## **CODING TEAM WORKFLOW BY FAIZAN**

STEP	INPUT	TOOL	USES	ОИТРИТ
1	Raw reads (fastq)	Fastqc	Check quality of reads	Summary html report
2	Raw reads(fastq)	Fastp	Improve raw reads quality	Trim reads(fastq)
3	Trim reads(fastq)	Spades	trimmed reads into longer contiguous sequences (contigs)	Assembled sequences(contigs .fasta)
4	Assembled sequences(contigs.fasta )	SeqSero2	predict the serotype	predicted serotype(result.txt)
5	Assembled sequences(contigs.fasta )	PlasmidFinde r	uses a database of known plasmid sequences to identify matches	predicted plamidtyping (result.txt)
6	Assembled sequences(contigs.fasta )	AMRFinderPl us	compares contigs against a database of known resistance genes	predicted amrgenes (result.txt)
7	Trim reads(fastq) + reference genome (fasta)	Snippy	variant calling and core genome alignment	variant calls (vcf) + consensus sequence based on the reference and the called variants(fasta) + BAM file of reads aligned to the reference genome (BAM) + Provides details on the process and results(Summary Stats)
8	BAM file of reads aligned to the reference genome (BAM)	Snippy-core	combine the outputs of multiple Snippy runs to create a core	A single multiple sequence alignment (MSA)

			genome alignment.	FASTA file of the core genome across all included samples. (fasta)
9	FASTA file of the core genome across all included samples. (fasta)	RAxML-NG	building phylogenetic trees using maximum likelihood estimation.	The tree is often in Newick format.(.tree) + logfiles
10	Combine file contain identifier ,serotype,plasmidtype,a mrtypes(csv) + treedata(.tree)	ggplot(R)	Visulization of plot	Final figure