Title: Interactions between color and *Symbiodinium* in *Montipora capitata* across environmental gradients in Kāne‘ohe Bay, O‘ahu, Hawai‘i

Running Page Head: Distribution of *Symbiodinium* and Color

Authors: T. Innis1, R. Cunning2, R. Ritson-Williams2, C. B. Wall2, R. D. Gates2

1Northeastern University, Marine Science Center, 430 Nahant Rd, Nahant, MA 01908, USA

2University of Hawai‘i at Mānoa, Hawai‘i Institute of Marine Biology, PO Box 1346, Kāne‘ohe, HI 96744, USA

\*Corresponding author: teegan.innis@gmail.com

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**ABSTRACT**

Scleractinian corals form mutualistic symbioses with genetically and functionally diverse photosynthetic dinoflagellates in the genus *Symbiodinium* that confer varying levels of performance and stress-tolerance to their hosts. Variation in bleaching susceptibility among conspecifics is linked to symbiont community variability, yet the factors driving dissimilar symbiont associations within a population are poorly understood. To investigate potential drivers, we characterized symbiont communities in *Montipora capitata* (N = 707 colonies) across the biophysical regions, reef types, and depth range of Kāne‘ohe Bay (Hawai‘i, USA), where this dominant reef-builder associates with *Symbiodinium* clades C (C31) and/or D (D1a) and occurs as two color morphs (brown and orange). The prevalence of C- and D-dominated colonies (69 vs. 41 % of colonies respectively) did not vary across bay regions, reef types or among individual reefs. However, there was a significant influence of depth on both symbiont-dominance and color, with orange, D-dominated colonies more prevalent in shallower, high-light environments, and brown, C-dominated colonies more prevalent in deeper, low-light environments. Though either color morph could be dominated by either clade, brown colonies were more likely to be C-dominated regardless of depth, while orange colonies were more likely to be D-dominated above 3.53 m and C-dominated below, revealing a significant interaction between host color and symbiosis ecology. This work reveals that symbiont community variability may arise either from holobiont phenotypic plasticity or differential survival across light gradients, with significant implications for predicting coral responses to climate change stressors and informing potential management applications such as selective breeding of robust corals.

**INTRODUCTION**

The persistence of coral reefs is essential to sustain ecosystem services worth billions of dollars annually, including shoreline protection, critical habitat formation, and support of tourist economies (Barbier et al. 2011). Reef construction is made possible by corals’ mutualistic endosymbioses with photosynthetic dinoflagellates (*Symbiodinium* spp.; Muscatine & Porter 1977), yet the vulnerability of these symbioses to collapse at high temperatures (coral ‘bleaching’; Jokiel & Coles 1977) forecasts a bleak future for coral reefs in warming oceans (Hoegh-Guldberg et al. 2007).  
 However, significant variability exists in coral susceptibility to thermal stress, even among individual colonies. Indeed, one colony may bleach severely while neighboring conspecifics appear unaffected, which in many cases is due to association with different types of *Symbiodinium* (Glynn et al. 2001; Jones et al. 2008; Cunning et al. 2016)*.* Heterogeneous symbiont associations are observed among individuals in many, but not all, coral populations including *Pocillopora* spp. in the eastern Pacific (Glynn et al. 2001), *Orbicella* spp. in the Caribbean (Thornhill et al. 2006), *Acropora* spp. in Taiwan (Chen et al. 2005) and Australia (Jones et al. 2008), and *Montipora capitata* in Hawai‘i (Stat et al. 2011; Cunning et al. 2016). Different *Symbiodinium* types (Pochon & Gates 2010) display different capacities to utilize light energy (Rowan 2004; Iglesias-Prieto et al. 2004) and nutrients (Baker et al. 2013), which may lead to differences in photosynthate supply (Cantin et al. 2009), host growth (Little et al. 2004), and bleaching susceptibility (Glynn et al. 2001). Therefore, understanding the factors that drive symbiont community assembly within (and variability among) individuals is critical to understand coral performance and function.

