human reproduction

ORIGINAL ARTICLE Embryology

Artificial oocyte activation with SrCl₂ or calcimycin after ICSI improves clinical and embryological outcomes compared with ICSI alone: results of a randomized clinical trial

Mohamed Fawzy^{1,*}, Mai Emad¹, Ali Mahran², Mohamed Sabry³, Ahmed N. Fetih⁴, Hazem Abdelghafar³, and Salah Rasheed³

¹IbnSina IVF Centre, IbnSina Hospital, Sohag 15322, Egypt ²Department of Dermatology, Venereology and Andrology, Faculty of Medicine, Assiut University, AG 71515, Egypt ³Department of Obstetrics and Gynecology, Sohag University, Sohag 82524, Egypt ⁴Department of Obstetrics and Gynecology, Faculty of Medicine, Assiut University, AG 71515, Egypt

*Correspondence address. IVF Laboratory Director (IbnSina and Banoon IVF Centres), IbnSina Hospital, 146El Aref Square, Sohag, Egypt. Tel: +20-101-112-2286; E-mail: drfawzy001@me.com

Submitted on February 28, 2018; resubmitted on June 30, 2018; accepted on July 9, 2018

STUDY QUESTION: Are pregnancy and birth rates affected by artificial oocyte activation (AOA) with SrCl₂ or calcimycin after ICSI for couples with male-factor infertility linked to abnormal sperm morphology or for couples with previous ICSI cycles of unexplained low fertilization or inadequate fertilization associated with impaired oocyte morphology?

SUMMARY ANSWER: AOA with either SrCl₂ or calcimycin can improve the rates of clinical pregnancy, ongoing pregnancy and live birth compared with ICSI alone, and the two agents have diverse effects for different subgroups of patients.

WHAT IS KNOWN ALREADY: ICSI is a successful treatment for infertility, but not in all individuals. AOA has potential to overcome inadequate fertilization in ICSI. Calcimycin and SrCl₂ are candidate agents for AOA, but their effectiveness remains to be compared.

STUDY DESIGN, SIZE, DURATION: This study was a randomized, open-label, three-arm, parallel-group, double-centre, superiority trial conducted between April 2015 and January 2016. The study evaluated the effects of AOA with calcimycin or SrCl₂ for clinical pregnancy rates after ICSI and included 343 couples divided into three groups.

PARTICIPANTS/MATERIALS, SETTING, METHODS: Couples were included if they had two previous ICSI cycles of no or low fertilization (0–30%) with unknown causes or impaired oocyte morphology. Male-factor infertility cycles (frozen–thawed sperm, surgically retrieved sperm or ejaculates contained <10 millions spermatozoa/ml) undergoing their first ICSI attempt were also included if they had 100% abnormal sperm morphology (including globozoospermia and tapered-head). Couples were randomized to undergo ICSI with SrCl₂ AOA, ICSI with calcimycin AOA or ICSI alone, with clinical pregnancy as the primary endpoint. Effect sizes were summarized as absolute rate differences (ARDs) and odds ratios (ORs), with precision evaluated by 95% CIs.

MAIN RESULTS AND THE ROLE OF CHANCE: Both $SrCl_2$ and calcimycin AOA improved clinical pregnancy rates compared to ICSI alone (49, 42 and 27%; ARD 22, 95% CI: 9–33; P = 0.0007 and ARD 16, 95% CI: 3–27; P = 0.014). $SrCl_2$ and calcimycin AOA were also superior to ICSI alone on the rates of ongoing pregnancy (42, 36 and 23%; P = 0.0019 and P = 0.023) and live birth (40, 33 and 18%; P = 0.0002 and P = 0.012). Among couples with previous ICSI cycles of low fertilization, AOA with $SrCl_2$ (but not with calcimycin) was superior to ICSI alone for rates of clinical pregnancy (ARD 35 percentage points (pp), P = 0.0007), ongoing pregnancy (ARD 27 pp, P = 0.009) and live birth (ARD 37 pp, P = 0.002). Among couples affected by male-factor infertility, AOA with calcimycin (but not with $SrCl_2$) was superior to ICSI alone for rates of clinical pregnancy (ARD 22 pp, P = 0.006), ongoing pregnancy (ARD 19 pp, P = 0.013) and live birth (ARD 17 pp, P = 0.02).

LIMITATIONS, REASONS FOR CAUTION: This study was an open-label trial, and this design might have introduced bias, although randomization methods were used. The study did not include a longitudinal follow-up, so further evidence is required to demonstrate the safety of AOA.

WIDER IMPLICATIONS OF THE FINDINGS: The decision to use SrCl₂ or calcimycin for AOA after ICSI may depend on whether the activation failure originates in the oocyte or the sperm.

STUDY FUNDING/COMPETING INTEREST(s): The study received no funding and the authors have no conflicts of interest to declare.

TRIAL REGISTRATION NUMBER: NCT02424214.

TRIAL REGISTRATION DATE: 22 April 2015.

DATE OF FIRST PATIENT'S ENROLMENT: 27 April 2015.

Key words: oocyte activation / calcium ionophore / calcium oscillation / failed fertilization / male factor infertility

Introduction

IVF and ICSI are effective treatments for infertility (Palermo et al., 1992; Steptoe and Edwards, 1992; Kissin et al., 2014). Successful fertilization requires oocyte activation, which depends on a proper interaction between the gametes (Yeste et al., 2016). Activation failure results in poor fertilization rates (Flaherty et al., 1998; Cheng et al., 2011; Clift and Schuh, 2013).

Oocyte activation triggers a cascade of events including Ca²⁺ oscillation, leading to fertilization and embryo development (Yeste et al., 2016). In animal models, the duration, amplitude and frequency of Ca²⁺ oscillation affect the normal oocyte activation and embryo development (Sfontouris et al., 2015). The precise mechanisms underlying oocyte activation remain elusive. A likely mechanism is that the phospholipase C zeta (PLCC) of a fertilizing spermatozoon produces inositol-1,4,5trisphosphate within the oocyte which interacts with receptors in the endoplasmic reticulum, causing Ca²⁺ oscillation (Ridgway et al., 1977; Steinhardt et al., 1977; Miyazaki et al., 1992, 1993; Wassarman et al., 2001; Ducibella et al., 2002; Saunders et al., 2002; Horner and Wolfner, 2008; Kashir et al., 2010; Amdani et al., 2013; Nomikos et al., 2013; Escoffier et al., 2014). Reports suggest that abnormal sperm morphology reduces the rates of fertilization and implantation (De Vos et al., 2003; Lu et al., 2012; Greco et al., 2013). The oocyte also contributes to the activation events via identified and unidentified factors (Tesarik et al., 2002; Eldar-Geva et al., 2003; Heindryckx et al., 2005, 2008; Combelles et al., 2010; Yelumalai et al., 2015).

