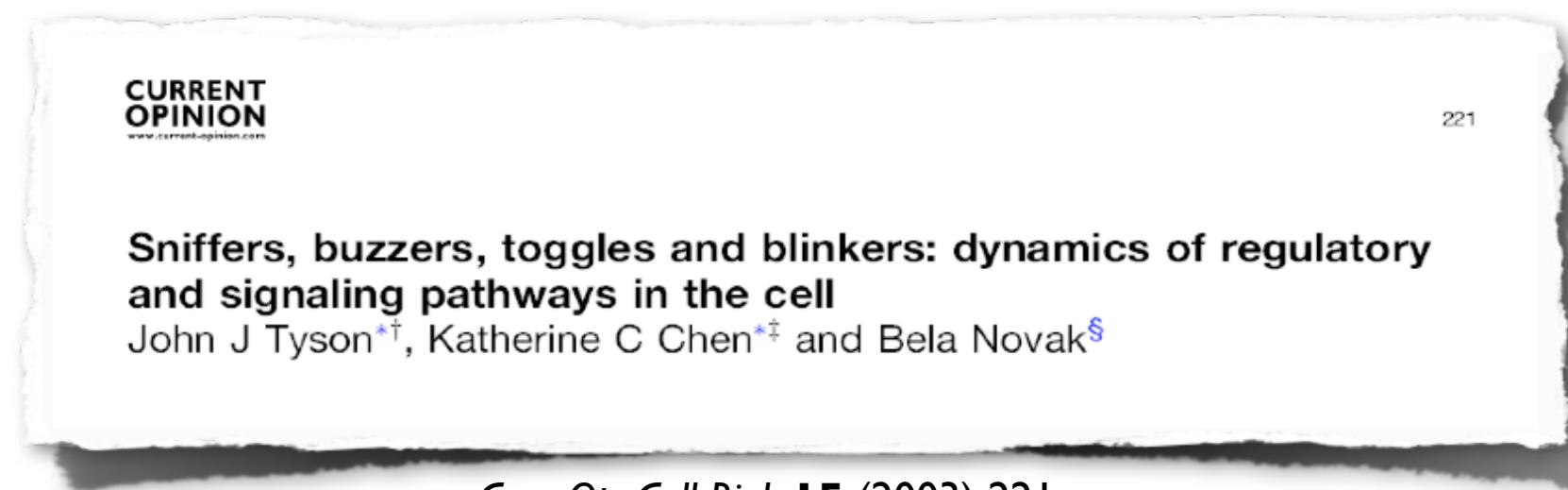


# V24 – Kinetic Motifs in Signaling Pathways

- Types of kinetic motifs in signaling pathways
- Application to cell cycle
- Circadian clocks



*Curr. Op. Cell Biol.* **15** (2003) 221

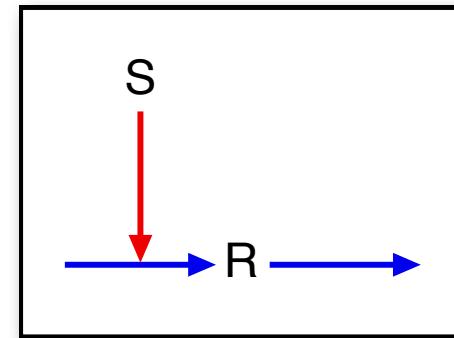
# Linear Response

E.g., protein synthesis and degradation (see lecture V8)

S = signal (e.g., concentration of mRNA)

R = response (e.g., concentration of a protein)

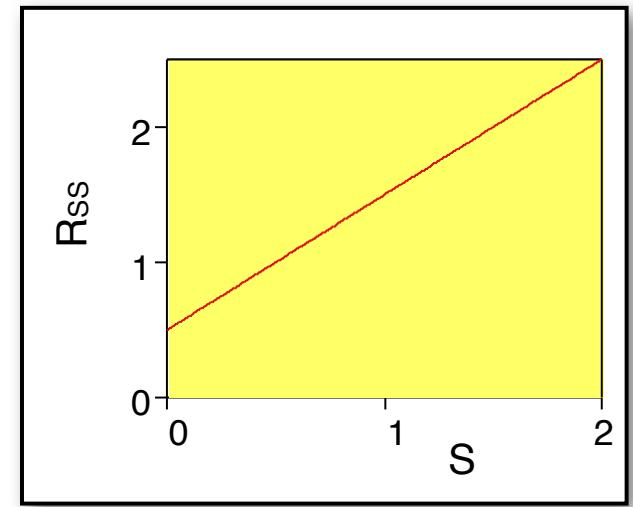
$$\frac{dR}{dt} = k_0 + k_1 S - k_2 R$$



At steady state (which implies  $S = \text{const}$ ):

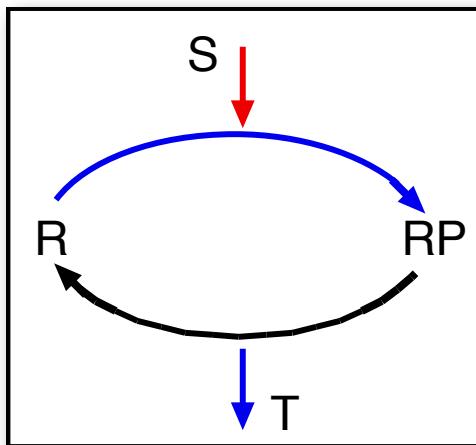
$$\left. \frac{dR}{dt} \right|_{R=R_{ss}} = 0 \Rightarrow R_{ss} = \frac{k_0 + k_1 S}{k_2} = \frac{k_0}{k_2} + \frac{k_1}{k_2} S$$

$R_{ss}$  linearly dependent on S



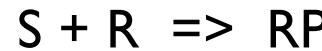
$$k_0 = 1, k_1 = k_2 = 2$$

# phosphorylation/dephosphorylation



„forward“: R is converted to phosphorylated form RP

„backward“: RP can be dephosphorylated again to R



$$\text{with } R_{\text{tot}} = R + RP$$

↑  
phosphorylated form

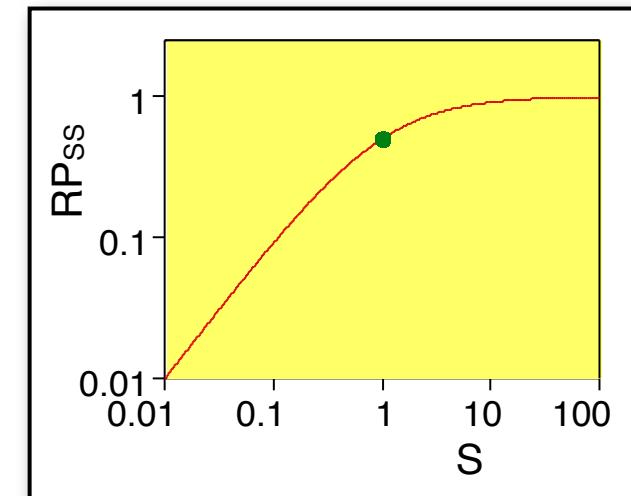
$$\frac{dRP}{dt} = k_1 SR - k_2 RP = k_1 S (R_{\text{tot}} - RP) - k_2 RP$$

Find steady state for RP: linear until saturation

$$RP_{ss} = \frac{k_1 R_{\text{tot}} S}{k_1 S + k_2} = \frac{R_{\text{tot}} S}{S + k_2/k_1} = \frac{R_{\text{tot}} S}{S + S_0}$$

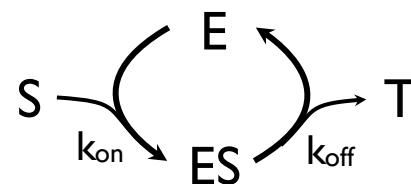
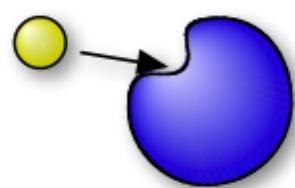
Output T proportional to RP level:

$$\frac{dT}{dt} = k_2 RP$$



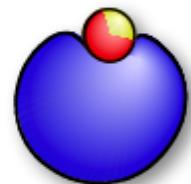
$$R_{\text{tot}} = 1, S_0 = 1$$

# Enzyme: Michaelis-Menten-kinetics

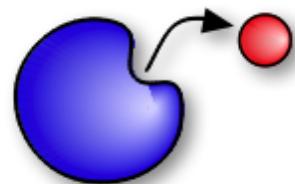


Reaction rate:

$$V = k_{off}ES$$



Steady state:  $k_{on}E \cdot S = k_{off}ES$



$$ES = \frac{k_{on} E \cdot S}{k_{off}} = \frac{E \cdot S}{K_M}$$

Total amount of enzyme is constant:

$$E_T = E + ES \quad \Rightarrow \quad ES = E_T \frac{S}{S + K_M}$$

turnover:  $V = V_{max} \frac{S}{S + K_M}$

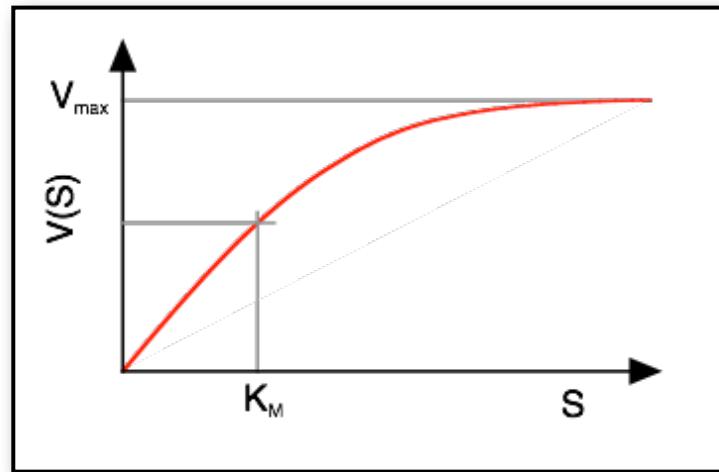
# The MM-equation

Effective turnover according to MM:

$$V = V_{max} \frac{S}{S + K_M}$$

$$V_{max} = k_{off} E_T$$

$$K_M = \frac{k_{off}}{k_{on}}$$



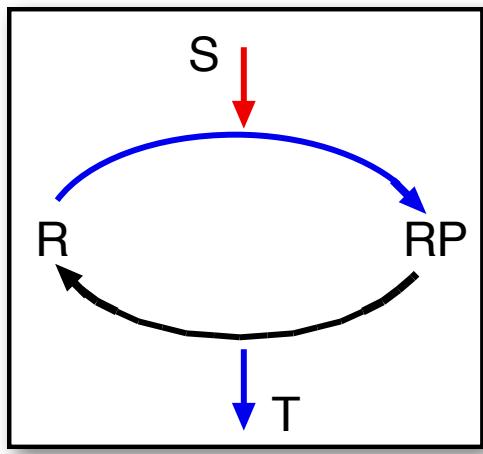
- Pro:
- analytical formula for turnover
  - curve can be easily interpreted:  $V_{max}$ ,  $K_M$
  - enzyme concentration can be ignored

Cons:

less kinetic information

$$k_{on}, k_{off}, E_T \Rightarrow V_{max}, K_M$$

# Sigmoidal Characteristics with MM kinetics



Same topology as before with Michaelis-Menten kinetics for phosphorylation and dephosphorylation.

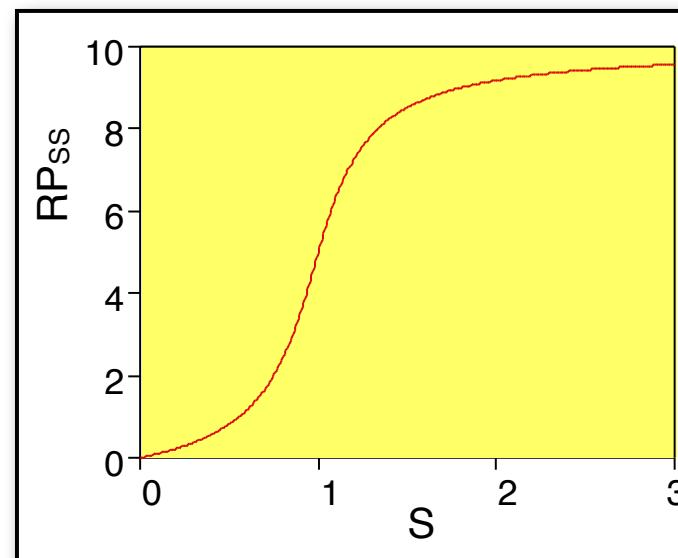
$$\frac{dRP}{dt} = \frac{k_1 S (R_t - RP)}{R_0 + (R_t - RP)} - \frac{k_2 RP}{RP_0 + RP} \stackrel{!}{=} 0$$

$$V = V_{max} \frac{S}{S + K_M}$$
 this means that  $S = R_t - RP$   
 $K_M = R_0$