In coral populations where multiple *Symbiodinium* types are observed, they often co-occur in individual colonies, where one type is typically numerically dominant with background populations of the other(s) (Silverstein et al. 2012). The presence of multiple symbionts implies a potential for “shuffling” (i.e. temporal change in the relative abundance of types) in response to changing environmental conditions (Rowan & Knowlton 1995; Jones et al. 2008), and suggests that a colony’s symbiont community structure may reflect competition or differential performance of symbionts under prevailing abiotic conditions. Indeed, temperature and light may influence which symbiont establishes dominance in a mixed community (Cunning et al. 2015), and thus spatial variability in symbiont dominance within populations may reflect variability in the external environment (Sampayo et al. 2007; Bongaerts et al. 2015).

However, frequent occurrence of neighboring conspecifics with dissimilar symbionts (Cunning et al. 2016) suggests factors other than the environment are also important. A colony’s symbionts may reflect which cells were transmitted to the larva from the mother colony (Padilla-Gamino et al. 2012), or may be otherwise genetically determined (Poland and Coffroth 2016). Alternatively, the symbiont community may reflect an emergent interaction of abiotic and biotic factors, as particular host traits may directly or indirectly influence symbiont community dynamics. One such trait is colony color morph, which arises from host-produced fluorescent proteins (Kelmanson & Matz 2003) that modify intracellular light environments (Salih et al. 2000) and may correlate with symbiont type (Frade et al. 2008; LaJeunesse and Thornhill 2013).

Here, we investigate potential biotic and abiotic drivers of *Symbiodinium* community variability in *Montipora capitata* inKāne‘ohe Bay (Hawaiʻi, USA), where this dominant reef-builder may harbor clade C (C31; bleaching-susceptible) and/or D (D1a; bleaching-resistant) symbionts (LaJeunesse & Thornhill 2011, Stat et al. 2011, Cunning et al. 2016) and occur as distinct brown and orange color morphs (LaJeunesse, Thornhill, et al. 2004). The mosaic of patch and fringing reefs in Kāneʻohe Bay occurs across regions of varying of oceanic influence, water turnover time, and freshwater input (Smith et al. 1981; Bahr et al. 2015), while shallow depths and high turbidity produce strong vertical irradiance clines. These environmental gradients create an ideal study system to explore the interactions of environment, color morph, and *Symbiodinium* community variability. Elucidating these patterns may provide significant insight into symbiosis ecology and the power to predict distributions of individuals with differential responses to environmental change.

**MATERIALS AND METHODS**

*Study Design and Sampling*

Colonies of *Montipora capitata* in Kāne‘ohe Bay were tagged and sampled between 7 June and 12 August 2016. Sampling effort was distributed among three bay regions (north, central, and south) and across five patch reefs and three fringing reefs within each region. At each reef, 10 colonies from the windward slope, reef top, and leeward slope (patch reefs only) were sampled, for a total of 30 samples from each patch reef and 20 samples from each fringe reef. Additional colonies below ~10m were targeted across all sites to increase replication in deeper environments, and a deeper submerged patch reef was also sampled south of the Hawai‘i Institute of Marine Biology (HIMB) (n = 25 reefs; Fig. 1). In total, 16 patch reefs and 9 fringing reefs were sampled across Kāne’ohe Bay resulting in 25 collection sites and a sample size of 707 colonies.

To ensure random sampling within each targeted area (e.g. reef top, slope), weights with attached floats were thrown from the surface across a distance of approximately 20 m. The closest colony of *M. capitata* to each float was tagged and sampled, and colony depth was recorded using a depth gauge. Photographs with color standard were taken of each colony to later assess color morph of each colony (Fig. 2). A tissue biopsy (<1 cm2) was collected from the upper surface of each colony.

*Sample Processing*

Tissue biopsies were placed in 500 μL DNA buffer (5M NaCl, 0.5M EDTA) with 1% sodium dodecyl sulfate, and DNA was extracted following a CTAB-chloroform protocol (dx.doi.org/10.17504/protocols.io.dyq7vv) at HIMB. The relative abundances of clades C and D *Symbiodinium* within each DNA sample were measured using quantitative PCR assays targeting actin loci specific to each clade (Cunning & Baker 2013). All samples were assayed in duplicate 10 μL reactions for 40 cycles on a StepOnePlus platform (Applied Biosystems) with a fluorescence threshold of 0.01 and a baseline interval of cycles 15 - 22. The ratio of clade C to D in each sample was calculated by the formula C:D=2^(CTC-CTD), where CTC and CTD are the threshold cycles for clade C and D amplifications, normalized for fluorescence intensity and locus gene copy number (Cunning et al. 2016). Symbiont clades detected in only one technical replicate were not considered as present in a colony.