Artificial oocyte activation (AOA) can repair a defective activation (Yeste et al., 2016), improving ICSI outcomes (Montag et al., 2012). Mechanical, electrical and chemical activation serve as AOA methods (Vanden Meerschaut et al., 2012; 2014). Chemical AOA is the prominent, and it induces either a single, prolonged wave or pulsatile waves of intracytoplasmic Ca²⁺ (Yeste et al., 2016; Murugesu et al., 2017). Calcimycin, ionomycin, puromycin and 6-dimethylaminopurine increase membrane permeability to extracellular Ca²⁺, inducing a single, prolonged Ca²⁺ wave (Rybouchkin et al., 1997; Ebner et al., 2015; Yeste et al., 2016). Strontium chloride (SrCl₂), phorbol esters or anhydrous alcohol release the Ca²⁺ from the endoplasmic reticulum in pulsatile waves (Kline and Kline, 1992; Kishikawa et al., 1999; Kim et al., 2014; Yeste et al., 2016).

AOA with calcimycin or SrCl₂ can improve outcomes for ICSI cycles of impaired fertilization (Yanagida et al., 2006; Chen et al., 2010; Kim

et al., 2014; Murugesu et al., 2017). Variable studies including retrospective designed analyses, historical comparisons or relatively underpowered studies have synthesized this evidence. Most of the studies have only examined calcimycin AOA, although SrCl₂ has shown potential for AOA (Kim et al., 2014; Yeste et al., 2016). Recently, Lu et al. (2018) showed that SrCl₂ AOA cannot provoke calcium oscillation or parthenogenic division for human oocytes. However, Lu et al. used in vitro matured oocytes or oocytes characterized by smooth endoplasmic reticulum. In addition, there has been no comparative randomized clinical trial (RCT) to evaluate the relative effectiveness of calcimycin and SrCl₂. Whether the sperm or the oocyte is most responsible for the activation defects remains unidentifiable from a clinical perspective (Murugesu et al., 2017).

To address these questions, we compared $SrCl_2$ and calcimycin AOA with each other and with ICSI alone in this prospective RCT, identifying the rate of clinical pregnancy as a primary endpoint. To determine whether the sperm or the oocyte is most responsible, infertile couples included were those undergoing ICSI cycles for previous low fertilization (<30%) but normal semen parameters or first ICSI cycles for male-factor infertility with abnormal sperm morphology.

Materials and Methods

Trial design

The study adopted a randomized, open-label, three-arm, parallel-group, double-centre, superiority trial design (NCT02424214, registered at www. ClinicalTrials.gov). Two private IVF facilities in Egypt (Ibnsina IVF Centre, Suhag and Banoon IVF Centre, Asyut) conducted the trial. The trial included a screening period of 18 months (from the time of getting the consent to the start of assignment of participants). Research instructors disclosed the details to eligible couples. From January to April 2015, research instructors recalled participants for randomization and enrolment. The first couple was recruited on 30 April 2015, and the last oocyte retrieval was performed on 17 January 2016. Participants provided written informed consent. The ethics committees of the centres approved the study (Number 09; 02/2013). Independent experts and safety committees oversaw this study.

Participants

Women were 18–40 years of age with a BMI of $<31 \, \text{kg/m}^2$. Inclusion criteria were deliberately broad to include couples with two previous ICSI cycles of no or low fertilization (0–30%) or with male-factor infertility

1638 Fawzy et al.

undergoing their first ICSI cycle. We classified previous cycles of low fertilization as 'unexplained low fertilization' or as 'low fertilization associated with impaired oocyte morphology', but within normal sperm limits. Impaired morphology of oocytes, according to the Istanbul Consensus, was non-conformity with the following definition: a spherical structure in a uniform zona pellucida with a uniform translucent cytoplasm free of inclusions and with a size-proper polar body (Alpha Scientists in Reproductive Medicine and ESHRE Special Interest Group of Embryology, 2011). The male-factor infertility cycles (frozen-thawed sperm, surgically retrieved sperm or ejaculates containing <10 million spermatozoa/ml) undergoing their first ICSI attempt had 100% abnormal sperm morphology (including globozoospermia and tapered-head). Couples had primary infertility with no previous completed pregnancy or abortion. Exclusion criteria were couples with endometriosis, thin endometrial lining or pinhead sperm defects.

Randomization and masking

Using an online randomization tool (http://www.openepi.com/Random/Random.htm), independent data managers assigned participants to SrCl₂ AOA, calcimycin AOA or ICSI alone with a 1:1:1 allocation ratio. Block randomization was performed to ensure equal representation of couples in the study groups. Each allocation was sealed in a sequentially numbered, opaque envelope. On 27 April 2015, independent instructors recorded each envelope number in each woman's case-record file. On Day 21 of the cycle preceding ICSI, research instructors opened the envelope and assigned the participant to the relevant group. Immediately, the instructors transferred the allocation result to the laboratory team, keeping gynaecologists and participants unaware of the result until the embryo transfer.

Stimulation protocol, embryo transfer and luteal-phase support

Pituitary downregulation was attainable by subcutaneous GnRH-a (Decapeptyl 0.1 mg, Ferring, Saint-Prex, Switzerland) from Day 21 of the cycle preceding ICSI. On Day 2 of the ICSI cycle, women received 225–300 IU of rFSH (Gonal-F, Merck Serono) and hMG injections (Menogon, Ferring) with a 2:1 ratio, with the dose adjusted according to the response. When three follicles measured 18 mm diameter, final maturation was induced by 10 000 IU human chorionic gonadotropin (hCG, Choriomon, IBSA, Lugano, Switzerland). Oocytes were collected 37 h later and the metaphase II (MII) were inseminated by ICSI (Palermo et al., 1992). On Day 5, specified clinicians followed a standard protocol, transferring one to two blastocysts for each woman. For luteal phase support, women began intramuscular progesterone (100-mg, Prontogest, IBSA) from the day after oocyte retrieval to the 12th week of gestation, unless the biochemical pregnancy test was negative.

