

1 **STATE OF THE ART OF UNBOUND CEFTRIAXONE AS A PHARMACODYNAMIC TOOL: ARE WE READY FOR**
2 **ITS IMPLEMENTATION IN CLINICAL PRACTICE?**

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26 **Running head:** Unbound ceftriaxone in clinical practice

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28 **Abstract:**

29 **Background:** Ceftriaxone is pivotal in treating severe infections; however, modeling unbound plasma
30 ~~clearance (CL) and distribution (Vd) is challenging due to the lack of reliable data. Furthermore, the pharmacokinetics (PK) of ceftriaxone are highly variable, and the unbound fraction (fu) is not well characterized.~~

31 **Objectives:** This study aimed to (1) ~~(1)~~ predict CEF_u from CEF_{tot}, (2) ~~(2)~~ determine optimal thresholds for CEF_{tot}
32 trough concentration in plasma, (3) perform an external validation of published models, and ~~(3)~~ (4) analyze
33 factors influencing CEF_{tot} trough concentration and the probability of target attainment (PTA) ~~in a retrospective cohort.~~

34 **Methods:** CEF_u predictions based on CEF_{tot} were ~~assessed/evaluated~~ using ~~previously published models previously published in the literature~~, considering
35 both normal albumin levels (concentrations >35 g/L) and hypoalbuminemia (<20 g/L). Using the minimum inhibitory concentration (MIC) of 1 mg/L, optimal CEF_u thresholds for
36 MIC of 1 mg/L were calculated to ~~obtain/achieve~~ CEF_u plasma concentrations with fT > 1xMIC 100% and fT > 4xMIC 100%.

37 External validation was conducted using prospective data (62 samples). Retrospective data, comprising 408
38 CEF_{tot} and 222 patients, were ~~extracted from the Clinical Data Warehouse~~ analyzed to ~~deeply identify~~ significant predictors of CEF_u trough concentrations and
39 PTA based on ~~analyzed the evaluated~~ models.

40 **Results:** ~~Nine publications were retained for CEF_u modeling.~~ Optimal CEF_u trough concentration thresholds ranged from 1.8 mg/L to 169 mg/L (1xMIC) and from
41 6.6 mg/L to 56.2 mg/L (4xMIC). External validation suggested that some published models predicted well
42 CEF_u. In the retrospective cohort, PTA varied from 94.4% to 98.7% for 1xMIC and from 66.9% to 97.3% for
43 4xMIC. Age, daily dose, albuminemia and creatininemia were significant predictors of CEF_{tot} concentration.

44 ~~Notably, combining model 1 with model 2 improved PTA compared to model 1, model 2, or model 3. The combination of model 1 and model 2 improved PTA compared to model 1 or model 2.~~

45 **Conclusions:** ~~It is imperative to subject~~ Modeling or quantifying CEF_u may enhance patient outcomes but requires standardized
46 analytical approaches and further investigation. CEF_u concentration modeling requires external validation before considering implementation in clinical practice.

INTRODUCTION:

Ceftriaxone (CEF) is a widely-used third-generation beta-lactam antibiotic in the cephalosporin class. It plays a crucial role in preventing and treating severe infections like meningitis, pneumonia, osteoarticular infections, soft tissue infections, and endocarditis. In emergency medical settings, CEF is often the preferred choice for antimicrobial therapy due to its rapid and broad-spectrum activity. (1)-¹- However, determining the optimal dosing regimen for individual patients is a challenge, primarily because of the drug's total ceftriaxone (CEF_{tot}) exhibits nonlinear pharmacokinetics (PK), in contrast to unbound ceftriaxone (CEF_u), which follows linear PK due to saturable protein binding.^{2,3} (2, 3) In cases of severe sepsis and septic shock, the PK of CEF_{tot} undergoes significant modifications, with due to decreased albumin levels, hypotension, and increased renal clearance. (4) Moreover, the pharmacodynamic (PD) properties of antibiotics and the potential changes in their PK in such critical conditions is essential for tailoring individualized dosing regimens. (5) Since CEF has a high, saturable binding affinity to plasma proteins, particularly especially albumin. Consequently, in a saturable manner, increases in total ceftriaxone (CEF_{tot}) concentration and/or decreases in albumin from normal initial plasma levels increase the fraction of unbound CEF_u. This higher CEF_u level increases leads to a higher in-apparent volume of distribution and enhanced clearance resulting in a lower overall drug exposure. (6) Such reductions that may compromise time-dependent PD target of CEF.⁻⁶

Interestingly, in the pharmacokinetic realm of CEF, the therapeutic drug monitoring (TDM) predominantly aims to assess the concentration of CEF_u, whereas however, CEF's activity is related to the unbound fraction of CEF_u, which has high variability both within and between patients, exhibiting high between-patient variability. Understanding the dynamics and implications of this variability could potentially unravel provide crucial insights into for optimizing dosing regimens and improving therapeutic outcomes.

Several research teams have endeavored to model CEF_u using diverse mathematical formulas that incorporate albumin levels/concentrations. Nevertheless, these models have been developed within specific populations, such as adult and pediatric intensive care units, and have suffered from a lack of external validation. Given the potential of CEF_u quantitation and modeling to offer new insights into PK, efficacy, and toxicity, this study's objectives are threefold/fourfold: (1) to predict CEF_u in plasma from CEF_{tot}, based on existing formula in the literature; (2) to establish optimal CEF_{tot} thresholds in plasma to achieve a predefined CEF_u/CEF_{tot} target of 1 mg/L; (3) perform an external validation of the previously published models; and (34) to scrutinize

73 predictors of $CE_{T_{1/2}}$ and the probability of target attainment (PTA), based on [therapeutic drug monitoring \(TDM\)](#) in a comprehensive
74 retrospective cohort.

75 RESULTS

76 Literature Search

77 A total of 23 publications were identified. Among these, 14 publications were excluded for the
78 following reasons: 3 were outside the scope, (7–9) 5 lacked quantitation of CEF_u (10–14) and 5 did not involve
79 modeling of CEF_u based on CEF_{tot}. (15–19) ^{15–19}. Notably, one publication met the criteria but could not be used
80 due to discrepancies in the PK parameters. (20) ^{15–19, 20–25}. As a result, a total of 9 publications were retained for CEF_u
81 modeling (Table 1). CEF_{tot} is defined as the sum of CEF_u and bound ceftriaxone (CEF_b) (Eq.1). Six of the 9
82 publications (21–26) used a non-linear protein-binding model (Eq.2) for CEF_b resulting in Eq.3. In this
83 equation, Bmax and Kd represented the maximum protein binding capacity and the dissociation constant,
84 expressed in mg/L or mM. Solving for CEF_u from Eq.3 yielded Eq.4 which was used by these authors.

$$85 \text{ CEF}_{tot} = \text{CEF}_u + \text{CEF}_b \quad (\text{Eq.1})$$

$$86 \text{ CEF}_b = \frac{Bmax \times \text{CEF}_u}{Kd + \text{CEF}_u} \quad (\text{Eq.2})$$

$$87 \text{ CEF}_{tot} = \text{CEF}_u + \frac{Bmax \times \text{CEF}_u}{Kd + \text{CEF}_u} \quad (\text{Eq.3})$$

$$88 \text{ CEF}_u = \frac{1}{2} \left((\text{CEF}_{tot} - Bmax - Kd) + \sqrt{(\text{CEF}_{tot} - Bmax - Kd)^2 + 4 \times Kd \times \text{CEF}_{tot}} \right) \quad (\text{Eq.4})$$

89 Two publications employed the calculation of the unbound fraction (fu) using either a polynomial
90 (27) ²⁷ or an exponential approach. (28) ²⁸. Additionally, one publication devised its own transformation to predict
91 CEF_u from CEF_{tot}. (29) ²⁹. For the quantitation of CEF_u, UF and equilibrium dialysis (ED) were used in 6 and 2
92 publications respectively whereas the method employed was not explicitly defined in one publication. (23) ⁴⁰. It
93 is worth noting that 8 out of the 9 formulas used albuminemia as a significant predictor of Bmax.

94 ~~Unbound ceftriaxone~~ Ceftriaxone free fraction modeling & CEF_{tot} optimal thresholds

95 The prediction of CEF ~~free fraction~~ according to CEF_{tot} is depicted in Figure 1. The Ullidmolins model
96 is distinguished by its linear relationship between CEF_u ~~free fraction~~ and CEF_{tot}. The Standing model estimated
97 the higher CEF_u ~~free fraction~~ values according to based on CEF_u, both in cases of normal albumin levels and hypoalbuminemia (Figure 1A) and
98 normal albuminemia (Figure 1B). Substantial disparities emerged for with increasing CEF_u concentrations exceeding 100 mg/L (Figure 1A).

99 However, for the models developed by Bos, Dreesen, Gijsen, Gregoire, Hartman, Heffernan and Leegwater displayed similar
100 CEF_u free fraction predictions were observed within the lower range of CEF_{tot} (from 0 to 100.75 mg/L), especially in the presence of normal
101 albuminemia levels (35 g/L) (Figure 1B). However, in hypoalbuminemia (20 g/L), differences in the predicted CEF
102 free fraction become more pronounced across the full range of predicted concentrations (Figure 1A). For
103 further analysis, a calculator for determining CEF_u concentrations from CEF_{tot} and albumin concentrations is
104 available at: <https://github.com/ThomasDufлот/Ceftriaxone-AAC>.

105 Of note, differences in predicted CEF_u became more pronounced with hypoalbuminemia (20 g/L),
106 spanning the entire range of predicted concentrations (Figure 1C and 1D). To facilitate further analysis of
107 these models, a calculator of free ceftriaxone concentrations from total concentration and albuminemia is
108 included as additional material (Supplementary files S3 – CEF_u Calculator).