*Data Analysis*

Each colony was categorized as C- or D-dominated based on its numerically most abundant symbiont clade. The proportions of clades C and D present in colonies dominated by each clade and color morph were compared using Chi-Squared analyses. Chi-Squared tests were then used to assess differences in the proportion of C- vs. D-dominated colonies, orange vs. brown colonies and one vs. two symbiont clade assemblages between bay regions (northern, central and southern), reef types (patch vs. fringe), and each individually sampled reef. Logistic regressions were used to model the probability of C- vs. D-dominance and orange vs. brown color as a function of depth, with depth data corrected for differences in mean sea level using daily tide tables for Moku o Lo‘e, Kāne‘ohe Bay at 6-minute intervals (NOAA, "Tide Predictions - MOKU O LOE 1612480 Tidal Data Daily View - NOAA Tides & Currents", 2016). Spatial autocorrelation among reefs in the composition of color morph and dominant symbiont association (based on Bray-Curtis dissimilarity) was assessed by a Mantel Test after a Multinomial Logistic Regression was performed to account for the influence of depth. All analyses were performed in R v.3.2.2 (R Core Team 2016).

**RESULTS**

*Overall prevalence of symbionts and color morphs*

Samples of *Montipora capitata* contained *Symbiodinium* clades C and D either exclusively or in combination. Across all samples, 53 % contained only clade C, 1.2 % contained only clade D and 45.8 % contained a mixture of clades C and D. Clade C was the dominant symbiont in 61 % of colonies, and of these C-dominated colonies, most (86.6 %) occurred without background clade D. The remaining 49 % of colonies were dominated by clade D; but in striking dissimilarity, D rarely occurred without background clade C (3.3% of D-dominated colonies; Fig. 3).

Of all colonies sampled, 54.5 % were orange and 45.5 % were brown. There was a significant correlation between color and dominant symbiont (*p* < 0.001), with clade C-dominance in 95.3 % of brown colonies but only 33.0 % of orange colonies (Fig. 4). The relative abundance of all four color-symbiont combinations among colonies sampled throughout Kāneʻohe Bay was 43.3 % Brown-C, 36.5 % Orange-D, 18.0 % Orange-C, and 2.2 % Brown-D.

*Spatial Distribution*

Depth proved to be the significant driving factor for symbiont dominance (*p* < 0.001) and color morph (*p* < 0.001) among colonies of *M. capitata*. Colonies were more likely to be C-dominated at depths below 1.29 m, while D-dominated colonies were more common at depths above 1.29 m. A higher probability of a colony being orange was observed in colonies at depths < 3.64 m, whereas brown color morphs were more prevalent at depths > 3.64 m. Brown colonies were more likely to be dominated by clade C *Symbiodinium* across all depths, but orange colonies were more likely to be dominated by clade D at depths shallower 2.75 m and clade C at depths deeper than 2.75 m (*p* < 0.001; Fig. 5).

The proportion of colonies that were C- vs. D-dominated did not differ among regions of the bay (*p* = 0.14), individual reefs (*p* = 0.07) or between patch and fringing reef types (*p* = 0.37) initially. However, after adjusting for the influence of depth on spatial patterns, bay region was a significant predictor of a colony’s likelihood of being C- or D-dominated and brown or orange in coloration concurrently (*p* < 0.01; Fig. 6). D-dominated brown colonies were more abundant in the southern (*n* = 9) bay area than they were central (*n* = 3) and northern (*n* = 3) regions, though this trend is likely due to the trifling subset of samples with only 15 brown colonies dominated by clade D.