Sperm preparation, oocyte retrieval, denudation and ICSI

Fresh, frozen–thawed and surgically retrieved sperm were prepared at room temperature by density-gradient centrifugation (PureSperm, Nidacon, Mölndal, Sweden) (Björndahl et al., 2010). Samples were then washed and incubated in AllGrade Wash medium (LifeGlobal, Guilford, CT, USA). Follicles were aspirated into, and handled in, Global HEPES medium (LifeGlobal) at 37°C, using a tube warmer. With chemical and mechanical tools (80 IU hyaluronidase, LifeGlobal; Denudation Pipette, Vitrolife, Göteborg, Sweden), oocytes were denuded at 39 h after hCG. Immediately, ICSI was performed at 37°C, in Global HEPES (LifeGlobal).

Culture protocol and embryo scoring

In the two centres and across the study, laboratory variables were controlled using: only Minc-1000 incubators (Cook, USA); Global Total medium (LifeGlobal); $37\pm0.1\,^{\circ}\text{C}$ culture temperature verified by a thermocouple (Chino, Tokyo, Japan); $7.25\pm0.02\,\text{pH}$ validated weekly by a blood gas analyser (Epocal, Ottawa, Canada); group culture of three oocytes; 15 μ l medium droplet in micro-droplet dish (Vitrolife) overlaid by 5 ml oil (NidOil, Nidacon); continuous culture from Day 0 through Day 5/6, without medium renewal; premixed gas (7.5% CO $_2$, 5% O $_2$ and 87.5% N $_2$); and the same batches of disposables. Fertilization check and embryo grading were performed on Days I, 3 and 5 of culture (Alpha Scientists in Reproductive Medicine and ESHRE Special Interest Group of Embryology, 2011).

Interventions

Immediately after ICSI, AOA was performed and comprised incubation for I h in 10 mM SrCl $_2$ (Sigma, St Louis, MO, USA) diluted in Global Total medium or incubation for 20 min in a commercial calcimycin (Cult-active Ca-lonophore; Gynemed, Lensahn, Germany). AOA droplets were 200 μ l each in 60-mm dishes (353652, BD Falcon, USA) overlaid with 10 ml oil (NidOil) and were equilibrated overnight at 37°C and in an atmosphere of 7.5% CO $_2$, 5% O $_2$, and 87.5% N $_2$ (pH 7.25 \pm 0.02).

Study outcomes

The primary endpoint was clinical pregnancy rate (positive heartbeat on ultrasonography at ≥ 4 weeks after ET). Secondary endpoints included the rates of (i) fertilization (fertilized oocytes with two pronuclei per MII oocyte injected); (ii) high-quality embryos on Day 3 (seven or eight blastomeres of equal-size and <10% fragmentation by volume); (iii) formed blastocyst on Day 5/6 from fertilized oocytes; (iv) high-quality blastocysts (blastocysts $\geq 3.1.1$ grade, per fertilized oocyte); (v) biochemical pregnancy (positive b-hCG at ≥ 14 days following ET); (vi) ongoing pregnancy (pregnancy following week 12 of gestation); (vii) multiple pregnancy (\geq two foetuses with a heartbeat); (viii) implantation (sacs with a heartbeat); (ix) miscarriage (pregnancy loss before gestational week 12); and (x) live birth (viable neonate at \geq 30 weeks of gestation).

Statistical analysis

Previous cycles of low fertilization for women younger than 40 years of age showed a 16% clinical pregnancy rate (Murugesu et al., 2017). This trial was powered to detect a 14% rise in pregnancy rate (16–30%) with 80% power at 5% significance level, requiring 342 participants (114 per arm) to undergo SrCl₂ AOA, calcimycin AOA or ICSI alone and reach the primary endpoint. An alternative was to enrol up to 450 women, to allow for any significant dropout. The independent data and safety committees performed an interim analysis at 343 participants, advising that no more enrolment is needed. Data were analysed using binomial logistic regression with log link, one-way ANOVA, Fisher's exact and Chi-square tests where appropriate. Between-clinician and between-embryologist differences were controlled by Fleiss' Kappa analysis (McHugh, 2012). Results are reported as an intentionto-treat and per indication analyses and are presented as an absolute rate difference (ARD), odds ratio (OR) with 95% CI) or means with SD where appropriate. P < 0.05 was regarded as statistically significant. All analyses were performed using SPSS (version 22, IBM. Armonk, NY, USA).

Results

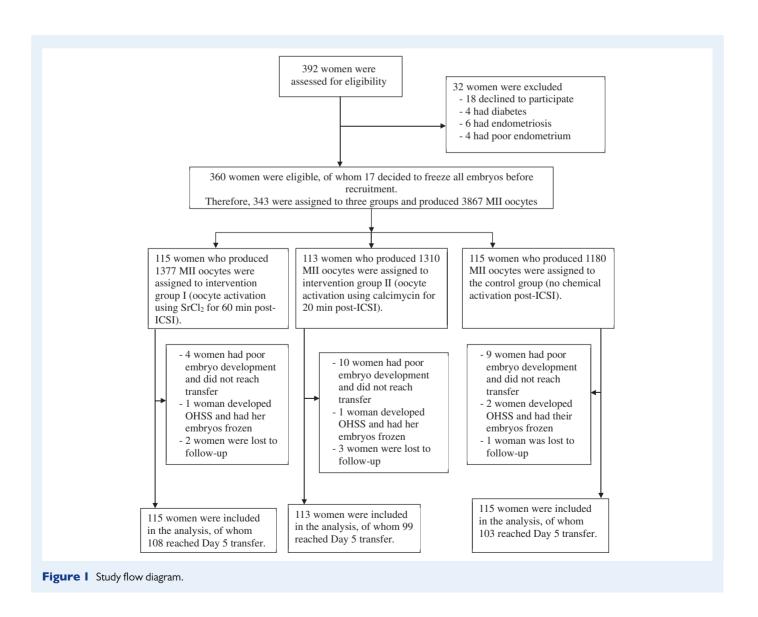
Between 27 April 2015 and 17 January 2016, we recruited 343 participants ($SrCl_2 AOA = 115$, calcimycin AOA = 113 and ICSI alone = 115)

after excluding 49 (Fig. I). Patient and cycle characteristics were similar in the three groups (Table I).