109 For achieving fT > MIC 100% under normal albumin conditions (35 g/L), substantial variability was
110 observed across models, with the Ulldemolins model requiring the lowest CEF_{tot} concentration (3.3 mg/L)
111 and the Gijsen model the highest (16.9 mg/L). This variability was even more pronounced when targeting fT
112 > 4xMIC 100%: the Ulldemolins model suggested a threshold as low as 13.1 mg/L, while the Heffernan model
113 indicated a much higher concentration of 56.2 mg/L. Across models, CEF_{tot} thresholds showed marked
114 differences depending on the PD target, with the mean CEF_{tot} concentration for fT > MIC 100% at 11.4±5.3
115 mg/L, and 35.8±14.5 mg/L for fT > 4xMIC 100%. Notably, as shown in Table 2, the coefficient of variation for
116 thresholds under normal albumin conditions was 48.3%.

117 In cases of hypoalbuminemia (20 g/L), the required CEF_{tot} concentrations decreased overall, yet
118 variability across models remained high. For fT > MIC 100%, the Standing model estimated the lowest CEF_{tot}
119 threshold (1.8 mg/L), while the Gregoire model required the highest concentration (15.1 mg/L). Similarly, for
120 fT > 4xMIC 100%, the lowest CEF_{tot} concentration was 6.6 mg/L (Standing model), and the highest was 50.1
121 mg/L (Gregoire model), as outlined in Table 2. Hypoalbuminemia notably intensified between-model
122 variability, with the mean CEF_{tot} threshold concentrations averaging 7.2±4.1 mg/L for fT > MIC 100% and
123 23.4±13.0 mg/L for fT > 4xMIC 100%. The coefficient of variation increased to 62.8% under hypoalbuminemic

conditions, highlighting the complex impact of reduced albumin on target attainment and model-dependent

[illegible]

External validation and comparison of predictive performance

A total of 59 patients (26 women and 33 men) receiving ceftriaxone treatment were included in the external validation study, with 62 plasma samples collected in total. The albumin concentrations, CEF_{tot}, and CEF_u measurements spanned ranges of 18.9–37.5 g/L, 2.9–259.0 mg/L, and 0.14–94.70 mg/L, respectively.

Upon analysis, the models developed by Gregoire, Hartman, and Heffernan provided the most accurate predictions, with favorable metrics across MSE, MPE, RMSE, RMSE%, and R^2 (Table 3 and Figure 2). This result was further supported by Bland-Altman plots of signed and relative differences, which showed the lowest variability (Supplementary Figure S1). An overview of significant differences between models is provided in Supplementary Table S1.

Evaluation of CEFu prediction on a retrospective cohort of patient treated by ceftriaxone

From the retrospective cohort, a total of 408 CEF_{tot} plasma samples and 376 albumin levels were available, originating from 222 patients. Since some patients had multiple CEF_{tot} quantifications (ranging from 1 to 14 samples per patient), patients- and dosing-related variables were categorized accordingly (Table 3). The median albumin concentration was 27.0 g/L, below the normal range. Albumin concentrations ranged from a minimum of 10.4 g/L to a maximum of 42.7 g/L. Among the patients, 27 out of 376 (7.2%) had albumin concentrations below 20 g/L. Most patients were hospitalized in the infectious diseases department, followed by the medical ICU and cardiology. The primary indication for treatment was infectious endocarditis, mainly caused by *Enterococcus faecalis* (53%), often treated in combination with

The frequency of target attainment (TA) showed minimal variability across different models for a PD target of $fT > 1 \times MIC$ 100%, with a mean frequency of $97.3 \pm 1.0\%$. The lowest PTA value was 95.7% (Gregoire model), while the highest was 98.7% (Ulldemolins model) (Figure 3A). For a more stringent PD

Commenté [DT1]: Revoir en parlant 1) de la variabilité inter-model et 2) de l'effet de l'hypoalbuminémie sur les cut-off

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149 target of $ft > 4 \times MIC$ 100%, the mean PTA dropped to $86.4 \pm 9.7\%$, ranging from a minimum of 67.8% (Gregoire
150 model) to a maximum of 97.3% (Ulldemolins model) (Figure 3A).

151 The concordance matrix for the $ft > 1 \times MIC$ 100% target indicated strong agreement between models,
152 with concordance rates above 97% (Figure 3B). In contrast, the concordance matrix for the $ft > 4 \times MIC$ 100%
153 target revealed three distinct groups: the Gregoire and Heffernan models (93% concordance), the Standing
154 and Ulldemolins models (99%), and a cluster of Hartman, Gijsen, Dreesen, Leegwater, and Bos models, each
155 with concordance rates of 96% or higher (Figure 3C). did not appear to exhibit significant differences across
156 various models for a pharmacodynamic (PD) target of $ft > MIC$ 100%. The mean frequency was $97.3 \pm 1.0\%$,
157 with a minimum value of 95.7% (Gregoire model) and a maximum value of 98.7% (Ulldemolins model) (Figure
158 2A). For a PD target of $ft > 4 \times MIC$ 100%, the mean frequency of TA was $86.4 \pm 9.7\%$. The minimum value was
159 67.8% (Gregoire model), and the maximum value was 97.3% (Ulldemolins model) (Figure 2A). Concordance
160 matrix between models for a PD target of $ft > MIC$ 100% indicated a high degree of concordance ($>97\%$)
161 (Figure 2B). However, the concordance matrix for a PD target of $ft > 4 \times MIC$ 100% revealed 3 distinct blocks:
162 Gregoire and Heffernan models (93%), Standing and Ulldemolins models (99%), and Hartman, Gijsen,
163 Dreesen, Leegwater, and Bos models ($\geq 96\%$) (Figure 2C).

164 In addition, clinical, demographic, and biological data from the retrospective cohort were analyzed
165 as predictors of CEF_{tot} concentration at trough. Simple linear mixed-effects regression identified age
166 ($p=0.033$), intake dose ($p<0.001$), daily dose ($p=0.001$), albumin ($p=0.002$), and creatininemia ($p=0.021$) as
167 significant predictors. After integrating these predictors into a full model and applying backward variable
168 selection, the final model retained age ($p=0.005$), daily dose ($p<0.001$), albumin ($p=0.009$), and creatininemia
169 ($p<0.001$) as key predictors of CEF_{tot} concentration at trough (Table 4).

170 Additionally, clinical, demographic, and biological data from the retrospective cohort were analyzed
171 and CEF_{tot} trough concentrations were 52.6 ± 33.5 mg/L for the 1g x1/day regimen, 62.9 ± 44.6 mg/L for 2g x1/day,
172 84.2 ± 38.5 mg/L for 1g x2/day, and 126.3 ± 69.1 mg/L for 2g x2/day (Figure 4). The 2g x2/day regimen showed
173 significantly higher CEF_{tot} trough concentrations than the other dosing regimens ($p<0.001$). Notably, the 1g
174 and 2g x1/day regimens showed no significant differences in trough concentrations between the two regimens ($p=0.15$).

x2/day regimen produced higher CEF_{tot} trough concentrations than both the 2g x1/day regimen (p=0.029) and the 1g x1/day regimen (p<0.001). However, no statistically significant difference was observed between the 1g x1/day and 2g x1/day regimens (p=0.522) (Figure 4).

ANOVA demonstrated that CEFt trough concentrations increased with dosing regimen. The mean CEFt concentrations were 52.6±33.5 mg/L for 1g x1 /day, 62.9±44.6 mg/L for 2g x1 /day, 84.2±38.5 mg/L for 1g x2 /day, and 126.3±69.1 mg/L for 2g x2 /day (Figure 3). Significantly higher CEFt trough concentrations were observed for the 2g x2 /day regimen when compared to other dosing regimens (p<0.001). Interestingly, the 1g x2 /day regimen exhibited higher CEFt trough concentrations than 2g x1 /day (p=0.029) and 1g x1 /day (p<0.001), but no statistically significant differences were noted between 1g x1 /day and 2g x1 /day (p=0.522) (Figure 3).

PTA curves further corroborated these results, showing that increasing dose improved PTA. It is worth noting that 1g x2 /day resulted in higher PTA than 2g x1/day. Model used to predict CEFu had a pronounced impact on PTA for each dosing regimen. The range of MIC adequately covered (PTA > 90%) for 1g of CEF once daily varied from 1 mg/L (Gregoire model) to 8 mg/L (Ulldemolins model) for the same plasma CEFt concentration. Increasing the dosing regimen tended to elevate the CEFt trough concentration, resulting in broader coverage of higher MIC values, with the 2g twice daily regimen achieving coverage ranging from slightly below 4mg/L for the Gregoire model to 16 mg/L for the Standing and Ulldemolins models (Figure 4).

The primary aim of the present study was to conduct a comprehensive analysis of literature data concerning CEF_u, both in terms of quantification and modeling, in order to perform an external validation and to assess its relevance and potential applicability in clinical practice. The study revealed several noteworthy findings, despite the presence of significant limitations.

