## FORWARD GENETIC APPROACHES FOR MENDELIAN DISEASE MODELING IN MICE

Exome sequencing for variant discovery module for "Big Data Skills Training for Professors BD2K"

May 24, 2017

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#### Forward Genetics in Mice

#### **ADVANTAGES**

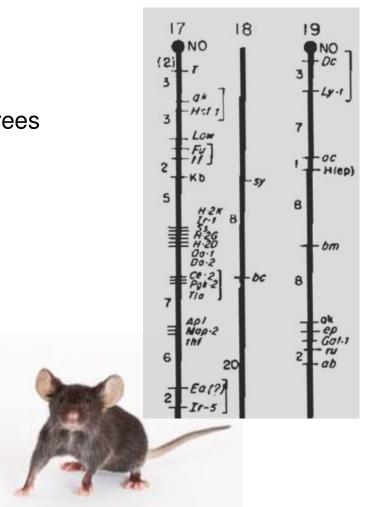
- unbiased, phenotype to gene
- selective breeding, large consanguineous pedigrees
- genetically defined inbred strain backgrounds
- mutation rates
  - spontaneous, ~6 x 10<sup>-6</sup> / locus / generation

#### RESOURCES AT THE JACKSON LABORATORY

- scale
- quality control for 'phenodeviants'

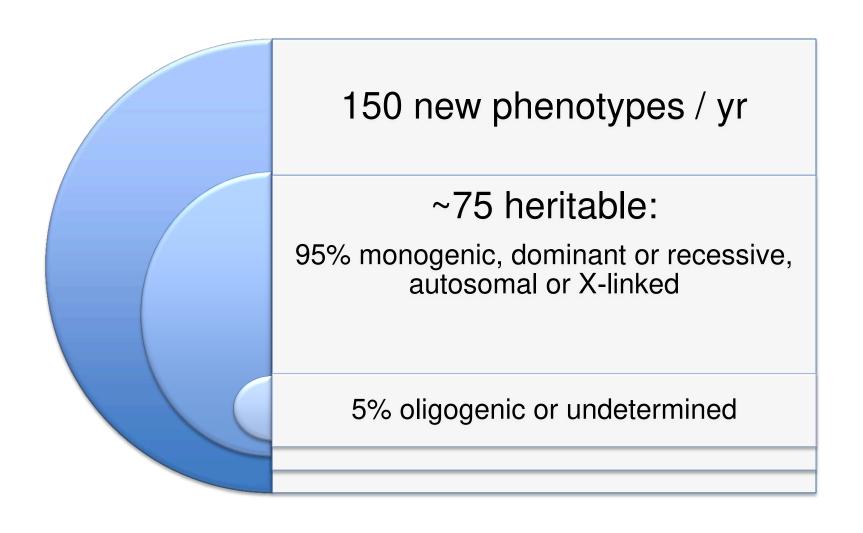
#### KEY TECHNOLOGIES

- High-throughput sequencing
- Common inbred strain reference genomes (Sanger Mouse Genomes Project)

