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(342) 78. The method of embodiment 74, wherein the first target region is within 600 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(343) 79. The method of embodiment 74, wherein the first target region is within 500 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(344) 80. The method of embodiment 74, wherein the first target region is within 500 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(345) 81. The method of embodiment 74, wherein the first target region is within 400 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(346) 82. The method of embodiment 74, wherein the first target region is within 400 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(347) 83. The method of embodiment 74, wherein the first target region is within 300 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(348) 84. The method of embodiment 74, wherein the first target region is within 300 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(349) 85. The method of embodiment 74, wherein the first target region is within 200 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(350) 86. The method of embodiment 74, wherein the first target region is within 200 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(351) 87. The method of embodiment 74, wherein the first target region is within 100 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(352) 88. The method of embodiment 74, wherein the first target region is within 100 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(353) 89. The method of embodiment 59, wherein the TSS is a preS2 RNA TSS.

(354) 90. The method of embodiment 89, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the preS2 RNA TSS.

(355) 91. The method of embodiment 89, wherein the preS2 RNA TSS is located at nucleotide 3159 of SEQ ID NO: 1082 or at nucleotide 3159 of SEQ ID NO: 1083.

(356) 92. The method of embodiment 89, wherein the first target region is within 600 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(357) 93. The method of embodiment 89, wherein the first target region is within 600 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(358) 94. The method of embodiment 89, wherein the first target region is within 500 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(359) 95. The method of embodiment 89, wherein the first target region is within 500 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(360) 96. The method of embodiment 89, wherein the first target region is within 400 base pairs of

nucleotide 3159 in SEQ ID NO: 1082.

(361) 97. The method of embodiment 89, wherein the first target region is within 400 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(362) 98. The method of embodiment 89, wherein the first target region is within 300 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(363) 99. The method of embodiment 89, wherein the first target region is within 300 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(364) 100. The method of embodiment 89, wherein the first target region is within 200 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(365) 101. The method of embodiment 89, wherein the first target region is within 200 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(366) 102. The method of embodiment 89, wherein the first target region is within 100 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(367) 103. The method of embodiment 89, wherein the first target region is within 100 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(368) 104. The method of embodiment 89, wherein the TSS is an HBx RNA TSSs.

(369) 105. The method of embodiment 104, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the HBx RNA TSS.

(370) 106. The method of embodiment 105, wherein the HBx RNA TSS is located at a nucleotide within the sequence of nucleotides 1243-1338 of SEQ ID NO: 1082 or nucleotides 1243-1338 of SEQ ID NO: 1083.

(371) 107. The method of embodiment 105, wherein the first target region is within 600 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(372) 108. The method of embodiment 105, wherein the first target region is within 600 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(373) 109. The method of embodiment 105, wherein the first target region is within 500 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(374) 110. The method of embodiment 105, wherein the first target region is within 500 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(375) 111. The method of embodiment 105, wherein the first target region is within 400 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(376) 112. The method of embodiment 105, wherein the first target region is within 400 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(377) 113. The method of embodiment 105, wherein the first target region is within 300 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(378) 114. The method of embodiment 105, wherein the first target region is within 300 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(379) 115. The method of embodiment 105, wherein the first target region is within 200 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(380) 116. The method of embodiment 105, wherein the first target region is within 200 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(381) 117. The method of embodiment 105, wherein the first target region is within 100 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(382) 118. The method of embodiment 105, wherein the first target region is within 100 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(383) 119. The method of embodiment 105, wherein the first target region is within 600 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(384) 120. The method of embodiment 105, wherein the first target region is within 500 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(385) 121. The method of embodiment 105, wherein the first target region is within 500 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(386) 122. The method of embodiment 105, wherein the first target region is within 400 base pairs of

nucleotide 1338 in SEQ ID NO: 1082.

(387) 123. The method of embodiment 105, wherein the first target region is within 400 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(388) 124. The method of embodiment 105, wherein the first target region is within 300 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(389) 125. The method of embodiment 105, wherein the first target region is within 300 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(390) 126. The method of embodiment 105, wherein the first target region is within 200 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(391) 127. The method of embodiment 105, wherein the first target region is within 200 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(392) 128. The method of embodiment 105, wherein the first target region is within 100 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(393) 129. The method of embodiment 105, wherein the first target region is within 100 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(394) 130. The method of any one of embodiments 1-129, wherein the reduction is a reduction in the number of HBV viral episomes.

(395) 131. The method of embodiment 130, wherein the reduction is a reduction in the number of cccDNA genomes.

(396) 132. The method of embodiment 130, wherein the reduction is a reduction in total HBV DNA.

(397) 133. The method of any one of embodiments 1-129, wherein the reduction is a reduction in the replication of the HBV genome.

(398) 134. The method of any one of embodiments 1-129, wherein the reduction is a reduction in a level of expression of a protein product encoded by the HBV genome.

(399) 135. The method of embodiment 130, wherein the reduction is a reduction in a level of HBsAg.

(400) 136. The method of embodiment 130, wherein the reduction is a reduction in a level of HBeAg.

(401) 137. The method of any one of embodiments 1-129, wherein the reduction is a reduction of total HBV DNA of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and wherein the reduction is maintained for at least 14 days after the contacting or the administering.

(402) 138. The method of any one of embodiments 1-129, wherein the reduction is a reduction of HBeAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and wherein the reduction is maintained for at least 14 days after the contacting or the administering.

(403) 139. The method of any one of embodiments 1-129, wherein the reduction is a reduction of HBsAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and wherein the reduction is maintained at or below that level for at least 14 days after the contacting or the administering.

(404) 140. The method of any one of embodiments 137-139, wherein the reduction is a reduction of at least 90%.

(405) 141. The method of any one of embodiments 137-139, wherein the reduction is a reduction of at least 95%.

(406) 142. The method of any one of embodiments 137-139, wherein the reduction is a reduction of at least 99%.

(407) 143. The method of any one of embodiments 137-139, wherein the reduction is a reduction of at least 99.9%.

(408) 144. The method of any one of embodiments 140-143, wherein the reduction is maintained for at least 14 days after the contacting or the administering.

(409) 145. The method of embodiment 144, wherein the reduction is maintained for at least 21 days.

(410) 146. The method of embodiment 144, wherein the reduction is maintained for at least 28 days.

(411) 147. The method of embodiment 144, wherein the reduction is maintained for at least 35 days.

(412) 148. The method of embodiment 144, wherein the reduction is maintained for at least 42 days.

(413) 149. The method of embodiment 144, wherein the reduction is maintained for at least 56 days.

(414) 150. The method of embodiment 144, wherein the reduction is maintained for at least 70 days.



(415) 151. The method of embodiment 144, wherein the reduction is maintained for at least 84 days.

(416) 152. The method of embodiment 144, wherein the reduction is maintained for at least 112 days.

(417) 153. The method of embodiment 144, wherein the reduction is maintained for at least 140 days.

(418) 154. The method of embodiment 144, wherein the reduction is maintained for at least 168 days.

(419) 155. The method of embodiment 144, wherein the reduction is maintained for at least 6 months.

(420) 156. The method of embodiment 144, wherein the reduction is maintained for at least 7 months.

(421) 157. The method of embodiment 144, wherein the reduction is maintained for at least 8 months.

(422) 158. The method of embodiment 144, wherein the reduction is maintained for at least 9 months.

(423) 159. The method of embodiment 144, wherein the reduction is maintained for at least 12 months.

(424) 160. The method of embodiment 144, wherein the reduction is maintained for at least 18 months.

(425) 161. The method of embodiment 144, wherein the reduction is maintained for at least 24 months.

(426) 162. The method of any one of embodiments 1-161, wherein the method does not comprise contacting the HBV gene or genome with a nucleoside or nucleotide analog (NUC) or wherein the method does not comprise administering a NUC to the subject.

(427) 163. The method of any one of embodiments 1-162, wherein the method further comprises contacting the HBV gene or genome with a nucleoside or nucleotide analog (NUC) or wherein the method further comprises administering a NUC to the subject.

(428) 164. The method of any one of embodiments 1-163, wherein the first DNA binding domain comprises a CRISPR-Cas protein.

(429) 165. The method of embodiment 164, wherein the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region.

(430) 166. The method of embodiment 165, wherein the gRNA comprises a sequence selected from a gRNA provided herein, preferably wherein the gRNA comprises a sequence provided in Table 12 or 13.

(431) 167. The method of any one of embodiments 1-164, wherein the first DNA binding domain comprises a zinc-finger protein.

(432) 168. The method of embodiment 167, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1 or Table 18.

(433) 169. The method of embodiment 167 or 168, wherein the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein.

(434) 170. The method of any one of embodiments 1-169, wherein the transcriptional repressor domain comprises ZIM3.

(435) 171. The method of any one of embodiments 1-170, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(436) 172. The method of embodiment 171, wherein the first DNMT domain comprises a sequence of a DNMT domain provided herein.

(437) 173. The method of any one of embodiments 1-172, wherein the epigenetic editing system further comprises a second DNMT domain or a nucleic acid encoding thereof.

(438) 174. The method of embodiment 173, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(439) 175. The method of embodiment 173 or 174, wherein the second DNMT domain comprises a sequence of a DNMT domain provided herein.

(440) 176. The method of any one of embodiments 173-175, wherein the epigenetic editing system comprises a fusion protein or a nucleic acid encoding thereof, and wherein the fusion protein comprises the first DNA binding domain, the first DNMT domain, the repressor domain and the second DNMT domain.

(441) 177. The method of embodiment 176, wherein the fusion protein further comprises a nuclear localization sequence (NLS).

(442) 178. The method of embodiment 177, wherein the fusion protein comprises a sequence of a fusion protein provided herein.

(443) 179. The method of any one of embodiments 1-178, wherein the epigenetic editing system further comprises a second DNA binding domain or a nucleic acid encoding a second DNA binding domain,

wherein the second DNA binding domain binds a second target region of the HBV genome.

(444) 180. The method of embodiment 179, wherein the second target region is a target region recited in any of embodiments 22-129.

(445) 181. The method of embodiment 179 or 180, wherein the second DNA binding domain comprises a CRISPR-Cas protein.

(446) 182. The method of any one of embodiments 1-180, wherein the epigenetic editing system comprises at least one CRISPR-Cas DNA binding domain and at least two different gRNAs.

(447) 183. The method of embodiment 182, wherein the epigenetic editing system comprises a first gRNA binding the first HBV target region and a second gRNA binding a second HBV target region, wherein the first and second target regions are not identical.

(448) 184. The method of embodiment 183, wherein the first gRNA comprises a gRNA sequence provided herein, e.g., a sequence provided in Table 12 or 13, and wherein the second gRNA comprises a different gRNA sequence provided herein, e.g., a sequence provided in Table 12 or 13.

(449) 185. The method of embodiment 179, wherein the second DNA binding domain comprises a zinc-finger protein.

(450) 186. The method of embodiment 185, wherein the zinc-finger protein of the second DNA binding domain comprises a zinc-finger motif with a sequence selected from a zinc finger motif sequence provided herein, e.g., a zinc finger motif provided in Table 1.

(451) 187. The method of embodiment 185 or 186, wherein the zinc-finger protein of the second DNA binding domain comprises a sequence of a zinc finger motif provided in Table 1.

(452) 188. The method of any one of embodiments 179-187, wherein the epigenetic editing system comprises a first fusion protein or a first nucleic acid encoding thereof and a second fusion protein or a second nucleic acid encoding thereof, wherein the first fusion protein comprises the first DNA binding domain and the first DNMT domain, and wherein the second fusion protein comprises the second DNA binding domain and the transcriptional repressor domain.

(453) 189. The method of embodiment 188, wherein the first fusion protein comprises a sequence of a fusion protein provided herein.

(454) 190. The method of embodiment 188 or 189, wherein the second fusion protein comprises a sequence of a fusion protein provided herein.

(455) 191. The method of any one of embodiments 179-190, wherein the epigenetic editing system further comprises a third DNA binding domain or a nucleic acid encoding a third DNA binding domain, wherein the third DNA binding domain binds to a third target region of the HBV genome, optionally, wherein the third DNA binding domain comprises at least one CRISPR-Cas DNA binding domain, optionally wherein the epigenetic editing system comprises a third gRNA comprising a sequence complementary to a strand of a third HBV target region, optionally wherein the third gRNA comprises a gRNA sequence provided herein, optionally, a gRNA sequence provided in Table 12 or 13, optionally, wherein the third DNA binding domain is comprised in a fusion protein comprising a DNMT domain and a transcriptional repressor domain, optionally, wherein the fusion protein is a fusion protein provided herein.

(456) 192. A method, comprising administering an epigenetic editing system to a subject, wherein the subject is characterized by the presence of detectable levels of HBV DNA, HBsAg, and/or HBeAg in the plasma of the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding the same, wherein the first DNA binding domain binds a first target region of an HBV gene or genome, wherein the administering results in a reduction of the level of HBV DNA, the level of HBsAg, and/or the level of HBsAg in the plasma of the subject, wherein the reduction of the level of HBV DNA, of the level of HBsAg, and/or of the level of HBsAg in the plasma of the subject, is at least 90% (a 1-log reduction) compared to the respective level observed or observable in the plasma of the subject prior to the administering, and wherein the 1-log reduction is maintained for at least 14 days after the administering.

(457) 193. The method of embodiment 192, wherein the reduction of the level of HBV DNA in the plasma of the subject is at least 90% (a 1-log reduction).

(458) 194. The method of embodiment 192, wherein the reduction of the level of HBV DNA in the plasma of the subject is at least 99% (a 2-log reduction).

(459) 195. The method of embodiment 192, wherein the reduction of the level of HBsAg in the plasma of the subject is at least 90% (a 1-log reduction).

(460) 196. The method of embodiment 192, wherein the reduction of the level of HBsAg in the plasma of the subject is at least 99% (a 2-log reduction).

(461) 197. The method of embodiment 192, wherein the reduction of the level of HBeAg in the plasma of the subject is at least 90% (a 1-log reduction).

(462) 198. The method of embodiment 192, wherein the reduction of the level of HBeAg in the plasma of the subject is at least 99% (a 2-log reduction).

(463) 199. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 21 days.

(464) 200. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 28 days.

(465) 201. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 35 days.

(466) 202. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 42 days.

(467) 203. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 56 days.

(468) 204. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 70 days.

(469) 205. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 84 days.

(470) 206. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 112 days.

(471) 207. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 140 days.

(472) 208. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 168 days.

(473) 209. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 6 months.

(474) 210. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 9 months.

(475) 211. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 12 months.

(476) 212. The method of any one of embodiments 192-198, wherein the reduction is maintained for at least 24 months.

(477) 213. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype A.

(478) 214. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype B.

(479) 215. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype C.

(480) 216. The method of any one of embodiments 192-212, wherein the HBV genome comprises, HBV genotype D.

(481) 217. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype E.

(482) 218. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype F.

(483) 219. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype G.

(484) 220. The method of any one of embodiments 192-212, wherein the HBV genome comprises HBV genotype H.

(485) 221. The method of any one of embodiments 192-212, wherein the HBV genome comprises a sequence with at least 80%, at least 90%, at least 95%, at least 99%, or greater than 99% sequence identity to an HBV genome sequence provided herein.

(486) 222. The method of any one of embodiments 192-221, wherein the first target region is located in a region of the HBV genome within nucleotides 0-303 of an HBV genome provided herein.

(487) 223. The method of any one of embodiments 192-221, wherein the first target region is located within nucleotides 0-303 of SEQ ID NO: 1082.

(488) 224. The method of any one of embodiments 192-221, wherein the first target region is located within nucleotides 0-303 of SEQ ID NO: 1083.

