Bayesian Data Analysis

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Outline

- 🚺 Normal linear regression (recall)
- Departures from Normality: Poisson data
- Departures from Normality: Binary or Bernoulli Data
- Basic structure of GLMs
- Frequentist inference for GLMs
- Bayesian inference for the Poisson GLM
- Bayesian inference for the Binomial GLM
- Categorical Covariates
- Computation of posterior and predictive probabilities
- Bayesian GLM: from JAGS to INLA



Section 1

Normal linear regression (recall)

Normal linear regression (recall)

We label Y=response variable, x's=covariates. (Y=random variable, y=observed value.)

$$\hookrightarrow \mathbb{E}[Y_i] = \mu_i = \beta_0 + \beta_1 x_{i,1} + \ldots + \beta_p x_{i,p}, i=1,\ldots,n.$$

 \hookrightarrow Abbreviated notation for $\mu_i = \mathbf{x}_i^T \boldsymbol{\beta}$, where

$$\mathbf{x}_{i} = \begin{bmatrix} 1 \\ x_{i,1} \\ \vdots \\ x_{i,n} \end{bmatrix} \quad \boldsymbol{\beta} = \begin{bmatrix} \beta_{0} \\ \beta_{1} \\ \vdots \\ \beta_{n} \end{bmatrix}$$

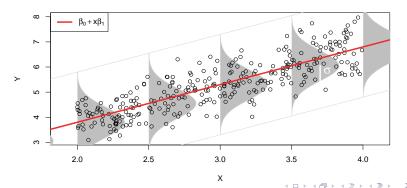
 \hookrightarrow This is a model for the **mean** structure of a random variable Y.

Normal linear regression

 \hookrightarrow We further assumed a probability distribution of Y_i given \mathbf{x}_i and β :

$$Y_i | \mathbf{x}_i, \boldsymbol{\beta}, \sigma^2 \sim \text{Normal} \left(\mathbf{x}_i^T \boldsymbol{\beta}, \sigma^2 \right), i = 1, \dots, n$$

With a common variance σ^2 .



Normal linear regression

 \hookrightarrow We can then estimate β and σ^2 by frequentist (classical) methods, namely maximum likelihood:

$$\hat{\boldsymbol{\beta}} = (X^T X)^{-1} X^T \mathbf{y}$$

$$\hat{\sigma}^2 = \frac{\sum_{i=1}^{n} (y_i - \mathbf{x}_i^T \hat{\boldsymbol{\beta}})^2}{n}$$

where

$$X = \begin{bmatrix} 1 & x_{1,1} & x_{1,2} & \dots & x_{1,p} \\ 1 & x_{2,1} & x_{2,2} & \dots & x_{2,p} \\ \vdots & \vdots & \vdots & \vdots & \vdots \\ 1 & x_{n,1} & x_{n,2} & \dots & x_{n,p} \end{bmatrix}$$

Normal linear regression

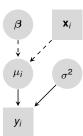
 \hookrightarrow Or by Bayesian estimation of β and σ^2 , with the posterior distribution:

$$p(\boldsymbol{\beta}, \sigma^2 \mid \mathbf{y}) = \frac{p(\boldsymbol{\beta}, \sigma^2, \mathbf{y})}{m(\mathbf{y})} = \frac{f(\mathbf{y} \mid \boldsymbol{\beta}, \sigma^2)\pi(\boldsymbol{\beta}, \sigma^2)}{\int \dots \int f(\mathbf{y} \mid \boldsymbol{\beta}, \sigma^2)\pi(\boldsymbol{\beta}, \sigma^2)d\beta_0 \dots d\sigma^2}$$

$$\propto f(\mathbf{y} \mid \boldsymbol{\beta}, \sigma^2)\pi(\boldsymbol{\beta}, \sigma^2)$$

Normal linear regression

$$\begin{array}{rcl} y_i \mid \mu_i, \sigma^2, \mathbf{x}_i & \sim & \mathsf{N}(\mu_i, \sigma^2), & \mathsf{i} = 1, \dots, \mathsf{n} \\ \mu_i & = & \mathbf{x}_i^T \beta \\ \beta_j & \sim & \mathsf{N}(0, \sigma_{\beta_j}^2), & \mathsf{j} = 0, \dots, \mathsf{p} \\ \sigma & \sim & \mathit{Unif}(0, u_\sigma) \end{array}$$



Section 2

Departures from Normality: Poisson data

Departures from Normality: Poisson data

→ An example is the number of "satellite" male crabs that a female Horseshoe crab attracts in addition to a "primary" male crab, y, compared to the width of the carapace (the larger outer shell), x.

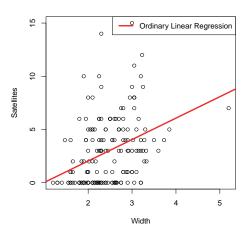




¹pictures NWF; Jane Brockwell

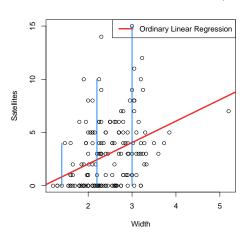
Departures from Normality: Poisson data

Female Horseshoe Crab's Satellites vs Carapace Width



Departures from Normality: Poisson data

Female Horseshoe Crab's Satellites vs Carapace Width



- → Discrete response
- Heteroskedasticity (non-constant variance)

Departures from Normality: Poisson data

- \hookrightarrow Frequentist usual Normal linear regression assumes that the variance of y_i is a constant, i.e., $Var(y_i) = \sigma^2$, but in a Poisson distribution $Var(y_i) = E(y_i) = \mu_i$.
- \hookrightarrow But modeling μ_i as in the Normal model does not ensure positive mean, i.e.

$$\mu_i = \beta_0 + \beta_1 x_i > 0$$

is not quaranteed.

