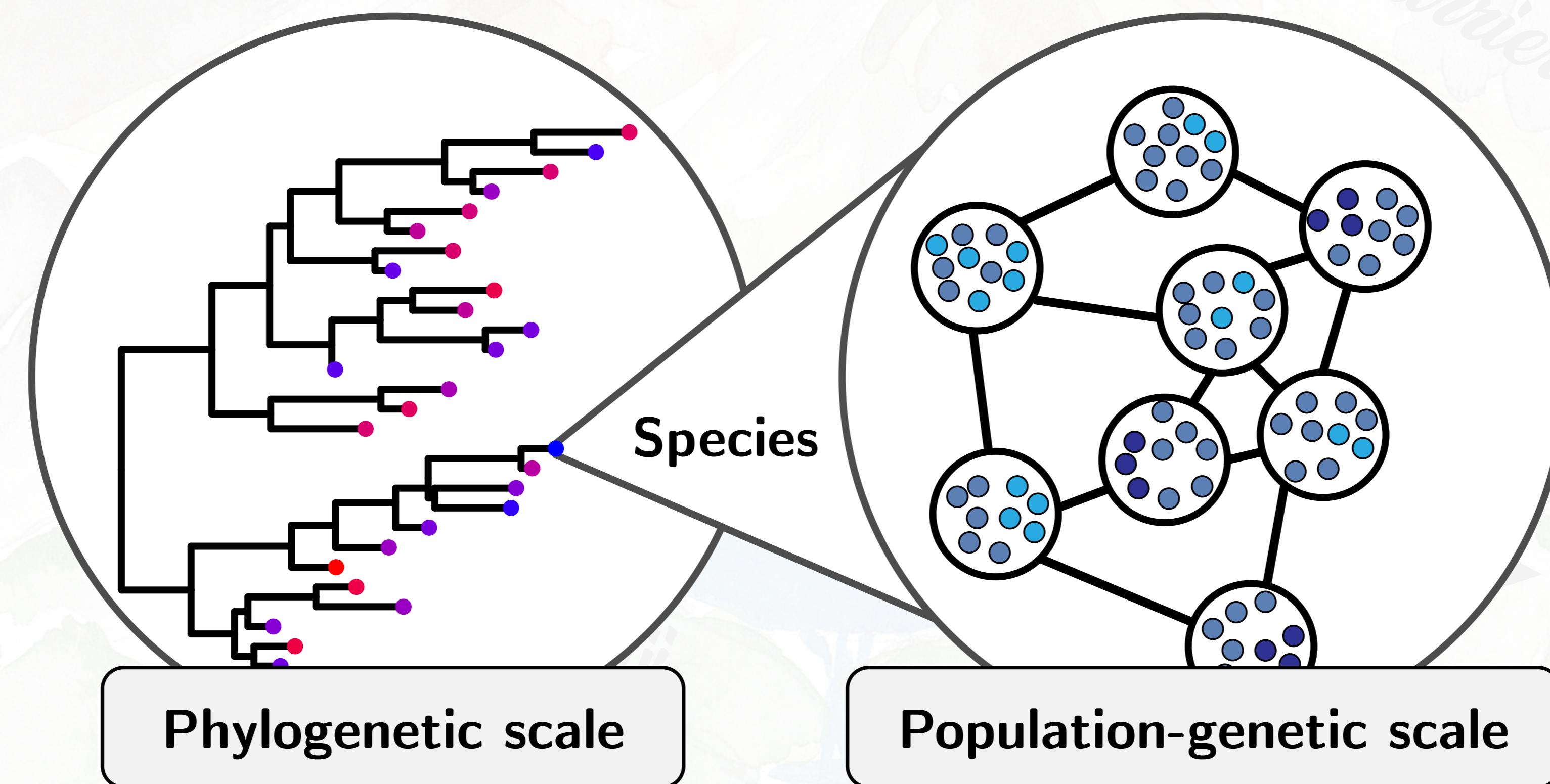
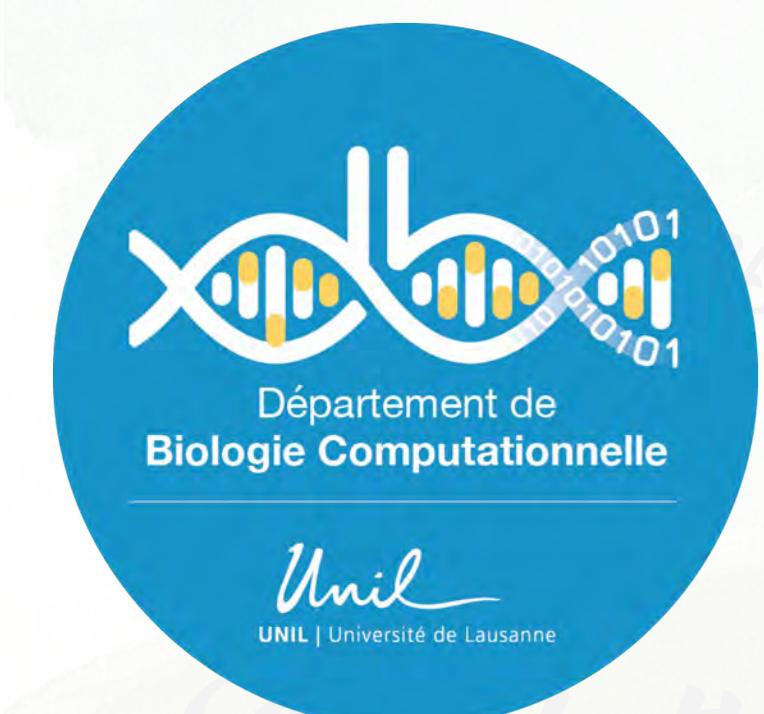
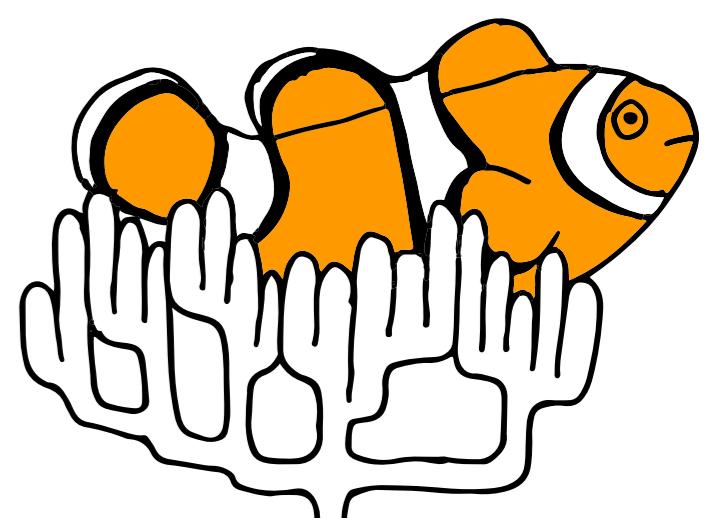


Predicting Selection on Traits and Sequences: Contrast Across Evolutionary Scales



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What tools to bridge evolutionary scales?