Overall (Table II), the clinical pregnancy rate was significantly higher with SrCl₂ and calcimycin AOA than with ICSI alone (49, 42 and 27%; ARD 22, 95% CI: 9-33 and ARD 16, 95% CI: 3-27). SrCl₂ and calcimycin AOA also significantly increased the biochemical pregnancy rate compared with ICSI alone. The ongoing pregnancy rate was considerably higher with SrCl₂ and calcimycin AOA than with ICSI alone (42, 36 and 23%; ARD 19, 95% CI: 7-30 and ARD 14, 95% CI: 2-25). The implantation rate was significantly also higher with SrCl₂ and calcimycin AOA than with ICSI alone (32, 28 and 18%; ARD 14, 95% CI: 5-23 and ARD 10, 95% CI: 2-19). These results culminated in significantly higher rates of live birth for SrCl₂ and calcimycin AOA than for ICSI alone (40, 33 and 18%; ARD 22, 95% CI: 10-33 for SrCl₂ and ARD 14, 95% CI: 3-25). The rates of cancelled transfer, chemical pregnancy, miscarriage and multiple pregnancy were similar between the groups. Both SrCl₂ and calcimycin AOA resulted in similar rates of ongoing pregnancy, implantation and live birth (Supplementary Table SV).

Among couples with previous low fertilization, SrCl₂ AOA significantly improved the clinical pregnancy rate compared with ICSI alone (ARD 35, 95% CI: 15–52; Supplementary Table SI) or calcimycin AOA (ARD 30, 95% CI: 10–47; Supplementary Table SV), whereas calcimycin AOA and ICSI alone were comparable (ARD 5, 95% CI:–14 to 24; Supplementary Table SI). SrCl₂ AOA was superior to ICSI alone regarding the rates of biochemical pregnancy, ongoing pregnancy and implantation, resulting in a higher live birth rate (ARD 37, 95% CI: 16–53; Supplementary Table SI). SrCl₂ AOA was also superior to calcimycin AOA, resulting in higher rates of ongoing pregnancy and live birth (ARD 22, 95% CI: 2–40 and ARD 26, 95% CI: 6–44; Supplementary Table SV). Only the implantation rate was significantly higher with calcimycin AOA than with ICSI alone (Supplementary Table SI).

Among couples of male-factor infertility with abnormal sperm, calcimycin AOA significantly improved the clinical pregnancy rate compared with ICSI alone (ARD 22, 95% CI: 6–36; Supplementary Table SII), whereas SrCl₂ AOA and ICSI alone were comparable (ARD 12, 95% CI:–3 to 26). Calcimycin AOA was superior to ICSI alone regarding the rates of biochemical pregnancy, clinical pregnancy



1640 Fawzy et al.

Table I Baseline characteristics by trial group.

	Artificial oocy	te activation (AOA) gro	oups	
	SrCl ₂ AOA	Calcimycin AOA	Control (no AOA)	
	(n = 115)	(n=113)	(n = 115)	
Age (years): mean ± SD	31 ± 5.1	32.4 ± 5.2	32 ± 5.2	
BMI (kg/m 2): mean \pm SD	28.4 ± 2.7	28.4 ± 2.7	28.3 ± 2.7	
Duration of infertility (years): mean \pm SD	6 ± 2.7	6 ± 2.5	6 ± 2.2	
Basal FSH (IU/L): mean \pm SD	6 ± 1.2	6 ± 1.3	6 ± 2	
Antral follicle count: mean \pm SD	15.0 ± 2.2	14.0 ± 2.1	14.3 ± 2.3	
Total FSH/hMG: mean ± SD	2245 ± 244	2294 ± 195	2264 ± 227	
Impaired-fertilization cycles:				
Previous fertilization failure: n (%)	7 (6)	5 (4)	7 (6)	
Previous low fertilization ($<$ 30%) with average oocyte quality: n (%)	14 (12)	13 (12)	12 (11)	
Previous low fertilization ($<$ 30%) associated with impaired oocyte quality: n (%)	26 (23)	27 (23)	24 (21)	
Male-factor infertility cycles:				
Frozen–thawed sperm of affected-morphology samples: n (%)	22 (19)	24 (21)	23 (20)	
Surgically retrieved sperm of affected-morphology samples: n (%)	19 (17)	21 (19)	22 (19)	
Globozoospermia: n (%)	4 (3)	3 (3)	5 (4)	
100% tapered-head sperm: n (%)	23 (20)	20 (18)	22 (19)	
Number of oocytes collected: mean \pm SD	14 ± 9.4	13 ± 9.1	12 ± 8.0	
Number of mature oocytes: mean \pm SD	13 ± 6.5	12 ± 8.0	II ± 6.2	
Number of embryos transferred: mean \pm SD	1.7 ± 0.5	1.6 ± 0.5	1.6 ± 0.5	

 $Comparison \ of the \ differences \ between \ the \ three \ groups \ was \ performed \ by \ one-way \ ANOVA \ and \ the \ chi-square \ test \ where \ appropriate.$

and ongoing pregnancy, implantation, live birth and live-birth implantation, whereas $SrCl_2$ AOA and ICSI alone were comparable, as were $SrCl_2$ and calcimycin AOA were comparable (Supplementary Tables SII and SV).

SrCl₂ AOA resulted in significantly greater rates of fertilization, top-quality embryo and compaction on Day 3, blastocyst development, high-quality blastocysts and cryopreservation compared with ICSI alone (all P < 0.0001; Table III). Calcimycin AOA also resulted in significantly higher rates of fertilization, blastocyst formation and cryopreservation compared with ICSI alone (P = 0.003, P = 0.026 and P < 0.0001; Table III). The embryological outcomes for the subgroup analyses are shown in Supplementary Tables SIII and SIV.

