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ChIP-seq Data Reveals an Association between Androgen Receptor Protein and **NORAD Promoter**

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BACKGROUND

Androgens and Androgens Receptor (AR) in Prostate Cancer

- Prostate cancer is the most common cancer that occurs in men [1].
- Androgens and AR are primary factors of the disease progression
 - Androgens trigger prostate cancer cells to divide + multiply.
 - AR is activated by binding to androgens.

AR



Androgens

Activate

- Apoptosis
- Cell proliferation
- Invasion
- To suppress the cancer, androgen deprivation therapy can block androgen synthesis [2].
- The cancer can cause AR mutation
 - survive without binding to androgens [3]

Non-Coding RNA Activated by DNA Damage (NORAD)

- Highly abundant + conserved long non-coding RNA
- Promotes metastasis + cell proliferation of prostate cancer cells [4]
- Reduces response to chemotherapy

death rate of cancer patients

Castration of mice

Key observation

The expression of **NORAD** and **AR** (C. Fletcher, personal communication)



Hypothesis:

NORAD expression may be regulated by **AR**

Aim:

 To investigate an interaction and relationship between NORAD and AR protein

MATERIALS AND METHODS

• RIP-seq, RNA-seq, and ChIP-seq data of LNCaP cell line

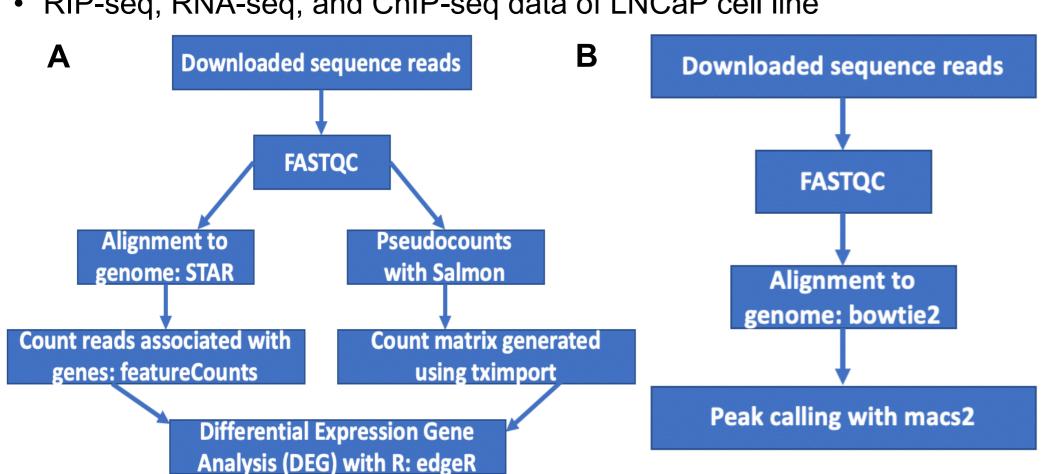


Figure 1. Data analysis pipelines. (A) Analysis pipeline for RIP-seq and RNA-seq data. (B) Analysis workflow for ChIP-seq data.

RESULTS B A 40 nonsignificant 30 LogFC MT-ATP6 -Log₁₀FDR 30 -Log₁₀FDR FDR MT-ND2 significant MT-CO2 MT-ND1 10 NORAD 10 -1 0 1 -1 0 1 LogFC LogFC

Figure 2. DEG analysis from RIP-seq data using STAR and Salmon aligners. The log ratio of expression levels between anti-AR and anti-IgG samples (x axis) is plotted against -logFDR. (A) A volcano plot depicting DEG genes using STAR. NORAD was not significant (FDR = 0.25, logFC = -0.52). (B) Volcano plot representation of DEG analysis using Salmon. NORAD was also not significant (FDR = 0.18, logFC= -0.81). Significantly differentially-expressed genes between the two conditions based on the specified criteria of FDR < 0.05 and |log2FC| >= 1.0.

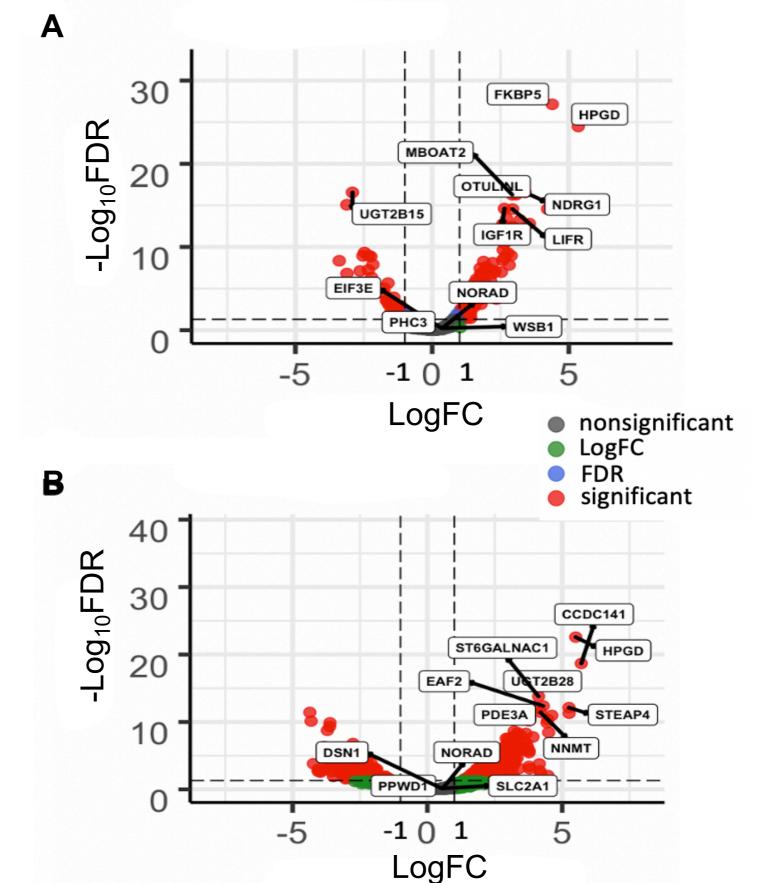


Figure 3. DEG analysis from RNA-seq data using STAR and Salmon aligners.

