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# R COURSE

VITOR SOUSA, LAURENT EXCOFFIER

Day 3, Linear models

### Linear models with R



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In this session, we are going to examine in details a simple linear model

Linear regression

At the end of this document you will also find information on additional basic linear models, but we will not go over them here.

- One factor ANOVA (one way ANOVA)
- Two factor ANOVA (two way ANOVA)
- ANCOVA

### Linear regression

 $u^{^{\mathsf{b}}}$ 

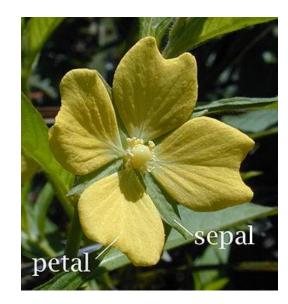
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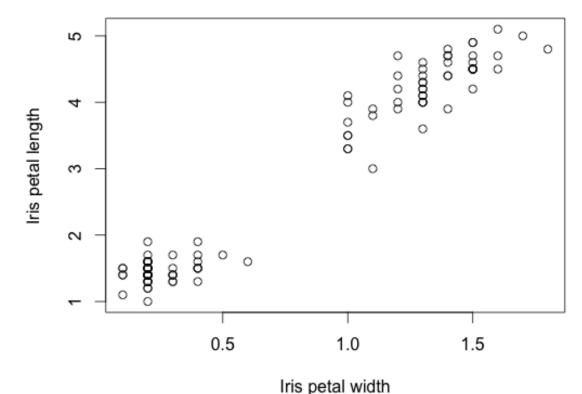
Linear regression is a statistical method aiming at modeling a **linear relationship between two quantitative variables**, usually for making predictions.

The variable to be explained is sometimes called the **response or dependent variable**. The other variable is sometimes called the **explanatory or independent variable**.

For instance we have the following relationship between petal width and

length in iris





### Linear regression



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Suppose that we have a given variable Y that we want to predict from an explanatory variable X.

We can write a simple linear model as

$$Y = \beta_0 + \beta_1 X + \varepsilon$$

The  $\varepsilon$  variable represents noise or stochastic errors (e.g. in measurements), and the parameters  $\beta_0$  and  $\beta_1$  are unknown.

We usually want to estimate these parameters from n observations of couples  $(x_1, y_1), (x_2, y_2), ..., (x_n, y_n)$  and a given observation  $y_i$  can be predicted as

$$y_i = \beta_0 + \beta_1 x_i + \varepsilon$$

**This is our model**, and we estimate the parameters  $(\beta_0; \beta_1)$  from the data.

# Regression line



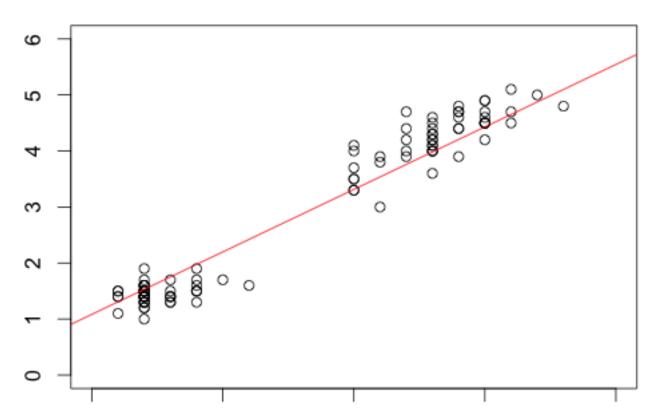
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 $b_0$  corresponds to the y-axis intercept at x=0 and  $b_1$  represents the slope of the **regression line** given by

$$f(x) = \hat{\beta}_0 + \hat{\beta}_1 x$$

which is plotted below in red. The predicted values of y are given by

$$\hat{y}_i = \hat{\beta}_0 + \hat{\beta}_1 x_i$$



### Residuals



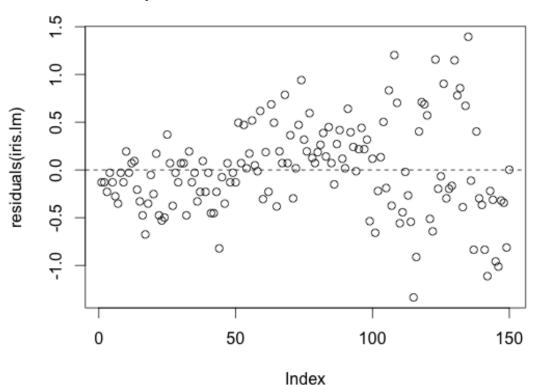
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The **residuals**  $\varepsilon_i$  represent the difference between the observed and predicted values of the Y dependent variable.

They are given by

$$\hat{\varepsilon}_i = y_i - \hat{y}_i$$

Their examination allows one to check the adjustment of the model to the data and to see if there are any aberrant points.



### Formulae



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R is using a specific notation to describe relationships between variables. These are called formulae. They use the tilde sign "~" like in

$$Y \sim X$$

which describes here a linear relationship between the dependent variable *Y* and the explanatory variable *X*.

This is a compact notation that R is using instead of

$$Y = \beta_0 + \beta_1 X + \varepsilon$$

One can use formulae to describe more complex relationships between variables as well as more complex models, which can be linear or non-linear, with or without interaction terms between variables.