The literature search yielded a total of nine models for CEF_u modeling based on CEF_{tot}. It should be noted that some data were found to be unsuitable for modeling purposes and were challenging to obtain. Some data were deemed irrelevant⁽²⁰⁾ while others exhibited inconsistencies, including discrepancies between the original manuscript and supplementary model.⁽²¹⁾ However, it is important to clarify that

201 these discrepancies did not appear to impact the overall conclusions drawn in these studies. Furthermore, it
202 is worth highlighting that although efforts were made to accurately translate the models from the literature
203 for use in this study, errors in data interpretation cannot be ruled out. To promote transparency and
204 reproducibility, the code used in this study is available on GitHub at
205 <https://github.com/ThomasDufлот/Ceftriaxone-AAC>. ~~To ensure transparency in our approach, the code~~
206 ~~employed has been provided as supplementary material, thereby facilitating the reproducibility of this work.~~

207 A notable degree of variability was observed between models when examining CEF_u modeling and
208 the determination of optimal thresholds based on minimum inhibitory concentration (MIC). with higher
209 variability of the thresholds when hypoalbuminemia occurs. ~~One~~ An interesting key factor is the variation in the
210 studied population among the different research studies. These differences included the age of patients,
211 their critical illness status, the number of samples collected, the timing of sample collection, the presence or
212 absence of hypoalbuminemia, and the use of cardiopulmonary bypass. Additionally, the method employed
213 for sample processing, such as UF or ED, introduced another source of variability. It is noteworthy that a
214 recent paper reported significant differences in the parameters B_{max} and K_d between *in vitro* UF and *in vivo*
215 IV microdialysis. (30) ³⁰—The choice between UF and ED is particularly important, as it influences the
216 determination of the free drug fraction. While ED is regarded as the gold standard method, it is also known
217 for its time-consuming nature. Conversely, UF is a more straightforward approach but is sensitive to a range
218 of analytical conditions. Both UF and ED are influenced by temperature, and UF is particularly affected by
219 centrifugation speed and time. (31) It is also important to note that these analytical considerations may vary
220 depending on the physico-chemical properties of the drug being studied. ~~Furthermore, both UF and ED are~~
221 ~~influenced by temperature, with centrifugation speed and time also serving as critical variables for~~
222 ~~ultrafiltration.~~ ³¹

223 The significance of external validation in ensuring the reliability of the study's findings cannot be
224 overstated. Consequently, the predictive performance of the models under investigation was rigorously
225 assessed, despite the limited sample size in this single-center prospective cohort (N=62). Although this
226 limitation is acknowledged, the use of combined fit metrics provided valuable insights, revealing that certain

227 models demonstrated a higher degree of reliability compared to others. It is important to interpret these
228 results with caution, as their validation requires replication and further extensive investigation.

229 Interestingly, although the Gregoire, Heffernan, and Hartman models demonstrated satisfactory
230 metrics during external validation, we observed differences in concordance between Hartman and the other
231 two models. Gregoire and Heffernan formed a concordance group with high similarity (93%), while Hartman
232 showed lower concordance—85% with Heffernan and 79% with Gregoire. This intriguing result may be
233 attributed to Hartman's higher MSE and MPE. Given the greater variability in MPE for the Hartman model,
234 we hypothesize that the Gregoire and Heffernan models offer better predictive performance, with Heffernan
235 being the strongest overall due to its lowest MPE, RMSE, RMSE%, and highest R².

236 Transitioning from modeling concepts to clinical implications, the primary objective was to ascertain
237 whether the CEF dosing regimen was sufficient to achieve the therapeutic objectives. Although the
238 retrospective cohort study possessed evident limitations, it has been observed that when employing a CEF_u
239 threshold of 1 mg/L, PTA is 95.7% across all models. Consequently, the level of concordance among these
240 models was relatively high. However, as the thresholds increased, inter-model variability may start to impact
241 clinical conclusions regarding the effectiveness of CEF.

242 Balancing the limitations of retrospective data, it is crucial to emphasize that the significant
243 predictors of CEF_{tot} identified in this cohort, including age, plasma albumin, plasma creatinine, and dose, have
244 been previously highlighted in the literature.^(18, 26)^{18,26} As demonstrated in the mixed effects regression
245 analysis (Table 4), daily dose was treated as a continuous variable, revealing that CEF_{tot} CEF_u levels
246 concentrations increase with higher doses. Of note, a population pharmacokinetic (popPK) analysis would
247 allow a more thorough evaluation of covariates and CEF_u estimation using nonlinear mixed-effects modeling.
248 However, due to the retrospective nature of our data, the low number of patients with repeated ceftriaxone
249 concentration measurements, and the heterogeneity of our patient population (encompassing both ICU and
250 general ward patients), developing a popPK model for our cohort would likely introduce bias and yield
251 inconclusive results. Notably, ~~it~~ Despite this limitation, it is interesting to observe that CEF_{tot} CEF_u
252 concentrations were significantly higher with a dosing regimen of 1g administered twice daily compared to

253 2g administered once daily. This suggests that dividing the daily dose or employing continuous infusion may
254 represent more effective approaches for achieving the therapeutic target but may also elevate the risk of
255 toxicity.^(32, 33)^{32,33} ~~In conclusion~~addition, the observed disparities in the probability of target attainment
256 (PTA) curves among the various models under evaluation may lead to divergent conclusions regarding the
257 optimal therapeutic management and dosage adjustments for ceftriaxone.

258 To conclude, determining CEF_u offers an intriguing opportunity to enhance our understanding of
259 CEF's PK and PD, as recent publications have emphasized. In line with this, the current study has strived to
260 provide comprehensive results based on several available models, enabling fellow researchers to improve
261 their collective understanding of this topic. From a clinician's perspective, targeting 4 times the MIC during
262 the interdose period is essential for treating serious infections like infective endocarditis. However, achieving
263 this target depends on the model used, and nutritional status plays a crucial role in dose optimization, with
264 a balance between inefficacy and toxicity. Analysis of dosing regimens in the retrospective cohort revealed
265 that splitting a dose twice daily is more effective than once daily administration. Nevertheless, it is important
266 to stress the need for standardized analytical considerations and rigorous external validation to establish
267 CEF_u as a robust PD biomarker in clinical practice. In summary, the application of CEF_u in clinical practice may
268 face challenges due to potential analytical biases, which warrant further investigation.

269 MATERIALS AND METHODS

270 Literature Search

271 A systematic review of population pharmacokinetic (PK) models for both ~~CEF_{tot}~~ ~~CEF_t~~ and ~~CEF_u~~ ~~CEF_f~~
272 was conducted using Pubmed, covering the period from January 2000 up to December 2022. The terms
273 "population", "pharmacokinetics", "free", "unbound" and "ceftriaxone" were selected for the literature
274 review and combined to obtain the following search query:

275 - Population AND pharmacokinetics AND ceftriaxone AND (free OR unbound) AND
276 (("2000/01/01"[Date - Publication] : "2022/12/31"[Date - Publication]))

277 The query was not limited by age groups or medical conditions, but articles included were required
278 to be in English. Informations from the selected articles were collected, including the number of patients and
279 samples, the studied population, the method used to quantitate ~~CEF_u, CEF_t~~, the formula used to predict the
280 relationship between ~~CEF_{tot}, CEF_t~~ and CEF_u, and the values of each parameter of the formula. Formulas were
281 retained for further analysis if all variable and parameter values were provided, allowing for comprehensive
282 CEF_u modeling.

283 **Total (CEF_{tot}) and unbound (CEF_u) quantitation**

284 Chemicals and reagents

285 Ceftriaxone and the internal standard ceftriaxone-d4 were purchased from Alsachim® (Illkirch-
286 Graffenstaden - France). HPLC-grade methanol and water were supplied by Carlo Erba Reagents® (Val de
287 Reuil, France). Centrifugal filter units (Amicon® Ultra 0.5 mL 30K) for CEF_u determination were provided by
288 Merck Millipore (Cork, Ireland).

289 Sampling and analysis

290 Blood samples for therapeutic drug monitoring (TDM) purposes were collected using dry collection
291 tubes and were promptly subjected to centrifugation at 1,700 x g for 10 min. For the determination of CEF_u,
292 500 µL of serum was processed through ultrafiltration (UF) utilizing centrifugal filter units at room
293 temperature, following the manufacturer's guidelines (centrifuged at 14,000 x g for 10 minutes). The
294 resulting filtrates underwent the same sample preparation procedure as CEF_{tot}. Detailed analytical
295 procedures for quantifying both CEF_{tot} and CEF_u can be found in Supplementary Material S1.

296

297 **Unbound ceftriaxone (CEF_u) modeling**

298 For each model, the concentration of CEF_u was modeled as a function of ~~CEF_{tot}, CEF_t~~ both under normal
299 albumin ~~levels-concentrations~~ (35 g/L) and hypoalbuminemia (20 g/L). To fully appreciate the non-linear
300 relationship between CEF_u and ~~CEF_{tot}, CEF_t~~, figures were generated with ~~CEF_{tot}, CEF_t~~ concentrations ranging from 0 to
301 300 mg/L, commonly observed in clinical practice.

Determination of optimal total ceftriaxone (~~CEF_{tot}~~, ~~CEF_{ft}~~) thresholds

A target minimum inhibitory concentration (MIC) of 1 mg/L, which is considered the breakpoint concentration of ceftriaxone against *Enterobacteriaceae* by the European Committee on Antimicrobial Susceptibility Testing,⁽³⁴⁾ was employed to establish ~~CEF_{tot}~~, ~~CEF_{ft}~~ thresholds based on MIC. These thresholds were calculated for the criteria of achieving FT > MIC 100% and FT > 4 × MIC 100%.

External validation and comparison of predictive performance

For external validation, performance metrics, including signed error, relative error, mean signed error (MSE), mean percentage error (MPE), root mean square error (RMSE), root mean square error of percentage (RMSE%) and determination coefficient (R²) were calculated and employed to compare the predictive performance of each formula. Analysis of variance (ANOVA) followed by Tukey Honest Significant Differences tests were performed for between model comparison. Bland-Altman plots for each model for both signed and relative differences were drawn for a full representation of the data.

Studied population

EDSaN solution,⁽³⁵⁾ a Clinical Data Warehouse (CDW), was used to identify and extract trough plasma CEF concentration requests for trough plasma CEF concentration, spanning from 2016 to 2022. These requests were subsequently obtained, along with the relevant patient data, from the CDW. The extracted data encompassed various blood biology elements, in addition to clinical and demographic data. Moreover, information regarding the CEF dosing regimen was manually retrieved from the medical records of the patients.

Ethics

The French Data Protection Authority (CNIL) approved the construction and the usage of the Rouen University Hospital Clinical Data Warehouse (decision DT-2020-007), based on a declaration compatible with the General Data Protection Regulation applicable in France. Following national rules, a global public information was issued and individual information provided for each new patient in the hospital. In addition,

327 the prospective study was conducted following approval from our local ethics committee (approval number
328 E2024-19, obtained on February 29, 2024).— Due to the non-interventional nature of the study, written
329 informed consent was not mandatory according to the national regulatory framework.