Quadratic equation for RP

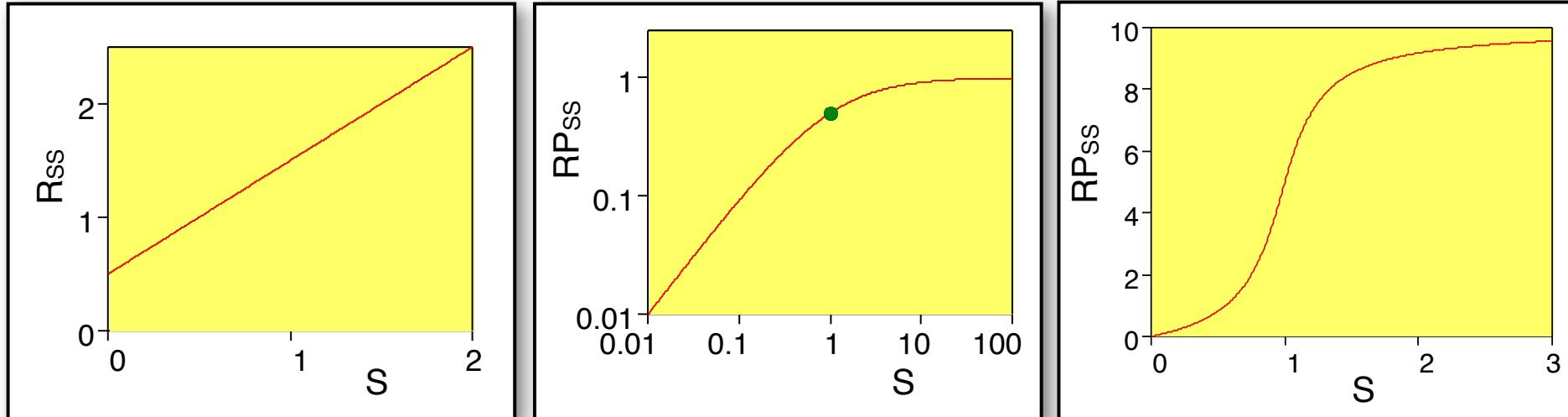
$$k_2 RP(R_0 + (R_t - R_p)) = k_1 S(R_t - RP)(RP_0 + RP)$$

=> sigmoidal characteristics  
(threshold behavior)  
often found in signalling cascades



$$R_t = 10, R_0 = RP_0 = 1, k_1 = k_2 = 1$$

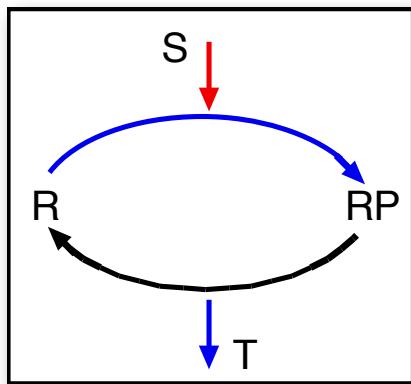
# Graded Response



Linear, hyperbolic, and sigmoidal characteristic give the same steady state response independent of the previous history  
=> no hysteresis

BUT: In fast time-dependent scenarios,  
delay may lead to a modified response

# Time-dependent Sigmoidal Response



Direct implementation:

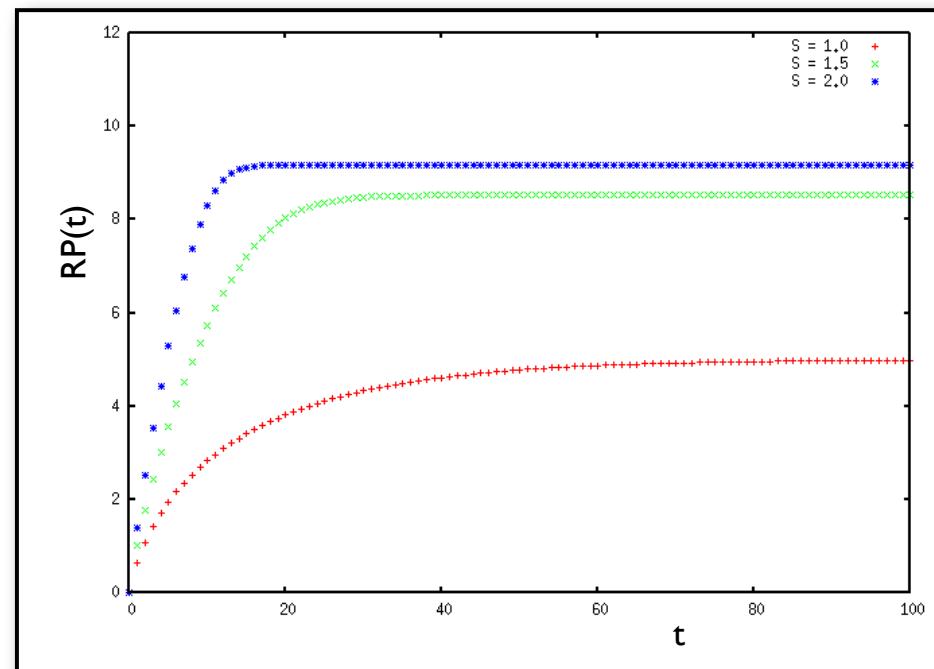
$$v_1 = \frac{Sk_1R}{R_0 + R} \quad v_2 = \frac{k_2RP}{RP_0 + RP}$$

Parameters:  $k1 = 1 \text{ (mol s)}^{-1}$ ,  $k2 = 1 \text{ s}^{-1}$ ,  $R_0 = RP_0 = 1 \text{ mol}$

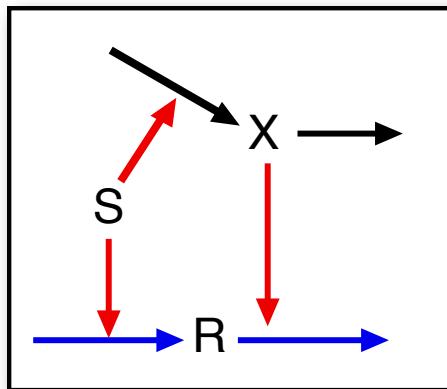
Initial conditions:  $R = 10 \text{ mol}$ ,  $RP = 0$

Time courses for  
 $S = 1, 1.5, \text{ and } 2$ ,  
 $RP(0) = 0$ :

equilibrium is reached  
faster for  
stronger signal



# Adaption - „sniffer“



Linear response modulated by a second species X

$$\frac{dX}{dt} = k_3 S - k_4 X$$

$$\frac{dR}{dt} = k_1 S - k_2 X R$$

Steady state:  $R_{ss}$  independent of S

$$X_{ss} = \frac{k_3}{k_4} S$$

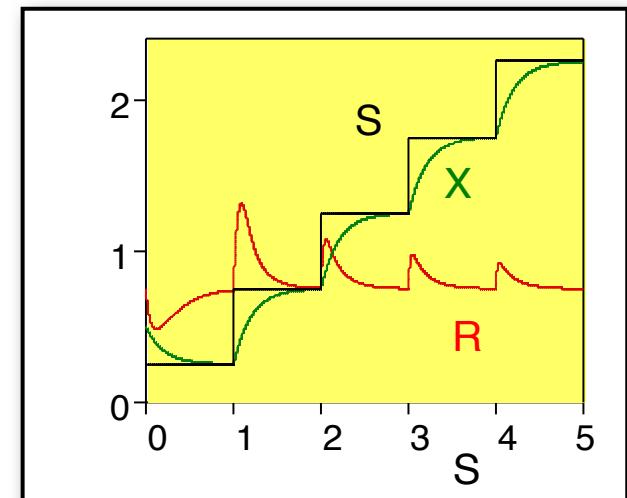
$$R_{ss} = \frac{k_1 k_4}{k_2 k_3}$$

R changes transiently when S changes, then goes back to its basal level.

found in smell, vision, chemotaxis, ...

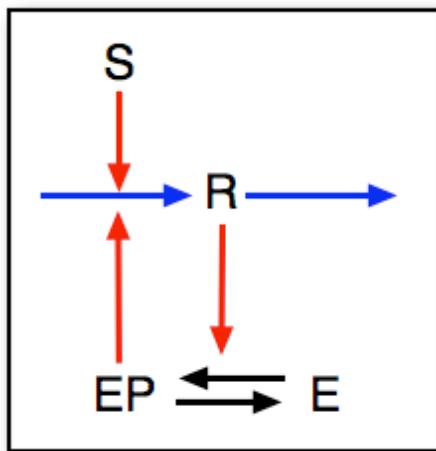
Note: response strength  $\Delta R$  depends on rate of change of S.

=> non-monotonous relation for  $R(S)$



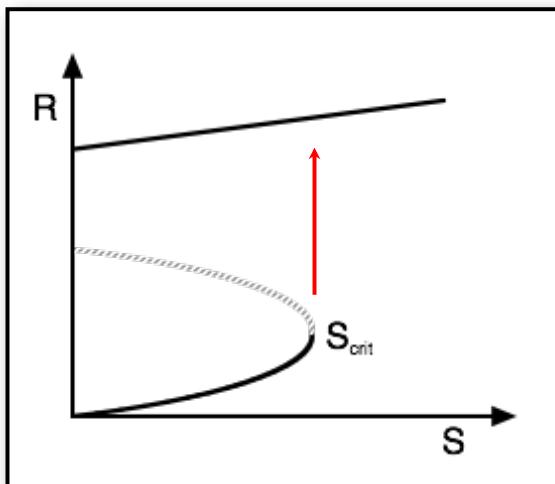
$$k_1 = 30, k_2 = 40, k_3 = k_4 = 5$$

# Positive Feedback



$$\frac{dR}{dt} = k_4 EP(R) + k_1 S - k_2 R$$

$$\frac{dEP}{dt} = \frac{k_3 R E}{EP_0 + EP} - \frac{k_5 EP}{E_0 + E}$$

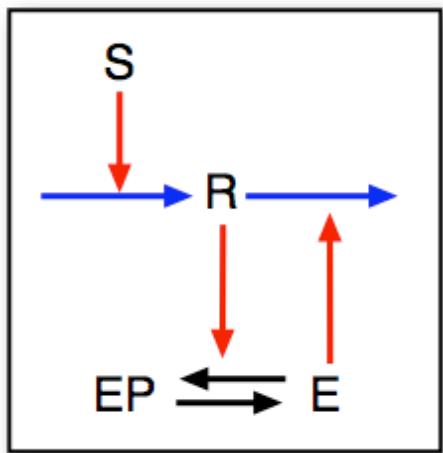


Feedback via R and EP  
=> high levels of R will stay

**"one-way switch"** via bifurcation

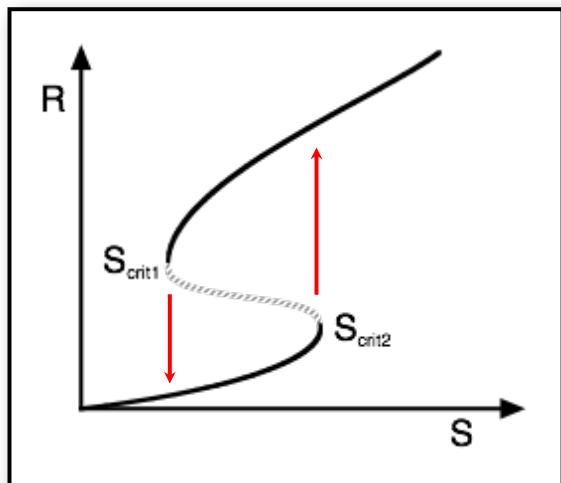
Found in processes that are "final":  
frog oocyte maturation, apoptosis, ...

# Mutual Inhibition - Toggle Switch



$$\frac{dR}{dt} = k_1 S - k_2 R - k_4 E(R)$$

$$\frac{dEP}{dt} = \frac{k_3 R E}{EP_0 + EP} - \frac{k_5 EP}{E_0 + E}$$

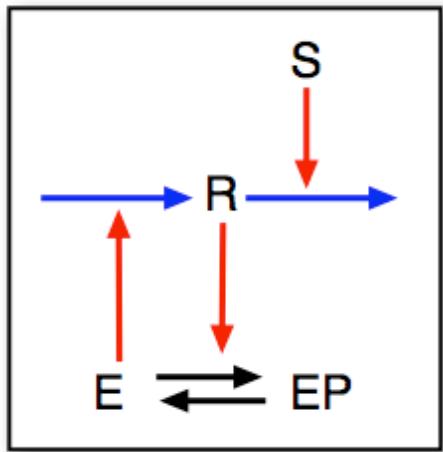


Sigmoidal "threshold" in  $E \leftrightarrow EP$  leads to bistable response (hysteresis):  
**toggle switch** (dt. *Kippschalter*)

Converts continuous external stimulus into two well defined stable states:

