# An Open Source Tool for Determination of Pharmacogenomics Haplotypes from Diverse and Complex Data



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#### 1 Introduction

#### 1.1 Background

- Growing availability of genomic data has increased the opportunity for precision medicine with pharmacogenomics (PGx).
- Variation in methods drives challenge in data interpretation for PGx, creating a need for accessible and configurable bioinformatics tools for determination of clinically relevant haplotypes and structural variations (SV) within pharmacogenes.
- Efforts by existing authorities (CPIC, PharmVar, PharmGKB) to standardize definitions are sporadically implemented.
- An existing open source solution for pharmacogenomics haplotyping and structural variation determination does not exist.

#### 1.2 Objectives

Create an open source software tool that could determine pharmacogenomics haplotypes based on standardized inputs from pharmacogenomics authority organizations and provide estimation of breakpoints + copy gain/loss for genes with known structural variations (e.g. *CYP2D6*).

## 2 Development of hiMoon PGx

#### 2.1 Implementation

- Written in Python, requires Python 3.6+.
- Heavy utilization of Pysam for VCF and BAM parsing.
- CNV detection optimized with Numba.

## 2.2 Haplotype Identification

- Considers all possible haplotypes H (from library) in each gene.
- Tests adherence of each subject S variant v (AA=0, AB=1, BB=2) to candidate haplotype.
- Determines all possible 2-way haplotype combinations  $(S_{H_1}, S_{H_2})$  such that:

$$egin{aligned} 1. \ f(S_H,0) &= 1 \ 2. \ f(S_{H_1} \cap S_{H_2},2) &= 1 \end{aligned}$$

Where:

$$f(S_H,x)=rac{|\{S_H|v>x\}|}{|S_H|}$$

- Reports diplotypes where the number of used variants in both haplotypes is maximized.
- Suggests that novel haplotype exists if additional variants that are not used in reported diplotype.

## 2.3 Haplotype Libraries

- Haplotype libraries are provided by the user, and follow the same format as the tabdelimited files available from PharmVar.
- Users may also create their own files in this format for genes not annotated in PharmVar.

## 3 Development of hiMoon PGx Continued

#### 3.1 Structural Variation Determination

**Method 1:** Maximum penalized likelihood estimation method (MPLE, below), adapted from *SeqCNV* method published by Chen, et al (DOI: 10.1186/s12859-017-1566-3).

• Seeks to maximize the following, where  $p_i$  is the probability of a given read originating from the unknown sample,  $t_i$  is the number of reads from the unknown sample for a given region,  $c_i$  is the number of reads from the control sample, and  $\lambda$  is the Bayesian information criterion (BIC).

$$PL = \sum_i (t_i ln(p_i) + c_i ln(1-p_i)) - 2\lambda$$

- User defines regions to estimate breakpoints and copy number in an unknown sample relative to a control sample.
- Predicts copy gain if coverage ratio in highest likelihood region is > 1.4, or copy loss if < 0.6.

Method 2: Control region ratio determination (quick CNV).

- Copy number is estimated based on a highly similar region in the same sample (e.g. CYP2D6 = Target; CYP2D8P = Comparator).
- Depth of coverage is normalized to the length of the region, and a log ratio of the case:control is calculated.
- Usually a ratio  $\leq$  0 copy loss and ratio > 0.5 = copy gain.