 \hookrightarrow One solution to this is to model the logarithm of μ_i :

$$ln(\mu_i) = \beta_0 + \beta_1 x_i$$

Then

$$\mu_i = \exp(\beta_0 + \beta_1 x_i) > 0$$



Departures from Normality: Poisson data

 \hookrightarrow The likelihood of the Poisson model with mean $\mu_i = \exp(\beta_0 + \beta_1 x_i)$ is:

$$f(\mathbf{y} \mid \beta_0, \beta_1, \mathbf{x}) \quad \propto \quad \prod_i e^{-\mu_i} \mu_i^{y_i} = \prod_i e^{-\exp(\beta_0 + \beta_1 x_i)} \left(\exp(\beta_0 + \beta_1 x_i) \right)^{y_i}$$

 \hookrightarrow Under the frequentist paradigm, maximum (log)likelihood with respect to β_0 and β_1 :

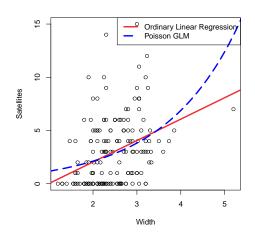
$$\ln(f(\mathbf{y} \mid \beta_0, \beta_1, \mathbf{x})) \quad \propto \quad \sum_i \left(-\exp(\beta_0 + \beta_1 x_i) + y_i(\beta_0 + \beta_1 x_i)\right)$$

 \hookrightarrow Under the Bayesian paradigm, a prior distribution for β_0 and β_1 must be defined and the posterior of the parameters is found in the usual way:

$$p(\beta_0, \beta_1 \mid \mathbf{y}, \mathbf{x}) \propto f(\mathbf{y} \mid \beta_0, \beta_1, \mathbf{x}) \pi(\beta_0, \beta_1)$$

Departures from Normality: Poisson data

Female Horseshoe Crab's Satellites vs Carapace Width



Section 3

Departures from Normality: Binary or Bernoulli
Data

Departures from Normality: Binary or Bernoulli Data

- \hookrightarrow Binary Data: Success or Failure, 1 or 0: $Y \sim \text{Bernoulli}(\theta)$. Thus $\mathbb{E}[Y] = \mu = \theta$.
- \hookrightarrow The probability of success, θ , may be a function of covariates.
- Can you think of some examples of Binary Data and corresponding covariates that could affect θ?
- \hookrightarrow For example, is a randomly chosen Scot in favour of Scottish independence? Let θ = Pr(in favour) = Pr(Y = 1). What factors might affect θ ?
- \hookrightarrow Suppose x=Age is a factor. What might be wrong with the following model?

$$\mu_i \equiv \theta_i = \beta_0 + \beta_1 Age_i$$

Departures from Normality: Binary or Bernoulli Data

 \hookrightarrow Again a solution for this is to construct a 1:1 function of μ , $g(\mu)$, as a linear combination of covariates,

$$g(\mu_i) \equiv g(\theta_i) = \eta_i = \beta_0 + \beta_1 Age_i$$

where the inverse of g maps into the allowable range of θ :

$$0 < [g^{-1}(\beta_0 + \beta_1 Age_i) = \theta_i] < 1$$

 \rightarrow A particular function that can be used for this case is the *logit* function, $\ln(x/(1-x))$:

$$g(\theta_i) = \ln(\theta_i/(1-\theta_i)) = \beta_0 + \beta_1 Age_i$$

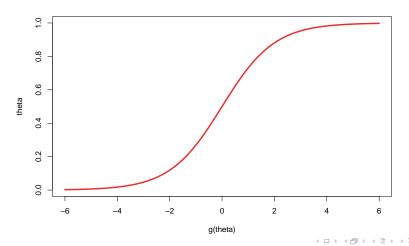
Then

$$0 < [g^{-1}(\beta_0 + \beta_1 Age_i) \equiv \theta_i = \frac{1}{1 + \exp(-\beta_0 - \beta_1 Age_i)}] < 1$$



Departures from Normality: Binary or Bernoulli Data

Inverse-Logit function



Departures from Normality: Binary or Bernoulli Data

 \hookrightarrow The likelihood of the Bernulli model with probability of success $\theta_i = \frac{1}{1 + \exp(-\beta_0 - \beta_1 \chi_i)}$ is:

$$f(\mathbf{y} \mid \beta_0, \beta_1, \mathbf{x}) = \prod_i \theta_i^{y_i} (1 - \theta_i)^{(1 - y_i)} = \prod_i \left(1 + e^{-\beta_0 - \beta_1 x_i} \right)^{-y_i} \left(1 + e^{\beta_0 + \beta_1 x_i} \right)^{-(1 - y_i)}$$

 \hookrightarrow Under the frequentist paradigm, maximum (log)likelihood with respect to β_0 and β_1 :

$$ln(f(\mathbf{y} \mid \beta_0, \beta_1, \mathbf{x})) = \sum_i (y_i(\beta_0 + \beta_1 x_i) - ln(1 + exp(\beta_0 + \beta_1 x_i)))$$

 \hookrightarrow Under the Bayesian paradigm, a prior distribution for β_0 and β_1 must be defined and the posterior of the parameters is found in the usual way:

$$p(\beta_0, \beta_1, | \mathbf{y}, \mathbf{x}) \propto f(\mathbf{y} | \beta_0, \beta_1, \mathbf{x}) \pi(\beta_0, \beta_1)$$



Section 4

Basic structure of GLMs

Basic Structure

Generalised Linear Models extend the Normal Linear Model in two ways:

- 1 The distribution of Y is a member of the Exponential Family Distributions.
- **2** A 1:1 'link' function of $\mathbb{E}[Y] = \mu$, $g(\mu)$, is a linear combination of covariates, $g(\mu) = \mathbf{x}^T \boldsymbol{\beta}$

$$Y \sim f(y \mid \mu, ...)$$

 $g(\mu) = \eta$
 $\eta = \mathbf{x}^T \boldsymbol{\beta}$

Exponential Family of Distributions

- Exponential Family of Distributions include Normal, Poisson, Bernoulli, Binomial, Multinomial, Exponential, Gamma, Beta, Lognormal, Dirichlet.

$$f_Y(y \mid \theta) = h(y) \exp (\eta(\theta) \cdot T(y) - A(\theta))$$

→ Precise Definition for Multiple Parameter Case

$$f_Y(y \mid \theta) = h(y) \exp \left(\sum_{i=1}^s \eta_i(\theta) T_i(y) - A(\theta) \right)$$

- $\hookrightarrow h(y)$ and T(y) are functions of y
- $\hookrightarrow \eta(\theta)$ and $A(\theta)$ are functions of θ
- → More on this topic can be found in Generalised Regression Models course.
- → NOTE: Exponential family distributions have conjugate priors!