- lac operon in bacteria
- activation of M-phase promoting factor in frog eggs

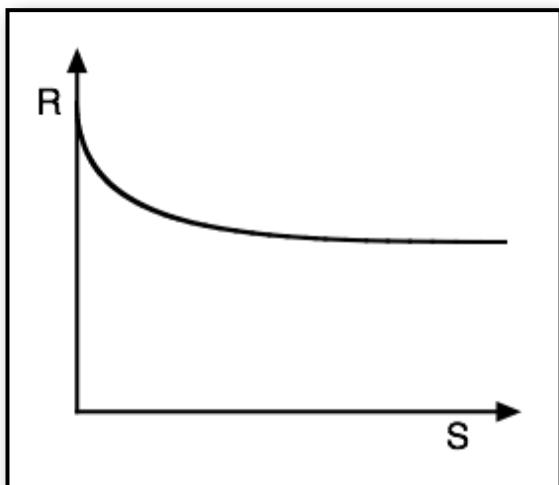
# Negative Feedback



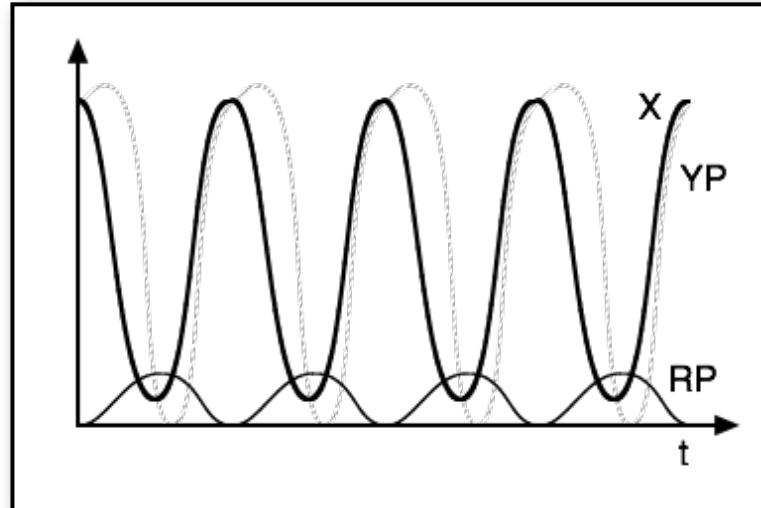
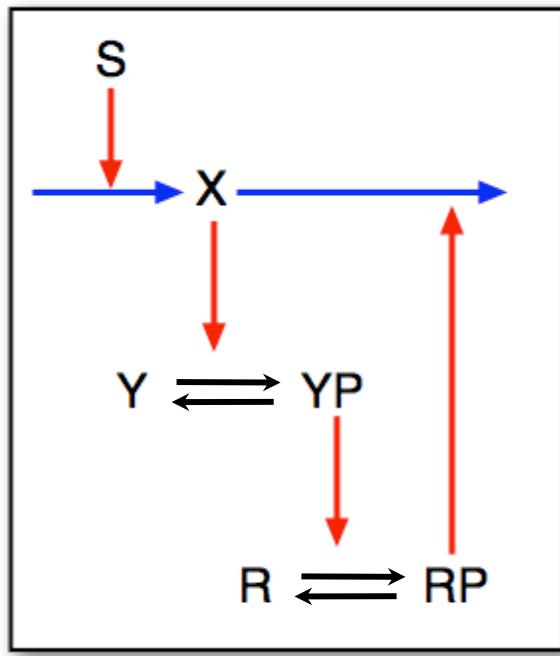
S controls the "demand" for R

=> **homeostasis**

found in biochemical pathways,  
no transient changes in R for steps in S  
(cf. "sniffer")



# Negative Feedback with Delay



Cyclic activation  $X \Rightarrow YP \Rightarrow RP \Rightarrow X$   
=> **Oscillations** (in a range of  $S$ )

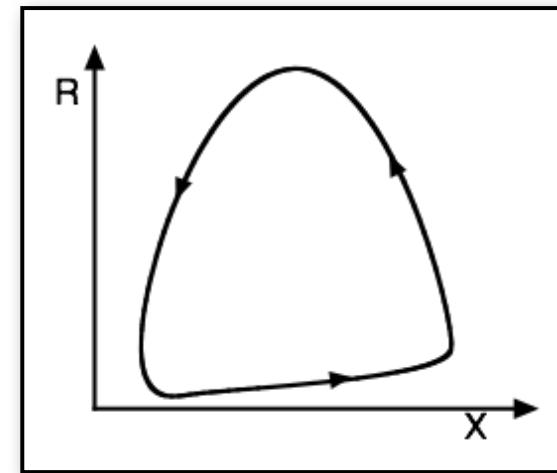
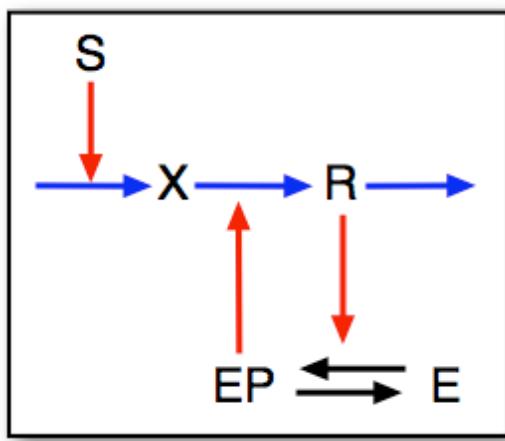
$$\frac{dX}{dt} = k_0 + k_1 S - k_2 X - k_7 R P X$$

$$\frac{dYP}{dt} = \frac{k_3 X Y}{Y_0 + Y} - \frac{k_4 Y P}{Y P_0 + Y P}$$

$$\frac{dR P}{dt} = \frac{k_5 Y P R}{R_0 + R} - \frac{k_6 R P}{R P_0 + R P}$$

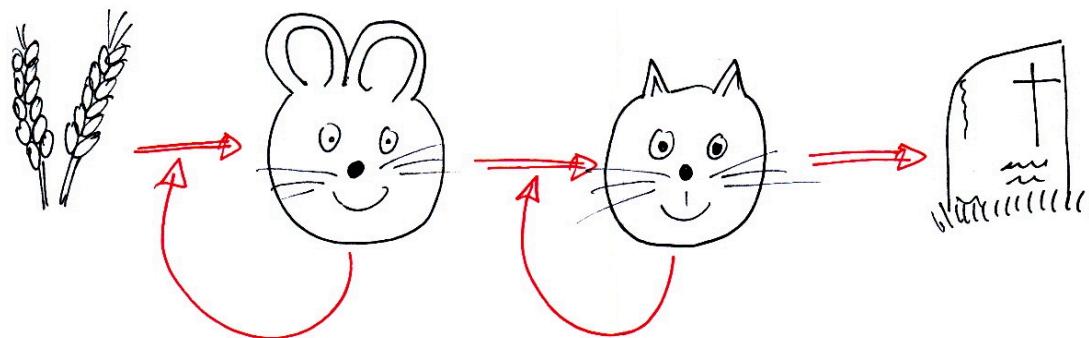
Proposed mechanism  
for **circadian clocks**

# Substrate-Depletion Oscillations

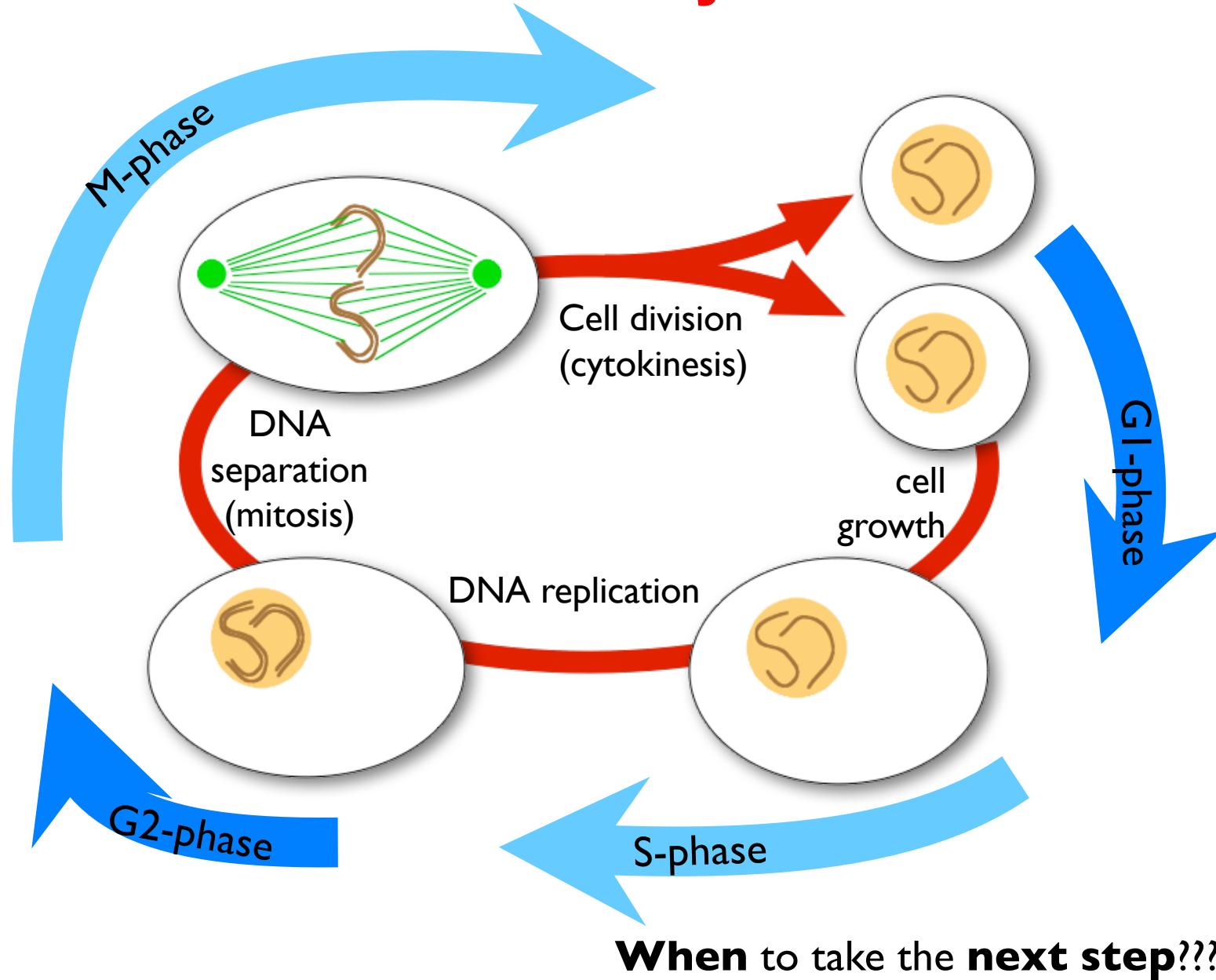


$R$  is produced in an **autocatalytic** reaction from  $X$ , finally **depleting**  $X$ ...

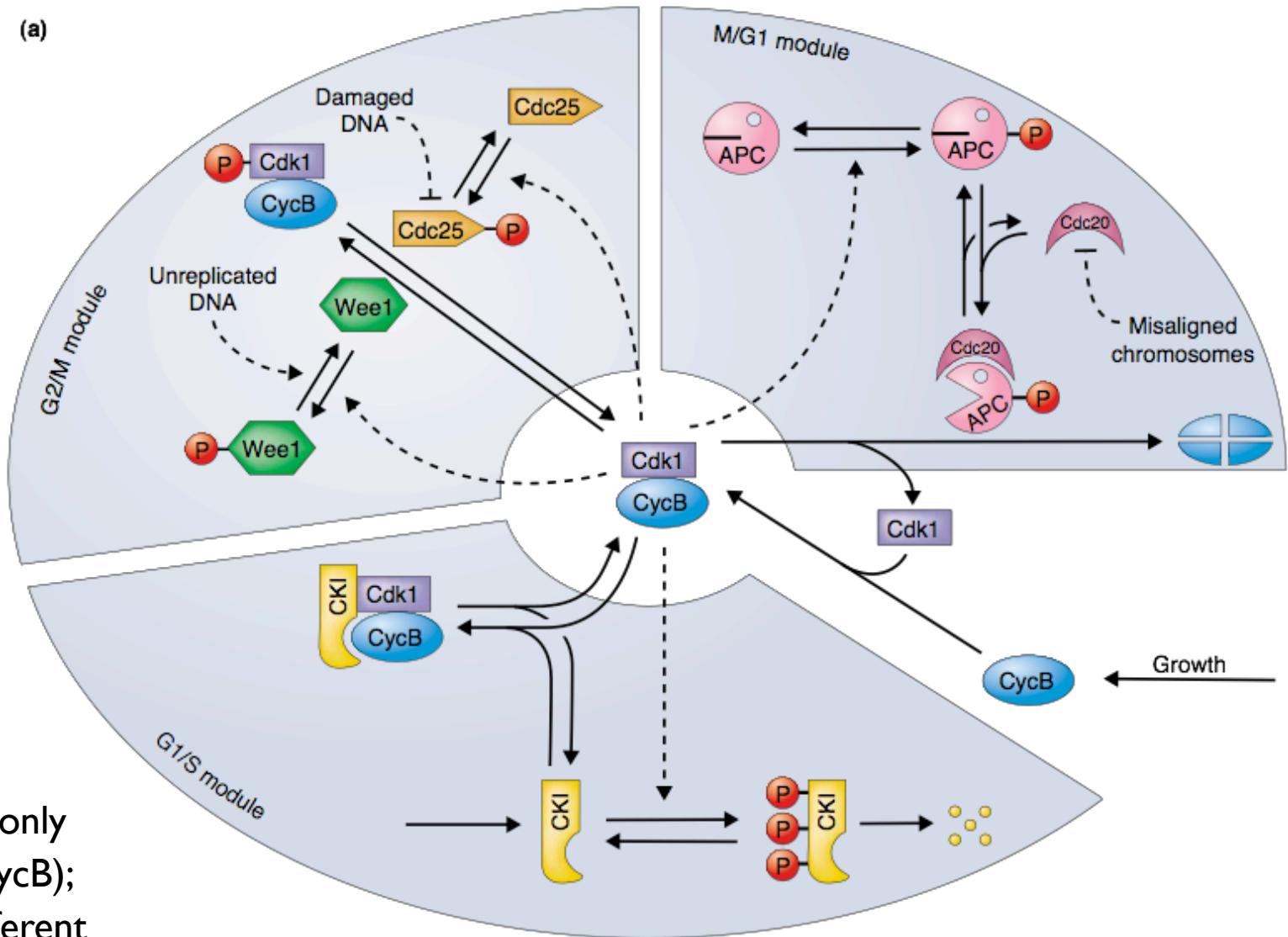
Similar to Lotka-Volterra system (autocatalysis for  $X$ , too):