(489) 225. The method of any one of embodiments 192-221, wherein the first target region is located in a region of the HBV genome within nucleotides 1000-2448 of an HBV genome provided herein.

(490) 226. The method of any one of embodiments 192-221, wherein the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1082.

(491) 227. The method of any one of embodiments 192-221, wherein the first target region is located within nucleotides 1000-2448 of SEQ ID NO: 1083.

(492) 228. The method of any one of embodiments 192-221, wherein the first target region is located in a region of the HBV genome within nucleotides 2802-3182 of an HBV genome provided herein.

(493) 229. The method of any one of embodiments 192-221, wherein the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1082.

(494) 230. The method of any one of embodiments 192-221, wherein the first target region is located within nucleotides 2802-3182 of SEQ ID NO: 1083.

(495) 231. The method of any one of embodiments 192-221, wherein the first target region of the HBV genome is located in an HBV CpG island (CGI).

(496) 232. The method of embodiment 231, wherein the CGI is an HBV canonical CGI.

(497) 233. The method of embodiment 231, wherein the CGI is canonical CGI-I.

(498) 234. The method of embodiment 231, wherein the CGI is canonical CGI-I of HBV genotype D.

(499) 235. The method of embodiment 233, wherein CGI-I spans nucleotides 186-288 of SEQ ID NO: 1082.

(500) 236. The method of embodiment 233, wherein CGI-I spans nucleotides 186-288 of SEQ ID NO: 1083.

(501) 237. The method of embodiment 231, wherein the CGI is canonical CGI-II.

(502) 238. The method of embodiment 231, wherein the CGI is canonical CGI-II HBV genotype D.

(503) 239. The method of embodiment 238, wherein the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1082.

(504) 240. The method of embodiment 238, wherein the CGI is CGI II spans nucleotides 1,217-1,670 of SEQ ID NO: 1083.

(505) 241. The method of embodiment 231, wherein the CGI is canonical CGI-III.

(506) 242. The method of embodiment 231, wherein the CGI is canonical CGI-III HBV genotype D.

(507) 243. The method of embodiment 242, wherein the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1082.

(508) 244. The method of embodiment 242, wherein the CGI is CGI-III spans nucleotides 2,282-2,448 of SEQ ID NO: 1083.

(509) 245. The method of any one of embodiments 192-221, wherein the first target region of the HBV genome is located in a promotor.

(510) 246. The method of embodiment 245, wherein the first target region of the HBV genome is located in the sp1 promotor.

(511) 247. The method of embodiment 245, wherein the first target region of the HBV genome is located in sp2 promotor.

(512) 248. The method of embodiment 245, wherein the first target region of the HBV genome is located in cp promotor.

(513) 249. The method of embodiment 245, wherein the first target region of the HBV genome is located in xp promoter.

(514) 250. The method of any one of embodiments 192-221, wherein the first target region of the HBV genome is located in an enhancer region.

(515) 251. The method of embodiment 250, wherein the first target region of the HBV genome is located in Enh I.

(516) 252. The method of embodiment 250, wherein the first target region of the HBV genome is located in Enh II.

(517) 253. The method of any one of embodiments 192-221, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a transcript.

(518) 254. The method of embodiment 253, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a pgRNA transcript.

(519) 255. The method of embodiment 253, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a preCore RNA transcript.

(520) 256. The method of embodiment 253, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a preS RNA transcript.

(521) 257. The method of embodiment 253, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes an S RNA transcript.

(522) 258. The method of embodiment 253, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes an HBx RNA transcript.

(523) 259. The method of any one of embodiments 192-221, wherein the first target region of the HBV genome is within 1000, 900, 800, 700, 600, 500, 400, 300, 200, or 100 base pairs (bp) of an HBV transcription start site (TSS).

(524) 260. The method of embodiment 259, wherein the TSS is a pg RNA TSS.

(525) 261. The method of embodiment 260, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the pg RNA TSS.

(526) 262. The method of embodiment 260, wherein the pg RNA TSS is located at nucleotide 1820 of SEQ ID NO: 1082 or at nucleotide 1820 of SEQ ID NO: 1083.

(527) 263. The method of embodiment 260, wherein the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(528) 264. The method of embodiment 260, wherein the first target region is within 600 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(529) 265. The method of embodiment 260, wherein the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(530) 266. The method of embodiment 260, wherein the first target region is within 500 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(531) 267. The method of embodiment 260, wherein the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(532) 268. The method of embodiment 260, wherein the first target region is within 400 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(533) 269. The method of embodiment 260, wherein the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(534) 270. The method of embodiment 260, wherein the first target region is within 300 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(535) 271. The method of embodiment 260, wherein the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1082.

(536) 272. The method of embodiment 260, wherein the first target region is within 200 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(537) 273. The method of embodiment 260, wherein the first target region is within 100 base pairs of nucleotide 1820 in SEQ ID NO: 1082 or wherein the first target region is within 100 base pairs of nucleotide 1820 in SEQ ID NO: 1083.

(538) 274. The method of embodiment 259, wherein the TSS is a preC RNA TSS.

(539) 275. The method of embodiment 274, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the preC RNA TSS.

(540) 276. The method of embodiment 274, wherein the preC RNA TSS is located at nucleotide 1791 of SEQ ID NO: 1082 or at nucleotide 1791 of SEQ ID NO: 1083.

(541) 277. The method of embodiment 274, wherein the first target region is within 600 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(542) 278. The method of embodiment 274, wherein the first target region is within 600 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(543) 279. The method of embodiment 274, wherein the first target region is within 500 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(544) 280. The method of embodiment 274, wherein the first target region is within 500 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(545) 281. The method of embodiment 274, wherein the first target region is within 400 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(546) 282. The method of embodiment 274, wherein the first target region is within 400 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(547) 283. The method of embodiment 274, wherein the first target region is within 300 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(548) 284. The method of embodiment 274, wherein the first target region is within 300 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(549) 285. The method of embodiment 274, wherein the first target region is within 200 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(550) 286. The method of embodiment 274, wherein the first target region is within 200 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(551) 287. The method of embodiment 274, wherein the first target region is within 100 base pairs of nucleotide 1791 in SEQ ID NO: 1082.

(552) 288. The method of embodiment 274, wherein the first target region is within 100 base pairs of nucleotide 1791 in SEQ ID NO: 1083.

(553) 289. The method of embodiment 259, wherein the TSS is a preS2 RNA TSS. 290. The method of embodiment 289, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the preS2 RNA TSS. 291. The method of embodiment 289, wherein the preS2 RNA TSS is located at nucleotide 3159 of SEQ ID NO: 1082 or at nucleotide 3159 of SEQ ID NO: 1083.

(554) 292. The method of embodiment 289, wherein the first target region is within 600 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(555) 293. The method of embodiment 289, wherein the first target region is within 600 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(556) 294. The method of embodiment 289, wherein the first target region is within 500 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(557) 295. The method of embodiment 289, wherein the first target region is within 500 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(558) 296. The method of embodiment 289, wherein the first target region is within 400 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(559) 297. The method of embodiment 289, wherein the first target region is within 400 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(560) 298. The method of embodiment 289, wherein the first target region is within 300 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(561) 299. The method of embodiment 289, wherein the first target region is within 300 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(562) 300. The method of embodiment 289, wherein the first target region is within 200 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(563) 301. The method of embodiment 289, wherein the first target region is within 200 base pairs of

nucleotide 3159 in SEQ ID NO: 1083.

(564) 302. The method of embodiment 289, wherein the first target region is within 100 base pairs of nucleotide 3159 in SEQ ID NO: 1082.

(565) 303. The method of embodiment 289, wherein the first target region is within 100 base pairs of nucleotide 3159 in SEQ ID NO: 1083.

(566) 304. The method of embodiment 259, wherein the TSS is an HBx RNA TSSs.

(567) 305. The method of embodiment 304, wherein the first target region is within 600, within 500, within 400, within 300, within 200, or within 100 base pairs of the HBx RNA TSS.

(568) 306. The method of embodiment 304, wherein the HBx RNA TSS is located at a nucleotide within the sequence of nucleotides 1243-1338 of SEQ ID NO: 1082 or nucleotides 1243-1338 of SEQ ID NO: 1083.

(569) 307. The method of embodiment 304, wherein the first target region is within 600 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(570) 308. The method of embodiment 304, wherein the first target region is within 600 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(571) 309. The method of embodiment 304, wherein the first target region is within 500 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(572) 310. The method of embodiment 304, wherein the first target region is within 500 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(573) 311. The method of embodiment 304, wherein the first target region is within 400 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(574) 312. The method of embodiment 304, wherein the first target region is within 400 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(575) 313. The method of embodiment 304, wherein the first target region is within 300 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(576) 314. The method of embodiment 304, wherein the first target region is within 300 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(577) 315. The method of embodiment 304, wherein the first target region is within 200 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(578) 316. The method of embodiment 304, wherein the first target region is within 200 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(579) 317. The method of embodiment 304, wherein the first target region is within 100 base pairs of nucleotide 1243 in SEQ ID NO: 1082.

(580) 318. The method of embodiment 304, wherein the first target region is within 100 base pairs of nucleotide 1243 in SEQ ID NO: 1083.

(581) 319. The method of embodiment 304, wherein the first target region is within 600 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(582) 320. The method of embodiment 304, wherein the first target region is within 500 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(583) 321. The method of embodiment 304, wherein the first target region is within 500 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(584) 322. The method of embodiment 304, wherein the first target region is within 400 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(585) 323. The method of embodiment 304, wherein the first target region is within 400 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(586) 324. The method of embodiment 304, wherein the first target region is within 300 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(587) 325. The method of embodiment 304, wherein the first target region is within 300 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(588) 326. The method of embodiment 304, wherein the first target region is within 200 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(589) 327. The method of embodiment 304, wherein the first target region is within 200 base pairs of

nucleotide 1338 in SEQ ID NO: 1083.

(590) 328. The method of embodiment 304, wherein the first target region is within 100 base pairs of nucleotide 1338 in SEQ ID NO: 1082.

(591) 329. The method of embodiment 304, wherein the first target region is within 100 base pairs of nucleotide 1338 in SEQ ID NO: 1083.

(592) 330. The method of any one of embodiments 192-329, wherein the reduction is a reduction in the number of HBV viral episomes.

(593) 331. The method of embodiment 330, wherein the reduction is a reduction in the number of cccDNA genomes.

(594) 332. The method of embodiment 330, wherein the reduction is a reduction in total HBV DNA.

(595) 333. The method of any one of embodiments 192-329, wherein the reduction is a reduction in the replication of the HBV genome.

(596) 334. The method of any one of embodiments 192-329, wherein the reduction is a reduction in a level of expression of a protein product encoded by the HBV genome.

(597) 335. The method of embodiment 330, wherein the reduction is a reduction in a level of HBsAg.

(598) 336. The method of embodiment 330, wherein the reduction is a reduction in a level of HBeAg.

(599) 337. The method of any one of embodiments 192-329, wherein the reduction is a reduction of total HBV DNA of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and wherein the reduction is maintained for at least 14 days after the contacting or the administering.

(600) 338. The method of any one of embodiments 192-329, wherein the reduction is a reduction of HBeAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and wherein the reduction is maintained for at least 14 days after the contacting or the administering.

(601) 339. The method of any one of embodiments 192-329, wherein the reduction is a reduction of HBsAg of at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or at least 99.9%, and wherein the reduction is maintained at or below that level for at least 14 days after the contacting or the administering.

(602) 340. The method of any one of embodiments 337-339, wherein the reduction is a reduction of at least 90%.

(603) 341. The method of any one of embodiments 337-339, wherein the reduction is a reduction of at least 95%.

(604) 342. The method of any one of embodiments 337-339, wherein the reduction is a reduction of at least 99%.

(605) 343. The method of any one of embodiments 337-339, wherein the reduction is a reduction of at least 99.9%.

(606) 344. The method of any one of embodiments 340-343, wherein the reduction is maintained for at least 14 days after the contacting or the administering.

(607) 345. The method of embodiment 344, wherein the reduction is maintained for at least 21 days.

(608) 346. The method of embodiment 344, wherein the reduction is maintained for at least 28 days.

(609) 347. The method of embodiment 344, wherein the reduction is maintained for at least 35 days.

(610) 348. The method of embodiment 344, wherein the reduction is maintained for at least 42 days.

(611) 349. The method of embodiment 344, wherein the reduction is maintained for at least 56 days.

(612) 350. The method of embodiment 344, wherein the reduction is maintained for at least 70 days.

(613) 351. The method of embodiment 344, wherein the reduction is maintained for at least 84 days.

(614) 352. The method of embodiment 344, wherein the reduction is maintained for at least 112 days.

(615) 353. The method of embodiment 344, wherein the reduction is maintained for at least 140 days.

(616) 354. The method of embodiment 344, wherein the reduction is maintained for at least 168 days.

(617) 355. The method of embodiment 344, wherein the reduction is maintained for at least 6 months.

(618) 356. The method of embodiment 344, wherein the reduction is maintained for at least 7 months.

(619) 357. The method of embodiment 344, wherein the reduction is maintained for at least 8 months.

(620) 358. The method of embodiment 344, wherein the reduction is maintained for at least 9 months.

(621) 359. The method of embodiment 344, wherein the reduction is maintained for at least 12 months.

(622) 360. The method of embodiment 344, wherein the reduction is maintained for at least 18 months.



(623) 361. The method of embodiment 344, wherein the reduction is maintained for at least 24 months.

(624) 362. The method of any one of embodiments 192-361, wherein the method does not comprise contacting the HBV gene or genome with a nucleoside or nucleotide analog (NUC) or wherein the method does not comprise administering a NUC to the subject.

(625) 363. The method of any one of embodiments 192-362, wherein the method further comprises contacting the HBV gene or genome with a nucleoside or nucleotide analog (NUC) or wherein the method further comprises administering a NUC to the subject.

(626) 364. The method of any one of embodiments 192-363, wherein the first DNA binding domain comprises a CRISPR-Cas protein.

(627) 365. The method of embodiment 364, wherein the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region.

(628) 366. The method of embodiment 365, wherein the gRNA comprises a sequence selected from a gRNA provided herein, preferably wherein the gRNA comprises a sequence provided in Table 12 or 13.

(629) 367. The method of any one of embodiments 192-364, wherein the first DNA binding domain comprises a zinc-finger protein.

(630) 368. The method of embodiment 367, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1 or Table 18.

(631) 369. The method of embodiment 367 or 368, wherein the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein.

(632) 370. The method of any one of embodiments 192-369, wherein the transcriptional repressor domain comprises ZIM3.

(633) 371. The method of any one of embodiments 192-370, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(634) 372. The method of embodiment 371, wherein the first DNMT domain comprises a sequence of a DNMT domain provided herein.

(635) 373. The method of any one of embodiments 1-372, wherein the epigenetic editing system comprises the fusion protein provided in SEQ ID NO: 1248 or the fusion protein provided in SEQ ID NO: 1252 and at least one guide RNA, wherein the guide RNA is the guide RNA provided as gRNA #003, gRNA #007, gRNA #008, gRNA #009, gRNA #011, or gRNA #015 herein.

(636) 374. An epigenetic editing system for use in the method of any one of embodiments 1-373, comprising: a fusion protein or a nucleic acid encoding the fusion protein, wherein the fusion protein comprises: (a) a DNA-binding domain that binds a target region of a HBV gene or genome, (b) a first DNA methyltransferase (DNMT) domain, and (c) a transcriptional repressor domain.

(637) 375. The epigenetic editing system of embodiment 374, wherein the fusion protein comprises a sequence of a fusion protein provided herein.