Two Examples: Exponential Family of Distributions

$$f_Y(y \mid \theta) = h(y) \exp(\eta(\theta) \cdot T(y) - A(\theta))$$

 \hookrightarrow Poisson(θ) ²

$$f_Y(y \mid \theta) = \frac{\exp(-\theta)\theta^y}{y!} = \frac{1}{y!} \exp(\ln(\theta) \cdot y - \theta)$$

 \hookrightarrow Exponential(θ)³

$$f_Y(y \mid \theta) = \theta \exp(-\theta y) = \exp(-\theta \cdot y - (-\ln(\theta)))$$

²where h(y)=1/y!; $\eta(\theta)=\ln(\theta)$; T(y)=y; and $A(\theta)=\theta$.

³where h(y)=1; $\eta(\theta)=-\theta$; T(y)=y; and $A(\theta)=-\ln(\theta)$.

Link Function

$$g(\mu) = \eta = \mathbf{x}_i^T \boldsymbol{\beta}$$

- \hookrightarrow The Link Function g is not a unique function.
- $\hookrightarrow g$ is usually (but not always) defined so that $g^{-1}\left(\mathbf{x}_{i}^{T}\beta\right)$ yields only allowable values for μ .
 - For example, the "identity" link function $g(\mu)=\mu$ can be specified for a Poisson but that does not ensure that $g^{-1}(\mathbf{x}_i^T\boldsymbol{\beta})=\mathbf{x}_i^T\boldsymbol{\beta}>0$. In practice, some constraints must be added to the estimates $\mathbf{x}_i^T\boldsymbol{\beta}$.

Exponential Family of Distributions and Link Functions

Some examples of link functions for certain distributions:

Distribution
$$g(\mu) = \mathbf{x}^t \beta$$

Normal (μ, σ^2) Identity: $g(\mu) = \mu$

Poisson (μ) Log: $g(\mu) = \ln(\mu)$

Bernoulli (μ) Logit: $g(\mu) = \ln(\mu/(1-\mu))$

C-log-log: $g(\mu) = \ln(-\ln(1-\mu))$

Probit: $g(\mu) = \Phi^{-1}(\mu)$

Probit link: $\Phi(x) = \text{CDF}$ for a Standard Normal random variable:

$$\Phi(x) = \int_{-\infty}^{x} \frac{1}{\sqrt{2\pi}} \exp\left(\frac{-x^2}{2}\right)$$

 $\Phi: \mathcal{R}^1 \to [0, 1]$ and is 1:1, thus $\Phi^{-1}: [0, 1] \to \mathcal{R}^1$.



Distribution

Section 5

Frequentist inference for GLMs

Frequentist Inference with R

- → We will first discuss how to do frequentist inference for GLMs using the R function glm, and then show how to carry out a parallel Bayesian analysis using JAGS.
- → We will look at two examples: a Poisson model (using the Horseshoe crabs data) and a Binomial model

```
glm(formula, family = gaussian, data, weights, subset,
na.action, start = NULL, etastart, mustart, offset, ...)
```

Frequentist Inference with R

 formula is an expression for the mean structure that is similar to that for normal linear regression

 \hookrightarrow While interpretation of the right-hand side is the same as for lm: $\mathbf{x}^T \beta$, the left-hand side is $g(\mu)$ (i.e. the function of $\mathbb{E}[Y \mid \mathbf{x}]$).

```
formula = ``g(\mu)'' \sim x1 + x2
```

Frequentist Inference with R

→ family specifies what exponential family distribution is used:

```
family=gaussian # the default value
family=poisson
family=binomial
```

Each family has an implicit link function, $g(\mu)$ used by default. For example, here are the default link functions made explicit for some distributions:

```
gaussian(link = "identity")
poisson(link = "log")
binomial(link = "logit")
Gamma(link = "inverse")
inverse.gaussian(link = "1/mu^2")
```

One can override these defaults by typing an alternative link function between brackets.
 For example:

```
glm(formula=y ~ x, family=poisson(link=sqrt))
```

Frequentist Inference: Poisson and Horseshoe crabs

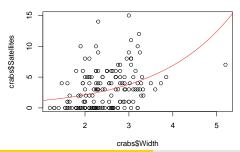
→ The crabs data set has 6 attributes:

```
head(crabs)[,-1]
## Colour Spine Weight Width Satellites
## 1 med both.bad 28.3 3.05 8
## 2 dk.med both.bad 26.0 2.60 4
## 3 dk.med both.bad 25.6 2.15 0
## 4 dk one.bad 21.0 1.85 0
## 5 med both.bad 29.0 3.00 1
## 6 lt.med one.bad 25.0 2.30 3
```

To begin we just focus on modelling the number of satellites as a function of carapace width.

Frequentist Inference: Poisson and Horseshoe crabs

R code for Poisson model



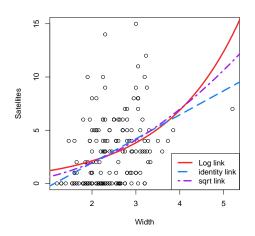
Frequentist Inference: Poisson and Horseshoe crabs

Different link functions:

 Different link functions lead to different estimated parameters with very different interpretations.

Frequentist Inference: Poisson and Horseshoe crabs

Prediction for different link functions:



Frequentist Inference: Binomial and Dead Beetles

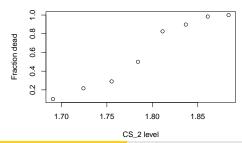
→ Bliss (1935) presented results of an experiment on the fraction of confused flour beetles (Tribolium confusum) that were dead 5 hours after being exposed to gaseous Carbon Disulphide (CS₂). Eight different concentrations of CS₂ were used and about 60 beetles were exposed to each concentration.