**DISCUSSION**

*Symbiodinium* communities in colonies of *Montipora capitata* were highly variable with some hosting clade C or clade D exclusively, and others hosting mixtures of both clades in different relative proportions. Most mixtures were heavily dominated by one clade with low ‘background’ amounts of the other (Fig. 4), though more evenly mixed assemblages also occurred. While C and D each dominated colonies frequently (61 and 49 %, respectively), they showed very different patterns at ‘background’ levels: clade D was usually absent from C-dominated colonies, while clade C was almost always present in D-dominated colonies. This distribution may indicate that clade C often competitively excludes clade D (Baker et al. 2013), but D almost never excludes C. Such a competitive dynamic is consistent with observations in other coral species that associate with both C- and D-types, where the C-type is a better performer than the D-type under favorable conditions (Cantin et al. 2009; Baker et al. 2013). Nevertheless, the presence of D-dominated colonies suggests D may outperform C under certain conditions or microenvironments and forms a stable community once established.

Differential performance of these symbionts is also supported by the contrasting depth distributions of C- and D-dominance (Fig. 5). While colonies dominated by either symbiont were found at all depths, shallow colonies were more likely to be D-dominated (~70 % < 1 m), whereas deeper colonies were much more likely to be C-dominated (~99 % > 10 m). While this depth zonation may correlate with gradients in temperature, freshwater lensing, or suspended particles, vertical light attenuation is likely to be the strongest driving factor. Indeed, particular *Symbiodinium* types may be adapted to particular light regimes (Iglesias-Prieto et al. 2004), and clade D specifically may tolerate high light (Yuyama et al. 2016). Thus, the relative performance of C and D types may change along a light gradient, leading to differential outcomes in symbiont dominance. While depth zonation in symbiont dominance has been observed in many coral species across tens of meters (Bongaerts et al. 2015), the shallow transition from a higher frequency of D- to C-dominated colonies observed here (1.29 m) likely reflects the high turbidity and rapid attenuation of light in Kāne‘ohe Bay (Grigg 1965).

While depth alone was a significant predictor of clade C- or D-dominance, there was also a significant interactive effect of depth and colony color morph. While deeper colonies (> 10 m) of both colors had < 5 % chance of being D-dominated, this probability rose to 80 % in shallow orange colonies, but only to 20 % in shallow brown colonies. Thus, symbiont community structure may be more responsive to the environment in orange colonies compared to brown. Alternatively, differential survival of different color-symbiont pairings across a depth gradient may give rise to these distributions. Overall, these patterns do not support a fixed relationship between color morph and symbiont type (LaJeunesse et al. 2004), but rather complex interactions between the environment, colony color, and symbiont community structure (Frade et al. 2008).

However, after accounting for the strong influence of depth, little spatial variability in these traits was evident. No significant spatial differences were observed across regions of the bay, individual reefs or reef types, consistent with the lack of spatial variation in symbiont prevalence previously observed across sites and regions (Stat et al. 2011). The proportions of different color-symbiont pairings did not change throughout the bay despite strong differences in oceanic influence and water turnover time (Smith et al. 1981; Lowe et al. 2009). Temperature and light are similar across the bay (ZENODO), although slightly higher light in the south end of the bay (Cunning et al. 2016) correlated with the slightly higher abundance of brown-D colonies. Moreover, there were no interactions between geography and depth, indicating that the strong influence of depth on these patterns was consistent throughout the bay. Overall, the overwhelming influence of depth on these distributions suggests that light is the most important factor driving the relationship between symbiont dominance and color.

Color morph variation in other species has been linked to differential expression of fluorescent proteins (Kelmanson & Matz 2003), indicating the possibility for epigenetic or environmental control. In this way, differences in irradiance may drive differential expression of fluorescent pigments (Takabayashi & Hoegh-Guldberg 1995), which may subsequently create an internal spectral niche (Salih et al. 2000) that favors a certain symbiont (Frade et al. 2008). Alternatively, different light levels may directly mediate the competitive dominance of a certain symbiont, which may subsequently modify colony color with its own pigments (Oswald et al. 2007) or by mediating host pigment expression (DeSalvo et al. 2010). Moreover, other microbial members of the holobiont may contribute to color: indeed, bacterial communities may differ between orange and brown morphs of *M. capitata* (Shore-Maggio et al. 2015), and cyanobacterial phycoerithrin has been linked to orange coloration in *Montastraea cavernosa* (Lesser et al. 2004).