# The Cell Cycle



# Simplified Version of Cell Cycle Control System



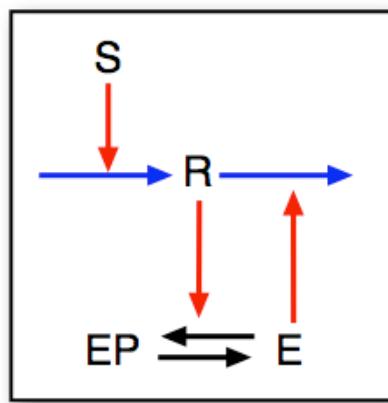
cdc =  
"cell division cycle"

Cdk1: cyclin  
dependent kinase I

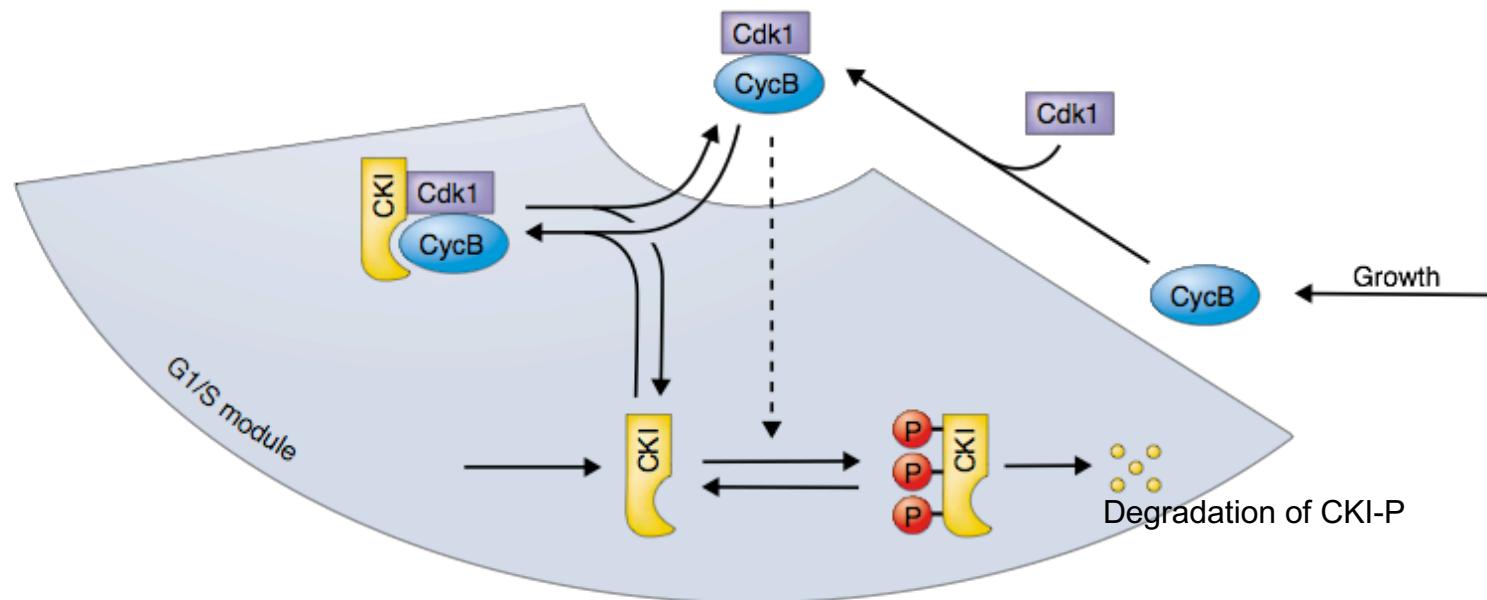
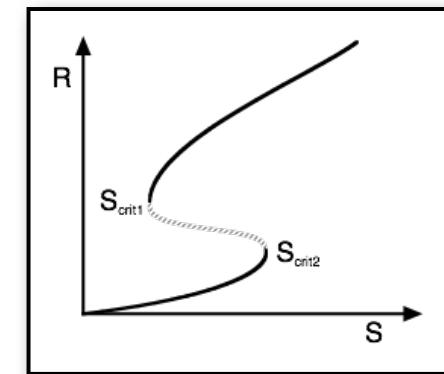
Simplification: assume only  
one type of cyclins (CycB);  
in reality there are different  
ones

Tyson et al, *Curr. Op. Cell Biol.* **15** (2003) 221

# G1 $\Rightarrow$ S — Toggle Switch

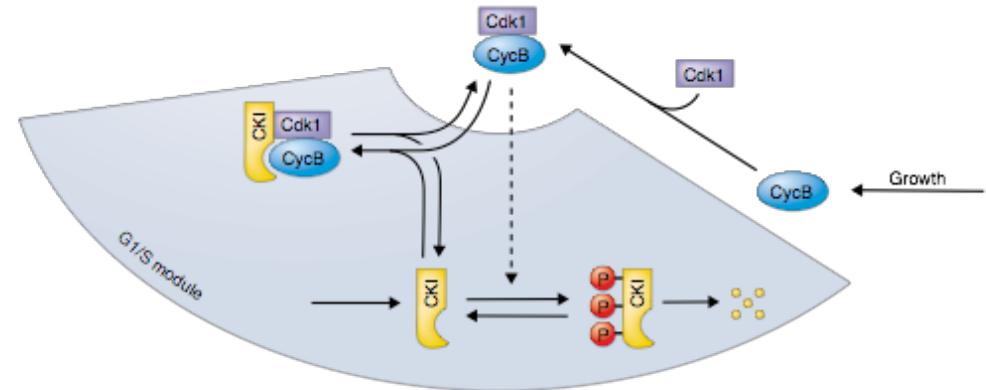
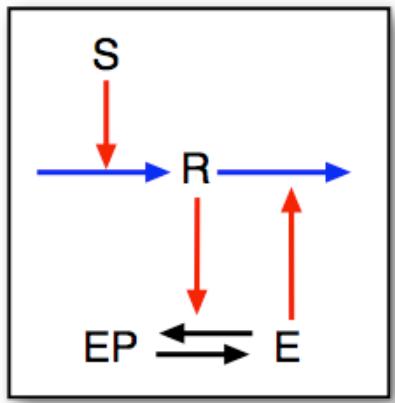


Mutual inhibition between  
Cdk1-CycB and CKI  
(cyclin kinase inhibitor)

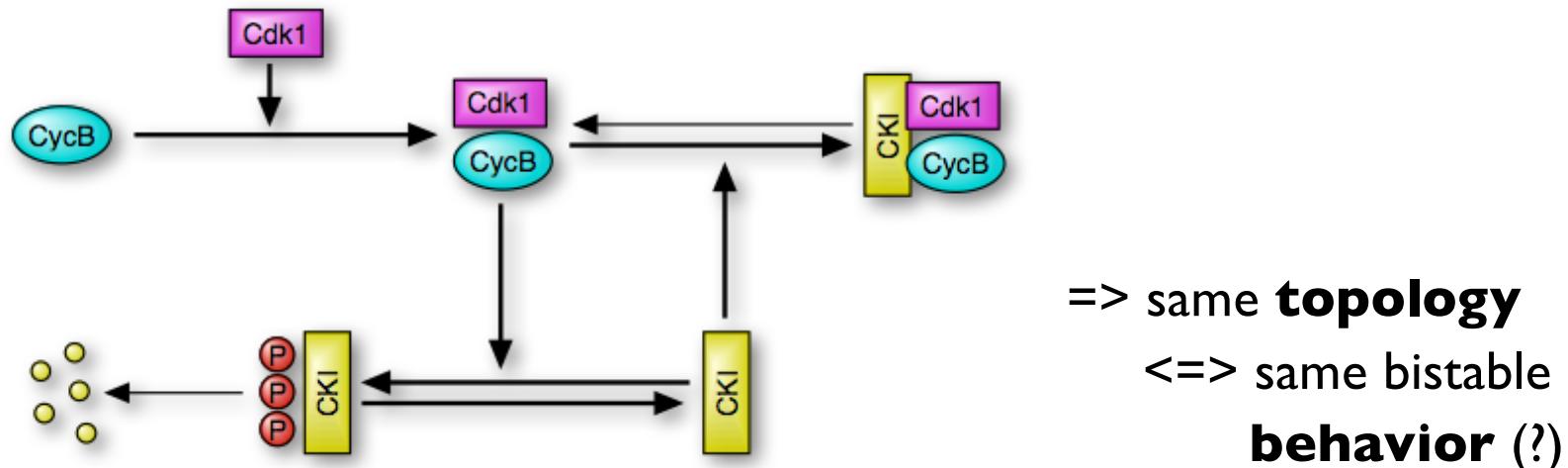


Tyson et al, *Curr. Op. Cell Biol.* 15 (2003) 221

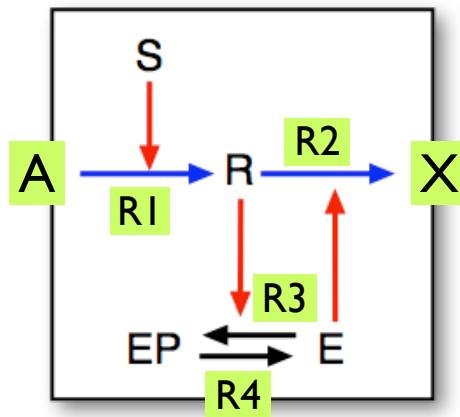
# Mutual Inhibition



Assume: CycB:Cdk1:CKI is stable  $\Leftrightarrow$  dissociation is very slow



# Rate Equations: Toggle Switch



Stoichiometric  
matrix  
"(C)" = catalyst

	R1	R2	R3	R4
A	-1			
S	(C)			
R	1	-1	(C)	
E		(C)	-1	1
EP			1	-1
X		1		

$$\frac{dR1}{dt} = k_1 A S$$

$$\frac{dR2}{dt} = k_2 R E$$

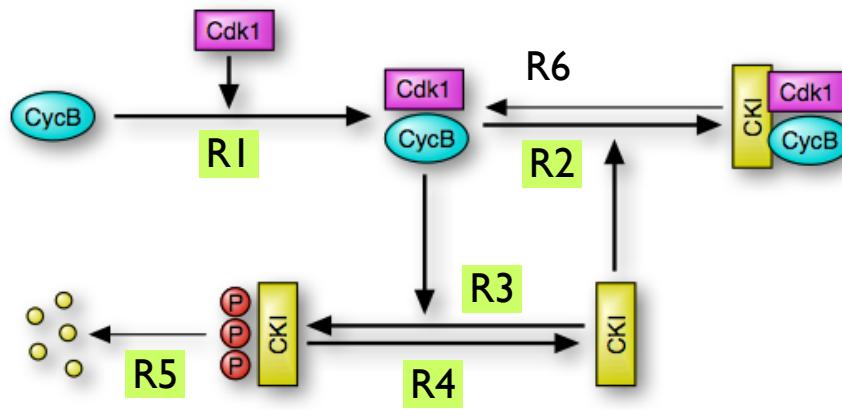
$$\frac{dR3}{dt} = \frac{k_3 R E}{E_0 + E}$$

$$\frac{dR4}{dt} = \frac{V_4 EP}{EP_0 + EP}$$

$$\frac{dR}{dt} = \frac{dR1}{dt} - \frac{dR2}{dt} = k_1 A S - k_2 R E$$

$$\frac{dE}{dt} = \frac{dR4}{dt} - \frac{dR3}{dt}$$

# Rate Equations: G1/S Module



$$\frac{dR1}{dt} = k_1 [\text{CycB}] [\text{Cdk1}]$$

$$\frac{dR2}{dt} = k_2 [\text{CycB:Cdk1}] [\text{CKI}]$$

$$\frac{dR3}{dt} = \frac{k_3 [\text{CycB:Cdk1}] [\text{CKI}]}{K_3 + [\text{CKI}]}$$

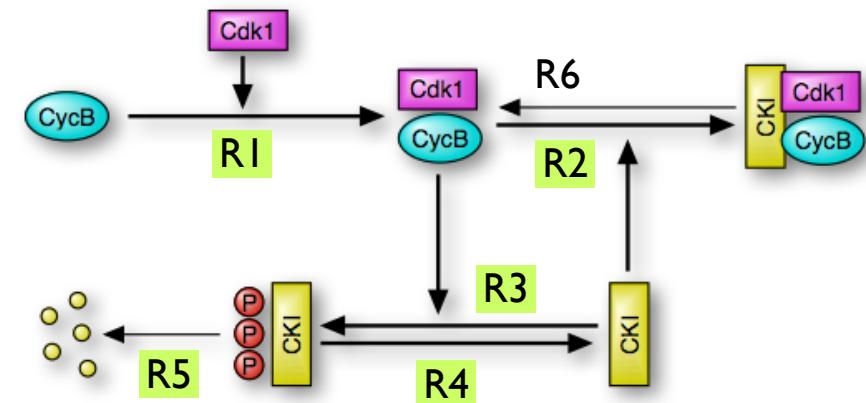
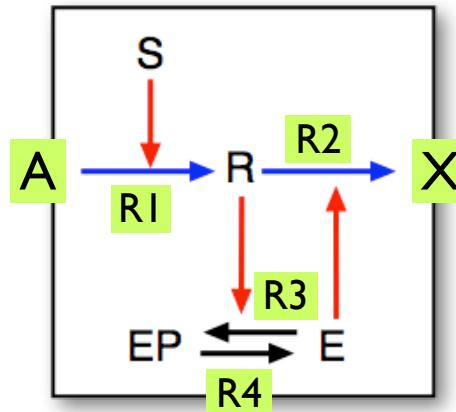
$$\frac{dR4}{dt} = \frac{V_4 [\text{CKI:P}_3]}{K_4 + [\text{CKI:P}_3]}$$