→ Fraction dead vs exposure:



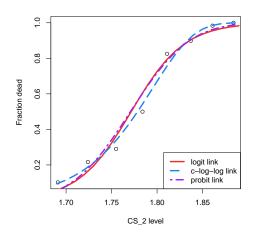


Frequentist Inference: Binomial and Dead Beetles

```
n.exposed \leftarrow c(59, 60, 62, 56, 63, 59, 62, 60)
CS2.level <- c(1.691, 1.724, 1.755, 1.784, 1.811, 1.837, 1.861, 1.884)
v.dead \leftarrow c(6, 13, 18, 28, 52, 53, 61, 60)
beetles <- data.frame (n.exposed, CS2.level, y.dead)
m.logit <- qlm(cbind(y.dead, n.exposed-y.dead) ~ CS2.level,
              family=binomial(link=logit), data = beetles)
m.cloglog<- <pre>glm(cbind(y.dead, n.exposed-y.dead) ~ CS2.level,
                family=binomial(link=cloglog), data = beetles)
m.probit <- qlm(cbind(y.dead, n.exposed-y.dead) ~ CS2.level,
               family=binomial(link=probit), data = beetles)
tabb2<-(round(rbind(coef(m.logit),coef(m.cloglog),coef(m.probit)),3))
row.names(tabb2) <-c("logit", "cloglog", "probit")</pre>
colnames(tabb2) <-c("beta_0", "beta_1 (CS2.level)")</pre>
tabb2
##
     beta 0 beta 1 (CS2.level)
## logit -60.740
                              34.286
## cloalog -39.522
                              22.015
## probit -34.956
                                19.741
```

Frequentist Inference: Poisson and Horseshoe crabs

Prediction for different link functions:



Section 6

Bayesian inference for the Poisson GLM

Bayesian Inference

- \hookrightarrow As usual, for the Bayesian approach we need to specify priors for the unknown parameters. In the case of GLMs, that is primarily priors for the β s in the model for the expected response via the link function formulation: $g(\mu) = \mathbf{x}^T \beta$.
- \hookrightarrow The posterior distribution for β :

$$p(\beta \mid \mathbf{y}) \propto f(\mathbf{y} \mid \beta)\pi(\beta)$$

But now the β will appear in the likelihood via the link function. For example, with a Poisson and a single covariate:

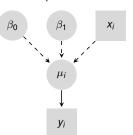
$$\rho(\mathbf{y} \mid \beta_0, \beta_1, \mathbf{x}) \quad \propto \quad \prod_i e^{-\mu_i} \mu_i^{y_i} = \exp\left(-\sum_{i=1}^n \mu_i\right) \prod_i \mu_i^{y_i} \\
= \quad \exp\left(-\sum_i g^{-1}(\beta_0 + \beta_1 x_i)\right) \prod_i \left(g^{-1}(\beta_0 + \beta_1 x_i)\right)^{y_i}$$

Bayesian Inference: Poisson and Horseshoe Crabs

→ The Poisson full model for the Horseshoe Crabs and its DAG representation:

$$y_i \mid \mu_i, \mathbf{x}_i \sim \text{Poisson}(\mu_i), \quad i = 1, ..., n$$

 $\log(\mu_i) = \beta_0 + \beta_1 \textit{Width}_i$
 $\beta_j \sim \text{N}(0, \sigma_{\beta_j}^2), \quad j = 0, 1$



Bayesian Inference: Parameters and Priors

Data Centering

- High correlation between parameters can slow the MCMC. With many correlated parameters, the time necessary to generate enough effective samples for each parameter might be too long.
- \hookrightarrow Centering the covariates usually helps $(x-\overline{x})$, especially by reducing the dependence of β_0 with the rest of the β s.

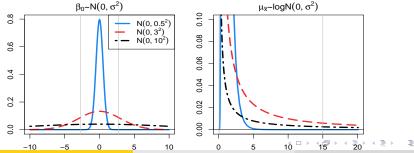
Prior Selection

- → The selection of "non-informative" priors is highly dependent on the possible values of the response variable, the covariates and the link function.
- \hookrightarrow Numerical issues might arise with too sparse priors for the β s and extreme initial values (see, for example, the problem of separation in logistic regression⁴)
- \hookrightarrow On the other hand, too concentrated priors can be too informative or even restrictive to the support of μ .
- \hookrightarrow A useful diagnostic for priors for GLMs is to examine the induced priors for the mean parameter μ for typical and extreme individuals.

⁴Gelman A, Carlin JB, Stern HS, et al. Bayesian Data Analysis, Third Edition, CRC:Press. Chapter 16, section 16.3 🔻 🕠 🤉 🕒

Bayesian Inference: Parameters and Priors

- $\hookrightarrow \log(\mu_i) = \beta_0 + \beta_1(x_i \overline{x})$ or equivalently $\mu_i = e^{\beta_0 + (x_i \overline{x})\beta_1}$
- \hookrightarrow If the covariates are centered, β_0 determines the expected value of y_i for a typical individual $(x_i = \overline{x})$. $\mu_{\overline{x}} = q^{-1} (\beta_0 + \beta_1(\overline{x} \overline{x})) = q^{-1} (\beta_0) = e^{\beta_0}$
- \hookrightarrow For example, usual values for the number of Horseshoe crab satellites (y_i) are between 0 and 15. With a log-link function, we could try to be uninformative by giving high probability density for the induced variable $\mu_{\overline{x}} = e^{\beta_0}$ on the interval [1/15, 15] and surroundings; or equivalentely for β_0 on the interval [- In(15), In(15)].