(638) 376. The epigenetic editing system of embodiment 374 or 375, wherein the DNA-binding domain is a CRISPR-Cas DNA binding domain, and wherein the epigenetic editing system comprises at least gRNA provided herein.

(639) 377. The epigenetic editing system of embodiment 374, wherein the epigenetic editing system comprises the fusion protein provided in SEQ ID NO: 1248 or the fusion protein provided in SEQ ID NO: 1252 and at least one guide RNA, wherein the guide RNA is the guide RNA provided as gRNA #003, gRNA #007, gRNA #008, gRNA #009, gRNA #011, or gRNA #015 herein.

(640) 378. An epigenetic editing system comprising: 1. a first fusion protein or a nucleic acid encoding the first fusion protein, wherein the first fusion protein comprises a first DNA binding domain and a first DNMT domain, wherein the first DNA binding domain binds a first target region of a HBV genome, and 2. a second fusion protein or a nucleic acid encoding the second fusion protein, wherein the second fusion protein comprises a second DNA binding domain and a transcriptional repressor domain, wherein the second DNA binding domain binds a second target region of the HBV genome.

(641) 379. The epigenetic system of embodiment 378, wherein the epigenetic editing system is capable of reducing a number of the HBV viral episome, replication of the HBV, or expression of a gene product encoded by the HBV genome, wherein said reduction is at least about 20% compared to contacting the

HBV genome with a suitable control.

(642) 380. The epigenetic system of embodiment 378 or 379, wherein the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA.

(643) 381. The epigenetic system of embodiments 378-380, wherein the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H.

(644) 382. The epigenetic system of embodiments 378-381, wherein the HBV genome comprises a sequence with at least 80% identity to an HBV genome provided herein.

(645) 383. The epigenetic system of embodiments 378-381, further comprising a third fusion protein or a nucleic acid encoding the third fusion protein, wherein the third fusion protein comprises a third DNA binding domain and a second DNMT domain, wherein the third DNA binding domain binds a third target region of the HBV genome.

(646) 384. The epigenetic system of embodiment 383, wherein the first target region, the second target region or the third target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome provided herein.

(647) 385. The epigenetic system of embodiment 383, wherein the first target region, the second target region or the third target region of the HBV genome is located in a CpG island.

(648) 386. The epigenetic system of embodiment 383, wherein the first target region, the second target region or the third target region of the HBV genome is located in a promotor.

(649) 387. The epigenetic system of embodiment 383, wherein the first target region, the second target region or the third target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a precore mRNA, a preS mRNA, a S mRNA, and a X mRNA.

(650) 388. The epigenetic system of embodiment 383, wherein the first DNA binding domain, the second DNA binding domain or the third DNA binding domain comprises a CRISPR-Cas protein.

(651) 389. The epigenetic system of embodiment 388, wherein the epigenetic editing system further comprises a first gRNA that comprises a region complementary to a strand of the first target region, a second gRNA that comprises a region complementary to a strand of the second target region or a third RNA that comprises a region complementary to a strand of the third target region.

(652) 390. The epigenetic system of embodiment 389, wherein the first gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 or 13, the second gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 or 13, and/or the third gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 or 13.

(653) 391. The epigenetic system of embodiment 383, wherein the first DNA binding domain, the second DNA binding domain or the third DNA binding domain comprises a zinc-finger protein.

(654) 392. The epigenetic system of embodiment 391, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein.

(655) 393. The epigenetic system of embodiment 391 or 392, wherein the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1.

(656) 394. The epigenetic system of embodiments 378-393, wherein the transcriptional repressor domain comprises ZIM3.

(657) 395. The epigenetic system of embodiments 378-394, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(658) 396. The epigenetic system of embodiment 395, wherein the first DNMT domain comprises a sequence of a DNMT provided herein.

(659) 397. The epigenetic system of embodiment 383, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(660) 398. The epigenetic system of embodiment 397, wherein the second DNMT domain comprises a sequence of a DNMT domain provided herein.

(661) 399. The epigenetic system of embodiment 378-398, wherein the first fusion protein comprises a sequence of a fusion protein provided herein.

(662) 400. The epigenetic system of embodiments 378-399, wherein the second fusion protein comprises a sequence of a fusion protein provided herein.

(663) 401. The epigenetic system of embodiments 383-399, wherein the third fusion protein comprises a sequence of a fusion protein provided herein.

(664) 402. The method of any one of embodiments 1-401, wherein the epigenetic editing system comprises a nucleic acid sequence provided in Table 18.

(665) LISTING #2 of Exemplary Embodiments:

(666) 1. A method of modifying an epigenetic state of a hepatitis B virus (HBV) gene or genome, comprising contacting the HBV gene or genome with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, and wherein the contacting results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by the HBV gene or genome, wherein the reduction is at least about 20% compared to contacting the HBV gene or genome with a suitable control or without contacting the HBV gene or genome with the epigenetic editing system.

(667) 2. A method of treating an HBV infection in a subject comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, and wherein the administering results in a reduction of: number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by an HBV gene or genome, wherein the reduction is at least about 20% compared to administering a suitable control or without administering the epigenetic editing system.

(668) 3. A method of modulating expression of an HBV gene or genome comprising contacting the HBV gene or genome with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, and wherein the contacting results in a reduction of expression of a gene product encoded by the HBV gene or genome, optionally, wherein the gene product is a nucleic acid or a protein, wherein the reduction is at least about 20% compared to contacting the HBV gene or genome with a suitable control or without contacting the HBV gene or genome with the epigenetic editing system.

(669) 4. A method of inhibiting viral replication in a cell infected with an HBV comprising contacting the cell with an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the epigenetic editing system targets a target region of an HBV gene or genome, and wherein the contacting results in a reduction of number of HBV viral episomes or replication of the HBV gene or genome, wherein the reduction is at least about 20% compared to contacting the cell with a suitable control or without contacting the cell with the epigenetic editing system.

(670) 5. A method of inhibiting viral replication in a subject infected with an HBV comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the epigenetic editing system targets a target region of the HBV gene or genome, and wherein the administering results in a reduction of number of HBV viral episomes, replication of the HBV gene or genome, or expression of a protein product encoded by an HBV gene or genome, wherein the reduction is at least about 20% compared to administering a suitable control or without administering the epigenetic editing system.

(671) 6. The method of embodiment 2 or 5, wherein the reduction is at least about 30%, about 40%, about 50%, about 60% or about 70% compared to administering the suitable control.

(672) 7. The method of any one of embodiments 1, and 3-4, wherein the reduction is at least about 30%, about 40%, about 50%, about 60% or about 70% compared to contacting with the suitable control.

(673) 8. The method of any one of embodiments 1-7, wherein the reduction is maintained for at least 6 days, 19 days, 27 days, 42 days, or 168 days.

(674) 9. The method of embodiment 4, wherein the contacting further results in a reduction of a protein product.

(675) 10. The method of embodiment 5, wherein the administering further results in a reduction of a protein product.

(676) 11. The method of any one of embodiments 1-2 and 9-10, wherein the protein product comprises a HBe antigen.

(677) 12. The method of any one of embodiments 1-2 and 9-10, wherein the protein produce comprises a HBs antigen.

(678) 13. The method of any one of embodiments 1-12, wherein the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA.

(679) 14. The method of any one of embodiments 1-13, wherein the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H.

(680) 15. The method of any one of embodiments 1-14, wherein the HBV genome comprises a sequence with at least 80% identity to an HBV genome sequence provided herein.

(681) 16. The method of embodiment 15, wherein the first target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome provided herein.

(682) 17. The method of any one of embodiments 1-15, wherein the first target region of the HBV genome is located in a CpG island.

(683) 18. The method of any one of embodiments 1-15, wherein the first target region of the HBV genome is located in a promotor.

(684) 19. The method of any one of embodiments 1-15, wherein the first target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a precore mRNA, a preS mRNA, a S mRNA, and a X mRNA.

(685) 20. The method of any one of embodiments 1-19, wherein the first DNA binding domain comprises a CRISPR-Cas protein.

(686) 21. The method of any one of embodiments 1-20, wherein the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region.

(687) 22. The method of embodiment 21, wherein the gRNA comprises a sequence selected from a gRNA provided herein, e.g., in Table 12 and/or 13.

(688) 23. The method of any one of embodiments 1-19, wherein the first DNA binding domain comprises a zinc-finger protein.

(689) 24. The method of embodiment 23, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1 or Table 18.

(690) 25. The method of embodiment 23 or 24, wherein the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein.

(691) 26. The method of any one of embodiments 1-25, wherein the transcriptional repressor domain comprises ZIM3.

(692) 27. The method of any one of embodiments 1-26, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(693) 28. The method of embodiment 27, wherein the first DNMT domain comprises a sequence of a DNMT domain provided herein.

(694) 29. The method of any one of embodiments 1-28, wherein the epigenetic editing system further comprises a second DNMT domain or a nucleic acid encoding thereof.

(695) 30. The method of embodiments 29, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(696) 31. The method of embodiment 30, wherein the second DNMT domain comprises a sequence of a DNMT domain provided herein.

(697) 32. The method of any one of embodiments 29-31, wherein the epigenetic editing system comprises a fusion protein or a nucleic acid encoding thereof, and wherein the fusion protein comprises

the first DNA binding domain, the first DNMT domain, the repressor domain and the second DNMT domain.

(698) 33. The method of embodiment 32, wherein the fusion protein further comprises a nuclear localization sequence (NLS).

(699) 34. The method of embodiment 33, wherein the fusion protein comprises a sequence of a fusion protein provided herein.

(700) 35. The method of any one of embodiments 1-34, wherein the epigenetic editing system further comprises a second DNA binding domain or a nucleic acid encoding thereof, wherein the second DNA binding domain binds a second target region of the HBV genome.

(701) 36. The method of embodiment 35, wherein the second target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182.

(702) 37. The method of embodiment 35, wherein the second target region of the HBV genome is located in a CpG island.

(703) 38. The method of embodiment 35, wherein the second target region of the HBV genome is located in a promotor.

(704) 39. The method of embodiment 35, wherein the second target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a precore mRNA, a preS mRNA, a S mRNA, and a X mRNA.

(705) 40. The method of any one of embodiments 35-39, wherein the second DNA binding domain comprises a CRISPR-Cas protein.

(706) 41. The method of embodiment 40, wherein the epigenetic editing system further comprises a second gRNA that comprises a region complementary to a strand of the second target region.

(707) 42. The method of embodiment 41, wherein the gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., a sequence provided in Table 12 and/or 13.

(708) 43. The method of any one of embodiments 35-39, wherein the second DNA binding domain comprises a zinc-finger protein.

(709) 44. The method of embodiment 43, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif sequence provided herein, e.g., a zinc finger motif provided in Table 1 and/or 18.

(710) 45. The method of embodiment 43 or 44, wherein the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1 and/or 18.

(711) 46. The method of any one of embodiments 35-45, wherein the epigenetic editing system comprises a first fusion protein or a first nucleic acid encoding thereof and a second fusion protein or a second nucleic acid encoding thereof, wherein the first fusion protein comprises the first DNA binding domain and the first DNMT domain, and wherein the second fusion protein comprises the second DNA binding domain and the transcriptional repressor domain.

(712) 47. The method of embodiment 46, wherein the first fusion protein comprises a sequence of a fusion protein provided herein.

(713) 48. The method of embodiment 46, wherein the second fusion protein comprises a sequence of a fusion protein provided herein.

(714) 49. The method of any one of embodiments 46-48, wherein the epigenetic editing system further comprises a third DNA binding domain or a nucleic acid encoding thereof, wherein the third DNA binding domain binds to a third target region of the HBV genome.

(715) 50. The method of embodiment 49, wherein the third target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182.

(716) 51. The method of embodiment 49, wherein the third target region of the HBV genome is located in a CpG island.

(717) 52. The method of embodiment 49, wherein the third target region of the HBV genome is located in a promotor.

(718) 53. The method of embodiment 49, wherein the third target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a precore mRNA, a preS mRNA, a S mRNA, and a X mRNA.

(719) 54. The method of any one of embodiments 49-53, wherein the third DNA binding domain comprises a CRISPR-Cas protein.

(720) 55. The method of embodiment 54, wherein the epigenetic editing system further comprises a third gRNA that comprises a region complementary to a strand of the third target region.

(721) 56. The method of embodiment 55, wherein the third gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., of a gRNA sequence provided in Table 12 and/or 13.

(722) 57. The method of any one of embodiments 49-53, wherein the third DNA binding domain comprises a zinc-finger protein.

(723) 58. The method of embodiment 57, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein.

(724) 59. The method of embodiment 57 or 58, wherein the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1 and/or 18.

(725) 60. The method of any one of embodiments 49-59, wherein the epigenetic editing system further comprises a second DNMT domain or a nucleic acid encoding thereof.

(726) 61. The method of embodiment 60, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(727) 62. The method of embodiment 61, wherein the epigenetic editing system comprises a third fusion protein or a nucleic acid encoding thereof, wherein the third fusion protein comprises the third DNA binding domain and the second DNMT domain.

(728) 63. The method of embodiment 62, wherein the third fusion protein comprises a sequence of a fusion protein provided herein.

(729) 64. An epigenetic editing system comprising: a fusion protein or a nucleic acid encoding the fusion protein, wherein the fusion protein comprises: (a) a DNA-binding domain that binds a target region of a HBV gene or genome, (b) a first DNA methyltransferase (DNMT) domain, and (c) a transcriptional repressor domain.

(730) 65. The epigenetic system of embodiment 64, wherein the epigenetic editing system is capable of reducing a number of the HBV viral episome, replication of the HBV, or expression of a gene product encoded by the HBV gene or genome, wherein said reduction is at least about 20% compared to contacting the HBV gene or genome with a suitable control.

(731) 66. The epigenetic system of embodiment 64 or 65, wherein the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA.

(732) 67. The epigenetic system of any one of embodiments 64-66, wherein the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H.

(733) 68. The epigenetic system of any one of embodiments 64-67, wherein the HBV genome comprises a sequence with at least 80% identity to an HBV genome sequence provided herein.

(734) 69. The epigenetic system of any one of embodiments 64-68, wherein the target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome sequence provided herein.

(735) 70. The epigenetic system of any one of embodiments 64-68, wherein the target region of the HBV genome is located in a CpG island.

(736) 71. The epigenetic system of any one of embodiments 63-68, wherein the target region of the HBV genome is located in a promotor.

(737) 72. The epigenetic system of any one of embodiments 63-68, wherein the target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a precore mRNA, a preS mRNA, a S mRNA, and a X mRNA.

(738) 73. The epigenetic system of embodiments 63-72, wherein the DNA binding domain comprises a CRISPR-Cas protein.

(739) 74. The epigenetic system of embodiment 73, wherein the epigenetic editing system further comprises a gRNA that comprises a region complementary to a strand of the target region.

(740) 75. The epigenetic system of embodiment 74, wherein the gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., in Table 12 and/or 13.

(741) 76. The epigenetic system of any one of embodiments 63-72, wherein the DNA binding domain comprises a zinc-finger protein.

(742) 77. The epigenetic system of embodiment 76, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein.

(743) 78. The epigenetic system of embodiment 76 or 77, wherein the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1 and/or 18.

(744) 79. The epigenetic system of any one of embodiments 63-78, wherein the transcriptional repressor domain comprises a sequence of a transcriptional repressor provided herein.

(745) 80. The epigenetic system of any one of embodiments 63-79, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(746) 81. The epigenetic system of embodiment 80, wherein the DNMT domain comprises a sequence of a DNMT domain provided herein.

(747) 82. The epigenetic system of any one of embodiments 63-81, wherein the fusion protein further comprises a second DNMT domain.