Let's examine the use of formulae and linear regression using the iris data set

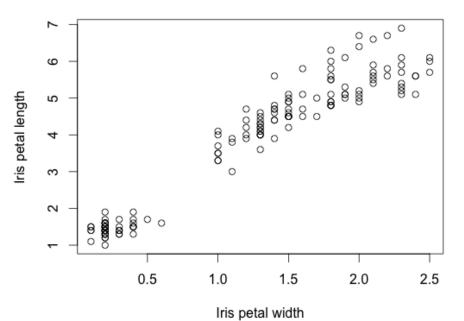


# Relation between iris petal width and length

```
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```

```
data(iris)
str(iris)
'data.frame': 150 obs. of 5 variables:
  Sepal.Length: num 5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
  Sepal.Width: num 3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
 $ Petal.Length: num
                              1.3
                                  1.5 1.4 1.7
                                              1.4 1.5 1.4 1.5 ...
  Petal.Width : num
                      0.2
                         0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1
  Species
               : Factor w/ 3 levels "setosa", "versicolor", ...: 1 1
 1 1 1 ...
plot(iris$Petal.Width,iris$Petal.Length, xlab="Iris petal width",
  vlab="Iris petal length" )
```





# Relation between iris petal width and length



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A simple linear regression of a dependent variable Y on an explanatory variable X (abbreviated as: regression of Y on X) is simply computed in R with the **Im function**, which expects a formula as input.

If we want to regress iris petal length on iris petal width we can simply state:

```
iris.lm=lm(Petal.Length ~ Petal.Width, data=iris)
```

which is a slightly clearer notation than

```
iris.lm=lm(iris$Petal.Length ~ iris$Petal.Width)
```

Let's have a look at the returned results found in iris.lm

```
names(iris.lm)
[1] "coefficients" "residuals" "effects" "rank" "fitted.values" "assign"
[7] "qr" "df.residual" "xlevels" "call" "terms" "model"
```



### Examining the estimated parameters



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```
summary(iris.lm)
Call:
lm(formula = Petal.Length ~ Petal.Width, data = iris)
Residuals:
    Min 10 Median 30 Max
-1.33542 -0.30347 -0.02955 0.25776 1.39453
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 1.08356 0.07297 14.85 <2e-16 ***
Petal.Width 2.22994 0.05140 43.39 <2e-16 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
Residual standard error: 0.4782 on 148 degrees of freedom
Multiple R-squared: 0.9271, Adjusted R-squared: 0.9266
F-statistic: 1882 on 1 and 148 DF, p-value: < 2.2e-16
```



# $u^{t}$

# Examining the significance of the estimated parameters with the **summary** function

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```
summary(iris.lm)
Call:
lm(formula = Petal.Length ~ Petal.Width, data = iris)
Residuals:
                                                    Intercept significantly
    Min 10 Median
                                30
                                        Max
                                                    different from zero
-1.33542 - 0.30347 - 0.02955 0.25776 1.39453
Coefficients:
                                                            Slope significantly
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 1.08356 0.07297 14.85 <2e-16 ***
                                                            different from zero
Petal.Width 2.22994 0.05140 43.39 <2e-16 ***
               0 \***' 0.001 \**' 0.01 \*' 0.05 \.' 0.1 \' 1
Signif. codes:
Residual standard error: 0.4782 on 148 degrees of freedom
Multiple R-squared: 0.9271, Adjusted R-squared: 0.9266
F-statistic: 1882 on 1 and 148 DF, p-value: < 2.2e-16
```

There is thus a significant relationship between petal width and length!





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```
summary(iris.lm)
Call:
lm(formula = Petal.Length ~ Petal.Width, data = iris)
Residuals:
    Min
           10 Median
                                30
                                        Max
                                              Proportion of variance of Y
-1.33542 - 0.30347 - 0.02955 0.25776 1.39453
                                                 explained by model
Coefficients:
                                                            Test of the difference
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 1.08356 0.07297 14.85
                                         <2e-16 ***
                                                           between our model and
Petal.Width 2.22994 0.05140 43.39
                                         <2e-16 ***
                                                           one with only the y-axis
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
                                                                 intercept
Residual standard error: 0.4782 on 148 degrees of freedom
Multiple R-squared: 0.9271, Adjusted R-squared: 0.9266
F-statistic: 1882 on 1 and 148 DF, p-value: < 2.2e-16
```

# Plotting the regression line



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```
The estimated coefficients (\beta_0; \beta_1) are given by
```

```
iris.lm$coefficients
(Intercept) Petal.Width
    1.083558    2.229940
```

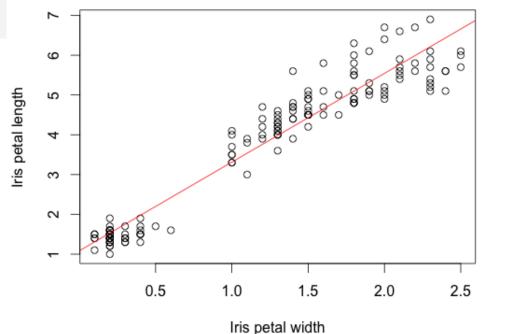
The first column is  $\beta_0$  (intercept) and the second is  $\beta_1$  (slope)

You can directly plot the regression line on top of the scatter plot with abline, which knows how to handle the coefficients of an Im analysis

(note that coef(iris.lm) is equivalent to iris.lm\$coefficients)

abline (coef (iris.lm), col="red")





# Plotting the residuals



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We can easily plot the residuals of the regression analysis