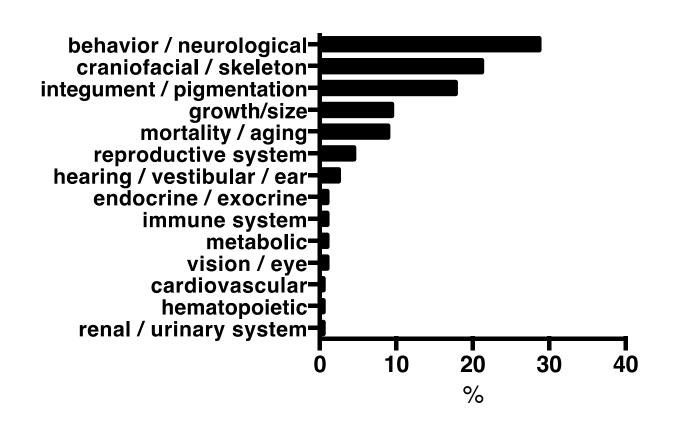




# Annual recovery of "phenodeviants" from a production scale vivarium

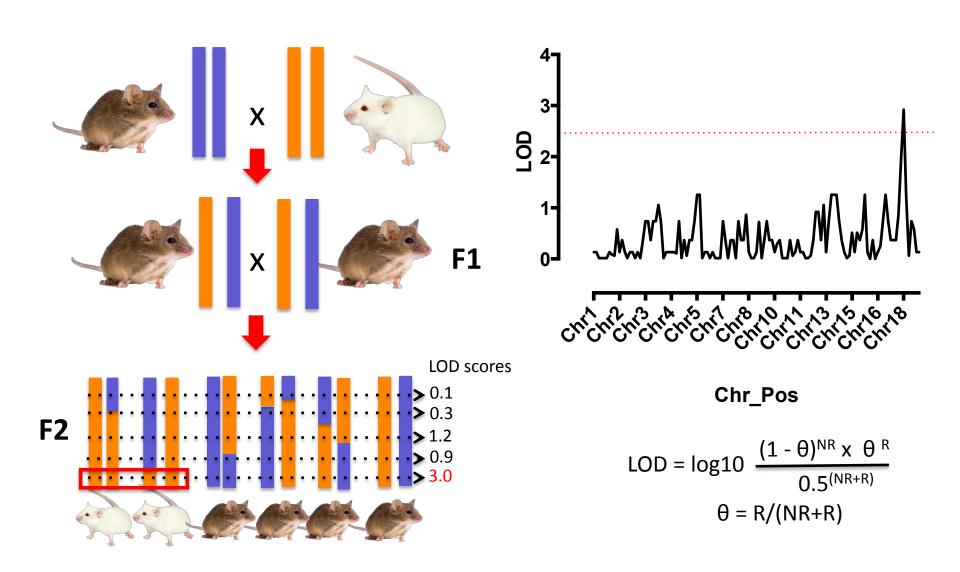


### Heritable Mendelian phenotypes at the The Jackson Laboratory mouse "clinic"

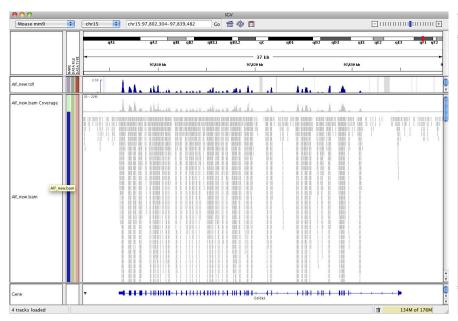


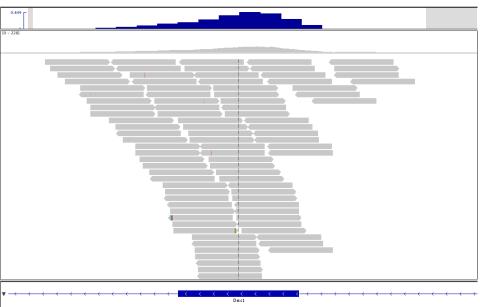
Mammalian Phenotype Ontology: Smith CL and Eppig JT., Mamm Genome. 2012 Oct;23(9-10):653-68

### Mapping cross, recessive trait

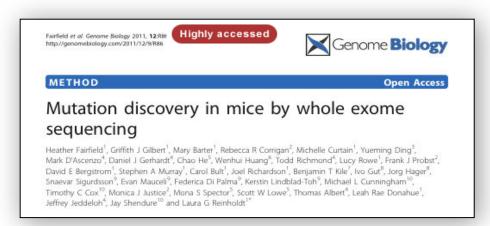


# New technologies, resources, impact

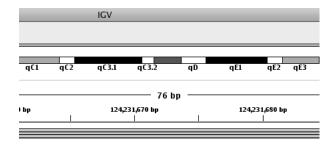


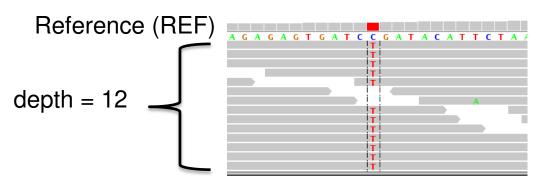


- Based on Mouse Gene Catalog (curated by Mouse Genome Database), non-redundant gene predictions from NCBI, Ensembl and Vega
- 203,225 exonic regions ~33,000 genes and miRNAs