A combination of theoretical models and empirical studies.

Empirical studies

Genes and sites under adaptation at the phylogenetic scale also exhibit adaptation at the population-genetic scale

Thibault Latrille^{a,b,c,1}, Nicolas Rodrigue^d, and Nicolas Lartillot^a

Bridging Time Scales in Evolutionary Biology



Diego A. Hartasánchez, Thibault Latrille, Marina Brasó-Vives, and Arcadi Navarro

Equal contribution: DAH & TL

Inferring Long-Term Effective Population Size with Mutation-Selection Models

Thibault Latrille^{ID,*1,2}, Vincent Lanore,¹ and Nicolas Lartillot¹

Estimating the proportion of beneficial mutations that are not adaptive in mammals

T. Latrille, J. Joseph, D. A. Hartasánchez, N. Salamin

This article is a preprint

Detecting diversifying selection for a trait from within and between-species genotypes and phenotypes

T. Latrille¹, M. Bastian², T. Gaboriau¹, N. Salamin¹

An Improved Codon Modeling Approach for Accurate Estimation of the Mutation Bias

Thibault Latrille^{ID,*1,2} and Nicolas Lartillot^{ID¹}

Quantifying the impact of changes in effective population size and expression level on the rate of coding sequence evolution

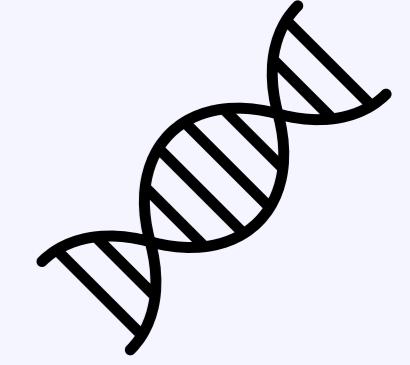
T. Latrille^{a,b,*}, N. Lartillot^a

Theoretical models

— PhD

— Postdoc

Sequences

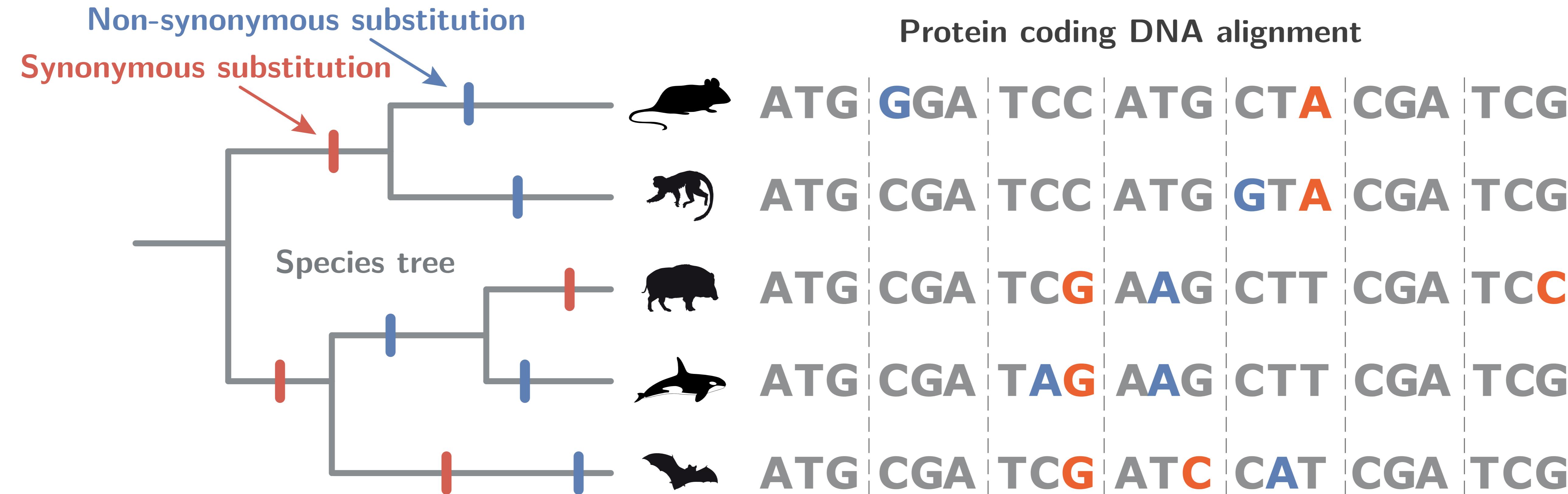


Part I

Can we predict the rate of protein evolution?

How to quantify changes in protein evolution?

With both synonymous and non-synonymous substitutions.