```
plot(residuals(iris.lm)); abline(h=0, lty=2)
```

We see that the variance of the residuals increases for larger indices

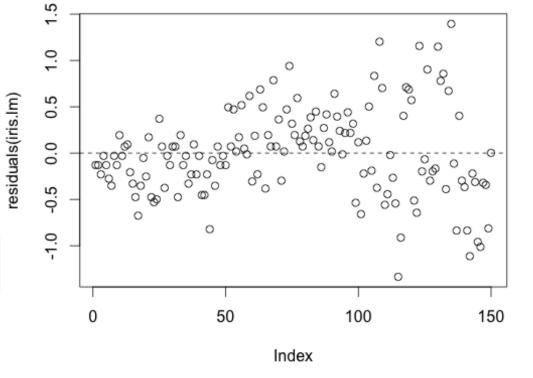
(heteroscedasticity)

Note that the sum of residuals should be zero by definition

### But

sum(residuals(iris.lm))
[1] -2.94556e-15

(not zero due to rounding errors in R)





# Plotting the residuals (advanced)



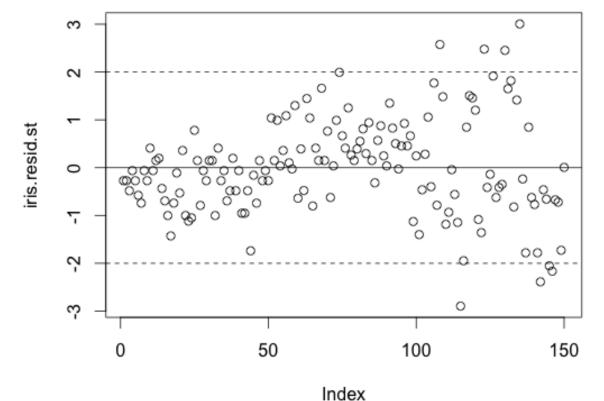
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We can **standardize the residuals** with the **rstudent** function and plot the limits of a 95% CI for the residuals.

```
iris.resid.st=rstudent(iris.lm)
plot(iris.resid.st)
abline(h=c(2,0,-2), lty=c(2,1,2))
```

We have about 8/150 points (5.33%) outside the 95% CI when we were

expecting 5%.



# Making predictions



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Since we have built a model, we could use this model to be able to predict the petal length of a given flower with known petal width

Suppose we have a flower with petal width of 0.8 cm, what would be its expected length?

# Making predictions

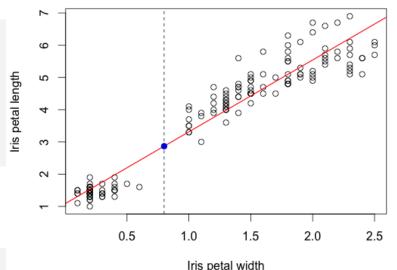


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Since we have built a model, we could use this model to be able to predict the petal length of a given flower with known petal width

Suppose we have a flower with petal width of 0.8 cm, what would be its expected length?

We can use the **predict** function for this, but it needs the new X value to be given as a **data frame** similar to that analysed with Im



### Check...

```
abline(v=0.8,lty=2)
points(0.8,predict(iris.lm, newXval), pch=19, col="blue")
```

# Confidence intervals (advanced notion)



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We can also use the predict function to get confidence intervals

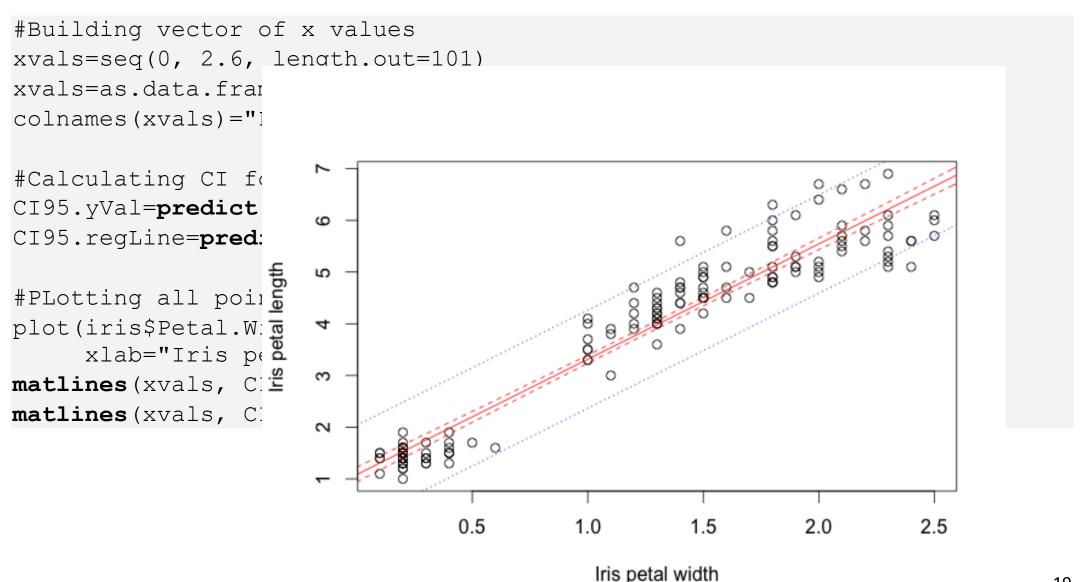
```
#Building vector of x values
xvals=seq(0, 2.6, length.out=101)
xvals=as.data.frame(xvals)
colnames(xvals)="Petal.Width"
#Calculating CI for y values and regression line
CI95.yVal=predict(iris.lm, xvals, interval="pred", level=0.95)
CI95.regLine=predict(iris.lm, xvals, interval="conf", level=0.95)
#Plotting all points and lines
plot(iris$Petal.Width, iris$Petal.Length,
     xlab="Iris petal width", ylab="Iris petal length")
matlines (xvals, CI95.yVal, lty=c(0,3,3), col="blue")
matlines (xvals, CI95.regLine, lty=c(1,2,2), col="red")
```



### Confidence intervals



We can also use the predict function to get confidence intervals



# One factor ANalysis Of Variance (ANOVA)



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In a one factor ANOVA, one studies the relationship between a **quantitative variable**, say Y, and **a categorical (qualitative) variable**, say A.

Even though name suggests that we are analyzing the variance, this analysis compares the means of Y for the different A categories (taking into account their variances)

It can thus be considered as an **extension of a t-test to more than two categories**.

# One factor ANalysis Of Variance (ANOVA)



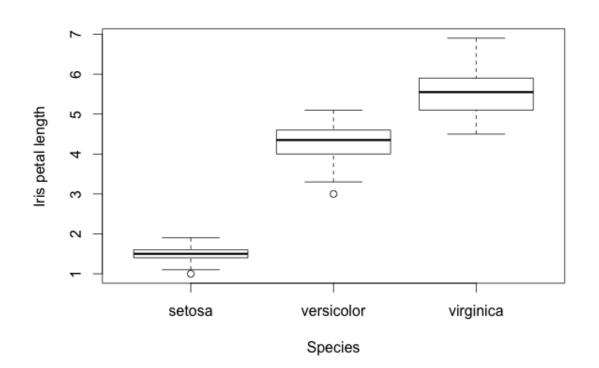
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For instance, taking the iris data set, we could want to compare the petal lengths among the different iris species.

As suspected before, there are obvious differences between the three species for petal length

An ANOVA formalizes such a test





### ANOVA with Im



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ANOVAs are extremely simply performed in R. One uses the same syntax as for linear regression.