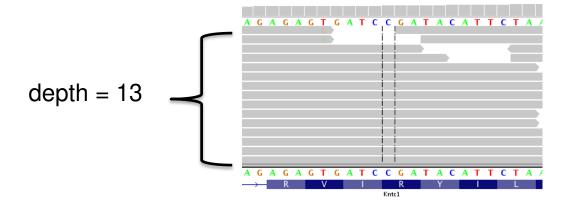
#### Variant features



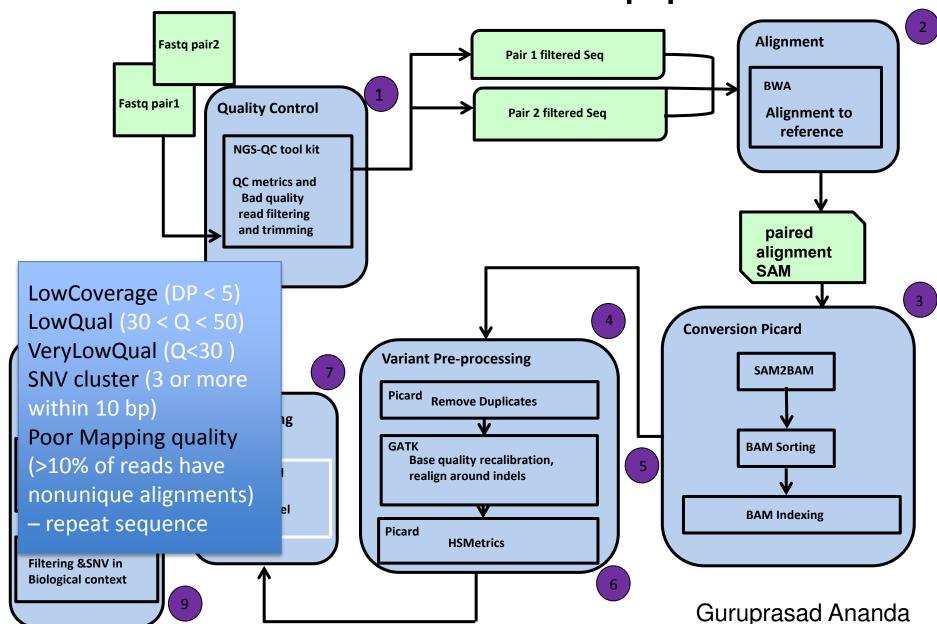


allele ratio (REF/ALT) = 0/12

allele frequency (var %) (for non reference allele) = 1.0



### Mouse exome CIVET pipeline

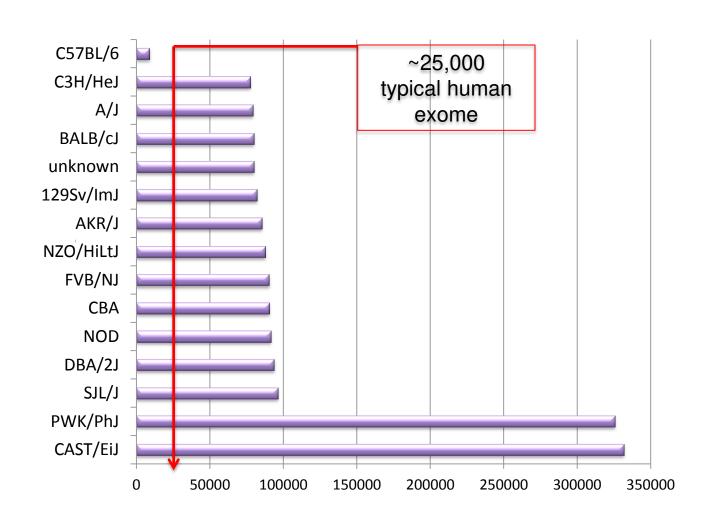


### Variant caller format (VCF)

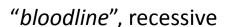
#reference	_=file:///data/s	nared/genor	nes/Mus_mus	culus/UCSC	C/mm10_3-18-2	013/Sequence	WholeGe	.nomeFasta/gr	⊿nome.fa		السساء	,	السسساء		/	<u> </u>	<u> </u>	
#source_2	0140024.1=vcf	i-annotate(r7	97) -a /data/s'	nared/mmr	r/dbSNP.mm10	tab.gz -c CHRC	M,FROM,	ro,id			.(	4	į J		11			
#CHROM	POS	ID	REF	ALT	QUAL	FILTER	IFO	FORMAT	SJL			1	(L					_[
hr10	3120209	rs29378454	A	G	2004.77	PASS	/ C=2	AF=1.00	AN=2	BaseQRankSi	DP=77	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
hr10	3120231	rs29358647	A	G	2049.77	PASS	/C=2	AF=1.00	AN=2	BaseQRankSi	DP=85	Dels=0.00	FS=3.740	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
hr10	3120311	rs29371433	C	T	1125.77	PASS	/ C=2	AF=1.00	AN=2	BaseQRankSi	DP=43	Dels=0.00	FS=6.453	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
chr10	3125031	rs47392805	G	A	625.77	PASS	/C=2	AF=1.00	AN=2	DP=25	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=57.17	MQ0=0	QD=25.03
hr10	3125055	rs48707627	G	C	509.77	PASS	/C=2	AF=1.00	AN=2	DP=17	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=55.80	MQ0=0	QD=29.99
hr10	3125193	rs50658327	C	T	234.78	PASS	C=2	AF=1.00	AN=2	DP=8	Dels=0.00	FS=0.000	HaplotypeSci	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0	QD=29.35
hr10	3268447	rs29364458	A	G	674.77	PASS	/ C=2	AF=1.00	AN=2	DP=24	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=53.32	MQ0=0	QD=28.12
hr10	3268450	rs29346449	G	A	614.77	PASS	C=2	AF=1.00	AN=2	DP=22	Dels=0.00	FS=0.000	HaplotypeSci	MLEAC=2	MLEAF=1.00	MQ=53.62	MQ0=0	QD=27.94
hr10	3268460		TG	T	668.73	PASS	C=2	AF=1.00	AN=2	DP=19	FS=0.000	HaplotypeSci	MLEAC=2	MLEAF=1.00	MQ=54.01	MQ0=0	QD=35.20	RPA=3,2
chr10	3268474	rs29368948	C	T	620.77	PASS	C=2	AF=1.00	AN=2	DP=21	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=54.68	MQ0=0	QD=29.56
chr10	3545733	rs29315555	A	G	845.81	PASS	C=2	AF=1.00	AN=2	BaseQRankSi	DP=35	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
chr10	3554546	rs29376248	T	C	450.77	PASS	C=2	AF=1.00	AN=2	DP=15	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0	QD=30.05
hr10	3554631	rs29383399	C	T	497.77	PASS	C=2	AF=1.00	AN=2	BaseQRankSi	DP=20	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