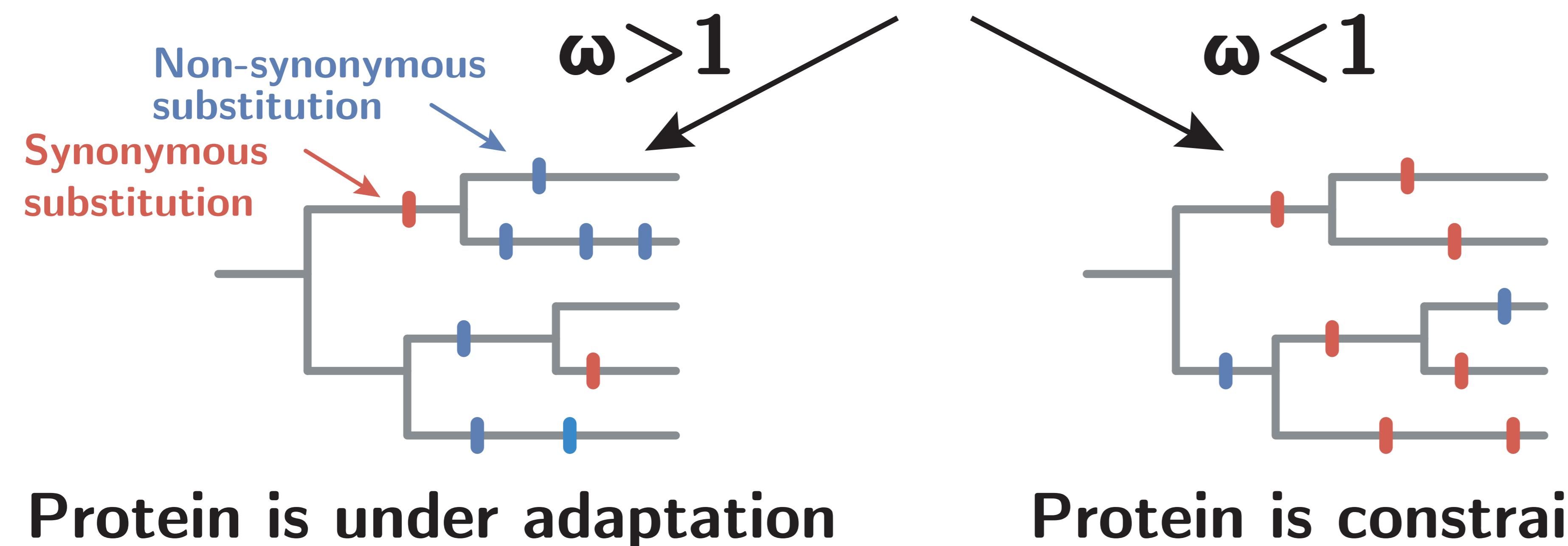
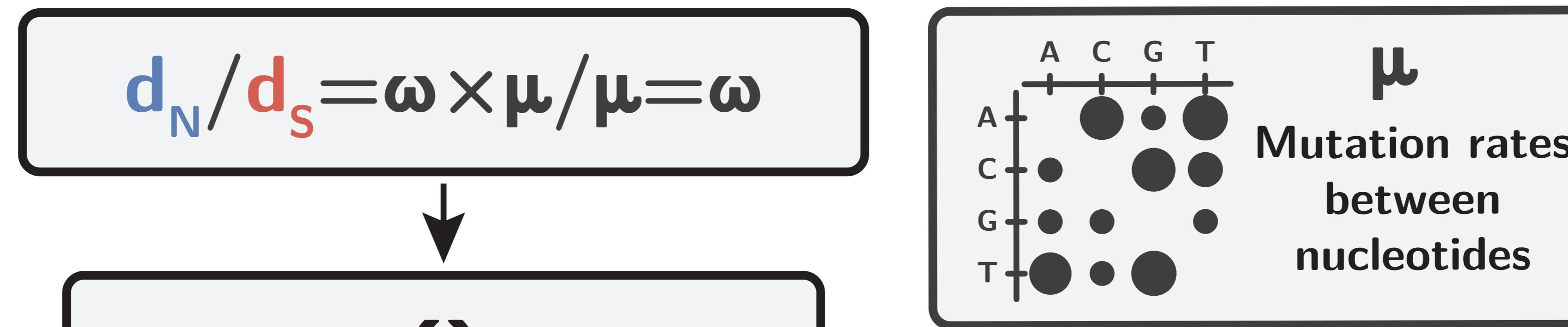
- **Non-synonymous** substitutions are reflecting the effect of mutation, selection and drift.
- **Synonymous** substitutions are considered selectively neutral, reflecting the mutational processes.

King & Jukes (1969); Kimura (1983); Goldman & Yang (1994); Muse & Gaut (1994).

How to measure the rate of protein evolution?

$d_N/d_S = \omega$ as the rate of protein evolution.

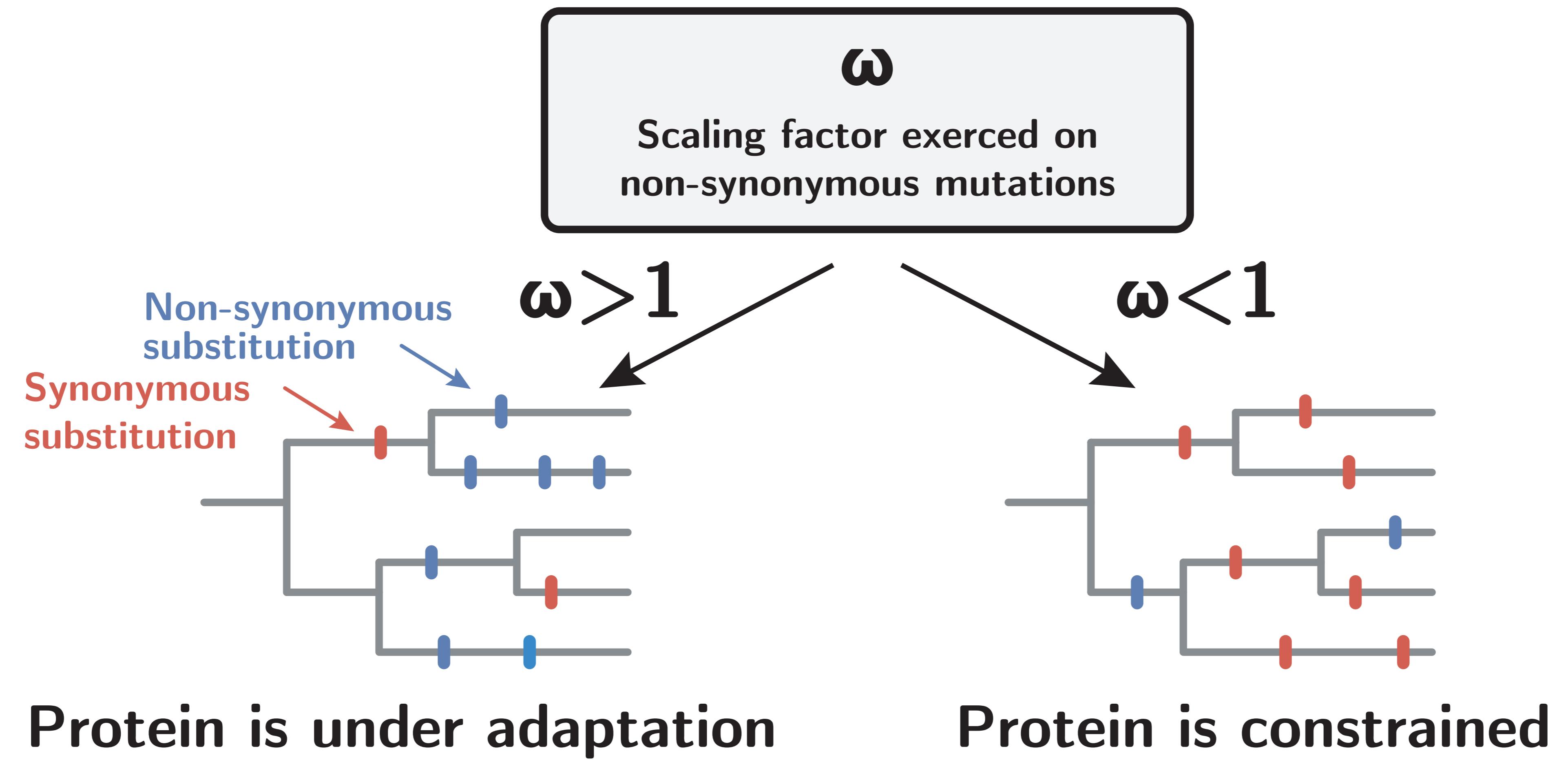
$$\begin{cases} d_S = \mu \text{ for synonymous substitutions.} \\ d_N = \omega \times \mu \text{ for non-synonymous substitutions.} \end{cases}$$



ω can be interpreted as the average fixation probability of non-synonymous mutations, relative to neutral mutations.

What are the predictors of ω ?