```
iris.anova=lm(Petal.Length ~ Species, data=iris)
```

Im automatically performs the ANOVA when the formula describes a quantitative variable as a function of a categorical variable.

### ANOVA with Im



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ANOVAs are extremely simply performed in R. One uses the same syntax as for linear regression.

```
iris.anova=lm(Petal.Length ~ Species, data=iris)
```

Im automatically performs the ANOVA when the formula describes a quantitative variable as a function of a categorical variable.

The results of the analysis can be visualized by the anova helper function

### ANOVA with Im



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ANOVAs are extremely simply performed in R. One uses the same syntax as for linear regression.

```
iris.anova=lm(Petal.Length ~ Species, data=iris)
```

Im automatically performs the ANOVA when the formula describes a quantitative variable as a function of (a) categorical variable(s).

The results of the analysis can be visualized with the anova helper function

```
anova (iris.anova)
                                                             Probability that the
Analysis of Variance Table
                                                           observed means of the
Response: Petal.Length
                                                           different species are as
            Df Sum Sq Mean Sq F value
                                            Pr (>F)
                                                          different by chance alone
             2 437.10 218.551 1180.2
                                        < 2.2e-16 ***
Species
                27.22
Residuals 147
                         0.185
Signif. codes:
                 0 \***' 0.001 \**' 0.01 \*' 0.05 \.' 0.1 \' 1
```

### ANOVA as a linear model



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Actually an ANOVA analysis can be expressed as a linear model where a given observation *j* of the variable Y in group *i* can be expressed as

$$y_{ij} = \mu + \alpha_i + \varepsilon_{ij}$$

with:

 $\mu$ : global mean effect

 $\alpha_i$ : additional effect of being in group i

 $\mathcal{E}_{ij}$ : residual for observation j

This model is contrasted to the model

$$y_{ij} = \mu + \varepsilon_{ij}$$

where it is assumed that there is no effect of the modality, or in other words that all groups have the same mean.

# ANOVA estimated parameters



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The parameters of the model can be examined with the **summary** function

```
summary(iris.anova)
Call:
lm(formula = Petal.Length ~ Species, data = iris)
Residuals:
  Min 10 Median 30 Max
-1.260 -0.258 0.038 0.240 1.348
Coefficients:
                Estimate Std. Error t value Pr(>|t|)
(Intercept) 1.46200 0.06086 24.02 <2e-16 ***
Speciesversicolor 2.79800 0.08607 32.51 <2e-16 ***
Speciesvirginica 4.09000 0.08607 47.52 <2e-16 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.4303 on 147 degrees of freedom
Multiple R-squared: 0.9414, Adjusted R-squared: 0.9406
F-statistic: 1180 on 2 and 147 DF, p-value: < 2.2e-16
```

# ANOVA estimated parameters



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The parameters of the model can be examined with the summary function