	R1	R2	R3	R4	R5	R6
CycB	-I					
Cdk1	-I					
CycB:Cdk1	I	-I	(C)			I
CKI		-I	-I	I		I
CKI:P <sub>3</sub>			I	-I		
CKI:P <sub>3</sub>					-I	
CycB:Cdk1:CKI		I				-I

$$\frac{d[\text{CycB:Cdk1}]}{dt} = \frac{dR1}{dt} - \frac{dR2}{dt} + \frac{dR6}{dt}$$

$$\frac{d[\text{CKI}]}{dt} = \frac{dR4}{dt} - \frac{dR3}{dt} - \frac{dR2}{dt} + \frac{dR6}{dt}$$

# Comparison: Matrices

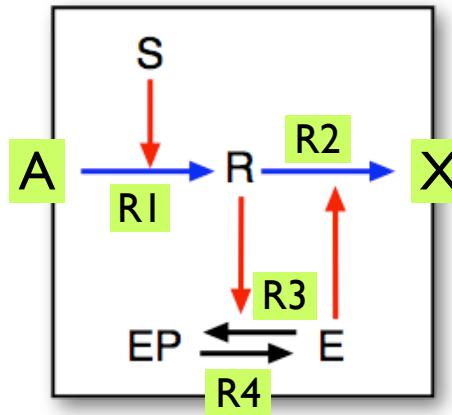


	R1	R2	R3	R4
A	-I			
S	(C)			
R	I	-I	(C)	
E		(C)	-I	I
EP			I	-I
X		I		

	R1	R2	R3	R4	R5	R6
CycB	-I					
Cdk1	-I					
CycB:Cdk1	I	-I	(C)			I
CKI		-I	-I	I		I
CKI:P <sub>3</sub>			I	-I		
CKI:P <sub>3</sub>					-I	
CycB:Cdk1:CKI		I				-I

Difference: catalysts vs. substrates

# Comparison: Equations



$$\frac{dR1}{dt} = k_1 A S$$

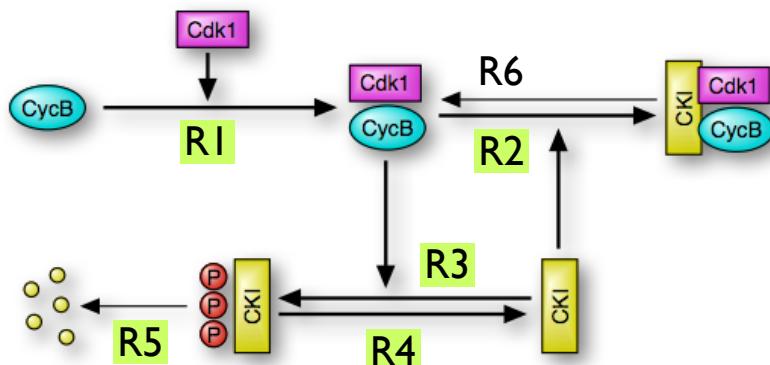
$$\frac{dR2}{dt} = k_2 R E$$

$$\frac{dR3}{dt} = \frac{k_3 R E}{E_0 + E}$$

$$\frac{dR4}{dt} = \frac{V_4 EP}{EP_0 + EP}$$

$$\frac{dR}{dt} = \frac{dR1}{dt} - \frac{dR2}{dt} = k_1 A S - k_2 R E$$

$$\frac{dE}{dt} = \frac{dR4}{dt} - \frac{dR3}{dt} = \frac{k_3 R E}{E_0 + E} - \frac{V_4 EP}{EP_0 + EP}$$



$$\frac{dR1}{dt} = k_1 [\text{CycB}] [\text{Cdk1}]$$

$$\frac{dR2}{dt} = k_2 [\text{CycB:Cdk1}] [\text{CKI}]$$

$$\frac{dR3}{dt} = \frac{k_3 [\text{CycB:Cdk1}] [\text{CKI}]}{K_3 + [\text{CKI}]}$$

$$\frac{dR4}{dt} = \frac{V_4 [\text{CKI:P}_3]}{K_4 + [\text{CKI:P}_3]}$$

$$\frac{d[\text{CycB:Cdk1}]}{dt} = \frac{dR1}{dt} - \frac{dR2}{dt} + \frac{dR6}{dt}$$

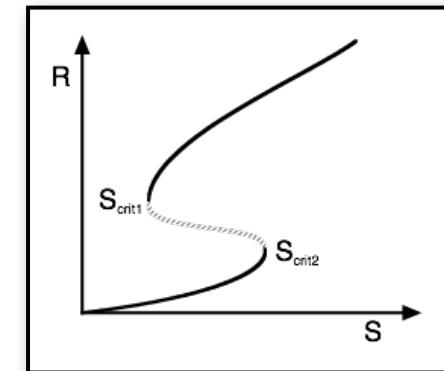
$$\frac{d[\text{CKI}]}{dt} = \frac{dR4}{dt} - \frac{dR3}{dt} - \frac{dR2}{dt} + \frac{dR6}{dt}$$

Rename species => same rate equations => same behavior

# Predicted Behavior: G1 $\Rightarrow$ S

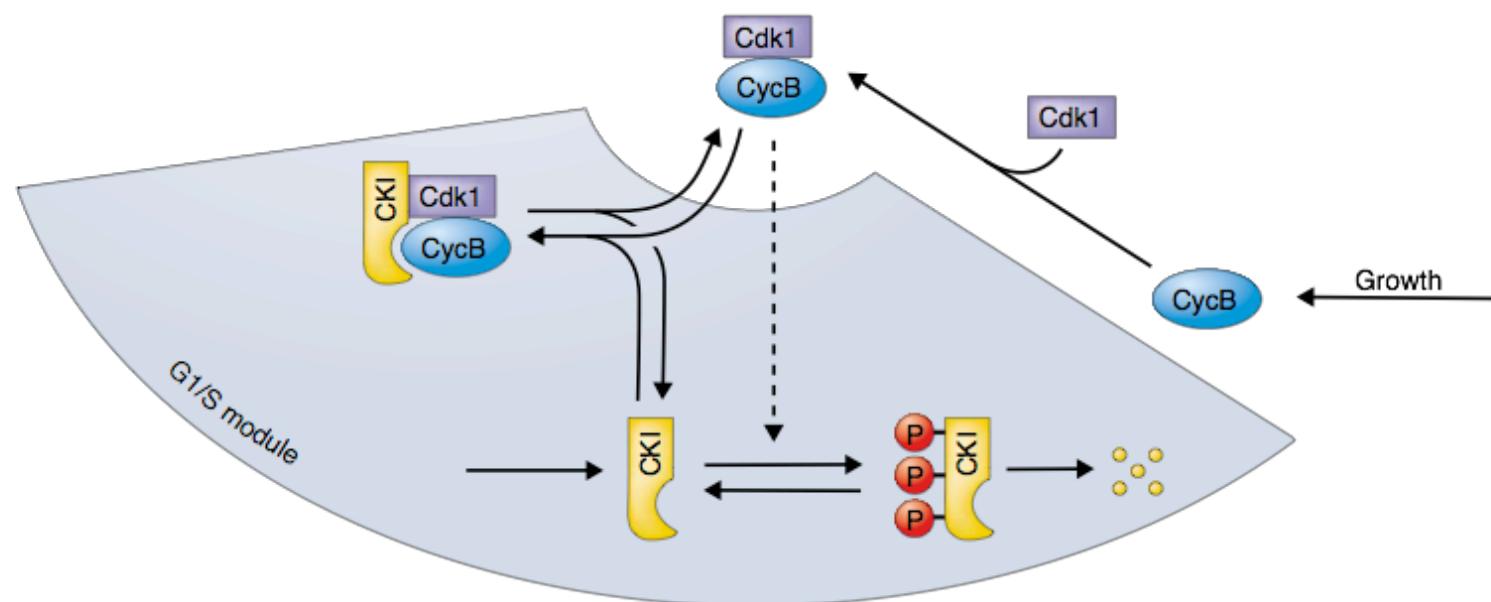
Signal: cell growth = concentration of CycB, Cdk1

Response: activity (concentration) of CycB:Cdk1



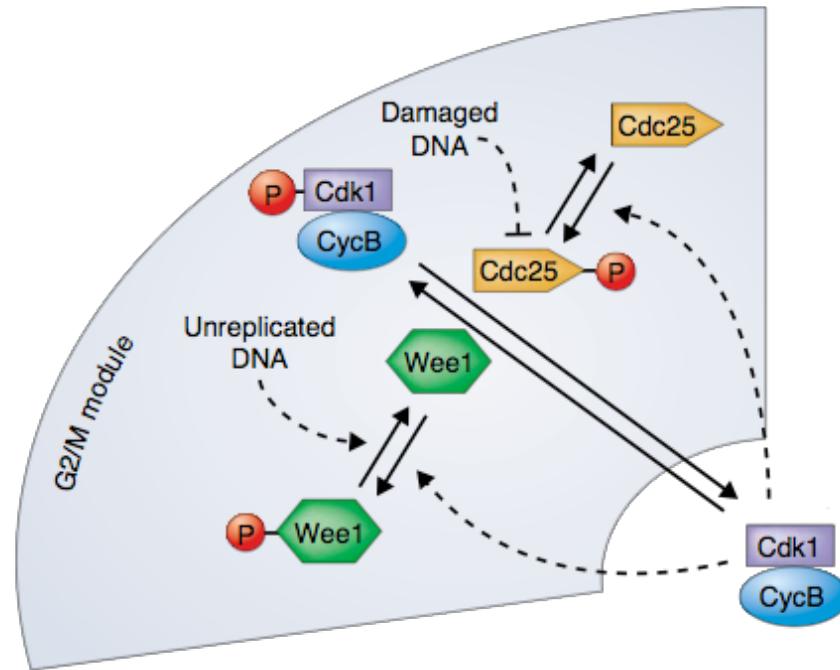
Toggle switch:

=> above critical cell size, CycB:Cdk1 activity will switch on



Tyson et al, *Curr. Op. Cell Biol.* 15 (2003) 221

# G2 => M



## Dual toggle switch:

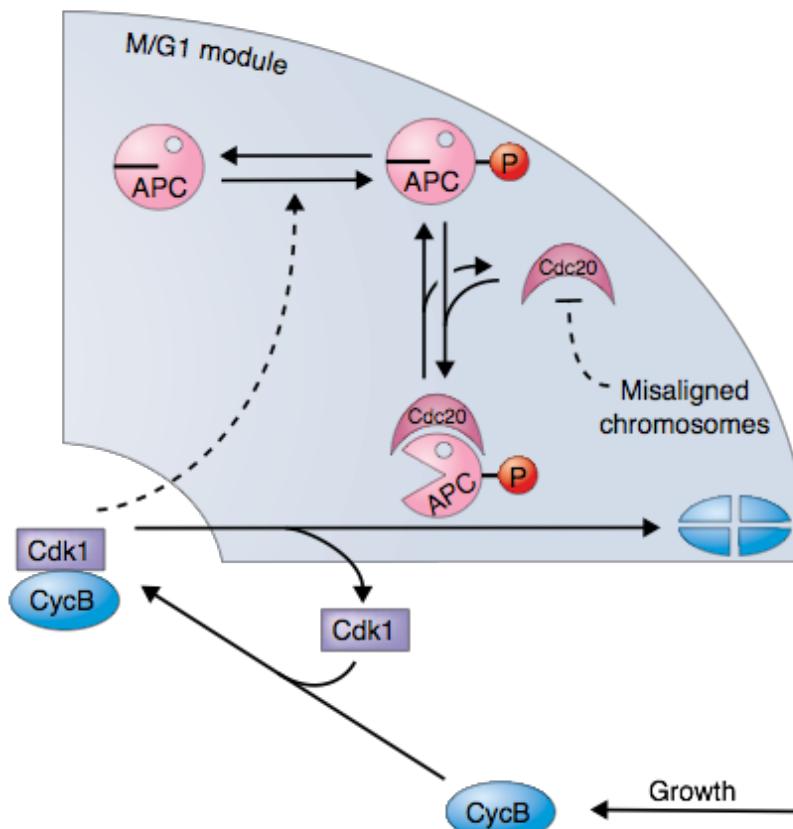
- **mutual activation** between CycB:Cdk1 and Cdc25 (phosphatase that activates the dimer)
- **mutual inhibition** between CycB:Cdk1 and Wee1 (kinase that inactivates the dimer)

=> when the cell **grows** further during the second gap phase G2, the activity of CycB:Cdk1 will **increase** by a further **step**

# M => G1

## Negative feedback loop oscillator

- i) CycB:Cdk1 activates anaphase promoting complex (APC)
- ii) APC-P activates Cdc20
- iii) Cdc20:APC-P degrades CycB