Few genes/sites under adaptation ($\omega>1$), a majority are constrained ($\omega<1$).

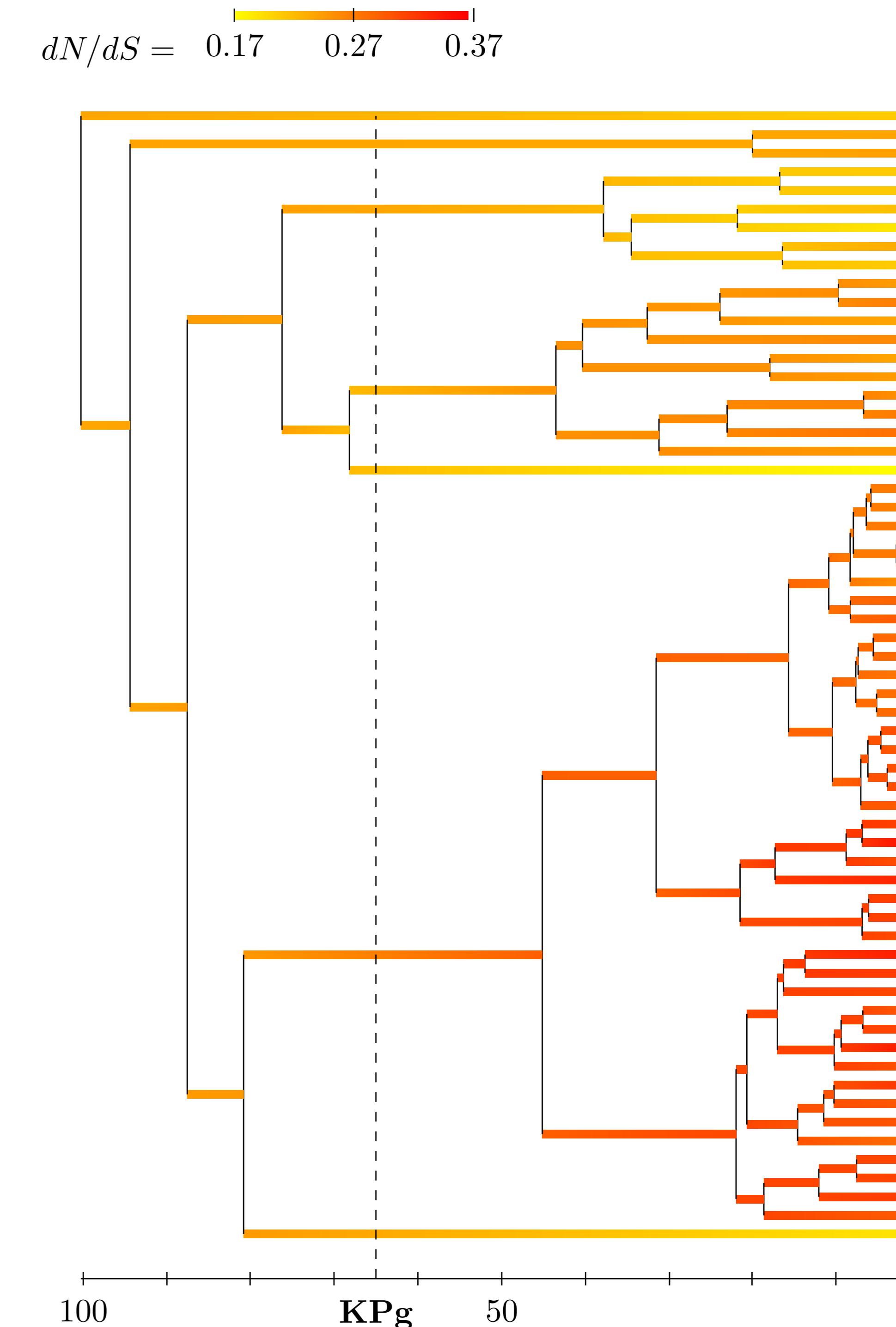
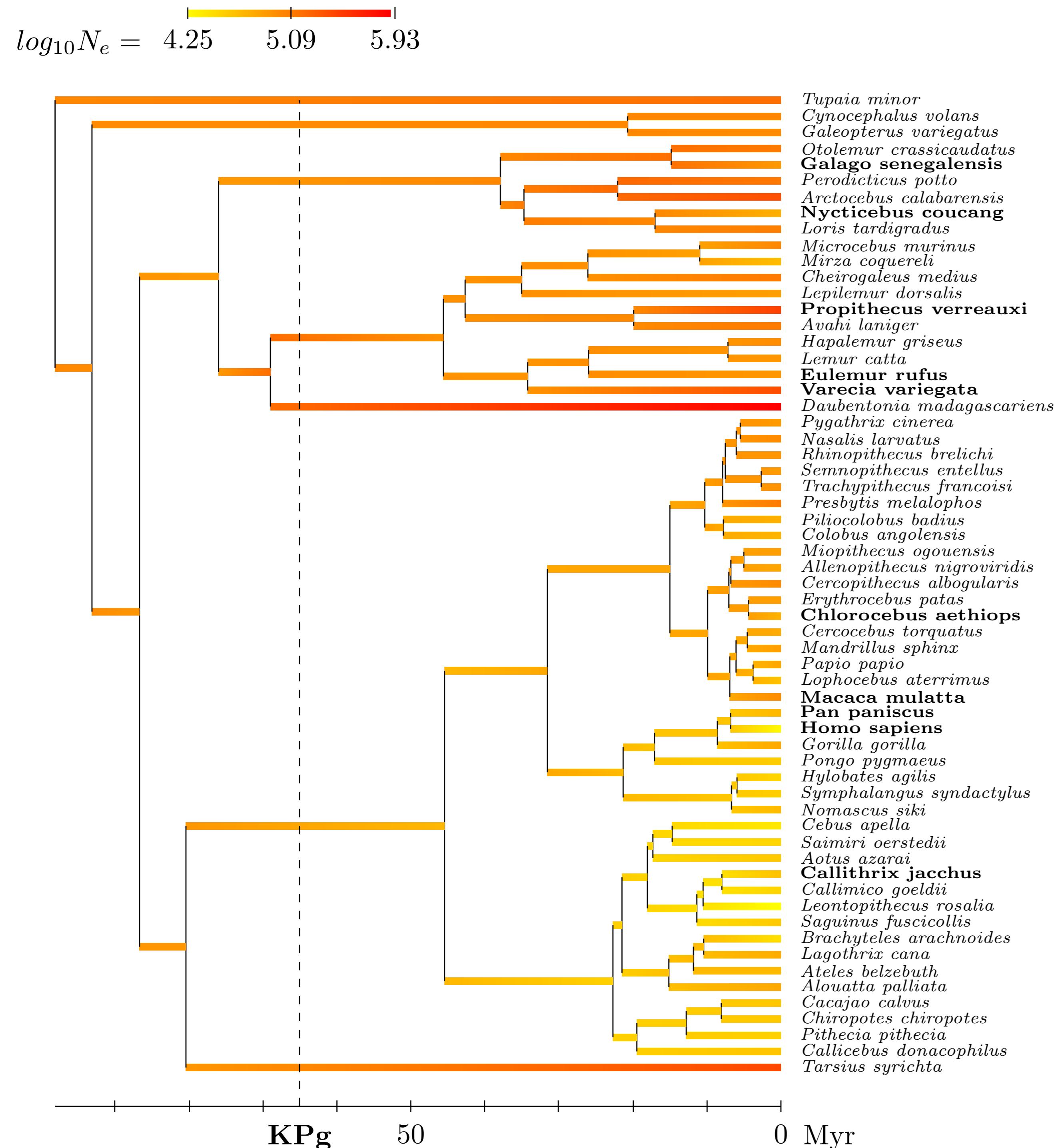