330 **Statistical analysis**

331 ~~CEF_{tot} modeling~~Statistical analysis was performed using R software v4.2.2³⁶, (36) RStudio v2024.4.2.764, (37) and the
332 following packages: ggplot2 v3.4.3³⁵, (38) ggsci v2.3.9², (39) ³⁹ggpubr v0.5.6⁰, (40) reshape2 v1.4.4, (41) ~~and~~ cowplot
333 v1.1.4³⁷, (42) forcats v1.0.0, (43) dplyr v1.1.4, (44) flextable v0.9.6, (45) gridExtra v2.3, (46) gt v0.11.1, (47)
334 gtsummary v2.0.3, (48) officer v0.6.6, (49) qqcorrplot v0.1.4.1, (50) exact2x2 v1.6.9, (51) lmerTest v3.1.3,
335 (52) RColorBrewer v1.1.3 (53) and multcomp v1.4.26. (54)

336 Concerning patient and sample-related variables, continuous and categorical variables were
337 presented as medians with the interquartile range (IQR) and n (%) respectively, where "n" corresponds to
338 the number of non-missing observations.

339 Predictors of CEF_{tot} trough concentration were examined through linear mixed effects models using
340 Satterthwaite's degrees of freedom for p-value computation, considering multiple measurements for the
341 same patient. Subsequently, all predictors with a p-value < 0.05 were integrated into a full model. Irrelevant
342 variables were eliminated from the full model using backward variable selection, guided by the Akaike
343 Information Criterion.

344 Analysis of variance (ANOVA), followed by Tukey's ~~honestly-Honestly significant~~ Significant difference
345 Differences method for post hoc pairwise comparison, was conducted to assess the impact of the main
346 ceftriaxone dosing regimen on CEF_{tot}, ~~CEF_{tr}~~ trough concentrations. PTA was then calculated for each model,
347 stratified by dosing regimen, across various minimum inhibitory concentration (MIC) values ranging from
348 0.125 to 32 mg/L.

349 ~~Site: https://github.com/ThomasDufлот/Ceftriaxone-AAC~~
350 https://github.com/ThomasDufлот/Ceftriaxone-AAC. In order to maintain patient privacy; age, sex, admission

351 dates and co-morbidities have been removed from the raw data of studied population and considered as

352 "NA" ~~.(Supplementary files S1).~~

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359 The authors declare that they have no known competing financial interests or personal relationships
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361 **References :**

- 362 1. Kollef MH, Shorr AF, Bassetti M, Timsit J-F, Micek ST, Michelson AP, Garnacho-Montero J. 2021.
363 Timing of antibiotic therapy in the ICU. Crit Care 25:360.
- 364 2. Patel IH, Chen S, Parsonnet M, Hackman MR, Brooks MA, Konikoff J, Kaplan SA. 1981.
365 Pharmacokinetics of ceftriaxone in humans. Antimicrob Agents Chemother 20:634–641.
- 366 3. EldougDoug MW, Youssef DM, El-Shal AS, Sharaf YA, Raparla S, Jasti BR, Elnahas HM. 2023. Evaluation
367 of ceftriaxone pharmacokinetics in hospitalized Egyptian pediatric patients. Eur J Pediatr 182:4407–
368 4420.
- 369 4. Roberts JA, Lipman J. 2009. Pharmacokinetic issues for antibiotics in the critically ill patient. Crit Care
370 Med 37:840–851; quiz 859.
- 371 5. Novy E, Martinière H, Roger C. 2023. The Current Status and Future Perspectives of Beta-Lactam
372 Therapeutic Drug Monitoring in Critically Ill Patients. Antibiotics (Basel) 12:681.
- 373 6. Ulldemolins M, Roberts JA, Rello J, Paterson DL, Lipman J. 2011. The effects of hypoalbuminaemia on
374 optimizing antibacterial dosing in critically ill patients. Clin Pharmacokinet 50:99–110.

- 375 7. Lode H, File TM, Mandell L, Ball P, Pypstra R, Thomas M, 185 Gemifloxacin Study Group. 2002. Oral
376 gemifloxacin versus sequential therapy with intravenous ceftriaxone/oral cefuroxime with or without
377 a macrolide in the treatment of patients hospitalized with community-acquired pneumonia: a
378 randomized, open-label, multicenter study of clinical efficacy and tolerability. *Clin Ther* 24:1915–
379 1936.
- 380 8. Serafino Wani RL, Filson SA, Chattaway MA, Godbole G. 2016. Invasive shigellosis in MSM. *Int J STD*
381 *AIDS* 27:917–919.
- 382 9. Rambaud A, Gaborit BJ, Deschanvres C, Le Turnier P, Lecomte R, Asseray-Madani N, Leroy A-G,
383 Deslandes G, Dailly É, Jolliet P, Boutoille D, Bellouard R, Gregoire M, Nantes Anti-Microbial Agents
384 PK/PD (NAMAP) study group. 2020. Development and validation of a dosing nomogram for amoxicillin
385 in infective endocarditis. *J Antimicrob Chemother* 75:2941–2950.
- 386 10. Nathan BR, Scheld WM. 2003. The efficacy of trovafloxacin versus ceftriaxone in the treatment of
387 experimental brain abscess/cerebritis in the rat. *Life Sci* 73:1773–1782.
- 388 11. Marsot A. 2020. Population pharmacokinetic models of first choice beta-lactam antibiotics for severe
389 infections treatment: What antibiotic regimen to prescribe in children? *J Pharm Pharm Sci* 23:470–
390 485.
- 391 12. Perry TR, Schentag JJ. 2001. Clinical use of ceftriaxone: a pharmacokinetic-pharmacodynamic
392 perspective on the impact of minimum inhibitory concentration and serum protein binding. *Clin*
393 *Pharmacokinet* 40:685–694.
- 394 13. Simon N, Dussol B, Sampol E, Purgus R, Brunet P, Lacarelle B, Berland Y, Bruguerolle B, Urien S. 2006.
395 Population pharmacokinetics of ceftriaxone and pharmacodynamic considerations in haemodialysed
396 patients. *Clin Pharmacokinet* 45:493–501.

- 397 14. Wang Y-K, Wu Y-E, Li X, Tian L-Y, Khan MW, Tang B-H, Shi H-Y, Zheng Y, Hao G-X, van den Anker J, You
398 D-P, Zhao W. 2020. Optimal Dosing of Ceftriaxone in Infants Based on a Developmental Population
399 Pharmacokinetic-Pharmacodynamic Analysis. *Antimicrob Agents Chemother* 64:e01412-20.
- 400 15. Blumer JL, Reed MD, Kaplan EL, Drusano GL. 2005. Explaining the poor bacteriologic eradication rate
401 of single-dose ceftriaxone in group a streptococcal tonsillopharyngitis: a reverse engineering solution
402 using pharmacodynamic modeling. *Pediatrics* 116:927–932.
- 403 16. Tsai D, Stewart P, Goud R, Gourley S, Hewagama S, Krishnaswamy S, Wallis SC, Lipman J, Roberts JA.
404 2016. Total and unbound ceftriaxone pharmacokinetics in critically ill Australian Indigenous patients
405 with severe sepsis. *Int J Antimicrob Agents* 48:748–752.
- 406 17. Meletiadis J, Turlej-Rogacka A, Lerner A, Adler A, Tacconelli E, Mouton JW, the SATURN Diagnostic
407 Study Group. 2017. Amplification of Antimicrobial Resistance in Gut Flora of Patients Treated with
408 Ceftriaxone. *Antimicrob Agents Chemother* 61:e00473-17.
- 409 18. Tang Girdwood S, Dong M, Tang P, Stoneman E, Jones R, Yunger T, Ostermeier A, Curry C, Forton M,
410 Hail T, Mullaney R, Lahni P, Punt N, Kaplan J, Vinks AA. 2022. Population Pharmacokinetic Modeling of
411 Total and Free Ceftriaxone in Critically Ill Children and Young Adults and Monte Carlo Simulations
412 Support Twice Daily Dosing for Target Attainment. *Antimicrob Agents Chemother* 66:e0142721.
- 413 19. Meenks SD, le Noble JLML, Foudraine NA, de Vries F, Neef K, Janssen PKC. 2022. Population
414 pharmacokinetics of unbound ceftriaxone in a critically ill population. *Int J Clin Pharmacol Ther*
415 60:373–383.
- 416 20. Cheng V, Abdul-Aziz MH, Burrows F, Buscher H, Cho Y-J, Corley A, Gilder E, Kim H-S, Lim SY,
417 McGuinness S, Parke R, Reynolds C, Rudham S, Wallis SC, Welch SA, Fraser JF, Shekar K, Roberts JA,
418 ASAP ECMO Investigators. 2022. Population Pharmacokinetics and Dosing Simulations of Ceftriaxone
419 in Critically Ill Patients Receiving Extracorporeal Membrane Oxygenation (An ASAP ECMO Study). *Clin*
420 *Pharmacokinet* 61:847–856.