```
summary(iris.anova)
                                                               Here the first group (I. setosa)
Call:
                                                                  is taken as a reference
lm(formula = Petal.Length ~ Species, data = iris)
                                                            and \partial_{\text{setosa}} = 0. It implies that the
Residuals:
                                                            mean petal length of I. setosa is M
   Min
            10 Median
                            30
                                   Max
-1.260 -0.258 0.038
                         0.240
                                 1.348
                                             a versicolor
                                                               a virginica
Coefficients:
                    Estimate Std. Error t value Pr(>|t|)
                     1.46200
                                  0.06086
                                             24.02 <2e-16 ***
(Intercept)
                    2.79800
                                  0.08607
                                             32.51 <2e-16 ***
Speciesversicolor
                                             47.52 <2e-16 ***
Speciesvirginica
                     4.09000
                                  0.08607
                                                                            Probability of
                  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Signif. codes:
                                                                           simpler model
Residual standard error: 0.4303 on 147 degrees of freedom
                                                                             \mathbf{y}_{ii} = \mathcal{M} + \mathcal{C}_{ii}
Multiple R-squared: 0.9414, Adjusted R-squared:
F-statistic: 1180 on 2 and 147 DF, p-value: < 2.2e-16
```

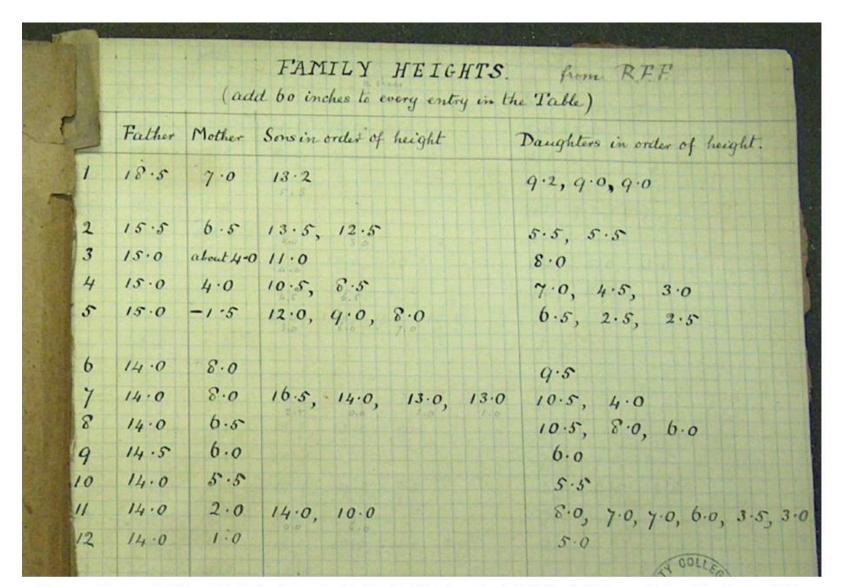


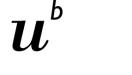
Figure 1. Photograph of the entries for the first 12 families listed in Galton's notebook. Published with the permission of the Director of Library Services of University College London.



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Hanley (2004) The American Statistician



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# Advanced linear models (for your own information, but not examined in this course)

### Multi-factor ANOVA with interaction

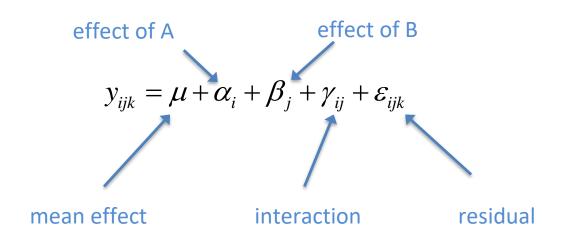


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In this analysis, one wants to model one quantitative variable (Y) as a function of several qualitative variables (A, B,...). Here we shall consider only two factors A and B.

In this case, we need to take into account the separate effects of the variables A and B, as well as their joint effects (interaction effects)

The model we will consider can thus be written as



# R modeling of 2-factor ANOVA



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The previous model can be simply specified in R with the following formula

An equivalent and perhaps more explicit formulation would be

$$Y \sim A+B+A:B$$

Where A:B specifies explicitly the interaction term to be tested

If one does not want to test the interaction (assumes that factors act additively only), then one can use the following formula

# $u^{^{\scriptscriptstyle b}}$

# Example of 2-factor ANOVA with interaction

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We shall analyze the MASS::cabbages data set, which reports data from a cabbage field experiment.

Load the MASS library and the cabbages data set

```
require(MASS)
data(cabbages)
```



### Look at its structure

```
str(cabbages)
'data.frame': 60 obs. of 4 variables:
    $ Cult : Factor w/ 2 levels "c39", "c52": 1 1 1 1 1 1 1 1 1 1 1 1 ...
    $ Date : Factor w/ 3 levels "d16", "d20", "d21": 1 1 1 1 1 1 1 1 1 1 1 ...
    $ HeadWt: num    2.5 2.2 3.1 4.3 2.5 4.3 3.8 4.3 1.7 3.1 ...
    $ VitC : int 51 55 45 42 53 50 50 52 56 49 ..
```

Cult: 2 two cabbage cultivars (plants selected for some desirable characteristics)

Date: 3 planting dates

HeadWt: Cabbage head weight (kg)

VitC: amount of ascorbic acid (Vit C) received (information discarded here)

Question: How does the head weight depend on Cult and Date?

### Look at the data



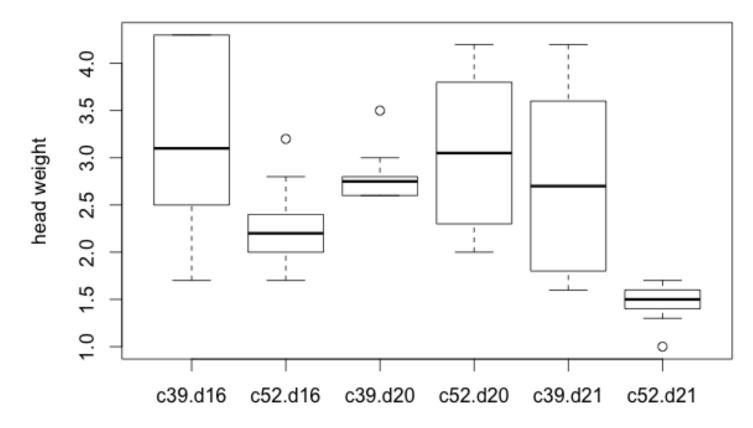
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We can also use a formula in boxplot to specify we would like to plot all types of weight for all combinations of Cult and Date

```
boxplot(HeadWt ~ Cult*Date, data=cabbages)
```



The mean weights of the different combinations are quite different



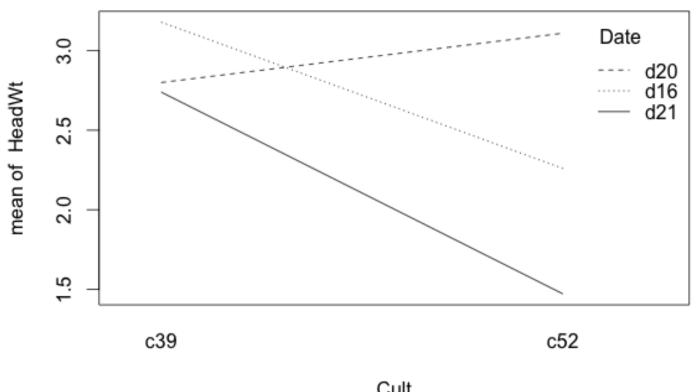
### Look at the data



Use an interaction plot to visualize how the means of the different combinations of factors differ