- A very few genes have $\omega>1$.
Kosiol *et al* (2008).
- But we can detect sites with $\omega>1$.
Nieslen & Yang (1998); Enard *et al* (2016).
- Some branches can have a transient $\omega>1$.
Yang & Nielsen (1998); Zhang & Nielsen (2005).

- Lower ω for highly expressed proteins.
Drummond (2005); Zhang & Yang (2015).
- Lower ω for buried sites inside a protein.
Ramsey *et al* (2011); Echave *et al* (2016).
- Lower ω for short-lived and smaller species.
Popadin *et al* (2007); Lanfear *et al* (2010).

Is effective population size (N_e) predicting ω ?

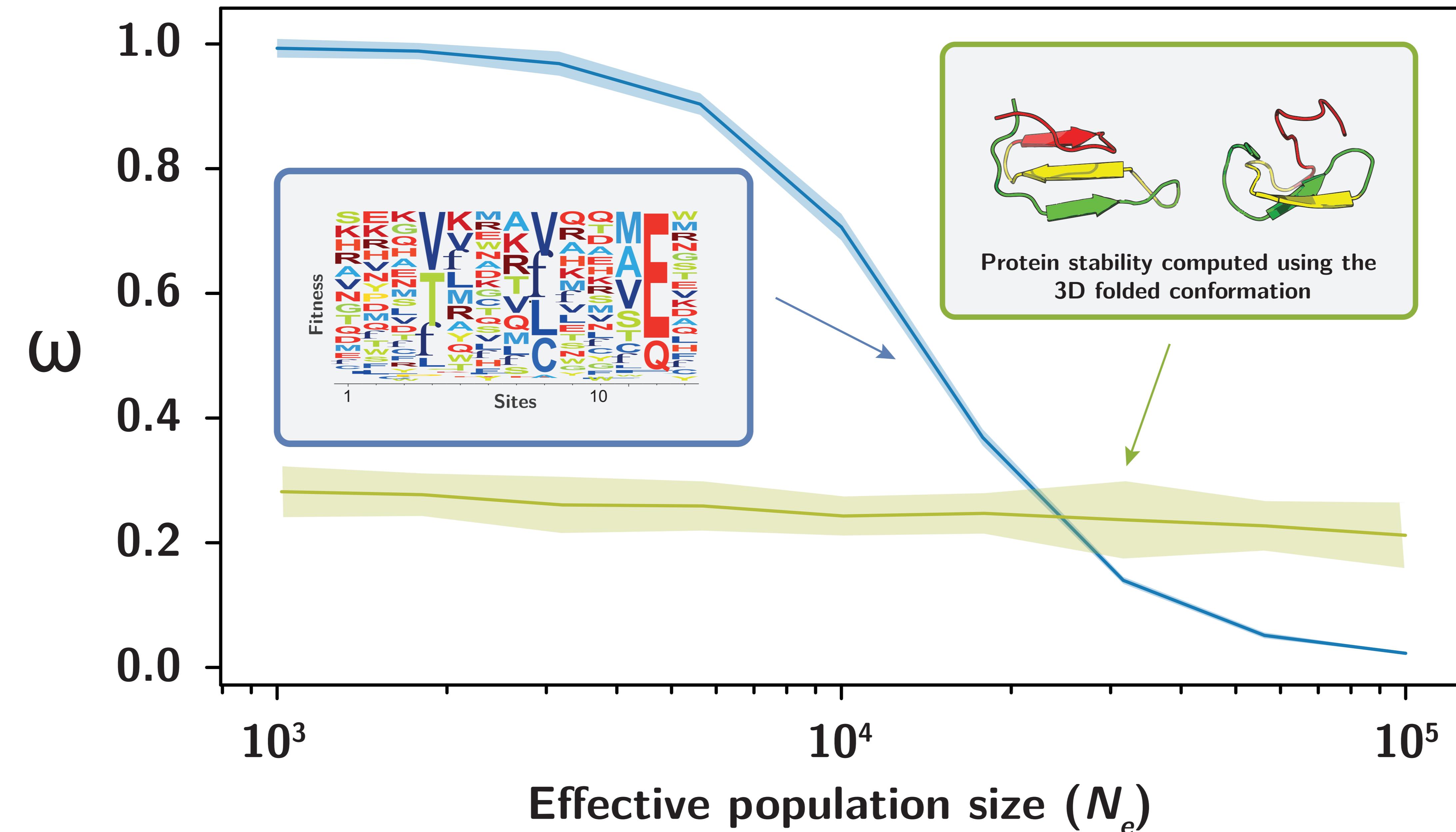
Higher N_e results in lower ω due to better efficacy of selection ($r=-0.58$).



Brevet & Lartillot (2021)

Can we theoretically use ω to predict N_e ?

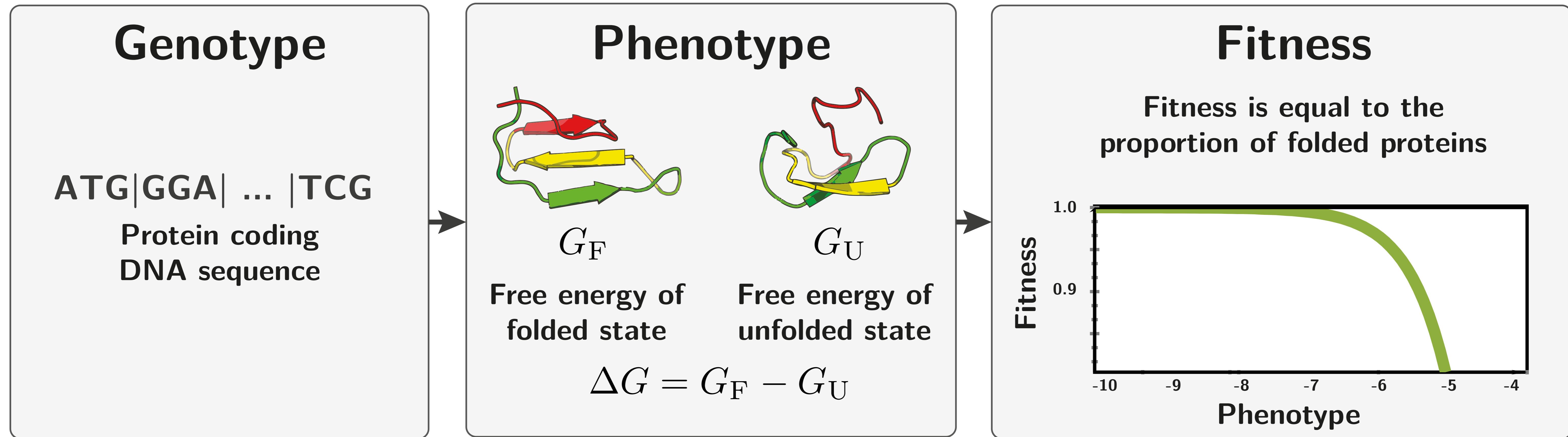
Not directly because the relationship depends on the model of protein evolution.