chr10	3872653	rs33849715	T	C	637.77	PASS	C=2	AF=1.00	AN=2	DP=21	Dels=0.00	FS=0.000	HaplotypeSci	MLEAC=2	MLEAF=1.00	MQ=57.77	MQ0=0	QD=30.37
hr10	3872657	rs29364945	G	A	711.77	PASS	/C=2	AF=1.00	AN=2	DP=25	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=58.13	MQ0=0	QD=28.47
hr10	3933583	rs29358987	C	T	1011.77	PASS	C=2	AF=1.00	AN=2	BaseQRankS	DP=40	Dels=0.00	FS=2.636	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
hr10	3978708	rs29362746	rs29320259	T	С	582.77	ASS	AC=2	AF=1.00	AN=2	BaseQRankSi	DP=21	Dels=0.00	FS=3.222	HaplotypeSc	MLEAC=2	MLEAF=1.00	J MQ=57.70
chr10	3978709	rs29362746	G	Α	593.77	PASS	C=2	AF=1.00	AN=2	BaseQRankSi	DP=21	Dels=0.00	FS=3.222	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=57.70	MQ0=0
chr10	3978749		T	TTTTG	467.75	PASS	C=1	AF=0.500	AN=2	BaseQRankSi	DP=14	FS=6.990	HaplotypeSc	MLEAC=1	MLEAF=0.50	MQ=56.95	MQ0=0	MQRankSum
chr10	4007800	rs29378605	rs222352237	T	C	684.77	ASS	AC=2	AF=1.00	AN=2	BaseQRankSi	DP=28	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00
hr10	4032345	rs29322875	T	С	525.28	PASS	C=1	AF=0.500	AN=2	BaseQRankS	DP=26	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=1	MLEAF=0.50	MQ=60.00	MQ0=0
chr10	4041894	rs47215848	G	T	941.77	PASS	C=2	AF=1.00	AN=2	BaseQRankSi	DP=34	Dels=0.00	FS=4.523	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
hr10	4048178	rs51676436	C	T	1479.77	PASS	/C=2	AF=1.00	AN=2	BaseQRankSi	DP=60	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
hr10	4103364	rs29333656	C	A	1495.77	PASS	C=2	AF=1.00	AN=2	BaseQRankS	DP=61	Dels=0.00	FS=10.685	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0
hr10	4155568		CTT	C	556.73	PASS	C=2	AF=1.00	AN=2	DP=13	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=57.23	MQ0=0	QD=42.83	RPA=3,1
chr10	4155580	rs29320956	A	G	396.77	PASS	C=2	AF=1.00	AN=2	DP=14	Dels=0.00	FS=0.000	HaplotypeSc	MLEAC=2	MLEAF=1.00	MQ=55.69	MQ0=0	QD=28.34
hr10	4166804	rs29357681	. c	T	104.03	LowCoverage	C=2	AF=1.00	AN=2	DP=4	Dels=0.00	FS=0.000	HaplotypeSci	MLEAC=2	MLEAF=1.00	MQ=60.00	MQ0=0	QD=26.01
hr10	4368518	rs51768590	A	C	177.77	PASS	C=1	AF=0.500	AN=2	BaseQRankSi	DP=101	Dels=0.00	FS=4.671	HaplotypeSci	MLEAC=1	MLEAF=0.50	MQ=55.20	MQ0=0
hr10	4368528	rs227191183	AE	G	287.77	PASS	C=1	AF=0.500	AN=2	BaseQRankSi	DP=129	Dels=0.00	FS=11.550	HaplotypeSci		MLEAF=0.50		MQ0=0
hr10		rs258468563	1000	T	100000000000000000000000000000000000000	The second secon	C=1	AF=0.500	AN=2	BaseQRankSi			The state of the s	HaplotypeSc		MLEAF=0.50		MQ0=0