## Behavior:

at a critical cell size

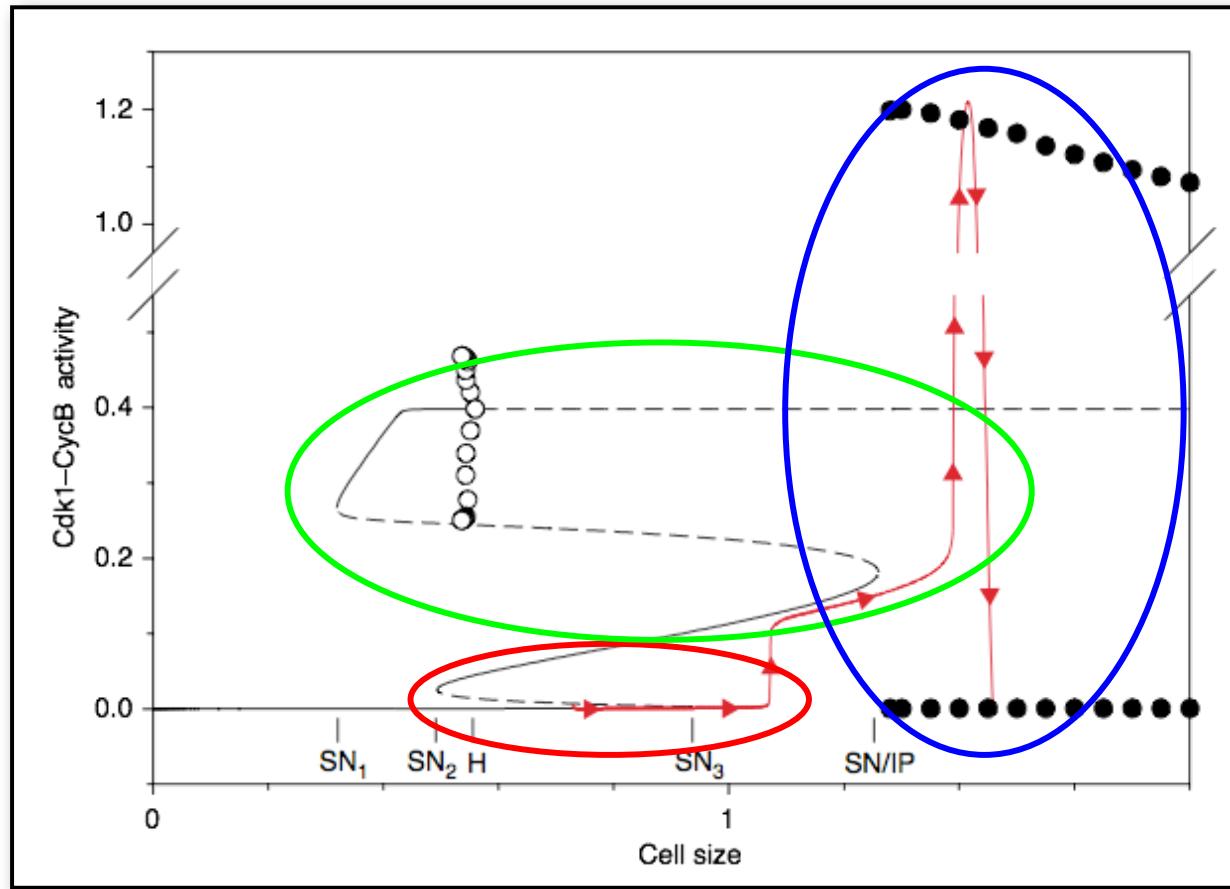
CycB:Cdk1 activity increases and **decreases** again

=> at low CycB:Cdk1 level, the G1/S toggle switches off again,

=> cell cycle completed

Tyson et al, *Curr. Op. Cell Biol.* **15** (2003) 221

# Overall Behavior



G1/S toggle => bistability

M/G1 oscillator

G2/M toggle => bistability

Tyson et al, *Curr. Op. Cell Biol.* **15** (2003) 221

# Circadian clocks in mammals and plants

Most organisms (animals, plants, fungi and cyanobacteria) enhance their fitness by coordinating their development with daily environmental changes through molecular timekeepers (circadian clocks)

**Mammals** display circadian rhythms in behavioural and physiological processes, such as

- sleep
- feeding
- blood pressure and
- metabolism

Roles in **plants** e.g.:

- opening of flowers in the morning and their closure at night

Circadian rhythms are guided by **external light–dark signals** that are integrated through intrinsic central and peripheral molecular clocks

McClung Plant Cell 18, 792 (2006)

# Circadian rhythms

- (1) Circadian rhythms are the subset of biological rhythms with period of 24 h.  
The term circadian combines the Latin words “circa” (about) and “dies” (day).
- (2) Circadian rhythms are **endogenously generated** and **self-sustaining**.

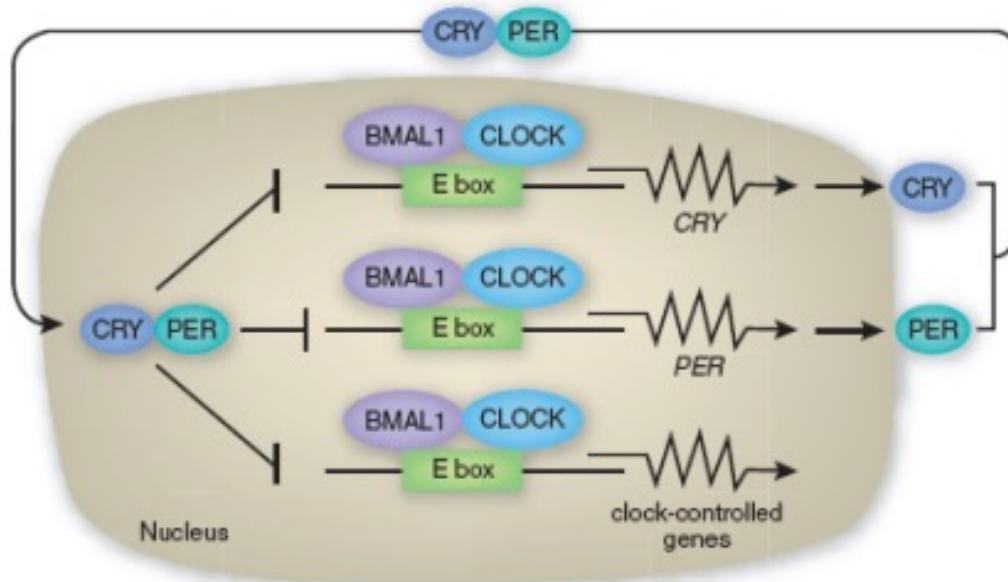
They persist under constant environmental conditions, typically constant light (or dark) and constant temperature.

Under these controlled conditions, the free-running period of **24 h** is observed.

- (3) For all circadian rhythms the **period** remains relatively **constant** over a range of ambient temperatures.

This is thought to be one property of a general mechanism that buffers the clock against changes in cellular metabolism.

# Basic molecular elements of mammalian clocks



This is the **minimal scheme** for the mammalian clock.

It requires several interconnecting transcriptional, translational and post-translational loops to achieve gene expression with circadian periodicity

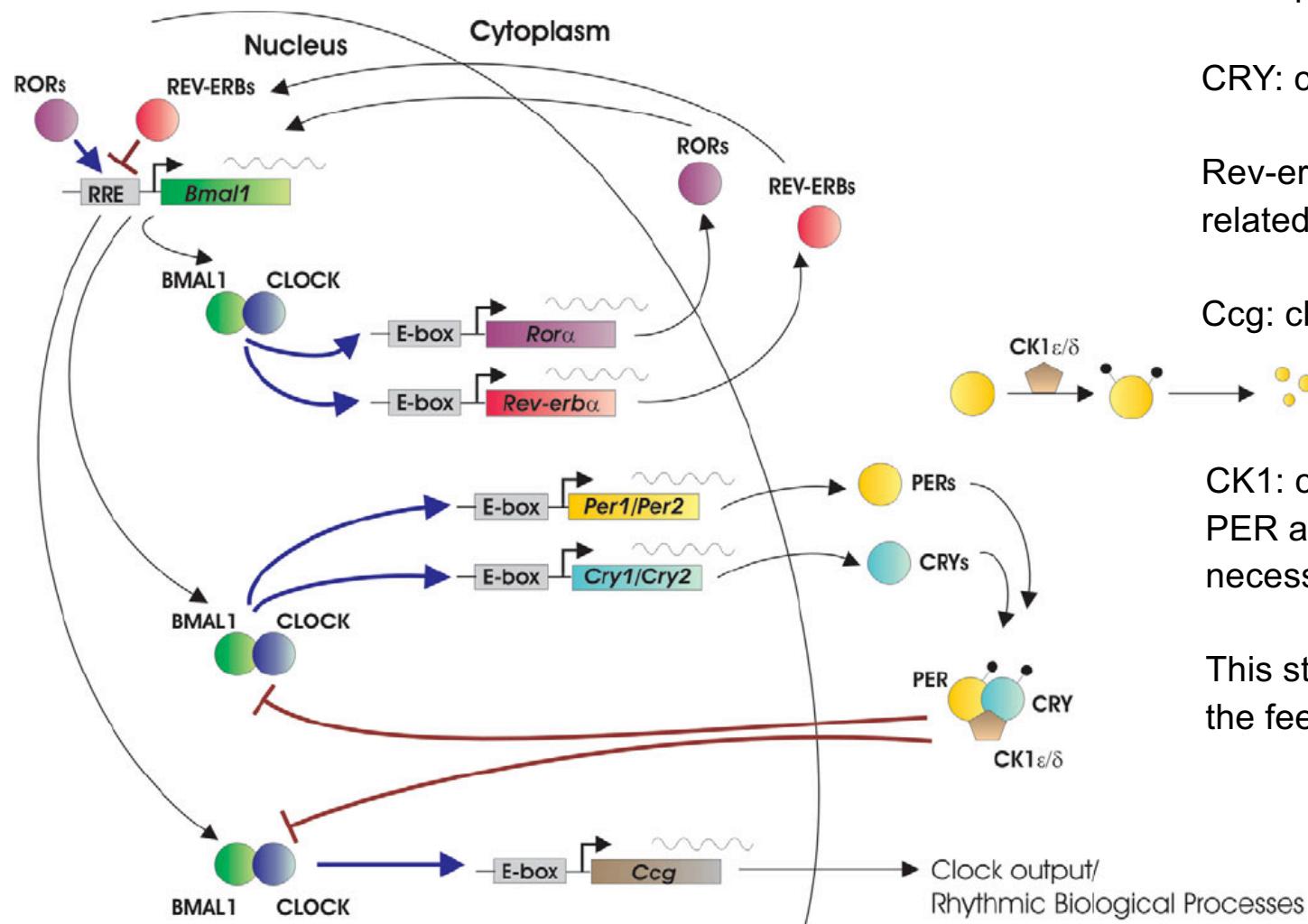
(a) 2 TFs **CLOCK** and **BMAL1** heterodimerize.

(b) BMA1:CLOCK binds to the **E-boxes** in the promoters of -the *PER* and *CRY* genes, - and of clock-controlled genes, and activate their transcription.

(c) The translated PER and CRY proteins dimerize in the cytosol, enter the nucleus and **inhibit** CLOCK-BMAL1– activated transcription.

Sancar,  
Nat. Struct. Mol. Biol. 15, 23 (2008)

# Circuit of circadian rhythms in mammals

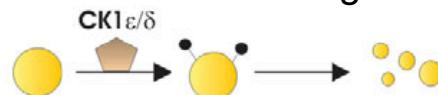


PER: period

CRY: cryptochrome

Rev-erb, ROR: retinoic acid-related orphan nuclear receptors

Ccg: clock-controlled genes



CK1: casein kinase; phosphorylates PER and CRY;  
necessary for their dimerization

This step serves to slow down the feed-back cycle.

Figure 1. A network of transcriptional–translational feedback loops constitutes the mammalian circadian clock.

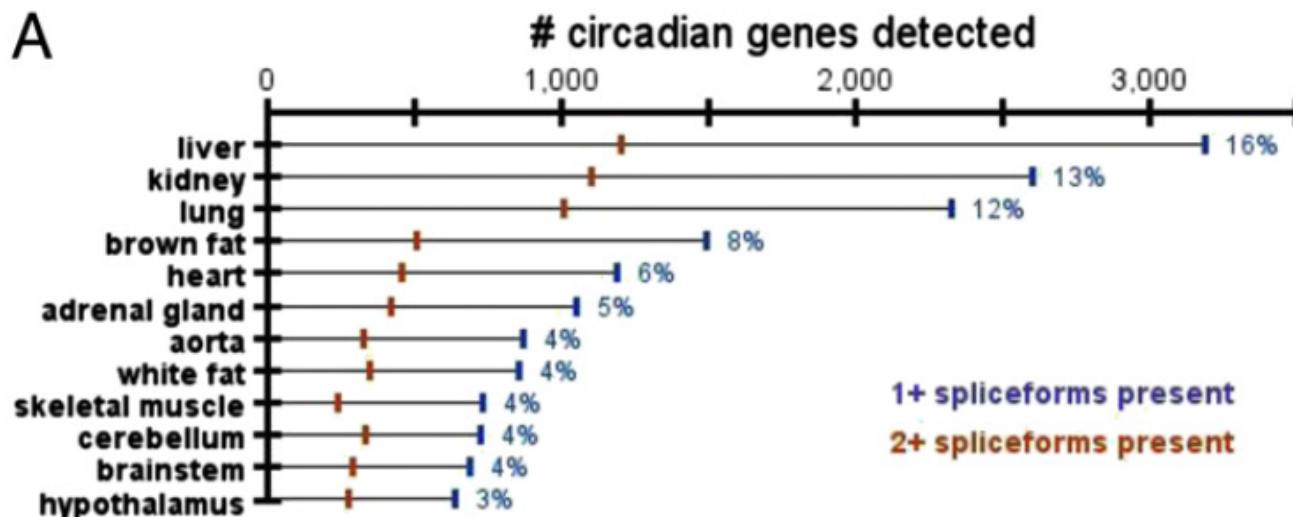
# Are circadian rhythms relevant for bioinformatics?