- 421 21. Standing JF, Ongas MO, Ogwang C, Kagwanja N, Murunga S, Mwaringa S, Ali R, Mturi N, Timbwa M,
422 Manyasi C, Mwalekwa L, Bandika VL, Ogutu B, Waichungo J, Kipper K, Berkley JA, FLACSAM-PK Study
423 Group. 2018. Dosing of Ceftriaxone and Metronidazole for Children With Severe Acute Malnutrition.
424 Clin Pharmacol Ther 104:1165–1174.
- 425 22. Bos JC, Prins JM, Mistício MC, Nunguiane G, Lang CN, Beirão JC, Mathôt RAA, van Hest RM. 2018.
426 Pharmacokinetics and pharmacodynamic target attainment of ceftriaxone in adult severely ill sub-
427 Saharan African patients: a population pharmacokinetic modelling study. J Antimicrob Chemother
428 73:1620–1629.
- 429 23. Leegwater E, Kraaijenbrink BVC, Moes DJ a. R, Purmer IM, Wilms EB. 2020. Population
430 pharmacokinetics of ceftriaxone administered as continuous or intermittent infusion in critically ill
431 patients. J Antimicrob Chemother 75:1554–1558.
- 432 24. Heffernan AJ, Curran RA, Denny KJ, Sime FB, Stanford CL, McWhinney B, Ungerer J, Roberts JA,
433 Lipman J. 2021. Ceftriaxone dosing in patients admitted from the emergency department with sepsis.
434 Eur J Clin Pharmacol 77:207–214.
- 435 25. Hartman SJF, Upadhyay PJ, Hagedoorn NN, Mathôt RAA, Moll HA, van der Flier M, Schreuder MF,
436 Brüggemann RJ, Knibbe CA, de Wildt SN. 2021. Current Ceftriaxone Dose Recommendations are
437 Adequate for Most Critically Ill Children: Results of a Population Pharmacokinetic Modeling and
438 Simulation Study. Clin Pharmacokinet 60:1361–1372.
- 439 26. Dreesen E, Gijzen M, Elkayal O, Annaert P, Debaveye Y, Wauters J, Karlsson MO, Spriet I. 2022.
440 Ceftriaxone dosing based on the predicted probability of augmented renal clearance in critically ill
441 patients with pneumonia. J Antimicrob Chemother 77:2479–2488.
- 442 27. Grégoire M, Dailly E, Le Turnier P, Garot D, Guimard T, Bernard L, Tattevin P, Vandamme Y-M, Hoff J,
443 Lemaitre F, Verdier M-C, Deslandes G, Bellouard R, Sébille V, Chiffolleau A, Boutoille D, Navas D,

444 Asseray N. 2019. High-Dose Ceftriaxone for Bacterial Meningitis and Optimization of Administration
445 Scheme Based on Nomogram. *Antimicrob Agents Chemother* 63:e00634-19.

446 28. Ulldemolins M, Bastida C, Llauradó-Serra M, Csajka C, Rodríguez A, Badia JR, Martín-Loeches I, Soy D.
447 2021. Once-daily 1 g ceftriaxone optimizes exposure in patients with septic shock and
448 hypoalbuminemia receiving continuous veno-venous hemodiafiltration. *Eur J Clin Pharmacol*
449 77:1169–1180.

450 29. Gijssen M, Dreesen E, Van Daele R, Annaert P, Debaveye Y, Wauters J, Spriet I. 2021.
451 Pharmacokinetic/Pharmacodynamic Target Attainment Based on Measured versus Predicted
452 Unbound Ceftriaxone Concentrations in Critically Ill Patients with Pneumonia: An Observational
453 Cohort Study. *Antibiotics (Basel)* 10:557.

454 30. Sanz-Codina M, Wicha SG, Wulkersdorfer B, Al Jalali V, Van Os W, Vossen MG, Bauer M, Lackner E,
455 Dorn C, Zeitlinger M. 2023. Comparison of ultrafiltration and microdialysis for ceftriaxone protein-
456 binding determination. *J Antimicrob Chemother* 78:380–388.

457 31. Metsu D, Lanot T, Fraissinet F, Concordet D, Gayraud V, Averseng M, Ressault A, Martin-Blondel G,
458 Levade T, Février F, Chatelut E, Delobel P, Gandia P. 2020. Comparing ultrafiltration and equilibrium
459 dialysis to measure unbound plasma dolutegravir concentrations based on a design of experiment
460 approach. *Sci Rep* 10:12265.

461 32. Heffernan AJ, Sime FB, Kumta N, Wallis SC, McWhinney B, Ungerer J, Wong G, Joynt GM, Lipman J,
462 Roberts JA. 2022. Multicenter Population Pharmacokinetic Study of Unbound Ceftriaxone in Critically
463 Ill Patients. *Antimicrob Agents Chemother* 66:e0218921.

464 33. Alasmari F, Alasmari MS, Muwainea HM, Alomar HA, Alasmari AF, Alsanea S, Alshamsan A, Rasool MF,
465 Alqahtani F. 2023. Physiologically-based pharmacokinetic modeling for single and multiple dosing
466 regimens of ceftriaxone in healthy and chronic kidney disease populations: a tool for model-informed
467 precision dosing. *Front Pharmacol* 14:1200828.

- 468 34. Comité de l'Antibiogramme de la Société Française de Microbiologie. Société Française de
469 Microbiologie. [https://www.sfm-microbiologie.org/boutique/_comite-de-lantibiogramme-de-la-sfm-](https://www.sfm-microbiologie.org/boutique/_comite-de-lantibiogramme-de-la-sfm-ca-sfm-v1-0-juin-2024/)
470 [ca-sfm-v1-0-juin-2024/](https://www.sfm-microbiologie.org/boutique/_comite-de-lantibiogramme-de-la-sfm-ca-sfm-v1-0-juin-2024/). Retrieved 13 November 2024.
- 471 35. Pressat-Laffouilhère T, Balayé P, Dahamna B, Lelong R, Billey K, Darmoni SJ, Grosjean J. 2022.
472 Evaluation of Doc'EDS: a French semantic search tool to query health documents from a clinical data
473 warehouse. BMC Med Inform Decis Mak 22:34.
- 474 36. R Core Team. 2023. R: A Language and Environment for Statistical Computing. R Foundation for
475 Statistical Computing, Vienna, Austria. <https://www.R-project.org/>.
- 476 37. RStudio Team. 2023. RStudio: Integrated Development Environment for R. RStudio, PBC, Boston, MA.
477 <https://posit.co/download/rstudio-desktop/>.
- 478 38. Wickham H. 2016. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.
479 <https://ggplot2.tidyverse.org>.
- 480 39. Xiao N. 2024. ggsci: Scientific Journal and Sci-Fi Themed Color Palettes for "ggplot2." [https://CRAN.R-](https://CRAN.R-project.org/package=ggsci)
481 [project.org/package=ggsci](https://CRAN.R-project.org/package=ggsci).
- 482 40. Kassambara A. 2023. ggpubr: "ggplot2" Based Publication Ready Plots. [https://CRAN.R-](https://CRAN.R-project.org/package=ggpubr)
483 [project.org/package=ggpubr](https://CRAN.R-project.org/package=ggpubr).
- 484 41. Wickham H. 2007. Reshaping Data with the reshape Package. Journal of Statistical Software 21:1–20.
- 485 42. Wilke CO. 2024. cowplot: Streamlined Plot Theme and Plot Annotations for "ggplot2."
486 <https://CRAN.R-project.org/package=cowplot>.
- 487 43. Wickham H. 2023. forcats: Tools for Working with Categorical Variables (Factors). [https://CRAN.R-](https://CRAN.R-project.org/package=forcats)
488 [project.org/package=forcats](https://CRAN.R-project.org/package=forcats).
- 489 44. Wickham H, François R, Henry L, Müller K, Vaughan D. 2023. dplyr: A Grammar of Data Manipulation.
490 <https://CRAN.R-project.org/package=dplyr>.

491 45. Gohel D, Skintzos P. 2024. flextable: Functions for Tabular Reporting. [https://CRAN.R-](https://CRAN.R-project.org/package=flextable)
492 [project.org/package=flextable](https://CRAN.R-project.org/package=flextable).

493 46. Auguie B. 2017. gridExtra: Miscellaneous Functions for “Grid” Graphics. [https://CRAN.R-](https://CRAN.R-project.org/package=gridExtra)
494 [project.org/package=gridExtra](https://CRAN.R-project.org/package=gridExtra).

495 47. Iannone R, Cheng J, Schloerke B, Hughes E, Lauer A, Seo J, Brevoort K, Roy O. 2024. gt: Easily Create
496 Presentation-Ready Display Tables. <https://CRAN.R-project.org/package=gt>.

497 48. Sjoberg DD, Whiting K, Curry M, Lavery JA, Larmarange J. 2021. Reproducible Summary Tables with
498 the gtsummary Package. The R Journal 13:570–580.

499 49. Gohel D, Moog S. 2024. officer: Manipulation of Microsoft Word and PowerPoint Documents.
500 <https://CRAN.R-project.org/package=officer>.

501 50. Kassambara A. 2023. ggcorrplot: Visualization of a Correlation Matrix using “ggplot2.”
502 <https://CRAN.R-project.org/package=ggcorrplot>.

503 51. Fay MP. 2010. Confidence intervals that match Fisher’s exact or Blaker’s exact tests. Biostatistics
504 11:373–374.

505 52. Kuznetsova A, Brockhoff PB, Christensen RHB. 2017. lmerTest Package: Tests in Linear Mixed Effects
506 Models. Journal of Statistical Software 82:1–26.

507 53. Neuwirth E. 2022. RColorBrewer: ColorBrewer Palettes. [https://CRAN.R-](https://CRAN.R-project.org/package=RColorBrewer)
508 [project.org/package=RColorBrewer](https://CRAN.R-project.org/package=RColorBrewer).

509 54. Hothorn T, Bretz F, Westfall P. 2008. Simultaneous Inference in General Parametric Models.
510 Biometrical Journal 50:346–363.

512

513

514 1. Kollef MH, Shorr AF, Bassetti M, *et al.* Timing of antibiotic therapy in the ICU. *Crit Care* 2021; **25**: 360.

515 2. De Paepe P, Belpaire FM, Buylaert WA. Pharmacokinetic and pharmacodynamic considerations when treating patients with sepsis and septic shock. *Clin*

516 *Pharmacokinet* 2002; **41**: 1135–51.

517 3. Lizza BD, Raush N, Micek ST. Antibiotic Optimization in the Intensive Care Unit. *Semin Respir Crit Care Med* 2022; **43**: 125–30.

518 4. Roberts JA, Lipman J. Pharmacokinetic issues for antibiotics in the critically ill patient. *Crit Care Med* 2009; **37**: 840–51; quiz 859.

519 5. Novy E, Martinière H, Roger C. The Current Status and Future Perspectives of Beta Lactam Therapeutic Drug Monitoring in Critically Ill Patients. *Antibiotics*

520 *(Basel)* 2023; **12**: 681.