Disentangling the complex relationships between the environment, symbiont communities, and color will require additional manipulative and experimental work, particularly to address whether these traits are plastic and dynamic within individuals. This information is critical to determine whether the primary mechanism driving the ecological distributions of these traits occurs at the colony level, where dynamic color and symbiont communities within each colony are shaped by the environment (phenotypic plasticity, or ‘polyphenism’), or at the population level, where genetically constrained color and symbiont dominance drive differential survival across environmental gradients (‘polymorphism’, *sensu* Kelmanson & Matz 2003). A third possibility, in which these traits are environmentally-determined in early ontogeny (e.g., during larval development or settlement) and remain fixed throughout a colony’s lifetime, involves both of these scenarios, and potentially implicates epigenetic control and/or alternate stable states. We suggest that observations of mixed symbiont assemblages in varying proportions (Fig. 4) are more consistent with phenotypic plasticity, and although direct evidence for temporal dynamism in these traits in *M. capitata* individuals is currently lacking, colonies of other coral species have shown environmentally-induced changes in both symbiont dominance (Baker 2001) and color (Todd et al. 2002).

The ecological distributions of color and symbiont dominance identified here suggest that multiple symbiont types may compete within individual corals with outcomes determined interactively by attributes of the host (color) and the abiotic environment (light). Understanding the drivers of symbiont dominance and variability within and among colonies helps predict disturbance ecology and bleaching susceptibility; the high probability of clade C dominance in deeper, brown colonies suggests that these are likely to be most sensitive in response to a given level of environmental stress, while shallow, orange colonies may be the most robust. However, shallow, brown, C-dominated colonies, as ecological outliers, may also be specially adapted to high light environments. These patterns can inform management practices such as selective breeding efforts by helping to identify the most robust individuals.

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**LITERATURE CITED**

Bahr KD, Bruno J, Jokiel PL, Toonen RJ (2015) The unnatural history of Kane’ohe Bay: coral reef resilience in the face of centuries of anthropogenic impacts. PeerJ

Baker, A. C. (2001). Ecosystems: reef corals bleach to survive change. *Nature*, *411*(6839), 765-766

Baker, D. M., Andras, J. P., Jordán-Garza, A. G., & Fogel, M. L. (2013). Nitrate competition in a coral symbiosis varies with temperature among Symbiodinium clades. *The ISME journal*, *7*(6), 1248-1251

Barbier EB, Hacker SD, Kennedy C, Koch EW, Stier AC, Silliman BR (2011) The value of estuarine and coastal ecosystem services. Ecol Monogr 81:169–193

Bongaerts P, Carmichael M, Hay KB, Tonk L, Frade PR, Hoegh-guldberg O (2015) Prevalent endosymbiont zonation shapes the depth distributions of scleractinian coral species. Proc R Soc Open Sci 2:1–11

Cantin NE, Oppen MJH Van, Willis BL, Mieog JC, Negri AP (2009) Juvenile corals can acquire more carbon from high-performance algal symbionts. Coral Reefs 28:405–414

Chen, C. A., Wang, J. T., Fang, L. S., & Yang, Y. W. (2005). Fluctuating algal symbiont communities in Acropora palifera (Scleractinia: Acroporidae) from Taiwan. *Marine Ecology Progress Series*, *295*, 113-121

Cunning R, Baker AC (2013) Excess algal symbionts increase the susceptibility of reef corals to bleaching. Nat Clim Chang 3:259–262

Cunning, R., Silverstein, R. N., & Baker, A. C. (2015). Investigating the causes and consequences of symbiont shuffling in a multi-partner reef coral symbiosis under environmental change. In *Proc. R. Soc. B* 282:20141725

Cunning R, Ritson-Williams R, Gates RD (2016) Patterns of bleaching and recovery of Montipora capitata in Kaneohe Bay, Hawaii, USA. Mar Ecol Prog Ser 551:131–139