option for soft filter

# Total exome variants by strain background

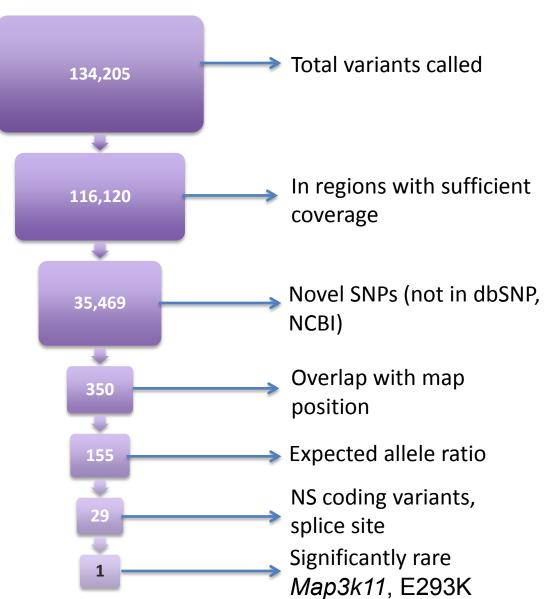


# Stepwise reduction of candidate variants, single sample



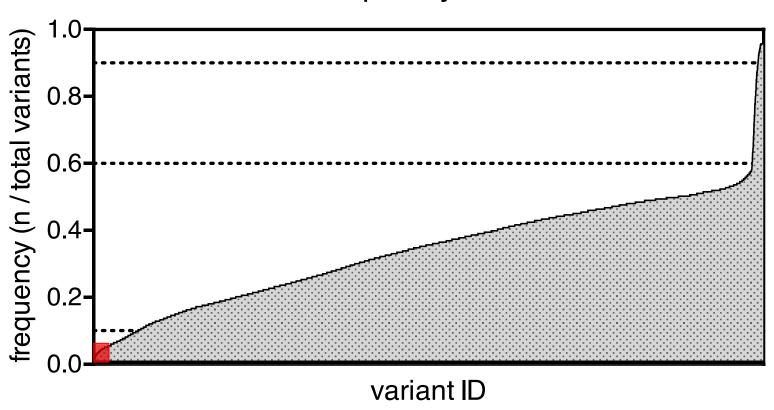


- reduced bone mass
- abnormal tooth pulp

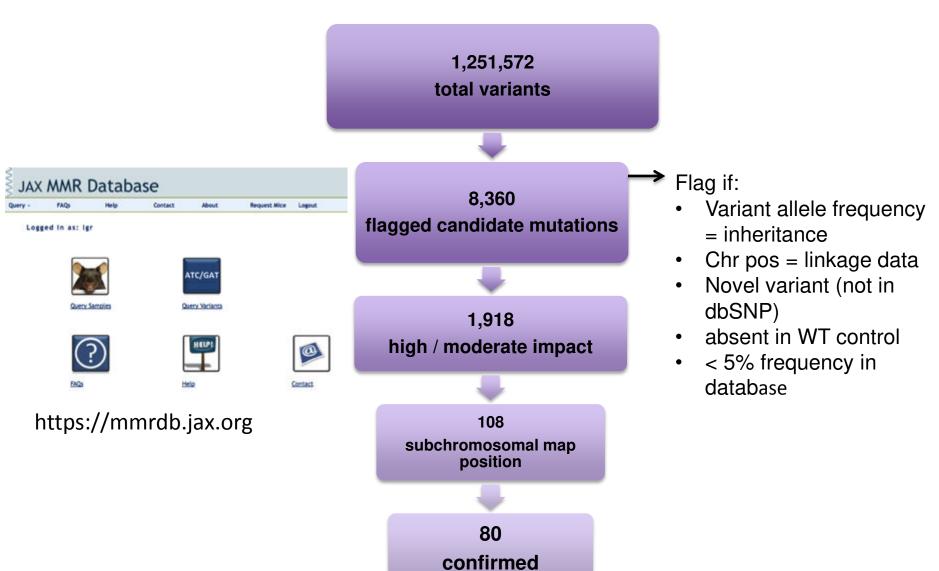


### Exome variant frequency distribution, >200 mouse exomes

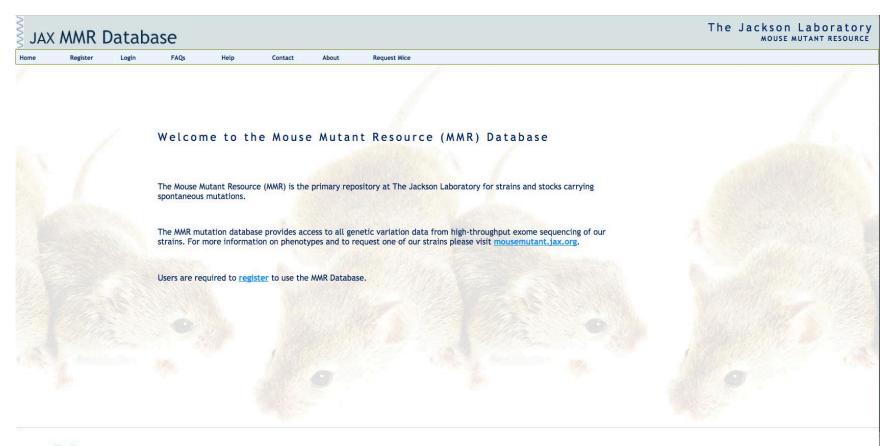
variant frequency, 200 exomes



# Exome variants $\rightarrow$ putative causative mutations



#### Demo



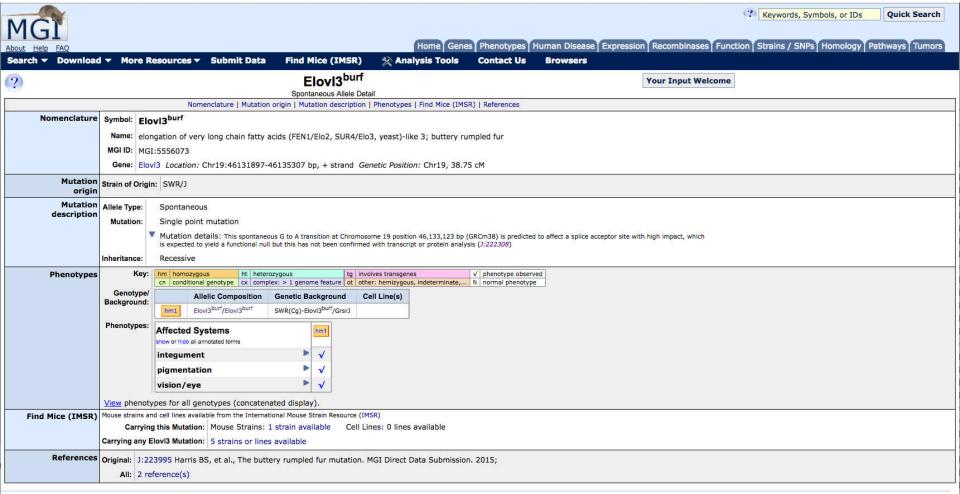




### Candidate gene analysis

- MGI batch query alleles, phenotypes
- MouseMine gene expression
- Validation PCR genotyping of samples from the colony (segregation analysis)
- Allele or complementation tests (limited)
- Data release...