```
with (cabbages, interaction.plot(Cult, Date, HeadWt))
```

Note the use of the with function, and the order of the variables (explanatory first, and dependent last)



34 Cult

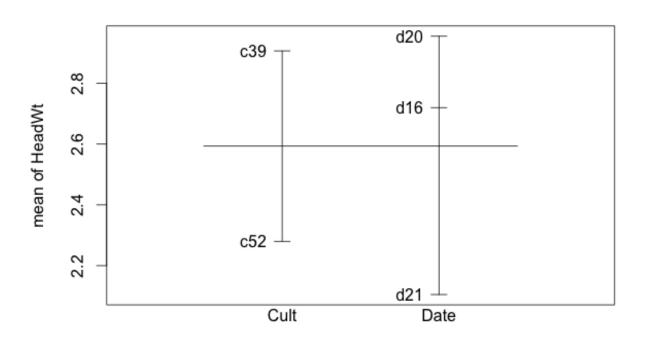
### Look at the data



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Finally, another way to plot differences in means of single categories is to use the **plot.design** function

plot.design(cabbages)



### ANOVA results



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### Let's perform our 2 factor ANOVA (2-way ANOVA):

#### **ANOVA** results



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#### Let's perform our 2 factor ANOVA:

```
>cab.anova=lm(HeadWt ~ Cult*Date, data=cabbages)
                                                        Interaction term is significant:
>anova(cab.anova)
                                                          It implies that the weight
Analysis of Variance Table
                                                         depends on the interaction
Response: HeadWt
                                                         between the cultivar and the
               Sum Sq Mean Sq F value Pr(>F)
                                                               planting date
Cult
               5.8907 5.8907 12.4969 0.0008451
                                                   * * *
            2 7.7063 3.8532 8.1744 0.0007920
Date
Cult:Date
               6.8863 3.4432 7.3046 0.0015571
Residuals 54 25.4540 0.4714
                 0 \***' 0.001 \**' 0.01 \*' 0.05 \.' 0.1 \' 1
Signif. codes:
```

### Distinguishing between models



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We have seen that all our factors were significant, but we might want to compare the fit of this model with that of simpler ones, for instance let's compare

HeadWt ~ Cult \* Date

with

HeadWt ~ Cult + Date

```
hw1=lm(HeadWt ~ Cult*Date, data=cabbages)
hw2=lm(HeadWt ~ Cult+Date, data=cabbages)
#Performs an analysis of variance on associated model residuals
anova(hw2,hw1)
Analysis of Variance Table

Model 1: HeadWt ~ Cult + Date
Model 2: HeadWt ~ Cult * Date
Res.Df RSS Df Sum of Sq F Pr(>F)
1 56 32.340
2 54 25.454 2 6.8863 7.3046 0.001557 **
```





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We have seen that all our factors were significant, but we might want to compare the fit of this model with that of simpler ones, for instance let's compare

HeadWt ~ Cult \* Date

with

HeadWt ~ Cult + Date

#### Residual sums of