DeSalvo, M. K., Estrada, A., Sunagawa, S., & Medina, M. (2012). Transcriptomic responses to darkness stress point to common coral bleaching mechanisms. *Coral Reefs*, *31*(1), 215-228

Frade PR, Englebert N, Faria J, Visser PM, Bak RPM (2008) Distribution and photobiology of Symbiodinium types in different light environments for three colour morphs of the coral Madracis pharensis: Is there more to it than total irradiance? Coral Reefs 27:913–925

Frade PR, Jonghe F de, Vermuelen F, Bleuswuk J van, Bak RPM (2008) Variation in symbiont distribution between closely related coral species over large depth ranges. Mol Ecol 17:691–703

Glynn, P. W., Maté, J. L., Baker, A. C., & Calderón, M. O. (2001). Coral bleaching and mortality in Panama and Ecuador during the 1997–1998 El Niño–Southern Oscillation event: spatial/temporal patterns and comparisons with the 1982–1983 event. *Bulletin of Marine Science*, *69*(1), 79-109

Grigg RW (1965) Ecological studies of black corals in Hawaii. Pacific Sci 19:244–259

Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., Gomez, E., ... & Knowlton, N. (2007). Coral reefs under rapid climate change and ocean acidification. *science*, *318*(5857), 1737-1742

Iglesias-Prieto R, Beltrá N VH, Lajeunesse TC, Reyes-Bonilla H, Thomé PE (2004) Different algal symbionts explain the vertical distribution of dominant reef corals in the eastern Pacific. Proc R Soc London B Biol Sci 271:1757–1763

Jokiel, P. L., & Coles, S. L. (1977). Effects of temperature on the mortality and growth of Hawaiian reef corals. *Marine Biology*, *43*(3), 201-208

Jones AM, Berkelmans R, Oppen MJH Van, Mieog JC, Sinclair W (2008) A community change in the algal endosymbionts of a scleractinian coral following a natural bleaching event: field evidence of acclimatization. Proc R Soc London B Biol Sci 275:1359–1365

Kelmanson I V., Matz M V. (2003) Molecular basis and evolutionary origins of color diversity in great star coral Montastraea cavernosa (Scleractinia: Faviida). Mol Biol Evol 20:1125–1133

LaJeunesse TC, Bhagooli R, Hidaka M, DeVantier L, Done T, Schmidt GW, Fitt WK, Hoegh-Guldberg O (2004) Closely related Symbiodinium spp. differ in relative dominance in coral reef host communities across environmental, latitudinal and biogeographic gradients. MEPS 284:147–161

LaJeunesse TC, Thornhill DJ (2011) Improved resolution of reef-coral endosymbiont (Symbiodinium) species diversity, ecology, and evolution through psbA non-coding region genotyping. PLoS One 6

LaJeunesse TC, Thornhill DJ, Cox EF, Stanton FG, Fitt WK, Schmidt GW (2004) High diversity and host specificity observed among symbiotic dinoflagellates in reef coral communities from Hawaii. Coral Reefs 23:596–603

Lesser, M. P., Mazel, C. H., Gorbunov, M. Y., & Falkowski, P. G. (2004). Discovery of symbiotic nitrogen-fixing cyanobacteria in corals. *Science*, *305*(5686), 997-1000

Little AF, Oppen M van, Willis BL (2004) Flexibility in algal endosymbiosis: shapes growth in reef corals. Science (80- ) 304:1492–1494

Lowe, R. J., Falter, J. L., Monismith, S. G., & Atkinson, M. J. (2009). Wave-driven circulation of a coastal reef-lagoon system. *Journal of Physical Oceanography*, *39*(4), 873-893

Muscatine, L., & Porter, J. W. (1977). Reef corals: mutualistic symbioses adapted to nutrient-poor environments. *Bioscience*, *27*(7), 454-460

Oswald, F., Schmitt, F., Leutenegger, A., Ivanchenko, S., D'Angelo, C., Salih, A., ... & Matz, M. V. (2007). Contributions of host and symbiont pigments to the coloration of reef corals. *Febs Journal*, *274*(4), 1102-1122