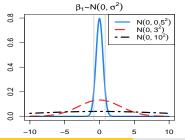


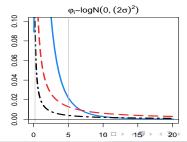
Bayesian Inference: Parameters and Priors

 \hookrightarrow And for the prior of β_1 , with a log-link function, we have that

$$\mu_i = e^{\beta_0 + (x_i - \overline{x})\beta_1} = \mu_{\overline{x}} e^{(x_i - \overline{x})\beta_1} \equiv \mu_{\overline{x}} \varphi_i$$

- \hookrightarrow Therefore β_1 and x_i determine a factor φ_i that indicates, for any individual i, how many times larger μ_i is with respects of $\mu_{\overline{\nu}}$.
- \hookrightarrow Usual values of the width of Horseshoe crabs are between 1 and 5, so $\max |x_i \overline{x}| \approx 2$. For extreme values of x_i , $\mu_{\overline{x}}$ will be multiplied or divided by a factor $\varphi_i = e^{2\beta_1}$.
- \hookrightarrow If we consider that an extreme individual can have a mean at most $\varphi_i = 5$ times the mean of a typical individual, then $\beta_1 = \ln(\varphi_i)/|x_{extreme} \overline{x}|$ would be in $[-\ln(5)/2, \ln(5)/2]$.





Bayesian Inference: Poisson and Horseshoe Crabs

- Using JAGS, we repeat the analysis of the number of Horseshoe Crab Satellites as a function of carapace width, again assuming a Poisson distribution, in a Bayesian framework.
- \hookrightarrow The log link function is used and the covariate will be centred: $\log(\mu) = \beta_0 + \beta_1(Width \overline{Width})$.
- \hookrightarrow The priors for the β s will be Normal(0, τ), where τ =0.01.
- → For JAGS we need three components: a data block, initial values for the parameters, and a
 model statement.

Bayesian Inference: Poisson and Horseshoe Crabs

→ Data block and Initial Values

Bayesian Inference: Poisson and Horseshoe Crabs

→ Model statement

```
crabs.model <- "model {
 # Hyperparameters
 beta.mii.0 <-0
 beta.tau.0 <- 0.01
 # prior
 beta0 ~ dnorm(beta.mu.0, beta.tau.0)
 beta1 ~ dnorm(beta.mu.0,beta.tau.0)
#Likelihood
 for(i in 1:n) {
     # Note: link function on LHS of fn assignment
     log(mu[i]) <- beta0+beta1*(Width[i]-mean(Width[]))</pre>
     Satellites[i] ~ dpois(mu[i])
```

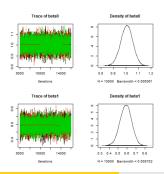
Bayesian Inference: Poisson and Horseshoe Crabs

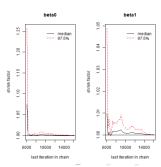
→ Call from R to JAGS:

Bayesian Inference: Poisson and Horseshoe Crabs

 \hookrightarrow Diagnostics before looking at the summary statistics

```
# Trace plots and density
plot(results.crabs.B)
# Brooks-Gelman-Rubin statistic (want a value near 1)
gelman.plot(results.crabs.B)
```





Bayesian Inference: Poisson and Horseshoe Crabs

 \hookrightarrow Other measures of "mixing"- effective sample size given 10,000

```
effectiveSize(results.crabs.B[[1]][,"beta0"])

## var1
## 5254.064

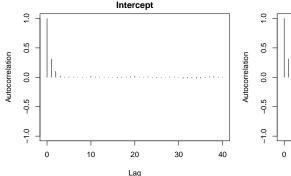
effectiveSize(results.crabs.B[[1]][,"beta1"])

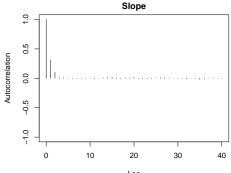
## var1
## 5236.409
```

Bayesian Inference: Poisson and Horseshoe Crabs

→ And autocorrelation within each chain.

```
autocorr.plot(results.crabs.B[[1]][,"beta0"],main="Intercept")
autocorr.plot(results.crabs.B[[1]][,"beta1"],main="Slope")
```





Bayesian Inference: Poisson and Horseshoe Crabs

 \hookrightarrow Now look at the posteriors and compare to MLEs:

- → Monte Carlo error: error in the calculated mean due to random sampling. Can be reduced by increasing the chain length
- → A "rule of thumb" (in the OpenBUGS manual) is to have a chain length such that the MC error is no more than 1/20th the size of the standard deviation of the parameter. ⁵

⁵The reduction is not proportional; in this case, double chain length reduces MC error by 28% $_{\odot}$

Bayesian Inference: Poisson and Horseshoe Crabs

\hookrightarrow Interpretation

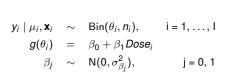
```
# Joinning all the chains in one data.frame
results.crabs.output <- do.call(rbind.data.frame, results.crabs.B)
#interpretation
cat("E[exp(beta0)] is",round(mean(exp(results.crabs.output$beta0)),3))
## E[exp(beta0)] is 2.739
cat("E[exp(beta1)] is",round(mean(exp(results.crabs.output$beta1)),3))
## E[exp(beta1)] is 1.801</pre>
```

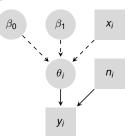
- → 2.739 is the posterior expected number of satellite male crabs for a female with an average carapace width
- → For a unit increase in the carapace width of a female crab, the expected number of her satellite male crabs increases of 1.801 times, i.e. a 80% increase.

Section 7

Bayesian inference for the Binomial GLM

Bayesian Inference: Binomial and Dead Beetles





Bayesian Inference: Parameters and Priors

- $\hookrightarrow g(\theta_i) = ln(\theta_i/(1-\theta_i)) = \beta_0 + \beta_1(x_i \overline{x})$
- $\hookrightarrow \ \beta_0$ is the log odds of beetle death for the mean concentration \overline{x} of CS₂
- \hookrightarrow β_1 is the log odds-ratio (i.e. the expected change in the log odds) of beetle death for a one-unit increase in the concentration x of CS_2
- \hookrightarrow When setting priors we have to consider the minimum distance in x, $x_u x_l$, for which we think the change of probability can happen. We then set a prior that ensures that $|\beta_1|$ can be larger than 10 over that minimum change distance.

$$-5 \approx \ln(0.005/(1 - 0.005)) = \beta_0 + \beta 1(x_I - \overline{x})$$

$$5 \approx \ln(0.995/(1 - 0.995)) = \beta_0 + \beta 1(x_I - \overline{x})$$

- \hookrightarrow For the prior of β_0 , we have to ensure that $|\beta_0|$ can be larger than $(\overline{x} x_m)|\beta_1|^6$ where $x_m = (x_u + x_l)/2$.
- → When in doubt, try vaguer priors and check sensitivity. If results do not change, you are probably being uninformative enough.