The log ratio of expression levels between untreated and treated androgen samples (x axis) is plotted against -logFDR. Significantly differentiallyexpressed genes between the two conditions based on the specified criteria of FDR < 0.05 and |log2FC| >= 1.0.

(A) A volcano plot depicting DÉG analysis using STAR. NORAD was not significant (FDR = 0.59, logFC = 0.36)

(B) Volcano plot representation of DEG analysis using Salmon. NORAD was not considered significant as FDR = 0.74 and logFC = 0.56.

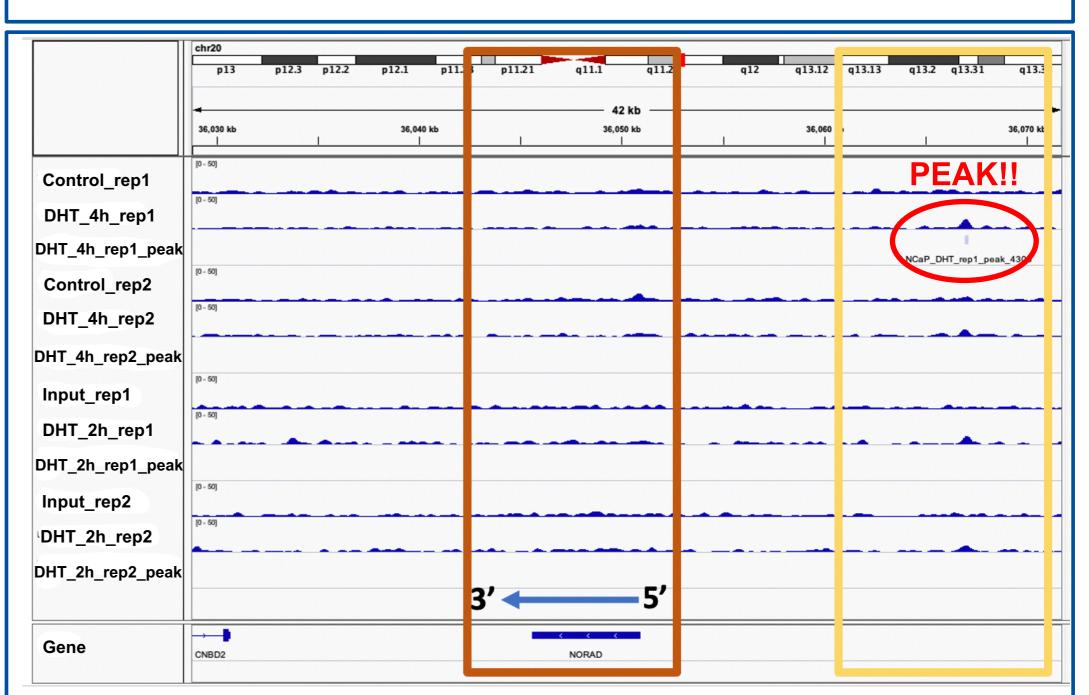


Figure 4. Integrative Genomics Viewer of ChIP-seq data. The x-axis represents the human reference genome, whereas the y-axis represents the number of sequence reads from the samples that match the human genome. Peak calling analysis was conducted using macs2. Mapped reads of treatment (DHT 2h and DHT_4h) and control (Control and Input) samples were compared in order to identify peak regions that aligned with the human genome. Unlike DHT_2h, DHT_4h samples were treated with 10 nM DHT for four hours, whereas DHT_2h samples were treated with 100 nM DHT for two hours. DHT_4h_rep1_peak, DHT_4h_rep2_peak, DHT_2h_rep1_peak, and DHT_2h_rep2_peak represent peaks that indicate AR binding sites in the DNA sequence when FDR < 0.05. One peak was identified in the DHT_4h_rep1 sample (red circle); no other peaks were identified (yellow box)

CONCLUSION

- There are no supporting evidences from RIP-seq and RNA-seq data that NORAD is associated with AR protein
- ChIP-seq in LNCaP showed a peak at the area of NORAD promoter possible AR binding upstream of NORAD



Future work:

- To clarify AR binds to the NORAD promoter
 - ChIP-PCR for AR association with NORAD promoter and enhancer regions across a timecourse of androgen treatments and in different prostate cancer cell lines.

REFERENCES

- 1. C. Hoey et al., Circulating miRNAs as non-invasive biomarkers to predict aggressive prostate cancer after radical prostatectomy. J Transl Med 17, 173 (2019). 2. B. C. Thomas, D. E. Neal, Androgen deprivation treatment in prostate cancer. BMJ 346, e8555 (2013).
- 3. T. M. Amaral, D. Macedo, I. Fernandes, L. Costa, Castration-resistant prostate cancer: mechanisms, targets, and treatment. Prostate Cancer 2012, 327253 (2012). 4. H. Zhang, H. Guo, Long non-coding RNA NORAD induces cell proliferation and migration in prostate cancer. J Int Med Res 47, 3898-3904 (2019).