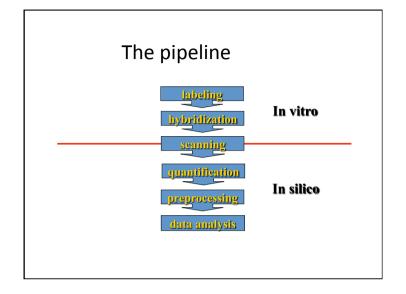
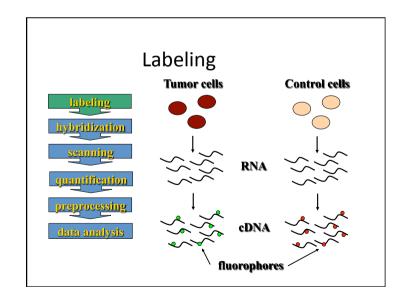
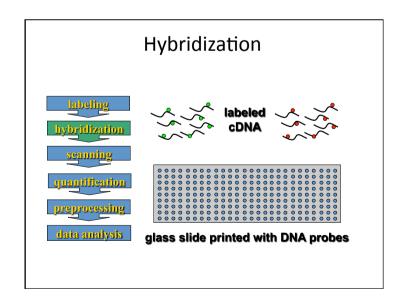
## Preprocessing of Dual-channel Microarray Data

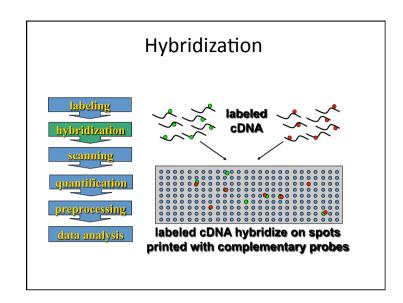
Vincent Detours
IRIBHM - Free University of Brussels