#### MGI Direct Data Submission



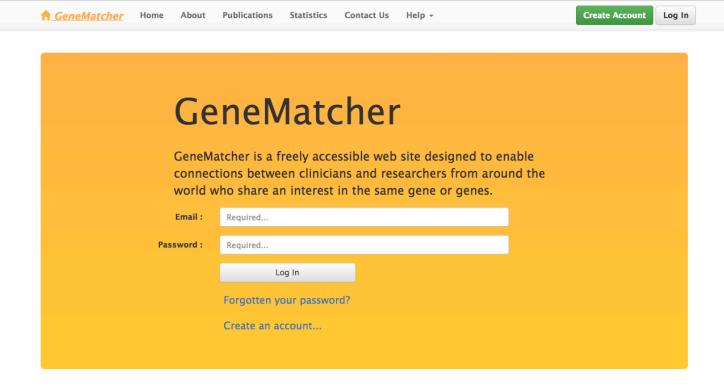
Contributing Projects:

Citing These Resources
Funding Information
Warranty Disclaimer & Copyright Notice
Send questions and comments to User Support

Mouse Genome Database (MGD), Gene Expression Database (GXD), Mouse Tumor Biology (MTB), Gene Ontology (GO), MouseCyc



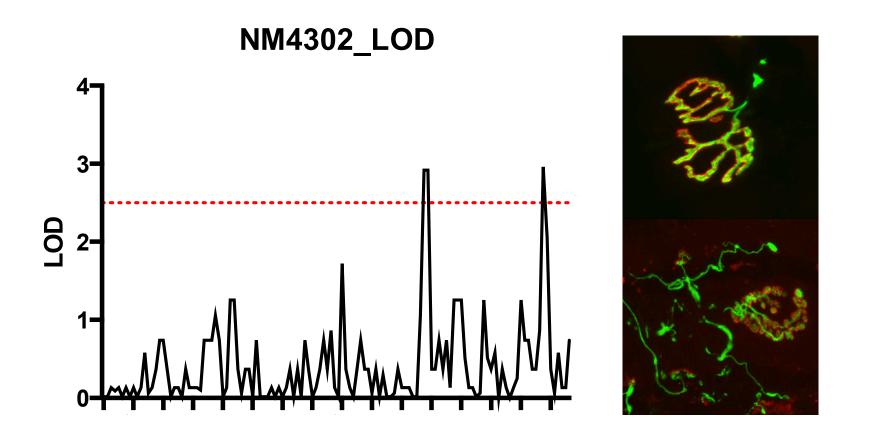
### Connecting with clinicians



Centers for Mendelian Genomics Centers for Mendelian Genomics

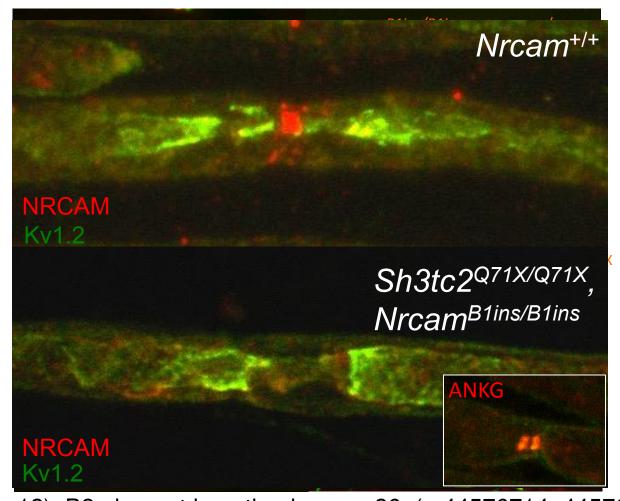
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# NM4302, progressive paralysis and death at 5 months



Robert Burgess, Greg Cox, Dave Schroeder

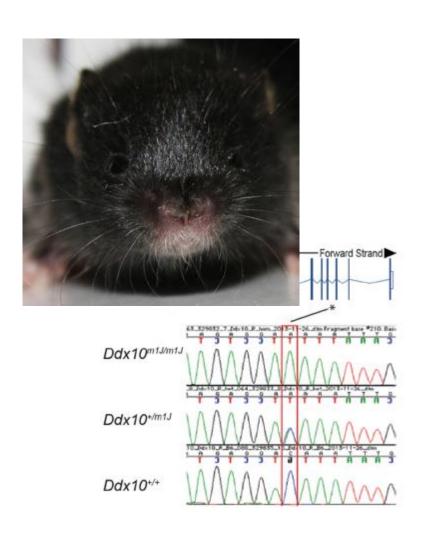
## NM4302, progressive paralysis and death at 5 months