521 6. Uldemolins M, Roberts JA, Rello J, Paterson DL, Lipman J. The effects of hypoalbuminaemia on optimizing antibacterial dosing in critically ill patients. *Clin*

522 *Pharmacokinet* 2011; **50**: 99–110.

523 7. Lode H, File TM, Mandell L, *et al.* Oral gemifloxacin versus sequential therapy with intravenous ceftriaxone/oral cefuroxime with or without a macrolide in the

524 treatment of patients hospitalized with community-acquired pneumonia: a randomized, open-label, multicenter study of clinical efficacy and tolerability. *Clin*

525 *Ther* 2002; **24**: 1915–36.

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Model	Population	CEFu	Formula	Parameter values	Reference
A	Severely ill sub-Saharan African adults (N=88 patients, 277 samples for CEF_{tot} and 276 samples for CEF_u).	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 0.12 \times \left(\frac{ALB}{0.42}\right)^{1.3}$ mM $Kd = 0.092$ mM	Bos et al(22)
B	Children with severe acute malnutrition (N=81 children, 244 samples for CEF_{tot} , CEF_t and CEF_u)	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 22.89 \times \left(\frac{ALB}{33.75}\right)^{-0.26}$ mg/L $Kd = 0.56$ mg/L	Standing et al(21)
C	Adults with suspected or proven bacterial meningitis (N=153 patients, 301 samples for CEF_{tot} , CEF_t and 214 for CEF_u)	UF	$fu = 1 - ((5.10^{-9} \times (CEF_{tot}^{2.3}) + (6.10^{-7} \times (CEF_{tot}^{2.2}) - 0.0004 \times CEF_{tot} + 0.9393))$ $CEF_u = fu \times CEF_{tot}$		Gregoire et al(27)
D	Critically ill adults (N=55 patients, 110 samples for CEF_{tot} and CEF_u)	-	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 113 \times (1 + (0.04 \times ALB - 29))$ mg/L $Kd = 11.5$ mg/L	Leegwater et al(23)
E	Critically ill adults with pneumonia (N=31 patients, 72 samples for CEF_{tot} and CEF_u)	ED	$CEF_u = Bmax \times \left(\left(\frac{ALB}{0.44}\right)^{0.26} \times CEF_{tot} + CEF_{tot}^{Kd}\right)$	$Bmax = -0.64 \times \left(\frac{ALB}{0.44}\right)^{0.26}$ mM $Kd = 1.09$ mM	Gijssen et al(29)
F	Critically ill children (N=45 patients, 205 samples for CEF_{tot} and 45 samples for CEF_u)	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 223 \times \left(\frac{ALB}{27}\right)^1$ mg/L $Kd = 30.3$ mg/L	Hartman et al(25)
G	Adults with septic shock, hypoalbuminemia and hemodiafiltration (N=50 patients, 50 samples for CEF_{tot} and CEF_u)	UF	$fu = 1 \times e^{-0.82 \times \left(\frac{ALB}{24.2}\right)}$ $CEF_u = fu \times CEF_{tot}$		Ulldemolins et al(28)
H	Critically ill adults with augmented clearance (N=33 patients, 259 samples for CEF_{tot} and 76 for CEF_u)	ED	$CEF_{tot} = CEF_u + \frac{CEF_u \times n \times ALB}{CEF_u + Kd}$	$n = 0.771$ $Kd = 0.053$ mM	Dreesen et al(26)
I	Critically ill adults (N=36 patients, 267 samples for CEF_{tot} and 207 samples for CEF_u)	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = ALB \times 0.82 \times 8.34 \times 1000$ mg/L $Kd = \frac{K_{off}}{K_{on}} = \frac{18537}{1290} = 14.37$ mg/L	Heffernan et al(24)

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TABLE 1: Description of the different formulas used to predict unbound ceftriaxone (CEF_u) from total ceftriaxone (CEF_{tot}) levels/concentrations.

Model	Population	CEFu	Formula	Parameter values	Reference
A	Severely ill sub-Saharan African adults (N=88 patients, 277 samples for CEF_{tot} and 276 samples for CEF_u).	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 0.12 \times \left(\frac{ALB}{0.42}\right)^{1.3}$ mM $Kd = 0.092$ mM	Bos et al(22)
B	Children with severe acute malnutrition (N=81 children, 244 samples for CEF_{tot} and 244 samples for CEF_u).	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 22.89 \times \left(\frac{ALB}{33.75}\right)^{-0.26}$ mg/L $Kd = 0.56$ mg/L	Standing et al(21)
C	Adults with suspected or proven bacterial meningitis (N=153 patients, 301 samples for CEF_{tot} and 214 for CEF_u).	UF	$fu = 1 - ((5.10^{-9} \times (CEF_{tot})^{3.3}) + (6.10^{-7} \times (CEF_{tot})^{2.2}) - 0.0004 \times CEF_{tot} + 0.9393)$ $CEF_u = fu \times CEF_{tot}$		Gregoire et al(27)
D	Critically ill adults (N=55 patients, 110 samples for CEF_{tot} and 110 for CEF_u).	-	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 113 \times (1 + (0.04 \times ALB - 29))$ mg/L $Kd = 11.5$ mg/L	Leegwater et al(23)
E	Critically ill adults with pneumonia (N=31 patients, 72 samples for CEF_{tot} and 72 for CEF_u).	ED	$CEF_u = Bmax \times \left(\frac{ALB}{0.44}\right)^{0.26} \times CEF_{tot} + CEF_{tot}^{Kd}$	$Bmax = -0.64 \times \left(\frac{ALB}{0.44}\right)^{0.26}$ mM $Kd = 1.09$ mM	Gijzen et al(29)
F	Critically ill children (N=45 patients, 205 samples for CEF_{tot} and 45 samples for CEF_u).	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = 223 \times \left(\frac{ALB}{27}\right)^1$ mg/L $Kd = 30.3$ mg/L	Hartman et al(25)
G	Adults with septic shock, hypoalbuminemia and hemodiafiltration (N=50 patients, 50 samples for CEF_{tot} and 50 for CEF_u).	UF	$fu = 1 \times e^{-0.82 \times \left(\frac{ALB}{24.2}\right)}$ $CEF_u = fu \times CEF_{tot}$		Ulldemolins et al(28)
H	Critically ill adults with augmented clearance (N=33 patients, 259 samples for CEF_{tot} and 76 for CEF_u).	ED	$CEF_{tot} = CEF_u + \frac{CEF_u \times n \times ALB}{CEF_u + Kd}$	$n = 0.771$ $Kd = 0.053$ mM	Dreesen et al(26)
I	Critically ill adults (N=36 patients, 267 samples for CEF_{tot} and 207 samples for CEF_u).	UF	$CEF_{tot} = CEF_u + \frac{CEF_u \times Bmax}{CEF_u + Kd}$	$Bmax = ALB \times 0.82 \times 8.34 \times 1000$ mg/L $Kd = \frac{K_{off}}{K_{on}} = \frac{18537}{1290} = 14.37$ mg/L	Heffernan et al(24)

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528 ALB: Albuminemia, Bmax: Maximum binding capacity , ~~CEF_{tot}~~ ~~CE_{ff}~~: Total ceftriaxone, CEF_u: Unbound Ceftriaxone, ED: Equilibrium dialysis, fu: fraction unbound, Kd:
529 Dissociation constant, Koff: Dissociation rate constant, Kon: Association rate constant, UF: Ultrafiltration.
530

TABLE 2: Total ceftriaxone thresholds for a MIC of 1 mg/L in case of normal albumin ~~level~~ concentration (35 g/L) and hypoalbuminemia (20g/L).

Model	fT > MIC 100%		fT > 4 x MIC 100%	
	Normal albumin level <u>concentration</u>	Hypoalbuminemia <u>(% decrease)</u>	Normal albumin level <u>concentration</u>	Hypoalbuminemia <u>(% decrease)</u>
Bos	15.7	8.1 <u>(-48%)</u>	43.3	23.0 <u>(-46%)</u>
Dreesen	8.5	5.3 <u>(-38%)</u>	31.0	19.5 <u>(-37%)</u>
Gijssen	16.9	9.1 <u>(-46%)</u>	36.0	23.3 <u>(-35%)</u>
Gregoire	15.1	15.1 <u>(0%)</u>	50.1	50.1 <u>(0%)</u>
Hartman	10.3	6.3 <u>(-39%)</u>	37.8	23.3 <u>(-38%)</u>
Heffernan	16.6	10.0 <u>(-40%)</u>	56.2	33.8 <u>(-40%)</u>
Leegwater	12.3	6.8 <u>(-45%)</u>	40.2	22.7 <u>(-44%)</u>
Standing	4.0	1.8 <u>(-55%)</u>	14.6	6.6 <u>(-55%)</u>
Ulldemolins	3.3	2.0 <u>(-39%)</u>	13.1	7.9 <u>(-40%)</u>

fT: Fraction of time, MIC: Minimal inhibitory concentration.