Latrille et al. (2021)

What is the expected relationship between ω and N_e ? (1/4)

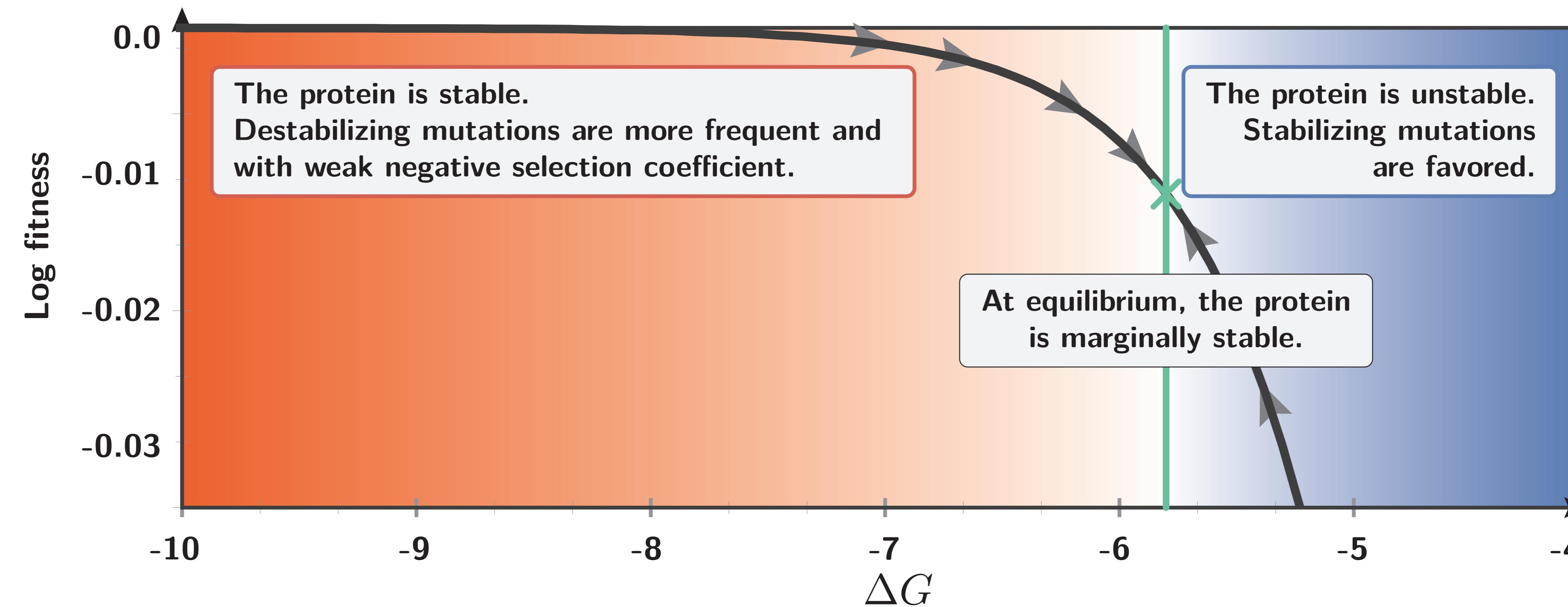
We first need to define a genotype-phenotype-fitness relationship.



Miyazawa and Jernigan (1985), Williams et al (2006), Goldstein (2011), Pollock et al (2012)

What is the expected relationship between ω and N_e ? (2/4)

Then we need to find the equilibrium and ω at this equilibrium.

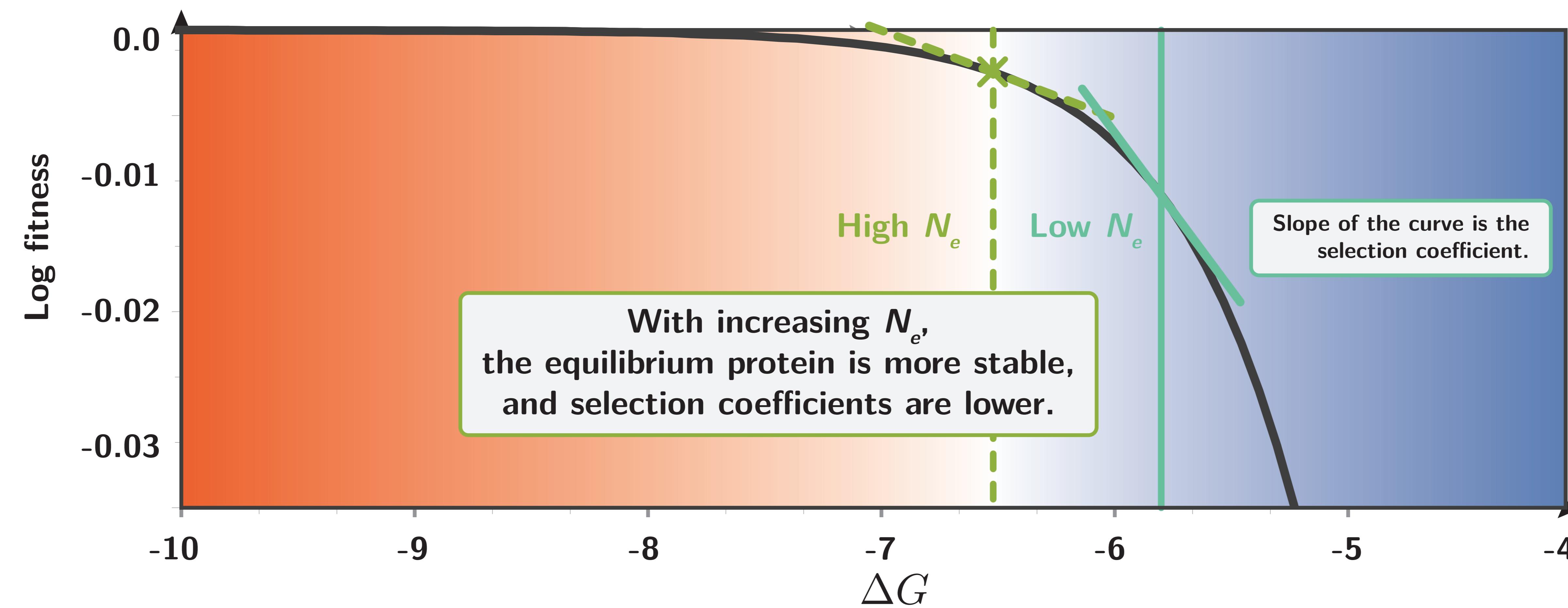


- The optimal stability of proteins is never achieved.
- Marginal stability is the default expectation of the mutation-selection balance even under directional selection for stability.