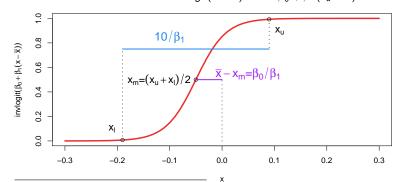
⁶To work out this relationship, start by adding the two equations in this slide → ⟨≡ → □ ✓ ००

Bayesian Inference: Binomial and Dead Beetles

 \hookrightarrow β_1 determines how fast the change in probability happens with respects to x. β_0 determines the horizontal displacement of this change. ⁷

$$-5 \approx logit(0.005) = \beta_0 + \beta 1(x_l - \overline{x})$$

$$5 \approx logit(0.995) = \beta_0 + \beta 1(x_l - \overline{x})$$



⁷Example with $\beta_0 = 1.75$ and $\beta_1 = 35$



Bayesian Inference: Binomial and Dead Beetles

- → For the Bayesian analysis of beetle mortality rate, we need three components: data, initial values, and model statement (with priors and likelihood).
- The election of different initial values allows us to detect possible multimodalities of the model.
- \hookrightarrow If one has an idea of the range of possible values for β_0 and β_1 , five initial values corresponding to the 0.05, 0.25, 0.5, 0.75, 0.95 quantiles are one possibility.
- → Another option is to randomly generate the initial values (from the priors or narrower distributions) using a function that generates a list of random initial values.

Bayesian Inference: Binomial and Dead Beetles

→ Model Statement

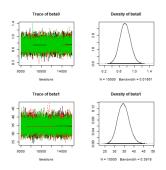
```
beetles.model <- "model {
  #Hyperparameters
  beta.mu.0 <-0
 beta.tau.0 <- 0.0001
  # prior
  beta0 ~ dnorm(beta.mu.0,beta.tau.0)
  beta1 ~ dnorm(beta.mu.0.beta.tau.0)
  #Likelihood
  for(i in 1:n) {
    logit(mu[i]) <- beta0+beta1*(CS2.level[i]-mean(CS2.level[]))</pre>
    y.dead[i] ~ dbin(mu[i],n.exposed[i])
```

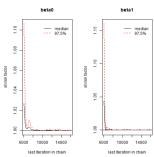
Bayesian Inference: Binomial and Dead Beetles

```
# Run JAGS to the completion of the "adaption" stage
results.beetles.A <- jags.model(file=textConnection(beetles.model),
data=beetles.data, inits=beetles.inits, n.chains=3)
# Burn-in of 5000 iterations
update(results.beetles.A, n.iter=5000)
# Longer run for making inferences, assuming chains have converged
results.beetles.B <- coda.samples(results.beetles.A,
variable.names=c("beta0","beta1"),n.iter=10000)</pre>
```

Bayesian Inference: Binomial and Dead Beetles

```
# Trace plots and density
plot(results.beetles.B)
# Brooks-Gelman-Rubin statistic (want a value near 1)
gelman.plot(results.beetles.B)
```





Bayesian Inference: Binomial and Dead Beetles

 $\hookrightarrow \ \, \text{Effective sample size}$

```
effectiveSize(results.beetles.B[[1]][,"beta0"])

## var1
## 4985.016

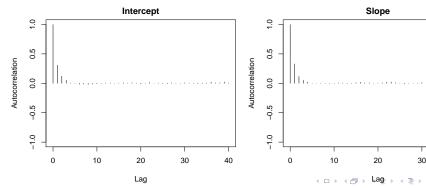
effectiveSize(results.beetles.B[[1]][,"beta1"])

## var1
## 5046.473
```

Bayesian Inference: Binomial and Dead Beetles

 \hookrightarrow Autocorrelation Function (ACF).

```
autocorr.plot (results.beetles.B[[1]][,"beta0"], main="Intercept")
autocorr.plot (results.beetles.B[[1]][,"beta1"], main="Slope")
```



Bayesian Inference: Binomial and Dead Beetles

 \hookrightarrow Now look at the posteriors and compare to MLEs:

```
## Mean SD Naive SE Time-series SE
## beta0 0.7561 0.140 0.0008065 0.0011320
## beta1 0.3464 0.029 0.0001689 0.0002375
```

Bayesian Inference: Binomial and Dead Beetles

 \hookrightarrow Interpretation

```
# Joinning all the chains in one data.frame
results.beetles.output <- do.call(rbind.data.frame, results.beetles.B)
#interpretation
ilogit=function(x) { 1/(1+exp(-x))}
#
cat("E[ilogit(beta0)] is",round(mean(ilogit(results.beetles.output$beta0))
## E[ilogit(beta0)] is 0.68
cat("E[exp(beta1)] is",round(mean(exp(results.beetles.output$beta1)),3))
## E[exp(beta1)] is 1.415</pre>
```

Section 8

Categorical Covariates

Bayesian Analysis: Categorical Covariates

- → Sometimes a covariate is categorical with K possible values. For example, there are K=4 crab carapace colours: light medium, medium, dark medium, and dark.
- \hookrightarrow To include such covariates in a regression model, one must create K-1 "indicator" variables
- → The effect of indicator ("dummy") variables is to shift the model intercept up or down, and there are different ways to construct indicators.

$$g(\mu) = \beta_0 + \beta_1 (Width - \overline{Width}) + \beta_2 I_{LtMed} + \beta_3 I_{Med} + \beta_4 I_{DkMed}$$

Thus, for a light medium crab, $g(\mu) = \beta_0 + \beta_1 (Width - \overline{Width}) + \beta_2$ and similarly for the other cases.