(748) 83. The epigenetic system of embodiment 82, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(749) 84. The epigenetic system of any one of embodiments 63-83, wherein the fusion protein further comprises a nuclear localization sequence (NLS).

(750) 85. The epigenetic system of embodiment 84, wherein the fusion protein comprises a sequence of a fusion protein provided herein.

(751) 86. An epigenetic editing system comprising: a first fusion protein or a nucleic acid encoding the first fusion protein, wherein the first fusion protein comprises a first DNA binding domain and a first DNMT domain, wherein the first DNA binding domain binds a first target region of a HBV genome, and a second fusion protein or a nucleic acid encoding the second fusion protein, wherein the second fusion protein comprises a second DNA binding domain and a transcriptional repressor domain, wherein the second DNA binding domain binds a second target region of the HBV genome.

(752) 87. The epigenetic system of embodiment 86, wherein the epigenetic editing system is capable of reducing a number of the HBV viral episome, replication of the HBV, or expression of a gene product encoded by the HBV genome, wherein said reduction is at least about 20% compared to contacting the HBV genome with a suitable control.

(753) 88. The epigenetic system of embodiment 86 or 87, wherein the HBV genome is a covalently closed circular DNA (cccDNA) or an HBV integrated DNA.

(754) 89. The epigenetic system of any one of embodiments 86-88, wherein the HBV genome comprises HBV genotype A, HBV genotype B, HBV genotype C, HBV genotype D, HBV genotype E, HBV genotype F, HBV genotype G or HBV genotype H.

(755) 90. The epigenetic system of any one of embodiments 86-89, wherein the HBV genome comprises a sequence with at least 80% identity to an HBV genome provided herein.

(756) 91. The epigenetic system of any one of embodiments 86-89, further comprising a third fusion protein or a nucleic acid encoding the third fusion protein, wherein the third fusion protein comprises a third DNA binding domain and a second DNMT domain, wherein the third DNA binding domain binds a third target region of the HBV genome.

(757) 92. The epigenetic system of embodiment 91, wherein the first target region, the second target region or the third target region is located in a region of the HBV genome within nucleotide 0-303, 1000-2448 or 2802-3182 of an HBV genome provided herein.

(758) 93. The epigenetic system of embodiment 91, wherein the first target region, the second target region or the third target region of the HBV genome is located in a CpG island.

(759) 94. The epigenetic system of embodiment 91, wherein the first target region, the second target region or the third target region of the HBV genome is located in a promotor.

(760) 95. The epigenetic system of embodiment 91, wherein the first target region, the second target region or the third target region of the HBV genome is located in a section of the HBV genome that encodes a transcript selected from the group consisting of a pgRNA, a precore mRNA, a preS mRNA, a S mRNA, and a X mRNA.

(761) 96. The epigenetic system of embodiment 91, wherein the first DNA binding domain, the second DNA binding domain or the third DNA binding domain comprises a CRISPR-Cas protein.

(762) 97. The epigenetic system of embodiment 96, wherein the epigenetic editing system further comprises a first gRNA that comprises a region complementary to a strand of the first target region, a second gRNA that comprises a region complementary to a strand of the second target region or a third RNA that comprises a region complementary to a strand of the third target region.

(763) 98. The epigenetic system of embodiment 97, wherein the first gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 and/or 13, the second gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 and/or 13, and/or the third gRNA comprises a sequence selected from a gRNA sequence provided herein, e.g., provided in Table 12 and/or 13.

(764) 99. The epigenetic system of embodiment 91, wherein the first DNA binding domain, the second DNA binding domain or the third DNA binding domain comprises a zinc-finger protein.

(765) 100. The epigenetic system of embodiment 99, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from a zinc finger motif provided herein.

(766) 101. The epigenetic system of embodiment 99 or 100, wherein the zinc-finger protein comprises a sequence of a zinc finger motif provided in Table 1 and/or 18.

(767) 102. The epigenetic system of any one of embodiments 86-101, wherein the transcriptional repressor domain comprises ZIM3.

(768) 103. The epigenetic system of any one of embodiments 86-102, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(769) 104. The epigenetic system of embodiment 103, wherein the first DNMT domain comprises a sequence of a DNMT provided herein.

(770) 105. The epigenetic system of embodiment 91, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(771) 106. The epigenetic system of embodiment 105, wherein the second DNMT domain comprises a sequence of a DNMT domain provided herein.

(772) 107. The epigenetic system of any one of embodiment 86-106, wherein the first fusion protein comprises a sequence of a fusion protein provided herein.

(773) 108. The epigenetic system of any one of embodiments 86-107, wherein the second fusion protein comprises a sequence of a fusion protein provided herein.

(774) 109. The epigenetic system of any one of embodiments 91-107, wherein the third fusion protein comprises a sequence of a fusion protein provided herein.

(775) 110. The method of any one of embodiments 1-63, wherein the epigenetic editing system comprises a nucleic acid sequence provided in Table 18.

(776) 111. A method of treating an HDV infection in a subject comprising administering an epigenetic editing system to the subject, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the contacting results in a reduction of: number of HDV viral episomes, replication of the HDV gene or genome, or expression of a protein product encoded by the HDV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control.

(777) 112. A method of inhibiting viral replication in a cell infected with an HDV comprising administering an epigenetic editing system, wherein the epigenetic editing system comprises a first DNA binding domain, a first DNMT domain, and a transcriptional repressor domain or one or more nucleic acid molecules encoding thereof, wherein the first DNA binding domain binds a first target region of a HBV gene or genome, and wherein the epigenetic editing system targets a target region of the HBV gene or genome, and wherein the contacting results in a reduction of number of HDV viral episomes or replication of the HDV gene or genome, wherein said reduction is at least about 20% compared to administering a suitable control.

(778) 113. The method of embodiment 111 or 112, wherein the first DNA binding domain comprises a CRISPR-Cas protein.



(779) 114. The method of embodiment 113, wherein the epigenetic editing system further comprises a first guide RNA (gRNA) that comprises a region complementary to a strand of the first target region.

(780) 115. The method of embodiment 114, wherein the gRNA comprises a sequence selected from a gRNA provided herein, e.g., in Table 12 and/or 13.

(781) 116. The method of embodiment 111 or 112, wherein the first DNA binding domain comprises a zinc-finger protein.

(782) 117. The method of embodiment 116, wherein the zinc-finger protein comprises a zinc-finger motif with a sequence selected from any zinc finger or zinc finger motif provided herein, e.g., in Table 1 and/or 18.

(783) 118. The method of embodiment 116 or 117, wherein the zinc-finger protein comprises a sequence of any of the zinc finger epigenetic repressors provided herein.

(784) 119. The method of any one of embodiments 111-118, wherein the transcriptional repressor domain comprises ZIM3.

(785) 120. The method of any one of embodiments 111-119, wherein the first DNMT domain is a DNMT3A domain or a DNMT3L domain.

(786) 121. The method of embodiment 120, wherein the first DNMT domain comprises a sequence of a DNMT domain provided herein.

(787) 122. The method of any one of embodiments 111-121, wherein the epigenetic editing system further comprises a second DNMT domain or a nucleic acid encoding thereof.

(788) 123. The method of embodiment 122, wherein the second DNMT domain is a DNMT3A domain or a DNMT3L domain.

(789) 124. The method of embodiment 123, wherein the second DNMT domain comprises a sequence of a DNMT domain provided herein.

(790) 125. The method of any one of embodiments 122-123, wherein the epigenetic editing system comprises a fusion protein or a nucleic acid encoding thereof, and wherein the fusion protein comprises the first DNA binding domain, the first DNMT domain, the repressor domain and the second DNMT domain.

(791) 126. The method of embodiment 125, wherein the fusion protein further comprises a nuclear localization sequence (NLS).

(792) 127. The method of embodiment 126, wherein the fusion protein comprises a sequence of a fusion protein provided herein.

(793) 128. The method of any one of embodiments 111-127, wherein the first DNA binding domain binds a target region of an HBV gene or genome encoding or controlling expression of an S-antigen.

(794) In order that the present disclosure may be better understood, the following examples are set forth. These examples are for purposes of illustration only and are not to be construed as limiting the scope of the present disclosure in any manner.

## EXAMPLES

### Example 1: Selection of Target HBV Sequences for Epigenetic Silencing

(795) Target sequences were manually and computationally designed using the representative HBV genome sequences (SEQ ID Nos. 1082, 1083) as a reference:

(796) While target site design focused on CpG islands identified within the HBV genome, target sites outside of HBV CpG islands were also considered.

(797) Table 2 presents some representative target sites that were identified as suitable for targeting with an epigenetic repressor.

(798) Target domains identified above that are adjacent to a PAM sequence, e.g., an *S. pyogenes* Cas9 PAM sequence, can be targeted by a CRISPR-based epigenetic repressor, e.g., an epigenetic repressor comprising a dCas9 DNA-binding domain. For example, target sites 1-143 are suitable for dCas9-based epigenetic repressor targeting. FIG. 1 provides an overview over the position of the target sites identified in the HBV genome.

(799) Target sites were analyzed for conservation across HBV genotypes A-E (FIGS. 2 and 3). Some target sites were identified that were well conserved across two or more, or in some cases all, HBV genotypes. Targeting such conserved sites allows for silencing different genotypes with the same

epigenetic repressor.

#### Example 2: Guide RNA Assays in HepAD38 HBV Cells

(800) The HepAD38 cell line expresses the HBV genome under a doxycycline-inducible promoter (see, e.g., Ladner et al., Inducible expression of human hepatitis B virus (HBV) in stably transfected hepatoblastoma cells: a novel system for screening potential inhibitors of HBV replication. *Antimicrob. Agents Chemother.* 41:1715-1720(1997), incorporated herein by reference).

(801) Results are shown in FIGS. 4A and B.

#### Example 3: Guide RNA Assays in HepG2-NTCP Cells

(802) HepG2 cells were engineered by lentiviral transduction to express the human NTCP receptor which is used by hepatitis B virus (HBV) to infect the cells.

(803) HBV viral particles were produced using the HepAD38 cell line. HepAD38 is a subclone, derived from HepG2 cell line, that expresses HBV genome (genotype D subtype ayw) under the transcriptional control of a tetracycline-responsive promoter in a TET-OFF system.

(804) A triple combination of Engineered Transcriptional Repressors (ETRs) consisting of three plasmids expressing dCas9-KRAB, dCas9-DNMT3A and dCas9-DNMT3L was used in combination with one or more of the designed sgRNAs.

(805) LNPs were formulated using GENVOY ILM Lipid Mix (Precision Nanosystem) and the formulator Nanoassembler Spark (Precision Nanosystem). LNPs were formulated according to the manufacturer's recommendations with Nitrogen:Phosphate (NP) ratio equal to 6 and flow rate ratio (FRR) 2:1. The RNA payload was diluted to a final concentration of 350 ng/uL in the PNI formulation buffer. The ETRs, dCas9-KRAB, dCas9-DNMT3A, dCas9-DNMT3L and each of the 121 sgRNA were mixed at 1:1:1:4 ratio. The RNA mix, the Genvoy lipid mix (25 mM) and PBS were loaded each in the dedicated chambers of the Spark cartridge and formulated. The quality of the formulated LNPs was evaluated quantifying the packaged mRNA using Quant-it™ RiboGreen RNA Assay Kit (Thermo Fisher) and sizing the LNP by Dynamic Light Scattering (Zetasizer, Malvern Panalytic).

(806) HepG2-NTCP cells were plated at 20,000 cells/well in collagen coated 96 well plates. After 24 h cells were infected with HBV at 5,000 multiplicity of genome equivalent (MGE) and 16 h after viral inoculum was removed, cells were washed with PBS, and fresh media was added. Three days post-infection, using LNPs, each sgRNA and the mRNAs encoding each of the components of the triple constructs of ETRs (dCas9-KRAB, dCas9-DNMT3A, dCas9-DNMT3L) were delivered. Three days after, LNP was removed, medium was replaced, and cells were maintained in complete medium for three days.

(807) Viral antigens HBeAg and HBsAg were quantified 6 days after LNP removal using ELISA assays. Data were normalized to a non-targeting guide designed against the mouse PCSK9 and control 3.2 gRNA was used as positive control. Cells viability assay were performed and normalized to non-targeting control.

(808) The Table below provides amino acid sequences of exemplary epigenetic editors used in the gRNA screen (the ETR constructs):

(809) TABLE-US-00013 TABLE 6 amino acid sequences of exemplary epigenetic editors  
SEQ ID NO Description Amino acid sequence 476 dCas9:G:KRAB

MYPYDVPDYASPKKKRKVEASDKKYSIGLAIGTNSVGWAVITDEYKVPSKKEK  
VLGNTDRHSIKKNLIGALLEDGETAEATRLKRTARRRYTRRKNRICYLQEIF  
SNEMAKVDDSFHRLSEFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLR  
KKLVDSTDKADLRLIYLALAHMIKERGHFLIEGDLNPDNSDVKLFIQLVQTY  
NQLFEENPINASGVDAKAILSARLSKSRRLNLIQPLGEEKKNGLFGNLIALS  
LGLTPNFKSNEDLAEDAKLQLSKDTYDDDLDNLLAQIGDQYADLELAAKNLS  
AILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPKEYKEIF  
FDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKORT  
FDNGSIPHQIHLGELHAILRRQEDFYFPLKDNREKIEKILTRIPYYVGPLAR  
GNSRFAWMTRKSEETITPWNFEVVDKGASAQSFIERMTNEDKNLPNEKVLPK  
HSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLFKTNRKVTQKQ  
LKEDYFKKIECFDSVEISGVEDRENASLGTYHDLLKIIKDKDELNEENEDIL