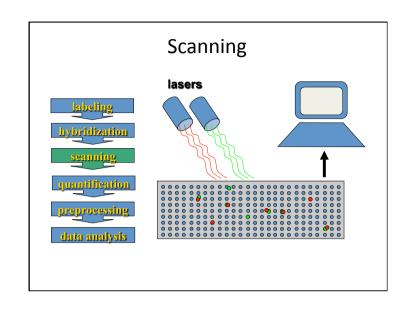
vdetours@ulb.ac.be

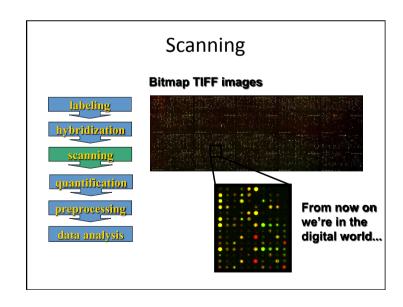


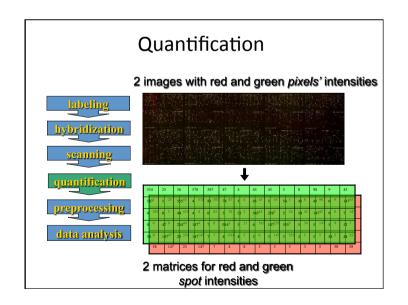


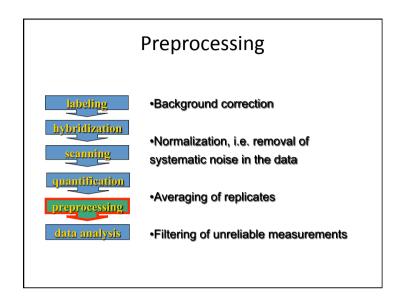


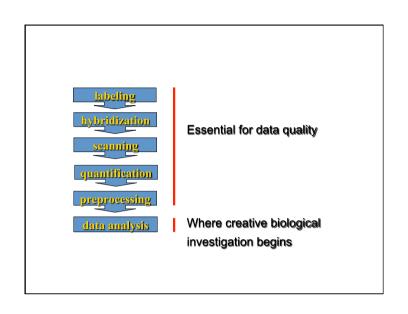


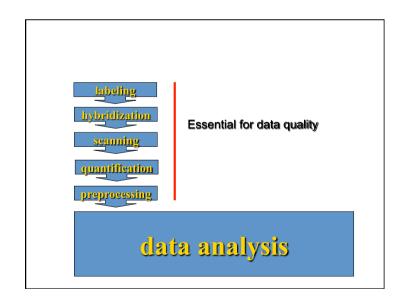


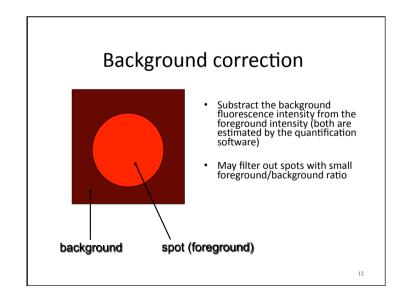


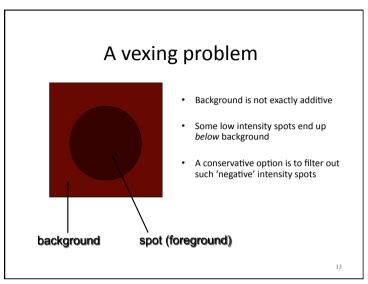


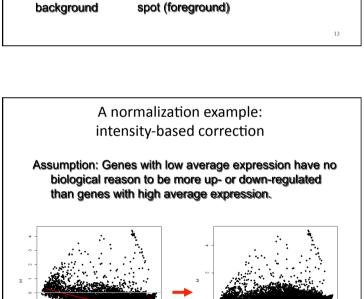








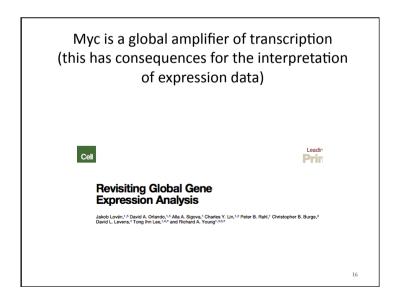


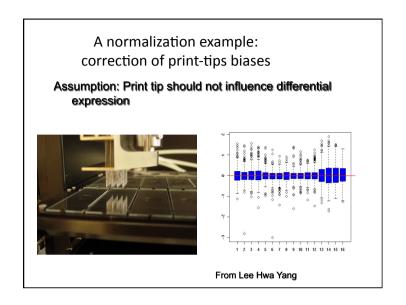


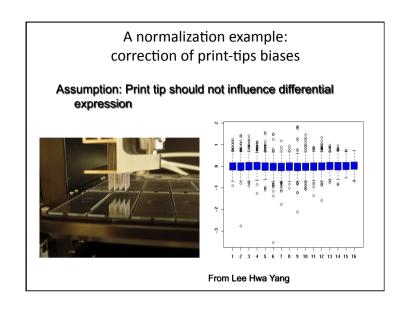
## Some assumptions about how 'good' data should look like are needed in order to detect and correct for systematic biases. Normalization algorithms typically assume that Gene regulation is independent of non biological variables (e.g., the location of a spot on the slide), Regulation levels averaged across large numbers of genes is constant (#up = #down).

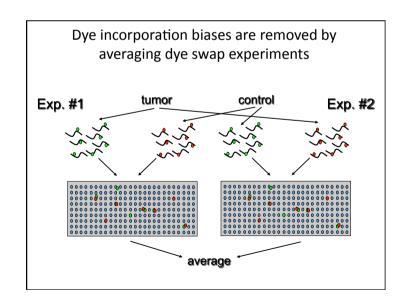
Normalization relies on (possibly false) hypothesis

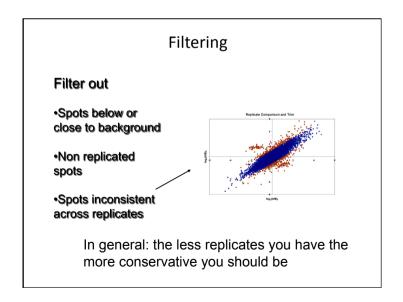
If these assumptions are not true—they clearly aren't in some biological contexts—only comparisons of expression ratios are valid.

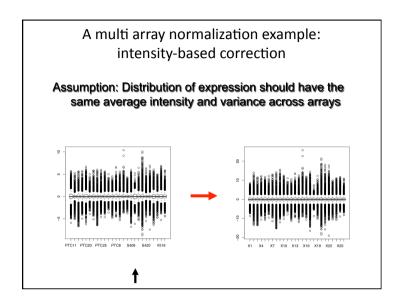












## **CONCLUSION**

Your analysis should not depend on tweaking the preprocessing.

Cross-study, cross-platform, comparison will ultimately be the gold standard, thus

OPEN AND UNRESTRICTED ACCESS TO RAW DATA IS THE KEY.

## So, what preprocessing is best?

- Hard to say in the absence of well accepted benchmark data sets!
- A reasonable pipeline is:
  - background correction
  - intensity normalization
  - space normalization
  - dye swap averaging

22