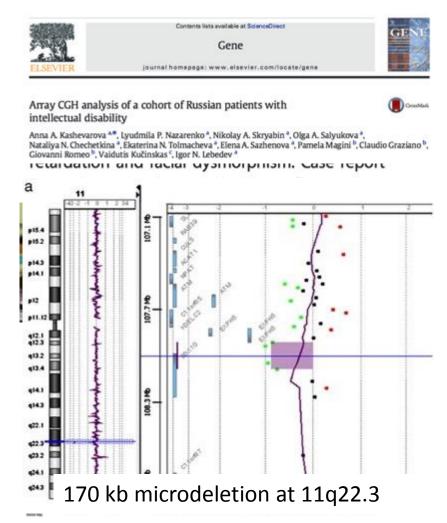


Nrcam (Chr12): B2 element insertion in exon 26 (g.44576714\_44576715insB2) Sh3tc2 (Chr18): C to T mutation (c.211C>T), results in p.Q71X

Rob Burgess, Greg Cox, Abby Tadenev, The Jackson Laboratory

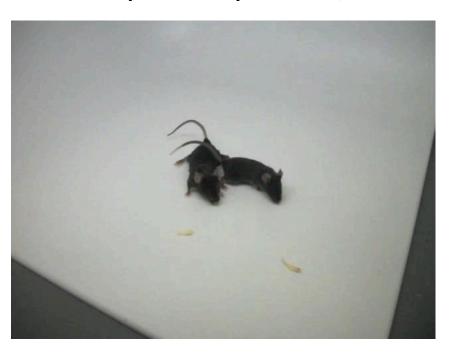
#### Ddx10 – DEAD box polypeptide10





#### *Fdxr* – ferredoxin reductase

#### nm4877, *Fdxr*<sup>m1J</sup>, R389Q



#### **Clinical data – Ambry Genetics**

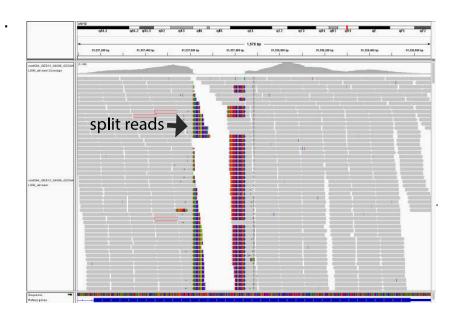
- Recessive missense mutations in three pedigrees
- Suspected mitochondrial disorder with polyneuropathy, gait defects, vision impairment

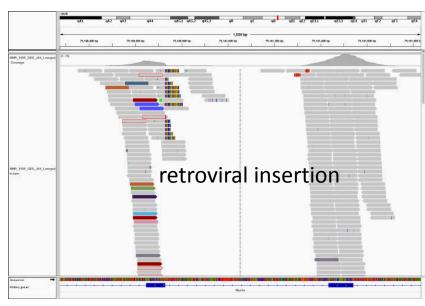
### What are we missing?

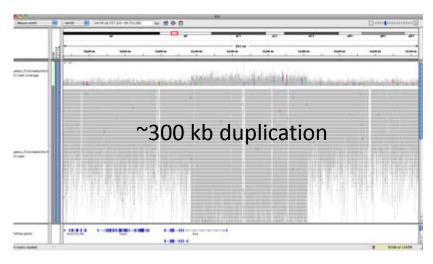
Exome sequencing of mice with proven Mendelian phenotypes provides putative causative mutations with a success rate of ~55%.

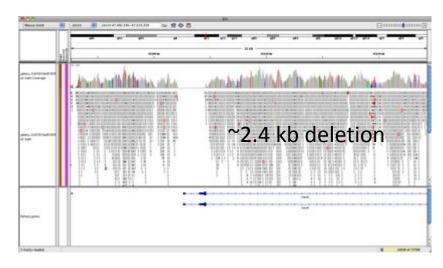
- Poor gene annotation
- Structural mutations
- Non-coding mutations