 \hookrightarrow The category without a dummy variable (Dark) will be the reference category. This means that $\exp(\beta_0)$ is now interpreted as the expected number of satellite male crabs for a female crab with a dark carapace of average width.



Bayesian Inference: Categorical Covariates

→ Data block 8

Bayesian Analysis: Categorical Covariates

→ Model Statement

```
crabs.mult.model <- "model {
  #Hyperparameters
 beta.mu.0 <- 0
 beta.tau.0 <-0.001
  # prior
 beta0
              ~ dnorm(beta.mu.0,beta.tau.0)
 beta.width
              ~ dnorm(beta.mu.0,beta.tau.0)
 beta.lt.med ~ dnorm(beta.mu.0,beta.tau.0)
 beta.med
              ~ dnorm(beta.mu.0,beta.tau.0)
 beta.dk.med ~ dnorm(beta.mu.0,beta.tau.0)
  #Likelihood
  for(i in 1:n) {
    log(mu[i]) <- beta0+beta.width*(Width[i]-mean(Width[])) +</pre>
                beta.lt.med*lt.med.Ind[i] + beta.med*med.Ind[i] +
                beta.dk.med*dk.med.Ind[i]
    Satellites[i] ~ dpois(mu[i])
  1 11
```

Bayesian Inference: Categorical Covariates

 \hookrightarrow Call from R to JAGS:

Bayesian Inference: Categorical Covariates

 $\,\hookrightarrow\,$ Now look at the summary of the posteriors:

```
summarv(results.crabs.mult.B)
##
                         SD
                             Naive SE Time-series SE
               Mean
  beta.dk.med 0.0103 0.18561
                            0.0010716
                                         0.0058468
  beta.lt.med 0.4553 0.21197
                            0.0012238 0.0059372
  beta.med 0.2581 0.16901 0.0009758 0.0059801
  beta.width 0.5443 0.06831 0.0003944 0.0005911
  beta0 0.8159 0.15520 0.0008960 0.0054734
##
##
                          25%
                                50%
                 2.5%
                                         75%
                                             97.5%
  beta.dk.med -0.34411 -0.1164 0.006755 0.1345 0.3839
  beta.lt.med 0.04373 0.3120 0.452149 0.5967 0.8727
  beta.med -0.06382 0.1417 0.255108 0.3699 0.5993
## beta.width 0.40846 0.4989 0.544873 0.5907 0.6761
## beta0 0.49562 0.7127 0.821404 0.9232 1.1067
```

 $\,\hookrightarrow\,$ We can immediately spot an increasing pattern towards lighter carapace colours.

Bayesian Inference: Categorical Covariates

\hookrightarrow Interpretation

```
# Joinning all the chains in one data.frame
results.crabs.mult.output <- do.call(rbind.data.frame, results.crabs.mult.B)
#interpretation
cat("E[exp(beta_LightMed)] is",
    round(mean(exp(results.crabs.mult.output$beta.lt.med)),3))

## E[exp(beta_LightMed)] is 1.613
#... similarly for the other colours</pre>
```

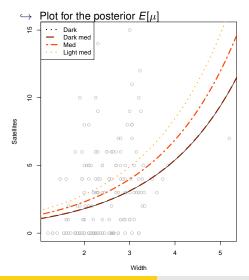


Bayesian Analysis: Categorical Covariates

 \hookrightarrow Plot for the posterior $E[\mu]$ (R code)

```
x < - seq(1, 6, 0.1)
m.crabs <- summary(results.crabs.mult.B)$statistics[,"Mean"]</pre>
plot(crabs$Width, crabs$Satellites, col="gray", xlab="Width", ylab="Satellites")
PostMean mu=matrix(NA, 4, length(x))
for(k in 1:length(x)){
PostMean mu[1,k]=mean(exp(results.crabs.mult.output$beta0+
results.crabs.mult.output$beta.width*(x[k]-mean(crabs$Width))))
PostMean_mu[2,k]=mean(exp(results.crabs.mult.output$beta0+
results.crabs.mult.output$beta.width*(x[k]-mean(crabs$Width))+
results.crabs.mult.output$beta.dk.med))
# . . .
lines (x, PostMean mu[1,], lwd=4, ltv=3)
lines(x,PostMean mu[2,],col="orangered4", lwd=3, lty=5)
legend("topleft", c("Dark", "Dark med", "Med", "Light med"), col = c("black",
"orangered4", "orangered", "orange"), lty = c(1,5,6,3), lwd = 3)
```

Bayesian Analysis: Categorical Covariates



Section 9

Computation of posterior and predictive probabilities

Bayesian Analysis: Direct calculations using the simulations

```
library(runjags)
fit.crabs <- as.data.frame(combine.mcmc(results.crabs.mult.B))
# or use function do.call as shown in slide 71</pre>
```

 \hookrightarrow It can be used to calculate probabilities, e.g. $Pr(\beta_{Med} > 0|y)$.

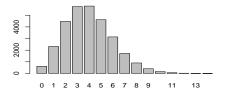
```
mean(fit.crabs$beta.med>0)
## [1] 0.9388333
```

 \hookrightarrow One can also obtain simulations for the predictive distribution $\int f(y^*|\beta)p(\beta|y)d\beta$ by plugging-in posterior samples of β into the likelihood of the new observation y^* , for example $p(y^*|y, Width^* = 3, Colour^* = Med)$:

Bayesian Analysis: Direct calculations using the simulations

 \hookrightarrow In this way we can plot the distribution $p(y^*|y, Width^* = 3, Colour^* = Med)$

```
barplot (table (y_star) )
```



 \hookrightarrow And calculate predictive probabilites such as $Pr(y^* \ge 3|y, \textit{Width}^* = 3, \textit{Colour}^* = \textit{Med})$

```
mean (y_star>=3)
## [1] 0.7530667
```