```
hw1=lm(HeadWt ~ Cult*Date, data=cabbages)
                                                  squares
hw2=lm(HeadWt ~ Cult+Date, data=cabbages)
#Performs an analysis of variance on associated model residuals
anova (hw2, hw1)
Analysis of Variance Table
                                                      Smaller mean
Model 1: HeadWt ~ Cult + Date
                                                                     The more complex
residuals in more
 Res.Df
            RSS Df Sum of Sq
                                      Pr(>F)
                                                                     model better fits
                                                     complex model is
      56 32.340
                                                                         the data
      54 25.454
                      6.8863 7.3046 0.001557 **
                                                        significant
```

### Analysis of covariance - ANCOVA



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ANCOVA is about the analysis of a quantitative variable to be explained by quantitative and qualitative variables.

The simplest model would be that of a quantitative variables Y to be explained by another quantitative variable X and a categorical variable A with I categories.

Since the relationship between Y and X may depend on the category of A, one performs separate regressions for each of the I categories.

The model for the *i*-th category would thus look like

$$y_{i,j} = \alpha_i + \gamma_i x_{i,j} + \varepsilon_{i,j}$$

### Analysis of covariance - ANCOVA



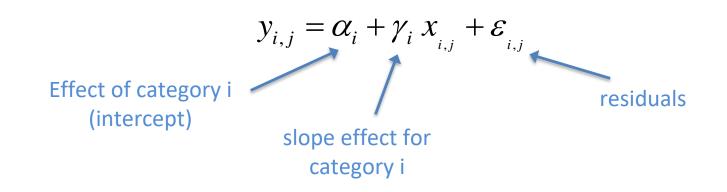
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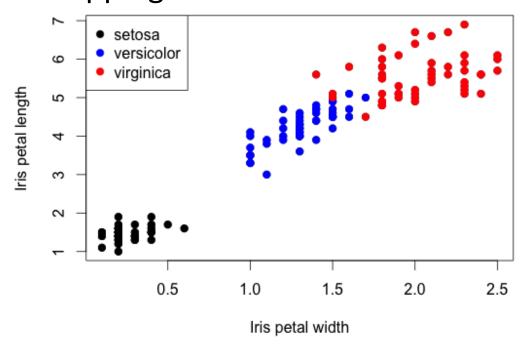
### ANCOVA applied example



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Let's consider the iris example again.

We have previously analyzed the relationship between petal length and petal width, by pooling data from the three species, but the three species have almost non-overlapping distributions.



We could thus ask whether the relationship between these two quantitative variables differs between species.

### Defining the ANCOVA model in R



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We would thus perform an ANCOVA to test if the relationship between the size and the shape of the petals is different in the three species

This is straightforward in R as we just have to analyze the model

#### Petal.Length ~ Petal.Width\*Species

R will understand that we need to do an ANCOVA since we want to explain a quantitative variable by both a quantitative and a qualitative variable

### Performing the ANCOVA



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#### We can again use the lm function to perform the ANCOVA



### Performing the ANCOVA



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#### We can again use the Im function to perform the ANCOVA

iris.ancova=lm(Petal.Length ~ Petal.Width\*Species, data=iris) anova (iris.ancova) All effects are significant: Analysis of Variance Table Petal length depends on i) petal width, ii) species, Response: Petal.Length and iii) the interaction Df Sum Sq Mean Sq F value Pr(>F) between species and 1 430.48 430.48 3294.5561 < 2.2e-16 \*\*\* Petal.Width width 13.01 6.51 49.7891 < 2.2e-16 \*\*\* Species Petal.Width:Species 2 2.02 1.01 7.7213 0.0006525 \*\*\* Residuals 18.82 0.13 144



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## The interpretation of the summary is not obvious and requires some explanations

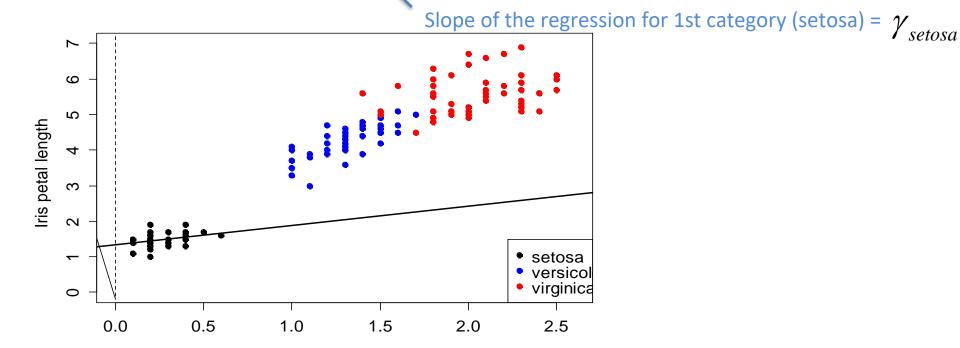
```
summary(iris.ancova)
Call:
lm(formula = Petal.Length ~ Petal.Width * Species, data = iris)
Residuals:
    Min
          10 Median 30
                                      Max
-0.84099 - 0.19343 - 0.03686 0.16314 1.17065
Coefficients:
                            Estimate Std. Error t value Pr(>|t|)
                              1.3276
                                        0.1309 10.139 < 2e-16 ***
(Intercept)
Petal.Width
                             0.5465
                                        0.4900 1.115 0.2666
Speciesversicolor
                             0.4537
                                        0.3737 1.214 0.2267
Speciesvirginica
                             2.9131
                                        0.4060 7.175 3.53e-11 ***
Petal.Width:Speciesversicolor
                            1.3228
                                        0.5552
                                               2.382 0.0185 *
Petal.Width:Speciesvirginica
                              0.1008
                                        0.5248
                                                0.192
                                                       0.8480
Residual standard error: 0.3615 on 144 degrees of freedom
Multiple R-squared: 0.9595, Adjusted R-squared: 0.9581
F-statistic: 681.9 on 5 and 144 DF, p-value: < 2.2e-16
```



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#### Let's concentrate on the coefficients:

Intercept of the regression for 1st category (setosa) =  $\alpha_{setosa}$ Coefficients: Estimate Std. Error t value Pr(>|t|) (Intercept) 1.3276 0.1309 10.139 \* \* \* < 2e-16 Petal.Width 0.5465 0.4900 1.115 0.2666 Speciesversicolor 0.4537 0.3737 1.214 0.2267 Speciesvirginica 2.9131 0.4060 7.175 3.53e-11 \* \* \* Petal.Width:Speciesversicolor 1.3228 0.5552 2.382 0.0185 \* 0.8480 Petal.Width:Speciesvirginica 0.1008 0.5248 0.192



Iris petal width

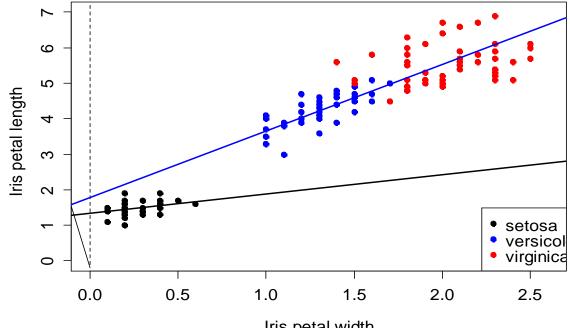


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#### Let's concentrate on the coefficients:

Difference in intercept between versicolor and setosa =  $lpha_{versicolor} - lpha_{setosa}$ Coefficients: Estimate Std. Error t value Pr(>|t|) (Intercept) 10.139 < 2e-16 \*\*\*1.3276 0.1309 Petal.Width 0.5465 0.4900 1.115 0.2666 Speciesversicolor 0.4537 0.3737 1.214 0.2267 Speciesvirginica 2.9131 0.4060 7.175 3.53e-11 \* \* \* Petal.Width:Speciesversicolor 1.3228 0.5552 2.382 0.0185 \* 0.8480 Petal.Width:Speciesvirginica 0.1008 0.5248 0.192





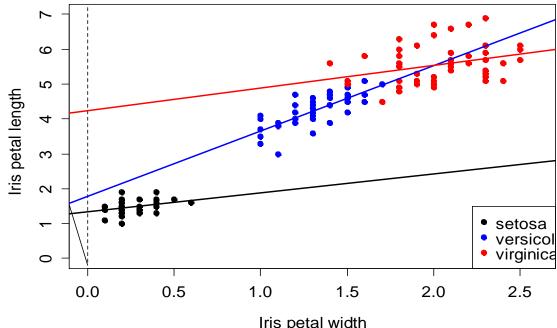


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#### Let's concentrate on the coefficients:

Difference in intercept between virginica and setosa =  $lpha_{virginica} - lpha_{setosa}$ Coefficients: Estimate Std. Error t value Pr(>|t|) 10.139 < 2e-16 \*\*\*(Intercept) 1.3276 0.1309 Petal.Width 0.5465 0.4900 1.115 0.2666 Speciesversicolor 0.4537 0.3737 1.214 0.2267 Speciesvirginica 2.9131 0.4060 7.175 3.53e-11 \* \* \* Petal.Width:Speciesversicolor 1.3228 0.5552 2.382 0.0185 \* 0.1008 0.192 Petal.Width:Speciesvirginica 0.5248 0.8480

Difference in slope between virginica and setosa =  $\gamma_{virginica} - \gamma_{setosa}$ 



### Interpretation of ANCOVA results



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Our ANCOVA analysis of the iris data show the following results:

1) Petal length significantly depends on petal width, on the species, and on the interaction between petal width and the species.

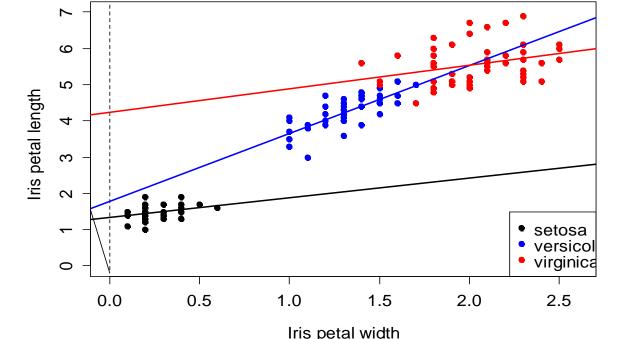
2) The relationships between petal length and width differs between species

a. In **setosa**, length does not depend on width (slope is not significant)

b. In versicolor, the intercept is the same as in setosa but the slope is different

c. In virginica, the intercept is not the same as in setosa but the slope is the same: virginica petals are significantly longer than in setosa for the same

width



#### Exercice



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Try to perform the ancova analysis on the iris data set and plot the regression lines on top of the plot as shown in the former slide



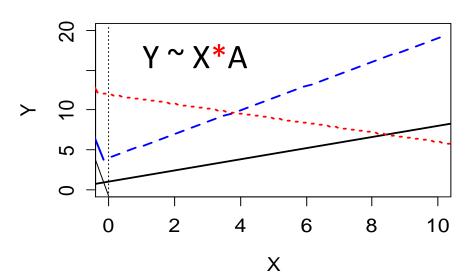
### Comparing models

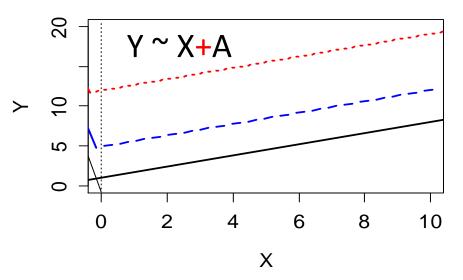


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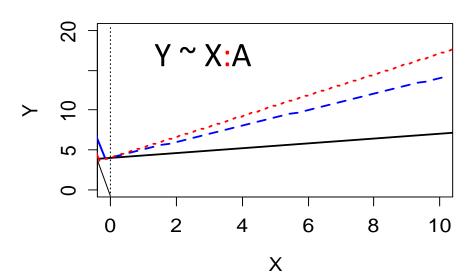


#### Same slopes, different intercepts





#### Same intercepts, different slopes



### Comparing models



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#### We can build the different iris models in R very simply