EDIVLTTLTFEDREMIEERLKTYAHLEDDKVMKQLKRRRYTGWGRLSRKLING  
IRDKQSGKTILDFLKSDGFANRNFMQLIHDDSLTFKEDIQKAQVSGQGDSLHE  
HIANLAGSPAIIKKGILQTVKVVDDELVKVMGRHKPENIVIAMARENQTTQKGQK  
NSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLQNGRDMYVDQEL  
DINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKMN  
YWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQI  
LDSRMNTKYDENDKLIREVKVITLKSCLVSDERKDFQFYKVVREINNYHHAHDA  
YLNNAVVGTAIIKKYPKLESEFVYGDYKVYDVVRKMIKSEQEIGKATAKYFFYS  
NIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVN  
IVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVV  
AKVEKGKSKKLKSVKELLGITIMERSSSFENPIDFLEAKGYKEVKKDLIIKLP  
KYSLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDN  
EQKQLFVEQHKHYLDEIIIEQISEFSKRVLADANLDKVL SAYNKH RDKPIREQ  
AENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ SITGLYET  
RIDLSQLGGDSPKKKRKVGV D **GSGGG**ALSPQHSAVTQGSIIKNKEGMDAKSLT  
AWSRTLVTFKDVFVDETREEWKLLDTAQQIVYRNVMLENYKNLVSLGYQLTKP  
DVILRLEKGEEPWLVEREIHQETHPDSETAFEIKSSV\* YPYDVPDYA - HA-Tag (SEQ ID  
NO: 479) **GSGGG** - Linker (SEQ ID NO: 480) 477 dCas9:G:DNMT3A  
MYPYDVPDYASPKKKRKVEASDKKYSIGLAIGTNSVGWAVITDEYKVPSKKFK  
VLGNTDRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRKNRICYLQEIF  
SNEMAKVDDSFHRL EESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLR  
KKLVDSTDKADLR LIYLALAHMIKFRGHFLIEGDLNPDNSDV DKLFIQLVQTY  
NQLFEENPINASGVDAKAILSARLSKSRRENLI AQLPGEKKNGLEGNLIALS  
LGLTPNFKSNFDLAEDAKLQLSKDTYDDDLDNLLAQIGDQYADLFLAAKNLSD  
AILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVROQLPEKYKEIF  
FDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKORT  
FDNGSIPHQIHLGELHAILRRQEDFY PFLKDNREKIEKILTFRIPYYVGPLAR  
GNSRFAWMTRKSEETITPWNFE EVVDKGASAQSFIERMTNEDKNLPNEKVLPK  
HSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLEKTNRKVTVKQ  
LKEDYFKKIECFDSVEISGVEDRENASLGTYHDLLKIIKDKDEL DNEENEDIL  
EDIVLTTLTFEDREMIEERLKTYAHLEDDKVMKQLKRRRYTGWGRLSRKLING  
IRDKQSGKTILDELKSDGFANRNFMQLIHDDSLTFKEDIQKAQVSGQGDSLHE  
HIANLAGSPAIIKKGILQTVKVVDDELVKVMGRHKPENIVIAMARENQTTQKGQK  
NSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYYLONGRDMYVDQEL  
DINRLSDYDVDAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKMN  
YWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQI  
LDSRMNTKYDENDKLIREVKVITLKSCLVSDFRKDFQFYKVVREINNYHHAHDA  
YLNNAVVGTAIIKKYPKLESEFVYGDYKVYDVVRKMIKSEQEIGKATAKYFFYS  
NIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVN  
IVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGEDSPTVAYSVLVV  
AKVEKGKSKKLKSVKELLGITIMERSSSFENPIDFLEAKGYKEVKKDLIIKLP  
KYSLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPEDN  
EQKOLFVEQHKHYLDEIIIEQISEFSKRVLADANLDKVL SAYNKH RDKPIREQ  
AENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ SITGLYET  
RIDLSQLGGDSPKKKRKVGV D **GSGGG**TYGLLRREDWPSRLQMFFANNHDQEF  
DPPKVYPPVPAEK RKPIRVLSLEDGIATGLLV LKDLGIQVDRIYASEVCEDSI  
TVGMVRHQGKIMYVGDVRSVTQKHIQEWGPFDLVIGGSPCNDLSIVNPARKGL  
YEGTGRLFFEFYRLLHDARPKEGDDRPFFWLFENVVAMGVSDKRDISR FLESN  
PVMIDAKEVSA AHRARYFWGNLPGMNRPLASTVNDKLELQECLEHGRIAKESK  
VRTITTRSNSIKQGKDQHFPVFMNEKEDILWCTEMERVFGFPVHYTDVSNMSR  
LARQRLLGRSWSVPVIRHLFAPLKEYFACV\* YPYDVPDYA - HA-Tag (SEQ ID NO:  
479) **GSGGG** - Linker (SEQ ID NO: 480) 478 dCas9:G:hDNMT3L

MYPYDVPDYAIPKPKRKVEASDKKYSIGLAIGTNSVGWAVITDEYKVPSPKKEK  
 VLGNTRHSIKKNLIGALLFDSGETAEATRLKRTARRRYTRRKNRICYLQEIF  
 SNEMAKVDDSFHRLSEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLR  
 KKLVDSTDKADLRLIYLALAHMIKFRGHFLIEGDLNPDNSDVKLFIQLVQTY  
 NQLFEENPINASGVDAKAILSARLSKSRRLLENLIAQLPGEKKNGLGNLIALS  
 LGLTPNFKSNEDLAEDAKLQLSKDTYDDDLNLLAQIGDQYADLFLAAKNLSD  
 AILLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPKEYKEIF  
 FDQSKNGYAGYIDGGASQEEFYKFIKPILEKMDGTEELLVKLNREDLLRKORT  
 FDNGSIPHQIHLGELHAILRRQEDFYPELKDNREKIEKILTFRIPYYVGPLAR  
 GNSRFAWMTRKSEETITPWNFEEVVDKGASASQSFIERMTNEDKNLPNEKVLPK  
 HSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVDLLEKTNRKVTVKQ  
 LKEDYFKKIECFDSVEISGVEDRENASLGTYHDLLKIIKDKDELNEENEDIL  
 EDIVLTTLTFEDREMIEERLKYAHLEDDKVMKQLKRRRYTGWGRLSRKLING  
 IRDKQSGKTILDELKSDGFANRNFQMQLIHDDSLTFKEDIQKAQVSGQGDSLHE  
 HIANLAGSPAIIKKGILQTVKVVDELVKVMGRHKPENIVIEMARENQTTQKGQK  
 NSRERMKRIEEGIKELGSQILKEHPVENTQLQNEKLYLYLQNGRDMYVDQEL  
 DINRLSDYDVAIVPQSFLKDDSIDNKVLTRSDKNRGKSDNVPSEEVVKMKMN  
 YWRQLLNAKLITQRKFDNLTKAERGGLSELDKAGFIKRQLVETRQITKHVAQI  
 LDSRMNTKYDENDKLIREVKVITLKSCLVSDERKDFQFYKVVREINNYHHAHDA  
 YLNAVVGTAIIKKYPKLESEFVYGDYKVYDVRKMIKSEQEIGKATAKYFFYS  
 NIMNFFKTEITLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVL SMPQVN  
 IVKKTEVQTGGFSKESILPKRNSDKLIARKKDWDPKKYGGEDSPTVAYSVLV  
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 KYSLFELENGRKRMLASAGELQKGNELALPSKYVNFYLYLASHYEKLKGS PEDN  
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 RIDLSQLGGDSPKKKRKVGV D**GSGGG**MAAIPALDPEAEPSMDVILVGSSELSS  
 SVSPGTGRDLIAYEVKANQRNIEDICICCGSLQVHTQHPLFEGGICAPCKDKF  
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 VHAMS NWVCYLCLPSSRSGLLQRRRKWRSQ LKAFYDRESENPLEMFETVPVWR  
 RQPVRVLSLFEDIKKELTSLGFLESGSDPGQLKHVVDVTDTVRKDVEEWGPED  
 LVYGATPPLGHTCDRPPSWYLFQFHRL LQYARPKPGSPRPFFWMFVDNLVLNK  
 EDLDVASR FLEM EPVTIPDVHGGSLQNAV RVWSNIPAIRSRHWALVSEEELSL  
 LAQNKQSSKLA AKWPTKL VKNCELPLREYFKYFSTELTSSL\* YPYDVPDYA - HA-Tag  
 (SEQ ID NO: 479) **GSGGG** - Linker (SEQ ID NO: 480) 479 HA-Tag YPYDVPDYA  
 480 linker GSGGG

(810) The Table below provides amino acid sequences and polynucleotide sequences of exemplary epigenetic editors

(811) TABLE-US-00014 TABLE 7 sequences of exemplary epigenetic editors

SEQ ID	NO	Description	Sequence
481	PLA001	amino acid	MPKKKRRKVPKKKRRKVYNHDQEFDPKVPVPAEKRRKPIRVLSLEDGIATG

LLVLKDLGIQVDRYIASEVCEDSITVGMVRHQGKIMYVG D VRSVTQKHIQE  
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 RPFFWLFENVVAMGVSDKRDISRFLSNPVMIDAKEVSA AHRARYFWGNLP  
 GMNRPLASTVNDKLELQECLEHGRIAKFSKVRTITTRSNSIKQGKDQHFPV  
 FMNEKEDILWCTEMERVFGFPVHYTDVSNMSRLARQRL LGRSWSVPVIRHL  
 FAPLKEYFACVSSGNSNANSRGPSESSGLVPLSLRGSHMAAIPALDPEAEP  
 SMDVILVGSSELSSSVSPGTGRDLIAYEVKANQRNIEDICICCGSLQVHTQ  
 HPLFEGGICAPCKDKFLDALFLYDDDGYQSYCSICCSGETLLICGNPDCTR  
 CYCFECVDSL VGPGTSGKVHAMS NWVCYLCLPSSRSGLLQRRRKWRSQ LKA  
 FYDRESENPLEMFETVPVWRRQPVRVLSLFEDIKKELTSLGFLESGSDPGQ  
 LKHVVDVTDTVRKDVEEWGPFDLVYGATPPLGHTCDRPPSWYLFQFHRL LQ

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CTGCTGGTGCTGAAGGATCTGGGCATCCAGGTGGACCGGTACATCGCCTCC  
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GAGTACTTCACCGTGTATAACGAGCTGACCAAAGTGAAATACGTGACCGAG  
GGAATGAGAAAAGCCCGCCTTCTGAGCGGCGAGCAGAAAAAGGCCATCGTG  
GACCTGCTGTTCAAGACCAACCGGAAAAGTGACCGTGAAGCAGCTGAAAGAG  
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CGGCTGAAAACCTATGCCACCTGTTCGACGACAAAGTGATGAAGCAGCTG  
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GACGGCTTCGCCAACAGAACTTCATGCAGCTGATCCACGACGACAGCCTG  
ACCTTTAAAGAGGACATCCAGAAAGCCCAGGTGTCCGGCCAGGGCGATAGC  
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CAGCTGCAGAACGAGAAGCTGTACCTGTACTACCTGCAGAATGGGCGGGAT  
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GAAGAGGTCGTGAAGAAGATGAAGAACTACTGGCGGCAGCTGCTGAACGCC  
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CCCAAGAAGAAAAGGAAGGTCCCAAAGAAAAAAGAAAGGTGTGA

(812) Table 8 below lists components of the fusion polypeptide PLA001 and their corresponding amino acid position in the fusion polypeptide sequence (SEQ ID No. 481) set forth in Table 7.

(813) TABLE-US-00015 TABLE 8 annotation of PLA001 amino acid sequence Type Start End Length  
SV40 NLS CDS 2 8 7 SV40 NLS CDS 9 15 7 DNMT3A CDS 17 317 301 Linker CDS 318 344 27  
DNMT3L full- CDS 345 730 386 length XTEN80 CDS 731 810 80 dCas9 CDS 811 2180 1370 NLS  
CDS 2181 2187 7 XTEN16 CDS 2188 2208 21 ZN627 CDS 2211 2290 80 FLAG CDS 2293 2300 8  
SV40 NLS CDS 2302 2308 7 SV40 NLS CDS 2309 2315 7

(814) Table 9 below lists components of the polynucleotide encoding the fusion polypeptide PLA001 and their corresponding nucleotide position in the polynucleotide sequence (SEQ ID No. 482) set forth in Table 7.

(815) TABLE-US-00016 TABLE 9 annotation of PLA001 polynucleotide sequence Name Type  
Minimum Maximum Length SV40 NLS CDS 4 24 21 SV40 NLS CDS 25 44 20 DNMT3A CDS 49 951  
903 Linker CDS 952 1032 81 DNMT3L full- CDS 1033 2190 1158 length XTEN80 CDS 2191 2430  
240 dCas9 CDS 2431 6540 4110 NLS CDS 6541 6561 21 XTEN16 CDS 6562 6624 63 ZN627 CDS  
6631 6870 240 FLAG CDS 6877 6900 24 SV40 NLS CDS 6904 6924 21 SV40 NLS CDS 6925 6945  
21

(816) Table 10 below lists components of the fusion polypeptide PLA002 and their corresponding amino acid position in the fusion polypeptide sequence (SEQ ID No. 483) set forth in Table 7.

(817) TABLE-US-00017 TABLE 10 annotation of PLA002 amino acid sequence Name Type Minimum  
Maximum Length SV40 NLS CDS 2 8 7 SV40 NLS CDS 9 15 7 DNMT3A CDS 17 317 301 Linker  
CDS 318 344 27 DNMT3L full- CDS 345 730 386 length XTEN80 CDS 731 810 80 dCas9 CDS 811  
2180 1370 NLS CDS 2181 2187 7 XTEN16 CDS 2188 2208 21 ZIM3 CDS 2211 2310 100 FLAG CDS  
2313 2320 8 SV40 NLS CDS 2322 2328 7 SV40 NLS CDS 2329 2335 7

(818) Table 11 below lists components of the polynucleotide encoding the fusion polypeptide PLA002 and their corresponding nucleotide position in the polynucleotide sequence (SEQ ID No. 484) set forth in Table 7.

(819) TABLE-US-00018 TABLE 11 annotation of PLA002 polynucleotide sequence Name Type

Minimum Maximum Length SV40 NLS CDS 4 24 21 SV40 NLS CDS 25 45 21 DNMT3A CDS 49 951  
903 Linker CDS 952 1032 81 DNMT3L full- CDS 1033 2190 1158 length XTEN80 CDS 2191 2430  
240 dCas9 CDS 2431 6540 4110 NLS CDS 6541 6561 21 XTEN16 CDS 6562 6624 63 ZIM3 CDS  
6631 6930 300 FLAG CDS 6937 6960 24 SV40 NLS CDS 6964 6984 21 SV40 NLS CDS 6985 7005  
21 stop terminator 7006 7008 3  
(820) Table 12 below provides gRNA sequence tested.  
(821) TABLE-US-00019 TABLE 12 Exemplary gRNA sequences Target SEQ domain SEQ IDs  
sequence IDs gRNA sequence 333 CCTGCTGGTG 1093  
CCUGCUGGUGGCUCCAGUUCGUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAU  
GCTCCAGTTC  
AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 334  
CTGAACTGGA 1094  
CUGAACUGGAGCCACCAGCAGUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAU  
GCCACCAGCA  
AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 335  
CCTGAACTGG 1095  
CCUGAACUGGAGCCACCAGCGUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAU  
AGCCACCAGC  
AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 336  
CCTCGAGAAG 1096  
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TCTCGAGGAT  
AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 338  
CGTCAATCTT 1098  
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GUCAAUCUUCUCGAGGAUUGGUUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAU  
TCGAGGATTG  
AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 340  
AACATGGAGA 1100  
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CGCAGAGTCT 1104

CGCAGAGACUCGUGGGUUUAAGAGCUAAGCUGGAAACAGCAUAGCAAGUUUAAAU  
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AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 345  
CACCACGAGT 1105  
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TGGACTTCTC 1106  
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ACGTAAACAA  
AAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUUUU 405  
TGCCGTTCCG 1165  
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ACCGACACG  
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CGATTGGTGG 1234