Discussion

To our knowledge, this RCT is the first to compare $SrCl_2$ and calcimycin AOA after ICSI to ICSI alone for cycles of previous low fertilization or male-factor infertility. Overall, this trial demonstrated significantly higher rates of clinical pregnancy and live birth for either $SrCl_2$ or calcimycin AOA after ICSI compared with ICSI alone, whereas $SrCl_2$ and calcimycin AOA were comparable. Our findings concur with a recent meta-analysis (Murugesu et al., 2017).

Among couples with previous ICSI cycles of low fertilization, SrCl₂ AOA was more effective than ICSI alone, whereas calcimycin AOA was comparable to ICSI alone. In this subgroup, neither sperm entry nor ICSI appears sufficient to initiate the oocyte activation. This is likely to relate the activation defects to the oocyte (Miyara et al., 2003;

Ebner et al., 2006; Xing et al., 2011; Kilani and Chapman, 2014). Reports suggest that ICSI with SrCl₂ AOA appears to stimulate pulsatile waves of Ca²⁺ oscillation from the endoplasmic reticulum during fertilization and further development (Kline and Kline, 1992; Kishikawa et al., 1999; Kim et al., 2014; Yeste et al., 2016). Although SrCl₂ AOA did not show signs of Ca²⁺ oscillation or parthenogenetic division for in-vitro matured oocytes (Lu et al., 2018), this situation may differ from the MII oocytes undergoing ICSI with AOA. Therefore, given its potential to improve ICSI outcomes, further studies are required to identify whether SrCl₂ AOA activates the oocyte by Ca²⁺ oscillation or other mechanisms.

For calcimycin AOA and previous ICSI cycles of low fertilization, our findings showed that calcimycin AOA and ICSI alone were comparable. Our results can be explained as calcimycin might be less effective than ${\rm SrCl_2}$ because it induces a single ${\rm Ca^{2+}}$ wave that is not converted into oscillation. Although our observations contradict a recent meta-analysis (Murugesu et al., 2017) yet concur with an earlier one (Sfontouris et al., 2015), the conclusions from the meta-analyses seem to be not substantial due to the heterogenous designs of their studies. Therefore, calcimycin AOA appears to be not beneficial for ICSI cycles with previous impaired fertilization, but normal sperm parameters.

Among couples with male-factor infertility, calcimycin AOA was superior to $SrCl_2$ AOA or ICSI alone. It is likely that when the problem results from male-factor infertility, Ca^{2+} oscillation is functional in the oocyte, but it has insufficient stimulation by the ICSI alone with affected sperm (Vanden Meerschaut et al., 2013; Ferrer-Buitrago et al., 2018). Calcimycin AOA provides additional Ca^{2+} influx from the

Table II Clinical Outcomes in the trial groups by intention-to-treat analysis.

	Artificial oocyte activation (AOA) groups (%)				Absolute rate difference, percentage points (95% CI)		Odds ratio (95% CI)	
	SrCl ₂ AOA (n = 115)	Calcimycin AOA (n = 113)	Control (no AOA) (n = 115)	SrCl ₂ to control	Calcimycin to control	SrCl ₂ to control	Calcimycin to control	
Cancelled transfer rate	7/115 (6)	14/113 (12)	12/115 (10)	-4.3 (-12 to 3)	2 (-6.5 to 11)	0.56 (0.21–1.47) P = 0.23	1.21 (0.54–2.75) P = 0.64	
Biochemical pregnancy rate	62/115 (54)	61/113 (54)	39/115 (34)	20 (7–32)	20 (7–32)	2.28 (1.34–3.88) P = 0.002	2.29 (1.34–3.90) P = 0.002	
Clinical pregnancy rate	56/115 (49)	48/113 (42)	31/115 (27)	22 (9–33)	16 (3–27)	2.57 (1.48–4.46) P = 0.0007	2.0 (1.15–3.49) P = 0.014	
Chemical pregnancy rate	6/115 (5)	13/113 (11)	8/115 (7)	-1.7 (-8 to 5)	5 (-3 to 13)	0.74 $(0.25-2.19)$ $P = 0.58$	1.74 (0.69–4.37) P = 0.24	
Ongoing pregnancy rate	48/115 (42)	41/113 (36)	26/115 (23)	19 (7–30)	14 (2–25)	2.45 (1.38–4.35) P = 0.0019	1.95 (1.10–3.49) P = 0.023	
Miscarriage rate	14/115 (12)	20/113 (18)	13/115 (11)	0.9 (-8 to 9.4)	6.4 (-2.9 to 15.7)	1.09 $(0.49-2.43)$ $P = 0.84$	1.69 (0.79–3.58) P = 0.17	
Multiple pregnancy rate	5/115 (4)	4/II3 (4)	2/115 (2)	2.6 (-2.4 to 8.2)	1.8 (-3.1 to 7.2)	2.57 $(0.49-13.5)$ $P = 0.45$	2.07 (0.37–11.6) P = 0.44	
Implantation rate	61/190 (32)	52/184 (28)	33/182 (18)	14 (5–23)	10 (2–19)	2.14 $(1.31-3.47)$ $P = 0.0019$	1.78 (1.08–2.92) P = 0.022	
Live birth rate	46/115 (40)	37/113 (33)	21/115 (18)	22 (10–33)	14 (3–25)	2.98 (1.63–5.45) P = 0.0002	2.18 (1.18–4.03) P = 0.012	
Live-birth-implantation rate	46/190 (24)	37/184 (20)	21/182 (12)	13 (5–20)	9 (I-I6)	2.45 (1.39–4.30) P = 0.0015	1.93 (1.08–3.45) P = 0.025	

The logistic regression analysis verified that there was no association between the primary outcome and the confounding variables including each centre's cycle characteristics, except for $SrCl_2$ AOA and Calcimycin AOA. Chemical pregnancy indicates a positive pregnancy test with no gestational sac identified 15 days after the test.

surrounding medium, boosting the initiation of Ca^{2+} oscillation (Yeste et al., 2016). In this situation, $SrCl_2$ may have less effect because the defect is not with the Ca^{2+} oscillation machinery itself, just the initiation of oscillation. Our observations concur with a recent meta-analysis (Murugesu et al., 2017) as it also included cycles with affected sperm function. Therefore, calcimycin AOA appears promising for ICSI cycles with abnormal sperm morphology.