Taverna & Goldstein (2002)

What is the expected relationship between ω and N_e ? (3/4)

Then we derive how changes in N_e shift the equilibrium.



- Selection coefficient is dependent on the position in the fitness landscape.
- We can then derive the relationship between N_e and ω as a function of the microscopic molecular parameters of the model.

Cherry (1998); Goldstein (2013).

What is the expected relationship between ω and N_e ? (4/4)

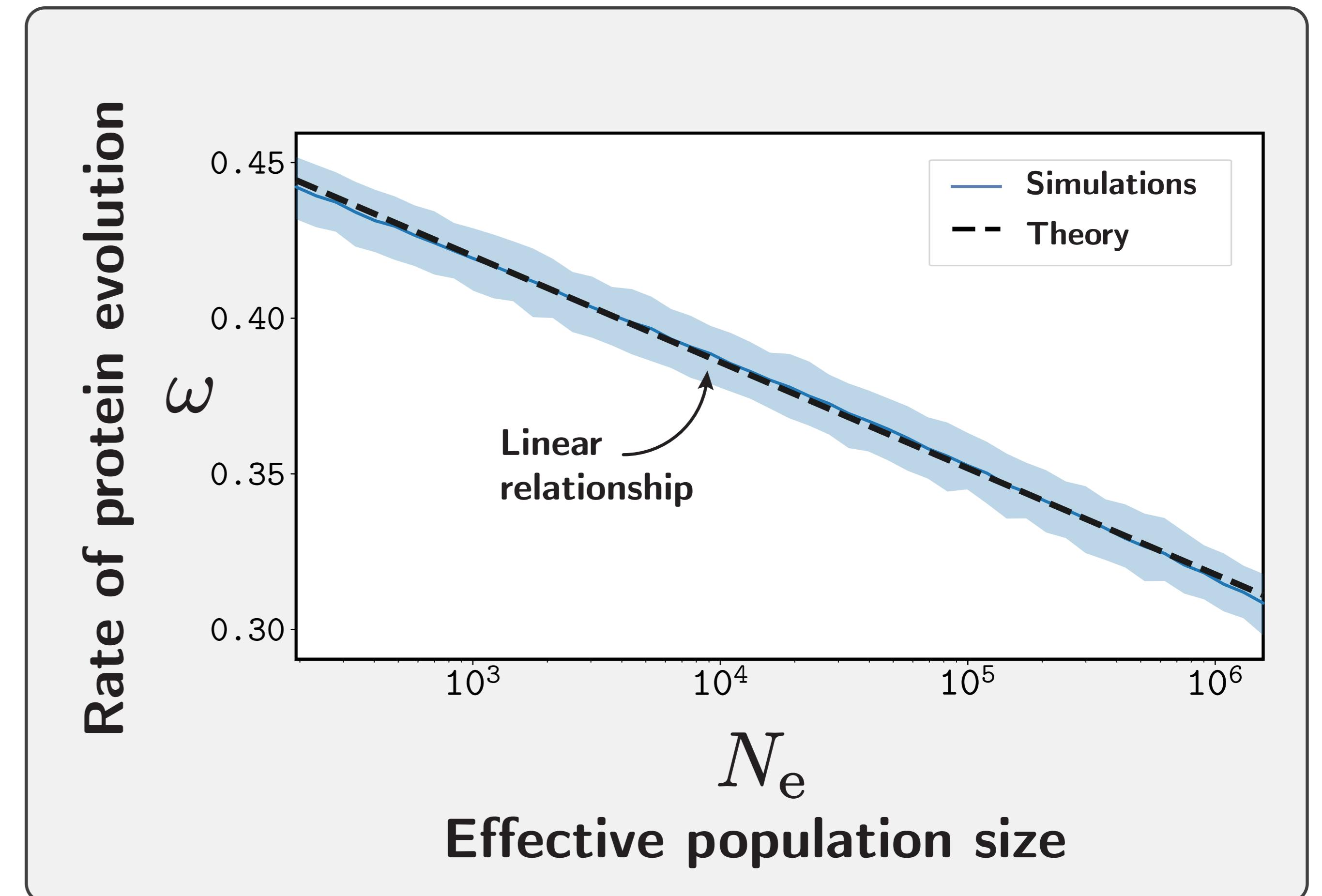
Negative linear relationship between ω and $\log(N_e)$.

Given:

- T the temperature.
- n the number of sites in the protein.
- $\Delta\Delta G > 0$ the destabilizing effect of a mutation.
- x the proportion of destabilizing sites (phenotype).
- $f(x)$ the phenotype-fitness map.
- x^* the equilibrium of x .

The response in ω after a change in N_e is:

$$\frac{d\omega}{d \ln(N_e)} \simeq - \frac{\frac{\partial \ln f(x^*)}{\partial x^*}}{\frac{\partial^2 \ln f(x^*)}{\partial x^{*2}}} \simeq - \frac{T}{n \times \Delta\Delta G}.$$



Latrille & Lartillot (2021)

What is the relationship between ω and expression level?