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(822) TABLE-US-00020 TABLE 13 Exemplary target domain sequences and effect on  
HbeAg and HbsAg expression guide RNA Associated HbeAg HbsAg guide RNA  
(%expression of (%expression of SEQ name (if Target domain non targeting non targeting  
IDs applicable) sequence control) control) 334 gRNA#001 CTGAACTGGAGCCACCAGCA  
27.77203753 23.4507853 335 gRNA#002 CCTGAACTGGAGCCACCAGC 41.3794605 42.3814023  
333 CCTGCTGGTGGCTCCAGTTC 65.36067834 43.2303179 336 CCTCGAGAAGATTGACGATA  
82.8943107 72.648219 337 TCGTCAATCTTCTCGAGGAT 45.82985382 59.7223204 338  
CGTCAATCTTCTCGAGGATT 70.38176383 73.1313979 339 GTCAATCTTCTCGAGGATTG  
51.92713248 54.330978 340 AACATGGAGAACATCACATC 79.31612772 80.8981286 341  
AACATCACATCAGGATTCCT 41.40633262 37.5509299 342 CTAGACTCTGCGGTATTGTG  
48.56267424 41.5330827 345 gRNA#003 CACCACGAGTCTAGACTCTG 44.43853541 40.8553881  
343 TACCGCAGAGTCTAGACTCG 49.18078863 56.151898 344 CGCAGAGTCTAGACTCGTGG  
52.41583101 57.2264647 346 TGGACTTCTCTCAATTTTCT 49.58564481 51.1350719 347  
GGACTTCTCTCAATTTTCTA 76.16671739 79.1684976 348 GACTTCTCTCAATTTTCTAG  
49.79317156 54.1540479 349 ACTTCTCTCAATTTTCTAGG 69.66968253 77.4650531 350  
CGAATTTTGGCCAAGACACA 53.53282063 54.0024954 371 gRNA#004  
CACAGAAAGGCCTTGTAAGT 42.35590319 41.6928086 370 CACTTTCTCGCCAACTTACA  
53.25960148 55.120666 373 gRNA#005 GGGCAACGGGGTAAAGGTTC 36.54111842 42.8120918  
375 gRNA#006 GTTGCCGGGCAACGGGGTAA 41.20322042 38.1885911 377  
CTGGCCGTTGCCGGGCAACG 57.27834882 60.830473 372 TGAACCTTTACCCCGTTGCC  
48.16509881 60.952804 378 CCTGGCCGTTGCCGGGCAAC 56.34234102 65.50842 379  
ACCTGGCCGTTGCCGGGCAA 54.10829257 53.324749 374 TTTACCCCGTTGCCCGGCAA  
56.72089131 62.6906255 380 GCACAGACCTGGCCGTTGCC 42.46818432 47.3720079 381  
GGCACAGACCTGGCCGTTGC 72.65381719 77.2400091 376 CCCGTTGCCCGGCAACGGCC  
50.93018919 61.086777 382 GCAAACACTTGGCACAGACC 57.0196485 69.491449 383  
GGGTTGCGTCAGCAAACACT 49.73518831 54.7510029 384 TTTGCTGACGCAACCCCCAC  
41.79724731 50.0362297 385 CTGACGCAACCCCCACTGGC 36.90727137 36.8247762 386  
TGACGCAACCCCCACTGGCT 46.49501492 59.6959921 387 GACGCAACCCCCACTGGCTG  
40.09200943 51.4756937 388 AACCCCCACTGGCTGGGGCT 61.82883278 79.8761795 390  
gRNA#007 TCCGCAGTATGGATCGGCAG 26.33655968 33.7255842 391 gRNA#008  
AGGAGTTCCGCAGTATGGAT 28.49512897 40.080391 389 gRNA#009  
TCCTCTGCCGATCCATACTG 28.45399116 42.735093 392 CGGCTAGGAGTTCCGCAGTA  
56.5241517 66.9060644 393 gRNA#010 TGCGAGCAAAACAAGCGGCT 41.5479747 40.5350018  
395 CCTGCTGCGAGCAAAACAAG 36.4525077 50.516964 394 CCGCTTGTTTTGCTCGCAGC  
108.4014077 90.5082399 396 TGTTTTGCTCGCAGCAGGTC 68.78508191 75.7537996 397  
GCAGCACAGCCTAGCAGCCA 78.73231487 68.3785588 398 TGCTAGGCTGTGCTGCCAAC  
59.52249922 69.0333267 401 CGTCCCGCGCAGGATCCAGT 52.51634701 49.5876502 399  
GCTGCCAACTGGATCCTGCG 75.81794218 89.0162904 400 CTGCCAACTGGATCCTGCGC  
77.79441236 73.9461516 402 AAACAAAGGACGTCCCGCGC 67.52500576 72.6685954 404  
CGCCGACGGGACGTAAACAA 77.77475148 70.288774 403 GTCCTTTGTTTACGTCCCGT  
94.99070926 103.867949 406 AGGTGCGCCCCGTGGTCGGT 68.80565242 65.4335257 407  
AGAGAGGTGCGCCCCGTGGT 42.18514493 55.1199635 408 GTAAAGAGAGGTGCGCCCCG  
53.39922155 55.7151401 410 CGGGGAGTCCGCGTAAAGAG 52.63946411 66.9249801 409  
GGGGCGCACCTCTCTTTACG 72.81702761 66.4993545 411 gRNA#011  
CAGATGAGAAGGCACAGACG 32.31425506 44.762352 413 GGCAGATGAGAAGGCACAGA

59.89738655 59.2550525 415 ACACGGTCGGCAGAGAGA 41.29188182 52.515655 412  
 GTCTGTGCCTTCTCATCTGC 70.71073836 72.0049046 416 GAAGCGAAGTGCACACGGTC  
 31.51588976 59.2847924 417 GAGGTGAAGCGAAGTGCACA 53.23795933 54.7085711 419  
 GGTCTCCATGCGACGTGCAG 98.80315853 94.871871 418 CTTCACCTCTGCACGTCGCA  
 76.66072308 76.4195077 421 GTCCTCTTATGTAAGACCTT 50.06169791 63.8903663 422  
 AGTCCTCTTATGTAAGACCT 54.84793515 62.0058784 420 TGCCCAAGGTCTTACATAAG  
 65.64906417 79.7359246 423 GTCTTACATAAGAGGACTCT 65.0201597 62.5458243 424  
 AATGTCAACGACCGACCTTG 53.64938718 65.5805852 425 TTTGAAGTATGCCTCAAGGT  
 68.9199506 80.763234 426 gRNA#012 AGTCTTTGAAGTATGCCTCA 30.45840615 47.6679105 427  
 AAGACTGTTTGTTTAAAGAC 75.19137394 74.1370789 428 AGACTGTTTGTTTAAAGACT  
 66.21290133 75.2309845 429 CTGTTTGTTTAAAGACTGGG 63.52924235 72.0972239 430  
 GTTTAAAGACTGGGAGGAGT 52.01423199 66.8961386 431 TCTTTGTACTAGGAGGCTGT  
 51.48581844 68.9533809 432 AGGAGGCTGTAGGCATAAAT 37.69681736 56.2655965 433  
 GTGAAAAAGTTGCATGGTGC 82.88524703 98.0043703 434 GCAGAGGTGAAAAAGTTGCA  
 31.73533955 53.6210823 435 gRNA#013 AACAAGAGATGATTAGGCAG 30.51551968 43.8402184  
 436 gRNA#014 GACATGAACAAGAGATGATT 15.37394867 25.9017005 437  
 AGCTTGAGGCTTGAACAGT 84.06388656 100.433196 441 gRNA#015  
 CCACCCAAGGCACAGCTTGG 22.57628478 29.4502561 443 AAGCCACCCAAGGCACAGCT  
 38.69686132 57.447646 438 CAAGCCTCCAAGCTGTGCCT 57.03790348 55.3144232 439  
 AAGCCTCCAAGCTGTGCCTT 101.2197916 108.433992 442 AGCTGTGCCTTGGGTGGCTT  
 62.50798441 75.5245296 444 GCTGTGCCTTGGGTGGCTTT 63.60985011 68.2127614 445  
 CTGTGCCTTGGGTGGCTTTG 58.80930094 60.2093595 446 TAGCTCCAAATTCTTTATAA  
 81.50792369 102.062484 447 GTAGCTCCAAATTCTTTATA 57.5300482 84.4089935 448  
 TAAAGAATTTGGAGCTACTG 55.34840957 67.1682598 449 ATGACTCTAGCTACCTGGGT  
 70.72899714 69.314819 450 CACATTTCTTGTCTCACTTT 135.7647935 119.430868 451  
 TAGTTTCCGGAAGTGTTGAT 52.38647155 59.8621336 452 CGTCTAACAACAGTAGTTTC  
 84.81350809 79.1119745 453 ACTACTGTTGTTAGACGACG 50.34753433 57.5139945 454  
 CTGTTGTTAGACGACGAGGC 47.03375963 53.0434947 455 CGAGGGAGTTCTTCTTCTAG  
 36.81318989 50.1844755 456 GCGAGGGAGTTCTTCTTCTA 68.04429109 71.2738682 457  
 gRNA#016 GGCGAGGGAGTTCTTCTTCT 35.40374342 49.4263836 459  
 GACCTTCGTCTGCGAGGCCA 28.35732375 53.108582 460 AGACCTTCGTCTGCGAGGCCG  
 41.45363172 58.2048965 461 GATTGAGACCTTCGTCTGCG 63.13599738 73.3793991 458  
 CTCCCTCGCCTCGCAGACGA 41.73812486 56.4066766 462 GATTGAGATCTTCTGCGACG  
 134.1434937 133.039909 463 GTCGCAGAAGATCTCAATCT 44.87633493 58.0732445 464  
 TCGCAGAAGATCTCAATCTC 70.59684886 75.0458487 465 gRNA#017  
 ATATGGTGACCCACAAAATG 41.36374656 46.043276 466 TTTGTGGGTCACCATATTCT  
 66.33644682 65.6466534 467 gRNA#018 TTGTGGGTCACCATATTCTT 48.06595023 41.7714626  
 468 GCTGGATCCAACCTGGTGGTC 65.83430344 69.3357339 469 CACCCCAAAAGGCCTCCGTG  
 21.63462413 23.5507547 471 gRNA#019 CCTGAGGGCTCCACCCCAAA 45.40727826 44.6869573  
 470 CCTTTTGGGGTGGAGCCCTC 50.06807456 31.73417 472 GGGGTGGAGCCCTCAGGCTC  
 64.29444481 64.1755302 473 GGGTGGAGCCCTCAGGCTCA 44.19826805 53.1051257 474  
 CGATTGGTGGAGGCAGGAGG 65.52555289 60.9306557 475 gRNA#020  
 CTCATCCTCAGGCCATGCAG 35.40063237 17.5286587

(823) In vitro silencing was observed in an HepG2-NTCP infection model with gRNAs targeting CpG islands with ETRs (FIG. 5A-FIG. 5B). A primary screen was conducted using LNPs of quality within expected parameters and a pilot experiment with a single guide (FIG. 6-FIG. 8). Results demonstrated that 48 gRNAs showed less than 50% expression of HBeAg at day 6 compared to non-targeting control (FIG. 9) and 28 gRNAs showed less than 50% expression of HBsAg at day 6 compared to non-targeting control (FIG. 10). HBsAg and HBeAg expression was positively correlated as shown in FIG. 11.

#### Example 4: Zinc Finger Repressors for Silencing HBV

(824) Zinc finger repressors targeting epigenetic target sites identified in the HBV genome were designed. Table 1 above provides amino acid sequences of zinc finger and its corresponding motif



sequences and target sequences of the zinc finger.

(825) Zinc finger repressors described in Table 1 are tested in an HBV infection model, e.g., in HepG2 cells as described herein, and efficient repression of HBV is confirmed for the zinc finger repressors provided in Table 1.

#### Example 5: Further In Vitro Evaluation of gRNAs

(826) A CRISPR-Off single construct encoding PLA002, consisting of KRAB, DNMT3A, DNMT3L, and dCas9, was used in combination with one or more of the designed sgRNAs for the in vitro assays described in this example.

(827) HepG2-NTCP cells were infected with HBV for 4 days, following procedures similar as those in Example 3, and were then transfected with CRISPR-off construct and individual exemplary gRNAs (as indicated in Table 13) formulated in a research-grade LNP. At Day 6 post-transfection HBsAg and HBeAg protein expression in the supernatant was evaluated by ELISA, as depicted in FIG. 12A. Results from this experiment are shown in FIG. 12B. All of the tested gRNAs led to reduction of HBsAg and HBeAg levels in the supernatant. Positive control used in this experiment is a gRNA against HBV genome that was previously shown to reduce antigens ~50%.

(828) In another experiment, the integrated HBV cell line, PLC/PRF/5, was used to evaluate activity of gRNAs. The PLC/PRF/5 cells were transfected with CRISPR-off (PLA002) and individual gRNAs using a commercial lipid-based transfection reagent. As depicted in FIG. 13A, four days after transfection HBsAg protein expression in the supernatant was evaluated by ELISA. Results from this experiment are shown in FIG. 13B. Target conservation was evaluated in silico and target conservation was defined as 100% gRNA-DNA match.

(829) In a further experiment, primary human hepatocytes (PHH) derived from humanized mice were infected with HBV for 4 days and then transfected with CRISPR-off (PLA002) and individual gRNAs formulated in a research-grade LNP, GenVoy LNPs. As depicted in FIG. 14A, at Day 6 post-infection HBsAg and HBeAg protein expression in the supernatant was evaluated by ELISA. Results from this experiment are shown in FIG. 14B. Positive control used in this experiment is an HBV gRNA that was previously shown to reduce antigens ~50%. The data suggested strong in vitro silencing by certain gRNAs at Day 6 after transfection. In a second PHH experiment, depicted in FIG. 14C, post-infection HBsAg and HBeAg protein expression in the supernatant was evaluated by ELISA at Day 12 after delivery of 100 ng of payload (1:1 effector to guide RNA ratio) in research-grade LNPs. Epigenetic editors repress HBsAg and HBeAg secretion in HBV infected PHH cells at this time point, as well. Results are shown in FIG. 14D. Sequences of the exemplary gRNAs that were tested in this example are listed in Table 13.

#### Example 6: In Vivo Silencing of HBV in HBV Rodent Models

(830) Two different HBV rodent models were tested in this study. As shown in FIG. 15, in one set of experiments, a non-transgenic model of persistent HBV infection in immunocompetent mice was used, which was established by administering an adeno-associated viral vector (AAV) that contains HBV Genotype D DNA into the mice. The administration of the AAV-HBV vector resulted in expression of hepatitis B surface antigen (HBsAg), hepatitis B e antigen (HBeAg), and high levels of serum HBV DNA in the mice. In another set of experiments, a transgenic mouse model of persistent HBV infection was used, whose genome was engineered to integrate HBV Genotype A DNA, resulting in expression of HBsAg and HBeAg, and circulating viral DNA in the mice.

(831) Both mouse models were used to test 6 different treatment groups as shown in FIG. 15. At certain times (such as 7, 14, 28, and 35 days) after single administration of 3 mg/kg of the LNPs that were loaded with the CRISPR-off construct and respective gRNAs, WT-Cas9 construct and gRNA, or control vehicle, mouse serum was extracted for analysis of HBsAg, HBeAg, and HBV DNA. Later the mice were sacrificed, and their livers were collected for further analysis.

(832) As shown in FIG. 16, in transgenic mouse model, durable (~1 month) and efficacious (~2 Log) DNA and HBsAg reduction was observed with CRISPR-Off/gRNA #011 treatment. And compared to Cas9 cutter, CRISPR-Off, when administered in combination with gRNA #011, showed similar circulating viral DNA reduction, but superior HBsAg and HBeAg reduction.

(833) Reduction of HBV markers in AAV-HBV model was also observed with administration of certain

exemplary constructs. As shown in FIG. 17, overall results in AAV8-HBV model are similar to the Tg-HBV mouse model. About 1 log DNA and HBsAg antigen reduction was observed with administration of CRISPR-Off and gRNA #011.

(834) Effects of redosing of certain exemplary constructs were also tested. In the same experiments as above, among the six transgenic mice receiving administration of “CRISPR-off+ gRNA #016” (CRISPR-off construct and gRNA gRNA #016), three were administered with a dose of “CRISPR-off+ gRNA #016” on Day 35, and the other three were administered with “CRISPR-off+ gRNA #011” on Day 35. As shown in FIG. 18A, redosing either with a less effective gRNA (gRNA #016 in this case) or with a more effective gRNA (gRNA #011 in this case) enhanced the silencing of all HBV marker, as shown by reduction of circulating HBV DNA, HBsAg, and HBeAg on Day 42. Redosing the gRNA #016-treated group with gRNA #011 (more effective gRNA) resulted in a more substantial reduction than redosing with gRNA #016 (less effective gRNA).

(835) Single-dose experiments were continued to 168 days, as shown in FIG. 18B. Results show durable and progressive reduction of viral antigens achieving -2.7 log DNA and -2.8 log HBsAg more than five months after single administration of an epigenetic editor (CRISPR-off with gRNA #011). Five out of six animals tested had undetectable HBV DNA and HBsAg 168 days after a single dose of an epigenetic editor.

