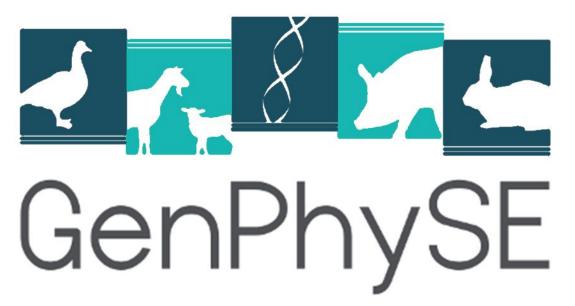
## INRAO>

## Environmental genomics of honey bees through the direct sequencing of hive products



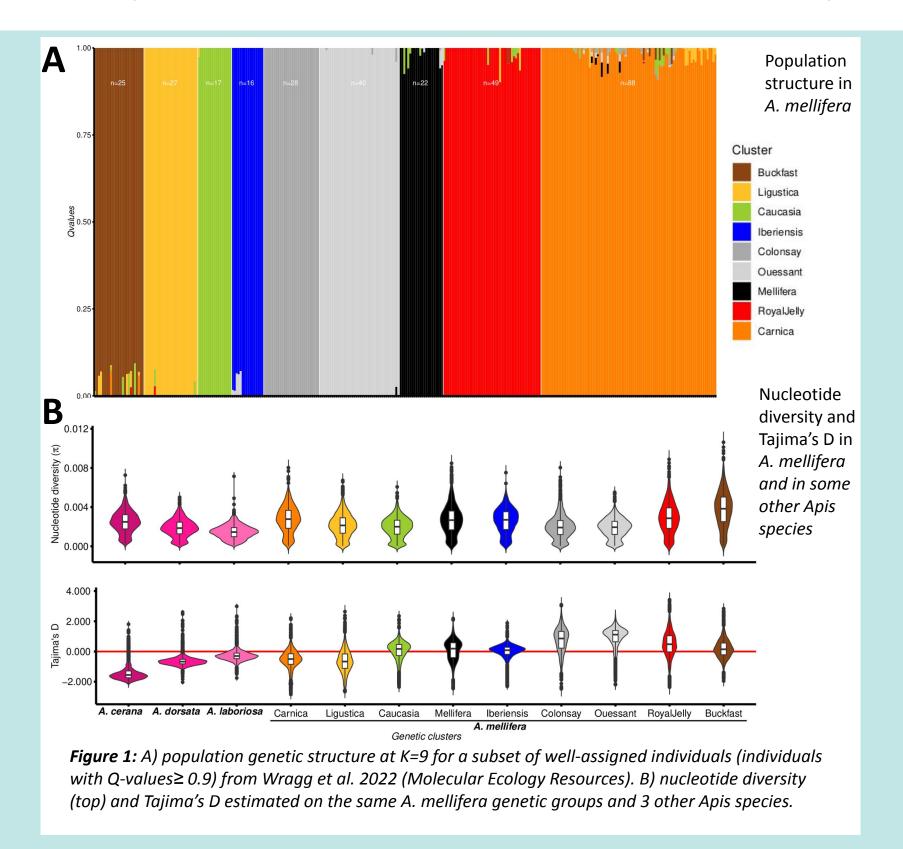


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Our research team located at INRAE in Toulouse, France specializes in the description of the genetic diversity and adaptation of honey bees.

During the last decade, we have put considerable effort into collecting and whole genome sequencing of hundreds of haploid males to describe the population structure and levels of diversity in Europe (Fig. 1).



Recently, Bovo et al (2020) sequenced three honey samples allowing access to whole-genome sequences of the contributing honey bees. These results support highly variable yields among the three samples, with 1 to 65% of reads effectively corresponding to bee DNA. To our knowledge, no study successfully sequenced bee DNA from beeswax samples.

