

First decade of single-cell sequencing

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Milestones of single cell sequencing since 2009

Transcriptome

2009: scRNA-seq

□ nature methods

mRNA-Seq whole-transcriptome analysis of a single cell

Fuchou Tang, Catalin Barbacioru, Yangzhou Wang, Ellen Nordman, Clarence Lee, Nanlan Xu, Xiaohui Wang, John Bodeau, Brian B Tuch, Asim Siddiqui, Kaiqin Lao [™] & M Azim Surani [™]

Genome

2011: scWGS



nature

ublished: 13 March 20

Tumour evolution inferred by single-cell sequencing

Nicholas Navin, Jude Kendall, Jennifer Troge, Peter Andrews, Linda Rodgers, Jeanne McIndoo, Kerry Cook, Asya Stepansky, Dan Levy, Diane Esposito, Lakshmi Muthuswamy, Alex Krasnitz, W. Richard McCombie, James Hicks & Michael Wigler

2012: scExome-seq



Log in

Single-Cell Exome Sequencing Reveals Single-Nucleotide Mutation Characteristics of a Kidney Tumor

Xun Xu 14 • Yong Hou 14 • Xuyang Yin 14 • ... Michael Dean A ☑ • Yingrui Li A ☑ • Jun Wang A

Multiomics

2015: scG&T (genome & transcriptome)

nature methods

G&T-seq: parallel sequencing of singlecell genomes and transcriptomes

lain C Macaulay ☑, Wilfried Haerty, Parveen Kumar, Yang I Li, Tim Xiaoming Hu, Mabel J Teng, Mubeen Goolam, Nathalie Saurat, Paul Coupland, Lesley M Shirley, Miriam Smith, Niles Van der Aa, Ruby Banerjee, Peter D Ellis, Michael A Quail, Harold P Swerdlow, Magdalena Zernicka-Goetz, Frederick J Livesey, Chris P Ponting ☑ & Thierry Voet ☑

2018: scNMT-seq (transcriptome & epigenome)

nature communications

Article | Open Access | Published: 22 February 2018

scNMT-seq enables joint profiling of chromatin accessibility DNA methylation and transcription in single cells

Stephen J. Clark $\[egin{array}{ll} \Box Argelaguet, ChantrioInt-Andreas Kapourani, Thomas M. Stubbs, Heather J. Lee, Celia Alda-Catalinas, Felix Krueger, Guido Sanguinetti, Gavin Kelsev, John C. Marioni <math>\[egin{array}{ll} \Box Argelaguet Britania & Argelaguet Brit$

2017: scTrio-seq (genome & transcriptome & epigenome

Cell Research

Single-cell multi-omics sequencing of mouse early embryos and embryonic stem cells

Fan Guo oxtimes , Lin Li, Jingyun Li, Xinglong Wu, Boqiang Hu, Ping Zhu, Lu Wen & Fuchou Tang oxtimes

Epigenome

2013: scHi-C

nature

Published: 25 September 2013

Single-cell Hi-C reveals cell-to-cell variability in chromosome structure

Takashi Nagano, Yaniv Lubling, Tim J. Stevens, Stefan Schoenfelder, Eitan Yaffe, Wendy Dean, Ernest D. Laue ☑, Amos Tanay ☑ & Peter Fraser ☑

2015: scChIP-seq

v nature biotechnology

Single-cell ChIP-seq reveals cell subpopulations defined by chromatin state

Assaf Rotem, Oren Ram, Noam Shoresh, Ralph A Sperling, Alon Goren, David A Weitz ☑ & Bradley E Bernstein ☑

2015: scATAC-seq

nature

Published: 17 June 2015

Single-cell chromatin accessibility reveals principles of regulatory variation

Jason D. Buenrostro, Beijing Wu, Ulrike M. Litzenburger, Dave Ruff, Michael L.
Gonzales, Michael P. Snyder, Howard Y. Chang ≅ & William J. Greenleaf ≅

2017: scNOMe-seq

Simultaneous measurement of chromatin accessibility, DNA methylation, and nucleosome phasing in single cells

Sebastian Pott*

2017: scCOOL-seq

Cell Research

Single-cell multi-omics sequencing of mouse early embryos and embryonic stem cells

Fan Guo [⊡], Lin Li, Jingyun Li, Xinglong Wu, Boqiang Hu, Ping Zhu, Lu Wen & Fuchou Tang [⊡]