## A circadian gene expression atlas in mammals: Implications for biology and medicine

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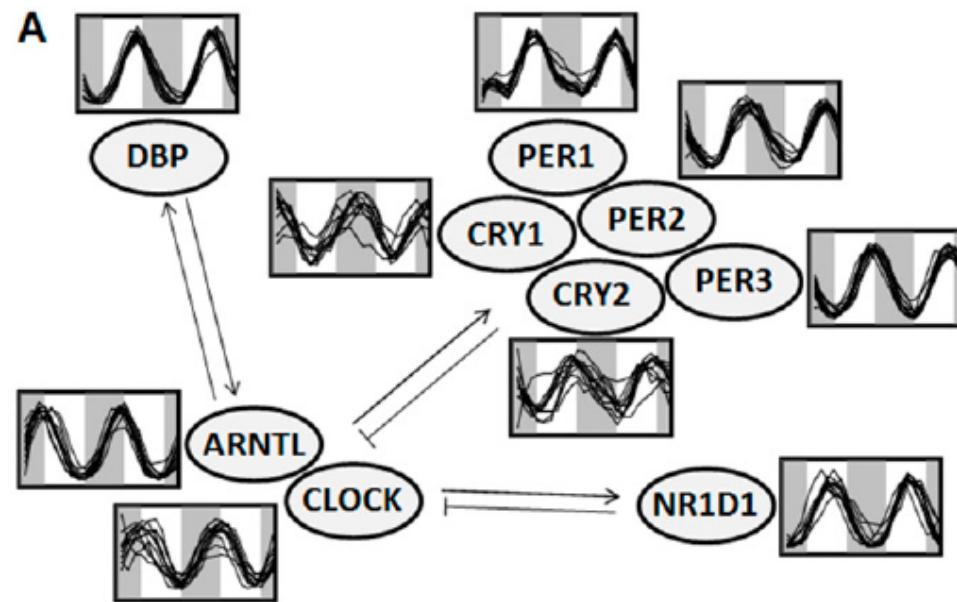
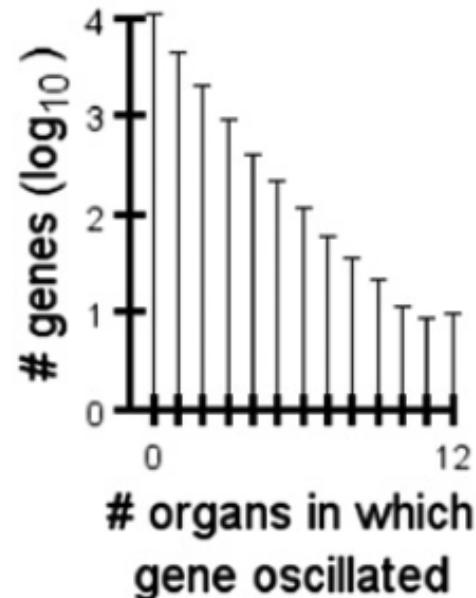
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- RNA-seq and DNA arrays to quantify transcriptomes of 12 mouse organs at 2 hour/6 hour intervals
- **Circadian genes:** defined as genes that oscillate with 24 hour-period (project on sine/cosine functions)



Liver contained most circadian genes (-> metabolism), Brain tissue the fewest („the brain never sleeps“)

# Globally oscillating genes in mouse tissue



Only 10 genes oscillated in all organs:

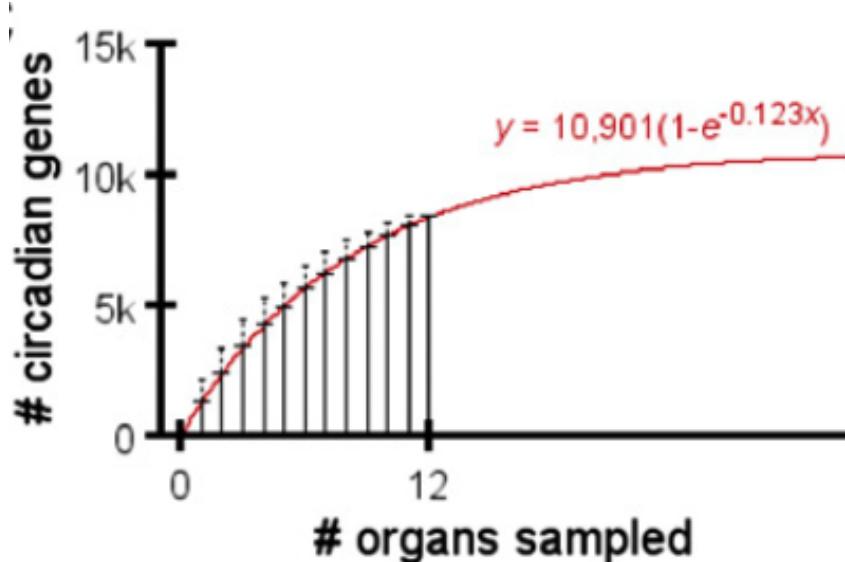
*Arntl*, *Dbp*, *Nr1d1*, *Nr1d2*, *Per1*, *Per2*, and *Per3* (core clock factors – **as expected**), and *Usp2*, *Tsc22d3*, and *Tspan4*.

*Usp2* - Ubiquitin carboxyl-terminal hydrolase 2

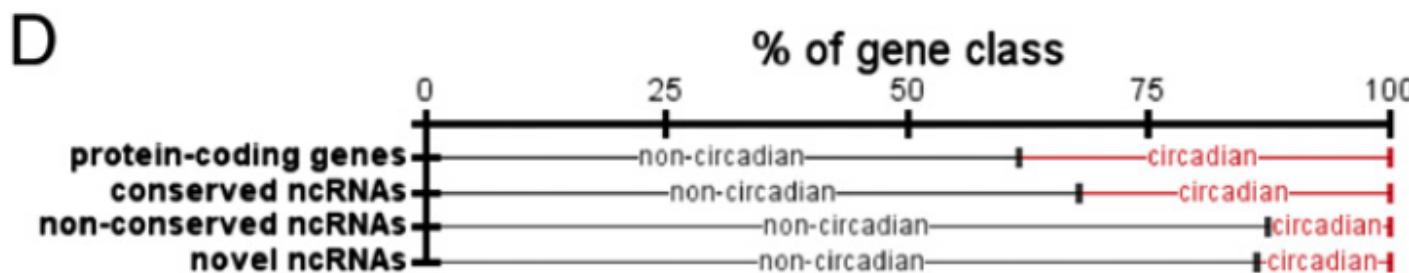
*Tsc22d3* - TSC22 domain family protein 3

*Tspan4* - The protein encoded by this gene is a member of the transmembrane 4 superfamily, also known as the tetraspanin family.

# Overlap of genes/organs (B), how many expected (C)?



Extrapolation shows that 55% of all genes are expected to show circadian expression in some organ.



Also non-coding RNAs show circadian expression (at lower frequencies).

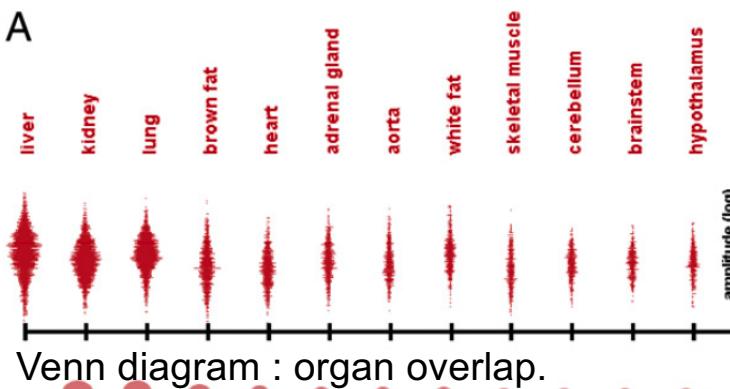
No individual ncRNA oscillated in more than five organs.

(ncRNA expression is known to be organ-specific).

Conserved ncRNAs means that they are conserved between human and mouse.

# (A) Phases + overlap, (B) similarity

A

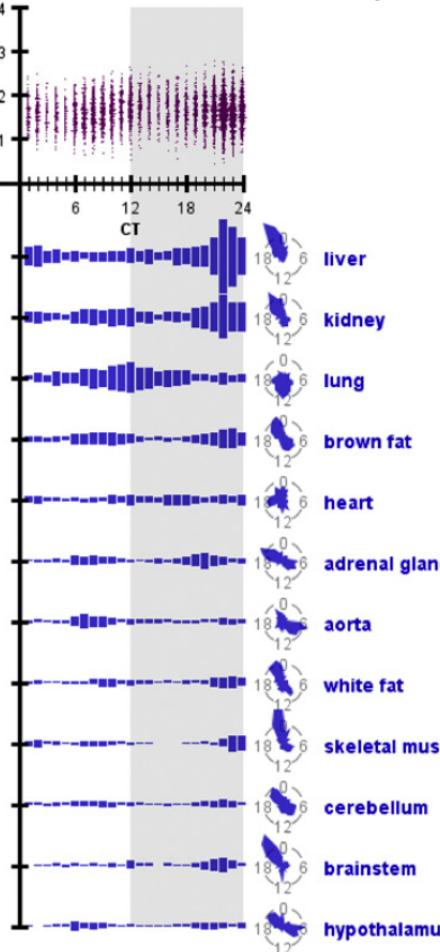


Venn diagram : organ overlap.

Time-dependent profiles.

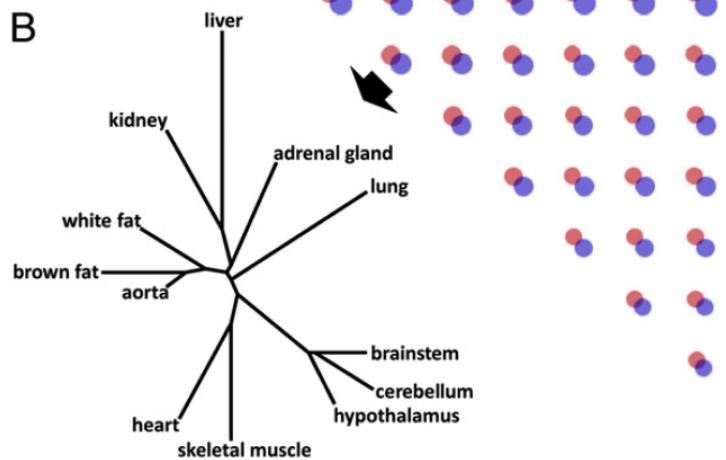
Top: all organs

Below: individual organs.



Most circadian genes show organ-specific expression (small overlap).