(836) Redosing experiments were also conducted in AAV-HBV mouse model, as shown in FIG. 19. Dosing with two different gRNAs (gRNA #016 and gRNA #011) further decreased all HBV markers. These data suggest of a potential enhanced activity when two HBV regions are targeted.

(837) Sequences of the exemplary gRNAs that were tested in this example are listed in Table 13.

#### Example 7: Evaluation of ZFP in HepG2-NTCP Cells

(838) In this example, ZF-off single constructs encoding a fusion protein consisting of KRAB, DNMT3A, DNMT3L, and an exemplary zinc finger motif of choice, were tested. Sequences of the exemplary zinc fingers that were tested in this example are listed in Table 18, as are sequences for plasmids yielding a subset of the ZF-off single construct fusion proteins.

(839) Certain exemplary ZF-off constructs were formulated in a research-grade LNP. HepG2-NTCP cells were infected with HBV for 4 days and then transfected with the ZF-off loaded LNPs. As depicted in FIG. 20A, at Day 6 post-infection HBsAg and HBeAg protein expression in the supernatant was evaluated by ELISA. FIG. 20B shows the results as measured by percentage reduction in HBV antigens as compared to non-targeting control. Positive control used in this experiment is a HBV gRNA previously shown to reduce antigens ~50%. FIG. 21A shows the results of the top ten ZF-off constructs that lead to the most reduction in HBV antigens. FIG. 21B shows the results for all constructs in the screen.

(840) Table 14 and 15 below show the raw data from these experiments, listed with the mRNA number yielding the zinc finger motif.

(841) TABLE-US-00021

TABLE 14 % HBsAg expression relative to non-targeting control	Trial#	1	2	3
4 5 6 7 8 Non-targ control	100	100	100	100
Pos control	54	59	68	61
mRNA0001	10	19	25	
mRNA0002	12	2	8	12
mRNA0003	10	11	14	15
mRNA0004	10	28	13	39
mRNA0005	3	5	1	8
mRNA0006	4	12	8	19
mRNA0007	97	86	60	66
mRNA0008	68	69	65	64
mRNA0009	65	67	74	98
mRNA0010	84	69	66	73
mRNA0011	67	50	60	59
mRNA0012	59	61	70	92
mRNA0013	97	70	66	71
mRNA0014	60	81	66	74
mRNA0015	81	73	77	129
mRNA0016	120	78	71	77
mRNA0017	75	77	82	82
mRNA0018	78	84	93	131
mRNA0019	107	107	77	100
mRNA0020	77	99	60	116
mRNA0021	32	49	68	66
mRNA0022	71	66	51	56
mRNA0023	65	71	76	41
mRNA0024	109	89	86	92
mRNA0025	86	92	90	82
mRNA0026	77	88	81	104
mRNA0027	128	77	80	81
mRNA0028	71	67	59	66
mRNA0029	48	47	40	57
mRNA0030	109	82	76	75
mRNA0031	46	32	41	27
mRNA0032	50	59	52	73
mRNA0033	61	62	46	50
mRNA0034	51	24	41	25
mRNA0035	30	25	24	34
mRNA0036	16	22	19	19
mRNA0037	54	43	42	46
mRNA0038	19	23	13	29
mRNA0039	28	46	37	36
mRNA0040	88	78	83	80
mRNA0041	103	92	100	
mRNA0042	99	91	99	
mRNA0043	93	89	97	
mRNA0044	98	100	95	
mRNA0045	100	96	95	
mRNA0046	94	83	92	
mRNA0047	97	77	99	
mRNA0048	96	94	90	
mRNA0049	88	87	89	
mRNA0050	87	87	85	
mRNA0051	106	104	114	
mRNA0052	104	101	107	
mRNA0053	88	86	92	
mRNA0054	98	102	91	
mRNA0055	101	96	100	
mRNA0056	99	107	108	
mRNA0057	101	102	104	
mRNA0058	110	104	102	

mRNA0059 100 91 98 mRNA0060 94 103 100 mRNA0061 104 96 103 mRNA0062 106 98 104  
mRNA0063 96 86 99  
(842) TABLE-US-00022 TABLE 15 % HBeAg expression relative to non-targeting control Trial# 100  
100 100 100 Non-targ control 100 100 100 100 Pos control 26 36 41 53 43 43 34 54 mRNA0001 12 19  
22 23 mRNA0002 15 8 17 20 mRNA0003 11 9 13 12 mRNA0004 10 17 9 27 mRNA0005 1 1 -1 3  
mRNA0006 5 8 7 13 mRNA0007 95 78 59 65 mRNA0008 64 67 60 65 mRNA0009 65 64 81 98  
mRNA0010 84 68 69 70 mRNA0011 65 51 51 67 mRNA0012 64 61 74 96 mRNA0013 92 74 73 79  
mRNA0014 58 85 58 76 mRNA0015 82 83 78 124 mRNA0016 108 81 72 80 mRNA0017 72 77 72 80  
mRNA0018 55 55 71 93 mRNA0019 71 79 51 87 mRNA0020 34 36 32 52 mRNA0021 32 40 55 55  
mRNA0022 77 64 53 65 mRNA0023 60 69 72 43 mRNA0024 98 76 87 84 mRNA0025 91 86 82 92  
mRNA0026 78 97 87 102 mRNA0027 117 62 68 74 mRNA0028 75 59 58 71 mRNA0029 31 32 22 45  
mRNA0030 124 86 79 77 mRNA0031 42 23 27 20 mRNA0032 46 57 57 82 mRNA0033 56 51 44 76  
mRNA0034 42 21 41 18 mRNA0035 22 22 24 39 mRNA0036 13 17 16 13 mRNA0037 50 35 34 35  
mRNA0038 12 16 13 25 mRNA0039 29 45 39 36 mRNA0040 93 73 80 82 mRNA0041 80 63 111  
mRNA0042 114 94 98 mRNA0043 98 91 99 mRNA0044 91 115 108 mRNA0045 71 55 62 mRNA0046  
76 66 63 mRNA0047 55 55 45 mRNA0048 66 63 78 mRNA0049 83 59 52 mRNA0050 51 55 49  
mRNA0051 55 49 49 mRNA0052 56 57 66 mRNA0053 92 60 57 mRNA0054 50 55 56 mRNA0055 83  
88 74 mRNA0056 61 69 112 mRNA0057 106 73 65 mRNA0058 66 65 65 mRNA0059 69 66 71  
mRNA0060 59 94 101 mRNA0061 111 81 68 mRNA0062 28 33 41 mRNA0063 65 55 31

Example 8. Dose Response Testing of Viral Antigens in HepG2-NTCP Cells

(843) In this example, top ZF fusion proteins were tested in 5-point dose response assay for HBsAg and HBeAg. The 5 dosage points were 200ng, 150ng, 100ng, 50ng, and 25ng. Experimental schematic and results are shown in FIG. 22.

Example 9. Testing for Durable Repression of HBsAg in HepG2.2.15 Cells

(844) In this example, top ZF and CRISPR-off fusion proteins with guide RNAs were tested for durable repression of HBsAg. Active ZFPs and CRISPR-off editors showed durable silencing through Day 27 with 50ng treatment. Experimental schematic and results are shown in FIGS. 23A-23C.

Example 10. Testing of Silencing of HBsAg in a Second Model for Int-HBV

(845) In this example, top ZF fusion proteins were tested for repression of HBsAg in PLC/PRF/5 cells. A subset of the ZFPs silenced HBsAg in this second model. Experimental schematic and results are shown in FIG. 24. 1. Testing ZF Fusion Proteins and CRISPR-off with guide RNAs for Specificity  
(846) In this example, ZF fusion proteins targeting HBV exhibiting significant silencing were profiled for specificity in HepG2-NTCP at day 19. All comparisons were performed against a non-targeting ZFP control. An exemplary result for the ZF fusion protein with mRNA0001 zinc finger motif is shown in FIG. 25A. CRISPR-off with guide RNAs were similarly profiled. HepG2-NTCP cells were transfected with 100 ng of total payload using GenVoy™ LNP at a 1:1 gRNA:effector ratio. Cells were split every 3-4 days and collected at day 15 post-treatment for specificity assessments, including RNA-seq and methylation array. DESeq2 was used to identify differential gene expression. As shown in FIG. 25B, little to no changes were observed above chosen thresholds (absolute[log 2[fold change]]>1 and -log 10[adjusted p-value]>5) as expected for effectors targeting HBV DNA. For methylation array, the Infinium MethylationEPIC v2.0 array was used, and DMRs were identified using Bumhunter. EE3, EE4, and EE5 had a result of DMR=0. Results are shown in FIGS. 25C-25D.

Example 11. In Vivo Analysis of ZF-Off Constructs

(847) Ten ZF-Off constructs as well as vehicle-only and CRISPR-Off controls were administered to AAV-HBV mice at 1 mg/kg as shown in the schematic in FIG. 26. Table 16 shows the zinc finger motifs for each experimental group; the corresponding plasmid from Table 18, comprising the nucleic acid encoding the ZF-Off construct, was administered. Plasma from the mice was tested at Days 7, 14, 21, and 28 post dose for HBV DNA, HBsAg, and HBeAg. The livers were collected for further analysis. Results are shown in FIG. 27. The ZF-Off construct with the ZF motif from mRNA0004 showed more than a 1.5 log reduction in HBV DNA, a >2 log reduction in HSbsAg, and a >2 log reduction of HBeAg, all sustained up to 28 days from the dose.

(848) TABLE-US-00023 TABLE 16 Experimental groups for in vivo testing of ZF-Off constructs. ZF

motif in construct Group administered N 1 mRNA0001 6 2 mRNA0002 6 3 mRNA0003 6 4  
mRNA0005 6 5 mRNA0006 6 6 mRNA0038 6 7 mRNA0004 6 8 mRNA0039 6 9 mRNA0021 6  
10 mRNA0037 6

Example 12. Zinc Finger Protein Multiplexing Study in an AAV-HBV and Tg-HBV Mouse Model  
(849) AAV-HBV mice are injected with a single administration at 0.5 mg/kg of one, two, or three ZF  
fusion proteins, delivered as mRNA, in LNPs (schematic, FIG. 28) in accordance with Table 17. HBV  
DNA, HBsAg, and HBeAg are assayed in plasma at one or more time points, and the mouse liver is  
collected for further analysis.

(850) TABLE-US-00024 TABLE 17 Multiplexing sample groups. Group ZF\_Off-1 ZF\_Off-2 ZF\_Off-3  
1 mRNA0004 mRNA0021 — 2 mRNA0004 mRNA0003 — 3 mRNA0004 mRNA0038 — 4  
mRNA0004 mRNA0021 mRNA0003 5 mRNA0004 mRNA0038 mRNA0003 6 mRNA0004  
mRNA0021 mRNA0038 7 mRNA0004 mRNA0001 — 8 mRNA0004 mRNA0039 — 9 mRNA0004  
— — 10 Vehicle — —

Example 13. Dose Response for CRISPR-Off Constructs in an AAV In Vivo Model

(851) A single dose of CRISPR-Off (SEQ ID NO: 1248) mRNA with guide RNA #008 as well as  
vehicle-only control was tested via 1:1 mRNA:guide RNA administration to AAV-HBV mice at 0.5  
mg/kg, 1 mg/kg, or 3 mg/kg in LNPs as shown in the schematic in FIG. 29. Plasma from the mice was  
tested for HBsAg at thirteen time points through 186 days after injection. Results are shown in FIG. 30.  
The highest dose administered showed an approximately 3.3 log reduction in HBsAg, sustained through  
186 days after the dose.

Example 14. Dose Response for CRISPR Off Constructs in Tg In Vivo Model

(852) A single dose of CRISPR-Off (SEQ ID NO: 1248) mRNA with guide RNA #008 as well as  
vehicle-only control was tested via 1:1 mRNA:guide RNA administration to Tg-HBV mice at 0.5 mg/kg,  
1 mg/kg, or 3 mg/kg in LNPs as shown in the schematic in FIG. 31. Plasma from the mice was tested for  
HBsAg at thirteen time points through 186 days after injection. Results are shown in FIG. 32. The  
highest dose administered showed an approximately 2.6 log reduction in HBsAg, sustained through 196  
days after the dose.

(853) A second dose response experiment in Tg-HBV model using CRISPR-Off (SEQ ID NO: 1248)  
mRNA with guide RNA #008 formulated in LNPs was conducted, with administrations at 0.2 mg/kg, 0.5  
mg/kg, 1 mg/kg, or 3 mg/kg of 1:1 mRNA:guide RNA. A vehicle-only control was also used. In this  
experiment, plasma was tested for HBV DNA, HBsAg, and HBeAg at 13 time points through 207 days  
after injection. Results are shown in FIG. 32. The HBsAg results for individual mice at the final time  
point of 207 days after injection are plotted in FIG. 33. All of the mice in the 0.5 mg/kg, 1 mg/kg, and 3  
mg/kg group had reduced HBsAg at Day 207 as compared to vehicle only control. Alanine transaminase  
(ALT) level in the mice was also tested at 207 days and found to be comparable to that of healthy  
untreated mice for all treatment groups.

Example 15. Guide RNA Testing in AAV-HBV Mice

(854) Six guide RNAs were tested for relative efficacy using CRISPR-Off (SEQ ID NO: 1248) in a 28-  
day, single-dose study. CRISPR-Off construct mRNA and one of gRNA #003, gRNA #007, gRNA #008,  
gRNA #009, gRNA #011, and gRNA #015 was delivered at 1:1 mRNA:guide RNA at 1 mg/kg. Controls  
included vehicle only, CRISPRi with gRNA #008 (not shown), and wild type Cas9 with gRNA #011  
(not shown). HBV DNA and HBsAg was measured over 28 days. Results are shown in FIG. 34. Most of  
the single guide treatments tested in this experiment resulted in decreased HBV DNA and HBsAg versus  
vehicle only control.

Example 16. Durability Study for ZF-Off in AAV-HBV In Vivo Model: Single and Re-Dose

(855) Mice were injected with a single dose ZF-Off construct (SEQ ID NO: 36) mRNA at 1 mg/kg in  
LNPs. HBV DNA and HBsAg were measured from plasma over a period of 168 days. Results are shown  
in FIG. 35A. The treatment resulted in a sustained reduction of greater than 2 log in HBV DNA and  
similar sustained reduction in HBsAg.

(856) In another study, mice were injected with the ZF-Off construct (SEQ ID NO: 36) mRNA at 1  
mg/kg for three doses: Day 0, Day 21, and Day 42. HBV DNA and HBsAg were measured from plasma  
over a period of 225 days. Results are shown in FIG. 35B. Results were similar to those of the previous

single-dose experiment and in this experiment sustained over 225 days.

#### Example 17. Re-Dosing Studies for CRISPR-Off in AAV-HBV In Vivo Model

(857) AAV-HBV mice were dosed with either a single dose or three doses, all at 1 mg/kg in LNPs, of CRISPR-Off (SEQ ID NO: 1248) mRNA with gRNA #008 at a 1:1 ratio of mRNA: guide RNA. For the single dose condition, the dose was administered at Day 0. For the three-dose condition, the doses were administered at Day 36, Day 57, and Day 78. A vehicle-only control was also administered. Plasma measurements of HBV DNA, HBsAg, and HBeAg were taken through Day 168 for the single-dose condition, and through Day 261 for both the three-dose condition and the vehicle control. Results are shown in FIG. 36. Re-dosing with CRISPR-Off further improved and sustained the durability of the modulation of these HBV biomarkers.