TABLE 3: MSE, MPE, RMSE, RMSE% and R² of the external validation (N=62)

Model	Mean Signed	Mean Percentage	Root Mean	RMSE%	R-squared (R²)
	Error (MSE)	Error (MPE)	Square Error		
			(RMSE)		
Bos	-18.32	-110.81	30.34	168.67	0.76
Dreesen	-4.63	-56.39	9.47	83.45	0.82
Gijsen	-2.52	-38.11	9.72	77.19	0.75
Gregoire	6.63	31.29	13.90	43.74	0.66
Hartman	0.13	-21.92	7.94	52.42	0.84
Heffernan	3.21	14.55	8.17	37.00	0.86
Leegwater	-8.76	-59.20	16.71	95.97	0.81
Standing	-28.79	-255.08	39.10	298.03	0.71
Ulldemolins	-15.93	-205.95	19.07	246.80	0.73

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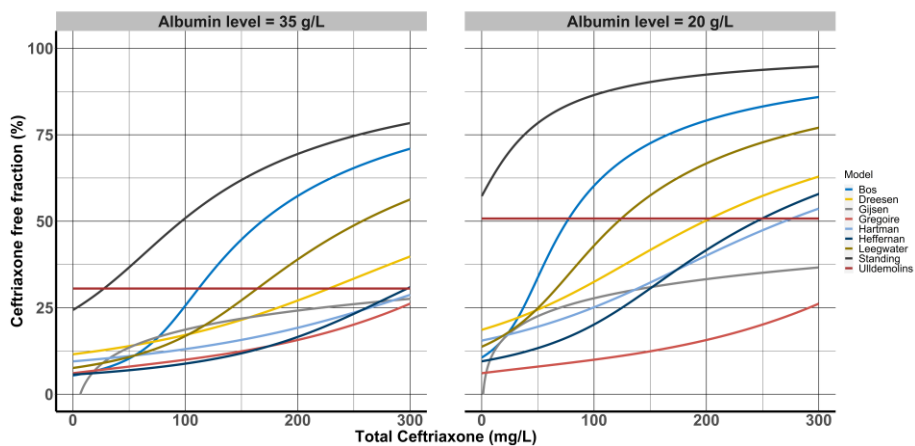
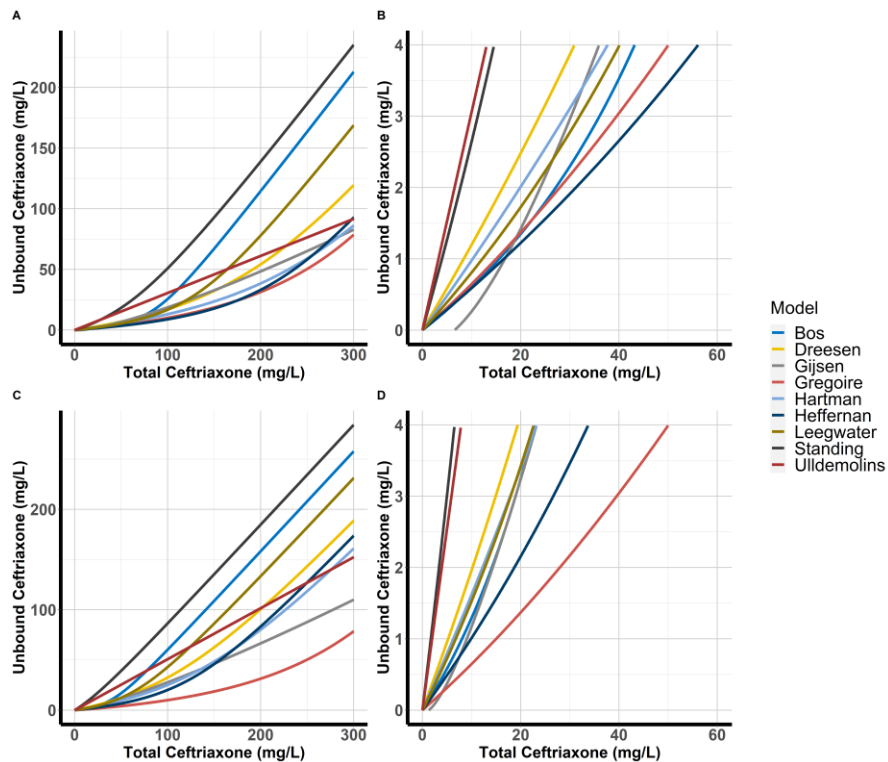
Patient related variables	n	Overall (N=222)	Dosing related variables	n	Overall (N=408)
Age, years	222	74 [63 - 82]	Total Ceftriaxone (mg/L)	408	69 [43 - 105]
Sex	222		Albumin (g/dL)	376	27.0 [23.4 - 30.1]
Man		139 (62.6%)	Bilirubin (μmol /L)	403	6 [5 - 11]
Department	221		Creatinine (μmol/L)	407	121 [81 - 202]
ID		53 (24.0%)	C-reactive protein (mg/L)	404	49 [22 - 97]
Medical ICU		45 (20.4%)	Neutrophils (x10 ⁹ /L)	408	6.3 [4.7 - 9.2]
Cardiology		41 (18.6%)	GGT (U/L)	404	92 [42 - 171]
Medicine		22 (10.0%)	Hemoglobin (g/dL)	408	9.5 [8.6 - 10.9]
Other		18 (8.1%)	Proteides-Protein (g/L)	407	65 [60 - 69]
Geriatrics		17 (7.7%)	ASAT (U/L)	404	30 [22 - 43]
Nephrology		11 (5.0%)	ALAT (U/L)	404	24 [16 - 41]
HGE		8 (3.6%)	Urea (mmol/L)	407	11 [6 - 17]
Neurology		6 (2.7%)	Concomitant Antibiotics	404	
BMI, (kg/m ²)	166	28 [24 - 32]	Betalactams		225 (55.7%)
Diabetes	219	79 (36.1%)	Both		71 (17.6%)
Hypertension	220	134 (60.9%)	Other		52 (12.9%)
Kidney failure	220	54 (24.5%)	None		50 (12.4%)
Hepatic failure	220	15 (6.8%)	Aminoglycosides		6 (1.5%)
Heart failure	222	58 (26.1%)	Dosing regimen	355	
Infection	218		1g once a day		87 (24.5%)
Endocarditis		86 (39.4%)	1g twice daily		83 (23.4%)
Bacteremia		44 (20.2%)	2g twice daily		79 (22.3%)
Other		37 (17.0%)	2g once daily		75 (21.1%)
UTI		24 (11.0%)	Other		31 (8.7%)
ProbabilisticSuspected		14 (6.4%)			
Pneumopathy		13 (6.0%)			
Length of stay (days)	222	27 [17 - 46]			
Bacteria	183				
<i>E. faecalis</i>		97 (53.0%)			
Other		37 (20.2%)			
<i>E. coli</i>		33 (18.0%)			
<i>K. pneumoniae</i>		9 (4.9%)			
<i>S. pneumoniae</i>		7 (3.8%)			

Data are expressed as median [IQR] for continuous variables and as n (%) for categorical variables. ALAT: L-alanine aminotransferase, ASAT: L-aspartate aminotransferase, BMI: Body mass index, *E. coli*: *Escherichia coli*, *E. faecalis*: *Enterococcus faecalis*, GGT: Gamma-Glutamyl Transferase, HGE: Hepato-gastro-enterology, ICU: Intensive care unit, ID: Infectious diseases, *K.pneumoniae*: *Klebsiella pneumoniae*, N: Total number of observations, n: Number of non-missing observations, *S. pneumoniae*: *Streptococcus pneumoniae*, UTI: urinary tract infection.

TABLE 4: Predictors of total ceftriaxone concentration.

Predictors	Unadjusted ^a		Full model ^b (N=325)		Final model ^c (N=325)	
	$\beta \pm \text{s.e}$	P-value ^d	$\beta \pm \text{s.e}$	P-value ^d	$\beta \pm \text{s.e}$	P-value ^d
Age (per year increase)	0.445 \pm 0.208	0.033	0.523 \pm 0.229	0.024	0.618 \pm 0.216	0.005
Weight (per kilogram increase)	0.336 \pm 0.159	0.036	0.296 \pm 0.162	0.068		
Sex (ref=woman)	6.17 \pm 6.49	0.342				
Diabetes (ref=no)	10.65 \pm 6.24	0.089				
Hypertension (ref=no)	3.02 \pm 6.25	0.630				
Intake dose (per gram increase)	24.92 \pm 4.88	<0.001	-10.00 \pm 7.39	0.177		
Daily dose (per gram increase)	16.60 \pm 1.95	0.001	20.38 \pm 3.04	<0.001	17.57 \pm 1.90	<0.001
Albumin (per gram/L increase)	1.86 \pm 0.58	0.002	1.36 \pm 0.60	0.023	1.53 \pm 0.57	0.009
Bilirubin (per $\mu\text{mol/L}$ increase)	-0.017 \pm 0.07	0.818				
Creatininemia (per $\mu\text{mol/L}$ increase)	0.045 \pm 0.020	0.021	0.100 \pm 0.020	<0.001	0.108 \pm 0.020	<0.001
Urea (per mmol/L increase)	0.28 \pm 0.30	0.346				

^a Simple linear mixed effects regression, ^b Multiple linear mixed effects regression for variables with P-value below 0.05, ^c Backward variable selection from the full model, ^d Satterthwaite's degrees of freedom for p-value computation. β : coefficient estimate, ref: reference, s.e: standard error.



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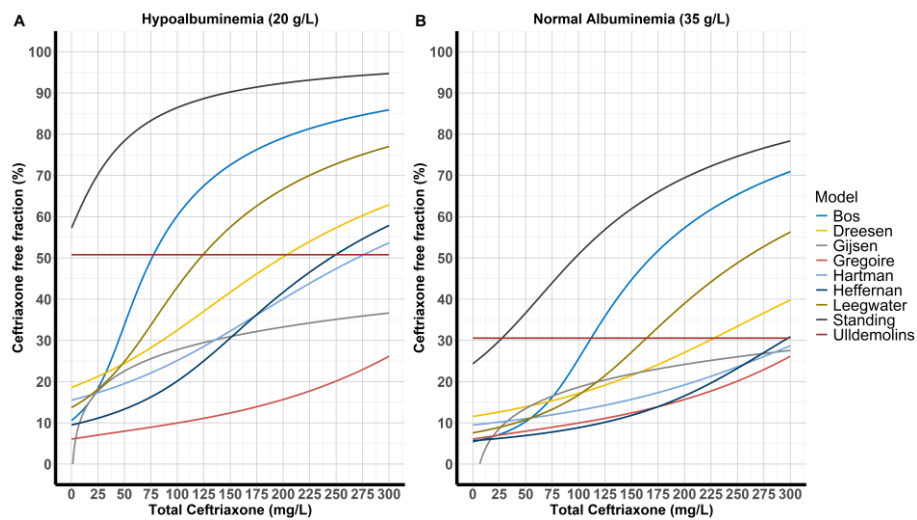


Figure 1: Modeling of ceftriaxone free fraction based on total ceftriaxone in the case of hypoalbuminemia (20 g/L - 1A) and normal albumin level concentration (35 g/L - 1B).