Section 10

Bayesian GLM: from JAGS to INLA

- \hookrightarrow We repeat the analysis of the crab data with categorical covariates.
- → In INLA, the log link function is the only available option for Poisson likelihood. This is the
 default link, so it does not have to be specified explicitly
- → In INLA, the K-1 indicators are automatically created by the system when we use categorical variables in the regression formula

From JAGS to INLA: Categorical Covariates

→ Now look at the summary of the posteriors:

```
summary (m2.I)
##
                      sd 0.025quant 0.5quant 0.975quant
                                                     mode kld
               mean
             0.828 0.149
                             0.523
                                     0.832
                                              1.109
   (Intercept)
                                                     0.841
  Width.ctr 0.546 0.068
                             0.410 0.547
                                              0.678
                                                     0.548
  Colourdk.med 0.003 0.180
                            -0.344 0.000
                                              0.364 -0.004
  Colourlt.med 0.453 0.208
                            0.045 0.452
                                              0.863 0.452
## Colourmed 0.248 0.163 -0.062 0.244
                                              0.578 0.237
```

→ The summary statistics are essentially identical to what we got from JAGS.

- \hookrightarrow Now we are going to plot the posterior mean of μ , as a function of the width, for a set of equally spaced widths, separately for each 4 colours.
- → This can be done by including new rows in the dataset with the response variable (Satellites) set to NA.

```
#Create a sequence of widths at which the posterior means
#of the fitted values is computed
x = seq(1, 6, 0.1)
lx=length(x)
#Create new rows in the dataset for these widths.
#for all 4 colours, with the response Satellites set to NA
newdata <- rbind(data.frame(Width=x,Colour=as.factor("dk")),</pre>
        Satellites=NA), data.frame(Width=x, Colour=as.factor("dk.med"),
Satellites=NA), data.frame(Width=x,Colour=as.factor("med"),
Satellites=NA), data.frame(Width=x, Colour=as.factor("lt.med"),
Satellites=NA))
#Join the new rows with the original dataframe,
#and include the centered width covariate
newdata=rbind (newdata, data.frame (Satellites=crabs$Satellites,
              Width=crabs$Width, Colour=crabs$Colour))
newdata$Width.ctr=newdata$Width-mean(crabs$Width)
```

- → We fit the model in INLA. Note that we need to set control.predictor = list(compute = TRUE, link=1)
- \hookrightarrow **link=1** tells INLA to compute the marginals and means of the fitted values μ_i for the rows where the response is set to NA
- \hookrightarrow normally, when the response is set to NA, the statistics in **inla.model\$summary.fitted.values** will refer to the linear predictor η_i , and not μ_i

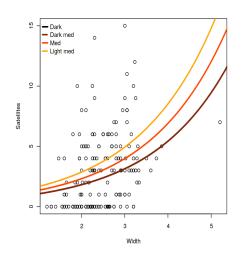
```
#whenever we use a model with link function that is not the identity
#and we want to compute the posterior marginal and mean of mu_i
#and not eta_i, we need to set link=1 in control.predictor

m3.I <- inla(formula = Satellites ~ Width.ctr+Colour, family="poisson",
data=newdata, control.fixed=prior.beta,
control.predictor = list(compute = TRUE, link=1),
control.compute=list(config=TRUE))</pre>
```

From JAGS to INLA: Categorical Covariates

→ We extract the posterior mean of the fitted values by m3.l\$summary.fitted.values\$mean, and plot them.

```
#Rows 1:1x contain the results for colour Light med,
#rows lx+1:2*lx contain the results for colour Med, etc.
fittedvaluesm=m3.I$summary.fitted.values$mean
plot (crabs$Width, crabs$Satellites, col="black", xlab="Width",
vlab="Satellites")
legend("topleft", c("Dark", "Dark med", "Med", "Light med"),
col = c("black", "orangered4", "orangered", "orange"),
       lty = c(1, 5, 6, 3), lwd = 3, bty = "n")
lines(x, fittedvaluesm[1:lx], col="black",lwd=3)
lines (x, fittedvaluesm[(lx+1):(2*lx)], col="orangered4", lwd=3, lty=5)
lines (x, fittedvaluesm[(2*lx+1):(3*lx)], col="orangered", lwd=3, lty=6)
lines(x, fittedvaluesm[(3*lx+1):(4*lx)], col="orange", lwd=3, ltv=3)
```



From JAGS to INLA: posterior and predictive probabilities

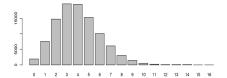
 \hookrightarrow We look at the posterior and posterior predictive probabilities

```
#We have already included the Width=3, Colour = Med
#in the newdata dataframe in row 51*2+21=123
#We only include this single linear predictor
#and the regression coefficient for colour med
#in the output of inla.posterior.sample by including
#selection=list(Predictor=123, Colourmed=1)
nbsamp=100000
crab.samples=inla.posterior.sample(n=nbsamp, result=m3.I,
            selection=list (Predictor=123, Colourmed=1))
#POSTERIOR PROBABILITY OF beta_MED > 0
beta.med.samples=inla.posterior.sample.eval(function(...) {Colourmed},
crab.samples)
mean (beta.med.samples>0)
#0.93691
```

From JAGS to INLA: posterior and predictive probabilities

```
\rightarrow and P(y^*|y, width^* = 3, Colour^* = Med)
```

```
predictor.samples=inla.posterior.sample.eval(function(...) {Predictor},
crab.samples)
post.pred.samples=rpois(n=nbsamp, lambda=exp(predictor.samples))
barplot (table (post.pred.samples) )
mean (post.pred.samples>=3)
#0.75814
```



The R code containing all of the analyses in Lecture 4 using JAGS and INLA is available on Learn.

Bayesian Analysis: further readings on Bayesian GLMs

- → Reich BJ, Ghosh SK. Bayesian Statistical Methods. CRC press, 2019. Chapter 4, Section 4.3 and subsections.
- Gelman A, Carlin JB, Stern HS, et al. Bayesian Data Analysis, Third Edition. CRC press, 2014. Chapter 16, Introduction, Section 16.1, 16.2 "Offsets" subsection, and 16.4.