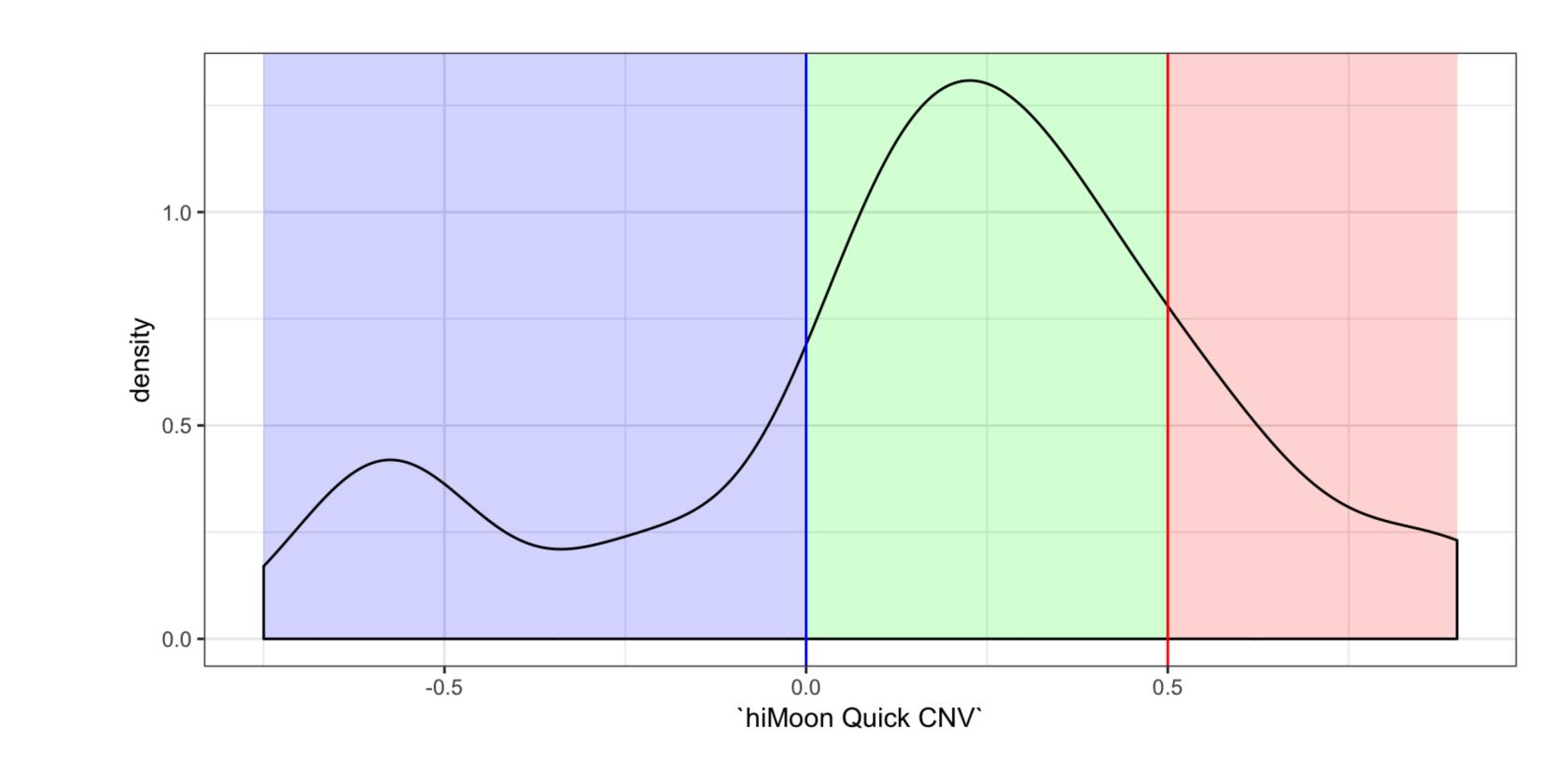


Figure 3.1: Bimodal distribution of average read depth shows delineation between copy loss and normal/gain.

#### 3.2 Testing

- Whole exome sequencing BAM files were obtained from fifteen 1000 genomes samples.
- hiMoon was used to estimate star alleles and copy number with quick CNV.
- Note that poor coverage in many areas leads to erroneous star allele calls relative to the known calls generated using higher fidelity genotyping methods.
- Haplotype and structural variation carried out in 100 samples from 1000X + coverage of *CYP2D6* with ~200bp flanks.

#### 4 Results

## 4.1 Quick CNV and CYP2D6 haplotype calling with 1000 Genomes WES data

- Whole exome data applied to the full *CYP2D6* translation table from PharmVar provides sufficient coverage for calling most star alleles.
- Inadequate coverage of intronic regions and gene flanks leads to ambiguities.
- hiMoon successfully called star alleles in 10/15 samples, and noted coverage ambiguity in 5/5 cases of the miscalled samples.
- Copy number was correctly called in all samples.

## 4.2 MPLE CNV and allele calls for 100 high coverage samples

Table 4.1: Concordance with star alleles as reported by Pratt, et al. (DOI: 10.1016/j.jmoldx.2015.08.005)

• Evidence of structural variation (partial/comple	ete
gain or loss) in $> 15\%$ of samples.	

- Non-ambiguous call obtained in 42/100 samples.
- Prevalence of ambiguous star allele calls highlights

  a possible need to subset translation tables due to NA12717

  the need for extensive gene coverage.

#### 4.3 Future Directions

- Ongoing development to determine maximum NA18565 \*10/(\*10 [\*36]) resolution for breakpoints, currently >100bp.
- Further validation with other clinical relevant NA18959 \*2/(\*10 [\*36])
- Phts NA07000 \*9/\*2 (\*35) 0.1314 \*2/
  Le to NA12717 \*1/\*1 0.1550 \*1/
  NA20509 \*4/\*35 0.2490 \*2/
  NA19239 \*15/\*17 0.2751 \*2/\*
  NA12006 \*4/\*41 0.2948 \*10/\*1
  NA19007 \*1/\*1 0.3619 \*1/
  NA12878 \*3/\*4 0.4406 \*3/
  NA12878 \*3/\*4 0.5197 \*10/\*10x
  NA19785 \*1/\*2(XN) 0.6282 \*34/\*39(x)
- genes with common structural variation.
  Potential development of clinical outflow to allow integration with clinical practice guidelines.
- Transition Numba optimized CNV functions to Cython to improve speed with high coverage samples, currently up to 2 minutes per samples.

## 5 Conclusion

- Accessible, fast estimation of star alleles, easy to use, and tunable for data from sequencing, array, or targeted genotyping.
- Standardized library (inputs directly from PharmVar or formatted per PharmVar specifications).
- Reference agnostic (accepts VCF/BAM aligned to any reference, as long as library matches).
- Accurate calling of haplotypes + structural variation for complex pharmacogenes (e.g. CYP2D6).
- Conservative reporting with acknowledgement of possible novel discoveries.

## 6 Disclosures

All authors are affiliated with Ariel Precision Medicine. hiMoon is developed in coordination of resources from Shenandoah University and Ariel Precision Medicine.