Regarding embryo development, AOA with $SrCl_2$ or calcimycin gave significantly better results than conventional ICSI in the overall and subgroup analyses. The effect sizes for $SrCl_2$ were larger than for calcimycin. Both AOA protocols were superior to ICSI alone for the blastocyst cryopreservation rate, suggesting that, besides the clinical improvements observed, AOA might improve the cumulative pregnancy rate. The improvements in embryo development could have resulted from restoration of Ca^{2+} oscillation to physiological or supraphysiological status. However, the mechanisms by which this restoration might have occurred are beyond this study, and whether

possible supraphysiological Ca²⁺ oscillation might have epigenetic effects remains a subject for further investigation.

Previous studies (Murugesu et al., 2017) have shown that the clinical pregnancy rate is 16% for ICSI cycles with previous low fertilization. This rate appears relatively lower than that we observed in our facilities and we reported for this group. In our study, the rigorous control for the laboratory and clinical confounders, the relatively young age of women included (~32), and the short time of conduct seemed to be reflected in the observed outcomes in the three groups.

To limit the sample size required, the margin of superiority was set at 14%, which is a relatively high. This margin was considered because although calcimycin and $SrCl_2$ appear safe, this safety has not been proved unequivocally (Yeste et al., 2016).

Our study has several strengths. It represents, to our knowledge, the first RCT with an appropriate sample size to compare SrCl₂ to calcimycin for AOA. Although clinical pregnancy was the primary outcome, ongoing pregnancy and live birth rates were also assessed. Our

Table III Embryological outcomes in the trial groups.

	Artificial oocyte activation (AOA) groups (%)			Absolute rate difference, percentage points (95% CI)		Odds ratio (95% CI)	
	SrCl ₂ AOA (n = 115)	Calcimycin AOA (n = 113)	Control (no AOA) (n = 115)	SrCl₂ to control	Calcimycin to control	SrCl ₂ to control	Calcimycin to control
Maturation rate: MII oocytes/collected oocytes	1377/ 1581 (87)	1310/1483 (88)	1180/1329 (89)	-1.7 (-4.1 to 0.7)	-0.45 (-2.8 to 1.9)	0.85 (0.68–1.07) P = 0.16	0.96 (0.76–1.21) P = 0.71
Fertilization rate: 2PN oocytes/injected MII oocytes	1168/ 1377 (85)	874/1310 (67)	720/1180 (61)	24 (20–27)	5.7 (1.9–9.5)	3.57 (2.96–4.31) <i>P</i> < 0.0001	1.28 (1.09–1.51) P = 0.003
Cleavage rate: cleaved embryos/fertilized oocytes	1134/ 1168 (97)	854/874 (98)	702/720 (98)	-0.4 (-1.9 to 1.2)	0.2 (-1.3 to 1.8)	0.86 (0.48–1.53) P = 0.597	1.09 (0.5–72.09) P = 0.78
Top-quality Day 3 embryos/fertilized oocytes	1021/ 1168 (87)	513/874 (59)	413/720 (57)	30 (26–34)	1.3 (3.5–6.0)	5.16 (4.11–6.48) <i>P</i> < 0.0001	1.06 (0.87–1.29) P = 0.59
Compaction rate: compacted Day 3 embryos/fertilized oocytes	518/1168 (44)	212/874 (24)	189/720 (26)	18 (14–22)	-2 (-6.3 to 2.3)	2.24 (1.83–2.74) <i>P</i> < 0.0001	0.89 (0.72–1.13) P = 0.36
Blastocyst formation rate: blastocysts/fertilized oocytes	722/1168 (62)	447/874 (51)	328/720 (46)	17 (12–21)	5.6 (0.7–10.5)	1.93 (1.60–2.34) P < 0.0001	1.25 (1.03–1.52) P = 0.026
High-quality blastocysts/fertilized oocytes	475/1168 (41)	232/874 (27)	195/720 (27)	14 (9–18)	-0.5 (-0.5 to 0.4)	1.85 (1.51–2.26) P < 0.0001	0.97 (0.78–1.22) P = 0.806
Cryopreservation rate: vitrified blastocysts/fertilized oocytes	518/1168 (44)	251/874 (29)	133/720 (18)	26 (22–30)	6. I (6–I4)	3.52 (2.82–4.39) P < 0.0001	1.7 (1.40–2.26) P < 0.0001

2PN, two pronuclei; MII, metaphase II.

findings provide insight into the relative suitability of $SrCl_2$ and calcimycin for AOA for particular causes of infertility, which can improve the outcomes for subgroups of patients who experience poor outcomes of ICSI.

Our study also has several limitations. The open-label trial design might have introduced bias, although randomization, at least partly, addressed this possibility. The study did not include a longitudinal follow-up period, so further evidence regarding the safety of AOA is still required. The study included couples with different indications for ICSI, rather than focusing on a single subgroup, but post hoc analyses were performed to determine the particular effects of AOA in these subgroups.

In conclusion, in our study population, ICSI followed by AOA with SrCl₂ or calcimycin was associated with significantly better embryological and clinical outcomes than ICSI alone. Calcimycin AOA appears effective in overcoming defects related to abnormal sperm morphology, whereas SrCl₂ AOA appears to improve outcomes for ICSI cycles characterized by previous impaired fertilization. In consideration of the current lack of knowledge regarding the long-term safety and transgenerational effects of AOA, we do not recommend this treatment for the general ICSI population. Instead, we recommend that more studies are performed, to further elucidate the mechanisms that underlie our findings, in particular those related to SrCl₂ AOA. We also recommend that a multicentre trial is conducted to identify the cumulative live birth rate and to assess the health of the offspring as primary outcomes. Until these studies have been conducted, we recommend that AOA should be used cautiously, and the advantages and disadvantages regarding its use should be weighed in each case.

Supplementary data

Supplementary data are available at Human Reproduction online.

Acknowledgements

The authors thank the IbnSina IVF laboratory team for their dedicated efforts during the study, especially Dr Ahmed AlAboudy and Dr Mostafa Ail.

Authors' roles

M.F. created the concept and design of the study, was the primary investigator and wrote the article. M.E. participated in the investigation as well as the collection, analysis and interpretation of the data. All other authors participated in critically reviewing the concept upon which the study was conducted and participated in the investigation. All authors critically revised the manuscript and gave their final approval of the version for publication.