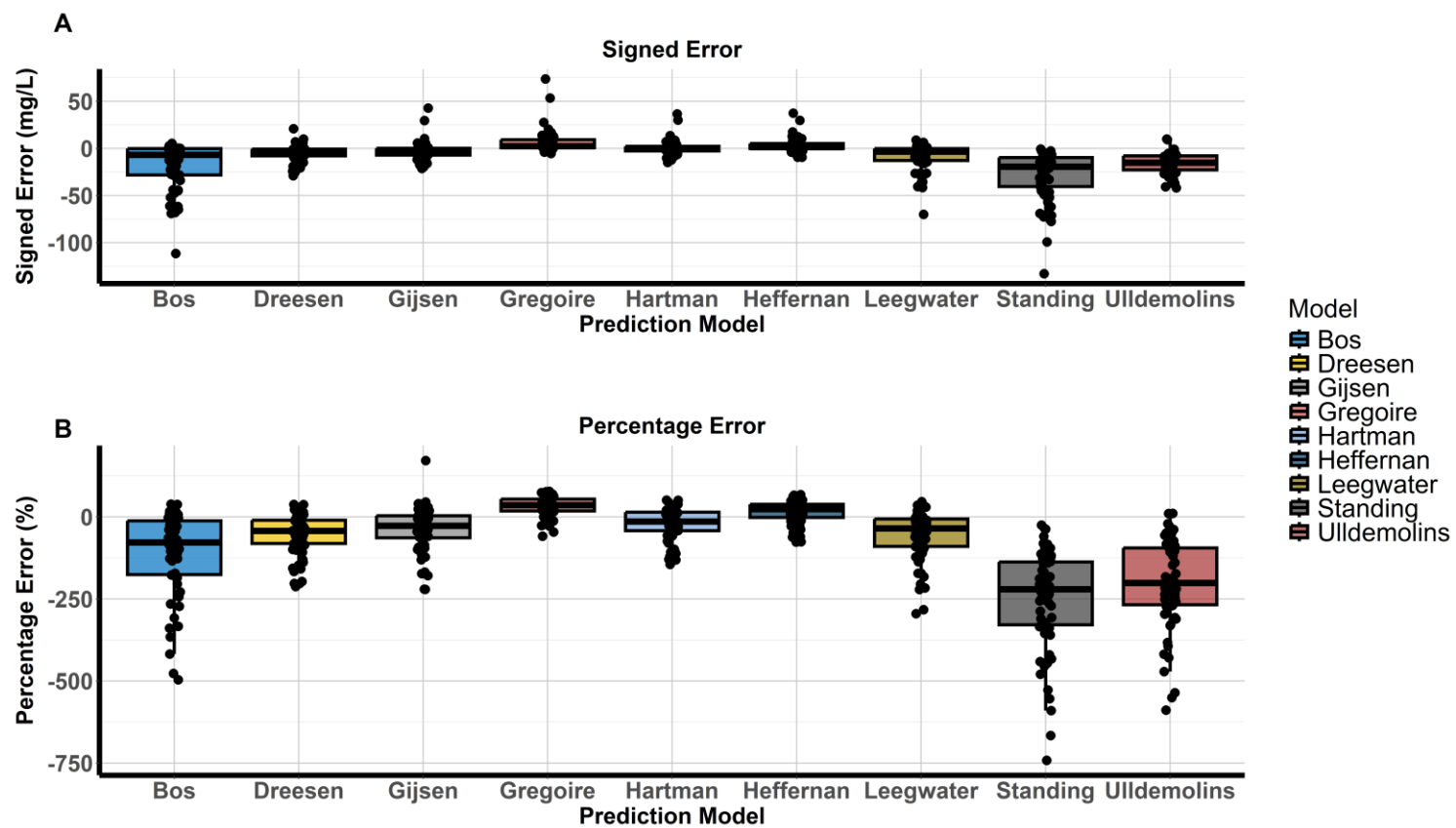


Figure 2: Signed (A) and percentage error (B) for the 9 studied model during external validation (N=62)

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~~and hypoalbuminemia (20 g/L)~~ Modeling of unbound ceftriaxone based on total ceftriaxone according to the retained models from the litterature in the case of normal albumin level (35 g/L, panels A and B) and of hypoalbuminemia (20 g/L, panels C and D).

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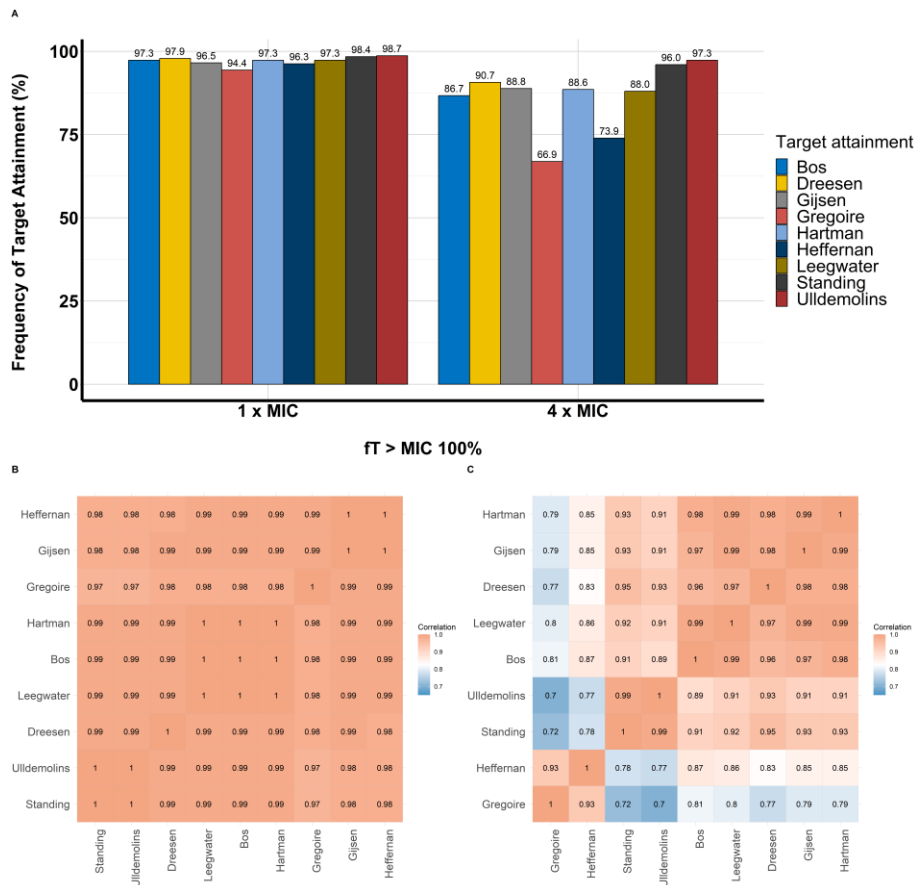


Figure 23: Frequency of target attainment for 1 x MIC and 4 x MIC using ft > MIC 100% according to the different models **(A)** and concordance matrix for 1 x MIC **(B)** and 4 x MIC **(C)**.

Figure 43: Boxplots of total ceftriaxone trough concentration (mg/L) according to dosing regimen. * $p < 0.05$, *** $p < 0.001$. N=324 observations (87, 83, 75 and 79 observations respectively).

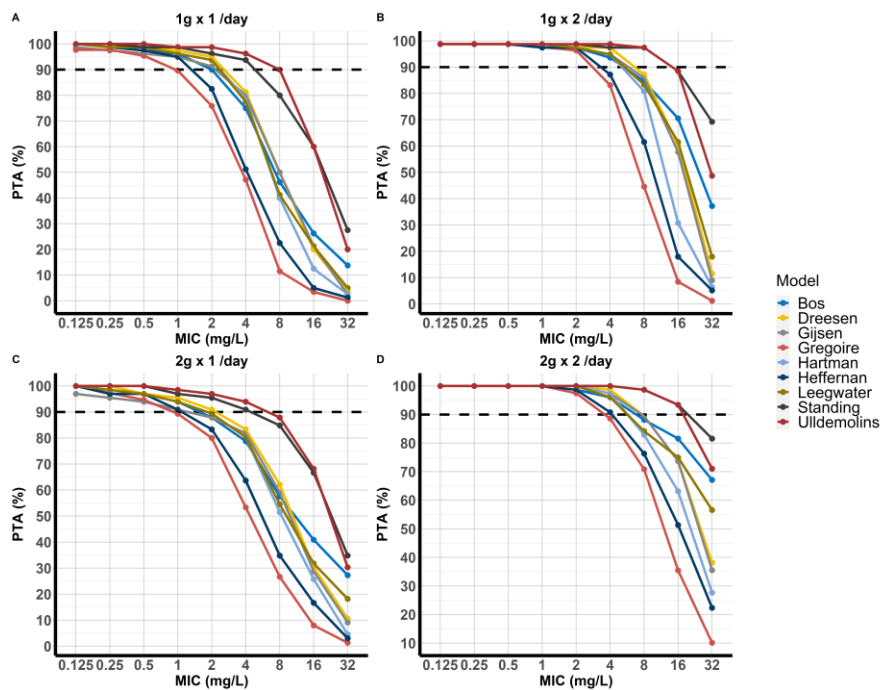


Figure 45: Probability of target attainment curves according to the different models and stratified by dosing regimen. N=87 for (A), N=83 for (B), N=75 for (C) and N=79 for (D). Horizontal dotted lines indicate 90% PTA values. MIC: Minimum inhibitory concentration, PTA: Probability of target attainment (ft > MIC 100%).