The sequencing of drones has many advantages, including a direct access to haplotypes. Collecting drones however requires regular visits to the hives, which can be an issue for large regional sampling. In addition, campaigns could be difficult or impossible to organize for practical and/or geopolitical issues.

Worldwide collection of samples

iversity of non-local honey

samples: Algeria, India,

Nepal, Sweden, ...

diversity of plants)

(honeys from a large

+ 2 beeswax samples from

Does direct sequencing of hive products represent a good strategy for future population genomic and metagenomic investigations?

The direct sequencing of honey allows to: ES - Perform non-lethal sampling - Reduce the carbon footprints U of our research (especially ADVANTA

associated with the sampling) - Have easy access to many samples given the importance of the market across the globe

- Access to some other genetic information (pollinated plant species, microbiome, Varroa, ...)

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**DNA** extraction for each **Transhumance of beehives** beehive sample & QC Black locust Sweet chestnut Lavander (Castanea) (Robinia) (Lavandula) - Fine tune a DNA extraction July 2023 June 2023 August 2023 protocol to yield high quality

> DNA from comb honey and also additional honeypot samples

> - Develop a specific protocol for DNA extraction from beeswax

- Overall DNA quantification

using Nanodrop & Picogreen

nuclear loci

- Specific amplification of honey bee DNA using mitochondrial and

Bee DNA amplification & shotgun sequencing

1: Wax Sample 1 - Our protocol 1 2: Wax S.1 - Our protocol 2 : Wax S.2 - Our protocol 1 4: Wax S.2 - Our protocol 2 5: T+ Bee Individual 1 5: T+ Bee Individual 2 7: Honey S.1 - CTAB Protocol 1 8: Honey S.1 - CTAB Protocol 2 9: Honey S.1 - CTAB Protocol 3 10: Honey S.1 - Our protocol 1 11: Honey S.1 - Our protocol 2 12: Honey S.1 - Our protocol 3 TM: 100bp DNA ladder

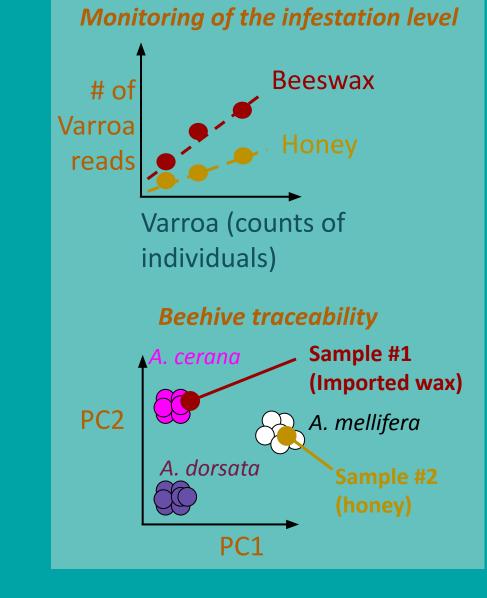
Product: Honey Wax DNA yield (ng/μL)

We successfully extracted and dosed DNA, and we were able to amplify mtDNA from several honeys and beeswaxes sample.

We also successfully identified bee DNA among the Illumina sequencing reads, including from wax samples, a world premiere to the best of our knowledge.

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These results are promising, pointing to a number of possible avenues. Thanks to the larger diversity of the project samples, we will perform a metagenomic analysis of all DNAs present in honey and wax. Reads associated with plant and Varroa DNA will be especially studied. We will especially try to monitor colony health by studying the infestation level of Varroa along a beekeeping season.



We also plan to analyse in detail the population structure of the samples, and to compare them to those already described (Fig. 1) in order to trace the geographical origin of the honey bees. This topic is crucial given that trade channels remain poorly traced, especially for beeswax. Chemical testing of wax is indeed not resolutive enough to allow this tracing. Consequently, direct shotgun sequencing of beewax could be a promising solution.

BACKGR

LIMITS

CURRENT