2015: scWGBS-seq

Cell Reports

2015: scDamID-seq

Cell

Genome-wide Maps of Nuclear Lamina Interactions in Single Human Cells

Joo Kod A ≅ - Ludo Page - Sandra S. de Vires - ... Job Dekker - Alexander van Oudermaarden

2015: scDNase-seq

nature

Genome-wide detection of DNase I hypersensitive sites in single cells and FFPE tissue samples

Wenfei Jin, Qingsong Tang, Mimi Wan, Kairong Cui, Yi Zhang, Gang Ren, Bing Ni, Jeffrey Sklar, Teresa M. Przytycka, Richard Childs, David Levens & Keji Zhao [⊡]

2018: scMNase-seq

nature

Letter | Published: 26 September 2018

Principles of nucleosome organization revealed by single-cell micrococcal nuclease sequencing

Binbin Lai, Weiwu Gao, Kairong Cui, Wanli Xie, Qingsong Tang, Wenfei Jin, Gangqing Hu, Bing Ni & Keji Zhao [™]

2019: scCUT&RUN

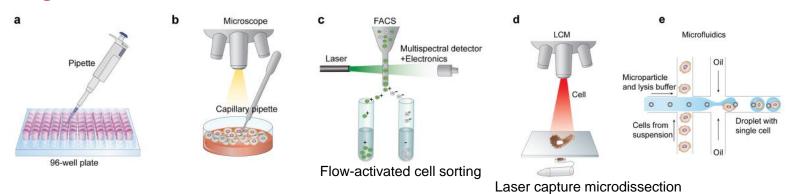


Profiling of Pluripotency Factors in Single Cells and

Sarah J. Hainer A. □ «Ana Bošković «Kurtis N. McCannell » Oliver J. Rando «Thomas G. Fazzio A. 4

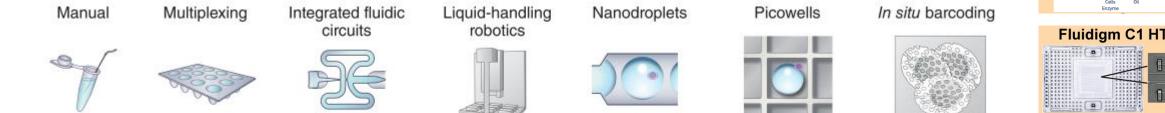
Typical single-cell experimental/technical pipeline

Step 1: Single cell isolation



Step 2: Library preparation (cell capture and lysis, PCR-based barcoding)