```
full_model=lm(Petal.Length ~ Petal.Width*Species, data=iris)
same_slope=lm(Petal.Length ~ Petal.Width*Species, data=iris)
same_intercept=lm(Petal.Length ~ Petal.Width:Species, data=iris)
```

and we can perform model comparison by an ANOVA on the residuals



## $u^{t}$

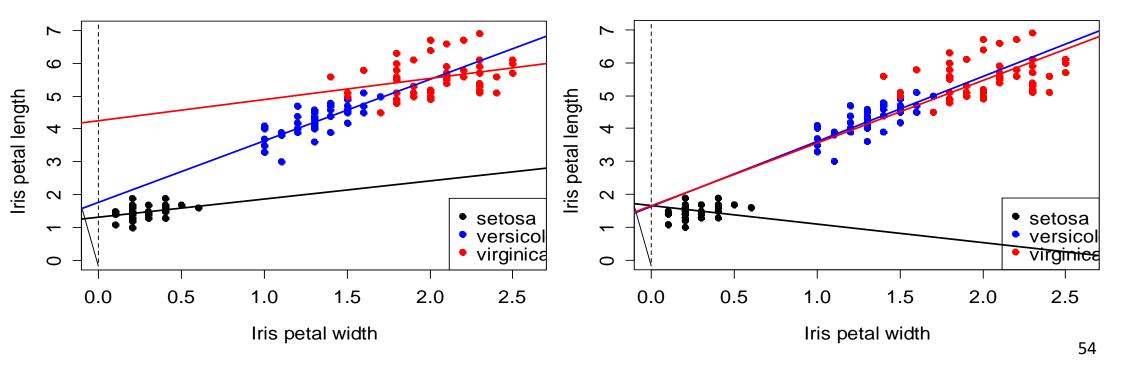
# Comparing full model and model with same intercept

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```
anova(same_intercept, full_model)
Analysis of Variance Table

Model 1: Petal.Length ~ Petal.Width:Species
Model 2: Petal.Length ~ Petal.Width * Species
Res.Df RSS Df Sum of Sq F Pr(>F)

1 146 25.563
2 144 18.816 2 6.7474 25.82 2.614e-10 ***
```



### $u^{t}$

# Comparing full model and model with same slope

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```
anova(same_slope, full_model)
Analysis of Variance Table

Model 1: Petal.Length ~ Petal.Width + Species
Model 2: Petal.Length ~ Petal.Width * Species
   Res.Df   RSS Df Sum of Sq   F   Pr(>F)
1   146 20.833
2   144 18.816 2   2.0178 7.7213 0.0006525 ***
---
Signif. codes: 0 \***' 0.001 \**' 0.05 \'.' 0.1 \' 1
```