Funding

The research was not funded by any agency.

Conflict of interest

The authors have no conflicts of interest to declare.

REFERENCES

- Alpha Scientists in Reproductive Medicine and ESHRE Special Interest Group of Embryology. Alpha Scientists in Reproductive Medicine and ESHRE Special Interest Group of Embryology. The Istanbul consensus workshop on embryo assessment: proceedings of an expert meeting. *Hum Reprod* 2011;**26**:1270–1283.
- Amdani SN, Jones C, Coward K. Phospholipase C zeta (PLCζ): oocyte activation and clinical links to male factor infertility. *Adv Biol Regul* 2013; **53**:292–308
- Björndahl L, Mortimer D, Barratt CLR, Castilla JA, Menkveld R, Kvist U, Alvarez JG, Haugen TB. A Practical Guide to Basic Laboratory Andrology. Cambridge, UK: Cambridge University Press, 2010.
- Chen J, Qian Y, Tan Y, Mima H. Successful pregnancy following oocyte activation by strontium in normozoospermic patients of unexplained infertility with fertilisation failures during previous intracytoplasmic sperm injection treatment. *Reprod Fertil Dev* 2010; **22**:852–855.
- Cheng D, Li J, Guo C-C, Xiong C-L. [Failed fertilization after ICSI: causes and countermeasures]. Zhonghua Nan Ke Xue 2011;17:1131–1134.
- Clift D, Schuh M. Restarting life: fertilization and the transition from meiosis to mitosis. *Nat Rev Mol Cell Biol* 2013;**14**:549–562.
- Combelles CMH, Morozumi K, Yanagimachi R, Zhu L, Fox JH, Racowsky C. Diagnosing cellular defects in an unexplained case of total fertilization failure. *Hum Reprod* 2010;**25**:1666–1671.
- De Vos A, Van De Velde H, Joris H, Verheyen G, Devroey P, Van Steirteghem A. Influence of individual sperm morphology on fertilization, embryo morphology, and pregnancy outcome of intracytoplasmic sperm injection. *Fertil Steril* 2003;**79**:42–48.
- Ducibella T, Huneau D, Angelichio E, Xu Z, Schultz RM, Kopf GS, Fissore R, Madoux S, Ozil JP. Egg-to-embryo transition is driven by differential responses to Ca2+ oscillation number. *Dev Biol* 2002;**250**:280–291.
- Ebner T, Montag M, Montag M, Van Der Ven K, Van Der Ven H, Ebner T, Shebl O, Oppelt P, Hirchenhain J, Krüssel J et al. Live birth after artificial oocyte activation using a ready-to-use ionophore: a prospective multicentre study. Reprod Biomed Online 2015;30:359–365.
- Ebner T, Moser M, Tews G. Is oocyte morphology prognostic of embryo developmental potential after ICSI? *Reprod Biomed Online* 2006; 12: 507–512.
- Eldar-Geva T, Brooks B, Margalioth EJ, Zylber-Haran E, Gal M, Silber SJ. Successful pregnancy and delivery after calcium ionophore oocyte activation in a normozoospermic patient with previous repeated failed fertilization after intracytoplasmic sperm injection. *Fertil Steril* 2003;**79**: 1656–1658.
- Escoffier J, Yassine S, Lee HC, Martinez G, Delaroche J, Coutton C, Karaouzéne T, Zouari R, Metzler-Guillemain C, Pernet-Gallay K et al. Subcellular localization of phospholipase Cz in human sperm and its absence in DPY19L2-deficient sperm are consistent with its role in oocyte activation. Mol Hum Reprod 2014;21:157–168.
- Ferrer-Buitrago M, Dhaenens L, Lu Y, Bonte D, Meerschaut F Vanden, Sutter P De, Leybaert L, Heindryckx B. Human oocyte calcium analysis predicts the response to assisted oocyte activation in patients experiencing fertilization failure after ICSI. *Hum Reprod* 2018;33: 416–425.
- Flaherty SP, Payne D, Matthews CD. Fertilization failures and abnormal fertilization after intracytoplasmic sperm injection. *Hum Reprod* 1998;**13**: 155–164.
- Greco E, Scarselli F, Fabozzi G, Colasante A, Zavaglia D, Alviggi E, Litwicka K, Varricchio MT, Minasi MG, Tesarik J. Sperm vacuoles negatively affect outcomes in intracytoplasmic morphologically selected sperm injection in terms of pregnancy, implantation, and live-birth rates. *Fertil Steril* 2013;**100**:379–385.

1644 Fawzy et al.

Heindryckx B, De Gheselle S, Gerris J, Dhont M, De Sutter P. Efficiency of assisted oocyte activation as a solution for failed intracytoplasmic sperm injection. Reprod Biomed Online 2008; 17:662–668.