B

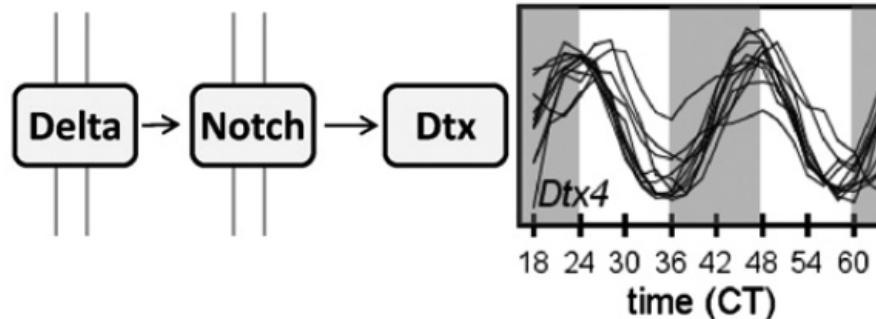


**Peaks often at dawn and dusk.**

Cluster tissues by similarity of peak phases

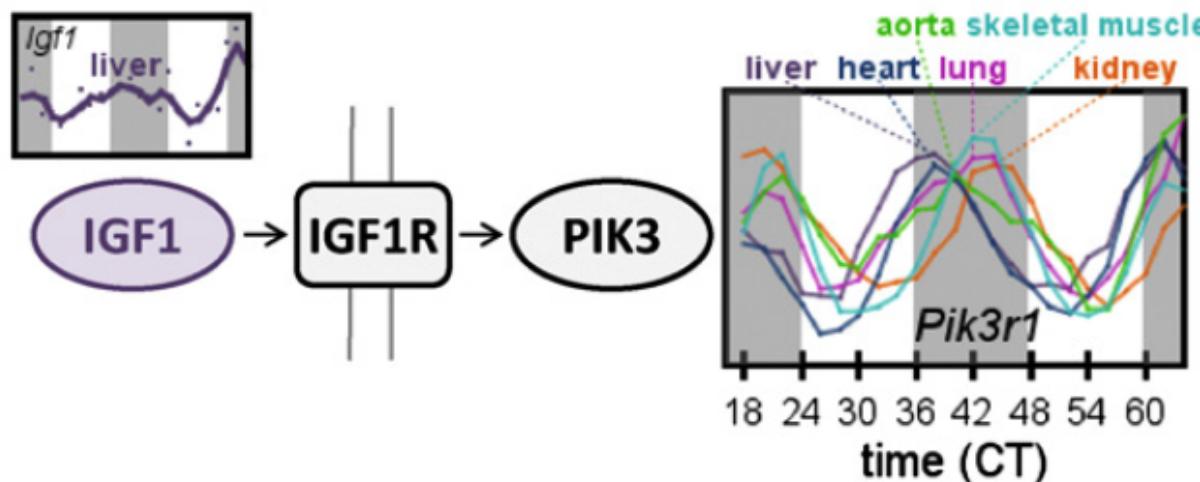
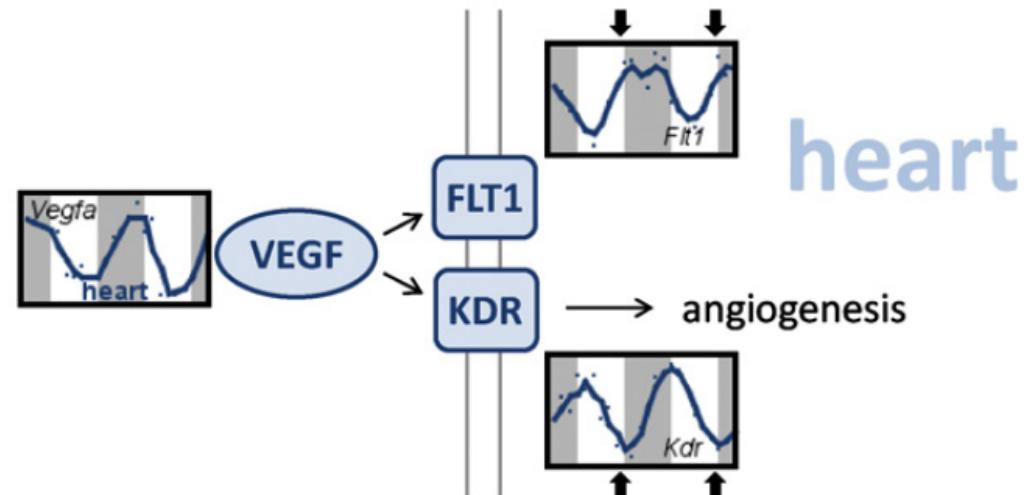
Tree in panel B shows that developmentally related organs tend to share circadian genes .

# Three Examples



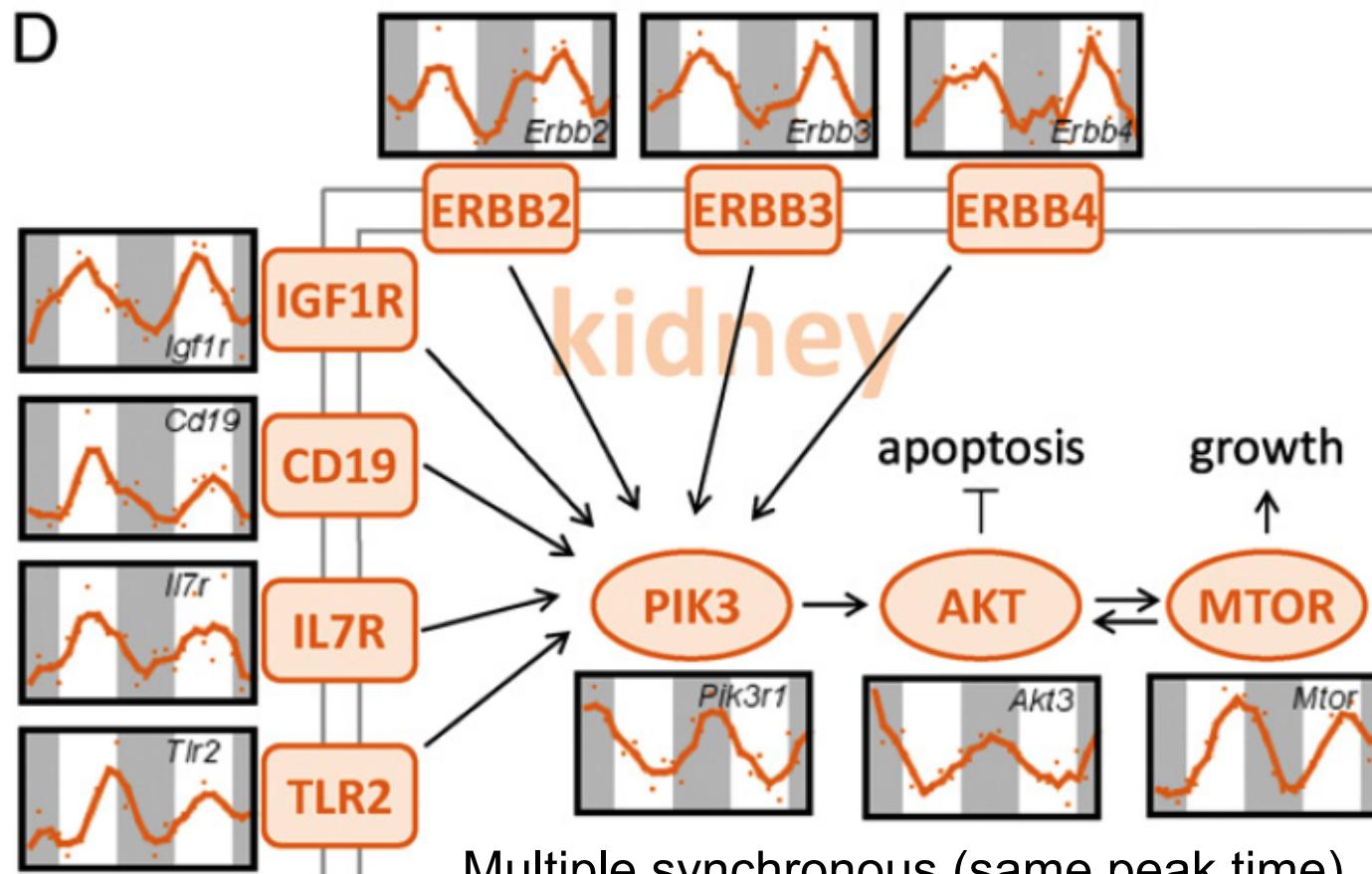
(1) *Dtx4*, a Notch pathway E3 ubiquitin ligase, oscillated in phase with *Arntl* in all organs

(2) Two VEGF-receptors FLT1 and KDR are expressed alternatively. Arrows: times of anti-phasing.



(3) IGF1 is most produced in liver  $\rightarrow$  peaks at the same time throughout body. However PIK3r1 (regulatory subunit for PIK3) peaks at different times in different organs.

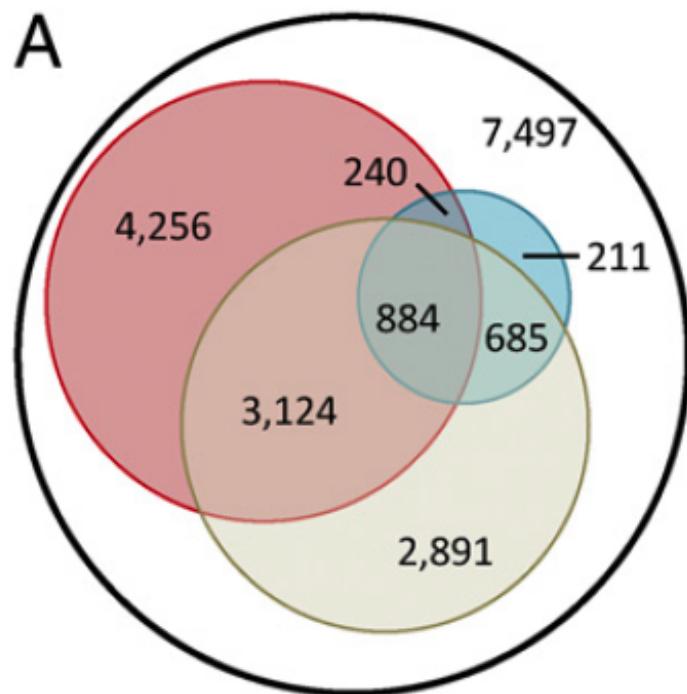
# Multiple coordinated pathways control PIK3-AKT-MTOR



Multiple synchronous (same peak time)  
receptors feed into PIK3-AKT-MTOR  
pathway that controls growth and  
apoptosis.

All of them oscillate only in **kidney!**  
Bioinformatics III

# Many drug-targets show circadian expression



- all genes
- circadian genes
- drug targets
- genes associated with disease

Relevance: drug response will differ significantly depending on day/night time of application

Unclear whether these effects are taken into account during clinical studies

# Relevance: mouse → humans, drugs

Table 1. Drugs of the top-100 best-seller list that target circadian genes and have half-life < 6h

Rank	Sales, \$	Trade name	Indications	Circadian-gene targets	Organs in which targets oscillate
2	1.46 b	Nexium	Gastritis, GERD, Esophagitis	<i>Atp4a</i>	L
5	1.28 b	Advair Diskus	Asthma, Chronic obstructive pulmonary di...	<i>Serpina6, Pgr, Nr3c2, Adrb2, Pla2g4a</i>	Lu, H, L, K, S, A
11	794 m	Rituxan	Rheumatoid arthritis, Non-Hodgkin's lymph...	<i>Fcgr2b, Ms4a1, Fcgr3</i>	L, K, S
20	538 m	Diovan	Hypertension, Heart failure	<i>Slc22a6, Agtr1a, Slco1b2, Car4, Kcnma...</i>	H, AG, L, K, S
27	431 m	Vyvanse	Attention deficit hyperactivity disorder	<i>Adra1b</i>	L
32	392 m	Tamiflu	Influenza	<i>Neu2, Neu1, Ces1g, Slc22a8, Slc15a1, ...</i>	Lu, L, BF, K, C
33	383 m	Ritalin	Attention deficit hyperactivity disorder	<i>Slc6a4</i>	AG, K
37	348 m	AndroGel	Hypogonadism	<i>Slc22a4, Slc22a3, Ar, Cyp1a1, Cyp2b10...</i>	Lu, H, BS, WF, AG...
38	346 m	Lidoderm	Pain	<i>Slc22a5, Cyp2b10, Egfr, Abcb1a</i>	Lu, H, AG, BF, L,...
44	304 m	Seroquel XR	Bipolar disorder, Major depressive disor...	<i>Htr2c, Htr1b, Htr2a, Chrm2, Drd4, Adr...</i>	Lu, H, BS, WF, AG...
45	289 m	Viagra	Erectile dysfunction	<i>Cyp1a1, Pde6g, Abcc5, Abcc10, Pde5a, ...</i>	Lu, H, BS, WF, AG...
47	281 m	Niaspan	Hyperlipidemia	<i>Slco2b1, Slc22a5, Qprt, Slc16a1</i>	Lu, H, BS, AG, WF...
48	279 m	Humalog	Diabetes mellitus T2	<i>Igf1r</i>	K
49	274 m	Alimta	Mesothelioma, Nonsmall cell lung cancer	<i>Tyms, Atic, Gart, Slc29a1</i>	Lu, H, BS, BF, L,...
54	267 m	Combivent	Asthma, Chronic obstructive pulmonary di...	<i>Slc22a5, Slc22a4, Chrm2, Adrb1, Adrb2</i>	Lu, H, BS, BF, K,...
56	262 m	ProAir HFA	Asthma, Chronic obstructive pulmonary di...	<i>Adrb1, Adrb2</i>	Lu, K, S
62	240 m	Janumet	Diabetes mellitus T2	<i>Slc47a1, Slc22a2, Prkab1, Abcb1a, Dpp4</i>	H, BS, AG, Hy, L,...
66	236 m	Toprol XL	Hypertension, Heart failure	<i>Slc22a2, Adrb1, Adrb2, Abcb1a</i>	Lu, H, AG, BF, L,...
71	220 m	Vytorin	Hyperlipidemia	<i>Hmgcr, Cyp2b10, Soat1, Abcc2, Anpep, ...</i>	Lu, H, BS, AG, BF...
78	209 m	Aciphex	Gastritis, GERD, Esophagitis	<i>Cyp1a1, Atp4a, Abcg2</i>	Lu, H, BS, WF, L,...
90	189 m	Lunesta	Insomnia	<i>Ptgs1, Tspo, Gabra3</i>	Lu, H, AG, K
98	173 m	Prilosec	Gastritis, GERD, Esophagitis	<i>Cyp1a1, Atp4a, Abcg2, Cyp1b1, Abcb1a</i>	Lu, H, BS, WF, AG...
99	171 m	Focalin XR	Attention deficit hyperactivity disorder	<i>Slc6a4</i>	AG, K

Rank and sales are based on USA 2013 Q1 data from [Drugs.com](#). A, aorta; AG, adrenal gland; BF, brown fat; BS, brainstem; C, cerebellum; H, heart; Hy, hypothalamus; K, kidney; L, liver; Lu, lung; S, skeletal muscle; WF, white fat.

About half of top-100 drugs have half lives < 6 hours!