Padilla-Gamiño, J. L., Pochon, X., Bird, C., Concepcion, G. T., & Gates, R. D. (2012). From parent to gamete: vertical transmission of Symbiodinium (Dinophyceae) ITS2 sequence assemblages in the reef building coral Montipora capitata. *PLoS One*, *7*(6)

Pochon X, Gates RD (2010) A new Symbiodinium clade (Dinophyceae) from soritid foraminifera in Hawai’i. Mol Phylogenet Evol 56:492–497

Poland, D. M., & Coffroth, M. A. Trans-generational specificity within a cnidarian–algal symbiosis. *Coral Reefs*, 1-11

Rowan R (2004) Thermal adaptation in reef coral symbionts. Nat Publ Gr 430:742

Rowan R, Knowltono N, Paine RT (1995) Intraspecific diversity and ecological zonation in coral-algal symbiosis. Proc Natl Acad Sci 92:2850–2853

Salih A, Larkum A, Cox G, Kühl M, Hoegh-Guldberg O (2000) Fluorescent pigments in corals are photoprotective. Nature 408:850–853

Sampayo EM, Franceschinis L, Hoegh-Guldberg O, Dove S (2007) Niche partitioning of closely related symbiotic dinoflagellates. Mol Ecol 16:3721–3733

Shore-Maggio, A., Runyon, C. M., Ushijima, B., Aeby, G. S., & Callahan, S. M. (2015). Differences in Bacterial Community Structure in Two Color Morphs of the Hawaiian Reef Coral Montipora capitata. *Applied and environmental microbiology*, *81*(20), 7312-7318

Silverstein RN, Correa AMS, Baker AC (2012) Specificity is rarely absolute in coral-algal symbiosis: implications for coral response to climate change. Proc Biol Sci 279:2609–18

Smith S V., Kimmerer WJ, Laws EA, Brock RE, Walsh TW (1981) Kaneohe Bay Sewage Diversion Experiment: perspectives on ecosystem responses to nutritional perturbation.

Stat M, Bird CE, Pochon X, Chasqui L, Chauka LJ, Concepcion GT, Logan D, Takabayashi M, Toonen RJ, Gates RD (2011) Variation in Symbiodinium ITS2 sequence assemblages among coral colonies. PLoS One 6:1–13

Takabayashi, M., & Hoegh-Guldberg, O. (1995). Ecological and physiological differences between two colour morphs of the coral Pocillopora damicornis. *Marine Biology*, *123*(4), 705-714

Thornhill, D. J., Fitt, W. K., & Schmidt, G. W. (2006). Highly stable symbioses among western Atlantic brooding corals. *Coral Reefs*, *25*(4), 515-519

Todd, P., Sidle, R., & Chou, L. (2002). Plastic corals from Singapore: 2. *Coral reefs*, *21*(4), 407-408

Yuyama, I., Harii, S., & Hidaka, M. (2012). Algal symbiont type affects gene expression in juveniles of the coral Acropora tenuis exposed to thermal stress. *Marine environmental research*, *76*, 41-47

**FIGURE CAPTIONS**

Fig 1. Collection reef locations in Kāne’ohe Bay, O’ahu, Hawai’i, USA

Fig. 2. *Montipora capitata* colonies of both color morphs: orange (left) and brown (right). Photo credit: Raphael Ritson-Williams

Fig. 3. Proportion of occurrence of *Symbiodinium* clades C and D in *Montipora capitata* colonies per dominant symbiont clade

Fig. 4. Percentage of clade D in all colonies of *Montipora capitata*. Bar colors indicate colony color morph

Fig. 5. (Top) Bars indicate the proportion of clade-dominance in all colonies grouped by 1m depth intervals. Line indicates the probability of clade D-dominance as a function of depth. (Middle) Bars indicate the proportion of occurrence of each color morph in all colonies grouped by 1m depth intervals. Line indicates the probability of occurrence of the orange color morph as a function of depth. (Bottom) Probability of clade D-dominance for all colonies of each color morph as a function of depth

Fig. 6. Latitudinal geographic distribution of *Symbiodinium* and color morph in *Montipora capitata* across Kāne’ohe Bay, O’ahu, Hawai’i, USA