Brennecke et al. 201364



Klein et al. 201534

Macosko et al. 201540

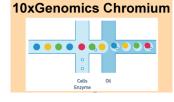


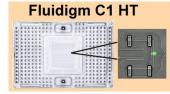
Tang et al. 200918

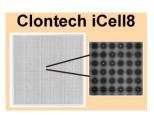
Islam et al. 201124



Jaitin et al. 201433





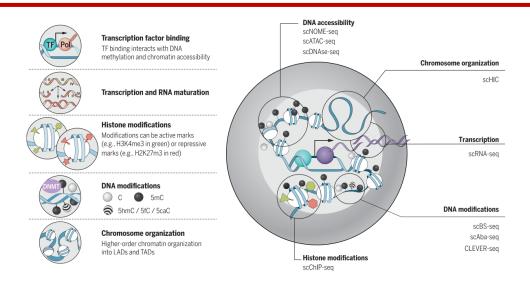


Cao et al. 201751

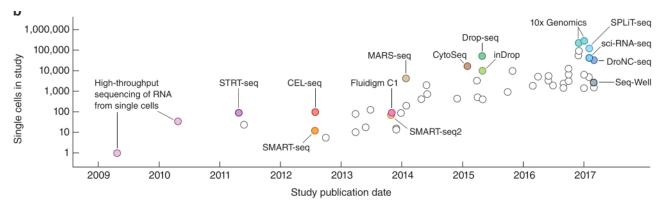
Rosenberg et al. 201752

Bose et al. 201543

More challenges for single-cell genome/epigenome sequencing

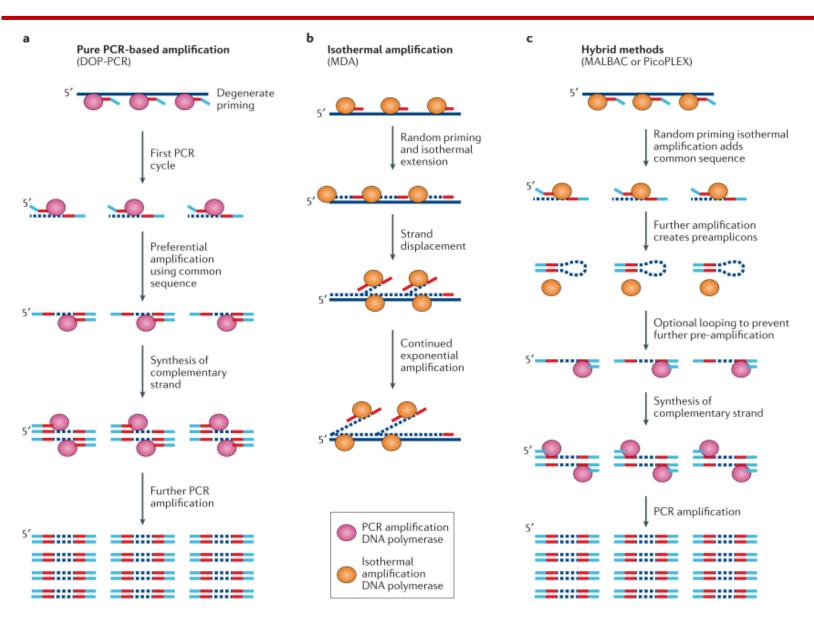


Development of scRNA-seq



- A single diploid human cell (~3GB genome size)
 - o DNA
 - Two copies: paternal and maternal
 - ~6 pg
 - o RNA
 - 360,000 mRNA molecules; 12,000 different transcripts
 - 10-30 pg total RNA, 1-5% of them are mRNA
- > Sequencer
 - Nanogram microgram
- Solution
 - PCR amplification
 - Pool many single cells cell barcoding
 - PCR-based barcoding
 - PCR-free barcoding (transposase)

Typical single-cell genome/epigenome sequencing: amplification first



	PCR-based (DOP-PCR)	Isothermal (MDA)	Hybrid (MALBAC or PicoPLEX)
False-negative rate (coverage and allelic dropout)	High	Low	Intermediate
Non-uniformity	Low	High	Low
False-positive rate (amplification error rate)	High	Low	Intermediate

Lähnemann et al. Genome Biology (2020) 21:3

Genome Biology

REVIE

Open Access

Eleven grand challenges in single-cell data science



David Lähnemann^{1,2,3}, Johannes Köster^{1,4}, Ewa Szczurek⁵, Davis J. McCarthy^{6,7}, Stephanie C. Hicks⁸, Mark D. Robinson⁹ , Catalina A. Vallejos ^{10,11}, Kieran R. Campbell ^{1,21,31,4}, Niko Beerenwinkel^{1,5,16}, Ahmed Mahfouzl^{7,18}, Luca Pinellol^{1,20,21}, Pavel Skums²², Alexandros Stamatakis^{2,32,4}, Camille Stephan-Otto Attolini²⁵, Samuel Aparicol^{3,26}, Jasmijin Baaijens²⁷, Marleen Balvert^{27,28}, Buys de Barbanson^{29,30,31}, Antonio Cappuccio³², Giacomo Corleone³³, Bas E. Dutilh^{28,34}, Buys de Barbanson^{29,30,31}, Victor Guyey³⁵, Rens Holmer³⁶, Katharina Jahn^{1,5,16}, Thamar Jessurun Lobo³⁵, Emma M. Keizer³⁷, Indu Khatri³⁸, Szymon M. Kielbasa³⁹, Jan O. Korbel⁴⁰, Alexey M. Kozlov²³, Tzu-Hao Kuo³, Boudewijn P.F. Lelieveldr^{41,42}, Ion I. Mandoiuf⁴³, John C. Marioni^{44,5,46}, Tobias Marschall^{47,48}, Felix Mölder^{1,49}, Amir Niknejad^{50,51}, Lukasz Raczkowski⁵, Marcel Reinders^{17,18}, Jeroen de Ridder^{23,03}, Antoine-Ermanuel Saliba³², Antonios Somarakis⁴², Oliver Stegle^{40,46,53}, Fabian J. Theis⁵⁴, Huan Yang⁵⁵, Alex Zelikovsky^{56,57}, Alice C. McHardy³, Benjamin J. Raphael¹⁸, Sohrab P. Shah⁵⁹ and Alexander Schönhuth^{27,28}*