- Heindryckx B, Van der Elst J, De Sutter P, Dhont M. Treatment option for sperm- or oocyte-related fertilization failure: assisted oocyte activation following diagnostic heterologous ICSI. Hum Reprod 2005;20: 2237–2241.
- Horner VL, Wolfner MF. Transitioning from egg to embryo: triggers and mechanisms of egg activation. Dev Dyn 2008;237:527–544.
- Kashir J, Heindryckx B, Jones C, De Sutter P, Parrington J, Coward K. Oocyte activation, phospholipase C zeta and human infertility. Hum Reprod Update 2010;16:690–703.
- Kilani S, Chapman MG. Meiotic spindle normality predicts live birth in patients with recurrent in vitro fertilization failure. Fertil Steril 2014;101:403–6.
- Kim JW, Kim SD, Yang SH, Yoon SH, Jung JH, Lim JH. Successful pregnancy after SrCl₂ oocyte activation in couples with repeated low fertilization rates following calcium ionophore treatment. Syst Biol Reprod Med 2014; **60**:177–182.
- Kishikawa H, Wakayama T, Yanagimachi R. Comparison of oocyteactivating agents for mouse cloning. *Cloning* 1999;1:153–159.
- Kissin DM, Jamieson DJ, Barfield WD. Monitoring health outcomes of assisted reproductive technology. *N Engl J Med* 2014;**371**:91–93.
- Kline D, Kline JT. Repetitive calcium transients and the role of calcium in exocytosis and cell cycle activation in the mouse egg. *Dev Biol* 1992;**149**: 80–89.
- Lu Y, Gao H, Li B, Zheng Y, Ye Y, Qian Y, Xu C, Huang H, Jin F. Different sperm sources and parameters can influence intracytoplasmic sperm injection outcomes before embryo implantation. *J Zhejiang Univ Sci B* 2012; **13**:1–10.
- LuYReddyRBuitragoMFVander Jeught MNeupaneJDe VosWHVan Den AbbeelELiermanSDe SutterPHeindryckxB2018Strontium fails to induce Ca2+ release and activation in human oocytes despite the presence of functional TRPV3 channels
- McHugh ML. Interrater reliability: the kappa statistic. *Biochem Med* 2012; **22**:276–282.
- Miyara F, Aubriot FX, Glissant A, Nathan C, Douard S, Stanovici A, Herve F, Dumont-Hassan M, LeMeur A, Cohen-Bacrie P et al. Multiparameter analysis of human oocytes at metaphase II stage after IVF failure in nonmale infertility. Hum Reprod 2003; 18:1494–1503.
- Miyazaki S, Shirakawa H, Nakada K, Honda Y. Essential role of the inositol 1,4,5-trisphosphate receptor/Ca2+ release channel in Ca2+ waves and Ca2 + oscillations at fertilization of mammalian eggs. *Dev Biol* 1993;**158**:62–78.
- Miyazaki S, Yuzaki M, Nakada K, Shirakawa H, Nakanishi S, Nakade S, Mikoshiba K. Block of Ca2+ wave and Ca2+ oscillation by antibody to the inositol 1,4,5-trisphosphate receptor in fertilized hamster eggs. *Science* 1992;**257**:251–255.
- Montag M, Köster M, van der Ven K, Bohlen U, van der Ven H. The benefit of artificial oocyte activation is dependent on the fertilization rate in a previous treatment cycle. *Reprod Biomed Online* 2012;**24**:521–526.
- Murugesu S, Saso S, Jones BP, Bracewell-Milnes T, Athanasiou T, Mania A, Serhal P, Ben-Nagi J. Does the use of calcium ionophore during artificial oocyte activation demonstrate an effect on pregnancy rate? A meta-analysis. *Fertil Steril* 2017;108:468–482.e3.

- Nomikos M, Kashir J, Swann K, Lai FA. Sperm PLCζ: from structure to Ca2+ oscillations, egg activation and therapeutic potential. *FEBS Lett* 2013:**587**:3609–3616.
- Palermo G, Joris H, Devroey P, Van Steirteghem AC. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. *Lancet* 1992;340:17–18.
- Ridgway EB, Gilkey JC, Jaffe LF. Free calcium increases explosively in activating medaka eggs. *Proc Natl Acad Sci USA* 1977;**74**:623–627.
- Rybouchkin AV, De Sutter P, Van Der Straeten F, Dhont M, Quatacker J. Fertilization and pregnancy after assisted oocyte activation and intracytoplasmic sperm injection in a case of round-headed sperm associated with deficient oocyte activation capacity. Fertil Steril 1997;68:1144–1147.
- Saunders CM, Larman MG, Parrington J, Cox LJ, Royse J, Blayney LM, Swann K, Lai FA. PLC zeta: a sperm-specific trigger of Ca(2+) oscillations in eggs and embryo development. *Development* 2002;**129**:3533–3544.
- Sfontouris IA, Nastri CO, Lima MLS, Tahmasbpourmarzouni E, Raine-Fenning N, Martins WP. Artificial oocyte activation to improve reproductive outcomes in women with previous fertilization failure: a systematic review and meta-analysis of RCTs. *Hum Reprod* 2015;**30**:1831–1841.
- Steinhardt R, Zucker R, Schatten G. Intracellular calcium release at fertilization in the sea urchin egg. Dev Biol 1977;58:185–196.
- Steptoe PC, Edwards RG. Birth after the reimplantation of a human embryo. *Arch Pathol Lab Med* 1992;**116**:321.
- Tesarik J, Rienzi L, Ubaldi F, Mendoza C, Greco E. Use of a modified intracytoplasmic sperm injection technique to overcome sperm-borne and oocyte-borne oocyte activation failures. *Fertil Steril* 2002;**78**:619–624.
- Vanden Meerschaut F, Leybaert L, Nikiforaki D, Qian C, Heindryckx B, De Sutter P. Diagnostic and prognostic value of calcium oscillatory pattern analysis for patients with ICSI fertilization failure. Hum Reprod 2013;28:87–98.
- Vanden Meerschaut F, Nikiforaki D, De Gheselle S, Dullaerts V, Van Den Abbeel E, Gerris J, Heindryckx B, De Sutter P. Assisted oocyte activation is not beneficial for all patients with a suspected oocyte-related activation deficiency. *Hum Reprod* 2012;**27**:1977–1984.
- Vanden Meerschaut F, Nikiforaki D, Heindryckx B, De Sutter P. Assisted oocyte activation following ICSI fertilization failure. Reprod Biomed Online 2014;28:560–571.
- Wassarman PM, Jovine L, Litscher ES. A profile of fertilization in mammals. Nat Cell Biol 2001; 3:E59–E64.
- Xing X, Zhao H, Li M, Sun M, Li Y, Chen ZJ. Morphologically abnormal oocytes not capable of fertilization despite repeated strategies. *Fertil Steril* 2011;**95**:2435.e5–7.
- Yanagida K, Morozumi K, Katayose H, Hayashi S, Sato A. Successful pregnancy after ICSI with strontium oocyte activation in low rates of fertilization. *Reprod Biomed Online* 2006; **13**:801–806.
- Yelumalai S, Yeste M, Jones C, Amdani SN, Kashir J, Mounce G, Silva SJM Da, Barratt CL, McVeigh E, Coward K. Total levels, localization patterns, and proportions of sperm exhibiting phospholipase C zeta are significantly correlated with fertilization rates after intracytoplasmic sperm injection. *Fertil Steril* 2015;**104**:561–568.e4.
- Yeste M, Jones C, Amdani SN, Patel S, Coward K. Oocyte activation deficiency: a role for an oocyte contribution? Hum Reprod Update 2016;22: 23–47.