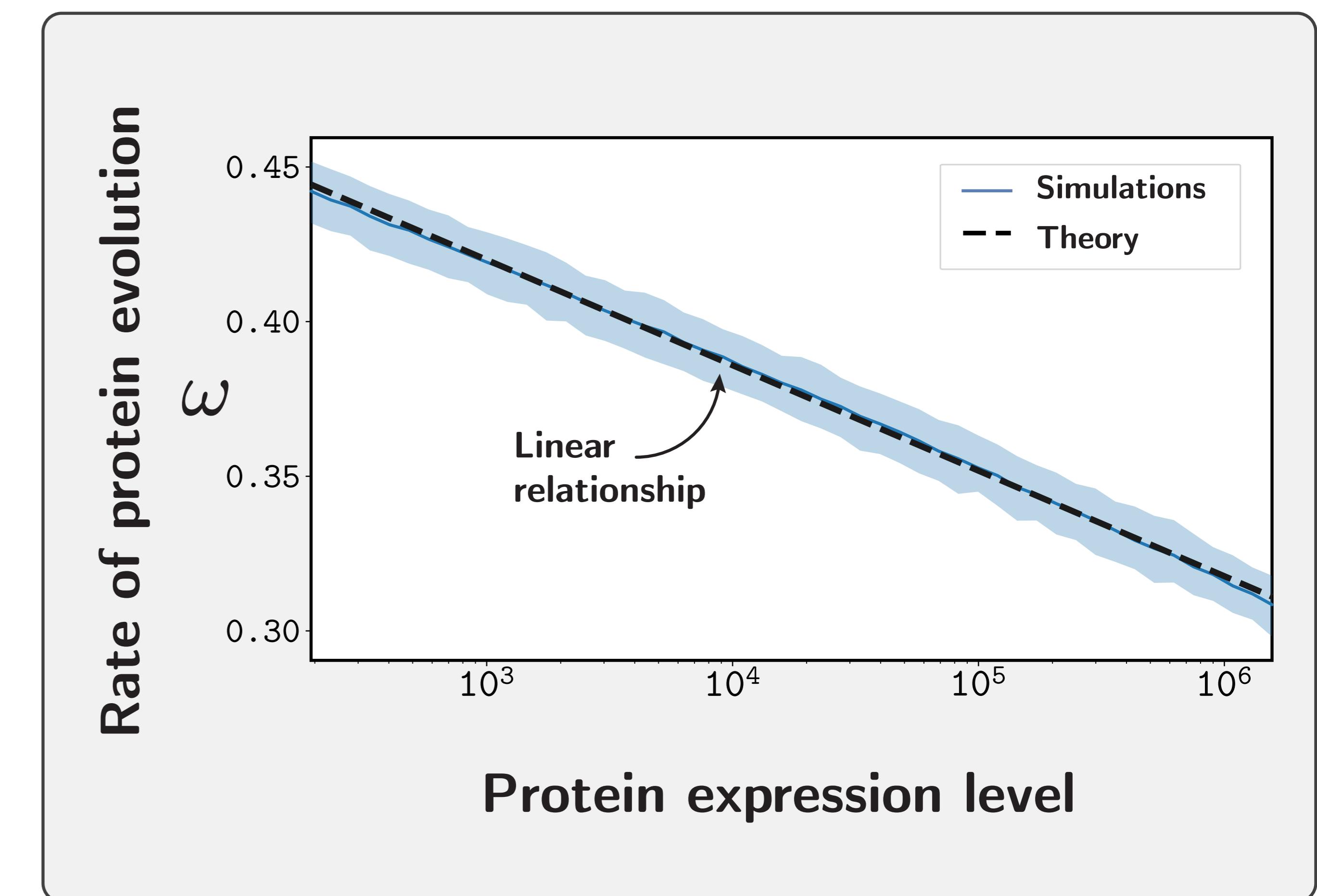
Negative linear relationship between ω and log of expression level.

If misfolded proteins are toxic, the decrease in fitness is proportional to protein expression level.

- T the temperature.
- n the number of sites in the protein.
- $\Delta\Delta G > 0$ the destabilizing effect of a mutation.
- x the proportion of destabilizing sites (phenotype).
- $f(x)$ the phenotype-fitness map.
- x^* the equilibrium of x .

The response in ω after a change in protein expression

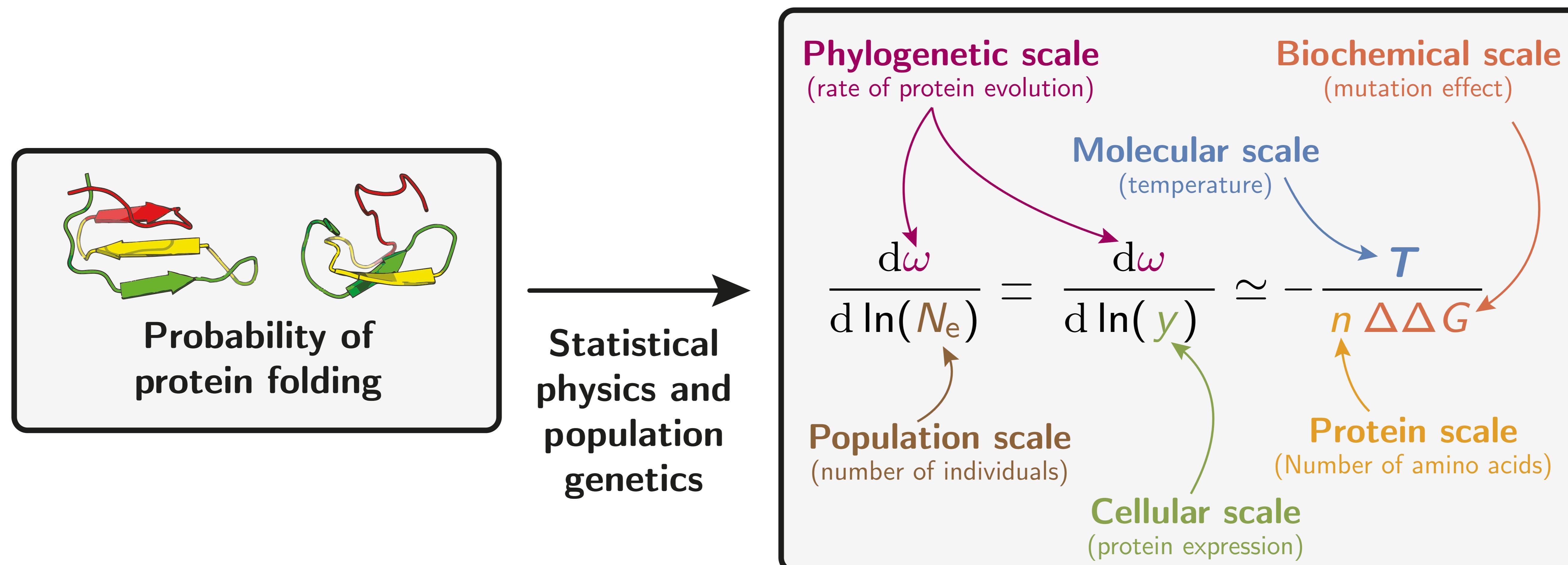
$$\frac{d\omega}{d \ln(y)} \simeq - \frac{\frac{\partial \ln f(x^*)}{\partial x^*}}{\frac{\partial^2 \ln f(x^*)}{\partial x^{*2}}} \simeq - \frac{T}{n \times \Delta\Delta G}.$$



Latrille & Lartillot (2021)

Can theoretical models of protein folding predict rate of evolution?

Models form a bridge across different scales and can be tested.



Chapter I

Can we predict the rate of protein evolution (ω)?

- With a theoretical model for selection on protein folding, ω is linearly decreasing with N_e and expression level (on log scale).
- This model forms a bridge across different scales and can be tested.
- In our model, there is no adaptation possible, ω is always < 1 .
- How to detect adaptation when proteins are generally constrained?

Quantifying the impact of changes in effective population size and expression level on the rate of coding sequence evolution

T. Latrille ^{a,b,*}, N. Lartillot ^a