(858) In another study, AAV-HBV mice were dosed with either a single dose of CRISPR-Off (SEQ ID NO: 1248) mRNA with gRNA #008 with an updated modification pattern (SEQ ID NO: 1249) (1:1 ratio mRNA: guide RNA) in LNPs at 3 mg/kg, or three doses of the same epigenetic editor, each at 1 mg/kg. Both groups received a dose at Day 0, and the three-dose group also received a dose at Day 14 and at Day 28. A vehicle-only control was also administered. HBsAg and HBeAg were measured from plasma through 126 days. Results are shown in FIG. 37. Near-additive pharmacology was demonstrated with the repeat dosing.

#### Example 18. Testing CRISPR-Off and Guide RNA Modifications in an AAV-HBV In Vivo Model

(859) AAV-HBV mice were dosed with a single dose of either CRISPR-Off (SEQ ID NO: 1248) mRNA with gRNA #008 or an updated CRISPR-Off variant (SEQ ID NO: 1252) mRNA with gRNA #008 with an updated modification pattern (SEQ ID NO: 1249), with a 1:1 ratio of mRNA to guide RNA at either 0.5 mg/kg or 1 mg/kg, delivered in LNPs. A vehicle only control was also administered. HBsAg was measured in plasma over 28 days. Results are shown in FIG. 38. The updated CRISPR-Off variant with guide RNA modifications demonstrated 1.5× potency over the previous lead epigenetic editor.

#### Example 19. Methylation Studies for CRISPR-Off with Various Guide RNAs

(860) HepG2.2.15 cells were dosed at 1 nanogram (ng)/microliter (100 ng) of 1:1 CRISPR-Off (SEQ ID NO: 1248) mRNA with various single guide RNAs in LNPs with commercial apolipoprotein E (to aid LNP entry). Methylation profiles were performed on the HBV genome samples as well as controls: for gRNA #008, untreated samples and treated with CRISPRi and wild type Cas9. For other gRNAs tested, an untreated sample (APOE only) was used as a control. Results for gRNA #008, gRNA #003, gRNA #007, gRNA #009, gRNA #011, and gRNA #015 are shown in FIGS. 39A, 39B, 39C, 39D, 39E, and 39F, respectively. A control for the application of an off-target PCSK9 guide RNA is shown in FIG. 39G.

#### Example 20. Specificity studies for CRISPR-Off and ZF Off

(861) HepG2.2.15 cells were transfected with either ZF-Off (SEQ ID NOs: 36 and 73) mRNA or CRISPR-Off (SEQ ID NO: 1248) mRNA with gRNA #008 in research-grade LNPs. RNA-Seq was conducted to determine differentially expressed genes (DEGs), and the Twist panel was used to determine differentially methylated regions (DMRs) at CpG-enriched sites. Differentially expressed genes (DEG) and differentially methylated regions (DMR) are defined based on literature reviews, software recommendations, sequencing depth and controls DEGs are genes that have  $\geq 2$ -fold change and with adjusted p-value  $\leq 1e-05$ . DMRs are defined as regions with a minimum of 10 CpGs, with 5× coverage, p-value of  $\leq 1e-10$  and min average change in methylation (beta)  $\geq 20\%$ . Results are shown in FIG. 40. Silencing data for same samples was also obtained. Results are shown in FIG. 41.

#### Example 21. Dose Response of Guide RNAs In Vitro

(862) An 8-point dose-response (two-fold dilution with from 4 ng/μL (400ng) to 0.031 ng/μL (3.1 ng)) was generated using HepG2.2.15 cells treated with LNPs with CRISPR-Off effector (SEQ ID NO: 1248), delivered as mRNA, and each of four gRNAs co-formulated in a 1:1 ratio. HBsAg and HBeAg were measured over six days. Results are shown in FIG. 42.

#### Example 22. Dose Response of CRISPR-Off Variant In Vitro

(863) HepG2.2.15 cells transfected via Messenger Max with CRISPR-Off effector (SEQ ID NO: 1252), delivered as mRNA, and gRNA #008 with updated modification pattern (SEQ ID NO: 1249) was used to generate a 9-point dose-response (200-0.8 ng) curve. HBsAg and HBeAg were measured over 6 days. Results are shown in FIG. 43.

#### Example 23. Multiplexing Study in AAV-HBV and Tg-HBV Mouse Models

(864) AAV-HBV and Tg-HBV mice are injected with a single administration at 0.5 mg/kg of one, two, three, or four guide RNAs targeting regions listed in Table 12 and Table 13 with CRISPR-Off (SEQ ID NO: 1248 or 1252) mRNA formulated in LNPs.

(865) Amongst others, the following gRNAs are combined: (1) gRNA #008 and gRNA #011; (2) gRNA #008 and gRNA #003; (3) gRNA #008 and gRNA #015; (4) gRNA #008, gRNA #011, and gRNA #015; (6) gRNA #008, gRNA #011, and gRNA #003. Treatment with a single guide RNA, e.g., gRNA #008 or gRNA #011 serves as a positive control, and treatment with vehicle or with a non-targeting guide as a negative control.

(866) One or more of HBV DNA, HBsAg, and HBeAg are assayed in plasma of the mice at one or more time points after administration, and the mouse liver is collected for further analysis. Combinations of multiple guides yield silencing at least as robust as treatment with single guides. In some cases, more robust silencing with multiple guides as compared to treatment with a single guide is observed.

#### Example 24. Testing mRNA: Guide RNA Ratios In Vivo

(867) AAV-HBV mice are treated with CRISPR-Off effector (SEQ ID NO: 1252) mRNA with guide RNA (SEQ ID NO: 1249) in ratios including 1:1, 1:1.5, 2:1, 1:2, and 1:3 mRNA:guide RNA formulated into LNPs and administered at 0.5 mg/kg. 5 or 6 mice per study group are used. An optimized ratio of effector and guide RNA is identified that results in durable reduction of one or more HBV biomarkers, e.g., plasma level measurements of HBV DNA, HBsAg, and HBeAg of greater than 2 log below the observed control plasma level.

#### Example 25. Combination Treatment with Epigenetic Editor In Vivo

(868) Tg-HBV mice are dosed with Entecavir (ETV) at 0.1 mg/kg for 14 days followed by CRISPR-Off with guide RNA at 1 mg/kg in a single intravenous dose. HBV DNA and HBsAg are measured in plasma for 112 days. HBV DNA levels drop after ETV treatment and there is slight synergism in the CRISPR-Off with guide with ETV group. After ETV withdrawal, the CRISPR-Off with guide maintains sustained reduction of DNA comparable to a group treated with CRISPR-Off and guide RNA alone. The addition of ETV does not affect HBsAg.

#### Example 26. Stable HBV Silencing Via Epigenetic Editing in Non-Transgenic Mouse Model of Persistent HBV Infection

(869) A non-transgenic model of persistent HBV infection (AAV-HBV) in immunocompetent mice was used, which was established by administering an adeno-associated viral vector (AAV) that contains HBV Genotype D DNA into the mice. The administration of the AAV-HBV vector resulted in expression of hepatitis B surface antigen (HBsAg), hepatitis B e antigen (HBeAg), and high levels of serum HBV DNA in the mice.

(870) The CRISPR-off and ZF-off constructs are tested. Constructs are delivered via IV administration of mRNA/gRNA (CRISPR-Off) or mRNA (ZF-Off) formulated into a lipid nanoparticle (LNP) at 2.5 mg/kg and 0.5 mg/kg for CRISPR-Off and ZF-Off, respectively. Some constructs are formulated in LNP compositions as described in PCT/US2014/070882, US20220402862A1, and/or US20230203480A1. A subset of the mice are re-dosed at two weeks after the first dose; a second subset are re-dosed at one month after the first dose. The readouts are circulating viral DNA, HBsAg, and HBeAg, tested using mouse plasma at one or more time points (such as 7, 14, 28, and 35 days). A durable and significant reduction in the levels of one or more of HBV DNA, HBsAg, and HBeAg is observed for some constructs.

(871) Longer-term durability is tested over three to six months using the HBV DNA, HBsAg, and HBeAg markers. Progressive and durable reduction in one or more of these markers is seen with delivery of some constructs. The mice are sacrificed and livers are collected for further analysis, and durable silencing is confirmed by at least 2 log reduction of HBsAg and HBV DNA.

#### Example 27: Stable HBV Silencing Via Epigenetic Editing in Transgenic Mice Expressing Viral HBV DNA

(872) A transgenic mouse model of persistent HBV infection (Tg-HBV) was used, whose genome was engineered to integrate HBV Genotype A DNA, resulting in expression of HBsAg and HBeAg, and circulating viral DNA in the mice.

(873) The CRISPR-off and ZF-off constructs are tested. Constructs are delivered via IV administration of mRNA/gRNA (CRISPR-Off) or mRNA (ZF-Off) formulated into LNP at 2.5 mg/kg and 0.5 mg/kg for CRISPR-Off and ZF-Off, respectively. Some constructs are formulated in LNP compositions as described in US20220402862A1, and/or US20230203480A1. A subset of the mice are re-dosed at two weeks after the first dose; a second subset are re-dosed at one month after the first dose. The readouts are circulating viral DNA, HBsAg, and HBeAg, tested using mouse plasma at one or more time points (such as 7, 14, 28, and 35 days). A durable and significant reduction in the levels of one or more of HBV DNA, HBsAg, and HBeAg is observed for some constructs.

(874) Longer-term durability is tested over three to six months using the HBV DNA, HBsAg, and HBeAg markers. Progressive and durable reduction in one or more of these markers is seen with delivery of some constructs. The mice are sacrificed and livers are collected for further analysis, and durable silencing is confirmed by at least 2 log reduction of HBsAg and HBV DNA.

#### SEQUENCES

(875) The SEQ ID NOs (SEQ) of nucleotide (nt) and amino acid (aa) sequences described in the present disclosure are listed in Table 18 below.

(876) TABLE-US-00025 TABLE 18 Sequence listing. SEQ Description Sequence 1 *S. pyogenes*  
WT ATGGATAAGAAATACTCAATAGGCTTAGATATCGGCACAAATAGCGTCGGATGG Cas9  
Sequence GCGGTGATCACTGATGAATATAAGGTTCCGTCTAAAAAGTTCAAGGTTCTGGGA (nt)  
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*pyogenes* WT MDKKYSIGLDIGTNSVGWAVITDEYKVP SKKFKVLGNTDRHSIKKNLIGALLED  
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DWLAYIQELRN 22 inactive HFAsCpf1

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DWLAYIQELRN 23 inactive

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[illegible]



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*H. parainfluenzae*  
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*Arabidopsis* DRM1  
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OMe modified nucleotide, \* indicates a phosphorothioate bond) 1250 CRISPR-Off

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1 plasmid

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 VNFTQGEWQRLNPEQRNLYRDVMLENYSNLVSVGQGETTKPDVILRLEQGKEPW  
 LEEEEVLGSGRAEKNGDIGGQIWKPDKVKELSAKRPAATKKAGQAKKK

(877) TABLE-US-00026 TABLE 19 Annotation of PLA003 amino acid sequence Name Type Minimum  
 Maximum Length SV40 NLS CDS 2 8 7 SV40 NLS CDS 9 15 7 DNMT3A CDS 17 317 301 Linker  
 CDS 318 344 27 DNMT3L full- CDS 345 730 386 length XTEN80 CDS 731 810 80 dCas9 CDS 811  
 2180 1370 NLS CDS 2181 2187 7 XTEN16 CDS 2188 2208 21 ZIM3 CDS 2211 2310 100 SV40 NLS  
 CDS 2313 2319 7 SV40 NLS CDS 2320 2326 7

(878) TABLE-US-00027 TABLE 20 Annotation of PLA003 polynucleotide sequence Name Type  
 Minimum Maximum Length SV40 NLS CDS 4 24 21 SV40 NLS CDS 25 45 21 DNMT3A CDS 49 951  
 903 Linker CDS 952 1032 81 DNMT3L full- CDS 1033 2190 1158 length XTEN80 CDS 2191 2430  
 240 dCas9 CDS 2431 6540 4110 NLS CDS 6541 6561 21 XTEN16 CDS 6562 6624 63 ZIM3 CDS  
 6631 6930 300 SV40 NLS CDS 6937 6957 21 SV40 NLS CDS 6958 6978 21 stop terminator 6979  
 6981 3

## Claims

1. A method, comprising administering an epigenetic editing system to a subject, wherein the subject comprises detectable levels of HBV DNA, HBsAg, and/or HBeAg in plasma of the subject, wherein the epigenetic editing system comprises a) a fusion protein comprising i) a dCas9 protein domain, ii) a DNMT3A domain and iii) a human KRAB domain or one or more nucleic acid molecules encoding the fusion protein, and b) a guide RNA (gRNA) comprising a nucleic acid base sequence selected from the group consisting of SEQ ID NOs: 1093-1110, 1130-1164, 1166-1173, 1175-1199, 1201-1235, and 1249, wherein the nucleic acid base sequence comprises a region complementary to a strand of a target region wherein the target region comprises a sequence selected from the group consisting of SEQ ID NOs: 333-350, 370-404, 406-413, 415-439, and 441-475.

2. The method of claim 1, wherein the fusion protein further comprises a DNMT3L domain.
  3. The method of claim 1, wherein the dCas9 protein domain is from *Streptococcus pyogenes* or *Staphylococcus aureus*.
  4. The method of claim 3, wherein the dCas9 protein domain comprises a sequence selected from the group consisting of SEQ ID NOs: 12, 13, 27, 28 and 29.
  5. The method of claim 3, wherein the dCas9 protein domain is from *Streptococcus pyogenes*.
  6. The method of claim 4, wherein the dCas9 protein domain comprises the sequence of SEQ ID NO: 12.
  7. The method of claim 1, wherein the DNMT3A domain comprises a sequence selected from the group consisting of SEQ ID NOs: 1028 and 1029.
  8. The method of claim 1, wherein the DNMT3A domain comprises a sequence of SEQ ID NO: 1029.
  9. The method of claim 1, wherein the human KRAB domain comprises a sequence selected from the group consisting of SEQ ID NOs: 495, 551, 536, 537, 707, and 717.
  10. The method of claim 1, wherein the human KRAB domain is a ZIM3 KRAB domain.
  11. The method of claim 10, wherein the ZIM3 KRAB domain comprises the sequence of SEQ ID NO: 495.
  12. The method of claim 1, wherein the target region comprises the sequence of SEQ ID NO: 391.
  13. The method of claim 1, wherein the target region comprises the sequence of SEQ ID NO: 392.
  14. The method of claim 1, wherein the gRNA comprises the nucleic acid base sequence of SEQ ID NO: 1151 or 1249.
  15. The method of claim 2, wherein the DNMT3L domain is a human DNMT3L domain.
  16. The method of claim 2, wherein the DNMT3L domain comprises a sequence selected from the group consisting of SEQ ID NOs: 1032-1035.
  17. The method of claim 16, wherein the DNMT3L domain comprises the sequence of SEQ ID NO: 1033.
  18. The method of claim 1, wherein the dCas9 protein domain comprises a sequence of SEQ ID NO: 12, the DNMT3A domain comprises a sequence of SEQ ID NO: 1029, and the human KRAB domain comprises a sequence of SEQ ID NO: 495.
  19. The method of claim 1, wherein the dCas9 protein domain comprises a sequence with at least 99% sequence identity to the sequence of SEQ ID NO: 12, the DNMT3A domain comprises a sequence with at least 99% sequence identity to the sequence of SEQ ID NO: 1029, and the human KRAB domain comprises a sequence with at least 99% sequence identity to the sequence of SEQ ID NO: 495.
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