

New path towards a better rice architecture

Cell Research (2017) 27:1189–1190. doi:10.1038/cr.2017.115; published online 12 September 2017

New plant type (NPT) or ideal plant architecture (IPA) is an attractive way of increasing yield potential by promoting high resource use efficiency combined with better lodging resistance. In a recent paper in *Cell Research*, Wang *et al.* describe how a QTL they identified could bring about the desired NPT architecture by elucidating the role of its encoded gene in controlling the stability of IPA1/OsSPL14, a previously reported NPT protein, in the context of ubiquitination.

Decades after the Green Revolution, plant breeders are still faced with the challenge to increase the yield potential of rice for the sake of sustaining a booming global population [1]. One of the popular approaches used by breeders to attain higher yield is to breed plants with ideal plant architecture, leading to the New Plant Type (NPT) concept. Unlike the semi-dwarf plants of the Green Revolution [2, 3], NPT plants have less tillers but more grains per panicle coupled with sturdier stems, giving rise to higher yield potential and better lodging resistance.

In a recent paper in *Cell Research*, Wang *et al.* [4] report a gene involved in the NPT architecture. The gene was identified through QTL analysis and positional cloning using japonica-indica crosses and was named as rice *OTUB1* (*OsOTUB1*) because of its homology to human *OTUB1* (ovarian tumor domain-containing ubiquitin aldehyde-binding protein 1). Plants showing the NPT phenotype had five unique DNA polymorphisms in *OsOTUB1* relative to conventional plants, and such polymorphisms did not alter the amino acid se-

quence. The involvement of *OsOTUB1* in the NPT architecture was confirmed by the following observations. First, a near isogenic line (NIL) carrying an allele with downregulated *OsOTUB1* expression showed the typical NPT phenotypes such as enhanced size of the shoot apical meristem (SAM), increased grain number per panicle due to increased primary and secondary rachis number, increased culm diameter and 1 000-grain weight, and reduced tiller number per plant. Second, similar phenotypes were observed in plants with disrupted *OsOTUB1* function by CRISPR/Cas9. Third, Huang *et al.* [5] also recently reported the null allele mutant (*wtg1*) of *OsOTUB1* and the same grain phenotypes as *OsOTUB1* loss-of-function mutants were observed, further confirming that *OsOTUB1* negatively regulates these traits. In contrast, *OsOTUB1* overexpression induced aberrant phenotypes such as dwarfism, malformed panicle with decreased grain number, necrotic leaves, and decreased tiller number per plant, which are not completely opposite to the phenotypes of the loss-of-function mutants, and thus, may indicate the side-effect of *OsOTUB1* overproduction.

Previous studies revealed that the human OTUB1 is a Lysine48 (K48)-specific deubiquitinating enzyme [6], whereas this protein was also reported to directly interact with the ubiquitin (Ub)-conjugating enzyme (E2), UBC13, thereby inhibiting ubiquitin transfer [7]. In contrast to human OTUB1, *OsOTUB1* has cleavage activity for both K48- and K63-linked Ub chains; but like human OTUB1, *OsOTUB1* also physically interacts with rice UBC13 (*OsUBC13*).

Overexpression of *OsUBC13* also induced the NPT phenotypes, whereas its knockdown plants showed similar phenotypes with *OsOTUB1* overexpressors. They further searched for proteins interacting with *OsOTUB1*, and identified SQUAMOSA promoter-binding protein-like (SPL) transcription factor 14 (*OsSPL14*) or IPA1, which is known to control plant architecture associated with reduced tiller number, thickened culm and enhanced grain number [8, 9]. They revealed that the NPT architecture controlled by *OsOTUB1* depends on *IPA1/OsSPL14* function, and that the NPT architecture is governed by the antagonistic relationship between the two genes as *OsOTUB1* promotes the degradation of IPA1/*OsSPL14*. One of the most interesting findings in this study is that *OsOTUB1*-mediated deubiquitination of K63-linked Ub chain is required for the degradation of IPA1/*OsSPL14*, whereas the deubiquitination of K48-linked Ub-IPA1/*OsSPL14* makes it resistant to proteasome-dependent degradation just like the case of human OTUB1. This is the reason why they called the cleavage of K63-linked poly-Ub as a non-canonical type of regulation.

In relation to the Ub-mediated degradation of IPA1/*OsSPL14*, another IPA1/*OsSPL14*-interacting protein, IPI1, a RING-finger E3 ligase was reported [10]. Just like *OsOTUB1*, overexpression and knockout of *IPI1* causes decrease and increase in grain number per panicle, respectively, suggesting a possible similarity in their function in the Ub-mediated degradation of IPA1/*OsSPL14* (Figure 1). Interestingly, IPI1 was found to have a tissue-specific dual nature; that is, it promotes proteasome-dependent degra-