Errors/issues during WGA

Loss of coverage

MDA: 84%

MALBAC: 72%

DOP-PCR: 39%

- Decreased coverage uniformity
- Allelic imbalance/allelic dropout
- Errors during PCR amplification

Single-cell genome sequencing with PCR-free barcoding

nature methods

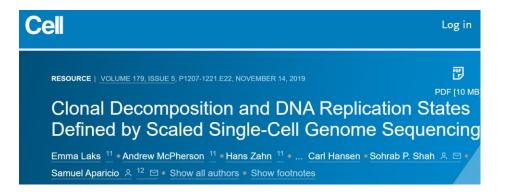
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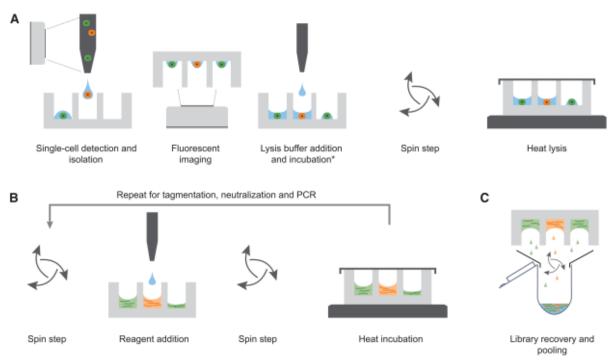
Published: 09 January 2017

Scalable whole-genome single-cell library preparation without preamplification

Hans Zahn, Adi Steif, Emma Laks, Peter Eirew, Michael VanInsberghe, Sohrab P Shah ☑, Samuel Aparicio ☑ & Carl L Hansen ☑



Barcoding first, then PCR amplication



- Increase coverage uniformity compared to pre-amplification methods
 - Copy number variation analysis
- ➤ On average, <15% genome coverage per single cell

Two types of PCR-free barcoding methods

DNA ligase-based (more steps, low efficiency)

Transposase-based (simpler, faster, high efficiency)

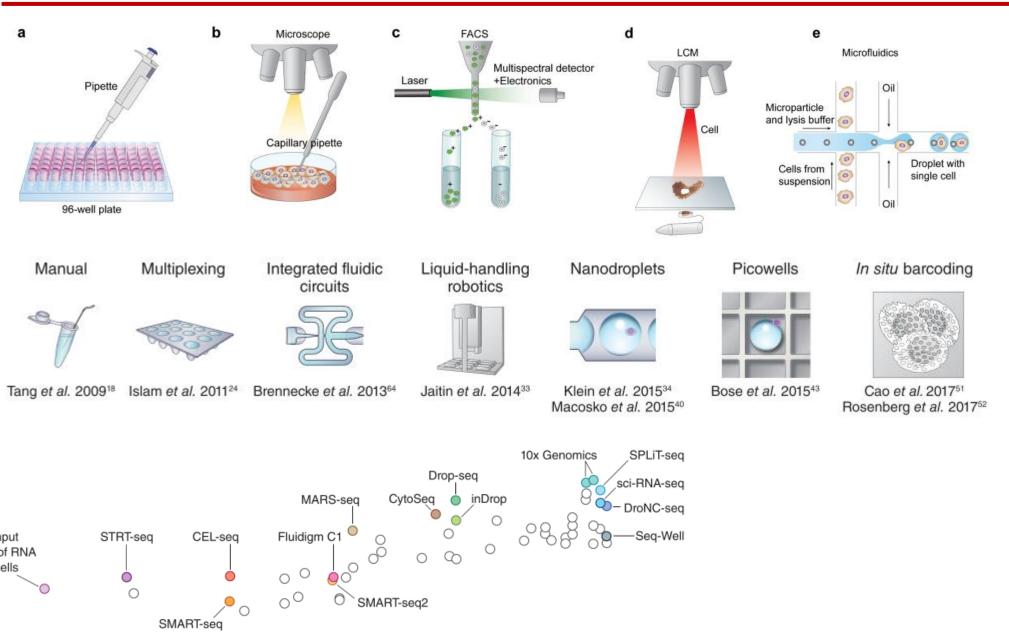
Tn5 transposase complex with adaptors



- > Single-cell single-molecule long-read sequencing (transposase-based barcoding + PacBio/ONT)
 - An automatic platform (e.g., 10X): large cell number
 - Cost: thousands of Tn5 complex with different barcodes

Thanks

Single-cell isolation



Hwang, et al. Exp Mol Med (2018); Svensson, et al. Nat Protoc (2018)