#### Structural mutations

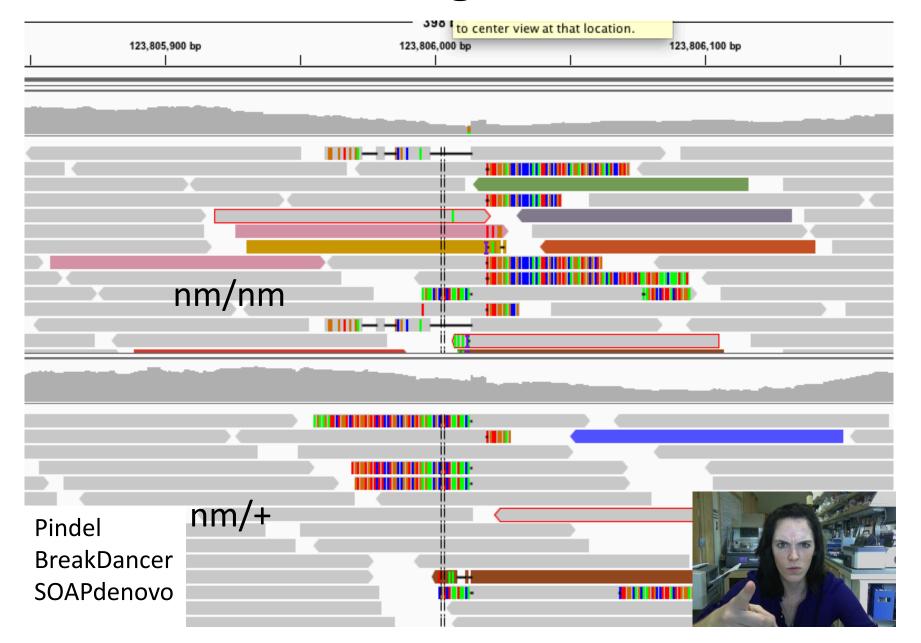








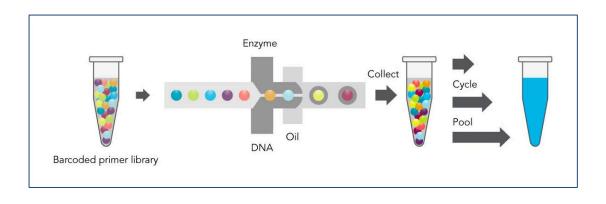
### Non-coding mutation



## Approaches for solving failed exomes

- RNA Seq
  - correct tissue and relevant time points, live mice
- CGH
  - works well for CNV detection > ~5 Kb
- Whole genome sequencing → 100 pending, stay tuned
- 10X Genomics → synthetic long reads

# 10X Genomics – synthetic long read technology

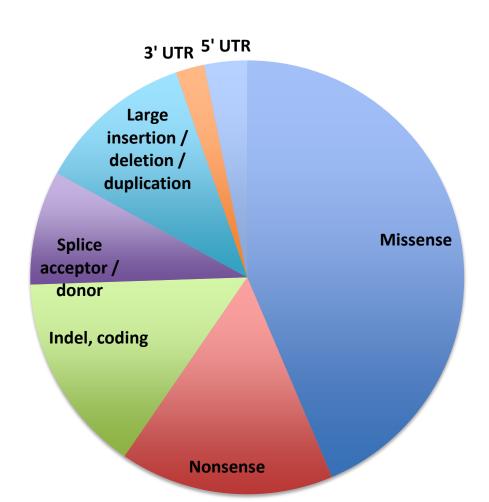


A B C

### Summary

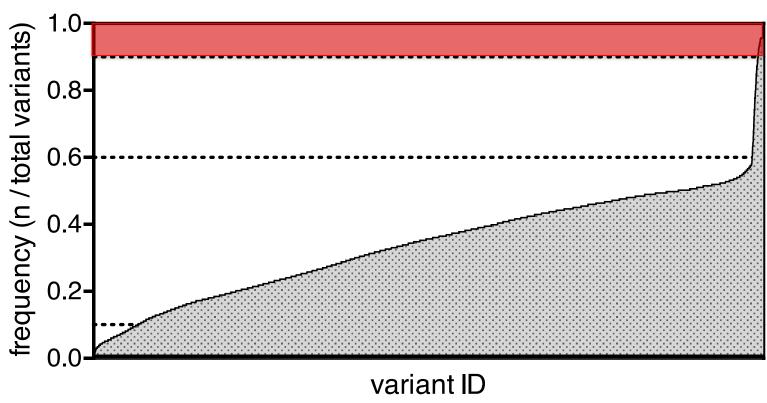
- Using exome sequencing, our rate of causative gene discovery in spontaneously arising mouse models of Mendelian disease has increased >10X.
- ~10% are the first phenotypic alleles and over 80% are informative for human Mendelian disease modeling.
- Unique resource of exome recalcitrant, Mendelian disease causing mutations.
- Using both forward and reverse genetic approaches we are creating new mouse models with unprecedented speed and precision.

# Putative pathogenic mutation spectrum, 91 spontaneous mutations



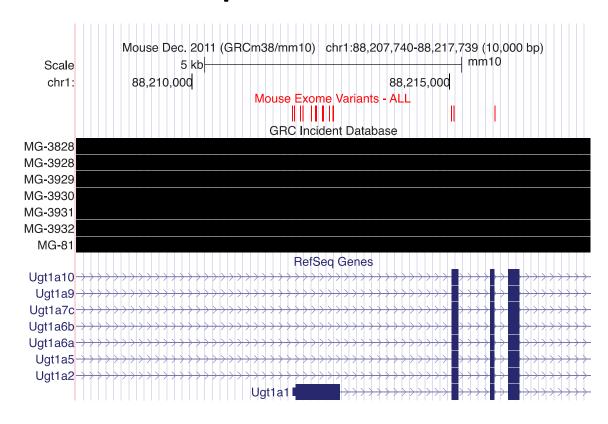
#### Exome variant frequency distribution

#### variant frequency, 200 exomes



All variants passing soft filter

# Variants with allele frequency >95% map to regions with reported assembly problems



### Acknowledgements



JAX Genetic Resources Steve Murray, JAX KOMP Cat Lutz, JAX Repository

#### Resource for Comparative Mendelian Genomics, Mouse Mutant Resource

David Bergstrom Heather Fairfield Belinda Harris Son Yong Karst Melissa Berry

Reinholdt Lab
Candice Byers
Anne Czechanski
Whitney Martin

#### JAX Mammalian Genetics Research

Rob Burgess Greg Cox Kevin Seburn

### JAX Computational Sciences

Anuj Srivastava Guru Ananda Roger Liu Anu Lakshminaryana Centers for Mendelian Genomics Baylor/Hopkins David Valle

#### **Ambry Genetics**

Nara Sobreira

Deepali Shinde Kelly Hagman Sha Tang Zoe Powis