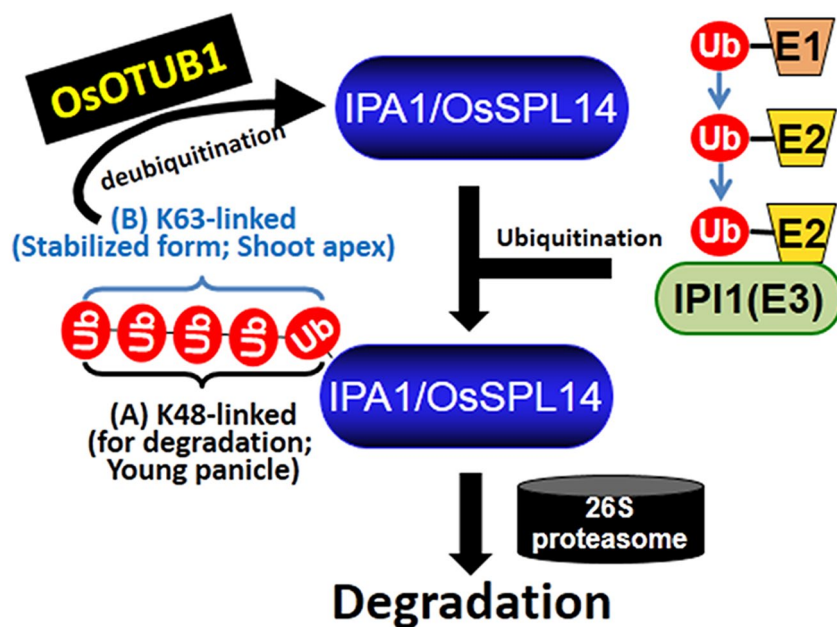


Figure 1 Model of IPA1/OsSPL14 degradation mediated by OsOTUB1 and IPI1 (E3). IPA1/OsSPL14 is a key factor regulating plant architecture in rice. Its ubiquitination is promoted by IPI1, an E3 ligase that can produce K48- or K63-linked Ub-IPA1/OsSPL14. The K48-linked type (A) mainly occurs in the young panicle and is degraded by the 26S-proteasome. In contrast, K63-linked type (B) mainly produced in the shoot apex resists proteasome-dependent degradation. OsOTUB1 has a deubiquitinating activity for the K63-linked poly-Ub, which promotes the production of K48-linked Ub-IPA1/OsSPL14 and consequently, leads to the degradation of IPA1/OsSPL14. In the case of loss of function of OsOTUB1 or IPI1, IPA1/OsSPL14 protein accumulates, leading to NPT plants.

dation of IPA1/OsSPL14 in young panicles and at the same time, promotes its stability in shoot apices. Consequently, *IPI1* knockout plants showed increased tiller number compared to the *IPA1/OsSPL14* overexpressors and *OsOTUB1* mutants, which show decreased tiller number characteristic of NPT plants. About this dual nature of IPI1, Wang *et al.* [10] explained that IPI1 ubiquitinates IPA1/OsSPL14 in two different manners; that is, K48-linked ubiquitination in panicles and K63-linked ubiquitination in shoot apices, which further supports the discussion above that IPA1/OsSPL14 with K63-linked Ub resists proteasome-dependent degradation.

Every existing rice architectural

concept has some drawbacks, and every attempt at increasing yield is subject to a trade-off. Indeed, most genes inducing large panicle, such as *IPA1/OsSPL14* [4] and *SCM3/OsTBI* [11], can increase the grain number per panicle, but at the same time, decrease the tiller number per plant. In the same manner, the *OsOTUB1* discussed by Wang *et al.* also appears to follow the same trade-off trend, but with good results in terms of high grain yield and lodging resistance. However, since breeders are under constant pressure to innovate, it is tempting to identify other genetic mechanisms that can break or attenuate such trade-off relationship in the future. To explore this possibility, the *IPI1* discussed here could be a good

candidate as it does not appear to sacrifice tiller number for increased grains per panicle. Furthermore, the paper about *OsOTUB1* by Wang *et al.* [4], alongside with the paper about *IPI1* [10], strongly suggests that the mode of ubiquitination of IPA1/OsSPL14 (K48- or K63-linked) regulates its stability in an organ-specific manner and consequently controls the NPT or ideal rice architecture, although the detailed molecular mechanism for this has not yet been fully clarified. Further studies should therefore be done not only in terms of basic science dealing with the Ub-mediated proteasome pathway but also molecular breeding for us to find better ways of increasing rice yield.

Reynante L Ordonio¹,
Makoto Matsuoka²

¹Plant Breeding and Biotechnology Division,
Philippine Rice Research Institute, Maligaya,
Science City of Munoz 3119, The Philippines;
²Bioscience and Biotechnology Center, Nagoya
University, Chikusa, Nagoya 464 8601

Correspondence: Makoto Matsuoka
Email: makoto@nuagr1.agr.nagoya-u.ac.jp

References

- Hirano K, Ordonio RL, Matsuoka M. *Proc Jpn Acad Ser B Phys Biol Sci* 2017; **93**:220-233.
- Sasaki A, Ashikari M, Ueguchi-Tanaka M, *et al. Nature* 2002; **416**:701-702.
- Spielmeier W, Ellis MH, Chandler PM. *Proc Natl Acad Sci USA* 2002; **99**:9043-9048.
- Wang S, Wu K, Qian Q, *et al. Cell Res* 2017; **27**:1142-1156.
- Huang K, Wang D, Duan P, *et al. Plant J* 2017; **91**:849-860.
- Sun X-X, Challagundla KB, Dai M-S. *EMBO J* 2012; **31**:576-92.
- Wiener R, Zhang X, Wang T, *et al. Nature* 2012; **483**:618-622.
- Jiao Y, Wang Y, Xue D, *et al. Nat Genet* 2010; **42**:541-544.
- Miura K, Ikeda M, Matsubara A, *et al. Nat Genet* 2010; **42**:545-549.
- Wang J, Yu H, Xiong G, *et al. Plant Cell* 2017; **29**: 697-707.
- Yano K, Ookawa T, Aya K, *et al. Mol Plant* 2015; **8**:303-314.